



Article

Treatment of Winery Wastewater by Combined Almond Skin Coagulant and Sulfate Radicals: Assessment of HSO_5^- Activators

Nuno Jorge ^{1,2} , Ana R. Teixeira ² , Lisete Fernandes ², Sílvia Afonso ³, Ivo Oliveira ³, Berta Gonçalves ³ , Marco S. Lucas ² and José A. Peres ^{2,*}

¹ Escuela Internacional de Doctorado (EIDO), Campus da Auga, Campus Universitario de Ourense, Universidade de Vigo, As Lagoas, 32004 Ourense, Spain

² Centro de Química de Vila Real (CQVR), Departamento de Química, Universidade de Trás-os-Montes e Alto Douro (UTAD), Quinta de Prados, 5000-801 Vila Real, Portugal

³ Centre for the Research and Technology of Agro-Environmental and Biological Sciences (CITAB), Universidade de Trás-os-Montes e Alto Douro (UTAD), Quinta de Prados, 5000-801 Vila Real, Portugal

* Correspondence: jperes@utad.pt

Abstract: The large production of wine and almonds leads to the generation of sub-products, such as winery wastewater (WW) and almond skin. WW is characterized by its high content of recalcitrant organic matter (biodegradability index < 0.30). Therefore, the aim of this work was to (1) apply the coagulation–flocculation–decantation (CFD) process with an organic coagulant based on almond skin extract (ASE), (2) treat the organic recalcitrant matter through sulfate radical advanced oxidation processes (SR-AOPs) and (3) evaluate the efficiency of combined CFD with UV-A, UV-C and ultrasound (US) reactors. The CFD process was applied with variation in the ASE concentration vs. pH, with results showing a chemical oxygen demand (COD) removal of 61.2% (0.5 g/L ASE, pH = 3.0). After CFD, the germination index (GI) of cucumber and corn seeds was $\geq 80\%$; thus, the sludge can be recycled as fertilizer. The SR-AOP initial conditions were achieved by the application of a Box–Behnken response surface methodology, which described the relationship between three independent variables (peroxymonosulfate (PMS) concentration, cobalt (Co^{2+}) concentration and UV-A radiation intensity). Afterwards, the SR-AOPs were optimized by varying the pH, temperature, catalyst type and reagent addition manner. With the application of CFD as a pre-treatment followed by SR-AOP under optimal conditions (pH = 6.0, [PMS] = 5.88 mM, [Co^{2+}] = 5 mM, T = 343 K, reaction time 240 min), the COD removal increased to 85.9, 82.6 and 80.2%, respectively, for UV-A, UV-C and US reactors. All treated wastewater met the Portuguese legislation for discharge in a municipal sewage network ($\text{COD} \leq 1000 \text{ mg O}_2/\text{L}$). As a final remark, the combination of CFD with SR-AOPs is a sustainable, safe and clean strategy for WW treatment and subproduct valorization.

Keywords: almond skin extract; Box–Behnken; coagulation–flocculation; response surface methodology; sludge valorization; SR-AOPs; wine production



Citation: Jorge, N.; Teixeira, A.R.; Fernandes, L.; Afonso, S.; Oliveira, I.; Gonçalves, B.; Lucas, M.S.; Peres, J.A. Treatment of Winery Wastewater by Combined Almond Skin Coagulant and Sulfate Radicals: Assessment of HSO_5^- Activators. *Int. J. Environ. Res. Public Health* **2023**, *20*, 2486. <https://doi.org/10.3390/ijerph20032486>

Academic Editor: Paul B. Tchounwou

Received: 30 December 2022

Revised: 26 January 2023

Accepted: 27 January 2023

Published: 30 January 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

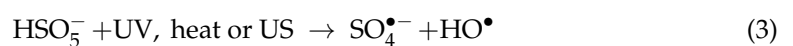
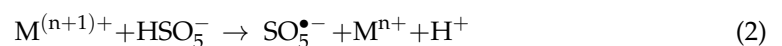
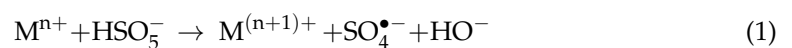
The agro-industries account for 70% of all freshwater withdrawals worldwide, with the wine industry covering three sectors of the economy: agriculture, manufacturing and trade [1]. Due to the consumer demand for quality wines, washing and disinfection operations are necessary, and it is estimated that a winery produces about 1.3 to 1.5 kg of residue per liter of wine produced, 75% of which is winery wastewater (WW). Without a proper treatment process, the WW environmental impact is enormous due to the pollution of the water, degradation of the soil, damage to the vegetation, release of odors into the air, eutrophication of water resources, and consumption of oxygen from the rivers and lakes, leading to the suffocation of aquatic and amphibious life [2,3].

The almond (*Prunus dulcis* (Miller) D.A. Webb) is a member of the genus *Prunus*. The widespread distribution of the almond is related to its low chilling requirement, which makes almonds very suitable for regions with mild winters and dry, hot summers. World almond production has increased from 3.1 million to 4.1 million metric tons from 2011 to 2020 [4]. The almond industrial processes lead to the generation of large amounts of waste related to the removal of shells and skins [5]. Therefore, in this work, it is proposed to valorize the almond skin as a coagulant to treat WW via the CFD process.

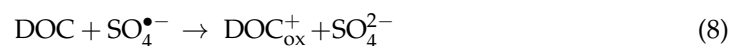
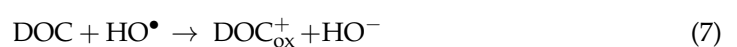
The CFD process is a straightforward and economical process that has the capability to remove organic and inorganic substances and colloidal particles, depending on operational conditions, coagulant type and wastewater characteristics [6]. Although almond skin has never been used before in WW treatment, it has been observed that the extraction of active compounds from plants achieved significant results in wastewater treatment [7,8] and the sludge could be recycled for fertilization [9].

Considering the chemical composition of WW and the presence of recalcitrant organic matter in WW, one possible treatment strategy could be the use of advanced oxidation processes (AOPs). Among AOPs, hydroxyl-based AOPs (HR-AOPs) or sulfate-based AOPs (SR-AOPs) can be applied. In AOPs, hydroxyl radicals (HO^\bullet) are generated by a number of processes. These radicals have an oxidation potential of 2.8 V, they are non-selective, reacting with the pollutants and oxidizing them to CO_2 , H_2O and partially oxidized species [10,11].

In the past years, persulfates, such as peroxymonosulfate (PMS, HSO_5^-) and peroxydisulfate (PDS, $\text{S}_2\text{O}_8^{2-}$), have attracted increasing attention because they are much more stable than hydrogen peroxide [12]. In addition, the sulfate radicals ($\text{SO}_4^{\bullet-}$) produced by the activation of PMS and PDS (1) are more selective than HO^\bullet radicals for the oxidation of compounds with carbon-carbon double bonds and benzene rings, and (2) have a higher oxidation potential than HO^\bullet radicals ($E^0 = 2.5\text{--}3.1\text{ V}$) [13,14]. PMS is the active ingredient of triple potassium salt ($\text{KHSO}_5 \bullet 0.5\text{KHSO}_4 \bullet 0.5\text{K}_2\text{SO}_4$): with an oxidation potential of 1.82 V, it is stable at ambient temperature, easy to handle and the bond energy is estimated to be in the range of 140–213.3 kJ/mol [15]. To generate the $\text{SO}_4^{\bullet-}$ radicals, the PMS can be activated by metal catalysts, heat, UV, visible light, ultrasound (US), alkali and photo-catalytic activation (h^+/e^-) (Equations (1)–(6)) [16].



One of the major problems detected in the treatment of WW is the presence of high values of turbidity, total suspended solids, total polyphenols and dissolved organic carbon (DOC) [17,18], which affect the efficiency of the AOPs due to radical scavenging (Equations (7) and (8)) [19]. In a search performed in the Web of Science and Scopus, results showed only 11 works published with the application of sulfate radicals for the treatment of WW. A background for the activation of PMS in real WW has not been established, particularly for WW with high contents of turbidity and TSS.



Considering this background, the main aim and novelty of this work is the production of a coagulant from almond skin to apply in a CFD process for WW treatment and to recycle the sludge as fertilizer. To decrease more recalcitrant organic matter, the aim is to use a response surface methodology (RSM) Box–Behnken design to define the best SR-AOP to

adopt for WW treatment. The influence of pH, temperature, transition metals, single vs. multiple addition of reagents and different radiation sources (UV-A, UV-C and ultrasound) is studied in the removal of organic carbon.

2. Material and Methods

2.1. Reagents

Potassium peroxymonosulfate (PMS) was acquired from Alfa Aesar (Ward Hill, Massachusetts, US), aluminum sulfate 18-hydrate (10% *w/w*, $\text{Al}_2(\text{SO}_4)_3 \cdot 18\text{H}_2\text{O}$) was obtained from Scharlau (Barcelona, Spain), cobalt(II) sulfate heptahydrate, iron(II) sulfate heptahydrate ($\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$) and manganese(II) sulfate monohydrate ($\text{MnSO}_4 \cdot \text{H}_2\text{O}$) were acquired from Panreac (Barcelona, Spain), Zinc(II) sulfate heptahydrate was acquired from Merck, (Darmstadt, Germany), copper(II) sulfate 2-hydrate ($\text{CuSO}_4 \cdot 2\text{H}_2\text{O}$) was acquired from Honeywell Riedel-de-HaënTM (Charlotte, North Carolina, USA), magnesium sulfate heptahydrate ($\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$), 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical, 6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid (Trolox), catechin, gallic acid, caffeic acid and sodium chloride (NaCl) were purchased from Sigma-Aldrich (St. Louis, MO, USA). For pH adjustment, sodium hydroxide (NaOH) was used from Labkem (Barcelona, Spain) and sulfuric acid (H_2SO_4 , 95%) from Scharlau (Barcelona, Spain). Deionized water was used to prepare the respective solutions.

2.2. Analytical Determinations

Different physical–chemical parameters were determined to characterize the WW, including turbidity, total suspended solids (TSS), volatile suspended solids (VSS), chemical oxygen demand (COD), biological oxygen demand (BOD_5), dissolved organic carbon (DOC) and total polyphenols (TPh). The main WW characteristics are shown in Table 1.

Table 1. Physicochemical characteristics of winery wastewater (mean \pm SD).

Parameter	Values	
	WW1	WW2
pH (Sorensen scale)	3.95 \pm 0.20	3.61 \pm 0.24
Conductivity ($\mu\text{S cm}^{-1}$)	45 \pm 2.3	285 \pm 14.3
Turbidity (NTU)	69 \pm 4	649 \pm 32
Total suspended solids—TSS (mg L^{-1})	200 \pm 10	1405 \pm 70
Dissolved organic carbon—DOC (mg C L^{-1})	138 \pm 7	976 \pm 49
Total nitrogen—TN (mg N L^{-1})	3.4 \pm 0.2	10.7 \pm 0.5
Chemical oxygen demand—COD ($\text{mg O}_2 \text{ L}^{-1}$)	616 \pm 31	4925 \pm 246
Biochemical oxygen demand— BOD_5 ($\text{mg O}_2 \text{ L}^{-1}$)	163 \pm 8	1438 \pm 72
Biodegradability— BOD_5/COD	0.26 \pm 0.01	0.29 \pm 0.02
Total polyphenols—TPh ($\text{mg gallic acid L}^{-1}$)	1.90 \pm 0.1	49.5 \pm 2.5
Absorbance at 254 nm (diluted 1:25)	0.102 \pm 0.005	0.198 \pm 0.010
Absorbance at 254 nm (diluted 1:10)	0.124 \pm 0.006	0.356 \pm 0.018

The COD analysis was carried out in a COD reactor from Macherey-Nagel (Düren, Germany), and a HACH DR 2400 spectrophotometer (Loveland, CO, USA) was used for colorimetric measurements. The BOD_5 was determined according to the 5-day BOD test (Standard Method 5210B) using a respirometric OxiTop[®] IS 12 system (WTW, Yellow Springs, OH, USA). The pH was determined via a 3510 pH meter (Jenway, Cole-Parmer, UK) and conductivity was determined via a portable conductivity meter, VWR C030 (VWR, V. Nova de Gaia, Portugal), in accordance with the methodology of the Standard Methods [20]. The turbidity was determined via a 2100N IS Turbidimeter (Hach, Loveland, CO, USA), and the total suspended solids were measured through spectrophotometry according to Standard Method 2540D using a HACH DR/2400 portable spectrophotometer (Hach, Loveland, CO, USA). The total polyphenols were measured using the Folin–Ciocalteu method, adapted by Singleton and Rossi [21]. UV-vis measurements were performed using a Jasco V-530

UV/VIS spectrophotometer. The total nitrogen (TN) and DOC samples were analyzed via direct injection of the filtered samples into a Shimadzu TOC-L_{CSH} analyzer (Shimadzu, Kyoto, Japan) equipped with an ASI-L autosampler, provided with an NDIR detector, and calibrated with standard solutions of potassium phthalate.

2.3. Preparation of Almond Skin Extract (ASE) and Characterization Methodologies

Almond samples (*Prunus dulcis*) from cultivars commonly produced in Trás-os-Montes, northeastern Portugal, were obtained directly from producers located in this region, transported to the laboratory and the skin was removed from the almond. The skin was ground to a fine powder using a 150 W Princess grinder with 2 knife blades (Deluxe, Netherlands). All ground skins were sieved through a 0.4 mm sieve, and the resulting fraction with particle size less than 0.4 mm was used in the coagulation experiments. The extract was prepared by the addition of 12.5 g of prepared powder to 250 mL of a NaCl 1M solution, and the suspension was stirred for 5 h at ambient temperature (298 K) to extract the coagulation-active compounds. Finally, the suspension was allowed to rest for 5 min to sediment the solid parts, and the extract was collected through decantation [22]. The crude extracts were stored in the refrigerator (278 K) and used the following day to avoid aging phenomena and improve reproducibility. The ASE presented a pH of 5.0 ± 0.25 .

