



# Impact of thermal processing on dietary flavonoids

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Flavonoids are widely distributed in natural products and foods as a class of polyphenols. They processed diverse bioactivities, including anti-inflammation activity, antiaging activity, and antioxidant activity. The foods rich in flavonoids are usually consumed after thermal processing. However, flavonoids are commonly vulnerable under thermal processing, and it could cause various influences on their stability and bioactivities. Therefore, in this review, the effects of thermal processing on thermal stability and bioactivities of dietary flavonoids from different food sources were first summarized. The strategies to improve thermal stability of dietary flavonoids were then discussed. Noticeably, the effect of some of the promising thermal technologies on dietary flavonoids was also clarified preliminarily in the current review. The promising thermal technologies may be an alternative to conventional thermal processing technologies.

## Addresses

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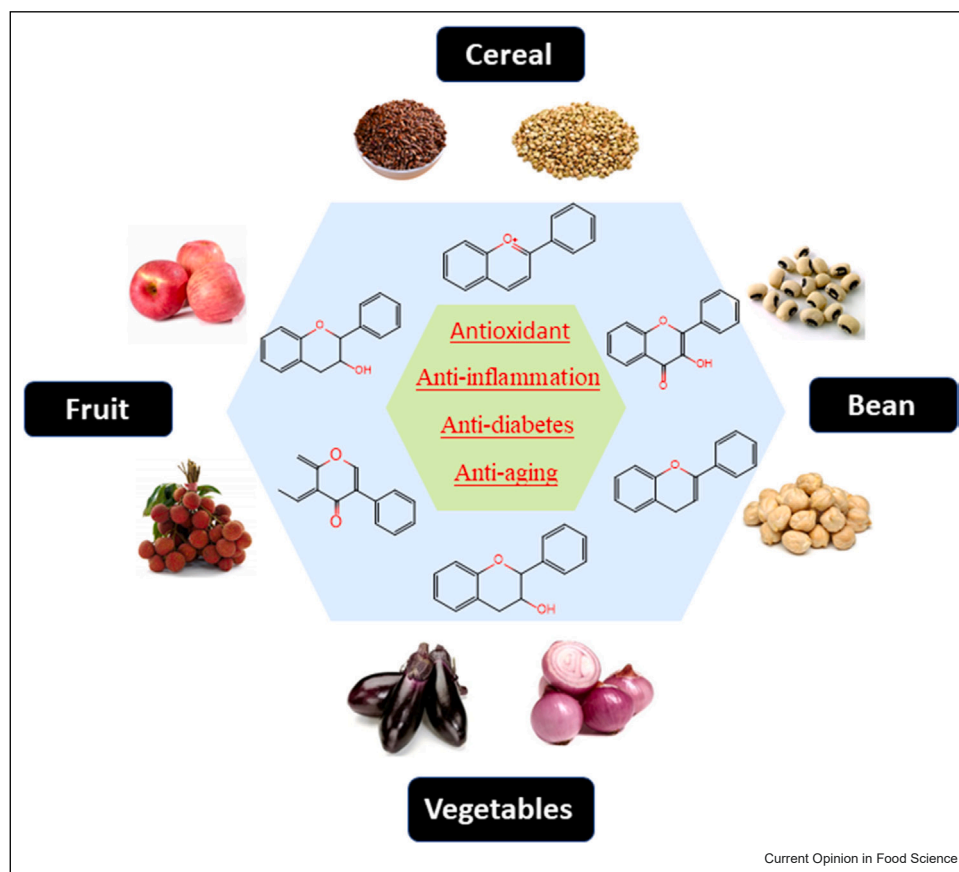
## Introduction

Flavonoids are a class of polyphenols, with the typical chemical structures containing A, B, C three (C6–C3–C6) rings, including flavanols, flavones, isoflavones, flavanones, flavan-3-ols, and anthocyanidins [1–4]. Flavonoids mainly form fruits, vegetables, cereals and beans, have profitable advantages on human health, including anti-inflammation activity, antiaging activity, antioxidant activity, and antidiabetes activity (Figure 1) [5–10].

To meet the safety and shelf-life-stable requirements, the processing is inevitable for foods before delivering to consumers. Flavonoids are commonly vulnerable under different processing conditions, especially thermal processing. Thermal processing is a kind of widely used processing technique in the food industry and household cooking. The application of thermal processing has been favored by the important technological developments experienced over the last few years. During thermal processing, undesirable microorganisms and spoilage enzymes were inactivated, and gastrointestinal digestion of food generally enhanced [11, 12]. On the other hand, the quality of food (color, taste, nutrients, and bioactive compounds) is usually impaired by long-time heating, resulting in reduced bioavailability and organoleptic property destruction [13].

