

Impact of gene polymorphisms of gonadotropins and their receptors on human reproductive success

Livio Casarini^{1,2}, Daniele Santi^{1,3} and Marco Marino^{1,2}

¹Unit of Endocrinology, Department of Biomedical, Metabolic and Neural Sciences and ²Center for Genomic Research, University of Modena and Reggio Emilia, Via G. Campi, 287, 41125 Modena, Italy and

³Azienda USL of Modena, NOCSAE, Via P. Giardini 1355, 41126 Modena, Italy

Correspondence should be addressed to L. Casarini; Email: livio.casarini@unimore.it

Abstract

Gonadotropins and their receptors' genes carry several single-nucleotide polymorphisms resulting in endocrine genotypes modulating reproductive parameters, diseases, and lifespan leading to important implications for reproductive success and potential relevance during human evolution. Here we illustrate common genotypes of the gonadotropins and gonadotropin receptors' genes and their clinical implications in phenotypes relevant for reproduction such as ovarian cycle length, age of menopause, testosterone levels, polycystic ovary syndrome, and cancer. We then discuss their possible role in human reproduction and adaptation to the environment.

Gonadotropins and their receptors' variants are differently distributed among human populations. Some hints suggest that they may be the result of natural selection that occurred in ancient times, increasing the individual chance of successful mating, pregnancy, and effective post-natal parental cares. The gender-related differences in the regulation of the reproductive endocrine systems imply that many of these genotypes may lead to sex-dependent effects, increasing the chance of mating and reproductive success in one sex at the expenses of the other sex. Also, we suggest that sexual conflicts within the FSH and LH-choriogonadotropin receptor genes contributed to maintain genotypes linked to subfertility among humans. Because the distribution of polymorphic markers results in a defined geographical pattern due to human migrations rather than natural selection, these polymorphisms may have had only a weak impact on reproductive success. On the contrary, such genotypes could acquire relevant consequences in the modern, developed societies in which parenthood attempts often occur at a later age, during a short, suboptimal reproductive window, making clinical fertility treatments necessary.

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Introduction

Follicle-stimulating hormone (FSH) and luteinizing hormone (LH) are glycoproteins produced by the pituitary regulating development and reproductive functions in both men and women. On the contrary, human choriogonadotropin (hCG) is the human placental hormone managing pregnancy. Gonadotropins share a common α subunit together with the thyroid-stimulating hormone, while having a unique β subunit, specific for the receptor located in the gonads. The FSH receptor (FSHR) and the common LH/hCG receptor (LHCGR) belong to the superfamily of the G protein-coupled receptors. They are characterized by an extracellular domain, seven transmembrane domains joined by three intra- and extracellular loops, and an intracellular, C-terminal domain. Upon hormone binding with the extracellular portion, the intracellular domain triggers the activation of multiple signaling pathways by interacting with specific molecules, such

as G proteins or β -arrestins (Simoni *et al.* 1997, Ascoli *et al.* 2002, Gloaguen *et al.* 2011).

Gonadotropins and their receptor genes carry several single-nucleotide polymorphisms (SNPs), resulting in several genotypes differently distributed among human populations and affecting sex-related reproductive features and diseases by modulating signal transduction (Casarini *et al.* 2011). These genotypes are evolutionarily old and have accompanied humans during their ancient migrations throughout the continents. However, the impact of these SNPs on human reproductive success and evolution is unclear and was recently debated (Grigороva *et al.* 2007, Simoni & Casarini 2014).

Polymorphisms of the *FSHR* and *FSHB* genes

The *FSHR* carries about 2000 SNPs but only a few of these are known as modulators of gonadal response. One of the most common *FSHR* polymorphisms is

rs6166 (NCBI SNPs database ID; <http://www.ncbi.nlm.nih.gov>) consisting of the nucleotide change A to G at position 2039 from the gene transcription start codon (c.2039A>G) and resulting in the amino acid change N to S at position 680 of the protein chain (p.N680S). rs6166 is in strong linkage disequilibrium with the SNP rs6165 (c.919A>G, p.T307A), at least in Caucasians and Asians, resulting in two discrete FSHR isoforms. p.N680S is close to the C-terminal intracellular region of the receptor and modulates serum FSH levels and gonadal response in both women and men (Lledo *et al.* 2013, Grigorova *et al.* 2014, Simoni & Casarini 2014). Women carriers of the p.N680S S homozygous genotype have higher serum FSH levels during the follicular phase and lower progesterone levels in the luteal phase than the carriers of different genotypes, while p.N680S N homozygous males are characterized by higher testes volume than p.N680S S homozygous men. It was suggested that the FSHR p.N680S S variant is functionally 'resistant' to FSH stimulation; the p.N680S polymorphism modulates cell signaling resulting in differential gene expression and steroidogenesis in cultured human lutein granulosa cells as recently demonstrated *in vitro* (Casarini *et al.* 2014).

Interestingly, the cumulative effect of p.N680S together with other FSHR polymorphisms (e.g., rs1394205; -29G>A) was proposed, leading to genotypes linked with lower fertility (Simoni & Casarini 2014, Grigorova *et al.* 2014). The -29G>A SNP falls within the 5'-UTR of the *FSHR* gene, 29 nucleotides upstream the ATG codon. The *in vitro* transcriptional activity of the -29G>A A variant is lower than that of the -29G>A G genotype in Chinese hamster ovary cells transfected with the *FSHR* promoter and was found to be associated with hypertension (Nakayama *et al.* 2006), lower estradiol levels in women (Achrekar *et al.* 2009), and higher serum FSH levels (Achrekar *et al.* 2009, Grigorova *et al.* 2014).

The FSH β subunit is encoded by the *FSHB* gene, which carries about 24 SNPs, but only the rs10835638 (-211G>T), located in the promoter region of the gene (-211G>T, rs10835638), was extensively studied in association with serum FSH levels and reproductive parameters in males (Grigorova *et al.* 2008). In particular, -211G>T T homozygous Baltic, Italian, and German men have lower FSH levels and testis volume compared to carriers of other genotypes (Grigorova *et al.* 2008, 2014, Tüttelmann *et al.* 2012). The promoter region of the *FSHB* gene is a putative target of a transcription regulatory element and is highly conserved among placental mammals (Grigorova *et al.* 2008), suggesting that the T nucleotide at position -211 affects the *FSHB* gene transcription leading to low hormone levels. Interestingly, the studies performed in males and females are contradictory; -211G>T T homozygous women were shown to have elevated FSH, LH, and reduced progesterone levels compared

with carriers of other genotypes, suggesting a gender-specific, compensatory regulation of the gonadotropin secretion (Schüring *et al.* 2013). Further elucidations may be provided by genotype-phenotype association studies focusing on the cumulative effect of *FSHB* together with *FSHR* gene SNPs, revealing how they affect the sex-related modulation of hormone levels and reproductive parameters. Taken together, the combination of SNPs within the *FSHB* and *FSHR* genes account for a substantial proportion of the total normal phenotypic variance in male and female reproductive parameters (Tüttelmann *et al.* 2012, La Marca *et al.* 2013, Grigorova *et al.* 2014, Simoni & Casarini 2014).

Polymorphisms of the *LHCGR* gene and *LHB/CGB* gene cluster

Several inactivating mutations of the *LHCGR* were associated with peculiar phenotypes such as the 46,XY disorder of sex development, primary amenorrhea and anovulation in women (Powell *et al.* 2003), and undescended testes and androgen deficiency in men (Simoni *et al.* 2008), revealing the crucial role of this receptor in human sex development and reproduction. *LHCGR* harbors at least 300 known polymorphisms but only a few of them lead to relevant effects (Casarini *et al.* 2011).

