



University of Groningen

Targeted Lung Denervation modulates the mucosal epithelial transcriptome in COPD

Srikanthan, Karthi; Kistemaker, Loes; Slebos, Dirk Jan; Gesierich, Wolfgang; Darwiche, Kaid; Bonta, Peter; Deslee, Gaetan; Shah, Pallav; Gosens, Reinoud

Published in: **ERJ Open Research**

DOI: 10.1183/23120541.00146-2022

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version Publisher's PDF, also known as Version of record

Publication date: 2022

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA): Srikanthan, K., Kistemaker, L., Slebos, D. J., Gesierich, W., Darwiche, K., Bonta, P., Deslee, G., Shah, P., & Gosens, R. (2022). Targeted Lung Denervation modulates the mucosal epithelial transcriptome in COPD. ERJ Open Research, 8(4), [00146-2022]. https://doi.org/10.1183/23120541.00146-2022

Copyright Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: https://www.rug.nl/library/open-access/self-archiving-pure/taverneamendment.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.



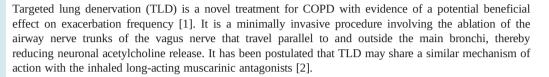
Targeted lung denervation modulates the mucosal epithelial transcriptome in COPD

To the Editor:

Copyright ©The authors 2022

This version is distributed under the terms of the Creative Commons Attribution Non-Commercial Licence 4.0. For commercial reproduction rights and permissions contact permissions@ersnet.org

Received: 9 Aug 2022 Accepted: 22 Aug 2022



In a double-blind, randomised, sham-controlled study (AIRFLOW-2; clinicaltrials.gov identifier NCT02058459), a significant reduction (p<0.001) in respiratory adverse events, which included COPD exacerbations, at 3–6.5 months post-procedure was reported in the TLD arm. Furthermore, over 12.5 months of follow-up, the risk of a severe COPD exacerbation requiring hospitalisation, as assessed *via* time-to-first event analysis, was lower for the treatment group (p=0.039) [1].

We conducted a substudy to evaluate the post-treatment airway mucosal transcriptome using next-generation RNA sequencing (seq) of mucosal brush samples. The aim of this study was to explore gene expression changes between the post-treatment TLD and sham-control patients and provide hypotheses for future investigation into the mechanisms of TLD.

Airway mucosal brush samples collected at the 3-month follow-up visit in both sham-control group and treatment group were used for this study. For each patient, three brushes were collected from the right lower lobe, which constituted one sample. Samples were processed using methods as described previously [3, 4]. Data analysis was performed on a short-read dataset obtained using Illumina next-generation sequencing technology. RNA-seq was conducted using the Illumina NovaSeq 6000 sequencer by GenomeScan (www.genomescan.nl/). The procedure included data quality control, adapter trimming, alignment of short reads and feature counting. Library preparation was checked by calculating ribosomal (and globin) content. Checks for possible sample and barcode contamination were performed and a set of standard quality metrics for the raw dataset was determined using quality control tools (FstQC v0.34 and FastQA). Prior to alignment, the reads were trimmed for adapter sequences using Trimmomatic v0.30. To align the reads of each sample, the human reference GRCh37.75 was used. 25 samples (13 and 12 in the TLD and sham-control arms, respectively) passed quality control and were used in the analysis.

Differential gene analysis was performed using DESeq2 package v1.24.0 on the R platform v3.6.0. The full gene set was filtered for low read counts by excluding all genes with an average fragment per kilobase million (FPKM) of <1. A t-test was then performed for the filtered read counts for each gene (TLD *versus* sham-control); genes were ranked by nominal p-value and the corresponding q-values were calculated to correct for multiple testing. Unfortunately, no genes met the 0.25 false discovery rate (FDR) criterion for transcriptome-wide significance. As the main objective of this initial analysis was to generate hypotheses for future investigation, genes with a nominal p-value of <0.05 were included for the hypergeometric distribution overrepresentation analysis.



