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1 Signatures of purifying selection and site-specific positive selection on the mitochondrial DNA of
2 dromedary camels (*Camelus dromedarius*)

3
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18
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20
21

22 **Key words:**

23 adaptive divergence, mtDNA protein-coding genes, signatures of selection, PAML
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31 **Abstract**

32 The two species of the Old World Camelini tribe, dromedary and Bactrian camels, show
33 superior adaptability to the different environmental conditions they populate, e.g. desert,
34 mountains and coastal areas, which might be associated with adaptive variations on their
35 mitochondrial DNA. Here, we investigate signatures of natural selection in the 13-
36 mitochondrial protein-coding genes of different dromedary camel populations from the
37 Arabian Peninsula, Africa and southwest Asia. The full mitogenome sequences of 42
38 dromedaries, 38 domestic Bactrian, 29 wild Bactrian camels and 31 samples representing the
39 New World Lamini tribe reveal species-wise genetic distinction among Camelidae family
40 species, with no evidence of geographic distinction among dromedary camels. We observe
41 gene-wide signals of adaptive divergence between the Old World and New World camels, with
42 evidence of purifying selection among Old World camel species. Upon comparing the different
43 Camelidae tribes, 27 amino acid substitutions across ten mtDNA protein-coding genes were
44 found to be under positive selection, in which, 24 codons were defined to be under positive
45 adaptive divergence between Old World and New World camels. Seven codons belonging to
46 three genes demonstrated positive selection in dromedary lineage. A total of 89 codons were
47 found to be under positive selection in Camelidae family based on investigating the impact of
48 amino acid replacement on the physiochemical properties of proteins, including equilibrium
49 constant and surrounding hydrophobicity. These mtDNA variants under positive selection in
50 the Camelidae family might be associated with their adaptation to their contrasting
51 environments.

52

53

54

55 **Introduction**

56 The Camelidae family is divided into two tribes: Old World camels (Camelini) and New World
57 camels (Lamini). The Camelini tribe is composed of two domestic species: single-humped
58 dromedary camels (*Camelus dromedarius*) and two-humped Bactrian camels (*Camelus*
59 *bactrianus*), and a two-humped wild Bactrian camel species (*Camelus ferus*). The Lamini tribe,
60 which diverged from the Camelini tribe approximately 16.3 million years ago, is composed of
61 two domestic species: Llama (*Lama glama*) and Alpaca (*Vicugna pacos*), and two wild species:
62 Guanaco (*Lama guanicoe*) and Vicuna (*Vicugna Vicugna*) (Wu et al., 2014, Burger et al.,
63 2019).

64

65 The camel species of these two tribes populate contrasting environmental niches. Dromedary
66 camels, also known as Arabian camels, are mainly distributed throughout hot arid regions of
67 Africa, Arabian Peninsula and southwest Asia, while some populations inhabit the mountains
68 and coast of the west, southwest and southeast of the Arabian Peninsula (Almathen et al., 2018).
69 Domestic and wild Bactrian camels are widespread in central Asia, including Mongolia, China
70 and Kazakhstan, throughout cold plains and rocky mountains reaching altitudes up to 4000m
71 (Burger et al., 2019). Camels of the Lamini tribe populate the high-altitude mountains of South
72 America which can reach 7000m in height (Wu et al., 2014).

73

74 These contrasting environmental conditions might be associated with different adaptations
75 associated with energy production and thermoregulation. For example, under water stress,
76 dromedary camels show adaptive heterothermy, during which their body temperature
77 fluctuates by 6°C – 8°C, between 34°C to more than 40°C, to avoid water loss through
78 perspiration during high temperatures (Tibary and El Allali, 2020). While the environmental

79 niche occupied by Lamini species is mainly characterized by high altitude hypoxia, for which
80 Lamini camels exhibit superior adaptability (Beall, 2007).

81

82 The mitochondria are involved in producing ATP molecules as an energy source and
83 generating heat to maintain body temperature (Blier et al., 2001). This organelle carries its own
84 genetic material comprising 13 protein-coding genes (*ND1*, *ND2*, *ND3*, *ND4*, *ND4L*, *ND5*,
85 *ND6*, *CYTB*, *COX-1*, *COX-2*, *COX-3*, *ATP6* and *ATP8*), 22 tRNA and 2 rRNA genes. The
86 encoded proteins are part of the electron transport chain (Complex I – Complex V) embedded
87 in the inner-membrane of each mitochondrion, which have been found to be targets of selective
88 pressures to meet these two primary physiological functions (Blier et al., 2001).

89

90 The analysis of mtDNA protein-encoding genes has been shown to be effective at identifying
91 footprints of purifying and adaptive (positive) selection in a range of species. Ruiz-Pesini *et al.*
92 (2004) report that in humans living within the arctic zone, several mitochondrial genes exhibit
93 signatures of adaptive selection, including *ND2*, *ND4* and *ATP6*. Signatures of positive
94 selection in arctic dwelling hare have also been identified in mitochondrial genes, including
95 *ATP8*, *CYTB* and *ND4* (Melo-Ferreira *et al.*, 2014). To date, analyses of the mitogenomes of
96 Old World camels has largely concerned diversity analyses of partial mtDNA sequences, which
97 indicates limited phylogeographic structuring among the dromedary camels in Asia and Africa
98 (Almathen et al., 2016, Alaqeely et al., 2021). A study by Mohandesan et al. (2017) analyzed
99 the full mitogenome of the three Old World camel species, revealing signals of gene-wide
100 purifying selection including positive selection in 18 sites or “codons” among the three species.
101 However, only a limited number of samples (≤ 10) were analysed for each species.

