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1-6-2023

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Repli

# Reply to Bareille et al. Are Viscoelastometric Assays of Old Generation Ready for Disposal? Comment on "Volod et al. Viscoelastic Hemostatic Assays: A Primer on Legacy and New Generation Devices. J. Clin. Med. 2022, 11, 860"

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Citation: Volod, O.; Bunch, C.M.;
Miller, J.; Moore, E.E.; Moore, H.B.;
Kwaan, H.C.; Patel, S.S.; Wiarda, G.;
Aboukhaled, M.; Thomas, S.G.; et al.
Reply to Bareille et al. Are
Viscoelastometric Assays of Old
Generation Ready for Disposal?
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Viscoelastic Hemostatic Assays: A
Primer on Legacy and New
Generation Devices. J. Clin. Med.
2022, 11, 860". J. Clin. Med. 2023, 12,
478. https://doi.org/10.3390/
jcm12020478

Academic Editor: Emmanuel Favaloro

Received: 6 December 2022 Revised: 16 December 2022 Accepted: 27 December 2022 Published: 6 January 2023



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We are pleased to see that Bareille et al. have written a Commentary: "Are viscoelastometric assays of old generation ready for disposal?" [1] in response to our paper published in the Special Edition of the Journal of Clinical Medicine by Volod et al. [2].

The authors have expressed concern regarding our perceived excessive enthusiasm for the utilization of VHAs to define hemostatic derangement in many clinical settings and for what they call the "so called" COVID Associated Coagulopathy (CAC). We feel that it is necessary to provide a rebuttal to their commentary.

This rebuttal focuses on two points.

The first concerns Bareille et al.'s disagreement with our emphasis on the term COVID Associated Coagulopathy (CAC).

The second point addresses Bareille et al.'s objection that we have failed to point out that these legacy and post-legacy VHA devices are not "interchangeable".

Regarding the first point, the authors referred to our review of the use of VHAs in COVID-19 and our use of the term "coagulopathy" to define the universally observed proclivity of these patients to not only present with clotting anomalies but also to simultaneously hemorrhage. They object to our use of the term "coagulopathy" and they suggest replacing it with their term of "disordered hemostasis". They further note that this "disordered hemostasis" can often cause patients to "exhibit both thrombotic and hemorrhagic complications". They express concern that we did not address this duality. In fact, we mentioned this phenomenon that uniquely defines COVID-19 patients. In our manuscript on page 13, we specifically state that these patients can "clot and bleed at the same time" thus agreeing with Bareille et al.'s characterization of the singular nature of the "disordered hemostasis" in COVID-19 patients. During the peak two years of the COVID-19 pandemic, our group and others relied heavily on adjunctive VHAs to guide the anticoagulation of patients who had a COVID-associated coagulopathy or "disordered homeostasis", as Barielle et al. would prefer to call this unique hemostatic derangement. However, the term CAC has been used by many hematologic societies to describe the unique characteristic of these patients to "clot and bleed at the same time" [3–9].

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Whether the term "CAC" or "disordered hemostasis" is used to define this unique characteristic of COVID-19 patients to "clot and bleed at the same time", as we have written in our paper, matters little as we and many others have used VHAs to thread the needle and avoid the complications of under- and over-coagulation at either end of the hemostatic spectrum of the "so called" CAC or "disordered hemostasis" as they describe it [10]. Given the demonstrated benefit of VHAs to guide heparin anticoagulation, the FDA approved VHAs specifically for the purpose to assist clinicians in this difficult task of anticoagulating patients with CAC [11]. Clearly, CAC is a singular and authentic entity where patients with fibrinolysis shutdown clot and bleed at the same time [12,13].

Finally, a recent review of the literature using the MeSH term COVID associated coagulopathy reveals that this term was a standard descriptor for the singular observation of these patients to develop early clots that required delicate and closely monitored anticoagulation [8,14–37].

Our second point concerns Bareille et al.'s statement that we have not emphasized enough the lack of interchangeability of the legacy and post-legacy devices. We state the following regarding the interchangeability of the TEG 5000 and the TEG6s on page 4, Table 1: "For the TEG® 6s and the TEG® 5000, the manufacturer (Haemonetics, Braintree, Massachusetts) states that any difference in the values is a consequence of the Clinical and Laboratory Standards Institute (CLSI) methodology used for the TEG® 6s values. Additionally, the manufacturer's ranges are not globally prescriptive, as every laboratory must establish their own reference intervals. The values for clinically significant fibrinolysis vary greatly in the literature".

