

Uniform bathymetric zonation of marine benthos on a Pan-Arctic scale

Vedenin A.A.¹, Mironov A.N.¹, Bluhm B.A.², Käß M.^{3,4}, Degen R.⁵, Galkin S.V.¹, Gebruk A.V.¹

¹ – Shirshov Institute of Oceanology, Russian Academy of Sciences

² – UiT, The Arctic University of Norway

³ – Alfred-Wegener-Institute for Polar and Marine Research

⁴ – University of Oldenburg

⁵ – University of Vienna

Corresponding author:

Vedenin A.A., urasterias@gmail.com

Abstract

While numerous regional studies of bathymetric zonation of benthic fauna globally have been done, few large-scale analyses exist, and no ocean-scale studies have focused on the Arctic Ocean to date. In the present work we, hence, examined bathymetric zonation of macro- and megabenthos over a depth range spanning from the shelf to the abyssal plain (14 – 5416 m) and regionally extending from the Fram Strait to the Beaufort Sea (as a whole hereafter called the Central Arctic). Based on 104 quantitative (box-corers and grabs) and 37 semi-quantitative (trawls) samples compiled from different studies we evaluated bathymetric zonation patterns in abundance, biomass and diversity, and also compared species composition among samples. Abundance and biomass decreased with depth from >3000 ind. m^{-2} and >40 g ww m^{-2} to ~ 130 ind. m^{-2} and <1 g ww m^{-2} corroborating previous studies. Diversity showed a parabolic pattern, peaking at ~ 100 - 600 m. Cluster analysis revealed four (macrofauna) and five (megafauna) groups of benthic assemblages, including three that covered the upper and lower continental slope and the abyssal plains with relatively little overlap (named the *Lower Shelf – Upper Slope 1*, the *Lower Slope* and the *Abyss*). Substantial changes in benthic community composition were observed at depths 650-950 m (between the *Lower Shelf – Upper Slope 1* and the *Lower Slope*) and 2600-3000 m (between the *Lower Slope* and the *Abyss*), so we interpreted these two depth horizons as major bathymetric boundaries. The first boundary (650-950 m) corresponds to the transition from sublittoral to bathyal fauna consistent with previous studies. The second boundary (2600-3000 m) reflects a decrease in benthic abundance, biomass and diversity within the Central Arctic abyssal plain. Bathymetric patterns and species overturn of benthos were relatively uniform throughout the entire Central Arctic continental slope and abyssal plain. For some regions of the Arctic Ocean, foremost for the area north from Greenland and Canadian Archipelago, benthic data are still unavailable and further research is needed.

Key Words: Central Arctic; Bathymetric zonation; Benthic assemblages; Biogeography

1. Introduction

Multiple lines of evidence suggest we can expect that on-going climate change and continuous decline of sea ice cover will lead to shifts in the Arctic marine ecosystem, eventually affecting benthic biota (Wassmann et al., 2011; Jansen et al., 2020). In particular, changes in species distributions, benthic abundance and biomass are expected due to sea ice losses followed by regionally enhanced algal blooms and subsequent increase of carbon uptake and storage (Meredith et al., 2019). To better understand possible changes, and perhaps predict them, a solid understanding of large-scale *current* patterns, derived from comparable and standardized data on benthic fauna, across all regions of the Arctic Ocean are required.

Bathymetric distribution of macro- and megabenthic communities was addressed by numerous studies worldwide (Hedgpeth, 1957; Grassle & Morse-Porteous, 1987; Golikov et al., 1990; Rex & Etter, 2010; Watling et al., 2013), but most consider small-scale basins or regions on scales of hundreds of kilometers at best. Past studies have examined depth-associated changes for specific seas (for example, Denisenko et al., 2003; Budaeva et al., 2008; Käß et al., 2019; Vedenin et al., 2015; Ravelo et al., 2020). Few investigations included the deep Central Arctic Ocean (e.g. Kröncke, 1994; Kröncke, 1998; Deubel, 2000), a still poorly sampled area. From the spotty data Sirenko et al. (1998) produced a schematic map of the types of benthic communities replacing one another with depth from coastal regions to the base of the continental slope along a wide band of Eurasian Arctic from the Barents Sea to the Chukchi Sea. More recently, Piepenburg et al. (2011) and Bluhm et al. (2011; 2020) demonstrated some degree of uniformity of bathymetric patterns of benthos distribution around the entire Arctic Ocean.

To date, however, few studies have attempted an integration of benthic fauna data from different Arctic regions and those that have gone beyond a single region showed somewhat differing results or focused on one benthic size fraction only. For example, from the Barents Sea to the Laptev Sea macrobenthic communities replaced one another relatively uniformly with depth in five main types of benthic communities corresponding to the shelf, upper slope, mid-slope, lower slope and abyssal plain (Vedenin et al., 2018). These same authors reported a peak in species richness at a depth of 100-300 m, which is shallower than the bathymetric richness peak reported in other bathymetric studies (e.g. Wlodarska-Kowalczyk et al., 2004 and a meta-analysis by Bluhm et al., 2011). Combining data from different regions processed by different authors into one data set can introduce bias owing to differences in sampling methods, protocols and taxonomy, as was summarized in Bluhm et al. (2011) and Vedenin et al. (2018). Yet, such integration must be the goal if we are to attempt to provide a comparable understanding of benthic bathymetric zonation based on quantitative taxonomical structure at the scale of the Arctic Ocean.

In this study, despite the difficulties mentioned, we pooled different data sets on benthic communities in the Arctic Ocean to analyze patterns of bathymetric distribution over a broad depth range

73 (from 14 to 5416 m). We hypothesize that the major patterns of bathymetric distributions of macro- and
74 megabenthos remain relatively constant across all regions of the Central Arctic Ocean. We use the
75 approximate position of bathymetric biogeographic boundaries as well as bathymetric trends in
76 abundance, biomass, and diversity as metrics. Based on earlier findings we also hypothesize that
77 bathymetric patterns in diversity differ from known patterns in other areas of the World Ocean. Our
78 geographic scope is large-scale- including the Barents, Kara and Greenland Seas, the Yermak Plateau, the
79 Beaufort Sea, and the entire Eurasian deep-sea basins.

80

Materials & Methods

2.1. Study Area

We focused on an extensive area of the Arctic Ocean extending across the continental shelf, continental slope, and abyssal plain. The sampled shelf areas vary in depth and size and include the broad and Barents Sea that extends down to ~500 m depth, the shallow and broad Kara and Laptev Seas, and the shallow and narrow Beaufort Sea. The continental slopes vary in steepness and the depth of the abyssal plains ranges from ~2500 to 4500 m. Several basins and a series of ridges, including the Gakkel, Lomonosov, Mendeleev and Alpha Ridges divide the abyssal Arctic Ocean north of Eurasia from that off of North America. The shallow waters of the focal areas differ markedly from one another, mainly because of differences in river run-off, which is especially prominent on the Siberian (Kara and Laptev Seas) shelf and in the Beaufort Sea. At ~100-700 m, warmer and more saline waters of Atlantic origin flow through the eastern Fram Strait into the central Arctic Ocean, thereby warming the water masses along the slopes of the Eurasian Arctic (Wassmann et al., 2019). The Atlantic layer gradually decreases in temperature as it travels eastward and into the Amerasian Basin (Bluhm et al., 2020). Below the Atlantic layer, cold Arctic deep-sea waters straddle the entire Arctic Ocean at depths ~>700-800 m (Bluhm et al., 2020; see Meltofte, 2013 for detailed hydrology of the Arctic Ocean). Sea ice covers vast areas of the deep-sea Arctic temporarily or permanently although mean sea ice extent is decreasing rapidly (Wassmann et al., 2011; Jansen et al., 2020).

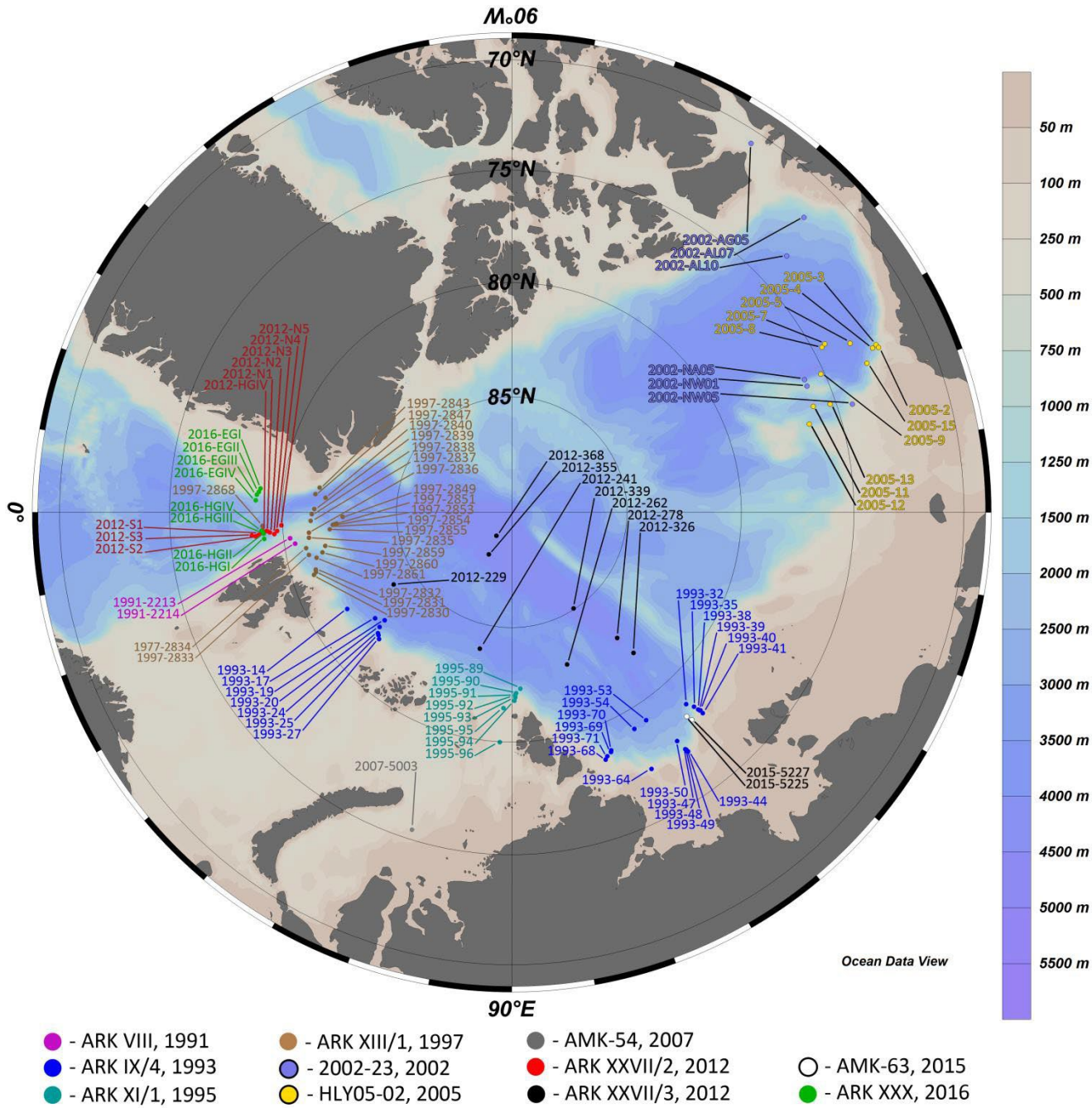
In particular, our focal areas include Fram Strait, the Yermak Plateau, the Northern Barents Sea slope, the Kara Sea, the Laptev Sea, the Beaufort Sea and adjacent Canada Basin and the deep-sea Nansen and Amundsen basins (Fig. 1, 2). Water depths range from 10s of meters in the Kara and Laptev Seas to over 5000 meters (Molloy Deep in the Fram Strait as our deepest point at 5416 meters). Hereafter we refer to this focal area as the Central Arctic.

2.2. Material processing

The total pool of invertebrate macro- and megafauna samples included 141 stations taken during 13 expeditions. We divided all samples into quantitative and semi-quantitative based on the sampling gear used. Overall we obtained 106 quantitative stations using grabs and corers: the 'Okean' grab (0.1 or 0.25 m² sampling area), USNEL Box-corer and Multibox-corer (Lisitsyn & Udintsev, 1955; Hessler & Jumars, 1974; Gerdes, 1990) (Fig. 1). The sampling area per station varied from 0.023 m² to 0.75 m² depending on the number of subsamples taken from each gear (Table S1), the mesh sizes varied from 0.2 to 0.5 mm. The final quantitative dataset was compiled from 11 expeditions: the 1991 expedition, where extensive box-corer sampling was surveyed in the Central Arctic Ocean across the Nansen and Amundsen Basins (Kröncke, 1994; Kröncke, 1998); the 1995 expedition, where multiple box-corer transects were collected and published for the continental slope in the Kara and Laptev seas (Anisimova et al., 2003; Sirenko et

116
117
118
119
120
121
122
123

al., 2004); the 1997 expedition to the Yermak Plateau with box-corers presented by [Degen et al. \(2014\)](#); the 2002 and 2005 expeditions to the Beaufort Sea slope and abyssal plain, the results of macrobenthic surveys using box corers were published by [Bluhm et al. \(2005\)](#) and [MacDonald et al. \(2011\)](#); the 2007 and 2015 expeditions where a series of grab samples on the shelf and slope of the Kara and Laptev Seas was obtained ([Vedenin et al. 2015; 2018](#)); the 1993 and 2012 surveys in the Central Arctic were performed using Multibox-corer ([Vedenin et al., 2018](#)). the 2012 and 2016 surveys of macrobenthos using box-corers at the Long-Term Ecological Research observatory “HAUSGARTEN” in the Fram Strait ([Vedenin et al., 2016; Käß et al., 2019](#)).



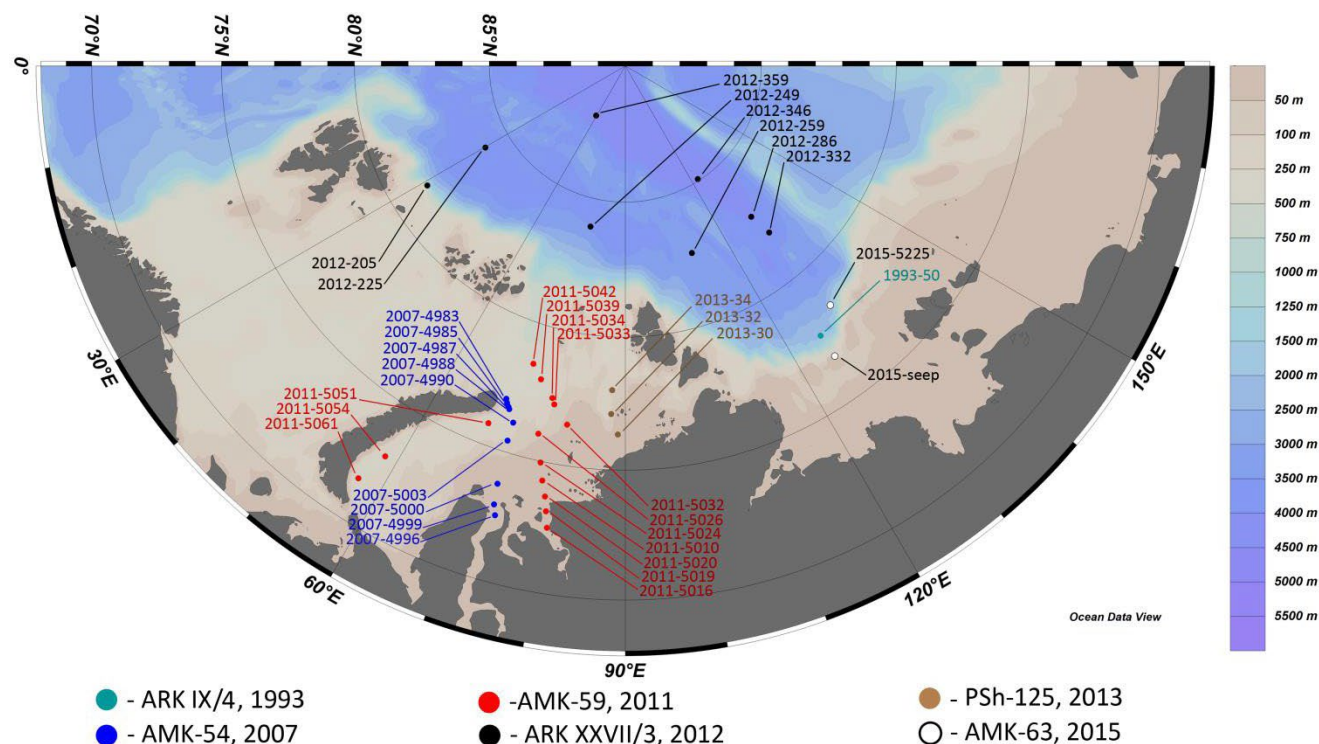
124
125
126
127

Fig. 1. Study area indicating the 104 quantitative stations across the Arctic. Specific expeditions are marked by colour. ARK – RV ‘Polarstern’; 2002-23 – CCGS ‘Louis S. St. Laurent’; HLY05-02 – USCGC ‘Healy’; AMK – RV ‘Akademik Mstislav Keldysh’.

128
129
130
131
132
133
134
135
136
137
138
139
140
141
142

The above listed publications provide species lists with abundance and biomass data (wet weight) standardized to square meter from these stations. The original samples were re-sorted and re-identified by a single pool of experts [except for samples in [Anisimova et al. \(2003\)](#), [Bluhm et al. \(2005\)](#) and [Macdonald et al. \(2010\)](#)] to avoid bias due to possible misidentification of benthic organisms. All taxa names were verified with use of the World Register of Marine Species database <http://marinespecies.org/>.

In addition, we obtained material from 37 semi-quantitative samples using Sigsbee and Agassiz trawls ([Eleftheriou & McIntyre, 2005](#)) (Fig. 2), samples were sieved through 1 mm mesh sized sieve, both total number of individuals and percentage values of abundance for each taxon was calculated. The expeditions included trawl surveys in the Kara and Laptev seas shelf and adjacent areas of the deep-sea Central Arctic; again, species lists from trawl samples were previously published ([Kim et al., 2006](#); [Vedenin et al., 2015](#); [Vedenin et al., 2018](#); [Rybakova et al., 2019](#); [Vedenin et al., 2020](#); [Udalov et al., 2020](#)). A complete list of all stations with the expedition name, year, coordinates, depth and sampling area is given in Appendix Table S1.



143
144
145
146
147
148
149
150
151

Fig. 2. Study area indicating the 37 semi-quantitative stations across the Eurasian Arctic. Specific expeditions are marked by colour. ARK – RV ‘Polarstern’; AMK – RV ‘Akademik Mstislav Keldysh’; PSh – RV ‘Professor Shtokman’.

2.3. Statistics

We used abundance, biomass and diversity indices calculated for quantitative samples as summary metrics for each station. For semi-quantitative (trawl) samples we calculated diversity indices only because it is impossible to normalize the abundance and biomass per square area ([Eleftheriou & McIntyre, 2005](#)).

Diversity metrics included number of taxa per sample, Shannon index (Shannon & Weaver, 1963), Pielou's evenness (Pielou, 1966; McCune et al., 2002) and Hurlbert rarefaction per 100 individuals (ES-100) (Hurlbert, 1971) which is suited to compare different sample sizes (Magurran, 2004). In addition, because of the low macrobenthic abundance in some samples, we extrapolated diversity using Hill numbers ($q = 0$) for 50 individuals (further referred to as 'Hill 50 extrapolated'). Chao et al. (2014) introduced this method and provided the detailed algorithms for extrapolation (see Appendix Table S2). We calculated Hill numbers using original non-transformed number of individuals in each sample.

We estimated similarity between all quantitative and (separately) among all trawl samples using Bray-Curtis similarity based on sample abundances (macrofauna) and number of individuals in trawl haul (megafauna):

$$C_{BC} = 200 \frac{\sum_1^{S_{12}} \min \{x_i, y_i\}}{\sum_1^{S_1} x_i + \sum_1^{S_2} y_i},$$

where C_{BC} is the similarity value, x_i and y_i are the abundances of i -th species in sample 1 and 2, S_1 and S_2 are the species richness of sample 1 and 2 and $\min \{x_i, y_i\}$ is the smaller abundance value of i -th species shared for samples 1 and 2 (Bray & Curtis, 1957; Clarke et al., 1996). Noting the sensitivity of Bray-Curtis to sample size differences (Chao et al., 2006) we also employed two other indices – the Morisita-Horn similarity index:

$$C_{MH} = 2 \frac{\sum_1^{S_{12}} \left(\frac{x_i y_i}{S_1 S_2} \right)}{\sum_1^{S_1} \left(\frac{x_i}{S_1} \right)^2 + \sum_1^{S_2} \left(\frac{y_i}{S_2} \right)^2},$$

and the quantitative Sørensen similarity index:

$$C_S = \frac{2UV}{U + V},$$

Where U and V are total relative abundances of those species in sample 1 and, respectively, sample 2, that are shared, i.e. present in both samples (Horn 1966; Chao et al., 2006).

Quantitative samples were analyzed untransformed and square-root transformed depending on specific metric used (Bray-Curtis similarity was calculated based on the untransformed data to accentuate the quantitative differences in species composition); semi-quantitative samples (data on simple number of individuals of each taxon) were fourth-root transformed to reduce the dominant taxa bias, as one-two taxa often contribute over 50% of the total abundance in the trawl samples (Galkin & Vedenin, 2015).

Hierarchical clusters were generated using the UPGMA-algorithm, i.e. group-average (Clarke et al., 1996). The similarity profile routine (SIMPROF) was used at the significance level of 0.01, to test for statistical significance of clusters (Clarke & Warwick, 2001). We identified different benthic assemblages based on quantitative faunal similarity in the dendrogram by slicing above the statistically insignificant nodes outlined with the SIMPROF routine. Identified assemblages were tested with Permutational multivariate analysis of variance (PERMANOVA) (Andersen, 2005). Hereafter we avoid the term

184 ‘benthic community’ because of the debate over the specific definition of this term, and replace it with
185 ‘benthic assemblage’. Non-parametric Kruskal-Wallis tests followed by Dunn’s post-hoc tests verified
186 differences between identified assemblages for abundance, biomass and diversity (Marshall, 2019).
187 Similarity percentages analysis based on the Bray-Curtis dissimilarity (SIMPER) was used for identifying
188 character species contributing most to cluster differences (Clarke & Warwick, 1994). We then plotted the
189 identified benthic assemblages along the depth axis to visualize bathymetric structure within the study
190 area.

191 Statistical analysis used PAST 3.0, Primer V6, IBM SPSS statistics and Microsoft Excel 2010
192 software (Clarke & Warwick, 2001; Hammer et al., 2001; Marshall, 2019). Hill numbers, Morisita-Horn
193 and quantitative Sørensen similarity indices were calculated using original Python 3.8 scripts using
194 NumPy, Pandas, Scipy, Scikit-bio, Scikit-learn, Math, and Matplotlib libraries
195 (<https://www.python.org/downloads/release/python-380/>). The scripts are available in Appendix S3.
196 Maps were built using Ocean Data View software (Schlitzer, 2020).

3. Results

3.1. Quantitative samples

Abundance and biomass values varied greatly, from 12 ind. m⁻² (at st. 2012-229) to 8861 ind. m⁻² (st. 1993-31) and, from 0.004 g ww m⁻² (st. 2002-AL10) to 2316.500 g ww m⁻² (st. 1995-93), respectively. The number of species per station varied from 2 (at several stations, including sts. 1997-2833, 1997-2847, 1997-2849-7, 1997-2868 and 2012-229) to 120 (st. 1995-93), recognizing taxon numbers are affected by area sampled. Sampling area was smallest at st. 1993-20 (0.023 m²) and largest at sts. 2015-5225 and 2015-5227 (0.750 m²), see Table S1 for details. Smaller variation characterized diversity indices: ES-100 differed from 2 (same stations as for lowest number of species per station) to 44 (st. 1993-40); Hill 50 extrapolated numbers varied from 2 (st. 2012-229) to 30 (st. 1997-2837); Pielou's evenness varied from 0.48 (st. 2005-5) to 1 (st. 2012-229) and Shannon index varied from 0.58 (st. 1997-2868) to 3.93 (st. 1993-40). For details, see Appendix Table S5. A complete list of individual taxa abundances per station is given in Appendix Table S6.

Cluster analysis with SIMPROF test based on Bray-Curtis similarity identified four distinct clusters (= benthic assemblages) at 6% similarity, including all but two stations (Fig. 3). We identified the clusters as the *Lower Shelf - Upper Slope 1*, *Lower Shelf - Upper Slope 2*, *Lower Slope* and *Abyss* assemblages. *Lower Shelf - Upper Slope 2* was defined based on lower similarity level (4) due to taxonomical peculiarity (described in this subsection below) (Fig. 3). The names of the clusters were chosen according to the geographic and bathymetric position of the station, explained below (Fig. 4). PERMANOVA showed significant results with p -value < 0.001 (Fig. 3). Non-parametric Kruskal-Wallis test found significant differences between most clusters in terms of the community parameters, including abundance, biomass and diversity metrics (Table 1).

