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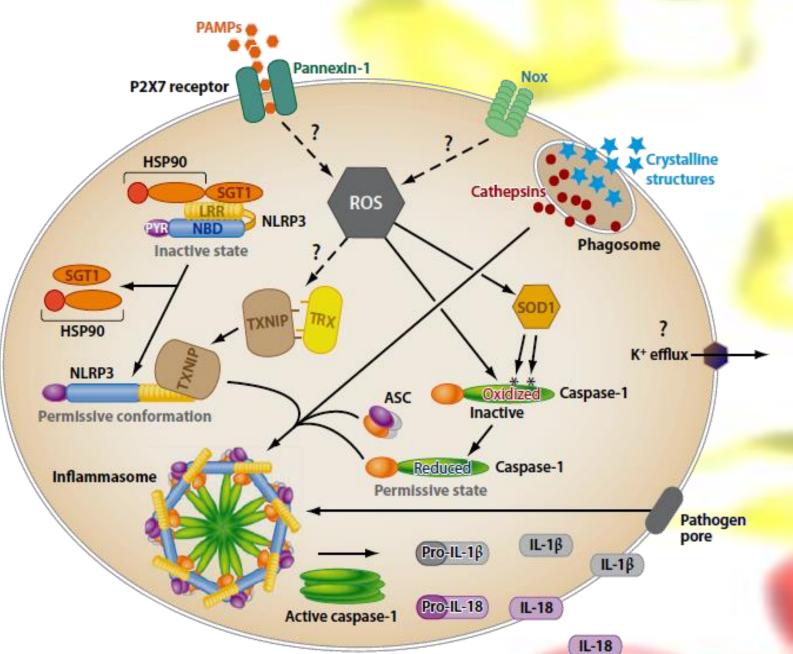
Synthesis and preliminary evaluation of model compounds targeting the NLRP3 inflammasome pathways



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Introduction

Inflammasomes have recently emerged as key mediators of inflammation and immunity. Four inflammasome complexes have been described to date. The most intensely studied is the NLRP3 inflammasome formed by the nucleotide-binding domain leucine-rich repeat family member NLRP3 and the adapter protein ASC, its assembling is triggered by a diverse series of endogenous and exogenous stimuli. (figure 1).



NLRP3 assembly leads to caspase-1 activation which causes the maturation and secretion of the proinflammatory cytokines IL-1β and IL-18.

Fig. 1. Model of inflammasome activation.Coordination of a manifold series of signals culminates in the activation of the inflammasome.

A series of autosomal and dominant or *de novo* mutations of nlrp3 lead to auto-inflammatory syndromes known as CAPS (cryopyrin–associated periodic syndromes) which are characterized by high levels of IL-1 β release and associated sterile systemic inflammation. The onset of chronic inflammation has also been linked to a wide range of metabolic disorders, such as type 2 diabetes, atherosclerosis, and Alzheimer's disease.²

Design of inhibitors of NLRP3-related pathways

Few small molecules inhibitors of NLRP3 inflammasome-related effects have been described, among them parthenolide, Bay 11-7082 and bromoxone are the most studied.

These molecules share the ability to behave as Michael acceptors. Although Michael acceptors are traditionally shunned in modern drug discovery, many biologically relevant and druggable pathways are targeted by thiol-reactive compounds.³

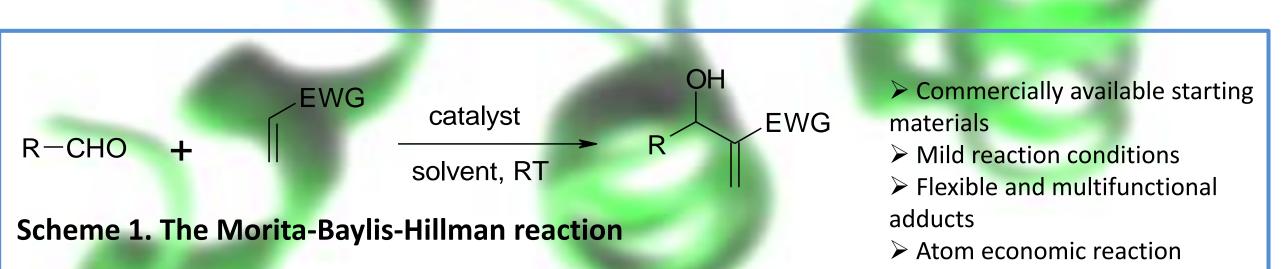
To explore the use of Michael acceptors as pharmaco-chemical tools modulating inflammatory pathways we designed some model compounds (figure 2).

