

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,200

Open access books available

169,000

International authors and editors

185M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Asymptomatic Dengue and Silent Transmission

Pavithra Dilakshini Dayananda and B.G.D. Nissanka K. de Silva

Abstract

With over 90% of infected proportions being asymptomatic to dengue, their possible contribution to silent transmission has generated much attention in epidemic and non-epidemic settings. The challenges in identifying the true asymptomatic representation, owing to no clinical symptoms, have limited scientific knowledge of the asymptomatic dengue, its viral kinetics, immune mechanisms and underlying protective mechanisms in action. The chapter gives an overview of dengue, and its asymptomatic counterparts. It elaborates on the current knowledge in immunity, and immunopathology in symptomatic cases and provides postulations on possible protective mechanisms responsible for the asymptomatic nature of individuals. The chapter further discusses the importance of identifying the asymptomatic proportion in a community and the challenges in diagnosis. It highlights the major role, that asymptomatic carriers play in silent transmission, and its implications and further discuss the possible measures to minimize the transmission risk.

Keywords: dengue, dengue without symptoms, asymptomatic, dengue transmission, silent transmission, transmission risk

1. Introduction

Dengue is considered the most prevalent arthropod-borne viral disease in the world, causing more than 90 million cases and approximately 40,000 deaths per year [1, 2]. Causative agent- Dengue virus (DENV) is a single-stranded RNA virus of the Genus Flavivirus, which is comprised of 4 closely related, antigenically discrete serotypes, DENV1, DENV2, DENV3 and DENV4. However, in 2013 a 5th DENV serotype (DENV5) also has been reported [3]. DENV is transmitted by *Aedes* mosquitoes, mainly *Aedes aegypti* and *Aedes albopictus*. The virus and its vectors are widespread in over 100 countries worldwide, both tropical and subtropical [4]. Since there is no specific medication other than clinical management, the prevention of the disease relies mainly on vector control, and vaccine development is urgently required. Currently, a live attenuated vaccine, chimeric yellow fever 17D—tetravalent dengue vaccine (CYD-TDV), has been licensed for clinical use in some countries, and many candidate vaccines; including live attenuated vaccines, inactivated vaccines, recombinant subunit vaccines, viral vectored vaccines, and DNA vaccines are still under research and development [5].

Many factors have contributed to the expansion of dengue spread such as population growth, urbanization, inadequate water management, poor waste management, lack of effective mosquito control and increased global travel. Changes in global climatic patterns are believed to have expanded the vector habitat range and resulting increased epidemic activity may have caused an increase in the rate of viral genetic change and the emergence of strains or genotypes with greater epidemic potential [6–8].

Dengue has a wide spectrum of clinical outcomes ranging from asymptomatic to symptomatic; resulting in asymptomatic infections, undifferentiated fevers, Dengue Fever (DF), Dengue Hemorrhagic Fever (DHF) and Dengue Shock Syndrome (DSS) [1, 9]; alternatively, they can be classified as dengue with warning signs (DWS), Dengue without warning signs (DWOS) and severe dengue (SD) as suggested by WHO [10]. Each year, 390 million DENV infections occur globally and an estimated 300 million result in asymptomatic/mildly symptomatic [11–14].

Primary Dengue infections are often observed to be asymptomatic and are known to generate immunity to the homologous DENV strain. However, 90% of DWS are known to reportedly occur following a second exposure to a heterologous strain of DENV [15]. It has been observed that sequential or secondary DENV infections are more likely to produce severe diseases [16, 17]. Cross-protection from previous infections and the neutralizing antibody levels also seem to play a major role in determining the level of severity of the disease [18, 19].

2. DWOS; asymptomatic infections

The incidence of dengue infection has been rising over the last five decades [20] and the majority of infected individuals are known to have no or insufficient symptoms to result in clinical presentation [12, 14]. Mildly symptomatic or subclinical infections are mostly referred to as DENV infections without major symptoms requiring medical attention, while the patients who have virologically or serologically confirmed dengue with no reported or detected symptoms are named dengue ‘asymptomatic’ patients [12].

Although asymptomatic infections are considered more frequent than symptomatic, the relative number of cases is observed to be vary according to the year of infection, geographical area, the epidemiological context, the immunological status of individuals, and the circulating serotypes and viral strains [21]. The ratios of asymptomatic to symptomatic cases have been shown to vary from 2.1:1 to 13:1 around the world [22] making this change in the proportion of symptomatic to asymptomatic infections, one of the main contributors to the rise in the dengue incidence.

A study carried out in Thailand during 1980–1981 reported a ratio of asymptomatic to symptomatic cases as 6.1:1, while this ratio was refined as 5.5:1 for DENV 1 cases, 4.5:1 for DENV 2 cases and for entirely asymptomatic for DENV4 cases [16]. During the period of 1998–2000, a similar survey in Northern Thailand reported a ratio of 1.1:1 [23]. A study carried out in Central America reported a ratio of 13:1 during the period of 2001–2003 when DENV2 was dominant and a ratio of 6.1 when DENV1 was frequent [24]. It has been reported that the Health authorities in Singapore assumed ratios between 2:1 and 10:1 during the period of 2006–2007 [25]. Studies from Sri Lanka have shown that the asymptomatic to symptomatic ratio was 3.4:8.4 between 2008 and 2010 [22]. In a pediatric dengue cohort study in Nicaragua, a wide variation from 16.5:1 in 2006–2007 [26] to 1.2:1 in 2009–2010 [27] has been shown. All these studies reveal the extensive variation of asymptomatic to

symptomatic ratios, where the differences might be attributed to the extrinsic and intrinsic factors of the host and the circulating virus types [24, 28].

Clinically apparent dengue is frequently studied with various research objectives. However, the science behind asymptomatic dengue is inadequately investigated mainly due to the challenges in diagnosing asymptomatic cases on time. Thus, understanding the host factors such as the role of immunity on the lack of clinical symptoms and or the other protective mechanisms for being symptomless has mainly been based on careful investigation of symptomatic cases.

2.1 Dengue; immunity

Once an immune-susceptible host meets with the infection, an acute, self-limiting febrile systematic syndrome is known to develop usually within initial 4–7 days. This is known to be associated with strong innate and adaptive immune responses [29].

2.1.1 Innate immune response

At the site of the mosquito bite, Langerhans cells, dermal cells and interstitial dendritic cells of the innate immune system become the initial targets for DENV [29]. The three cell types, monocytes, macrophages, and dendritic cells which are tolerable for DENV infection act as the main phagocytic cells of the innate immune system, responsible for detecting and removing hostile pathogens. These three major phagocytic cells also function as antigen-presenting cells critical for the initiation, expansion, and polarization of adaptive cellular immunity [30]. All these innate immunity mechanisms are triggered immediately upon pathogen invasion and play a significant role in managing pathogenic infection. The killing of target cells is associated with inflammatory cytokine and or chemokine responses [30]. However, DENV has evolved successfully to suppress innate immunity and to infect the host productively using passive and active evasive strategies, which have a negative effect on the subsequent production of antigen-specific adaptive immunity to these viruses [31].

2.1.2 Adaptive immune response (cell-mediated immune response)

Many studies have shown that the adaptive immune response to DENV has protective as well as detrimental aspects [32]. Dengue-induced immune enhancement plays a major role in the clinical manifestations of dengue disease. The imbalanced and deregulated, cell-mediated immunity is considered as a major component in severe dengue conditions [29]. It is hypothesized that DENV infection of monocytes and macrophages increases T cell activation, leading to the release of cytokines and chemical mediators such as Tumor Necrosis Factors (TNF), Interleukins (IL), Platelet Activating Factors (PAF), complement components and histamines causing increased vascular permeability, plasma leakage, shock and malfunction of the coagulatory system resulting in hemorrhage and shock [33, 34]. In this phenomenon, DENV infection of dendritic cells strongly activates CD4+ and CD8+ T cells which produce a surplus of cytokines, which recruit numerous other cytokines and chemical mediators that further increase the vascular permeability of the host [34, 35].

2.1.3 Adaptive immune response (humoral response)

Individuals infected with DENV generate serum antibody titers that provide long-term protection against future homotypic infections with the same serotype [32].

Infection with DENV also builds a degree of cross-protective immunity against the other three DENV serotypes by means of heterotypic (cross-reactive) IgG antibodies which usually persist for a duration of several months to a few years [36]. The produced heterotypic antibody titers are known to reduce over long time periods of approximately 4 to 20 years [37]. However, conversely, the homotypic IgG antibody titers are known to increase over time which could be due to the preferential survival of long-lived memory B cells producing homotypic antibodies [37].

2.2 Dengue; immunopathogenesis

The postulated hypotheses on dengue immunopathogenesis include the antibody enhancement theory, cross-reactive memory T cells activation and the original antigenic sin where all in a way cause either an overproduction or a skewed profile of cytokine release [38, 39].