Cluster

- Lower Shelf - Upper Slope 1
- Lower Slope
- ◆ Abyss
- ▶ Lower Shelf - Upper Slope 2

PERMANOVA

Permutations: 1000
 Test statistics: 5.97
 p-value: $9.99 \cdot 10^{-4}$
 R²: 0.20

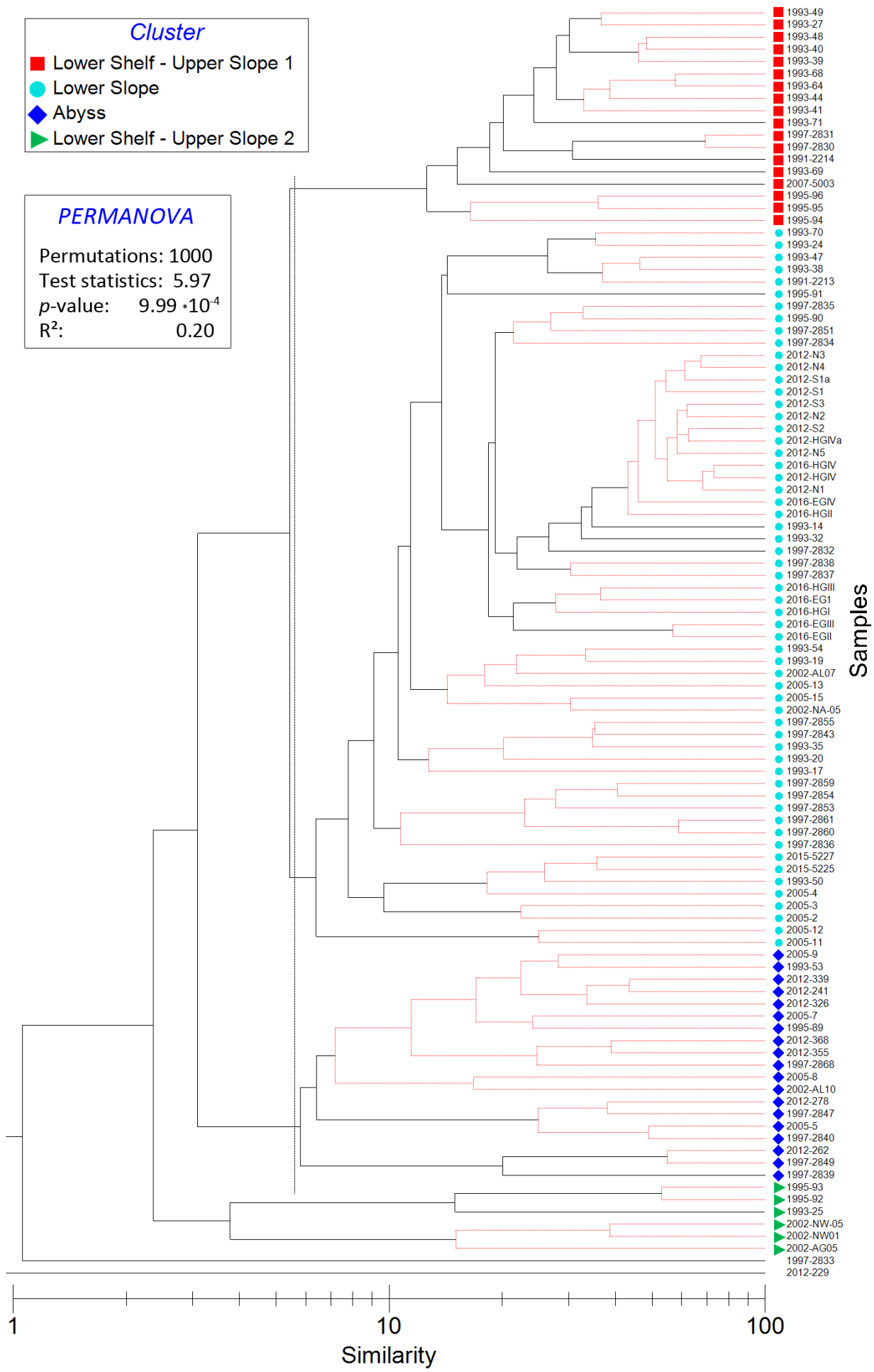


Fig. 3. Cluster analysis of quantitative stations using Bray-Curtis similarity index with SIMPROF results. Red lines indicate branches and nodes not statistically significant at $p < 0.01$. Marker colour and shape indicate benthic assemblages, defined at a similarity level of 6 percent; similarity is plotted on a logarithmic scale. Enclosed the PERMANOVA results are shown.

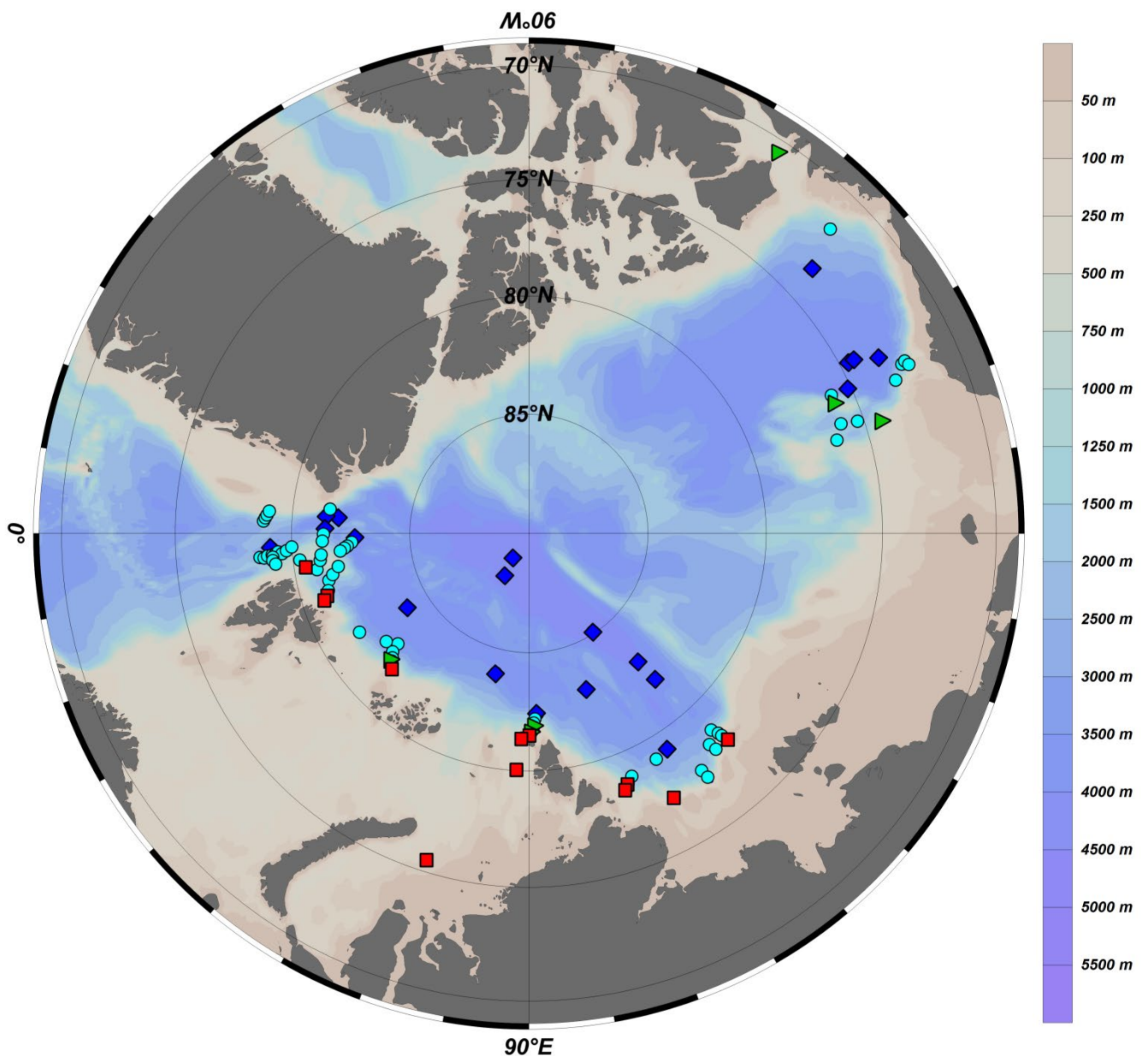
Morisita-Horn or Sørensen similarity indices yielded similar results, with the *Lower Shelf - Upper Slope 1* assemblage containing almost identical stations to the Bray-Curtis dendrogram. However, compared to the Bray-Curtis dendrogram, several stations shifted between the *Lower Slope* and *Abyss* assemblages, whereas the *Lower Shelf - Upper Slope 2* assemblage became indistinct (Supplementary 5, Fig. S-1, S-2). Given that all 3 indices yielded generally comparable results, and that some of the statistical methods work only with Bray-Curtis similarity, we based plots and figures in the main text on Bray-Curtis similarities.

The benthic assemblages were distributed within specific depth ranges, with three forming consistent depth bands that spanned the study area – the *Lower Shelf - Upper Slope 1*, the *Lower Slope* and the *Abyss* (Fig. 4).

Table 1. Mean values of benthic abundance, biomass and diversity of each assemblage and results of Kruskal-Wallis and Dunn's post-hoc tests for quantitative samples.

Group	Mean values \pm SD				Kruskal-Wallis		Dunn's post-hoc comparisons
	(1) <i>Lower Shelf - Upper Slope 1</i>	(2) <i>Lower Slope</i>	(3) <i>Abyss</i>	(4) <i>Lower Shelf - Upper Slope 2</i>	Chi square	p	
Quantitative samples							
Number of samples	18	59	19	6	-	-	-
Abundance (ind m ⁻²)	3845 \pm 2020	805 \pm 453	134 \pm 156	3694 \pm 2765	73.08	<0.001	1-2; 1-3; 1-4; 2-3; 2-4
Biomass (g ww m ⁻²)	44 \pm 75	4 \pm 5	0.8 \pm 1.1	633 \pm 901	57.63	<0.001	1-2; 1-3; 2-3; 2-4
Species number	70 \pm 23	28 \pm 20	8 \pm 6	57 \pm 45	51.15	<0.001	1-2; 1-3; 2-3; 3-4
ES-100	34 \pm 8	20 \pm 10	8 \pm 6	24 \pm 12	45.61	<0.001	1-2; 1-3; 2-4; 3-4
Hill 50 extrapolated	24 \pm 5	18 \pm 6	11 \pm 8	17 \pm 7	29.66	<0.001	1-2; 1-3; 1-4; 2-3

Numbers in Dunn's post-hoc comparisons column indicate corresponding group of benthic assemblages, revealed with Bray-Curtis similarity.



242 Fig. 4. Distribution of groups of benthic assemblages (color-shape-coded) based on quantitative samples
 243 (Bray-Curtis similarity). Colours and shapes of markers are the same as in Fig. 3.
 244
 245

246 Each of the identified assemblages was generally weakly structured within clusters on the
 247 dendrogram. One exception included station groups of high similarity ($> 50\%$) in the Fram Strait area
 248 (2012 and 2016 stations on Fig. 1, e.g. HGIV, S1-S3, N1-N5) within the *Lower Slope* assemblage. Second
 249 exception were the Beaufort Sea stations (2002 and 2005 stations on Fig. 1) grouped separately within the
 250 *Lower Shelf – Upper Slope 2* group and, to a lesser extent, within the *Lower Slope* group (Fig. 3).

251 The SIMPER analysis based on Bray-Curtis similarity identified the most characteristic species
 252 according to their mean contribution to differences between the assemblages (taxa with cumulative
 253 contribution up to 70% are shown, Table 2). In the *Lower Shelf - Upper Slope 1* group the bivalve *Yoldiella*
 254 *solidula* was the most characteristic species, followed by the polychaetes *Prionospio cirrifera*, *Spiophanes*
 255 *kroyeri* and *Tharyx* sp. The polychaetes *Galathowenia fragilis*, *Myriochele heeri*, *Chaetozone jubata* and

256 *Prionospio* sp. contributed most to the *Lower Slope* cluster. The polychaetes *Anobothrus laubieri*,
257 *Ophelina opisthobranchiata*, *Aricidea* spp. and *Tharyx* sp. and sponge *Thenia abyssorum* were primarily
258 responsible for separating the *Abyss* group. In the *Lower Shelf - Upper Slope 2* assemblage, the
259 polychaetes *Bushiella (Jugaria) similis*, sponges *Demospongiae* gen. sp. and the bryozoans *Tubulipora*
260 *fruticosa* were of primary importance (Table 2).

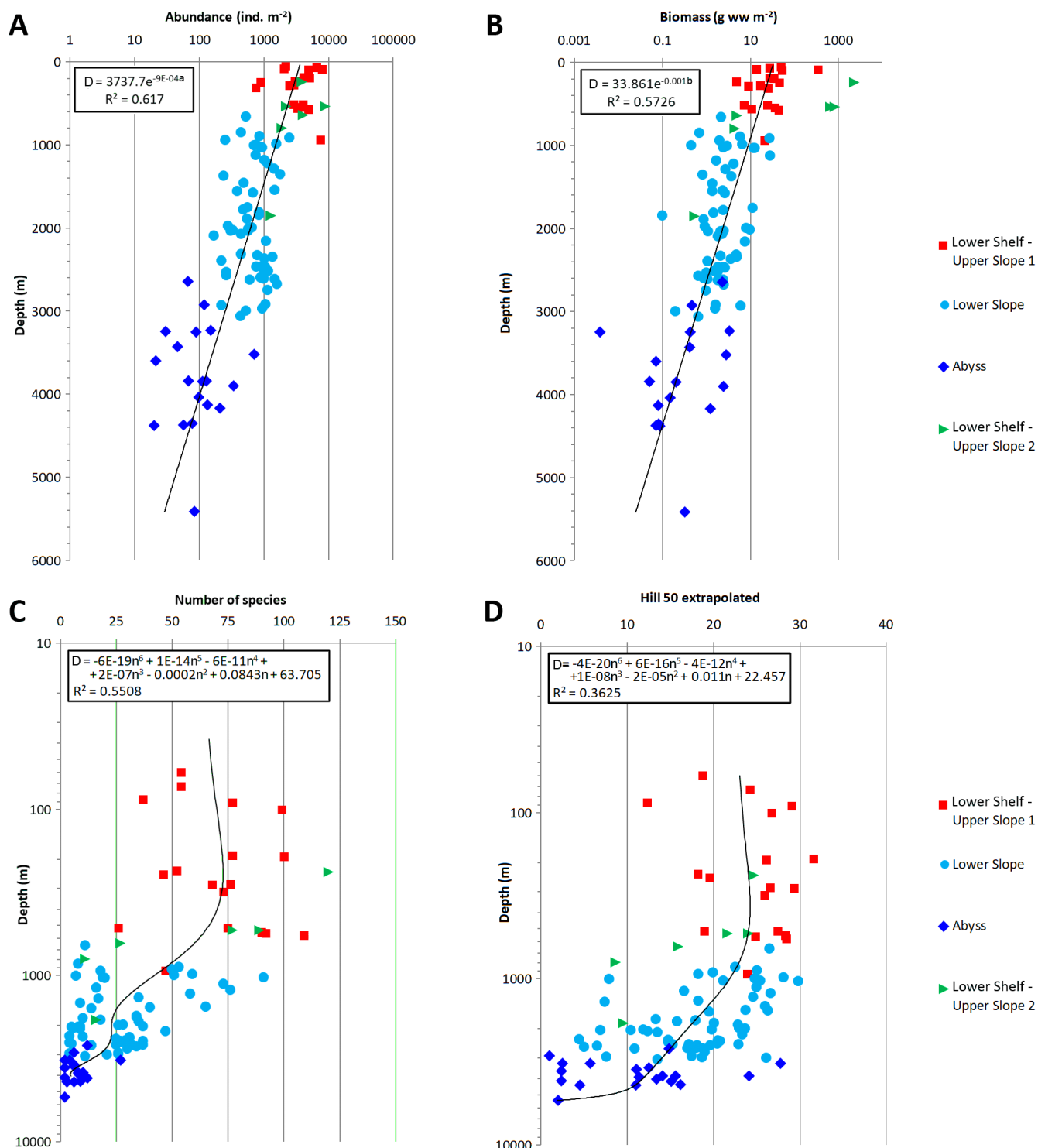
261 Changes in the summary metrics with depth indicated several patterns. Abundance and biomass
262 gradually decreased with depth (Fig. 5 a,b) except for samples from the Lower Shelf - Upper Slope 2
263 assemblage. Exponential trend lines produced the best approximations. The species number and
264 extrapolated Hill 50 values changed parabolically with maximum values in the ~100-500 m depth range
265 (Fig. 5 c,d). For diversity values, polynomial trend lines provided the best approximations given by the
266 equations and R² values (Fig. 5). The Hill 50 values of *Lower Slope* and *Abyss* stations decreased with
267 depth more slowly than other diversity measures due to extrapolation of diversity considering the
268 potentially unsampled taxa (Fig. 5 d, Fig. 6). The 'true' rarefactions (Fig. 6) are denoted with continuous
269 lines ending with large circles, with dashed lines showing the extrapolated rarefaction with larger values
270 of expected number of species per 50 individuals (Fig. 6).

272
273
274

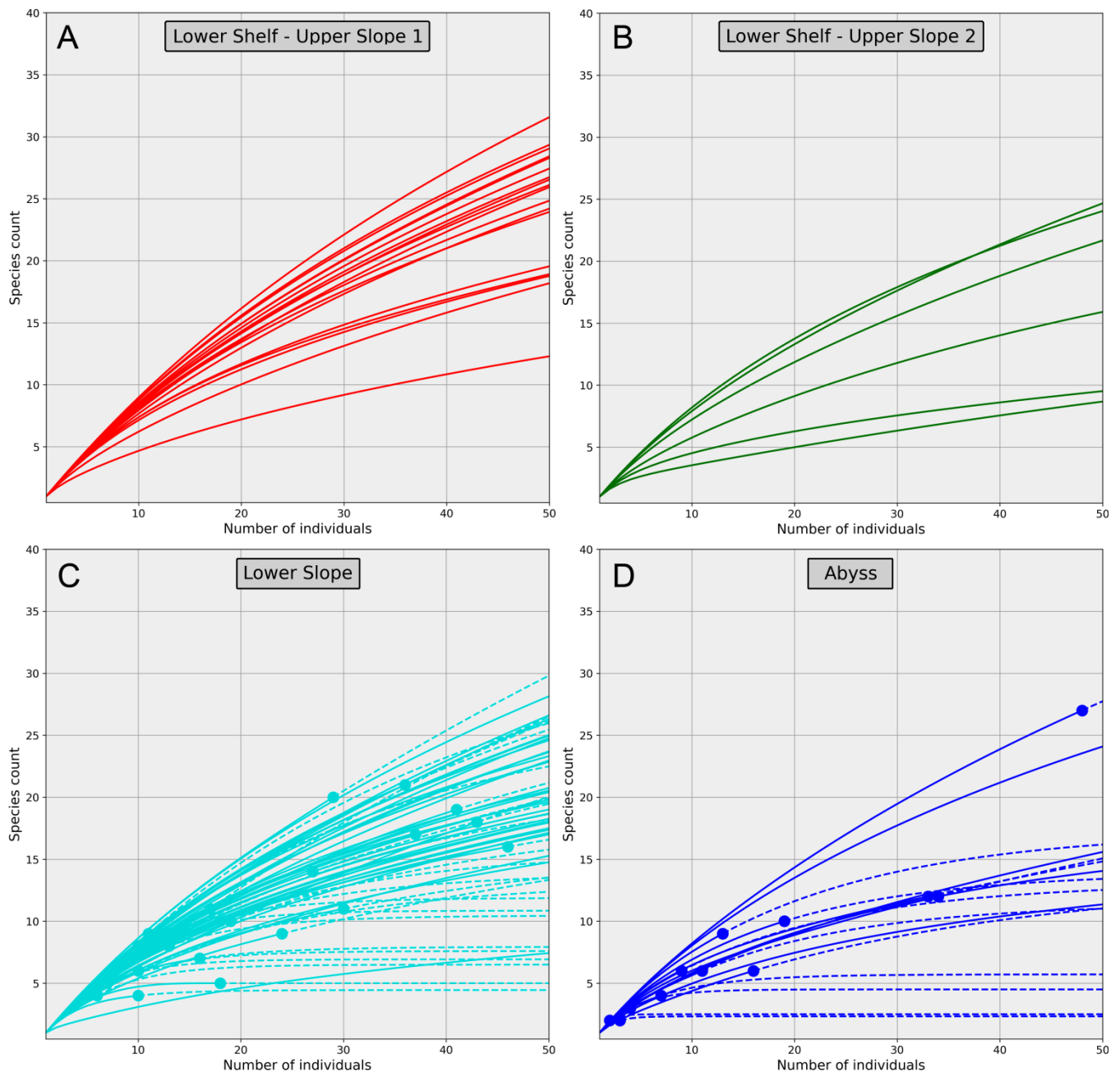
Table 2. Results of SIMPER analysis of quantitative samples with values of average abundance, average dissimilarity and percentage contribution of taxa within retrieved groups of assemblages individually and cumulatively. First several most significant taxa in each assemblage are marked in grey.

Group	Species	Average abundance (ind. m ⁻²)				Average Dissimilarity	Dissimilarity/SD	Contribution (%)	Cumulative (%)
		Lower Shelf - Upper Slope 1	Lower Slope	Abyss	Lower Shelf - Upper Slope 2				
Pol	<i>Bushiella (Jugaria) similis</i>	0	0	0	363.1	5.6	0.8	5.7	5.7
Pol	<i>Siboglinum hyperboreum</i>	0	12.3	1.1	338.9	4.5	0.5	4.6	11.4
Por	<i>Demospongiae</i> gen.sp.	0.1	0.4	3.6	203.1	2.7	0.5	2.8	16
Pol	<i>Prionospio cirrifera</i>	196.3	23	0	21.5	2.7	0.8	2.8	18.8
Cni	<i>Stephanoscyphus</i>	2	6.5	0	141.7	2.5	0.4	2.5	21.5
Pol	<i>Tharyx</i> sp.	173.4	26.6	4.3	15.3	2.4	1	2.5	24.1
Biv	<i>Yoldiella solidula</i>	255.6	0.7	0	3.3	2.2	0.6	2.2	26.6
Pol	<i>Chone duneri</i>	124.3	0.1	0	70.4	2	0.5	2.1	28.8
Pol	<i>Galathowenia fragilis</i>	0	65.9	0.8	0	1.8	0.5	1.8	30.8
Pol	<i>Polychaeta</i> gen.sp.	0.4	8	0	127.7	1.7	0.4	1.7	32.7
Pol	<i>Myriochele heeri</i>	61.7	43.9	0.3	0.3	1.6	0.6	1.6	34.4
Por	<i>Tetilla infrequens</i> (sensu Koltun)	0	0	0	114.3	1.4	0.7	1.5	36
Pol	<i>Anobothrus laubieri</i>	14.6	1.9	45.9	1.5	1.4	0.4	1.4	37.5
Pol	<i>Spiophanes kroeyeri</i>	137.4	0.4	0	0	1.2	0.4	1.2	38.9
Biv	<i>Medicula ferruginosa</i>	131.2	0.1	0	0	1.1	0.4	1.2	40.1
Pol	<i>Galathowenia oculata</i>	57.4	24.3	0.2	0.3	1.1	0.6	1.2	41.2
Sip	<i>Nephasoma</i> sp.	64.4	16.8	0	6.2	1.1	0.7	1.2	42.4
Cum	<i>Ectonodiastylis nimia</i>	89.7	0.5	0	0	1.1	0.3	1.1	43.5
Pol	<i>Chaetozone setosa</i>	94.4	0.5	0	29.2	1.1	0.7	1.1	44.6
Por	<i>Geodia</i> spp.	0	0	0	137	1.1	0.8	1.1	45.7
Pol	<i>Notoproctus oculatus</i>	89.6	1.1	0	22.9	1	0.6	1.1	46.8
Oph	<i>Ophiacantha bidentata</i>	9.7	0	0	77.5	1	0.7	1.1	47.9
Pol	<i>Pholoe</i> sp.	92.5	0.6	0	1.8	1	0.6	1	48.9
Pol	<i>Chaetozone jubata</i>	0	36.5	2.9	0	1	0.6	1	50
Bry	<i>Tubulipora fruticosa</i>	4.2	0.2	0	182.5	0.9	0.4	1	51
Pol	<i>Aricidea (Strelzovia) abbranchiata</i>	10.2	31	3.9	0	0.9	0.4	1	51.9
Pol	<i>Prionospio</i> sp.	1	34.1	0	0	0.9	0.4	0.9	52.9
Pol	<i>Proclea graffi</i>	83.6	0.8	0	0.3	0.9	0.6	0.9	53.8
Pol	<i>Melinnopsis arctica</i>	40.4	18.7	0	0	0.9	0.4	0.9	54.7
Nem	<i>Nemertea</i> gen.sp.	64.7	9.7	2.3	2.1	0.9	0.8	0.9	55.6
Pol	<i>Maldane arctica</i>	67.5	5.9	1.2	0	0.8	0.4	0.9	56.5
Amp	<i>Harpinia mucronata</i>	55.7	4.6	0.3	0	0.8	0.6	0.8	57.3
Por	<i>Sycon</i> sp.	17.5	4.3	0	96.9	0.8	0.4	0.8	58.1
Por	<i>Polymastiidae</i> gen.spp.	12.8	14.1	0	10.1	0.7	0.5	0.7	58.9
Pol	<i>Terebellides atlantis</i>	20.6	11.2	5.6	0	0.6	0.6	0.7	59.6
Pol	<i>Scoletoma fragilis</i>	76.1	0.5	0	0.7	0.6	0.4	0.7	60.3
Por	<i>Craniella cranium</i>	2.1	0	0	95	0.6	0.6	0.6	60.9
Pol	<i>Ophelina opisthobranchiata</i>	0	21.5	3.7	0	0.6	0.4	0.6	61.6
Bry	<i>Exidmonea atlantica disticha</i>	0	0	0	119.6	0.6	0.5	0.6	62.2
Oph	<i>Ophiocten sericeum</i>	61.9	0.2	0	12.8	0.6	0.5	0.6	62.8
Pol	<i>Apomatus globifer</i>	0	0.1	0	59.1	0.6	0.4	0.6	63.4
Bry	<i>Diplosolen intricarium</i>	0	0	0	95.2	0.6	0.6	0.6	64
Pol	<i>Aricidea nolani</i> (tax. inquirend.)	14	1.5	2.3	20.8	0.5	0.5	0.6	64.6
Pol	<i>Ophelina cylindrica data</i>	21.5	3.2	0	11.7	0.5	0.5	0.5	65.1
Tan	<i>Akanthophoreus gracilis</i>	59.3	1.3	0	4	0.5	0.6	0.5	65.7
Pol	<i>Euchone</i> sp.	23.3	1.7	0	44.6	0.5	0.6	0.5	66.2
Bry	<i>Cyclostomatida</i> gen.sp.	3.6	0.8	0.3	19.4	0.5	0.4	0.5	66.7
Biv	<i>Batharca frielei</i>	3.2	12.6	0	20.8	0.5	0.5	0.5	67.3
Por	<i>Thenea abyssorum</i>	0	8.7	8.6	0	0.5	0.5	0.5	67.8
Biv	<i>Yoldiella annenkovae</i>	8.9	15.5	0.8	0	0.5	0.4	0.5	68.3
Pol	<i>Clymenura polaris</i>	55.4	1.2	0	1.5	0.5	0.4	0.5	68.8
Pol	<i>Terebellides</i> sp.	17.6	0.4	0	12	0.5	0.6	0.5	69.3
Amp	<i>Byblis minuticornis</i>	41.4	1.4	0	0	0.5	0.3	0.5	69.8
Pol	<i>Praxillura longissima</i>	43.5	4.7	0	0	0.5	0.6	0.5	70.3