102

103 Here, we investigated the mtDNA protein-coding genes of dromedary camels (n = 42)
104 distributed along Africa, Arabian Peninsula and southwest Asia for signatures of selection at
105 gene-wide and site-specific levels. Publicly available full mitogenome sequences of camels
106 representing domestic Bactrian (n = 38), wild Bactrian (n = 29) and New World Lamini (n =
107 31) were also included for comparison.

108

109 **Materials and Methods**

110 **Whole genome sequence data from the Camelidae family**

111 Genomic DNA of seven dromedary camels, which included two from Kuwait and five from
112 Saudi Arabia, was extracted from 5ml blood samples using the DNeasy[®] Blood and Tissue kit
113 (Qiagen). Whole-genome sequencing of DNA was performed on the Illumina Hiseq 2000
114 platform using 150 bp paired-end libraries with 270 bp insert size. Sequencing of the Kuwaiti
115 samples was performed by the BGI in China, and at Macrogen in South Korea for the Saudi
116 samples (Table S1).

117

118 Whole-genome sequencing data for 102 samples was retrieved from the European Nucleotide
119 Archive. This included: 21 dromedary samples from the Arabian Peninsula (Bioproject
120 accession number: PRJEB47650) (Bahbahani and Almathen, 2022); four dromedaries from
121 Iran, 31 domestic Bactrian camels, and 19 wild Bactrian camels from Central Asia (Bioproject
122 accession number: PRJNA383081) (Ming et al., 2020); and 27 camel samples belong to the
123 Lamini tribe from South America: Llama (n = 7), Vicuna (n = 6), Guanaco (n = 7), and Alpaca
124 (n = 7) (Bioproject accession number: PRJNA612032) (Fan et al., 2020) (Table S2).

125

126

127

128 **Raw sequence read processing and full mtDNA sequence generation**

129 The raw sequencing reads were processed using Fastp v0.22 (Chen et al., 2018). This included
130 trimming sequencing adapters, and discarding reads if: (1) 50% or more of the read bases were
131 uncertain; (2) 40% or more of the read bases were low quality (base $Q_{\text{phred}} \leq 20$); (3) reads
132 length was shorter than 15 bases; or (4) read complexity was less than 30%. The remaining
133 high-quality reads of the dromedary, domestic Bactrian, wild Bactrian, and Lamini camels
134 were aligned to reference mtDNA sequences for their respective species: dromedary
135 (NC_009849.1); domestic Bactrian (NC_009628.2); wild Bactrian (NC_009629.2); and
136 Alpaca (NC_002504.1). Reads were aligned using the *bwa-mem* algorithm implemented in
137 Burrows-Wheeler Aligner (BWA) (Li and Durbin, 2010). The resulting alignments were
138 coordinate-sorted using the *SortSam* option, and PCR-duplicates were removed using the
139 *MarkDuplicates* and *REMOVE_DUPLICATES=true* options in the Picard tools v1.119
140 (<http://broadinstitute.github.io/picard/index.html>).

141

142 Single Nucleotide Polymorphisms (SNPs) were called using the *HaplotypeCaller* algorithm
143 implemented in Genome Analysis Toolkit (GATK) v4.1 (McKenna et al., 2010). Quality
144 control filtering criteria were applied on the per-sample variants using the *VariantFilteration*
145 algorithm in GATK. This included: (1) excluding variants with low quality by depth (QD) (QD
146 < 2); (2) excluding variants with root mean square of mapping quality for all reads of a site less
147 than 40 ($MQ < 40.0$); (3) excluding variants with base quality score less than 30 ($QUAL < 30$);
148 (4) excluding variants with high probability of allele-specific strand bias between forward and
149 reverse strand ($FS > 60$); (5) excluding variants with bias in mapping quality between the reads
150 supporting the reference and alternative alleles ($MQRankSum < -12.5$); and (6) excluding
151 variants with bias in the position of the alternative allele towards the ends of the reads
152 ($ReadPosRankSum < -8$). SNPs with depth of coverage ranging between two reads and three

153 standard deviation (SD) from the mean depth of coverage across samples were retained. Fasta
154 files of the full mtDNA sequences of each sample were generated using
155 *FastaAlternateReferenceMaker* algorithm in GATK v4.1. ([File S1](#))
156

157 **mtDNA diversity statistics and molecular phylogeny**

158 The generated mtDNA sequences were aligned with the full mtDNA sequences downloaded
159 from NCBI's Genbank using the *Clustal W* algorithm implemented in MEGA v10 (Kumar et
160 al., 2018). The NCBI sequences included: dromedary camels (n = 10), domestic Bactrian
161 camels (n = 7), wild Bactrian camels (n = 10), Llama (n = 1), Vicuna (n = 1), Guanaco (n = 1),
162 and Alpaca (n = 1) (Table S3). The number of variable sites, parsimony-informative and
163 singletons, were calculated with DnaSP v6 (Librado and Rozas, 2009) for the 140 mtDNA
164 sequences. Haplotype (H_d) and nucleotide (π) diversities were calculated for dromedaries,
165 domestic Bactrian, wild Bactrian, and Lamini samples, separately, using DnaSP. Tajima's D
166 neutrality index and Fu and Li's F statistics were also calculated with DnaSP to determine
167 historical population expansions and contractions of the different Camelidae species.