The chemical spectrum of the almond powder (AP) was obtained via Fourier transform infrared spectroscopy (FTIR), and the KBr sample was analyzed using an IRAffinity-1S Fourier transform infrared spectrometer (Shimadzu, Kyoto, Japan) with the infrared spectrum in transmission mode recorded in the $4000\text{--}400\text{ cm}^{-1}$ frequency region. The microstructural characterization of the almond powder was carried out with scanning electron microscopy (FEI QUANTA 400 SEM/ESEM, Fei Quanta, Hillsboro, WA, USA) and the chemical composition was estimated using energy dispersive X-ray spectroscopy (EDS/EDAX, PAN'alytical X'Pert PRO, Davis, CA, USA). The pH of the point of zero charge (pH_{PZC}) was determined according to the method described by Oussalah et al. [23], in which 50 mL of 0.01 M NaCl solutions were adjusted to a pH range of 2–12. Then 200 mg of almond skin powder was added to each NaCl solution. The suspensions were stirred for 48 h at room temperature, and the final pH of the solutions (pH_f) was determined. The pH_{PZC} was obtained from the plot of ($\text{pH}_f - \text{pH}_i$) versus pH_i .

For the preparation of the methanolic extract, 40 mg of AP was weighed and mixed by vortexing with 1 mL of 70% methanol. The mixtures were heated at $70\text{ }^\circ\text{C}$ for 30 min and centrifuged at 13,000 rpm at $1\text{ }^\circ\text{C}$ for 15 min (Eppendorf Centrifuge 5804 R, Hamburg, Germany). The supernatants were collected and filtered with Spartan filters (0.2 mm) into HPLC amber vials. The methodology of Singleton and Rossi (1965) [21] was used for the quantification of total phenolics in a 96-well microplate. Total phenolics results were expressed as mg gallic acid equivalent (GAE)/g f.w. The total flavonoid content was determined in a 96-well microplate using the colorimetric method described in Dewanto et al. [24]. The total flavonoid content was expressed as mg catechin equivalent (CE)/g f.w. The total ortho-diphenol content was determined in a 96-well microplate, in accordance with the methodology of Soufi et al. [25], and the results were expressed as mg of caffeic acid equivalents/g of dry weight. The 2,2-diphenyl-1-picrylhydrazyl (DPPH) antioxidant activity assay was performed through spectrophotometry, as described by Siddhraj and Becker [26], in a 96-well microplate. The DPPH was expressed as μg trolox equivalent/g f.w.

2.4. Coagulation–Flocculation–Decantation Experiments

The coagulation–decantation–flocculation process was performed in a Jar test device (ISCO JF-4, Louisville, KY, USA), with four mechanical agitators powered by a regulated speed engine. The mixing of the ASE with WW samples was performed under a fast mix of 150 rpm/3 min and a slow mix of 20 rpm/20 min, at ambient temperature (298 K). Four different ASE concentrations (0.1, 0.5, 1.0 and 2.0 g/L) were tested against four different pH

levels (3.0, 6.0, 9.0 and 11.0), and, after a sedimentation time of 4 h, samples were retrieved for analysis.

2.5. Box–Behnken Experimental Design

A Box–Behnken design was employed to assess the effect of different parameters on the UV-A LED SR-AOP treatment of WW1, such as the concentration of PMS (mM, X_1), the concentration of Co^{2+} (mM, X_2) and radiation intensity (W m^{-2} , X_3) under fixed conditions (COD = 616 mg O_2/L , temperature = 323 K, pH = 6.0, time = 120 min). For this study, 15 experiments were performed in triplicate. The levels considered for the Box–Behnken design are listed in Table 2, with three replicates at the center of the design (SR2, SR10 and SR11). Experiments were randomized to maximize the efforts of unexplained variability in the observed response due to external factors.

Table 2. Symbols and coded factor levels for the considered variables.

Independent Variables	Code	Levels		
		−1	0	1
[PMS] (mM)	X_1	0	5	10
$[\text{Co}^{2+}]$ (mM)	X_2	0	2.5	5.0
Radiation	X_3	0	16.35	32.70

2.6. SR-AOP Set-Up

The experiments were performed on WW2 in a beaker with 500 mL capacity under constant agitation (350 rpm). The temperature of the reaction was controlled with a heating plate (Nahita blue model 692/1, Navarra, Spain) equipped with temperature probe. The following variables were studied: (1) pH (3.0–11.0), (2) temperature (298–363 K), (3) catalyst type (Zn^{2+} , Al^{3+} , Co^{2+} , Cu^{2+} , Fe^{2+} , Mg^{2+} and Mn^{2+}) and (4) reagent addition (single vs. multiple addition). Finally, three radiation sources were used:

- (1) A UV-A LED system composed of 12 indium gallium nitride (InGaN) LED lamps (Roithner APG2C1-365E LEDs) with a $\lambda_{\text{max}} = 365$ nm. Each UV-A LED had a nominal consumption of 1.4 W when the current was 350 mA, with an optical power of 135 mW and an opening angle of 120° , making any shadow zone impossible. The radiation was emitted in continuous mode for all 12 UV-A LEDs and was controlled using a power MOSFET in six different current settings, resulting in irradiance levels from 5.2 to 32.7 W m^{-2} measured at a 5 cm distance with a UVA Light Meter (Linshang model LS126A);
- (2) A Heraeus TNN 15/32 lamp (14.5 cm in length and 2.5 cm in diameter) mounted in the axial position inside the reactor, with 15 W power. The spectral output of the low-pressure mercury vapor lamp emitted mainly (85–90%) at 253.7 nm and about 7–10% at 184.9 nm;
- (3) An ultrasonic system (Vibracell Ultrasonic processor VCX 500, Sonics & Materials Inc., Danbury, CT, USA) with 500 W power, equipped with a titanium alloy probe (136 mm diameter, 13 mm) and a temperature control probe. For temperature control, a water jacket was installed.

To determine the removal percentage of the parameters, Equation (9) was applied, as follows [27,28]:

$$X_i(\%) = \frac{C_i - C_f}{C_i} \times 100 \quad (9)$$

where C_i and C_f are the initial concentrations and 100 is the conversion factor.

2.7. Phytotoxicity Tests

Phytotoxicity tests were performed via the germination of *Zea mays* (corn) and *Cucumis sativa* (cucumber) seeds (standard species recommended by the US EPA, the US FDA and the OECD [29]). Seeds were immersed in a 10% sodium hypochloride solution for 10 min

to ensure surface sterility, then they were soaked in pure water. One piece of filter paper (Whatman filter paper 9 cm, Maidstone, UK) was put into each 100 mm × 15 mm Petri dish, and 5 mL of test medium was added [30]. Seeds were then transferred onto the filter paper, with 10 seeds per dish and a 1 cm or larger distance between each seed. Petri dishes were covered and sealed with tape and placed in a controlled atmosphere with a constant temperature (25 °C), maintained during the course of the experiment with a WTM TS 608-G/2-i (Weilheim, Germany). After 7 days of darkness and 7 days of light, the germination index was determined by Equation (10), in accordance with Varnero et al. [31] and Tiquia and Tam [32], as follows:

$$GI(\%) = \frac{\bar{N}_{SG,T}}{\bar{N}_{SG,B}} \times \frac{\bar{L}_{R,T}}{\bar{L}_{R,B}} \times 100 \quad (10)$$

where GI is the germination index, $\bar{N}_{SG,T}$ is the arithmetic mean of the number of germinated seeds in each extract (wastewater), $\bar{N}_{SG,B}$ is the arithmetic mean of the number of germinated seeds in standard solution (distilled water), $\bar{L}_{R,T}$ is the mean root length of each extract (wastewater), and $\bar{L}_{R,B}$ is the mean root length in control (distilled water). If $GI \leq 50\%$, then there was a high concentration of phytotoxic substances; if $80\% < GI < 50\%$, then there was a moderate presence of phytotoxic substances; and if $GI \geq 80\%$, then there were no phytotoxic substances (or they existed in very small dosages).

2.8. Statistical Analysis

The coefficients corresponding to the model equation were obtained using Minitab Statistical Software 2018 (State College, PA, USA). All experiments were performed in triplicate, and a one-way ANOVA was carried out to determine any significant differences ($p < 0.05$) using the Tukey test. Results are presented as mean ± standard deviation (SD).

3. Results and Discussion

3.1. Characterization of Almond Skin Powder

Before the production of the almond skin extract (ASE), it was necessary to characterize the powder obtained from the almond skin. Figure 1a shows the chemical spectrum of the AS by FTIR. The band at 3460 cm^{-1} can be attributed to O-H and N-H stretching vibrations of hydroxyl groups and amide A of polypeptides and amino acids, respectively. The O-H stretching is also associated with the water fraction and with the polyphenol content of the samples [33]. The band at 3190 cm^{-1} is related to the stretching vibration of C=CH cis olefinic groups of unsaturated fatty acids [34]. The bands at 2924 and 2895 cm^{-1} correspond to the symmetric and asymmetric stretching vibrations of aliphatic CH_2 functional groups, respectively, which are linked with the saturated fatty acid fraction [35]. The band at 1608 cm^{-1} corresponds with the stretching vibration of cis C=C of unsaturated acyl groups [36]. The band at 1508 cm^{-1} corresponds to the bending vibration of the N-H functional group mainly observed in Amide I and Amide II of protein compounds [33]. The bands at 1450 and 1255 cm^{-1} are associated with the presence of CH bending vibrations in CH_2 and CH_3 , respectively [37]. Finally, a peak at 1016 cm^{-1} is related to the stretching vibration of C-O functional groups characteristic of the carbohydrate fraction [36].

The SEM images (Figure 1b) show that the almond skin powder presents dark spaces that correspond to empty spaces, similar to findings in other works involving plant-based materials [38]. These porous materials allow the adsorption of the NaCl solution and the desorption of material from the powder to the exterior, a necessary characteristic to produce the ASE.

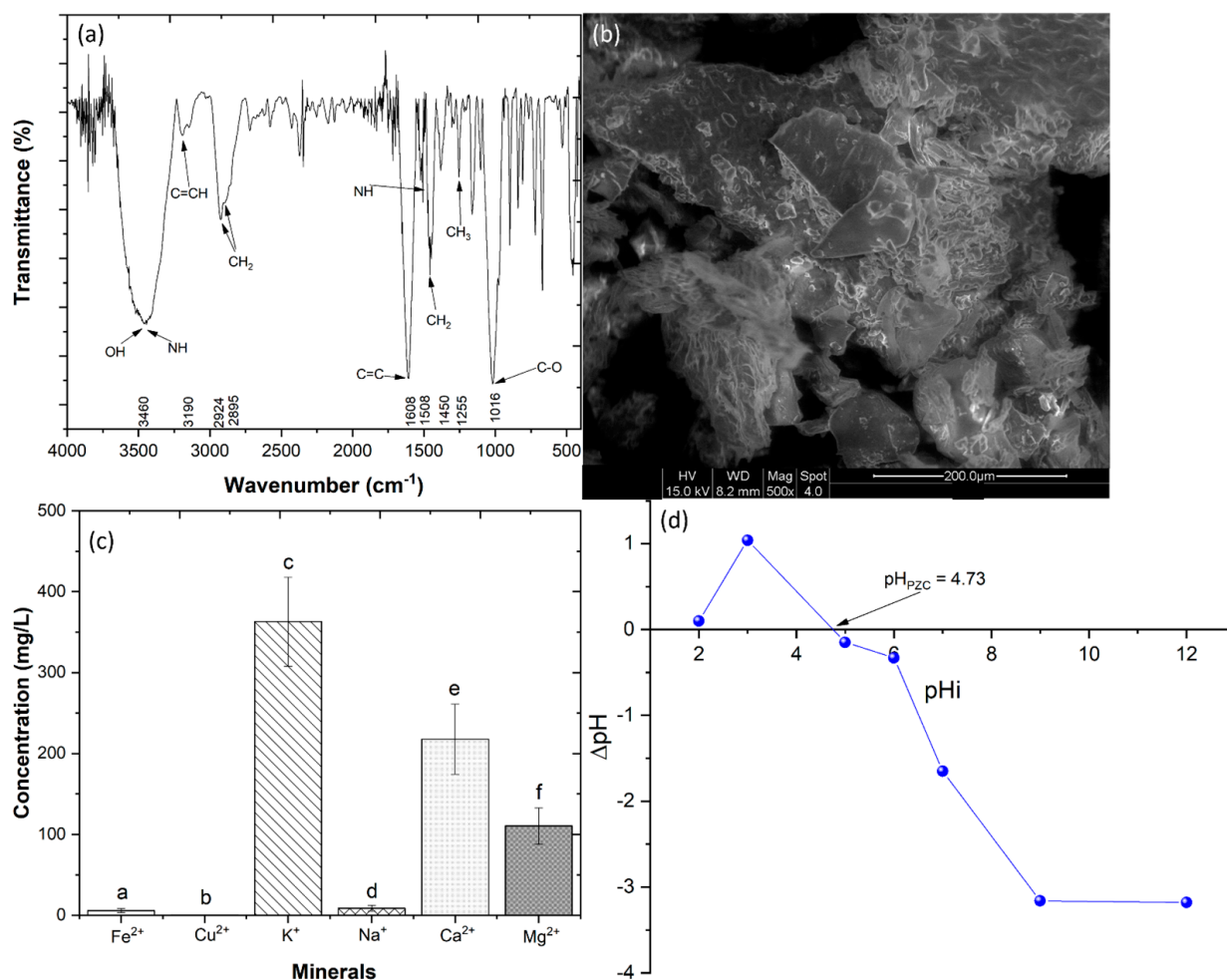


Figure 1. Almond skin powder characterization: (a) FTIR spectrum, (b) SEM image (500 \times), (c) mineral analysis and (d) pH_{PZC} of almond powder. The pH_i is the initial pH value of the solution, and ΔpH is the difference between final pH values (after contact with the AP) and pH_i values. Means in bars with different letters represent significant differences ($p < 0.05$) between different minerals.

