Henry Ford Health Henry Ford Health Scholarly Commons

Hematology/Oncology Articles

Hematology-Oncology

2-9-2023

Association between the choice of the conditioning regimen and outcomes of allogeneic hematopoietic cell transplantation for myelofibrosis

Guru Subramanian Guru Murthy

Soyoung Kim

Noel Estrada-Merly

Muhammad Bilal Abid

Mahmoud Aljurf

See next page for additional authors

Follow this and additional works at: https://scholarlycommons.henryford.com/ hematologyoncology_articles

Authors

Guru Subramanian Guru Murthy, Soyoung Kim, Noel Estrada-Merly, Muhammad Bilal Abid, Mahmoud Aljurf, Amer Assal, Talha Badar, Sherif M. Badawy, Karen Ballen, Amer Beitinjaneh, Jan Cerny, Saurabh Chhabra, Zachariah DeFilipp, Bhagirathbhai Dholaria, Miguel Angel Diaz Perez, Shatha Farhan, Cesar O. Freytes, Robert Peter Gale, Siddhartha Ganguly, Vikas Gupta, Michael R. Grunwald, Nada Hamad, Gerhard C. Hildebrandt, Yoshihiro Inamoto, Tania Jain, Omer Jamy, Mark Juckett, Matt Kalaycio, Maxwell M. Krem, Hillard M. Lazarus, Mark Litzow, Reinhold Munker, Hemant S. Murthy, Sunita Nathan, Taiga Nishihori, Guillermo Ortí, Sagar S. Patel, Marjolein Van der Poel, David A. Rizzieri, Bipin N. Savani, Sachiko Seo, Melhem Solh, Leo F. Verdonck, Baldeep Wirk, Jean A. Yared, Ryotaro Nakamura, Betul Oran, Bart Scott, and Wael Saber



Association between the choice of the conditioning regimen and outcomes of allogeneic hematopoietic cell transplantation for myelofibrosis

by Guru Subramanian Guru Murthy, Soyoung Kim, Noel Estrada-Merly, Muhammad Bilal Abid, Mahmoud Aljurf, Amer Assal, Talha Badar, Sherif M. Badawy, Karen Ballen, Amer Beitinjaneh, Jan Cerny, Saurabh Chhabra, Zachariah DeFilipp, Bhagirathbhai Dholaria, Miguel Angel Diaz Perez, Shatha Farhan, Cesar O. Freytes, Robert Peter Gale, Siddhartha Ganguly, Vikas Gupta, Michael R. Grunwald, Nada Hamad, Gerhard C. Hildebrandt, Yoshihiro Inamoto, Tania Jain, Omer Jamy, Mark Juckett, Matt Kalaycio, Maxwell M. Krem, Hillard M. Lazarus, Mark Litzow, Reinhold Munker, Hemant S. Murthy, Sunita Nathan, Taiga Nishihori, Guillermo Ortí, Sagar S. Patel, Marjolein van der Poel, David A. Rizzieri, Bipin N. Savani, Sachiko Seo, Melhem Solh, Leo F. Verdonck, Baldeep Wirk, Jean A. Yared, Ryotaro Nakamura, Betul Oran, Bart Scott, and Wael Saber

Received: September 6, 2022. Accepted: February 1, 2023.

Citation: Guru Subramanian Guru Murthy, Soyoung Kim, Noel Estrada-Merly, Muhammad Bilal Abid, Mahmoud Aljurf, Amer Assal, Talha Badar, Sherif M. Badawy, Karen Ballen, Amer Beitinjaneh, Jan Cerny, Saurabh Chhabra, Zachariah DeFilipp, Bhagirathbhai Dholaria, Miguel Angel Diaz Perez, Shatha Farhan, Cesar O. Freytes, Robert Peter Gale, Siddhartha Ganguly, Vikas Gupta, Michael R. Grunwald, Nada Hamad, Gerhard C. Hildebrandt, Yoshihiro Inamoto, Tania Jain, Omer Jamy, Mark Juckett, Matt Kalaycio, Maxwell M. Krem, Hillard M. Lazarus, Mark Litzow, Reinhold Munker, Hemant S. Murthy, Sunita Nathan, Taiga Nishihori, Guillermo Ortí, Sagar S. Patel, Marjolein van der Poel, David A. Rizzieri, Bipin N. Savani, Sachiko Seo, Melhem Solh, Leo F. Verdonck, Baldeep Wirk, Jean A. Yared, Ryotaro Nakamura, Betul Oran, Bart Scott, and Wael Saber. Association between the choice of the conditioning regimen and outcomes of allogeneic hematopoietic cell transplantation for myelofibrosis. Haematologica. 2023 Feb 9. doi: 10.3324/haematol.2022.281958 [Epub ahead of print]

Publisher's Disclaimer.

E-publishing ahead of print is increasingly important for the rapid dissemination of science. Haematologica is, therefore, E-publishing PDF files of an early version of manuscripts that have completed a regular peer review and have been accepted for publication. E-publishing of this PDF file has been approved by the authors. After having E-published Ahead of Print, manuscripts will then undergo technical and English editing, typesetting, proof correction and be presented for the authors' final approval; the final version of the manuscript will then appear in a regular issue of the journal. All legal disclaimers that apply to the journal also pertain to this production process.

Title page

Title: Association between the choice of the conditioning regimen and outcomes of allogeneic hematopoietic cell transplantation for myelofibrosis

Running head: Conditioning regimen in myelofibrosis

<u>Authors:</u> Guru Subramanian Guru Murthy¹, Soyoung Kim^{2, 3}, Noel Estrada-Merly³, Muhammad Bilal Abid⁴, Mahmoud Aljurf⁵, Amer Assal⁶, Talha Badar⁷, Sherif M. Badawy^{8,9}, Karen Ballen¹⁰, Amer Beitinjaneh¹¹, Jan Cerny¹², Saurabh Chhabra³, Zachariah DeFilipp¹³, Bhagirathbhai Dholaria¹⁴, Miguel Angel Diaz Perez¹⁵, Shatha Farhan¹⁶, Cesar O. Freytes¹⁷, Robert Peter Gale¹⁸, Siddhartha Ganguly¹⁹, Vikas Gupta²⁰, Michael R. Grunwald²¹, Nada Hamad²², Gerhard C. Hildebrandt²³, Yoshihiro Inamoto²⁴, Tania Jain²⁵, Omer Jamy²⁶, Mark Juckett²⁷, Matt Kalaycio²⁸, Maxwell M. Krem²⁹, Hillard M Lazarus³⁰, Mark Litzow³¹, Reinhold Munker²³, Hemant S. Murthy³², Sunita Nathan³³, Taiga Nishihori³⁴, Guillermo Ortí³⁵, Sagar S. Patel³⁶, Marjolein van der Poel³⁷, David A Rizzieri³⁸, Bipin N Savani³⁹, Sachiko Seo⁴⁰, Melhem Solh⁴¹, Leo F. Verdonck⁴², Baldeep Wirk⁴³, Jean A. Yared⁴⁴, Ryotaro Nakamura⁴⁵, Betul Oran⁴⁶, Bart Scott⁴⁷, Wael Saber³

Affiliations:

¹Division of Hematology & Oncology, Medical College of Wisconsin, Milwaukee, WI; ²Division of Biostatistics, Institute for Health and Equity, Medical College of Wisconsin, Milwaukee, WI; ³CIBMTR® (Center for International Blood and Marrow Transplant Research), Department of Medicine, Medical College of Wisconsin, Milwaukee, WI; ⁴Divisions of Hematology/Oncology, & Infectious Diseases, Department of Medicine,

Medical College of Wisconsin, Milwaukee, WI; ⁵Department of Oncology, King Faisal Specialist Hospital Center & Research, Rivadh, Saudi Arabia; ⁶Columbia University Irving Medical Center, Department of Medicine, Bone Marrow Transplant and Cell Therapy Program, New York, NY; ⁷Mayo Clinic, Jacksonville, FL; ⁸Division of Hematology, Oncology and Stem Cell Transplantation, Ann & Robert H. Lurie Children's Hospital of Chicago, Chicago, IL; ⁹Department of Pediatrics, Northwestern University Feinberg School of Medicine, Chicago, IL; ¹⁰Division of Hematology/Oncology, University of Virginia Health System, Charlottesville, VA; ¹¹Division of Transplantation and Cellular Therapy, University of Miami Hospital and Clinics, Slyvester Comprehensive Cancer Center, Miami, FL; ¹²Division of Hematology/Oncology, Department of Medicine, University of Massachusetts Medical Center, Worcester, MA; ¹³Hematopoietic Cell Transplant and Cellular Therapy Program, Massachusetts General Hospital, Boston, MA; ¹⁴Vanderbilt University Medical Center, Nashville, TN; ¹⁵Department of Hematology/Oncology, Hospital Infantil Universitario Niño Jesus, Madrid, Spain; ¹⁶Henry Ford Health System Stem Cell Transplant & Cellular Therapy Program, Detroit, MI; ¹⁷University of Texas Health Science Center at San Antonio, San Antonio, TX; ¹⁸Haematology Research Centre, Department of Immunology and Inflammation, Imperial College London, London, UK; ¹⁹Division of Hematological Malignancy and Cellular Therapeutics, University of Kansas Health System, Kansas City, KS; ²⁰ MPN Program, Princess Margaret Cancer Centre, University of Toronto, Toronto, ON, Canada; ²¹Department of Hematologic Oncology and Blood Disorders, Levine Cancer Institute, Atrium Health, Charlotte, NC; ²²St. Vincent Hospital, Darlinghurst, NSW, Australia; ²³Markey Cancer Center, University of Kentucky,

Lexington, KY; ²⁴Division of Hematopoietic Stem Cell Transplantation, National Cancer Center, Tokyo, Japan; ²⁵John Hopkins University School of Medicine, Baltimore, MD; ²⁶University of Alabama at Birmingham, Birmingham, AL; ²⁷University of Minnesota Blood and Marrow Transplant Program – Adults; ²⁸Cleveland Clinic Taussig Cancer Institute, Cleveland Clinic, Cleveland, OH; ²⁹Kansas City VA Medical Center, Kansas City, MO; ³⁰University Hospitals Cleveland Medical Center, Case Western Reserve University, Cleveland, OH; ³¹Division of Hematology and Transplant Center, Mayo Clinic Rochester, Rochester, MN; ³²Division of Hematology-Oncology, Blood and Marrow Transplantation Program, Mayo Clinic, Jacksonville, FL; ³³Section of Bone Marrow Transplant and Cell Therapy, Rush University Medical Center, Chicago, IL; ³⁴Department of Blood & Marrow Transplant and Cellular Immunotherapy (BMT CI), Moffitt Cancer Center, Tampa, FL; ³⁵Vall d'Hebron University Hospital, Barcelona, Spain; ³⁶Blood and Marrow Transplant Program, Huntsman Cancer Institute, University of Utah, Salt Lake City, UT; ³⁷Department of Internal Medicine, Division of Hematology, GROW School for Oncology and Developmental Biology, Masstricht University Medical Center, Maastricht, Netherlands; ³⁸Division of Hematologic Malignancies and Cellular Therapy, Duke University, Durham, NC; ³⁹Division of Hematology/Oncology, Department of Medicine, Vanderbilt University Medical Center, Nashville, TN; ⁴⁰Department of Hematology and Oncology, Dokkyo Medical University, Tochigo, Japan; ⁴¹The Blood and Marrow Transplant Group of Georgia, Northside Hospital, Atlanta, GA; ⁴²Department of Hematology/Oncology, Isala, Clinic, Zwolle, The Netherlands; ⁴³Bone Marrow Transplant Program, Penn State Cancer Institute, Hershey, Pennsylvania; ⁴⁴Transplantation & Cellular Therapy Program, Division of

Hematology/Oncology, Department of Medicine, Greenebaum Comprehensive Cancer Center, University of Maryland, Baltimore, MD; ⁴⁵Department of Hematology & Hematopoietic Cell Transplantation, City of Hope, Duarte, CA; ⁴⁶Department of Stem Cell Transplantation, Division of Cancer Medicine, The University of Texas MD Anderson Cancer Center, Houston, TX; ⁴⁷Fred Hutchinson Cancer Research Center, Seattle, WA

Corresponding Author

Guru Subramanian Guru Murthy MD, MS Division of Hematology & Oncology Medical College of Wisconsin Milwaukee, WI Email: gmurthy@mcw.edu Ph: 414-805-4600 Fax number: 414-805-4606

Abstract Word Count: 219

Text Word Count: 2607

Figures: 3 (main), 12 (supplement)

Tables: 2 (main), 7 (supplement)

Reference Count: 27

<u>Key words</u>: Myelofibrosis, Stem cell transplantation, Conditioning regimen, Allogeneic, Conditioning intensity

Acknowledgement:

Funding: The CIBMTR is supported primarily by Public Health Service U24CA076518 from the National Cancer Institute (NCI), the National Heart, Lung and Blood Institute (NHLBI) and the National Institute of Allergy and Infectious Diseases (NIAID): HHSH250201700006C from the Health Resources and Services Administration (HRSA); and N00014-20-1-2705 and N00014-20-1-2832 from the Office of Naval Research; Support is also provided by Be the Match Foundation, the Medical College of Wisconsin, the National Marrow Donor Program, and from the following commercial entities: AbbVie; Accenture; Actinium Pharmaceuticals, Inc.: Adaptive Biotechnologies Corporation; Adienne SA; Allovir, Inc.; Amgen, Inc.; Astellas Pharma US; bluebird bio, inc.; Bristol Myers Squibb Co.; CareDx; CSL Behring; CytoSen Therapeutics, Inc.; Daiichi Sankyo Co., Ltd.; Eurofins Viracor, DBA Eurofins Transplant Diagnostics; Fate Therapeutics; Gamida-Cell, Ltd.; Gilead; GlaxoSmithKline; HistoGenetics; Incyte Corporation; Iovance; Janssen Research & Development, LLC; Janssen/Johnson & Johnson; Jasper Therapeutics; Jazz Pharmaceuticals, Inc.; Kadmon; Karius; Karyopharm Therapeutics; Kiadis Pharma; Kite Pharma Inc; Kite, a Gilead Company; Kyowa Kirin International plc; Kyowa Kirin; Legend Biotech; Magenta Therapeutics; Medac GmbH; Medexus; Merck & Co.; Millennium, the Takeda Oncology Co.; Miltenyi Biotec, Inc.; MorphoSys; Novartis Pharmaceuticals Corporation; Omeros Corporation; Oncolmmune, Inc.; Oncopeptides, Inc.; OptumHealth; Orca Biosystems, Inc.; Ossium Health, Inc; Pfizer, Inc.; Pharmacyclics, LLC; Priothera; Sanofi Genzyme; Seagen, Inc.; Stemcyte; Takeda Pharmaceuticals; Talaris Therapeutics; Terumo Blood and Cell Technologies; TG Therapeutics; Tscan; Vertex; Vor Biopharma; Xenikos BV.

DATA USE STATEMENT

CIBMTR supports accessibility of research in accord with the National Institutes of Health (NIH) Data Sharing Policy and the National Cancer Institute (NCI) Cancer Moonshot Public Access and Data Sharing Policy. The CIBMTR only releases deidentified datasets that comply with all relevant global regulations regarding privacy and confidentiality.

