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October 06-29

THE ULTIMATE WEBINAR SERIES IN GENE EXPRESSION STUDIES







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Saliva-based testing for diagnosis of SARS-CoV-2 infection: A meta-analysis.

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Conflicts of Interest: No conflicts of interest

Keywords: saliva specimen, SARS-CoV-2, nucleic acid amplification, meta-analysis

To the Editor,

Diagnostic testing of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is primarily conducted from an upper respiratory specimen, like nasopharyngeal swab (NPS) or oropharyngeal swab (OPS) obtained by the health-care personnel. Recent systematic reviews concluded that non-invasive saliva might serve as an alternative specimen type to NPS, but evidence was still limited (1, 2). These systematic reviews included studies with confirmed coronavirus disease (COVID-19) patients, therefore

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1002/jmv.26613.

addressing different population and context (i.e. utility of test during some follow-up) compared to testing patients without COVID-19.

The Food and Drug Administration has authorized at least six saliva-based tests for emergency use for SARS-CoV-2 diagnosis (3). Nonetheless, the World Health Organization does not currently recommend saliva specimen for routine diagnostic use (4).

We conducted a meta-analysis of diagnostic accuracy studies that compared salivabased index test to NPS or OPS based reference test in patients without confirmed SARS-CoV-2 at enrollment. PubMed was used (latest search as of September 15) with a search strategy: *saliva and diagnosis and (COVID-19 or COVID 19 or SARS-COV-2 or SARS-COV 2)*. In addition, medRxiv and bioRxiv were used, and references checked from published studies on the topic. Studies with less than 50 patients, studies with patients with confirmed SARS-CoV-2 infection, and studies without relevant diagnostic data for sensitivity and specificity were excluded. Stata version 16.0 with a userwritten module (5) was used for bivariate modeling to obtain average sensitivity and specificity with 95% confidence interval (CI).

Fourteen studies (16 cohorts) were included (6-19). A total number of patients for sensitivity and specificity comparison was 5863 (median 158, interquartile range 94 to 234) (Supplemental Table 1). Mean or median age of patients ranged from 33.5 to 44.9 years in six studies (seven cohorts) where information was available, and three studies included some pediatric patients. A consecutive enrollment of patients was mentioned in one study and no patients were excluded from one study's other cohort. One study estimated specificity of saliva-based test in a subsample of patients, which excluded most of the test negative patients.

Nine studies reported polymerase chain reaction (PCR) cycle threshold (Ct) for positivity, and it varied from 37 to 45. Out of 10 studies that reported specific SARS-CoV-2 target genes, four focused on single E or N target genes.

A prevalence of SARS-CoV-2 infection varied from 0.3% to 78.4% (median 20.8%, interquartile range 7.1% to 37.2%) with NPS or OPS across the 16 cohorts. Average sensitivity was 0.85 (95% CI 0.77 to 0.91) and average specificity 0.99 (95% CI 0.98 to 1.00) with saliva-based index test compared to NPS or OPS based reference test (Figure 1). Positive and negative likelihood ratio was 90 (95% CI 35 to 234) and 0.15 (95% CI 0.10 to 0.23), respectively. Between-study heterogeneity is visualized in Supplemental Figure 1.
Two studies (two cohorts) (13, 19) were outliers on model check, and exclusion of these studies did not markedly affect average sensitivity and specificity, as did not

when Ct for positivity was adjusted with meta-regression.

Our meta-analysis shows that saliva-based nucleic acid amplification tests have lower sensitivity and comparable specificity in diagnostic testing of SARS-CoV-2 infection among nearly 5900 asymptomatic or symptomatic patients without confirmed COVID-19 diagnosis at study enrollment, which reflects a typical testing setting in general population.

Some reasons, like lower viral loads in saliva specimens, might explain why saliva is an inferior specimen type compared to swab specimen for the detection of SARS-COV-2. Studies included in our analyses tended to report higher Ct values (suggesting lower viral loads) for saliva specimens although not systematically evaluated.

Czumbel and coworkers highlighted conditions to be standardized for optimal salivabased testing, such as sample collection, transportation and test methods (1). Nonetheless, saliva-based testing can offer benefits, including no need of health-care workers for sample collection. According to one study, only a small proportion of selfcollected saliva samples were deemed unsuitable (20).

