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REVIEW

Glycoconjugation: An approach to cancer therapeutics

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Abstract

Cancer constitutes the second leading cause of death globally and is considered to have been responsible for an estimated 9.6 million fatalities in 2018. Although treatments against gastrointestinal tumors have recently advanced, those interventions can only be applied to a minority of patients at the time of diagnosis. Therefore, new therapeutic options are necessary for advanced stages of the disease. Glycosylation of antitumor agents, has been found to improve pharmacokinetic parameters, reduce side effects, and expand drug half-life in comparison with the parent compounds. In addition, glycosylation of therapeutic agents has been proven to be an effective strategy for their targeting tumor tissue, thereby reducing the doses of the glycodrugs administered to patients. This review focusses on the effect of the targeting properties of glycosylated antitumor agents on gastrointestinal tumors.

Key words: Glycosylation; Gastrointestinal cancers; Antitumoral agents; Therapeutic strategies; Drug targeting

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Core tip: In nature, glycosylation has proven an effective strategy for expanding the biologic information of biomolecules by adding a new level of structural diversity. The high specificity of the interaction with carbohydrates and the overexpression of carbohydrate receptors in tumoral cells that can be specifically targeted by glycodrugs enable a selective administration of those agents to the tumor tissues. Accordingly, the glycosylation of antitumor agents has been found to improve pharmacokinetic parameters, reduce side effects, expand drug half-life, and reduce the dosage of the consequent glycoderivatives.



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INTRODUCTION

Cancer is the second leading cause of death worldwide, having been responsible for an estimated of 9.6 million deaths in 2018^[1,2]. Moreover, 21.6% of all tumors worldwide are gastrointestinal cancers at more than 26000 cases per year^[3]. Despite considerable research efforts in recent years, the impact of conventional strategies including surgery, radiotherapy, and chemotherapy - on the prognosis of tumors has been only moderate^[4,5]. The anticancer drugs used in chemotherapy usually target cells that are proliferating. These chemodrugs can be grouped according to their main role: antitumor antibiotics, alkylating agents, topoisomerase inhibitors, DNAcomplexing agents, mitotic inhibitors, hormones, and immunotherapeutic agents. The activity of those drugs is essential for predicting side effects; moreover, what is remarkable is that most of the available anticancer drugs have the disadvantage of lacking systems for delivery to the target organ or tissue. Consequently, the majority of the administered drug remains circulating in the bloodstream, thus increasing the side effects on noncancerous cells^[6].

The selective targeting of therapeutic agents has several advantages, such as increasing the concentration of the drugs in the tumor and reducing the concentration in other tissues^[7]. Carbohydrates *per se*, exhibit a high solubility in water, a low toxicity, and a high biocompatibility; thus constituting an attractive system for facilitating drug-delivery. Glycosylated compounds can be targeted to a broad range of cellular receptors because of the specificity of interaction with cell-surface carbohydrates. Accordingly, a number of glycoconjugated antitumoral agents have been reported to selectively deliver the parent drugs to the desired sites^[8,9]. In 1971, Rogers *et al*^[10] demonstrated for the first time the usefulness of targeting proteins that bear carbohydrates as ligands. To our knowledge, no therapeutic targeting system is currently on the market, though many candidates aimed at that end nevertheless exist. This review will therefore focus primarily on the synthesis of the most relevant glycosylated therapeutic agents and the effect of those derivatives on gastrointestinal tumors.

GLYCOTARGETING SYSTEMS MEDIATED BY RECEPTORS AND TRANSPORTERS

The most widely studied drug-transporting and receptor-targeting glycoligands are the glucose-transporters (GLUTs) and the lectin receptors as described below.

GLUT

The energy produced from glucose metabolism is essential for sustaining mammalian-cell life. The end products of that metabolic pathway are lactate and, upon full oxidation in the mitochondria, $CO_2^{[11]}$.

In tumors and other proliferating cells, the rate of glucose uptake increases considerably and lactate is produced, regardless of the availability of oxygen and functional mitochondria. The tendency of tumoral tissues to anaerobically metabolize large amounts of glucose, in comparison with noncancerous tissue, is known as the Warburg Effect^[12,13].

The GLUT is a transport protein of the facilitator family involved in glucose translocation across the cell membrane. Although, GLUT transporters are expressed in almost every cell type^[14], tumor cells express a large number of glucose transporters that are related to poor prognosis^[15]. At present, 14 different GLUTs-1 to 14 - have been described^[16]. GLUT-1, in particular, is known to be overexpressed in tumor cells, including those of liver, pancreas, and stomach^[17]. These transporters can specifically recognize and transport different sugars, such as glucose, mannose, galactose, 2-deoxyglucose and glucosamine analogs^[18].

Therefore, as a result of the Warburg effect, designing and developing glycosylbased targeted drugs is a subject of high/considerable/widespread interest^[19,20].



Lectin receptors

Lectins are defined as proteins, usually linked to carbohydrates^[21-23], that are present in plants and animals and involved in many biologic processes - including cell growth, differentiation, signalling, adhesion and migration, and apoptosis^[24-26]. Lectins can act as receptors, either for binding oligosaccharides to cell membranes or free-floating glycans involving monosaccharides in order to mediate signal transduction and/or drug transport^[27].

Asialoglycoprotein receptors (ASGPRs), including asialoglycoprotein receptors 1 and 2-ASGPR1 and ASGPR2 - are expressed on the surface of hepatocytes and stomach and gallbladder epithelia^[28] and preferentially interact with the sugars D-galactose and L-rhamnose. The molecular mechanism consists in the internalization of the receptor-ligand complex through a clathrin-mediated endocytosis. Once inside the cell, the ligands are released, enabling the recycling of ASGPR receptors back into the plasma membrane. The quick cycling of internalized receptors is the key process that maintains their concentration on the cell surface. Owing to the high specificity in the binding of galactose and rhamnose to ASGPR receptors, these interactions result a promising approach to drug targeting^[29].

