



https://helda.helsinki.fi

Co-inoculation of biochar and arbuscular mycorrhizae for growth promotion and nutrient fortification in soybean under drought conditions

Jabborova, Dilfuza

2022-07-22

Jabborova , D , Annapurna , K , Azimov , A , Tyagi , S , Pengani , K R , Sharma , P , Vikram , K V , Poczai , P , Nasif , O , Ansari , M J & Sayyed , R Z 2022 , ' Co-inoculation of biochar and arbuscular mycorrhizae for growth promotion and nutrient fortification in soybean under drought conditions ' , Frontiers in plant science , vol. 13 . https://doi.org/10.3389/fpls.2022.947547

http://hdl.handle.net/10138/354177 https://doi.org/10.3389/fpls.2022.947547

cc_by publishedVersion

Downloaded from Helda, University of Helsinki institutional repository.

This is an electronic reprint of the original article.

This reprint may differ from the original in pagination and typographic detail.

Please cite the original version.

Check for updates

OPEN ACCESS

EDITED BY Iftikhar Ali, State Key Laboratory of Molecular Developmental Biology, Institute of Genetics and Developmental Biology (CAS), China

REVIEWED BY

Ravish Choudhary, Indian Agricultural Research Institute (ICAR), India Abdul Gafur, SMF Corporate R&D, Indonesia

*CORRESPONDENCE

Dilfuza Jabborova dilfuzajabborova@yahoo.com Kannepalli Annapurna annapurna96@gmail.com Peter Poczai peter.poczai@helsinki.fi R. Z. Sayyed sayyedrz@gmail.com

SPECIALTY SECTION

This article was submitted to Plant Abiotic Stress, a section of the journal Frontiers in Plant Science

RECEIVED 18 May 2022 ACCEPTED 30 June 2022 PUBLISHED 22 July 2022

CITATION

Jabborova D, Annapurna K, Azimov A, Tyagi S, Pengani KR, Sharma P, Vikram KV, Poczai P, Nasif O, Ansari MJ and Sayyed RZ (2022) Co-inoculation of biochar and arbuscular mycorrhizae for growth promotion and nutrient fortification in soybean under drought conditions. *Front. Plant Sci.* 13:947547.

doi: 10.3389/fpls.2022.947547

COPYRIGHT

© 2022 Jabborova, Annapurna, Azimov, Tyagi, Pengani, Sharma, Vikram, Poczai, Nasif, Ansari and Sayyed. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or

reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s)

are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Co-inoculation of biochar and arbuscular mycorrhizae for growth promotion and nutrient fortification in soybean under drought conditions

Dilfuza Jabborova^{1,2*}, Kannepalli Annapurna^{2*}, A. Azimov¹, Swati Tyagi², Kedharnath Reddy Pengani², Prakriti Sharma², K. V. Vikram², Peter Poczai^{3*}, Omaima Nasif⁴, Mohammad Javed Ansari⁵ and R. Z. Sayyed^{6*}

¹Institute of Genetics and Plant Experimental Biology, Uzbekistan Academy of Sciences, Tashkent, Uzbekistan, ²Division of Microbiology, ICAR-Indian Agricultural Research Institute, New Delhi, India, ³Finnish Museum of Natural History, University of Helsinki, Helsinki, Finland, ⁴Department of Physiology, College of Medicine and King Khalid University Hospital, King Saud University, Riyadh, Saudi Arabia, ⁵Department of Botany, Hindu College, Moradabad, Mahatma Jyotiba Phule Rohilkhand University, Bareilly, India, ⁶Department of Microbiology, PSGVP Mandal's S. I. Patil Arts, G. B. Patel Science and S. T. K. V. Sangh Commerce College, Shahada, India

Drought is significant abiotic stress that affects the development and yield of many crops. The present study is to investigate the effect of arbuscular mycorrhizal fungi (AMF) and biochar on root morphological traits, growth, and physiological traits in soybean under water stress. Impact of AMF and biochar on development and root morphological traits in soybean and AMF spores number and the soil enzymes' activities were studied under drought conditions. After 40 days, plant growth parameters were measured. Drought stress negatively affected soybean growth, root parameters, physiological traits, microbial biomass, and soil enzyme activities. Biochar and AMF individually increase significantly plant growth (plant height, root dry weight, and nodule number), root parameters such as root diameter, root surface area, total root length, root volume, and projected area, total chlorophyll content, and nitrogen content in soybean over to control in water stress. In drought conditions, dual applications of AMF and biochar significantly enhanced shoot and root growth parameters, total chlorophyll, and nitrogen contents in soybean than control. Combined with biochar and AMF positively affects AMF spores number, microbial biomass, and soil enzyme activities in water stress conditions. In drought stress, dual applications of biochar and AMF increase microbial biomass by 28.3%, AMF spores number by 52.0%, alkaline phosphomonoesterase by 45.9%, dehydrogenase by 46.5%, and fluorescein diacetate by 52.2%, activities. The combined application of biochar and AMF enhance growth, root parameters in soybean and soil enzyme activities, and water stress tolerance. Dual applications with biochar and AMF benefit soybean cultivation under water stress conditions.

KEYWORDS

AMF, biochar, drought stress, fluorescein diacetate, phosphomonoesterase, plant growth, soybean

Introduction

Soybean is an important food legume crop globally. Seeds of soybean contain a higher content of protein (35–40%), an essential source of oil (18–22%), carbohydrate (34%), macronutrients [phosphorus (P), nitrogen (N), potassium (K)] and micronutrients such as iron (Fe), zinc (Zn), and manganese (Mn) (de Vargas et al., 2018). Soybean contains essential amino acids [histidine, lysine, threonine, phenylalanine, cysteine, glycine, ornithine, proline, and serine (Tessari et al., 2016)]. Soybean is known to stimulate soil fertility by symbiotic biological nitrogen fixation (Ciampitti and Salvagiotti, 2018).

Drought can negatively affect soybean growth, development, yield, and nitrogen fixation (Basal and Szabo, 2020; Kannepalli et al., 2021). Water stress reduces the germination of the plant (Ilyas et al., 2020; Khan et al., 2020), growth of the plant (Sheteiwy et al., 2021), and yield (Li et al., 2018; Kour et al., 2019) in legumes. Several investigated that drought stress decreases the number of nodules per plant, the number of pods in legumes (Yücel et al., 2010; Shamsizadeh et al., 2014; Ilyas et al., 2020), and decreases root length, root volume, and yield of grain (Habibzadeh et al., 2014).

