

Manuscript version: Author's Accepted Manuscript

The version presented in WRAP is the author's accepted manuscript and may differ from the published version or Version of Record.

Persistent WRAP URL:

http://wrap.warwick.ac.uk/173211

How to cite:

Please refer to published version for the most recent bibliographic citation information. If a published version is known of, the repository item page linked to above, will contain details on accessing it.

Copyright and reuse:

The Warwick Research Archive Portal (WRAP) makes this work by researchers of the University of Warwick available open access under the following conditions.

Copyright © and all moral rights to the version of the paper presented here belong to the individual author(s) and/or other copyright owners. To the extent reasonable and practicable the material made available in WRAP has been checked for eligibility before being made available.

Copies of full items can be used for personal research or study, educational, or not-for-profit purposes without prior permission or charge. Provided that the authors, title and full bibliographic details are credited, a hyperlink and/or URL is given for the original metadata page and the content is not changed in any way.

Publisher's statement:

Please refer to the repository item page, publisher's statement section, for further information.

For more information, please contact the WRAP Team at: wrap@warwick.ac.uk.

Diffusive Molecular Communication with a Spheroidal Receiver for Organ-on-Chip Systems

Hamidreza Arjmandi¹, Mohamad Zoofaghari², Mitra Rezaei¹, Kajsa Kanebratt³, Liisa Vilen³, David Janzen⁴, Peter Gennemark^{3,5}, and Adam Noel¹

¹School of Engineering, University of Warwick, Coventry, UK.

²Electrical Engineering Department, Yazd University, Yazd, Iran.

³ Drug Metabolism and Pharmacokinetics, Research and Early Development, Cardiovascular, Renal and Metabolism (CVRM), Bio-Pharmaceuticals, R&D, AstraZeneca, Gothenburg, Sweden.

 ${}^4Clinical\ Pharmacology\ and\ Quantitative\ Pharmacology, Clinical\ Pharmacology\ and\ Safety\ Sciences,$

AstraZeneca AB R&D Gothenburg, Gothenburg, Sweden.

⁵Department of Biomedical Engineering, Linköping University, Sweden.

Abstract—Realistic models of the components and processes are required for molecular communication (MC) systems. In this paper, a spheroidal receiver structure is proposed for MC that is inspired by the 3D cell cultures known as spheroids being widely used in organ-on-chip systems. A simple diffusive MC system is considered where the spheroidal receiver and a point source transmitter are in an unbounded fluid environment. The spheroidal receiver is modeled as a porous medium for diffusive signaling molecules, then its boundary conditions and effective diffusion coefficient are characterized. It is revealed that the spheroid amplifies the diffusion signal, but also disperses the signal which reduces the information communication rate. Furthermore, we analytically formulate and derive the concentration Green's function inside and outside the spheroid in terms of infinite series-forms that are confirmed by a particle-based simulator (PBS).

Index Terms—Molecular communication, Spheroid, Organ-onchip, Diffusion.

I. INTRODUCTION

Molecular communication (MC) is a bio-inspired mechanism that is envisioned to realize micro- and nano-scale communication systems using molecules as information carriers [1]. Despite many efforts by the MC community to model the components of MC systems, more realistic models are required. Elements and structures of *in vitro* environments such as organs-on-chip could be used to improve model realism, potentially contribute to MC research and development, and to provide mechanistic insight into the biology of the organs (on the chip).

Existing literature over-simplifies the MC receiver in a biological environment and does not sufficiently account for the interactions of signaling molecules with the environment and the biological or biosynthetic receivers and transmitters (i.e., cells). The passive receiver, in which information molecules freely diffuse in the receiver's space and the movement of molecules is not affected, is the most common model used in previous works [2]. Such a simple model is often used to facilitate analysis of other aspects of a molecular communication system, e.g., the environment boundary [3]. The passive model

may be relevant for small and hydrophobic molecules (which are repelled from water molecules) that easily pass through a cell membrane. However, most extracellular molecules are too large or too hydrophilic to traverse the cell membrane [4] and also cells usually react with signaling molecules (either directly on the surface or in the intracellular environment).

To address the limitations of the passive receiver model, some works have considered a reception mechanism across the receiver (cell) membrane activating a signaling pathway (i.e., a series of chemical reactions controlling a cell function). These works have studied the effects of various types of reactions across the membrane, as well as the size, number, spatial distribution, and/or saturation of the receptors on the cell membrane [5]–[11].

