

Technical Report

Bilateral Project: ACIAR-Establishing sustainable solutions to cassava diseases in mainland South-East Asia.

Plant Health Initiative WP2: Validation of Artificial Intelligence tools for disease recognition at large scale, available via a digital platform supporting surveillance and breeding efforts for resistance evaluations

A protocol for measuring Cassava Bacterial Blight disease progression using machine-learning image analysis

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Abstract

Cassava (*Manihot esculenta* Crantz), a starchy root crop, is the fourth most important source of dietary carbohydrate in developing countries and important source of industrial raw material in tropical regions especially in southeast Asia (SEA). *Xanthomonas phaseoli* pv. *manihotis* (Xpm) formerly known as *Xanthomonas axonopodis* pv. *manihotis* causes cassava bacterial blight (CBB) disease, a major bacterial disease of cassava worldwide. To improve CBB diagnostics and support breeding for resistance programs, we standardized a screen house protocol for the reliable, simple, space- and time-efficient and high throughput tool based on machine learning image analysis using the PhenotyperCV software. In addition, we sequenced several bacterial isolates using Nanopore technology and their pathogenicity profile was evaluated by inoculation on 3-month old plants, under greenhouse conditions. Disease symptoms were measured to calculate the area under the disease progress curve (AUDPC) as described below. This protocol allows us to identify distinct pathogenic Xpm strains and evaluate CBB at different DPI by performing symptoms measurements in an automated way. AUDPC curves are used for pathogenic characterization of additional isolates and subsequent varietal resistance screenings.

Protocol v1.0:

1. Inoculation: Through infiltration, 3 technical replicates per strain onto the leaf lobe.



2. Take photos (0,4,6,9,12 days post inoculation) image device at the same conditions. use X
– Rite Passport color card to gray color correction.



3. Choose clear symptoms to create a collage.

- Select affected area manually (GIMP or IMAGE J)
- Create a mask: Convert picture to black and white. Only affected area white background black.



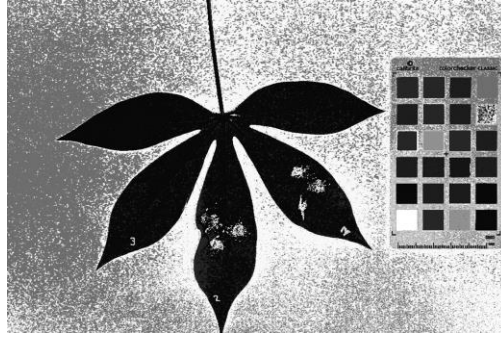
4. Machine training:

- Use software Phenotyper (Berry et al., 2018).
- Train naive Bayes or SVM using ML_CREATE specifying classifier type:

```
PhenotyperCV -m=ML_CREATE -method=bayes -i=collage.png -b=mask.png -class=output_bayes_classifier.yaml
```

Check the classifier prediction using either ML_PRED specifying classifier type

Prediction image: Affected area showed in white:



5. Routine measuring process with Phenotyper:

1. Click only the gray scale color chips from black to white
2. Feature prediction of input image
3. Threshold the prediction to isolate features

Click on the features to measure. If there are different types of features that you'd like to consider separate, currently only three different features are supported and you can specify them by which type of mouse click you use. Left = red, right = green, middle = blue