Drought negatively affects plant nutrients (Tuteja and Gill, 2013) and physiological properties such as photosynthesis (Asha et al., 2021), relative water content (Sánchez-Blanco et al., 2004), and photosynthetic rate (Hashem et al., 2019) in many plants. The P content of leaves in mungbean (*Vigna radiata* L.) decreased by water stress (Habibzadeh et al., 2014). Hashem et al. (2019) investigated water stress reduced P uptake and total N in chickpea (*Cicer arietinum* L.). Drought decreases total chlorophyll contents in chickpea varieties (Khadraji and Ghoulam, 2017). Islam et al. (2011) investigated, that drought tress decreased the transpiration rate and stomatal conductance in cowpea *Vigna unguiculata*. Drought strongly affects urease, β -glycosidase, and phosphatase of soil and soil microbial activities (Sardans and Peñuelas, 2005; Nguyen et al., 2018).

Biochar contains organic matter, nutrients, and biologically important compounds (Dume et al., 2016). Numerous studies

on biochar applications enhance bioenergy production, carbon sequestration, and soil fertility (Ok et al., 2015). Biochar improved soil structure and water holding capacity in water stress (Bamminger et al., 2016). It positively impacted on plant growth and development (Sun et al., 2020; Mannan et al., 2021), nodulation (Sun et al., 2020), and yield (Liu et al., 2017) in legume crops under normal conditions and under drought stress. Biochar application increased N fixation and total biomass in Trifolium pratense L. (Mia et al., 2014). Numerous studies have informed biochar improved plant nutrients (Ahmed et al., 2021) and physiological traits under unstressed and water stress in many crops (Jabborova et al., 2021a,b,c; Christou et al., 2022). Mannan et al. (2021) investigated the application of biochar-enhanced soil nutrients may have enhanced soybean drought tolerance. In addition, biochar enhanced soil urease, catalase, and urease activities (Chen et al., 2020).

Arbuscular mycorrhizal fungi (AMF) are the most useful soil microbes and symbiosis relationship with roots of plant (Brundrett and Tedersoo, 2018; Jabborova et al., 2021d,f). In drought stress, AMF helps increase growth, root parameters, nodulation, and yield in legume crops (Hao et al., 2019; Haghighi and Saharkhiz, 2022). Ahmed et al. (2000) reported that AMF increased pods number and nodule number in faba bean (Vicia faba L.). AMF improves physiological properties (Hashem et al., 2019) and mineral nutrients, especially P and N. They help tolerate drought stress in legumes (Hashem et al., 2019). Numerous studies have informed that AMF improves host plants' development, water uptake, and nutrient in water stress (Baum et al., 2015; Zhao et al., 2015). The AMF promoted stomatal conductance and relative water content in host plants under drought stress (Augé et al., 2015). Dual application with biochar and AMF inoculation improves the photosynthetic rate, and plant growth in chickpea under water stress (Hashem et al., 2019). This study was investigated to evaluate the impact of AMF and biochar on soybean growth, root parameters, chlorophyll content, and soil enzymatic activities in drought conditions.

Materials and methods

Arbuscular mycorrhizal fungi, biochar, soil, and seed

The soil was collected from the farms of the Indian Agricultural Research Institute. The studied soil had the following agrochemical properties: pH 8.0, EC 0.45 ds/m, SOC 0.41%, nitrogen 167 kg/ha, P 40.3 kg/ha, P 788 kg/ha. Biochar used in the experiment was produced at 400–500°C from woody biomass. Soybean seed was procured from the "Division of Vegetable Science" and AMF from Microbiology, IARI, India.

Experimental design

The effect of AMF and biochar on the growth and root morphological traits in soybean was investigated in a net house at the Division of Microbiology, IARI. Four treatments as control, AMF alone, biochar alone, and combined with AMF and biochar were used for the experiment. Mixing the soil with 1.0% biochar used a pot experiment. AMF biofertilizer consists of 100 spores/g and 1,200 IP/g. Seed of soybean was cultivated into plastic pots, including soil (5.0 kg). Within 40 days of plant growth in drought conditions (50% of the field capacity) were maintained. After 40 days, plant height, nodule number, dry shoot weight, and dry root weight were measured. All the experiments were performed in triplicates.

Root parameters measurement

Soybean root morphological traits such as total root length, projected area, root surface area, root volume, and root diameter were evaluated. All roots were spread out and detected using a scanning system (Expression 4990, Epson, CA, United States) with a blue board as a background.

Chlorophyll, N, and C contents of leaves measurement

Chlorophyll content were analyzed (SPAD-502 meter) using the leaves of soybean. Elemental Analyzer (CHNS) Eurovictor determined C and N.

Determination of arbuscular mycorrhizal fungi spores number and microbial biomass in soil

The AMF spores number was counted using a stereomicroscope by Dare et al. (2013). The biomass C was analyzed on those described by Vance et al. (1987).

The absorbance of the resulting supernatant liquid was measured at 280 nm.

Determination of enzymes activities in soil

Alkaline phosphatase enzyme activities were analyzed according to the method of Tabatabai and Bremner (1969). The fluorescein diacetate (FDA) hydrolytic activity was determined as per the Green et al. (2006) method. The dehydrogenase activity (DHA) was analyzed by Casida et al. (1964). The experimental data were analyzed with the StatView Software using ANOVA. The magnitude of the *F* value determined the significance of the effect of treatment (P < 0.05 < 0.001).

Results

In water stress, biochar alone significantly greater the height of the plant by 28.9%, the dry weight of roots by 51.5%, and a number of nodules (per plant) by 34.9% than the control (**Table 1**). The AMF alone significantly enhances the plant's height by 20.0%, the dry weight of roots by 53.8%, and the number of nodules (per plant) by 69.2% over the control under water stress. However, the dual application of AMF and biochar was applied, the plant height by 31.1%, the dry weight of root by 69.2%, and the number of nodules per plant by 63.9% over control under drought conditions.

The biochar exhibited a significant positive impact on the root morphological traits of soybean (Table 2). Over the control, root surface area and total root length were suddenly promoted by biochar alone, which was significantly enhanced by 56.8% and 54.9% under drought stress. Biochar alone substantially increases the projected area by 26.2%, root volume by 35.2%, and the root diameter by 45.8% more than the control. Over to the control, total root length and, surface area were enhanced due to the application of AMF alone by 27.4 and 32.5% in drought conditions. In drought stress, AMF alone visibly increased projected area by 18.5% and root diameter by 22.9%. However, AMF and biochar significantly developed root morphological parameters under drought conditions. The maximum values of total root length were observed in dual applications with AMF and biochar in water stress. Combined biochar application with AMF treatment significantly increases the root volume by 85.2%, root diameter by 72.9%, root surface area by 66.7%, and total root length by 61.9% more than the control. While in water stress conditions, this treatment discernible improved the projected area by 35.4% over the control.