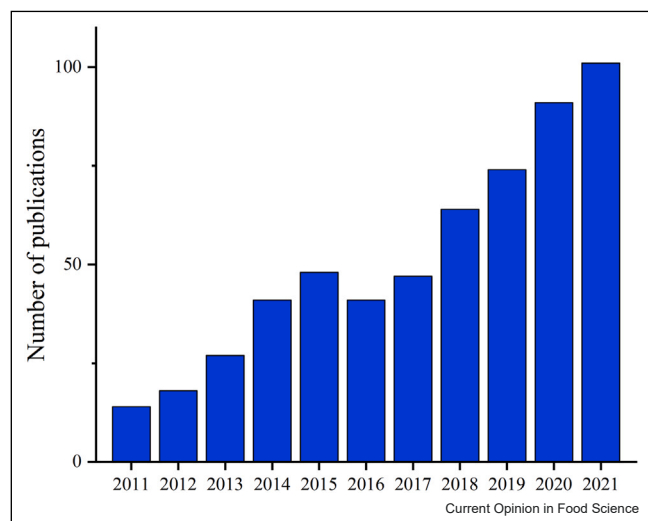
Usually, the dietary flavonoids are vulnerable under thermal processing, and their bioactivity usually impaired after thermal processing, due to their lower stability. Therefore, researches related to the impact of thermal processing on dietary flavonoids were explored to clarify the thermal stability of flavonoids and bioactivity of dietary flavonoids after thermal processing, and establish strategies to improve thermal stability of dietary flavonoids. The factors, such as thermal processing techniques, heating temperature, heating time, and food matrices, influenced thermal stability of dietary flavonoids variously [14–16]. Some strategies or new techniques were applied to improve thermal stability of dietary flavonoids, which is becoming a research hotspot [17–19]. The number of publications (indexed by Pubmed) on “thermal processing and flavonoid” sharply increased since 2011 (Figure 2). Therefore, this review will mainly focus on the impacts of thermal processing on dietary flavonoids.

Figure 1



The main resources, chemistry structures, and bioactivities of dietary flavonoids.

Figure 2



The number of publications on "thermal processing and flavonoid" indexed by Pubmed database since 2011.

### Influence factor of thermal processing on the thermal stability of dietary flavonoids

The intake of dietary flavonoids is conducive to reduce the risk of diseases, partly attributed to intake of dietary flavonoids therein. The roasting, barking, steaming, frying, and grilling are common thermal processing techniques in food industries (Table 1). However, the thermal stability of dietary flavonoids was easily influenced by different factors, such as thermal processing techniques, heating temperature, heating time, and food matrices. The impacts of thermal processing on flavonoid content in various food matrices under different conditions are shown in Table 1.

The thermal stability of dietary flavonoids under different thermal conditions was distinct. For example, only frying (170 °C for 10 min) could increase total flavonoid content (TFC) of dark-purple eggplant (*Solanum melongena*), on the contrary, barking (180 °C for 30 min), boiling (100 °C for 20 min), and grilling (120 °C for 10 min) decreased its TFC [20] (Table 1). Contrary to eggplant, barking (180 °C for 30 min), frying (140 °C for

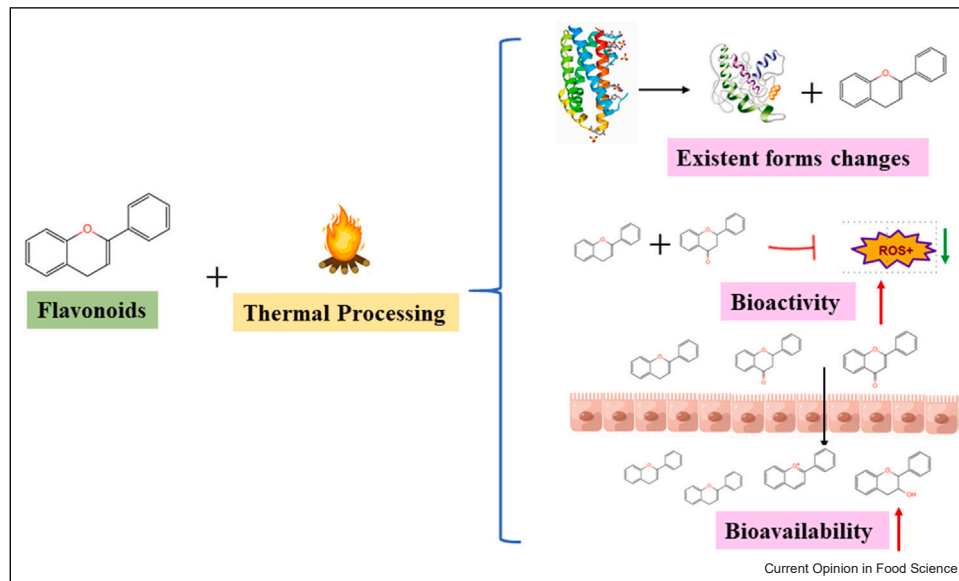
**Table 1**  
**Impacts of thermal processing on flavonoid content in various foods.**

