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# Combining ability and heterosis studies for grain iron and zinc concentrations in pearl millet [*Cenchrus americanus* (L). Morrone]

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Iron (Fe) and zinc (Zn) deficiency has been identified as a major food-related health issue, affecting two billion people globally. Efforts to enhance the Fe and Zn content in food grains through plant breeding are an economic and sustainable solution to combat micronutrient deficiency in resource-poor populace of Asia and Africa. Pearl millet, Cenchrus americanus (L). Morrone, considered as a hardy nutri-cereal, is the major food crop for millions of people of these nations. As an effort to enhance its grain mineral content, an investigation was conducted using line x tester analysis to generate information on the extent of heterosis, gene action, combining ability for grain yield potential, and grain mineral nutrients (Fe and Zn). The partitioning of variance attributable to parents indicated that the lines and testers differed significantly for the traits studied. For most of the attributes, hybrids that were superior to the parents in the desired direction in terms of per se performance were identified. The analysis of combining ability variance indicated the preponderance of both additive and non-additive genetic effects. Thus, reciprocal recurrent selection can be used to develop a population with highgrain Fe and Zn contents. The Fe and Zn content in grain exhibited a highly significant and positive association between them, whereas the Fe and Zn contents individually showed a negative, albeit weak, correlation with grain yield and a moderate positive relation with grain weight. This indicates that mineral nutrient contents in grains can be improved without significant compromise on yield. The consistency of these trends across the environment suggests that these findings could be directly used as guiding principles for the genetic enhancement of Fe and Zn grain content in pearl millet.

#### KEYWORDS

combining ability, grain iron and zinc density, gene action, heterosis,  $\mathsf{L}\times\mathsf{T}$  mating design, pearl millet

# **1** Introduction

Micronutrient malnutrition or hidden hunger results from the deficiency of one or more micronutrients. Globally, about two-thirds of the population is reported to be suffering from this hidden hunger due to the dearth of essential micronutrients in their diet (Kumar et al., 2018). Cereal grains are the main source of proteins, carbohydrates, and micronutrients in human nutrition. The human body requires about two dozen of essential elements to ensure optimal health. For humans, several beneficial mineral elements such as iron (Fe), zinc (Zn), boron (B), copper (Cu), manganese (Mn), and cobalt (Co) are necessary in trace quantities (Kumar et al., 2015). Of these, Fe and Zn are primarily deficient in human diets (Mahendrakar et al., 2019). Mineral malnutrition is a major diet-related health issue for people in the developing world due to inadequate access to and affordability of horticultural products (fruits and vegetables) and meat. Reliance on carbohydrate dominated cereal grain-based diets further exacerbates the problem. The consequences of malnutrition can be catastrophic, resulting in mental retardation, impaired fitness, low productivity, lethargy, and decreased stamina (Kumar et al., 2016).

Fe and Zn requirements can be fulfilled through several ways. Food fortification and dietary supplementation are commonly adopted strategies to address malnutrition. However, these solutions are impractical and entail a significant expenditure (Jeong and Guerinot, 2008). Crop biofortification *via* plant breeding techniques has a potential to solve the issue of Fe-Zn deficiency in a sustainable and lucrative manner. Biofortification through genetic improvement has resulted in tangible increase in Fe levels of many crops like corn (*Zea mays*), wheat (*Triticum aestivum*), sorghum (*Sorghum bicolor*), pearl millet (*Cenchrus americanus*), and cowpea (*Vigna unguiculata*).

Pearl millet is the most important cereal crops next to rice, wheat, corn, barley, and sorghum with a very high photosynthetic efficiency and dry matter output being a C<sub>4</sub> species. It can be cultivated in even the most unfavorable agro-ecologies where other cereals, such as rice or wheat, do not thrive and fail to generate economic outputs. It can play a significant role not just contributing to the food and nutritional welfare of the poor in the pearl millet-growing areas of India and Sub-Saharan Africa but can have a potential health benefit for the wealthier as well (Rai et al., 2012). India is the largest producer of pearl millet in the world with an area of 6.93 million ha with an average production of 8.61 million tones and productivity of 1,243 kg/ ha during 2018-2019 (Directorate of Millets Development, 2020). It is a rich source of fiber and minerals-notably Fe, Zn, and calciumthus, rightly said as "nutri-cereal". Despite these health benefits, it is less popular as a staple due to lack of awareness about its nutritious value, proper processing technologies, and remunerative prices for farmers. This is apparent from the geographical distribution of pearl millet intake across India. About 19% to 63% of the total intake of Fe and 16% to 56% of the total intake of Zn was restricted to the states of Rajasthan, Maharashtra, and Gujarat (Rao et al., 2006). High genetic diversity for Fe and Zn available in breeding lines, improved populations, and germplasm were already reported in pearl millet offers good opportunities for breeding pearl millet varieties with increased levels of these micronutrients (Velu et al., 2007; Velu et al., 2008; Rai et al., 2012).

Combining ability analysis is considered helpful in determining the right combination of parents, which would generate heterotic hybrid and superior segregants in the succeeding generations. Such studies also throw some light on nature and magnitude of the gene action involved in the inheritance of target traits, which further dictate the breeding method to be adopted in segregating generations for trait improvement. There are a number of strategies for determining the breeding value of lines in terms of its combining ability and genetic composition. The line × tester study suggested by Kempthorne (1957) is the widely used method to determine the combining ability of the parent and general combining ability (GCA) and specific combining ability (SCA) effects and variances for crosses of various traits. Bearing this in mind, the present research work was undertaken to elucidate the combining ability of grain minerals and yield components to ascertain the nature and magnitude of gene actions of parental lines and hybrids developed in a line × tester design.

# 2 Materials and methods

## 2.1 Plant materials

Three CMS lines along with 10 restorers (testers) of pearl millet were used for the present study. These three CMS lines were crossed with 10 restorers in line  $\times$  tester mating design as suggested by Kempthorne (1957) and reviewed by Dabholkar (1992), yielding 30 hybrids in *Kharif* (2018). The details of parents along with their pedigree are given in Table 1. Thus, a total of 44 genotypes including 13 parents, 30 crosses, and one check (Pusa1201) constituted the experimental material for combining ability study. The parentals used for hybrid development were of diverse origin with a wide variation for grain Fe and Zn densities, grain yield, and other agronomic traits.