168
169 We identified the best-fitting DNA evolution model according to the Bayesian Information
170 Criterion (BIC) implemented in MEGA software which was the GTR (General Time
171 Reversible) model with gamma correction parameter = 0.56. A neighbor-joining distance tree
172 was constructed using MEGA, employing 1000 bootstrap replications to assess the mtDNA
173 relationship among the camel species. ggtree package (Yu, 2020) for R software version 4.2
174 (R Core Team, 2022) was used to plot the tree.

175

176

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178 **Gene-wide signatures of selection analyses**

179 Signatures of selection were investigated on the 13 mtDNA protein-coding genes at a gene-
180 wide level using three approaches: McDonald-Kreitman (MK); neutrality index (NI); and ω
181 ratio tests using DnaSP. The MK test measures the ratio of nonsynonymous (P_n) to
182 synonymous (P_s) substitutions within species as compared to the ratio of nonsynonymous (K_n)
183 to synonymous (K_s) substitutions between species. A $K_n/K_s > P_n/P_s$ indicates a signal of
184 positive selection, while a $K_n/K_s < P_n/P_s$ and $K_n/K_s = P_n/P_s$ are signals of purifying selection
185 and neutrality, respectively (McDonald and Kreitman, 1991). NI calculates the ratio of
186 polymorphic sites within species to the fixed sites between species at nonsynonymous and
187 synonymous substitutions ($NI = (P_n * K_s) / (K_n * P_s)$). Whereby $NI = 1$ equates no selection, NI
188 > 1 indicates purifying selection, while $NI < 1$ indicates positive selection. The ω ratio is the
189 ratio of non-synonymous substitutions per non-synonymous sites (D_n) to the synonymous
190 substitutions per synonymous sites (D_s). A D_n/D_s ratio > 1 indicates positive selection, while
191 a D_n/D_s ratio < 1 indicates purifying selection, and $D_n/D_s = 1$ indicates no selection. Two-
192 tailed Fisher's exact test P -values were computed to define statistically significant signals for
193 the MK and NI tests. Each of these analyses were conducted at three levels: (1) comparing
194 dromedary camels ($n = 42$) to domestic and wild Bactrian ($n = 67$); (2) comparing the
195 dromedary camels ($n = 42$) to the New World Lamini camels ($n = 31$); and (3) comparing Old
196 World camels (dromedary and Bactrian) ($n = 109$) to the New World Lamini camels ($n = 31$).

197

198 **Site-specific signatures of selection analyses**

199 Signatures of selection at the site-specific level were identified along the 13 mtDNA protein-
200 coding genes using the site models implemented in the CODEML package of Phylogenetic
201 Analysis by Maximum Likelihood (PAML) v4.9 (Yang, 1997, Yang, 2007, Yang et al., 2000).
202 Several different models implemented in the CODEML package were evaluated. The one ratio

203 (M0) model allows a single ω for all codons. The nearly neutral model (M1a) assumes two
204 classes of codons: one with $0 \leq \omega_0 < 1$ and a proportion of codons p_0 , while the second class
205 assumes $\omega_1 = 1$ and a proportion of codons $p_1 = 1 - p_0$. Model M2a (positive selection) is an
206 extension of the M1a model with a third class that allows for $\omega_2 > 1$ and a proportion of codons
207 $p_2 = 1 - p_1 - p_0$. Model M3 (discrete) uses by default three discrete classes to model the
208 heterogeneity of ω between codons. The M7 and M8 models assume a beta distribution of ω
209 over codons with two beta function parameters (p and q). The M7 model does not allow for
210 codons under positive selection by constraining ω to be in the interval $(0, 1)$. In contrast, the
211 M8 model allows for codons with $\omega_1 > 1$ with a proportion of codons p_1 . After calculating the
212 log likelihood value (L) of each model fitted to our data, likelihood ratio tests (LRTs) were
213 conducted between the M1a–M2a and M7–M8 models to test for positive selection, and
214 between the M0–M3 models to test for variable selection pressures among codons. The statistic
215 for each LRT is defined as twice the log likelihood difference between two models ($2\Delta L$). This
216 statistic was compared to a χ^2 distribution with a degree of freedom (d.f.) equal to the difference
217 in the number of parameters between the two models (d.f. = 2 for M1a–M2a and M7–M8, and
218 d.f. = 4 for M0–M3). A Bayes Empirical Bayes (BEB) approach was used to identify codons
219 under positive selection. Sites with BEB posterior probability > 0.5 were considered to be under
220 positive selection, with a value > 0.95 considered as extreme selection (Yang et al., 2005). The
221 site model analyses were conducted at the three different levels described previously (1:
222 dromedary with domestic and wild Bactrian; 2: dromedary with New World Lamini; 3: Old
223 World camels with New World Lamini).