275 Taxa with cumulative contribution < 70% are shown. First several most significant taxa in each assemblage are marked
 276 in grey. Por – Porifera; Cni – Cnidaria; Pol – Polychaeta; Sip – Sipuncula; Nem – Nemertea; Biv – Bivalvia; Cum –
 277 Cumacea; Amp – Amphipoda; Tan – Tanaidacea; Bry – Bryozoa; Oph – Ophiuroidea.
 278
 279



280
 281 Fig. 5. Total abundance (ind. m⁻², a), biomass (g ww m⁻², b), species number (c) and Hill 50 extrapolated values (d) in
 282 quantitative Arctic benthic samples in relation to depth. Colour and shape of markers correspond to assemblages as in
 283 Figs. 3 and 4. Axes of abundance, biomass in upper plots and depth in lower plots are logarithmic. Trend line equations
 284 and R² values are shown in boxes (D – depth; a – abundance; b – biomass; n – species number or rarefaction per 50
 285 individuals).
 286

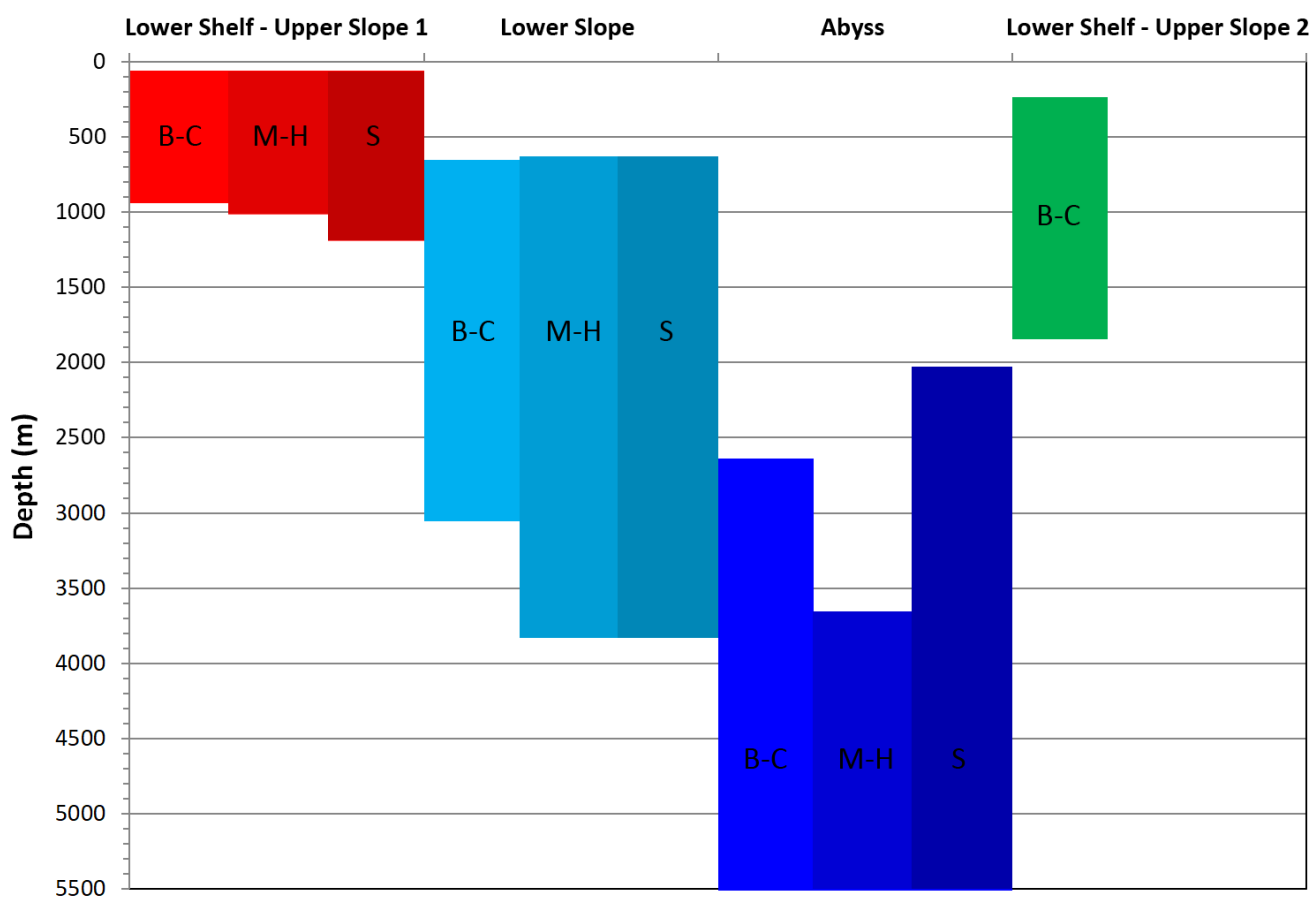


287
 288 Fig. 6. Rarefaction curves for the quantitative samples up to 50 individuals with extrapolation based on the Hill
 289 numbers ($q = 0$). A – *Lower Shelf - Upper Slope 1* assemblage; B – *Lower Shelf - Upper Slope 2* assemblage; C –
 290 *Lower Slope* assemblage; D – *Abyss* assemblage. Colour corresponds to assemblages as in Figs. 3-5. Continuous lines
 291 indicate true (sample-sized) rarefaction; circles indicate the end of sample in case of <50 ind m^{-2} ; dashed lines indicate
 292 the extrapolated rarefaction.

293
 294 Benthic assemblages largely replaced one another with depth except the *Lower Shelf - Upper Slope*
 295 *2* assemblage that sits within the depth range of the *Lower Shelf - Upper Slope 1* and *Lower Slope*
 296 assemblages (Fig. 7). The Bray-Curtis similarity dendrogram identified depth ranges of community
 297 clusters as follows: 60-942 m for the *Lower Shelf - Upper Slope 1* group; 239-1850 m for the *Lower Shelf*
 298 *- Upper Slope 2* group; 657-3054 m for the *Lower Slope* group and 2644-5416 m for the *Abyss* group (Fig.
 299 7). The depth ranges differed some for the Morisita-Horn index: 60-1026 m for the *Lower Shelf - Upper*
 300 *Slope 1* group; 640-3848 m for the *Lower Slope* group and 3677-5416 m for the *Abyss* group; the *Lower*

301 *Shelf - Upper Slope 2* group stations split into few clusters not forming a single assemblage (Fig 7,
 302 Supplementary 5, Fig. S-1). For the quantitative Sørensen similarity index the depth ranges were a bit
 303 more overlapping between assemblages: 60-1216 m for the *Lower Shelf - Upper Slope 1* group; 640-3848
 304 m for the *Lower Slope* group and 2027-5416 m for the *Abyss* group; the *Lower Shelf - Upper Slope 2*
 305 stations overlapped the *Lower Shelf - Upper Slope 1* group and the *Lower Slope* group (Fig 7,
 306 Supplementary 5, Fig. S-2).

307



308

309 Fig. 7. Bathymetric distribution of groups of Arctic benthic assemblages based on quantitative samples (three
 310 different similarity indices). Colours as in Figs. 3-5. B-C – depth ranges from the Bray-Curtis similarity
 311 dendrogram; M-H – depth ranges from the Morisita-Horn similarity dendrogram; S – depth ranges from the
 312 quantitative Sørensen similarity dendrogram.

313

314

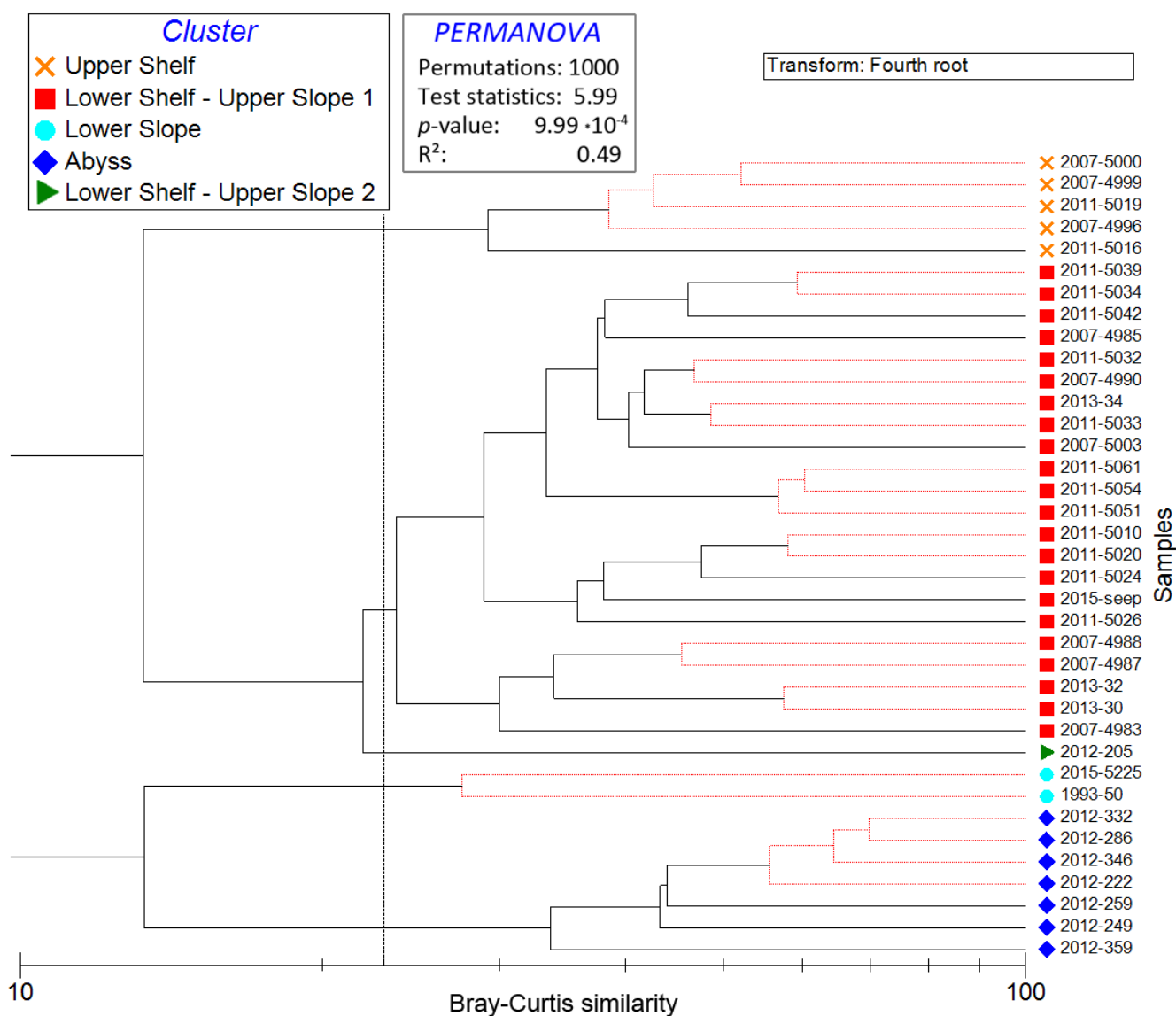
3.2. Semi-quantitative samples

315 The number of taxa per semi-quantitative sample varied from 8 (st. 2012-259) to 199 (st. 2015-seep), the
 316 diversity index values varied from 8 (st. 2012-259) to 39 (st. 2012-205) for ES-100, from 0.23 (st. 2007-
 317 4996) to 0.98 (st. 2012-359) for Pielou's evenness and from 0.8 (st. 2007-4996) to 3.5 (st. 2012-205) for
 318 Shannon diversity (Supplementary 3). A complete list of individual taxa with percentage abundance at
 319 each station appears in Supplementary 6.

320 The cluster analysis based on the Bray-Curtis similarity index with SIMPROF test identified five
 321 distinct clusters (= benthic assemblages), corresponding to the same cluster names introduced in the

322 previous section except the *Upper Shelf*, since no shelf stations were included in the quantitative data set
 323 (Fig. 8). PERMANOVA showed significant results with p -value < 0.001 (Fig. 8). Other similarity indices
 324 demonstrated similar dendrogram topology, except of the *Upper Shelf*, *Lower Shelf - Upper Slope 2*
 325 clustered within the *Upper Shelf*, *Lower Shelf - Upper Slope 1* assemblage (Morisita-Horn index –
 326 Supplementary 5, Fig. S-3). Furthermore, *Lower Slope* stations clustered together with the *Abyss* stations
 327 (Sørensen index – Supplementary 5, Fig. S-4).

328



329

330

331

332

333

334

335

336

337

Fig. 8. Cluster analysis of semi-quantitative Arctic benthos samples using Bray-Curtis similarity index with SIMPROF results. Branches and nodes not statistically significant at $p < 0.01$ are shown with red lines. Colour and shape indicate benthic assemblages, defined at the similarity level of 22 (vertical line); similarity axis is logarithmic. Enclosed the PERMANOVA results are shown.

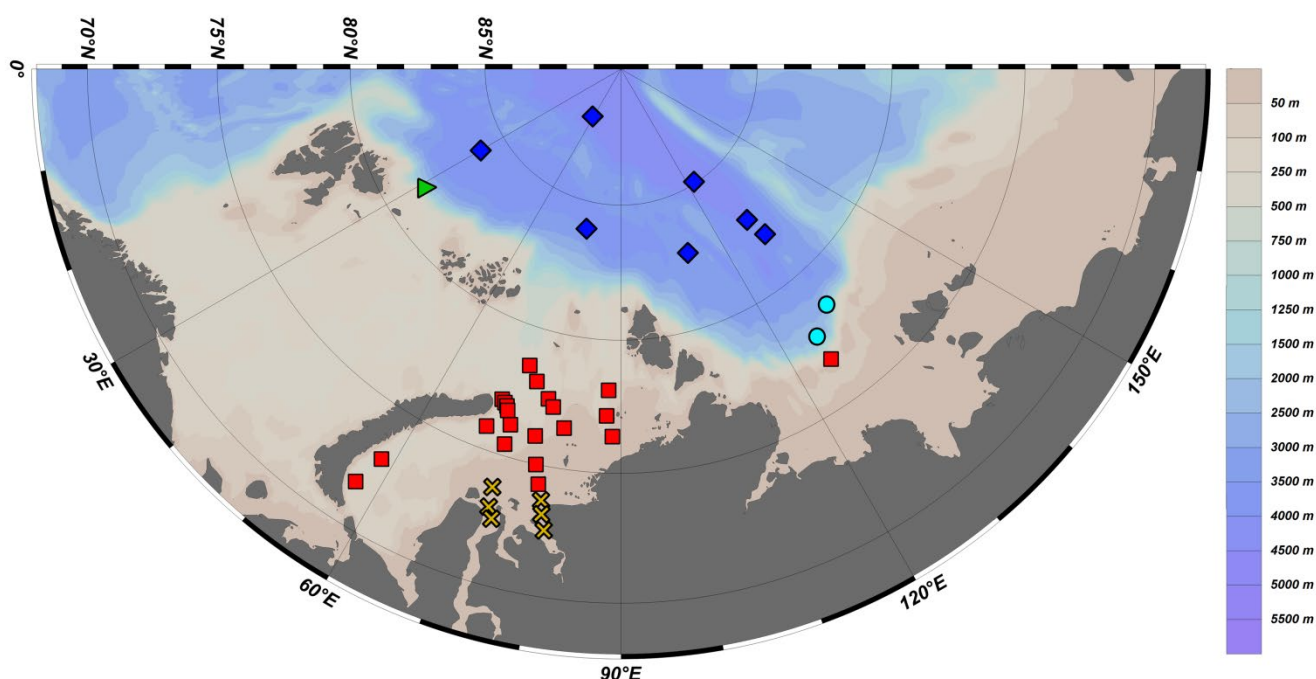
Kruskal-Wallis tests indicated significant difference between benthic assemblages for diversity indices (Table 3), though with lower p -values compared to quantitative samples. The *Lower Shelf – Upper Slope 1* and the *Abyss* groups appeared widely distributed across the entire area. The *Upper Shelf*, *Lower*

338 Shelf - Upper Slope 2 and Lower Slope groups were more localized, resulting from the limited geographic
 339 spread of samples in those assemblages (Fig. 9).

340 Table 3. Mean values of benthic diversity of each assemblage and results of Kruskal-Wallis and Dunn's post-hoc
 341 tests for semi-quantitative samples.

Group	Mean values \pm SD					Kruskal-Wallis		Dunn's post-hoc comparisons
	(1) Upper Shelf	(2) Lower Shelf - Upper Slope 1	(3) Lower Slope	(4) Abyss	(5) Lower Shelf - Upper Slope 2	Chi square	p	
Number of samples	7	20	2	7	1	-	-	-
Species number	55 \pm 36	98 \pm 34	50 \pm 33	19 \pm 6	133	22.82	<0.001	1-2; 2-4; 4-5
ES-100	16 \pm 7	24 \pm 8	22 \pm 6	11 \pm 2	39	17.46	0.0016	2-4; 4-5
Hill 50 extrapolated	10 \pm 3	15 \pm 5	16 \pm 3	11 \pm 6	24	11.24	0.0240	1-2; 1-5; 2-4; 4-5

342 Numbers in Dunn's post-hoc comparisons column indicate corresponding group of benthic assemblages, revealed
 343 with Bray-Curtis similarity.
 344



345 Fig. 9. Distribution of semi-quantitative samples coloured according to benthic assemblages. Colours and shapes
 346 of markers as in Fig. 3-7.
 347
 348

349 SIMPER analysis revealed a suite of taxa from different phyla and classes responsible for
 350 differences among assemblages (Table 4). As in Table 2, all taxa are listed according to their mean
 351 contribution to assemblage differences (only taxa with cumulative contribution up to 70% are shown).
 352 The bivalve *Portlandia arctica*, the mysid *Mysis oculata*, the polychaete *Spio* cf. *filicornis* and the
 353 ophiuroid *Stegophiura nodosa* were dominant in relative abundance and separated the *Upper Shelf* cluster.
 354 The *Lower Shelf - Upper Slope 1* group differed from other clusters by the high relative abundances of
 355 the ophiuroids *Ophiocten sericeum* and, to a lesser extent, *Ophiacantha bidentata* and *Ophiura robusta*
 356 and the bivalve *Yoldiella solidula*. The prevalence of the bivalve *Bathyarca frielei*, the isopod *Saduria*
 357 *sabini*, the gastropod *Mohnia danielsseni* and, to a lesser extent, the holothurian *Kolga hyalina*
 358 differentiated the *Lower Slope* group. The extreme dominance of *K. hyalina* and high relative abundances

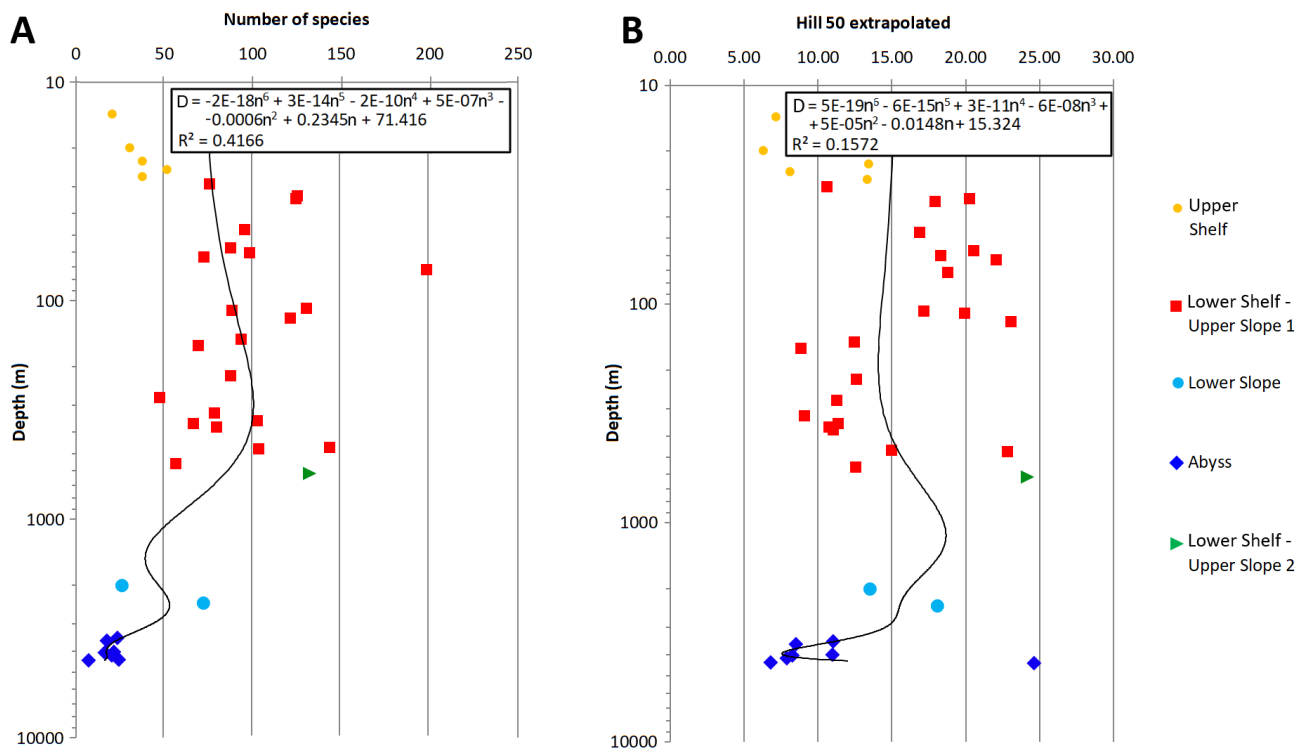
of the actiniarian *BathypHELLIA margaritacea*, the polychaete *Anobothrus laubieri* and, occasionally the ophiuroid *Ophiostriatus striatus* differentiated the *Abyss* group. The *Lower Shelf – Upper Slope 2* group, represented by a single station, contained high proportional abundances of the polychaete *Notoproctus oculatus*, the sipunculid *Nephasoma diaphanes*, the sponge *Geodia barretti* and Scyphozoa polyps (*Stephanoscyphus*) (Table 4).

Diversity parameters in semi-quantitative samples also yielded a parabolic relationship with depth with maximum species numbers and ES-100 at depths of ~100-600 m (Fig. 10).

Table 4. Results of SIMPER analysis of semi-quantitative Arctic benthos samples with values of average percentage abundance, average dissimilarity and percentage contribution within retrieved groups of assemblages.