Our meta-analysis has limitations. First, one author conducted search and extracted data from the included studies. Second, no risk of bias assessment was done. Third, average sensitivity and specificity warrants cautious interpretation due to unclear or varying thresholds for positivity.

In conclusion, saliva-based test appears to be a less sensitive specimen type compared to upper respiratory specimens for SARS-CoV-2 nucleic acid amplification. This highlights the need for proper validation of diagnostic tests that rely on specimen types less frequently deployed. Saliva-based tests might require fewer resources, less technical expertise, and cause less discomfort to patients compared upper respiratory specimens. Thus, the inferior performance of saliva-based tests may be overcome by their better utility in specific settings.

References

1. Czumbel LM, Kiss S, Farkas N, et al. Saliva as a Candidate for COVID-19 Diagnostic Testing: A Meta-Analysis. Front Med 2020;7:465.

2. Peeters E, Singh SKDA, Vandesompele J, et al. Rapid systematic review of the sensitivity of SARS-CoV-2 molecular testing on saliva compared to nasopharyngeal swabs. medRxiv 2020 https://doi.org/10.1101/2020.08.05.20168716.

3. Ali F, Sweeney DA. No One Likes a Stick up Their Nose: Making the Case for Saliva-Based Testing for COVID-19. Clin Infect Dis 2020 Sep 2;ciaa1314. doi: 10.1093/cid/ciaa1314

4. https://www.who.int/publications/i/item/diagnostic-testing-for-sars-cov-2, accessed September 26, 2020

5. Dwamena B. MIDAS: Stata module for meta-analytical integration of diagnostic test accuracy studies. https://ideas.repec.org/c/boc/bocode/s456880.html (accessed September 26, 2020)

6. Becker D, Sandoval E, Amin A, et al. Saliva is less sensitive than nasopharyngeal swabs for COVID-19 detection in the community setting. medrxiv 2020 https://doi.org/10.1101/2020.05.11.20092338

7. Bhattacharya D, Parai D, Rout UK, et al. Saliva as a potential clinical specimen for
diagnosisofSARS-CoV-2.medRxiv2020doi:https://doi.org/10.1101/2020.09.11.20192591

8. Byrne RL, Kay GA, Kontogianni K, et al. Saliva offers a sensitive, specific and noninvasive alternative to upper respiratory swabs for SARS-CoV-2 diagnosis. medRxiv 2020 https://doi.org/10.1101/2020.07.09.20149534

9. Caulley L, Corsten M, Eapen L, et al. Salivary Detection of COVID-19. Ann Intern Med 2020 Aug 28. doi: 10.7326/M20-4738

10. Griesemer SB, Van Slyke G, Ehrbar D, et al. Evaluation of specimen types and saliva stabilization solutions for SARS-CoV-2 testing. medrxiv 2020 https://doi.org/10.1101/2020.06.16.20133041

11. Hanson KE, Barker AP, Hillyard DR, et al. Self-Collected Anterior Nasal and Saliva Specimens versus Healthcare Worker-Collected Nasopharyngeal Swabs for the Molecular Detection of SARS-CoV-2. J Clin Microbiol 2020 Aug 12:JCM.01824-20. doi: 10.1128/JCM.01824-20

12. Landry ML, Criscuolo J, Peaper DR. Challenges in use of saliva for detection of SARS CoV-2 RNA in symptomatic outpatients. J Clin Virol 2020;130:104567. doi: 10.1016/j.jcv.2020.104567

13. Moreno-Contreras J, Espinoza MA, Sandoval-Jaime C, et al. Saliva sampling is an excellent option to increase the number of SARS CoV2 diagnostic tests in settings with supply shortages. J Clin Microbiol 2020 Jul 23:JCM.01659-20. doi: 10.1128/JCM.01659-20

14. Pasomsub E, Watcharananan SP, Boonyawat K, et al. Saliva sample as a noninvasive specimen for the diagnosis of coronavirus disease 2019: a cross-sectional study. Clin Microbiol Infect 2020 May 15:S1198-743X(20)30278-0. doi: 10.1016/j.cmi.2020.05.001 15. Rutgers Clinical Genomics Laboratory. Accelerated emergency use authorization (EUA) summary SARS-CoV-2 assay. https://www.fda.gov/media/136875/download (accessed Sep 3 2020).