Antibody-dependent enhancement (ADE); The leading hypothesis is that DHF occurs via ADE of a DENV infection [40–43]. Preexisting heterologous, cross-reactive antibodies from a previous infection (or maternal antibodies in infants) recognize and bind to heterologous DENV in a secondary or tertiary infection (primary infection in infants), are unable to neutralize this virus, either because they are non-neutralizing, or due to inadequate avidity or occupancy [41, 44–46]. These non-neutralizing antibody–virus complexes are known to increase the infection of monocytes via their Fc receptors, dramatically increasing viral replication and load, thereby causing DHF. Weak and non-neutralizing cross-reactive antibodies induced from immunodominant B cell epitopes are known to comprise the majority of the humoral immune response to DENV infection [38, 47, 48].

Cross-reactive memory T cells activation and Original antigenic sin; the second hypothesis indicates that there is a highly skewed cellular response to heterologous DENV infection motivated by low affinity, cross-reactive memory CD4⁺ and CD8⁺ T cells [49–51]. Cross-reactive T cell epitopes have been identified across the DENV proteome, however, immunodominant CD8⁺ T cell epitopes in dengue non-structural protein NS3 have been found strongly associated with DHF [15, 52, 53]. The common theme throughout DENV immune enhancement is the concept of “original antigenic sin,” which describes the shift in the hierarchy of immunodominance that occurs when previous exposure to cross-reactive antigens alters and inhibits the subsequent immune response to related antigens, either as a new infection or by vaccination [49]. Both humoral and cellular responses are known to be plagued by such misdirected or inappropriate heterotypic immunity [54].

All these mechanisms are known to increase the activation of immune cells, resulting in impaired immune responses or cytokine storms that cause endothelium dysfunction and increase vascular permeability. However, multiple host and viral factors seem to be influencing the determination of the disease severity of DENV infections via favorable and unfavorable interactions thus have triggered much research interest [55].

2.3 DWOS; asymptomatic infections - protective mechanisms

Fundamental immunological differences in the immune responses associated with symptomatic and asymptomatic infections have been studied [56, 57]. The kinetics of asymptomatic infections are known to differ from the symptomatic infections in the magnitude of the viremia and the rate of clearance [58]. The protective mechanisms that contribute to the lack of clinical manifestations in individuals, that remain asymptomatic

or inapparent compared to symptomatic dengue cases have been found interesting and are still being investigated. Epidemiological risk factors such as age, duration between consecutive dengue infections, DENV serotypes of previous infections, concentrations of pre-existing heterotypic neutralizing antibodies, interactions between viral genotype within the serotype and resulting immune responses are known to be associated with these subclinical outcomes after the dengue infection [12, 23, 59–61].

In general, it is regarded that secondary infections are associated with more severe disease due to the phenomenon of ADE and/or cross-reactive T cells [62]. However, post-secondary infections are known to induce different immune responsiveness in susceptible hosts and have also been found to impact upon inapparent rates [63]. Thus, the role of previous infection in perhaps decreasing or increasing the risk of infection causing it inapparent needs to be further investigated [12, 64]. Alexander et al. (2020) have studied the impact of frequent immune boosting; that occurs as a result of frequent disease exposure in dengue-endemic areas, on the fluctuating symptomatic and asymptomatic ratios [28]. It has been reported that antibodies play a greater role than immune cells in heterologous DENV infections [65]. Neutralizing antibodies seem to play a major role in this and it is evident that, the individuals who are previously exposed to DENV, manifest clinical symptoms differently due to the presence of pre-existing neutralizing antibodies resulting in asymptomatic or inapparent dengue status on many occasions. Furthermore, high concentrations of neutralizing antibodies against DENV infection have been frequently observed in asymptomatic individuals [61, 66–68].

Variations in immune reactions to the virus have been reported in dengue asymptomatic and symptomatic patients. In a study carried out by Simon-Loriere et al. (2017), the inflammatory pathways and innate immune responses were found similar in asymptomatic and symptomatic diseases. However, the expression of proteins related to antigen presentation and subsequent T and B cell activation pathways were found differently regulated, independent of the viral load or previous DENV infections. Asymptomatic individuals have been found to have increased T cell responses with feedback regulation compared to symptomatic counterparts [57]. According to his findings, asymptomatic infections seem to be determined by increased activation of the adaptive immune response and properly controlled mechanisms leading to the removal of viral infection without excessive immune activations [57].

Furthermore, apart from immune status, host genetic factors are considered to have an impact on the protective mechanisms in asymptomatic diseases [56], which involves a complex network of genes that are expressed differentially in the asymptomatic or inapparent individuals. A polymorphism in Fc gamma receptors (FcγRIIA) has been found to be associated with inapparent infections compared to symptomatic infections with DF or DHF in the Cuban population [56]. Moreover, according to the studies of [67] a broad down-regulation of host defense response (innate, adaptive, cytokines and matrix metalloprotease) genes in asymptomatic individuals against symptomatic patients. A selective up-regulation of distinct genes which are associated with protection has been observed [66]. However, these observations warrant further investigations in order to correlate their expression with conferring protection against clinical dengue infections.

2.4 DWOS; asymptomatic infections- detection

Detection of DWOS or asymptomatic infections is known to be challenging. The symptomatic dengue can be clinically suspected based on the symptoms and

a confirmatory laboratory diagnosis will provide a definite diagnosis. Detection of asymptomatic cases happens only based on laboratory diagnosis, since there are insufficient or no clinical cues for infection [68].

Direct diagnostic methods such as molecular and antigen-detecting methods are not usually considered convenient to detect asymptomatic infections owing to the shorter period of viremia after the infection. Serological tests such as HAI, ELISA and PRNT have been accepted as suitable methods to detect DWOS and have been frequently applied in detecting asymptomatic dengue cases than the direct diagnostic methods [68]. However, direct methods to detect acute infection and indirect methods; mostly the serological methods and further, mosquito inoculation techniques have also been incorporated in many studies for detecting asymptomatic infection in high-risk cohorts (**Figure 1**) [69–71].

Surveillance studies for DWOS or asymptomatic infections are carried out in the general population over a long period of time with frequent blood sampling and testing [16, 23] and by screening the dengue high-risk groups [71].

2.5 Dengue transmission

Transmission of DENV among human hosts occurs through horizontal and vertical transmission pathways. In horizontal transmission, viruses are transmitted among individuals of the same generation. Human-to-mosquito transmission is known as the most common mode of horizontal transmission, while transmission through blood transfusion [25, 70, 72–76] and organ transplants [25, 77] have also been infrequently reported. In addition to these transmission modes, a few cases of nosocomial transmission through needle stick injury and mucocutaneous exposure have also been reported [78, 79]. The difficulties in differentiating non-vector transmission from vector or mosquito transmission in dengue-endemic areas could be the result of the observed infrequency of records of these cases [80].

Studies have been carried out to investigate the possible sexual transmission of DENV. So far, cases of DENV in semen [81, 82], and vaginal secretions [83] have been

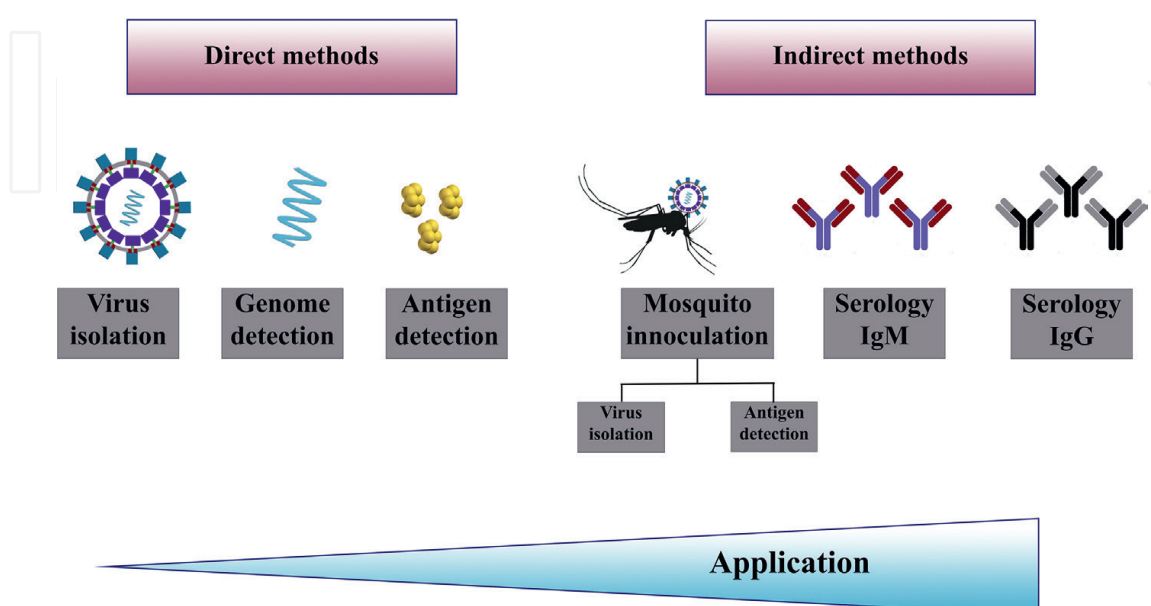


Figure 1. Schematic diagram illustrating the application of diagnostic tests for the detection of asymptomatic dengue.

rarely recorded. However, in 2019, two cases of possible sexual transmission were reported in Spain and South Korea [84, 85]. Thus, though plausible, sexual transmission of dengue is considered extremely rare and uncommon in endemic communities [86].

