



## Short communication

# Synthesis and biological evaluation of fluorinated 1,5-diarylpyrrole-3-alkoxyethyl ether derivatives as selective COX-2 inhibitors endowed with anti-inflammatory activity



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## ABSTRACT

A series of substituted 1,5-diarylpyrrole-3-alkoxyethyl ethers were previously synthesized and the potential anti-inflammatory and antinociceptive activities of these compounds were evaluated *in vivo*. The compounds displayed a very good activity against both carrageenan-induced hyperalgesia and oedema in the rat paw test. Therefore, in a very preliminary test, compounds (**8a,b**) showed antiproliferative activity in the HaCaT (aneuploid immortal keratinocyte from adult human skin) cell models. On these basis, our research continued with the synthesis of fluorinated derivatives (**8c,d**, **9b-d**, and **10b-d**) with the aim of improving the pharmacokinetic profile of the previous active compounds. Substitution of a hydrogen atom by a fluorine atom may change the conformational preferences of the molecules due to stereo-electronic effects and also fluorine atom may be able to exert the metabolic obstruction reducing the "first-pass effect". Compound **10b** exhibited a prominent *in vivo* anti-inflammatory and antinociceptive activities, in addition its antiproliferative power in an *in vitro* model of human skin cancer is herein described.

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## 1. Introduction

Prostaglandin G/H synthase enzymes (PGHS), commonly known as cyclooxygenases (COX), are responsible for the formation of important biological mediators called prostanoids, which play a critical role in various biological processes [1]. Arachidonic acid (AA) is a 20 carbon unsaturated fatty acid distributed throughout the lipid bilayer of the cell membranes. Phospholipase A2 (PLA<sub>2</sub>)

enzymes cleave membrane bound arachidonate for the conversion into bioactive precursors. AA can be metabolized through COX pathway via a two-step process: the first step involves conversion of AA to prostaglandin G<sub>2</sub> (PGG<sub>2</sub>), a 9,11-endoperoxide-15-hydroperoxide, and in the second step peroxidase reduces PGG<sub>2</sub> to prostaglandin H<sub>2</sub> (PGH<sub>2</sub>). Specific PG synthases further metabolize PGH<sub>2</sub> to give structurally related bioactive lipids, including PGs PGE<sub>2</sub>, PGD<sub>2</sub>, PGF<sub>2α</sub>, prostacycline PGI<sub>2</sub> and thromboxane A<sub>2</sub> (TxA<sub>2</sub>) [2].

Non-Steroidal Anti-Inflammatory Drugs (NSAID), which include both traditional NSAIDs (*t*NSAIDs) and selective inhibitors of COX-2 (COXib) (Fig. 1), relieve pain and inflammation suppressing the COX function of PGHS and the consequent formation of PGE<sub>2</sub> and

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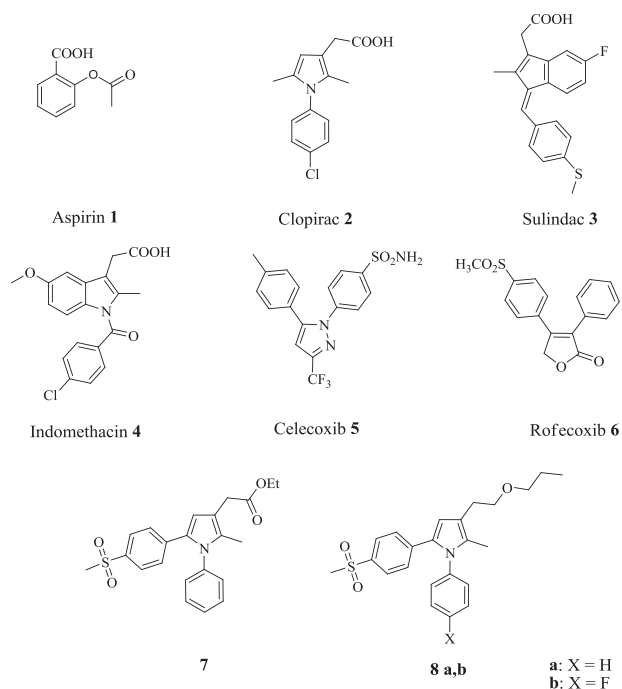


Fig. 1. Structures of reference compounds.

prostacyclin (PGI<sub>2</sub>), along with other prostanoids [1].

Long-term therapy with tNSAIDs generates adverse gastrointestinal complications ranging from stomach erosions and silent intestinal ulcerations to life-threatening complications [3]. Selective inhibitors of COX-2 depress prostacyclin (PGI<sub>2</sub>), an atheroprotective agent, but not COX-1-derived thromboxane A<sub>2</sub> (TXA<sub>2</sub>), a proaggregatory and vasoconstrictor mediator, which might predispose patients to heart attack and stroke [4]. In 2004, Rofecoxib, (4-[4-(methylsulfonyl)phenyl]-3-phenyl-2(5H)-furanone), marketed as Vioxx was withdrawn from the market due to increased risk for cardiovascular thrombotic events in a long term-therapy [5].

Moreover, the involvement of COX-2 in cancer development was previously evidenced by pharmacological analysis of PGs in different human cancers. PGE<sub>2</sub> specifically exerts carcinogenic effects in the human body. It was found that premalignant lesions and established cancers produce excessive quantities of PGE<sub>2</sub> and this enhances tumor cell growth and increases tumor invasiveness [1]. COX-2 is up-regulated in a number of epithelial cancers, including those originating in the colon and rectum, stomach, breast, prostate, and lung [6]. In the context of cutaneous tissues, increased expression of COX-2 and production of its primary product PGE<sub>2</sub> have been reported to increase cell growth and decrease apoptosis. PGE<sub>2</sub> is also thought to be responsible for different skin cancers, such as squamous cell and basal cell carcinoma [7].