They point out that the older as well as the post-legacy new generation VHAs are not interchangeable. We agree. We clearly state on page 5 the following in our paper regarding comparison of these two devices: "...the thresholds for intervention are not uniform and depend on local standards that determine the triggers for intervention. VHA guidance of BCTs, and pro-hemostatic agents with simplified algorithms, facilitate goal-directed BCT and HAT for bleeding and anticoagulated patients". We specifically address the idea of interchangeability with the following on page 6: "although the TEG® 6s and the TEG® 5000 use slightly different mechanisms to assess similar coagulation components, there is a small difference in absolute values of respective parameters of the two devices. Therefore, the absolute values are not interchangeable".

The authors are to be commended for their extensive review of the literature of the latest post-legacy generation devices and for their very complete table, which bolsters their argument as well as ours for the lack of interchangeability of these new devices. This table will be a useful instrument for those who wish to compare the efficacy of these devices much as our Table 3 on page 7, which compared the same devices to the cup and pin methods, will be. However, it remains the case that our Table 3 and the authors' table comparing the devices again manifest what has been a central tenet in the literature since the inception of the VH: that these VHA devices, whether legacy or post-legacy, cannot be used in a comparative setting and that each institution must establish its own standardization. In our legend for Table 3 in our manuscript, where we broadly describe the various types of legacy and post-legacy VHAs in our "primer" on VHAs, we refer to the literature that clearly states that the parameters for these many devices are not interchangeable.

On a technical point, regarding the comparison of legacy and post-legacy devices, the authors assert that we do not identify Sonoclot as a legacy system. However, we state on page 5: "The TEG® 5000 and the ROTEM® delta systems are considered cup and pin "legacy" systems. Another legacy linear motion system is called Sonoclot®; however, it is not a rotation based system".

In summary, we are very grateful for the commentary that Bareille et al. have given on our manuscript and we point out that for the special edition of the JCM devoted to VHAs our "primer" paper by Volod et al. was of central importance because of its purpose of providing an introduction for experts and nonexperts to viscoelastic testing in the clinical and research settings. We believe that Bareille et al.'s characterization of our enthusiasm

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as excessive contradicts the extraordinary experience and growth of these devices not just during the pandemic but also recently in the care of severe trauma patients and in patients with post-partum hemorrhage, (PPH), extracorporeal mechanical oxygenation (ECMO) as well as in surgical and medical critical care medicine [38–46].

The authors have also invoked the objection that before the widespread use of these VHA devices is undertaken we need larger RCTs to confirm their worth. We would like to point out that the activated patrial thromboplastin time (aPTT) test was developed around the time of TEG. If one peruses how aPTT and heparin therapeutic ranges were established, they would be surprised by the lack of large RCTs that established the norms for guiding heparin anticoagulation. Nevertheless, standard algorithms have been universally adopted by hematologists and other specialties.

In fact, the "1.5 to 2.5 times control" UFH therapeutic range was established is instructive regarding the history of RCTs in offering guidance for heparin anticoagulation. In 1972, a lower risk of recurrent thromboembolism occurrence among patients anticoagulated with heparin within an aPTT of 1.5 to 2.5 times the mean of the laboratory reference range was reported in the original retrospective analysis by Basu et al. [47]. An experimental rabbit model of thrombus extension supported the 1.5 to 2.5 therapeutic range in a subsequent study using results with the same aPTT reagents [48]. However, randomized controlled trials have not confirmed the clinical relevance of this therapeutic range. However, using a protamine–heparin titration assay and 0.35 to 0.70 IU/mL with a factor Xa assay, an aPTT of 1.5 to 2.5 times control was found to be equivalent to a heparin level of 0.2 to 0.4 IU/mL [49]. Levine et al. formed the basis for the recommendation of a 0.3 to 0.7 IU/mL therapeutic range for UFH using an anti-Xa assay by demonstrating this relationship in a randomized controlled trial study [45,49,50].

The more recent literature confirms the lack of large RCTs supporting specific algorithms regarding the aPPT or anti-Xa levels to guide heparin therapy. An authoritative review concerning guidance for the practical management of the heparin anticoagulants in the treatment of venous thromboembolism in 2016 stated that: "Despite the standard practice to monitor heparin through coagulation laboratory testing, the body of evidence supporting monitoring is surprisingly weak". As a result, the guidelines and algorithms are still in a state of evolution [51].

Bareille et al. also harken to the use of "rapid coagulation tests" as a substitute for the VHAs without noting that the very citations they use for these rapid tests (which claim the capability of obtaining actionable results in 20 min) have not been successfully reproduced despite the antiquated references by the authors dating from 2010 and 2013 [52,53].