The *LHCGR* variant 18insLQ, consisting of the insertion of six nucleotides in frame in exon 1 and falling near the N-terminus of the mature receptor, was associated with early onset of breast cancer and short disease-free survival. This is consistent with increased *LHCGR* 18insLQ sensitivity and plasma membrane expression (1.9-fold lower hCG half-effective concentration and 1.4-fold higher expression levels than WT *LHCGR*, respectively; Piersma *et al.* 2006). Interestingly, *LHCGR* 18insLQ has a high frequency among Northern-European Caucasians that are characterized by a higher prevalence of breast cancer compared to other ethnic groups, leading to the speculation that the *LHCGR* genotype may be linked to disease risk (Casarini *et al.* 2011).

Only a few other *LHCGR* SNPs provided significant clinical findings so far. The SNP rs2293275 (c.942G>A, p.S312N), which falls within exon 10 of the *LHCGR* gene, might affect the trafficking and stability of the receptor resulting in an impaired spermatogenesis in men (Simoni *et al.* 2008) and an increased risk of developing polycystic ovary syndrome (PCOS) in women (Thathapudi *et al.* 2015). Lastly, the polymorphic *LHCGR* variant rs4073366 (c.3442-20797C>G) occur about 142 bp downstream of *LHCGR*18insLQ. The C allele was associated with an approximately threefold increased risk of developing ovarian hyperstimulation syndrome in adult women undergoing procedures for assisted reproduction (O'Brien *et al.* 2013).

Few *LHB* gene variants are known. The so-called V-LH variant was discovered in Finland and is in the double

amino acid exchange p.W8R and p.I15T of *LHB* (Pettersson *et al.* 1992). V-LH shows a lower circulatory half time and bioactivity *in vivo* than the 'classical' LH, possibly compensated by increased transcriptional levels of the LH β subunit due to SNPs within the promoter *LHB* region, which are in linkage disequilibrium with p.W8R and p.I15T (Jiang *et al.* 1999). Curiously, V-LH may be a protective agent from symptomatic PCOS in obese women, among which it is less frequent compared to healthy women and non-obese PCOS patients (Tapanainen *et al.* 1999).

While the genes encoding the FSH β and LH β are present in all vertebrates, the CG β -coding genes exist only in primates and equids, likely as result of repeated duplications of an ancestral *LHB* gene (Henke & Gromoll 2008). The human genome carries eight *CGB* genes contiguous with the *LHB* gene on chromosome 19; subsequently, frame-shift mutations and nucleotide insertions resulted in 24 additional codons for *CGB*. The *LHB/CGB* gene cluster spans about 40 kbp and carries several SNPs; especially, polymorphic variants of the *CGB5* were associated with recurrent spontaneous abortions in Chinese and Caucasian women (Rull *et al.* 2008, Sun & Ji 2014).

Gonadotropin variants and implications in disease and menopause

Although further investigations are needed to elucidate the molecular mechanisms underlying the modulatory effects of SNPs within *FSHR* and *FSHB* genes on reproductive parameters and diseases, their pathophysiological relevance and clinical outcomes were widely described in the literature. On the contrary, the pathophysiological implications of SNPs belonging to the *LHCGR* gene and the *LHB/CGB* gene cluster are poorly understood.

Polycystic ovarian syndrome

PCOS is a common endocrine disorder affecting 4–10% of women of reproductive age. A wide number of candidate genes were found to be potential markers of the disease (Chen *et al.* 2011, Shi *et al.* 2012). PCOS women are characterized by heterogeneous sub-fertile phenotypes and related clinical features. Hyperandrogenism, metabolic syndrome, insulin resistance, and anovulation are some of the main clinical aspects of PCOS, which may be the result of endocrine adaptation to ancestral environmental conditions (Corbett & Morin-Papunen 2013, Casarini & Brigante 2014). Several studies searched evolutionary explanations for the origin of PCOS, suggesting that the energy saving resulting from less-ovulatory reproductive systems and insulin resistant phenotypes may be advantageous during seasons of food shortage or high energy demand, when indeed the

anovulation risk increases (Vitzthum *et al.* 2004, Vitzthum 2009, Corbett & Morin-Papunen 2013). However, theories supporting natural selection of PCOS phenotypes were downsized in favor of genetic drift; this issue is still debated and needs further investigation (Casarini & Brigante 2014). Gonadotropins and their receptors are logical candidate genes involved in the pathogenesis of the disease due to their crucial role in folliculogenesis and hormone regulation. However, conflicting data exist in the literature, because of the polygenic nature of the disease and the ethnic differences in the prevalence of lifestyle-related symptoms.

Alzheimer's disease

Alzheimer's disease is a progressive, neurodegenerative disorder characterized by neuronal and synaptic loss, neurofibrillary tangles located in neuronal cytoplasm, and deposition of amyloid in neuritic plaques. Genome-wide association studies (GWAS) suggested that SNPs within the *FSHR* and *LHCGR* genes may contribute to the pathogenesis of the disease (Sun *et al.* 2014). Especially, the polymorphism rs4073366 (c.161+28G>C) located within the first intron of the *LHCGR* gene was associated with a protective effect from the disease risk in males (Haasl *et al.* 2008).

Cancer

Gonadotropins activate multiple intracellular signaling pathways that may result in proliferative or anti-apoptotic events in primary cells and cell lines. In addition, gonadotropin receptors are expressed in several tumor cells (Mertens-Walker *et al.* 2012); thus, the possible link between hormone level and cancer risk was proposed.

FSHR p.N680S was indicated as a possible modulator of ovarian cancer (Yang *et al.* 2006, Ludwig *et al.* 2009) as well as *LHCGR* polymorphism 18insLQ, which may be linked with breast cancer risk (Powell *et al.* 2003). Some studies suggested that *LHB* SNPs are risk factors for cryptorchidism (Kaleva *et al.* 2005) and testicular cancer (Elkins *et al.* 2003). Interestingly, SNPs within gonadotropin genes were linked to papillary thyroid cancer risk (Schonfeld *et al.* 2012), revealing possible cross-activity among these molecules and their receptors.

Menopausal age

A link between menopausal age and SNPs in gonadotropins and their receptors' genes was suggested, providing a wide spectrum of candidate markers and conflicting, ethnicity-related results. Several loci associated with age at natural menopause were identified by meta-analyzing 22 GWAS in women of European ancestry (Stolk *et al.*

2012, Perry *et al.* 2014). This statistically powerful analysis identified top SNPs located within three out of 17 genomic regions in strong linkage disequilibrium with *FSHB*, *STARD1*, and *BCAR4* genes in Caucasians, suggesting that they are involved in the hormonal regulation of follicle recruitment and exhaustion, but further confirmation in other ethnic groups are required. Interestingly, women with PCOS have a later onset of menopause compared to normo-ovulatory women (Tehrani *et al.* 2010), likely resulting from the protective effect of high anti-Müllerian hormone levels for ovarian reserve, extending the reproductive lifespan in spite of less ovulatory cycles. Taken together, SNPs in the gonadotropins and their receptors' genes modulate fertility of both sexes and may affect the lifespan and reproductive health.

Limitations

Owing to the polygenic regulation and the modulatory effects of lifestyle on reproductive traits (Sharma *et al.* 2013), genotype–phenotype associations need to be well characterized in different, appropriately sized sample groups and independently confirmed to avoid methodological biases. However, the medical literature often provides conflicting results. Although the link between the FSHR SNP p.N680S and serum FSH levels or ovarian response was repeatedly observed (Simoni & Casarini 2014), other studies failed to find the same associations (Binder *et al.* 2012, Mohiyiddeen *et al.* 2013, Trevisan *et al.* 2014), suggesting that the endocrine features are modulated by several factors such as age or ethnicity. However, studies using suboptimal sample groups characterized by subfertility or endocrine dysfunction (e.g., premenopausal women or poor responders to gonadotropin treatments) should be carefully evaluated. Proper sample sizes and combined genotype analysis are required to detect significant and clinically relevant associations. For example, to unmask the effects of the p.N680S polymorphism on serum FSH levels in men, a combined model taking into account the *FSHB* promoter SNP –211G>T may be necessary (Tüttelmann *et al.* 2012). Association studies of polygenic traits should be replicated in different sample groups rigorously established and corroborated by *in vitro* evidences. Finally, mathematical corrections weighing the sample size from different investigations should provide the optimal verification; therefore, meta-analysis may be a safe and reliable tool to further confirm *in vivo* association studies.