## 2.2 Field trials

### 2.2.1 Crossing

Three A-lines (ICMA-02555, ICMA-07999, and ICMA-12222) were crossed with 10 diverse R-lines at IARI, New Delhi during *Kharif* (2018). Individual healthy plants were tagged and used for making plant  $\times$  plant crosses to produce the F<sub>1</sub>s. Before stigma emergence both the male and female parents were covered with brown bags. On the day of crossing, bagged panicles of female parents were checked for complete stigma emergence and bagged male parent panicles for pollen shedding. Pollens from target parent were collected between 10:00 am and 11:30 am and were dusted on the female parent's panicle in which stigma has fully emerged. The pollinated panicle was immediately covered to prevent outcrossing. The details of cross such as parents and date of pollination were mentioned on the tag. Crossed panicles were finally collected at maturity (40 days after pollination) and dried. Then, panicles were threshed to get F<sub>1</sub> (hybrid) seed.

#### 2.2.2 Evaluation of parents and hybrids

Hybrids and parents were planted during the summer season 2019 (E1) at ICRISAT, Patancheru, and during the rainy season 2019

TABLE 1 Origin and parentage of B-lines (maintainer counterparts of A-lines) and restorer lines (R-lines).

SI no.	Line	Parentage
B lines		
1	ICMB-02555	CMV87901-175-2-3-2-B-1
2	ICMB-07999	(HTBC 48-B-1-1-1-5 × B-bulk)-25-1-B-B
3	ICMB-12222	(ICMB 95444 × ICMB 92111)-4-B-4-3-B-B
R lines		
4	A5RT-17/4	(IPC 107 × ICMV 91059 S1-14-2-1-1-2)-6-2-5-B-B-6-1
5	A5RT-17/5	(IPC 1617 × SDMV 90031-S1-84-1-1-1)-28-1-1-2-B-B-2-1-3
6	A5RT-17/6	(IPC 1617 × SDMV 90031-S1-84-1-1-1)-28-1-1-2-B-B-2-2-1
7	A5RT-17/7	(IPC 1617 × SDMV 90031-S1-84-1-1-1)-28-1-1-2-B-B-2-3-1
8	A5RT-17/8	(IPC 1617 × SDMV 90031-S1-84-1-1-1)-28-1-1-2-B-B-2-6-1
9	A5RT-17/16	(IPC 1268 × ICMV 91059 S1-58-2-2-2-1)-7-3-4-1-B-B-B-4-1
10	A5RT-17/19	(IPC 107 × ICMV 91059 S1-14-2-1-1-2)-19-1-2-2-3-B-5-2-1
11	A5RT-17/25	[(IPC 337 × SDMV 90031-S1-84-1-1-1) × RCB-2-S1-144-2-2-2-1-1-1]-2-4-2-3-3-B-B-3-1
12	A5RT-17/26	[(IPC 337 × SDMV 90031-S1-84-1-1-1) × RCB-2-S1-144-2-2-2-1-1-1]-2-4-2-3-3-B-B-5-4
13	A5RT-17/29	(IPC 107 × SDMV 90031-S1-84-1-1-1)-1-1-2-2-B-B-B

(E2) at IARI, New Delhi. A replicated (twice) randomized complete block design was followed at both the locations. Each genotype was sown in two rows of 2 m length with a spacing of 75 cm between rows in the rainy season in 2019 and 60 cm in the summer season in 2019. The recommended agronomic practices for healthy crop growth were adopted. Panicles were harvested at maturity from five randomly selected plants, sun-dried to appropriate moisture content (10%– 13%) followed by manual threshing for the preparation of grain samples. After taking the grain weight [considered as grain yield per plant (GYPP) in grams], 1000-grain weight were counted from each sample and weighed to estimate 1,000-grain weight. The average value of five plants per each replication was used for downstream analysis. At each step care, it was ensured that grain did not come in contact with metal containers or instruments to generate reliable estimates of micronutrients.

# 2.3 Estimation of grain iron, zinc, and phytate contents

Fe and Zn contents in cleaned grain samples were measured using energy-dispersive x-ray fluorescence (XRF). The XRF machine model X-Supreme 8000 (M/s Oxford Inc., USA) was used for the purpose of following the method by Govindaraj et al. (2016). Phytic acid (phytate) was determined by an assay unique to the measurement of phosphorous release based on available phosphorous from phytate, myo-inositol (phosphate)n, and monophosphate esters by phytase and alkaline phosphatase using the Phytic Acid/Total Phosphorus Assay Kit (Megazyme, 2016).

## 2.4 Molar ratio of phytate/mineral

The molar ratios of Phy/mineral were first determined by converting the quantities of phytic acid, Fe/Zn in mg  $g^{-1}$  to mmol  $g^{-1}$  using a molecular mass unit (phytate, 660.8 g/mol; Fe, 55.8 g/mol; Zn, 65.38 g/mol). These values were then used to derive corresponding molar ratios.

## 2.5 Statistical analysis

Replication-wise data of all the traits were analyzed using the software Analysis of Genetic Designs with R for Windows (AGD-R) (Rodríguez et al., 2015). Line × tester model for female × male hybrid was executed to generate estimates of the GCA for lines and testers as well as the SCA effects for hybrids. Furthermore, a mixed-effect model was adopted with genotypes (lines, testers, and hybrids) and environments as fixed effects, and interactions (site × line, site × tester, line × tester, and site × line × tester) and replication as random effects. The ANOVA model considered for the combining ability analysis through line × tester design is as follows (Arunachalam, 1974; Dabholkar, 1999):

$$Y_{ijk} = \mu + g_i + g_j + s_{ij} + r_k + e_{ijk}$$

where  $Y_{ijk}$  is the mean trait value measured on cross made between ith line and jth tester in the kth replication;  $\mu$  is the grand mean;  $g_i$  is the GCA effect of the ith parent;  $g_j$  is the GCA effect of the parent j;  $s_{ij}$  is the SCA effect of cross i  $\times$  j;  $r_k$  is the replication effect; and  $e_{ijk}$  is the environmental effect. Three estimates of heterosis, i.e., mid-parent

heterosis (MPH), better-parent heterosis (BPH), and standard heterosis (SH), were calculated for individual environments and the mean of two environments (Hallauer and Miranda, 1981). The tests of significance for MPH and BPH were done *via* "t" test.

# **3** Results

### 3.1 Genetic variability

Analysis of variance for combining ability (Table 2) revealed that the variance due to lines and tester was significant for all characters under recorded at both environments and combined over environment except for the grain yield per plant in summer. The source of variance due to line  $\times$  tester interaction was also significant for all traits in the individual environments and pooled environment except for grain yield per plant in summer season. Variance for the environments was highly significant for all the characters under study except for grain phytate content.