Generally, water stress decreased the N and C content of leaves in soybean (**Table 3**). Biochar treatment increases the N content by 15.7% and the C content by 52.8% of leaf under water stress. Over the control under drought stress, the application

Treatments	Plant height (cm)	Shoot dry weight (g)	Root dry weight (g)	Nodule number
Control	30.00 ± 0.57	1.56 ± 0.01	0.13 ± 0.01	16.67 ± 1.52
Biochar	$38.67 \pm 0.58^{*}$	1.62 ± 0.01	$0.21\pm0.01^{*}$	$24.00\pm1.00^{\ast}$
AMF	$36.00 \pm 1.00^{*}$	1.59 ± 0.01	$0.20\pm0.01^{*}$	$21.67 \pm 1.53^{*}$
Biochar + AMF	$39.33 \pm 0.18^{*}$	1.70 ± 0.01	$0.22\pm0.02^*$	$27.33 \pm 0.58^{*}$

TABLE 1 The plant height, shoot dry weight, root dry weight, and nodule number in soybean as affected by biochar and AMF under drought condition.

Data are means of three replicates (n = 3). Asterisk differed significantly at *P < 0.05, **P < 0.01, ***P < 0.001.

TABLE 2 Root morphological traits in soybean as affected by biochar and AMF under drought condition.

Treatments	Total root length (cm)	Projected area (cm ²)	Root surface area (cm ²)	Root volume (cm ³)	Root diameter (mm)
Control	109.54 ± 12.72	14.57 ± 0.96	5.66 ± 0.29	0.88 ± 0.03	0.48 ± 0.01
Biochar	$171.76 \pm 1.53^{*}$	$18.40\pm0.5^{*}$	$8.77\pm0.94^{\ast}$	$1.19 \pm 0.02^{**}$	$0.70\pm0.01^*$
AMF	$139.57 \pm 13.77^*$	$17.27 \pm 0.11^{*}$	$7.50\pm0.08^{*}$	$0.96\pm0.01^*$	$0.59\pm0.06^{*}$
Biochar + AMF	$177.44 \pm 5.75^{**}$	$19.74 \pm 0.99^{**}$	$9.44 \pm 0.81^{**}$	$1.63 \pm 0.07^{**}$	$0.83 \pm 0.03^{**}$

Data are means of three replicates (n = 3). Asterisk differed significantly at *P < 0.05, **P < 0.01, ***P < 0.001.

of AMF alone promoted the N content by 10.0% and the C content by 48.4%. The C and N content reached a maximum at dual applications with AMF and biochar than to all treatments. In water stress, dual AMF and biochar treatment suggestively promoted the C content by 59.3% and N content by 16.5% more than the control.

Water stress declines total chlorophyll content in soybean. In water stress, biochar alone significantly enhances the chlorophyll content in soybean by 12.9%. The application of AMF alone increased chlorophyll content by 14.9% over the control. Highest values of the chlorophyll content were observed at the dual application of AMF, and biochar increased significantly by 16.5% (**Figure 1**).

The number of spores in AMF, increases from 34.7% in drought stress. The application of applying biochar treatment stimulated the number of spores significantly in AMF under water stress conditions. The biochar alone increases the number of spores of AMF considerably by 20.6%, respectively. The dual application of AMF and biochar was more beneficial in enhancing the number of AMF spores in the soil. However, dual AMF and biochar meaningfully improved the number of AMF spores by 52.0% over the control under drought conditions (**Figure 2**).

TABLE 3 The nitrogen and carbon content of leaf in soybean as affected by biochar and AMF under drought condition.

Treatments	N (%)	C (%)
Control	2.459 ± 0.09	28.789 ± 2.01
Biochar	$2.846 \pm 0.05^{*}$	$43.998 \pm 1.22^{*}$
AMF	$2.705 \pm 0.02^{*}$	$42.749 \pm 5.36^{*}$
Biochar + AMF	$2.867 \pm 0.03^{*}$	$45.889 \pm 1.34^{*}$

Data are means of three replicates (n=3). A sterisk differed significantly at *P<0.05, ** P<0.01, *** P<0.001.



In drought stress, AMF and biochar alone significantly improved the microbial biomass, it increased 14.1 and 20.7%, respectively under drought conditions over the control (**Figure 3**). In drought stress, dual with biochar and AMF increase microbial biomass by 28.3% in soil compared with the control. The combined application of AMF and biochar under drought stress led to the maximum microbial biomass compared to all treatments.

Under drought stress, a single biochar application significantly enzyme activities of soil (**Table 4**). The dehydrogenase, alkaline phosphomonoesterase, and FDA activities were enhanced by 23.6, 27.7, and 30.8% when the soil was amended by biochar under water stress. Compared to the control, the application of AMF alone enhanced significantly by 34.0% under water stress. Under water stress, the application of AMF alone, dehydrogenase, and FDA activities increased by 31.5 and 38.1% over the control. In drought conditions,



FIGURE 2

The AMF spore numbers in soil affected by biochar and AMF under drought conditions. C, control; BC, biochar; AMF, arbuscular mycorrhizal fungi; BC + AMF, biochar + arbuscular mycorrhizal fungi. *P = 0.01.



TABLE 4 Soil enzymes activities in soil affected by biochar and AMF under drought condition.

Treatments	Alkaline phosphomon- oesterase (µg g ⁻¹ h ⁻¹)	Dehydrogenase activity $(\mu g g^{-1} h^{-1})$	FDA (fluorescein diacetate) activity (µg g ⁻¹ h ⁻¹)
Control	45.67 ± 0.59	38.00 ± 1.01	40.33 ± 0.58
Biochar	$58.33\pm0.58^{\ast}$	$47.00\pm1.00^{\ast}$	$52.77\pm2.08^{\ast}$
AMF	$61.20\pm2.63^{\ast}$	$50.00\pm2.20^{\ast}$	$55.73 \pm 1.00^{\ast}$
Biochar + AMF	$66.67 \pm 1.53^{*}$	$55.67 \pm 2.30^{**}$	$61.40 \pm 1.10^{**}$

Data are means of three replicates (n = 3). Asterisk differed significantly at *P < 0.05, **P < 0.01, ***P < 0.001.

dual applications with AMF and biochar increase soil enzyme activities were greatly better than all treatments. In drought stress, the dual applications of biochar and AMF treatments resulted in 45.9% more alkaline phosphomonoesterase activities than the control. The dual applications with AMF and biochar treatment promote significant dehydrogenase enzyme activity by 46.5% and FDA enzyme activity by 52.2% in soil over the control in drought conditions.