Precision-based medicine (PBM) adopts the concept of describing a hemostatic phenotype defined by VHA and common coagulation tests (CCTs) to guide anticoagulation and blood component therapy (BCT) and hemostatic adjunctive therapy (HAT) in many clinical situations. The "one size fits all" methodology of awaiting large RCTs to confirm the validity of VHAs would have deprived the medical community of the rapid advances that have occurred during the pandemic and that led to the early approval of VHAs by the FDA for use in guiding the anticoagulation of COVID-19 patients [11,38,54–56].

We agree with the authors that large RCTs can be complementary in affirming the clinical utility of VHAs (provided appropriate data quality control). During the pandemic, clinicians who relied on an evidence-based medicine approach were forced to practice with evolving data concerning the escalation of anticoagulation in the COVID-19 patients who clearly presented with a very high rate of thrombosis [57]. Therefore, during this early period of the pandemic without the benefit of RCTs many institutions and hematologic societies provided diverse and evolving guidelines regarding the escalation of prophylactic anticoagulation to intermediate or therapeutic doses of heparin for patients with COVID-19 pneumonitis without macrovascular thrombosis, often relying on VHAs to assist in guiding the anticoagulation of these patients who, as we stated, would "clot and bleed at the same time" [58–74].

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However, as the COVID-19 pandemic has illustrated, the reliance on hypotheses generated by smaller studies during times of epidemiologic uncertainty has provided us with a unique opportunity to advance the cause of precision-based guidance for the treatment of patients with not just COVID-19-related coagulopathies but of all severe coagulopathies related to liver transplantation, cardiac surgery, trauma resuscitation, ECMO, PPH and other areas of medical and surgical coagulopathy where the assessment of a patient's hemostatic phenotype requires not just CCTs but also VHA analysis [44].

**Author Contributions:** Conceptualization, O.V., C.M.B., J.M., E.E.M., H.B.M., H.C.K., S.S.P., G.W., M.A., S.G.T., D.F., L.E., A.T. and M.M.W.; writing—original draft preparation, O.V., C.M.B., J.M., E.E.M., A.T. and M.M.W.; writing—review and editing, O.V., C.M.B., J.M., E.E.M., H.B.M., H.C.K., S.S.P., G.W., M.A., S.G.T., D.F., L.E., A.T. and M.M.W.; supervision, O.V., C.M.B., J.M., E.E.M. and M.M.W. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: The Authors would like to thank Matthew R. Walsh, for his editorial assistance.

Conflicts of Interest: The authors declare no conflict of interest.