7. Data analysis: The software Phenotyper generate a .txt file:

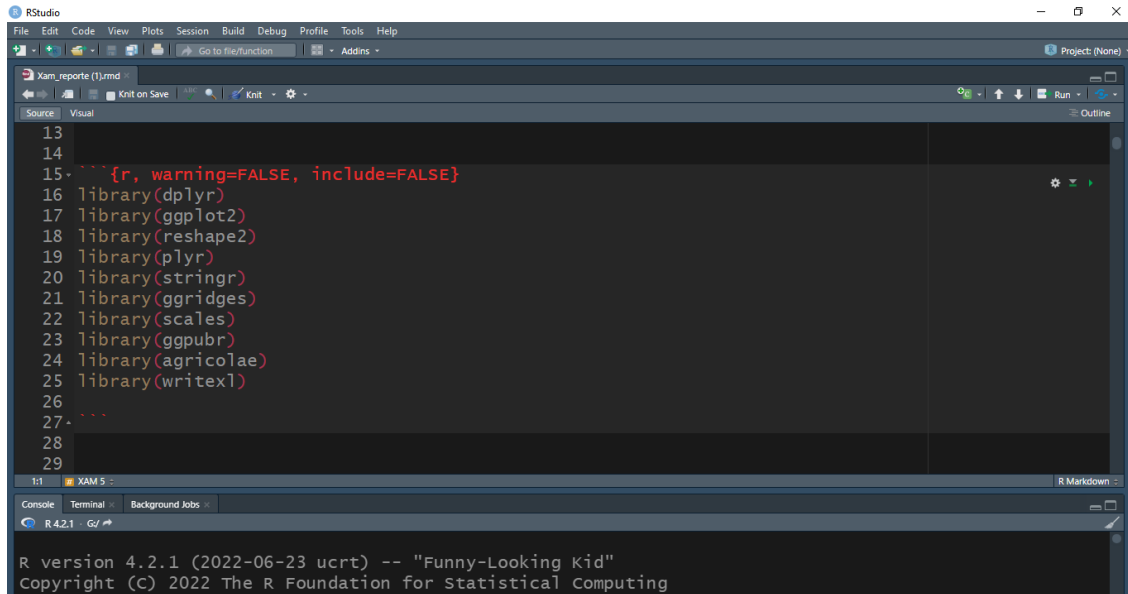
```

file:///C:/Users/.../phenotyper/.../pilesassay5.txt (File build/dspac_phenotyper)
phenotyper.txt x pilesassay5.txt x
/home/.../Assay/014/OM81-2/Img167.jpg blue 0 81 10 15.5 0.89555 12.9515 5 6 673 8 1007 6 674 1007 6 5.74247 3.42862 30.5698 0.897703 0.749148 0.597805 1.67406 1.33333 0
/home/.../Assay/014/OM81-2/Img167.jpg blue 1 81 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img167.jpg blue 2 82 12 6.5 1.84615 7.62843 5 720 583 620 937 5 720 178 820 94 5 22667 2.27914 86.017 1.0628 2.46061 0.435228 2.29765 1.15314 0
/home/.../Assay/014/OM81-2/Img167.jpg green 0 81 504 55.0 0.91192 121.811 29 25 965 754 1065 72 14 965 668 1065 31 27 9984 22.4883 0.627378 0.426841 0.803199 1.24502 1.29689 0
/home/.../Assay/014/OM81-2/Img167.jpg green 1 81 825 862.5 0.95622 146.811 33 36 962 778 1001 72 15 962 664 1001 95 33 3967 29.8357 42.6572 0.461794 0.461001 0.89373 1.1193 1.68188 0
/home/.../Assay/014/OM81-2/Img167.jpg green 2 81 771 798 0.966165 134 983 32 34 961 807 954 899 28 961 353 954 2 32 8342 29 1054 11 9138 0 421741 0 53175 0 911607 1 90761 1 60797 0
/home/.../Assay/014/OM81-2/Img167.jpg red 0 81 54 78 0.89288 52.8746 5 24 1151 11 968 389 6 1150 44 959 211 22 6918 5 09641 178 82 1 24532 0 250237 0 224592 4 45251 1 35898 0
/home/.../Assay/014/OM81-2/Img167.jpg red 1 81 170 339 5 0.22975 93 3206 24 25 1167 69 867 679 12 1166 97 871 373 24 1721 20 5387 170 644 0 548361 0 112537 0 849695 1 17696 1 23398 0
/home/.../Assay/014/OM81-2/Img167.jpg red 2 81 115 373 0 0.307898 149 535 22 25 1074 9 855 522 13 1074 83 857 633 24 4824 18 6853 14 9891 0 688164 0 6646278 0 763215 1 31025 0 950073 0
/home/.../Assay/014/OM81-2/Img168.jpg blue 0 40 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img168.jpg blue 1 40 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img168.jpg green 0 40 179 495 0.361610 181 279 29 26 796 246 663 57 10 793 52 859 761 32 5581 18 4978 141 698 0 92358 0 806449 0 568147 1 76611 1 16478 0
/home/.../Assay/014/OM81-2/Img168.jpg green 1 40 9 4 5.2 2.4264 4 3 720 667 842 889 6 720 604 842 898 3 28837 2 20648 68 6435 0 813682 2 98212 0 668961 1 49486 1 85664 0
/home/.../Assay/014/OM81-2/Img168.jpg green 2 40 8 2 0.6667 5 4 2 783 5 759 5 4 -1 -1 -1 -1 -1 4.62124 -1 -1 1.10992 0
/home/.../Assay/014/OM81-2/Img168.jpg red 0 40 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img168.jpg red 1 40 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img168.jpg red 2 40 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img169.jpg blue 0 41 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img169.jpg blue 1 41 0 3 5 2.8971 0 41421 3 3 540 125 829 125 5 1 -1 -1 -1 -1 -1 2.4435 -1 -1 1.0
/home/.../Assay/014/OM81-2/Img169.jpg blue 2 41 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img169.jpg green 0 41 358 657.5 0.53219 199 973 31 30 729 466 1086 9 13 729 83 1068 75 28 1828 25 8499 23 1751 0 40688 0 189985 0 917224 1 0.9825 1 27756 0
/home/.../Assay/014/OM81-2/Img169.jpg green 1 41 388 636 5 0.60984 187 338 29 30 807 722 983 996 13 807 730 1209 12 6562 93 8304 1 88849 0 158283 0 411937 2 42755 0 950073 0
/home/.../Assay/014/OM81-2/Img169.jpg green 2 41 28 22 1.27273 17 6569 6 7 711 393 959 7 711 426 959 896 7 53372 3 94958 146 046 0 975445 1 1286 0 524254 1 98747 1 88840 0