During the last decade a large number of COX inhibitors have been described. In particular, very recently novel chemical entities have been identified by different approaches such as: i) rational design providing novel scaffolds based on 4-phenylpyrimidine-2(1H)-thiones [8] and, indole based peptidomimetics [9], or ii) computational procedures providing novel and structurally unrelated COX-2 inhibitors [10].

Within a large programme devoted to the design and synthesis of new anti-inflammatory agents based on a diaryl heterocyclic scaffold, we previously synthesized different series of 1,5-diarylpyrrole-3-acetic esters **7** [11] and 3-alkoxyethyl ethers **8a,b** [12] as new selective COX-2 inhibitors, in which the pyrroloacetic

subunit, reminiscent of indomethacin and clopirac was conjugated with vicinal diaryl moiety mimicking the main feature of COXib (Fig. 1).

These compounds were shown to be potent and highly selective COX-2 inhibitors in *in vitro* cell culture assays [12–14]. The potential anti-inflammatory and antinociceptive activities of these compounds were also evaluated *in vivo*, demonstrating a very good activity against both carrageenan induced hyperalgesia and oedema in rat paw test. Furthermore, compounds **8a,b** showed, in a very preliminary screening, a clear inhibition of cell proliferation on HaCaT cells with non-toxic effects in the range between 10 and 100 μM [7].

Encouraged by these data, we focused our attention on the preparation of two homologous series of differently fluorinated 3-substituted-1,5-diarylpyrrole alkoxyethyl ethers (**8c,d**; **9b-d** and **10b-d**) (Fig. 2) with the aim of investigating the biological effects that can be elicited by the insertion of a fluorine atom on different portion of the previously synthesized COX-2 inhibitors. Compound **10b**, which displays the best biological profile in terms of affinity and selectivity toward COX-2, as well as percent inhibition of the enzyme (cell culture assay), was also investigated for its *in vivo* anti-inflammatory, analgesic and antiproliferative activity, displaying interesting results.

## 2. Results and discussion

### 2.1. Chemistry

The synthetic procedure used to obtain final compounds **8c,d**, **9b-d** and **10b-d** is depicted in Scheme 1. Briefly, the suitable alcohols **11c-d** were treated with propyl iodide in NaOH 50% (w/w) solution, following the previously reported procedure [12] to obtain final compounds **8c,d**. Using the same procedure, derivatives **11b-d** were treated with the proper hydroxyalkyl bromide in its tetrahydropyranyl protected form [15] (see Supplementary Material) to yield compounds **12b-d** and **13b-d** that were successively deprotected to give hydroxyalkyl ethers **14b-d** and **15b-d**. These derivatives were then activated by tosylation and treated with Bu<sub>4</sub>N<sup>+</sup>F<sup>-</sup> to afford fluorinated compounds **9b-d** and **10b-d** in satisfactory yield. Purity of compounds **8c,d**, **9b-d**, **10b-d** was assessed by RP-HPLC and was found to be higher than 95% (see Supplementary Material for further details).

### 2.2. Structure-activity relationship, molecular modeling studies and biological evaluation

In this limited series of compounds the presence of an additional fluorine atom in the 3-alkoxyalkyl side-chain with respect to compounds **8a,b** would allow us to assess if the contemporary presence of fluorine atoms in two different portions of the molecule could influence the interaction of these new inhibitors with the COX-2 active site. In particular, compounds **8c,d** were synthesized

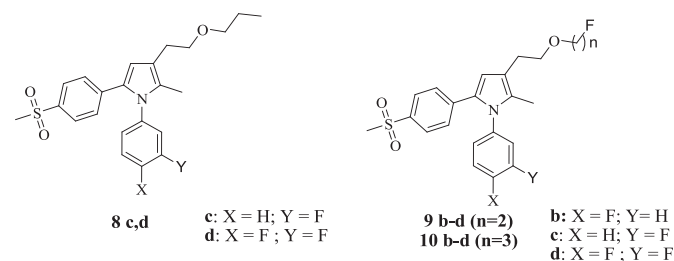
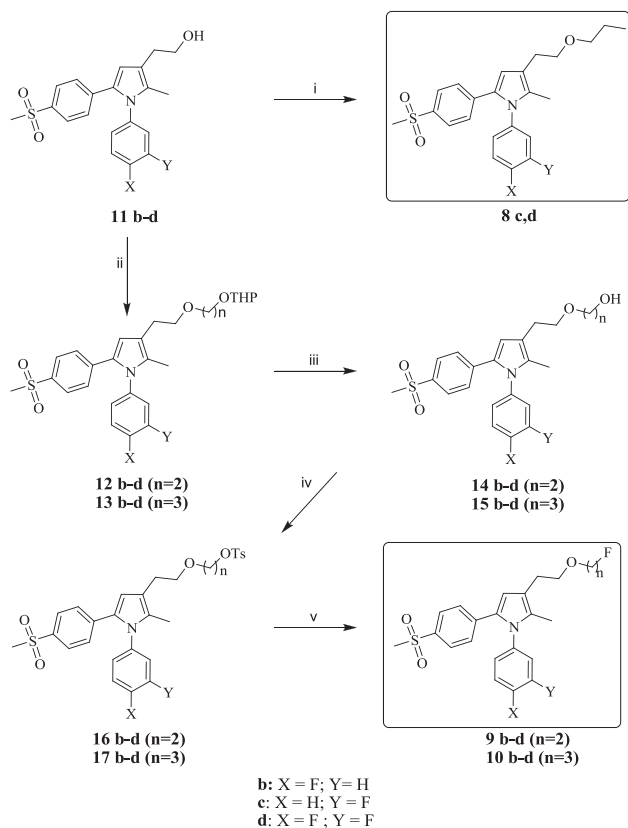


Fig. 2. Structure of fluorinated compounds **8c,d**, **9b-d** and **10b-d**.