Figure 1c shows major concentrations of potassium (K^+), calcium (Ca^{2+}) and magnesium (Mg^{2+}), which are linked to the stress resistance of the plants [39]. The chemical analysis presented in Table S1 shows that AS revealed a total phenolic concentration of 6.51 ± 0.62 mg GAE/g, a flavonoid concentration of 4.30 ± 0.16 mg CE/g, an O-diphenol concentration of 0.65 ± 0.05 mg CAE/g and a DPPH concentration of 8.93 ± 0.34 μ g Trolox/g. These results are in agreement with the work of Oliveira et al. [40], who observed similar concentrations in almonds. These results showed that the skin of the almond has antioxidant capacities.

Figure 1d shows the effect of pH on almond skin powder. The pH_{PZC} value determined for the ASP was 4.73. This means that at a $pH < 4.73$, the surface of the almond skin powder is positively charged, and the biosorption of anionic particles will be favored [41]. However, at a $pH > 4.73$, the surface of the ASP is negatively charged, increasing the attraction of cations [23].

3.2. Coagulation–Flocculation–Decantation Experiments

In this section, almond skin extract (ASE) was applied for the treatment of WW2 via the CFD process. Table 1 shows that WW2 has a complex composition with a high content of organic matter, polyphenols and turbidity. In order to optimize the CFD process and to understand the behavior of ASE, the pH of the wastewater and concentration of ASE were varied. The results showed that the ASE efficiency was affected by the pH of the wastewater. From Figure 2a, results show that a pH of 3.0 achieved a turbidity removal

within the range of 94.8 to 96.8% and a TSS removal within the range of 96.9 to 98.1%. This turbidity and TSS removal decreased as the pH increased. On the other hand, the variation in ASE concentration showed little difference regarding turbidity and TSS removal. These results are not in agreement with the work of Hussain Haydar [42], who observed that the natural coagulant *Opuntia stricta* achieved high TSS removal at pH 10.3. Figure 2b shows the influence of ASE on TPh removal. The importance of TPh removal lies in the reduction in the color, which is caused mainly by the polyphenols [27]. The results showed that pH was the main factor that influenced TPh removal, with the highest removal observed at pH 3.0. These results agree with the pH_{PHZ} values, which showed that a $pH < 4.73$ increased the attraction of the proteins present in the ASE and the negatively charged polyphenols, such as tannins. As the pH increased above the pH_{PZC} , the electrostatic repulsion between the proteins in the ASE and the polyphenols increased, explaining the low removal [43]. The COD and DOC removals were studied (Figure 2c), with results showing high removals with 0.5 g/L ASE at pH 3.0 (61.2 and 56.8%, respectively). These results showed that ASE was able to reduce the organic carbon in suspension as well as the dissolved organic carbon. Considering the low levels of organic carbon removed in other works [44], the use of ASE proved to be competitive. Regarding the pH of the WW2 (3.61), the cost of pH changing is avoided, thus this becomes a cheaper process for wastewater treatment.

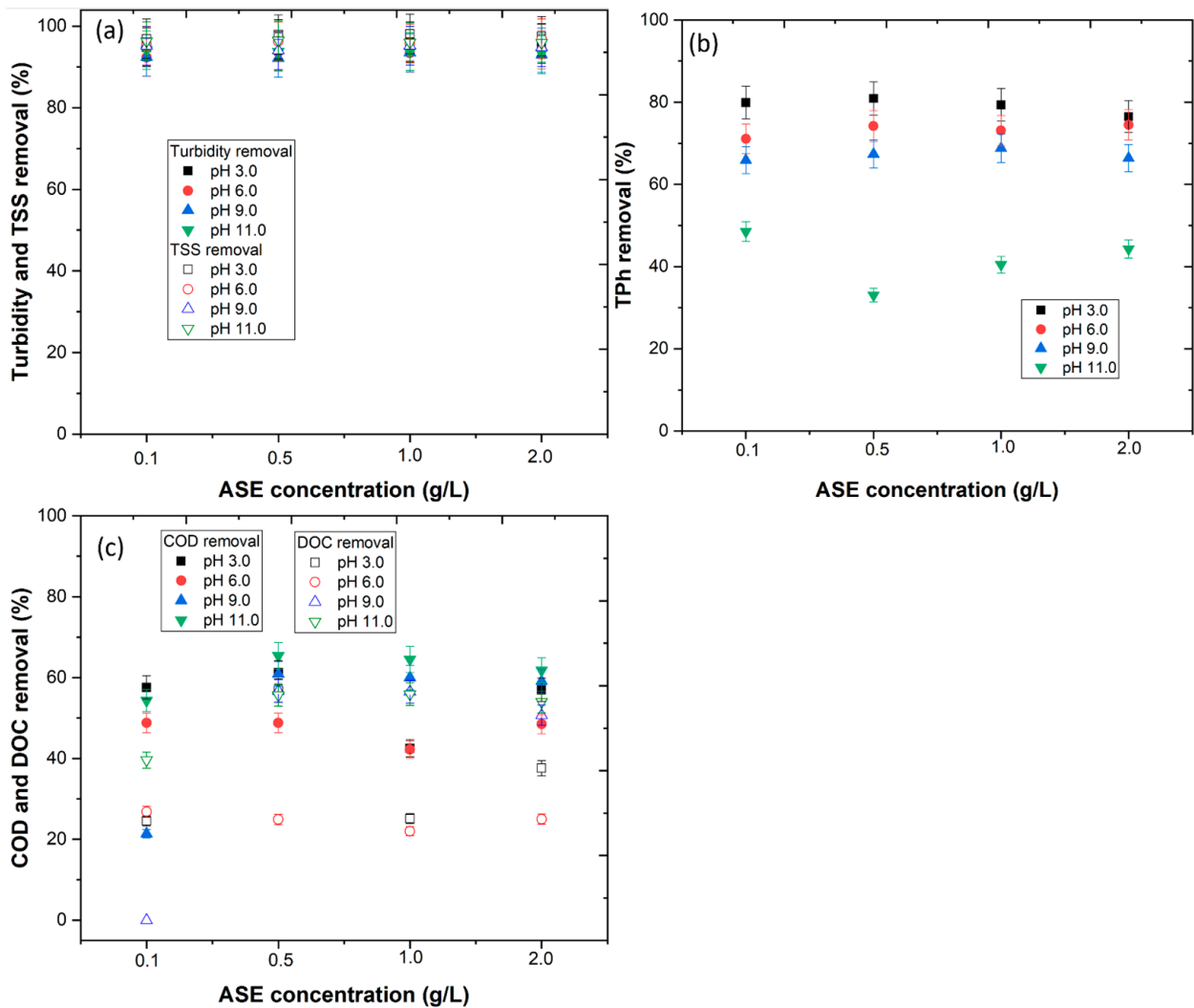


Figure 2. Effect of ASE concentration (g/L) vs. pH in (a) turbidity and TSS removal, (b) TPh removal, and (c) COD and DOC removal from winery wastewater. Operational conditions: fast mix (rpm/min) = 150/3, slow mix (rpm/min) = 20/20, sedimentation time of 4 h.

In order to decrease the environmental impact of sludge generation, this work tested the possibility of recycling the sludge as fertilizer. The recovered sludge was applied as a substrate for cucumber and corn seed germination. The results showed that the application of 1.0 and 2.0 g/L ASE at pH 3.0 and 6.0 could have a toxic effect on seed germination (Figure 3a,b). At the operational conditions selected (0.5 g/L ASE at pH 3.0), the results showed a GI \geq 80% with good radicular growth; thus, the sludge can be recycled as fertilizer. These results are in agreement with the work of Jorge et al. [9], who observed that the sludge of WW could be recycled as fertilizer if organic coagulants were applied.

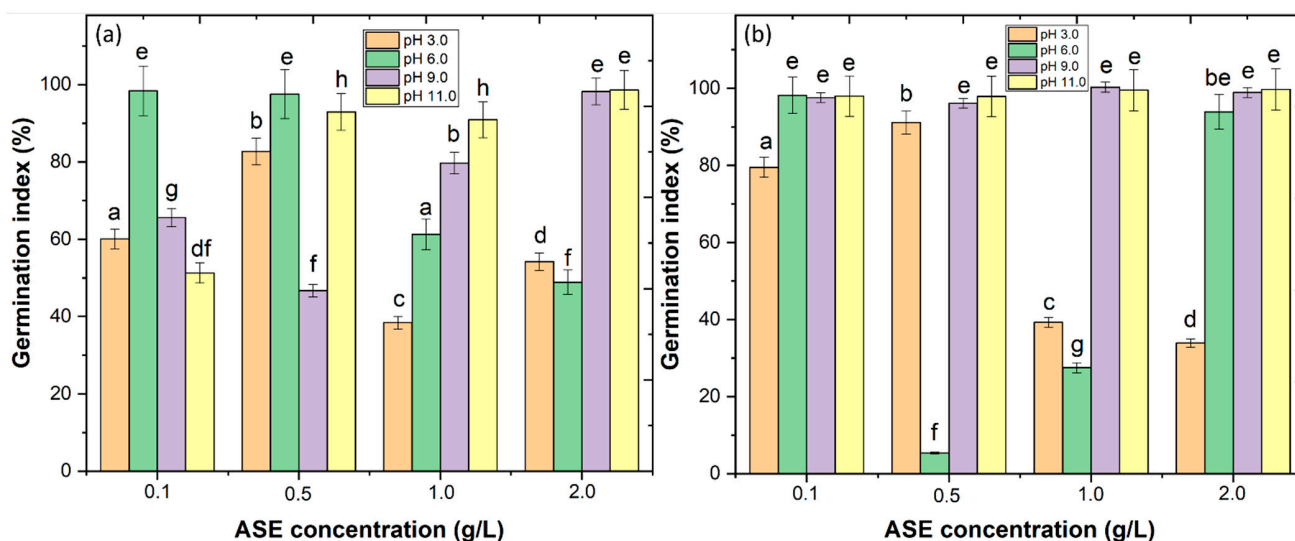


Figure 3. Effect of ASE concentration vs. pH in (a) germination index of cucumber and (b) corn seeds. Means in bars with different letters represent significant differences ($p < 0.05$) within each ASE concentration by comparing the pH.

3.3. SR-AOP Optimization through Response Surface Methodology

In the previous section, ASE showed high efficiency in the removal of turbidity and TSS; however, the coagulant showed limited capacity in the removal of COD. Therefore, in this section, the removal of COD by SR-AOPs was studied. Considering the limited information available regarding the oxidation of organic matter from WW by PMS and Co^{2+} , it was necessary to create a model that can help reach the concentration of PMS and Co^{2+} and, at the same time, allow the study of different SR-AOPs. In this section, a Box–Behnken design was applied to achieve the initial conditions of PMS and Co^{2+} concentrations. WW1 was used for this model due to its low COD, thus reducing the reagent requirement. The assessment of the COD removal was performed throughout a range of SR-AOP conditions ($n = 15$) based on distinct combinations of PMS concentration (X_1 : 0–10 mM), Co^{2+} concentration (X_2 : 0–5 mM) and radiation (X_3 : 0–32.70 W m^{-2}). The results of the 15 runs are shown in Table 3. The range of PMS and Co^{2+} concentrations was consistent with those previously assayed in the treatment of agro-industrial wastewaters by other authors [45,46]. The LED lamps were selected as radiation sources due to (1) the LED efficiency, (2) the LED price and (3) the maximum emission wavelength [47,48].

Before optimizing the treatment process, it is necessary to understand which SR-AOP is best fitted to reduce the COD present in the WW. The RSM model allowed the study of different variables to understand how they affect the generation of sulfate radicals ($\text{SO}_4^{\bullet-}$). In Figure 4a, the removal efficiencies of different oxidation systems, which include (1) PMS, (2) UV-A, (3) Co^{2+} , (4) Co^{2+} + UV-A, (5) PMS + UV-A, (6) PMS + Co^{2+} and (7) PMS + Co^{2+} + UV-A, can be observed. The reactions involving Co^{2+} , PMS, Co^{2+} + UV-A and UV-A reached a COD removal of 7.7, 10.1, 18.3 and 23.1%, respectively. The low removal observed with the application of Co^{2+} and Co^{2+} + UV-A was expected since cobalt alone is not able to generate radicals [49]. The UV-A radiation alone was revealed to be inefficient, which is

in agreement with the work of Jorge et al. [18]. The application of PMS alone achieved a reduced COD removal, a result that could be related to the low PMS oxidation potential (1.82 V) [16]. The PMS + UV-A reached higher COD removal than PMS and UV-A alone (33.7%), which is in agreement with the work of Huang et al. [50]. These results could be due to the conversion of the PMS by the UV-A radiation into $SO_4^{\bullet-}$ radicals. The highest COD removals were observed with the application of PMS + Co^{2+} and PMS + Co^{2+} + UV-A (44.3 and 62.9%, respectively). The results showed that the activation of PMS by cobalt was higher than the activation by UV-A radiation. It was also observed that a synergy effect occurs with the application of PMS + Co^{2+} + UV-A. This synergy could be linked to the regeneration of Co^{3+} to Co^{2+} by UV-A radiation, which further enhances the conversion of PMS and the generation of $SO_4^{\bullet-}$ radicals.