Previous works have considered the receiver as a single cell (or machine) whose surface is reacting with molecules. However, cells do not normally live in isolation but in populations with other cells of the same or of different types. This is true *in vivo* but also in many *in vitro* systems. In particular, tissues and tumors in multi-cellular organisms and biofilms of microorganisms are common natural instances whereas spheroids, organoids, tumoroids, and cell islets are well-known instances in biological experimental setups. This inspires the design of MC transceivers based on a population of (biological or biosynthetic) cells. In [12], the authors consider an implantable drug delivery system inside a spherical tumor microenvironment releasing drug molecules and obtain the spatiotemporal drug concentration profile in the surrounding tissue

One realistic receiver for MC is a spheroid structure, which is a 3D cell aggregation in a spherical shape that is widely used in organ-on-chip systems. Spheroids are constructed by various methods aiming to emulate the internal physiological activities of an organ [13]. In this paper, we consider a diffusive MC system with a spheroidal receiver and a point source transmitter. We model the spheroidal receiver as a porous medium for the diffusion signal and characterize its effective diffusion coefficient and implied boundary condi-

tion. We introduce and characterize two important processing features of the spheroid: *amplification* and *dispersion* of the diffusion signal. Furthermore, we obtain the concentration Green's function in an unbounded environment in the presence of the spheroidal receiver. Our results are confirmed by a particle based simulator (PBS).

The rest of the paper is organized as follows. Section II describes the spheroid structure and diffusive MC system. In Section III, the spheroid Green's function is provided. Results and discussions are presented in Section IV. Finally, Section V concludes the paper.

II. SYSTEM MODEL

A. Spheroid model

A spheroid with radius R_s formed by N_c cells is considered. The spheroid interior space is comprised of the cells and the void space between the cells (i.e., extracellular environment). Given that the volume of a cell within the spheroid is V_c , the volumes of the cell matrix and the void space inside the spheroid are given by V_cN_c and $V_s-V_cN_c$, respectively, where $V_s=\frac{4}{3}\pi R_s^3$ is the spheroid volume.

We model the spheroid structure as a porous medium with volume V_s whose porosity, ϵ , is defined as the ratio of the void space to the whole spheroid volume, i.e.,

$$\epsilon = 1 - \frac{N_c V_c}{V_s}. (1)$$

We assume that the spheroid is in a fluid medium that surrounds it and fills its void space. The diffusive signaling molecules of type A in the medium can diffuse into the void space of the spheroid and stimulate the spheroid's cells through a transmembrane mechanism, e.g., by binding to receptors.

Inside the spheroid, the molecules diffuse via the curved paths of the void space among the cells, which leads to a shorter net displacement of the molecules in a given time interval. Thus, macroscopic diffusion within the spheroid effectively differs from the diffusion within the free fluid outside the spheroid. Since the molecules traverse a shorter net path within the spheroid, the effective diffusion coefficient is smaller than the diffusion coefficient in the free fluid medium and molecules are more likely to be observed and sensed by the spheroid's cells. Given the diffusion coefficient D for molecules A in the free fluid, the effective diffusion coefficient within the spheroid is given by

$$D_{\text{eff}} = -\frac{\epsilon}{\tau}D,\tag{2}$$

where τ is the tortuosity, a measure of the transport properties of the porous medium, and is modeled as a function of spheroid porosity as $\tau = \frac{1}{\epsilon^{0.5}}$ [14].

To describe the environment geometry, we use the spherical coordinate system where (r,θ,φ) denote radial, elevation, and azimuth coordinates, respectively. At the border of the two diffusive environments with different diffusion coefficients, we have a flow continuity condition as

$$D_{\text{eff}} \frac{\partial c_s(\bar{r}, t)}{\partial r} = D \frac{\partial c_o(\bar{r}, t)}{\partial r}, \tag{3}$$

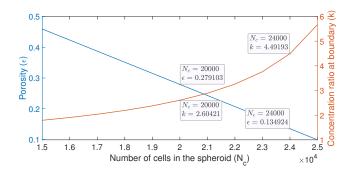


Fig. 1. Porosity (ϵ) and concentration ratio at the boundary (k) versus the number of cells inside the spheroid (N_c) .

and another boundary condition that is generally modeled as [15, Ch. 3]

$$c_s(\bar{r}, t) = kc_o(\bar{r}, t), \tag{4}$$

where $\bar{r} \in \partial \Omega$, $\partial \Omega$ denotes the boundary region of the spheroid, and c_s and c_o denote the concentration function inside and outside the spheroid, respectively. The constant k is a function of porosity of the medium and should be determined experimentally. But, the literature on mathematical analysis of tumor spheroids simply assumes continuity of concentration, i.e., k=1, by neglecting the porosity of the medium [16], [17]. However, in the results section, we show using a PBS that for two ideal diffusion environments with diffusion coefficients D and $D_{\rm eff}$, we have $k=\sqrt{\frac{D}{D_{\rm eff}}}$. Thus, for $k\neq 1$, a concentration discontinuity (i.e., jump) occurs at the boundary.