| Order | Food sources  | Flavonoids   | Processing method   | Contents of flavonoids   | Reference                                  |
|-------|---|--|---|--|--|
| 1     | Eggplant  | TFC  | Baking (180 °C for 30 min)<br>Boiling (100 °C for 20 min)<br>Frying (170 °C for 10 min)<br>Grilling (120 °C for 10 min) | TFC <sup>a</sup> ↓<br>TFC ↓<br>TFC ↑<br>TFC ↓  | Martini et al. [20]                        |
| 2     | Onions  | TFC  | Baking (180 °C for 30 min)<br>Boiling (100 °C for 30 min)<br>Frying (140 °C for 8 min)<br>Grilling (110 °C for 15 min)  | TFC ↑<br>TFC ↓<br>TFC ↑<br>TFC ↓   | Cattivelli et al. [16]                     |
| 3     | Mulberry juice  | TFC  | Heating by water bath (25, 35, 45, 60, and 100 °C for 60 min)   | TFC ↓ in 15–45 °C treatment, TFC ↑ in 45–100 °C treatment  | Li et al. [14]                             |
| 4     | Blueberry ( <i>Vaccinium</i> spp.) puree  | TAC  | Heating (at 40, 50, 60, 70, 80, 90, and 100 °C for 20 min)  | TAC <sup>b</sup> under 40–80 °C treatment, TAC ↑ under 90 °C treatment, and TAC ↓ under 100 °C treatment   | Zhang et al. [8,21]                        |
| 5     | Teff ( <i>Eragrostis tef</i> )  | TFC  | Roasting (180 °C ± 20 °C for 5–10 min)  | TFC ↑ in roasting for 7.5 min, TFC ↓ in roasting for 10 min  | Kataria, Sharma, & Dar [22]                |
| 6     | Buckwheat bran  | TFC, rutin, isoquercetin, and quercetin            | Baking (180 °C for 30 min)  | TFC, isoquercetin, and quercetin ↓ rutin   | Ge & Wang [23]                             |
| 7     | Buckwheat flour   | TFC, rutin, isoquercetin, and quercetin            | Baking (180 °C for 30 min)  | TFC, rutin, isoquercetin; quercetin ↓  |  |
| 8     | Sesame (ganzhi9, ganzhi17, j9014, ezhi7, and luozhi18)<br>Sesame (Liaozhi8)     | TFC  | Roasting (240 °C for 20 min)  | TFC  | Chen et al. [24]                           |
| 9     | Thai rice bran (Kiaw Ngu and Leum Pua)  | Rutin, myricetin, quercetin-3-glucuronide, and TFC | Water-bath heat treatment (100 °C for 15 min)   | TFC ↓<br>Rutin, myricetin, TFC ↓, and quercetin-3-glucuronide ↑<br>Rutin ↓ myricetin, TFC, and quercetin-3-glucuronide ↑<br>Rutin, myricetin, TFC, and quercetin-3-glucuronide ↓ | Peanparkdee, Patrawart, & Iwamoto [25]     |
| 10    | Canned lychee pulp  | TFC  | Heat treatment (70 °C and 121 °C for 30 min)  | TFC ↓  | Wang et al., [26]                          |
| 11    | Pomelo ( <i>Citrus maxima</i> ) juice<br>Common buckwheat and tartary buckwheat | TFC<br>Rutin, quercetin, and TFC                   | Pasteurization<br>Steaming (120 °C for 20 min)  | TFC ↓<br>TFC of common buckwheat, TFC of tartary buckwheat ↓, rutin ↓, quercetin ↑   | Basumatary et al. [27]<br>Chen et al. [28] |
| 12    | Samh ( <i>Mesembryanthemum forsskaei</i> Hochst) seeds                          | TFC  | Roasting (150 °C for 20 min)<br>Roasting (150 °C for 10 min)  | Rutin ↑, quercetin ↑, TFC<br>TFC ↓   | Ahmed et al. [29]                          |
| 13    | Jack bean ( <i>Canavalia ensiformis</i> (L.) DC)                                | Anisoyl kaempferol glycosides                      | Heating (95 °C for 0, 2, 4, 8, 16, and 24 h)  | Transferred to another two isomers   | Sutedja et al. [30]                        |

<sup>a</sup> TFC= total flavonoid content;

<sup>b</sup> TAC= total anthocyanin content; ↓ indicates content decreased, ↑ indicates content increased, - indicates content does not change.

Figure 3



Effects of thermal processing on the biological bioactivity of dietary flavonoids.