Vertical transmission occurs when the virus is transmitted from mothers to their offspring, through intrapartum transmission [87–89] or transmission at the onset of delivery [80, 87]. Although DENV virus particles have been found in breast milk, the studies are insufficient to conclude the transmission of DENV via breast milk [90], however considering the benefits and immunological protection from breast milk to infants, breastfeeding in DENV-infected mothers are encouraged in dengue -endemic regions [91, 92].

2.5.1 Human to mosquito transmission of DENV

A susceptible female *Aedes* mosquito acquires a DENV infection after it consumes a blood meal from a dengue viremic person. When viremic blood reaches the mosquito midgut, the extracellular virus binds to undefined receptors on the cellular surface of the midgut epithelium. Once the virus is capable of successfully infecting and replicating in midgut epithelial cells, a new progeny of viruses is shed into the hemocoel, where it can, later on, disseminate and infect secondary tissues, legs, brain and salivary glands [93]. The duration of the viral incubation between the time of ingestion and reaching the salivary glands, where mosquitoes become infectious is known as the extrinsic incubation period (EIP), which is generally considered as 8–12 days [94]. Upon adequate viral replication in the salivary glands, the mosquito becomes a potential vector to transmit DENV to a new host during the next probing or feeding event [93].

The factors influencing the transmission of DENV from humans to mosquitoes include viral, host, vector and environmental aspects. In terms of host factors, viral titer in the human plasma and duration of human infectiousness are considered. The amount of viral titer circulating in the blood of an infected human influences the possibility of a mosquito becoming infected after a blood meal. Mosquito infectious dose, or the viremia in humans that is required to infect 50% of mosquitoes differs between viral serotypes [95]. A dose-response relationship is generally observed with an increasing number of DENV RNA copies [96]. The period between infection and the onset of infectiousness in a human is called the Intrinsic Incubation Period (IIP). The intrinsic incubation period of a human varies, and it is typically considered as 4–7 days [94]. It is estimated that onward transmission results from mosquitoes biting during the pre-symptomatic phase of DENV infections in most cases than, during the post-symptomatic period [14]. Further, it is also reflected that, patients with a high early viremia have a greater probability of having an extended duration of DENV infectiousness. Furthermore, host immune factors [96] and host stimuli for mosquito attraction such as body temperature, body odor, blood type [87, 97], etc. are also known to contribute as host factors for dengue transmission.

As for vectors, diurnal and crepuscular biting behavior of both *Aedes* mosquito species [98], anthropophilic nature, considerable flying span, and highly domesticated nature, especially of the primary vector, *Ae. aegypti* mosquitoes [99] have made them excellent vectors in disease transmission. Mosquito susceptibility to infection and vector competence (VC), which elaborates on mosquito infection, dissemination and onward transmission of the virus, plays a major role in transmission. Relative vector competence of two major vectors *Ae. aegypti* and *Ae. albopictus* have been extensively studied. *Ae. albopictus* are known, more susceptible to midgut infection than *Ae. aegypti*, but the ability to disseminate the virus of *Ae. aegypti* has been found greater

suggesting a greater potential for transmission in nature [100]. The susceptibility for DENV in mosquitoes of different geographical strains has been reported [101, 102] and population-specific differences in the susceptibility with each serotype, have revealed consistent patterns of high and low infection [103]. Further, differential susceptibility by different viral isolates of genotypes within the same serotype in a single geographical population has also been reported [101, 104, 105].

Dengue virus is also known to manipulate the biology and behaviors of the infected host to facilitate virus transmission [93, 106]. Studies on the blood-feeding behavior of DENV-infected mosquitoes have investigated the time duration of probing and feeding [107], transmission efficiency during probing [108], and motivation and avidity to feed [106, 109] and revealed the relationships of such in disease spread.

Environmental factors have been known to play a major role in dengue transmission via mosquito vectors [110]. The temperature has been known to have implications in altering mosquito VC to transmit viruses. The lower temperatures are known to induce slow virus replication and high temperatures are known to induce increased virus replication resulting in reduced EIPs [100, 111]. Changes in the humidity levels are also known to intervene with the vector competency of vector mosquitoes, which affect DENV transmission [110]. Research interest in the factors contributing to DENV mosquito transmission is ongoing and in-depth studies are warranted [93].

2.6 Silent transmission; vector and non-vector transmission

Studies on vector transmission of DENV from asymptomatic patients are rare and the level of mosquito infectiousness has not been adequately investigated [69]. It was long assumed that people with inapparent and asymptomatic infections fail to infect mosquitoes and have low viremia levels. Many studies have reported lower viremia in asymptomatic infections than those of symptomatic infections but also in detectable levels [58, 69, 112–115].

It has been shown that people with asymptomatic infections have had 100-fold lower infectious doses of viruses to mosquitoes that eventually have resulted in larger viral loads in infected mosquitoes [116]. This was also evident to us in a study carried out in Sri Lanka, where silent transmission from asymptomatic individuals (with no detectable viremia or sometimes no detectable antigen levels), to vector mosquitoes (with detectable antigen levels) was observed [71]. A recent study has reported a slower viral decay rate in asymptomatic subjects compared to symptomatic individuals, enabling the asymptomatic cases more available for silent transmission [58]. Furthermore, studies to evaluate mosquito infectivity of asymptomatic subjects have shown a significant increase in mosquito infectiousness among asymptomatic cases than the symptomatic cases (**Figure 2**) [18, 69], postulating that strong immunological response and high cytokine levels during symptomatic illness reduce human infectiousness to mosquitoes in symptomatic dengue cases [69].

Non-vector transmission of DENV via atypical routes such as blood transfusion, organ transplant and intrapartum transmission has been confirmed in many studies, and the possibility of these transmission routes originating from asymptomatic, pre-symptomatic or subclinical cases has also been discussed [12, 73, 117–120]. Furthermore, vertically transmitted dengue in a neonate born to a mother with asymptomatic dengue infection has been reported in a recent case study from Sri Lanka, and instances, where such cases are misdiagnosed owing to no maternal history in asymptomatic mothers have been discussed [121].

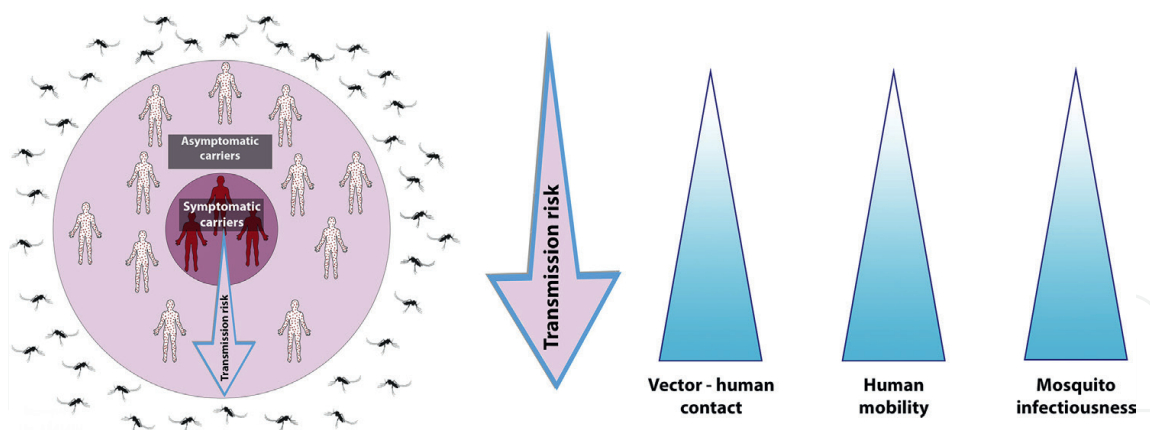


Figure 2. Schematic diagram illustrating the increasing transmission risk towards the asymptomatic proportion of an infected community.

According to the modeling analysis of Bosh et al., (2018), it has been suggested that inapparent infections contribute appreciably to DENV transmission and its disease burden [14]. Further, their finding that approximately one-quarter of an individual's infectiousness occurs prior to symptom onset, supports the hypothesis that a large proportion of human to-mosquito transmission is silent [14]. Collectively, these evidences show that asymptomatic cases play a major role in silent transmission, having a high transmission risk compared to symptomatic cases (**Figure 2**). Furthermore, the fact that the vector- host contact is considerably high in asymptomatic carriers through their daily routines compared to symptomatic cases, who will be hospitalized or less accessible should also be accounted with great concern. In addition, human mobility is also known to play a key role in the spread [122, 123], thus silent transmission of dengue via uninterrupted daily routings of these asymptomatic or mildly symptomatic carriers can be identified as a key factor contributing to the dengue spread than the symptomatic cases [69, 100] (**Figure 2**).