**Scheme 1.** Synthesis of compounds **8c,d**, **9b-d**, and **10b-d**. Reagents and conditions: (i) Propyl iodide,  $(\text{Bu}_4\text{N}^+)\text{Br}^-$ , Sodium hydroxide, 50% (w/w) aqueous solution, 60 °C, 72 h; (ii)  $\text{Br}(\text{CH}_2)_n\text{OTHP}$ ,  $(\text{Bu}_4\text{N}^+)\text{Br}^-$ , Sodium hydroxide, 50% (w/w) aqueous solution, 60 °C, 72 h (iii) PPTS, MeOH, 55 °C, 20 h; (iv) DMAP, DIPEA, TsCl, DCM, r.t., 4 h; (v)  $(\text{Bu}_4\text{N}^+)\text{F}^-$  in THF, 1 M; dry THF, 80 °C, 3 h.

in order to evaluate if the number and the different position of fluoro substituent in the pendant N1 aromatic ring could lead to biological results comparable with those previously reported for compounds **8a** and **8b** [12].

The effects of a fluorine atom as a substituent on physical and chemical properties, as lipophilicity, electronic properties, hydrogen bonding, and metabolic stability are well known [16,17]. The difference in electronegativity between carbon and fluorine generates a large dipole moment in this bond that, when combined with the electrostatic distribution of a specific molecule, may contribute to the molecule's ability to establish intermolecular interactions.

By means of *in vitro* cell culture (J774 murine macrophage) assay, the title compounds were evaluated for their inhibitory potency and selectivity on both COX-isofoms and the results are resumed in Table 1.

Results showed that in compounds **8c,d**, the presence of a fluoro substituent in *meta* and *in meta/para* position of the pendant N1 ring did not alter the inhibitory potency and selectivity with respect to previously reported analogues **8a,b**. In compounds sharing general structures **9** and **10**, the replacement of a hydrogen atom with fluorine in the side chain coupled with the presence of mono- or difluoro-substitution on the N1 pendant ring led to contrasting effects on the inhibitory activity of the resulting compounds. Accordingly, for *meta*-substituted compounds **9c** and **10c**, a considerable decrease of COX-2 inhibition was observed whereas the *para*-fluoro derivative **10b**, showed a significantly improved inhibitory activity ( $\text{IC}_{50} = 0.007 \mu\text{M}$ ) when compared with that of

1,5-diarylpyrrole-3-propoxyethyl-, 3-fluoroethoxyethyl-, and 3-fluoropropoxyethyl ethers **8a-d**, **9c,d**, and **10c,d**, respectively, the  $\text{IC}_{50}$  of which ranges from 0.018 to 0.030  $\mu\text{M}$ . The *in vitro* potency and selectivity showed by compound **10b** appears more striking when compared to that of celecoxib (**5**) the  $\text{IC}_{50}$  of which being 0.079  $\mu\text{M}$  (11-fold less potent and much more less selective than **10b**).

Next, the new fluorinated derivatives were analysed by means of molecular docking calculations using Glide software (Glide version 5.7, Schrödinger, LLC, New York, NY, 2011), with the aim of investigating the potential binding modes of such inhibitors within the COX-2 binding site. The selected compounds **9b** and **10b** differing for one carbon atom in the fluorinated tail, showed a similar binding mode to those found for a previously reported COX-2 inhibitors [18]. In particular, compound **9b** strongly interacts with the COX-2 active site by a series of polar contacts with the backbone of Phe518 and with the side-chain of Arg120. Moreover, one  $\pi-\pi$  stacking was established between Tyr355 and **9b** (Fig. 3). The fluorophenyl moiety interacts with a series of hydrophobic residues as depicted in Fig. 3B. These interactions of **9b** into COX-2 binding site result in docking XPscore of  $-8.47 \text{ kcal/mol}$  and in estimated free binding energy ( $\Delta G_{\text{bind}}$ ) of  $-97.74 \text{ kcal/mol}$ .

As depicted in Fig. 4, compound **10b** showed a similar binding mode with respect to that found for **9b**. In fact, **10b** established the same hydrophobic/polar contacts previously described. Also, the docking score and the estimated free binding energy resulted comparable to those calculated for **9b** (XPscore  $-9.27 \text{ kcal/mol}$ ;  $\Delta G_{\text{bind}} -100.97 \text{ kcal/mol}$ ). Remarkably, all the newly synthesized compounds reported in Table 1 showed a comparable binding mode as well as the docking score and the estimated  $\Delta G_{\text{bind}}$  (data not shown), confirming the strong affinity of this kind of scaffold for COX-2 enzyme.