16. Skolimowska K, Rayment M, Jones R, et al. Non-invasive saliva specimens for the diagnosis of COVID-19: caution in mild outpatient cohorts with low prevalence. Clin Microbiol Infect 2020 Jul 18:S1198-743X(20)30421-3. doi: 10.1016/j.cmi.2020.07.015

17. Vaz SN, Santana DS, Netto EM, et al. Saliva is a reliable, non-invasive specimen for SARS-CoV-2 detection. Braz J Infect Dis 2020 Aug 31:S1413-8670(20)30115-X. doi: 10.1016/j.bjid.2020.08.001

18. Williams E, Bond K, Zhang B, Putland M, Williamson DA. Saliva as a Noninvasive Specimen for Detection of SARS-CoV-2. J Clin Microbiol 2020;58:e00776-20. doi: 10.1128/JCM.00776-20

19. Yokota I, Shane PY, Okada K, et al. Mass screening of asymptomatic persons forSARS-CoV-2usingsaliva.medrxiv2020https://doi.org/10.1101/2020.08.13.20174078

20. Guest JL, Sullivan PS, Valentin-Graves M, et al. Suitability and Sufficiency of Telehealth Clinician-Observed, Participant-Collected Samples for SARS-CoV-2 Testing: The

iCollect Cohort Pilot Study. JMIR Public Health Surveill 2020;6:e19731.

Figure Legends

Figure 1. Sensitivity and specificity of saliva-based index test compared to nasopharyngeal or oropharyngeal swab-based reference test for diagnostic testing of SARS-CoV-2 infection with nucleic acid amplification, based on 14 studies (16 cohorts).

STUDY		SENSITIVITY (95% CI)	STUDY		SPECIFICITY (95% CI)
Becker		0.40 (0.12 - 0.74)	Becker		0.98 (0.89 - 1.00)
Bhattacharya	–	0.91 (0.81 - 0.97)	Bhattacharya	— •	1.00 (0.79 - 1.00)
Byrne	- +	0.86 (0.57 - 0.98)	Byrne	÷	1.00 (0.96 - 1.00)
Caulley		0.61 (0.47 - 0.74)	Caulley	, in the second se	0.99 (0.99 - 1.00)
Griesemer (Albany)	_∎ _¦	0.50 (0.21 - 0.79)	Griesemer (Albany)	ļ.	1.00 (0.98 - 1.00)
Griesemer (New Rochelle)	,	0.87 (0.78 - 0.93)	Griesemer (New Rochelle)	4	0.99 (0.95 - 1.00)
Hanson	—	0.94 (0.86 - 0.98)	Hanson	,	0.98 (0.95 - 0.99)
Landry	-#	0.85 (0.68 - 0.95)	Landry	-#	0.98 (0.92 - 1.00)
Moreno-Contreras	-	0.79 (0.65 - 0.89)	Moreno-Contreras	-	0.78 (0.70 - 0.85)
Pasomsub	-#	0.84 (0.60 - 0.97)	Pasomsub		0.99 (0.96 - 1.00)
Rutgers		1.00 (0.88 - 1.00)	Rutgers		1.00 (0.88 - 1.00)
Skolimowska	-	0.83 (0.59 - 0.96)	Skolimowska	, in the second	0.99 (0.95 - 1.00)
Vaz		0.94 (0.86 - 0.98)	Vaz	-	0.98 (0.92 - 1.00)
Williams	÷	0.85 (0.69 - 0.94)	Williams		0.98 (0.89 - 1.00)
Yokota (airport)	_	0.80 (0.28 - 0.99)	Yokota (airport)	į.	1.00 (1.00 - 1.00)
Yokota (contact)	i i	0.93 (0.80 - 0.98)	Yokota (contact)	-	0.95 (0.89 - 0.98)
OVERALL	ò	0.85 (0.77 - 0.91)	OVERALL	å	0.99 (0.98 - 1.00)