Conflict of interest:

Dr. Guru Subramanian Guru Murthy reports the following outside the submitted work -Cardinal Health (Honoraria), TG Therapeutics (Advisory board), Gilead (Consultancy), Cancerexpert now (Consultancy), Qessential (Consultancy), Techspert (Consultancy), DAVA Oncology (Honoraria), and Curio science (Honoraria), all outside the submitted work.

Dr. Assal reports research funding from Incyte Corporation.

Dr. Badar reports honorarium from Pfizer Hematology and Oncology.

Dr. Cerny reports participation on a Data Safety Monitoring Board for Allovir, Inc.; financial relationships with Actinium Pharmaceuticals, Bluebird Bio Inc., Dynavax Pharma, Atyr Pharmac, Gamida Cell, Miragen Therapeutics, Mustang Bio, Novavax, Ovid Therapeutics, Sorrento Therapeutics, TG Therapeutics, Vaxart Inc, and Veru Inc., outside the submitted work.

Dr. Dholaria reports institutional research funding with Takeda, Janssen, Angiocrine, Pfizer, Poseida, MEI, Sorrento; consultancy with Jazz, Celgene, and Gamida Cell.

Dr. Ganguly reports financial relationships as speaker with Seattle Genetics and KITE Pharma; and advisory board member with Kadmon, BMS, Sanofi, Astrazeneca, Kite, Daiichi Sankyo and Astellas.

Dr.Gupta reports consultancy work for Novartis, Incyte, BMS-Celgene, Sierra Oncology, Morphosys, Pfizer, and Takeda, and received research grant through institution from Novartis and Incyte.

Dr. Grunwald reports having worked as a PI with multiple pharmaceutical sponsors and as both consultant and PI for Incyte (manufacturer of ruxolitinib).

Dr. Hamad reports as advisory board member of Novartis.

Dr. Jain reports honoraria for advisory board participation for Care Dx. Bristol Myers Squibb, and Incyte; honoraria for lecture at APP Oncology Summit.

Dr. Nishihori reports research support (clinical trial support) to the institution by Novartis; research support (drug supply only) to the institution for clinical trial by Karyopharm. Dr. Orti reports financial relationships with Incyte (grant), BMS (personal fees), Incyte

(personal fees), Novartis (personal fees), and Pfizer (personal fees).

Dr.Munker reports research support and stock ownership with Incyte.

Dr. Rizzieri reports as a consultant and on speaker bureau for Incyte (makers of ruxolitinib used for treatment).

<u>Role of the Funder/Sponsor:</u> The funding organizations had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

<u>Disclaimer</u>: The views expressed in this article do not reflect the official policy or position of the NIH, the Department of the Navy, the Department of Defense, the HRSA, or any other agency of the US Government.

<u>Contribution</u>: G.M., W.S., S.K, N.E conceived and designed the study, collected and assembled the data, and wrote the manuscript; all authors performed data analysis and interpretation of data; and G.M., W.S., S.K, provided final approval of the manuscript. G.M and W.S had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Presented in part as an abstract at the American Society of Hematology Annual Meeting Dec 2021.

ABSTRACT

Allogeneic hematopoietic cell transplantation (allo-HCT) remains the only curative treatment for myelofibrosis. However, the optimal conditioning regimen either with reduced intensity conditioning (RIC) or myeloablative conditioning (MAC) is not well known. Using the Center for International Blood and Marrow Transplant Research database, we identified adults aged ≥18 years with myelofibrosis undergoing allo-HCT between 2008-2019 and analyzed the outcomes separately in the RIC and MAC cohorts based on the conditioning regimens used. Among 872 eligible patients, 493 underwent allo-HCT using RIC (Fludarabine/busulfan=166, Fludarabine/melphalan=327) and 379 using MAC (Fludarabine/busulfan=247, Busulfan/cyclophosphamide=132). In multivariable analysis with RIC, Fludarabine/melphalan was associated with inferior overall survival (HR 1.80, 95% CI 1.15-2.81, p=0.009), higher early non-relapse mortality (HR 1.81, 95% CI 1.12-2.91, p=0.01) and higher acute graft versus host disease (GVHD) (grade II-IV- HR 1.45, 95% CI 1.03-2.03, p=0.03; grade III-IV HR 2.21, 95%CI 1.28-3.83, p=0.004) compared to Fludarabine/busulfan. In the MAC setting, Busulfan/cyclophosphamide was associated with a higher acute GVHD (grade II-IV HR 2.33, 95% CI 1.67-3.25, p<0.001; grade III-IV HR 2.31, 95% CI 1.52-3.52, p<0.001) and inferior GVHD-free relapse-free survival (GRFS) (HR 1.94, 95% CI 1.49-2.53, p<0.001) as compared to Fludarabine/busulfan. Hence, our study suggests that Fludarabine/busulfan is associated with better outcomes in RIC (better overall survival, lower early non-relapse mortality, lower acute GVHD) and MAC (lower acute GVHD and better GRFS) in myelofibrosis.

Introduction:

Myelofibrosis is a chronic myeloproliferative neoplasm arising either de-novo (primary) or secondary to antecedent essential thrombocytosis or polycythemia vera. Despite the recent advances in disease biology and treatment options such as Janus Activating Kinase (JAK) inhibitors, allogeneic hematopoietic cell transplantation (allo-HCT) remains the only potentially curative option ¹⁻³. The availability of reduced intensity conditioning (RIC) and the choice of donors have expanded the scope of allo-HCT for these patients who are often older adults ⁴. While several factors influence outcomes of allo-HCT, conditioning intensity and conditioning regimen are aspects that could be tailored to improve the outcomes. Currently, both myeloablative conditioning (MAC) and RIC platforms are available for allo-HCT in myelofibrosis ⁵⁻¹². A large study from the European Group for Blood and Marrow Transplant (EBMT) compared the outcomes of allo-HCT with RIC versus MAC in myelofibrosis and demonstrated comparable results with both approaches, but better graft-versus-host disease (GVHD)-and relapse-free survival (GRFS) with MAC⁹. However, the optimal conditioning regimen either with RIC or MAC is not well known. While some studies have previously compared different RIC regimens with varying results ¹⁰⁻¹², similar comparative studies with MAC are lacking and no studies have demonstrated a survival difference based on the conditioning regimen. Hence, we sought to determine the outcomes of allo-HCT for myelofibrosis based on the choice of the conditioning regimen, separately with RIC and MAC.

Methods:

Study objective

Our objectives were to compare the overall survival, disease free survival, non-relapse mortality, relapse, incidence of acute GVHD, chronic GVHD and GRFS based on the choice of the conditioning regimen used with RIC or MAC.

Data source

CIBMTR is a combined research program of the Medical College of Wisconsin and the National Marrow Donor Program. It comprises a voluntary network of more than 450 transplantation centers worldwide that contribute data on consecutive allo-HCT to a centralized statistical center ¹³. Observational studies conducted by the CIBMTR are performed in compliance with all applicable federal regulations pertaining to the protection of human research participants. Patients provided written informed consent for research. The institutional review boards of the Medical College of Wisconsin and the National Marrow Donor Program approved this study.

Study population

Adults aged ≥18 years with a diagnosis of myelofibrosis (chronic phase) who underwent allo-HCT between the period 2008-2019 and data reported to the CIBMTR were identified. The cohort was then selected to focus on the most common conditioning regimens used in RIC [Fludarabine/busulfan vs. Fludarabine/melphalan] and MAC [Fludarabine/busulfan vs. Busulfan/cyclophosphamide] setting (Supplemental Figure S1). Conditioning regimens were classified in the CIBMTR dataset based on prior published data ^{14, 15}. The donor groups included matched related donors, 8/8 (HLA-A, -B, -C and -DRB1) matched unrelated donors and 7/8 matched unrelated donors. Key exclusion criteria were allo-HCT from haploidentical donor, syngeneic donor, cord

blood, and ex-vivo T-cell depleted or CD34 selected grafts. In addition, 51 patients in Fludarabine/busulfan MAC group who received post-transplant cyclophosphamide (post-Cy) were excluded as there were no such corresponding patients in Busulfan/cyclophosphamide MAC group.

Statistical analysis:

Baseline characteristics were summarized using descriptive statistics with median and range for continuous variables and proportions for categorical variables. Outcomes were compared separately in RIC and MAC cohorts based on the conditioning regimens. Definitions of the outcomes are provided in Supplement. Cumulative incidence estimates were calculated for competing risks outcomes including acute GVHD, chronic GVHD, non-relapse mortality, and relapse. Kaplan-Meier method was used to estimate the probabilities for survival. To evaluate for other relevant factors that could influence the outcomes, multivariable Cox regression analysis was used (see below for the variables included). The proportional hazards assumption was examined and covariates that violate the proportional hazards assumption were added as timedependent covariates. In the absence of binary endpoints, hazard ratio (HR) and confidence limits were reported. A pairwise comparison within the non-reference groups was also performed in multivariable models to demonstrate their effect and shown as contrasts. Variables included in multivariable analysis were age, race/ethnicity, disease subtype (primary vs. post ET or PV), dynamic international performance scoring system (DIPSS) score, hematopoietic cell transplantation comorbidity index (HCT-CI), karnofsky performance scale (KPS), systemic symptoms, splenic radiation, splenomegaly, interval from diagnosis to allo-HCT, ruxolitinib use pre-transplant, donor-

recipient HLA-match, gender match, CMV match, stem cell source, GVHD prophylaxis (tacrolimus based vs. cyclosporine based vs. post-Cy vs. others), use of antithymocyte globulin (ATG)/alemtuzumab, and year of transplant. A stepwise selection method was used to identify the final model with a significance level of 0.05 and only variables reaching that statistical significance were shown. In addition, adjusted univariate estimates were provided for outcomes that were significantly associated with conditioning regimen. Fine and Gray model was used for analysis of non-relapse mortality, GVHD and relapse ¹⁶. Center effect was tested using the score test proposed by Commenges and Andersen and marginal Cox models were used for further adjustments ¹⁷. Center effect was noted to be significant only for chronic GVHD and was adjusted accordingly. Missing category was included in the models as one group to avoid loss of data and power ¹⁸. All analyses were performed at a two-sided significance level of 0.05 using SAS 9.4 (SAS Institute, Cary, NC).

Results:

Baseline characteristics

Of 872 eligible patients, 493 underwent allo-HCT using RIC (Fludarabine/busulfan=166, Fludarabine/melphalan=327) and 379 using MAC (Fludarabine/busulfan=247, Busulfan/cyclophosphamide=132). Key baseline characteristics of the patients are summarized (Table 1, Supplemental Tables S1, S2; unadjusted univariate estimates in Tables S3, S4). In the RIC cohort, compared to Fludarabine/busulfan patients, Fludarabine/melphalan patients had longer median interval from diagnosis to allo-HCT (37 vs. 22 months, p=0.02), lower proportion with antithymocyte globulin/alemtuzumab

use (25% vs 52%, p<0.01), and higher proportion with pre-transplant ruxolitinib use (61% vs 49%, p=0.03). In MAC cohort, compared to Fludarabine/busulfan patients, Busulfan/cyclophosphamide patients had younger age (median age 55 vs 60 years p<0.01), higher proportion with low-intermediate risk disease (61% vs 54%, p=0.03), higher proportion with bone marrow graft (12% vs 4%, p<0.01), lower proportion with antithymocyte globulin/alemtuzumab use (5% vs 45%, p<0.01), and lower proportion with pre-transplant ruxolitinib use (43% vs 59%, p<0.01). Median follow-up of the cohort was 26 (3-150) months.

Overall survival

In multivariable analysis (Table 2), overall survival in RIC setting was significantly worse with Fludarabine/melphalan (HR 1.80, 95%CI 1.15-2.81, p=0.009, 2-year adjusted overall survival- 54.4% vs. 60.9%) as compared to Fludarabine/busulfan (Figure 1). In the MAC setting, overall survival was not significantly different between based on the conditioning regimen (Busulfan/cyclophosphamide- HR 1.14, 95%CI 0.75-1.71, p=0.54) (Figure 2). Other factors significantly associated with overall survival were donor-recipient HLA- match (higher risk with unrelated donors in MAC) and the use of antithymocyte globulin/alemtuzumab (higher risk in RIC) (Supplemental Tables S5, S6).

Disease free survival

In multivariable analysis (Table 2), disease free survival was not significantly different based on the conditioning regimen used in RIC (Fludarabine/melphalan- HR 1.03, 95%CI 0.77-1.38, p=0.85) or MAC (Busulfan/cyclophosphamide- HR 1.03, 95%CI 0.77-1.38, p=0.83) settings (Supplemental Figures S2 and S3). Other factors significantly

associated with disease free survival were Karnofsky performance status (higher risk with lower score in MAC) and pre-transplant ruxolitinib use (higher risk in MAC) (Supplemental Tables S5 and S6).

Non-relapse mortality

In RIC setting, there was a significantly higher risk of early non-relapse mortality with Fludarabine/melphalan as compared to Fludarabine/busulfan (17.4% vs. 4.3%, HR 1.81, 95%Cl 1.12-2.91, p=0.01). Beyond 6 months the risk of non-relapse mortality was low with fludarabine/melphalan (HR 0.46, 95%Cl 0.23-0.91, p=0.02) (Table 2, Supplemental Figure S4) (Cut-off of 6 months was chosen due to non-proportional hazard). No significant differences in non-relapse mortality were seen with MAC based on the conditioning regimens (Busulfan/cyclophosphamide- HR 1.36, 95%Cl 0.83-2.21, p=0.22) (Supplemental Figure S5). The other factor significantly associated with non-relapse mortality was donor-recipient HLA-match (higher risk with unrelated donors in MAC) (Supplemental Tables S5, S6).

<u>Relapse</u>

The risk of relapse was not significantly different based on the conditioning regimen used in RIC or MAC (RIC- Fludarabine/melphalan HR 0.85, 95%Cl 0.64-1.12, p=0.25; MAC - Busulfan/cyclophosphamide HR 0.92, 95%Cl 0.64-1.32, p=0.65) (Supplemental Figures S6, S7, Supplemental Tables S5, S6). Other factors significantly associated with relapse were karnofsky performance status (higher risk with poor score in MAC), pre-transplant ruxolitinib use (higher risk in MAC) and year of transplant (higher risk with recent period in RIC).

<u>GVHD</u>

In the RIC setting, Fludarabine/melphalan was associated with a significantly higher risk of acute GVHD grade II-IV (Fludarabine/melphalan 40%, Fludarabine/busulfan 35.3%, HR 1.45, 95%CI 1.03-2.03, p=0.03) and grade III-IV (Fludarabine/melphalan 21.8%, Fludarabine/busulfan 12.1%, HR 2.21, 95%CI 1.28-3.83, p=0.004) (Supplemental Figures S8, S9). In the MAC setting, Busulfan/cyclophosphamide was associated with a significantly higher risk of acute GVHD grade II-IV (Busulfan/cyclophosphamide 58.9%, Fludarabine/busulfan 34.4%; HR 2.33, 95%CI 1.67-3.25, p<0.001) and grade III-IV (Busulfan/cyclophosphamide 32.6%, Fludarabine/busulfan 11.9%; HR 2.31, 95%CI 1.52-3.52, p<0.001) (Supplemental Figures S10, S11). Chronic GVHD was significantly associated with donor-recipient HLA-match (higher risk with 7/8 matched unrelated donors in RIC) and pre-transplant ruxolitinib use (lower risk in MAC), but not by the conditioning regimen (Supplemental Tables S5, S6).