Population genetics of gonadotropins and gonadotropin receptors' polymorphisms

Previous studies demonstrated that the Africa holds the highest human genetic variability worldwide (Cann *et al.* 2002, Ramachandran *et al.* 2005, Li *et al.* 2008).

Consistently with the routes of ancient human migrations, genetic variability decreases together with the distance from Africa, and oppositely to the genetic diversity, determining the current distribution of several sex-related genetic markers (Casarini & Brigante 2014). Because natural selection contributed poorly to the distribution of human genotypes worldwide (Li *et al.* 2008), it is reasonable that slightly different hormonal levels and menstrual cycle duration may have only a marginal impact on the selection of sex-related genotypes, compared to other, more determinant phenotypic features, such as skin pigmentation or sickle cell anemia (Liu *et al.* 2013).

On the other hand, a full explanation of human reproductive success may not merely rely on human migrations or genetic drift, and the evolutionary role of the SNPs in gonadotropin and their receptors' genes was debated (Grigorova *et al.* 2007, Simoni & Casarini 2014). It was estimated that about 20% of Caucasians carry a 'less favorable' *FSHB/FSHR* genotype in terms of serum FSH levels and FSHR expression and activity, which are enriched in sub-fertile subjects previously studied (Simoni & Casarini 2014). Especially, ovarian cycle length depends, at least in part, on the combination of *FSHB* and *FSHR* genotypes, which affect the sensibility threshold to FSH. This results in heterogeneity in menstrual cycle length and, consequently, a theoretical difference in the total number of cycles that can be calculated in about ± 30 –40 ovarian cycles during the reproductive lifespan depending on the *FSHR* genotype. FSHR p.N680S S homozygous women have longer ovarian cycles than p.N680S N homozygous women (Greb *et al.* 2005). In fact, the FSHR variant carrying the amino acid serine at position 680 is more abundant in South-Central Asians and Oceanians (Simoni & Casarini 2014) who are characterized by an overall longer cycle duration than women of East Asian, European, or African ancestry (Vitzthum 2009). This is consistent with the lower steroidogenic potential of the FSHR p.N680S homozygous S compared to the homozygous N genotype (Casarini *et al.* 2014). Most importantly, this suggests that some women have a lower number of ovulations for months of exposure, potentially resulting in slightly lower reproductive potential but preserving the individual from unnecessary energy expenditure to maintain overall fitness (Simoni & Casarini 2014). However, because women with low cycle variability have a higher conception rate than those with longer but irregular cycle duration, pregnancy success depends on cycle quality rather than length (Vitzthum 2009).

Prenatal maternal investments give a key contribution in maintaining progeny (Vitzthum 2009), suggesting that the genotype of *LHB/CGB* gene cluster is important to optimize the birth rate across human evolution. Protective effect from recurrent miscarriage was associated with some SNPs located in both the *CGB5* and *CGB8* genes, which encode the major fraction of

CGB-mRNA transcripts (Rull *et al.* 2008) reflecting their importance in physiological adaptation to pregnancy. The genomic region embedding the *CGB2*, *CGB5*, and *CGB8* promoter genes is featured by high heterozygosity and increased frequencies of the derived alleles in non-African populations (Fig. 1). On the contrary, ancestral alleles of *CGB2*, *CGB5*, and *CGB8* promoter genes achieve the highest frequencies among individuals of African ancestry (Fig. 1). Moreover, high heterozygosity in non-Africans suggests that balancing selection accompanied ancient human migrations (Rull *et al.* 2008). Taken together, this is consistent with the concept that genotypic (thus, phenotypic) variability improves the persistence of a population in a given habitat (Forsman 2014), providing more flexible reproductive features, such as endocrine adaptation to the new environmental conditions (Cornelius *et al.* 2013) reasonably encountered out from Africa. Interestingly, the analysis of the *LHB/CGB* cluster sequences from several human populations revealed selective pressures among Africans compared to humans in other continents (Fig. 2). The Cross Population Extended Haplotype Homozygosity (XP-EHH) test (Sabeti *et al.* 2007), a measure of natural selection that takes into account the SNPs frequencies within a genomic region, is higher when calculated for the *LHB/CGB* gene cluster of individuals from Africa compared to other populations. Because African populations maintained high homozygosity for the *LHB* gene and *CGB2*, *CGB5*, and *CGB8* promoter genes (Fig. 1), this was likely an advantageous condition in (but not out from) Africa. This conflicts with the concept that Africa, where human species arose, holds the highest heterozygosity and genetic variability (Cann *et al.* 2002, Ramachandran *et al.* 2005, Li *et al.* 2008). Also, because chorionic gonadotropin is massively produced exclusively in pregnant females, the *CGB* gene cluster is reasonably the result of selection

acting only in women, providing an interesting model to study sex-related aspects of the human evolution. However, the contribution of males in the selection of *LHB/CGB* cluster genotypes should not to be excluded, at least in Africans; paternal transmission of methylated SNPs within *CGB5* promoter results in the loss of bi-allelic expression, leading to the failure of pregnancy by impairment of placental–maternal interface (Uusküla *et al.* 2011). In addition, a role of certain *CGβ* transcripts in the male reproductive system was proposed (Parrott *et al.* 2011) suggesting that paternal inheritance of *LHB/CGB* cluster genotypes was important for pregnancy in daughters.

An evolutionary role of pregnancy may consist in protecting from disease risk due to long-term exposure to physiologic pituitary gonadotropins (Meier-Abt *et al.* 2015) and a link between fertility and lifespan was indeed observed (Kuningas *et al.* 2011); it is plausible, even if speculative, that a longer lifespan could provide a wider reproductive window. However, the impact of life duration in human evolution remains unclear, because the mean life expectancy was overall <40 years worldwide until the beginning of the 20th century, mainly due to causes unrelated to hormonal features (e.g., infectious diseases, famines, etc.; Christensen *et al.* 2009), thus suggesting that the reproductive lifespan had mild beneficial effects for human reproduction.

Postnatal parental care is important for progeny growth, improving reproductive success (Vitzthum 2009). Because sexual behavior and fatherhood are linked to testosterone levels in men (Gettler *et al.* 2013), the functional significance of hormonal changes in mammalian males was debated (Saltzman & Ziegler 2014). While high testosterone levels favor the male in acquiring sex partners, increased paternal care was associated with low testosterone levels in humans (Perini *et al.* 2012, Pollet *et al.* 2013). Therefore, genotypes