Shareable abstract (@ERSpublications)

This study shows that TLD reduces airway epithelial expression of genes related to acetylcholine processing and airway inflammation, which may help to elucidate the mechanism for its effect of reducing severe exacerbations in COPD https://bit.ly/3dWcqZk

Gene set enrichment analysis uses the whole unfiltered gene set and ranks it according to expression

levels. It then uses a database of gene sets to find biologically related pathways that are significantly

Cite this article as: Srikanthan K, Kistemaker L, Slebos D-J, *et al.* Targeted lung denervation modulates the mucosal epithelial transcriptome in COPD. *ERJ Open Res* 2022; 8: 00146-2022 [DOI: 10.1183/23120541.00146-2022].

expressed in the up- or downregulated ends of the ranked gene list. We used the Reactome database (https://reactome.org/), of which four gene sets were significantly downregulated in the TLD arm. The top two downregulated gene sets were related to acetylcholine, the main neurotransmitter involved in the parasympathetic nervous system: "Reactome highly calcium permeable postsynaptic nicotinic acetylcholine receptors" (p=0.008, q=0.047) and "Reactome acetylcholine binding and downstream events" (p=0.03, q=0.04).

In order to perform the HGD analysis, the FPKM >1 genes were separated into upregulated and downregulated genes. Of the upregulated genes, 40 had a nominal p<0.05, but no Reactome gene sets were found to be overrepresented on HGD analysis. Of the downregulated genes, 991 had a nominal p<0.05 and several immunity-related gene sets were overrepresented. On closer inspection, these immunity-related gene sets shared a number of common genes. These 35 genes were fed into the StringDB website (https://string-db.org/), which performs cluster analysis based on known protein–protein interactions. A large cluster of genes was noted to relate to the ubiquitin-proteasome system, which is the cellular machinery used to dispose of misfolded proteins (figure 1).

In the present study, whole-transcriptome sequencing of mucosal brush samples has shown reduced expression of acetylcholine-related genes after TLD. This suggests reductions in mucosal acetylcholine pathways, which may have been indirectly induced by TLD itself. These acetylcholine-related gene sets refer to nicotinic receptors, which are known to be expressed in airway epithelial cells and may even correlate with the development of airway obstruction in COPD [5, 6]. No genes were significantly differentially expressed after correction for multiple testing. However, when genes with nominal p<0.05 were analysed with HGD and StringDB, we identified a cluster of genes common to several overrepresented immunity-related gene sets that relate to the ubiquitin-proteasome system (UPS).

The UPS removes denatured, misfolded, damaged or improperly translated proteins from cells. It is a highly complex pathway that is involved in many important cellular processes, including the regulation of immune and inflammatory responses as well as the cellular response to stress [7]. The UPS is known to regulate CD8⁺ T-cell response to viral infections, activate the NF-κB pathway and manage oxidative stress [8]. It is noted that all three processes are relevant to the pathophysiology of COPD and are increased during periods of exacerbation [9, 10]. Given that COPD exacerbations were reduced in the TLD arm of the AIRFLOW-2 study, we postulate that the cluster of downregulated UPS-related genes is a surrogate marker of reduced airway inflammation and oxidative stress, suggestive of a potential anti-inflammatory mechanism of TLD. Baseline data (table 1) show that there was a tendency toward more severe airways obstruction and worse health-related quality of life in the treatment group. One would expect airways inflammation to be worse in this group with upregulation of immunity-related gene expression. However, our results show the converse, and would support the hypothesis that TLD has suppressed inflammatory pathways. As mentioned previously, there were no significantly differentially expressed genes after the FDR correction was applied. This may be due to low expression levels of genes potentially modifiable by TLD that would require a higher number of biological replicates to attain statistical significance. Correcting for multiple testing is an important step in analysing gene expression data in order to minimise the number of false positive results. By using nominal p-values (<0.05) instead of FDR q-values as our threshold for differential expression, the risk of Type 1 error is amplified. However, we feel this is an acceptable adjustment to make given the aim of this study is to merely generate hypotheses for future TLD studies. Another major limitation was the lack of baseline data and therefore no between-group analyses. This makes it more difficult to interpret the data as a true treatment effect and leaves more room for confounding factors.