The variance due to hybrids and their interaction with environments (H  $\times$  E) were highly significant for all characters. However, the contribution of  $H \times E$  interactions to the variability relative to those due to genetic differences among the hybrids was low for grain Fe content (24%), grain Zn content (23%), grain yield per plant (40%), grain phytate contents (20%), phytate:Fe molar ratio (21%), and phytate: Zn molar ratio (21.4%) and higher for 1000-grain weight (50.3%). Further partitioning of H × E interactions showed that L × E interactions contributed more to the variability relative to line effects, which were 15% for grain Fe content, 19% for grain Zn content, 35% for grain yield per plant, 0.5% for grain phytate contents, 0.2% for phytate:Fe molar ratio, and 0.2% for phytate:Zn molar ratio. However, for thousand grain weight, line effects contributed 42% more to the variability relative to L  $\times$  E interactions effects. The T × E interaction variance relative to those due to tester effects were 6.4% for grain Fe content, 10% for grain Zn content, 36% for grain yield per plant, 25.4% for grain phytate contents, 4% for phytate:Fe molar ratio, 11.11% for phytate:Zn molar ratio, and 60% for thousand grain weight.

The L × T × E interaction variance relative to that due to L × T interaction was 1% for grain Fe content, 46% for grain Zn content, 53% for grain yield per plant, 26% for grain phytate contents, 80% for phytate:Fe molar ratio, 42% for phytate:Zn molar ratio, and 46% for thousand grain weight. The line × tester interactions contributed more to the variance of test weight than those of lines and testers. The magnitude of  $\sigma^2$ SCA was quite higher than the magnitude of  $\sigma^2$ GCA for Fe, Zn, phytate, phytate:Fe/Zn molar ratio, and grain yield.

# 3.2 Parental performance *per se* and combining ability

The grain Fe content varied from 49.34 to 66.73  $\mu$ g/g among the lines and from 39.85 to 65.12  $\mu$ g/g among the testers (Table 3). Two lines (ICMA-07999 and ICMA-12222) showed highly significant GCAs in the positive direction, and one line (ICMA-02555) had significant GCAs in the negative direction for grain Fe content.

Similarly, four testers had highly significant GCAs toward positive side, and six testers had significant GCAs for grain Fe content in the negative direction (Table 3). Grain Zn content varied between 44.72 and 57.32  $\mu$ g/g among lines and from 38.38 to 60.86  $\mu$ g/g among testers. All the three lines had significant positive GCA effects for grain Zn content. Among testers, four testers had highly significant GCAs toward positive side, and six testers had significant negative GCAs for grain Zn content Figure 1.

*Per se* performance in terms of grain yield per plant varied between 48.80 and 68.23 g in lines and from 39.45 to 54.44 g in testers. One line each had highly significant positive and negative GCAs for grain yield per plant. None of the testers had significant positive or negative GCA effects. Thousand grain weight ranged from 8.983 to 12.275 g in lines and from 6.45 to 14.3 g among testers. None of the lines had either significant positive or negative GCA effects. Only one tester had significant GCA effect toward negative side in thousand grain weight.

Grain phytate contents varied from 0.079 to 0.121 mg  $g^{-1}$  for lines and from 0.075 to 0.091 mg  $\rm g^{-1}$  for testers. Significant negative GCA effect was observed for only one line. Among testers, two testers showed significant GCA effect in the positive direction, and two testers had significant GCA effects in the negative direction. Mean values of bioavailable Fe (phytate:Fe molar ratio) varied between 0.112 and 0.2 among lines and from 0.113 to 0.181 among tester genotypes. Only one line had significant GCA effect in the negative direction for phytate:Fe molar ratio. Five testers had a significant GCA effect toward positive side, and four had significant GCA effects toward negative side for the phytate:Fe molar ratio. For bioavailable Zn (phytate:Zn molar ratio), the mean value varied between 0.146 and 0.235 among lines and from 0.129 to 0.206 in tester genotypes. Among three lines, one had significant GCA effect toward positive side, and one line had significant negative GCA effect. Among testers, four testers had a significant GCA effect in the positive direction, and three testers had significant GCA effect in the negative direction.

## 3.3 Heterosis and combining ability

Grain Fe content for hybrids ranged from 47.9 to 95.6  $\mu g/g$  over environments. Among all the 30 hybrids, 23 hybrids displayed significant positive MPH, and negative heterosis was observed among the remaining. Similarly, 22 hybrids had significant positive BPH, and five hybrids showed significant negative heterosis. Of the 30 cross-combinations 11 crosses showed a significant SCA effect in the positive direction, and eight crosses had a significant SCA effect in the negative direction. The highest significant positive SCA effect was exhibited by the cross ICMA-02555 × A5RT-17/25 and ICMA-12222 × A5RT-17/16. Similarly, grain Zn content for hybrids ranged from 44.3 to 75.96 µg/g. Among all the 30 hybrids, 15 hybrids showed significant BPH in the positive direction, whereas four hybrids had significant heterosis in the negative direction. Similarly, for MPH, 18 hybrids had significant heterosis in the positive direction, and two hybrids had significant heterosis in the negative direction. Significant positive SCA effect was observed for 12 of the 30 hybrids, whereas significant negative SCA effect for 10 hybrids. The highest significant positive SCA effect was exhibited by the hybrids ICMA-12222  $\times$ A5RT-17/5 and ICMA-02555 × A5RT-17/8.

TABLE 2 Mean squares and genetic components for traits studied in line x tester trial of pearl millet in the 2019 summer season (E1) and 2019 rainy season (E2).