Table 3. Box–Behnken design: effect of operational variables on COD removal yield ([PMS] = 0–10 mM; [Co^{2+}] = 0–5 mM; radiation = 0–32.70 $W\ m^{-2}$). Operational conditions: COD = 616 $mg\ O_2/L$, temperature = 323 K, pH = 6.0, reaction time = 120 min.

Experiments	Coded Level			Response Values	
	[PMS] (mM)	[Co^{2+}] (mM)	Radiation	COD Removal (%) Observed	COD Removal (%) Predicted
SR1	5	5.0	32.70	54	49.1
SR2	5	2.5	16.35	46.4	46.4
SR3	0	5.0	16.35	16.8	14.4
SR4	5	5.0	0.00	44.3	42.5
SR5	10	5.0	16.35	58.7	67.8
SR6	5	0.0	32.70	33.7	35.5
SR7	0	0.0	16.35	23.1	14.0
SR8	10	2.5	32.70	62.9	58.7
SR9	10	2.5	0.00	52.5	45.2
SR10	5	2.5	16.35	46.4	46.4
SR11	5	2.5	16.35	46.4	46.4
SR12	5	0.0	0.00	10.1	14.9
SR13	10	0.0	16.35	24.6	27.0
SR14	0	2.5	32.70	18.3	25.6
SR15	0	2.5	0.00	7.7	11.9

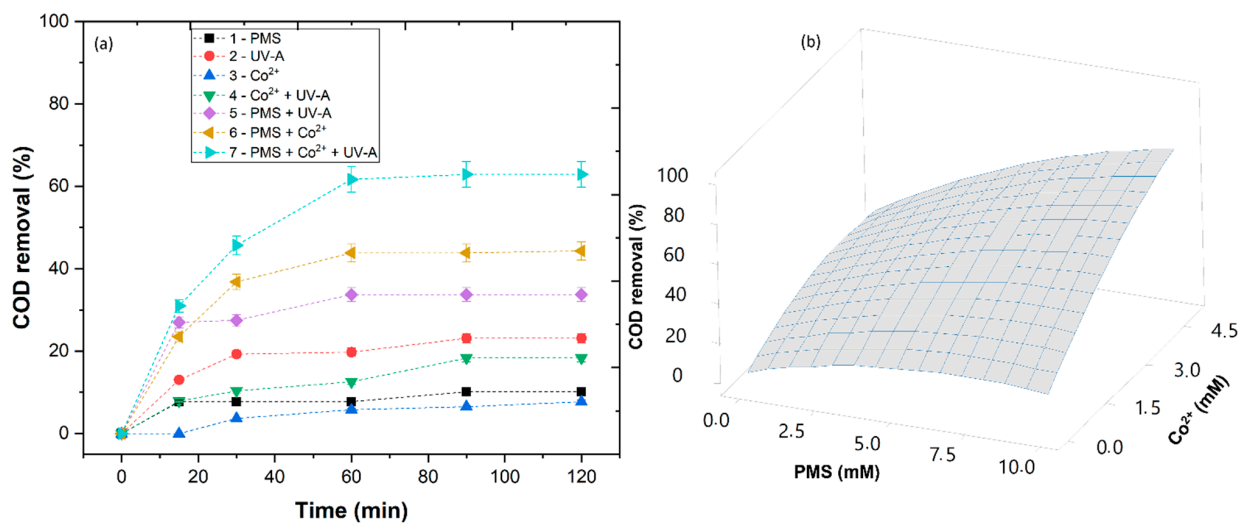
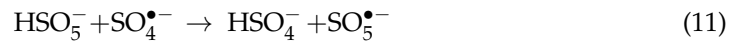
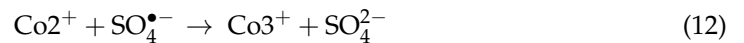


Figure 4. SR-AOP optimization: (a) experiments with different oxidation systems in the reduction in COD from WW1, (b) response surface methodology. Operational conditions: COD = 616 $mg\ O_2/L$, pH = 6.0, [PMS] = 5 mM, [Co^{2+}] = 2.5 mM, temperature = 323 K, radiation UV-A 32.7 $W\ m^{-2}$, reaction time = 120 min.

The RSM model also contributes to the optimization of PMS and Co^{2+} concentrations. The increase in PMS concentration up to 5 mM showed an increase in COD removal due to the higher generation of $\text{SO}_4^{\bullet-}$ radicals. As the PMS concentration increased to 10 mM, results showed a reduction in COD removal (Figure 4b) due to the excess of PMS in the solution, which competed with the organic matter for the $\text{SO}_4^{\bullet-}$ radicals (Equation (11)) [51]. These results are in agreement with the work of Govindan et al. [52], who observed that high concentrations of PMS led to the consumption of $\text{SO}_4^{\bullet-}$ radicals, decreasing the degradation of the contaminant pentachlorophenol.



The Co^{2+} concentration was observed to have a significant effect on COD removal. Comparing the results obtained in SR1 and SR8, when the PMS: Co^{2+} ratio decreased from 1:0.5 to 1:0.25, the COD removal increased. The excess of Co^{2+} present in SR1 was observed to compete for $\text{SO}_4^{\bullet-}$ radicals, decreasing the efficiency of the oxidation process (Equation (12)) [53]. These results are in agreement with the work of Rodríguez-Chueca et al. [45], who observed that an excess of cobalt led to the consumption of $\text{SO}_4^{\bullet-}$ radicals in the treatment of WW.



The quadratic model developed in this work permitted the adjustment of the theoretical values of COD removal to observed values with a low deviation (Table 3), suggesting a successful application of the RSM methodology. From the RSM model, the polynomial equation (Equation (13)) was obtained, and the regression coefficient (R^2) for this method was 0.931, which means that the model matches the COD removal adequately.

$$\text{COD} = 0.50 + 4.47X_1 + 7.64X_2 + 1.018X_3 - 0.316X_1X_1 - 1.234X_2X_2 - 0.0118X_3X_3 + 0.808X_1X_2 - 0.0006X_1X_3 - 0.085X_2X_3 \tag{13}$$

The regression coefficients of the intercept, linear, quadratic and interaction terms of the model were determined with the application of the least squares method. The effect of linear, quadratic or interaction coefficients on the response was studied via analysis of variance (ANOVA) (Table 4). The degree of significance of each factor is represented by its p -value, which indicates that the regression models for COD removal were statistically relevant with a level of significance of $p = 0.027 < 0.05$ (Table S2). The model did not display a significant lack of fit ($p > 0.05$), with $R^2 = 92.04\%$ (Table S3); thus, it can be considered a well-fitting model for the described variables. These statistical analyses revealed that the most important variables for the COD removal from the WW were the PMS (X_1) and Co^{2+} (X_2) (Table 4). Throughout this statistical model, the most relevant conditions were obtained: [PMS] = 5.88 mM, [Co^{2+}] = 5 mM, radiation intensity = 32.7 W m^{-2} .

Table 4. F and p -values for selected responses for each obtained coefficient.

Variable	X_1	X_2	X_3	X_1X_1	X_1X_2	X_1X_3	X_2X_2	X_2X_3	X_3X_3
F -value	29.59	11.36	4.95	3.08	5.48	0.00	2.95	0.65	0.50
p -value	*	*	n.s.	n.s.	n.s.	n.s.	n.s.	n.s.	n.s.

X_1 : PMS (mM); X_2 : Co^{2+} (mM); X_3 : radiation. n.s.: non-significant. Significant at * $p < 0.05$.

3.4. SR-AOPs Applied to a High Load WW

The operational conditions obtained with the RSM were applied in the treatment of WW2, with a COD of 4925 mg O_2/L . The advantage of creating a model with a WW with a lower COD content is the application of a lower concentration of reagents, decreasing both costs and scavenging reactions. The role of different parameters such as pH, temperature, transition metals, the dosing procedure of reagents and radiation sources were investigated to establish the optimal conditions for the treatment of WW2 with SR-AOPs.

Initially, the pH was varied from 3.0 to 11.0 over 240 min (Figure 5a). The results showed a COD removal of 47.3, 57.0, 59.4 and 56.4%, respectively, for pH levels 3.0, 6.0, 9.0

and 11.0. The low COD removal observed at pH 3.0 was consistent with the non-productive reactions under highly acidic pH levels (Equation (6)), resulting in reduced production of $\text{SO}_4^{\bullet-}$ radicals, which is in agreement with the work of Huling et al. [54], who observed no considerable differences between pH 3.0 and 6.0 in the degradation of methyl tert-butyl ether (MTBE) by persulfate oxidation. The results showed a favorable range from 6.0 to 9.0, which was in agreement with Yi et al. [55]. Above pH 9.0, a decrease in COD removal was observed. These results were due to the fact that the alkaline solution inhibited the dissolution of Co^{2+} while promoting the complexation and deposition of Co^{2+} . In this way, pH 11.0 inhibited the dissolution of Co^{2+} , slowing down the activation of PMS and reducing the catalytic activity [13,56]. Considering the initial pH of the WW (4.81) and considering that the difference in COD removal between pH 6.0 and 9.0 was not significant, pH 6.0 is a better choice, since a lower content of NaOH is required to be spent, which is in agreement with the work of Rodríguez-Chueca et al. [46], who observed that pH 6.5 could achieve significant COD removal from WW.

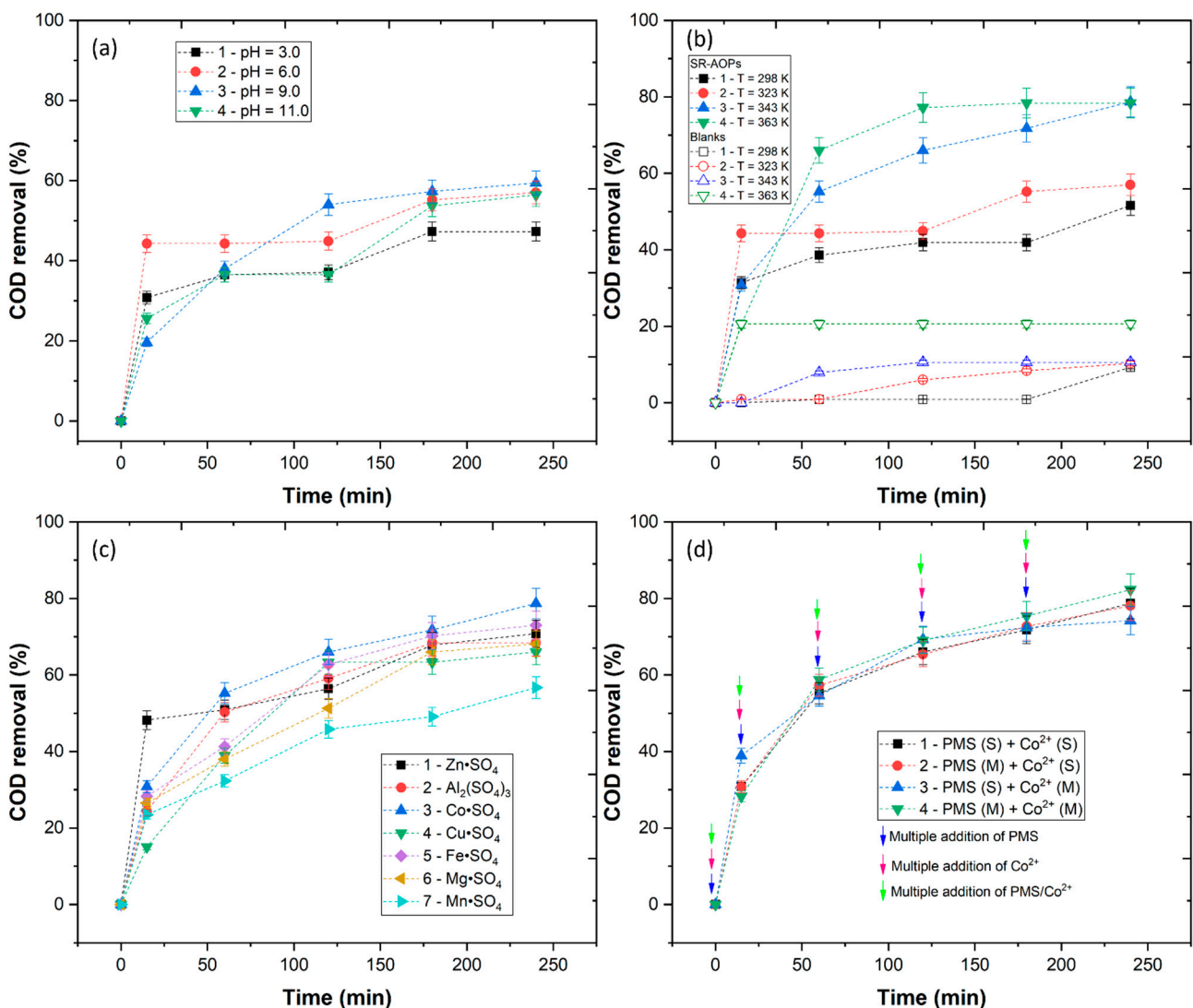


Figure 5. COD removal in the optimization of (a) pH ([PMS] = 5.88 mM, [Co²⁺] = 5 mM, radiation UV-A, T = 323 K), (b) temperature (SR-AOPs: pH = 6.0, [PMS] = 5.88 mM, [Co²⁺] = 5 mM, radiation UV-A; blanks: pH = 6.0, radiation UV-A), (c) type of metal catalyst (pH = 6.0, [PMS] = 5.88 mM, [Mⁿ⁺] = 5 mM, radiation UV-A, T = 343 K) and (d) dosing of PMS and Co²⁺. S—single addition, M—multiple addition (pH = 6.0, [PMS] = 5.88 mM, [Co²⁺] = 5 mM, radiation UV-A, T = 343 K).