Cellular material in bronchial brush samples is unlikely to be purely epithelial in origin. For example, there is likely to be a significant inflammatory cell component within these samples. Bulk RNA-seq methods, such as that used in this study, are unable to resolve these differences in cell origin. Therefore, it must be noted that the gene expression results presented here are likely to have been influenced, in part, by nonepithelial cells. This would act to dilute any signal induced by TLD, which is more likely to modulate gene expression in the epithelium than in luminal inflammatory cells. This issue may be mitigated in future studies by using single-cell RNA-seq, a powerful tool that can assess gene expression at the level of individual cells instead of a global average across all cells in any given sample.

In conclusion, we observed trends in reduced acetylcholine-related gene expression after targeted lung denervation, potentially serving as novel indirect evidence that to some degree, denervation has occurred in these patients. Trends were also seen in downregulation of genes related to immunity and inflammation,

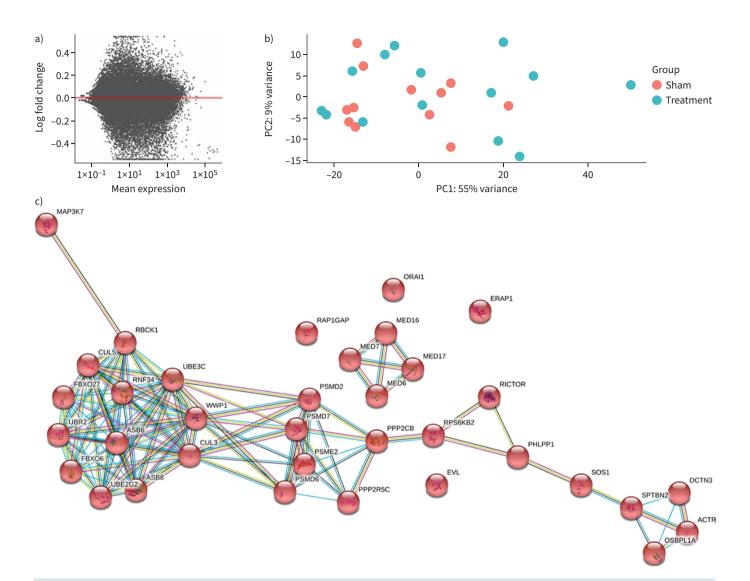


FIGURE 1 a) Mean average plot showing only modest log fold changes; high variance across the range of expression levels; and a lack of any significantly differentially expressed genes. The low number of biological replicates (*i.e.* participants sampled) may account for this. b) Principal component (PC) analysis plot showing horizontal clustering of the sham cases to the left with PC1 accounting for over half of the total variance within the sequencing read count dataset. This is suggestive of a significant difference in gene expression between the two groups. c) StringDB output for HGD-overrepresented downregulated genes. The large cluster of genes on the left relate to the ubiquitin-proteasome system.

TABLE 1 Baseline data with an indication that the treatment group had more severe airways o	bstruction as
well as worse health-related quality of life	

	TLD	Sham
Age (years)	62	62.5
Body mass index (kg·m ⁻²)	25	25.5
Smoking (pack-years)	37	42.5
FEV ₁ (L)	0.92	1.4
FEV ₁ (%)	40	44
SGRQ-C	51	44
CAT	18	15
mMRC	2	2

TLD: targeted lung denervation; FEV₁: forced expiratory volume in 1 s; SGRQ-C: St George's Respiratory Questionnaire for COPD patients; CAT: COPD Assessment Test; mMRC: modified Medical Research Council dyspnoea scale.

specifically the UPS. This may reflect reduced airway inflammation and oxidative stress after TLD, and perhaps explain the mechanism behind its effect on reducing exacerbations in COPD.