		Mean sum of squares											
Source of Variation	df <sup>+</sup>		Fe (µgg <sup>-1</sup> )			Zn (µgg⁻	<sup>1</sup> )	TGW (g)			GYPP (g)		
		E1	E2	Combined	E1	E2	Combined	E1	E2	Combined	E1	E2	Combined
Environment	1			11688.49**			7160.61**			63.73**			2539.75**
Replications	(1) 2	1.550000	32.08**	16.82*	1.05	2.68	1.86	2.14	0.00	1.07	2.5	180.359**	98.17**
Hybrid	29	339.7**	620.45**	772.12**	171.39**	263.79**	352.51**	1.44**	5.67**	4.73**	10.444	39.897**	35.65**
Lines	(2) 2	334.97**	65.077**	347.33**	136.83**	333.21**	397.76**	0.31	4.49**	1.98**	28.66*	312.922**	252.77**
Testers	(9) 9	708.78**	1360.91**	1945.41**	248.24**	449.51*	636.51**	0.71	2.52**	2.02**	14.214	12.625	19.72*
Lines × Testers	(18) 18	155.62**	311.94**	232.67**	136.81**	163.22**	205.47**	1.94**	7.37**	6.40**	6.535	23.19*	19.48**
Hybrid × Environment	29			187.99**			82.68**			2.38**			14.69*
Line × Environment	2			52.70**			72.28**			2.81**			88.85**
Tester × Environment	9			124.26**			61.24**			1.21**			7.11
Line  imes Tester  imes Environment	18			234.88**			94.55**			2.92**			10.24
Error	(29) 58	2.93	6.75	4.84	2.93	1.78	2.47	0.54	0.24	0.4	6.304	9.836	8.26
						·	Genetic Comp	onents					
Genotype Variance		168.37	306.85	5597.89	84.11	131.01	2555.66	0.45	2.72	34.31	2.40	14.51	258.46
Line Variance		8.97	0.00	2.87	0.00	8.50	4.81	0.00	0.00	0.00	1.11	14.49	5.83
Tester Variance		92.19	174.83	142.73	18.57	47.72	35.92	0.00	0.00	0.00	1.28	0.00	0.02
LinexTester Variance		76.35	152.59	56.96	66.82	80.72	50.75	0.70	3.57	1.50	0.45	6.15	2.81
Additive Variance		227.11	380.72	332.85	42.68	124.11	90.72	0.00	0.00	0.00	4.82	20.61	9.97
Dominance Variance		305.38	610.37	227.83	267.29	322.88	203.00	2.80	14.27	6.01	1.80	24.62	11.23
Contributions of lines		6.80	0.72	3.10	5.51	8.71	7.79	1.48	5.45	2.89	18.93	54.09	38.65
Contributions of testers		64.76	68.07	78.20	44.95	52.88	56.04	15.25	13.81	13.26	42.24	9.82	26.52
Contributions of lxt		28.44	31.21	18.70	49.54	38.40	36.18	83.27	80.74	83.86	38.84	36.09	34.83

TABLE 2. CONTINUED

Source of Variation	df	Mean sum of squares								
			PA (mgg⁻¹)		PA/Fe				PA/Zn	
		E1	E2	Combined	E1	E2	Combined	E1	E2	Combined
Environment	1			0.000026			0.025**			0.051**
Replications	(1) 2	0.00000659	0.0002482*	0.00015**	0.0000001	0.0005713**	0.00029**	0.0000028	0.0007966*	0.00046**
Hybrid	29	0.00010438**	0.00021115**	0.00026**	0.0011854**	0.0007508**	0.0016**	0.0024542**	0.0011905**	0.003**
Lines	(2) 2	0.00042237*	0.00050228	0.00092**	0.0001666	0.0001608	0.00032**	0.0031733	0.0029251	0.0061**
Testers	(9) 9	0.000088	0.00023921	0.00026**	0.0024944**	0.0015719**	0.0039**	0.0035351	0.0014439	0.0045**
Lines × Testers	(18) 18	0.00007724**	0.00016477**	0.000193**	0.0006442**	0.0004058**	0.00058**	0.0018339**	0.0008711**	0.0019**
Hybrid × Environment	29			0.000051**			0.00033**			0.00064**
Line × Environment	2			0.000004			0.00000055			0.000007
Tester × Environment	9			0.000066**			0.00014**			0.0005**
$Line \times Tester \times Environment$	18			0.000049*			0.00046**			0.00079**
Error	(29) 58	0.0000085	0.0000400	0.000022	0.0000433	0.0000643	0.0000449	0.0000809	0.0001143	0.000084
				1	G	enetic Components			1	
Genotype Variance		4.78E-05	8.77E-05	0.001914	0.00057	0.00035	0.011632702	0.00119	0.00055	0.021752776
Line Variance		1.73E-05	1.69E-05	0.000018	0.00000	0.00000	0	0.00007	0.00010	0.000104488
Tester Variance		1.79E-06	1.24E-05	0.000006	0.00031	0.00019	0.000278941	0.00028	0.00010	0.000213954
LinexTester Variance		3.42E-05	6.45E-05	0.000043	0.00030	0.00018	0.000134601	0.00088	0.00039	0.000457707
Additive Variance		3.35E-05	5.72E-05	0.000044	0.00067	0.00043	0.000630135	0.00077	0.00039	0.000669491
Dominance Variance		1.37E-04	2.58E-04	0.000170	0.00121	0.00071	0.000538405	0.00350	0.00156	0.001830829
Contributions of lines		27.91	16.41	21.53	0.97	1.48	1.38	8.92	16.95	13.65
Contributions of testers		26.16	35.16	32.19	65.30	64.98	65.19	44.70	37.64	40.38
Contributions of lxt		45.93	48.43	46.28	33.73	33.55	33.43	46.38	45.42	45.97

\*, \*\*Significant at the P < 0.05 and 0.01 probability level, respectively. <sup>†</sup>Figures in parentheses indicate individual environment degrees of freedom.

TABLE 3 Performance per se of the lines and testers and their general combining ability (GCA) effects for traits studied in pearl millet across two environments.

	Fe (µg g <sup>-1</sup> )		Zn (μg g <sup>-1</sup> )		TGW (g)		GYPP (g)		PA (mg g <sup>-1</sup> )		PA/Fe		PA/Zn	
Parent	Mean (ppm)	GCA	Mean (ppm)	GCA	Mean (ppm)	GCA	Mean (ppm)	GCA	Mean (ppm)	GCA	Mean (ppm)	GCA	Mean (ppm)	GCA
Lines														
ICMA- 02555	66.728	-3.39**	57.325	2.1**	11.643	-0.23	48.802	2.74**	0.085	-0.005**	0.112	0.002	0.146	-0.0137**
ICMA- 07999	49.343	1.95**	44.715	1.52**	8.983	0.21	68.229	0.550	0.079	0.0046**	0.136	0.001	0.176	0.003
ICMA- 12222	51.380	1.43**	51.290	-3.62**	12.275	0.017	58.290	2.19**	0.121	0.000	0.200	-0.0032*	0.235	0.01023**
							Teste	rs						
A5RT- 17/4	65.125	13.4**	57.078	12.41**	13.905	0.061	43.424	-0.040	0.083	-0.003	0.113	-0.02**	0.145	-0.033**
A5RT- 17/5	52.500	-7.63**	51.045	-2.79**	11.564	-0.09	46.337	1.680	0.089	0.0064**	0.145	0.019**	0.174	0.0204**
A5RT- 17/6	51.800	-8.07**	44.795	-1.44*	6.455	-0.36	46.526	0.310	0.080	0.005*	0.131	0.017**	0.176	0.0087**
A5RT- 17/7	56.068	-10**	53.265	-6.37**	12.570	0.40	43.608	-1.060	0.084	0.000	0.129	0.015**	0.161	0.021**
A5RT- 17/8	52.533	-10.9**	49.238	-8.31**	7.397	-0.8**	41.054	0.330	0.091	-0.0046*	0.148	0.01**	0.184	0.018**
A5RT- 17/16	39.850	-12.9**	41.328	-6.11**	13.776	0.39	43.900	1.940	0.086	-0.003	0.181	0.015**	0.205	0.006
A5RT- 17/19	41.450	-8.9**	38.383	-6.1**	10.742	0.31	54.444	0.160	0.080	-0.0059**	0.163	0.003	0.206	0.002
A5RT- 17/25	53.728	15.99**	60.863	7.83**	14.304	-0.42	40.000	-1.150	0.075	-0.001	0.119	-0.0232**	0.129	-0.0221**
A5RT- 17/26	54.388	16.72**	59.203	7.49**	9.705	0.17	43.271	-2.340	0.085	-0.001	0.134	-0.0252**	0.150	-0.021**
A5RT- 17/29	52.695	12.39**	49.090	3.4**	7.291	0.33	39.454	0.170	0.085	0.0073**	0.139	-0.0112**	0.172	0.000

\* and \*\*, significant at the P< 0.05 and 0.01 probability level, respectively.