/home/.../Assay/014/OM81-2/Img170.jpg red 0 41 7 2 5 2 0 5 41421 4 2 1080 71 877 571 4 -1 -1 -1 -1 -1 3.88888 -1 -1 1.85664 0
/home/.../Assay/014/OM81-2/Img170.jpg red 1 41 67 178 5 0 37529 72 8331 29 14 1129 49 818 433 8 1129 33 829 772 31 209 12 6562 93 8304 1 88849 0 158283 0 411937 2 42755 0 950073 0
/home/.../Assay/014/OM81-2/Img170.jpg red 2 41 67 54 5 1 22938 23 4853 18 5 1844 33 740 582 8 1044 42 746 218 16 5675 0 39232 95 4886 1 11319 1 52649 0 388483 2 62879 1 85788 0
/home/.../Assay/014/OM81-2/Img171.jpg blue 0 25 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img171.jpg blue 1 25 7 6 1.6667 13 942 5 7 489 535 4 413 540 11 9845 1 14675 0 8445 0 794551 0 517152 0 684344 1 46125 0 792481 0
/home/.../Assay/014/OM81-2/Img171.jpg blue 2 25 16 11 1.45455 13 6569 4 5 545 562 595 5 545 765 595 4 51642 3 39538 0 0 63884 1 87862 0 796987 1 25618 1 23448 0
/home/.../Assay/014/OM81-2/Img171.jpg green 0 25 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img171.jpg green 1 25 3 6 0 5 25 1868 27 234 40 31 829 187 823 24 18 828 41 822 847 32 757 26 4387 73 1275 0 513478 0 884834 0 88817 1 15185 1 63197 0
/home/.../Assay/014/OM81-2/Img171.jpg green 2 25 253 583 5 0 43550 186 972 21 42 678 162 784 996 13 676 739 794 164 28 3637 24 2753 61 7842 0 53681 0 8989447 0 855910 1 16534 1 123269 0
/home/.../Assay/014/OM81-2/Img171.jpg blue 0 25 26 72 5 0 358821 47 2471 18 11 1072 62 689 154 7 1078 81 698 797 27 6534 21 4494 120 493 0 668951 0 146364 0 77565 1 28924 0 8648508 0
/home/.../Assay/014/OM81-2/Img171.jpg red 1 25 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img171.jpg red 2 25 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img171.jpg blue 0 41 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img171.jpg blue 1 41 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img171.jpg blue 2 41 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img171.jpg red 0 41 67 543 1 22389 99 7613 30 23 750 478 965 91 9 747 588 960 396 40 1606 21 2388 83 387 1 83938 0 8849891 0 459482 2 17466 0 86356 0
/home/.../Assay/014/OM81-2/Img171.jpg red 1 41 287 885 5 0 3563 277 259 39 30 817 899 938 317 13 817 395 938 317 13 2018 25 271 105 938 0 494138 0 8469151 0 75988 1 11737 1 27991 0
/home/.../Assay/014/OM81-2/Img171.jpg red 2 41 518 985 0 5 564667 276 234 40 31 829 187 823 24 18 828 41 822 847 32 757 26 4387 73 1275 0 513478 0 884834 0 88817 1 15185 1 63197 0
/home/.../Assay/014/OM81-2/Img172.jpg blue 0 41 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img172.jpg blue 1 41 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img172.jpg blue 2 41 0 0 -nan 0 0 -nan -nan 0 -1 -1 -1 -1 -1 -1 -1 -nan 0
/home/.../Assay/014/OM81-2/Img172.jpg green 0 41 489 888 5 0 559366 238 887 37 35 824 446 981 041 12 825 487 980 225 34 72 28 2242 116 286 0 611783 0 115351 0 81291 1 23815 1 34782 0
/home/.../Assay/014/OM81-2/Img172.jpg green 1 41 562 648 0 815242 166 826 37 28 891 609 982 546 10 891 391 982 961 11 7388 23 1851 80 8765 0 734173 0 237378 0 738495 1 18893 1 34255 0
/home/.../Assay/014/OM81-2/Img172.jpg green 2 41 170 544 5 0 348136 179 249 33 29 821 637 867 943 11 821 361 865 87 31 3228 22 9316 54 2788 0 738252 0 8684826 0 727938 1 37469 1 8666 0