Fig. 1 demonstrates the porosity (ϵ) and boundary concentration ratio (k) as a function of the number of the cells, $15000 < N_c < 25000$ in the spheroid when the spheroid radius and cell volume are assumed to be $R_s = 275~\mu{\rm m}$ and $V_c = 3.14 \times 10^{-15}~{\rm m}^3$, respectively [18]. As observed in Fig. 1, the boundary concentration ratio increases exponentially with an increase in N_c . For $N_c = 24000$, which is the approximate number of cells in HepaRG spheroids reported in [18], we have $\epsilon = 0.13$ and correspondingly k = 4.49. This value of k suggests a large concentration discontinuity at the spheroid boundary.

We assume that the molecules may bind to cell receptors through a chain of reactions leading to product molecules ${\cal E}$ that are counted by cells, i.e.,

$$A + B \Longrightarrow ... \longrightarrow E,$$
 (5)

where B and E may represent the receptor molecules and the final products in the cells, respectively.

A real mature spheroid is tightly formed by a large number of cells, resulting in a small porosity. Then, a signaling molecule moving within the void space would be very close to the cells' boundaries and frequently exposed to bind to receptors. Thus, we assume that molecules within the void space of the spheroid are homogeneously available for transmembrane reactions.

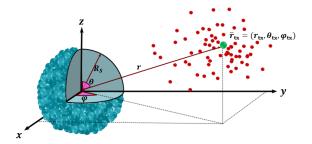


Fig. 2. System model schematic where the red, greenish-blue, and green spheres represent the signaling molecule, cell, and point source transmitter, respectively.

B. Diffusive MC system with spheroidal receiver

A point-to-point diffusive MC system is considered in which the spheroid is assumed to be the receiver. The spheroid might model a cancer tumor site that should receive drug molecules; it might model the spheroids in a organ-on-chip system that receive nutrient molecules to survive; alternatively, it could be a transceiver nanomachine for information exchange in a MC system.

The spheroid is considered in an unbounded fluid medium and the center of the spheroid is chosen as the origin of the spherical coordinate system. A point source transmitter is located at an arbitrary point $\bar{r}_{\rm tx} = (r_{\rm tx}, \theta_{\rm tx}, \varphi_{\rm tx})$ outside the spheroid. The transmitter uses signaling molecules of type A. A schematic illustration of the system model is represented in Fig. 2. The released molecules move randomly in the environment following Brownian motion. Their movements are assumed to be independent of each other. The diffusing molecules, which are exposed to the receptors in the spheroid, may bind and generate molecules E that are counted within the cells in the spheroidal receiver.

The received signal can be defined according to the application. Generally, the cells might respond differently and react to the generated E molecules. A cell's response might be a continuous function of product molecule concentration. Then, the received signal in the spheroid may be defined as the aggregate number of generated E molecules inside the cell. In this case, the concentration profile of E molecules is obtained as a function of signaling A molecules based on (5). Then, given the uniform density of cells within the spheroid, the received signal is given by

$$\int_0^{R_s} 4\pi c_E(\bar{r}, t) V_c \frac{N_c}{V_s} r^2 \mathrm{d}r,\tag{6}$$

where c_E represents the product molecule concentration. Correspondingly, the received signal can be defined as the total generation rate of the product E molecules inside the spheroid, i.e., the time derivative of (6). Alternatively, an individual cell might be activated and respond according to a stimulus threshold. For example, if the spheroid is a tumor receiving drug molecules, then each cell might die by receiving a concentration of the drug higher than a threshold. Then, the

received signal may be defined as the ratio of killed cells receiving the drug molecules, i.e.,

$$\int_0^{R_s} 4\pi I_{c_E(\bar{r},t)>\mathcal{T}} V_c \frac{N_c}{V_s} r^2 \mathrm{d}r,\tag{7}$$

where $I_{(\cdot)}$ is the indicator function and \mathcal{T} is the threshold above which an individual cell is activated.

III. GREEN'S FUNCTION BOUNDARY VALUE PROBLEM

To analyze the presented diffusive MC system, we formulate the corresponding Green's function boundary value problem for diffusion in the environment. We assume that the point source transmitter, located at an arbitrary point $\bar{r}_{\rm tx}=(r_{\rm tx},\theta_{\rm tx},\varphi_{\rm tx})$ outside the spheroid, has an instantaneous molecule release rate of $\delta(t-t_0)$ molecule 1/s, where $\delta(\cdot)$ is the Dirac delta function. This impulsive point source is represented by the function $S(\bar{r},t,\bar{r}_{\rm tx},t_0)=\frac{\delta(r-r_{\rm tx})\delta(\theta-\theta_{\rm tx})\delta(\varphi-\varphi_{\rm tx})\delta(t-t_0)}{r^2\sin\theta}$ s^-1 m^-3. Given the source $S(\bar{r},t,\bar{r}_{\rm tx},t_0)$, the molecular diffusion out of the spheroid is described by the partial differential equation (PDE) [19]