Among the lectin-based receptors, the rhamnose-binding lectin receptor ligates specifically to rhamnose and is highly expressed on various tumor-cell types, like the cell-culture lines KB (from a human squamous-cell carcinoma), PC3 (from a human prostatic small-cell carcinoma), HT-29 (from a human colon adenocarcinoma) and MCF-7 (a breast adenocarcinoma)^[30]. Although the molecular mechanism involved in the transmission of messages by rhamnose-binding lectin receptor has not been yet studied in humans, the strategy for using rhamnosylated anticancer molecules as novel candidates for pharmacological applications has recently been explored^[29].

ANTITUMORAL DRUG GLYCOCONJUGATES

Below we present several antitumoral agents as informative examples of such modifications - namely, ifosfamide, chlorambucil, and paclitaxel. The drugs were linked to various carbohydrate moieties and the antitumoral effects evaluated on gastrointestinal tumors. The cytotoxic activities of the synthesized glycoconjugates were then compared to those of the corresponding parent nonglycosylated molecules (Table 1).

Ifosfamide

Ifosfamide - whose cytotoxic metabolite of in plasma is ifosforamide - is an alkylating agent has been bound to β-D-glucose to form glufosfamide^[31,32]. This glycoconjugate was the first molecule bearing a sugar to be explicitly designed and evaluated as a cancer-targeting cytotoxic compound. Within tumor cells, glufosfamide is metabolized by glucosidases to form ifosforamide. This cytotoxic metabolite, in turn, forms DNA crosslinks, therefore inhibiting DNA replication and cell growth^[33]. Moreover, treatment with GLUT-1 inhibitors reduced the anticancer efficiency of glufosfamide, suggesting that the drug conjugate was internalized into cells via the GLUT-1 transporter. Finally, glufosfamide was less myelotoxic and presented a higher antitumour activity both *in vitro* and *in vivo* than the parent aglycone^[52].

In 1997, the first human clinical trial to test glufosfamide was carried out in Europe, and the results obtained with 20 pancreatic-cancer patients were reported^[34]. Two cases evidenced a good response to the treatment, 10 resulted in stable disease, while 8 patients failed to respond. More significantly, one pancreatic-cancer patient in a different trial experienced a complete remission for over 4 years. Since pancreatic-cancer biopsy samples had been found to overexpress GLUT-1; in 2010, Chiorean *et al*^[35] performed a phase-II study of glufosfamide plus the nucleoside analog gemcitabine, the standard chemotherapeutic treatment in pancreatic cancer. Glufosfamide and gemcitabine in combination yielded a modest response in two trials on pancreatic-adenocarcinoma patients. In conclusion, glufosfamide appeared to constitute an effective cytotoxic agent exhibiting high antitumor selectivity that was due to an active interaction with the transporter GLUT-1.

Chlorambucil

Chlorambucil is an antineoplasic drug within the class of alkylating agents that is used to treat various forms of cancer^[36]. The reactive radical ethylenimonium forms after alkylation that interferes in DNA, RNA, and protein synthesis. The first report of chlorambucil synthesis was over five decades ago^[37]. Goff *et al*^[38] (2010) synthesized and evaluated a 63-member library of chlorambucil-based neoglycosides in ten different human-carcinoma cell-culture systems, including lung, colorectal, liver,

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Table 1 List of the glycoconjugates and its effects

Agent	Activity	Sugar moiety	Efficacy glycoconjugates compared to aglicone
Ifosfamide	Alkalating agent	Glucose	<i>In vitro</i> (less toxic) and <i>in vivo</i> (reduced tumor size). Clinical trials ongoing.
Chlorambucil	Alkalating and DNA-complexing agent	D-threose	HT29 and HCT15 (showed 8-12 fold, and 15-fold, respectively, improved activities targeting cancers cell lines over the parent drug).
Geldanamycin	HSP90 inhibitor	Glucose	Glucose-GA showed anticancer activity with IC50 of 70.2-380.9 nM in SW620, HT29, MCF-7 and K562 cancer cells by-glucosidase activation inside of the tumor cells.
Geldanamycin	HSP90 inhibitor	Galactose	SW620, HT29, MCF-7 and K562 (anticancer activity of galactose-GE conjugate increased by 3- to 40-fold when incubated with galactosidase over the parent drug).
Emodin	Tyrosine kinase inhibitor	D-rhamnose	A594, HepG2, OVCAR-3, Hela, K562 and SGC-790 (cell proliferation was inhibited and EM-d-Rha conjugate displayed IC ₅₀ values in low µmolar ranges).
Paclitaxel	Mitotic inhibitor	Glucose	NCI-H838, MES-SA, HCT-116, and NPC-TW01 (cell proliferation was inhibited the conjugated presented higher cytotoxicity, induced tubulin aggregation and chromosomal condensation compared to paclitaxel).
Doxorubicin	Antitumor antibiotic	Galactose	<i>In vitro</i> viability of HepG2 (hepatocarcinoma) and MCF-7 (breast cancer) tumor cells incubated with DOX was higher than that of Gal-DOX. <i>In vivo</i> experiments showed that tumor size in Gal-DOX- treated groups was greatly reduced in comparison to the DOX-treated group.
Doxorubicin	Antitumor antibiotic	2-amino-2-deoxy-D-glucose and succinic acid	<i>In vitro</i> and <i>in vivo</i> studies showed that 2DG-SUC-ADM induced a higher level of apoptosis and higher inhibition rates in MCF-7 and HepG2 tumoral cells than the parent aglycone ADM.

breast, prostate, central nervous system, and ovarian cell lines. The synthesis consists of several chemical steps to perform chlorambucil-based libraries for chemoselective glycosylation. On the basis of this study, the neoglycosides, D-glucuronolactonide and D-threoside were selected as the most potent antitumoral agents. D-threoside glycoside manifested an 8-fold higher efficacy in general, with a respective 12-fold, and 15-fold greater effectiveness in targeting the malignant cell lines HT-29, and HCT-15 (from colorectal adenocarcinomas) over the parent drug. The authors concluded that D-threoside was the most active chlorambucil neoglycoside among the compounds tested. In summary, a novel panel of glycoconjugates were designed and synthesized through the use of common metabolic carbohydrates that are preferentially recognized by cell-membrane receptors, therefore favoring the uptake of the glycodrugs^[38]. The specific mechanisms and the receptors or transporters involved in this process remain to be elucidated.