```

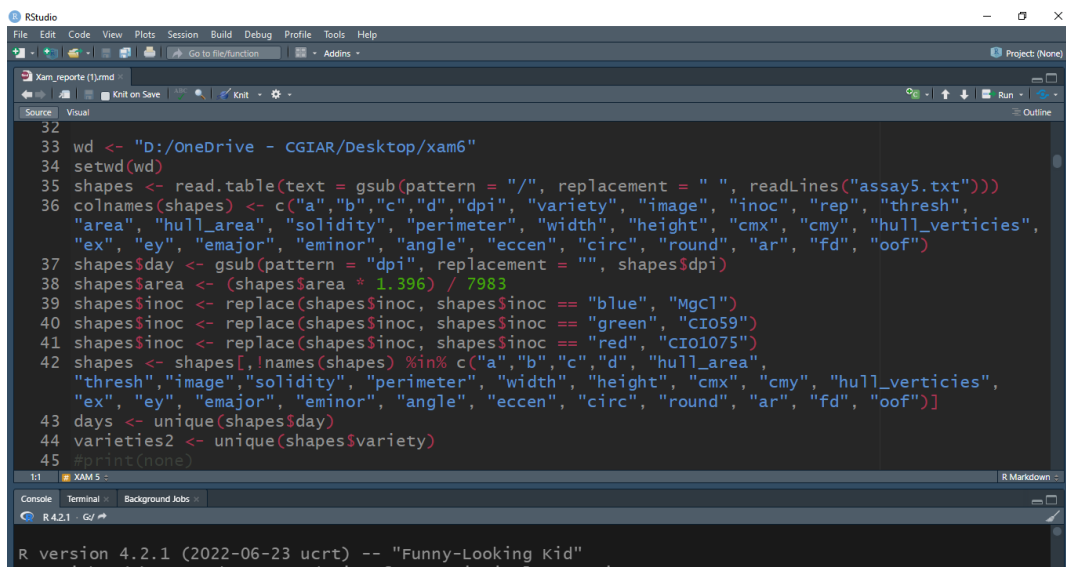
The measuring value in the .txt file is in term of pixels.

8. The automated statistical analysis is performed using R Studio Software and the report is generated with the R Markdown tool.



```
RStudio
File Edit Code View Plots Session Build Debug Profile Tools Help
Xam_reporte (1).rmd
Source Visual
13
14
15- {r, warning=FALSE, include=FALSE}
16 library(dplyr)
17 library(ggplot2)
18 library(reshape2)
19 library(plyr)
20 library(stringr)
21 library(ggribes)
22 library(scales)
23 library(ggpubr)
24 library(agricolae)
25 library(writexl)
26
27-
28
29
R version 4.2.1 (2022-06-23 ucrt) -- "Funny-Looking Kid"
Copyright (C) 2022 The R Foundation for Statistical Computing
```