224

225 Branch-site models of positive selection, implemented in the CODEML package (Zhang et al.,
226 2005), were also tested on the mtDNA protein-coding genes to detect codons under positive
227 selection in a specific lineage, the “foreground lineage”, but which remained neutral or under

228 purifying selection in the other lineages, the “background lineages”. The dromedary camels
229 were set as the foreground lineage, while the other camel populations were set as background
230 lineages. On the branch-site models, Model A assumes positive selection by defining four
231 classes of codons. Class 0 includes codons with $0 \leq \omega_0 < 1$ throughout the tree. Class 1 includes
232 codons with $\omega_1 = 1$ throughout the tree. Class 2a includes codons with $0 \leq \omega_0 < 1$ in the
233 background lineages, and $\omega_2 \geq 1$ in the foreground lineage. Class 2b includes codons with $\omega_1 =$
234 1 in the background lineages, and $\omega_2 \geq 1$ in the foreground lineages. The null model, which
235 does not assume positive selection, is the same as Model A, but fixes $\omega_2 = 1$. As in the site
236 models, the log likelihood value (L) was calculated for each model fitted to our data, and LRTs
237 were conducted between Model A and the null model to test for positive selection. The
238 calculated log likelihood statistic for the LRT was compared to a χ^2 distribution (d.f. = 1). A
239 BEB approach was used to identify codons under positive selection as in the site models.

240

241 Codons under positive selection were also determined using TreeSAAP v3.2 (Woolley et al.,
242 2003). TreeSAAP identifies selective influences based on 31 structural and biochemical
243 properties. It accounts for the impact magnitude of amino acid changes on physiochemical
244 properties of the protein, and tests if the observed degree of amino acid substitution deviates
245 from the neutral expectation. If the change in magnitude ≥ 6 and the goodness-of-fit test P -
246 value < 0.001 , it is considered a strong indication of positive selection in the physiochemical
247 property tested. For estimating the significance of changes, the software outputs two main
248 values: category value and statistical z-score value (equivalent to P -value). The category values
249 were numbered from (1 to 8); 1 is the most conservative amino acid category and 8 is the most
250 radical value. Category values ≥ 6 and z-score value ≥ 3.09 are considered as significant signals
251 of positive selection. A phylogenetic tree for each mitochondrial protein-coding gene in the

252 different species of the Camelidae family was constructed using MEGA and used as an input
253 for TreeSAAP.

254

255 **Results**

256 **Camelidae full mtDNA diversity statistics and molecular phylogeny**

257 The alignment of the 140 full mtDNA sequences returned 3,520 variable sites, of which 191
258 were singletons and 3,329 were parsimony-informative. Higher nucleotide and haplotype
259 diversities were calculated for Lamini samples ($H_d = 0.998$ and $\pi = 0.0254$) than for those of
260 the Camelini tribe (Table 1). Among the Camelini tribe, relatively lower nucleotide diversity
261 and higher haplotype diversity values were observed in dromedaries ($\pi = 0.00109$ and $H_d =$
262 0.997) than in domestic Bactrian ($\pi = 0.00124$ and $H_d = 0.989$) and wild Bactrian camels ($\pi =$
263 0.00115 and $H_d = 0.781$). Negative Tajima's D and Fu and Li's F statistics were calculated for
264 dromedary camels (Tajima's D = -2.24, Fu and Li's F = -2.87) and domestic Bactrian camels
265 (Tajima's D = -1.223, Fu and Li's F = -2.42), which deviated significantly from neutrality in
266 dromedaries. Positive Tajima's D and Fu and Li's F statistics were calculated for wild Bactrian
267 camels (Tajima's D = 2.655, Fu and Li's F = 2.067) and Lamini camels (Tajima's D = 0.167,
268 Fu and Li's F = 0.524), which deviated significantly from neutrality in the wild Bactrian
269 camels.

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276 **Table 1: Haplotype and nucleotide diversities, Tajima's D and Fu and Li's F statistics for**
 277 **dromedary, domestic Bactrian, wild Bactrian and Lamini camels included in the study.**

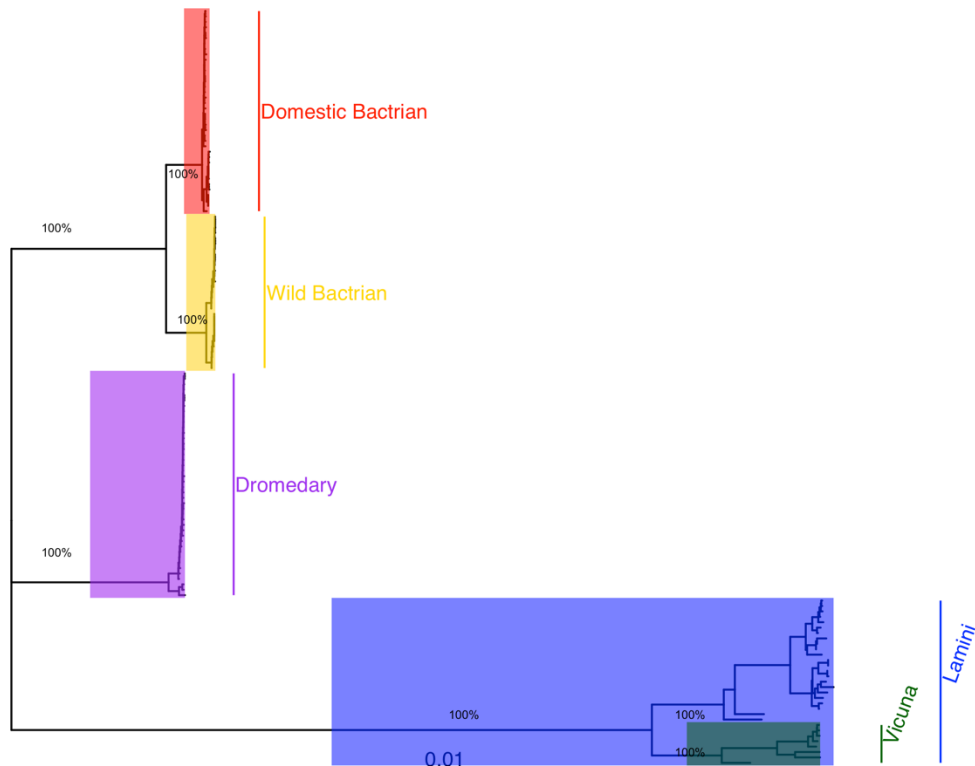
	<i>C. dromedarius</i>	<i>C. bactrianus</i>	<i>C. ferus</i>	<i>Lamini</i>
Haplotype diversity (Hd)	0.997	0.989	0.781	0.998
Nucleotide diversity (π)	0.00109	0.00124	0.00124	0.0254
Tajima's D	-2.24*	-1.22	2.655*	0.167
Fu and Li's F	-2.87*	-2.42	2.067*	0.524