For grain yield per plant, hybrids mean value varied from 43.2 to 54.2 g. Among 30 hybrids, 10 hybrids displayed significant BPH in the negative direction, and no hybrid had significant positive heterosis. Similarly, for MPH, only one hybrid had significant negative heterosis. Of the 30 cross-combinations, none of the crosses displayed a significant positive or negative SCA effect for grain yield per plant. Similarly, thousand grain weight varied from 8.98 to 12.28 g. Twelve of the 30 hybrids showed significant MPH in the positive direction, and none of the hybrids had significant negative heterosis. Similarly, for BPH, only one hybrid had significant negative heterosis, and six hybrids had significant positive heterosis. Of the 30 hybrids, three hybrids showed a significant SCA effect in the positive direction, and four hybrids showed a significant SCA effect in the negative direction. The highest significant positive SCA effect was exhibited by the hybrids ICMA-02555 × A5RT-17/7 and ICMA-12222 × A5RT-17/25.

Averaged over the environments, grain phytate contents varied from 0.076 to 0.11 mg g<sup>-1</sup>. Five of the 30 hybrids displayed significant positive MPH, and two hybrids had significant heterosis in the negative direction. Similarly, 10 hybrids had significant heterosis in

the negative direction, and five hybrids had significant BPH in the positive direction. Of the 30 hybrids, five hybrids showed a significant SCA effect toward positive side, and three hybrids showed a significant SCA effect toward negative side. The highest significant SCA effect in the positive direction was exhibited by the hybrids ICMA-07999 × A5RT-17/5 and ICMA-07999 × A5RT-17/26, and the highest significant negative SCA effect was exhibited by the hybrids ICMA-02555 × A5RT-17/25 and ICMA-12222 × A5RT-17/26.

For bioavailable Fe (phytate:Fe molar ratio), the *per se* performance of hybrids varied between 0.071 and 0.148. Among 30 hybrids, 18 hybrids showed significant MPH in the negative direction, and zero hybrids had significant positive BPH. Similarly, 22 hybrids had significant heterosis in the negative direction, and no hybrid had significant positive BPH. Of the 30 hybrids, five hybrids showed a significant SCA effect in the positive direction, and five hybrids showed a significant positive SCA effect was exhibited by the hybrids ICMA-02555 × A5RT-17/16 and ICMA-12222 × A5RT-17/25, and the highest significant negative SCA effect was exhibited by the hybrids ICMA-02555 × A5RT-17/25 and ICMA-12222 × A5RT-17/

5. Similarly, for bioavailable Zn estimate (phytate:Zn molar ratio), the mean value ranged from 0.11 to 0.22. Among 30 hybrids, 17 hybrids showed significant negative BPH, and only one hybrid had significant positive BPH. Similarly, nine hybrids had significant heterosis in the negative direction, and only one hybrid had significant BPH in the positive direction. Of the 30 hybrids, seven hybrids showed a significant SCA effect toward positive side and seven hybrids showed a significant positive SCA effect toward negative side. The highest significant positive SCA effect was exhibited by the hybrids ICMA-07999 × A5RT-17/5 and ICMA-12222 × A5RT-17/7, and the highest significant negative SCA effect was exhibited by the hybrids ICMA-12222 × A5RT-17/7.

Both the hybrids ICMA-02555 × A5RT-17/8 and ICMA-02555 × A5RT-17/8 proved to be the best specific combiner over all the environments for grain Fe and grain Zn content. Similarly, ICMA-12222 × A5RT-17/5 for bioavailable Fe estimate (phytate:Fe molar ratio) and bioavailable Zn estimate (phytate:Zn molar ratio) and ICMA-02555 × A5RT-17/25 is the best specific combiner over all the environments for grain phytate contents. The hybrids, namely, ICMA-02555 × A5RT-17/26 and ICMA-07999 × A5RT-17/25,

showed a high *per se* performance, significant heterobeltiosis, SH, and a significant positive SCA effect involving both good and poor GCA. Table 4 shows the best five hybrids for the pearl millet attributes examined in two environments for performance *per se*, SCA, BPH, and SH. There were significant and negative correlations between performance *per se* of hybrids and mid-parental values for Fe (r = 0.43), and, for Zn, it is positively correlated (r = 0.36 for Zn) across environment (Figure 2). Similarly, a highly significant positive correlation was observed between performance *per se* of the hybrids, and their SCA was observed for both Fe density (r = 0.46) and Zn density (r = 0.52) across environment (Figure 3).

# 4 Discussion

The combining ability analysis revealed the presence of genetic variation among parents and hybrids for the traits studied. Substantial variability was observed for the Fe and Zn content among parents, which has also been reported earlier (Velu et al., 2007; Rai et al., 2012; Govindraj et al., 2013; Kanatti et al., 2014). In comparison with the

TABLE 4 Performance per se, specific combining ability (SCA), better-parent heterosis (BPH) and standard heterosis (SH) of five top rank hybrids for the traits studied of pearl millet across two environments.