9. The .txt file generated by Phenotyper should be uploaded in the R Markdown tools. The data could be replaced from pixels to square centimeters; the bacterial strain ID should be assigned using a color code.



```
RStudio
File Edit Code View Plots Session Build Debug Profile Tools Help
Xam_reporte (1).rmd
Source Visual
32
33 wd <- "D:/OneDrive - CGIAR/Desktop/xam6"
34 setwd(wd)
35 shapes <- read.table(text = gsub(pattern = "/", replacement = " ", readLines("assay5.txt")))
36 colnames(shapes) <- c("a", "b", "c", "d", "dpi", "variety", "image", "inoc", "rep", "thresh",
37 "area", "hull_area", "solidity", "perimeter", "width", "height", "cmx", "cmy", "hull_verticies",
38 "ex", "ey", "emajor", "eminor", "angle", "eccen", "circ", "round", "ar", "fd", "oof")
39 shapes$day <- gsub(pattern = "dpi", replacement = "", shapes$dpi)
40 shapes$area <- (shapes$area * 1.396) / 7983
41 shapes$inoc <- replace(shapes$inoc, shapes$inoc == "blue", "MgC1")
42 shapes$inoc <- replace(shapes$inoc, shapes$inoc == "green", "CI059")
43 shapes$inoc <- replace(shapes$inoc, shapes$inoc == "red", "CI01075")
44 shapes <- shapes[,!names(shapes) %in% c("a", "b", "c", "d", "hull_area",
45 "thresh", "image", "solidity", "perimeter", "width", "height", "cmx", "cmy", "hull_verticies",
46 "ex", "ey", "emajor", "eminor", "angle", "eccen", "circ", "round", "ar", "fd", "oof")]
47 days <- unique(shapes$day)
48 varieties2 <- unique(shapes$variety)
49 #print(none)
R version 4.2.1 (2022-06-23 ucrt) -- "Funny-Looking Kid"
Copyright (C) 2022 The R Foundation for Statistical Computing
```

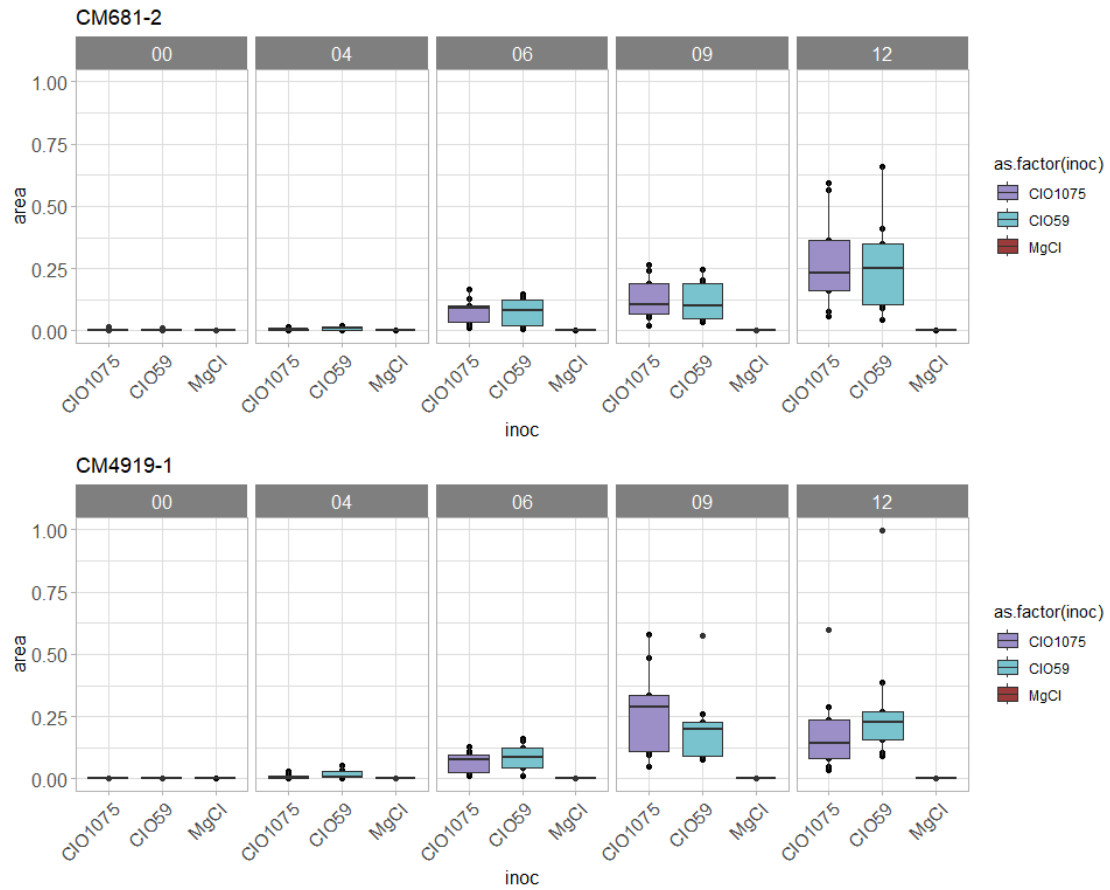
10. The report is generated as a word document as follows:

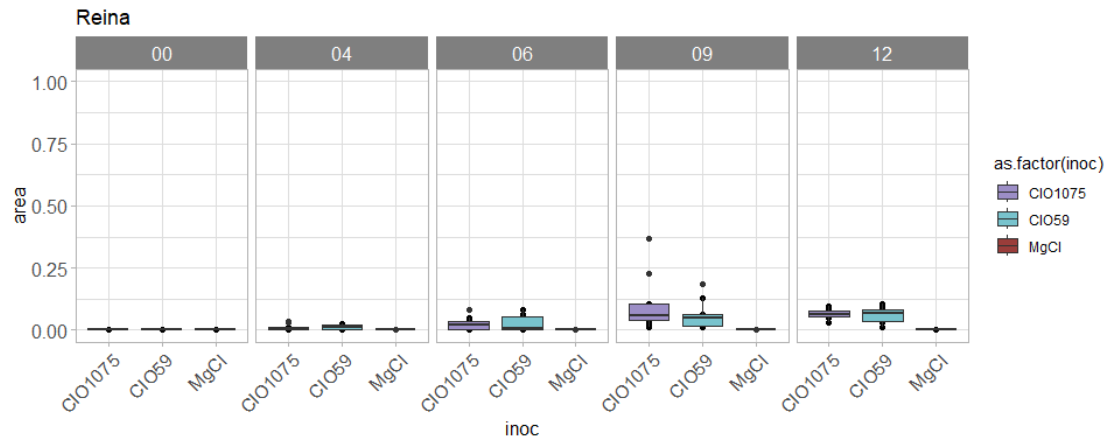
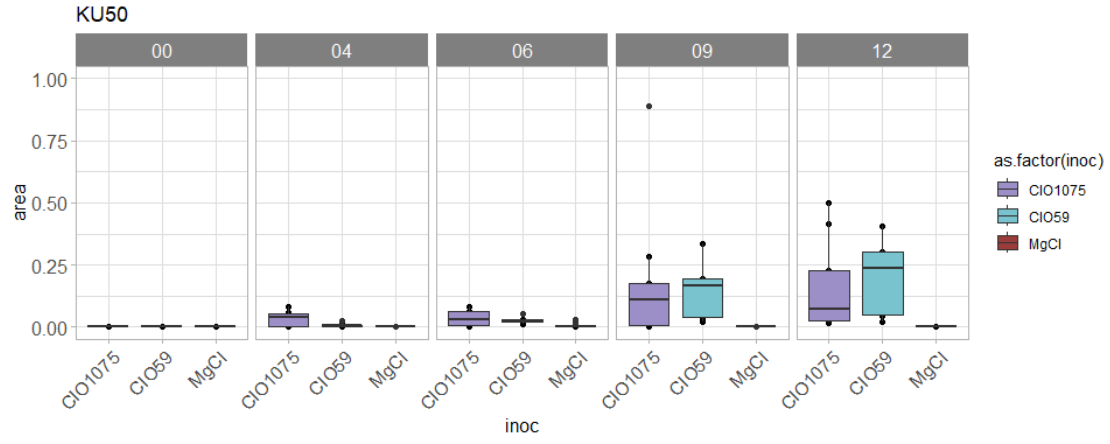
XPM 5