Group	Species	Average Abundance (%)					Average Dissimilarity	Dissimilarity/SD	Contribution (%)	Cumulative (%)
		Upper Shelf	Lower Shelf - Upper Slope 1	Lower Slope	Abyss	Lower Shelf - Upper Slope 2				
Biv	<i>Portlandia arctica</i>	59.1	6	0	0	0	11	0.6	11.2	11.2
Pol	<i>Notoproctus oculatus</i>	0	0.1	0	0	24.7	5.7	2	5.8	17
Oph	<i>OphiocTen sericeum</i>	0.01	14.4	3.6	0	0	4.7	0.8	4.9	21.9
Biv	<i>Bathyarca frielei</i>	0	0	33	0	0	4.7	0.8	4.9	26.8
Sip	<i>Nephasoma diaphanes</i>	0	8.1	0	0	11.3	4.2	1.9	4.3	31.1
Mys	<i>Mysis oculata</i>	11.2	0.1	0	0	0	3.6	0.7	3.6	34.7
Hol	<i>Kolga hyalina</i>	0	0	3.6	48.8	0	3.5	3.1	3.6	38.3
Oph	<i>Ophiacantha bidentata</i>	0	6.8	0	0	0.7	2.4	1.9	2.5	40.8
Biv	<i>Yoldiella solidula</i>	0	7.4	0	0	0.7	2.4	1.8	2.4	43.2
Pol	cf. <i>Spio filicornis</i>	5.5	0	0	0	0	2.4	0.5	2.4	45.6
Oph	<i>Ophiura robusta</i>	0	4.4	0	0	0.2	2.2	1.8	2.2	47.8
Cni	<i>Stephanoscyphus</i>	0	4.7	0	0.1	4.4	2.1	1.5	2.1	49.9
Por	<i>Geodia barretti</i>	0	0	0	0	8.4	1.9	3.3	2	51.9
Cni	<i>BathypHELLIA margaritacea</i>	0	0	6.9	13.5	0	1.9	0.8	2	53.9
Oph	<i>Stegophiura nodosa</i>	3.4	0	0	0	0	1.9	0.5	1.9	55.8
Iso	<i>Saduria sabini</i>	2.0	0.3	2.7	0	0	1.6	1	1.6	57.4
Gas	<i>Mohnia danielsseni</i>	0	0	4.0	0.1	0	1.2	1	1.3	58.7
Biv	<i>Portlandia aestuariorum</i>	2.7	0	0	0	0	1.2	0.5	1.2	59.9
Pol	<i>Myriochele heeri</i>	0	0.9	1.3	0	3.5	1.1	1.9	1.2	61.1
Pol	<i>Anobothrus laubieri</i>	0	0	0	13.4	0.7	1.1	1.3	1.2	62.2
Hol	<i>Elpidia heckeri</i>	0	0	6.1	4.1	0	1.1	0.8	1.1	63.3
Biv	<i>Dacrydium vitreum</i>	0	5.1	0	0	0	0.9	0.3	1	64.3
Pol	<i>Siboglinum hyperboreum</i>	0	0	6.2	0	0	0.9	0.8	0.9	65.2
Iso	<i>Saduria sibirica</i>	1.8	0.1	0.6	0	0	0.8	0.8	0.9	66.1
Por	<i>Thenea muricata</i>	0	0	0	0	3.5	0.8	3.3	0.8	66.9
Pol	<i>Galathowenia oculata</i>	0	0.04	0	0	3.2	0.8	2	0.8	67.6
Hol	<i>Elpidia glacialis</i>	0	4	0	0	0	0.7	0.3	0.8	68.4
Hol	<i>Myriotrochus theeli</i>	0	0	4.9	0	0	0.7	0.8	0.7	69.1
Pol	<i>Notomastus</i> sp.	0	0	1.3	0	0.4	0.6	1.6	0.6	69.8
Biv	<i>Similipecten greenlandicus</i>	0	1.8	0	0	0	0.6	0.5	0.6	70.4

Taxa with cumulative contribution <70% are shown. First several most significant taxa in each assemblage are marked in grey. Por – Porifera; Cni – Cnidaria; Pol – Polychaeta; Sip – Sipuncula; Gas – Gastropoda; Biv – Bivalvia; Mys – Mysida; Iso – Isopoda; Hol – Holothuroidea; Oph – Ophiuroidea.



373
374
375
376
377
378

Fig. 10. Values of species number (a) and Hill 50 extrapolated (b) in semi-quantitative samples in relation to depth. Colour corresponds to clusters as in Figs. 3-8. Depth axis is logarithmic. Trend line equations and R^2 values are shown in rectangles (D – depth; n – species number or rarefaction per 50 individuals).

379
380
381
382
383
384
385
386
387
388
389
390
391

The benthic assemblages identified for semi-quantitative samples did not overlap by depth. The depth ranges of the benthic assemblages based on Bray-Curtis similarity were as follows: 14-27 m for the *Upper Shelf*; 29-553 m for the *Lower Shelf - Upper Slope 1* group; 615 m for the *Lower Shelf - Upper Slope 2* group; 1992-2390 m for the *Lower Slope* group and 3472-4380 m for the *Abyss* group. For the Morisita-Horn index identified depth ranges of 14-27 m for the *Upper Shelf* group; 29-615 m for the *Lower Shelf - Upper Slope 1* group; 1992-2390 m for the *Lower Slope* group and 3472-4380 m for the *Abyss* group; the *Lower Shelf - Upper Slope 2* assemblage fell within the *Lower Shelf - Upper Slope 1* cluster. The depth ranges for the quantitative Sørensen similarity index were the same as for the Bray-Curtis index except that the *Lower Slope* and the *Abyss* assemblages shared a single cluster (Supplementary 5, Fig. S-3, S-4). Unlike groups based on quantitative samples, large depth gaps separated the groups of benthic assemblages based on semi-quantitative samples, likely reflecting the lower sample size and geographic coverage. Bathymetric distribution of clusters and rarefaction curves are available in Supplementary Figures S-5 and S-6.

4. Discussion

4.1. Benthic assemblages along a depth gradient

Our analysis done within the Central Arctic was the first one to analyse the bathymetric distribution of the benthic assemblages across a large geographic area (from the Fram Strait to the Beaufort Sea) and a large depth range (from 14 to 5416 m). We identified five groups of benthic assemblages, and all except the *Lower Shelf – Upper Slope 2* group replaced, rather than transitioned into, one another with depth. This is especially interesting considering the sampling area of each sample varied over 30-fold (from 0.023 to 0.750 m²), apparently not decisively affecting the dendrogram structure (Figs. 3, 8). This zonation pattern spanned the entire analysed area, based either on quantitative (for macrofauna) or semi-quantitative (for megafauna) samples and based on three different quantitative similarity indices (Figs. 3-10). The five assemblages we identified largely aligned with those described in previous smaller-scale regional studies, (Deubel, 2000; Sirenko et al., 2004; Vedenin et al. 2018; Käß et al., 2019; Ravelo et al., 2020) with impressively little bathymetric or geographic structure and variation within each assemblage. The exceptions were the Fram Strait area, which resembled the *Lower Slope* group, and the Beaufort Sea stations that grouped separately within the *Lower Shelf - Upper Slope 2* and, to a lesser extent, within the *Lower Slope* groups (Fig. 3, Supplementary 5). Three of the described assemblages (the *Lower Shelf – Upper Slope 1*, the *Lower Slope* and the *Abyss*) formed consistent bands of zonation across entire study area. The distributional patterns of all five assemblages likely relate to environmental gradients at corresponding depths such as water masses, organic carbon fluxes, and near-bottom currents (Watling et al., 2013) which we discussed below, acknowledging that the exact factors remain hypothetical.

The *Upper Shelf* group identified from semi-quantitative trawl samples partly corresponds to the *Portlandia arctica* and *Ennucula tenuis* bivalve community previously described for the Kara, Laptev, and East-Siberian Seas (Filatova & Zenkevich 1957; Deubel, 2000; Petryashev et al. 2004; Sirenko & Denisenko 2010; Vedenin et al. 2018). A similar assemblage type was reported from the Beaufort Sea coastal areas (Carey & Ruff, 1977). The low overall diversity of this assemblage corresponds with high values of abundance and biomass (Petryashev et al., 2004; Vedenin et al., 2018). The *Upper Shelf* group distribution likely relates to high sedimentation rate from the freshwater inflow over the wide and shallow Siberian shelves (Weber, 1989; Fütterer & Galimov, 2003; Flint et al., 2019; Vedenin et al., 2021). The shallow Chukchi Sea inflow shelf (of which we did not include samples) experiences little of this freshwater influence, and higher diversity (Blanchard et al., 2013; Schonberg et al., 2014; Grebmeier et al., 2015), so the pattern is not pan-Arctic but rather characteristic for interior shelves (Williams & Carmack 2015).

The *Lower Shelf – Upper Slope 1* group partly corresponds to two communities dominated by the ophiuroids *Ophiocten sericeum* and *Ophiopleura borealis*, both widely distributed across the Siberian and Beaufort shelves (Filatova & Zenkevich 1957; Petryashev et al., 2004; Ravelo et al., 2020). Near Franz-

427 Joseph Land, Svalbard and in the Chukchi Sea, *Ophiura robusta* ophiuroids and/or *Yoldiella solidula*
428 bivalves (Dahle et al., 2009) dominate communities at similar depths, along with the ophiuroids
429 *Ophiacantha bidentata* and *Ophiura sarsi* (Piepenburg et al., 1996; Bluhm et al., 2009). Vedenin et al.
430 (2018) and Deubel (2000) delineated this benthic assemblage as the UPPER SLOPE and HANG (=
431 ‘slope’) groups, respectively, for the Barents, Laptev and East-Siberian Seas shelf edge and slope.
432 Assemblages similar in taxonomic composition were described for the Beaufort Sea by Ravelo et al.
433 (2020). The quantitative and semi-quantitative samples used in this study identified *Y. solidula* and *O.*
434 *sericeum* as the dominant species in this area, and an assemblage of higher diversity than the *Upper Shelf*
435 group. This distribution likely relates to the higher number of ecological niches at the shelf edge and upper
436 slope and less stress from turbidity and low salinity associated with river discharge (Carney, 2005; Flint
437 et al., 2019). In addition, its lower limit roughly corresponds with the lower boundary of the Atlantic water
438 layer (Wassman et al., 2019; Bluhm et al., 2020). Hydrobiochemical connectivity of the entire Arctic
439 upper continental slope is given through boundary currents, cross-slope upwelling of nutrient rich water,
440 and down-welling particulate organic matter from upper waters (summarized in Bluhm et al., 2020).
441 Previous studies report faunal changes coinciding with the transition between the Atlantic layer and other
442 water masses for polychaetes, isopods, gastropods and fishes in the Norwegian Sea, where a particularly
443 steep hydrological gradient occurs (Svavarsson et al., 1990; Bergstad et al., 1999; Høisæter, 2010; Oug et
444 al., 2017).

445 The *Lower Slope* group in our study extended from the continental slope in the Fram Strait (at both
446 Eastern and Western sides) to the Beaufort Sea and corresponds to benthic communities described in
447 smaller-scale studies: *Galathowenia fragilis* - *Myriochele heeri* polychaete community in the Fram Strait
448 (Budaeva et al., 2008; Vedenin et al., 2016; Käß et al., 2019); Polychaeta-dominated community in the
449 Laptev Sea (Fütterer, 1993; Sirenko, 1998; Sirenko et al., 2004); the MID-SLOPE and LOWER SLOPE
450 (Vedenin et al., 2018) and RAND and RÜCKEN (= ‘margin’ and ‘ridge’; Deubel, 2000) communities
451 within the Barents and Laptev Seas and on Lomonosov Ridge. Oweniidae polychaetes and *Bathycarca*
452 *frielei* bivalves dominated our quantitative and semi-quantitative samples, respectively. The *Lower Slope*
453 group apparently spans a huge area around the Central Arctic Basin perimeter and including slopes of the
454 Greenland and Norwegian Seas at depths of ~700-3000 m (Nilsen & Holthe, 1985; Schnack, 1998;
455 Vedenin et al., 2016). Bluhm et al. (2020) suggested faunistic similarity along the entire continental slope
456 in the Arctic Ocean based on a smaller data set. Our data set supports this assertion, along with occasional
457 reports of communities dominated by polychaetes, for example Paraonidae and Cirratulidae (Paul &
458 Menzies, 1974) or *Prionospio* sp. and unidentified polychaetes (Bluhm et al., 2005) from the Canada
459 Basin continental slope. Environmental conditions defining the *Lower Slope* group distribution may
460 include weaker currents compared to the *Lower Shelf – Upper Slope 1* group and similar hydrographic
461 properties related to Arctic Deep Water (synthesized in Bluhm et al., 2020). Interestingly, the mentioned

462 *Galathowenia* and *Myriochele* polychaetes are strict deposit-feeders that build their tubes out of fragments
463 of plankton foraminifers and sponge spicules, depending therefore indirectly on the higher organic matter
464 supply from upper water layers (Nilsen & Holthe, 1985; Parapar, 2006). This factor might be the reason
465 for preventing Oweniidae distribution down to the abyssal plain.

466 The holothurians *Kolga hyalina* and *Elpidia heckeri* and the actiniarian *Bathypheilia margaritacea*
467 dominated the *Abyss* group, that apparently occupies the entire Central Arctic at depths >~3000 m, as
468 shown in numerous publications based on data from the Greenland, Nansen, Amundsen and Canada basins
469 (Gorbunov, 1946; Kröncke, 1992; Kröncke, 1994; Deubel, 2000; Sirenko et al., 2004; Soltwedel et al.,
470 2009; Vedenin et al., 2018; Rybakova et al., 2019). These dominants also often appear in bottom images
471 and in semi-quantitative samples such as in trawls (Macdonald et al., 2010; Rybakova et al., 2019; Zhulay
472 et al., 2019). Smaller quantitative gears demonstrate also high abundances of smaller polychaete species,
473 such as *Anobothrus laubieri*, *Ymerana pteropoda* and *Ophelina opisthobranchiata* (Table 2). The poor
474 trophic conditions likely resulted in the lowest values of abundance, biomass and diversity within the
475 *Abyss* group (described in Bluhm et al. 2011; Bluhm et al., 2015; Vedenin et al., 2018; Rybakova et al.,
476 2019). For megafauna, including the holothurians *K. hyalina* and *E. heckeri*, falls of *Melosira arctica*
477 algae colonies torn off the sea ice were shown to be important (if not primary) food source, that possibly
478 drives the distribution of these taxa across the Central Arctic abyssal plains (Boetius et al., 2013).

479 The *Lower Shelf – Upper Slope 2* group overlapped the *Lower Shelf – Upper Slope 1* and *Lower*
480 *Slope* groups in depth. The former differed from other clusters in our study by its greater biomass and
481 species number and the prevalence of filter-feeders, such as sponges (mostly *Geodia* spp. and *Thenaea*
482 *muricata*), polychaetes (*Bushiella similis*) and bryozoans (*Tubulipora* spp.). This benthic assemblage
483 corresponds to the community of filter-feeders described by Sirenko et al. (2004) from a few areas around
484 the Severnaya Zemlya archipelago. Furthermore, the taxonomic composition of this group resembled the
485 benthic ecosystems in several areas in the northern Barents Sea and around Svalbard (Sswat et al., 2015).
486 The similarity within this assemblage was lower than for the other assemblages because of some
487 taxonomic differences between the Eurasian samples (northern slopes of Barents and Kara seas) and the
488 Amerasian samples (northern slopes of Beaufort Sea and Northwind Ridge / Chukchi Plateau, see Fig. 3).
489 However, we decided to retain the *Lower Shelf – Upper Slope 2* as a separate assemblage because of
490 similarity in filter-feeders composition. Strong near-bottom currents enriched with suspended food
491 particles usually favour the development of communities dominated by filter-feeders, as shown for
492 communities on the shelf edge of the North-Eastern Beaufort Sea (Pisareva et al., 2015; Rand et al., 2018).
493 This type of benthic assemblages may occur widely across the Arctic continental slope, though with
494 uneven distribution given its strong dependence on near-bottom hydrodynamics (Thomson, 1982).

495 496 4.2. Benthic abundance, biomass and taxon richness

497 Previous studies have documented gradual decrease of abundance and biomass of macrobenthos with
498 depth for different areas of the Arctic Ocean (Rex & Etter, 2010; Macdonald et al., 2010; Bluhm et al.,
499 2011; Bluhm et al., 2015; Bluhm et al., 2020; Vedenin et al., 2018). Our study extends earlier work by
500 Vedenin et al. (2018) with three times more stations and several fold extension of the study region,
501 however, we find similar patterns of depth-related changes (Fig. 5). One notable difference links to
502 different diversity patterns using extrapolated rarefactions based on Hill numbers, which indicated higher
503 diversity values for the *Lower Slope* and for the *Abyss* assemblages (Fig. 6, Supplementary 3). We also
504 identified a cluster of *Lower Shelf–Upper Slope 2* stations, absent in Vedenin et al. (2018). Decreases in
505 abundance and biomass with depth likely relate to decreased food availability at the seabed with increasing
506 ocean depth, a trend common for the global ocean and especially prominent in the Arctic Ocean largely
507 covered with seasonal sea-ice (Wei et al., 2010 a; Degen et al., 2015; Vedenin et al., 2018; Käb et al.,
508 2019; Górska et al., 2020; Oleszczuk et al., 2021). Exceptions to this depth-related decline are rare and
509 probable exist only where oceanographic anomalies disrupt typical patterns of food supply and oxygen
510 concentrations, such as the East Pacific off Peru where biomass peaks at mid-depths of 800-1000 m
511 (Rowe, 1971).

512 Trends in diversity with depth in the ocean differ from trends in abundance and biomass. Maximum
513 values of species richness and diversity often occur at mid- and low-bathyal depths from 2000 m to 4000
514 m (Rex, 1981; Flach & de Bruin, 1999; Gage et al., 2000; Gebruk et al., 2010; Brown & Tatje, 2014).
515 However, in some locations, gradual decrease in diversity with depth (similar to the abundance and
516 biomass trends) occur (Rex & Etter, 2010). Such a pattern may relate to the biogeographic history of a
517 basin. For example, in basins with limited deep-water exchange such as those with shallow straits or sills
518 where geologically recent impoverishment or extinction of the benthic ecosystem occurred, recolonization
519 could follow from shallow waters or more slowly from adjacent deep-sea areas (Rex & Etter, 2010). The
520 Mediterranean Sea and Arctic Ocean illustrate this process. The former was recolonized through the
521 Gibraltar Strait after complete fauna extinction during the Messinian salinity crisis (Krijgsman et al.,
522 1999). In the Eastern Mediterranean, maximum benthic diversity occurs in shallow waters and decreases
523 with depth (Tselepides & Eleftheriou, 1992; Tselepides et al., 2000). Several lines of evidence point to
524 depletion (and possible extinction) of the benthic fauna of the Arctic Ocean during the Pleistocene
525 glaciations, given that primary production dropped dramatically or even ceased beneath the ice shield
526 (Menzies, 1962; Dunbar, 1968; Menzies, 1973; Renaud et al., 2008; Kędra et al., 2015). Following
527 glaciation, recolonization took place from the Atlantic through the Fram Strait and from the Pacific
528 through the Bering Strait, involving mostly shallow-water species (Vermeij, 1991; Nesis, 2001; Mironov,
529 2013). Therefore species numbers (and biodiversity) decrease with depth in the high Arctic, as shown by
530 Włodarska-Kowalczyk et al. (2004), Bluhm et al. (2011) and Käb et al. (2019). However, recent analysis
531 of a subset of the samples used in the present study identified a diversity peak at depths 100-300 m

(Vedenin et al., 2018), a depth range not included in these previous deep-sea studies. Our study extended the sampling area westward to the Yermak Plateau and Fram Strait (Fig. 4), and we found a similar parabolic pattern of diversity change with depth. Based on semi-quantitative trawl samples on the Siberian shelf and slope (Fig. 9), diversity peaked at 100-600 m. Vedenin et al. (2018) linked the diversity maximum at 100-300 m on the Laptev Sea shelf and slope to river run-off that impacts ocean stratification and primary productivity above the shallow shelf. However, this explanation cannot apply to the Svalbard area. A possible reason for peak diversity values can be the warmer Atlantic waters inflowing at ~100-800 m, enriching this area with organic carbon as well as Atlantic boreal species (Wassmann et al., 2019; Bluhm et al., 2020). For example, the inflow of zooplankton biomass contributed by *Calanus* spp. alone might exceed a million tons C year⁻¹ (Wassmann et al., 2015; Basedow et al., 2018). Our dataset lacks samples from <100 m depth near Svalbard, so the local diversity trends could be different.

4.3. Bathymetric boundaries in the Arctic benthic fauna

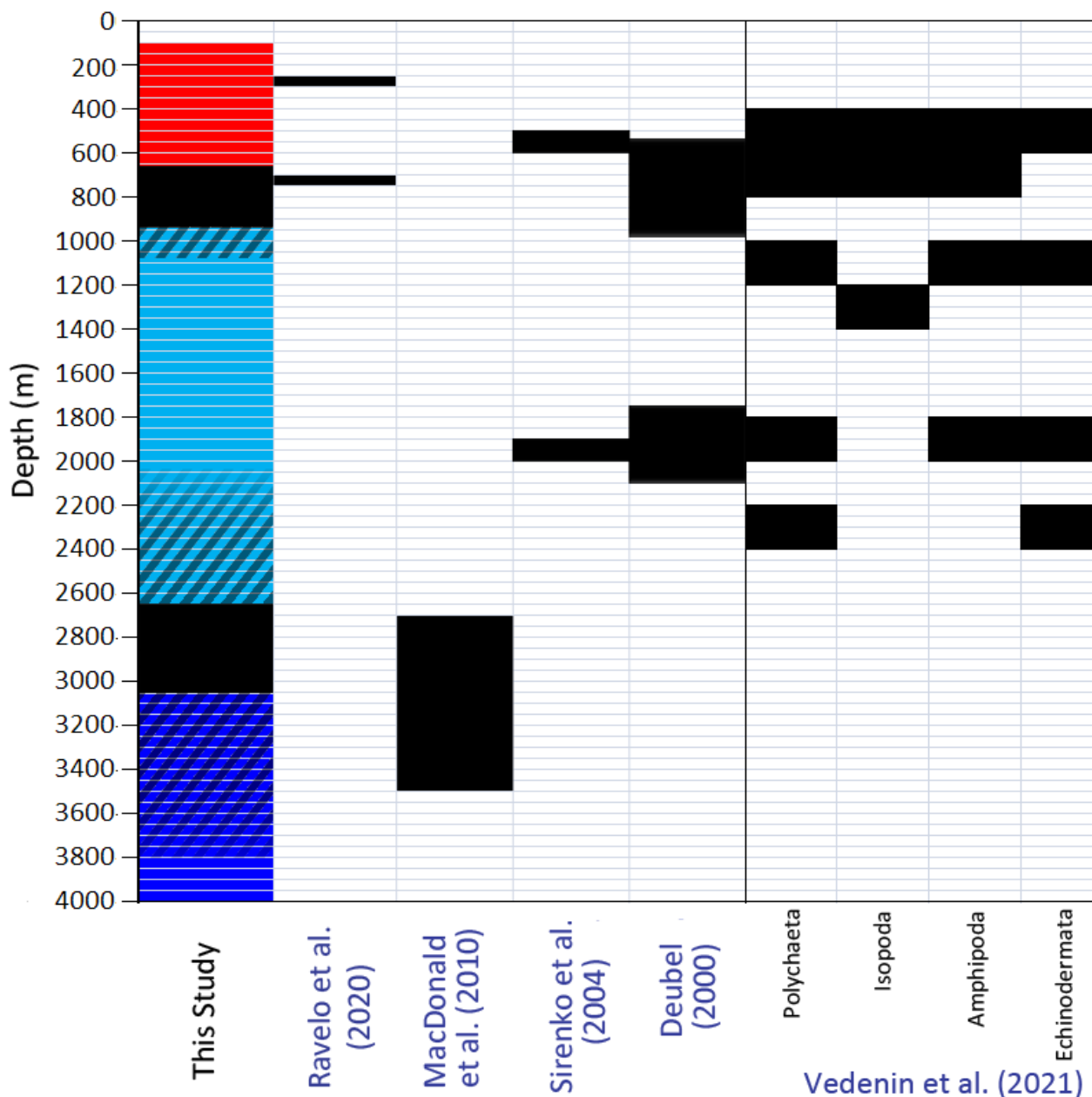
Different researchers have used different approaches to describe biogeographic zonation, including bathymetric patterns. Mironov (2013) distinguished three main types: based on the distribution of species (1), communities (2) and environmental parameters (3). Multiple studies show major bathymetric boundaries in the global ocean, based on species distributions near 200 m, corresponding to the boundary between sublittoral and bathyal faunas, and near 3000 m at the boundary between bathyal and abyssal faunas (Carney, 2005; Howell et al., 2002; Krayushkina, 2000; Mironov, 1986; Vinogradova, 1962). The 200-m boundary roughly corresponds to the lower limit of the photic zone and to the shelf break (Carney, 2005; Wei et al., 2010 b), whereas the 3000-m boundary likely reflects decreasing food availability and significant changes in bottom topography, such as the transition from continental slope to continental rise and abyssal plain (Thistle, 2003; Wei et al., 2010 b; Watling et al., 2013). Many studies also identify a boundary at 800-1200 m depth (Gage, 1986; Svavarsson, 1990; Howell et al., 2002), explaining this boundary by peculiarities of bottom topography, near-bottom currents and sediment changes, that affect the distribution of benthos in case of isopods (Svavarsson, 1990) and distributional paths of plankton larvae in case of echinoderms (Gage, 1986). Most of these publications were based on data from the Pacific, Atlantic and the Norwegian Sea. In the high Arctic, only Vedenin et al. (2021) used a species-based approach to bathymetric zonation. Based on data on three macrofaunal taxa - Polychaeta, Crustacea and Echinodermata, we identified two main boundaries, the first at 450-800 m between the sublittoral and bathyal faunas and at 1800-2000 m between the bathyal and abyssal faunas. Our study, based on the distribution of groups of benthic assemblages rather than species, identified the boundary between sublittoral and bathyal faunas at depths of ~650-950 m (between the *Lower Shelf – Upper Slope 1* and *Lower Slope* groups) based on Bray-Curtis similarity or at ~650-1200 m based on Sørensen similarity. Thus, two different methods support a strong boundary around ~450-1200 m. We refrained from defining

567 a boundary between the *Upper Shelf* and *Lower Shelf – Upper Slope 1* groups because of the small sample
568 set, and because previously published data suggest a deeper position for biogeographic boundaries in the
569 Arctic (Mironov, 2013; Vedenin et al., 2021). As we already mentioned, a possible environmental driver
570 for this boundary is the transition from the warm Atlantic layer to the deep and cold Arctic waters that
571 lies at approximately 700-800 m (Wassman et al., 2019; Bluhm et al., 2020).

572 Species distributions indicated a boundary at ~2000 m (Vedenin et al., 2021). When considering the
573 distribution of benthic assemblages based on quantitative samples from our study, similar species
574 appeared to dominate within a depth range of 650-3000 m (based on Bray-Curtis similarity) or 650-3800
575 m (based on Morisita-Horn and Sørensen indices) for the *Lower Slope* assemblage. The semi-quantitative
576 (trawl) material we have is relatively scarce, with large depth gaps. However, Sirenko et al. (2004)
577 reported a boundary at ~1900-2000 m between the slope ‘polychaete community’ and the deeper
578 community dominated mainly by the holothurians *Kolga hyalina* and *Elipida heckeri*. Deubel (2000)
579 reported boundaries at ~530-990 m and at 1740-2100 m based on quantitative macrofaunal data. For the
580 Beaufort Sea and Canada Basin stations in this study, we identified a transition between the different
581 deep-sea communities at ~1850-3200 m (Bluhm et al., 2005) and at ~2700-3500 m (MacDonald et al.,
582 2010). We lack any bathymetric transects on the slope north of Ellesmere Island and Greenland but
583 identify different bathymetric boundaries in the Arctic Ocean based on our own data and on previous
584 studies (Fig. 11).