Discussion

Drought decreases plant growth parameters and nodule number. Several researchers have reported drought stress reduced plant growth, nodulation, and legume yield (Du et al., 2020; Jabborova et al., 2021e). Rusmana et al. (2020) informed that the growth and yield in *Glycine max* L. Merr. were decreased by water stress.

Numerous studies found that water stress reduced K, P, and N contents in plants (Dimkpa et al., 2017; Hashem et al., 2019) and physiological traits such as relative water content, carotenoids, and chlorophyll contents in legumes (Chowdhury et al., 2016; Kenawy et al., 2022). Ohashi et al. (2006) reported that the photosynthetic pigments in soybean decreased in drought. Geng et al. (2015) and Mariotte et al. (2015) informed water stress declined microbial activity and fungal traits in soil.

Biochar increased plant growth and root morphological traits in soybean under drought in the present study (Tables 1, 2). Similarly, Hashem et al. (2019) and Batool et al. (2015) found that biochar enhanced plant growth parameters in chickpea in water stress. In addition, de Melo Carvalho et al. (2013) reported biochar increased rice biomass in drought conditions. Addition of biochar significantly promotes chlorophyll, N, and C contents in soybean in drought conditions. Several studies show that biochar improved nutrient uptake (Akhtar et al., 2014) and plant physiological traits (Lyu et al., 2016) in various plants under drought stress. The present experiment investigated that biochar improves a significant number of AMF spores, microbial biomass, enzyme activities in soil in a significant number of AMF spores, microbial biomass, and enzyme activities in soil under water stress. Li and Cai (2021) investigated that biochar promoted microbial biomass by 38.0 and 65.9% in water stress. Identical observations recorded by Ahmad et al. (2014) found a helpful impact of biochar on microbial biomass activity.

Numerous studies have demonstrated that inoculation of microorganisms help plants to overcome drought stress and grow well under drought conditions (Kalam et al., 2020; Basu et al., 2021; Hamid et al., 2021; Khan et al., 2021; Sagar et al., 2022). Islam et al. (2022) reported the abundance of mycorrhizae in various host plants and rhizosphere soil. Bastami et al. (2021) reported positive impacts of mycorrhizal fungi and organic fertilizers on quantitative and qualitative traits in Satureja species. Researchers have established that AMF improves host plants' development and water uptake in water stress conditions (Quiroga et al., 2017; Vafa et al., 2021). Identical data were informed by Hashem et al. (2019). Under drought-stressed conditions, AMF improved growth and biomass production, nutrient and water acquisition (Fallah et al., 2021; Jabborova et al., 2021e; Najafi et al., 2021; Saboor et al., 2021). Inoculated with AMF to maize plants significantly increased height by 36.32% and dry weight by 75.73% than the control in water stress (Begum et al., 2019; Rawat et al., 2019).

Arbuscular mycorrhizal fungi inoculation improved plant nutrients in plants under water stress (Baum et al., 2015; Zhao et al., 2015). AMF-inoculated enhanced uptake of minerals "Mg, N, and K" in cucumber in drought conditions (Wang et al., 2008). AMF enhances the significant P content of maize in water stress (Zhao et al., 2015). Similarly, as confirmed by Abdel-Salam et al. (2018), AMF improved damask's chlorophyll content under water stress. Similarly, Gong et al. (2013) reported that inoculated AMF significantly enhances chlorophyll content and stomatal conductance compared with non-mycorrhizal seedlings in drought. AMF alone considerably promoted the number of AMF spores number, the microbial biomass, and soil enzyme activities in a previous study. Similar findings were reported by Li and Cai (2021) AMF stimulated soil microbial biomass and promoted spores number of AMF in water stress (Hashem et al., 2019).

Combining biochar and AMF under drought stress promoted plant growth and root parameters in plants under drought conditions (Gujre et al., 2021; Mesbah et al., 2021; Ndiate et al., 2021). Budi and Setyaningsih (2013) reported that AMF and biochar promote shoot dry weight in Melia azedarach Linn. Harun et al. (2021) informed that combined with AMF and biochar improved "shoot and root biomass" and root length in Annona muricata L. Dual applications of AMF and biochar enhance N, C, and chlorophyll content in drought conditions. Similarly, Li and Cai (2021) investigated dual biochar, and AMF improved the P content of maize in water stress. Dual applications with AMF and biochar improve the significant number of AMF spores, microbial biomass, and soil enzyme activities in drought. Hashem et al. (2019) and He et al. (2020) informed that combined application of biochar, and AMF meaningfully stimulated the total number of AMF spores under drought stress conditions. Li and Cai (2021) combined with AMF and biochar greatly improved soil microbial activity and phosphatase activity in soil under water stress.

Conclusion

Biochar could improve plant growth, root morphological parameters, and soil enzyme activities in water stress conditions. Applications of biochar alone and AMF alone affected soybean tolerance to drought stress. AMF alone promoted plant growth, root morphological traits, chlorophyll content, AMF spores number, and microbial biomass under drought conditions. Dual applications with AMF and biochar showed the best results in drought stress. Combined with AMF and biochar improved plant growth, root morphological traits, and microbial biomass in drought conditions. The dual applications with biochar and AMF can decrease the effects of water stress, helping to improve soybean growth, yield, and soil enzyme activities under drought conditions.

Data availability statement

The original contributions presented in this study are included in the article/supplementary material, further inquiries can be directed to the corresponding authors.

Author contributions

DJ: conceptualization, methodology, and writing the original draft. KA, AA, ST, KP, PS, and KV: methodology. DJ, KA, ON, MA, and RS: formal analysis and writing—review and editing. PP: conceptualization, formal analysis, open access fund acquisition, and writing—review and editing. ON: writing—review and editing and fund acquisition. All authors contributed to the article and approved the submitted version.

Funding

This project was supported by the Department of Biotechnology, Government of India (DBT), TWAS, and Researchers Supporting Project number (RSP-2022/257) King Saud University, Riyadh, Saudi Arabia. This project was supported by the Researchers Supporting Project number (RSP-2022/257), King Saud University, Riyadh, Saudi Arabia, Department of Biotechnology, Government of India (DBT), and TWAS. Open access was supported by Helsinki University Library.

Acknowledgments

We thank our colleagues at the Division of Microbiology, IARI, New Delhi, India for the support. PP is thankful to Helsinki University Library for open access support.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Abdel-Salam, E., Alatar, A., and El-Sheikh, M. A. (2018). Inoculation with arbuscular mycorrhizal fungi alleviates harmful effects of drought stress on damask rose. *Saudi J. Biol. Sci.* 25, 1772–1780. doi: 10.1016/j.sjbs.2017.10.015

Ahmad, M., Rajapaksha, A. U., Lim, J. E., Zhang, M., Bolan, N., Mohan, D., et al. (2014). Biochar as a sorbent for contaminant management in soil and water: a review. *Chemosphere* 99, 19–33. doi: 10.1016/j.chemosphere.2013.10.071

Ahmed, F. E., Yagoub, S. O., and Elsheikh, E. A. (2000). Effects of mycorrhizal inoculation and phosphorus application on the nodulation, mycorrhizal infection and yield components of faba bean grown under two different watering regimes. *Univ. Khartoum J. Agr. Sci.* 8, 107–16.