Tue:t	Uubrid	Perf	ormance <i>per se</i>		SCA	ווחס	CC Duca 1201	
Ifall	пурпа	F1 (hybrid)	P1 (line)	P2 (tester)	JCA	DPT	CC Pusa 1201	
Fe (µg g <sup>-1</sup> )	ICMA-02555 × A5RT-17/25	95.025	66.728	53.728	9.46**	42.41 **	125.55 **	
	ICMA-12222 × A5RT-17/16	70.070	51.380	39.850	8.58**	36.38 **	66.32 **	
	ICMA-02555 × A5RT-17/26	94.600	66.728	54.388	8.3**	41.77 **	124.54 **	
	ICMA-02555 × A5RT-17/8	66.475	66.728	52.533	7.88**	-0.38	57.79 **	
	ICMA-07999 × A5RT-17/16	62.113	49.343	39.850	0.103	25.88 **	47.43 **	
Zn (µgg <sup>-1</sup> )	ICMA-12222 × A5RT-17/5	65.965	51.290	51.045	13.43**	28.61 **	96.44 **	
	ICMA-02555 × A5RT-17/8	63.208	57.325	49.238	10.47**	10.26 *	88.23 **	
	ICMA-02555 × A5RT-17/26	75.968	57.325	59.203	7.41**	28.32 **	126.23 **	
	ICMA-12222 × A5RT-17/16	55.290	51.290	41.328	6.075**	7.8	64.65 **	
	ICMA-12222 × A5RT-17/4	73.225	51.290	57.078	5.49**	28.29 **	118.06 **	
	ICMA-02555 × A5RT-17/7	15.982	11.643	12.570	2.95**	27.14 **	7.84	
	ICMA-12222 × A5RT-17/25	13.858	12.275	14.304	1.4**	-3.12	-6.49	
TGW (g)	ICMA-07999 × A5RT-17/8	13.623	8.983	7.397	1.35**	51.66 **	-8.07	
	ICMA-02555 × A5RT-17/5	13.421	11.643	11.564	0.410	15.27	-9.44	
	ICMA-12222 × A5RT-17/8	12.899	12.275	7.397	0.820	5.08	-12.96	
	ICMA-02555 × A5RT-17/16	52.075	48.802	43.900	3.641	6.7	-4.46	
	ICMA-12222 × A5RT-17/29	54.160	58.290	39.454	2.557	-7.085	-0.64	
GYPP (g)	ICMA-07999 × A5RT-17/6	52.476	68.229	46.526	2.37	-23.08 **	-3.73	
	ICMA-02555 × A5RT-17/7	47.730	48.802	43.608	2.29	-2.19	-12.43	
	ICMA-12222 × A5RT-17/4	53.457	58.290	43.424	2.065	-8.29	-1.93	
$\mathbb{D} \wedge (m q q^{-1})$	ICMA-07999 × A5RT-17/4	0.083	0.079	0.083	-0.0103**	-0.12	-8.08	
PA (mg g-`)	ICMA-02555 × A5RT-17/25	0.076	0.085	0.075	-0.00975**	-10.8	-16	

(Continued)

Trait		Perf	ormance <i>per se</i>	2	<b>CCA</b>		CC Pusa 1201	
	Hybrid	F1 (hybrid)	P1 (line)	P2 (tester)	SCA	ВЬН		
	ICMA-12222 × A5RT-17/26	0.082	0.121	0.085	-0.0091**	-32.89 **	-9.44	
	ICMA-07999 × A5RT-17/8	0.085	0.079	0.091	-0.004	-6.45	-5.75	
	ICMA-12222 × A5RT-17/5	0.093	0.121	0.089	0.001	-23.5 **	3.22	
PA/Fe	ICMA-02555 × A5RT-17/25	0.071	0.112	0.119	-0.02**	-40.18 **	-32.47 **	
	ICMA-12222 × A5RT-17/5	0.11	0.200	0.145	-0.017**	-44.86 **	5.21	
	ICMA-02555 × A5RT-17/8	0.12	0.112	0.148	-0.01133*	-24.05 **	7.05	
	ICMA-12222 × A5RT-17/16	0.114	0.200	0.181	-0.017**	-43.14 **	8.481	
	ICMA-02555 × A5RT-17/26	0.08	0.112	0.134	-0.01*	-40.99 **	-24.52 *	
PA/Zn	ICMA-12222 × A5RT-17/5	0.144	0.235	0.174	-0.048**	-38.91 **	24.57	
	ICMA-02555 × A5RT-17/8	0.15	0.146	0.184	-0.0227**	-22.79 *	23.57	
	ICMA-07999 × A5RT-17/7	0.17	0.176	0.161	-0.015*	-3.23	47.82 **	
	ICMA-07999 × A5RT-17/4	0.12	0.176	0.145	-0.015*	-34.31 **	0.34	
	ICMA-02555 × A5RT-17/7	0.154	0.146	0.161	-0.014*	-4.34	33.64 *	

#### TABLE 4 Continued

 $^{\ast}$  and  $^{\ast\ast},$  significant at the P< 0.05 and 0.01 probability level, respectively.

individual contributing effects of lines and testers, the influence of line tester hybrid was more evident. For the traits studied, the variability among the hybrids due to SCA ( $\sigma^2$  SCA) was higher than the variability owing to GCA ( $\sigma^2$  GCA) with the predictability ratio being lesser than unity. This revealed that both Fe and Zn grain

content is mostly non-additive genetic control as well as the presence of superior crosses with significant SCA and higher heterosis (Nandaniya et al., 2016; Jeeterwal et al., 2017; Warrier et al., 2020). As a result, the material used in this study is best suited for use in the hybrid breeding program. Several earlier studies have reported the



FIGURE 1

Best general combining parents (line and tester) for both grain Fe and Zn content.

predominance of additive variance for grain Fe and Zn in pearl millet (Velu et al., 2011; Govindraj et al., 2013; Kanatti et al., 2014), maize (Arnold and Bauman, 1976; Long et al., 2004; Chen et al., 2007) and rice (Zhang et al., 2004). Both SCA ( $\sigma^2$  SCA) and GCA ( $\sigma^2$  GCA) were significant with higher  $\sigma^2$  SCA for GYPP, demonstrating the significance of both variances with a predominance of non-additive variance. Earlier Nandaniya et al. (2016) and Chittora and Patel (2016) revealed the significance of non-additive gene action for GYPP. According to Kathale et al. (2013) and Ansodariya et al. (2013), the relevance of both additive and non-additive gene action for 1,000-grain weight is significant.

Govindaraj et al. (2013) also observed substantial GCA in both directions for Fe and Zn grain densities. This clearly indicates that gene combinations in different lines were not similar. Likewise, the lines with significant negative GCA for Zn had an equally significant positive GCA for Fe content, and the testers with negative significant GCA for Zn had an equally significant negative GCA for grain Fe content. This showed that the mechanism of Fe and Zn filling in grain in testers is regulated by the same gene(s) or that the variability of both Fe and Zn might be controlled by a close genetic linkage (Kanatti et al., 2014)

This research also identified restorer lines (R) and maintainer lines (B) with a high–grain Fe and Zn content, which are also good general combiners. Furthermore, to be economically viable, genetic enhancement of grain Fe and Zn content must not compromise on grain yield. An earlier research of pearl millet found no negative relationship between these micronutrients and grain output (Gupta et al., 2009). However, in a later study comprising four different hybrid trials, grain Fe content was significantly negatively linked with grain production in two trials and uncorrelated in other two trials. In any of these four trials, Zn density did not correlate with grain yield (Rai et al., 2012). For better clarity, such character association studies must obviously be conducted employing a wide range of materials and diverse environments. The molar ratio of grain phytate:Fe to Zn and grain phytate content should have negative heterosis to increase the bioavailability of both Fe and Zn. Several parents had nonsignificant GCA effects for grain phytate, indicating that this trait has dominant genetic control and may be used to reduce grain phytate content. Few hybrids had lower mid-parent values for grain phytate and phytate:Fe/Zn molar ratios. These results indicate that these hybrids have lower levels of these traits with increased bioavailability of Fe and Zn. This finding is consistent with the findings of Brkic et al (2003), who reported some heterotic phytate-lowering effects in maize.