2.7 Seroprevalence and risk of antibody-dependent enhancement

Relatively high dengue seroprevalence among the dengue endemic communities around the world has been reported [13, 24, 124–129]. Comparative to the number of confirmed dengue cases, an increased level of dengue infection suggested by high IgG seropositivity in endemic areas, has revealed a vast majority of DENV infections [128]. Attributing to the fact that the majority of dengue infections in these communities are either asymptomatic or inapparent [11, 14, 16, 23, 65].

As a consequence of this, a considerable proportion of the population who are immune to a circulating dengue serotype/strain after an epidemic will be created. Co-circulation of several dengue serotypes in dengue-endemic areas has been reported [129, 130]. Worsening the situation, the prolonged seroprevalence in symptomatic and asymptomatic individuals has also been observed in studies [13]. Thus, this proportion would be at risk of developing ADE or severe dengue in a subsequent epidemic of a differing dengue serotype with non-neutralizing antibodies or neutralizing antibodies at sub-neutralizing levels (**Figure 3**).

Furthermore, the transmission of heterogeneous anti-dengue antibodies from symptomatic or asymptomatic cases through blood transfusion or organ transplant, and transmission of maternal antibodies to infants has also been suggested to enhance

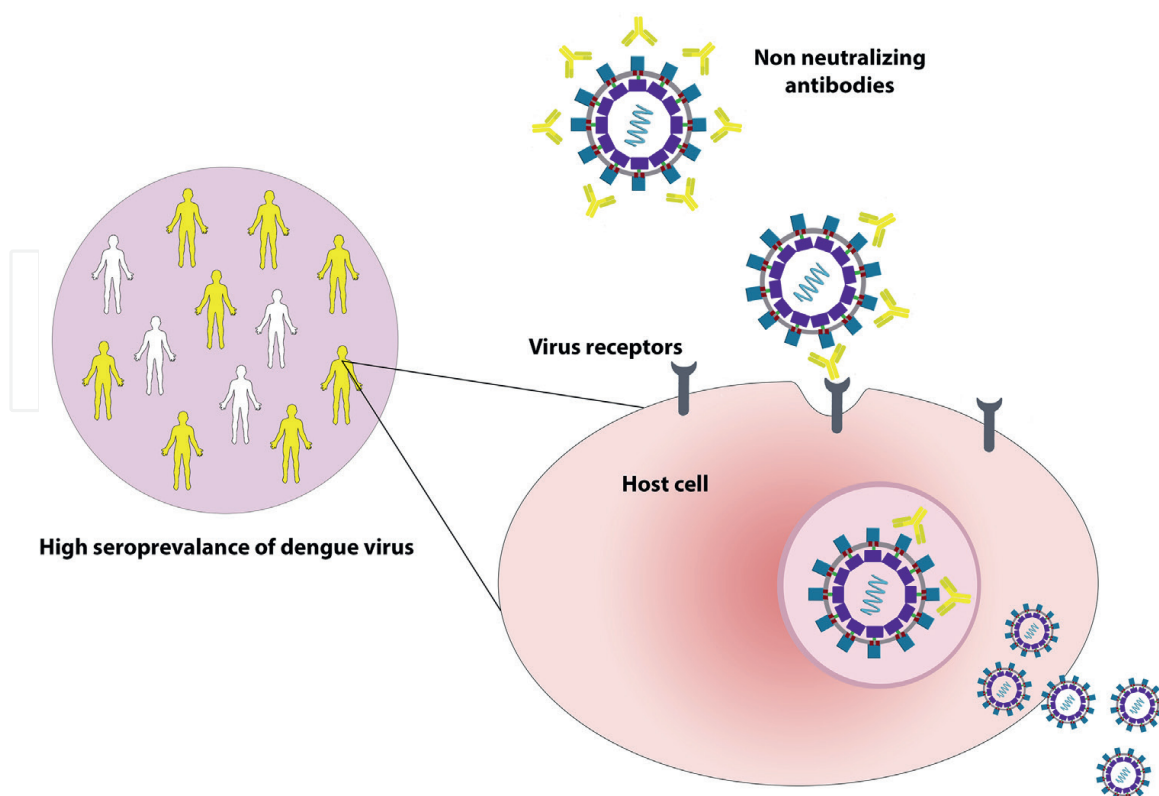


Figure 3. Schematic diagram depicting the high seroprevalence of dengue virus in dengue-endemic communities and the risk of developing antibody-dependent enhancement during subsequent infections.

the viral infectivity and virulence in recipients who will later be exposed to a heterotypic DENV infection [131].

The global incidence of DHF/DSS has increased more than 500-fold in recent years [11]. And the risk of dengue virus or its antibodies which can be transmitted through such different passive modes of transmission has been identified as a major counterpart.

2.8 Prevention; a way forward

Prevention from silent transmission through mosquito vectors can only be achieved by vector control, in a setting where asymptomatic carriers can only be identified by certain laboratory identification techniques. However, the routine fogging and insecticide spraying practices just after a case report (peri-domestic space spraying) can be taken as an initial step to reduce the risk of transmission from asymptomatic carriers. Although many studies have suggested the importance of screening the populations for DENV, and dengue seroprevalence, no available efficient measures and or diagnostic services for such events prevail in many dengue-endemic countries [75].

However, there is a need to incorporate integrated approaches including increasing awareness among the community, establishing routine diagnostic methods for screening asymptomatic carriers and incorporating preventive measures to reduce the exposure, which will eventually help in reducing the dengue burden [75].

Many endemic countries have identified the risk of transfusion-associated transmission from asymptomatic donors, and have adopted policies where they recommend screening of blood products. The positive blood donors will be deferred for periods depending on the endemicity of the region. Similarly, transplant guidelines in

some countries have recommended dengue screening prior to transplantation and a specific deferral period before taking up for transplant surgery if the donor or recipient is found positive for dengue.

Vaccines and herd immunity; though seems like the only promising solution, limited knowledge of immune responses against dengue infection, lack of human or animal model of disease, and suboptimal assay strategies to detect immune responses after infection or vaccination, which are some barriers to the vaccine and drug development. Furthermore, in addition to the protection against symptomatic infection, it is also important to assess protection against asymptomatic infection.

3. Conclusion

Asymptomatic or inapparent dengue infections provide a fundamental link in the chain of disease transmission in dengue-endemic communities. The knowledge gap in understanding the viral kinetics of asymptomatic individuals along with their immunoresponses must be urgently fulfilled and investigative studies on such should be encouraged. Understanding the presence and the prevalence of asymptomatic to symptomatic proportion of a community enables a glimpse of the targeted population and helps in introducing disease management strategies. The chapter highlights the increased transmission risk towards the asymptomatic carriers of the community, attributed to the increased vector-human contact, human mobility and mosquito infectiousness. All precautions must be taken to reduce dengue transmission in a community via vector and non-vector routes. Furthermore, the seroprevalence of a community must be routinely monitored and the vaccine efficacy in such settings depending on the endemicity, should be closely evaluated.

Acknowledgements

We appreciate the support of Dr. Dulan Jayasekara, for designing the figures for us.

Conflict of interest

The authors declare no conflict of interest.

IntechOpen

Author details


Pavithra Dilakshini Dayananda¹ and B.G.D. Nissanka K. de Silva^{2*}

1 Genetics and Molecular Biology Unit, Faculty of Applied Sciences, University of Sri Jayewardenepura, Nugegoda, Sri Lanka