278 *Significant values with P -value < 0.05

279

280 The neighbour-joining phylogenetic tree revealed species-wise mtDNA distinction among the
 281 Camelidae family, separate dromedary, Bactrian and Lamini camels from each other. Within
 282 the Lamini tribe, the Vicuna camels were differentiated from the other New World camels,
 283 while among the two-humped Old World camels we observed differentiation between the
 284 domestic and wild Bactrian camels. No geographic-genetic distinction was observed among
 285 the different dromedary camel populations (Fig. 1).

286



287

288 **Fig. 1. Neighbor joining phylogenetic tree of the Camelidae family based on the full mitogenome.**

289

290 **Gene-wide level signatures of selection**

291 Based on the McDonald-Kreitman (MK) test and Neutrality index (NI), significant signals of
 292 purifying selection were obtained in six genes; *ND1*, *COX1*, *ND4*, *ND5*, *ND6*, and *CYTB*, upon
 293 comparing dromedary with domestic and wild Bactrian camels. While non-significant signals
 294 of purifying and positive selection were found across the remaining genes (Table 2 and Table
 295 S4). Comparing dromedary with Lamini camels revealed significant signals of positive
 296 selection in *ND5*, *ATP6* and *COX3* (Table 2 and Table S4). Comparing Old World camels with
 297 Lamini camels showed significant signals of positive selection in *ND1*, *ND4L*, *ND5*, *ATP6* and
 298 *COX3* (Table 2 and Table S4).

299

300

301 **Table 2: Types of selection signals defined on the mitochondrial protein-coding genes based on**
 302 **McDonald-Kreitman test and Neutrality index on the three different analyses levels.**

	Dromedary with domestic and wild Bactrian		Dromedary with Lamini		Old World with Lamini	
Gene	Type of selection	<i>P</i> -value ¹	Type of selection	<i>P</i> - value ¹	Type of selection	<i>P</i> - value ¹
ND1	Purifying	0.019*	Positive	0.061	Positive	0.022*
ND2	Positive	0.471	Positive	0.531	Positive	0.748
COX1	Purifying	0.044*	Positive	1	Positive	0.390
COX2	Positive	1	Positive	0.327	Positive	0.070
ATP8	Purifying	1	Positive	0.158	Positive	0.603
ATP6	Positive	0.706	Positive	0.017*	Positive	0.041*
COX3	Purifying	0.302	Positive	0.015*	Positive	0.02*
ND3	Purifying	0.637	Positive	0.783	Positive	0.619
ND4L	Purifying	0.492	Positive	0.166	Positive	0.012*
ND4	Purifying	0.043*	Positive	0.577	Positive	0.295
ND5	Purifying	0.015*	Positive	0.038*	Positive	0.031*
ND6	Purifying	0.009**	Purifying	0.101	Positive	1
CYTB	Purifying	0.0003***	Purifying	0.532	Purifying	0.378

303 ¹The *P*-values are the significant values of the two-tailed fisher's exact test. *values with asterisk*
 304 *are significant with significant level of *P<0.05, **P<0.01, ***P<0.001.*
 305

306 The ratio of non-synonymous substitutions per non-synonymous sites (Dn) to the synonymous
 307 substitutions per synonymous sites (Ds) (ω ratio) was less than one, indicating purifying
 308 selection, on all the mtDNA protein-coding genes at the different analyses. Among the thirteen
 309 genes, *ATP8* showed the highest ω ratio, whilst, *COX1* and *COX2* showed the lowest ω ratios
 310 (Fig. S1 and Table S5).