On the basis of the findings, it is possible to infer that there is great genetic potential to increase the bioavailability of grain Fe/Zn content in the lines to provide an inexpensive and long-term solution to the iron-deficient population. Common and overlapping quantitative trait loci (QTL) for Fe and Zn grain density have been identified in wheat (Peleg et al., 2009; Singh et al., 2010), pearl millet (Kumar, 2011), rice (Stangoulis et al., 2007), and beans (Cichy et al., 2009; Blair et al., 2011). Diverse sources of pearl millet with high Fe and Zn grain density levels have been identified, providing valuable experimental materials for mapping population and identifying diverse physiological processes for optimal Fe and Zn accumulation in the grains. Using marker-assisted selection to introgress high Fe and Zn QTL and pyramiding them in parental lines may be the most effective strategy for breeding hybrids with a high Fe and Zn content in pearl millet. The combining ability tests and the manifestation of heterosis in F1 provided various cross-combinations that may be advised to enhance the population's Fe/Zn profile. A plant population with higher grain Fe/Zn content, enhanced Fe/Zn bioavailability, higher grain yields, and lower phytate concentration can be created by delaying the selection in crosscombinations until later generations.





FIGURE 3

Relationship between grain Fe and Zn densities of hybrids and their specific combining ability (SCA) effects in line x tester across two environments.

# Data availability statement

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding authors.

# Author contributions

TR and SS planned and designed the research. SS, TR, MS, and MM performed the field experiment. AS and MM helped in Fe and Zn estimation. TR, TS, and MM conducted recording and compilation of data. TR and MS conducted statistical analysis. TR and JM prepared the manuscript. CS edited and revised the manuscript for publication. All authors contributed to the article and approved the submitted version.

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# References

Ansodariya, V. V., Dangaria, C. J., and Savaliya, J. J. (2013). Combining ability analysis of grain yield and its component characters in pearl millet [*Pennisetum glaucum* (L.) r. br.]. *Prog. Res.* 8 (Special), 564–567. doi: 10.1016/j.jksus.2019.12.009

Arnold, J. M., and Bauman, L. F. (1976). Inheritance of and interrelationships among maize kernel traits and elemental contents 1. *Crop Sci.* 16 (3), pp.439-pp.440. doi: 10.2135/cropsci1976.0011183X001600030034x

Arunachalam, V. (1974). The fallacy behind the use of modified line x tester design. Indian J. @ Genet. Plant Breed 34, 280–287.

Blair, M. W., Astudillo, C., Rengifo, J., Beebe, S. E., and Graham, R. (2011). QTL analyses for seed iron and zinc concentrations in an intra-genepool population of Andean common beans (*Phaseolus vulgaris* l.). *Theor. Appl. Genet.* 122 (3), 511–521. doi: 10.1007/s00122-010-1465-8

Brkić, I., Šimić, D., Zdunić, Z., Jambrović, A., Ledenčan, T., Kovačević, V., et al. (2003). Combining abilities of corn-belt inbred lines of maize for mineral content in grain. *Maydica* 48, 293–297.

Chen, F., Chun, L., Song, J., and Mi, G. (2007). Heterosis and genetic analysis of iron concentration in grains and leaves of maize. *Plant Breed* 126 (1), 107–109. doi: 10.1111/j.1439-0523.2006.01289.x

Chittora, K., and Patel, J. A. (2016). Combining ability analysis for seed yield and yield component characters in pearl millet [*Pennisetum glaucum* (L) r. br.]. *Bioscan* 11, (4) 3161–3165.

Cichy, K. A., Caldas, G. V., Snapp, S. S., and Blair, M. W. (2009). QTL analysis of seed iron, zinc, and phosphorus levels in an Andean bean population. *Crop Sci.* 49 (5), 1742–1750. doi: 10.2135/cropsci2008.10.0605

Dabholkar, A. R. (1999). Elements Of Bio Metrical Genetics (revised And Enlarged Edition). Concept publishing company.

Directorate of Millets Development Department of agriculture, Co-operation & farmers welfare, ministry of agriculture & farmers welfare, government of India. Available at: http://millets.dacfw.nic.in/POP%20Pearl.html (Accessed 26. 05. 2020).

Govindaraj, M., Rai, K. N., Pfeiffer, W. H., Kanatti, A., and Shivade, H. (2016). Energydispersive X-ray fluorescence spectrometry for cost-effective and rapid screening of pearl millet germplasm and breeding lines for grain iron and zinc density. *Commun. Soil Sci. Plant Anal.* 47 (18), pp.2126–2134. doi: 10.1080/00103624.2016.1228938

Govindaraj, M., Rai, K. N., Shanmugasundaram, P., Dwivedi, S. L., Sahrawat, K. L., Muthaiah, A. R., et al. (2013). Combining ability and heterosis for grain iron and zinc densities in pearl millet. *Crop Sci.* 53 (2), pp.507–pp.517. doi: 10.2135/cropsci2012.08.0477

Gupta, S. K., Velu., G., Rai, K. N., and Sumalini, K. (2009). Association of grain iron and zinc content with grain yield and other traits in pearl millet [*Pennisetum glaucum* (L.) r. br.]. *Crop Improv.* 36, 4–7.

millet breeding), ICRISAT, Patancheru, Telangana (India), for providing the cytoplasmic male sterile lines and restorers belonging to  $A_5$  cytoplasm. The research work presented here is a part of Ph.D. thesis of Thribhuvan, R., submitted to ICAR- IARI, New Delhi -110012, India.

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Hallauer, A. R., and Miranda, J. B. (1981). Quantitative genetics in maize breeding. Iowa State Univ Press, Ames, IA. Jenkins, JN, WL Parrott, JC McCarty Jr & RL Shepherd, 1988a. Registration of three noncommercial germplasm lines of upland cotton tolerant to tobacco budworm *Crop Sci* 28, 869

Jeeterwal, R. C., Sharma, L. D., and Nehra, A. (2017). Combining ability studies through diallel analysis in pearl millet [*Pennisetum glaucum* (L.) r. br.] under varying environmental conditions. *J. Pharmacogn Phytochem.* 6 (4), 1083–1088.