### References

- 1. Bareille, M.; Lecompte, T.; Mullier, F.; Roullet, S. Are Viscoelastometric Assays of Old Generation Ready for Disposal? Comment on Volod et al. Viscoelastic Hemostatic Assays: A Primer on Legacy and New Generation Devices. *J. Clin. Med.* 2022, 11, 860. *J. Clin. Med.* 2023, 12, 477. [CrossRef]
- Volod, O.; Bunch, C.M.; Zackariya, N.; Moore, E.E.; Moore, H.B.; Kwaan, H.C.; Neal, M.D.; Al-Fadhl, M.D.; Patel, S.S.; Wiarda, G.; et al. Viscoelastic Hemostatic Assays: A Primer on Legacy and New Generation Devices. J. Clin. Med. 2022, 11, 860. [CrossRef] [PubMed]
- 3. Santos, F.; Marinho, R.; Carneiro, M.; Matos, R.; Pinho, N.; Moreira, L.; Seidi, N.; Coutinho, M.; Morais, S.; Cruz, E. Impact of COVID-19 associated coagulopathy in disease severity. *Res. Pract. Thromb. Haemost.* **2021**, *5* (Suppl. 2).
- 4. Kazi, S.; Othman, M.; Khoury, R.; Bernstein, P.; Thachil, J.; Ciantar, E.; Netto, M.; Ferrara, L.; Abdul-Kadir, R.; Malinowski, A. Isth registry on pregnancy and COVID-19 associated coagulopathy (COVPREG-COAG)-first report. *Res. Pract. Thromb. Haemost.* **2021**, *5* (Suppl. 2). [CrossRef]
- 5. Abdelaal, A.; Abu-Elfatth, A.; Bakkar, L.M.; El-Azeem, H.G.A.; Hetta, H.F.; Badawy, E.R. Assessment of COVID-19 associated coagulopathy and multiple hemostatic markers: A single center study in Egypt. *Infection* **2022**, 1–10. [CrossRef] [PubMed]
- 6. Thachil, J.; Tang, N.; Gando, S.; Falanga, A.; Cattaneo, M.; Levi, M.; Clark, C.; Iba, T. ISTH interim guidance on recognition and management of coagulopathy in COVID-19. *J. Thromb. Haemost.* **2020**, *18*, 1023–1026. [CrossRef]
- 7. Becker, R.C. COVID-19 update: Covid-19-associated coagulopathy. J. Thromb. Thrombolysis 2020, 50, 54–67. [CrossRef]
- 8. Leentjens, J.; van Haaps, T.F.; Wessels, P.F.; Schutgens, R.E.G.; Middeldorp, S. COVID-19-associated coagulopathy and antithrombotic agents-lessons after 1 year. *Lancet Haematol.* **2021**, *8*, e524–e533. [CrossRef]
- 9. Gerber, G.F.; Chaturvedi, S. How to recognize and manage COVID-19-associated coagulopathy. *Hematol. Am. Soc. Hematol. Educ. Program.* **2021**, 2021, 614–620. [CrossRef]
- Bunch, C.M.; Thomas, A.V.; Stillson, J.E.; Gillespie, L.; Khan, R.Z.; Zackariya, N.; Shariff, F.; Al-Fadhl, M.; Mjaess, N.; Miller, P.D.; et al. Preventing Thrombohemorrhagic Complications of Heparinized COVID-19 Patients Using Adjunctive Thromboelastography: A Retrospective Study. J. Clin. Med. 2021, 10, 3097. [CrossRef]
- 11. FDA. Coagulation Systems for Measurement of Viscoelastic Properties: Enforcement Policy During the Coronavirus Disease 2019 (COVID-19) Public Health Emergency: Guidance for Industry and Food and Drug Administration Staff; FDA: Silver Spring, MD, USA, January 2021.
- 12. Kwaan, H.C.; Lindholm, P.F. The Central Role of Fibrinolytic Response in COVID-19-A Hematologist's Perspective. *Int. J. Mol. Sci.* **2021**, 22, 1283. [CrossRef]
- 13. Walsh, M.M.; Khan, R.; Kwaan, H.C.; Neal, M.D. Fibrinolysis Shutdown in COVID-19-Associated Coagulopathy: A Crosstalk among Immunity, Coagulation, and Specialists in Medicine and Surgery. *J. Am. Coll. Surg.* **2021**, 232, 1003–1006. [CrossRef]
- 14. Asakura, H.; Ogawa, H. COVID-19-associated coagulopathy and disseminated intravascular coagulation. *Int. J. Hematol.* **2021**, 113, 45–57. [CrossRef] [PubMed]
- 15. Balagholi, S.; Dabbaghi, R.; Eshghi, P.; Mousavi, S.A.; Heshmati, F.; Mohammadi, S. Potential of therapeutic plasmapheresis in treatment of COVID-19 patients: Immunopathogenesis and coagulopathy. *Transfus. Apher. Sci.* **2020**, *59*, 102993. [CrossRef]
- 16. Cacciola, R.; Gentilini Cacciola, E.; Vecchio, V.; Cacciola, E. Cellular and molecular mechanisms in COVID-19 coagulopathy: Role of inflammation and endotheliopathy. *J. Thromb. Thrombolysis* **2022**, *53*, 282–290. [CrossRef] [PubMed]
- 17. Colling, M.E.; Kanthi, Y. COVID-19-associated coagulopathy: An exploration of mechanisms. *Vasc. Med.* **2020**, 25, 471–478. [CrossRef] [PubMed]

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18. Conway, E.M.; Mackman, N.; Warren, R.Q.; Wolberg, A.S.; Mosnier, L.O.; Campbell, R.A.; Gralinski, L.E.; Rondina, M.T.; van de Veerdonk, F.L.; Hoffmeister, K.M.; et al. Understanding COVID-19-associated coagulopathy. *Nat. Rev. Immunol.* **2022**, 22, 639–649. [CrossRef] [PubMed]