The temperature of the wastewater was observed by other authors to have an influence on the conversion of persulfate and the generation of $\text{SO}_4^{\bullet-}$ radicals [57,58]. Figure 5b shows the COD removal at different temperatures (298–363 K) using the optimal pH of 6.0. Before the application of the SR-AOPs, it was necessary to understand the effect of temperature without PMS and Co^{2+} , so blank experiments were performed. The results in Figure 5b show a maximum COD removal with the application of 363 K (20.6%). Thus, the organic matter appears to be stable at high temperatures. With the application of SR-AOPs, the results showed no significant differences between 298 and 323 K (51.6 and 57.0%, respectively); however, the COD increased significantly with 343 K (78.7%). In the work of Chen et al. [57], it was observed that persulfate was more easily converted in $\text{SO}_4^{\bullet-}$ radicals at 343 K, which agrees with the results obtained in this work. These results are also in agreement with the works of Rodríguez-Chueca et al. [59] and Jorge et al. [28], who observed that high temperatures are beneficial in activating PMS. Increasing the temperature to 363 K the COD removal is reduced (78.4%), which could suggest that temperatures above 343 K can inactivate the $\text{SO}_4^{\bullet-}$ radicals, decreasing the efficiency of the reaction. In previous studies, the reduction in organic matter via thermally activated PMS was observed to follow a pseudo first-order kinetic rate [46].

The effect of the nature of the metal catalyst was studied. In this section, seven different sulfate catalysts were tested (ZnSO_4 , $\text{Al}_2(\text{SO}_4)_3$, CoSO_4 , CuSO_4 , FeSO_4 , MgSO_4 and MnSO_4) to evaluate their effect on the activation of PMS (Figure 5c). The results showed the highest COD removal with catalysts CoSO_4 , FeSO_4 and ZnSO_4 , with 78.7, 73.0 and 70.8%, respectively. Cobalt has been reported as one of the most effective catalysts for the activation of PMS, promoting a high generation of $\text{SO}_4^{\bullet-}$ and HO^{\bullet} radicals [55,60]. The regeneration of Co^{3+} to Co^{2+} in the PMS/Co (Equation (2)) is thermodynamically feasible (0.82 V), fast and the process proceeds cyclically many times until PMS is consumed [13]. The application of FeSO_4 was shown to be as efficient as CoSO_4 . In the work of Latif et al. [61], iron was observed to be effective in the production of $\text{SO}_4^{\bullet-}$ radicals in the degradation of the organic contaminant bisphenol A (BPA). The ZnSO_4 catalyst was observed in this work to have a significant effect on the conversion of PMS in $\text{SO}_4^{\bullet-}$ and HO^{\bullet} radicals, which is in agreement with the work of Fang et al. [62], who observed that zinc could efficiently react with PMS to generate $\text{SO}_4^{\bullet-}$ and HO^{\bullet} radicals that degrade BPA. $\text{Al}_2(\text{SO}_4)_3$ was used as a catalyst in Al^{3+} /PMS reaction. The results showed a higher COD removal (68.4%) with the application of catalysts MgSO_4 , CuSO_4 and MnSO_4 (68.1, 66.0 and 56.7%, respectively). Aluminum sulfate is highly used in coagulation–flocculation–decantation processes, posing a low risk to the environment with a higher discharge limit (10 mg Al/L) than cobalt (3 mg Co/L) and iron (2 mg Fe/L) [63]. Thus, $\text{Al}_2(\text{SO}_4)_3$ can be considered an alternative to cobalt.

To complement this optimization, the dosing procedure of the reagents was investigated, in which the reagents were added in a single step at the beginning of the reaction (S) or five times in a multiple addition (M). Four different ways were then selected to understand the effect of single vs. multiple addition in COD removal: (1) PMS (S) + Co^{2+} (S), (2) PMS (M) + Co^{2+} (S), (3) PMS (S) + Co^{2+} (M) and (4) PMS (M) + Co^{2+} (M) (Figure 5d). The results showed that the application of PMS (S) + Co^{2+} (M) achieved the lowest COD removal (74.2%). These results could be due to a higher concentration of PMS present in solution and insufficient Co^{2+} present to generate the $\text{SO}_4^{\bullet-}$. In addition, the excess of PMS could have a scavenger effect (Equation (11)), decreasing the efficiency of the reaction, which could explain the decrease in COD removal. The applications of PMS (S) + Co^{2+} (S) and PMS (M) + Co^{2+} (S) showed similar results, with 78.7 and 78.1% COD removal, respectively. These results showed that the catalyst addition had a significant effect on the conversion of PMS and the generation of $\text{SO}_4^{\bullet-}$ radicals. With the application of PMS (M) + Co^{2+} (M) the COD removal reached 82.3%, the highest of all addition methods. In contrast with PMS (S) + Co^{2+} (S), the multiple dosing of reagents kept the concentrations of PMS and Co^{2+} low in the reactor, suppressing the rate of scavenging reactions and, as a consequence, a more gradual supply of $\text{SO}_4^{\bullet-}$ radicals were generated, resulting in a more significant COD removal. The influence of multiple dosages of reagents was reported by

other authors. For example, in the work of Sun et al. [64], it was observed that the addition of PMS in multiple dosages decreased the self-decomposition of the oxidant caused by the high concentrations of the addition in single mode.

In this work, the COD removal results were well-fitted into a pseudo first-order kinetic model ($\ln [\text{COD}]_t = -kt + \ln [\text{COD}]_0$), and the corresponding rate constants were 2.16×10^{-3} , 2.49×10^{-3} , 5.82×10^{-3} and $6.40 \times 10^{-3} \text{ min}^{-1}$ for, respectively, 298, 323, 343 and 363 K (Figure 6a). To have a better understanding of the influence of temperature on PMS activation, the Arrhenius equation ($\ln k = \ln A - E_a/RT$) was used, and a chart was plotted, fitting $\ln k$ vs. $1/T$ (Figure 6b), where A is the frequency factor, E_a is the activation energy, R is the universal gas constant and T is the absolute temperature [58]. The results showed a good fit of the data ($y = -1901.73X + 0.27647$, $R^2 = 0.967$). The average E_a ($16.07 \pm 0.58 \text{ kJ mol}^{-1}$) was observed to be lower than the activation of PMS reported by Rodriguez-Narvaez et al. [65] (29.9 kJ mol^{-1}) in the degradation of a contaminant by PMS/ Co^{2+} at a temperature of 343 K. In further experiments, a temperature of 343 K was selected, considering that above this temperature no significant COD removal was observed.

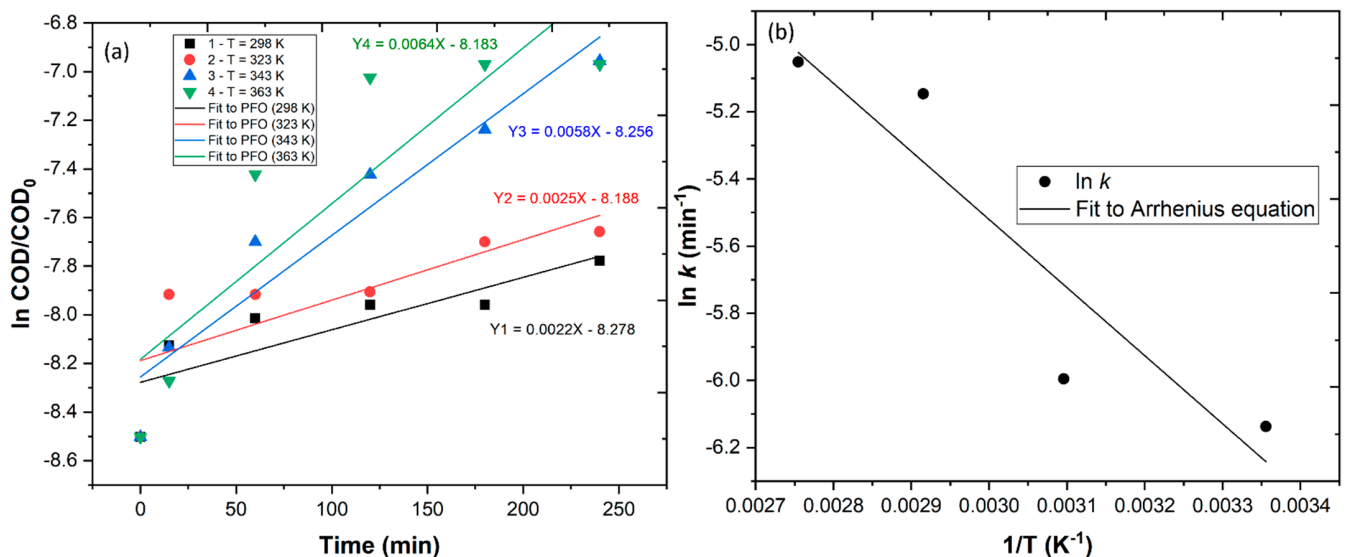


Figure 6. (a) Pseudo first-order of COD removal at different temperatures and (b) plot of $\ln k$ vs. $1/T$ for E_a estimation using the Arrhenius equation.

The PMS/ Co^{2+} /UV system was shown to be effective in the removal of organic matter. In this section, the optimized conditions (pH = 6.0, [PMS] = 5.88 mM, [Co^{2+}] = 5 mM, $T = 343 \text{ K}$, time = 240 min) were applied to the treatment of WW in three different reactors. A UV-A LED photosystem, a low-pressure UV-C mercury lamp and an ultrasound reactor were used. Figure 7a shows the COD and DOC removal after the application of the optimal PMS/ Co^{2+} /UV system in the three reactors. Results showed a COD removal of 82.3, 76.0 and 52.2%, respectively, and a DOC removal of 75.8, 64.1 and 38.8%, respectively, for UV-A, UV-C and US. The high mineralization observed with the UV-C reactor can be explained by the photolysis of the PMS, which generates one mole of $\text{SO}_4^{\bullet-}$ radicals and one mole of HO^{\bullet} radicals per mole of PMS (Equation (3)). In the work of Wang and Chu [66], it was observed that the system PMS/ Fe^{2+} /UV (254 nm) achieved the highest degradation of 2,4,5-trichlorophenoxyacetic acid, which was attributed to the decomposition of PMS by the UV-C radiation. However, in this work, it was observed that the application of a UV-A reactor achieved higher mineralization than the UV-C reactor. The activation of PMS with UV-A radiation was shown to be effective in the removal of micropollutants from the wastewater [67] and in the removal of organic carbon from WW [46], which is in agreement with the results obtained in this work. Although the photolysis of PMS is negligible with UV-A radiation [68], the application of Co^{2+} appears to be the contributing factor that

increased the efficiency of the reaction and higher mineralization of the organic carbon. The activation of PMS by ultrasound was also studied. The US can be used to generate $\text{SO}_4^{\bullet-}$ and HO^\bullet radicals by the decomposition of PMS (Equation (3)) [69]. The use of ultrasound radiation showed a high removal of organic carbon, although the results were shown to be lower than those of UV-A and UV-C radiation. These results are in agreement with the work of Lu et al. [70], who observed a 71.4% atrazine degradation in the PMS/US system.

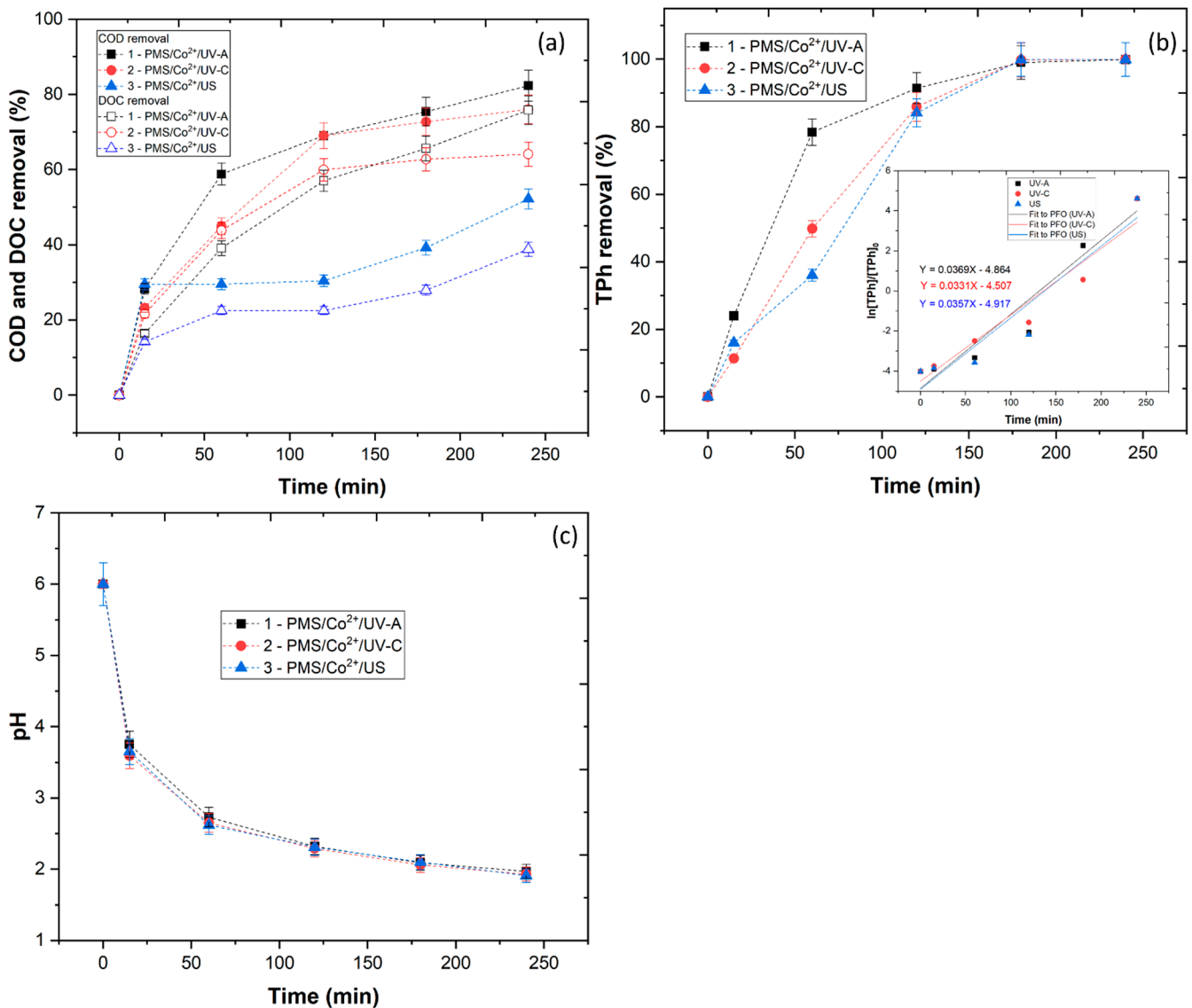


Figure 7. Evaluation of the PMS/Co²⁺/UV system with three radiation reactors (UV-A, UV-C and US) in (a) COD and DOC removal, (b) TPh removal and (c) pH. Operational conditions: $\text{pH}_i = 6.0$, $[\text{PMS}] = 5.88 \text{ mM}$, $[\text{Co}^{2+}] = 5 \text{ mM}$, $T = 343 \text{ K}$, time = 240 min.