On the basis of its very encouraging *in vitro* COX-2 inhibitory activity, compound **10b** was evaluated in a rat model of acute inflammatory pain. As shown in Table 2, inflammatory pain was induced by the intraplantar administration of carrageenan.

The anti-inflammatory effect of the new compounds emerged also by the measurement of the paw edema volume, it was significantly decreased by **10b** ( $3-20 \text{ mg kg}^{-1}$ ) (Table 2). To evaluate the pain reliever profile of the active compound **10b**, its antinociceptive efficacy and potency was assessed in the model of visceral pain induced by the intraperitoneal (i.p.) injection of a 0.6% acetic acid solution (Writhing test). The abdominal contractions induced by the acid solution (about 30 writhing in vehicle-treated animals, Table 3), were dose-dependently prevented by the administration p.o. of **10b**. The measurement performed 30 min after 20  $\text{mg kg}^{-1}$  **10b** treatment showed a number of writhing reduced to  $10.7 \pm 2.5$  (Table 3).

The efficacy of compound **10b** was also evaluated in the rat osteoarthritis model induced by the intra-articular injection of monoiodoacetate (MIA) (Table 4). The injection of MIA in the knee joint induces necrosis of chondrocytes with decrease of cartilage thickness and osteolysis [19]. Kobayashi et al. showed that MIA is able to disorganize chondrocytes and to promote cartilage erosion [20]. These alterations are comparable with joint damages typical of humans affected by osteoarthritis [21–23]. Briefly, on day 14 after MIA injection the pain threshold on the ipsilateral paw was decreased to  $39.2 \pm 3.2 \text{ g}$  in comparison to  $62.9 \pm 2.3 \text{ g}$  of control rats as evaluated by Paw pressure test. Compound administered p.o. was able to reduce articular pain starting from the dosage of  $3 \text{ mg kg}^{-1}$ . Twenty  $\text{mg kg}^{-1}$  of **10b** were effective up to 60 min after treatment reaching the threshold of  $54.5 \pm 3.5 \text{ g}$  (60 min, Table 4). The repeated treatment with **10b** significantly prevented MIA dependent hyperalgesia (before treatment). A further compound administration (after treatment, 30 min) did not induce an additive

**Table 1**  
*In vitro* inhibition (J774 murine macrophage assay) of COX-1 and COX-2 by compounds **8a-d**, **9b-d**, and **10b-d**.

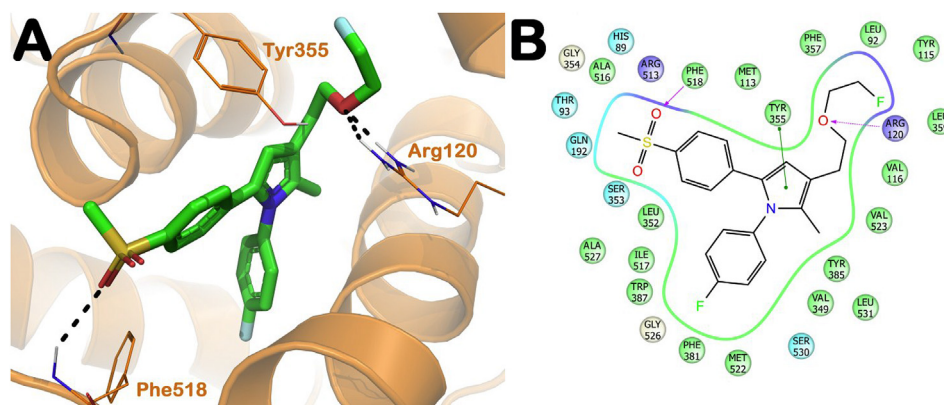
Cpd	X	Y	n	COX-1 IC <sub>50</sub> (μM) <sup>a</sup>	COX-2 IC <sub>50</sub> (μM) <sup>a</sup>	COX-1/COX-2 (SI) <sup>b</sup>
<b>8a<sup>c</sup></b>	H	F	—	>100	0.018	>5555
<b>8b<sup>c</sup></b>	F	F	—	>100	0.030	>3333
<b>8c</b>	H	F	—	>100	0.013	>7692
<b>8d</b>	F	F	—	>100	0.018	>5555
<b>9b</b>	F	H	2	>100	0.013	>7692
<b>9c</b>	H	F	2	>100	0.140	>714
<b>9d</b>	F	F	2	>100	0.029	>3448
<b>10b</b>	F	H	3	>100	0.007	>14,285
<b>10c</b>	H	F	3	>100	0.135	>2857
<b>10d</b>	F	F	3	>100	0.019	>5263
Celecoxib <sup>d</sup>				5.1	0.079	65

<sup>a</sup> Results are expressed as the mean ( $n = 3$ ) of the % inhibition of PGE<sub>2</sub> production by the test compounds with respect to control sample and IC<sub>50</sub> values were calculated by GraphPad Instat program.