$$D\nabla^2 c_o(\bar{r}, t | \bar{r}_{\rm tx}, t_0) + S(\bar{r}, t, \bar{r}_{\rm tx}, t_0) = \frac{\partial c_o(\bar{r}, t | \bar{r}_{\rm tx}, t_0)}{\partial t}, (8)$$

where $c_o(\bar{r}, t|\bar{r}_{\rm tx}, t_0)$ denotes the molecule concentration at point \bar{r} outside spheroid at time t. In the rest of the paper, we briefly write c_o instead of $c_o(\bar{r}, t|\bar{r}_{\rm tx}, t_0)$. In the spherical coordinate system and Fourier domain, (8) is re-written as

$$\frac{D}{r^{2}} \frac{\partial}{\partial r} \left(r^{2} \frac{\partial C_{o}}{\partial r} \right) + \frac{D}{r^{2} \sin \theta} \frac{\partial}{\partial \theta} \left(\sin \theta \frac{\partial C_{o}}{\partial \theta} \right) + \frac{D}{r^{2} \sin^{2} \theta} \frac{\partial^{2} C_{o}}{\partial \varphi^{2}} + \frac{\delta (r - r_{tx}) \delta (\theta - \theta_{tx}) \delta (\varphi - \varphi_{tx}) \delta (t - t_{0})}{r^{2} \sin \theta} = i \omega C_{o},$$
(9)

where C_o is the Fourier transform of c_o , i is the imaginary unit, and ω is the angular frequency variable in the Fourier transform². With a first order approximation of the spheroid cell receptor reaction chain (5), the effective diffusion inside the spheroid is modeled as

$$D_{\text{eff}} \nabla^2 C_s(\bar{r}, \omega | \bar{r}_{\text{tx}}, t_0) - \mathcal{K}(\omega) C(\bar{r}, \omega | \bar{r}_{\text{tx}}, t_0)$$

$$= i\omega C_s(\bar{r}, \omega | \bar{r}_{\text{tx}}, t_0),$$
(10)

where $\mathcal{K}(\omega)$ is the net impact on molecules A due to the binding process characterized by the reaction chain (5). As mentioned, the boundary conditions at the border of the spheroid are given by (3) and (4).

The concentration functions $c_s(\bar{r},t|\bar{r}_{\rm tx},t_0)$ and $c_o(\bar{r},t|\bar{r}_{\rm tx},t_0)$ that satisfy (8) and (10) subject to the boundary conditions (3) and (4) are called the concentration Green's functions (CGFs) of diffusion inside and outside the spheroid, respectively. We have solved the problem and obtained the series-form expressions (15) in the Appendix.

¹Note that the number of molecules does not have a dimension, so we have not represented it in the units.

 $^{^2}$ We use the small c and capital C to represent the concentration function in the time and Fourier domains, respectively.

IV. RESULTS

In this section, we first evaluate the boundary concentration ratio, k, with a particle based simulator (PBS). We verify the proposed analysis of the spheroid Green's function using this PBS. Furthermore, we compare the received signal at the spheroidal receiver with a transparent receiver to reveal the signal amplification and dispersion within the spheroid.

The geometric parameters considered in this section are based on the real hepatocyte spheroids used in [18] where the spheroid's radius is 275 μ m with $N_c \approx 24000$ cells. The volume of one hepatocyte cell is estimated as 3.14×10^{-15} m³. The reaction (5) inside the spheroid is simply assumed to be an irreversible first order reaction

$$A \xrightarrow{k_f} E,$$
 (11)

with $k_{\rm f}=0.01$ or $0.1~{\rm s}^{-1}$. Therefore, the generation rate of the E molecules inside the spheroid is obtained as $\frac{\partial c_E}{\partial t}=k_fc_s$ and correspondingly the term $\mathcal{K}(\omega)$ in (10) is simply $-k_f$.

The PBS is implemented in MATLAB (R2021b; The Math-Works, Natick, MA) where the time is divided into time steps of $\Delta t = 0.05$ s. The molecules released at the point source transmitter at $\bar{r}_{\rm tx} = (500 \ \mu {\rm m}, \pi/2, 0)$ move independently in the 3-dimensional space. The displacement of a molecule in Δt s is modeled as a Gaussian random variable with zero mean and variance $2D\Delta t$ in each dimension of Cartesian coordinates. For the PBS, the spheroid is simply a diffusion environment with effective coefficient $D_{\rm eff}$. If the calculated displacement vector of a molecule in a time slot passes the spheroid boundary, then the length of the portion of the vector inside the spheroid is updated based on the effective diffusion coefficient $D_{\rm eff}$, i.e., that part of the vector is scaled by the factor $\sqrt{\frac{D_{\mathrm{eff}}}{D}}.$ Considering the reaction (11) inside the spheroid, a molecule may be absorbed from the void space of the spheroid by the cells during a time step Δt s with approximate probability $k_f \Delta t$ [20]. To simulate the impulsive source, 10⁶ molecules are released at the transmitter location. To obtain a concentration at a given time and location, we assume a transparent sphere centered at that location with a small radius of 10 μm and count the number of molecules inside the sphere at that time. The concentration would then be the counted number of molecules divided by the volume of the sphere. The concentration value is normalized, i.e., is divided by 10^6 , to obtain the response to the impulsive source.