8 min), and grilling (110 °C for 15 min) improved TFC of onions, and boiling (100 °C for 30 min) reduced its TFC [16] (Table 1). It is hard to say that one certain thermal processing technique can improve or retain more dietary flavonoids. Obviously, the thermal processing method is a significant interfering factor influencing satiability of dietary flavonoids.

The temperature of thermal processing is one of the critical factors to affect thermal satiability of dietary flavonoids. For example, 25–45 °C thermal treatment, the TFC of mulberry juice was decreased, however, 45–100 °C thermal treatment increased its TFC [14] (Table 1). The total anthocyanin concentrations of blueberry (*Vaccinium* spp.) puree were changed strangely under different temperature treatment (Table 1) [21]. Processing time is another major factor affecting dietary flavonoid satiability. The TFC of teff (*Eragrostis tef*) was increased within 7.5 min of roasting, and decreased after 10 min, which indicated that thermal processing time may be one of the major factors affecting TFC in roasting [22] (Table 1).

Even at the same heating temperature and time, the thermal stability of dietary flavonoids was a noticeable difference in different food matrices. For example, barking at 180 °C for 30 min leads to decreasing of isoquercetin, quercetin, TFC in buckwheat bran, and increasing of isoquercetin, rutin, and TFC in buckwheat flour [23] (Table 1). The thermal satiability of different varieties was various, as evidenced by a marked reduction in TFC in Liaozhi8 sesame after roasting, however,

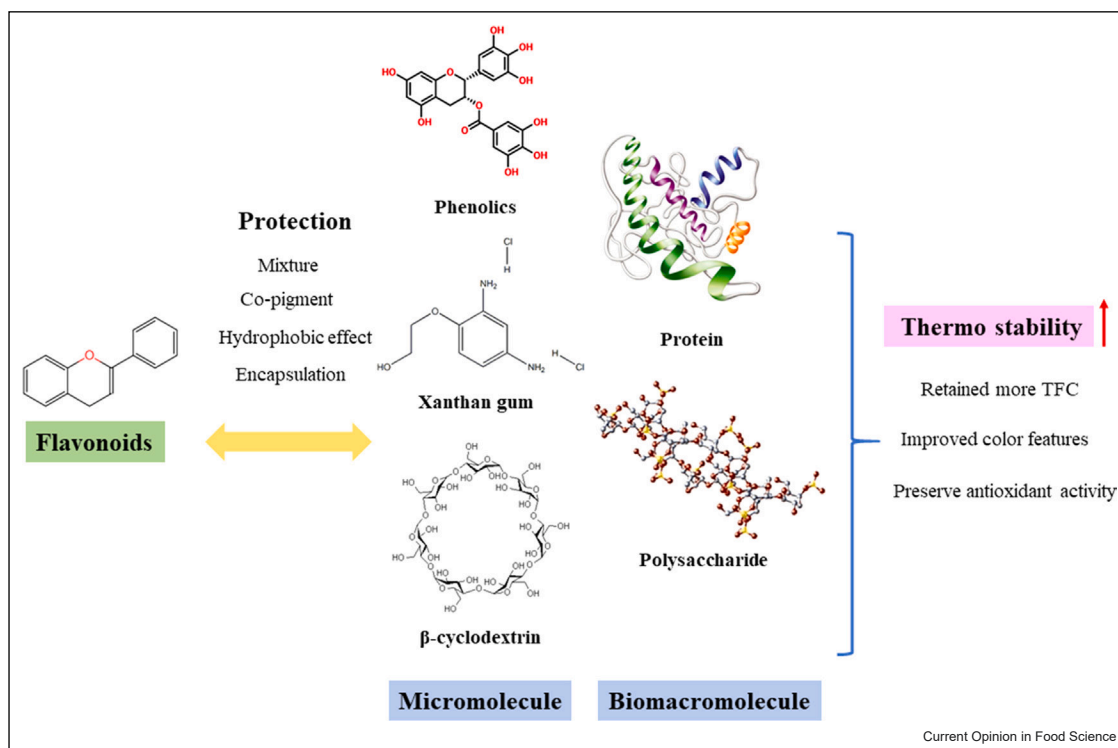
no significant influence on TFC change in ganzhi9, ganzhi17, ji9014, ezhi7, and luozhi18 sesame [24] (Table 1). The thermal stability of foods in different states is also diverse. It was also found that steaming could retain more anthocyanins than boiling for solid-state food, and boiling was preferable for liquid food [15].