Jeong, J., and Guerinot, M. L. (2008). Biofortified and bioavailable: the gold standard for plant-based diets. *Proc. Natl. Acad. Sci.* 105 (6), 1777-1778. doi: 10.1073/pnas.071233010

Kanatti, A., Rai, K. N., Radhika, K., Govindaraj, M., Sahrawat, K. L., and Rao, A. S. (2014). Grain iron and zinc density in pearl millet: combining ability, heterosis and association with grain yield and grain size. *SpringerPlus* 3 (1), 1–12. doi: 10.1186/2193-1801-3-763

Kathale, M. N., Jadhav, P. A., and Wadekar, P. B. (2013). Heterosis in pearl millet. *Bioinfolet* 10 (3B), 1002–1005.

Kempthorne, O. (1957). An introduction to genetic statistics (New York: John Wiley & Sons).

Kumar, S. (2011) Development of new mapping population and marker-assisted improvement of iron and zinc grain density in pearl millet [*Pennisetum glaucum* (L.) r. br.]. *Bikaner: Dissertation Swami Keshwanand Rajasthan Agric. University.* 

Kumar, S., Hash, C. T., Nepolean, T., Mahendrakar, M. D., Satyavathi, C. T., Singh, G., et al. (2018). Mapping grain iron and zinc content quantitative trait loci in an iniadiderived immortal population of pearl millet. *Genes* 9 (5), 248. doi: 10.3390/genes9050248

Kumar, S., Hash, C. T., Thirunavukkarasu, N., Singh, G., Rajaram, V., Rathore, A., et al. (2016). Mapping quantitative trait loci controlling high iron and zinc content in self and open pollinated grains of pearl millet [*Pennisetum glaucum* (L.) r. br.]. *Front. Plant Sci.* 7. doi: 10.3389/fpls.2016.01636

Kumar, S., Thirunavukkarasu, N., Singh, G., Sharma, R., and Kulkarni, K. S. (2015). "Biofortification for selecting and developing crop cultivars denser in iron and zinc," in *Nutrient use efficiency: From basics to advances* (New Delhi: Springer), 237–253.

Long, J. K., Bänziger, M., and Smith, M. E. (2004). Diallel analysis of grain iron and zinc density in southern African-adapted maize inbreds. *Crop Sci.* 44 (6), 2019–2026. doi: 10.2135/cropsci2004.2019

Mahendrakar, M. D., Kumar, S., Singh, R. B., Rathore, A., Potupureddi, G., Kishor, P. K., et al. (2019). Genetic variability, genotype× environment interaction and correlation analysis for grain iron and zinc contents in recombinant inbred line population of pearl millet [*Pennisetum glaucum* (L). r. br.]. *Ind. J. Genet. Plant Breed* 79 (03), 545–551. doi: 10.31742/IJGPB.79.3.3

Megazyme (2016). Available at: https://secure.megazyme.com/files/Booklet/K-PHYT\_DATA.pdf (Accessed 05.12.2019).

Nandaniya, K. U., Mungra, K. D., and Sorathiya, J. S. (2016). Assessment of combining ability for yield and micronutrients in pearl millet. *Electron. J. Plant Breed* 7 (4), 1084–1088. doi: 10.5958/0975-928X.2016.00149.6

Peleg, Z., Cakmak, I., Ozturk, L., Yazici, A., Jun, Y., Budak, H., et al. (2009). Quantitative trait loci conferring grain mineral nutrient concentrations in durum wheat  $\times$  wild emmer wheat RIL population. *Theor. Appl. Genet.* 119, 353–369. doi: 10.1007/s00122-009-1044-z

Rai, K. N., Govindaraj, M., and Rao, A. S. (2012). Genetic enhancement of grain iron and zinc content in pearl millet. Qual. Assur. Saf. Crop 4 (3), 119–125. doi: 10.1111/j.1757-837X.2012.00135x

Rao, P. P., Birthal, P. S., Reddy, B. V., Rai, K. N., and Ramesh, S. (2006). Diagnostics of sorghum and pearl millet grains-based nutrition in India. *Int. Sorghum Millets News.* 47, 93–96.

Rodríguez, F., Alvarado, G., Pacheco, Á, and Crossa J and Burgueño, J. (2015). AGD-r (Analysis of genetic designs with r for windows) version 4.0. *Int. Maize Wheat Improvement Center (CIMMYT)*.

Singh, K., Chhuneja, P., Tiwari, V. K., Rawat, N., Neelam, K., Aggarwal, R., et al. (2010). Mapping of QTL for grain iron and zinc content in diploid a genome wheat and validation of these loci in U and s genomes. *Paper presented at: Plant Anim. Genomes XVIII Conference San Diego*. Stangoulis, J. C., Huynh, B. L., Welch, R. M., Choi, E. Y., and Graham, R. D. (2007). Quantitative trait loci for phytate in rice grain and their relationship with grain micronutrient content. *Euphytica* 154 (3), 289–294. doi: 10.1007/s10681-006-9211-7

Velu, G., Rai, K. N., Muralidharan, V., Kulkarni, V. N., Longvah, T., and Raveendran, T. S. (2007). Prospects of breeding biofortified pearl millet with high grain iron and zinc content. *Plant Breed.* 126 (2), 182–185. doi: 10.1111/j.1439-0523.2007.01322.x

Velu, G., Rai, K. N., Muralidharan, V., Longvah, T., and Crossa, J. (2011). Gene effects and heterosis for grain iron and zinc density in pearl millet (Pennisetum glaucum (L.) r. br). *Euphytica* 180 (2), 251–259. doi: 10.1007/s10681-011-0387-0

Velu, G., Rai, K. N., Sahrawat, K. L., and Sumalini, K. (2008). Variability for grain iron and zinc contents in pearl millet hybrids. J. SAT Agric. Res. 6, 1-4.

Warrier, S. R., Patel, B. C., Kumar, S., and Sherasiya, S. A. (2020). Combining ability and heterosis for grain minerals, grain weight and yield in pearl millet and SSR markers based diversity of lines and testers. J. @ King Saud University-Science 32 (2), 1536–1543. doi: 10.1016/j.jksus.2019.12.009

Zhang, M. W., Guo, B. J., and Peng, Z. M. (2004). Genetic effects on fe, zn, Mn and p contents in indica black pericarp rice and their genetic correlations with grain characteristics. *Euphytica* 135 (3), 315–323. doi: 10.1023/B:EUPH.0000013340.98344.60