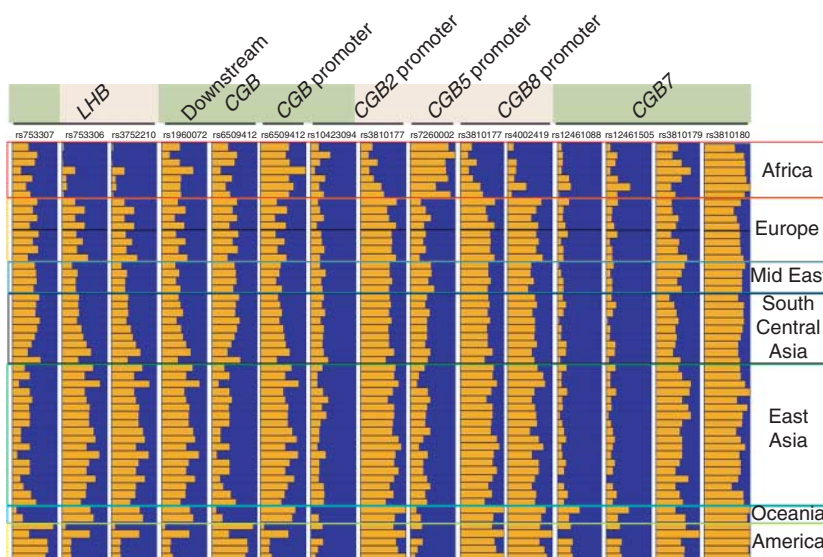


Figure 1 Allele frequencies of SNPs within *LHB/CGB* gene cluster in human populations. Data were obtained using the Human Genome Diversity Project (HGDP) selection browser (<http://hgdp.uchicago.edu/cgi-bin/gbrowse/HGDP>). Orange/blue bars indicate the proportion in percentage of the two alleles in the different human groups, which are represented by the colored lines in each column (please refer to the HGDP website for the populations order and name). The populations belonging to the same geographical area were grouped as indicated on the right side of the panel. SNPs ID are shown above each column and grouped by gene. Pink panels above the bars indicate when mean SNP frequencies of Africa are significantly different vs that of all other continents (Kruskal–Wallis and Dunn’s post-test; $P < 0.001$); nonsignificant differences are indicated by green panels (exceptions: Africa vs America for SNPs rs753306 and rs3752210, $P \geq 0.001$).

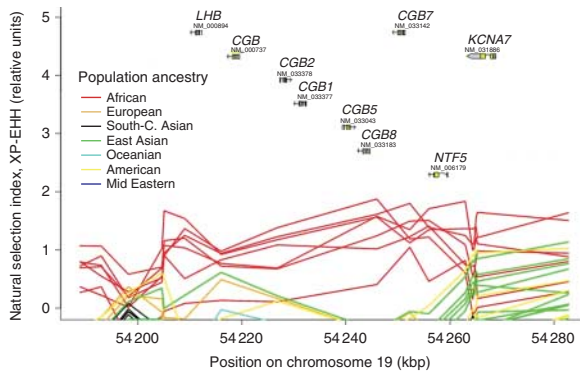


Figure 2 Analysis of the natural selection pressure sustained by the *LHB/CGB* gene cluster. The measure of natural selection was inferred from the gene cluster sequences of several human populations using the XP-EHH index (Sabeti *et al.* 2007) and represented on the Y-axis (relative units). The name, ID, and exon sequences (boxes and arrows) of each gene are indicated on the panel in proximity of their genomic position on chromosome 19 (X-axis). Red lines corresponding to measures of natural selection of the *LHB/CGB* cluster in Africans achieve higher levels than that of other populations, indicating that stronger natural selection occurs in Africans compared to other populations. The populations belonging to the same geographical area were grouped and colored as indicated in the legend (top-left side of the panel); please refer to the Human Genome Diversity Project (HGDP) selection browser for the population name list (<http://hgdp.uchicago.edu/cgi-bin/gbrowse/HGDP>). The calculation of the XP-EHH index was performed by the proper online tool available at the HGDP selection browser.

linked to low fertility may have provided an evolutionary advantage, especially when the adaptation to new environmental factors favored the need of cooperative behaviors among kin (Apicella *et al.* 2012), which should be plausibly strengthened during ancient migration of relatively small human groups. This may explain why the relatively recent SNP variants associated with lower fertile phenotypes, such as rs1394205 ($-29G>A$, *FSHR*) and rs10835638 ($-211G>T$, *FSHB*) (Grigорова *et al.* 2008, Tüttelmann *et al.* 2012), have higher frequencies among Northern European and native American populations than in Africa, where humans arose (Simoni & Casarini 2014). However, the current distribution of genotypes evolutionarily disadvantageous among humans may be due, at least in part, to social issues (e.g., patrilineal populations) that affect the genetic diversity by sex-biased transmission of reproductive success (Heyer *et al.* 2015).

Reproductive conflicts

Intralocus sexual conflict occurs when traits encoded by the same genetic locus result in opposite effects in males and females, in terms of reproductive success (Pennell & Morrow 2013). This was experimentally demonstrated in animal models, revealing that high levels of the sex hormone testosterone result in different, sex-related reproductive success in the bank vole *Myodes glareolus*

(Mills *et al.* 2012). In this model, high testosterone levels were oppositely associated with the reproductive success of sons and daughters; thus, genetic benefits of selecting reproductively successful males with high testosterone levels were lost with daughters. This may explain why genetic variants linked to sub-fertile phenotypes in females did not disappear during evolution. Because risk alleles may have been maintained in a population due to their beneficial effect in one sex (Gilks *et al.* 2014), GWAS of sex-specific reproductive disorders could be improved by including both sexes, rather than separate-sex analysis. Unfortunately, sex-related genetic disorders (e.g., PCOS) are usually investigated by excluding male samples. Using human genotypic data from both males and females, we recently observed that sexual conflict might explain the geographic distribution of PCOS risk alleles and the overall constant prevalence of the disease (Casarini & Brigante 2014). In particular, we observed that genotypes linked to hyperandrogenic phenotypes could have been evolutionarily favorable for males in challenging for food resources, although disadvantageous for females in which they are involved in PCOS pathogenesis. PCOS markers are SNPs located within several genomic regions, including *FSHR* and *LHCGR* genes (Chen *et al.* 2011, Shi *et al.* 2012). Because gonadotropin receptor genes are linked to testosterone levels and testes volume in men (Grigорова *et al.* 2014), they may be hot spots for intralocus sexual conflicts by oppositely modulating the reproductive parameters in a sex-dependent manner.

Even if speculative, the evolution of the *LHB/CGB* gene cluster may be a case of solved intralocus sexual conflict occurred via sexual dimorphism by gene duplication (Assis & Bachtrog 2013), resulting in the independent evolution of novel functions of the derived genes. In this sense, gestation and embryo development in primates are controlled by several copies of the *CGB* gene derived from the original *LHB* gene (Henke & Gromoll 2008, Nagirnaja *et al.* 2010), which, in turn, maintains the original physiologic functions exerted in the development, folliculogenesis, ovulation, and spermatogenesis in all animals but the primates. In primates, the number of *CGB* genes increase together with complexity of hemochorial placentation (Cole 2009), revealing that they have different, widely unknown roles in pregnancy and evolved separately. The *CGB1* and *CGB2* genes are highly conserved in humans and great apes, and a low number of SNPs map in the proximity of these genes. Owing to the low genetic variation of *CGB1* and *CGB2* genes, it is plausible that they are dedicated to the regulation of delicate stages such as embryo implantation and placental development (Hallast *et al.* 2007), which are crucial for pregnancy in all primates. Other *CGB* genes are abundantly transcribed in different gestational periods, suggesting that they may serve for further species-specific adaptations to later stages of pregnancy.