Geldanamycin

Geldanamycin (GE) is a potent anticancer antibiotic that inhibits the heat-shock protein 90 (Hsp90)^[39]. This protein is a molecular chaperon involved in the modulation of the activity of various protein kinases. Hsp90 has been found to be 2- to 10-fold more expressed in various human cancer cells than in normal tissues, suggesting the use of that protein as a possible target for cancer therapy^[40]. Although

GE has long been recognized as an inhibitor of tumor growth, the potential clinical utility of that agent is hampered by its severe side effects^[41,42]. To circumvent this problem, an approach involving a binary chemical step was designed by Cheng et al^[43] (2005), resulting in a series of glycosyl-GE derivatives. The enzyme-specific activation of these glycosylated prodrugs with galactose and glucose moieties was performed with α -galactosidase and β -glucosidase, respectively. The effect of the resulting derivatives was evaluated in different cancer-cell lines, including SW620, HT-29, MCF-7, and K-562 leukemia cells. In particular, the glucose-GE exhibited antitumor activity after cleavage of the glucose moiety by the β -glucosidase inside the tumor cells. The anticancer activity of the galactose-GE conjugate increased by 3- to 40-fold when incubated with exogenous galactosidase in vitro, but remained inactive in the absence of the added enzyme because, unlike intracellular glucosidases, galactosidases are present in only low or undetectable levels in serum^[43]. Consequently, the activation of the galactosyl-GE prodrugs by the exogenous enzyme would appear to be a necessary targeting approach, involving a strategy leading to a dual tactic consisting of the glycosylation of the enzyme as well as the drug, in order to direct the whole system to the target tissue^[8,9].

Emodin

Emodin (1,3,8-trihydroxy-6-methyl-anthraquinone) is a natural anthraquinone derivative found in the roots and rhizomes of numerous plants. As a tyrosine-kinase inhibitor, emodin inhibits cell growth in several types of tumor cells^[44-46] and regulates the expression of genes involved in cell apoptosis, oncogenesis, cell proliferation, and cancer-cell invasiveness and metastasis^[47-49]. The antitumor effects of emodin have been described, but the molecular mechanism has not been fully elucidated. The synthesis and design of emodin conjugated to D-rhamnose (EM-D-Rha), inhibited cell proliferation in a panel of different human-cancer-cell lines including A549 (lung carcinoma), HepG2 (hepatoma), OVCAR-3 (ovarian carcinoma), HeLa (cervical adenocarcinoma). EM-d-Rha also manifested lower IC₅₀ values and was 10-fold more cytotoxically effective than the aglycone on HepG2 cells, leading to a decrease in mitochondrial transmembrane potential and to an upregulation of the expression of apoptosis elements^[50].

Doxorubicin (Adriamycin)

Doxorubicin (DOX), the active compound in the trade drug named adriamycin (ADM), belongs to the anthracycline family, is one of the most powerful and widely used chemotherapeutic drugs, and as such is recognized by the World Health Organization^[51]. Unfortunately, apart from the drug's activity as an intercalating agent and topoisomerase-II inhibitor, DOX causes severe toxic effects such as nephrotoxicity, hepatotoxicity, and alopecia, compromising mainly heart tissues and the gastrointestinal tract as a result of the compound's systemic action. In view of the many efforts that have been made order to overcome these limitations, an increase in drug efficiency through the conjugation of DOX to a carbohydrate ligand that can specifically recognize tumoral cells is a promising approach^[52,53].

Ma et al^[54] (2015) conjugated galactose to DOX covalently to form the prodrug Gal-DOX and then evaluated the derivative's tumor-targeting capability in different cancer-cell lines. They found that Gal-Dox treatment increased cell death in HepG2 and MCF-7 cells compared to DOX. These results were the opposite when they treated normal L02 hepatocytes. They proposed that this difference was caused by the high specific binding of Gal to ASGPR1 receptors, which were overexpressed on the surface of HepG2 and MCF-7 tumor cells. Consistent with that conclusion, since those nonmalignant hepatocytes contained lower plasma-membrane levels of ASGPR1, the L02 cells maintained high cell viability, thus suggesting low toxicity of Gal-DOX. As to side effects, Gal-DOX is accumulated in heart tissue to a lesser extent than DOX, thus diminishing myocardial damage. In vivo results indicated a reduction in tumor size in the Gal-DOX-treated group compared to the DOX-treated group. Of notable relevance here is that the survival of the Gal-DOX-treated group was 100%, whereas the rate of DOX-treated group was 50%, thus supporting the conclusion that Gal-Dox preferentially targeted the tumor cells. These findings suggest that Gal-DOX is a promising drug for tumor- directed therapy[54]. In another study, ADM was conjugated with 2-amino-2-deoxy-D-glucose and succinic acid (2DG-SUC-ADM). There the investigators found that the ternary derivative was highly specific for tumor cells (MCF-7 and HepG2) via the GLUT-1 transporter, while exerting no significant adverse effects on normal cells. The action of the glycoconjugate was also confirmed in vivo, thus demonstrating that the glycosylated molecule could specifically target tumoral cells as opposed to the aglycone. These results suggest 2DG-SUC-ADM as a promising drug for targeting cancer cells^[55].