Ahmed, N., Basit, A., Bashir, S., Bashir, S., Bibi, I., Haider, Z., et al. (2021). Effect of acidified biochar on soil phosphorus availability and fertilizer use efficiency of maize (Zea mays L.). *J. King Saud Univ. Sci.* 33:101635. doi: 10.1016/j.jksus.2021. 101635

Akhtar, S. S., Li, G., Andersen, M. N., and Liu, F. (2014). Biochar enhances yield and quality of tomato under reduced irrigation. *Agric. Water Manage*. 138, 37–44. doi: 10.1016/j.agwat.2014.02.016

Asha, A. D., Nivetha, N., Krishna, G. K., Thakur, J. K., Rathi, M. S., Manjunatha, B. S., et al. (2021). Amelioration of short-term drought stress during different growth stages in Brassica juncea by rhizobacteria mediated maintenance of ROS homeostasis. *Physiol. Planta.* 72, 1880–1893. doi: 10.1111/ppl.13399

Augé, R. M., Toler, H. D., and Saxton, A. M. (2015). Arbuscular mycorrhizal symbiosis alters stomatal conductance of host plants more under drought than under amply watered conditions: a meta-analysis. *Mycorrhiza* 25, 13–24. doi: 10.1007/s00572-014-0585-4

Bamminger, C., Poll, C., Sixt, C., Högy, P., Wüst, D., Kandeler, E., et al. (2016). Short-term response of soil microorganisms to biochar addition in a temperate agroecosystem under soil warming. *Agric. Ecosyst. Environ.* 233, 308–317. doi: 10.1016/j.agee.2016.09.016

Basal, O., and Szabo, A. (2020). Physiomorphology of soybean as affected by drought stress and nitrogen application. *Scientifica* 2020, 1–7. doi: 10.1155/2020/6093836

Bastami, A., Amirnia, R., Sayyed, R. Z., and Enshasy, H. E. (2021). The effect of mycorrhizal fungi and organic fertilizers on quantitative and qualitative traits of two important Satureja species. *Agronomy* 11:1285. doi: 10.3390/agronomy11071285

Basu, A., Prasad, P., Das, S. N., Kalam, S., Sayyed, R. Z., Reddy, M. S., et al. (2021). Plant Growth Promoting Rhizobacteria (PGPR) as Green Bioinoculants: Recent Developments, Constraints, and Prospects. *Sustainability* 13:1140. doi: 10.3390/su13031140

Batool, A., Taj, S., Rashid, A., Khalid, A., Qadeer, S., Saleem, A. R., et al. (2015). Potential of soil amendments (Biochar and Gypsum) in increasing water use efficiency of *Abelmoschus esculentus* L. Moench. *Front. Plant Sci.* 6:733. doi: 10.3389/fpls.2015.00733

Baum, C., El-Tohamy, W., and Gruda, N. (2015). Increasing the productivity and product quality of vegetable crops using arbuscular mycorrhizal fungi: a review. *Sci. Horticult*. 187, 131–141. doi: 10.1016/j.scienta.2015.03.002

Begum, N., Ahanger, M. A., Su, Y., Lei, Y., Mustafa, N. S., Ahmad, P., et al. (2019). Improved drought tolerance by AMF inoculation in maize (Zea mays) involves physiological and biochemical implications. *Plants* 8:579. doi: 10.3390/ plants8120579

Brundrett, M. C., and Tedersoo, L. (2018). Evolutionary history of mycorrhizal symbioses and global host plant diversity. *New Phytol.* 220, 1108–1115. doi: 10. 1111/nph.14976

Budi, S. W., and Setyaningsih, L. (2013). Arbuscular mycorrhizal fungi and biochar improved early growth of neem (*Melia azedarach* Linn.) seedling under greenhouse conditions. *J. Manajemen Hutan Tropika* 19, 103–110.

Casida, L. E., Klein, D. A., and Santoro, T. (1964). Soil dehydrogenase activity. Soil Sci. 98, 371-376. doi: 10.1097/00010694-196412000-00004

Chen, H., Yang, X., Wang, H., Sarkar, B., Shaheen, S. M., Gielen, G., et al. (2020). Animal carcass-and wood-derived biochars improved nutrient bioavailability, enzyme activity, and plant growth in metal-phthalic acid ester co-contaminated soils: a trial for reclamation and improvement of degraded soils. *J. Environ. Manage.* 261:110246. doi: 10.1016/j.jenvman.2020.110246

Chowdhury, J. A., Karim, M. A., Khaliq, Q. A., Ahmed, A. U., and Khan, M. S. (2016). Effect of drought stress on gas exchange characteristics of four soybean genotypes. *Bangl. J. Agric. Res.* 41, 195–205. doi: 10.3329/bjar.v41i2.28215

Christou, A., Stylianou, M., Georgiadou, E. C., Gedeon, S., Ioannou, A., Michael, C., et al. (2022). Effects of biochar derived from the pyrolysis of either biosolids,

manure or spent coffee grounds on the growth, physiology and quality attributes of field-grown lettuce plants. *Environ. Technol. Innovat.* 26:102263. doi: 10.1016/j. eti.2021.102263

Ciampitti, I. A., and Salvagiotti, F. (2018). New insights into soybean biological nitrogen fixation. *Am. Soc. Agron.* 110, 1185–1196. doi: 10.2134/agronj2017.06. 0348

Dare, M. O., Abaidoo, R., Fagbola, O., and Asiedu, R. (2013). Diversity of arbuscular mycorrhizal fungi in soils of yam (Dioscorea spp.) cropping systems in four agroecologies of Nigeria. *Arch. Agron. Soil Sci.* 59, 521–531. doi: 10.1080/03650340.2011.653682

de Melo Carvalho, M. T., Madari, B. E., Bastiaans, L., van Oort, P. A., Heinemann, A. B., da Silva, M. A., et al. (2013). Biochar improves fertility of a clay soil in the Brazilian Savannah: short term effects and impact on rice yield. *J. Agric. Rural Dev. Tropics Subtrop.* 114, 101–107.