585 For the Barents and Laptev Seas previous work showed that benthic communities replace one
586 another with depth with little bathymetric overlap (Vedenin et al., 2018). Unlike these results, we found
587 significant depth overlap at ~650-950 m (or 650-1200 m by different similarity indices) between two
588 groups of benthic assemblages – the *Lower Shelf – Upper Slope 1* and *Lower Slope*. This overlap may
589 reflect regional differences in the depth ranges of the eurybathic species that dominated their
590 corresponding assemblages (discussed in Vermeij, 1991; Nesis, 2001; Vedenin et al., 2021). At greater
591 depths we found overlap between the *Lower Slope* and *Abyss* groups (2644-3054 m based on Bray-Curtis
592 similarity; 3677-3848 m based on Morisita-Horn similarity; 2027-3848 m based on Sørensen similarity).
593 No published evidence suggests any boundary at these depths in the Arctic, except for Vedenin et al.
594 (2018) and, possibly, MacDonald et al. (2010) (both data sets integrated in this study). Based on the
595 present and earlier studies we think that the boundary at this depth reflects abrupt changes in major
596 community characteristics, including abundance and diversity (Table 1, 3; Figs. 5, 10) at the continental
597 rise to abyssal seafloor transition, rather than significant changes in taxonomic composition (Boetius et
598 al., 1996; Klages et al., 2004; Degen et al., 2015). Therefore, reduced sensitivity of the Morisita-Horn and
599 Sørensen similarity indices to sample size (Chao et al., 2006), elucidated a markedly different depth range
600 of the *Lower Slope / Abyss* transition compared to the Bray-Curtis index.

601 Environmental gradients, briefly discussed earlier and in previous publications (Mironov, 2013;
 602 Watling et al., 2013; Vedenin et al., 2021) drive the boundaries elucidated by these different approaches.
 603 In particular, the boundary between the *Lower Shelf – Upper Slope 1* and the *Lower Slope* assemblages
 604 roughly corresponds to the lower boundary of the warm Atlantic water masses, while the boundary
 605 between the *Lower Slope* and the *Abyss* matches the transition from the continental rise to abyssal plain.
 606 However, the exact mechanisms remain unknown and require further studies.



607 Fig. 11. Bathymetric boundaries between the sublittoral and bathyal (black rectangles within ~450-900 m) and
 608 between the bathyal and abyssal (black rectangles within ~1700-2100 and ~2000 m) faunas identified in the Arctic
 609 in the present study (based on Bray-Curtis similarity) and in published research. Diagonal pattern indicate
 610 boundaries at different depth ranges revealed by different similarity indices (either Morisita-Horn or Sørensen).
 611 Colours in the left column are the same as in Fig. 3-10. MacDonald et al. (2010) did not sample shallower than 800
 612 m and Ravelo et al. (2020) sampled to 1000 m.
 613
 614

615 4.4. Pan-arctic extrapolation

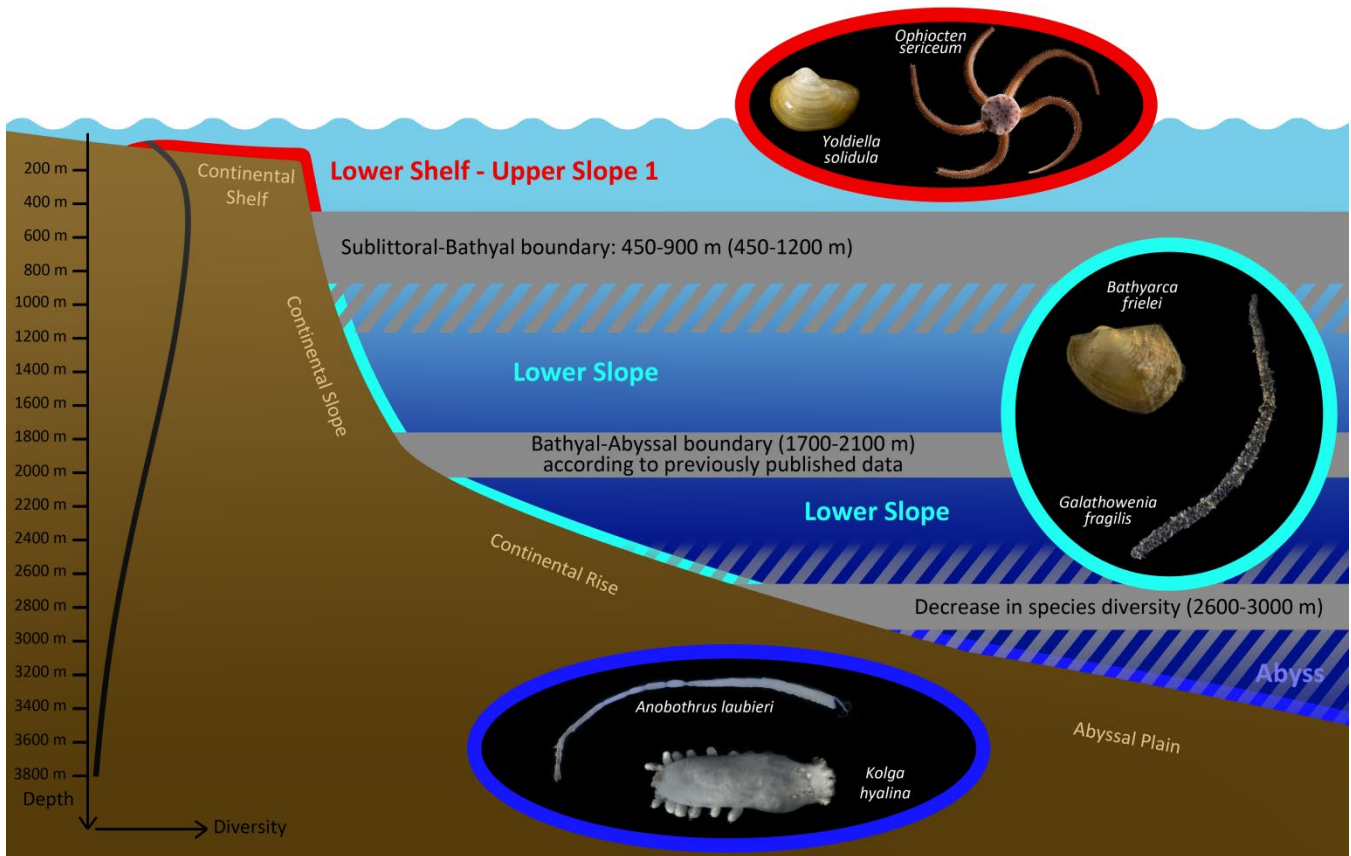
616 Based on combined data on bathymetric benthic fauna zonation across the Central Arctic slope and abyssal
617 zone, our study confirms major boundaries at ~450-900 m (Deubel, 2000; Sirenko et al., 2004; Bluhm et
618 al., 2020; Vedenin et al., 2021; this study) and at ~1700-2100 m (Deubel, 2000; Sirenko et al., 2004;
619 Vedenin et al., 2021). The entire macrofauna essentially changes at these pronounced biogeographic
620 boundaries. The less universal deeper boundary at ~2600-3000 m (MacDonald et al., 2010; present study)
621 can be distinguished based on cluster analysis and corresponds to the depth of a steep decrease in
622 abundance, biomass, and diversity within the Central Arctic rather than a taxonomic turnover (Klages et
623 al., 2004; Degen et al., 2015; Vedenin et al., 2018). The shelf benthic assemblages documented in our
624 study are unlikely to be Pan-arctic, given quite marked differences in species composition between the
625 Barents Sea, the Siberian seas and the Chukchi Sea within the Arctic (Sirenko et al., 1998; Grebmeier et
626 al., 2006; Mironov, 2013).

627 The bathymetric overlap between the deeper benthic assemblages increases when extending the area
628 from the smaller Barents and Laptev Sea area to almost the entire Arctic (Vedenin et al., 2018; this study).
629 Apparently, higher environmental variability (e.g. varying freshwater discharge, sea ice volume or
630 Atlantic/Pacific waters influence, Bluhm et al., 2020) over the broader study area contributes to this
631 overlap. However, general trends of replacement of benthic communities and their composition remain
632 similar at similar depths. Therefore, we suggest potential extrapolation of these bathymetric patterns
633 across the continental slope and abyssal plain around the entire Central Arctic Ocean. Similar bathymetric
634 patterns occur on both sides of the continental slope of the Fram Strait, despite the influence of different
635 water masses: The warm Norwegian Current bathes the eastern slope, whereas the western slope sits
636 beneath the cold East Greenland Current (Käb et al., 2019). Faunal composition deviates in the Beaufort
637 Sea where the upper slope differs slightly as a result of the admixture of species of Pacific origin with
638 those that disappear rapidly with the transition to the Atlantic layer (Bluhm et al., 2005; MacDonald et
639 al., 2010; Conlan et al. 2013; Zhulay et al., 2019; Ravelo et al., 2020). In addition, a significant data gap
640 remains on benthic communities from the continental slope of a large area north of Greenland and the
641 Canadian Arctic Archipelago (Bluhm et al., 2020). A comprehensive test of the hypothesis of the pan-
642 arctic uniformity of benthic bathymetric zonation will require adding data from this area, where remaining
643 multi-year sea ice has hindered access to date.

644 We propose a visualization of the benthic bathymetric boundaries in the Central Arctic (Fig. 12),
645 averaging the bathymetric profile for the Arctic Ocean and setting the shelf boundary at ~200 m (see
646 Jakobsson et al., 2012; Hay, 2016). Within the study area we propose relatively uniform patterns
647 including: (1) a parabolic change in species diversity that peaks at ~100-600 m, (2) bathymetric zonation
648 of benthic assemblages (based on cluster analysis), that changes with increasing depth, (3) major
649 biogeographic boundaries at ~450-900 m (or 450-1200 m depending on similarity indices) and ~1700-

650
651
652

2100 m, and a less universal boundary (recognised by community summary characteristics such as biomass and species richness) at around ~2600-3000 m (Fig. 12).



653
654
655
656
657
658
659
660
661

Fig. 12. Schematic of the benthic faunal bathymetric zonation in the Central Arctic based on the present study and published data. Upper and lower solid gray horizontal lines indicate boundaries at different depth ranges revealed by Bray-Curtis similarity index; diagonal pattern indicate boundaries revealed by different similarity indices (Morisita-Horn and Sørensen). Diversity trend is shown in the left. The seafloor contour and the outline of ellipses are coloured according to the benthic assemblages as in Figs. 3-11. Species characterizing the community clusters are shown in the corresponding ellipses (photos by A. Vedenin).

Acknowledgments

662
663
664
665
666
667
668
669
670
671

We thank Dr. Natalya Budaeva, Dr. Kirill Minin, Dr. Alexey Udalov, Dr. Vadim Mokievsky, Dr. Thomas Soltwedel and Dr. Vassily Spiridonov for their help with the literature search. Many thanks are due to Dr. Antje Boetius and Dr. Ingrid Krönke for providing original macrobenthic data from earlier expeditions. We are grateful to all the specialists who identified benthic organisms in the original samples – Dr. Nataliya Anisimova, Dr. Manuela Gusky, Dr. Hendrik Deubel. We acknowledge support from NOAA’s office of Ocean Exploration and Research for the original work conducted in the Beaufort Sea/Canada Basin, and from the Arctic SIZE project co-funded by UiT The Arctic University of Norway and the Tromsø Research Foundation. This research was supported by the Russian Foundation for Basic Research (RFBR) project #18-05-60228.

672
673
674
675
676
677
678
679
680
681
682
683
684
685
686
687
688
689
690
691
692
693
694
695
696
697
698
699
700
701
702
703
704
705
706
707
708
709
710
711
712
713
714
715
716
717
718
719
720
721
722
723
724
725
726
727
728
729
730
731
732
733

References

1. Anderson, M. J. (2005). Permutational multivariate analysis of variance. Department of Statistics, University of Auckland, Auckland, 26, 32-46.
2. Anisimova, N.V., Frolova, E.A., Lubin, P.A., Frolova, A.A., Denisenko, N.V., Panteleeva, N.N., Lubina, O.S. (2003). Species composition and quantitative distribution of macrobenthos in the Voronin trench and on the adjacent continental slope. In: Fauna of invertebrates of Kara, Barents and White Seas (ecology, biogeography), p. 79-92. [In Russian]
3. Basedow, S. L., Sundfjord, A., von Appen, W. J., Halvorsen, E., Kwasniewski, S., & Reigstad, M. (2018). Seasonal variation in transport of zooplankton into the Arctic basin through the Atlantic gateway, Fram Strait. *Frontiers in Marine Science*, 5, 194 p. <https://doi.org/10.3389/fmars.2018.00194>
4. Bergstad, O. A., Bjelland, O., Gordon, J. D. (1999). Fish communities on the slope of the eastern Norwegian Sea. *Sarsia*, 84(1), 67-78. <https://doi.org/10.1080/00364827.1999.10420452>
5. Blanchard, A. L., Parris, C. L., Knowlton, A. L., & Wade, N. R. (2013). Benthic ecology of the northeastern Chukchi Sea. Part I. Environmental characteristics and macrofaunal community structure, 2008–2010. *Continental Shelf Research*, 67, 52-66. <https://doi.org/10.1016/j.csr.2013.04.021>
6. Bluhm, B. A., MacDonald, I. R., Debenham, C., & Iken, K. (2005). Macro-and megabenthic communities in the high Arctic Canada Basin: initial findings. *Polar Biology*, 28(3), 218-231. <https://doi.org/10.1007/s00300-004-0675-4>
7. Bluhm, B. A., Iken, K., Hardy, S. M., Sirenko, B. I., & Holladay, B. A. (2009). Community structure of epibenthic megafauna in the Chukchi Sea. *Aquatic Biology*, 7(3), 269-293. <https://doi.org/10.3354/ab00198>
8. Bluhm, B. A., Ambrose, W. G. Jr., Bergmann, M., Clough, L. M., Gebruk, A. V., Hasemann, C., et al. (2011). Diversity of the Arctic deep-sea benthos. *Marine Biodiversity* 41, 87–107. <https://doi.org/10.1007/s12526-010-0078-4>
9. Bluhm, B. A., Kosobokova, K. N., & Carmack, E. C. (2015). A tale of two basins: An integrated physical and biological perspective of the deep Arctic Ocean. *Progress in Oceanography*, 139, 89-121. <https://doi.org/10.1016/j.pocean.2015.07.011>
10. Bluhm, B. A., Janout, M. A., Danielson, S. L., Ellingsen, I., Gavrilov, M., Grebmeier, J. M., ... & Carmack, E. C. (2020). The pan-Arctic continental slope: sharp gradients of physical processes affect pelagic and benthic ecosystems. *Frontiers in Marine Science*. <https://doi.org/10.3389/fmars.2020.544386>
11. Boetius, A., Grahl, C., Kröncke, I., Liebezeit, G., & Nöthig, E. M. (1996). Distribution of plant pigments in surface sediments of the Eastern Arctic. *Reports of Polar Research*, 212, 213-218.
12. Boetius, A., Albrecht, S., Bakker, K., Bienhold, C., Felden, J., Fernández-Méndez, M., ... & ARK27, R. P. (2013). Export of algal biomass from the melting Arctic sea ice. *Science*, 339(6126), 1430-1432. <https://doi.org/10.1126/science.1231346>
13. Bray, J. R., Curtis, J. T., (1957). An ordination of the upland forest communities of southern Wisconsin. *Ecological monographs*, 27(4), 325-349. <https://doi.org/10.2307/1942268>
14. Brown, A., & Thatje, S. (2014). Explaining bathymetric diversity patterns in marine benthic invertebrates and demersal fishes: physiological contributions to adaptation of life at depth. *Biological Reviews*, 89(2), 406-426. <https://doi.org/10.1111/brv.12061>
15. Budaeva, N. E., Mokievsky, V. O., Soltwedel, T., & Gebruk, A. V. (2008). Horizontal distribution patterns in Arctic deep-sea macrobenthic communities. *Deep-Sea Research Part I: Oceanographic Research Papers*, 55(9), 1167-1178. <https://doi.org/10.1016/j.dsr.2008.05.002>
16. Carey Jr, A. G., Ruff, R. E. (1977). Ecological studies of the benthos in the western Beaufort Sea with special reference to bivalve molluscs. *Polar oceans*. Arctic Institute of North America, Calgary, 505-530.
17. Carney, R.S. (2005). Zonation of deep biota on continental margins. In: (Gibson, R.N., Atkinson, R.J.A., Gordon, J.D.M. (Eds.) *Oceanography and Marine Biology: an annual review*, 43, p. 211-278.
18. Chao, A., Chazdon, R. L., Colwell, R. K., & Shen, T. J. (2006). Abundance-based similarity indices and their estimation when there are unseen species in samples. *Biometrics*, 62(2), 361-371.
19. Chao, A., Gotelli, N. J., Hsieh, T. C., Sander, E. L., Ma, K. H., Colwell, R. K., & Ellison, A. M. (2014). Rarefaction and extrapolation with Hill numbers: a framework for sampling and estimation in species diversity studies. *Ecological monographs*, 84(1), 45-67.
20. Clarke, K., Warwick, R., (1994). *An Approach to Statistical Analysis and Interpretation*. PRIMER-E, Plymouth.
21. Clarke, K.R., Warwick, R.M. (2001). *Changes in Marine Communities: An Approach to Statistical Analysis and Interpretation* (2nd ed.). PRIMER-E, Plymouth.
22. Clarke, K. R., Somerfield, P. J., & Chapman, M. G., (2006). On resemblance measures for ecological studies, including taxonomic dissimilarities and a zero-adjusted Bray–Curtis coefficient for denuded assemblages. *Journal of Experimental Marine Biology and Ecology*, 330(1), 55-80. <https://doi.org/10.1016/j.jembe.2005.12.017>
23. Cochrane, S. K., Denisenko, S. G., Renaud, P. E., Emblow, C. S., Ambrose Jr, W. G., Ellingsen, I. H., & Skarðhamar, J. (2009). Benthic macrofauna and productivity regimes in the Barents Sea—ecological implications in a changing Arctic. *Journal of Sea Research*, 61(4), 222-233. <https://doi.org/10.1016/j.seares.2009.01.003>
24. Conlan, K., Hendrycks, E., Aitken, A., Williams, B., Blasco, S., & Crawford, E. (2013). Macrofaunal biomass distribution on the Canadian Beaufort Shelf. *Journal of Marine Systems*, 127, 76-87. <https://doi.org/10.1016/j.jmarsys.2013.07.013>
25. Dahle, S., Anisimova, N. A., Palerud, R., Renaud, P. E., Pearson, T. H., & Matishov, G. G. (2009). Macrobenthic fauna of the Franz Josef Land archipelago. *Polar Biology*, 32(2), 169-180. <https://doi.org/10.1007/s00300-008-0516-y>

- 734 26. Degen, R., Vedenin, A., Gusky, M., Boetius, A., Brey, T. (2014): Macrobenthic abundance, biomass, productivity and
735 production during POLARSTERN cruise ARK-VIII/3. PANGAEA, <https://doi.org/10.1594/PANGAEA.828340>.
- 736 27. Degen, R., Vedenin, A., Gusky, M., Boetius, A., & Brey, T. (2015). Patterns and trends of macrobenthic abundance,
737 biomass and production in the deep Arctic Ocean. *Polar Research*, 34(1), 24008. <https://doi.org/10.3402/polar.v34.24008>
- 738 28. Denisenko, S. G., Denisenko, N. V., Lehtonen, K. K., Andersin, A. B., & Laine, A. O. (2003). Macrozoobenthos of the
739 Pechora Sea (SE Barents Sea): community structure and spatial distribution in relation to environmental conditions.
740 *Marine Ecology Progress Series*, 258, 109-123. <https://doi.org/10.3354/meps258109>
- 741 29. Deubel, H. (2000). Struktureigenschaften und Nahrungsbedarf der Zoobenthosgemeinschaften im Bereich des
742 Lomonossowrückens im Arktischen Ozean. *Berichte zur Polarforschung (Reports on Polar Research)*, 370.
- 743 30. Deubel H., Engel M., Fetzer I., Gagaev S., Hirche H.J., Klages M., Larionov V., Lubin P., Lubina O., Nothig E.M.,
744 Okolodkov Y., Rachor E. (2003) The southern Kara Sea ecosystem: Phytoplankton, zooplankton and benthos
745 communities influenced by river run-off. In: Ruediger S. (ed.) *Siberian River Runoff in the Kara Sea: characterization,*
746 *quantification variability and environmental significance. Proceedings in Marine Science*, 6, Elsevier, Amsterdam, pp
747 237–275.
- 748 31. Dunbar, M.J. (1968). *Ecological development in polar regions. A study in evolution.* Englewood Cliffs, N. J., Prentice
749 Hall, VIII, 119 p.
- 750 32. Eleftheriou, A., McIntyre, A. (2005). *Methods for the study of marine benthos.* Blackwell Science, Oxford, UK, 418 p.
- 751 33. Filatova, Z. A., Zenkevich, L. A. (1957). The quantitative distribution of the bottom fauna of the Kara Sea. *Trudy*
752 *Vsesoyuznogo Gidrobiologicheskogo Obshchestva*, 8, 3-67. [In Russian]
- 753 34. Flach, E., & de Bruin, W. (1999). Diversity patterns in macrobenthos across a continental slope in the NE Atlantic. *Journal*
754 *of Sea Research*, 42(4), 303-323. [https://doi.org/10.1016/S1385-1101\(99\)00034-9](https://doi.org/10.1016/S1385-1101(99)00034-9)
- 755 35. Flint, M. V., Poyarkov, S. G., Rimskii-Korsakov, N. A., Mirosnikov, A. Y. (2019). Ecosystems of the Siberian Arctic
756 Seas 2018 (Cruise 72 of the R/V Akademik Mstislav Keldysh). *Oceanology*, 59(3), 460-463.
757 <https://doi.org/10.1134/S0001437019030056>
- 758 36. Fredriksen, R., Christiansen, J. S., Bonsdorff, E., Larsen, L. H., Nordström, M. C., Zhulay, I., & Bluhm, B. A. (2020).
759 Epibenthic megafauna communities in Northeast Greenland vary across coastal, continental shelf and slope habitats. *Polar*
760 *Biology*, 43(10), 1623-1642. <https://doi.org/10.1007/s00300-020-02733-z>
- 761 37. Fütterer, D. (1993). The expedition ARCTIC'93: Leg ARK-IX/4 of RV "Polarstern" 1993. *Reports for Polar Research*,
762 149.
- 763 38. Fütterer, D., Galimov, E.M. (2003). Siberian river run-off into the Kara Sea: Characterization, quantification, variability
764 and environmental significance – An introduction. In: Stein R., Fahl K., Fütterer, D., Stepanets O. (Eds.) *Siberian river*
765 *run-off in the Kara Sea.* Amsterdam: Elsevier Science B.V., pp. 1-9.
- 766 39. Gage J. (1986). The benthic fauna of the Rockall Trough: regional distribution and bathymetric zonation. *Proceedings of*
767 *Royal Society of Edinburgh*, 88b: 159–174. <https://doi.org/10.1017/S026972700000453X>
- 768 40. Gage, J. D., Lamont, P. A., Kroeger, K., Paterson, G. L., & Vecino, J. L. G. (2000). Patterns in deep-sea macrobenthos at
769 the continental margin: standing crop, diversity and faunal change on the continental slope off Scotland. In *Island, Ocean*
770 *and Deep-Sea Biology* (pp. 261-271). Springer, Dordrecht.
- 771 41. Galkin, S. V., & Vedenin, A. A. (2015). Macrobenthos of Yenisei Bay and the adjacent Kara Sea shelf. *Oceanology*,
772 55(4), 606-613. <https://doi.org/10.1134/S0001437015040086>
- 773 42. Gebruk, A. V., Budaeva, N. E., & King, N. J. (2010). Bathyal benthic fauna of the Mid-Atlantic Ridge between the Azores
774 and the Reykjanes Ridge. *Journal of the Marine Biological Association of the United Kingdom*, 90(1), 1-14.
775 <https://doi.org/10.1017/S0025315409991111>
- 776 43. Gerdes, D. (1990). Antarctic trials with the multibox corer, a new device for benthos sampling. *Polar Records*, 26, 35–
777 38.
- 778 44. Golikov, A. N., Dolgolenko, M. A., Maximovich, N. V., & Scarlato, O. A. (1990). Theoretical approaches to marine
779 biogeography. *Marine Ecology Progress Series.*, 63(2), 289-301.
- 780 45. Gorbunov G.P. (1946). Bottom life of the Novosiberian shoalwaters and the central part of the Arctic Ocean. In: Gorbunov
781 G.P., Uschakov P.V. (Eds.) *Proceedings of the drifting expedition of Glavsevmorput on ice-breaker “G. Sedov” in 1937-*
782 *1940. Glavsevmorput, Moscow-Leningrad*, pp. 30-138. [In Russian]
- 783 46. Górska, B., Soltwedel, T., Schewe, I., Włodarska-Kowalczyk, M., 2020. Bathymetric trends in biomass size spectra,
784 carbon demand, and production of Arctic benthos (76-5561 m, Fram Strait). *Progress in Oceanography* 102370.
785 <https://doi.org/10.1016/j.pocean.2020.102370>
- 786 47. Grassle, J. F., & Morse-Porteous, L. S. (1987). Macrofaunal colonization of disturbed deep-sea environments and the
787 structure of deep-sea benthic communities. *Deep-Sea Research Part A. Oceanographic Research Papers*, 34(12), 1911-
788 1950. [https://doi.org/10.1016/0198-0149\(87\)90091-4](https://doi.org/10.1016/0198-0149(87)90091-4)
- 789 48. Grebmeier, J. M., Cooper, L. W., Feder, H. M., Sirenko, B. I. (2006). Ecosystem dynamics of the Pacific-influenced
790 Northern Bering and Chukchi Seas in the Amerasian Arctic. *Progress in Oceanography*, 71, 331–361.
791 <http://dx.doi.org/10.1016/j.pocean.2006.10.001>
- 792 49. Grebmeier, J. M., Bluhm, B. A., Cooper, L. W., Denisenko, S. G., Iken, K., Kędra, M., & Serratos, C. (2015). Time-series
793 benthic community composition and biomass and associated environmental characteristics in the Chukchi Sea during the
794 RUSALCA 2004–2012 Program. *Oceanography*, 28(3), 116-133.
795 <http://dx.doi.org/10.5670/oceanog.2015.61>