2 Department of Zoology, Faculty of Applied Sciences, University of Sri Jayewardenepura, Nugegoda, Sri Lanka

*Address all correspondence to: nissanka@sci.sjp.ac.lk

IntechOpen

© 2023 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

References

- [1] WHO Fact Sheet: Vector-Borne Diseases [Internet]. 2022. Available from: <https://www.who.int/news-room/fact-sheets/detail/vector-borne-diseases> [Accessed: October 20, 2022]
- [2] Kading RC, Brault AC, Beckham JD. Global perspectives on arbovirus outbreaks: A 2020 snapshot. *Tropical Medicine and Infectious Disease*. 2020;5(3):142. DOI: 10.3390/tropicalmed5030142
- [3] Mustafa MS, Rasotgi V, Jain S, Gupta VJ. Discovery of fifth serotype of dengue virus (DENV-5): A new public health dilemma in dengue control. *Medical journal armed forces India*. 2015;71(1):67-70. DOI: 10.1016/j.mjafi.2014.09.011
- [4] Dehghani R, Kassiri H. A review on epidemiology of dengue viral infection as an emerging disease. *Research Journal of Pharmacy and Technology*. 2021;14(4):2296-2301. DOI: 10.52711/0974-360X.2021.00406
- [5] Deng SQ, Yang X, Wei Y, Chen JT, Wang XJ, Peng HJ. A review on dengue vaccine development. *Vaccine*. 2020;8(1):63. DOI: 10.3390/vaccines8010063
- [6] Gubler D. Dengue and dengue hemorrhagic fever. *Clinical Microbiology Reviews*. 1998;11(3):480-496. DOI: 10.1128/CMR.11.3.480
- [7] Messer WB, Gubler DJ, Harris E, Sivananthan K, De Silva AM. Emergence and global spread of a dengue serotype 3, subtype III virus. *Emerging Infectious Diseases*. 2003;9(7):800. DOI: 10.3201%2F0907.030038
- [8] Dash PK, Sharma S, Soni M, Agarwal A, Sahni AK, Parida M. Complete genome sequencing and evolutionary phylogeography analysis of Indian isolates of dengue virus type 1. *Virus research*. 2015;195:124-134. DOI: 10.1016/j.virusres.2014.08.018
- [9] World Health Organization (WHO). *Strategies for the Prevention of Blindness in National Programmes: a Primary Health Care Approach*. 2nd ed. [Internet]. 1997. Available from: <https://www.apps.who.int/iris/handle/10665/41887> [Accessed: October 20, 2022]
- [10] World Health Organization (WHO). *Dengue: Guidelines for Diagnosis, Treatment, Prevention and Control*. New Edition. Geneva: World Health Organization [Internet] 2009. Available from: https://www.apps.who.int/iris/bitstream/handle/10665/44188/9789241547871_eng.pdf [Accessed December 18, 2019].
- [11] Bhatt S, Gething PW, Brady OJ, Messina JP, Farlow AW, Moyes CL, et al. The global distribution and burden of dengue. *Nature*. 2013;496(7446):504-507. DOI: 10.1038/nature12060
- [12] Grange L, Simon-Loriere E, Sakuntabhai A, Gresh L, Paul R, Harris E. Epidemiological risk factors associated with high global frequency of inapparent dengue virus infections. *Frontiers in Immunology*. 2014;5:280. DOI: 10.3389/fimmu.2014.00280
- [13] Luo S, Cui W, Li C, Ling F, Fu T, Liu Q, et al. Seroprevalence of dengue IgG antibodies in symptomatic and asymptomatic individuals three years after an outbreak in Zhejiang Province, China. *BMC Infectious Diseases*. 2018;18(1):92. DOI: 10.1186/s12879-018-3000-5

- [14] Ten Bosch QA, Clapham HE, Lambrechts L, Duong V, Buchy P, Althouse BM, et al. Contributions from the silent majority dominate dengue virus transmission. *PLoS Pathogens*. 2018;**14**(5):e1006965. DOI: 10.1371/journal.ppat.1006965
- [15] Mathew A, Rothman AL. Understanding the contribution of cellular immunity to dengue disease pathogenesis. *Immunological Reviews*. 2008;**225**(1):300-313. DOI: 10.1111/j.1600-065X.2008.00678.x
- [16] Burke DS, Nisalak A, Johnson DE, Scott RM. A prospective study of dengue infections in Bangkok. *The American Journal of Tropical Medicine and Hygiene*. 1988;**38**(1):172-180. DOI: 10.4269/ajtmh.1988.38.172
- [17] Vaughn DW, Green S, Kalayanarooj S, Innis BL, Nimmannitya S, Suntayakorn S, et al. Dengue viremia titer, antibody response pattern, and virus serotype correlate with disease severity. *The Journal of Infectious Diseases*. 2000;**181**(1):2-9. DOI: 10.1086/315215
- [18] Anderson KB, Endy TP, Thomas SJ. The dynamic role of dengue cross-reactive immunity: Changing the approach to defining vaccine safety and efficacy. *The Lancet Infectious Diseases*. 2018;**18**(10):e333-e338. DOI: 10.1016/S1473-3099(18)30126-9
- [19] Azami NA, Moi ML, Ami Y, Suzaki Y, Lim CK, Taniguchi S, et al. Genotype-specific and cross-reactive neutralizing antibodies induced by dengue virus infection: Detection of antibodies with different levels of neutralizing activities against homologous and heterologous genotypes of dengue virus type 2 in common marmosets (*Callithrix jacchus*). *Virology Journal*. 2018;**15**(1):1-2. DOI: 10.1186/s12985-018-0967-x
- [20] Murray NE, Quam MB, Wilder-Smith A. Epidemiology of dengue: Past, present and future prospects. *Clinical Epidemiology*. 2013;**5**:299. DOI: 10.2147%2FCLEPS34440
- [21] Chastel C. Eventual role of asymptomatic cases of dengue for the introduction and spread of dengue viruses in non-endemic regions. *Frontiers in Physiology*. 2012;**3**:70. DOI: 10.3389/fphys.2012.00070
- [22] Malavige GN, Jeewandara C, Ghose A, Somathilake G, Tissera H. Changing epidemiology of dengue in Sri Lanka—Challenges for the future. *PLoS Neglected Tropical Diseases*. 2021;**15**(8):e0009624. DOI: 10.1371/journal.pntd.0009624
- [23] Endy TP, Chunsuttiwat S, Nisalak A, Libraty DH, Green S, Rothman AL, et al. Epidemiology of inapparent and symptomatic acute dengue virus infection: A prospective study of primary school children in Kamphaeng Phet, Thailand. *American Journal of Epidemiology*. 2002;**156**(1):40-51. DOI: 10.1093/aje/kwf005
- [24] Balmaseda A, Hammond SN, Tellez Y, Imhoff L, Rodriguez Y, Saborío SI, et al. High seroprevalence of antibodies against dengue virus in a prospective study of schoolchildren in Managua, Nicaragua. *Tropical medicine & International Health*. 2006;**11**(6):935-942. DOI: 10.1111/j.1365-3156.2006.01641.x
- [25] Wilder-Smith A, Chen LH, Massad E, Wilson ME. Threat of dengue to blood safety in dengue-endemic countries. *Emerging Infectious Diseases*. 2009;**15**(1):8. DOI: 10.3201%2Fid1501.071097
- [26] Balmaseda A, Standish K, Mercado JC, Matute JC, Tellez Y,

- Saborío S, et al. Trends in patterns of dengue transmission over 4 years in a pediatric cohort study in Nicaragua. *The Journal of Infectious Diseases*. 2010;**201**(1):5-14. DOI: 10.1086/648592
- [27] Gordon A, Kuan G, Mercado JC, Gresh L, Avilés W, Balmaseda A, et al. The Nicaraguan pediatric dengue cohort study: Incidence of inapparent and symptomatic dengue virus infections, 2004-2010. *PLoS Neglected Tropical Diseases*. 2013;**7**(9):e2462. DOI: 10.1086/648592
- [28] Alexander LW, Ben-Shachar R, Katzelnick LC, Kuan G, Balmaseda A, Harris E, et al. Boosting can explain patterns of fluctuations of ratios of inapparent to symptomatic dengue virus infections. *Proceedings of the National Academy of Sciences*. 2013;**110**(14):e2013941118. DOI: 10.1073/pnas.2013941118
- [29] Guabiraba R, Ryffel B. Dengue virus infection: Current concepts in immune mechanisms and lessons from murine models. *Immunology*. 2014;**141**(2):143-156. DOI: 10.1111/imm.12188
- [30] Sun P, Kochel TJ. The battle between infection and host immune responses of dengue virus and its implication in dengue disease pathogenesis. *The Scientific World Journal*. 2013;**2013**. DOI: 10.1155/2013/843469
- [31] Morrison J, Aguirre S, Fernandez-Sesma A. Innate immunity evasion by dengue virus. *Viruses*. 2012;**4**(3):397-413. DOI: 10.3390/v4030397
- [32] Rothman AL. Immunity to dengue virus: A tale of original antigenic sin and tropical cytokine storms. *Nature Reviews Immunology*. 2011;**11**(8):532-543. DOI: 10.1038/nri3014
- [33] Chaturvedi U, Agarwal R, Elbishbishi E, Mustafa A. Cytokine cascade in dengue hemorrhagic fever: Implications for pathogenesis. *FEMS Immunology & Medical Microbiology*. 2000;**28**(3):183-188. DOI: 10.1111/j.1574-695X.2000.tb01474.x
- [34] Yeo A, Azhar N, Yeow W, Talbot C Jr, Khan M, Shankar E, et al. Lack of clinical manifestations in asymptomatic dengue infection is attributed to broad down-regulation and selective up-regulation of host defense response genes. *PLoS One*. 2014;**9**(4):e92240. DOI: 10.1111/j.1574-695X.2000.tb01474.x
- [35] Kurane I, Ennis FA. Cytokines in dengue virus infections: Role of cytokines in the pathogenesis of dengue hemorrhagic fever. *Seminars in Virology*. Dec 1994;**5**(6):443-448. DOI: 10.1006/smvy.1994.1050
- [36] Bonaparte M, Huleatt J, Hodge S, Zheng L, Lustig Y, DiazGranados CA, et al. Evaluation of dengue serological tests available in Puerto Rico for identification of prior dengue infection for prevaccination screening. *Diagnostic Microbiology and Infectious Disease*. 2020;**96**(3):114918. DOI: 10.1016/j.diagmicrobio.2019.114918
- [37] Guzman MG, Alvarez M, Rodriguez-Roche R, Bernardo L, Montes T, Vazquez S, et al. Neutralizing antibodies after infection with dengue 1 virus. *Emerging Infectious Diseases*. 2007;**13**(2):282. DOI: 10.3201%2F1302.060539
- [38] Crill WD, Hughes HR, Delorey MJ, Chang GJ. Humoral immune responses of dengue fever patients using epitope-specific serotype-2 virus-like particle antigens. *PLoS One*. 2009;**4**(4):e4991. DOI: 10.1371/journal.pone.0004991
- [39] Rathakrishnan A, Wang SM, Hu Y, Khan AM, Ponnampalavanar S, Lum LC,

et al. Cytokine expression profile of dengue patients at different phases of illness. *PLoS One*. 2012;7(12):e52215. DOI: 10.1371/journal.pone.0052215