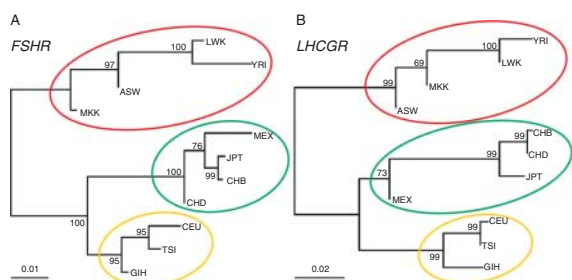


Figure 3 Phylogenetic analysis of the *FSHR* (A) and *LHCGR* (B) genes. SNPs frequencies were extracted from HapMap populations (<http://hapmap.ncbi.nlm.nih.gov>) and analyzed by the POPTREE2 Software (Takezaki *et al.* 2010). The populations belonging to the same geographical area were grouped by colored ovals (red, populations of African ancestry; green, East Asian/American; yellow, European/Caucasian/Central Asian), resulting in phylogenetic patterns of both the *FSHR* and *LHCGR* genotypes according to the continental distribution of the human groups. The populations were assigned to each continent depending on the major genetic component of their ancestry (Jia *et al.* 2014); ASW was assumed as African, CHD as East Asian, GIH as Central Asian, CEU as Caucasian from Europe despite they are USA residents. The measure of genetic distance Fixation index (F_{st}) is indicated by the bars below the trees (relative frequency; please refer to the author's software and article for references about genetic distance); the numbers throughout the trees are percentage values representing an index of reliability of the analysis, which is assumed significantly reliable when ≥ 70 –75 (relative units) (Takezaki *et al.* 2010). POPTREE2 Software was used with these default settings: F_{st} uncorrected, NJ, Bootstrap 100 000.

Phylogenesis

Owing to the polygenic modulation of the sexual features, it is overall difficult to quantify the real impact of each genotypic variant of the gonadotropins and their receptors' genes in human reproductive success (Casarini *et al.* 2011). The overall, worldwide distribution of genotypic markers results in a geographical pattern due to human migrations rather than selection (Ramachandran *et al.* 2005, Li *et al.* 2008). Human phylogenetic trees produced using SNP frequencies of the whole *FSHR* and *LHCGR* genes from the HapMap database (International HapMap Consortium 2003) by the POPTREE2 Software (Life Science Research Center, Kagawa University, Kagawa, Japan) (Takezaki *et al.* 2010) (Fig. 3) revealed, indeed, that the genotypic variants of both the genes are embedded in continent-specific groups, depending on the genetic ancestry of the populations (Jia *et al.* 2014). This suggests that human populations may be represented by three main *FSHR* and *LHCGR* genotypes peculiar of Africa, Eurasian, and East Asian-American continents, supporting that ancient human migrations gave the main contribution to the current genetic diversity. This analysis did not take into account that few SNPs may have contributed to the selection of peculiar phenotypes (e.g., *FSHR* p.N680S; rs6166) more than

others (e.g., non-synonymous or intronic polymorphic variants). However, the *FSHR* and *LHCGR* genes are characterized by genomic regions in high linkage disequilibrium (Simoni & Casarini 2014), except in Africans, suggesting that they were inherited together. Taken together, gonadotropin receptor gene variants seem to have accompanied humans during ancient migrations only weakly contributing to their reproductive success.

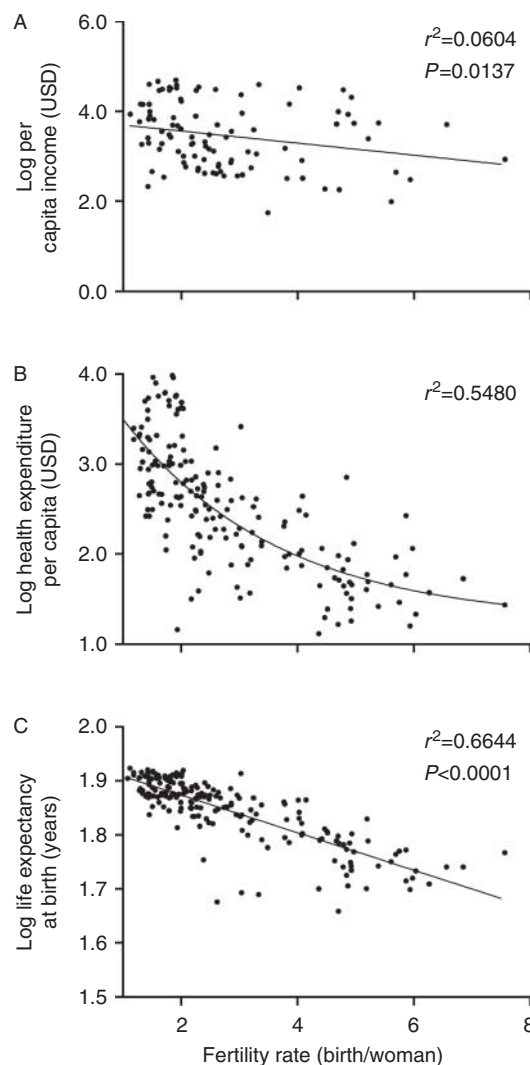


Figure 4 Relationship between fertility rate and socioeconomic current indexes in world countries. Fertility rate is represented as birth per woman and plotted against measures of socioeconomic status, i.e., per capita income (A), health expenditure per capita (B), and life expectancy at birth (C) (logarithmic Y-axis). Fertility rate is inversely related to all of these indexes, demonstrating that the countries in which people have a high standard of living are featured by a low number of births and *vice versa* (linear or non-linear regression were used where appropriate as best-fitting model; $P < 0.005$; calculation by GraphPad Prism, GraphPad Software, Inc., La Jolla, CA, USA). The graphs were obtained using data available at the World Bank Group website (<http://www.worldbank.org>), an observer at the United Nations Development Group.

Socioeconomic and cultural aspects of human reproduction

It is unclear how the endocrine genotypes and phenotypes affect human reproductive success in the modern, developed societies in which family structure, lifestyle, and health care deeply changed during the last century and appear now profoundly different from those of ancient times. Currently, different world regions differ widely in fertility rate. The number of births per woman is inversely related with socioeconomical indexes (per capita income, health expenditure, and life expectancy; Fig. 4), so that the highest income countries have the lowest fertility rate, and this is not depending on ethnicity (data available at the World Bank Group, <http://www.worldbank.org>). In low income countries, the mean fertility rate achieves six to eight births per woman. This means that reproductive success in current, developed human societies is merely depending on social and cultural aspects reflected by richness, health, trust in the future, etc., while it is poorly affected by the endocrine phenotype of the individuals. Couples of developed countries currently begin to search fertility and parenthood at late reproductive age, e.g., 35–40 years, when the reproductive success and birth rate are naturally low, mainly due to decreased ovarian reserve and/or metabolic disturbances that amplify the effects of sub-fertile phenotypes. This explains why several developed countries are currently characterized by population aging and demographic decline as compared to high fertility rates observed in the poorest countries (Bongaarts 2015). Therefore, the socioeconomic status is currently linked to reproductive success. In addition, in ancient human societies sexual activity aiming at conception were concomitant with the beginning of the fertile age and persisted for longer times, plausibly increasing the chance for parenthood as it continues to occur in the poorest countries. Endocrine and metabolic disorders such as hyperandrogenism or insulin resistance, which result in sub-fertile female phenotypes (Corbett & Morin-Papunen 2013), might significantly affect fertility in the modern, developed societies where conception attempts per individual are reasonably fewer compared to the ancient times. If so, then the genotypic features, irrelevant in the past, may be relevant to optimize fertility management in the modern societies, when an increasing number of 'reproductively aged' couples, characterized by a reduced fertile window, undergo clinical treatments for assisted reproduction.

Conclusions

An increasing number of studies progressively elucidate how polymorphic variants of gonadotropins and their receptors' genes modulate the human reproductive functions and diseases. Although traces of selective pressure on genes related to endocrine functions were

found, the effects of gonadotropins and their receptors' SNPs should normally have a relatively weak impact in human reproductive success. Peculiar endocrine genotypes may be linked to phenotypes leading to opposite, sex-related reproductive success, resulting in intralocus sexual conflicts and favoring the inheritance of alleles disadvantageous for one sex through the ancient human history. Thus, individuals from both sexes and proper sample sizes should be required in GWAS and evolutionary studies in the field of reproduction. The endocrine phenotypes related to subfertility may strengthen the decline of fertility in modern societies in which parenthood attempts are relegated in the last, short period of the fertile age.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review.