Author: JMP

Date: 2023-01-12

Boxplot Areas cm² (raw data) – Variety

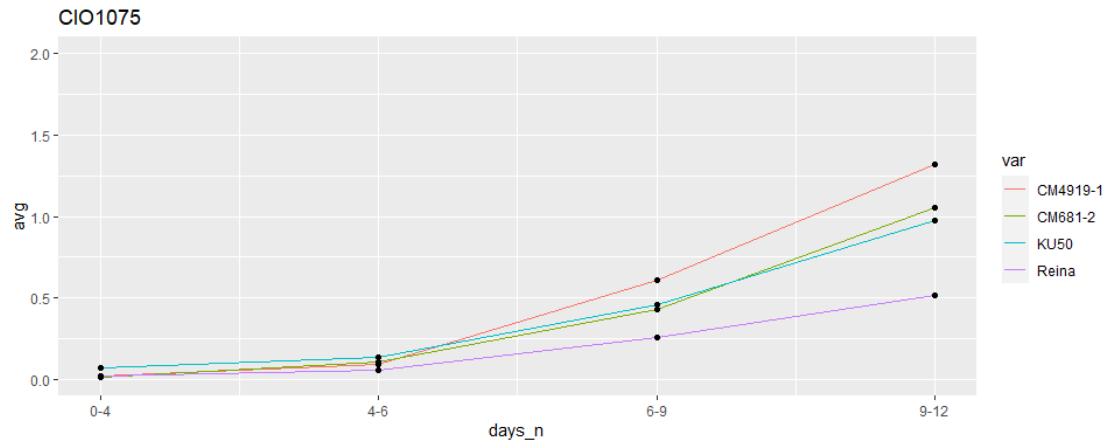
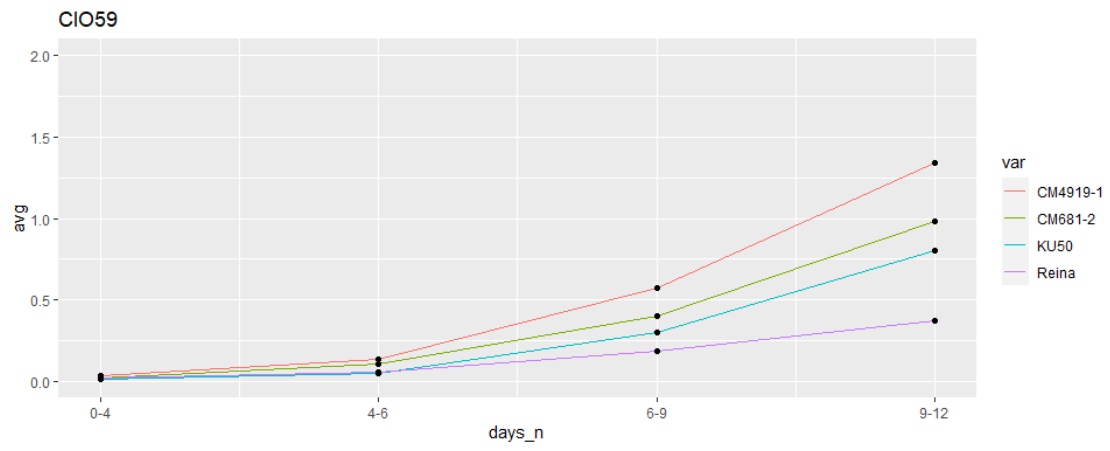
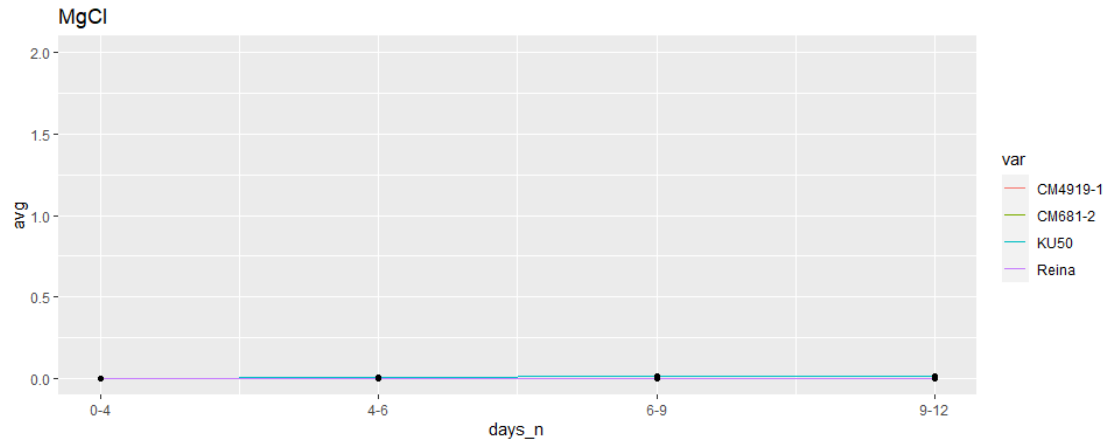




Boxplot Areas cm² (raw data) – Strain



Lines graphics for AUDPC by Strain



Variance analysis Test

	Df	Sum Sq	Mean Sq
rep	2	0.53798	0.26899

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
inoc	2	20.29297	10.14648	127.55	0.00024
Residuals	4	0.31819	0.07955	NA	NA

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
variety	3	4.8847	1.62823	10.3769	0.00001
inoc:variety	6	2.5452	0.42421	2.7035	0.01848
Residuals	90	14.1219	0.15691	NA	NA