Fig. 3 shows the concentration at inner and outer boundary points $c_s(275^-\mu\mathrm{m},0,0)$ and $c_o(275^+\mu\mathrm{m},0,0)$ of the spheroid given the impulsive source at the transmitter obtained from the PBS when $N_c=24000$. We have also plotted the concentration at the outer boundary scaled by the factor k to verify the boundary condition (4). The minor mismatch at the peaks is mainly due to the PBS procedure to approximate the concentration at a point. To compute the concentration at the boundary point $(275~\mu\mathrm{m},0,0)$, we have assumed a transparent sphere of radius $10~\mu\mathrm{m}$ centered at the boundary point and have counted the molecules in the left and right

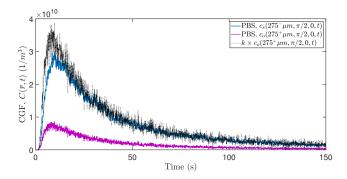


Fig. 3. CGF at the inner and outer boundaries versus time obtained from the PBS for $N_{\rm c}=24000$.

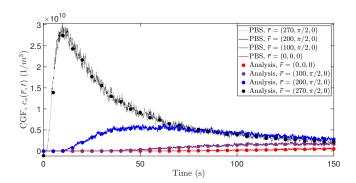


Fig. 4. CGF obtained at different points versus time obtained from analysis and PBS for $N_c=24000$.

hemisphere to approximate the concentration of the molecules at inner and outer points. This leads to slightly higher and lower approximations for the concentrations at the outer and inner boundary, respectively.

Fig. 4 depicts the concentration at different points inside the spheroidal receiver obtained from the analysis provided in the Appendix and also the PBS when $N_c=24000$. As observed, the PBS fully confirms the analytical results. The figure also indicates that the concentration signal weakens significantly at the points closer to the spheroid center and farther from the point source. In particular, it is observed that the concentration peak at the center is about 18 times smaller than that at the boundary.

Fig. 5 compares the received signal of the spheroidal and the transparent receivers to show how the spheroid structure affects the diffusion signal. We measure the received signal as the generation rate of the product E molecules all through the spheroid or transparent receiver. Note that the absorbing reaction with the same k_f is also considered within the "transparent" receiver. As observed, the spheroid structure amplifies the diffusion signal and delays the peak of the received signal, further revealing the dispersion of the diffusion signal inside the spheroid. These two effects of amplification and dispersion lead to a better received signal at the cost of a lower transmission rate that the receiver can perceive.

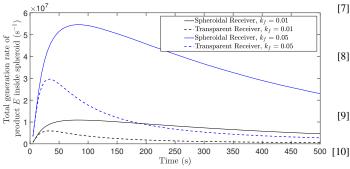


Fig. 5. Comparison of the received signal of the spheroidal receiver with the transparent receiver for $N_c = 24000$ and $k_f = \{0.01, 0.05\}$.

Conclusion

In this paper, we characterized the received diffusion signal by a spheroidal receiver inspired by spheroids used in organ-on-chip systems. We modeled the spheroid as a porous medium and its boundary condition with free fluid outside. We revealed that the diffusion signal is affected by the spheroid structure in two ways: amplification and dispersion. Both impacts are caused by the effective diffusion coefficient inside the spheroid which is smaller than the diffusion of molecules in the free fluid outside the spheroid. We also provided a series-form expression for the Green's function inside and outside the spheroid and confirmed it using PBS. The proposed spheroid model and analysis not only suggest a new MC receiver, but also support the analysis of organ-on-chip systems where the 3D cell culture spheroids are widely used and the tumors or tumoroids receiving drug molecules in in vivo and in vitro environments, respectively.

V. ACKNOWLEDGEMENT

This work was supported by the Engineering and Physical Sciences Research Council [grant number EP/V030493/1].