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References

- Achrekar SK, Modi DN, Desai SK, Mangoli VS, Mangoli RV & Mahale SD 2009 Poor ovarian response to gonadotrophin stimulation is associated with FSH receptor polymorphism. *Reproductive Biomedicine Online* **18** 509–515. (doi:10.1016/S1472-6483(10)60127-7)
- Apicella CL, Marlowe FW, Fowler JH & Christakis NA 2012 Social networks and cooperation in hunter-gatherers. *Nature* **481** 497–501. (doi:10.1038/nature10736)
- Ascoli M, Fanelli F & Segaloff DL 2002 The lutropin/choriogonadotropin receptor, a 2002 perspective. *Endocrine Reviews* **23** 141–174. (doi:10.1210/edrv.23.2.0462)
- Assis R & Bachtrog D 2013 Neofunctionalization of young duplicate genes in *Drosophila*. *PNAS* **110** 17409–17414. (doi:10.1073/pnas.1313759110)
- Binder H, Strick R, Zaherdoust O, Ditttrich R, Hamori M, Beckmann MW & Oppelt PG 2012 Assessment of FSHR variants and antiMüllerian hormone in infertility patients with a reduced ovarian response to gonadotropin stimulation. *Fertility and Sterility* **97** 1169–1175.e1. (doi:10.1016/j.fertnstert.2012.02.012)
- Bongaarts J 2015 Global fertility and population trends. *Seminars in Reproductive Medicine* **33** 5–10. (doi:10.1055/s-0034-1395272)
- Cann HM, de Toma C, Cazes L, Legrand MF, Morel V, Piouffre L, Bodmer J, Bodmer WF, Bonne-Tamir B, Cambon-Thomsen A *et al.* 2002 A human genome diversity cell line panel. *Science* **296** 261–262. (doi:10.1126/science.296.5566.261b)
- Casarini L & Brigante G 2014 The polycystic ovary syndrome evolutionary paradox: a genome-wide association studies-based, in silico, evolutionary explanation. *Journal of Clinical Endocrinology and Metabolism* **99** E2412–E2420. (doi:10.1210/jc.2014-2703)

- Casarini L, Pignatti E & Simoni M 2011 Effects of polymorphisms in gonadotropin and gonadotropin receptor genes on reproductive function. *Reviews in Endocrine & Metabolic Disorders* **12** 303–321. (doi:10.1007/s11154-011-9192-2)
- Casarini L, Moriondo V, Marino M, Adversi F, Capodanno F, Grisolia C, La Marca A, La Sala GB & Simoni M 2014 FSHR polymorphism p.N680S mediates different responses to FSH *in vitro*. *Molecular and Cellular Endocrinology* **393** 83–91. (doi:10.1016/j.mce.2014.06.013)
- Chen ZJ, Zhao H, He L, Shi Y, Qin Y, Shi Y, Li Z, You L, Zhao J, Liu J *et al.* 2011 Genome-wide association study identifies susceptibility loci for polycystic ovary syndrome on chromosome 2p16.3, 2p21 and 9q33.3. *Nature Genetics* **43** 55–59. (doi:10.1038/ng.732)
- Christensen K, Doblhammer G, Rau R & Vaupel JW 2009 Ageing populations: the challenges ahead. *Lancet* **374** 1196–1208. (doi:10.1016/S0140-6736(09)61460-4)
- Cole LA 2009 hCG and hyperglycosylated hCG in the establishment and evolution of hemochorial placentation. *Journal of Reproductive Immunology* **82** 112–118. (doi:10.1016/j.jri.2009.04.007)
- Corbett S & Morin-Papunen L 2013 The polycystic ovary syndrome and recent human evolution. *Molecular and Cellular Endocrinology* **373** 39–50. (doi:10.1016/j.mce.2013.01.001)
- Cornelius JM, Watts HE, Dingle H & Hahn TP 2013 Obligate versus rich patch opportunism: evolution and endocrine mechanisms. *General and Comparative Endocrinology* **176**–80. (doi:10.1016/j.ygcen.2013.04.003)
- Elkins DA, Yokomizo A, Thibodeau SN, J Schaid D, Cunningham JM, Marks A, Christensen E, McDonnell SK, Slager S, J Peterson B *et al.* 2003 Luteinizing hormone β polymorphism and risk of familial and sporadic prostate cancer. *Prostate* **56** 30–36. (doi:10.1002/pros.10220)
- Gilks WP, Abbott JK & Morrow EH 2014 Sex differences in disease genetics: evidence, evolution, and detection. *Trends in Genetics* **30** 453–463. (doi:10.1016/j.tig.2014.08.006)
- Gloaguen P, Crépieux P, Heitzler D, Poupon A & Reiter E 2011 Mapping the follicle-stimulating hormone-induced signaling networks. *Frontiers in Endocrinology* **2** 45. (doi:10.3389/fendo.2011.00045)
- Grigороva M, Rull K & Laan M 2007 Haplotype structure of FSHB, the β -subunit gene for fertility-associated follicle-stimulating hormone: possible influence of balancing selection. *Annals of Human Genetics* **71** 18–28. (doi:10.1111/j.1469-1809.2006.00299.x)
- Grigороva M, Punab M, Ausmees K & Laan M 2008 FSHB promoter polymorphism within evolutionary conserved element is associated with serum FSH level in men. *Human Reproduction* **23** 2160–2166. (doi:10.1093/humrep/den216)
- Grigороva M, Punab M, Punab AM, Poolamets O, Vihljajev V, Zilaitienė B, Erenpreiss J, Matulevičius V & Laan M 2014 Reproductive physiology in young men is cumulatively affected by FSH-action modulating genetic variants: FSHR –29G/A and c.2039 A/G, FSHB –211G/T. *PLoS ONE* **9** e94244. (doi:10.1371/journal.pone.0094244)
- Forsman A 2014 Effects of genotypic and phenotypic variation on establishment are important for conservation, invasion, and infection biology. *PNAS* **111** 302–307. (doi:10.1073/pnas.1317745111)
- Gettler LT, McDade TW, Agustín SS, Feranil AB & Kuzawa CW 2013 Do testosterone declines during the transition to marriage and fatherhood relate to men's sexual behavior? Evidence from the Philippines *Hormones and Behavior* **64** 755–763. (doi:10.1016/j.yhbeh.2013.08.019)