[40] Hawkes RA. Enhancement of the infectivity of arboviruses by specific antisera produced in domestic fowls. *Australian Journal of Experimental Biology and Medical Science*. 1964;42(4):465-482. DOI: 10.1038/icb.1964.44

[41] Halstead SB. Antibody, macrophages, dengue virus infection, shock, and hemorrhage: A pathogenetic cascade. *Reviews of Infectious Diseases*. 1989;11(Supplement_4):S830-S839. DOI: 10.1093/clinids/11.Supplement_4.S830

[42] Morens DM. Antibody-dependent enhancement of infection and the pathogenesis of viral disease. *Clinical Infectious Diseases*. 1994;19(3):500-512. DOI: 10.1093/clinids/19.3.500

[43] Thomas S, Redfern JB, Lidbury BA, Mahalingam S. Antibody-dependent enhancement and vaccine development. *Expert Review of Vaccines*. 2006;5(4):409-412. DOI: 10.1093/clinids/19.3.500

[44] Pierson TC, Fremont DH, Kuhn RJ, Diamond MS. Structural insights into the mechanisms of antibody-mediated neutralization of flavivirus infection: Implications for vaccine development. *Cell Host & Microbe*. 2008;4(3):229-238. DOI: 10.1016/j.chom.2008.08.004

[45] Guzman MG, Alvarez M, Halstead SB. Secondary infection as a risk factor for dengue hemorrhagic fever/dengue shock syndrome: An historical perspective and role of antibody-dependent enhancement of infection. *Archives of Virology*. 2013;158(7):1445-1459. DOI: 10.1007/s00705-013-1645-3

[46] Flipse J, Wilschut J, Smit JM. Molecular mechanisms involved in antibody-dependent enhancement of dengue virus infection in humans. *Traffic*. 2013;14(1):25-35. DOI: 10.1111/tra.12012

[47] Lai CY, Tsai WY, Lin SR, Kao CL, Hu HP, King CC, et al. Antibodies to envelope glycoprotein of dengue virus during the natural course of infection are predominantly cross-reactive and recognize epitopes containing highly conserved residues at the fusion loop of domain II. *Journal of Virology*. 2008;82(13):6631-6643. DOI: 10.1128/JVI.00316-08

[48] Crill W, Hughes H, Trainor N, Davis B, Whitney M, Chang G. Sculpting humoral immunity through dengue vaccination to enhance protective immunity. *Frontiers in Immunology*. 2012;3:334. DOI: 10.3389/fimmu.2012.00334

[49] Brehm MA, Pinto AK, Daniels KA, Schneck JP, Welsh RM, Selin LK. T cell immunodominance and maintenance of memory regulated by unexpectedly cross-reactive pathogens. *Nature Immunology*. 2002;3(7):627-634. DOI: 10.1038/ni806

[50] Welsh RM, Selin LK. No one is naive: The significance of heterologous T-cell immunity. *Nature Reviews Immunology*. 2002;2(6):417-426. DOI: 10.1038/nri820

[51] Welsh RM, Selin LK, Szomolanyi-Tsuda E. Immunological memory to viral infections. *Annual Review of Immunology*. 2004;22(1):711-743. DOI: 10.1146/annurev.immunol.22.012703.104527

[52] Duangchinda T, Dejnirattisai W, Vasanawathana S, Limpitikul W, Tangthawornchaikul N, Malasit P, et al. Immunodominant T-cell responses to dengue virus NS3 are associated

with DHF. Proceedings of the National Academy of Sciences. 2010;**107**(39):16922-16927. DOI: 10.1073/pnas.1010867107

[53] Friberg H, Burns L, Woda M, Kalayanarooj S, Endy TP, Stephens HA, et al. Memory CD8+ T cells from naturally acquired primary dengue virus infection are highly cross-reactive. *Immunology and Cell Biology*. 2011;**89**(1):122-129. DOI: 10.1038/icb.2010.61

[54] Welsh RM, Fujinami RS. Pathogenic epitopes, heterologous immunity and vaccine design. *Nature reviews Microbiology*. 2007;**5**(7):555-563. DOI: 10.1038/nrmicro1709

[55] Puerta-Guardo H, Biering SB, Harris E, Pavia-Ruz N, Vázquez-Prokopec G. Dengue Immunopathogenesis: A Crosstalk between Host and Viral Factors Leading to Disease: Part II—DENV Infection, Adaptive Immune Responses, and NS1 Pathogenesis. London, UK, London, UK: IntechOpen; 2020 Available from: <http://www.hdl.handle.net/10625/59929>

[56] García G, Sierra B, Pérez AB, Aguirre E, Rosado I, Gonzalez N, et al. Asymptomatic dengue infection in a Cuban population confirms the protective role of the RR variant of the FcγRIIIa polymorphism. *The American Journal of Tropical Medicine and Hygiene*. 2010;**82**(6):1153. DOI: 10.4269/ajtmh.2010.09-0353

[57] Simon-Lorière E, Duong V, Tawfik A, Ung S, Ly S, Casadémont I, et al. Increased adaptive immune responses and proper feedback regulation protect against clinical dengue. *Science Translational Medicine*. 2017;**9**(405):eaal5088. DOI: 10.1126/scitranslmed.aal5088

[58] Matangkasombut P, Manopwisedjaroen K, Pitabut N, Thaloengsok S, Suraamornkul S,

Yingtaweesak T, et al. Dengue viremia kinetics in asymptomatic and symptomatic infection. *International Journal of Infectious Diseases*. 2020;**101**:90-97. DOI: 10.1016/j.ijid.2020.09.1446

[59] Oh Ainle M, Balmaseda A, Macalalad AR, Tellez Y, Zody MC, Saborío S, et al. Dynamics of dengue disease severity determined by the interplay between viral genetics and serotype-specific immunity. *Science Translational Medicine*. 2011;**3**(114):114ra128. DOI: 10.1126/scitranslmed.3003084

[60] Montoya M, Gresh L, Mercado JC, Williams KL, Vargas MJ, Gutierrez G, et al. Symptomatic versus inapparent outcome in repeat dengue virus infections is influenced by the time interval between infections and study year. *PLoS Neglected Tropical Diseases*. 2013;**7**(8):e2357. DOI: 10.1371/journal.pntd.0002357

[61] Katzelnick LC, Montoya M, Gresh L, Balmaseda A, Harris E. Neutralizing antibody titers against dengue virus correlate with protection from symptomatic infection in a longitudinal cohort. *Proceedings of the National Academy of Sciences*. 2016;**113**(3):728-733

[62] Halstead SB, O'Rourke EJ. Dengue viruses and mononuclear phagocytes. I. Infection enhancement by non-neutralizing antibody. *The Journal of Experimental Medicine*. 1977;**146**(1):201-217. DOI: 10.1084/jem.146.1.201

[63] Olkowski S, Forshey BM, Morrison AC, Rocha C, Vilcarrromero S, Halsey ES, et al. Reduced risk of disease during postsecondary dengue virus infections. *The Journal of Infectious Diseases*. 2013;**208**(6):1026-1033. DOI: 10.1093/infdis/jit273