<u>GRFS:</u>

In the RIC setting, GRFS was not significantly different between Fludarabine/busulfan and Fludarabine/melphalan (HR 1.11, 95%Cl 0.90-1.35, p=0.32) (Supplemental Figure S12). However, in the MAC setting, Busulfan/cyclophosphamide was associated with significantly inferior GRFS (HR 1.94, 95%Cl 1.49-2.53, p<0.01) (2-year adjusted probability 5.1% vs. 19.4%) as compared to Fludarabine/busulfan (Table 2, Figure 3). Other factors significantly associated with GRFS included recipient age (in MAC) and donor-recipient HLA-match (higher risk with unrelated donors in MAC) (Supplemental Tables S5 and S6).

Engraftment:

The rates of neutrophil engraftment (30 days) were significantly better with Fludarabine/busulfan in RIC (Fludarabine/busulfan 95.1% vs. Fludarabine/melphalan 92.4%, p=0.006) and MAC (Fludarabine/busulfan 95.2% vs. Busulfan/cyclophosphamide 87.2%, p=0.02). The rate of platelet engraftment (100 days) was better with Fludarabine/busulfan in RIC setting (RIC - Fludarabine/busulfan 84.4% vs Fludarabine/melphalan 73.9%, p<0.001; MAC - Fludarabine/busulfan 86.1% vs. Busulfan/cyclophosphamide 83.7%, p=0.27).

Additional analyses:

In the RIC cohort, we investigated whether the outcomes differed based on the dose of melphalan (100 vs 140mg/m²) used in Fludarabine/melphalan group. As shown in Supplemental Table S7, the outcomes did not significantly vary based on the dose of melphalan (shown as contrasts between melphalan 100 vs 140mg/m²).

Discussion:

Our study highlights the significant differences in outcomes of allo-HCT for myelofibrosis based on the choice of the conditioning regimen. Fludarabine-busulfan conditioning was associated with superior overall survival, lower early non-relapse mortality and lower acute GVHD (all with RIC), and lower acute GVHD and superior GRFS with MAC. A key aspect of conditioning strategy is its ability be tailored in order to improve the outcomes. Events such as non-relapse mortality and GVHD that affect the morbidity and mortality after allo-HCT could be influenced by the conditioning strategy and efforts to minimize

these complications are vital to improve the long-term success. Although RIC and MAC platforms are clinically decided based on factors such as age, comorbidities, performance status, and other aspects that are often not modifiable, our results illustrate the influence of common conditioning regimens used in these settings and provides valuable information for choosing the appropriate regimen in clinical practice.

Prior retrospective studies have evaluated the impact of conditioning intensity and regimen in myelofibrosis, albeit with variable results and certain key differences compared to our study ⁵⁻¹². A study by Robin et al. included 160 patients with myelofibrosis from 2 European centers [Paris (Fludarabine-Busulfan) or Hamburg (Fludarabine-Melphalan)], but with antithymocyte globulin given for all patients who received Fludarabine-Busulfan conditioning¹¹. Another CIBMTR study by Gupta et al. included only patients with primary myelofibrosis and RIC (Fludarabine-TBI vs Fludarabine-Melphalan vs. Fludarabine-Busulfan) between 1997-2010 with a relatively younger patient population (median age 55 years)¹⁰. Hence, the differences in the study population, the nature of the cohort (registry vs. individual center based), treatment received and variations in time period included could have contributed to the differences in results noted between the current study and prior studies. To date, prospective studies of conditioning regimen in myelofibrosis are single arm or comparative studies with smaller sample size ^{19, 20, 21}. For example, a phase II study by Patriarca et al. prospectively compared fludarabine-busulfan and fludarabine-thiotepa for allo-HCT in sixty patients with myelofibrosis and showed similar outcomes with both these regimens ²¹. Hence, our study addresses the knowledge gap in this area using a

larger dataset with a comparison of commonly reported conditioning regimens. Unfortunately, due to the limited number of patients receiving other less common conditioning regimens such as fludarabine-thiotepa, these regimens could not be compared in our study. Additionally, given the results of a large EBMT study showing no difference in overall survival between MAC and RIC ⁹, we did not compare the outcomes of MAC versus RIC in our analysis which also helped to minimize the heterogeneity in comparisons.

Apart from the conditioning regimen, factors such as donor-recipient HLA-match, performance status and use of antithymocyte globulin/alemtuzumab influenced the outcomes similar to prior studies. The imbalances in baseline characteristics were adjusted in multivariable models and there were no significant interactions noted between the baseline characteristics and main effect (conditioning regimen). Antithymocyte globulin/alemtuzumab was associated with worse overall survival in RIC and was more commonly used with Fludarabine/busulfan regimens. Despite this, an early survival advantage was noted with Fludarabine/busulfan in RIC. The association between the outcomes and factors such as the route of busulfan administration (oral vs. intravenous, targeted vs. non-targeted; data not shown) and the dose of melphalan (in RIC) were also investigated and none was found. In MAC, ruxolitinib prior to allo-HCT was associated with higher risk of relapse, inferior disease-free survival, higher risk of acute GVHD and lower risk of chronic GVHD. Although prior studies indicate the feasibility and safety of ruxolitinib therapy prior to allo-HCT^{22, 23}, we could not evaluate the possible mechanisms behind these differences due to limited information on the

duration, dose, response, and other aspects of ruxolitinib therapy. Other factors such as the role of splenectomy, spleen size or splenic radiation therapy and their association with outcomes could not be evaluated due to the small number of patients with those interventions.

Despite the large sample size, our study is limited by the retrospective design and lack in-depth information on factors such as genomic mutations and therapies for myelofibrosis given pre- and post- allo-HCT that could affect the outcomes ^{24, 25}. The lack of detailed information on genomic mutations precluded further analyses and calculation of molecular risk scores (such as MIPSS70, MYSEC-PM etc). For example, study by Gagelman et al. investigated the prognostic significance of somatic mutations in myelofibrosis patients undergoing allo-HCT and identified that ASXL1 and non-CALR/MPL driver mutations were associated with poor outcomes. This study also established a prognostic model with variables such as patient age, performance status, white blood count, platelet count, HLA-mismatched donor and molecular mutations. However, due to the lack of information on these aspects, we could not apply this scoring system in our study ²⁴. We also could not assess the reasons behind the choice of individual conditioning regimens used for these patients, understanding that centers could have their preferences while choosing conditioning regimens. However, we evaluated for center-effects in multivariable analyses and adjustments were made accordingly. As our study mainly focused on patients with chronic phase myelofibrosis, the role of conditioning strategy in advanced phase disease (accelerated/blast phase) was not evaluated. Due to the nature of the GVHD reporting in the dataset, chronic

GVHD was analyzed as a whole outcome without further stratification (mild, moderate, severe).

Our study demonstrates that Fludarabine/busulfan based conditioning is associated with superior overall survival, lower early non-relapse mortality, and lower acute GVHD with RIC and lower acute GVHD and superior GRFS with MAC. The results provide valuable information for tailoring the conditioning strategies to minimize non-relapse mortality and GVHD and improve survival. Prospective comparative studies are warranted to confirm these results and identify the ideal conditioning regimen in myelofibrosis.

References

1. Tefferi A. Primary myelofibrosis: 2021 update on diagnosis, risk-stratification and management. Am J Hematol. 2021;96(1):145-162.

2. Tefferi A, Pardanani A. Myeloproliferative Neoplasms: A Contemporary Review. JAMA Oncol. 2015;1(1):97-105.

3. Harrison C, Kiladjian JJ, Al-Ali HK, et al. JAK inhibition with ruxolitinib versus best available therapy for myelofibrosis. N Engl J Med. 2012;366(9):787-798.

4. Phelan R, Arora M, Chen M. Current use and outcome of hematopoietic stem cell transplantation: CIBMTR US summary slides, 2020

5. Hernández-Boluda JC, Pereira A, Kröger N, et al. Determinants of survival in myelofibrosis patients undergoing allogeneic hematopoietic cell transplantation. Leukemia. 2021;35(1):215-224.

 Robin M, de Wreede LC, Wolschke C, et al. Long-term outcome after allogeneic hematopoietic cell transplantation for myelofibrosis. Haematologica. 2019;104(9):1782-1788.

7. Gowin K, Ballen K, Ahn KW, et al. Survival following allogeneic transplant in patients with myelofibrosis. Blood Adv. 2020;4(9):1965-1973.

8. Ballen KK, Shrestha S, Sobocinski KA, et al. Outcome of transplantation for myelofibrosis. Biol Blood Marrow Transplant. 2010;16(3):358-367.

McLornan D, Szydlo R, Koster L, et al. Myeloablative and Reduced-Intensity
 Conditioned Allogeneic Hematopoietic Stem Cell Transplantation in Myelofibrosis: A
 Retrospective Study by the Chronic Malignancies Working Party of the European
 Society for Blood and Marrow Transplantation. Biol Blood Marrow Transplant.
 2019;25(11):2167-2171.

10. Gupta V, Malone AK, Hari PN, et al. Reduced-intensity hematopoietic cell transplantation for patients with primary myelofibrosis: a cohort analysis from the center for international blood and marrow transplant research. Biol Blood Marrow Transplant. 2014;20(1):89-97.

11. Robin M, Porcher R, Wolschke C, et al. Outcome after Transplantation According to Reduced-Intensity Conditioning Regimen in Patients Undergoing Transplantation for Myelofibrosis. Biol Blood Marrow Transplant. 2016;22(7):1206-1211.

12. Jain T, Kunze KL, Temkit M, et al. Comparison of reduced intensity conditioning regimens used in patients undergoing hematopoietic stem cell transplantation for myelofibrosis. Bone Marrow Transplant. 2019;54(2):204-211.

13. Horowitz M. The role of registries in facilitating clinical research in BMT: Examples from the Center for International Blood and Marrow Transplant Research. Bone Marrow Transplant. 2008;42 Suppl 1:S1-S2.

14. Bacigalupo A, Ballen K, Rizzo D, et al. Defining the intensity of conditioning regimens: working definitions. Biol Blood Marrow Transplant. 2009;15(12):1628-1633.

15. Giralt S, Ballen K, Rizzo D, et al. Reduced-intensity conditioning regimen workshop: defining the dose spectrum. Report of a workshop convened by the center for international blood and marrow transplant research. Biol Blood Marrow Transplant. 2009;15(3):367-369.

16. Fine JP, Gray RJ. A proportional hazards model for the subdistribution of a competing risk. J Am Stat Assoc. 1999;94:496-509.

17. Commenges D, Andersen PK. Score test of homogeneity for survival data. Lifetime Data Anal. 1995;1(2):145-156.

18. Groenwold RHH, Dekkers OM. Missing data: the impact of what is not there. Eur J Endocrinol. 2020;183(4):E7-E9.

19. Rondelli D, Goldberg JD, Isola L, et al. MPD-RC 101 prospective study of reducedintensity allogeneic hematopoietic stem cell transplantation in patients with myelofibrosis. Blood. 2014;124(7):1183-1191.

20. Kroger N, Holler E, Kobbe G, et al. Allogeneic stem cell transplantation after reduced-intensity conditioning in patients with myelofibrosis: a prospective, multicenter study of the Chronic Leukemia Working Party of the European Group for Blood and Marrow Transplantation. Blood. 2009;114(26):5264-5270.

21. Patriarca F, Masciulli A, Bacigalupo A, et al. Busulfan- or Thiotepa-Based Conditioning in Myelofibrosis: A Phase II Multicenter Randomized Study from the GITMO Group. Biol Blood Marrow Transplant. 2019;25(5):932-940. 22. Shanavas M, Popat U, Michaelis LC, et al. Outcomes of Allogeneic Hematopoietic Cell Transplantation in Patients with Myelofibrosis with Prior Exposure to Janus Kinase 1/2 Inhibitors. Biol Blood Marrow Transplant. 2016;22(3):432-440.

23. Kröger N, Sbianchi G, Sirait T, et al. Impact of prior JAK-inhibitor therapy with ruxolitinib on outcome after allogeneic hematopoietic stem cell transplantation for myelofibrosis: a study of the CMWP of EBMT. Leukemia. 2021;35(12):3551-3560.

24. Gagelmann N, Ditschkowski M, Bogdanov R, et al. Comprehensive clinicalmolecular transplant scoring system for myelofibrosis undergoing stem cell transplantation. Blood. 2019;133(20):2233-2242.

25. Tamari R, Rapaport F, Zhang N, et al. Impact of High-Molecular-Risk Mutations on Transplantation Outcomes in Patients with Myelofibrosis. Biol Blood Marrow Transplant. 2019;25(6):1142-1151.

26. Przepiorka D, Weisdorf D, Martin P, et al. 1994 Consensus Conference on Acute GVHD Grading. Bone Marrow Transplant. 1995;15(6):825-828.

27. Sullivan KM, Shulman HM, Storb R, et al. Chronic graft-versus-host disease in 52 patients: adverse natural course and successful treatment with combination immunosuppression. Blood. 1981;57(2):267-276.

Characteristic	Reduced intensity conditioning			Myeloablative conditioning		
	Flu/Bu (n=166)	Flu/Mel (n=327)	P- Value	Flu/Bu (n=247)	Bu/Cy (n=132)	P- value
Age	63 (44-75)	63 (38- 78)	0.88	60 (27-74)	55 (24-67)	<0.01*
Disease type			0.22			0.85
Primary	132 (80)	242 (74)		191 (77)	100 (76)	
Post ET	14 (8)	45 (14)		20 (8)	13 (10)	
Post PV	20 (12)	40 (12)		36 (15)	19 (14)	
Median time from diagnosis to HCT (months, range)	22 (3-393)	37 (3- 594)	0.02*	25 (2-490)	38 (3-377)	0.41
DIPSS Score - no. (%)			0.07			0.03*
Low/Intermediate-1	71 (43)	107 (33)		134 (54)	80 (61)	
Intermediate-2/High	69 (42)	168 (51)		93 (38)	34 (26)	
Missing	26 (16)	52 (16)		20 (8)	18 (14)	
Donor type - no. (%)			0.75			0.15
HLA-identical sibling	48 (29)	94 (29)		79 (32)	53 (40)	
8/8-matched unrelated	107 (64)	205 (63)		142 (57)	62 (47)	
7/8 matched unrelated	11 (7)	28 (9)		26 (11)	17 (13)	
ATG/Alemtuzumab use			<0.01*			<0.01*
- no. (%)						
No	79 (48)	246 (75)		135 (55)	125 (95)	
Yes	87 (52)	81 (25)		112 (45)	7 (5)	
Graft type - no. (%)			0.84			<0.01*
Bone marrow	6 (4)	13 (4)		11 (4)	16 (12)	
Peripheral blood	160 (96)	314 (96)		236 (96)	116 (88)	
Pre-transplant			0.03*			<0.01*
ruxolitinib						
No	84 (51)	125 (38)		101 (41)	75 (57)	
Yes	82 (49)	201 (61)		146 (59)	57 (43)	
Missing	0	1		0)	0 ` ´	

Table 1: Key Baseline characteristics

*p<0.05-significant

Flu-Fludarabine; Bu-Busulfan; Mel- Melphalan; Cy- Cyclophosphamide; ET- Essential thrombocytosis; PV- polocythemia vera; HCT-hematopoietic cell transplantation; DIPSS- dynamic international prognostic scoring system; ATG – antithymocyte globulin