de Vargas, R. L., Schuch, L. O., Barros, W. S., Rigo, G. A., Szareski, V. J., Carvalho, I. R., et al. (2018). Macronutrients and micronutrients variability in soybean seeds. *J. Agric. Sci.* 10, 209–222. doi: 10.5539/jas.v10n4p209

Dimkpa, C. O., Bindraban, P. S., Fugice, J., Agyin-Birikorang, S., Singh, U., and Hellums, D. (2017). Composite micronutrient nanoparticles and salts decrease drought stress in soybean. *Agron. Sustain. Dev.* 37:5. doi: 10.1007/s13593-016-0412-8

Du, Y., Zhao, Q., Chen, L., Yao, X., and Xie, F. (2020). Effect of drought stress at reproductive stages on growth and nitrogen metabolism in soybean. *Agronomy* 10:302. doi: 10.3390/agronomy10020302

Dume, B., Mosissa, T., and Nebiyu, A. (2016). Effect of biochar on soil properties and lead (Pb) availability in a military camp in South West Ethiopia. *Afr. J. Environ. Sci. Technol.* 10, 77–85. doi: 10.5897/AJEST2015.2014

Fallah, M., Hadi, H., Amirnia, R., Ghorttapeh, A. H., Ali, T. K. Z., and Sayyed, R. Z. (2021). Eco-Friendly Soil Amendments Improve Growth, Antioxidant Activities, and Root Colonization in Lingrain (*Linum Usitatissimum* L.) Under Drought Condition. *PLoS One* 16:e0261225. doi: 10.1371/journal.pone.026 1225

Geng, S. M., Yan, D. H., Zhang, T. X., Weng, B. S., Zhang, Z. B., and Qin, T. L. (2015). Effects of drought stress on agriculture soil. *Nat. Haz.* 75, 1997–2011. doi: 10.1007/s11069-014-1409-8

Gong, M., Tang, M., Chen, H., Zhang, Q., and Feng, X. (2013). Effects of two Glomus species on the growth and physiological performance of Sophora davidii seedlings under water stress. *New For.* 44, 399–408. doi: 10.1007/s11056-012-9349-1

Green, V. S., Stott, D. E., and Diack, M. (2006). Assay for fluorescein diacetate hydrolytic activity: optimization for soil samples. *Soil Biol. Biochem.* 38, 693–701. doi: 10.1016/j.soilbio.2005.06.020

Gujre, N., Soni, A., Rangan, L., Tsang, D. C., and Mitra, S. (2021). Sustainable improvement of soil health utilizing biochar and arbuscular mycorrhizal fungi: a review. *Environ. Pollut.* 268:115549. doi: 10.1016/j.envpol.2020. 115549

Habibzadeh, Y., Evazi, A. R., and Abedi, M. (2014). Alleviation drought stress of mungbean (*Vigna radiata* L.) plants by using arbuscular mycorrhizal fungi. *Int. J. Agric. Sci. Nat. Resour.* 1, 1–6. doi: 10.18483/ijSci.651

Haghighi, T. M., and Saharkhiz, M. J. (2022). Mycorrhizal colonization and silicon nutrition mitigates drought stress in Licorice (Glycyrrhiza glabra L.) with morphophysiological and biochemical perspectives. *Indus. Crops Products* 178:114650. doi: 10.1016/j.indcrop.2022.114650

Hamid, B., Zaman, M., Farooq, S., Fatima, S., Sayyed, R. Z., Baba, Z. A., et al. (2021). Bacterial Plant Biostimulants: A Sustainable Way towards Improving Growth, Productivity, and Health of Crops. *Sustainability* 13:2856. doi: 10.3390/su13052856

Hao, Z., Xie, W., Jiang, X., Wu, Z., Zhang, X., and Chen, B. (2019). Arbuscular mycorrhizal fungus improves rhizobium–glycyrrhiza seedling symbiosis under drought stress. *Agronomy* 9:572. doi: 10.3390/agronomy9100572

Harun, N. S., Jaafar, N. M., and Sakimin, S. Z. (2021). The effects of rice husk biochar rate on arbuscular mycorrhizal fungi and growth of soursop (*Annona muricata* L.) seedlings. *Sustainability* 13, 1817. doi: 10.3390/su13041817

Hashem, A., Kumar, A., Al-Dbass, A. M., Alqarawi, A. A., Al-Arjani, A. B., Singh, G., et al. (2019). Arbuscular mycorrhizal fungi and biochar improves drought tolerance in chickpea. *Saudi J. Biol. Sci.* 26, 614–624. doi: 10.1016/j.sjbs. 2018.11.005

He, Y., Yao, Y., Ji, Y., Deng, J., Zhou, G., Liu, R., et al. (2020). Biochar amendment boosts photosynthesis and biomass in C3 but not C4

plants: a global synthesis. GCB Bioenerg. 12, 605-617. doi: 10.1111/gcbb.1 2720

Ilyas, N., Mumtaz, K., Akhtar, N., Yasmin, H., Sayyed, R. Z., Khan, W., et al. (2020). Exopolysaccharides Producing Bacteria for the Amelioration of Drought Stress in Wheat. *Sustainability* 12:8876. doi: 10.3390/su12218876

Islam, M., Al-Hashimi, A., Ayshasiddeka, M., Ali1, H., Enshasy, H. E., Dailin, D. J., et al. (2022). Prevalence of mycorrhizae in host plants and rhizosphere soil: a biodiversity aspect. *PLoS One* 17:e0266403. pone.0266403 doi: 10.1371/journal. pone.0266403

Islam, S., Calvin, K., and Willy, V. (2011). Physiological and morphological characteristics of cowpea genotypes to drought stress. *Arkansas Environ. Agric. Consum. Sci. J.* 42–45.

Jabborova, D., Kadirova, D., Narimanov, A., and Wirth, S. (2021a). Beneficial effects of biochar application on lettuce (*Lactuca sativa* L.) growth, root morphological traits and physiological properties. *Annals Phytomed.* 10, 93–100. doi: 10.21276/ap.2021.10.2.13

Jabborova, D., Wirth, S., Halwani, M., Ibrahim, M. F., Azab, I. H., El-Mogy, M. M., et al. (2021b). Growth response of ginger (*Zingiber officinale*), its physiological properties and soil enzyme activities after biochar application under greenhouse conditions. *Horticulturae* 7:250. doi: 10.3390/horticulturae7080250

Jabborova, D., Ma, H., Bellingrath-Kimura, S. D., and Wirth, S. (2021c). Impacts of biochar on basil (*Ocimum basilicum*) growth, root morphological traits, plant biochemical and physiological properties and soil enzymatic activities. *Sci. Horticult.* 290:110518. doi: 10.1016/j.scienta.2021.110518