For further quantitatively describing the impact of temperature on degradation, the kinetics of degradation of dietary flavonoids were investigated. The thermal degradation of anthocyanins followed a first-order kinetic model, and the kinetic rate constant increased with temperature raised [15, 31]. However, the thermal degradation of TFC in button mushroom (*Agaricus bisporus*) was followed by second-order kinetic model, with the activation energy 25.38 kJ/mol [32]. In the thermal processing, the dietary flavonoids seemly were rearrangement, hydrolysis, and cleavage of glycosidic bonds, which lead to degradation of the matrix, resulting in an apparent increase [30, 33].

### Effect of thermal processing on biological bioactivity of dietary flavonoids

Flavonoids have many biological activities, such as antioxidant, anti-inflammation, and prevention of cardiovascular disease activities. Dietary flavonoids are sensitive to thermal condition, however, some research found that thermal processing can promote its biological bioactivity (Figure 3). The thermal processing techniques decreased flavonoid contents of eggplants, however, promoted their bioavailability on the contrary [20].

Figure 4



Strategies to improve thermal stability of dietary flavonoids.

Antioxidant activities of six flavonoids (rutin, naringin, eriodictyol, mesquitol, luteolin, and 7-*O*-glucoside) improved after thermal processing (from 30 to 130 °C), and their thermal-product solutions increased activities of intracellular superoxide dismutase and glutathione peroxidase [34].

Even through thermal processing exerted negative effect inducing flavonoid degradation, some reactions occurred. The heat destroys cell walls and makes bound flavonoids hydrolyzed [30,35], which promoted the configuration of flavonoids from transformation of soluble flavonoids to insoluble-bound flavonoids, and flavonoid glycosides to aglycones [36,37]. The biological bioactivity of dietary flavonoids is closely related to their configuration, which may improve their biological bioactivity after thermal processing.

### Strategies to improve thermal stability of dietary flavonoids

The use of flavonoid-rich products in food industries and house-cooking is limited by their thermal stability. Considering the low thermal stability of flavonoids, therefore, the strategies to improve thermal stability of dietary flavonoids are encouraged [17,19]. The studies related to the effects of bioactive ingredient addition to improve the thermal stability of dietary flavonoids were

performed (Figure 4). The buckwheat in different concentrations partially replaced rice noodles, which exhibited a high TFC-retention rate of rice noodles after thermal processing [38]. In addition, the xanthan gum or phenolics supplementation also preserved thermal stability of dietary flavonoids after heat treatment [39,40]. The  $\beta$ -cyclodextrin microcapsule technology is a promising strategy to improve the thermal stability of mulberry flavonoids, by retaining 90% TFC after 100 °C heating treatment [14]. Recently, the use of biomacromolecule to improve the thermal stability of flavonoids has been widely studied, such as protein and polysaccharide (Figure 4). The pectic polysaccharide interacted with malvidin-3-*O*- $\beta$ -D-glucoside to improve its thermal stability [41]. The silkworm pupa protein-glucose conjugates combined with cyaniding 3-*O*-glucose via hydrophobic interactions and static quenching, and therefore enhancing its thermal stability over a pH range [42].

### The effects of promising thermal technologies on dietary flavonoids

The conventional thermal processing (e.g. barking, roasting, pasteurization, and steaming) is frequently queried, due to their low processing efficiency and penalty of nutrients in food. The novel thermal techniques, such as microwave, radiofrequency, and ohmic



heating with short time and low temperature, are regarded as promising thermal processes and possess great potential to replace conventional thermal processing. Microwave heating is usually utilized combined with conventional thermal processing techniques. The utilization of microwave heating can effectively shorten processing time and costs, and therefore can preserve more flavonoid contents than conventional thermal processing [43]. For example, with the aid of microwave (450 W) combined with hot-air drying of cherry tomato, the retention of TFC and lycopene was superior compared with conventional air-drying alone at 65 °C [44]. Radio-frequency heating is a kind of dielectric heating-based technique, which can easily penetrate into the inside of the food matrix, and seems to retain more of dietary flavonoids than conventional techniques [45]. Hot-air-assisted radio-frequency heating (100–105 °C, 15 min, and 110–115 °C, 6 min) that stabilized rice bran demonstrated higher free flavonoid content than extruded rice bran, which provides useful information for stabilization method for rice bran [46]. Ohmic heating is an electroconductive heating treatment, which is able to influence the internal of food matrix, and conducive to better retention of dietary flavonoids [47]. The ohmic heating is capable of disrupting food matrixes and modulating internal components, which could increase free flavonoid content in grape than other conventional techniques [48]. In general, the innovative thermal processing techniques probably retained more flavonoids in some situation, depending on its intensity and aims [••18]. The lower temperature and shorter time of food processing are conducive to the retention of flavonoids. Additionally, the equipment of some promising thermal technologies is not suitable for household cooking, and their application in household cooking needs to be further explored.