- [64] Clapham H, Hay J, Routledge I, Takahashi S, Choisy M, Cummings D, et al. Seroepidemiologic study designs for determining SARS-COV-2 transmission and immunity. *Emerging Infectious Diseases*. 2020;**26**(9):1978
- [65] Kyle JL, Harris E. Global spread and persistence of dengue. *Annual Review of Microbiology*. 2008;**62**(1):71-92. DOI: 10.3201%2Fcid2609.201840
- [66] Yeo AS, Azhar NA, Yeow W, Talbot CC Jr, Khan MA, Shankar EM, et al. Lack of clinical manifestations in asymptomatic dengue infection is attributed to broad down-regulation and selective up-regulation of host defence response genes. *PLoS One*. 2014;**9**(4):e92240. DOI: 10.1371/journal.pone.0092240
- [67] Corbett KS, Katzelnick L, Tissera H, Amerasinghe A, De Silva AD, de Silva AM. Preexisting neutralizing antibody responses distinguish clinically inapparent and apparent dengue virus infections in a Sri Lankan pediatric cohort. *The Journal of Infectious Diseases*. 2015;**211**(4):590-599. DOI: 10.1093/infdis/jiu481
- [68] Chatchen S, Sabchareon A, Sirivichayakul C. Serodiagnosis of asymptomatic dengue infection. *Asian Pacific Journal of Tropical Medicine*. 2017;**10**(1):11-14. DOI: 10.1016/j.apjtm.2016.12.002
- [69] Duong V, Lambrechts L, Paul RE, Ly S, Lay RS, Long KC, et al. Asymptomatic humans transmit dengue virus to mosquitoes. *Proceedings of the National Academy of Sciences*. 2015;**112**(47):14688-14693. DOI: 10.1073/pnas.1508114112
- [70] Tsai JJ, Lin PC, Tsai CY, Wang YH, Liu LT. Low frequency of asymptomatic dengue virus-infected donors in blood donor centers during the largest dengue outbreak in Taiwan. *PLoS One*. 2018;**13**(10):e0205248. DOI: 10.1371/journal.pone.0205248
- [71] Dayananda PD, De Silva DG, Silva D, Ranawaka R, Fernando L, de Silva BG. Asymptomatic Dengue Infection in Household Contacts of Dengue Patients and the Relationship to Immune Status of Patients and Mosquito Infectiousness. *Proceedings of the 22nd Annual Scientific Congress 2019*. Sri Lanka College of Pediatricians. 29th to 31st August 2019 at Hotel Cinnamon Grand, Colombo. 2019:61-62
- [72] Pozzetto B, Memmi M, Garraud O. Is transfusion-transmitted dengue fever a potential public health threat? *World Journal of Virology*. 2015;**4**(2):113. DOI: 10.5501%2Fwjv4.i2.113
- [73] Tambyah PA, Koay ES, Poon ML, Lin RV, Ong BK. Dengue hemorrhagic fever transmitted by blood transfusion. *New England Journal of Medicine*. 2008;**359**(14):1526-1527. DOI: 10.1056/NEJMc0708673
- [74] Linnen JM, Vinelli E, Sabino EC, Tobler LH, Hyland C, Lee TH, et al. Dengue viremia in blood donors from Honduras, Brazil, and Australia. *Transfusion*. 2008;**48**(7):1355-1362. DOI: 10.1111/j.1537-2995.2008.01772.x
- [75] Ashshi MA, Alghamdi S, El-Shemi AG, Almdani S, Refaat B, Mohamed AM, et al. Seroprevalence of asymptomatic dengue virus infection and its antibodies among healthy/eligible Saudi blood donors: Findings from holy Makkah city. *Virology: Research and Treatment*. 2017;**8**:1178122X17691261. DOI: 10.1177/1178122X17691261
- [76] Rafique I, Saqib MA, Munir MA, Qureshi H, Iqbal R, Ahmed W, et al. Asymptomatic dengue infection in adults of major cities of Pakistan. *Asian*

Pacific Journal of Tropical Medicine. 2017;**10**(10):1002-1006. DOI: 10.1016/j.apjtm.2017.09.013

[77] Wiwanitkit V. Unusual mode of transmission of dengue. *The Journal of Infection in Developing Countries*. 2010;**4**(01):051-054. DOI: 10.3855/jidc.145

[78] Chen R, Han GZ. Dengue in China: Comprehensive phylogenetic evaluation reveals evidence of endemicity and complex genetic diversity. *The American journal of tropical medicine and hygiene*. 2016;**94**(1):198. DOI: 10.4269%2Fajtmh.15-0546

[79] Chen LH, Wilson ME. Non-Vector Transmission of Dengue and Other Mosquito-Borne Flaviviruses. *Dengue Bulletin*. 2005;**29**:18-31

[80] Teo D, Ng LC, Lam S. Is dengue a threat to the blood supply? *Transfusion Medicine*. 2009;**19**(2):66-77. DOI: 10.1111/j.1365-3148.2009.00916.x

[81] Molton JS, Low I, Choy MM, Aw PP, Hibberd ML, Tambyah PA, et al. Dengue virus not detected in human semen. *Journal of Travel Medicine*. 2018;**25**(1):tay023. DOI: 10.1093/jtm/tay023

[82] Lalle E, Colavita F, Iannetta M, Teklè SG, Carletti F, Scorzolini L, et al. Prolonged detection of dengue virus RNA in the semen of a man returning from Thailand to Italy, January 2018. *Eurosurveillance*. 2018;**23**(18):18-00197

[83] Iannetta M, Lalle E, Musso M, Carletti F, Scorzolini L, D'Abramo A, et al. Persistent detection of dengue virus RNA in vaginal secretion of a woman returning from Sri Lanka to Italy, April 2017. *Eurosurveillance*. 2017;**22**(34):30600

[84] Norman FF, Henríquez-Camacho C, Díaz-Menendez M, Chamorro S, Pou D,

Molina I, et al. Imported arbovirus infections in Spain, 2009-2018. *Emerging Infectious Diseases*. 2020;**26**(4):658. DOI: 10.3201%2Fcid2604.190443

[85] Lee C, Lee H. Probable female to male sexual transmission of dengue virus infection. *Infectious Diseases*. 2019;**51**(2):150-152. DOI: 10.1080/23744235.2018.1521004

[86] Wilder-Smith A. Can dengue virus be sexually transmitted? *Journal of Travel Medicine*. 2019;**26**(3):tay157. DOI: 10.1093/jtm/tay157

[87] Rigau-Perez JG, Vorndam AV, Clark GG. The dengue and dengue hemorrhagic fever epidemic in Puerto Rico, 1994-1995. *The American Journal of Tropical Medicine and Hygiene*. 2001;**64**(1):67-74. DOI: 10.4269/ajtmh.2001.64.67

[88] Sirinavin S, Nuntnarumit P, Supapannachart S, Boonkasidecha S, Techasaensiri C, Yoksarn S. Vertical dengue infection: Case reports and review. *The Pediatric Infectious Disease Journal*. 2004;**23**(11):1042-1047. DOI: 10.1097/01.inf.0000143644.95692.0e

[89] Tran A, Chastel C. Mosquito-borne arboviruses and pregnancy: Pathological consequences for the mother and infant. A general review. *Bulletin de la Societe de Pathologie Exotique (1990)*. 2008;**101**(5):418-424.

[90] Barthel A, Gourinat AC, Cazorla C, Joubert C, Dupont-Rouzeyrol M, Descloux E. Breast milk as a possible route of vertical transmission of dengue virus? *Clinical Infectious Diseases*. 2013;**57**(3):415-417. DOI: 10.1093/cid/cit227

[91] Chheda S, Keeney SE, Goldman AS. Chapter 163 - Immunology of Human Milk and Host Immunity. In: Polin RA, Fox WW, Abman SH, et. al., editors.

Fetal and Neonatal Physiology (Third Edition). 2004;2:1610-1620. ISBN 9780721696546. DOI: 10.1016/B978-0-7216-9654-6.50166-1

[92] Gregory KE, Walker WA. Immunologic factors in human milk and disease prevention in the preterm infant. *Current Pediatrics Reports*. 2013;1(4):222-228. DOI: 10.1007/s40124-013-0028-2

[93] Carrington LB, Simmons CP. Human to mosquito transmission of dengue viruses. *Frontiers in Immunology*. 2014;5:290. DOI: 10.3389/fimmu.2014.00290

[94] Chan M, Johansson MA. The incubation periods of dengue viruses. *PLoS One*. 2012;7(11):e50972. DOI: 10.1371/journal.pone.0050972

[95] Novelo M, Hall MD, Pak D, Young PR, Holmes EC, McGraw EA. Intra-host growth kinetics of dengue virus in the mosquito *Aedes aegypti*. *PLoS Pathogens*. 2019;15(12):e1008218. DOI: 10.1371/journal.ppat.1008218

[96] Nguyen NM, Thi Hue Kien D, Tuan TV, Quyen NT, Tran CN, Vo Thi L, et al. Host and viral features of human dengue cases shape the population of infected and infectious *Aedes aegypti* mosquitoes. *Proceedings of the National Academy of Sciences*. 2013;110(22):9072. DOI: 10.1073/pnas.1303395110

[97] Takken W, Verhulst NO. Host preferences of blood-feeding mosquitoes. *Annual Review of Entomology*. 2013;58(1):433-453. DOI: 10.1146/annurev-ento-120811-153618

[98] Service, M.W. *Medical Entomology for Students*. 5th ed. Cambridge: Cambridge University Press; 2004. pp. 1-325

[99] Scott TW, Naksathit A, Day JF, Kittayapong P, Edman JD. A fitness

advantage for *Aedes aegypti* and the viruses it transmits when females feed only on human blood. *The American Journal of Tropical Medicine and Hygiene*. 1997;57(2):235-239

[100] Lambrechts L, Scott TW, Gubler DJ. Consequences of the expanding global distribution of *Aedes albopictus* for dengue virus transmission. *PLoS Neglected Tropical Diseases*. 2010;4(5):e646