- Greb RR, Grieshaber K, Gromoll J, Sonntag B, Nieschlag E, Kiesel L & Simoni M 2005 A common single nucleotide polymorphism in exon 10 of the human follicle stimulating hormone receptor is a major determinant of length and hormonal dynamics of the menstrual cycle. *Journal of Clinical Endocrinology and Metabolism* **90** 4866–4872. (doi:10.1210/jc.2004-2268)
- Haasi RJ, Ahmadi MR, Meethal SV, Gleason CE, Johnson SC, Asthana S, Bowen RL & Atwood CS 2008 A luteinizing hormone receptor intronic variant is significantly associated with decreased risk of Alzheimer's disease in males carrying an apolipoprotein E epsilon4 allele. *BMC Medical Genetics* **9** 37. (doi:10.1186/1471-2350-9-37)
- Hallast P, Rull K & Laan M 2007 The evolution and genomic landscape of CGB1 and CGB2 genes. *Molecular and Cellular Endocrinology* **260**–262 2–11. (doi:10.1016/j.mce.2005.11.049)
- Henke A & Gromoll J 2008 New insights into the evolution of chorionic gonadotropin. *Molecular and Cellular Endocrinology* **291** 11–19. (doi:10.1016/j.mce.2008.05.009)
- Heyer E, Brandenburg JT, Leonardi M, Toupance B, Balaresque P, Hegay T, Aldashev A & Austerlitz F 2015 Patrilineal populations show more male transmission of reproductive success than cognatic populations in Central Asia, which reduces their genetic diversity. *American Journal of Physical Anthropology* **157** 537–543. (doi:10.1002/ajpa.22739)
- International HapMap Consortium 2003 The International HapMap Project. *Nature* **426** 789–796. (doi:10.1038/nature02168)
- Jia J, Wei YL, Qin CJ, Hu L, Wan LH & Li CX 2014 Developing a novel panel of genome-wide ancestry informative markers for bio-geographical ancestry estimates. *Forensic Science International. Genetics* **8** 187–194. (doi:10.1016/j.fsigen.2013.09.004)
- Jiang M, Pakarinen P, Zhang FP, El-Hefnawy T, Koskimies P, Pettersson K & Huhtaniemi I 1999 A common polymorphic allele of the human luteinizing hormone β -subunit gene: additional mutations and differential function of the promoter sequence. *Human Molecular Genetics* **8** 2037–2046. (doi:10.1093/hmg/8.11.2037)
- Kaleva M, Virtanen H, Haavisto AM, Main K, Skakkebaek NE, Huhtaniemi I, Irjala K & Toppari J 2005 Does variant luteinizing hormone (V-LH) predispose to improper testicular position in late pregnancy? *Pediatric Research* **58** 447–450. (doi:10.1203/01.pdr.0000176918.68539.b4)
- Kuningas M, Altmäe S, Uitterlinden AG, Hofman A, van Duijn CM & Tiemeier H 2011 The relationship between fertility and lifespan in humans. *Age* **33** 615–622. (doi:10.1007/s11357-010-9202-4)
- La Marca A, Papaleo E, Alviggi C, Ruvalo G, De Placido G, Candiani M, Cittadini E, De Michele F, Moriondo V, Catellani V *et al.* 2013 The combination of genetic variants of the FSHB and FSHR genes affects serum FSH in women of reproductive age. *Human Reproduction* **28** 1369–1374. (doi:10.1093/humrep/det061)
- Li JZ, Absher DM, Tang H, Southwick AM, Casto AM, Ramachandran S, Cann HM, Barsh GS, Feldman M, Cavalli-Sforza LL *et al.* 2008 Worldwide human relationships inferred from genome-wide patterns of variation. *Science* **319** 1100–1104. (doi:10.1126/science.1153717)
- Liu X, Ong RT, Pillai EN, Elzein AM, Small KS, Clark TG, Kwiatkowski DP & Teo YY 2013 Detecting and characterizing genomic signatures of positive selection in global populations. *American Journal of Human Genetics* **92** 866–881. (doi:10.1016/j.ajhg.2013.04.021)
- Lledo B, Guerrero J, Turienzo A, Ortiz JA, Morales R, Ten J, Llacer J & Bernabeu R 2013 Effect of follicle-stimulating hormone receptor N680S polymorphism on the efficacy of follicle-stimulating hormone stimulation on donor ovarian response. *Pharmacogenetics and Genomics* **23** 262–268. (doi:10.1097/FPC.0b013e32835fe813)
- Ludwig AH, Murawska M, Panek G, Timorek A & Kuprynczyk J 2009 Androgen, progesterone, and FSH receptor polymorphisms in ovarian cancer risk and outcome. *Endocrine-Related Cancer* **16** 1005–1016. (doi:10.1677/ERC-08-0135)
- Meier-Abt F, Bentires-Alj M & Rochlitz C 2015 Breast cancer prevention: lessons to be learned from mechanisms of early pregnancy-mediated breast cancer protection. *Cancer Research* **75** 803–807. (doi:10.1158/0008-5472.CAN-14-2717)
- Mertens-Walker I, Baxter RC & Marsh DJ 2012 Gonadotropin signalling in epithelial ovarian cancer. *Cancer Letters* **324** 152–159. (doi:10.1016/j.canlet.2012.05.017)
- Mills SC, Koskela E & Mappes T 2012 Intralocus sexual conflict for fitness: sexually antagonistic alleles for testosterone. *Proceedings. Biological Sciences/The Royal Society* **279** 1889–1895. (doi:10.1098/rspb.2011.2340)
- Mohiyiddeen L, Newman WG, Cerra C, McBurney H, Mulugeta B, Roberts SA & Nardo LG 2013 A common Asn680Ser polymorphism in the follicle-stimulating hormone receptor gene is not associated with ovarian response to gonadotropin stimulation in patients undergoing *in vitro* fertilization. *Fertility and Sterility* **99** 149–155. (doi:10.1016/j.fertnstert.2012.08.037)
- Nagirajna L, Rull K, Uusküla L, Hallast P, Grigороva M & Laan M 2010 Genomics and genetics of gonadotropin β -subunit genes: unique FSHB and duplicated LHB/CGB loci. *Molecular and Cellular Endocrinology* **329** 4–16. (doi:10.1016/j.mce.2010.04.024)
- Nakayama T, Kuroi N, Sano M, Tabara Y, Katsuya T, Ogihara T, Makita Y, Hata A, Yamada M, Takahashi N *et al.* 2006 Mutation of the follicle-stimulating hormone receptor gene 5'-untranslated region associated with female hypertension. *Hypertension* **48** 512–518. (doi:10.1161/01.HYP.0000233877.84343.d7)