Jabborova, D., Annapurna, K., Choudhary, R., Bhowmik, S. N., Desouky, S. E., Selim, S., et al. (2021d). Interactive Impact of Biochar and Arbuscular Mycorrhizal on Root Morphology, Physiological Properties of Fenugreek (*Trigonella foenumgraecum L.*) and Soil Enzymatic Activities. *Agronomy* 11:2341. doi: 10.3390/ agronomy11112341

Jabborova, D., Annapurna, K., Al-Sadi, A. M., Alharbi, S. A., Datta, R., and Zuan, A. T. (2021f). Biochar and Arbuscular mycorrhizal fungi mediated enhanced drought tolerance in Okra (*Abelmoschus esculentus*) plant growth, root morphological traits and physiological properties. *Saudi J. Biol. Sci.* 28, 5490–5499. doi: 10.1016/j.sjbs.2021.08.016

Jabborova, D., Kannepalli, A., Davranov, K., Narimanov, A., Enakiev, Y., Syed, A., et al. (2021e). Co-inoculation of rhizobacteria promotes growth, yield, and nutrient contents in soybean and improves soil enzymes and nutrients under drought conditions. *Sci. Rep.* 11:22081. doi: 10.1038/s41598-021-01337-9

Kalam, S., Basu, A., Ahmad, I., Sayyed, R. Z., Enshasy, H. A. E., Dailin, D. J., et al. (2020). Recent understanding of soil Acidobacteria and their ecological significance: a critical review. *Front. Microbiol.* 11:580024. doi: 10.3389/fmicb. 2020.580024

Kannepalli, A., Davranov, D., Narimanov, A., Enakiev, Y., Syed, A., Elgorban, A. M., et al. (2021). Co-inoculation of rhizobacteria promotes growth, yield, and nutrient contents in soybean and improves soil enzymes and nutrients under drought conditions. *Sci. Rep.* 11:22081.

Kenawy, E. R., Rashad, M., Hosny, A., Shendy, S., Gad, D., and Saad-Allah, K. M. (2022). Enhancement of growth and physiological traits under drought stress in Faba bean (*Vicia faba* L.) using nanocomposite. *J. Plant Interact.* 17, 404–418. doi: 10.1080/17429145.2022.2038293

Khadraji, A., and Ghoulam, C. (2017). Effect of drought on growth, physiological and biochemical processes of chickpea-rhizobia symbiosis. *Legume Res.* 40, 94–99.

Khan, I., Awan, S. A., Ikram, R., Rizwan, M., Akhtar, N., Yasmin, H., et al. (2020). 24-Epibrassinolide regulated antioxidants and osmolyte defense and endogenous hormones in two wheat varieties under drought stress. *Physiol. Planta.* 172, 1–11. doi: 10.1111/ppl.13237

Khan, N., Ali, S., Shahi, M. A., Mustafa, A., Sayyed, R. Z., and Curaá, J. A. (2021). Insights into the Interactions among Roots, Rhizosphere and Rhizobacteria for Improving Plant Growth and Tolerance to Abiotic Stresses: A Review. *Cells* 10:1551. doi: 10.3390/cells10061551

Kour, D., Rana, K. L., Yadav, A. N., Yadav, N., Kumar, V., Kumar, A., et al. (2019). "Drought Tolerant Phosphorus Solubilizing Microbes: Biodiversity and Biotechnological Applications for Alleviation of Drought Stress in Plant," in *Plant Growth Promoting Rhizobacteria for Sustainable Stress Management Vol 1 Abiotic Stress Management*, eds R. Z. Sayyed, N. K. Arora, and M. S. Reddy (Singapore: Springer), 255–308. doi: 10.1007/978-981-13-6536-2_13

Li, M., and Cai, L. (2021). Biochar and Arbuscular Mycorrhizal Fungi Play Different Roles in Enabling Maize to Uptake Phosphorus. *Sustainability* 13:3244. doi: 10.3390/su13063244

Li, P., Zhang, Y., Wu, X., and Liu, Y. (2018). Drought stress impact on leaf proteome variations of faba bean (*Vicia faba* L.) in the Qinghai–Tibet Plateau of China. *3 Biotech* 8:110. doi: 10.1007/s13205-018-1088-3

Liu, L., Wang, Y., Yan, X., Li, J., Jiao, N., and Hu, S. (2017). Biochar amendments increase the yield advantage of legume-based intercropping systems over monoculture. *Agric. Ecosyst. Environ.* 237, 16–23. doi: 10.1016/j.agee.2016.12. 026

Lyu, S., Du, G., Liu, Z., Zhao, L., and Lyu, D. (2016). Effects of biochar on photosystem function and activities of protective enzymes in Pyrus ussuriensis Maxim. under drought stress. *Acta Physiol. Planta.* 38:220. doi: 10.1007/s11738-016-2236-1

Mannan, M. A., Mia, S., Halder, E., and Dijkstra, F. A. (2021). Biochar application rate does not improve plant water availability in soybean under drought stress. *Agric. Water Manage.* 253:106940. doi: 10.1016/j.agwat.2021. 106940

Mariotte, P., Robroek, B. J., Jassey, V. E., and Buttler, A. (2015). Subordinate plants mitigate drought effects on soil ecosystem processes by stimulating fungi. *Func. Ecol.* 29, 1578–1586. doi: 10.1111/1365-2435.12467

Mesbah, R., Ardakani, M. R., Moghaddam, A., and Rafiei, F. (2021). Correlation and path analysis of Tobacco (*Nicotiana tabacum* L.) yield vs root traits and relative water content as affected by Azotobacter, mycorrhizal symbiosis and biochar application under dry-land farming conditions. *Int. Agrophys.* 35, 319– 329. doi: 10.31545/intagr/143945

Mia, S., Van Groenigen, J. W., Van de Voorde, T. F., Oram, N. J., Bezemer, T. M., Mommer, L., et al. (2014). Biochar application rate affects biological nitrogen fixation in red clover conditional on potassium availability. *Agric. Ecosyst. Environ.* 191, 83–91. doi: 10.1016/j.agee.2014.03.011

Najafi, S., Nasi, H. N., Tuncturk, R., Tuncturk, M., Sayyed, R. Z., and Amirnia, R. (2021). Biofertilizer application enhances drought stress tolerance and alters the antioxidant enzymes in medicinal pumpkin (*Cucurbita pepo* convar. pepo var. *Styriaca*). *Horticulturae* 7:588. doi: 10.3390/horticulturae7120588

Ndiate, N. I., Saeed, Q., Haider, F. U., Liqun, C., Nkoh, J. N., and Mustafa, A. (2021). Co-Application of Biochar and Arbuscular mycorrhizal Fungi Improves Salinity Tolerance, Growth and Lipid Metabolism of Maize (*Zea mays L.*) in an Alkaline Soil. *Plants* 10:2490. doi: 10.3390/plants10112490