### Conclusion and perspective

The thermal processing is usually used to inactivate pathogens and undesirable enzymes, which is the most widespread processing method for food industry and household cooking. However, the dietary flavonoids are vulnerable, and easily degraded or converted into various derivatives under thermal processing. On the other hand, some kinds of flavonoids can be released from the food matrix during processing. The flavonoid contents and their bioactivities changed variously under different food matrices and thermal processing conditions. The variations of bioactivities were not in conformity with their contents and stabilities. However, the change mechanism of dietary flavonoids and their thermal-degradation products in food matrices during thermal processing were not studied thoroughly. This is probably because the reaction between dietary flavonoids and other components in food matrices is complicated and their change patterns are hard to clarify. In order to

maintain the stability of dietary flavonoids, the innovative thermal processing techniques with shorter time and lower temperature treatment were emerged, such as microwave, radiofrequency, and ohmic heating. Some equipment of promising thermal technologies may not be suitable for household cooking, which limited their application in household cooking. Overall, these innovative techniques need to be further studied in the food industry and household cooking to improve the thermal satiability of dietary flavonoids.

### Conflict of interest statement

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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### References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest.

1. Khan H, Ullah H, Tundis R, *et al.*: **Dietary flavonoids in the management of huntington's disease: mechanism and clinical perspective.** *eFood* 2020, **1**:38-52.
2. Lin S, Zhang H, Simal-Gandara J, Cheng K, Wang M, Cao H, Xiao J: **Investigation of new products of quercetin formed in boiling water via UPLC-Q-TOF-MS-MS analysis.** *Food Chem* 2022, **386**:132747.
3. Zhong R, Farag M, Chen M, He C, Xiao J: **Recent advances in the biosynthesis, structure–activity relationships, formulations, pharmacology, and clinical trials of fisetin.** *eFood* 2022, **3**:e3.
4. Cao H, Yi L, Zhong J, Högger P, Wang M, Prieto M, Simal-Gandara J, Xiao J: **Investigation of new products and reaction kinetics for myricetin in DMEM via an in situ UPLC-MS-MS analysis.** *Food Front* 2020, **1**:243-252.
5. Chen L, Yao M, Fan X, Lin X, Arroo R, Silva A, Sungthong B, Dragan S, Paoli P, Wang S, Teng H, Xiao J: **Dihydromyricetin attenuates streptozotocin-induced liver injury and inflammation in rats via regulation of NF-κB and AMPK signaling pathway.** *eFood* 2020, **1**:188-195.
6. Ganesan K, Quiles J, Daglia M, *et al.*: **Dietary phytochemicals modulate intestinal epithelial barrier dysfunction and autoimmune diseases.** *Food Front* 2021, **2**:357-382.
7. Teng H, Deng H, He Ym, *et al.*: **The role of dietary flavonoids for modulation of ATP binding cassette transporter mediated multidrug resistance.** *eFood* 2021, **2**:234-246.
8. Zhang H, Caprioli G, Hussain H, Khoi Le N, Farag M, Xiao: **A multifaceted review on dihydromyricetin resources, extraction, bioavailability, biotransformation, bioactivities, and food applications with future perspectives to maximize its value.** *eFood* 2021, **2**:164-184 (2021).