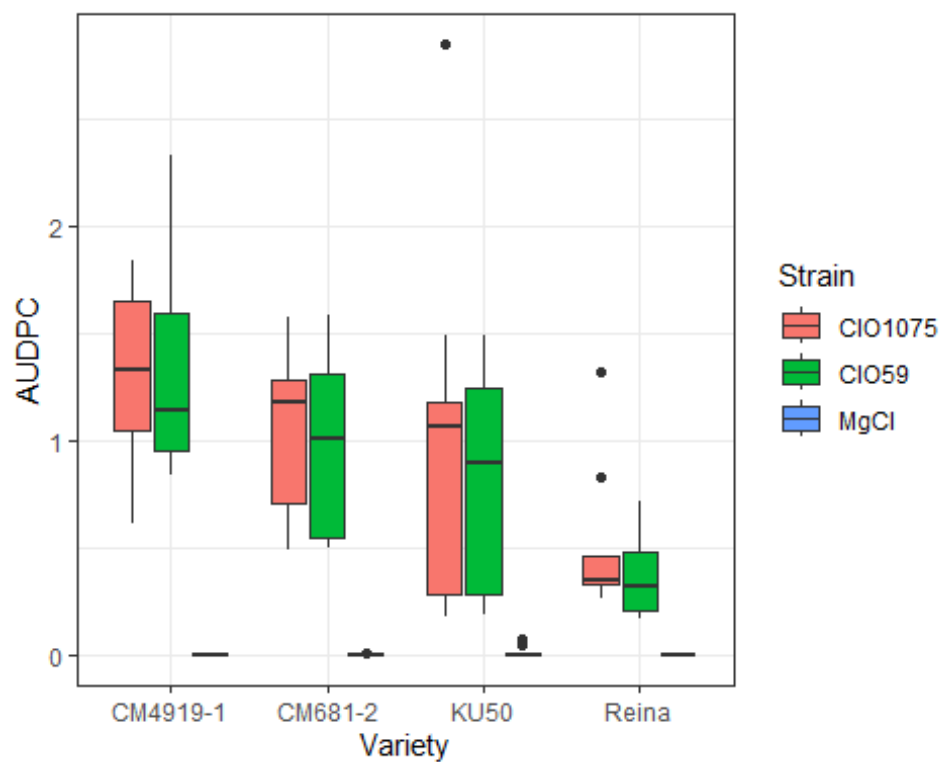
Comparing means Test

	as.numeric(acumAuc)	groups
CM4919-1	0.88773	a
CM681-2	0.68042	ab
KU50	0.59662	b
Reina	0.29575	c

	as.numeric(acumAuc)	groups
CIO59:CM4919-1	1.34408	a
CIO1075:CM4919-1	1.31732	a
CIO1075:CM681-2	1.05654	ab
CIO59:CM681-2	0.98403	ab
CIO1075:KU50	0.97317	ab
CIO59:KU50	0.80252	bc
CIO1075:Reina	0.51407	cd

	as.numeric(acumAuc)	groups
CIO59:Reina	0.37267	de
MgCl:KU50	0.01417	e
MgCl:CM4919-1	0.00178	e
MgCl:CM681-2	0.00068	e
MgCl:Reina	0.00051	e

Boxplot graphs for AUDPC (12DPI)



References

Lliott K, Berry JC, Kim H. et al. (2022). A comparison of ImageJ and machine learning based image analysis methods to measure cassava bacterial blight disease severity. *Plant Methods*, 18, 86.

Berry JC, Fahlgren N, Pokorny AA, Bart RS, Veley KM. (2018). An automated, high throughput method for standardizing image color profiles to improve image-based plant phenotyping. *PeerJ*. <https://doi.org/10.7717/peerj.5727>.

Restrepo S, Duque MC, Verdier V. (2000). Characterization of pathotypes among isolates of *Xanthomonas axonopodis* pv. *manihotis* in Colombia. *Plant Pathology*, 49(6), 680-687.

Zárate-Chaves CA, Gómez de la Cruz D, Verdier V, López CE, Bernal A, Szurek B. (2021). Cassava diseases caused by *Xanthomonas phaseoli* pv. *manihotis* and *Xanthomonas cassavae*. *Molecular Plant Pathology*, 22(12), 1520-1537.

CABI. (2020) *Xanthomonas axonopodis* pv. *manihotis* (cassava bacterial blight). In: Invasive species compendium. Available at: <https://www.cabi.org/isc/datasheet/56952>.

Bernal-Galeano V, Ochoa JC, Trujillo C, Rache L, Bernal A, López CA. (2018). Development of a multiplex nested PCR method for detection of *Xanthomonas axonopodis* pv. *manihotis* in cassava. *Tropical Plant Pathology*, 43(4), 341-350.