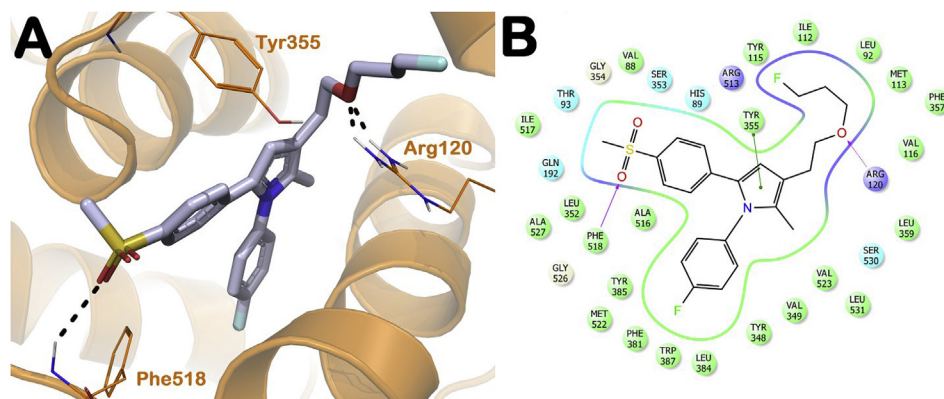
<sup>b</sup> IC<sub>50</sub> (COX-1)/IC<sub>50</sub> (COX-2).

<sup>c</sup> See Ref. [12].

<sup>d</sup> See Ref. [13].



**Fig. 3.** (A) Best docked pose for compound **9b** (green sticks) into COX-2 (orange cartoon PDB ID: 6COX) binding site. The key residues in the active site were represented by lines. H-bonds were represented by black dotted lines. The hydrogens were omitted for the sake of clarity. The picture was generated by means of PyMOL (The PyMOL Molecular Graphics System, v1.6-alpha; Schrodinger, LLC, New York, 2013). (B) Ligand interaction diagram (Maestro version 9.3, Schrödinger, LLC, New York, NY, 2012). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)



**Fig. 4.** (A) Best docked pose for compound **10b** (grey sticks) into COX-2 (orange cartoon PDB ID: 6COX) binding site. The key residues in the active site were represented by lines. H-bonds were represented by black dotted lines. The hydrogens were omitted for the sake of clarity. (B) Ligand interaction diagram. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

acute antihyperalgesic effect, suggesting that a repeated treatment with the present anti-inflammatory compounds could be able to prevent the joint damage and, consequently, pain [21–24].

In addition to the canonical anti-inflammatory and antinociceptive activities, many studies highlighted the potential of COX-2 inhibitors in the prevention of several cancer types such as

colon, breast, lung and prostate cancers [25–28] most likely by modulating the cell proliferation pathways. So, title compounds were evaluated to assess their effects on cell toxicity by means of LDH release assay and potential antiproliferative activity using HaCaT, aneuploid immortal keratinocyte cell line from adult human skin [7].

**Table 2**  
Effect of **10b** on carrageenan-induced acute inflammatory pain hypersensitivity (paw-pressure test) and paw edema.

Paw-pressure (g)			Paw volume (mL)					
Pre-treatment intraplantar	Treatment per os	Dose mg kg <sup>-1</sup>	Before treatment	After treatment				
				30 min	60 min	90 min	120 min	60 min
Vehicle	Vehicle		57.6 ± 2.2	63.8 ± 2.9	59.4 ± 3.3	62.7 ± 2.8	64.6 ± 3.2	1.45 ± 0.08
Carrageenan	Vehicle		33.9 ± 3.1	34.1 ± 2.3	35.4 ± 3.6	37.2 ± 3.4	34.5 ± 4.4	2.73 ± 0.07
Carrageenan	<b>10b</b>	3	30.5 ± 2.6	45.2 ± 4.6**	49.2 ± 3.7**	44.8 ± 3.0	36.2 ± 3.3	2.38 ± 0.06*
Carrageenan	<b>10b</b>	10	31.8 ± 2.7	51.7 ± 4.1**	53.5 ± 3.9**	47.2 ± 4.4**	38.1 ± 2.9	2.15 ± 0.08**
Carrageenan	<b>10b</b>	20	30.5 ± 2.0	53.9 ± 3.7**	56.3 ± 4.6**	44.9 ± 3.7**	35.5 ± 2.6	2.06 ± 0.05**

Carrageenan (Car, 1%, 100 µL) was injected in the left posterior sole 2 h before pain evaluation by the paw pressure test. Molecule or vehicle (1% carboxymethylcellulose) was administered intraplantarly at time 0 and measures were performed at time 30, 60, 90 and 120 min. The volume of the paw edema was measured at time 60 min. Values are the mean ± s.e.m. of 6 animals. \*P < 0.05 and \*\*P < 0.01 vs the carrageenan + vehicle group.

**Table 3**  
Effect of **10b** in the mouse abdominal contraction test (acetic acid 0.6%).

Treatment per os	Dose mg kg <sup>-1</sup>	Number of writhes 30 min	Number of writhes 60 min
Vehicle		30.5 ± 1.7	32.5 ± 2.4
<b>10b</b>	3	21.8 ± 2.5**	24.6 ± 2.7*
	10	11.4 ± 3.1**	17.7 ± 3.1**
	20	10.7 ± 2.5**	16.9 ± 2.7**

**10b** was administered 30 or 60 min before the experiment. 0.6% acetic acid was administered intraperitoneally 5 min before the observation. Each value represents the mean of 12 mice per group. \*P < 0.05 and \*\*P < 0.01 versus vehicle-treated mice.