Among the organic compounds that are present in the WW, polyphenols are linked to its toxicity, since aromatic compounds are difficult to degrade by microorganisms. Polyphenols have a significant contribution to the dark color of the WW, which limits the penetration of UV radiation and the catalyst regeneration [71]. In this section, the TPh removal was evaluated as a function of the three reactors, with results showing a near complete removal after the 240 min of reaction (Figure 7b). These results are linked with the generation of $\text{SO}_4^{\bullet-}$ and HO^\bullet radicals by the three systems, which are extremely powerful, non-selective and capable of oxidizing most organic compounds. The $\text{SO}_4^{\bullet-}$ and HO^\bullet radicals are capable of attacking the phenol rings of phenolic compounds, yielding benzoic

and cinnamic acids, flavonoids and anthocyanins, and then the rings of these compounds break up to give organic acids and finally CO₂ [28,72].

One of the main factors that affected the efficiency of organic carbon removal was the pH of the wastewater. Therefore, the pH of the reaction was monitored during the 240 min. The results in Figure 7c show that all three systems had a large drop in pH within the first 15 min, which could be linked to the production of SO₄^{•−} and HO[•] radicals. This fact was previously reported by Esteves et al. [73], who observed a high decrease in the pH in the treatment of high-strength olive mill wastewater using a Fenton-like oxidation process. The follow-up of the pH is important because the type and rate of the produced radicals are among the important effects of pH changes. At pH 6.0, the SO₄^{•−} radicals were predominant [74,75]. As the reaction unfolded, a significant reduction in the pH took place, caused by the hydrolysis of the SO₄^{•−} and the cobalt ions. Therefore, it could be perceived that a higher concentration of SO₄^{•−} radicals leads to a more significant pH decline, which is in agreement with the work of Li et al. [76].

3.5. CFD + PMS/Co²⁺/Radiation

In the previous section the legal limits for wastewater discharge in a sewage network were not achieved for the UV-C and US reactors (COD < 1000 mg O₂/L); therefore, in this section, the CFD + PMS/Co²⁺/radiation combined treatment was studied. The CFD process was performed as a pre-treatment followed by oxidation with SR-AOPs. Figure 8a shows the evolution of COD and DOC removal with the combined system. Results showed a COD removal of 85.9, 82.6 and 80.2% and a DOC removal of 83.3, 79.1 and 74.5%, respectively, for CFD/PMS/Co²⁺/UV-A, CFD/PMS/Co²⁺/UV-C and CFD/PMS/Co²⁺/US. Two factors were observed to increase the reactions' efficiency: (1) the removal of turbidity, TSS and TPh by the ASE, which clarified the wastewater, allowing better penetration of the radiation, and (2) the removal of the organic matter in suspension, which acted as a scavenger of HO[•] and SO₄^{•−} radicals. These results are in agreement with the work of Jaafarzadeh et al. [77], who observed that the application of CFD as a pre-treatment increased the efficiency of electro-activated HSO₅[−] to treat pulp and paper wastewater. In comparison to other works, such as Amor et al. [78], results showed that the application of the CFD/SR-AOP system achieved higher organic matter removal from WW with lower reagent consumption.

In order to evaluate the mineralization capacity of the different processes, an efficient parameter was applied, as a partial oxidation efficiency (μ_{partox}), which can be determined by Equations (14) and (15) [79], as follows:

$$\text{COD}_{\text{partox}} = \left(\frac{\text{COD}_0}{\text{DOC}_0} - \frac{\text{COD}}{\text{DOC}} \right) \times \text{DOC} \quad (14)$$

$$\mu_{\text{partox}} = \frac{\text{COD}_{\text{partox}}}{\text{COD}_0 - \text{COD}_t} \quad (15)$$

where μ_{partox} is the partial oxidation efficiency that is between 0 and 1. The μ_{partox} is 0 when only total oxidation occurs, and the value of 1 represents the ideal condition in which only partial oxidation occurs. In fact, total oxidation occurs in μ_{partox} < 0.5, while partial oxidation is dominant in μ_{partox} > 0.5. Figure 8b shows that with the application of UV-A and UV-C reactors, the reduction in organic carbon predominantly occurred via total oxidation reactions, becoming more pronounced at prolonged times, similar to the work of Papastefanakis et al. [80]. With application of the PMS/Co²⁺/US and CFD/PMS/Co²⁺/US systems, partial oxidation reactions occurred at 15 and 30 min, respectively, following total oxidation reactions until 240 min of reaction. These results can be explained by the high reduction in the organic matter by the HO[•] and SO₄^{•−} radicals, which increased the mineralization.

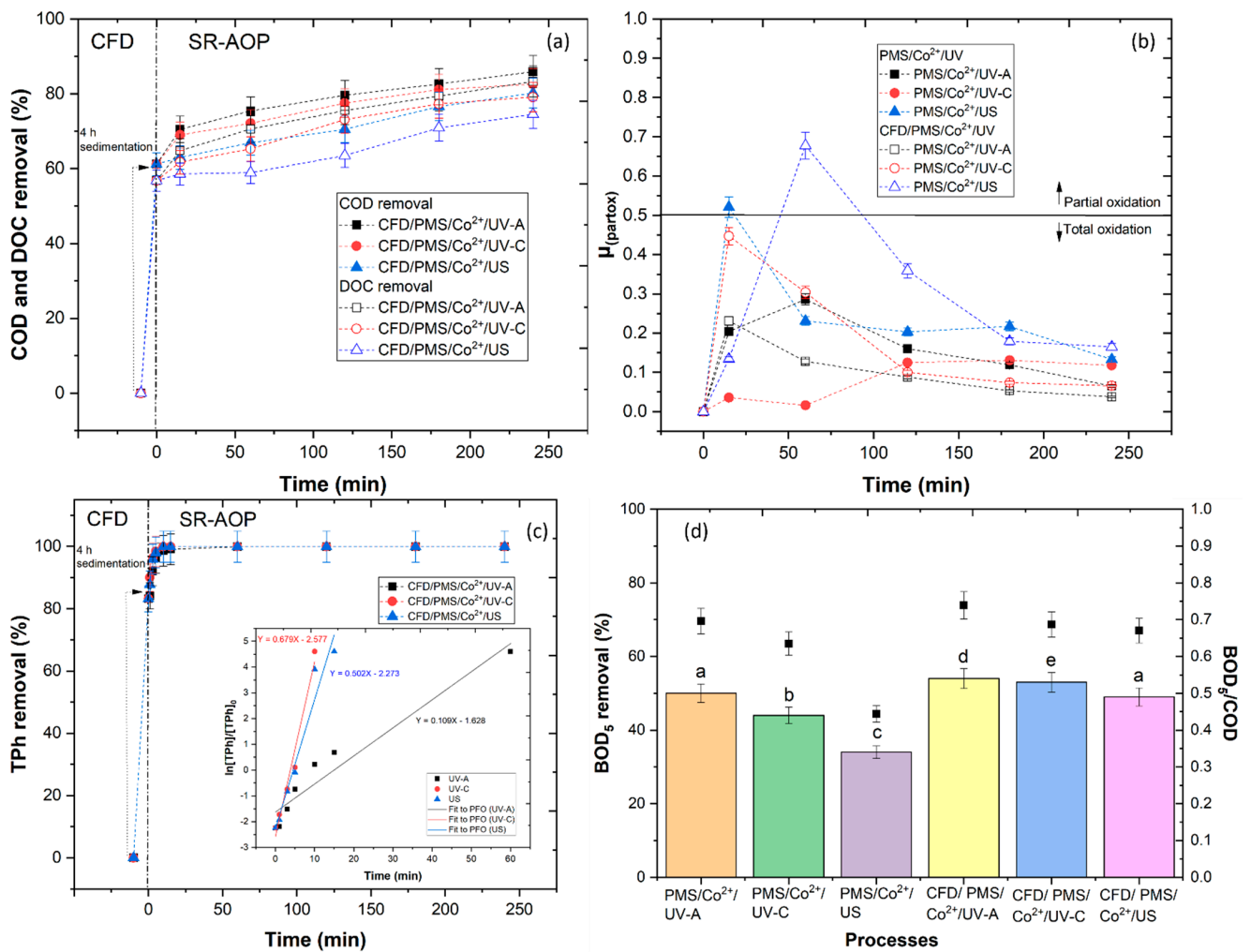


Figure 8. Effect of CFD/PMS/Co²⁺/radiation in (a) COD and DOC removal, (b) $\mu_{(partox)}$, (c) TPh removal and (d) BOD₅ removal and BOD₅/COD. Means in bars with different letters represent significant differences ($p < 0.05$) within BOD₅/COD by comparing treatment processes. CFD operational conditions: pH = 3.0, [ASE] = 0.5 g/L, fast mix (rpm/min) = 150/3, slow mix (rpm/min) = 20/20, sedimentation time 4 h. SR-AOP operational conditions: pH = 6.0, [PMS] = 5.88 mM, [Co²⁺] = 5 mM, radiation UV-A 32.7 W m⁻², UV-C 15 W, US 500 W, T = 343 K, reaction time = 240 min.

In Figure 8c, the effect of the combined treatment in the removal of TPh can be observed. The ASE was responsible for the removal of 83.2% of the TPh content from the WW, decreasing the dark color of the WW and allowing a better penetration of the radiation. With the application of the UV-A, UV-C and US reactors, the TPh removal rate increased significantly. These results were in agreement with the work of Jorge et al. [18], who observed that application of the CFD process boosted the TPh removal by the SR-AOPs.

Figure 8d shows the impact of the different treatment processes in the BOD₅ removal and, consequently, in the biodegradability. The applications of the PMS/Co²⁺/UV-A, PMS/Co²⁺/UV-C and PMS/Co²⁺/US achieved a BOD₅ removal of 69.6, 63.5 and 44.4%, respectively. With the application of the CFD process, the BOD₅ removal was increased to 73.9, 68.7 and 67.0%, respectively. The biodegradability was also evaluated, with results showing an increase from 0.31 (CFD process) to 0.54, 0.53 and 0.49, respectively. The combination of processes, was able to remove part of the recalcitrant matter, increasing the biodegradable fraction, and allowing a subsequent biological treatment. These results were in agreement with the works of Amor et al. [81] and Rodríguez-Chueca et al. [82], who observed that the combined photo-Fenton and CFD processes increased the biodegradability of crystallized-fruit wastewater.

4. Conclusions

Wine and almond production are two major Portuguese agro-industries with enormous weight in Portugal's economy. The WW generated from wine production is of environmental concern due to the high content of organic matter and polyphenols. From almond production, the skin, often neglected by the food industry, is used in this work to produce an almond skin extract (ASE). The results show that ASE achieves the highest results regarding turbidity, TSS, TPh and COD removal with the application of 0.5 g/L ASE at pH 3.0. It is concluded that the pH has a considerable effect on the ASE efficiency, considering the isoelectric point (4.73). The sludge generated by the treatment with ASE can be recycled as fertilizer, allowing its valorization. The response surface methodology (RSM) associated with a Box–Behnken design was revealed to be one of the most appropriate methods for the optimization of the basic conditions (PMS and Co^{2+} concentration and radiation) for COD removal from WW. The efficiency of the SR-AOP is concluded to be dependent on factors, such as pH, temperature, type of transition metal and manner of addition. Under the optimal conditions, pH = 6.0, [PMS] = 5.88 mM, [Co^{2+}] = 5.0 mM, T = 343 K, reaction time = 240 min, a COD removal was achieved of 82.3, 76.0 and 52.2%, respectively, for UV-A, UV-C and US reactors. The combination of CFD with SR-AOPs has a synergic effect, in which the ASE removes 61.2% of COD and the combined ASE/reactors remove 85.9, 82.6 and 80.2%, respectively, with UV-A, UV-C and US. Moreover, the combination of processes allows all reactors to achieve the Portuguese legal value of COD ($\leq 1000 \text{ mg O}_2/\text{L}$) for wastewater to be discharged as municipal wastewater in a wastewater treatment plant (WWTP).

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ijerph20032486/s1>, Table S1: Concentration of total phenolic, flavonoids, O-diphenol and DPPH present in almond skin extract; Table S2: Analysis of variance; Table S3: Model summary.