311

312

313 **Site-specific signatures of selection**

314 The LRT results from the M0-M3 models, which tests for variable selection pressures among
 315 codons, returned significant signals of selection both for dromedary with Lamini, and Old
 316 World with New World Lamini camels, in which a proportion of these sites had evolved under
 317 positive selection with ω value larger than 1 (Table S6). Twenty-seven amino acid substitutions
 318 in ten mtDNA protein-coding genes were identified to be under positive selection by Bayes
 319 Empirical Bayes (BEB) analyses. This included seven codons among dromedary with domestic
 320 and wild Bactrian, 21 codons among dromedary with Lamini, and 24 codons among Old World
 321 camels with New World Lamini camels (Table S7). Two of these codons are in genes for which
 322 positive selection models (M8 and M2a) fit the data significantly better than neutral models
 323 (Table 3). These are *COX2* (dromedary with Lamini, M8 and M2a) and *ND6* (Old World with
 324 New World Lamini camels, M8) (Table S7). Of particular note is codon 138 in *COX2* which,
 325 among dromedary with Lamini and Old World camels with New World Lamini, is under
 326 extreme selection based on the BEB posterior probability value.

327

328 **Table 3: Positively selected codons identified by the site-models based on the posterior probability**
 329 **of the Bayes Empirical Bayes (BEB).**

Gene	Codon	Amino acids in Dromedary	Amino acids in Bactrian	Amino acids in Lamini	Level of analysis
COX2	138	V	V	W	Dromedary with Lamini (BEB=0.996) (<i>P</i> -value <0.05)
					Old World with New World (BEB=0.996) (<i>P</i> -value <0.05)
ND6	102	I	S,A	L	Old World with New World (BEB=0.921) (<i>P</i> -value <0.01)

330 *P*-values are the significant values of the M8-M7 and M1a-M2a Likelihood ratio tests.

331

332

333 Based on the BEB posterior probability values of the branch-site model with dromedary camels
 334 as the foreground lineage, a total of seven codons within three genes were found to be under

335 positive selection. These include codons 34 and 66 in *ATP8*, codons 1 and 51 in *ATP6*, and
 336 codons 101, 116 and 133 in *ND6*. The three codons identified in *ND6* exhibit intra-species
 337 variation among the dromedaries. None of the genes carrying these codons have a significantly
 338 better fit to the positive selection model (Model A) than the neutral null model (Table 4 &
 339 Table S8).

340

341 **Table 4: Sites identified by branch-site models, based on Bayes Empirical Bayes (BEB), as**
 342 **potentially under selection.**

Gene	Codon	Amino acid in Foreground lineage	Amino acid in Background lineage	BEB Posterior probability	P-value
ATP8	34*	P	L, Y	0.795	$P > 0.05$
	66	L	Q	0.542	
ATP6	1*	V	M	0.513	$P > 0.05$
	51	Q	R	0.52	
ND6	101	I (V in Omani and Kenya samples)	I, M	0.833	$P > 0.05$
	116	I (V in Omani and Kenya samples)	I	0.909	
	133	S (F in Omani and Kenya samples)	S	0.901	

P-values are the significant values of the positive selection alternative and neutral null models likelihood ratio test of the branch-site models.

* Sites identified in Mohandesan et al., (2017) to be under positive selection.

343

344 We next sought to investigate the magnitude of the impact of amino acid replacement on the
345 physiochemical properties of the proteins. Using TreeSAAP we identified 89 codons in nine
346 genes to be under positive selection. These included eight codons in *ND1*, eleven codons in
347 *ND2*, two codons in *ATP6*, five codons in *COX3*, seven codons in *ND3*, seven codons in *ND4*,
348 twenty-seven codons in *ND5*, ten codons in *ND6*, and twelve codons in *CYTB* genes. Of these
349 codons, 71 were found to be under positive selection between dromedaries and Lamini.
350 Positive selection was also observed in 26 and 25 codons between dromedaries and domestic
351 Bactrian, and dromedaries and wild Bactrian camels, respectively. In particular, two amino
352 acid replacements (codon 9 in *ND4* and codon 101 in *ND6*) were found to be polymorphic
353 among dromedaries, differentiating the two Omani camels and a single Kenyan dromedary
354 from the other dromedaries. Out of the 31 physiochemical properties considered by TreeSAAP,
355 we identified significant changes in six properties: the equilibrium constant (ionization COOH)
356 (pK'); power to be at the middle of alpha helix (α_m); long range non-bonded energy (E_l); solvent
357 accessible reduction ratio (R_a); surrounding hydrophobicity (H_p); and alpha helical tendencies
358 (P_α) (Table S9).

359

360 **Discussion**

361 By analyzing the full mitogenomes of the Camelidae family species we have identified
362 signatures of positive adaptive divergence between Camelini and Lamini tribes. Within
363 Camelini species, signals of gene-wide purifying selection and site-specific positive selection
364 were also observed that might be associated with their adaptation to the different environmental
365 conditions they are populating.

366

367 Signals of past population expansion and/or positive selection in dromedary camels can be
368 observed based on the significant negative Tajima's D and F_i and Lu's F statistics. These
369 signals might relate to historical population growth and the continental-wise distribution of
370 dromedaries throughout Asia and Africa. While significant positive values in wild Bactrian
371 and non-significant negative values in domestic Bactrian camels potentially relates to the
372 restricted distribution of wild Bactrian camels, being found only in a protected area in the
373 Mongolian Gobi Desert, and the limited distribution of the domestic Bactrian camels in Central
374 Asia (Burger et al., 2019). These results are in agreement with those reported in Camelini
375 species by Mohandesan et al. (2017). Positive Tajima's D and F_u and Li's F statistics were
376 observed for New World Lamini species, which might also be related to the confined
377 distribution of these camels in South America (Fan et al., 2020).