- 19. Elieh Ali Komi, D.; Rahimi, Y.; Asghari, R.; Jafari, R.; Rasouli, J.; Mohebalizadeh, M.; Abbasi, A.; Nejadrahim, R.; Rezazadeh, F.; Shafiei-Irannejad, V. Investigation of the Molecular Mechanism of Coagulopathy in Severe and Critical Patients With COVID-19. *Front. Immunol.* 2021, 12, 762782. [CrossRef]
- 20. Gando, S.; Wada, T. Thromboplasminflammation in COVID-19 Coagulopathy: Three Viewpoints for Diagnostic and Therapeutic Strategies. *Front. Immunol.* **2021**, 12, 649122. [CrossRef]
- 21. Görlinger, K.; Levy, J.H. COVID-19-associated Coagulopathy. Anesthesiology 2021, 134, 366–369. [CrossRef]
- 22. Gu, S.X.; Tyagi, T.; Jain, K.; Gu, V.W.; Lee, S.H.; Hwa, J.M.; Kwan, J.M.; Krause, D.S.; Lee, A.I.; Halene, S.; et al. Thrombocytopathy and endotheliopathy: Crucial contributors to COVID-19 thromboinflammation. *Nat. Rev. Cardiol.* **2021**, *18*, 194–209. [CrossRef]
- 23. Iba, T.; Connors, J.M.; Levy, J.H. The coagulopathy, endotheliopathy, and vasculitis of COVID-19. *Inflamm. Res.* **2020**, *69*, 1181–1189. [CrossRef] [PubMed]
- 24. Iba, T.; Levy, J.H.; Connors, J.M.; Warkentin, T.E.; Thachil, J.; Levi, M. The unique characteristics of COVID-19 coagulopathy. *Crit. Care* **2020**, 24, 360. [CrossRef] [PubMed]
- 25. Iba, T.; Levy, J.H.; Levi, M.; Connors, J.M.; Thachil, J. Coagulopathy of Coronavirus Disease 2019. *Crit. Care Med.* 2020, 48, 1358–1364. [CrossRef] [PubMed]
- 26. Inoue, K.I.; Sagawa, T.; Takano, H. COVID-19 and Coagulopathy. Am. J. Respir. Cell Mol. Biol. 2021, 64, 764. [CrossRef] [PubMed]
- 27. Karimzadeh, S.; Dong, V.; Hassan, O.G.; Raut, A.; Fouda, A.; Parrill, A.; Eaton, K.; Huy, N.T. Covid-19-induced coagulopathy and observed benefits with anticoagulation. *Transfus. Apher. Sci.* **2020**, *59*, 102906. [CrossRef]
- 28. Katneni, U.K.; Alexaki, A.; Hunt, R.C.; Schiller, T.; DiCuccio, M.; Buehler, P.W.; Ibla, J.C.; Kimchi-Sarfaty, C. Coagulopathy and Thrombosis as a Result of Severe COVID-19 Infection: A Microvascular Focus. *Thromb. Haemost.* **2020**, 120, 1668–1679. [CrossRef]
- 29. Kohansal Vajari, M.; Shirin, M.; Pourbagheri-Sigaroodi, A.; Akbari, M.E.; Abolghasemi, H.; Bashash, D. COVID-19-related coagulopathy: A review of pathophysiology and pharmaceutical management. *Cell Biol. Int.* **2021**, *45*, 1832–1850. [CrossRef]
- 30. Lazzaroni, M.G.; Piantoni, S.; Masneri, S.; Garrafa, E.; Martini, G.; Tincani, A.; Andreoli, L.; Franceschini, F. Coagulation dysfunction in COVID-19: The interplay between inflammation, viral infection and the coagulation system. *Blood Rev.* **2021**, 46, 100745. [CrossRef]
- 31. Lippi, G.; Sanchis-Gomar, F.; Favaloro, E.J.; Lavie, C.J.; Henry, B.M. Coronavirus Disease 2019-Associated Coagulopathy. *Mayo Clin. Proc.* 2021, 96, 203–217. [CrossRef]
- 32. Memar Montazerin, S.; Najafi, H.; Shojaei, F.; Chi, G.; Goudarzi, S.; Sharfaei, S.; Kahe, F.; Hassanzadeh, S. COVID-19-associated coagulopathy: A concise review on pathogenesis and clinical implications. *Infez Med.* **2021**, 29, 1–9.