Reduced intensity conditioning				Myeloablative conditioning			
Outcome	HR	95% CI	p- value	Outcome	HR	95% CI	p-value
Overall survival** ≤ 6 months Flu/Bu Flu/Mel	1.00 1.80	Ref. 1.15-2.81	0.009*	Overall survival Flu/Bu Bu/Cy	1.00 1.14	Ref. 0.75-1.71	0.54
> 6 months Flu/Bu Flu/Mel	1.00 0.82	Ref. 0.53-1.26	0.35				
Disease free survival** ≤ 6 months Flu/Bu Flu/Mel > 6 months Flu/Bu	1.00 1.03 1.00	Ref. 0.77-1.38 Ref.	0.85	Disease free survival Flu/Bu Bu/Cy	1.00 1.03	Ref. 0.77-1.38	0.83
Flu/Mel	0.95	0.68-1.34					
NRM** ≤ 6 months Flu/Bu Flu/Mel > 6 months	1.00 1.81	Ref. 1.12-2.91	0.01*	NRM Flu/Bu Bu/Cy	1.00 1.36	Ref. 0.83- 2.21	0.22
Flu/Bu Flu/Mel	1.00 0.46	Ref. 0.23-0.91	0.02*				
Relapse Flu/Bu Flu/Mel	1.00 0.85	Ref. 0.64-1.12	0.25	Relapse Flu/Bu Bu/Cy	1.00 0.92	Ref. 0.64-1.32	0.65
Acute GVHD grade II-IV** ≤ 2 months Flu/Bu Flu/Mel	1.00 1.45	Ref. 1.03-2.03	0.03*	Acute GVHD grade II-IV** ≤ 2 months Flu/Bu Bu/Cy	1.00	Ref. 1.67-3.25	<0.001*
> 2 months Flu/Bu Flu/Mel	1.00 0.71	Ref. 0.43-1.17	0.18	> 2 months Flu/Bu Bu/Cy	1.00 0.88	Ref. 0.46-1.68	0.69
Acute GVHD grade III-IV** ≤ 2 months Flu/Bu Flu/Mel > 2 months	1.00 2.21	Ref. 1.28-3.83	0.004*	Acute GVHD grade III-IV Flu/Bu Bu/Cy	1.00 2.31	Ref. 1.52-3.52	<0.001*
Flu/Bu Flu/Mel	1.00 0.89	Ref. 0.64-1.24	0.48				

Table 2: Multivariable analysis of outcomes based on conditioning regimen

Chronic GVHD Flu/Bu Flu/Mel	1.00 0.91	Ref. 0.67-1.25	0.55	Chronic GVHD Flu/Bu Bu/Cy	1.00	Ref. 0.80-1.84	0.36
GRFS Flu/Bu Flu/Mel	1.00 1.11	Ref. 0.90-1.35	0.32	GRFS Flu/Bu Bu/Cy	1.00 1.94	Ref. 1.49-2.53	<0.001*

*p<0.05-significant **Outcomes separated by timepoints due to non-proportional hazard

Flu-Fludarabine; Bu-Busulfan; Mel- Melphalan; Cy- Cyclophosphamide; NRM – non-relapse mortality; GVHD- graft versus host disease; GRFS – GVHD free relapse free survival

Figure legends:

Figure 1: Overall survival with reduced intensity conditioning

Figure 2: Overall survival with myeloablative conditioning

Figure 3: GVHD-free Relapse-Free survival with myeloablative conditioning

Figure 1: Overall Survival with RIC

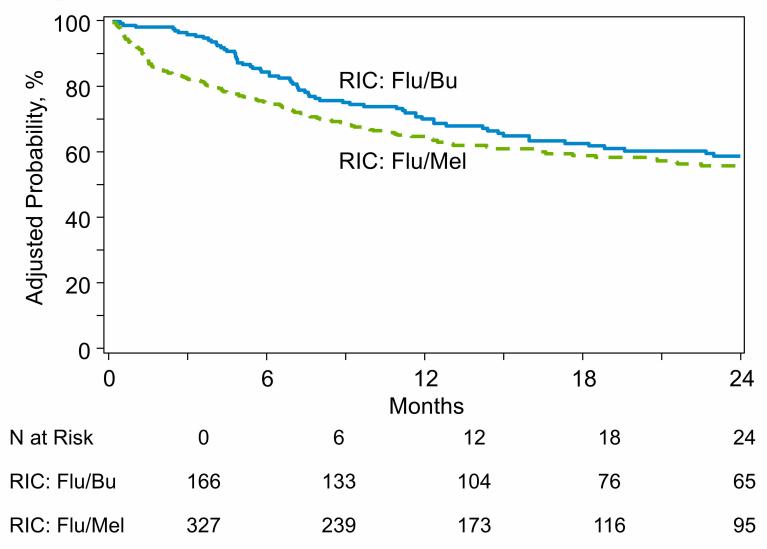


Figure 2: Overall Survival with MAC

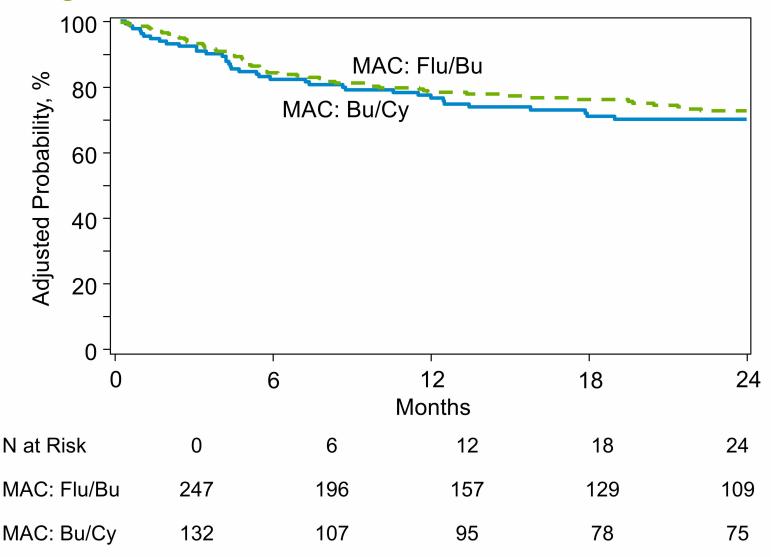
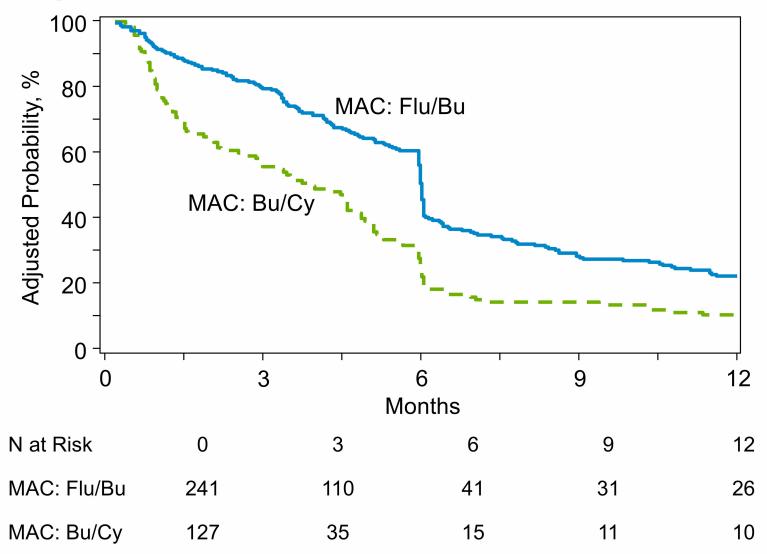


Figure 3- GVHD- and Relapse-Free Survival with MAC



Supplement:

Definition of outcomes:

Disease free survival was defined as time from allo-HCT to treatment failure (relapse or death from any cause).

Overall survival was defined as time from allo-HCT to death from any cause.

Acute GVHD ²⁶ and chronic GVHD ²⁷ were graded per standard criteria.

Relapse was defined as disease recurrence as reported by the centers to CIBMTR.

Non-relapse mortality was defined as death from any cause in the first 28 days after allo-HCT or death without evidence of disease recurrence beyond day 28.

GVHD-free relapse-free survival was defined as time from allo-HCT to death, post-transplant relapse, chronic GVHD requiring immunosuppressive treatment, or severe acute GVHD (aGVHD III-IV).

Characteristic	RIC: Flu/Bu F	RIC: Flu/Mel	P Value	
No. of patients	166	327		
Patient age - median (min-max)	63 (44-75)	63 (38-78)	0.88 ^a	
Age group - no. (%)			0.50 ^b	
30-39	0 (0)	1 (0)		
40-49	10 (6)	31 (9)		
50-59	43 (26)	77 (24)		
60-69	99 (60)	182 (56)		
>= 70	14 (8)	36 (11)		
Sex - no. (%)			0.40 ^b	
Male	105 (63)	194 (59)		
Female	61 (37)	133 (41)		
Region - no. (%)			<.01 ^b	
US	146 (88)	310 (95)		
Europe	12 (7)	2 (1)		
Australia/New Zealand	8 (5)	15 (5)		
Race/ethnicity - no. (%)			0.16 ^b	
Hispanic	3 (2)	17 (5)		
Non-Hispanic White	150 (90)	274 (84)		
Non-Hispanic Black/African American	4 (2)	9 (3)		
Other	7 (4)	14 (4)		
Missing	2 (1)	13 (4)		
HCT-CI - no. (%)			0.13 ^b	
0	23 (14)	72 (22)		
1	22 (13)	34 (10)		
2	29 (17)	52 (16)		
3	34 (20)	71 (22)		
4	24 (14)	40 (12)		
5	16 (10)	17 (5)		
6+	11 (7)	33 (10)		
Missing	7 (4)	8 (2)		
Karnofsky performance score - no. (%)			0.37 ^b	
90-100	79 (48)	152 (46)		
< 90	85 (51)	164 (50)		
Missing	2 (1)	11 (3)		
Disease type- no. (%)			0.22 ^b	
Primary Myelofibrosis	132 (80)	242 (74)		

Supplemental Table S1. Baseline characteristics with reduced intensity conditioning

Characteristic	RIC: Flu/Bu R	RIC: Flu/Bu RIC: Flu/Mel		
Post polycythemia vera	14 (8)	45 (14)		
Post Essential thrombocythemia	20 (12)	40 (12)		
Time from diagnosis to HCT - median (min-max)	22 (3-393) 3	57 (-99-594)	0.02 ^a	
Time from diagnosis to HCT - no. (%)			0.09 ^b	
<6 months	32 (19)	44 (13)		
6-11 months	34 (20)	50 (15)		
>=12 months	100 (60)	232 (71)		
Missing	0 (0)	1 (0)		
DIPSS Score - no. (%)			0.07 ^b	
Low/Intermediate-1	71 (43)	107 (33)		
Intermediate-2/High	69 (42)	168 (51)		
Missing	26 (16)	52 (16)		
Splenomegaly at HCT - no. (%)			0.21 ^b	
No	66 (40)	107 (33)		
Yes	82 (49)	168 (51)		
Splenectomy	8 (5)	16 (5)		
Missing	10 (6)	36 (11)		
Systemic symptoms prior to HCT no. (%)			0.19 ^b	
No	137 (83)	252 (77)		
Yes	19 (11)	58 (18)		
Missing	10 (6)	17 (5)		
Donor type - no. (%)			0.75 ^b	
HLA-identical sibling	48 (29)	94 (29)		
8/8 matched unrelated donor	107 (64)	205 (63)		
7/8 matched unrelated donor	11 (7)	28 (9)		
Donor/recipient sex match - no. (%)			0.81 ^b	
Male-Male	71 (43)	135 (41)		
Male-Female	39 (23)	81 (25)		
Female-Male	34 (20)	58 (18)		
Female-Female	21 (13)	52 (16)		
Missing	1 (1)	1 (0)		
Donor/recipient CMV serostatus - no. (%)			0.73 ^b	
+/+	54 (33)	102 (31)		
+/-	25 (15)	41 (13)		
-/+	39 (23)	70 (21)		
-/-	47 (28)	111 (34)		
Missing	1 (1)	3 (1)		
Graft type - no. (%)		. ,	0.84 ^b	

