



**PHYSICAL CHEMISTRY 2010**

**10th International Conference on  
Fundamental and Applied Aspects of  
Physical Chemistry**

Proceedings

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**The Conference is dedicated to the  
100th Anniversary of the academician Pavle Savić birthday  
and  
20th Anniversary of the Society of Physical Chemists of Serbia**

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**21-24 September 2010  
B E L G R A D E**

**ISBN 978-86-82475-17-0**

**Title:** Physical Chemistry 2010. (Proceedings)

**Editors:** S. Anić and Ž. Čupić

**Published by:** Society of Physical Chemists of Serbia, Studentski trg 12-16  
P.O.Box 47, 11158 Beograd, 218, Srbija

**Publisher:** Society of Physical Chemists of Serbia

**For Publisher:** S. Anić, President of Society of Physical Chemists of Serbia

**Printed by:** “Jovan” Printing and Publishing Company; 200 Copies;

Number of pages 16 + 388, **Format:** B5; Printing finished in September  
2010.

**Text and Layout:** “Jovan”

*200 - Copy printing*

*The Society of Physical Chemists of Serbia*

*in co-operation with*

*Institute of Catalysis, Bulgarian Academy of Sciences*

*Boreskov Institute of Catalysis, Siberian Branch of  
the Russian Academy of Sciences*

*Faculty of Physical Chemistry, University of Belgrade, Serbia*

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# Na<sup>+</sup>-DEPENDENT Ca<sup>2+</sup> ION FLUX INHIBITION BY 17 beta-ESTRADIOL IN CAUDATE NUCLEUS MITOCHONDRIA

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## Abstract

In this study the Ca<sup>2+</sup> ion flux modulation in the synaptosomal mitochondria isolated from caudate nucleus (CN) of the ovariectomised rats was examined. 17 beta-estradiol (E2), E2-conjugated to bovine serum albumin (E-BSA), estradiol receptor  $\alpha$  (ER $\alpha$ ) agonist 4,4',4''-(4-propyl-[1H]-pyrazole-1,3,5-triyl) trisphenol (PPT), ER $\beta$  agonist 2,3-bis(4-hydroxyphenyl)-propionitrile (DNP) and ER $\alpha/\beta$  antagonist 7 $\alpha$ ,17 $\beta$ -[9[(4,4,5,5,5-pentafluoropentyl)sulfinyl]nonyl]estra-1,3,5(10)-triene-3,17-diol (ICI 182,780) were used. The Ca<sup>2+</sup> efflux inhibition of about 27% was detected in the presence of 0.5 nmol/l E2, and of about 20% in the case of E-BSA. DNP (10 nmol/l) was as much potent Ca<sup>2+</sup> efflux inhibitor as E2, while PPT (10 nmol/l) hardly had any inhibitory effect (9% efflux decrease). When E2 binding to ER $\alpha$  and ER $\beta$  was prevented by 1  $\mu$ mol/l ICI 182,780, the Ca<sup>2+</sup> efflux inhibition of about 15% was detected. Our results suggest that E2 prevents Ca<sup>2+</sup> efflux from synaptosomal mitochondria due to ER $\beta$  activation rather than ER $\alpha$ . The involvement of the external E2 binding site on the mitochondrial membrane probably different from ER $\alpha/\beta$  should not be excluded because of Ca<sup>2+</sup> efflux inhibition detected in the presence of E-BSA and ICI 182,780. The Ca<sup>2+</sup> efflux modulation could be the mechanism through which E2 exerts its neuromodulatory role in specific brain structures.

## Introduction

Due to their Ca<sup>2+</sup> transport mechanisms mitochondria are organelles critical for Ca<sup>2+</sup> buffering in neuronal cells. The concentration of Ca<sup>2+</sup> in mitochondrial matrix ([Ca<sup>2+</sup>]<sub>m</sub>) regulates the respiration rate, ATP production, reactive oxygen species generation,  $\Delta\Psi_m$  collapse occurrence and also, it is a critical trigger for the permeability transient pore opening and therefore for apoptosis [1].