378

379 The phylogenetic analysis revealed inter-species genetic distinction between Camelini and
380 Lamini tribes. Within the Camelini tribe, dromedary camels separated from Bactrian camels,
381 which also show genetic distinction between domestic and the wild-type. However, neither
382 continental-wise nor population-wise genetic distinction was observed among the different
383 dromedary camel populations included in the study. The observed inter-species genetic
384 distinction between the Camelini and Lamini tribes is likely resulted from their divergence
385 about 16.3 Mya, which was followed by the later migration of the Camelini species to Eurasia,
386 about 6.5 – 7.5 Mya, and the Lamini tribe to South America about 3 Mya (Burger et al., 2019).
387 The later divergence between the dromedary and Bactrian camels, about 4.4 Mya, and the
388 different geographical distribution of these two species may explain the mitochondrial genetic
389 distinction observed between them. Such separation has also been observed previously at
390 autosomal level using whole genome sequence data (Ming et al., 2020, Bahbahani and
391 Almathen, 2022). The genetic distinction between domestic and wild Bactrian camels can

392 likely be attributed to the 1.1 Mya divergence between them (Burger et al., 2019). Genetic
393 separation is also observed between the wild vicuna species and the other Lamini species, with
394 *Lama* and *Vicugna* diverging around 2-3 Mya (Fan et al., 2020).

395

396 The lack of genetic distinction among the different dromedary camel populations may be due
397 to the historical purpose of this well-adapted species in transporting goods and people between
398 Asia and Africa, and throughout the Arabian Peninsula. Dromedary camels were initially used
399 in transportation along the “incense road” connecting the southern to the northern parts of
400 Arabian Peninsula. Dromedaries were used to transport goods between Africa and Asia during
401 the 1st millennium Before Common era (BC) via the Islands of Socotra. In parallel to
402 transporting goods, dromedaries were used to transfer people from Africa and different parts
403 of the Arabian Peninsula to Makkah during the annual Pilgrimage (Wilson, 1998). All of these
404 practices were associated with high interbreeding and gene flow between the dromedary camels
405 along the region, which have also been observed using whole genome sequence data
406 (Bahbahani and Almathen, 2022). Current breeding practices of local camel owners, which
407 rely on random mating between camels, further enhance gene flow between dromedary
408 populations.

409

410 Although the ω ratios indicate signals of purifying selection on all mtDNA protein-coding
411 genes at the varying levels of analysis, significant signals of positive selection were identified
412 on *ND1*, *ND4L*, *ND5*, *ATP6* and *COX3* upon comparing Old World camels with New World
413 Lamini. This indicates adaptive divergence between these two tribes, which might be attributed
414 to the different environmental conditions they inhabit. The various species of Camelini
415 populate the desert and mountainous areas of Asia and Africa, adapting to arid and semi-arid

416 conditions including tolerance to high temperature, and scarcity of food and water. In contrast,
417 Lamini species inhabit the high altitudes of south America, reaching more than 7000 m, and
418 thus face environmental challenges characterized with high altitude such as hypoxia (Beall,
419 2007, Wu et al., 2014). These environmental challenges present significant pressure on mtDNA
420 genes, for example to adapt to low oxygen concentration in the environment given the role of
421 mitochondria in oxygen consumption and production of ATP molecules. Upon comparing the
422 two domestic Old World camel species (dromedary and Bactrian), significant signals of
423 purifying selection were defined in six mtDNA protein-coding genes: *ND1*, *COX1*, *ND4*, *ND5*,
424 *ND6*, and *CYTB*, among which *ND1*, *ND5* and *ND6* have previously been identified to be under
425 purifying selection (Mohandesan et al., 2017). Here, purifying selection would act to eliminate
426 deleterious alleles that negatively affect the functionality of mitochondria.

427

428 The MK, NI and ω ratio analyses are considered conservative approaches to detect signatures
429 of selection given that mutations affecting specific codons in a gene that can be masked by
430 gene-wide purifying selection pressures (Crandall et al., 1999). The site-model BEB analysis
431 identified a total of 21 and 24 codons to be under positive selection, potentially associated with
432 adaptive divergence between dromedaries or Old World camels and New World camel species,
433 respectively. Seven codons under positive selection were identified when comparing
434 dromedary with domestic and wild Bactrian, indicative of adaptive divergence between these
435 two Old World camel species, potentially related to the different environmental niches they
436 occupy. Interestingly, seven codons identified by the branch-site model BEB analysis were
437 found to be under positive selection in dromedaries, indicating that these codons might be
438 associated with adaptation to desert environment in dromedaries. Among these sites, codon 1
439 in *ATP6* was found to be under positive selection in dromedaries using the branch-site models
440 by Mohandesan et al. (2017). Although all of these codons were defined to be under positive

441 selection based on BEB posterior probability, none of the genes fit the signature of selection
442 alternative model (Model A) better than the neutral null model, which was also observed by
443 Mohandesan et al. (2017).