- 33. Mitchell, W.B.; Davila, J.; Keenan, J.; Jackson, J.; Tal, A.; Morrone, K.A.; Silver, E.J.; O'Brien, S.; Manwani, D. Children and young adults hospitalized for severe COVID-19 exhibit thrombotic coagulopathy. *Pediatr. Blood Cancer* **2021**, *68*, e28975. [CrossRef] [PubMed]
- 34. Perico, L.; Benigni, A.; Casiraghi, F.; Ng, L.F.P.; Renia, L.; Remuzzi, G. Immunity, endothelial injury and complement-induced coagulopathy in COVID-19. *Nat. Rev. Nephrol.* **2021**, 17, 46–64. [CrossRef]
- 35. Plášek, J.; Gumulec, J.; Máca, J.; Škarda, J.; Procházka, V.; Grézl, T.; Václavík, J. COVID-19 associated coagulopathy: Mechanisms and host-directed treatment. *Am. J. Med. Sci.* **2022**, *363*, 465–475. [CrossRef] [PubMed]
- 36. Salabei, J.K.; Fishman, T.J.; Asnake, Z.T.; Ali, A.; Iyer, U.G. COVID-19 Coagulopathy: Current knowledge and guidelines on anticoagulation. *Heart Lung* **2021**, *50*, 357–360. [CrossRef] [PubMed]
- 37. Zhang, J.; Tecson, K.M.; McCullough, P.A. Endothelial dysfunction contributes to COVID-19-associated vascular inflammation and coagulopathy. *Rev. Cardiovasc. Med.* **2020**, *21*, 315–319. [CrossRef] [PubMed]
- 38. Walsh, M.; Thomas, S.; Kwaan, H.; Aversa, J.; Anderson, S.; Sundararajan, R.; Zimmer, D.; Bunch, C.; Stillson, J.; Draxler, D. Modern methods for monitoring hemorrhagic resuscitation in the United States: Why the delay? *J. Trauma Acute Care Surg.* **2020**, 89, 1018–1022. [CrossRef]
- 39. Bugaev, N.; Como, J.J.; Golani, G.; Freeman, J.J.; Sawhney, J.S.; Vatsaas, C.J.; Yorkgitis, B.K.; Kreiner, L.A.; Garcia, N.M.; Aziz, H.A. Thromboelastography and Rotational Thromboelastometry in Bleeding Patients with Coagulopathy: Practice Management Guideline from the Eastern Association for the Surgery of Trauma. *J. Trauma Acute Care Surg.* 2020, 89, 999–1017. [CrossRef]
- 40. Bell, S.; Roberts, T.; Pereira, J.F.M.; De Lloyd, L.; Amir, Z.; James, D.; Jenkins, P.; Collis, R.; Collins, P. The sensitivity and specificity of rotational thromboelastometry (ROTEM) to detect coagulopathy during moderate and severe postpartum haemorrhage: A prospective observational study. *Int. J. Obstet. Anesth.* 2022, 49, 103238. [CrossRef]
- 41. Curry, N.S.; Davenport, R. Transfusion strategies for major haemorrhage in trauma. Br. J. Haematol. 2019, 184, 508–523. [CrossRef]
- 42. Spahn, D.R.; Bouillon, B.; Cerny, V.; Duranteau, J.; Filipescu, D.; Hunt, B.J.; Komadina, R.; Maegele, M.; Nardi, G.; Riddez, L. The European guideline on management of major bleeding and coagulopathy following trauma. *Crit. Care* 2019, 23, 1–74. [CrossRef]
- 43. Curry, N.S.; Davenport, R.; Pavord, S.; Mallett, S.V.; Kitchen, D.; Klein, A.A.; Maybury, H.; Collins, P.W.; Laffan, M. The use of viscoelastic haemostatic assays in the management of major bleeding: A British Society for Haematology Guideline. *Br. J. Haematol.* 2018, 182, 789–806. [CrossRef] [PubMed]