Nguyen, L. T., Osanai, Y., Anderson, I. C., Bange, M. P., Tissue, D. T., and Singh, B. K. (2018). Flooding and prolonged drought have differential legacy impacts on soil nitrogen cycling, microbial communities and plant productivity. *Plant Soil* 431, 371–387. doi: 10.1007/s11104-018-3774-7

Ohashi, Y., Nakayama, N., Saneoka, H., and Fujita, K. (2006). Effects of drought stress on photosynthetic gas exchange, chlorophyll fluorescence and stem diameter of soybean plants. *Biol. Planta.* 50, 138–141. doi: 10.1007/s10535-005-0089-3

Ok, Y. S., Chang, S. X., Gao, B., and Chung, H. J. (2015). SMART biochar technology—a shifting paradigm towards advanced materials and healthcare research. *Environ. Technol. Innovat.* 4, 206–209. doi: 10.1016/j.eti.2015. 08.003

Quiroga, G., Erice, G., Aroca, R., Chaumont, F., and Ruiz-Lozano, J. M. (2017). Enhanced drought stress tolerance by the arbuscular mycorrhizal symbiosis in a drought-sensitive maize cultivar is related to a broader and differential regulation of host plant aquaporins than in a drought-tolerant cultivar. *Front. Plant Sci.* 8:1056. doi: 10.3389/fpls.2017.01056

Rawat, J., Saxena, J., and Sanwal, P. (2019). "Biochar: a sustainable approach for improving plant growth and soil properties," in *Biochar – An Imperative Amendment for Soil and the Environment*, eds V. Abrol and P. Sharma (London: IntechOpen). doi: 10.5772/intechopen.82151

Rusmana, R., Ningsih, E. P., and Justika, A. (2020). Growth and yield of various soy varieties (*Glycine max* L. Merr.) on drought stress. *J. Keteknikan Pertanian Tropis dan Biosistem.* 8, 228–235. doi: 10.21776/ub.jkptb.2020.008. 03.04

Saboor, A., Muhammad, A. A., Hussain, S., El Enshasy, H. E., Hussain, S., Ahmed, N., et al. (2021). Zinc nutrition and arbuscular mycorrhizal symbiosis effects on maize (*Zea mays* L.) growth and productivity. *Saudi J. Biol. Sci.* 28, 6339–6351. doi: 10.1016/j.sjbs.2021.06.096

Sagar, A., Yadav, S. S., Sayyed, R. Z., Sharma, S., and Ramteke, P. W. (2022). "Bacillus subtilis: A Multifarious Plant Growth Promoter, Biocontrol Agent, and Bioalleviator of Abiotic Stress," in Bacilli in Agrobiotechnology. Bacilli in Climate Resilient Agriculture and Bioprospecting, eds M. T. Islam, M. Rahman, and P. Pandey (Cham: Springer), doi: 10.1007/978-3-030-85465-2_24

Sánchez-Blanco, M. J., Ferrández, T., Morales, M. A., Morte, A., and Alarcón, J. J. (2004). Variations in water status, gas exchange, and growth in Rosmarinus officinalis plants infected with *Glomus deserticola* under drought conditions. *J. Plant Physiol.* 161, 675–682. doi: 10.1078/0176-1617-01191

Sardans, J., and Peñuelas, J. (2005). Drought decreases soil enzyme activity in a Mediterranean Quercus ilex L. forest. *Soil Biol. Biochem.* 37, 455–461. doi: 10.1016/j.soilbio.2004.08.004 Shamsizadeh, M., Shaban, M., and Motlagh, Z. R. (2014). Effect of Drought Stress and Zn Fertilizer on Some Root Characteristics of Chickpea Cultivars. *Int. J. Adv. Biol. Biom. Res.* 2, 2289–2293.

Sheteiwy, M. S., Ali, D. F., Xiong, Y. C., Brestic, M., Skalicky, M., Hamoud, Y. A., et al. (2021). Physiological and biochemical responses of soybean plants inoculated with Arbuscular mycorrhizal fungi and *Bradyrhizobium* under drought stress. *BMC Plant Biol.* 21:195. doi: 10.1186/s12870-021-02949-z

Sun, Q., Liu, Y., Liu, H., and Dumroese, R. K. (2020). Interaction of biochar type and rhizobia inoculation increases the growth and biological nitrogen fixation of *Robinia pseudoacacia* seedlings. *Forests* 11:711. doi: 10.3390/f11060711

Tabatabai, M. A., and Bremner, J. M. (1969). Use of p-nitrophenyl phosphate for assay of soil phosphatase activity. *Soil Biol. Biochem.* 1, 301–307. doi: 10.1016/0038-0717(69)90012-1

Tessari, P., Lante, A., and Mosca, G. (2016). Essential amino acids: master regulators of nutrition and environmental footprint? *Sci. Rep.* 6:26074. doi: 10. 1038/srep26074

Tuteja, N., and Gill, S. (2013). Climate Change and Plant Abiotic Stress Tolerance. Hoboken: John Wiley & Sons. doi: 10.1002/9783527675265 Vafa, N., Sohrabi, Y., Sayyed, R. Z., Suriani, N. L., and Datta, R. (2021). Effects of combinations of Rhizobacteria, mycorrhizae, and seaweeds on growth and yields in wheat cultivars under the influence of supplementary irrigation. *Plants* 10:811. doi: 10.3390/plants10040811

Vance, E. D., Brookes, P. C., and Jenkinson, D. S. (1987). An extraction method for measuring soil microbial biomass C. Soil Biol. Biochem. 19, 703–707. doi: 10.1016/0038-0717(87)90052-6

Wang, C., Li, X., Zhou, J., Wang, G., and Dong, Y. (2008). Effects of arbuscular mycorrhizal fungi on growth and yield of cucumber plants. *Commun. Soil Sci. Plant Anal.* 39, 499–509. doi: 10.1080/00103620701826738

Yücel, D. Ö, Anlarsal, A. E., Mart, D., and Yücel, C. (2010). Effects of drought stress on early seedling growth of chickpea (*Cicer arietinum* L.) genotypes. *World Appl. Sci. J.* 11, 478–485.

Zhao, R., Guo, W., Bi, N., Guo, J., Wang, L., Zhao, J., et al. (2015). Arbuscular mycorrhizal fungi affect the growth, nutrient uptake and water status of maize (*Zea mays* L.) grown in two types of coal mine spoils under drought stress. *Appl. Soil Ecol.* 88, 41–49. doi: 10.1016/j.apsoil.2014. 11.016