REFERENCES

- N. Farsad, et. al., "A comprehensive survey of recent advancements in molecular communication," in *IEEE Communications Surveys & Tutorials*, vol. 18, no. 3, pp.1887-919, Feb. 2016.
- [2] V. Jamali, et. al., "Channel modeling for diffusive molecular communication A tutorial review," in *Proceedings of the IEEE*, vol. 107, no. 7, pp. 1256-1301, July 2019.
- [3] H. Arjmandi, M. Zoofaghari and A. Noel, "Diffusive molecular communication in a biological spherical environment with partially absorbing boundary," in *IEEE Transactions on Communications*, vol. 67, no. 10, pp. 6858-6867, Oct. 2019.
- [4] D. Bi, et. al., "A survey of molecular communication in cell biology: Establishing a new hierarchy for interdisciplinary applications," in *IEEE Communications Surveys and Tutorials*, vol. 23, no. 3, pp. 1494-1545, 3rd quarter 2021.
- [5] N.V. Sabu, N. Varshney and A.K. Gupta, "3-D diffusive molecular communication with two fully-absorbing receivers: Hitting probability and performance analysis," in *IEEE Transactions on Molecular, Biological and Multi-Scale Communications*, vol. 6, no. 3, pp. 244-249, Dec. 2020.
- [6] M. Pierobon and I.F. Akyildiz, "Noise analysis in ligand-binding reception for molecular communication in nanonetworks," in *IEEE Transactions on Signal Processing*, vol. 59, no. 9, pp. 4168-4182, Sept. 2011.

- [7] A. Akkaya, H. B. Yilmaz, C.B. Chae and T. Tugcu, "Effect of receptor density and size on signal reception in molecular communication via diffusion with an absorbing receiver," in *IEEE Communications Letters*, vol. 19, no. 2, pp. 155-158, Feb. 2015.
- [8] A. Ahmadzadeh, H. Arjmandi, A. Burkovski and R. Schober, "Comprehensive reactive receiver modeling for diffusive molecular communication systems: Reversible binding, molecule degradation, and finite number of receptors," in *IEEE Transactions on NanoBioscience*, vol. 15, no. 7, pp. 713-727, Oct. 2016.
- [9] C. T. Chou, "Impact of receiver reaction mechanisms on the performance of molecular communication networks," in *IEEE Transactions on Nanotechnology*, vol. 14, no. 2, pp. 304-317, March 2015.
- [10] S. Lotter, M. Schfer, J. Zeitler and R. Schober, "Saturating receiver and receptor competition in synaptic DMC: Deterministic and statistical signal models," in *IEEE Transactions on NanoBioscience*, vol. 20, no. 4, pp. 464-479, Oct. 2021.
- [11] G. Genc, Y.E. Kara, T. Tugcu, A.E. Pusane, "Reception modeling of sphere-to-sphere molecular communication via diffusion," in *Nano* communication networks, vol. 16, pp. 69-80, Jan. 2018.
- communication networks, vol. 16, pp. 69-80, Jan. 2018.
 [12] M. M. Al-Zubi and A. S. Mohan, "Modelling of implantable drug delivery system in tumor microenvironment using molecular communication paradigm," in *IEEE Access*, vol. 7, pp. 141929-141940, 2019.
- [13] S. Daunys, et. al., "3D tumor spheroid models for in vitro therapeutic screening of nanoparticles," *InBio-Nanomedicine for Cancer Therapy*, Springer, Cham, pp. 243-270, 2021.
- [14] F. Sharifi, B. Firoozabadi, K. Firoozbakhsh, "Numerical investigations of hepatic spheroids metabolic reactions in a perfusion bioreactor," Frontiers in bioengineering and biotechnology, vol. 7, p.221, Sep. 2019.
- [15] J. Crank, The mathematics of diffusion, Oxford university press, 1979.
- [16] J.J. Klowss, et al., "A stochastic mathematical model of 4D tumour spheroids with real-time fluorescent cell cycle labelling," *Journal of the Royal Society Interface*, vol. 19, no. 189, Apr. 2022.
- [17] J. A. Bull, et al., "Mathematical modelling reveals cellular dynamics within tumour spheroids." *PLoS Computational Biology*, vol. 16, no. 8, Aug. 2020.
- [18] S. Bauer, et al., "Functional coupling of human pancreatic islets and liver spheroids on-a-chip: Towards a novel human ex vivo type 2 diabetes model," *Scientific reports*, vol. 7, no. 1, pp. 1-11, Nov. 2017.
- [19] P. Grindrod, *The theory and applications of reaction-diffusion equations:*Patterns and waves, Clarendon Press, 1996.
- [20] Y. Deng, et al., "Modeling and simulation of molecular communication systems with a reversible adsorption receiver," *IEEE Transactions on Molecular, Biological and Multi-Scale Communications*, vol. 1, no. 4, pp.347-362, Dec. 2015.
- [21] M. Zoofaghari, A. Etemadi, H. Arjmandi and I. Balasingham, "Modeling molecular communication channel in the biological sphere With arbitrary homogeneous boundary conditions," in *IEEE Wireless Communications Letters*, vol. 10, no. 12, pp. 2786-2790, Dec. 2021.