X = alkyl, cycloalkyl, aryl, substd aryl, heteroaryl

Fig. 2. General structure of designed Michael acceptors

Synthesis

Designed compounds were synthesized via the Morita-Baylis-Hillman (MBH) reaction (scheme 1).

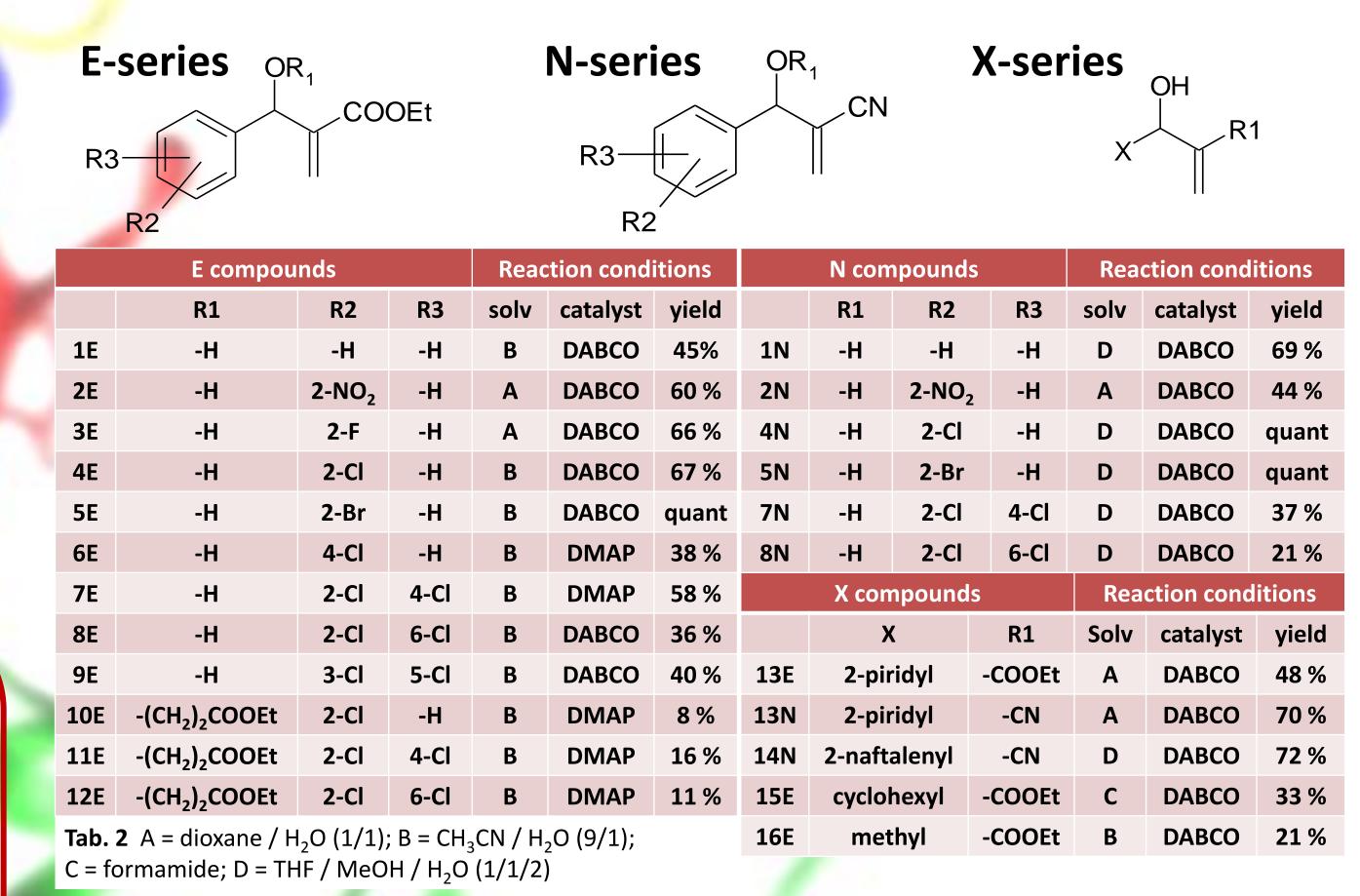


COOEt **COOEt** catalyst (1mmol) solvent, RT 1 mmol 3 mmol catalyst Time Yield a Yield a Solvent Solvent catalyst Time ✓ Use of polar protic (d) (d) CHCl₃ Et₃N CH₃CN/H₂O Et₃N medium is preferred **DMAP** 42 % CHCl₃ DMAP CH₃CN/H₂O 5 54 % CHCl₃ CH₃CN/H₂O 20 DBU DBU 25 % trace √ Use of DABCO or CHCl₃ CH₃CN/H₂O 20 < 2% lm lm **DMAP** gives best results 67 % CHCl₃ CH₃CN/H₂O DABCO **DABCO**