	Characteristic	RIC: Flu/Bu l	RIC: Flu/Mel	P Value
GVHD prophylaxis - no. (%) <.01 ^b Post-CY + other(s) 7 (4) 9 (3) TAC + MMF +- other(s) (except post-CY) 28 (17) 36 (11) TAC + MTX +- other(s) (except MMF, post-CY) 86 (52) 207 (63) TAC + other(s) (except MMF, MTX, post-CY) 7 (4) 36 (11) TAC + other(s) (except MMF, MTX, post-CY) 7 (4) 36 (11) TAC + other(s) (except MMF, post-CY) 23 (14) 24 (7) CSA + MMF +- other(s) (except MMF, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, MTX, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, MTX, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, MTX, post-CY) 0 (0) 1 (0) ATG/alemtuzumab use - no. (%) 6(4) 1 (0) ATG/alemtuzumab use - no. (%) <01 ^b <01 ^b No 79 (48) 246 (75) <01 ^b ATG alone 86 (52) 72 (22) Alemtuzumab alone 1 (1) 9 (3) <04 ^c ATG alone 86 (52) 72 (22) <01 ^a ATG dose (mg) - median (min-max) 400 (5-4900) 284 (4-2988) <td>Bone marrow</td> <td>6 (4)</td> <td>13 (4)</td> <td></td>	Bone marrow	6 (4)	13 (4)	
Post-CY + other(s) 7 (4) 9 (3) TAC + MMF +- other(s) (except post-CY) 28 (17) 36 (11) TAC + MTX +- other(s) (except MMF, post-CY) 86 (52) 207 (63) TAC + other(s) (except MMF, MTX, post-CY) 7 (4) 36 (11) TAC alone 4 (2) 6 (2) CSA + MMF +- other(s) (except post-CY) 4 (2) 2 (1) CSA + MTX +- other(s) (except post-CY) 23 (14) 24 (7) CSA + other(s) (except MMF, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, mTX, post-CY) 0 (0) 1 (0) ATG State 2 (4) 2 (2) Atter (s) Caste 7 (2) (0) ATG	Peripheral blood	160 (96)	314 (96)	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	GVHD prophylaxis - no. (%)			<.01 ^b
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Post-CY + other(s)	7 (4)	9 (3)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	TAC + MMF +- other(s) (except post-CY)	28 (17)	36 (11)	
TAC alone 4 (2) 6 (2) CSA + MMF +- other(s) (except post-CY) 4 (2) 2 (1) CSA + MTX +- other(s) (except MMF, post-CY) 23 (14) 24 (7) CSA + other(s) (except MMF, MTX, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, MTX, post-CY) 0 (0) 1 (0) CSA + other(s) (except MMF, MTX, post-CY) 0 (0) 1 (0) CSA + other(s) 6 (4) 1 (0) At other(s) 6 (4) 1 (0) ATG/alemtuzumab use - no. (%) <.01 ^b No 79 (48) 246 (75) Yes 87 (52) 81 (25) ATG alone 86 (52) 72 (22) Alemtuzumab alone 1 (1) 9 (3) No ATG or alemtuzumab 79 (48) 246 (75) ATG dose (mg) - median (min-max) 400 (5-4900) 284 (4-2988) <.01 ^a 0-5mg 2 (2) 4 (6) 0.43 ^b 0-5mg 2 (2) 4 (6) 0.43 ^b 0-5mg 2 (2) 4 (6) 0.43 ^b 0 75 (84) 80 0.43 ^b Busulfan Administration - no. (%) 7 (4) <td< td=""><td>TAC + MTX +- other(s) (except MMF, post-CY)</td><td>86 (52)</td><td>207 (63)</td><td></td></td<>	TAC + MTX +- other(s) (except MMF, post-CY)	86 (52)	207 (63)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	TAC + other(s) (except MMF, MTX, post-CY)	7 (4)	36 (11)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	TAC alone	4 (2)	6 (2)	
$\begin{array}{cccc} {\rm CSA} + {\rm other}({\rm s}) ({\rm except} {\rm MMF}, {\rm MTX}, {\rm post-CY}) & 0 (0) & 1 (0) \\ {\rm CSA} {\rm alone} & 1 (1) & 5 (2) \\ {\rm Other}({\rm s}) & 6 (4) & 1 (0) \\ \\ {\rm ATG}/{\rm alemtuzumab} {\rm use} - {\rm no}. (\%) & < {\rm O1}^{\rm b} \\ {\rm No} & 79 (48) & 246 (75) \\ {\rm Yes} & 87 (52) & 81 (25) \\ \\ {\rm ATG}/{\rm Alemtuzumab} - {\rm no}. (\%) & < {\rm O1}^{\rm b} \\ {\rm ATG} {\rm alone} & 86 (52) & 72 (22) \\ {\rm Alemtuzumab} {\rm alone} & 1 (1) & 9 (3) \\ {\rm No} {\rm ATG} {\rm or} {\rm alemtuzumab} & 79 (48) & 246 (75) \\ \\ {\rm ATG} {\rm dose} ({\rm mg}) - {\rm median} ({\rm min-max}) & 400 (5-4900) 284 (4-2988) \\ {\rm O-5mg} & 2 (2) & 4 (6) \\ {\rm 5-7.5mg} & 2 (2) & 0 (0) \\ {\rm 5-7.5mg} & 2 (2) & 0 (0) \\ {\rm 5-7.5mg} & 78 (91) & 65 (90) \\ \\ {\rm Melphalan} {\rm dose} - {\rm no}. (\%) & & & & & & \\ \\ {\rm Melphalan} {\rm dose} - {\rm no}. (\%) & & & & & & & \\ \\ {\rm Melphalan} {\rm domg/m}^2 & - 52 (16) \\ \\ {\rm Melphalan} {\rm domg/m}^2 & - 275 (84) \\ \\ \\ \\ \\ \\ \\ {\rm Busulfan} {\rm Administration} - {\rm no}. (\%) & & & & & & \\ \\ {\rm No} & 84 (51) 125 (38) \\ {\rm Yes} & 82 (49) 201 (61) \\ \\ \\ \\ {\rm Missing} & 0 (0) & 1 (0) \\ \\ \end{array} \right $	CSA + MMF +- other(s) (except post-CY)	4 (2)	2 (1)	
CSA alone1 (1)5 (2)Other(s)6 (4)1 (0)ATG/alemtuzumab use - no. (%)<.01b	CSA + MTX +- other(s) (except MMF, post-CY)	23 (14)	24 (7)	
Other(s) $6(4)$ $1(0)$ ATG/alemtuzumab use - no. (%) <01b No 79 (48) 246 (75) Yes 87 (52) 81 (25) ATG/Alemtuzumab - no. (%) <01b ATG alone 86 (52) 72 (22) Alemtuzumab alone 1 (1) 9 (3) 0.316^{-10} ATG dose (mg) - median (min-max) 400 (5-4900) 284 (4-2988) <0.13^{-10} ATG dose (mg) - no. (%) $2 (2)$ 4 (6) $5.7.5mg$ $2 (2)$ 0.43^{-10} $0.5mg$ $2 (2)$ 0.01^{-10} 0.43^{-10} 0.43^{-10} 0.43^{-10} $0.5mg$ $2 (2)$ 0.01^{-10} 0.43^{-10} 0.43^{-10} 0.43^{-10} $0.5mg$ $2 (2)$ 0.01^{-10} $0.4(5)$ $3 (4)$ $0.4(5)$ $0.4(6)$ $0.57.5mg$ $2 (2)$ 0.01^{-10} 0.32^{-10} 0.32^{-10} $0.4(5)$ $0.4(6)$ $0.57.5mg$ $2 (2)^{-10} (1)^{-10}$ 0.32^{-10} 0.32^{-10} 0.32^{-10} Melphalan 100mg/m ² $-52 (16)^{-10}$ 0.33^{-10} 0.33^{-10} 0.33^{-10} $0.33^$	CSA + other(s) (except MMF, MTX, post-CY)	0 (0)	1 (0)	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	CSA alone	1 (1)	5 (2)	
No79 (48)246 (75)Yes $87 (52)$ $81 (25)$ ATG/Alemtuzumab - no. (%)<.01b	Other(s)	6 (4)	1 (0)	
Yes $87 (52)$ $81 (25)$ ATG/Alemtuzumab - no. (%)<01b	ATG/alemtuzumab use - no. (%)			<.01 ^b
ATG/Alemtuzumab - no. (%) <.01 ^b ATG alone 86 (52) 72 (22) Alemtuzumab alone 1 (1) 9 (3) No ATG or alemtuzumab 79 (48) 246 (75) ATG dose (mg) - median (min-max) 400 (5-4900) 284 (4-2988) <.01 ^a 0-5mg 2 (2) 4 (6) 0.43 ^b 0-5mg 2 (2) 0 (0) >75.mg 0-5mg 2 (2) 0 (0) >75.mg 78 (91) 65 (90) Missing 4 (5) 3 (4) Melphalan 100mg/m ² - 52 (16) 0.03 ^b Melphalan 140mg/m ² - 52 (16) 0.03 ^b No 7 (4) - 0.03 ^b No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0)	No	79 (48)	246 (75)	
ATG alone $86 (52)$ $72 (22)$ Alemtuzumab alone $1 (1)$ $9 (3)$ No ATG or alemtuzumab $79 (48)$ $246 (75)$ ATG dose (mg) - median (min-max) $400 (5-4900)$ $284 (4-2988)$ $<.01^a$ ATG dose (mg) - no. (%) $2 (2)$ $4 (6)$ 0.43^b 0-5mg $2 (2)$ $4 (6)$ $5-7.5mg$ $2 (2)$ $0 (0)$ >7.5mg $78 (91)$ $65 (90)$ $Missing$ $4 (5)$ $3 (4)$ Melphalan dose - no. (%) $Melphalan 100mg/m^2$ $ 52 (16)$ Melphalan 140mg/m² $ 275 (84)$ 0.03^b Busulfan Administration - no. (%) 0.03^b $0 (0)$ 0.03^b No $84 (51)$ $125 (38)$ $7es$ $82 (49)$ $201 (61)$ No $84 (51)$ $125 (38)$ $7es$ $82 (49)$ $201 (61)$ Missing $0 (0)$ $1 (0)$ 0.36^b	Yes	87 (52)	81 (25)	
Alemtuzumab alone 1 (1) 9 (3) No ATG or alemtuzumab 79 (48) 246 (75) ATG dose (mg) - median (min-max) 400 (5-4900) 284 (4-2988) <.01 ^a ATG dose (mg) - no. (%) 0.43 ^b 0.43 ^b 0-5mg 2 (2) 4 (6) 0.43 ^b 0-5mg 2 (2) 0 (0) >7.5mg 2 (2) 0 (0) >7.5mg 2 (2) 0 (0) >7.5mg 3 (4) Melphalan dose - no. (%) 4 (5) 3 (4) Melphalan 100mg/m ² - 52 (16) 4 (5) Melphalan 140mg/m ² - 275 (84) 59 (96) Oral 7 (4) - 1V IV 159 (96) - 0.03 ^b No 84 (51) 125 (38) 125 (38) Yes 82 (49) 201 (61) 0.03 ^b No 82 (49) 201 (61) 0.36 ^b	ATG/Alemtuzumab - no. (%)			<.01 ^b
No ATG or alemtuzumab 79 (48) 246 (75) ATG dose (mg) - median (min-max) 400 (5-4900) 284 (4-2988) <.01 ^a ATG dose (mg) - no. (%) 0.43 ^b 0.43 ^b 0-5mg 2 (2) 4 (6) 5-7.5mg 2 (2) 0 (0) >7.5mg 78 (91) 65 (90) Missing 4 (5) 3 (4) Melphalan dose - no. (%) - 52 (16) Melphalan 100mg/m ² - 52 (16) Melphalan 140mg/m ² - 275 (84) Busulfan Administration - no. (%) 0.03 ^b Oral 7 (4) - IV 159 (96) - Ruxolitinib use prior to HCT- no. (%) 0.03 ^b No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b	ATG alone	86 (52)	72 (22)	
ATG dose (mg) - median (min-max) $400 (5-4900) 284 (4-2988)$ $<.01^a$ ATG dose (mg) - no. (%) $2 (2) 4 (6)$ 0.43^b $0-5mg$ $2 (2) 0 (0)$ $2(2) 0 (0)$ $5-7.5mg$ $2 (2) 0 (0)$ $>7.5mg$ $78 (91) 65 (90)$ Missing $4 (5) 3 (4)$ Melphalan dose - no. (%) $- 52 (16)$ Melphalan 100mg/m² $- 275 (84)$ Busulfan Administration - no. (%) 0.03^b Oral $7 (4) -$ IV $159 (96)$ No $84 (51) 125 (38)$ Yes $82 (49) 201 (61)$ Missing $0 (0) 1 (0)$ Splenic radiation prior HCT - no. (%) 0.36^b	Alemtuzumab alone	1 (1)	9 (3)	
ATG dose (mg) - no. (%) 0.43 ^b 0-5mg 2 (2) 4 (6) 5-7.5mg 2 (2) 0 (0) >7.5mg 78 (91) 65 (90) Missing 4 (5) 3 (4) Melphalan dose - no. (%) - 52 (16) Melphalan 100mg/m ² - 52 (16) Melphalan 140mg/m ² - 275 (84) Busulfan Administration - no. (%) 0ral 7 (4) Oral 7 (4) - IV 159 (96) - Ruxolitinib use prior to HCT- no. (%) 0.03 ^b No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0)	No ATG or alemtuzumab	79 (48)	246 (75)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		400 (5-4900)	284 (4-2988)	<.01 ^a 0.43 ^b
>7.5mg 78 (91) 65 (90) Missing 4 (5) 3 (4) Melphalan dose - no. (%) - 52 (16) Melphalan 140mg/m² - 52 (16) Melphalan 140mg/m² - 275 (84) Busulfan Administration - no. (%) - - Oral 7 (4) - IV 159 (96) - Ruxolitinib use prior to HCT- no. (%) 0.03 ^b No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b	0-5mg	· · ·	• •	
Missing 4 (5) 3 (4) Melphalan dose - no. (%) - 52 (16) Melphalan 100mg/m² - 52 (16) Melphalan 140mg/m² - 275 (84) Busulfan Administration - no. (%) - - Oral 7 (4) - IV 159 (96) - Ruxolitinib use prior to HCT- no. (%) 0.03 ^b No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b	•	• • •	• •	
Melphalan dose - no. (%) - 52 (16) Melphalan 100mg/m² - 52 (16) Melphalan 140mg/m² - 275 (84) Busulfan Administration - no. (%) - - Oral 7 (4) - IV 159 (96) - Ruxolitinib use prior to HCT- no. (%) 0.03 ^b No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b		· · ·	• •	
Melphalan 140mg/m² - 275 (84) Busulfan Administration - no. (%) 7 (4) - Oral 7 (4) - IV 159 (96) - Ruxolitinib use prior to HCT- no. (%) 0.03 ^b No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b	0	1 (0)	0(1)	
Busulfan Administration - no. (%) 7 (4) - Oral 7 (4) - IV 159 (96) - Ruxolitinib use prior to HCT- no. (%) 0.03 ^b No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b	Melphalan 100mg/m ²	-	52 (16)	
Oral 7 (4) - IV 159 (96) - Ruxolitinib use prior to HCT- no. (%) 0.03 ^b No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b	Melphalan 140mg/m ²	-	275 (84)	
IV 159 (96) - Ruxolitinib use prior to HCT- no. (%) 0.03 ^b No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b	Busulfan Administration - no. (%)			
IV 159 (96) - Ruxolitinib use prior to HCT- no. (%) 0.03 ^b No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b	Oral	7 (4)	-	
No 84 (51) 125 (38) Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b	IV	159 (96)	-	
Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b	Ruxolitinib use prior to HCT- no. (%)			0.03 ^b
Yes 82 (49) 201 (61) Missing 0 (0) 1 (0) Splenic radiation prior HCT - no. (%) 0.36 ^b	No	84 (51)	125 (38)	
Splenic radiation prior HCT - no. (%) 0.36 ^b	Yes	82 (49)	201 (61)	
	Missing	0 (0)	1 (0)	
	-	. ,		0.36 ^b
	No	161 (97)	308 (94)	
Yes 5 (3) 18 (6)	Yes		18 (6)	

Characteristic	RIC: Flu/Bu F	RIC: Flu/Bu RIC: Flu/Mel		
Missing	0 (0)	1 (0)		
Year of HCT - no. (%)			0.01 ^b	
2008	11 (7)	17 (5)		
2009	9 (5)	14 (4)		
2010	4 (2)	2 (1)		
2011	0 (0)	1 (0)		
2012	0 (0)	1 (0)		
2013	4 (2)	4 (1)		
2014	27 (16)	23 (7)		
2015	11 (7)	20 (6)		
2016	21 (13)	32 (10)		
2017	32 (19)	64 (20)		
2018	22 (13)	73 (22)		
2019	25 (15)	76 (23)		
Follow-up - median (range)	36 (3-143)	24 (3-122)		

Hypothesis testing: ^a Kruskal-Wallis test ^b Pearson chi-square test

Other race include: Asian (n= 15), Native Hawaiian or other Pacific Islander (n=4), More than 1 race (n=1)

RIC-Reduced intensity conditioning; Flu-Fludarabine; Bu-Busulfan; Mel- Melphalan; HCTCIhematopoietic cell transplantation comorbidity index; HCT-hematopoietic cell transplantation; DIPSS- dynamic international prognostic scoring system; CMV – cytomegalovirus; TACtacrolimus, CSA – cyclosporine; MMF- mycophenolate mofetil; MTX- methotrexate; CYcyclophosphamide; ATG – antithymocyte globulin; GVHD- graft versus host disease

Characteristic	MAC: Flu/Bu	MAC: Bu/Cy	P Value
No. of patients	247	132	
No. of centers	61	30	
Patient age - median (min-max)	60 (27-74)	55 (24-67)	<.01 ^a
Age group - no. (%)			<.01 ^b
18-29	1 (0)	1 (1)	
30-39	2 (1)	7 (5)	
40-49	25 (10)	24 (18)	
50-59	96 (39)	78 (59)	
60-69	110 (45)	22 (17)	
>= 70	13 (5)	0 (0)	
Sex - no. (%)			0.17 ^b
Male	126 (51)	77 (58)	
Female	121 (49)	55 (42)	
Region - no. (%)			0.62 ^b
US	240 (97)	130 (98)	
Canada	1 (0)	1 (1)	
Europe	2 (1)	0 (0)	
Australia/New Zealand	4 (2)	1 (1)	
Race/ethnicity - no. (%)			<.01 ^b
Hispanic	16 (6)	3 (2)	
Non-Hispanic White	209 (85)	113 (86)	
Non-Hispanic Black/African American	15 (6)	3 (2)	
Other	7 (3)	10 (8)	
Missing	0 (0)	3 (2)	
HCTCI - no. (%)			0.32 ^b
0	62 (25)	45 (34)	
1	38 (15)	21 (16)	
2	40 (16)	19 (14)	
3	47 (19)	26 (20)	
4	31 (13)	13 (10)	
5	12 (5)	5 (4)	
6+	16 (6)	2 (2)	
Missing	1 (0)	1 (1)	
Karnofsky performance score - no. (%)			0.42 ^b
90-100	151 (61)	89 (67)	