APPENDIX

In this Appendix, we solve the frequency domain PDEs (9) and (10) given the boundary conditions (3) and (4). The general solution of (9) and (10) is considered as [21]

$$C_q(r, \theta, \varphi, \omega | \bar{r}_{\rm tx}) = \sum_{n=0}^{\infty} \sum_{m=0}^{n} H_{mn} R_n^q(r, \omega) \cos(m(\varphi - \varphi_{\rm tx}))$$

$$\times P_{nm}(\cos \theta), \quad q \in \{s, o\}, \tag{15}$$

where $R_n^q(r,\omega)$ is the unknown radial function of r for the region $q \in \{s,m\}$, $\mathcal{F}(\theta,\varphi) = \cos(m\varphi)P_{nm}(\cos\theta)$ is the associated Fourier-Legendre function of the first kind with degree n and order m that satisfies the partial differential equation (PDE) in

$$\frac{D}{\sin \theta} \frac{\partial}{\partial \theta} \left(\sin \theta \frac{\partial \mathcal{F}(\theta, \varphi)}{\partial \theta} \right) + \frac{D}{\sin^2 \theta} \frac{\partial^2 \mathcal{F}(\theta, \varphi)}{\partial \varphi^2} + n(n+1)\mathcal{F}(\theta, \varphi) = 0,$$
(16)

and H_{mn} denotes the unknown coefficient corresponding to the mode mn of the response.

$$\sum_{n=0}^{\infty} \sum_{m=0}^{n} H_{mn} \frac{D}{r^{2}} \frac{\partial}{\partial r} (r^{2} \frac{\partial R_{n}^{o}(r,\omega)}{\partial r}) P_{nm}(\cos \theta) \cos(m(\varphi - \varphi_{tx}) + \sum_{n=0}^{\infty} \sum_{m=0}^{n} H_{mn} R_{n}^{o}(r,\omega) \frac{D}{r^{2} \sin \theta} \frac{\partial}{\partial \theta} (\sin \theta \frac{\partial P_{nm}(\cos \theta)}{\partial \theta}) + \frac{D}{r^{2} \sin^{2} \theta} \frac{\partial^{2} (\cos(m(\varphi - \varphi_{tx})))}{\partial \varphi^{2}} - (i\omega) \sum_{n=0}^{\infty} \sum_{m=0}^{n} H_{mn} R_{n}^{o}(r,\omega) P_{nm}(\cos \theta) \cos(m(\varphi - \varphi_{tx})) \\
= -\sum_{n=0}^{\infty} \sum_{m=0}^{n} L_{m} \frac{2n+1}{2} \frac{(n-m)!}{(n+m)!} \times P_{nm}(\cos \theta) P_{nm}(\cos \theta_{tx}) \cos(m(\varphi - \varphi_{tx})) \delta(\bar{r} - \bar{r}_{tx}), \tag{12}$$

$$\sum_{n=0}^{\infty} \sum_{m=0}^{n} H_{mn} \frac{D}{r^{2}} \frac{\partial}{\partial r} \left(r^{2} \frac{\partial R_{n}^{s}(r,\omega)}{\partial r}\right) P_{nm}(\cos \theta) \cos\left(m(\varphi - \varphi_{\text{tx}}) + \sum_{n=0}^{\infty} \sum_{m=0}^{n} H_{mn} R_{n}^{s}(r,\omega) \frac{D}{r^{2} \sin \theta} \frac{\partial}{\partial \theta} \left(\sin \theta \frac{\partial P_{nm}(\cos \theta)}{\partial \theta}\right) + \frac{D}{r^{2} \sin^{2} \theta} \frac{\partial^{2} (\cos(m(\varphi - \varphi_{\text{tx}})))}{\partial \varphi^{2}} - \left(\mathcal{K}(i\omega) + i\omega\right) \sum_{n=0}^{\infty} \sum_{m=0}^{n} H_{mn} R_{n}^{s}(r,\omega) P_{nm}(\cos \theta) \cos(m(\varphi - \varphi_{\text{tx}})) = 0.$$
(13)

$$\begin{bmatrix} j_n(k_1R_s) & -y_n(k_2R_s) & -j'_n(k_2R_s) & 0\\ D_{\text{eff}}k_1j_n(k_1R_s) & -Dk_2y'_n(k_2R_s) & -Dk_2j'_n(k_2R_s) & 0\\ 0 & y_n(k_2r_{tx}) & j_n(k_2r_{tx}) & -y_n(k_2r_{tx})\\ 0 & r_{tr}^2y'_n(k_2r_{tx}) & r_{tr}^2j'_n(k_2r_{tx}) & -r_{tr}^2y'_n(k_2r_{tx}), \end{bmatrix} \begin{bmatrix} G_n\\ A_n\\ B_n\\ D_n \end{bmatrix} = \begin{bmatrix} 0\\ 0\\ 0\\ 1 \end{bmatrix}$$

$$(14)$$

Also, the representation of $\delta(\varphi - \varphi_{\rm tx}) \frac{\delta(\theta - \theta_{\rm tx})}{\sin \theta}$ based on the aforementioned Fourier-Legendre basis functions is given by

$$\delta(\varphi - \varphi_{\rm tx}) \frac{\delta(\theta - \theta_{\rm tx})}{\sin \theta} = \sum_{n=0}^{\infty} \sum_{m=0}^{n} L_m \frac{2n+1}{2} \frac{(n-m)!}{(n+m)!}$$
(17)