The CN is an important brain structure necessary for learning and memory, particularly regarding feedback processing. Also, it prevents explosive activation of excitatory synapses by measuring the general activity of cerebral cortex and controlling the threshold potential. Permanently high neuronal activity in this brain region could be the reason for intensive movements of ions, especially Ca<sup>2+</sup> ions.

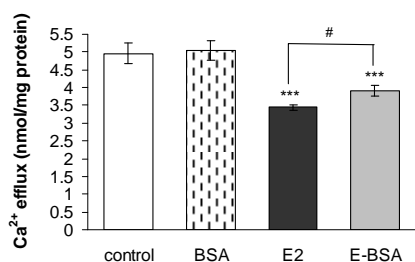
The steroid hormone E2 could modulate the activity of two main mitochondrial  $\text{Ca}^{2+}$  transport mechanisms: I)  $\text{Ca}^{2+}$  influx by the ruthenium red (RR) sensitive uniporter and II)  $\text{Ca}^{2+}$  efflux by the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger. In our previous work using synaptosomal mitochondria isolated from whole rat's brains, the  $\text{Na}^+$ -dependent  $\text{Ca}^{2+}$  efflux inhibition by physiological E2 concentrations was detected [2]. Therefore, the aim of present study was to determine if E2 could influence mitochondrial  $\text{Ca}^{2+}$  transport in CN and to estimate the contribution of  $\text{ER}\alpha$  and/or  $\text{ER}\beta$  in  $\text{Ca}^{2+}$  transport modulated by E2, using ERs antagonist and specific agonists. The observed effects on mitochondrial functions as well as on mitochondrial  $\text{Ca}^{2+}$  sequestration might be a consequence of membrane binding sites and/or mitochondrial estradiol receptors ( $\text{ER}\alpha$  and/or  $\text{ER}\beta$ ) activation.

## Experimental

Synaptosomal mitochondria used for  $\text{Ca}^{2+}$  transport measurements were isolated from the CN of ovariectomised (3 weeks prior to use) female rats. For  $\text{Ca}^{2+}$  transport monitoring mitochondria were preincubated at  $22^\circ\text{C}$  for 10 min in medium containing: 300 mmol/l mannitol, 10 mmol/l KCl, 1 mmol/l maleate, 5 mmol/l glutamate, 10 mmol/l Tris-HCl, pH 7.4. The influx of  $\text{Ca}^{2+}$  to synaptosomal mitochondria was initiated by adding 0.2 mmol/l  $\text{CaCl}_2$  (0.6  $\mu\text{Ci}$   $^{45}\text{CaCl}_2$ ). The reaction lasted 5 min and stopped by RR (17.5  $\mu\text{g}/\text{mg}$  protein). For  $\text{Ca}^{2+}$  efflux monitoring, mitochondria were loaded with  $\text{Ca}^{2+}$  in the same way and after adding RR, the  $\text{Ca}^{2+}$  efflux was initiated by adding NaCl (20 mmol/l) and 0.2 mmol/l EDTA and lasted 5 min. The different agents effects on Na-dependent  $\text{Ca}^{2+}$  efflux were measured by incubating  $\text{Ca}^{2+}$ -preloaded mitochondria with 0.5 nmol/l E2 (10 min), 10 nmol/l DNP (10 min), 10 nmol/l PPT (10 min) and 1  $\mu\text{mol}/\text{l}$  ICI 182,780 (20 min) before efflux initiation.

## Results and discussion

In the present study the  $\text{Ca}^{2+}$  movements through the CN mitochondrial membrane were monitored in order to determine the direct effect of E2 on mitochondrial  $\text{Ca}^{2+}$  flux.