- O'Brien TJ, Kalmin MM, Harralson AF, Clark AM, Gindoff I, Simmens SJ, Frankfurter D & Gindoff P 2013 Association between the luteinizing hormone/chorionic gonadotropin receptor (LHCGR) rs4073366 polymorphism and ovarian hyperstimulation syndrome during controlled ovarian hyperstimulation. *Reproductive Biology and Endocrinology* **11** 71. (doi:10.1186/1477-7827-11-71)
- Parrott AM, Sriram G, Liu Y & Mathews MB 2011 Expression of type II chorionic gonadotropin genes supports a role in the male reproductive system. *Molecular and Cellular Biology* **31** 287–299. (doi:10.1128/MCB.00603-10)
- Pennell TM & Morrow EH 2013 Two sexes, one genome: the evolutionary dynamics of intralocus sexual conflict. *Ecology and Evolution* **3** 1819–1834. (doi:10.1002/ece3.540)
- Perini T, Ditzen B, Hengartner M & Ehlert U 2012 Sensation seeking in fathers: the impact on testosterone and paternal investment. *Hormones and Behavior* **61** 191–195. (doi:10.1016/j.yhbeh.2011.12.004)
- Perry JR, Day F, Elks CE, Sulem P, Thompson DJ, Ferreira T, He C, Chasman DI, Esko T, Thorleifsson G *et al.* 2014 Parent-of-origin-specific allelic associations among 106 genomic loci for age at menarche. *Nature* **514** 92–97. (doi:10.1038/nature13545)
- Pettersson K, Ding YQ & Huhtaniemi I 1992 An immunologically anomalous luteinizing hormone variant in a healthy woman. *Journal of Clinical Endocrinology & Metabolism* **74** 164–171. (doi:10.1210/jcem.74.1.1727817)
- Piersma D, Berns EM, Verhoef-Post M, Uitterlinden AG, Braakman I, Pols HA & Themmen AP 2006 A common polymorphism renders the luteinizing hormone receptor protein more active by improving signal peptide function and predicts adverse outcome in breast cancer patients. *Journal of Clinical Endocrinology and Metabolism* **91** 1470–1476. (doi:10.1210/jc.2005-2156)
- Pollet TV, Cobey KD & van der Meij L 2013 Testosterone levels are negatively associated with fatherhood (corrected) in males, but positively related to offspring count in fathers. *PLoS ONE* **8** e60018. (doi:10.1371/journal.pone.0060018)
- Powell BL, Piersma D, Kevenaar ME, van Staveren IL, Themmen AP, Iacopetta BJ & Berns EM 2003 Luteinizing hormone signaling and breast cancer: polymorphisms and age of onset. *Journal of Clinical Endocrinology and Metabolism* **88** 1653–1657. (doi:10.1210/jc.2002-021585)
- Ramachandran S, Deshpande O, Roseman CC, Rosenberg NA, Feldman MW & Cavalli-Sforza LL 2005 Support from the relationship of genetic and geographic distance in human populations for a serial founder effect originating in Africa. *PNAS* **102** 15942–15947. (doi:10.1073/pnas.0507611102)
- Rull K, Nagirnaja L, Ulander VM, Kelgo P, Margus T, Kaare M, Aittomäki K & Laan M 2008 Chorionic gonadotropin β -gene variants are associated with recurrent miscarriage in two European populations. *Journal of Clinical Endocrinology and Metabolism* **93** 4697–4706. (doi:10.1210/jc.2008-1101)
- Sabeti PC, Varilly P, Fry B, Lohmueller J, Hostetter E, Cotsapas C, Xie X, Byrne EH, McCarroll SA, Gaudet R *et al.* 2007 Genome-wide detection and characterization of positive selection in human populations. *Nature* **449** 913–918. (doi:10.1038/nature06250)
- Saltzman W & Ziegler TE 2014 Functional significance of hormonal changes in mammalian fathers. *Journal of Neuroendocrinology* **26** 685–696. (doi:10.1111/jne.12176)
- Schonfeld SJ, Neta G, Sturgis EM, Pfeiffer RM, Hutchinson AA, Xu L, Wheeler W, Guénel P, Rajaraman P, de Vathaire F *et al.* 2012 Common genetic variants in sex hormone pathway genes and papillary thyroid cancer risk. *Thyroid* **22** 151–156. (doi:10.1089/thy.2011.0309)
- Schüring AN, Busch AS, Bogdanova N, Gromoll J & Tüttelmann F 2013 Effects of the FSH- β -subunit promoter polymorphism $-211G \rightarrow T$ on the hypothalamic–pituitary–ovarian axis in normally cycling women indicate a gender-specific regulation of gonadotropin secretion. *Journal of Clinical Endocrinology and Metabolism* **98** E82–E86. (doi:10.1210/jc.2012-2780)
- Sharma R, Biedenharn KR, Fedor JM & Agarwal A 2013 Lifestyle factors and reproductive health: taking control of your fertility. *Reproductive Biology and Endocrinology* **11** 66. (doi:10.1186/1477-7827-11-66)
- Shi Y, Zhao H, Shi Y, Cao Y, Yang D, Li Z, Zhang B, Liang X, Li T, Chen J *et al.* 2012 Genome-wide association study identifies eight new risk loci for polycystic ovary syndrome. *Nature Genetics* **44** 1020–1025. (doi:10.1038/ng.2384)
- Simoni M & Casarini L 2014 Mechanisms in endocrinology: Genetics of FSH action: a 2014-and-beyond view. *European Journal of Endocrinology* **170** R91–R107. (doi:10.1530/EJE-13-0624)
- Simoni M, Gromoll J & Nieschlag E 1997 The follicle-stimulating hormone receptor: biochemistry, molecular biology, physiology, and pathophysiology. *Endocrine Reviews* **18** 739–773. (doi:10.1210/edrv.18.6.0320)
- Simoni M, Tüttelmann F, Michel C, Böckenfeld Y, Nieschlag E & Gromoll J 2008 Polymorphisms of the luteinizing hormone/chorionic gonadotropin receptor gene: association with maldescended testes and male infertility. *Pharmacogenetics and Genomics* **18** 193–200. (doi:10.1097/FPC.0b013e3282f4e98c)
- Stolk L, Perry JR, Chasman DI, He C, Mangino M, Sulem P, Barbalic M, Broer L, Byrne EM, Ernst F *et al.* 2012 Meta-analyses identify 13 loci associated with age at menopause and highlight DNA repair and immune pathways. *Nature Genetics* **44** 260–268. (doi:10.1038/ng.1051)
- Sun Y & Ji X 2014 Association of rs7260002 of chorionic gonadotropin $\beta 5$ with idiopathic recurrent spontaneous abortion in Chinese population. *Journal of Assisted Reproduction and Genetics* **31** 1497–1500. (doi:10.1007/s10815-014-0321-1)
- Sun J, Song F, Wang J, Han G, Bai Z, Xie B, Feng X, Jia J, Duan Y & Lei H 2014 Hidden risk genes with high-order intragenic epistasis in Alzheimer's disease. *Journal of Alzheimer's Disease* **41** 1039–1056. (doi:10.3233/JAD-140054)
- Takezaki N, Nei M & Tamura K 2010 POPTREE2: software for constructing population trees from allele frequency data and computing other population statistics with Windows interface. *Molecular Biology and Evolution* **27** 747–752. (doi:10.1093/molbev/msp312)
- Tapanainen JS, Koivunen R, Fauser BC, Taylor AE, Clayton RN, Rajkova M, White D, Franks S, Anttila L, Pettersson KS *et al.* 1999 A new contributing factor to polycystic ovary syndrome: the genetic variant of luteinizing hormone. *Journal of Clinical Endocrinology & Metabolism* **84** 1711–1715. (doi:10.1210/jcem.84.5.5702)
- Thathapudi S, Kodati V, Erukambattu J, Addepally U & Qurratulain H 2015 Association of luteinizing hormone chorionic gonadotropin receptor gene polymorphism (rs2293275) with polycystic ovarian syndrome. *Genetic Testing and Molecular Biomarkers* **19** 128–132. (doi:10.1089/gtmb.2014.0249)
- Tehrani FR, Solaymani-Dodaran M, Hedayati M & Azizi F 2010 Is polycystic ovary syndrome an exception for reproductive aging? *Human Reproduction* **25** 1775–1781. (doi:10.1093/humrep/deq088)
- Trevisan CM, Peluso C, Cordts EB, de Oliveira R, Christofolini DM, Barbosa CP & Bianco B 2014 Ala307Thr and Asn680Ser polymorphisms of FSHR gene in human reproduction outcomes. *Cellular Physiology and Biochemistry* **34** 1527–1535. (doi:10.1159/000366356)
- Tüttelmann F, Laan M, Grigorova M, Punab M, Söber S & Gromoll J 2012 Combined effects of the variants FSHB $-211G \rightarrow T$ and FSHR 2039A $> G$ on male reproductive parameters. *Journal of Clinical Endocrinology and Metabolism* **97** 3639–3647. (doi:10.1210/jc.2012-1761)
- Uusküla L, Rull K, Nagirnaja L & Laan M 2011 Methylation allelic polymorphism (MAP) in chorionic gonadotropin $\beta 5$ (CGB5) and its association with pregnancy success. *Journal of Clinical Endocrinology & Metabolism* **96** E199–E207.
- Vitzthum VJ 2009 The ecology and evolutionary endocrinology of reproduction in the human female. *American Journal of Physical Anthropology* **140** 95–136. (doi:10.1002/ajpa.21195)
- Vitzthum VJ, Spielvogel H & Thornburg J 2004 Interpopulational differences in progesterone levels during conception and implantation in humans. *PNAS* **101** 1443–1448. (doi:10.1073/pnas.0302640101)
- Yang CQ, Chan KY, Ngan HY, Khoo US, Chiu PM, Chan QK, Xue WC & Cheung AN 2006 Single nucleotide polymorphisms of follicle-stimulating hormone receptor are associated with ovarian cancer susceptibility. *Carcinogenesis* **27** 1502–1506. (doi:10.1093/carcin/bgl014)

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