Supplemental Table S2. Baseline characteristics with myeloablative conditioning

Characteristic	MAC: Flu/Bu	MAC: Bu/Cy	P Value
< 90	92 (37)	42 (32)	
Missing	4 (2)	1 (1)	
Disease type - no. (%)			0.85 ^b
Primary Myelofibrosis	191 (77)	100 (76)	
Post polycythemia vera	20 (8)	13 (10)	
Post essential thrombocythemia	36 (15)	19 (14)	
Time from diagnosis to HCT - median (min-max)	25 (2-490)	38 (3-377)	0.41 ^a
Time from diagnosis to HCT - no. (%)			0.65 ^b
<6 months	38 (15)	24 (18)	
6-11 months	51 (21)	23 (17)	
>=12 months	158 (64)	85 (64)	
DIPSS Score - no. (%)			0.03 ^b
Low/Intermediate-1	134 (54)	80 (61)	
Intermediate-2/High	93 (38)	34 (26)	
Missing	20 (8)	18 (14)	
Splenomegaly at HCT - no. (%)			0.19 ^b
No	85 (34)	38 (29)	
Yes	133 (54)	73 (55)	
Splenectomy	10 (4)	12 (9)	
Missing	19 (8)	9 (7)	
Systemic symptoms prior to HCT- no. (%)			0.53 ^b
No	185 (75)	105 (80)	
Yes	51 (21)	21 (16)	
Missing	11 (4)	6 (5)	
Donor type - no. (%)			0.15 ^b
HLA-identical sibling	79 (32)	53 (40)	
8/8-matched unrelated	142 (57)	62 (47)	
7/8-matched unrelated	26 (11)	17 (13)	
Donor/recipient sex match - no. (%)			0.17 ^b
Male-Male	87 (35)	44 (33)	
Male-Female	72 (29)	31 (23)	
Female-Male	39 (16)	33 (25)	
Female-Female	49 (20)	24 (18)	
Donor/recipient CMV serostatus - no. (%)			0.37 ^b
+/+	66 (27)	26 (20)	
+/-	42 (17)	19 (14)	
_/+	64 (26)	40 (30)	

Characteristic	MAC: Flu/Bu	MAC: Bu/Cy	P Value
	71 (29)	46 (35)	
Missing	4 (2)	1 (1)	
Graft type - no. (%)			<.01 ^b
Bone marrow	11 (4)	16 (12)	
Peripheral blood	236 (96)	116 (88)	
GVHD prophylaxis - no. (%)			0.03 ^b
TAC + MMF +- other(s) (except post-CY)	26 (11)	5 (4)	
TAC + MTX +- other(s) (except MMF, post-CY)	193 (78)	111 (84)	
TAC + other(s) (except MMF, MTX, post-CY)	12 (5)	3 (2)	
TAC alone	4 (2)	0 (0)	
CSA + MMF +- other(s) (except post-CY)	3 (1)	2 (2)	
CSA + MTX +- other(s) (except MMF, post-CY)	9 (4)	11 (8)	
ATG/Alemtuzumab use - no. (%)			<.01 ^b
No	135 (55)	125 (95)	
Yes	112 (45)	7 (5)	
ATG/alemtuzumab - no. (%)			<.01 ^b
ATG + alemtuzumab	1 (0)	0 (0)	
ATG alone	107 (43)	6 (5)	
Alemtuzumab alone	4 (2)	1 (1)	
No ATG or alemtuzumab	135 (55)	125 (95)	
ATG dose (mg) - median (min-max)	338 (3-6000)	250 (150- 500)	0.32 ^a
ATG dose (mg) - no. (%)	3(3)	0(0)	0.10 ^{c[;]}
0-5mg 5-7.5mg	0(0)	0(0)	0110
>7.5mg	98(92)	4(67)	
Missing	6(6)	2(33)	
Busulfan pharmacokinetics performed - no. (%)			0.62 ^b
No	102 (41)	51 (39)	
Yes	145 (59)	81 (61)	
Busulfan Administration - no. (%)			0.02 ^b
Oral	2 (1)	6 (5)	
IV	245 (99)	126 (95)	
Ruxolitinib use prior to HCT - no. (%)			<.01 ^b
No	101 (41)	75 (57)	
Yes	146 (59)	57 (43)	
Splenic radiation prior HCT - no. (%)	007 (00)	404 (00)	0.07 ^b
No	237 (96)	131 (99)	

	MAC:	MAC:	
Characteristic	Flu/Bu	Bu/Cy	P Value
Yes	10 (4)	1 (1)	
Year of HCT - no. (%)			<.01 ^b
2008	9 (4)	15 (11)	
2009	12 (5)	21 (16)	
2010	5 (2)	8 (6)	
2011	2 (1)	6 (5)	
2012	1 (0)	3 (2)	
2013	9 (4)	6 (5)	
2014	35 (14)	6 (5)	
2015	27 (11)	15 (11)	
2016	25 (10)	11 (8)	
2017	47 (19)	15 (11)	
2018	39 (16)	15 (11)	
2019	36 (15)	11 (8)	
Follow-up - median (range)	25 (3-122)	49 (6-150)	

Hypothesis testing: ^a Kruskal-Wallis test ^b Pearson chi-square test

Other race include: Asian (n= 11), Native Hawaiian or other Pacific Islander (n=6), American Indian or Alaska Native (n=1), More than 1 race (n=1)

MAC-Myeloablative conditioning; Flu-Fludarabine; Bu-Busulfan; Cy- Cyclophosphamide; HCTCI-hematopoietic cell transplantation comorbidity index; HCT-hematopoietic cell transplantation; DIPSS- dynamic international prognostic scoring system; CMV – cytomegalovirus; TAC- tacrolimus, CSA – cyclosporine; MMF- mycophenolate mofetil; MTXmethotrexate; CY- cyclophosphamide; ATG – antithymocyte globulin; GVHD- graft versus host disease

	RIC	: Flu/Bu (N = 166)	RIC	: Flu/Mel (N = 327)	
Outcomes	Ν	Prob (95% Cl)	Ν	Prob (95% Cl)	P Value
Non-relapse mortality	164		321		0.21
100-day	Censored 10	4.3 (1.7-7.9)%	6	17.4 (13.5-21.8)%	<0.001
	Event 7		56		
	N at risk 148		261		
1-year	Censored 73	19.9 (14.1-26.5)%	146	27.3 (22.5-32.3)%	0.06
	Event 25		30		
	N at risk 51		86		
2-year	Censored 13	26.8 (20-34.2)%	28	28.7 (23.7-33.9)%	0.67
	Event 9		3		
	N at risk 27		52		
Relapse	164		321		0.08
1-year	Censored 37	48.8 (41-56.6)%	109	41.7 (36.2-47.2)%	0.14
	Event 78		129		
	N at risk 51		86		
2-year	Censored 21	49.6 (41.8-57.4)%	26	44 (38.4-49.7)%	0.26
	Event 1		5		
	N at risk 27		52		
Disease Free-Survival	164		321		0.99
1-year	Censored 5	31.3 (24.3-38.7)%	23	31.1 (26-36.4)%	0.96
	Event 110		215		
	N at risk 49		83		
2-year	Censored 12	23.6 (17.2-30.8)%	23	27.3 (22.2-32.6)%	0.40
	Event 10		8		
	N at risk 27		52		
Overall Survival	166		327		0.43
1-year	Censored 12	68.7 (61.3-75.7)%	45	65.6 (60.2-70.7)%	0.49
	Event 50		109		
	N at risk 104		173		
2-year	Censored 24	56.9 (48.7-64.8)%	60	56.7 (50.8-62.6)%	0.98
	Event 15		18		
	N at risk 65		95		
Acute GVHD II-IV	164		325		0.63
100-day	Censored 9	35.5 (28.3-43)%	42	40.1 (34.8-45.5)%	0.32
	Event 58		130		
	N at risk 98		154		

Supplemental Table S3. Univariable analysis with reduced intensity conditioning (Unadjusted, censored at 24 months)

		RIC	: Flu/Bu (N = 166)	RIC:	Flu/Mel (N = 327)	
Outcomes		Ν	Prob (95% CI)	Ν	Prob (95% Cl)	P Value
6 months	Censored	18	43.9 (36.3-51.6)%	14	43.6 (38.2-49)%	0.95
	Event	13		11		
	N at risk	68		130		
1-year	Censored	13	46.7 (38.9-54.5)%	31	45.4 (40-50.9)%	0.79
	Event	4		5		
	N at risk	50		96		
Acute GVHD III-IV	1	65		326		0.12
6 months	Censored	33	18.5 (12.9-24.9)%	64	22.8 (18.4-27.5)%	0.26
	Event	30		75		
	N at risk 1	103		190		
1-year	Censored	19	18.5 (12.9-24.9)%	49	23.9 (19.4-28.7)%	0.16
	Event	1		3		
	N at risk	83		141		
2-year	Censored	31	18.5 (12.9-24.9)%	61	23.9 (19.4-28.7)%	0.16
	Event	0		0		
	N at risk	51		74		
Chronic GVHD	1	65		323		0.15
1-year	Censored	56	44.6 (36.7-52.6)%	139	35.2 (29.8-40.8)%	0.05
	Event	68		104		
	N at risk	42		83		
2-year	Censored	15	50.1 (41.8-58.4)%	32	47.8 (41.7-54)%	0.66
	Event	6		24		
	N at risk	20		24		
GRFS	1	64		320		0.30
1-year	Censored	5	12.3 (7.6-17.9)%	19	11.6 (8.2-15.6)%	0.84
	Event 1	140		273		
	N at risk	19		28		
2-year	Censored	5	8.9 (4.8-14.2)%	7	4.7 (2.2-7.9)%	0.12
	Event	4		13		
	N at risk	10		8		

RIC-Reduced intensity conditioning; Flu-Fludarabine; Bu-Busulfan; Mel- Melphalan; GVHD-Graft-versus-host disease; GRFS- GVHD-Free Relapse-Free survival

	MAC	: Flu/Bu (N = 247)	MAC	: Bu/Cy (N = 132)	
Outcomes	Ν	Prob (95% Cl)	Ν	Prob (95% Cl)	P Value
Non-relapse mortality	242		132		0.28
100-day	Censored 12	6.2 (3.5-9.6)%	4	9.1 (4.8-14.6)%	0.32
	Event 15		12		
	N at risk 216		117		
1-year	Censored 102	15.7 (11.3-20.6)%	43	19 (12.8-26.2)%	0.42
	Event 22		13		
	N at risk 92		61		
2-year	Censored 24	16.8 (12.2-21.9)%	3	21.5 (14.8-29)%	0.28
	Event 2		3		
	N at risk 65		54		
Relapse	242		132		0.24
1-year	Censored 63	38.2 (32-44.6)%	29	32.9 (25.1-41.2)%	0.31
	Event 88		43		
	N at risk 92		61		
2-year	Censored 22	40.5 (34.1-47)%	5	33.7 (25.8-42.1)%	0.20
	Event 4		1		
	N at risk 65		54		
Disease-Free Survival	242		132		0.81
1-year	Censored 26	46.2 (39.7-52.6)%	4	48.1 (39.6-56.7)%	0.72
	Event 125		68		
	N at risk 91		60		
2-year	Censored 20	42.7 (36.3-49.3)%	2	44.8 (36.4-53.4)%	0.70
	Event 6		4		
	N at risk 65		54		
Overall Survival	247		132		0.71
1-year	Censored 38	78 (72.5-83.1)%	8	77.8 (70.4-84.5)%	0.96
	Event 52		29		
	N at risk 157		95		
2-year	Censored 38	72.1 (65.9-78)%	12	70.7 (62.4-78.3)%	0.77
	Event 10		8		
	N at risk 109		75		
Acute GVHD II-IV	244		129		<0.001
100-day	Censored 13	34.1 (28.3-40.2)%	7	58.9 (50.3-67.3)%	<0.001
	Event 83		76		
	N at risk 149		47		

Supplemental Table S4. Univariable analysis with myeloablative conditioning (Unadjusted, censored at 24 months)

	MAG	C: Flu/Bu (N = 247)	MAC	: Bu/Cy (N = 132)	
Outcomes	Ν	Prob (95% Cl)	Ν	Prob (95% CI)	P Value
6 months	Censored 18	43 (36.8-49.3)%	1	62.8 (54.2-71)%	<0.001
	Event 21		5		
	N at risk 111		41		
1-year	Censored 26	47.5 (41.1-53.9)%	1	62.8 (54.2-71)%	0.004
	Event 9		0		
	N at risk 75		40		
Acute GVHD III-IV	244		127		<0.001
6 months	Censored 39	16.6 (12.1-21.5)%	10	34.6 (26.6-43.2)%	<0.001
	Event 40		44		
	N at risk 168		75		
1-year	Censored 33	19.5 (14.6-24.8)%	8	34.6 (26.6-43.2)%	0.002
	Event 6		0		
	N at risk 127		67		
2-year	Censored	19.5 (14.6-24.8)%		34.6 (26.6-43.2)%	0.002
	Event				
	N at risk 82		53		
Chronic GVHD	242		131		0.29
1-year	Censored 73	46.1 (39.4-52.8)%	30	47.6 (38.9-56.3)%	0.78
	Event 100		61		
	N at risk 70		41		
2-year	Censored 21	53.3 (46.3-60.1)%	9	58.1 (49.1-66.8)%	0.40
	Event 12		11		
	N at risk 36		20		
GRFS	241		127		<0.001
1-year	Censored 14	19.2 (14.3-24.6)%	3	13.4 (8-19.9)%	0.15
	Event 186		109		
	N at risk 41		15		
2-year	Censored 6	14.9 (10.5-20)%	0	8.9 (4.5-14.7)%	0.09
	Event 9		5		
	N at risk 26		10		

MAC-Myeloablative conditioning; Flu-Fludarabine; Bu-Busulfan; Cy- Cyclophosphamide; GVHD-Graft-versus-host disease; GRFS-GVHD-Free Relapse-Free survival