Paclitaxel

Paclitaxel is a cytotoxic chemotherapeutic drug, classified as a "plant alkaloid," a "taxane" and an "antimicrotubule agent"[55,56]. The molecular basis for the action of paclitaxel consists in a binding to tubulin subunits, thus disrupting mitosis and causing cell death^[57-59]. Despite being widely used in the clinic, this drug has pronoinced side-effects; thus affecting both normal cells and tumors. Another problem with paclitaxel is its poor water solubility. Lin et al^[60] (2008) designed glycanbased paclitaxel prodrugs, consisting of 2'-paclitaxel conjugated with glucosyl or glucuronyl residues by an ester or an ether linkage. These glycodrugs not only had increased solubility in water compared to the parent compound, but exhibited higher selectivity against targeted cancer cells as well. Glucosyl-paclitaxel displayed the higher cytotoxicity and could induce tubulin aggregation and chromosomal condensation in a tumor-cell line^[60]. The cells overexpressing GLUTs favored the uptake of glucose-paclitaxel and facilitated the entrance of the bulky compounds. Therefore, the authors proposed the synthesis of glucoconjugates as an alternative approach for improving the directed delivery of drugs to cancer cells overexpressing GLUTs^[60].

5-fluorouracil

5-fluorouracil (5Fu) is part of a group of chemotherapeutic drugs known as antimetabolites. These compounds incorporate into normal macromolecules to produce a slightly different structure that interferes in the metabolism of the cancer cells. 5Fu is one of the most commonly used drugs to treat gastrointestinal and breast cancers, although the agent is mostly used in combination with other drugs like oxaliplatin^[61-63].

Davis and coauthors synthesized the prodrugs DOX and 5Fu capped by Lrhamnose and evaluated their use in a lectin-directed–enzyme-activated-prodrug therapy (LEAPT) system. The LEAPT system uses biocatalysts for site-selective drug delivery through the construction of novel glycosylated enzymes and prodrugs. The drugs capped with rhamnose synthesized by Davis and coworkers were released by an α-rhamnosidase, which enzyme is by itself glycosylated with galactose. The glycosylated enzyme specifically targets the ASGPR hepatic-receptors. The biodistribution revealed that the glycosylated enzyme became quickly sequestered to the liver, and to a lower extent to the kidney and bladder. Of essential relevance was that the co-administration of the prodrugs did not interfere in the colocalization of the LEAPT system^[8,9]. Finally, the authors demonstrated that the prodrug could be activated in the liver only by the presence of the prelocalized glycosylated enzyme^[8].

CARBOHYDRATE-BASED VACCINES

Within glycoscience new approaches are being undertaken for cancer therapy. Glycan-based vaccines have been developed for the specific enhancement of the immune response. The more complex task in formulating a cancer vaccine would be the selection of the appropriate antigen, which molecule should be exclusively expressed by cancer cells. Tumor-associated carbohydrate antigens, ought to be cellular components that are essential for malignant-cell survival in order to prevent the downregulation of the antigen and thus maintain the immune response. Tumorassociated carbohydrate antigens can be divided into two classes: glycoprotein antigens: The Thomsen Nouveau (Tn; GalNAc-a1-O-Ser/Thr), Thomsen-Friendreich (TF), and sialyl-Tn (sTn) linked to the hydroxyl group of serine or threonine residues of proteins and glycolipids^[64]. Several studies demonstrated that altered glycosylation and aberrant glycan structures helped tumor cells to circumvent immune surveillance. Since high-affinity T cells recognizing self-antigens are eliminated during development of the central immune system, tumor-associated carbohydrate antigen-directed cancer vaccines face the challenge of activating any remaining lowaffinity T cells[65-69].

Glycan-based vaccines may prove to be beneficial because unusual glycan motifs on glycoproteins can lead to vaccines with high specificity^[70]. Mucin 1 (MUC1) is an *O*-linked glycan transmembrane protein overexpressed in various tumors – such as lung, breast, pancreas, kidney, ovary, and colon – and has been demonstrated to be aberrantly glycosylated in cancer cells but highly glycosylated in normal cells. Up to the present, most vaccines have targeted nonglycosylated MUC1, although this approach did not prove to be cancer-specific. Thus, targeting cancer-associated glycopeptide epitopes in MUC1 would be a promising alternative possibility^[71-73]. New attempts have been made to develop glycosylation-based vaccines that target MUC1. In 2012, Madsen *et al*^[74] found that the addition of GalNAc residues to MUC1



to aid antigen uptake and major-histocompatability-complex-class-II presentation for the generation of a potent cancer-specific humoral response. In another investigation, Li *et al*^[75] designed and synthesized two linear trivalent glycopeptides of the immunedominant epitope of MUC1. The antibodies induced by glycosylated-MUC1-based vaccines had a stronger binding than those raised by nonglycosylated MUC1, thus, having the potential to overcome the weak immunogenicity of natural MUC1 glycopeptides^[75]. Further studies demonstrated that the antibodies elicited by a vaccine composed of the immunoadjuvant (Pam3CysSK4), a peptide T-helper epitope, and an aberrantly glycosylated MUC1 peptide, were significantly more lytic and more effective in tumor prevention than the unglycosylated control^[76]. Other glycan-based vaccines that lead to higher immune responses are presently under development. The most likely to be effective is a vaccine composed of MUC1 glycopeptide in combination with a T-helper peptide, while another uses a combination of MUC1 with the toll-like receptor 2. The last one is composed of MUC1, toll-like receptor 2/9, and a Th peptide^[70].

No clinical trials are currently being undertaken on gastrointestinal tumors, although a phase-III clinical trial is being carried out in patients with metastatic breast cancer that involves the glycan-based vaccine, sTn-KLH. The sTn-KLH–vaccine group evidenced an improved survival, relative to the overall survival among the patients treated without the vaccine^[77,78].