- 796 50. Hammer, Ø., Harper, D.A.T., Ryan, P.D. (2001). PAST: Paleontological Statistics Software Package for Education and
797 Data Analysis. *Palaeontologia Electronica*, 4(1), 9pp., 178kb. http://palaeo-electronica.org/2001_1/past/issue1_01.htm.
798 51. Hay, W. W. (2016). "Continental slope," in *Encyclopedia of Marine Geosciences*, eds J. Harff, M. Meschede, S. Petersen,
799 and J. Thiede (Berlin: Springer). https://doi.org/10.1007/978-94-007-6644-0_156-3
800 52. Hedgpeth, J. W. (1957). *Marine biogeography. Treatise on marine ecology and paleoecology*, 1, 359-382.
801 53. Hessler, R. R., & Jumars, P. A. (1974). Abyssal community analysis from replicate box cores in the central North Pacific.
802 *Deep-Sea Research*, 21(18), 209. [https://doi.org/10.1016/0011-7471\(74\)90058-8](https://doi.org/10.1016/0011-7471(74)90058-8)
803 54. Høisæter, T. (2010). The shell-bearing, benthic gastropods on the southern part of the continental slope off Norway.
804 *Journal of Molluscan Studies*, 76(3), 234-244. <https://doi.org/10.1093/mollus/eyq003>
805 55. Horn, H. S. (1966). Measurement of "overlap" in comparative ecological studies. *The American Naturalist*, 100(914),
806 419-424.
807 56. Howell K. L., Billett D. S. M., Tyler P. A. (2002). Depth-related distribution and abundance of seastars (Echinodermata:
808 Asteroidea) in the Porcupine Seabight and Porcupine Abyssal Plain, N.E. Atlantic. *Deep-Sea Research. Part I*, 49: 1901–
809 1920. [https://doi.org/10.1016/S0967-0637\(02\)00090-0](https://doi.org/10.1016/S0967-0637(02)00090-0)
810 57. Hurlbert, S.H., (1971). The nonconcept of species diversity: a critique and alternative parameters. *Ecology* 52 (4), 577–
811 586. <https://doi.org/10.2307/1934145>
812 58. Jakobsson, M., Mayer, L., Coakley, B., Dowdeswell, J. A., Forbes, S., Fridman, B., et al. (2012). The international
813 bathymetric chart of the Arctic Ocean (IBCAO) version 3.0. *Geophysical Research Letters*, 39, L12609.
814 <https://doi.org/10.1029/2012GL052219>
815 59. Jansen, E., Christensen, J. H., Dokken, T., Nisancioglu, K. H., Vinther, B. M., Capron, E., ... & Stendel, M. (2020). Past
816 perspectives on the present era of abrupt Arctic climate change. *Nature Climate Change*, 10(8), 714-721.
817 <https://doi.org/10.1038/s41558-020-0860-7>
818 60. Jørgensen, L. L., Pearson, T. H., Anisimova, N. A., Gulliksen, B., Dahle, S., Denisenko, S. G., & Matishov, G. G. (1999).
819 Environmental influences on benthic fauna associations of the Kara Sea (Arctic Russia). *Polar Biology*, 22(6), 395-416.
820 <https://doi.org/10.1007/s003000050435>
821 61. Käß, M., Vedenin, A., Hasemann, C., Brandt, A., & Soltwedel, T. (2019). Community structure of macrofauna in the
822 deep Fram Strait: A comparison between two bathymetric gradients in ice-covered and ice-free areas. *Deep-Sea Research*
823 *Part I: Oceanographic Research Papers*, 152, 103102. <https://doi.org/10.1016/j.dsr.2019.103102>
824 62. Kędra, M., Renaud, P. E., Andrade, H., Goszczko, I., & Ambrose, W. G. (2013). Benthic community structure, diversity,
825 and productivity in the shallow Barents Sea bank (Svalbard Bank). *Marine Biology*, 160(4), 805-819.
826 <https://doi.org/10.1007/s00227-012-2135-y>
827 63. Kędra, M., Moritz, C., Choy, E. S., David, C., Degen, R., Duerksen, S., ... & Węśławski, J. M. (2015). Status and trends
828 in the structure of Arctic benthic food webs. *Polar Research*, 34(1), 23775. <https://doi.org/10.3402/polar.v34.23775>
829 64. Klages, M., Boetius, A., Christensen, J. P., Deubel, H., Piepenburg, D., Schewe, I., & Soltwedel, T. (2004). The benthos
830 of Arctic seas and its role for the organic carbon cycle at the seafloor. In: *The organic carbon cycle in the Arctic Ocean*
831 (pp. 139-167). Springer, Berlin, Heidelberg.
832 65. Krayushkina, A. B. (2000). *Geography of the asteroids and holothurians of the Norwegian Sea. Benthos of the Russian*
833 *Seas and the Northern Atlantic*. Moscow: VNIRO Publishing House, 41. [In Russian]
834 66. Krijgsman, W., Hilgen, F. J., Raffi, I., Sierro, F. J., & Wilson, D. S. (1999). Chronology, causes and progression of the
835 Messinian salinity crisis. *Nature*, 400(6745), 652-655. <https://doi.org/10.1038/23231>
836 67. Kröncke, I. (1994). Macrobenthos composition, abundance and biomass in the Arctic Ocean along a transect between
837 Svalbard and the Makarov Basin. *Polar Biology*, 14(8), 519-529. <https://doi.org/10.1007/BF00238221>
838 68. Kröncke, I. (1998). Macrofauna communities in the Amundsen Basin, at the Morris Jesup Rise and at the Yermak Plateau
839 (Eurasian Arctic Ocean). *Polar Biology*, 19(6), 383-392. <https://doi.org/10.1007/s003000050263>
840 69. Lisitsyn, A.P., Udintsev, G.B. (1955). A new model of a bottom grab. *Trudy Vsesoyuznogo Gidrobiologicheskogo*
841 *Obshchestva*, 6, 217–222. [In Russian]
842 70. MacDonald, I. R., Bluhm, B. A., Iken, K., Gagaev, S., & Strong, S. (2010). Benthic macrofauna and megafauna
843 assemblages in the Arctic deep-sea Canada Basin. *Deep-Sea Research Part II: Topical Studies in Oceanography*, 57(1-2),
844 136-152. <https://doi.org/10.1016/j.dsr2.2009.08.012>
845 71. Magurran, A. E. (2004). *Measuring biological diversity*. MA, USA: Blackwell Science Ltd.
846 72. Marshall, E. (2019). Kruskal-Wallis in SPSS. Community project encouraging academics to share statistics support
847 resources. Available at: [https://www.studocu.com/row/document/bahcesehir-ueniversitesi/managerial-skills-and-](https://www.studocu.com/row/document/bahcesehir-ueniversitesi/managerial-skills-and-strategic-management/lecture-notes/kruskal-wallis-test-manual/6221473/view)
848 [strategic-management/lecture-notes/kruskal-wallis-test-manual/6221473/view](https://www.studocu.com/row/document/bahcesehir-ueniversitesi/managerial-skills-and-strategic-management/lecture-notes/kruskal-wallis-test-manual/6221473/view).
849 73. McCune, B., Grace, J.B., Urban, D.L. (2002). *Analysis of ecological communities. Mjmm software design*. Oregon:
850 Gleneden Beach, 28.
851 74. Meltofte, H., Barry, T., Berteaux, D., Bültmann, H., Christiansen, J. S., Cook, J. A., Dahlberg, A., Daniëls, F., Ehrlich,
852 D., Fjeldså, J., Friðriksson, F., Ganter, B., Gaston A.J., Gillespie, L.J., Grenoble, L., Hoberg, E.P., Kodkinson, I.D.,
853 Huntington, H.P., Ims, R.A., Josefson, A.B., Kutz, S.J., Kuzmin, S.L., Laidre K.L., Lassuy, D.R., Lewis, P.N., Lovejoy,
854 C., Michel, C., Mokievsky, V., Mustonen, T., Payer, D.C., Poulin, M., Reid, D.G., Reist, J.D., Tessler, D.F., Wrona, F.J.
855 (2013). *Arctic Biodiversity Assessment. Synthesis. Conservation of Arctic Flora and Fauna (CAFF)*.
856 75. Menzies, R.J. (1963). The abyssal fauna of the sea floor of the Arctic Ocean. *Proceedings of Arctic Symposium*, Arctic
857 Institute of North America, Hershey, Penn., pp. 46-66.

- 858 76. Menzies, R. J. (1973). Biological history of the Mediterranean Sea with reference to the abyssal benthos. *Rapports de la*
859 *Commission Internationale pour l'Exploration Scientifique de la Mer Méditerranée*, 1, 717-723.
- 860 77. Meredith, M., Sommerkorn, M., Cassotta, S., Derksen, C., Ekaykin, A., Hollowed, A., & Kofinas, G. (2019). Polar
861 Regions: IPCC Special Report on the Ocean and Cryosphere in a Changing Climate. IPCC.
862 <https://www.ipcc.ch/srocc/#home-chapter-3>.
- 863 78. Mironov, A. N. (1986). Vertical zonation of the sea urchins. *Zoological Journal*, 65(9): 1341-1349. [In Russian]
- 864 79. Mironov, A. N. (2013). Biotic complexes of the Arctic Ocean. *Invertebrate Zoology*, 10(1), 3-48.
- 865 80. Nesis, K. N. (2001). West-Arctic and East-Arctic distributional ranges of cephalopods. *Sarsia*, 86(1), 1-11.
866 <https://doi.org/10.1080/00364827.2001.10420456>
- 867 81. Nilsen, R., & Holthe, T. (1985). Arctic and Scandinavian Oweniidae (Polychaeta) with a description of *Myriochele*
868 *fragilis* sp. n., and comments on the phylogeny of the family. *Sarsia*, 70(1), 17-32.
869 <https://doi.org/10.1080/00364827.1985.10420615>
- 870 82. Oleszczuk, B., Grzelak, K., Kędra, M. (2021). Community structure and productivity of Arctic benthic fauna across depth
871 gradients during springtime. *Deep-Sea Research Part I: Oceanographic Research Papers* 170: 103457.
872 <https://doi.org/10.1016/j.dsr.2020.103457>
- 873 83. Oug, E., Bakken, T., Kongsrud, J. A., Alvestad, T. (2017). Polychaetous annelids in the deep Nordic Seas: Strong
874 bathymetric gradients, low diversity and underdeveloped taxonomy. *Deep Sea Research Part II: Topical Studies in*
875 *Oceanography*, 137, 102-112. <https://doi.org/10.1016/j.dsr2.2016.06.016>
- 876 84. Parapar, J. (2006). The genera *Myriochele* and *Myrioglobula* (Polychaeta, Oweniidae) in Icelandic waters with the
877 revision of type material of *Myriochele heeri* Malmgren, 1867, and the description of a new species. *Journal of Natural*
878 *History*, 40(9-10), 523-547. <https://doi.org/10.1080/00222930600711758>
- 879 85. Paul, A. Z., & Menzies, R. J. (1974). Benthic ecology of the high Arctic deep sea. *Marine Biology*, 27(3), 251-262.
880 <https://doi.org/10.1007/BF00391950>
- 881 86. Petryashov, V. V., Golikov, A. A., Schmid, M., Rachor, E. (2004). Macrobenthos of the Laptev Sea shelf. *Explorations*
882 *of the fauna of the seas*, 54(63), 9-27. [In Russian]
- 883 87. Pielou, E.C., (1966). The measurement of diversity in different types of biological collections. *J. Theor. Biol.* 13, 131–
884 144. [https://doi.org/10.1016/0022-5193\(66\)90013-0](https://doi.org/10.1016/0022-5193(66)90013-0).
- 885 88. Piepenburg, D., Chernova, N. V., Von Dorrien, C. F., Gutt, J., Neyelov, A. V., Rachor, E., ... & Schmid, M. K. (1996).
886 Megabenthic communities in the waters around Svalbard. *Polar Biology*, 16(6), 431-446.
887 <https://doi.org/10.1007/s003000050074>
- 888 89. Piepenburg, D., Archambault, P., Ambrose, W. G., Blanchard, A. L., Bluhm, B. A., Carroll, M. L., ... & Włodarska-
889 Kowalczyk, M. (2011). Towards a pan-Arctic inventory of the species diversity of the macro-and megabenthic fauna of
890 the Arctic shelf seas. *Marine Biodiversity*, 41(1), 51-70.
- 891 90. Pisareva, M. N., Pickart, R. S., Spall, M. A., Nobre, C., Torres, D. J., Moore, G. W. K., & Whitledge, T. E. (2015). Flow
892 of Pacific water in the western Chukchi Sea: Results from the 2009 RUSALCA expedition. *Deep Sea Research Part I:*
893 *Oceanographic Research Papers*, 105, 53-73. <https://doi.org/10.1016/j.dsr.2015.08.011>
- 894 91. Rand, K., Logerwell, E., Bluhm, B., Chenelot, H., Danielson, S., Iken, K., & Sousa, L. (2018). Using biological traits and
895 environmental variables to characterize two Arctic epibenthic invertebrate communities in and adjacent to Barrow
896 Canyon. *Deep Sea Research Part II: Topical Studies in Oceanography*, 152, 154-169.
897 <https://doi.org/10.1016/j.dsr2.2017.07.015>
- 898 92. Ravelo, A. M., Bluhm, B. A., Foster, N., & Iken, K. (2020). Biogeography of epibenthic assemblages in the central
899 Beaufort Sea. *Marine Biodiversity*, 50(1), 1-19. <https://doi.org/10.1007/s12526-019-01036-9>
- 900 93. Renaud P.E., Carroll M.L., Ambrose W.G., Jr. Duarte C.M. (2008). Effects of global warming on Arctic sea-floor
901 communities and its consequences for higher trophic levels. *Impacts of global warming on polar ecosystems*. Bilbao:
902 Fundación BBVA. pp. 139–175.
- 903 94. Rex, M. A. (1981). Community structure in the deep-sea benthos. *Annual Review of Ecology and Systematics*, 12(1),
904 331-353. <https://doi.org/10.1146/annurev.es.12.110181.001555>
- 905 95. Rex, M. A., & Etter, R. J. (2010). *Deep-sea biodiversity: pattern and scale*. Harvard University Press.
- 906 96. Rowe, G. T. (1971). Benthic biomass in the Pisco, Peru upwelling. *Investigacion Pesquera*, 35(1), 127-135.
- 907 97. Rybakova, E., Kremenetskaia, A., Vedenin, A., Boetius, A., & Gebruk, A. (2019). Deep-sea megabenthos communities
908 of the Eurasian Central Arctic are influenced by ice-cover and sea-ice algal falls. *PloS one*, 14(7), e0211009.
909 <https://doi.org/10.1371/journal.pone.0211009>
- 910 98. Schlitzer, R. (2020). Ocean Data View. <https://odv.awi.de/>.
- 911 99. Schonberg, S. V., Clarke, J. T., & Dunton, K. H. (2014). Distribution, abundance, biomass and diversity of benthic infauna
912 in the Northeast Chukchi Sea, Alaska: Relation to environmental variables and marine mammals. *Deep-Sea Research*
913 *Part II: Topical Studies in Oceanography*, 102, 144-163. <https://doi.org/10.1016/j.dsr2.2013.11.004>
- 914 100. Shannon, C., Weaver, W., (1963). *The Measurement Theory of Communication*. University of Illinois Press, Urbana, IL.
- 915 101. Sirenko, B. I. (1998). Marine fauna of the Arctic (after the expeditions of the Zoological Institute, Russian Academy of
916 Sciences). *Russian Journal of Marine Biology*, 24(6), 353-364. [In Russian]
- 917 102. Sirenko, B., Denisenko, S., Deubel, H., & Rachor, E. (2004). Deep water communities of the Laptev Sea and adjacent
918 parts of the Arctic Ocean. *Fauna and the ecosystems of the Laptev Sea and adjacent deep waters of the Arctic Ocean*.
919 *Explorations of the fauna of sea*. St. Petersburg: Zoological Institute of Russian Academy of Sciences, 54(62), 28-73.

- 920 103. Sirenko B.I., Denisenko S.G. (2010). Fauna of the East-Siberian Sea, distribution patterns and structure of bottom
921 communities. Explorations of the fauna of the Seas, 66. Zoological Institute RAS, Saint-Petersburg, pp. 248 [In Russian]
922 104. Soltwedel, T., Jaeckisch, N., Ritter, N., Hasemann, C., Bergmann, M., & Klages, M. (2009). Bathymetric patterns of
923 megafaunal assemblages from the arctic deep-sea observatory HAUSGARTEN. Deep-Sea Research Part I:
924 Oceanographic Research Papers, 56(10), 1856-1872. <https://doi.org/10.1016/j.dsr.2009.05.012>
925 105. Sswat, M., Gulliksen, B., Menn, I., Sweetman, A. K., & Piepenburg, D. (2015). Distribution and composition of the
926 epibenthic megafauna north of Svalbard (Arctic). Polar Biology, 38(6), 861-877. <https://doi.org/10.1007/s00300-015-1645-8>
927 106. Svavarsson J., Brattegard T. and Strömberg J.-O. (1990). Distribution and diversity pattern of asellote isopods (Crustacea)
928 in the deep Norwegian and Greenland Seas. Progress in Oceanography, 24: 297-310. [https://doi.org/10.1016/0079-6611\(90\)90039-5](https://doi.org/10.1016/0079-6611(90)90039-5)
929 107. Thistle, D. (2003). The deep-sea floor: an overview. In: Tyler, P. (ed.) Ecosystems of the deep oceans. Elsevier,
930 Amsterdam, pp. 5-37
931 108. Thomson, D. H. (1982). Marine benthos in the eastern Canadian High Arctic: multivariate analyses of standing crop and
932 community structure. Arctic, 61-74.
933 109. Tselepidis, A., & Eleftheriou, A. (1992). South Aegean (Eastern Mediterranean) Continental Slope Benthos:
934 Macroinfaunal—Environmental Relationships. In Deep-sea food chains and the global carbon cycle (pp. 139-156).
935 Springer, Dordrecht. https://doi.org/10.1007/978-94-011-2452-2_9
936 110. Tselepidis, A., Papadopoulou, K. N., Podaras, D., Plaiti, W., & Koutsoubas, D. (2000). Macrobenthic community
937 structure over the continental margin of Crete (South Aegean Sea, NE Mediterranean). Progress in Oceanography, 46(2-
938 4), 401-428. [https://doi.org/10.1016/S0079-6611\(00\)00027-6](https://doi.org/10.1016/S0079-6611(00)00027-6)
939 111. Udalov, A., Chikina, M., Azovsky, A., Basin, A., Galkin, S., Garlitska, L., ... & Mokievsky, V. (2020). Integrity of benthic
940 assemblages along the arctic estuarine-coastal system. Ecological Indicators, 107115.
941 <https://doi.org/10.1016/j.ecolind.2020.107115>
942 112. Vedenin, A. A., Galkin, S. V., & Kozlovskiy, V. V. (2015). Macrobenthos of the Ob Bay and adjacent Kara Sea shelf.
943 Polar Biology, 38(6), 829-844. <https://doi.org/10.1007/s00300-014-1642-3>
944 113. Vedenin, A., Budaeva, N., Mokievsky, V., Pantke, C., Soltwedel, T., & Gebruk, A. (2016). Spatial distribution patterns
945 in macrobenthos along a latitudinal transect at the deep-sea observatory HAUSGARTEN. Deep-Sea Research Part I:
946 Oceanographic Research Papers, 114, 90-98. <https://doi.org/10.1016/j.dsr.2016.04.015>
947 114. Vedenin, A., Gusky, M., Gebruk, A., Kremenetskaia, A., Rybakova, E., & Boetius, A. (2018). Spatial distribution of
948 benthic macrofauna in the Central Arctic Ocean. PloS one, 13(10), e0200121.
949 <https://doi.org/10.1371/journal.pone.0200121>
950 115. Vedenin, A. A., Kokarev, V. N., Chikina, M. V., Basin, A. B., Galkin, S. V., & Gebruk, A. V. (2020). Fauna associated
951 with shallow-water methane seeps in the Laptev Sea. PeerJ, 8, e9018. <https://doi.org/10.7717/peerj.9018>
952 116. Vedenin, A., Galkin, S., Mironov, A. N., & Gebruk, A. (2021). Vertical zonation of the Siberian Arctic benthos:
953 bathymetric boundaries from coastal shoals to deep-sea Central Arctic. PeerJ, 9, e11640.
954 <https://doi.org/10.7717/peerj.11640>
955 117. Vermeij G.J. 1991. Anatomy of an invasion: the trans-Arctic interchange. Paleobiology 17:281-307.
956 118. Vinogradova N.G., (1962). Vertical zonation in the distribution of deep-sea benthic fauna in the ocean. Deep-Sea
957 Research, 8: 245-250. [https://doi.org/10.1016/0146-6313\(61\)90025-9](https://doi.org/10.1016/0146-6313(61)90025-9)
958 119. Wassmann, P., Duarte, C. M., Agusti, S., & Sejr, M. K. (2011). Footprints of climate change in the Arctic marine
959 ecosystem. Global Change Biology, 17(2), 1235-1249. <https://doi.org/10.1111/j.1365-2486.2010.02311.x>
960 120. Wassmann, P., Kosobokova, K. N., Slagstad, D., Drinkwater, K. F., Hopcroft, R. R., Moore, S. E., ... & Berge, J. (2015).
961 The contiguous domains of Arctic Ocean advection: trails of life and death. Progress in Oceanography, 139, 42-65.
962 <https://doi.org/10.1016/j.pocean.2015.06.011>
963 121. Wassmann, P. F., Slagstad, D., & Ellingsen, I. (2019). Advection of mesozooplankton into the northern Svalbard shelf
964 region. Frontiers in Marine Science, 6, 458. <https://doi.org/10.3389/fmars.2019.00458>
965 122. Watling, L., Guinotte, J., Clark, M. R., & Smith, C. R. (2013). A proposed biogeography of the deep ocean floor. Progress
966 in Oceanography, 111, 91-112. <https://doi.org/10.1016/j.pocean.2012.11.003>
967 123. Weber J.R. (1989). Physiography and bathymetry of the Arctic Ocean seafloor. In: Herman Y. (ed.). The Arctic Seas.
968 Clymatology, Oceanography, Geology, and Biology. New York: Van Nostrand Reinhold Company, pp. 797-828
969 124. Wei, C. L., Rowe, G. T., Escobar-Briones, E., Boetius, A., Soltwedel, T., Caley, M. J., ... & Narayanaswamy, B. E. (2010
970 a). Global patterns and predictions of seafloor biomass using random forests. PloS one, 5(12), e15323.
971 <https://doi.org/10.1371/journal.pone.0015323>
972 125. Wei, C. L., Rowe, G. T., Hubbard, G. F., Scheltema, A. H., Wilson, G. D., Petrescu, I., ... & Wang, Y. (2010 b).
973 Bathymetric zonation of deep-sea macrofauna in relation to export of surface phytoplankton production. Marine Ecology
974 Progress Series, 399, 1-14. <https://doi.org/10.3354/meps08388>
975 126. Williams, W. J., & Carmack, E. C. (2015). The 'interior' shelves of the Arctic Ocean: Physical oceanographic setting,
976 climatology and effects of sea-ice retreat on cross-shelf exchange. Progress in Oceanography, 139, 24-41.
977 <http://doi.org/10.1016/j.pocean.2015.07.008>
978 127. Wlodarska-Kowalczyk, M., Kendall, M. A., Weslawski, J. M., Klages, M., & Soltwedel, T. (2004). Depth gradients of
979 benthic standing stock and diversity on the continental margin at a high-latitude ice-free site (off Spitsbergen, 79 N).
980 Deep-Sea Research Part I: Oceanographic Research Papers, 51(12), 1903-1914. <https://doi.org/10.1016/j.dsr.2004.07.013>
981
982

983
984
985

128. Zhulay, I., Iken, K., Renaud, P. E., & Bluhm, B. A. (2019). Epifaunal communities across marine landscapes of the deep Chukchi Borderland (Pacific Arctic). *Deep-Sea Research Part I: Oceanographic Research Papers*, 151, 103065. <https://doi.org/10.1016/j.dsr.2019.06.011>

986 **Supplementary captions**

987
988 Supplementary 1. Station data used in this study with the information about expedition names,
989 geographic regions, dates, station names, latitude, longitude, depths and sampling gears.

990
991 Supplementary 2. Python 3.8 script for calculating Hill numbers, the Morisita-Horn and quantitative
992 Sørensen similarity indices.

993
994 Supplementary 3. The values of abundance, biomass, number of species, Pielou's evenness, ES-100
995 and Shannon index for each station.

996
997 Supplementary 4. List of individual taxa abundances per station for quantitative samples.

998
999 Supplementary 5. Non-metric multidimensional scaling based on Bray-Curtis similarity index and
000 dendrogram plots based on Morisita-Horn and quantitative Sørensen similarity indices for
001 quantitative and semi-quantitative samples.