Karthi Srikanthan ¹, Loes Kistemaker², Dirk-Jan Slebos ³, Wolfgang Gesierich ⁴, Kaid Darwiche ⁵, Peter Bonta⁶, Gaetan Deslee⁷, Pallav Shah ^{1,9} and Reinoud Gosens^{8,9}

¹Royal Brompton and Harefield NHS Foundation Trust, National Heart and Lung Institute, Imperial College, London, UK. ²Aquilo BV, Groningen, The Netherlands. ³University Medical Center Groningen, Groningen, The Netherlands. ⁴Asklepios-Fachkliniken Munchen Gauting, Gauting, Germany. ⁵Department for Interventional Pneumology, Ruhrlandklinik – University Medicine, Essen, Germany. ⁶Academic Medical Centre, Amsterdam, The Netherlands. ⁷Department of Pulmonary Medicine, INSERM UMRS 1250, CHU of Reims, Reims, France. ⁸Department of Molecular Pharmacology, University of Groningen, Groningen, The Netherlands. ⁹These authors contributed equally.

Corresponding author: Karthi Srikanthan (k.srikanthan@nhs.net)

Provenance: Submitted article, peer reviewed.

Conflict of interest: L. Kistemaker is an employee of Aquilo BV. D-J. Slebos reports grants, and nonfinancial and other support from Nuvaira, Minneapolis, MN, USA, during the conduct of the study. W. Gesierich reports personal fees from PulmonX, grants from PneumrX/BTG and personal fees from AstraZeneca, outside the submitted work. P. Bonta reports support from Nuvaira during the conduct of the study. G. Deslee reports personal fees from Nuvaira during the conduct of the study. G. Deslee reports personal fees from Nuvaira during the conduct of the study. G. Deslee reports personal fees from Nuvaira during the conduct of the study. Respectively, Boehringer Ingelheim, Novartis, Chiesi and AstraZeneca, outside the submitted work. R. Gosens reports support from Nuvaira during the conduct of the study, and grants from Boehringer Ingelheim, Novartis and Aquilo, outside the submitted work; and is a scientific advisor for Aquilo. All other authors have nothing to disclose.

References

- Slebos D-J, Shah PL, Herth FJ, et al. Safety and adverse events after targeted lung denervation for symptomatic moderate to severe COPD (AIRFLOW): a multicenter randomized controlled trial 2019. Am J Respir Crit Care Med 2019; 200: 1477–1486.
- 2 Slebos D-J, Klooster K, Koegelenberg CFN, *et al.* Targeted lung denervation for moderate to severe COPD: a pilot study. *Thorax* 2015; 70: 411–419.
- 3 QIAGEN. RNeasy Mini Handbook. 2019. Available from: https://www.qiagen.com/gb/resources/resourcedetail? id=14e7cf6e-521a-4cf7-8cbc-bf9f6fa33e24
- 4 New England Biolabs. NEBNext Ultra II Directional RNA Library Prep Kit for Illumina. NEBNext Ultra II Directional RNA Library Prep Kit for Illumina. 2015. Available from: https://international.neb.com/products/ e7760-nebnext-ultra-ii-directional-rna-library-prep-kit-for-illumina#Protocols,%20Manuals%20&%20Usage_Manuals
- 5 Wilk JB, Shrine NRG, Loehr LR, *et al.* Genome-wide association studies identify CHRNA5/3 and HTR4 in the development of airflow obstruction. *Am J Respir Crit Care Med* 2012; 186: 622–632.
- 6 Lam DCL, Luo SY, Fu KH, *et al.* Nicotinic acetylcholine receptor expression in human airway correlates with lung function. *Am J Physiol Lung Cell Mol Physiol* 2016; 310: L232–L239.
- 7 Wang J, Maldonado MA. The ubiquitin-proteasome system and its role in inflammatory and autoimmune diseases. *Cell Mol Immunol* 2006; 3: 255–261.
- 8 McCarthy MK, Weinberg JB. The immunoproteasome and viral infection: a complex regulator of inflammation. *Front Microbiol* 2015; 6: 21.
- 9 Rahman I, Adcock IM. Oxidative stress and redox regulation of lung inflammation in COPD. *Eur Respir J* 2006; 28: 219–242.
- **10** Edwards MR, Bartlett NW, Clarke D, *et al.* Targeting the NF-κB pathway in asthma and chronic obstructive pulmonary disease. *Pharmacol Ther* 2009; 121: 1–13.