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44. Bunch, C.M.; Berquist, M.; Ansari, A.; McCoy, M.L.; Langford, J.H.; Brenner, T.J.; Aboukhaled, M.; Thomas, S.J.; Peck, E.; Patel, S.; et al. The Choice between Plasma-Based Common Coagulation Tests and Cell-Based Viscoelastic Tests in Monitoring Hemostatic Competence: Not an either-or Proposition. *Semin. Thromb. Hemost.* 2022, 48, 769–784. [CrossRef] [PubMed]

- 45. Volod, O.; Wegner, J. Viscoelastic Testing in the Management of Adult Patients on Mechanical Circulatory Support Devices with Focus on Extracorporeal Membrane Oxygenation. *Semin. Thromb. Hemost.* **2022**, *48*, 814–827. [CrossRef] [PubMed]
- 46. Collis, R.; Bell, S. The Role of Thromboelastography during the Management of Postpartum Hemorrhage: Background, Evidence, and Practical Application. *Semin. Thromb. Hemost.* 2022; ahead of print. [CrossRef]
- 47. Basu, D.; Gallus, A.; Hirsh, J.; Cade, J. A prospective study of the value of monitoring heparin treatment with the activated partial thromboplastin time. *N. Engl. J. Med.* **1972**, 287, 324–327. [CrossRef]
- 48. Chiu, H.M.; Hirsh, J.; Yung, W.L.; Regoeczi, E.; Gent, M. Relationship between the anticoagulant and antithrombotic effects of heparin in experimental venous thrombosis. *Blood* **1977**, *49*, 171–184. [CrossRef]
- 49. Levine, M.N.; Hirsh, J.; Gent, M.; Turpie, A.G.; Cruickshank, M.; Weitz, J.; Anderson, D.; Johnson, M. A randomized trial comparing activated thromboplastin time with heparin assay in patients with acute venous thromboembolism requiring large daily doses of heparin. *Arch. Intern. Med.* **1994**, 154, 49–56. [CrossRef]
- 50. Olson, J.D.; Arkin, C.F.; Brandt, J.T.; Cunningham, M.T.; Giles, A.; Koepke, J.A.; Witte, D.L. College of American Pathologists Conference XXXI on laboratory monitoring of anticoagulant therapy. Laboratory monitoring of unfractionated heparin therapy. *Arch. Pathol Lab. Med.* **1998**, 122, 782–798.
- 51. Smythe, M.A.; Priziola, J.; Dobesh, P.P.; Wirth, D.; Cuker, A.; Wittkowsky, A.K. Guidance for the practical management of the heparin anticoagulants in the treatment of venous thromboembolism. *J. Thromb. Thrombolysis* **2016**, *41*, 165–186. [CrossRef]
- 52. Chandler, W.L.; Ferrell, C.; Trimble, S.; Moody, S. Development of a rapid emergency hemorrhage panel. *Transfusion* **2010**, *50*, 2547–2552. [CrossRef]
- 53. Chandler, W.L. Emergency assessment of hemostasis in the bleeding patient. Int. J. Lab. Hematol. 2013, 35, 339–343. [CrossRef]
- 54. Görlinger, K.; Almutawah, H.; Almutawaa, F.; Alwabari, M.; Alsultan, Z.; Almajed, J.; Alwabari, M.; Alsultan, M.; Shahwar, D.; Yassen, K.A. The role of rotational thromboelastometry during the COVID-19 pandemic: A narrative review. *Korean J. Anesth.* **2021**, *74*, 91. [CrossRef] [PubMed]
- 55. Stettler, G.R.; Moore, E.E.; Moore, H.B.; Nunns, G.R.; Silliman, C.C.; Banerjee, A.; Sauaia, A. Redefining postinjury fibrinolysis phenotypes using two viscoelastic assays. *J. Trauma Acute Care Surg.* **2019**, *86*, 679–685. [CrossRef] [PubMed]
- 56. Walsh, M.; Grisoli, A.; Zackariya, N.; Thomas, A.V.; Sualeh, A. Randomized controlled trials and Cochrane analyses versus precision-based medicine for tranexamic acid and viscoelastic testing in trauma. *ANZ J. Surg.* **2020**, *90*, 415–416. [CrossRef] [PubMed]
- 57. Connors, J.M.; Levy, J.H. Thromboinflammation and the hypercoagulability of COVID-19. *J. Thromb. Haemost.* **2020**, *18*, 1559–1561. [CrossRef]
- 58. Lee, A.Y.; Connors, J.M.; Baumann Kreuziger, L.; Murphy, M.; Gernsheimer, T.; Lin, Y.; Huisman, M.; DeSancho, M. COVID-19 and Coagulopathy: Frequently Asked Questions. Available online: https://www.hematology.org/covid-19/covid-19-and-coagulopathy (accessed on 26 January 2021).
- 59. Bikdeli, B.; Madhavan, M.V.; Jimenez, D.; Chuich, T.; Dreyfus, I.; Driggin, E.; Nigoghossian, C.; Ageno, W.; Madjid, M.; Guo, Y.; et al. COVID-19 and Thrombotic or Thromboembolic Disease: Implications for Prevention, Antithrombotic Therapy, and Follow-Up: JACC State-of-the-Art Review. *J. Am. Coll Cardiol.* 2020, 75, 2950–2973. [CrossRef]
- 60. Spyropoulos, A.C.; Levy, J.H.; Ageno, W.; Connors, J.M.; Hunt, B.J.; Iba, T.; Levi, M.; Samama, C.M.; Thachil, J.; Giannis, D. Scientific and Standardization Committee communication: Clinical guidance on the diagnosis, prevention, and treatment of venous thromboembolism in hospitalized patients with COVID-19. *J. Thromb. Haemost.* **2020**, *18*, 1859–1865. [CrossRef]
- 61. Gomez, K.; Laffan, M.; Bradbury, C. Debate: Should the dose or duration of anticoagulants for the prevention of venous thrombosis be increased in patients with COVID-19 while we are awaiting the results of clinical trials? *Br. J. Anaesth* **2020**, 192, 459. [CrossRef]
- 62. Lemos, A.C.B.; do Espírito Santo, D.A.; Salvetti, M.C.; Gilio, R.N.; Agra, L.B.; Pazin-Filho, A.; Miranda, C.H. Therapeutic versus prophylactic anticoagulation for severe COVID-19: A randomized phase II clinical trial (HESACOVID). *Thromb. Res.* **2020**, *196*, 359–366. [CrossRef]
- 63. Cannegieter, S.C.; Klok, F.A. COVID-19 associated coagulopathy and thromboembolic disease: Commentary on an interim expert guidance. *Res. Pr. Thromb. Haemost.* **2020**, *4*, 439–445. [CrossRef]
- 64. Shah, A.; Donovan, K.; McHugh, A.; Pandey, M.; Aaron, L.; Bradbury, C.A.; Stanworth, S.J.; Alikhan, R.; Von Kier, S.; Maher, K. Thrombotic and haemorrhagic complications in critically ill patients with COVID-19: A multicentre observational study. *Crit. Care* 2020, 24, 1–10. [CrossRef]
- 65. Barnes, G.D.; Burnett, A.; Allen, A.; Blumenstein, M.; Clark, N.P.; Cuker, A.; Dager, W.E.; Deitelzweig, S.B.; Ellsworth, S.; Garcia, D.; et al. Thromboembolism and anticoagulant therapy during the COVID-19 pandemic: Interim clinical guidance from the anticoagulation forum. *J. Thromb. Thrombolysis* **2020**, *50*, 72–81. [CrossRef] [PubMed]
- 66. Demelo-Rodriguez, P.; Farfán-Sedano, A.I.; Pedrajas, J.M.; Llamas, P.; Sigüenza, P.; Jaras, M.J.; Quintana-Diaz, M.; Fernández-Capitán, C.; Bikdeli, B.; Jiménez, D. Bleeding risk in hospitalized patients with COVID-19 receiving intermediate-or therapeutic doses of thromboprophylaxis. *J. Thromb. Haemost.* 2021; ahead of print. [CrossRef]