N 166	HR 1.00	95% CI Lower Limit	95% CI Upper Limit	p-value	Overall p-value
166				p-value	
	1 00				
	1 00				
	1.00	Reference			0.009
326	1.80	1.15	2.81	0.009	
133	1.00	Reference			0.35
239	0.82	0.53	1.26	0.35	
324	1.00	Reference			0.03
168	1.39	1.03	1.88	0.03	
		95% CI	95% CI		
N	HR	Lower Limit	Upper Limit	p-value	Overall p-value
					0.85
164	1.00	Reference			
320	1.03	0.77	1.38	0.85	
					0.76
92	1.00	Reference			
181	0.95	0.68	1.34	0.76	
	239 324 168 N 164 320 92	 239 0.82 324 1.00 168 1.39 N HR 164 1.00 320 1.03 92 1.00 	239 0.82 0.53 324 1.00 Reference 168 1.39 1.03 95% CI N HR Lower 164 1.00 Reference 320 1.03 0.77 92 1.00 Reference	239 0.82 0.53 1.26 324 1.00 Reference 168 1.39 1.03 1.88 95% CI 95% CI 95% CI N HR Lower Limit Upper Limit 164 1.00 Reference 320 1.03 0.77 1.38 92 1.00 Reference	239 0.82 0.53 1.26 0.35 324 1.00 Reference168 1.39 1.03 1.88 0.03 NHRLowerUpper Limitp-value164 1.00 Reference320 1.03 0.77 1.38 0.85 92 1.00 Reference

Supplemental Table S5. Multivariable Analyses - reduced intensity conditioning

Relapse			95% CI	95% CI		Overall p-value
Variable	N	HR	Lower Limit	Upper Limit	p-value	
Main Effect						0.25
RIC: Flu/Bu	165	1.00	Reference			
RIC: Flu/Mel	320	0.85	0.64	1.12	0.25	
Year of transplant						0.01
2008-2014	116	1.00	Reference			
2015-2019	369	1.53	1.08	2.17	0.01	
Non-Relapse Mortality			95% CI	95% CI		
Variable	N	HR	Lower Limit	Upper Limit	p-value	Overal p-value
Main Effect (<=6 months)						0.01
RIC: Flu/Bu	164	1.00	Reference			
RIC: Flu/Mel	320	1.81	1.12	2.91	0.01	
Main Effect (> 6 months)						0.02
Main Effect (> 6 months) RIC: Flu/Bu	92	1.00	Reference			0.02
	92 181	1.00 0.46	Reference 0.23	0.91	0.02	0.02
RIC: Flu/Bu				0.91 95% Cl	0.02	0.02
RIC: Flu/Bu RIC: Flu/Mel			0.23		0.02 p-value	Overal
RIC: Flu/Bu RIC: Flu/Mel Acute GVHD II-IV	181	0.46	0.23 95% Cl Lower	95% Cl Upper		0.02 Overal p-value 0.03

RIC: Flu/Mel	324	1.45	1.03	2.03	0.03	
Main Effect (> 2 months)						0.18
Main Ellect (> 2 months)						0.10
RIC: Flu/Bu	111	1.00	Reference			
RIC: Flu/Mel	174	0.71	0.43	1.17	0.18	

Acute GVHD III-IV			95% CI	95% CI		
Variable	N	HR	Lower Limit	Upper Limit	p-value	Overall p-value
Main Effect (<=2 months)						0.004
RIC: Flu/Bu	165	1.00	Reference			
RIC: Flu/Mel	325	2.21	1.28	3.83	0.004	
Main Effect (>2 months)						0.48
RIC: Flu/Bu	143	1.00	Reference			
RIC: Flu/Mel	226	0.89	0.64	1.24	0.48	
Chronic GVHD (adjusted center effect)			95% CI	95% CI		Overall p-value
Variable	N	HR	Lower Limit	Upper Limit	p-value	
Main Effect						0.55
RIC: Flu/Bu	165	1.00	Reference			
RIC: Flu/Bu RIC: Flu/Mel	165 322	1.00 0.91	Reference 0.67	1.25	0.55	
				1.25	0.55	

8/8-matched unrelated donor	309	0.97	0.69	1.35	0.83	
7/8-matched unrelated donor	39	1.89	1.15	3.12	0.01	
Contrast						
8/8-matched unrelated vs 7/8- matched unrelated donor		0.51	0.30	0.88	0.01	
GRFS			95% CI	95% CI		Overall p-value
GRFS Variable	N	HR	95% CI Lower Limit	95% CI Upper Limit	p-value	
	Ν	HR	Lower	Upper	p-value	
Variable	N 164	HR 1.00	Lower	Upper	p-value	p-value
Variable Main Effect			Lower Limit	Upper	p-value 0.32	p-value

**RIC- reduced intensity conditioning; Flu/Bu – Fludarabine/busulfan; Flu/Mel-Fludarabine/melphalan; HCT- hematopoietic cell transplantation; ATG – antithymocyte globulin; GVHD- graft versus host disease; GRFS – GVHD-free relapse-free survival

The following factors were considered in the multivariable analysis - patient age, race/ethnicity, disease subtype, dynamic international performance scoring system (DIPSS) score, Hematopoietic Cell Transplantation Comorbidity Index (HCT-CI), Karnofsky Performance Scale (KPS), splenic radiation, splenomegaly at HCT, ruxolitinib use pre-transplant, donor-recipient HLA-match, gender match, CMV match, stem cell source, GVHD prophylaxis, use of antithymocyte globulin (ATG)/alemtuzumab, and year of transplant. Significant covariates are shown in table.

Overall Survival			95% CI	95% Cl		
Variable	Ν	HR	Lower Limit	Upper Limit	p-value	Overall p-value
Main Effect						0.54
MAC: Flu/Bu	247	1.00	Reference			
MAC: Bu/Cy	132	1.14	0.75	1.71	0.54	
Donor type						<0.001
HLA-identical sibling	132	1.00	Reference			
8/8-matched unrelated	204	1.78	1.09	2.89	0.02	
7/8-matched unrelated	43	3.33	1.83	6.06	<0.001	
Contrast						
8/8-matched unrelated vs 7/8- matched unrelated donor		0.53	0.32	0.89	0.01	
Disease Free Survival			95% CI	95% Cl		Overall p-value
Variable	N	HR	Lower Limit	Upper Limit	p-value	
Main Effect						0.83
MAC: Flu/Bu	238	1.00	Reference			
MAC: Bu/Cy	131	1.03	0.77	1.38	0.83	
Karnofsky score						0.006
90-100	238	1.00	Reference			

Supplemental Table S6. Multivariable Analyses for patients who received myeloablative conditioning

< 90	131	1.47	1.11	1.95	0.006	
Ruxolitinib use prior to HCT						0.04
No	170	1.00	Reference			
Yes	199	1.33	1.01	1.77	0.04	
Relapse			95% CI	95% Cl		Overall p-value
Variable	N	HR	Lower Limit	Upper Limit	p-value	
Main Effect						0.65
MAC: Flu/Bu	238	1.00	Reference			
MAC: Bu/Cy	131	0.92	0.64	1.32	0.65	
Karnofsky score						0.006
90-100	238	1.00	Reference			
< 90	131	1.61	1.14	2.26	0.006	
Ruxolitinib use prior to HCT						0.01
No	170	1.00	Reference			
Yes	199	1.56	1.10	2.21	0.01	
Non Relapse Mortality			95% CI	95% Cl		Overall p-value
Variable	N	HR	Lower Limit	Upper Limit	p-value	
Main Effect						0.22
MAC: Flu/Bu	242	1.00	Reference			

MAC: Bu/Cy	132	1.36	0.83	2.21	0.22	
Donor type						<0.001
HLA-identical sibling	131	1.00	Reference			
8/8-matched unrelated	201	2.66	1.36	5.18	0.004	
7/8-matched unrelated	42	5.33	2.44	11.61	<0.001	
Contrast 8/8-matched unrelated vs 7/8-						
matched unrelated donor		0.50	0.28	0.90	0.02	
Acute GVHD II-IV			95% CI	95% Cl		Overall p-value
Variable	N	HR	Lower Limit	Upper Limit	p-value	
Variable Main Effect (<=2 months)	Ν	HR			p-value	<0.001
	N 244	HR 1.00			p-value	<0.001
Main Effect (<=2 months)			Limit		p-value <0.001	<0.001
Main Effect (<=2 months) MAC: Flu/Bu MAC: Bu/Cy	244	1.00	Limit Reference	Limit		<0.001
Main Effect (<=2 months) MAC: Flu/Bu	244	1.00	Limit Reference	Limit		
Main Effect (<=2 months) MAC: Flu/Bu MAC: Bu/Cy Main Effect(> 2 months)	244 129	1.00 2.33	Limit Reference 1.67	Limit		
Main Effect (<=2 months) MAC: Flu/Bu MAC: Bu/Cy Main Effect(> 2 months) MAC: Flu/Bu	244 129 163	1.00 2.33 1.00	Limit Reference 1.67 Reference	Limit 3.25	<0.001	
Main Effect (<=2 months) MAC: Flu/Bu MAC: Bu/Cy Main Effect(> 2 months) MAC: Flu/Bu	244 129 163	1.00 2.33 1.00	Limit Reference 1.67 Reference	Limit 3.25	<0.001	
Main Effect (<=2 months) MAC: Flu/Bu MAC: Bu/Cy Main Effect(> 2 months) MAC: Flu/Bu MAC: Bu/Cy	244 129 163	1.00 2.33 1.00	Limit Reference 1.67 Reference	Limit 3.25	<0.001	0.69
Main Effect (<=2 months) MAC: Flu/Bu MAC: Bu/Cy Main Effect(> 2 months) MAC: Flu/Bu MAC: Flu/Bu MAC: Bu/Cy	244 129 163 53	1.00 2.33 1.00 0.88	Limit Reference 1.67 Reference 0.46	Limit 3.25	<0.001	0.69

Contrast

8/8-matched unrelated vs 7/8- matched unrelated donor		0.57	0.38	0.86	0.007	
Acute GVHD III-IV			95% CI	95% Cl		Overall p-value
Variable	N	HR	Lower Limit	Upper Limit	p-value	
Main Effect						<0.001
MAC: Flu/Bu	244	1.00	Reference			
MAC: Bu/Cy	127	2.31	1.52	3.52	<0.001	
Ruxolitinib use prior to HCT						0.02
No	170	1.00	Reference			
Yes	201	1.67	1.08	2.58	0.02	
Chronic GVHD (adjusted center effect)			95% CI	95% Cl		Overall p-value
Variable	N	HR	Lower Limit	Upper Limit	p-value	
Main Effect						
MAC: Flu/Bu		4.00	Reference			0.00
	242	1.00	Relefence			0.36
MAC: Bu/Cy	242 131	1.00 1.21	0.80	1.84	0.36	0.36
				1.84	0.36	0.36
MAC: Bu/Cy				1.84	0.36	0.36

GRFS			95% CI	95% Cl		Overall p-value
Variable	N	HR	Lower Limit	Upper Limit	p-value	
Main Effect						<0.001
MAC: Flu/Bu	241	1.00	Reference			
MAC: Bu/Cy	127	1.94	1.49	2.53	<.0001	
Recipient age						0.001
18-49	56	1.00	Reference			
50-59	171	0.74	0.53	1.03	0.07	
>=60	141	1.22	0.85	1.74	0.27	
Donor type						<0.001
HLA-identical sibling	127	1.00	Reference			
8/8-matched unrelated	198	1.31	1.02	1.69	0.03	
7/8-matched unrelated	43	2.41	1.66	3.51	<0.001	
Contrast						
Age 50-59 vs >=60		0.61	0.47	0.79	<0.001	
8/8-matched unrelated vs 7/8- matched unrelated donor		0.54	0.38	0.77	<0.001	

**MAC- myeloablative conditioning; Flu/Bu – Fludarabine/busulfan; Bu/Cy-Busulfan/cyclophosphamide; HCT- hematopoietic cell transplantation; ATG – antithymocyte globulin; GVHD- graft versus host disease; GRFS – GVHD-free relapse-free survival The following factors were considered in the multivariable analysis - patient age, race/ethnicity, disease subtype, dynamic international performance scoring system (DIPSS) score, Hematopoietic Cell Transplantation Comorbidity Index (HCT-CI), Karnofsky Performance Scale (KPS), splenic radiation, splenomegaly at HCT, ruxolitinib use pre-transplant, donor-recipient HLA-match, gender match, CMV match, stem cell source, GVHD prophylaxis, use of antithymocyte globulin (ATG)/alemtuzumab, and year of transplant. Significant covariates are shown in table.

Overall Survival			95% CI	95% CI		
Variable	N	HR	Lower Limit	Upper Limit	p-value	Overall p- value
Main Effect (<= 6 months)						0.02
RIC: Flu/Bu	166	1.00	Reference			
RIC: Flu/Mel-100	52	2.07	1.08	3.97	0.02	
RIC: Flu/Mel-140	274	1.75	1.11	2.77	0.01	
Main Effect (> 6 months)						0.45
RIC: Flu/Bu	133	1.00	Reference			
RIC: Flu/Mel-100	32	1.12	0.50	2.52	0.78	
RIC: Flu/Mel-140	207	0.78	0.49	1.22	0.27	
ATG/alemtuzumab						0.03
No	324	1.00	Reference			
Yes	168	1.38	1.03	1.88	0.03	
Contrast						
<=6 mon: RIC: Flu/Mel- 100 vs RIC: Flu/Mel-140		1.18	0.66	2.11	0.56	
>6 mon: RIC: Flu/Mel-100 vs RIC: Flu/Mel-140		1.44	0.64	3.21	0.37	
Disease Free Survival			95% CI	95% CI		
Variable	N	HR	Lower Limit	Upper Limit	p-value	Overall p- value

Supplemental Table S7. Multivariable Analyses with RIC – analyzed based on melphalan dose

Main Effect (<= 6 months)						
RIC: Flu/Bu	164	1.00	Reference			0.87
RIC: Flu/Mel-100	51	1.14	0.70	1.85	0.61	
RIC: Flu/Mel-140	269	1.01	0.75	1.37	0.94	
Main Effect (> 6 months)						0.91
RIC: Flu/Bu	92	1.00	Reference			
RIC: Flu/Mel-100	24	1.03	0.55	1.93	0.92	
RIC: Flu/Mel-140	157	0.94	0.66	1.33	0.72	
Contrast						
<=6 mon: RIC: Flu/Mel- 100 vs RIC: Flu/Mel-140		1.12	0.71	1.79	0.62	
>6 mon: RIC: Flu/Mel-100 vs RIC: Flu/Mel-140		1.10	0.60	2.01	0.76	
Relapse			95% CI	95% CI		
Variable	N	HR	Lower Limit	Upper Limit	p-value	Overall p-value
Main Effect						0.47
RIC: Flu/Bu	164	1.00	Reference			
RIC: Flu/Mel-100	51	0.76	0.45	1.30	0.32	
RIC: Flu/Mel-140	269	0.86	0.64	1.14	0.30	
Year of transplant						0.01
1						
2008-2014	116	1.00	Reference			

Contrast

RIC: Flu/Mel-100 vs RIC: Flu/Mel-140		0.89	0.53	1.48	0.66	
Non relapse mortality			95% CI	95% CI		
Variable	N	HR	Lower Limit	Upper Limit	p-value	Overall p- value
Main Effect (<=6 months)						0.03
RIC: Flu/Bu	164	1.00	Reference			
RIC: Flu/Mel-100	51	2.28	1.17	4.47	0.01	
RIC: Flu/Mel-140	269	1.71	1.05	2.80	0.03	
Main Effect (>6 months) RIC: Flu/Bu RIC: Flu/Mel-100 RIC: Flu/Mel-140	92 24 157	1.00 0.55 0.44	Reference 0.12 0.22	2.37 0.91	0.42 0.02	0.07
Contrast <=6 mon: RIC: Flu/Mel- 100 vs RIC: Flu/Mel- 140		1.33	0.74	2.38	0.33	
>6 mon: RIC: Flu/Mel- 100 vs RIC: Flu/Mel- 140		1.23	0.27	5.45	0.78	