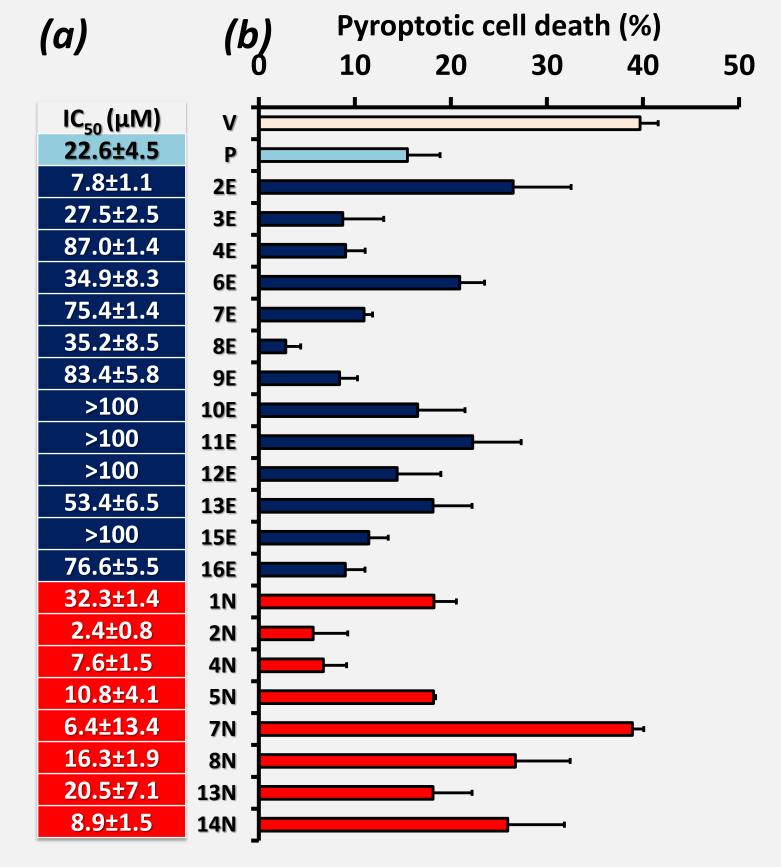
Optimisation of reaction conditions

CHCl₃ CH₃CN/H₂O PPh₃ PPh₃ 4 ✓ Long reaction times CHCl₃ **TMEDA** CH₃CN/H₂O TMEDA 20 trace are required 20 Dioxane/H₂O DABCO **Tab. 1** a) Isolated yields; b) reaction run on 10 mmol scale; c) complex THF/MeOH/H₂O DABCO 44 % mixture; d) run at 80 °C Solvents: ✓ Scale-up can be 65 % b CH₃CN/H₂O DABCO dioxane/ $H_2O(1/1)$; effectively obtained 40 % k CH₃CN/H₂O DABCO $CH_3CN/H_2O(9/1);$ THF/MeOH/H₂O (1/1/2)

The synthesis of **E- N- X-** series of compounds was obtained through reaction of ethylacrylate or acrylonitrile with different aldehydes.



Biological results



(a) Cytotoxicity

Immortalized human tubular epithelial cells were exposed to increasing compound concentrations (0.1-100 μM). After 72 h cell viability was evaluated by MTT assay. Data are means ± S.E.M. of at least three independent experiments run in quadruplicate.

(b) Pyroptotic cell death

PMA-differentiated THP-1 cells were stimulated with LPS. Cells were treated with vehicle alone (V), parthenolide (P) or synthesized compounds (10 µM; 1 h), then pulsed with ATP in serum-free medium. The culture supernatants were collected and assayed for LDH activity. Data are means ± S.E.M. of at least three independent experiments run in triplicate.

- ➤ Preliminary data identify a number of compounds able to inhibit pyroptotic cell death which is recognized to be related to NALP3 activation.
- Compounds **4E**, **8E**, **9E**, **16E** showed a better profile with respect to parthenolide, used as reference compound.

References

1) D. De Nardo et al. *Trends Immunol.* **2011**, *32*(*8*), 373-379. **2)** B. K. Davis et al. *Annu. Rev. Immunol.* **2011**, *29*, 707-735. **3)** S. Amslinger *ChemMedChem* **2010**, *5*, *351-356*.