9. Zhao C, Wan XZ, Zhou S, Cao H: **Natural polyphenols: a potential therapeutic approach to hypoglycemia.** *eFood* 2020, **1**:107-118.
  10. Zhu F, Li J, Ma Z, Li J, Du B: **Structural identification and in vitro antioxidant activities of anthocyanins in black chokeberry (*Aronia melanocarpa* Liot).** *eFood* 2021, **2**:201-208.
  11. Ho K, Redan B: **Impact of thermal processing on the nutrients, phytochemicals, and metal contaminants in edible algae.** *Crit Rev Food Sci Nutr* 2021, **62**:508-526.
  12. Cao H, Saroglu O, Karadag A, Diaconeasa Z, Zoccatelli G, Conte-Junior CA, Gonzalez-Aguilar GA, Ou J, Bai WB, Zamarioli CM, de Freitas LAP, Shpigelman A, Campelo PH, Capanoglu E, Hii CL, Jafari SM, Qi YP, Liao P, Xiao JB: **Available technologies on improving the stability of polyphenols in food processing.** *Food Front* 2021, **2**:109-139.
- It highlights the technologies on improving the stability of polyphenols in food processing.
13. Liu J, Bi J, McClements D, et al.: **Impacts of thermal and non-thermal processing on structure and functionality of pectin in fruit-and vegetable-based products: a review.** *Carbohydr Polym* 2020, **250**:116890.
  14. Li D, Zhu M, Liu X, Wang Y, Cheng J: **Insight into the effect of microcapsule technology on the processing stability of mulberry polyphenols.** *LWT* 2020, **126**:109144.
- It investigated the effect of microcapsule technology on the stability of polyphenols.
15. Zhang Y, Sun Y, Zhang H, Mai Q, Zhang B, Li H, Deng Z: **The degradation rules of anthocyanins from eggplant peel and antioxidant capacity in fortified model food system during the thermal treatments.** *Food Biosci* 2020, **38**:100701.
  16. Cattivelli A, Conte A, Martini S, Tagliacucchi D: **Influence of cooking methods on onion phenolic compounds bioaccessibility.** *Foods* 2021, **10**:1023, <https://doi.org/10.3390/foods1005102331>
  17. Choudhury N, Meghwal M, Das K: **Microencapsulation: an overview on concepts, methods, properties and applications in foods.** *Food Front* 2021, **2**:426-442.
- It summarized the concepts, methods, properties and applications of microencapsulation in foods.
18. Fu Y, Liu W, Soladoye OP: **Towards innovative food processing of flavonoid compounds: insights into stability and bioactivity.** *LWT* 2021, **150**:111968.
- It highlights the innovative food processing to improve the stability and bioactivity of flavonoids.
19. Jagtiani E: **Advancements in nanotechnology for food science and industry.** *Food Front* 2022, **3**:56-82.
  20. Martini S, Conte A, Cattivelli A, Tagliacucchi D: **Domestic cooking methods affect the stability and bioaccessibility of dark purple eggplant (*Solanum melongena*) phenolic compounds.** *Food Chem* 2021, **341**:128298.
  21. Zhang W, Shen Y, Li Z, Xie X, Gong ES, Tian J, Si X, Wang Y, Gao N, Shu C, Meng X, Li B, Liu RH: **Effects of high hydrostatic pressure and thermal processing on anthocyanin content, polyphenol oxidase and  $\beta$ -glucosidase activities, color, and antioxidant activities of blueberry (*Vaccinium* Spp.) puree.** *Food Chem* 2021, **342**:128564.
  22. Kataria A, Sharma S, Dar BN: **Changes in phenolic compounds, antioxidant potential and antinutritional factors of Teff (*Eragrostis tef*) during different thermal processing methods.** *Int J Food Sci Technol* 2021, <https://doi.org/10.1111/ijfs.15210>
  23. Ge RH, Wang H: **Nutrient components and bioactive compounds in tartary buckwheat bran and flour as affected by thermal processing.** *Int J Food Prop* 2020, **23**:127-137.
  24. Chen Y, Lin H, Lin M, Zheng Y, Chen J: **Effect of roasting and in vitro digestion on phenolic profiles and antioxidant activity of water-soluble extracts from sesame.** *Food Chem Toxicol* 2020, **139**:111239.
  25. Peanparkdee M, Patrawart J, Iwamoto S: **Physicochemical stability and in vitro bioaccessibility of phenolic compounds and anthocyanins from Thai rice bran extracts.** *Food Chem* 2020, **329**:127157.
  26. Wang Z, Wu G, Shu B, Huang F, Dong L, Zhang R, Su D: **Comparison of the phenolic profiles and physicochemical properties of different varieties of thermally processed canned lychee pulp.** *RSC Adv* 2020, **10**:6743-6751.
  27. Basumatary B, Nayak PK, Chandrasekar CM, Nath A, Nayak M, Kesavan RK: **Impact of thermo sonication and pasteurization on the physicochemical, microbiological and anti-oxidant properties of pomelo (*Citrus maxima*) juice.** *Int J Fruit Sci* 2020, **20**:S2056-S2073.
  28. Chen S, Wu L, Zhu H, Yao L, Wang L: **Effects of processing methods on phenolic compositions, anti-oxidant activities and  $\alpha$ -glucosidase inhibitory ability of two buckwheat varieties.** *Chem Pap* 2021, **75**:1029-1039.
  29. Ahmed IAM, Al Juhaimi FY, Osman MA, Al Maiman SA, Hassan AB, Alqah HA, Babiker EE, Ghafoor K: **Effect of oven roasting treatment on the antioxidant activity, phenolic compounds, fatty acids, minerals, and protein profile of Samh (*Mesembryanthemum forsskaei* Hochst) seeds.** *LWT* 2020, **131**:109825.
  30. Sutedja AM, Yanase E, Batubara I, Fardiaz D, Lioe HN: **Thermal stability of anisoyl kaempferol glycosides in jack bean (*Canavalia ensiformis* (L.) DC) and their effect on  $\alpha$ -glucosidase inhibition.** *J Agric Food Chem* 2022, **70**:2695-2700 (2022).
  31. Slavu M, Aprodu I, Milea SA, Enachi E, Răpeanu G, Bahrim GE, Stănciuc N: **Thermal degradation kinetics of anthocyanins extracted from purple maize flour extract and the effect of heating on selected biological functionality.** *Foods* 2020, **9**:1593, <https://doi.org/10.3390/foods911159335>.
- The degradation kinetics and isomerization of anthocyanins were investigated.
32. Singhal S, Rasane P, Kaur S, Singh J, Gupta N: **Thermal degradation kinetics of bioactive compounds in button mushroom (*Agaricus bisporus*) during tray drying process.** *J Food Process Eng* 2020, **43**:e13555.
  33. Dong J, Li S, Zhang J, Liu A, Ren J: **Thermal degradation of cyanidin-3-O-glucoside: mechanism and toxicity of products.** *Food Chem* 2022, **370**:131018.
  34. Ioannou I, Chekir L, Ghoul M: **Effect of heat treatment and light exposure on the antioxidant activity of flavonoids.** *Processes* 2020, **8**:1078, <https://doi.org/10.3390/pr809107836>
  35. Arfaoui L: **Dietary plant polyphenols: effects of food processing on their content and bioavailability.** *Molecules* 2021, **26**:2959, <https://doi.org/10.3390/molecules26102959>
  36. Li M, Chen X, Deng J, Ouyang D, Wang D, Liang Y, Chen Y, Sun Y: **Effect of thermal processing on free and bound phenolic compounds and antioxidant activities of hawthorn.** *Food Chem* 2020, **332**:127429.
  37. Xiao Y, Yang C, Xu H, Zhang J, Zhang L: **Study on the change of flavonoid glycosides to aglycones during the process of steamed bread containing tartary buckwheat flour and antioxidant,  $\alpha$ -glucosidase inhibitory activities evaluation in vitro.** *LWT* 2021, **145**:111527.
- It reported the transformation of flavonoid glycosides to aglycones during steaming bread.
38. Fu M, Sun X, Wu D, Meng L, Feng X, Cheng W, Tang X: **Effect of partial substitution of buckwheat on cooking characteristics, nutritional composition, and in vitro starch digestibility of extruded gluten-free rice noodles.** *LWT* 2020, **126**:109332.
  39. Erşan S, Müller M, Reuter L, Carle R, Müller-Maatsch J: **Co-pigmentation of strawberry anthocyanins with phenolic compounds from rooibos.** *Food Chem Mol Sci* 2022, **4**:100097.
  40. Zhao L, Pan F, Mehmood A, Zhang Y, Hao S, Rehman AU, Li J, Wang C, Wang Y: **Protective effect and mechanism of action of xanthan gum on the color stability of black rice anthocyanins in model beverage systems.** *Int J Biol Macromol* 2020, **164**:3800-3807.
  41. Fernandes A, Brandão E, Raposo F, Maricato É, Oliveira J, Mateus N, Coimbra M, de Freitas V: **Impact of grape pectic polysaccharides on anthocyanins thermostability.** *Carbohydr Polym* 2020, **239**:116240.