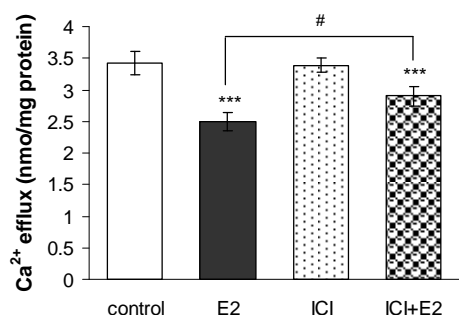


**Fig.1.** Effects of E2 and E-BSA *in vitro* on mitochondrial  $\text{Na}^+$ -dependent  $\text{Ca}^{2+}$  efflux.

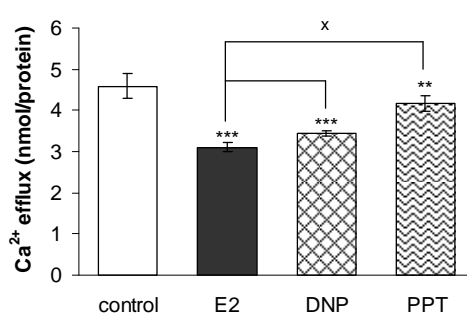
In our model system no effect of E2 on mitochondrial  $\text{Ca}^{2+}$  influx that occurs through the RR-sensitive uniporter was detected (data not shown). To verify if E2 affects  $\text{Na}^+$ -dependent  $\text{Ca}^{2+}$  efflux by acting directly on the mitochondrial membrane, the  $\text{Ca}^{2+}$ -preloaded mitochondria with 0.5 nmol/l E2 or membrane-impermeable E-BSA before efflux initiation were incubated. The efflux in control mitochondria and in the presence of E2, BSA or

E-BSA is presented in Figure 1. The  $\text{Ca}^{2+}$  efflux was decreased by 27% in the presence of E2. BSA did not affect  $\text{Ca}^{2+}$  efflux, while the same concentration of E-BSA reduced  $\text{Ca}^{2+}$  efflux by 20%, indicating inhibition due to an external binding site on the mitochondrial membrane, independent of E2 diffusion into mitochondrial matrix.

The involvement of mitochondrial ER $\alpha$ / $\beta$  during E2 modulation of  $\text{Ca}^{2+}$  efflux, also, was tested. When E2 binding to mitochondrial ER $\alpha$  and ER $\beta$  was prevented by ICI 182,780 pretreatment, the E2 inhibitory effect on  $\text{Ca}^{2+}$  efflux was lowered, but the  $\text{Ca}^{2+}$  efflux decrease of 15% comparing to control still persisted (Fig. 2). The mitochondria incubation with ER $\alpha$  agonist, PPT, led to no change in  $\text{Ca}^{2+}$  efflux with respect to control. However, the ER $\beta$  agonist DNP was almost as much potent inhibitor as E2 and inhibited  $\text{Ca}^{2+}$  efflux by 25% (Fig. 3).



**Fig.2.** Effects of E2 and ICI *in vitro* on mitochondrial Na<sup>+</sup>-dependent Ca<sup>2+</sup> efflux.



**Fig.3.** Effects of E2, DNP and PPT *in vitro* on mitochondrial Na<sup>+</sup>-dependent Ca<sup>2+</sup> efflux.

## Conclusion

In summary, our results confirm that E2 prevents  $\text{Ca}^{2+}$  efflux from caudate nucleus synaptosomal mitochondria by I) direct acting on still unknown membrane E2 binding sites (about 20% inhibition) and II) ER $\beta$  activation (about 25% inhibition), rather than ER $\alpha$ . Most likely, these are the mechanisms through which E2 could exert its neuromodulatory role in specific brain structures.

## Acknowledgements

This study was supported by Serbian Ministry of Sciences, Project No. 143044.

## References

- [1] R. Rizzuto, P. Pinton, D. Ferrari, M. Chami, G. Szabadkai, P.J. Magalhaes, F. Di Virgilio, T. Pozzan, *Oncogene*, 2003, **22**, 8619-8627.
- [2] A. Horvat, S. Petrovic, N. Nedeljkovic, J. V. Martinovic, G. Nikezic, *Gen. Physiol. Biophys.*, 2000, **19**, 59-71.