**Table 4**  
Effect of **10b** on monoiodoacetate-induced osteoarthritis hypersensitivity in the rat paw-pressure test.

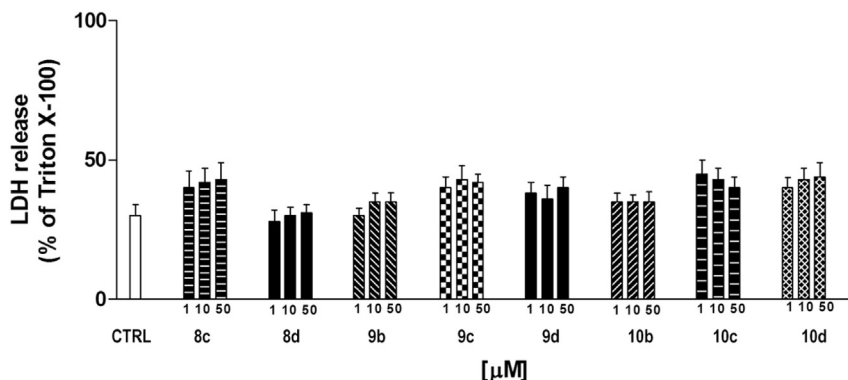
Pre-treatment intraplantar	Treatment per os	Dose mg kg <sup>-1</sup>	Before treatment	Paw-pressure (g)			
				After treatment			
				30 min	60 min	90 min	120 min
Vehicle	Vehicle		62.9 ± 2.3	60.5 ± 2.4	58.3 ± 4.0	66.3 ± 4.1	61.5 ± 3.2
MIA	Vehicle		39.2 ± 3.2	37.8 ± 3.3	36.1 ± 3.5	37.8 ± 3.1	39.2 ± 4.0
MIA	<b>10b</b>	3	36.5 ± 2.5	47.9 ± 4.2**	45.4 ± 3.8*	42.4 ± 3.5	39.0 ± 3.1
		10	38.8 ± 2.8	50.3 ± 4.4**	51.8 ± 3.7**	44.3 ± 3.8	40.6 ± 3.5
		20	37.8 ± 2.2	51.3 ± 3.3**	54.5 ± 3.5**	43.8 ± 3.1	37.1 ± 2.9

Effect of **10b** on monoiodoacetate (MIA)-induced unilateral osteoarthritis. MIA (2 mg/25 µL) was injected in the left posterior tibiotarsal articulation 14 days before pain evaluation by the paw pressure test. Molecules or vehicle were administered intraperitoneally at time 0 and measures were performed at time 30, 60, 90 and 120 min. Values are the mean ± s.e.m. of 6 rats. \*P < 0.05 and \*\*P < 0.01 in comparison with MIA + vehicle treated rats.

### 2.3. Effect of compounds **8c,d**, **9b-d**, and **10b-d** on cell toxicity and proliferation

Cytotoxicity of compounds was evaluated in HaCaT cells after treatment with doses ranging from 1 to 50 µM by means of LDH release. As shown in Fig. 5, 24 h after treatment no cytotoxicity was detected.

As shown in Fig. 6, the treatment with compounds **8c,d**, **9b-d**, and **10b-d** for 24 h, had a clear dose-dependent effect on cellular proliferation as measured by 5-bromo-2'-deoxyuridine (BrdU) incorporation. Indeed, while 1 µM did modify cellular proliferation, the treatment with higher albeit not toxic doses (10 and 50 µM) evidenced a drastic and significant cellular growth inhibition for **8d**, **9b**, **9c**, **9d**, **10b** and **10d** compared to the control cells (DMSO).

**Fig. 5.** Cytotoxicity of compounds **8c,d**, **9b-d**, and **10b-d** ranging from 1 to 50 µM after 24 h was evaluated in HaCaT cell line by LDH release assay. Data are expressed as percentage of Triton X-100 (100%) (averages of three different experiments, \*p < 0.05).

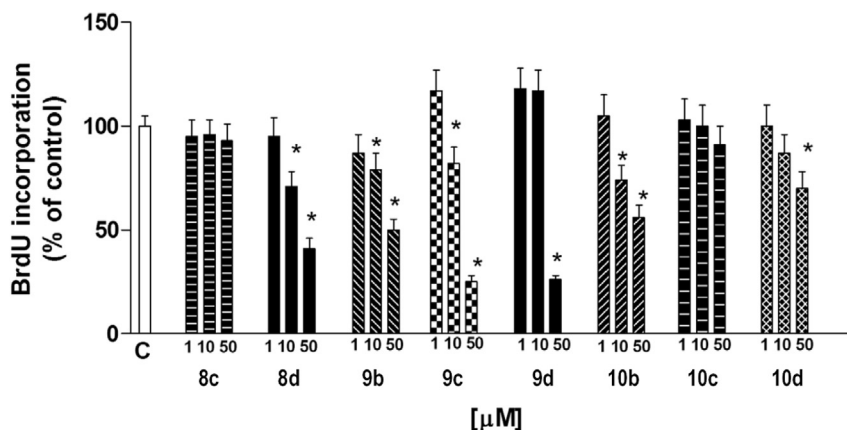


Fig. 6. The effect on cell proliferation of compounds **8c,d**, **9b-d**, and **10b-d** on HaCaT cells was evaluated by means of BrdU incorporation after 24 h of treatment. Data are expressed as percentage of control (averages of three different experiments, \* $p < 0.05$ ).

0.1%). Cells treated with **8c** and **10c** did not show any significant changes in BrdU incorporation. However, further investigation is required to confirm the antiproliferative activity showed by previously reported compounds **8a,b** [7].