The effects of polysaccharides on the thermostability of anthocyanins were investigated.

42. Attaribo T, Huang G, Xin X, Zeng Q, Zhang Y, Zhang N, Tang L, Sedjoah R, Zhang R, Lee K, Jin B, Gui Z: **Effect of the silkworm pupa protein–glucose conjugate on the thermal stability and antioxidant activity of anthocyanins.** *Food Funct* 2021, **12**:4132-4141.
43. Guzik P, Kulawik P, Zając M, Migdał W: **Microwave applications in the food industry: an overview of recent developments.** *Crit Rev Food Sci Nutr* 2021, **1**-20, <https://doi.org/10.1080/10408398.2021.1922871>
44. Wiset L, Poomsa-ad N, Onsaard W: **Drying characteristics and quality evaluation in microwave-assisted hot air drying of cherry tomato.** *Eng Appl Sci Res* 2021, **48**:724-731.
45. Ling B, Cheng T, Wang S: **Recent developments in applications of radio frequency heating for improving safety and quality of food grains and their products: a review.** *Crit Rev Food Sci Nutr* 2020, **60**:2622-2642.
46. Liao M, Damayanti W, Xu Y, Zhao Y, Xu X, Zheng Y, Jiao S: **Hot air-assisted radio frequency heating for stabilization of rice bran: enzyme activity, phenolic content, antioxidant activity and microstructure.** *LWT* 2020, **131**:109754.
47. Makroo HA, Rastogi NK, Srivastava B: **Ohmic heating assisted inactivation of enzymes and microorganisms in foods: a review.** *Trends Food Sci Technol* 2020, **97**:451-465.
48. Jesus MS, Ballesteros LF, Pereira RN, Genisheva Z, Carvalho AC, Pereira-Wilson C, Domingues L: **Ohmic heating polyphenolic extracts from vine pruning residue with enhanced biological activity.** *Food Chem* 2020, **316**:126298.