Author Contributions: Conceptualization, N.J., A.R.T., L.F., S.A. and I.O.; methodology, N.J., S.A. and I.O.; software, N.J.; validation, N.J., B.G., M.S.L. and J.A.P.; formal analysis, N.J.; investigation, N.J.; resources, N.J. and L.F.; data curation, N.J.; writing—original draft preparation, N.J.; writing—review and editing, N.J., B.G., M.S.L. and J.A.P.; visualization, N.J., B.G., M.S.L. and J.A.P.; supervision, M.S.L. and J.A.P.; project administration, M.S.L. and J.A.P.; funding acquisition, M.S.L. and J.A.P. All authors have read and agreed to the published version of the manuscript.

Funding: The authors are grateful for the financial support of the Project AgriFood XXI NORTE-01-0145-FEDER-000041 and Fundação para a Ciência e a Tecnologia (FCT) to CQVR (UIDB/00616/2020). Ana R. Teixeira also thanks the FCT for the financial support provided through the doctoral scholarship UI/BD/150847/2020.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: The authors are thankful to CITAB, supported by National Funds by FCT—Portuguese Foundation for Science and Technology, under the project UIDB/04033/2020. The authors are grateful for the support provided by the Unidade de Microscopia Eletrónica (UME) of UTAD.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Vlotman, D.E.; Key, D.; Bladergroen, B.J. Technological Advances in Winery Wastewater Treatment: A Comprehensive Review. *South Afr. J. Enol. Vitic.* **2022**, *43*, 58–80. [[CrossRef](#)]
2. Ioannou, L.A.; Puma, G.L.; Fatta-Kassinos, D. Treatment of Winery Wastewater by Physicochemical, Biological and Advanced Processes: A Review. *J. Hazard. Mater.* **2015**, *286*, 343–368. [[CrossRef](#)]

3. Jorge, N.; Santos, C.; Teixeira, A.R.; Marchão, L.; Tavares, P.B.; Lucas, M.S.; Peres, J.A. Treatment of Agro-Industrial Wastewaters by Coagulation-Flocculation-Decantation and Advanced Oxidation Processes—A Literature Review. *Eng. Proc.* **2022**, *19*, 33. [CrossRef]
4. FAOSTAT Almonds, with Shell. Available online: <https://www.fao.org/faostat/en/#home> (accessed on 26 October 2022).
5. Oliveira, I.; Meyer, A.S.; Afonso, S.; Sequeira, A.; Vilela, A.; Goufo, P.; Trindade, H.; Gonçalves, B. Effects of Different Processing Treatments on Almond (*Prunus dulcis*) Bioactive Compounds, Antioxidant Activities, Fatty Acids, and Sensorial Characteristics. *Plants* **2020**, *9*, 1627. [CrossRef]
6. Ahmad, A.; Kurniawan, S.B.; Abdullah, S.R.S.; Othman, A.R.; Hasan, H.A. Exploring the Extraction Methods for Plant-Based Coagulants and Their Future Approaches. *Sci. Total Environ.* **2022**, *818*, 151668. [CrossRef]
7. Madrona, G.S.; Serpelloni, G.B.; Vieira, A.M.S.; Nishi, L.; Cardoso, K.C.; Bergamasco, R. Study of the Effect of Saline Solution on the Extraction of the *Moringa oleifera* Seed's Active Component for Water Treatment. *Water Air Soil Pollut.* **2010**, *211*, 409–415. [CrossRef]
8. Teixeira, A.R.; Jorge, N.; Fernandes, J.R.; Lucas, M.S.; Peres, J.A. Textile Dye Removal by *Acacia dealbata* Link. Pollen Adsorption Combined with UV-A/NTA/Fenton Process. *Top. Catal.* **2022**, *65*, 1045–1061. [CrossRef]
9. Jorge, N.; Teixeira, A.R.; Lucas, M.S.; Peres, J.A. Agro-Industrial Wastewater Treatment with *Acacia dealbata* Coagulation/Flocculation and Photo-Fenton-Based Processes. *Recycling* **2022**, *7*, 54. [CrossRef]
10. Bello, M.M.; Raman, A.A.A.; Asghar, A. A Review on Approaches for Addressing the Limitations of Fenton Oxidation for Recalcitrant Wastewater Treatment. *Process. Saf. Environ. Prot.* **2019**, *126*, 119–140. [CrossRef]
11. Neyens, E.; Baeyens, J. A Review of Classic Fenton's Peroxidation as an Advanced Oxidation Technique. *J. Hazard. Mater.* **2003**, *98*, 33–50. [CrossRef] [PubMed]
12. Qi, C.; Liu, X.; Ma, J.; Lin, C.; Li, X.; Zhang, H. Activation of Peroxymonosulfate by Base: Implications for the Degradation of Organic Pollutants. *Chemosphere* **2016**, *151*, 280–288. [CrossRef] [PubMed]
13. Anipsitakis, G.P.; Dionysiou, D.D. Degradation of Organic Contaminants in Water with Sulfate Radicals Generated by the Conjunction of Peroxymonosulfate with Cobalt. *Environ. Sci. Technol.* **2003**, *37*, 4790–4797. [CrossRef] [PubMed]
14. Anipsitakis, G.P.; Dionysiou, D.D. Transition Metal/UV-Based Advanced Oxidation Technologies for Water Decontamination. *Appl. Catal. B Environ.* **2004**, *54*, 155–163. [CrossRef]
15. Wang, J.; Wang, S. Activation of Persulfate (PS) and Peroxymonosulfate (PMS) and Application for the Degradation of Emerging Contaminants. *Chem. Eng. J.* **2018**, *334*, 1502–1517. [CrossRef]
16. Giannakis, S.; Lin, K.A.; Ghanbari, F. A Review of the Recent Advances on the Treatment of Industrial Wastewaters by Sulfate Radical-Based Advanced Oxidation Processes (SR-AOPs). *Chem. Eng. J.* **2021**, *406*, 127083. [CrossRef]
17. Jorge, N.; Teixeira, A.R.; Marchão, L.; Lucas, M.S.; Peres, J.A. Application of Combined Coagulation–Flocculation–Decantation/Photo-Fenton/Adsorption Process for Winery Wastewater Treatment. *Eng. Proc.* **2022**, *19*, 22. [CrossRef]
18. Jorge, N.; Amor, C.; Teixeira, A.R.; Marchão, L.; Lucas, M.S.; Peres, J.A. Combination of Coagulation-Flocculation-Decantation with Sulfate Radicals for Agro-Industrial Wastewater Treatment. *Eng. Proc.* **2022**, *19*, 19. [CrossRef]
19. Vione, D.; Falletti, G.; Maurino, V.; Minero, C.; Pelizzetti, E.; Malandrino, M.; Ajassa, R.; Olariu, R.-I.; Arsene, C. Sources and Sinks of Hydroxyl Radicals upon Irradiation of Natural Water Samples. *Environ. Sci. Technol.* **2006**, *40*, 3775–3781. [CrossRef]
20. APHA; AWWA; WEF. *Standard Methods for the Examination of Water and Wastewater*, 20th ed.; American Public Health Association: Washington, DC, USA; American Water Works Association: Denver, CO, USA; Water Environment Federation: Alexandria, VA, USA, 1999.
21. Singleton, V.L.; Rossi, J.A. Colorimetry of Total Phenolics with Phosphomolybdic-Phosphotungstic Acid Reagents. *Am. J. Enol. Vitic.* **1965**, *16*, 144–158.
22. Šćiban, M.; Klačnja, M.; Antov, M.; Škrbić, B. Removal of Water Turbidity by Natural Coagulants Obtained from Chestnut and Acorn. *Bioresour. Technol.* **2009**, *100*, 6639–6643. [CrossRef]
23. Oussalah, A.; Boukerroui, A.; Aichour, A.; Djellouli, B. Cationic and Anionic Dyes Removal by Low-Cost Hybrid Alginate/Natural Bentonite Composite Beads: Adsorption and Reusability Studies. *Int. J. Biol. Macromol.* **2019**, *124*, 854–862. [CrossRef] [PubMed]
24. Dewanto, V.; Wu, X.; Adom, K.K.; Liu, R.H. Thermal Processing Enhances the Nutritional Value of Tomatoes by Increasing Total Antioxidant Activity. *J. Agric. Food Chem.* **2002**, *50*, 3010–3014. [CrossRef] [PubMed]
25. Soufi, O.; Romero, C.; Hayette, L. Ortho-Diphenol Profile and Antioxidant Activity of Algerian Black Olive Cultivars: Effect of Dry Salting Process. *Food Chem.* **2014**, *157*, 504–510. [CrossRef]
26. Siddhuraju, P.; Becker, K. Antioxidant Properties of Various Solvent Extracts of Total Phenolic Constituents from Three Different Agroclimatic Origins of Drumstick Tree (*Moringa Oleifera* Lam.) Leaves. *J. Agric. Food Chem.* **2003**, *51*, 2144–2155. [CrossRef] [PubMed]
27. Jorge, N.; Teixeira, A.R.; Matos, C.C.; Lucas, M.S.; Peres, J.A. Combination of Coagulation–Flocculation–Decantation and Ozonation Processes for Winery Wastewater Treatment. *Int. J. Environ. Res. Public Health* **2021**, *18*, 8882. [CrossRef]
28. Jorge, N.; Teixeira, A.R.; Guimarães, V.; Lucas, M.S.; Peres, J.A. Treatment of Winery Wastewater with a Combination of Adsorption and Thermocatalytic Processes. *Processes* **2022**, *10*, 75. [CrossRef]
29. OECD. *OECD Guidelines for the Testing of Chemicals: Terrestrial Plant Test*; OECD: Paris, France, 2004; Volume 208.
30. Lin, D.; Xing, B. Phytotoxicity of Nanoparticles: Inhibition of Seed Germination and Root Growth. *Environ. Pollut.* **2007**, *150*, 243–250. [CrossRef]