Many efforts have been made to develop carbohydrate-based vaccines, though until now, none have been approved for clinical use, and many cancer-vaccine candidates have failed in clinical trials. One reason could be the ability of tumor cells to escape from the endogenous immune response or downregulate the immune-target molecules. Nevertheless, an optimization of vaccine formulation in terms of receptoror antigen-targeting delivery systems may lead to significant improvements in the utilization of currently available carbohydrate antigens so as to open new perspectives for cancer treatment.

PERSPECTIVES

The conjugation of anticancer agents to carbohydrate-ligands that preferentially target tumor cells has resulted in the prediction that several conjugates would prove to have clinical efficacy. Specifically, glycoconjugation offers an improvement in targeting cancer cells, since are many sugar receptors are overexpressed in tumoral cells^[79,80] (Figure 1). Furthermore, the addition of a sugar moiety (*e.g.*, glucose, galactose, rhamnose) improves the water solubility and stability of the parent drugs. During recent years, this field has been emerging in translational medicine; but, in closing, we need to stress that glycodrug development is a rigorous process that still requires many steps to determine the true utility of that strategy.



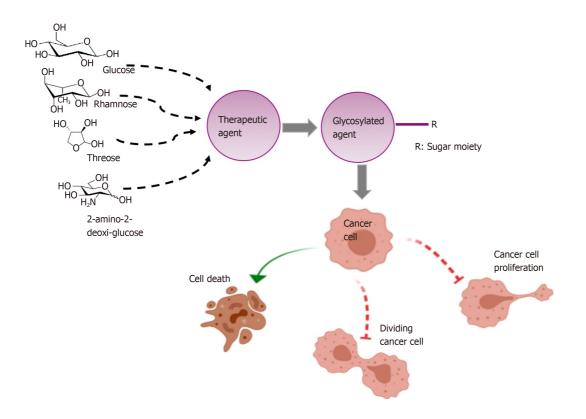


Figure 1 Glycoconjugation strategy. Different sugar moieties can be conjugated to therapeutic agents, with the aim at improving drug efficiency, including antiproliferative activity and growth inhibition, and upregulating cancer-cell death.

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REFERENCES

- Cancer Research UK. General cancer information. Available from: 1 w.cancerresearchuk.org/about-cancer/cancer-in-genera
- Global Burden of Disease Cancer Collaboration. Global Burden of Disease Cancer Collaboration, 2 Fitzmaurice C, Allen C, Barber RM, Barregard L, Bhutta ZA, Brenner H, Dicker DJ, Chimed-Orchir O, Dandona R, Dandona L, Fleming T, Forouzanfar MH, Hancock J, Hay RJ, Hunter-Merrill R, Huynh C, Hosgood HD, Johnson CO, Jonas JB, Khubchandani J, Kumar GA, Kutz M, Lan Q, Larson HJ, Liang X, Lim SS, Lopez AD, MacIntyre MF, Marczak L, Marquez N, Mokdad AH, Pinho C, Pourmalek F, Salomon JA, Sanabria JR, Sandar L, Sartorius B, Schwartz SM, Shackelford KA, Shibuya K, Stanaway J, Steiner C, Sun J, Takahashi K, Vollset SE, Vos T, Wagner JA, Wang H, Westerman R, Zeeb H, Zoeckler L, Abd-Allah F. Ahmed MB. Alabed S. Alam NK. Aldhahri SF. Alem G. Alemavohu MA. Ali R. Al-Raddadi R. Amare A, Amoako Y, Artaman A, Asayesh H, Atnafu N, Awasthi A, Saleem HB, Barac A, Bedi N, Bensenor I, Berhane A, Bernabé E, Betsu B, Binagwaho A, Boneya D, Campos-Nonato I, Castañeda-Orjuela C, Catalá-López F, Chiang P, Chibueze C, Chitheer A, Choi JY, Cowie B, Damtew S, das Neves J, Dey S, Dharmaratne S, Dhillon P, Ding E, Driscoll T, Ekwueme D, Endries AY, Farvid M, Farzadfar F, Fernandes J, Fischer F, G/Hiwot TT, Gebru A, Gopalani S, Hailu A, Horino M, Horita N, Husseini A, Huybrechts I, Inoue M, Islami F, Jakovljevic M, James S, Javanbakht M, Jee SH, Kasaeian A, Kedir MS, Khader YS, Khang YH, Kim D, Leigh J, Linn S, Lunevicius R, El Razek HMA, Malekzadeh R, Malta DC, Marcenes W, Markos D, Melaku YA, Meles KG, Mendoza W, Mengiste DT, Meretoja TJ, Miller TR, Mohammad KA, Mohammadi A, Mohammed S, Moradi-Lakeh M, Nagel G, Nand D, Le Nguyen Q, Nolte S, Ogbo FA, Oladimeji KE, Oren E, Pa M, Park EK, Pereira DM, Plass D, Qorbani M, Radfar A, Rafay A, Rahman M, Rana SM, Søreide K, Satpathy M, Sawhney M, Sepanlou SG, Shaikh MA, She J, Shiue I, Shore HR, Shrime MG, So S, Soneji S, Stathopoulou V, Stroumpoulis K, Sufiyan MB, Sykes BL, Tabarés-Seisdedos R, Tadese F, Tedla BA, Tessema GA, Thakur JS, Tran BX, Ukwaja KN, Uzochukwu BSC, Vlassov VV, Weiderpass E, Wubshet Terefe M, Yebyo HG, Yimam HH, Yonemoto N, Younis MZ, Yu C, Zaidi Z, Zaki MES, Zenebe ZM, Murray CJL, Naghavi M. Global, Regional, and National Cancer Incidence, Mortality, Years of Life Lost, Years Lived With Disability, and Disability-Adjusted Life-years for 32 Cancer Groups, 1990 to 2015: A Systematic Analysis for the Global Burden of Disease Study. JAMA Oncol 2017; 3: 524-548 [PMID: 27918777 DOI: 10.1001/jamaoncol.2016.5688]

Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: 3



GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018; **68**: 394-424 [PMID: 30207593 DOI: 10.3322/caac.21492]

- 4 **Taylor WF**, Jabbarzadeh E. The use of natural products to target cancer stem cells. *Am J Cancer Res* 2017; 7: 1588-1605 [PMID: 28744407]
- 5 Rao D, Parakrama R, Augustine T, Liu Q, Goel S, Maitra R. Immunotherapeutic advances in gastrointestinal malignancies. *NPJ Precis Oncol* 2019; 3: 4 [PMID: 30729176 DOI: 10.1038/s41698-018-0076-8]
- 6 **Skeel RT**. Antineoplastic drugs and biological response modifiers. In: Skeel RT. Handbook of cancer chemotherapy. Philadelpia: Lippincott Williams and Wilkins, 1999: 108-109
- 7 Sathornsumetee S, Rich JN. Molecularly targeted therapy in neuro-oncology. In: Aminoff MJ, Boller F, Swaab DF, editors. Handbook of Clinical Neurology. Elsevier, 2012: 255-278 [PMID: 22230448 DOI: 10.1016/B978-0-444-52138-5.00018-9]
- 8 Robinson MA, Charlton ST, Garnier P, Wang XT, Davis SS, Perkins AC, Frier M, Duncan R, Savage TJ, Wyatt DA, Watson SA, Davis BG. LEAPT: lectin-directed enzyme-activated prodrug therapy. *Proc Natl Acad Sci USA* 2004; 101: 14527-14532 [PMID: 15448212 DOI: 10.1073/pnas.0303574101]
- 9 Garnier P, Wang XT, Robinson MA, van Kasteren S, Perkins AC, Frier M, Fairbanks AJ, Davis BG. Lectin-directed enzyme activated prodrug therapy (LEAPT): Synthesis and evaluation of rhamnose-capped prodrugs. J Drug Target 2010; 18: 794-802 [PMID: 21047273 DOI: 10.3109/1061186X.2010.529909]
- 10 Rogers JC, Kornfeld S. Hepatic uptake of proteins coupled to fetuin glycopeptide. Biochem Biophys Res Commun 1971; 45: 622-629 [PMID: 5128172 DOI: 10.1016/0006-291x(71)90462-1]
- Annibaldi A, Widmann C. Glucose metabolism in cancer cells. Curr Opin Clin Nutr Metab Care 2010; 13: 466-470 [PMID: 20473153 DOI: 10.1097/MCO.0b013e32833a5577]
- 12 Warburg O. On the origin of cancer cells. *Science* 1956; **123**: 309-314 [PMID: 13298683 DOI: 10.1126/science.123.3191.309]
- 13 Gonzalez CD, Alvarez S, Ropolo A, Rosenzvit C, Bagnes MF, Vaccaro MI. Autophagy, Warburg, and Warburg reverse effects in human cancer. *Biomed Res Int* 2014; 2014: 926729 [PMID: 25197670 DOI: 10.1155/2014/926729]
- Uldry M, Thorens B. The SLC2 family of facilitated hexose and polyol transporters. *Pflugers Arch* 2004;
 447: 480-489 [PMID: 12750891 DOI: 10.1007/s00424-003-1085-0]
- 15 Kunkel M, Reichert TE, Benz P, Lehr HA, Jeong JH, Wieand S, Bartenstein P, Wagner W, Whiteside TL. Overexpression of Glut-1 and increased glucose metabolism in tumors are associated with a poor prognosis in patients with oral squamous cell carcinoma. *Cancer* 2003; 97: 1015-1024 [PMID: 12569601 DOI: 10.1002/cncr.11159]
- Thorens B, Mueckler M. Glucose transporters in the 21st Century. *Am J Physiol Endocrinol Metab* 2010;
 298: E141-E145 [PMID: 20009031 DOI: 10.1152/ajpendo.00712.2009]
- 17 Carvalho KC, Cunha IW, Rocha RM, Ayala FR, Cajaíba MM, Begnami MD, Vilela RS, Paiva GR, Andrade RG, Soares FA. GLUT1 expression in malignant tumors and its use as an immunodiagnostic marker. *Clinics (Sao Paulo)* 2011; 66: 965-972 [PMID: 21808860 DOI: 10.1590/s1807-59322011000600008]
- 18 Deng D, Xu C, Sun P, Wu J, Yan C, Hu M, Yan N. Crystal structure of the human glucose transporter GLUT1. Nature 2014; 510: 121-125 [PMID: 24847886 DOI: 10.1038/nature13306]
- 19 Calvaresi EC, Hergenrother PJ. Glucose conjugation for the specific targeting and treatment of cancer. Chem Sci 2013; 4: 2319-2333 [PMID: 24077675 DOI: 10.1039/C3SC22205E]
- 20 Coulibaly FS, Youan BC. Current Status of Lectin-Based Cancer Diagnosis and Therapy. AIMS Mol Sci 2017; 4: 1–27 [DOI: 10.3934/molsci.2017.1.1]
- 21 Cash HL, Whitham CV, Behrendt CL, Hooper LV. Symbiotic bacteria direct expression of an intestinal bactericidal lectin. *Science* 2006; 313: 1126-1130 [PMID: 16931762 DOI: 10.1126/science.1127119]
- 22 Thomas GB, Rader LH, Park J, Abezgauz L, Danino D, DeShong P, English DS. Carbohydrate modified catanionic vesicles: probing multivalent binding at the bilayer interface. J Am Chem Soc 2009; 131: 5471-5477 [PMID: 19323555 DOI: 10.1021/ja8076439]
- 23 Seeberger PH, Werz DB. Synthesis and medical applications of oligosaccharides. *Nature* 2007; 446: 1046-1051 [PMID: 17460666 DOI: 10.1038/nature05819]
- 24 Sharon N, Lis H. History of lectins: from hemagglutinins to biological recognition molecules. *Glycobiology* 2004; 14: 53R-62R [PMID: 15229195 DOI: 10.1093/glycob/cwh122]
- 25 Shahzad-ul-Hussan S, Cai M, Bewley CA. Unprecedented glycosidase activity at a lectin carbohydratebinding site exemplified by the cyanobacterial lectin MVL. J Am Chem Soc 2009; 131: 16500-16508 [PMID: 19856962 DOI: 10.1021/ja905929c]
- 26 Stowell SR, Arthur CM, Dias-Baruffi M, Rodrigues LC, Gourdine JP, Heimburg-Molinaro J, Ju T, Molinaro RJ, Rivera-Marrero C, Xia B, Smith DF, Cummings RD. Innate immune lectins kill bacteria expressing blood group antigen. *Nat Med* 2010; 16: 295-301 [PMID: 20154696 DOI: 10.1038/nm.2103]
- 27 Lerchen HG, Baumgarten J, Piel N, Kolb-Bachofen V. Lectin-Mediated Drug Targeting: Discrimination of Carbohydrate-Mediated Cellular Uptake between Tumor and Liver Cells with Neoglycoconjugates Carrying Fucose Epitopes Regioselectively Modified in the 3-Position. *Angew Chem Int Ed Engl* 1999; 38: 3680-3683 [PMID: 10649323 DOI:

10.1002/(SICI)1521-3773(19991216)38:24<3680::AID-ANIE3680>3.0.CO;2-9

- 28 Lis H, Sharon N. Lectins: Carbohydrate-Specific Proteins That Mediate Cellular Recognition. *Chem Rev* 1998; 98: 637-674 [PMID: 11848911 DOI: 10.1021/cr940413g]
- 29 Xu L, Liu X, Li Y, Yin Z, Jin L, Lu L, Qu J, Xiao M. Enzymatic rhamnosylation of anticancer drugs by an α-L-rhamnosidase from Alternaria sp. L1 for cancer-targeting and enzyme-activated prodrug therapy. *Appl Microbiol Biotechnol* 2019; 103: 7997-8008 [PMID: 31414160 DOI: 10.1007/s00253-019-10011-0]
- 30 Wang Y, Gao J, Gu G, Li G, Cui C, Sun B, Lou H. In situ RBL receptor visualization and its mediated anticancer activity for solasodine rhamnosides. *Chembiochem* 2011; 12: 2418-2420 [PMID: 21953983 DOI: 10.1002/cbic.201100551]
- 31 Stüben J, Port R, Bertram B, Bollow U, Hull WE, Schaper M, Pohl J, Wiessler M. Pharmacokinetics and whole-body distribution of the new chemotherapeutic agent beta-D-glucosylisophosphoramide mustard and its effects on the incorporation of [methyl-3H]-thymidine in various tissues of the rat. *Cancer Chemother Pharmacol* 1996; 38: 355-365 [PMID: 8674159 DOI: 10.1007/s002800050495]
- 32 Pohl J, Bertram B, Hilgard P, Nowrousian MR, Stüben J, Wiessler M. D-19575--a sugar-linked isophosphoramide mustard derivative exploiting transmembrane glucose transport. *Cancer Chemother Pharmacol* 1995; 35: 364-370 [PMID: 7850916 DOI: 10.1007/s002800050248]
- 33 Stenn KS. Metastasizing neoplastic cells. Am J Dermatopathol 1979; 1: 375-376 [PMID: 543531 DOI:

10.1097/00000372-197900140-00017]