J. Clin. Med. **2023**, 12, 478

67. Hartmann, J.; Ergang, A.; Mason, D.; Dias, J.D. The Role of TEG Analysis in Patients with COVID-19-Associated Coagulopathy: A Systematic Review. *Diagnostics* **2021**, *11*, 172. [CrossRef]

- 68. NIH ACTIV Trial of Blood Thinners Pauses Enrollment of Critically Ill COVID-19 Patients. Available online: https://www.nih.gov/news-events/news-releases/nih-activ-trial-blood-thinners-pauses-enrollment-critically-ill-covid-19-patients (accessed on 26 January 2021).
- Full-Dose Blood Thinners Decreased Need for Life Support and Improved Outcome in Hospitalized COVID-19 Patients. Available
  online: https://www.nih.gov/news-events/news-releases/full-dose-blood-thinners-decreased-need-life-support-improvedoutcome-hospitalized-covid-19-patients (accessed on 26 January 2021).
- 70. Ten Cate, H. Surviving COVID-19 with Heparin? N. Engl. J. Med. 2021, 385, 845–846. [CrossRef]
- 71. Słomka, A.; Kowalewski, M.; Żekanowska, E. Hemostasis in Coronavirus Disease 2019-Lesson from Viscoelastic Methods: A Systematic Review. *Thromb. Haemost.* **2021**, *121*, 1181–1192. [CrossRef] [PubMed]
- 72. Dauwerse, S.; Ten Cate, H.; Spronk, H.M.H.; Nagy, M. The Composition and Physical Properties of Clots in COVID-19 Pathology. *Diagnostics* **2022**, 12, 580. [CrossRef] [PubMed]
- 73. Lawler, P.R.; Goligher, E.C.; Berger, J.S.; Neal, M.D.; McVerry, B.J.; Nicolau, J.C.; Gong, M.N.; Carrier, M.; Rosenson, R.S.; Reynolds, H.R.; et al. Therapeutic Anticoagulation with Heparin in Noncritically Ill Patients with Covid-19. *N. Engl. J. Med.* **2021**, 385, 790–802. [CrossRef]
- 74. Goligher, E.C.; Bradbury, C.A.; McVerry, B.J.; Lawler, P.R.; Berger, J.S.; Gong, M.N.; Carrier, M.; Reynolds, H.R.; Kumar, A.; Turgeon, A.F.; et al. Therapeutic Anticoagulation with Heparin in Critically Ill Patients with COVID-19. *N. Engl. J. Med.* **2021**, *385*, 777–789. [CrossRef]

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