Acute GVHD II-IV			95% CI	95% CI		
Variable	N	HR	Lower Limit	Upper Limit	p-value	Overall p- value
Main Effect (<= 2 months)						0.10
RIC: Flu/Bu	164	1.00	Reference			
RIC: Flu/Mel-100	52	1.47	0.85	2.54	0.16	
RIC: Flu/Mel-140	272	1.44	1.01	2.04	0.04	
Main Effect(> 2 months)						0.34
RIC: Flu/Bu	111	1.00	Reference			
RIC: Flu/Mel-100	29	0.91	0.37	2.19	0.83	
RIC: Flu/Mel-140	145	0.67	0.39	1.14	0.14	
Contrast						
<=2 mon: RIC: Flu/Mel- 100 vs RIC: Flu/Mel- 140		1.02	0.61	1.69	0.92	
>2 mon: RIC: Flu/Mel- 100 vs RIC: Flu/Mel- 140		1.34	0.55	3.26	0.51	
Acute GVHD III-IV			95% CI	95% CI		
Variable	N	HR	Lower Limit	Upper Limit	p-value	Overall p- value
						0.005

Main Effect (<= 2

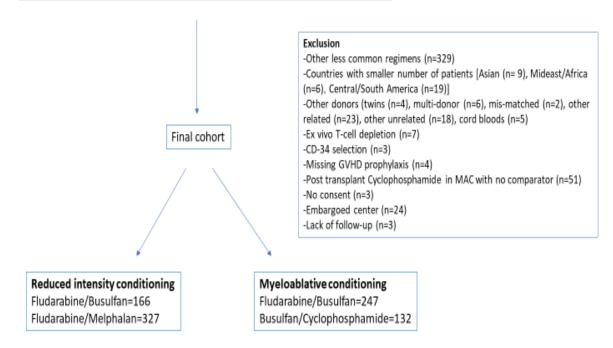
months)						
RIC: Flu/Bu	165	1.00	Reference			
RIC: Flu/Mel-100	52	1.31	0.51	3.35	0.57	
RIC: Flu/Mel-140	273	2.38	1.36	4.15	0.002	
Main Effect(>2 months)						0.42
RIC: Flu/Bu	143	1.00	Reference			
RIC: Flu/Mel-100	41	0.47	0.10	2.08	0.32	
RIC: Flu/Mel-140	185	0.67	0.32	1.39	0.28	
Contrast						
<=2 mon: RIC: Flu/Mel- 100 vs RIC: Flu/Mel- 140		0.55	0.23	1.27	0.16	
>2 mon: RIC: Flu/Mel-		0.00	0.20	1.27	0.10	
100 vs RIC: Flu/Mel- 140		0.70	0.16	3.11	0.64	
Chronic GVHD						
(adjusted center effect)			95% CI	95% CI		
			Lower	Upper		Overall p-
Variable	Ν	HR	Limit	Limit	p-value	value
Main Effect						0.73
RIC: Flu/Bu	165	1.00	Reference			
RIC: Flu/Mel-100	52	0.80	0.44	1.42	0.45	
RIC: Flu/Mel-140	270	0.92	0.67	1.27	0.64	

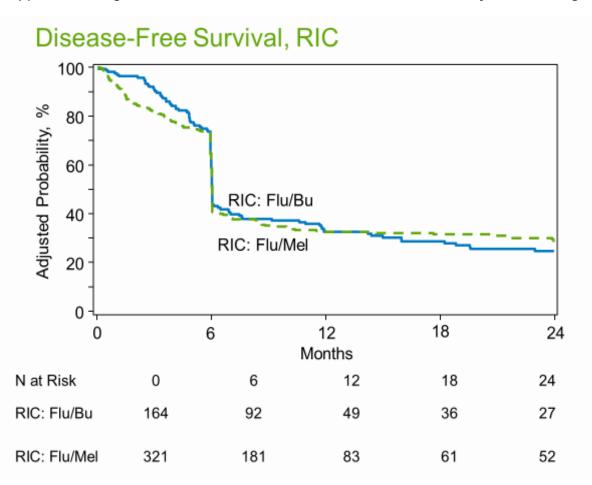
Donor type						0.03
HLA-identical sibling	139	1.00	Reference			
8/8-matched unrelated	309	0.97	0.70	1.34	0.86	
7/8-matched unrelated	39	1.92	1.17	3.16	0.009	
Contrast						
8/8-matched unrelated vs 7/8-matched unrelated		0.50	0.29	0.87	0.01	
RIC: Flu/Mel-100 vs RIC: Flu/Mel-140		0.86	0.50	1.48	0.59	
GRFS			95% CI	95% CI		
GRFS Variable	N	HR	95% CI Lower Limit	95% CI Upper Limit	p-value	Overall p- value
	N	HR	Lower	Upper	p-value	-
Variable	N 164	HR 1.00	Lower	Upper	p-value	value
Variable Main Effect			Lower Limit	Upper	p-value 0.67	value
Variable Main Effect RIC: Flu/Bu	164	1.00	Lower Limit Reference	Upper Limit		value
Variable Main Effect RIC: Flu/Bu RIC: Flu/Mel-100	164 51	1.00 1.07	Lower Limit Reference 0.76	Upper Limit	0.67	value
Variable Main Effect RIC: Flu/Bu RIC: Flu/Mel-100	164 51	1.00 1.07	Lower Limit Reference 0.76	Upper Limit	0.67	value

*RIC- reduced intensity conditioning; Flu/Bu – Fludarabine/busulfan; Flu/Mel-Fludarabine/melphalan; HCT- hematopoietic cell transplantation; ATG – antithymocyte globulin; GVHD- graft versus host disease; GRFS – GVHD-free relapse-free survival The following factors were considered in the multivariable analysis - patient age, race/ethnicity, disease subtype, dynamic international performance scoring system (DIPSS) score, Hematopoietic Cell Transplantation Comorbidity Index (HCT-CI), Karnofsky Performance Scale (KPS), splenic radiation, splenomegaly at HCT, ruxolitinib use pre-transplant, donor-recipient HLA-match, gender match, CMV match, stem cell source, GVHD prophylaxis, use of antithymocyte globulin (ATG)/alemtuzumab, and year of transplant. Significant covariates are shown in table.

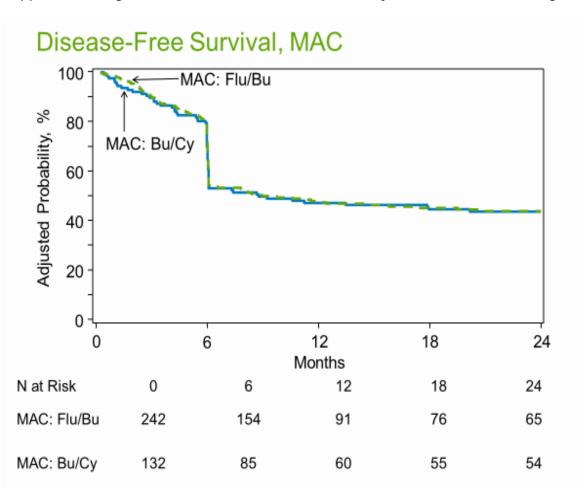
Cohort selection

Adults age ≥ 18 years with myelofibrosis undergoing first allogeneic hematopoietic cell transplantation from 2008-2019 and reported to the CIBMTR

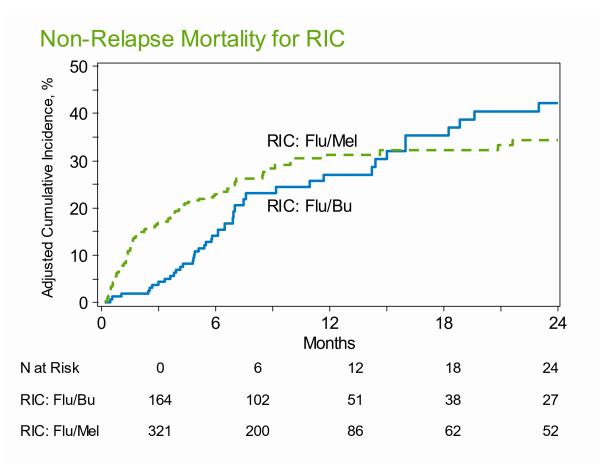




Supplemental Figure S2 – Disease free survival with reduced intensity conditioning

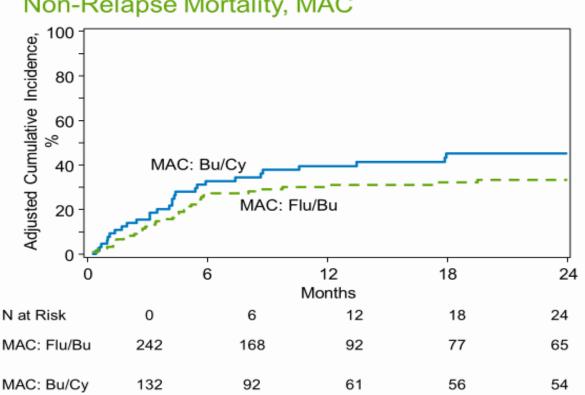


Supplemental Figure S3 – Disease free survival with myeloablative conditioning

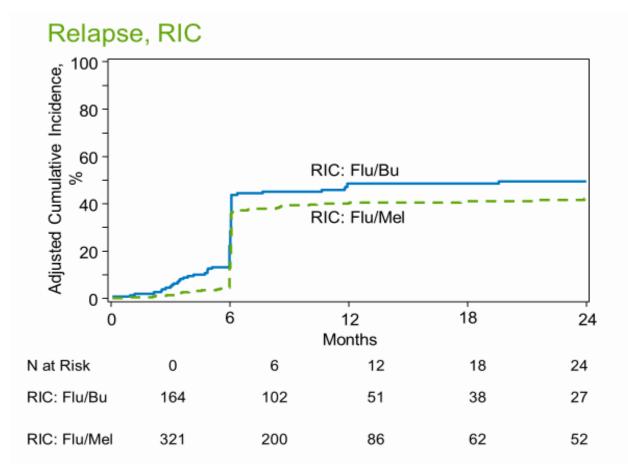


Supplemental Figure S4 – Non-relapse mortality with reduced intensity conditioning

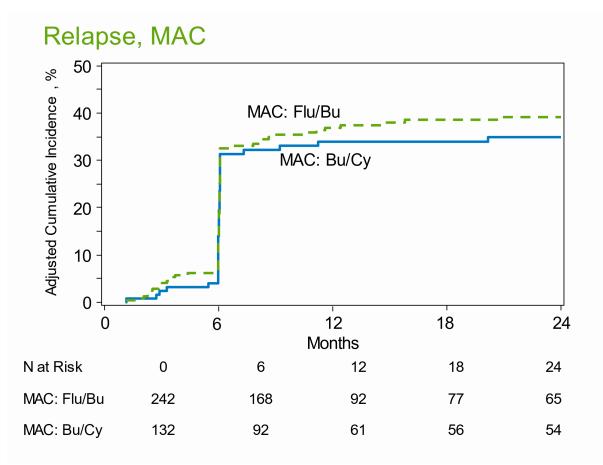
Supplemental Figure S5 – Non-relapse mortality with myeloablative conditioning



Non-Relapse Mortality, MAC

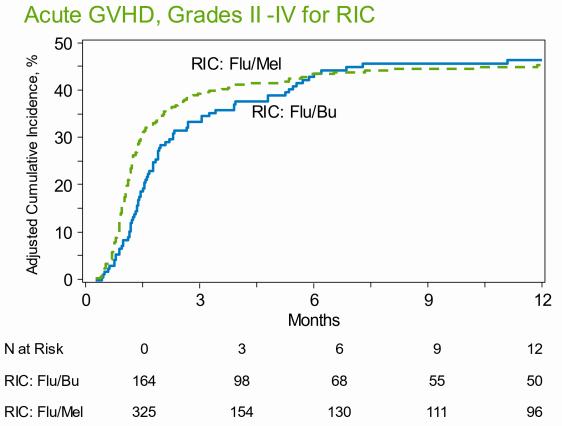


Supplemental Figure S6 – Relapse with reduced intensity conditioning

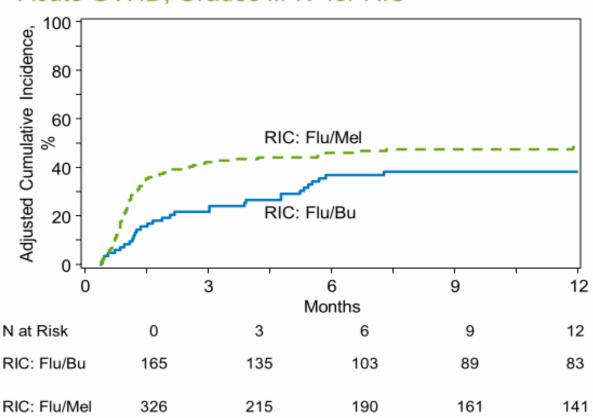


Supplemental Figure S7 – Relapse with myeloablative conditioning

Supplemental Figure S8 – Acute GVHD grade II-IV with reduced intensity conditioning

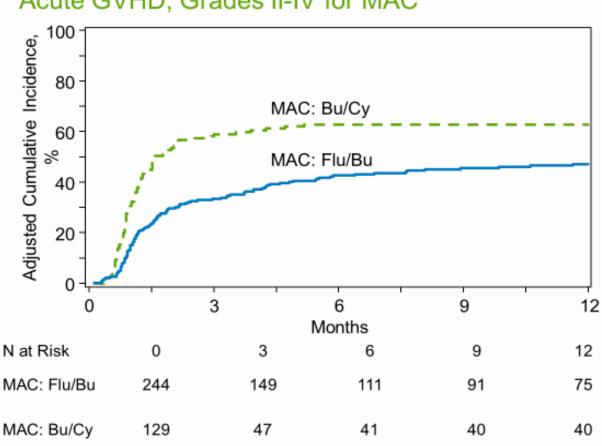


Supplemental Figure S9 – Acute GVHD grade III-IV with reduced intensity conditioning



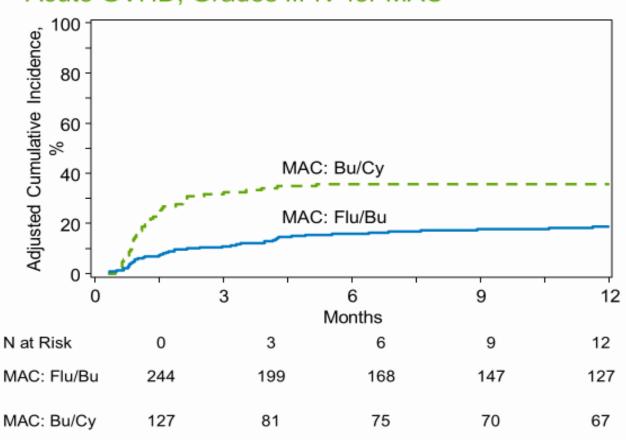
Acute GVHD, Grades III-IV for RIC

Supplemental Figure S10 – Acute GVHD grade II-IV with myeloablative conditioning



Acute GVHD, Grades II-IV for MAC

Supplemental Figure S11 – Acute GVHD grade III-IV with myeloablative conditioning



Acute GVHD, Grades III-IV for MAC

Supplemental Figure S12 – GVHD-free Relapse-free Survival with reduced intensity conditioning