### 3. Conclusions

In this work, we have designed, synthesized and tested a new series of fluorinated 1,5-diarylpyrrole-3-alkoxyethyl ether derivatives as COX-2 selective inhibitors. *In vitro* biological results are in full agreement with all the previously published data, demonstrating the high affinity of the 1,5-diarylpyrrole scaffold for COX-2 active site. The affinity was also confirmed by molecular docking calculations, proving that replacement of a hydrogen atom by fluorine in the alkyl side chain didn't modify steric and electronic properties of the newly conceived chemical entities. Compound **10b** demonstrated efficacy in the model of rat acute inflammatory pain and in the rat osteoarthritis model; thus, highlighting its favourable potential to turn into a novel anti-inflammatory and antinociceptive agents. Furthermore, the anti-proliferative but not toxic effect of the most promising compounds in the presented series on HaCaT cell suggests a possible application in the therapy of human skin disorders.

### 4. Methods

#### 4.1. Biological evaluation

##### 4.1.1. *In vitro* inhibition of COX-1 and COX-2 enzymes

The *in vitro* COX-1 and COX-2 inhibition of compounds **8a-d**, **9b-d**, and **10b-d** was evaluated through cell-based assay employing murine monocyte/macrophage J774 cell lines. The cell line was grown in DMEM supplemented with 2 mM glutamine, 25 mM HEPES, 100 units/mL penicillin, 100 μg/mL streptomycin, 10% fetal bovine serum (FBS), and 1.2% sodium pyruvate. Cells were plated in 24-well culture plates at a density of  $2.5 \times 10^5$  cells/mL and allowed to adhere at 37 °C in 5% CO<sub>2</sub> for 2 h. Immediately before the experiments, culture medium was replaced with fresh medium and cells were stimulated as described previously [12]. The evaluation of COX-1 inhibitory activity was achieved pre-treating cells with test compounds (100 μM) for 15 min and then incubating them at 37 °C for 30 min with 15 μM arachidonic acid to activate the constitutive COX. At the end of the incubation, the supernatants were collected for the measurement of PGE<sub>2</sub> levels by a radioimmunoassay (RIA). To evaluate COX-2 activity, cells were stimulated for 24 h with *Escherichia coli* lipopolysaccharide (LPS, 10 μg/mL) to

induce COX-2, in the absence or presence of test compounds (0.001–10 μM). The supernatants were collected for the measurement of PGE<sub>2</sub> by means of RIA. Celecoxib was utilized as a reference compound for the selectivity index. Triplicate wells were used for the various conditions of the treatment in the cell culture assay throughout the experiments. Results are expressed as the mean of three experiments, of the % inhibition of PGE<sub>2</sub> production by test compounds with respect to control samples. Data fit was obtained using the sigmoidal dose-response equation (variable slope) (GraphPad software). The IC<sub>50</sub> values were calculated by the GraphPad Instat program (GraphPad software).

#### 4.1.2. Animals

For all the experiments described below, male CD1 albino mice (23–25 g) or male Sprague–Dawley rats (Harlan, Varese, Italy) weighing approximately 200–250 g at the beginning of the experimental procedure, were used. Animals were housed in CeSAL (Centro Stabulazione Animali da Laboratorio, University of Florence) and used at least one week after their arrival. Four rats were housed *per cage* (size 26 × 41 cm); animals were fed a standard laboratory diet and tap water *ad libitum*, and kept at 23 ± 1 °C with a 12 h light/dark cycle, light at 7 a.m. All animal manipulations were carried out according to the European Community guidelines for animal care (DL 116/92, application of the European Communities Council Directive of 24 November 1986 (86/609/EEC). The ethical policy of the University of Florence complies with the Guide for the Care and Use of Laboratory Animals of the US National Institutes of Health (NIH Publication No. 85-23, revised 1996; University of Florence assurance number: A5278-01). Formal approval to conduct the experiments described was obtained from the Italian Ministry of Health (N°54/2014-B) and from the Animal Subjects Review Board of the University of Florence. Experiments involving animals have been reported according to ARRIVE guideline. All efforts were made to minimize animal suffering and to reduce the number of animals used.

#### 4.1.3. Carrageenan-induced acute inflammatory pain

One hundred μL of carrageenan solution (Sigma–Aldrich; 1% in saline) was injected intraplantarly into the rat left hind paw. Two h after carrageenan injection, compounds were *per os* (p.o.) administered and their anti-hypersensitive effect was measured over time by the paw pressure test. Control rats received 100 μL of vehicle (saline) solution intraplantarly and saline *i.p.*

#### 4.1.4. Abdominal contraction pain

At predetermined time intervals (30 and 60) after drug

administration, mice were injected intraperitoneally with a 0.6% (w/w) solution of acetic acid ( $10 \text{ mL kg}^{-1}$ ), to induce pain sensation evident from typical abdominal contractions of animals named 'writhing'. The number of induced writhings was counted for 10 min, starting 5 min after acetic acid injection [29].

#### 4.1.5. Monoiodoacetate-induced osteoarthritis

Unilateral osteoarthritis was also induced by injection of monoiodoacetate (MIA, Sigma–Aldrich) into the rat tibio-tarsal joint [30]. On day 0, rats were slightly anesthetized by 2% isoflurane, the left leg skin was sterilized with 75% ethyl alcohol and the lateral malleolus located by palpation; then, a 28-gauge needle was inserted vertically to penetrate the skin and turned distally for insertion into the articular cavity at the gap between the tibio-fibular and tarsal bone until a distinct loss of resistance was felt. 2 mg MIA in 25  $\mu\text{L}$  saline were delivered into the left articular cavity. The paw pressure was performed at day 14. The tested compounds were p.o. administered. Control rats received 25  $\mu\text{L}$  of saline solution (day 0) in the tibiotarsal joint and saline i.p. at day 14.