444

445 Several codon substitutions were found to be under positive selection by the TreeSAAP
446 software between dromedary and Lamini, and between dromedary and Bactrain. This approach
447 is considered more sensitive than ω ratio-based approaches since a single biochemically
448 adaptive physiochemical change can be too weak to be identified by gene-based approaches
449 where conservation scores are high (Hughes, 2007, McClellan, 2013). We observed magnitude
450 impacts on six physiochemical properties of proteins: the equilibrium constant (ionization
451 COOH), power to be at the middle of alpha helix, long range non-bonded energy, solvent
452 accessible reduction ratio, surrounding hydrophobicity, and alpha helical tendencies. Changes
453 in these properties can impact protein functions in several ways. An increase in alpha helical
454 tendency (P_α) may lead to long and rigid alpha helix, which renders interactions with amino
455 acid motifs more difficult. Decreasing this property makes the amino acid more flexible to an
456 open alpha helix, increasing the likelihood of amino acid interactions within a protein (Burkin
457 et al., 2000). Changes of the equilibrium constant (ionization COOH) (pK') and solvent
458 accessible reduction ratio (R_a) impacts amino acid water solubility, rendering it more hydrophilic
459 or hydrophobic, and is considered an important change when dehydrated, increasing longevity
460 by reducing reactive oxygen species (ROS) production (Beckstead et al., 2009). Increasing the
461 surrounding hydrophobicity (H_p) makes the surrounding area of the amino acid site
462 hydrophobic (Mohandesan et al., 2017). While changes in long-range non-bonded energy (E_i)
463 impacts amino acid interactions and may contribute of protein structure change.

464

465 The signals of selections characterized here were investigated based on gene-wide and site-
466 specific analyses. These analyses have taken the advantage of including large number of
467 samples in this study from the different species of the Camelidae family. These analyses can
468 be further improved upon including camel samples from more diverse geographical locations
469 with specific phenotypic traits, such as racing performance and milk production. Such
470 phenotypes may explain the polymorphic codons within the dromedary lineage, defined by the
471 branch-site models and TreeSAAP, in *ND6* and *ND4* genes.

472

473 **Conclusion**

474 In this study signatures of selection were investigated in the mitochondrial protein-coding
475 genes of different dromedary camel populations by comparing to Bactrian and Lamini camels,
476 revealing signals of adaptive divergence between Old World and New World camel species.
477 Gene-level purifying selection was identified among the Old World camels, and signals of
478 positive selection were identified at specific codons in Old World camels. These codons
479 provide a link between the mtDNA evolution in such camel species and their adaptation to the
480 diverse environmental conditions they populate.

481

482

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486

487

488 **Data Availability**

489 We have deposited the whole genome sequence data underlying these analyses at the European
490 Nucleotide Archive (Bioproject accession number: PRJEB53955).

491

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494 Arabian Peninsula.

495

496 **Conflict of interest**

497 The authors declare not competing of interest.

498

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Supporting information

610 **Fig. S1. The ω ratios for the 13 mtDNA protein-coding genes at three different analyses**
611 **levels.** 1) Dromedary camels (n=42) compared to Bactrian camels (n=67), 2) Dromedary
612 camels (n=42) compared to Lamini (n=31), 3) Dromedary and Bactrian camels (n=109)
613 compared to Lamini (n=31).

614

615 **File S1. Detailed script for mapping raw reads, calling variants and retrieving mtDNA sequences**

616

617 **Table S1. The seven dromedary samples with whole genome sequences from this study.**

618

619 **Table S2. Whole genome sequence data of the Camelidae family species included in the**
620 **study from European Nucleotide Archive.**

621

622 **Table S3. Full mtDNA sequences of Camelidae family species included in the study from**
623 **NCBI Genbank.**

624

625 **Table S4. McDonald-Kreitman (MK) test and Neutrality index (NI) values on the three**
626 **different analyses levels for the mtDNA protein-coding genes.** The values with asterisk are
627 significant with Fisher exact P-value * <0.05 , ** <0.01 , *** <0.001 .

628

629 **Table S5. The ω ratio (Dn/Ds) analyses for the 13 mtDNA protein-coding genes at three**
630 **different analyses levels.** A) Dromedary camels (n=42) compared to Bactrian camels (n=67),
631 B) Dromedary camels (n=42) compared to Lamini (n=31), C) Dromedary and Bactrian camels
632 (n=109) compared to Lamini (n=31).

633

634 **Table S6. Likelihood values and parameter estimates of the different site models**
635 **implemented in CODEML package for the mtDNA protein-coding genes in all three data**
636 **analysis levels.** n.s = non-significant.

637

638 **Table S7. Potential positively selected sites identified using the site-models based on Bayes**
639 **Empirical Bayes (BEB) in genes at the three different data analyses levels.** P-values are
640 the significant values of the M8-M7 Likelihood ratio test. n.s = non-significant.

641

642 **Table S8. Likelihood values and parameter estimates of the Branch-site models**
643 **implemented in CODEML package for the mtDNA protein-coding genes upon specifying**
644 **dromedary camels as foreground.**

645

646 **Table S9. TreeSAAP positive selection sites based on the impact on amino acids**
647 **physiochemical properties changes.** Pairwise comparisons between dromedary and Bactrian,
648 dromedary and wild Bactrian and dromedary and Lamini camels. The equilibrium constant
649 (ionization COOH) (pK'), power to be at the middle of alpha helix (αm), long range non-
650 bonded energy ($E1$), solvent accessible reduction ratio (R_a), surrounding hydrophobicity (H_p),
651 and alpha helical tendencies ($P\alpha$).

652