[101] Armstrong PM, Rico-Hesse R. Differential susceptibility of *Aedes aegypti* to infection by the American and southeast Asian genotypes of dengue type 2 virus. *Vector Borne and Zoonotic Diseases*. 2001;1(2):159-168

[102] Armstrong PM, Rico-Hesse R. Efficiency of dengue serotype 2 virus strains to infect and disseminate in *Aedes aegypti*. *The American Journal of Tropical Medicine and Hygiene*. 2003;68(5):539

[103] Rico-Hesse R. Dengue virus virulence and transmission determinants. *Curr Top Microbiol Immunol*. 2010;338:45-55. DOI: 10.1007/978-3-642-02215-9_4. PMID: 19802577; PMCID: PMC3057078

[104] Gubler D, Nalim S, Tan R, Saipan H, Saroso J. Variation in susceptibility to oral infection with dengue viruses among geographic strains of *Aedes aegypti*. *The American Journal of Tropical Medicine and Hygiene*. 1979;28(6):1045-1052

[105] Tardieux I, Poupel O, Lapchin L, Rodhain F. Variation among strains of *Aedes aegypti* in susceptibility to oral infection with dengue virus type 2. *The American journal of Tropical Medicine and Hygiene*. 1990;43(3):308-313

[106] Lima-Camara TN, Bruno RV, Luz PM, Castro MG, Lourenço-de-Oliveira R, Sorgine MH,

- et al. Dengue infection increases the locomotor activity of *Aedes aegypti* females. *PLoS One*. 2011;**6**(3):e17690
- [107] Sim S, Ramirez JL, Dimopoulos G. Dengue virus infection of the *Aedes aegypti* salivary gland and chemosensory apparatus induces genes that modulate infection and blood-feeding behavior. *PLoS Pathogens*. 2012;**8**(3):e1002631
- [108] Putnam JL, Scott TW. The effect of multiple host contacts on the infectivity of dengue-2 virus-infected *Aedes aegypti*. *The Journal of Parasitology*. Apr 1995;**81**(2):170-174. DOI: 10.1007/bf00931626
- [109] Maciel-de-Freitas R, Sylvestre G, Gandini M, Koella JC. The influence of dengue virus serotype-2 infection on *Aedes aegypti* (Diptera: Culicidae) motivation and avidity to blood feed. *PLoS One*. 2013;**8**(6):e65252. DOI: 10.1371/journal.pone.0065252
- [110] Morin CW, Comrie AC, Ernst K. Climate and dengue transmission: Evidence and implications. *Environmental Health Perspectives*. 2013;**121**(11-12):1264-1272. DOI: 10.1289/ehp.1306556
- [111] Xiao FZ, Zhang Y, Deng YQ, He S, Xie HG, Zhou XN, et al. The effect of temperature on the extrinsic incubation period and infection rate of dengue virus serotype 2 infection in *Aedes albopictus*. *Archives of Virology*. 2014;**159**(11):3053-3057. DOI: 10.1007/s00705-014-2051-1
- [112] Beckett CG, Kosasih H, Faisal I, Nurhayati, Tan R, Widjaja S, et al. Early detection of dengue infections using cluster sampling around index cases. *American Journal of Tropical Medicine and Hygiene*. Jun 2005;**72**(6):777-782
- [113] Reyes M, Mercado JC, Standish K, Matute JC, Ortega O, Moraga B, et al. Index cluster study of dengue virus infection in Nicaragua. *The American Journal of Tropical Medicine and Hygiene*. 2010;**83**(3):683. DOI: 10.4269/ajtmh.2010.10-0023
- [114] Dussart P, Baril L, Petit L, Beniguel L, Quang LC, Ly S, et al. Clinical and virological study of dengue cases and the members of their households: The multinational DENFRAME project. *PLoS Neglected Tropical Diseases*. 2012;**6**(1):e1482. DOI: 10.1371/journal.pntd.0001482
- [115] Ly S, Fortas C, Duong V, Benmarhnia T, Sakuntabhai A, Paul R, et al. Asymptomatic dengue virus infections, Cambodia, 2012-2013. *Emerging Infectious Diseases*. 2019;**25**(7):1354. DOI: 10.3201/eid2507.181794
- [116] Lambrechts L, Fansiri T, Pongsiri A, Thaisomboonsuk B, Klungthong C, Richardson JH, et al. Dengue-1 virus clade replacement in Thailand associated with enhanced mosquito transmission. *Journal of Virology*. 2012;**86**(3):1853-1861. DOI: 10.1128/JVI.06458-11
- [117] Chuang VW, Wong TY, Leung YH, Ma ES, Law YL, Tsang OT, et al. Review of dengue fever cases in Hong Kong during 1998 to 2005. *Hong Kong Medical Journal*. 2008;**14**(3):170
- [118] Stramer SL, Linnen JM, Carrick JM, Foster GA, Krysztof DE, Zou S, et al. Dengue viremia in blood donors identified by RNA and detection of dengue transfusion transmission during the 2007 dengue outbreak in Puerto Rico. *Transfusion*. 2012;**52**(8):1657-1666. DOI: 10.1111/j.1537-2995.2012.03566.x
- [119] Rosso F, Pineda JC, Sanz AM, Cedano JA, Caicedo LA. Transmission of dengue virus from deceased donors to solid organ transplant recipients: Case report and literature review. *Brazilian*

- Journal of Infectious Diseases. 2018;22:63-69. DOI: 10.1016/j.bjid.2018.01.001
- [120] Cedano JA, Mora BL, Parra-Lara LG, Manzano-Nuñez R, Rosso F. A scoping review of transmission of dengue virus from donors to recipients after solid organ transplantation. *Transactions of The Royal Society of Tropical Medicine and Hygiene*. 2019;113(8):431-436. DOI: 10.1093/trstmh/trz024
- [121] Gamhewage NC, Weerasekera M, Nazmy MH. Vertically transmitted dengue in a neonate born to a mother with asymptomatic dengue infection. *Sri Lanka Journal of Child Health*. 2019;48(2):168-169
- [122] Stoddard ST, Forshey BM, Morrison AC, Paz-Soldan VA, Vazquez-Prokopec GM, Astete H, et al. House-to-house human movement drives dengue virus transmission. *Proceedings of the National Academy of Sciences*. 2013;110(3):994-999. DOI: 10.1073/pnas.1213349110
- [123] Wesolowski A, Qureshi T, Boni MF, Sundsøy PR, Johansson MA, Rasheed SB, et al. Impact of human mobility on the emergence of dengue epidemics in Pakistan. *Proceedings of the National Academy of Sciences*. 2015;112(38):11887-11892. DOI: 10.1073/pnas.1504964112
- [124] Malavige GN, Fernando S, Aaskov J, Sivayogan S, Dissanayaka T, Peelawattage MK, Dabare M. Seroprevalence of Anti-dengue Virus Antibodies in Children in Colombo District, Sri Lanka. 2006. Available from: <http://www.who.int/iris/handle/10665/170265>
- [125] Tam CC, Tissera H, de Silva AM, De Silva AD, Margolis HS, Amarasinge A. Estimates of dengue force of infection in children in Colombo, Sri Lanka. *PLoS Neglected Tropical Diseases*. 2013;7(6):e2259. DOI: 10.1371/journal.pntd.0002259
- [126] Jeewandara C, Gomes L, Paranavitane SA, Tantirimudalige M, Panapitiya SS, Jayewardene A, et al. Change in dengue and Japanese encephalitis seroprevalence rates in Sri Lanka. *PLoS One*. 2015;10(12):e0144799. DOI: 10.1371/journal.pone.0144799
- [127] Jamjoom GA, Azhar EI, Kao MA, Radadi RM. Seroepidemiology of asymptomatic dengue virus infection in Jeddah, Saudi Arabia. *Virology: Research and Treatment*. 2016;7:VRT-S34187. DOI: 10.4137/VRT.S34187
- [128] Biswal S, Borja-Tabora C, Vargas LM, Velásquez H, Alera MT, Sierra V, et al. Efficacy of a tetravalent dengue vaccine in healthy children aged 4-16 years: A randomised, placebo-controlled, phase 3 trial. *The Lancet*. 2020;395(10234):1423-1433. DOI: 10.1016/S0140-6736(20)30414-1
- [129] Kanakaratne N, Wahala WM, Messer WB, Tissera HA, Shahani A, Abeysinghe N, et al. Severe dengue epidemics in Sri Lanka, 2003-2006. *Emerging Infectious Diseases*. 2009;15(2):192. DOI: 10.3201%2F1502.080926
- [130] Azhar E, Kao M, Niedrig M, Masri B, Godus A, Badierah R, Khan N, Almazrooa A, Ashshi A, Jamjoom G. Virological Diagnosis of Dengue Fever in Jeddah, Saudi Arabia: Comparison Between RT-PCR and Virus Isolation in Cell Culture. Available from: <https://www.edoc.rki.de/handle/176904/828#>
- [131] Ribas-Silva RC, Eid AA. Dengue antibodies in blood donors. *Revista brasileira de hematologia e hemoterapia*. 2012;34:193-195. DOI: 10.5581/1516-8484.20120048