#### 4.1.6. Paw pressure test

The pain threshold in the rat was determined with an analgesimeter (Ugo Basile, Varese, Italy) as described [31]. Briefly, a constantly increasing pressure was applied to a small area of the dorsal surface of the paw using a blunt conical probe. Pressure was increased until a vocalization or a withdrawal reflex occurred. The withdrawal threshold was expressed in grams, the test was repeated twice and the mean was considered as the value for each paw.

#### 4.1.7. Edema volume measurement

Rat paw volumes were measured using a plethysmometer. Sixty min after the injection of carrageenan, the paw volume of the ipsilateral hind paw was measured and compared with carrageenan + vehicle-treated controls. The results are reported as paw volume expressed in mL.

#### 4.1.8. Statistical analysis of behavioral measurements

Results were expressed as means  $\pm$  s.e.m. and the analysis of variance was performed by one way ANOVA. A Bonferroni's significant difference procedure was used as post-hoc comparison. P values of less than 0.05 or 0.01 were considered significant. Data were analyzed by means of the "Origin 9" software (OriginLab, Northampton, USA).

#### 4.1.9. Cell culture and treatments

HaCaT cells, (a cell line gift from Dr. F. Virgili), were grown in Dulbecco's modified Eagle's medium High Glucose (Lonza, Milan, Italy), supplemented with 10% FBS, 100 U/ml penicillin, 100 Iu/ml streptomycin and 2 mM L-glutamine as previously described [32]. Cell suspension containing 10 or 1105 viable cells/ml were used. Cells were incubated at 37 C for 24 h in 95% air/5%  $\text{CO}_2$  until 80% confluency. HaCaT cells were treated with different doses of title compounds at different time points (30 min, 1, 3 and 24 h). After treatment, cells or medium were collected for the several assays described below. Compounds were dissolved in DMSO at a concentration of 10 mM as a stock solution. The stock was diluted to the required concentrations directly in the medium. The final concentration of DMSO in culture medium during compounds treatment did not exceed 0.1% (v/v).

## 4.2. Computational details

All calculations performed in this work were carried out on Cooler Master Centurion 5 (Intel Core-i5 Quad CPU Q6600 @

2.40 GHz) with Ubuntu 10.04 LTS (long-term support) operating system running Maestro 9.2 and Maestro 9.3 (Schrödinger, LLC, New York, NY, 2011 and Maestro version 9.3, Schrödinger, LLC, New York, NY, 2012, respectively).

### 4.2.1. Molecular docking studies

**4.2.1.1. Molecules preparation.** Three-dimensional structures of compounds were built by means of Maestro (Maestro version 9.2, Schrödinger, LLC, New York, NY, 2011). Molecular energy minimizations were performed by means of MacroModel using the Optimized Potentials for Liquid Simulations-all atom (OPLS-AA) force field 2005 [33,34]. The solvent effects are simulated using the analytical Generalized-Born/Surface-Area (GB/SA) model [35], and no cutoff for nonbonded interactions was selected. Polak-Ribiere conjugate gradient (PRCG) method with 1000 maximum iterations and 0.001 gradient convergence threshold was employed. The compounds were treated by LigPrep application (LigPrep version 9.2, Schrödinger, LLC, New York, NY, 2011), implemented in Maestro suite 2011, generating the most probable ionization state of any possible enantiomers and tautomers at cellular pH value ( $7 \pm 0.5$ ).

**4.2.1.2. Protein preparation.** The three-dimensional structure of COX-2 was taken from PDB (PDB ID: 6COX) and imported into Schrödinger Maestro molecular modeling environment. The structure was submitted to protein preparation wizard implemented in Maestro suite 2011 (Protein Preparation Wizard workflow 2011) as previously reported [36,37]. This protocol through a series of computational steps, allowed us to obtain a reasonable starting structure of the proteins for molecular docking calculations by a series of computational steps. In particular, we performed diverse steps to (1) add missing residues or side-chains, (2) add hydrogens, (3) optimize the orientation of hydroxyl groups, Asn, and Gln, and the protonation state of His, and (4) perform a constrained refinement with the impref utility, setting the max RMSD of 0.30. The impref utility consists of a cycles of energy minimization based on the impact molecular mechanics engine and on the OPLS\_2005 force field [33,34].

**4.2.1.3. Molecular docking.** Docking studies were carried out by Glide (Grid-Based Ligand Docking with Energetics) using the ligands and the protein prepared as above-mentioned, applying Glide extra precision (XP) method [38,39]. Energy grid was prepared using default value of protein atom scaling factor (1.0 Å) within a cubic box centered on the crystallized inhibitor. After grid generation, the ligands and the crystallized inhibitor were docked into the enzymes with default parameters (no constraints were added). The number of poses entered to post-docking minimization was set to 50. Glide XP score was evaluated. The XP method correctly accommodated the crystallized inhibitors into the binding site (data not shown).

**4.2.1.4. Estimated free binding energy.** The Prime/MM-GBSA method implemented in Prime software (Prime version 3.0, Schrödinger, LLC, New York, NY, 2011) consists in computing the change between the free and the complex state of both the ligand and the protein after energy minimization. The technique was used on the docking complexes of the selected compounds presented in this study. The software was employed to calculate the free-binding energy ( $\Delta G_{\text{bind}}$ ) as previously reported by some of us [40–42].

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## Appendix A. Supplementary data

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.ejmech.2015.12.044>.

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