002
003 Supplementary 6. List of individual taxa abundances per station for semi-quantitative samples.

Table S1, continued

Expedition	Region	Date	Event	Station	Latitude	Longitude	Depth (m)	Gear	Sampling area (m ²)
ARK IX/4	Laptev Sea	1993	PS2468	1993-50	77.6900	125.9183	1992	Agassiz trawl	-
AMK-54	Kara Sea	2007	AMK-4983	2007-4983	76.9200	70.2700	555	Sigsbee trawl	-
AMK-54	Kara Sea	2007	AMK-4985	2007-4985	76.7833	70.6167	465	Sigsbee trawl	-
AMK-54	Kara Sea	2007	AMK-4987	2007-4987	76.6583	71.0483	275	Sigsbee trawl	-
AMK-54	Kara Sea	2007	AMK-4988	2007-4988	76.5883	71.2567	160	Sigsbee trawl	-
AMK-54	Kara Sea	2007	AMK-4990	2007-4990	76.1533	72.4967	110	Sigsbee trawl	-
AMK-54	Kara Sea	2007	AMK-4996	2007-4996	72.5718	73.7862	20	Sigsbee trawl	-
AMK-54	Kara Sea	2007	AMK-4999	2007-4999	72.9565	73.2947	27	Sigsbee trawl	-
AMK-54	Kara Sea	2007	AMK-5000	2007-5000	73.7530	72.9423	23	Sigsbee trawl	-
AMK-54	Kara Sea	2007	AMK-5003	2007-5003	75.4430	72.5155	60	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5016	2011-5016	72.5503	80.3420	14	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5019	2011-5019	73.1699	79.8608	25	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5020	2011-5020	73.7176	79.3896	29	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5010	2011-5010	74.2930	78.6251	33	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5024	2011-5024	74.9486	77.9020	34	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5026	2011-5026	75.9970	76.6741	63	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5032	2011-5032	76.5498	80.7481	57	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5033	2011-5033	77.2100	78.1277	120	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5034	2011-5034	77.4263	77.5674	220	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5039	2011-5039	78.0074	74.8968	364	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5042	2011-5042	78.4915	72.8047	472	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5051	2011-5051	75.8272	68.9850	351	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5054	2011-5054	72.9301	58.3447	376	Sigsbee trawl	-
AMK-59	Kara Sea	2011	AMK-5061	2011-5061	71.6692	57.0368	325	Sigsbee trawl	-
ARK XXVII/3	Central Arctic	2012	PS80/205	2012-205	81.4802	31.0252	615	Agassiz trawl	-
ARK XXVII/3	Central Arctic	2012	PS80/222	2012-222	84.0377	30.1620	4012	Agassiz trawl	-
ARK XXVII/3	Central Arctic	2012	PS80/249	2012-249	83.9673	77.6813	3470	Agassiz trawl	-
ARK XXVII/3	Central Arctic	2012	PS80/259	2012-259	82.7090	109.5777	3575	Agassiz trawl	-
ARK XXVII/3	Central Arctic	2012	PS80/286	2012-286	82.7780	129.8467	4159	Agassiz trawl	-
ARK XXVII/3	Central Arctic	2012	PS80/332	2012-332	81.9062	130.8432	4039	Agassiz trawl	-
ARK XXVII/3	Central Arctic	2012	PS80/346	2012-346	85.0688	122.6915	4354	Agassiz trawl	-
ARK XXVII/3	Central Arctic	2012	PS80/359	2012-359	87.8922	59.3887	4380	Agassiz trawl	-
PSh-125	Kara Sea	2013	125-30	2013-30	76.3533	88.8250	47	Sigsbee trawl	-
PSh-125	Kara Sea	2013	125-32	2013-32	77.1187	87.6292	149	Sigsbee trawl	-
PSh-125	Kara Sea	2013	125-34	2013-34	78.0150	87.6317	108	Sigsbee trawl	-
AMK-63	Laptev Sea	2015	AMK-5230	2015-seep	76.7715	125.8418	72	Sigsbee trawl	-
AMK-63	Laptev Sea	2015	AMK-5225	2015-5225	78.3747	130.6585	2390	Sigsbee trawl	-

Suppl 2

```
#####
```

```
''' Defenition of functions for diversity measurements.
```

```
Input - np.array of list of species abundances in one sample'''
```

```
import sys
```

```
import numpy as np
```

```
import scipy.stats
```

```
from math import comb
```

```
import random
```

```
def SpRich(x): #Number of species
```

```
    a = np.count_nonzero(x)
```

```
    return a
```

```
def sdi(x): #Shannon-Wiener index
```

```
    from math import log as ln
```

```
    def p(n, N):
```

```
        if n == 0:
```

```
            return 0
```

```
        else:
```

```
            return (float(n)/N) * ln(float(n)/N)
```

```
    N = sum(x)
```

```
    return -sum(p(n, N) for n in x)
```

```
def Hill(x,q): #Simple Hill number
```

```
    N = sum(x)
```

```
    def p(i):
```

```
        if i == 0:
```



```

    return 0
else:
    return (i/N)**q
if 1 !=0:
    return sum(p(i) for i in x)**(1/(1-q))
else: return sdi(x)

```

def Dm_0 (x): #Observed interpolated species rarefaction, $q = 0$, see Chao et al. 2014 for equations

```

n = round(sum(x))
S = SpRich(x)
l = []
noz = np.array([a for a in x if a != 0])
for m in range(1,n+1):
    a_list = []
    for i in range(len(noz)):
        a = comb((n-int(noz[i])),m)/comb(n,m)
        a_list.append(a)
    z = (sum(a_list))
    Dm = S - z
    l.append(Dm)
return np.array(l)

```

def Cind_0 (x): #Observed interpolated species sample coverage, $q = 0$

```

n = round(sum(x))
l = []
noz = np.array([a for a in x if a != 0])
for m in range(1,n-1):
    a_list = []
    for i in range(len(noz)):

```

```

    a = (int(noz[i])/n)*comb((n-int(noz[i])),m)/comb((n-1),m)

    a_list.append(a)

    z = (sum(a_list))

    Dm = 1 - z

    l.append(Dm)

return np.array(l)

```

def Dm_1(x,y): #Observed extrapolated species rarefaction, q = 0; y (individuals) - the upper limit of the extrapolation

```

    n = round(sum(x))

    S = SpRich(x)

    f1 = np.count_nonzero(x == 1)

    f2 = np.count_nonzero(x == 2)

    if f1 == 0:

        f0 = 0

    elif f1 != 0 and f2 == 0:

        f0 = ((n-1)/n)*f1*(f1-1)/2

    else:

        f0 = ((n-1)/n)*f1**2/(2*f2)

    l = []

    for m in range (0,y):

        if f0 ==0:

            z = 0

        else:

            z = f1/(n*f0+f1)

        Dnm = S + f0*(1-(1-z)**m)

        l.append(Dnm)

return np.array(l)

```

def Cind_1(x): #Observed extrapolated species sample coverage, q = 0

```

n = round(sum(x))
f1 = np.count_nonzero(x == 1)
f2 = np.count_nonzero(x == 2)
l = []
for m in range (0,n+1):
    Ci = 1-(f1/n)*(((n-1)*f1/(2*f2+f1*(n-1)))**m)
    l.append(Ci)
return np.array(l)

```

def pi_tuned(x): #Tuned probabilities of species occurrence in sample x

```

n = round(sum(x))
f1 = np.count_nonzero(x == 1)
f2 = np.count_nonzero(x == 2)
if f2 == 0:
    f0 = ((n-1)/n)*f1*(f1-1)/2
else:
    f0 = ((n-1)/n)*f1**2/(2*f2)
CindN = 1-(f1/n)*(((n-1)*f1/(2*f2+f1*(n-1)))
a_list = []
for i in range(len(x)):
    denom1 = (x[i]/n)*(1-x[i]/n)**n
    a_list.append(denom1)
denom = (sum(a_list))
l=[]
for i in range(len(x)):
    if x[i] != 0:
        lamb = (1-CindN)/denom
        p_i = (x[i]/n)*(1-lamb*((1-x[i]/n)**n))
    else:

```

```

    p_i = ((1-CindN)/(round(f0)))**2
    l.append(p_i)
return l

```

```

def pi_tuned_rev(x): #Reversed tuned probabilities for further randomization

```

```

    n = round(sum(x))
    f1 = np.count_nonzero(x == 1)
    f2 = np.count_nonzero(x == 2)
    if f2 == 0:
        f0 = ((n-1)/n)*f1*(f1-1)/2
    else:
        f0 = ((n-1)/n)*f1**2/(2*f2)
    CindN = 1-(f1/n)*((n-1)*f1/(2*f2+f1*(n-1)))
    a_list = []
    for i in range(len(x)):
        denom1 = (x[i]/n)*(1-x[i]/n)**n
        a_list.append(denom1)
    denom = (sum(a_list))
    l=[]
    for i in range(len(x)):
        if x[i] != 0:
            lamb = (1-CindN)/denom
            p_i = 1-(x[i]/n)*(1-lamb*((1-x[i]/n)**n))
        else:
            p_i = 1-((1-CindN)/(round(f0)))**2
        l.append(p_i)
    return l

```

```

def rand_gen(x,y): #Creates DataFrame with randomly generated samples of y replicates

```

```

population = x
sp_list = len(x)
prob_distr = pi_tuned_rev(x)
columns = np.array(range(1,len(x)))

rand_frame = pd.DataFrame({0: np.array(random.choices(population,
weights=prob_distr, k=sp_list))})

for i in range (1,y):

    rand_frame[i] = np.array(random.choices(population, weights=prob_distr, k=sp_list))

return rand_frame

```

```

def rand_df_curve(x,y,z): #Creates DataFrame with generated rarefactions up to y
individuals of z random replicates

```

```

    col_0 = rand_gen(x,z)[0]
    N_0 = (round(sum(col_0)))
    if N_0 >= y:
        curve_frame = pd.DataFrame({0: Dm_0(col_0)[:y]})
    elif N_0 < y:
        curve_frame = pd.DataFrame({0: np.concatenate((Dm_0(col_0),Dm_1(col_0, y-
N_0)[1:]),axis=None)})
    for i in range (1,z):
        col_i = rand_gen(x,z)[i]
        N_i = (round(sum(col_i)))
        if N_i >= y:
            curve_frame[i] = Dm_0(col_i)[:y]
        elif N_i < y:
            new_col1 = Dm_0(col_i)
            new_col2 = Dm_1(col_i, y-N_i)[1:]
            curve_frame[i] = np.concatenate((new_col1, new_col2), axis=None)
    return curve_frame

```

```

def confidence_interval(data, confidence): # To calculate Confidence interval

```

```

a = 1.0 * np.array(data)

n = len(a)

m, se = np.mean(a), scipy.stats.sem(a)

h = se * scipy.stats.t.ppf((1 + confidence) / 2., n-1)

return h

```

```

def mean_interval(data, confidence): # To calculate mean

```

```

    a = 1.0 * np.array(data)

    n = len(a)

    m, se = np.mean(a), scipy.stats.sem(a)

    h = se * scipy.stats.t.ppf((1 + confidence) / 2., n-1)

    return m

```

```

def mean(x,y): # mean curve up to y individuals, concatenated Dm_0 and Dm_1 in case you
deal with large samples

```

```

    N = (round(sum(x)))

    if N >= y:

        return Dm_0(x)[:y]

    else:

        return np.concatenate((Dm_0(x), Dm_1(x, y-N)[1:]), axis=None)

```

```

def CI_rarefaction_minus(x,y, confidence): # Lower confidence interval of rarefaction curve

```

```

    fr = rand_df_curve(x,y)

    N = (round(sum(x)))

    mean = np.concatenate((Dm_0(x), Dm_1(x, 100-N)[1:]), axis=None)

    y_graph = mean - confidence_interval(fr.T, confidence)

    return y_graph

```

```

def CI_rarefaction_plus(x,y, confidence): # Upper confidence interval of rarefaction curve

```

```

    fr = rand_df_curve(x,y)

```

```
N = (round(sum(x)))
```

```
mean = np.concatenate((Dm_0(x), Dm_1(x, 100-N)[1:]), axis=None)
```

```
y_graph = mean + confidence_interval(fr.T, confidence)
```

```
return y_graph
```

'''An example of the rarefaction curve for one of the stations (1995-89) used in this study'''

```
_1995_89_ = array([0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 4,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 7, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 1, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 7, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 1, 0, 0, 0, 0,
```

```
0, 0, 1, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 1, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 1, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 1, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 4, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 2, 0, 0, 0, 0, 0, 0, 0, 0, 0,
```

```
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 3, 0])
```

```
N = (round(sum(_1995_89_)))
```

```
S = SpRich(_1995_89_)
```

```
x1 = np.array(list(range(1,N+1)))
```

```
y1 = Dm_0(_1995_89_)
```

```
x2 = np.array(list(range(N,100)))
```

```
y2 = Dm_1(_1995_89_, 100-N)
```

```
x3 = N
```

```
y3 = S
```

```
fr = rand_df_curve(_1995_89_,100,10)
```

```
mean = np.concatenate((Dm_0(_1995_89_), Dm_1(_1995_89_, 100-N)[1:]), axis=None)
```

```
ci = confidence_interval(fr.T, 0.99)
```

```
x4 = np.array(list(range(1,100)))
```

```
y4 = mean-ci
```

```
y5 = mean+ci
```

```
#
```

```
'''
```

Or, in case of large samples when the sum of individuals exceeds usual values of 50 or 100, use this:

```
ci = confidence_interval(fr.T, 0.99)
```

```
mean_f = mean(_1995_89_, 100)
```

```
x4 = np.array(list(range(1,101)))
```



```

y4 = mean_f-ci
y5 = mean_f+ci
'''
#

fig = plt.figure(figsize =(15, 7))

plt.plot(x4,y4, color='orange', linewidth=2)
plt.plot(x4,y5, color='orange', linewidth=2)
plt.fill_between(x4, y4, y5, color='navajowhite')

plt.plot(x1,y1, color='darkorange', linewidth=4)
plt.plot(x2,y2, color='darkorange', linewidth=4, linestyle='dashed')
plt.plot(x3,y3, color='darkorange', marker='o', markersize=12)

plt.title('Rarefaction curve and extrapolation')
plt.ylim(0.5,25)
plt.xlim(1,50)
plt.grid(True)
plt.xlabel('Number of individuals')
plt.ylabel('Species count')
plt.show

#####

''' Defenition of similarity indices for cluster dendrograms.
Input - np.array of list of species abundances in each sample or pandas DataFrame'''

import pandas as pd

```

```

import numpy as np

import seaborn as sns

import matplotlib

import matplotlib.pyplot as plt

import scipy

import sklearn

import math

import os, re, sys

from math import log

def Br_Cur(data1, data2): #Bray-Curtis similarity index
    if len(data1) != len(data2):
        raise ValueError("Error while calculating Bray-Curtis similarity index. The two input
data lists must have the same length!\n")
    if sum(data1) == 0 or sum(data2) == 0:
        return -1
    A = sum(data1)
    B = sum(data2)
    z = 0
    for i, (x,y) in enumerate(zip(data1, data2)):
        if x!=0 and x<=y:
            z+=x
        elif y!=0 and x>=y:
            z+=y
    minAB = z
    if float(A+B) == 0:
        return 0
    else:
        return (2*minAB)/(A+B)

```

```
def rev_Br_Cur(data1, data2): #Bray-Curtis dissimilarity
```

```
    return (1-Br_Cur(data1, data2))
```

```
'''
```

```
the script partly taken from  
https://github.com/ngannguyen/immunoseq/blob/master/src/similarity.py
```

```
Jun 13 2012
```

```
'''
```

```
def morisitaHorn(data1, data2): #Morisita-Horn similarity index
```

```
    S1 = sum(data1)
```

```
    S2 = sum(data2)
```

```
    if len(data1) != len(data2):
```

```
        raise ValueError("Error while calculating MorisitaHorn similarity index. The two  
input data lists must have the same length!\n")
```

```
    if S1 == 0 or S2 == 0:
```

```
        return -1
```

```
    sum_xy = 0
```

```
    sum_x_sq = 0
```

```
    sum_y_sq = 0
```

```
    for i, x in enumerate(data1):
```

```
        y = data2[i]
```

```
        sum_xy += x*y
```

```
        sum_x_sq += x*x
```

```
        sum_y_sq += y*y
```

```
    n = 2*sum_xy/(S1*S2)
```

```
    d = (float(sum_x_sq)/(S1**2) + float(sum_y_sq)/(S2**2))
```

```
    return float(n)/d
```

```
def reversed_morisitaHorn(data1,data2): #Morisita-Horn dissimilarity
```

```
return (1-morisitaHorn(data1, data2))
```

```
def SoerAbIn(data1, data2): #Quantitative Soerensen similarity index
```

```
    if len(data1) != len(data2):
```

```
        raise ValueError("Error while calculating Soerensen similarity index. The two input  
data lists must have the same length!\n")
```

```
    if sum(data1) == 0 or sum(data2) == 0:
```

```
        return -1
```

```
    A = sum(data1)
```

```
    a = 0
```

```
    for i, (x,y) in enumerate(zip(data1, data2)):
```

```
        if y!=0:
```

```
            a+=x
```

```
    U = a/A
```

```
    B = sum(data2)
```

```
    b = 0
```

```
    for i, (x,y) in enumerate(zip(data1, data2)):
```

```
        if x!=0:
```

```
            b+=y
```

```
    V = b/B
```

```
    if float(U+V) == 0:
```

```
        return 0
```

```
    else:
```

```
        return float(2*U*V)/float(U+V)
```

```
def rev_SoerAbIn(data1, data2): #Quantitative Soerensen dissimilarity index
```

```
    return (1-SoerAbIn(data1, data2))
```

```
# Example of the dendrogram using Morisita-Horn similarity index.
```

#Input - DataFrame (named here as 'frame') with species as index names and stations as columns names

```
from scipy.spatial.distance import pdist, squareform
```

```
station_list = list(frame.columns)
```

```
sim_matr      =      pd.DataFrame(squareform(pdist(frame.T,      morisitaHorn)),  
columns=station_list, index=station_list) #non-transformed data
```

```
sim_matr2     =      pd.DataFrame(squareform(pdist(frame_trans.T, lambda u, v:  
morisitaHorn(u,v))), columns=station_list, index=station_list) #square-root transformed  
data
```

```
from scipy.cluster import hierarchy
```

```
ytdist = np.array(sim_matr2)
```

```
Z      =      hierarchy.linkage(ytdist,      'average',      metric=reversed_morisitaHorn,  
optimal_ordering=True)
```

```
plt.figure(figsize=(15, 20))
```

```
matplotlib.rcParams['lines.linewidth'] = 1
```

```
dn = hierarchy.dendrogram(Z,  
    p=100,  
    truncate_mode='level',  
    orientation='left',  
    labels=stations,  
    count_sort='ascending',  
    distance_sort='ascending',
```

```
show_leaf_counts=True,  
leaf_rotation=0,  
leaf_font_size=10,  
above_threshold_color='black')
```

```
plt.title('Cluster Analysis, Morisita-Horn index, sq-root transform, group average',  
fontdict={'fontsize':20}, pad=12);
```

```
plt.xlim()
```

```
plt.xticks(fontsize=12)
```

```
plt.xlabel('Morisita-Horn dissimilarity', fontsize=15)
```

```
plt.axvline(x=0.435, c='grey', lw=2, linestyle='dashed')
```

```
plt.savefig('Morisita_Horn_sq-root_average_stations.png', dpi=300, bbox_inches='tight')
```

Suppl. 3

Station	Abundance (ind. m ⁻²)	Biomass (g ww m ⁻²)	Species number	Pielou's evenness	ES(100)	Estimated diversity of 50 individuals	Shannon- Wiener index
Quantitative samples							
1991-2213	856	5.785	53	0.85	33.60	25.02	3.36
1991-2214	3292	10.701	92	0.78	35.91	24.84	3.52
1993-14	1051	1.633	26	0.80	20.88	17.50	2.60
1993-17	261	0.647	5	0.97	5.00	5.00	1.56
1993-19	522	0.196	21	0.93	20.31	26.13	2.84
1993-20	565	2.083	8	0.96	8.00	10.43	1.99
1993-24	891	2.416	19	0.89	18.20	21.16	2.61
1993-25	8861	650.529	77	0.80	33.38	24.04	3.46
1993-27	2487	8.959	68	0.89	39.98	29.34	3.75
1993-32	948	1.577	26	0.84	21.42	18.65	2.73
1993-35	1080	7.482	47	0.86	29.85	23.31	3.30
1993-38	1552	6.442	59	0.88	37.13	28.14	3.60
1993-39	2905	7.118	75	0.84	38.43	27.43	3.64
1993-40	4122	27.490	77	0.90	44.31	31.57	3.93
1993-41	6500	26.817	54	0.81	32.87	24.20	3.24
1993-44	7868	340.169	77	0.87	40.13	29.05	3.77
1993-47	915	11.832	91	0.83	41.47	25.45	3.76
1993-48	3888	35.869	90	0.84	40.96	28.29	3.76
1993-49	2914	16.855	76	0.82	37.88	26.53	3.56
1993-50	652	7.902	26	0.87	23.75	23.62	2.83
1993-53	149	3.261	27	0.91	26.38	27.74	2.99
1993-54	435	0.652	11	0.85	10.88	13.54	2.03
1993-64	5103	34.162	100	0.79	37.37	26.10	3.64
1993-68	4858	51.813	99	0.80	38.52	26.73	3.66
1993-69	4898	43.538	109	0.81	41.55	28.42	3.82
1993-70	1147	4.090	76	0.80	38.00	26.60	3.47
1993-71	3021	4.754	52	0.67	25.57	18.18	2.65
1995-89	66	2.300	12	0.88	12.00	14.82	2.19
1995-90	474	2.400	34	0.80	23.19	17.95	2.80
1995-91	752	28.100	73	0.81	35.74	24.96	3.47
1995-92	2206	818.800	89	0.71	32.15	21.66	3.17
1995-93	3897	2316.500	120	0.74	37.32	24.66	3.54
1995-94	2040	13.800	37	0.53	17.48	12.29	1.92
1995-95	750	25.200	73	0.81	38.03	25.93	3.46
1995-96	884	44.900	46	0.78	26.43	19.55	2.99
1997-2830	4092	23.848	26	0.85	20.98	18.91	2.77
1997-2831	3454	20.856	47	0.82	28.80	23.93	3.18
1997-2832	440	2.288	6	0.90	6.00	12.37	1.61
1997-2833	88	0.198	2	1.00	2.00	2.00	0.69
1997-2834	704	2.970	7	0.95	7.00	7.92	1.84
1997-2835	440	0.682	8	0.95	8.00	22.49	1.97
1997-2836	550	2.167	11	0.72	10.92	26.49	1.73
1997-2837	946	12.606	20	0.89	18.91	29.80	2.67
1997-2838	704	2.090	10	0.90	9.99	14.76	2.06
1997-2839	119	0.469	6	0.93	6.00	6.00	1.67
1997-2840	704	2.754	6	0.70	6.00	11.04	1.25
1997-2843	264	0.986	4	0.90	4.00	6.50	1.24
1997-2847	132	0.079	2	0.92	2.00	2.33	0.64
1997-2849	30	0.421	2	1.00	2.00	2.50	0.69
1997-2851	220	5.918	4	0.96	4.00	7.60	1.33
1997-2853	572	9.649	9	0.97	9.00	11.86	2.14
1997-2854	836	1.426	10	0.90	9.98	15.76	2.08
1997-2855	484	1.377	9	0.96	9.00	25.97	2.10
1997-2859	704	1.685	16	0.87	15.28	16.58	2.42
1997-2860	308	1.087	5	0.96	5.00	6.93	1.55
1997-2861	440	4.770	4	0.92	4.00	4.45	1.28
1997-2868	83	0.316	2	0.83	2.00	2.00	0.58
2007-5003	2174	48.835	54	0.73	25.29	18.77	2.91
2012-229	12	0.008	2	1.00	2.00	2.00	0.69
2012-241	46	0.411	6	0.93	6.00	12.52	1.66
2012-262	21	0.070	2	0.92	2.00	2.33	0.64
2012-278	209	1.212	12	0.83	11.87	15.06	2.06
2012-326	97	0.147	10	0.92	10.00	13.41	2.12
2012-339	78	0.081	9	0.95	9.00	16.18	2.10
2012-355	20	0.087	3	0.95	3.00	4.50	1.04
2012-368	57	0.071	6	0.88	6.00	11.01	1.58
2012-N5	1168	1.816	29	0.87	23.14	19.62	2.93
2012-N4	1592	2.509	34	0.78	22.07	17.37	2.74
2012-N3	1152	0.971	30	0.84	22.67	19.01	2.86
2012-N2	1008	1.478	27	0.81	21.48	18.08	2.67
2012-N1	1024	3.592	31	0.84	24.75	20.74	2.89
2012-HGIVa	984	2.628	27	0.76	20.68	20.41	2.52
2012-HGIVa	928	1.794	29	0.86	23.81	17.14	2.89
2012-51a	1456	2.380	32	0.82	23.05	18.69	2.86
2012-51	880	0.866	25	0.81	19.80	16.99	2.62
2012-52	1064	2.205	37	0.86	27.78	22.89	3.12
2012-53	1360	4.975	31	0.87	24.64	20.53	2.98
2015-5225	219	1.051	25	0.73	18.32	14.75	2.35
2015-5227	333	2.514	37	0.82	26.00	19.80	2.95
2016-EG1	764	0.452	51	0.82	33.50	24.76	3.24
2016-EGII	384	1.349	40	0.87	31.99	26.30	3.21
2016-EGIII	277	0.924	28	0.85	25.14	22.97	2.84
2016-EGIV	992	1.048	37	0.78	23.68	18.00	2.80
2016-HGI	1450	2.707	58	0.84	33.16	24.59	3.40
2016-HGII	1479	2.358	65	0.80	33.62	23.71	3.33
2016-HGIII	552	0.861	35	0.87	27.94	22.85	3.11
2016-HGIV	756	2.455	29	0.70	19.80	15.27	2.35
2002-AG05	4050	4.950	27	0.70	19.64	15.90	2.30
2002-AL07	675	2.600	14	0.93	13.87	19.47	2.45
2002-AL10	88	0.004	4	0.92	4.00	5.71	1.28
2002-NA05	1792	0.808	35	0.79	23.14	18.26	2.82
2002-NW01	1875	4.325	11	0.56	9.30	8.68	1.35
2002-NW05	1275	0.536	16	0.63	11.38	9.52	1.74
2005-2	2489	27.000	50	0.62	22.36	19.90	2.41
2005-3	833	0.100	18	0.76	16.01	20.06	2.20
2005-4	561	11.200	34	0.88	28.57	13.33	3.12
2005-5	339	2.400	8	0.48	7.60	11.36	1.00
2005-7	67	0.050	8	0.93	8.00	15.59	1.94
2005-8	128	0.050	10	0.90	10.00	14.06	2.07
2005-9	111	0.200	10	0.97	10.00	24.09	2.22
2005-11	239	3.700	17	0.90	16.67	7.43	2.55
2005-12	250	2.000	18	0.79	17.41	18.21	2.27
2005-13	167	1.800	9	0.77	8.98	13.49	1.69
2005-15	606	1.800	14	0.70	12.36	10.85	1.84