 $P_n^m(\cos\theta)P_n^m(\cos\theta_{\rm tx})\cos(m(\varphi-\varphi_{\rm tx})),$

where $L_0=\frac{1}{2\pi}$ and $L_m=\frac{1}{\pi}, m\geq 1$. Replacing C_q and $\delta(\varphi-\varphi_{\rm tx})\frac{\delta(\theta-\theta_{\rm tx})}{\sin\theta}$ in (9) and (10) by the corresponding series-form representations given by (15) and (17), respectively, leads to (12) and (13) at the top of this page. Matching the two sides of (12) yields

$$H_{mn} = L_m \frac{2n+1}{2} \frac{(n-m)!}{(n+m)!} P_n^m(\cos \theta_{\rm tx}), \qquad (18)$$

and

$$r^{2} \frac{\partial^{2} R_{n}^{o}(r,\omega)}{\partial r^{2}} + 2r \frac{\partial R_{n}^{o}(r,\omega)}{\partial r} + (k_{1}^{2}r^{2} - n(n+1))R_{n}^{o}(r,\omega)$$
$$= \delta(\bar{r} - \bar{r}_{tx}), \tag{19}$$

where $k_1=\sqrt{\frac{i\omega}{D}}.$ Similarly, (13) is reduced to

$$r^{2} \frac{\partial^{2} R_{n}^{s}(r,\omega)}{\partial r^{2}} + 2r \frac{\partial R_{n}^{s}(r,\omega)}{\partial r} + (k_{2}^{2}r^{2} - n(n+1))R_{n}^{s}(r,\omega)$$

$$= 0,$$
(20)

where $k_2 = \sqrt{\frac{-\mathcal{K}(i\omega) - i\omega}{D_{\mathrm{eff}}}}$. By applying (15) in the Fourier forms of boundary conditions (3) and (4), we obtain

$$D_{\text{eff}} \frac{\partial R_n^s(r,\omega)}{\partial r} \bigg|_{r=r_s} = D \frac{\partial R_n^o(r,\omega)}{\partial r} \bigg|_{r=r_s}, \qquad (21)$$

$$R_n^s(r,\omega)\Big|_{r=r_s} = k \times R_n^o(r,\omega)\Big|_{r=r_s}.$$
 (22)

To solve (19), we remove the source term $\delta(\bar{r} - \bar{r}_{\rm tx})$ at the right-hand side and consider the derivative discontinuity of

the CGF at $r = r_{\rm tx}$ that leads to the following corresponding boundary condition [21]³

$$r^{2} \frac{\partial R_{n}(r,\omega)}{\partial r} \bigg|_{r=r_{tx}^{+}} - r^{2} \frac{\partial R_{n}(r,\omega)}{\partial r} \bigg|_{r=r_{tx}^{-}} = 1.$$
 (23)

The solutions of the homogeneous form of the PDE of (19) and also (20) are then given by

$$R_n^s(r,\omega) = G_n j_n(kr), \ r < R_s \tag{24}$$

$$R_n^o(r,\omega) = \begin{cases} B_n j_n(kr) + A_n y_n(kr), & R_s < r < r_{\text{tx}} \\ D_n y_n(kr), & r > r_{\text{tx}} \end{cases}$$
(25)

where $j_n(\cdot)$ and $y_n(\cdot)$ are the *n*th order of the first and second types of spherical Bessel function, respectively. By applying the solution (24) to the boundary conditions (21), (22), and (23), and also the continuity condition of concentration at $\bar{r}_{\rm tx}$, i.e.,

$$\left. R_n^o(r,\omega) \right|_{r=r_{\rm tx}^+} = \left. R_n^o(r,\omega) \right|_{r=r_{\rm tx}^-},\tag{26}$$

the system of linear equations (14) is obtained by which the coefficients A_n , B_n , G_n , and D_n are calculated. Noteworthy, $j'(\cdot)$ and $y'(\cdot)$ in (14) are the derivative functions of $j(\cdot)$ and $y(\cdot)$ in terms of r, respectively. By having the coefficients A_n , B_n , G_n , and D_n , $R_n^q(r,\omega)$, $q \in \{s,o\}$ in (24) is known and the CGF is computed by taking the inverse Fourier transform of (15).

³The equivalent boundary condition of (23) is derived by integration of two sides of over $[r_{\rm tx}^+, r_{\rm tx}^-]$.