31. Varnero, M.T.; Rojas, C.; Orellana, R. Índices de Fitotoxicidad En Residuos Orgánicos Durante El Compostaje. *Rev. Cienc. Suelo Nutr. Veg.* **2007**, *7*, 28–37. [[CrossRef](#)]
32. Tiquia, S.M.; Tam, N.F.Y. Elimination of Phytotoxicity during Co-Composting of Spent Pig-Manure Sawdust Litter and Pig Sludge. *Bioresour. Technol.* **1998**, *65*, 43–49. [[CrossRef](#)]
33. Subramanian, A.; Harper, W.J.; Rodriguez-Saona, L.E. Rapid Prediction of Composition and Flavor Quality of Cheddar Cheese Using ATR-FTIR Spectroscopy. *J. Food Sci.* **2015**, *74*, C292–C297. [[CrossRef](#)]
34. Beltrán Sanahuja, A.; Moya, P.; Maestre Pérez, S.E.; Grané Teruel, N.; Martín Carratalá, M.L. Classification of Four Almond Cultivars Using Oil Degradation Parameters Based on FTIR and GC Data. *J. Am. Oil Chem. Soc.* **2009**, *86*, 51–58. [[CrossRef](#)]
35. Maqsood, S.; Benjakul, S. Synergistic Effect of Tannic Acid and Modified Atmospheric Packaging on the Prevention of Lipid Oxidation and Quality Losses of Refrigerated Striped Catfish Slices. *Food Chem.* **2010**, *121*, 29–38. [[CrossRef](#)]
36. Rohman, A.; Man, Y.C. Fourier Transform Infrared (FTIR) Spectroscopy for Analysis of Extra Virgin Olive Oil Adulterated with Palm Oil. *Food Res. Int.* **2010**, *43*, 886–892. [[CrossRef](#)]
37. García, A.V.; Beltran Sanahuja, A.; Garrigos Selva, M.D.C. Characterization and Classification of Almond Cultivars by Using Spectroscopic and Thermal Techniques. *J. Food Sci.* **2013**, *78*, C138–C144. [[CrossRef](#)]
38. Martins, R.B.; Jorge, N.; Lucas, M.S.; Raymundo, A.; Barros, A.I.; Peres, J.A. Food By-Product Valorization by Using Plant-Based Coagulants Combined with AOPs for Agro-Industrial Wastewater Treatment. *Int. J. Environ. Res. Public Health* **2022**, *19*, 4134. [[CrossRef](#)]
39. Martín, N.; Bernard, D. Calcium Signaling and Cellular Senescence. *Cell Calcium* **2018**, *70*, 16–23. [[CrossRef](#)]
40. Oliveira, I.; Meyer, A.S.; Afonso, S.; Aires, A.; Goufo, P.; Trindade, H.; Gonçalves, B. Phenolic and Fatty Acid Profiles, A-tocopherol and Sucrose Contents, and Antioxidant Capacities of Understudied Portuguese Almond Cultivars. *J. Food Biochem.* **2019**, *43*, e12887. [[CrossRef](#)]
41. dos Santos Escobar, O.; de Azevedo, C.F.; Swarowsky, A.; Adebayo, M.A.; Netto, M.S.; Machado, F.M. Utilization of Different Parts of *Moringa oleifera* Lam. Seeds as Biosorbents to Remove Acid Blue 9 Synthetic Dye. *J. Environ. Chem. Eng.* **2021**, *9*, 105553. [[CrossRef](#)]
42. Hussain, G.; Haydar, S. Textile Effluent Treatment Using Natural Coagulant *Opuntia Stricta* in Comparison with Alum. *CLEAN–Soil Air Water* **2021**, *49*, 2000342. [[CrossRef](#)]
43. Wang, Y.; Li, Y.; Zhang, Y.; Wei, W. Enhanced Brilliant Blue FCF Adsorption Using Microwave-Hydrothermal Synthesized Hydroxyapatite Nanoparticles. *J. Dispers. Sci. Technol.* **2020**, *41*, 1346–1355. [[CrossRef](#)]
44. Beltrán-Heredia, J.; Sánchez-Martín, J.; Muñoz-Serrano, A.; Peres, J.A. Towards Overcoming TOC Increase in Wastewater Treated with *Moringa oleifera* Seed Extract. *Chem. Eng. J.* **2012**, *188*, 40–46. [[CrossRef](#)]
45. Rodríguez-Chueca, J.; Amor, C.; Mota, J.; Lucas, M.S.; Peres, J.A. Oxidation of Winery Wastewater by Sulphate Radicals: Catalytic and Solar Photocatalytic Activations. *Environ. Sci. Pollut. Res.* **2017**, *24*, 22414–22426. [[CrossRef](#)] [[PubMed](#)]
46. Rodríguez-Chueca, J.; Amor, C.; Silva, T.; Dionysiou, D.D.; Li Puma, G.; Lucas, M.S.; Peres, J.A. Treatment of Winery Wastewater by Sulphate Radicals: HSO₅[−] /Transition Metal/UV-A LEDs. *Chem. Eng. J.* **2017**, *310*, 473–483. [[CrossRef](#)]
47. Ferreira, L.C.; Fernandes, J.R.; Rodriguez-Chueca, J.; Peres, J.A.; Lucas, M.S.; Tavares, P.B. Photocatalytic Degradation of an Agro-Industrial Wastewater Model Compound Using a UV LEDs System: Kinetic Study. *J. Environ. Manage.* **2020**, *269*, 110740. [[CrossRef](#)]
48. Jorge, N.; Teixeira, A.R.; Fernandes, J.R.; Oliveira, I.; Lucas, M.S.; Peres, J.A. Degradation of Agro-Industrial Wastewater Model Compound by UV-A-Fenton Process: Batch vs. Continuous Mode. *Int. J. Environ. Res. Public Health* **2023**, *20*, 1276. [[CrossRef](#)] [[PubMed](#)]
49. Li, J.; Lin, H.; Yang, L.; Zhang, H. Copper-Spent Activated Carbon as a Heterogeneous Peroxydisulfate Catalyst for the Degradation of Acid Orange 7 in an Electrochemical Reactor. *Water Sci. Technol.* **2016**, *73*, 1802–1808. [[CrossRef](#)] [[PubMed](#)]
50. Huang, Y.; Jia, Y.; Zuo, L.; Huo, Y.; Zhang, Y. Comparison of VUV/H₂O₂ and VUV/PMS (Peroxymonosulfate) for the Degradation of Unsymmetrical Dimethylhydrazine in Water. *J. Water Process Eng.* **2022**, *49*, 102970. [[CrossRef](#)]
51. He, B.; Yang, Y.; Liu, B.; Zhao, Z.; Shang, J.; Cheng, X. Degradation of Chlortetracycline Hydrochloride by Peroxymonosulfate Activation on Natural Manganese Sand Through Response Surface Methodology. *Environ. Sci. Pollut. Res.* **2022**, *29*, 82584–82599. [[CrossRef](#)]
52. Govindan, K.; Raja, M.; Noel, M.; James, E.J. Degradation of Pentachlorophenol by Hydroxyl Radicals and Sulfate Radicals Using Electrochemical Activation of Peroxomonosulfate, Peroxydisulfate and Hydrogen Peroxide. *J. Hazard. Mater.* **2014**, *272*, 42–51. [[CrossRef](#)]
53. Rastogi, A.; Al-Abad, S.R.; Dionysiou, D.D. Sulfate Radical-Based Ferrous–Peroxymonosulfate Oxidative System for PCBs Degradation in Aqueous and Sediment Systems. *Appl. Catal. B Environ.* **2009**, *85*, 171–179. [[CrossRef](#)]
54. Huling, S.G.; Ko, S.; Park, S.; Kan, E. Persulfate Oxidation of MTBE-and Chloroform-Spent Granular Activated Carbon. *J. Hazard. Mater.* **2011**, *192*, 1484–1490. [[CrossRef](#)] [[PubMed](#)]
55. Yi, Q.; Tan, J.; Liu, W.; Lu, H.; Xing, M.; Zhang, J. Peroxymonosulfate Activation by Three-Dimensional Cobalt Hydroxide/Graphene Oxide Hydrogel for Wastewater Treatment through an Automated Process. *Chem. Eng. J.* **2020**, *400*, 125965. [[CrossRef](#)]
56. Li, J.; Xu, M.; Yao, G.; Lai, B. Enhancement of the Degradation of Atrazine through CoFe₂O₄ Activated Peroxymonosulfate (PMS) Process: Kinetic, Degradation Intermediates, and Toxicity Evaluation. *Chem. Eng. J.* **2018**, *348*, 1012–1024. [[CrossRef](#)]

57. Chen, J.; Qian, Y.; Liu, H.; Huang, T. Oxidative Degradation of Diclofenac by Thermally Activated Persulfate: Implication for ISCO. *Environ. Sci. Pollut. Res.* **2016**, *23*, 3824–3833. [[CrossRef](#)]
58. Tan, C.; Gao, N.; Deng, Y.; An, N.; Deng, J. Heat-Activated Persulfate Oxidation of Diuron in Water. *Chem. Eng. J.* **2012**, *203*, 294–300. [[CrossRef](#)]
59. Rodríguez-Chueca, J.; Giannakis, S.; Marjanovic, M.; Kohantorabi, M.; Gholami, M.R.; Grandjean, D.; de Alencastro, L.F.; Pulgarín, C. Solar-Assisted Bacterial Disinfection and Removal of Contaminants of Emerging Concern by Fe²⁺-Activated HSO₅⁻ vs. S₂O₈²⁻ in Drinking Water. *Appl. Catal. B Environ.* **2019**, *248*, 62–72. [[CrossRef](#)]
60. Ahmadi, M.; Ghanbari, F. Combination of UVC-LEDs and Ultrasound for Peroxymonosulfate Activation to Degrade Synthetic Dye: Influence of Promotional and Inhibitory Agents and Application for Real Wastewater. *Environ. Sci. Pollut. Res.* **2018**, *25*, 6003–6014. [[CrossRef](#)]
61. Latif, A.; Kai, S.; Si, Y. Catalytic Degradation of Organic Pollutants in Fe (III)/Peroxymonosulfate (PMS) System: Performance, Influencing Factors, and Pathway. *Environ. Sci. Pollut. Res.* **2019**, *26*, 36410–36422. [[CrossRef](#)]
62. Fang, X.; Gan, L.; Wang, L.; Gong, H.; Xu, L.; Wu, Y.; Lu, H.; Han, S.; Cui, J.; Xia, C. Enhanced Degradation of Bisphenol A by Mixed ZIF Derived CoZn Oxide Encapsulated N-Doped Carbon via Peroxymonosulfate Activation: The Importance of N Doping Amount. *J. Hazard. Mater.* **2021**, *419*, 126363. [[CrossRef](#)]
63. Grehs, B.W.N.; Lopes, A.R.; Moreira, N.F.F.; Fernandes, T.; Linton, M.A.O.; Silva, A.M.T.; Manaia, C.M.; Carissimi, E.; Nunes, O.C. Removal of Microorganisms and Antibiotic Resistance Genes from Treated Urban Wastewater: A Comparison between Aluminium Sulphate and Tannin Coagulants. *Water Res.* **2019**, *166*, 115056. [[CrossRef](#)]
64. Sun, J.; Li, X.; Feng, J.; Tian, X. Oxone/Co²⁺ Oxidation as an Advanced Oxidation Process: Comparison with Traditional Fenton Oxidation for Treatment of Landfill Leachate. *Water Res.* **2009**, *43*, 4363–4369. [[CrossRef](#)] [[PubMed](#)]
65. Rodríguez-Narvaez, O.M.; Pacheco-Alvarez, M.O.A.; Wróbel, K.; Páramo-Vargas, J.; Bandala, E.R.; Brillas, E.; Peralta-Hernandez, J.M. Development of a Co²⁺/PMS Process Involving Target Contaminant Degradation and PMS Decomposition. *Int. J. Environ. Sci. Technol.* **2020**, *17*, 17–26. [[CrossRef](#)]
66. Wang, Y.R.; Chu, W. Photo-Assisted Degradation of 2,4,5-Trichlorophenoxyacetic Acid by Fe(II)-Catalyzed Activation of Oxone Process: The Role of UV Irradiation, Reaction Mechanism and Mineralization. *Appl. Catal. B Environ.* **2012**, *123–124*, 151–161. [[CrossRef](#)]
67. Guerra-Rodríguez, S.; Ribeiro, A.R.L.; Ribeiro, R.S.; Rodríguez, E.; Silva, A.M.; Rodríguez-Chueca, J. UV-A Activation of Peroxymonosulfate for the Removal of Micropollutants from Secondary Treated Wastewater. *Sci. Total Environ.* **2021**, *770*, 145299. [[CrossRef](#)]
68. Ao, X.; Li, Z.; Zhang, H. A Comprehensive Insight into a Rapid Degradation of Sulfamethoxazole by Peroxymonosulfate Enhanced UV-A LED/g-C₃N₄ Photocatalysis. *J. Clean. Prod.* **2022**, *356*, 131822. [[CrossRef](#)]
69. Yin, R.; Guo, W.; Wang, H.; Du, J.; Zhou, X.; Wu, Q.; Zheng, H.; Chang, J.; Ren, N. Enhanced Peroxymonosulfate Activation for Sulfamethazine Degradation by Ultrasound Irradiation: Performances and Mechanisms. *Chem. Eng. J.* **2018**, *335*, 145–153. [[CrossRef](#)]
70. Lu, Y.; Xu, W.; Nie, H.; Zhang, Y.; Deng, N.; Zhang, J. Mechanism and Kinetic Analysis of Degradation of Atrazine by US/PMS. *Int. J. Environ. Res. Public Health* **2019**, *16*, 1781. [[CrossRef](#)]
71. Lucas, M.S.; Peres, J.A.; Li, G. Treatment of Winery Wastewater by Ozone-Based Advanced Oxidation Processes (O₃, O₃/UV and O₃/UV/H₂O₂) in a Pilot-Scale Bubble Column Reactor and Process Economics. *Sep. Purif. Technol.* **2010**, *72*, 235–241. [[CrossRef](#)]
72. Marchão, L.; Fernandes, J.R.; Sampaio, A.; Peres, J.A.; Tavares, P.B.; Lucas, M.S. Microalgae and Immobilized TiO₂/UV-A LEDs as a Sustainable Alternative for Winery Wastewater Treatment. *Water Res.* **2021**, *203*, 117464. [[CrossRef](#)]
73. Esteves, B.M.; Rodrigues, C.S.D.; Maldonado-hódar, F.J.; Madeira, L.M. Treatment of High-Strength Olive Mill Wastewater by Combined Fenton-like Oxidation and Coagulation/Flocculation. *J. Environ. Chem. Eng.* **2019**, *7*, 103252. [[CrossRef](#)]
74. Norzaee, S.; Taghavi, M.; Djahed, B.; Kord Mostafapour, F. Degradation of Penicillin G by Heat Activated Persulfate in Aqueous Solution. *J. Environ. Manage.* **2018**, *215*, 316–323. [[CrossRef](#)] [[PubMed](#)]
75. Pouran, S.R.; Raman, A.A.A.; Daud, W.M.A.W. Review on the Application of Modified Iron Oxides as Heterogeneous Catalysts in Fenton Reactions. *J. Clean. Prod.* **2014**, *64*, 24–35. [[CrossRef](#)]
76. Li, M.; Yang, X.; Wang, D.; Yuan, J. Enhanced Oxidation of Erythromycin by Persulfate Activated Iron Powder–H₂O₂ System: Role of the Surface Fe Species and Synergistic Effect of Hydroxyl and Sulfate Radicals. *Chem. Eng. J.* **2017**, *317*, 103–111. [[CrossRef](#)]
77. Jaafarzadeh, N.; Ghanbari, F.; Alvandi, M. Integration of Coagulation and Electro-Activated HSO₅⁻ to Treat Pulp and Paper Wastewater. *Sustain. Environ. Res.* **2017**, *27*, 223–229. [[CrossRef](#)]
78. Amor, C.; Rodríguez-Chueca, J.; Fernandes, J.L.; Domínguez, J.R.; Lucas, M.S.; Peres, J.A. Winery Wastewater Treatment by Sulphate Radical Based-Advanced Oxidation Processes (SR-AOP): Thermally vs UV-Assisted Persulphate Activation. *Process. Saf. Environ. Prot.* **2019**, *122*, 94–101. [[CrossRef](#)]
79. Jaafarzadeh, N.; Omidinasab, M.; Ghanbari, F. Combined Electrocoagulation and UV-Based Sulfate Radical Oxidation Processes for Treatment of Pulp and Paper Wastewater. *Process. Saf. Environ. Prot.* **2016**, *102*, 462–472. [[CrossRef](#)]
80. Papastefanakis, N.; Mantzavinos, D.; Katsaounis, A. DSA Electrochemical Treatment of Olive Mill Wastewater on Ti/RuO₂ Anode. *J. Appl. Electrochem.* **2010**, *40*, 729–737. [[CrossRef](#)]

81. Amor, C.; Lucas, M.S.; Pirra, A.J.; Peres, J.A. Treatment of Concentrated Fruit Juice Wastewater by the Combination of Biological and Chemical Processes. *J. Env. Sci. Health Part A* **2012**, *47*, 1809–1817. [[CrossRef](#)]
82. Rodríguez-Chueca, J.; Amor, C.; Fernandes, J.R.; Tavares, P.B.; Lucas, M.S.; Peres, J.A. Treatment of Crystallized-Fruit Wastewater by UV-A LED Photo-Fenton and Coagulation-Flocculation. *Chemosphere* **2016**, *145*, 351–359. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.