Suppl 3 continued

Semiquantitative samples							
Station	Abundance (ind. m ⁻²)	Biomass (g ww m ⁻²)	Species number	Pielou's evenness	ES(100)	Estimated diversity of 50 individuals	Shannon- Wiener index
1993-50	-	-	27	0.73	17.21	13.52	2.42
2007-4983	-	-	57	0.45	18.94	12.52	1.81
2007-4985	-	-	144	0.48	22.68	14.96	2.37
2007-4987	-	-	48	0.49	16.03	11.24	1.92
2007-4988	-	-	70	0.32	12.29	8.82	1.37
2007-4990	-	-	89	0.67	30.58	19.92	3.02
2007-4996	-	-	31	0.23	9.05	6.27	0.79
2007-4999	-	-	38	0.58	18.47	13.28	2.11
2007-5000	-	-	38	0.57	17.66	13.39	2.08
2007-5003	-	-	99	0.61	27.14	18.29	2.82
2011-5016	-	-	21	0.46	9.50	7.13	1.40
2011-5019	-	-	52	0.32	11.91	8.08	1.27
2011-5020	-	-	76	0.40	16.98	10.58	1.73
2011-5010	-	-	126	0.64	29.36	20.26	3.12
2011-5024	-	-	125	0.54	28.65	17.93	2.61
2011-5026	-	-	73	0.73	31.85	22.03	3.15
2011-5032	-	-	88	0.64	33.24	20.52	2.89
2011-5033	-	-	122	0.69	34.47	23.06	3.31
2011-5034	-	-	88	0.46	19.68	12.56	2.06
2011-5039	-	-	67	0.35	16.48	10.74	1.49
2011-5042	-	-	104	0.72	35.08	22.82	3.33
2011-5051	-	-	103	0.46	16.90	11.37	2.15
2011-5054	-	-	80	0.45	17.29	11.03	1.99
2011-5061	-	-	79	0.43	11.82	9.08	1.86
2012-205	-	-	133	0.71	39.25	24.19	3.48
2012-222	-	-	22	0.48	14.25	10.96	1.49
2012-249	-	-	24	0.52	14.78	11.01	1.64
2012-259	-	-	18	0.60	10.96	8.48	1.73
2012-286	-	-	21	0.54	10.45	7.88	1.64
2012-332	-	-	17	0.55	10.96	8.28	1.55
2012-346	-	-	25	0.31	9.33	6.77	0.99
2012-359	-	-	8	0.98	8.00	24.61	2.04
2013-30	-	-	96	0.52	27.67	16.87	2.39
2013-32	-	-	94	0.48	18.06	12.44	2.16
2013-34	-	-	131	0.57	27.26	17.13	2.76
2015-seep	-	-	199	0.55	28.98	18.79	2.91
2015-5225	-	-	73	0.64	25.92	18.10	2.76

Suppl. 5

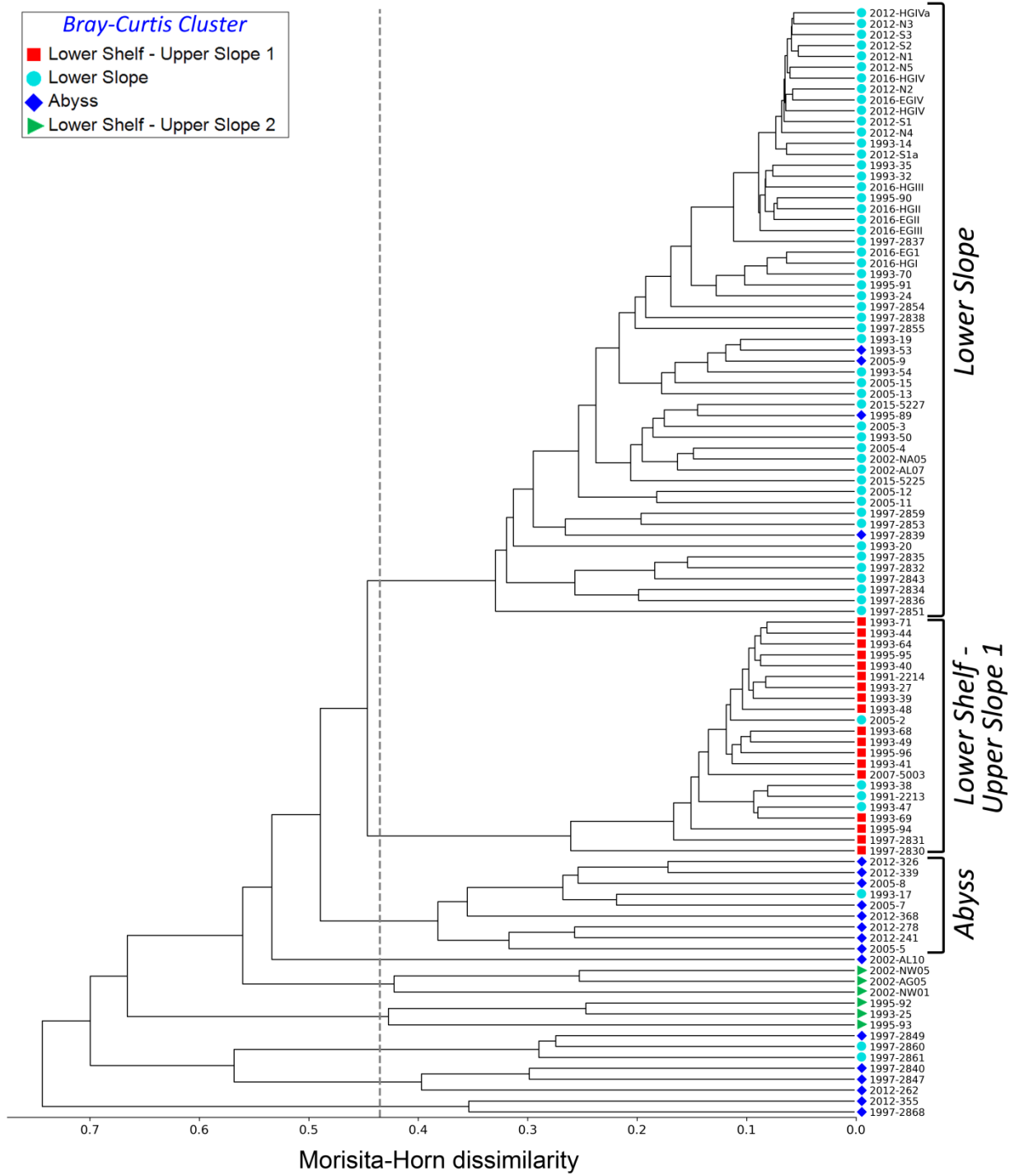


Fig. S-1. Cluster analysis of quantitative stations using Morisita-Horn dissimilarity (square-root transformation). Colour labels for stations are the same as for the Bray-Curtis similarity dendrogram. Dashed line indicates slice at the dissimilarity level of 0.43.

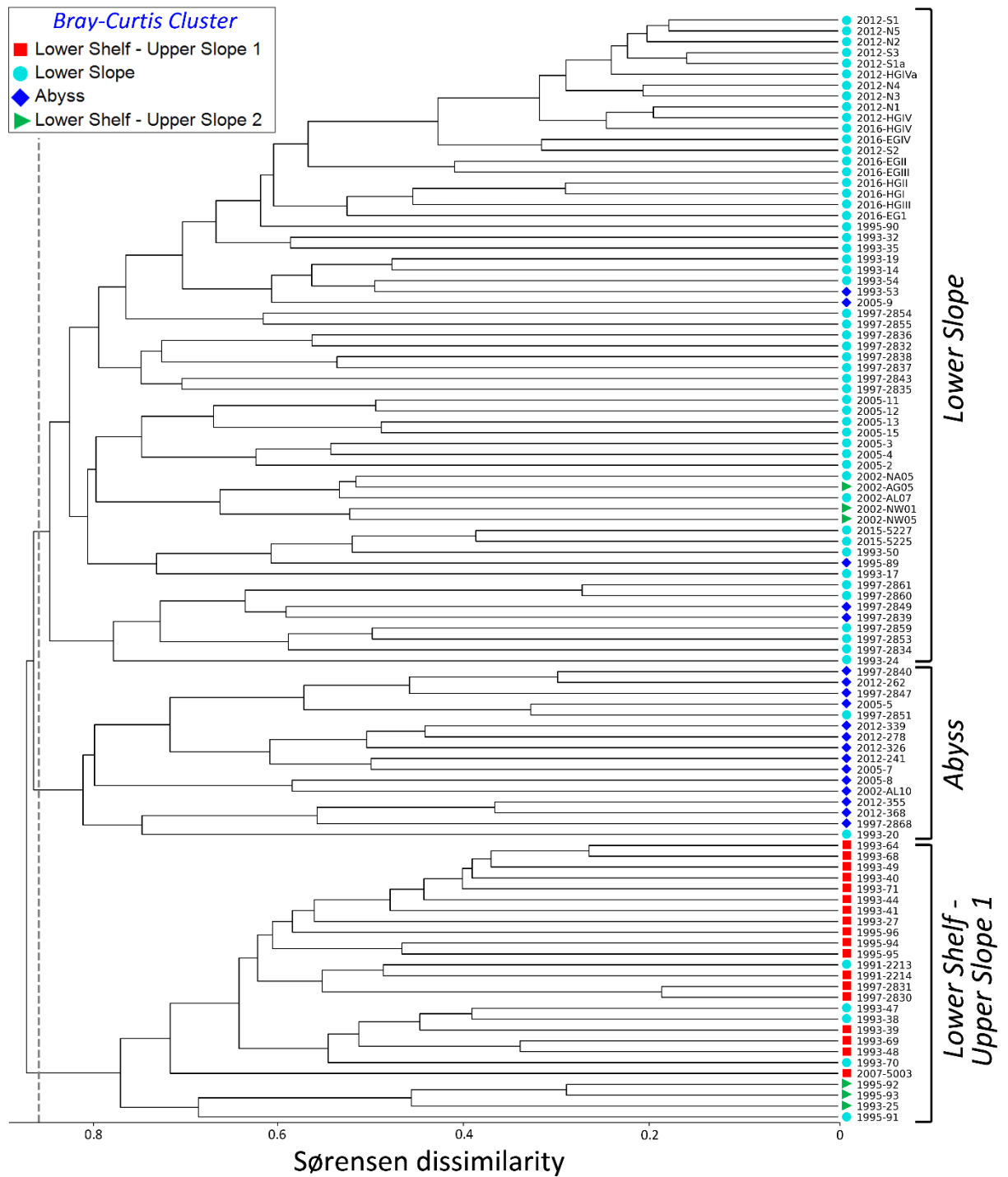


Fig. S-2. Cluster analysis of quantitative stations using quantitative Sørensen dissimilarity (square-root transformation). Colour labels for stations are the same as for the Bray-Curtis similarity dendrogram. Dashed line indicates slice at the dissimilarity level of 0.85.

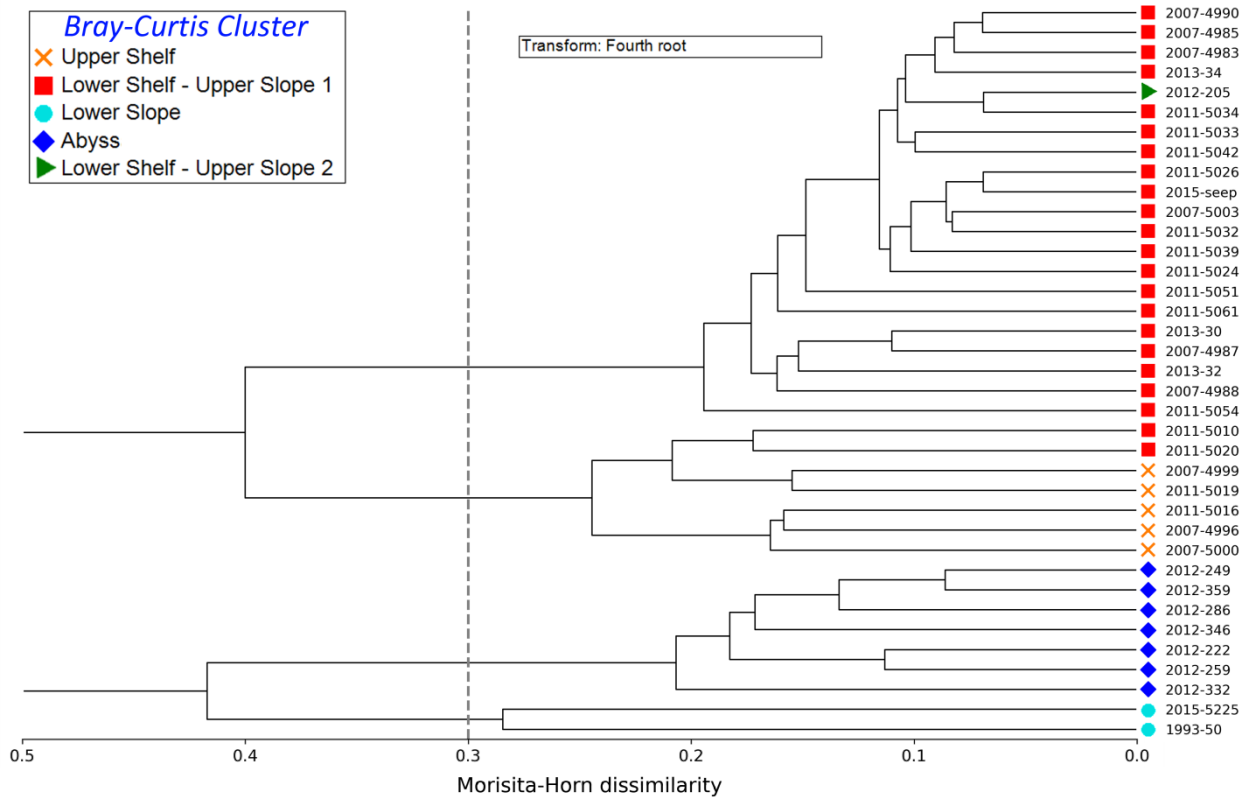


Fig. S-3. Cluster analysis of semi-quantitative stations using Morisita-Horn dissimilarity (4-th root transformation). Colour labels for stations are the same as for the corresponding Bray-Curtis similarity dendrogram. Dashed line indicates slice at the dissimilarity level of 0.3.

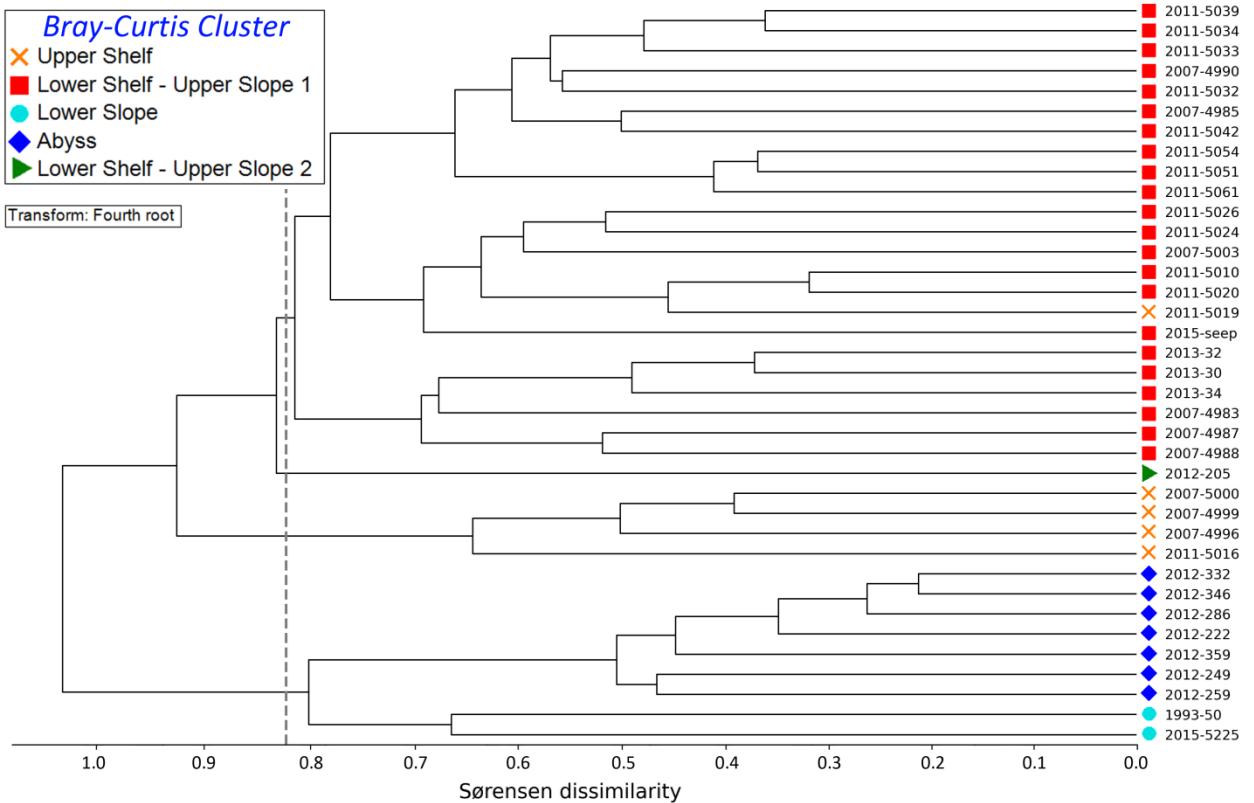


Fig. S-4. Cluster analysis of quantitative stations using quantitative Sørensen dissimilarity (square-root transformation). Colour labels for stations are the same as for the corresponding Bray-Curtis similarity dendrogram. Dashed line indicates slice at the dissimilarity level of 0.83.

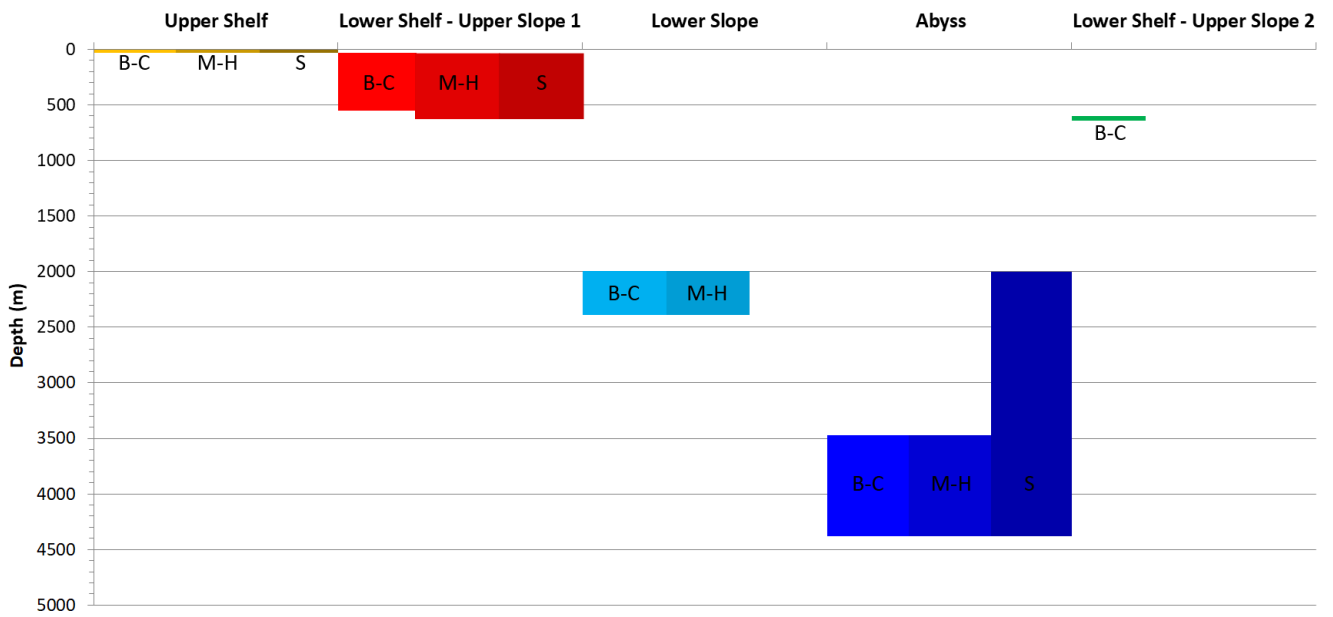


Fig. S-5. Bathymetric distribution of groups of Arctic benthic assemblages based on semi-quantitative samples (three different similarity indices). Colours as in Figs. 8-10. B-C – depth ranges from the Bray-Curtis similarity dendrogram; M-H – depth ranges from the Morisita-Horn similarity dendrogram; S – depth ranges from the quantitative Sørensen similarity dendrogram.

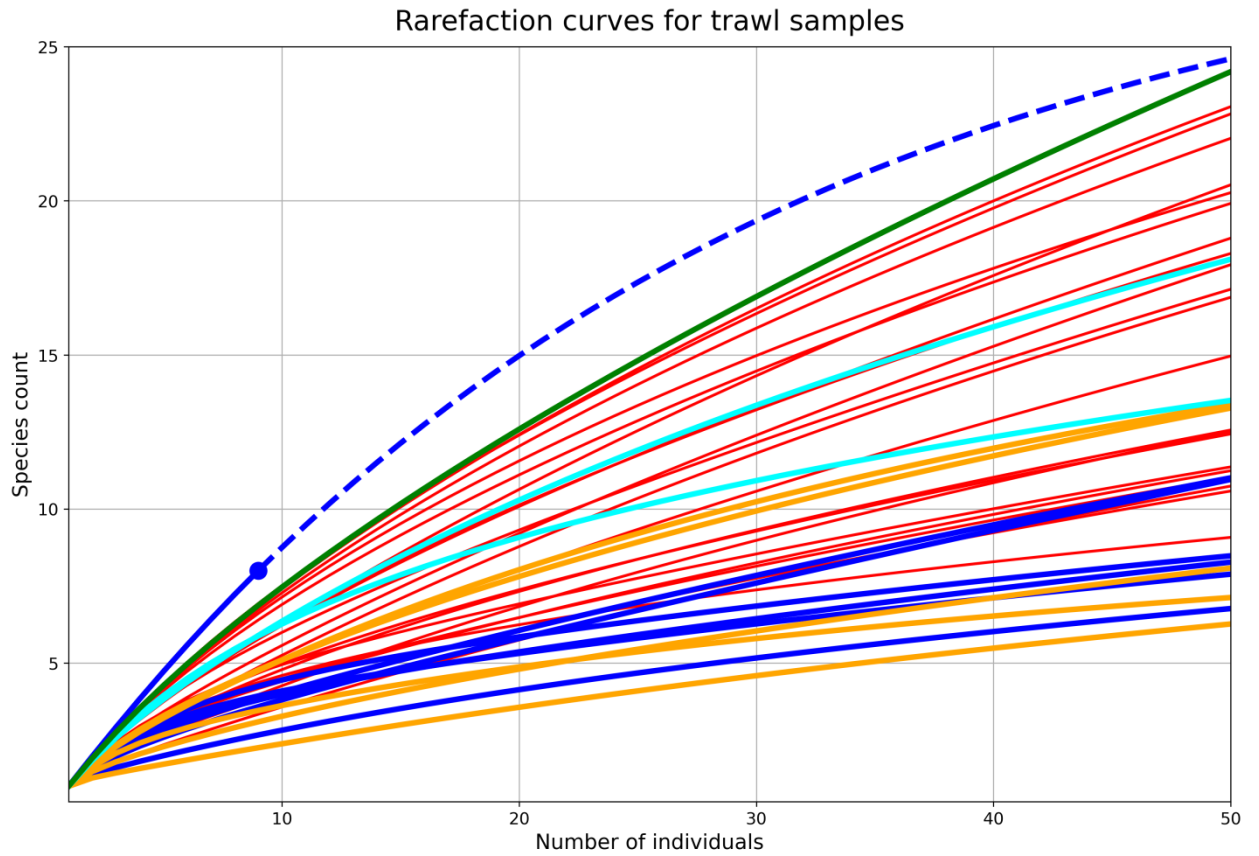


Fig. S-6. Rarefaction curves for the quantitative samples up to 50 individuals with extrapolation based on the Hill numbers ($q = 0$). Colours as in Figs. 8-10. Continuous lines indicate true (sample-sized) rarefaction; circles indicate the end of sample; dashed lines indicate the extrapolated rarefaction.