- 34 Briasoulis E, Judson I, Pavlidis N, Beale P, Wanders J, Groot Y, Veerman G, Schuessler M, Niebch G, Siamopoulos K, Tzamakou E, Rammou D, Wolf L, Walker R, Hanauske A. Phase I trial of 6-hour infusion of glufosfamide, a new alkylating agent with potentially enhanced selectivity for tumors that overexpress transmembrane glucose transporters: a study of the European Organization for Research and Treatment of Cancer Early Clinical Studies Group. *J Clin Oncol* 2000; 18: 3535-3544 [PMID: 11032596 DOI: 10.1200/JCO.2000.18.20.3535]
- 35 Chiorean EG, Dragovich T, Hamm J, Barrios CH, Gorini CF, Langmuir VK, Kroll S, Jung DT, Tidmarsh GT, Loehrer PJ. A phase 2 trial of glufosfamide in combination with gemcitabine in chemotherapy-naive pancreatic adenocarcinoma. *Am J Clin Oncol* 2010; **33**: 111-116 [PMID: 19687729 DOI: 10.1097/COC.0b013e3181979204]
- 36 Hartley JA, Bingham JP, Souhami RL. DNA sequence selectivity of guanine-N7 alkylation by nitrogen mustards is preserved in intact cells. *Nucleic Acids Res* 1992; 20: 3175-3178 [PMID: 1620613 DOI: 10.1093/nar/20.12.3175]
- 37 Zhang HJ, Jinks RN, Wishart AC, Battelle BA, Chamberlain SC, Fahrenbach WH, Kass L. An enzymatically enhanced recording technique for Limulus ventral photoreceptors: physiology, biochemistry, and morphology. *Vis Neurosci* 1994; 11: 41-52 [PMID: 8011582 DOI: 10.1017/s0952523800011093]
- 38 Goff RD, Thorson JS. Assessment of chemoselective neoglycosylation methods using chlorambucil as a model. J Med Chem 2010; 53: 8129-8139 [PMID: 20973561 DOI: 10.1021/jm101024j]
- 39 Neckers L. Hsp90 inhibitors as novel cancer chemotherapeutic agents. Trends Mol Med 2002; 8: S55-S61 [PMID: 11927289 DOI: 10.1016/s1471-4914(02)02316-x]
- 40 Ferrarini M, Heltai S, Zocchi MR, Rugarli C. Unusual expression and localization of heat-shock proteins in human tumor cells. Int J Cancer 1992; 51: 613-619 [PMID: 1601523 DOI: 10.1002/ijc.2910510418]
- 41 Supko JG, Hickman RL, Grever MR, Malspeis L. Preclinical pharmacologic evaluation of geldanamycin as an antitumor agent. *Cancer Chemother Pharmacol* 1995; 36: 305-315 [PMID: 7628050 DOI: 10.1007/BF00689048]
- 42 Sausville EA. Combining cytotoxics and 17-allylamino, 17-demethoxygeldanamycin: sequence and tumor biology matters. Commentary re: P. Münster *et al.*, Modulation of Hsp90 function by ansamycins sensitizes breast cancer cells to chemotherapy-induced apoptosis in an RB- and schedule-dependent manner. Clin. Cancer Res., 7: 2228-2236, 2001. *Clin Cancer Res* 2001; 7: 2155-2158 [PMID: 11489788]
- 43 Cheng H, Cao X, Xian M, Fang L, Cai TB, Ji JJ, Tunac JB, Sun D, Wang PG. Synthesis and enzymespecific activation of carbohydrate-geldanamycin conjugates with potent anticancer activity. *J Med Chem* 2005; 48: 645-652 [PMID: 15658879 DOI: 10.1021/jm049693a]
- 44 Chen YC, Shen SC, Lee WR, Hsu FL, Lin HY, Ko CH, Tseng SW. Emodin induces apoptosis in human promyeloleukemic HL-60 cells accompanied by activation of caspase 3 cascade but independent of reactive oxygen species production. *Biochem Pharmacol* 2002; 64: 1713-1724 [PMID: 12445860 DOI: 10.1016/s0006-2952(02)01386-2]
- 45 Su YT, Chang HL, Shyue SK, Hsu SL. Emodin induces apoptosis in human lung adenocarcinoma cells through a reactive oxygen species-dependent mitochondrial signaling pathway. *Biochem Pharmacol* 2005; 70: 229-241 [PMID: 15941563 DOI: 10.1016/j.bcp.2005.04.026]
- 46 Srinivas G, Anto RJ, Srinivas P, Vidhyalakshmi S, Senan VP, Karunagaran D. Emodin induces apoptosis of human cervical cancer cells through poly(ADP-ribose) polymerase cleavage and activation of caspase-9. *Eur J Pharmacol* 2003; **473**: 117-125 [PMID: 12892828 DOI: 10.1016/s0014-2999(03)01976-9]
- 47 Huang Q, Shen HM, Shui G, Wenk MR, Ong CN. Emodin inhibits tumor cell adhesion through disruption of the membrane lipid Raft-associated integrin signaling pathway. *Cancer Res* 2006; 66: 5807-5815 [PMID: 16740720 DOI: 10.1158/0008-5472.CAN-06-0077]
- 48 Kwak HJ, Park MJ, Park CM, Moon SI, Yoo DH, Lee HC, Lee SH, Kim MS, Lee HW, Shin WS, Park IC, Rhee CH, Hong SI. Emodin inhibits vascular endothelial growth factor-A-induced angiogenesis by blocking receptor-2 (KDR/Flk-1) phosphorylation. *Int J Cancer* 2006; **118**: 2711-2720 [PMID: 16388516 DOI: 10.1002/ijc.21641]
- 49 Cha TL, Qiu L, Chen CT, Wen Y, Hung MC. Emodin down-regulates androgen receptor and inhibits prostate cancer cell growth. *Cancer Res* 2005; 65: 2287-2295 [PMID: 15781642 DOI: 10.1158/0008-5472.CAN-04-3250]
- 50 Xing JY, Song GP, Deng JP, Jiang LZ, Xiong P, Yang BJ, Liu SS. Antitumor effects and mechanism of novel emodin rhamnoside derivatives against human cancer cells in vitro. *PloS One* 2015; 10: e0144781 [PMID: 26682731 DOI: 10.1371/journal.pone.0144781]
- 51 The WHOQOL Group. The World Health Organization quality of life assessment (WHOQOL): position paper from the World Health Organization. Soc Sci Med 1995; 41: 1403-1409 [PMID: 8560308 DOI: 10.1016/0277-9536(95)00112-K]
- 52 Mahato R, Tai W, Cheng K. Prodrugs for improving tumor targetability and efficiency. Adv Drug Deliv Rev 2011; 63: 659-670 [PMID: 21333700 DOI: 10.1016/j.addr.2011.02.002]
- 53 de Bono JS, Ashworth A. Translating cancer research into targeted therapeutics. *Nature* 2010; 467: 543-549 [PMID: 20882008 DOI: 10.1038/nature09339]
- 54 Ma Y, Chen H, Su S, Wang T, Zhang C, Fida G, Cui S, Zhao J, Gu Y. Galactose as Broad Ligand for Multiple Tumor Imaging and Therapy. *J Cancer* 2015; 6: 658-670 [PMID: 26078797 DOI: 10.7150/jca.11647]
- 55 Cao J, Cui S, Li S, Du C, Tian J, Wan S, Qian Z, Gu Y, Chen WR, Wang G. Targeted cancer therapy with a 2-deoxyglucose-based adriamycin complex. *Cancer Res* 2013; 73: 1362-1373 [PMID: 23396585 DOI: 10.1158/0008-5472.CAN-12-2072]
- 56 Rooseboom M, Commandeur JN, Vermeulen NP. Enzyme-catalyzed activation of anticancer prodrugs. Pharmacol Rev 2004; 56: 53-102 [PMID: 15001663 DOI: 10.1124/pr.56.1.3]
- 57 Abraham S, Guo F, Li LS, Rader C, Liu C, Barbas CF, Lerner RA, Sinha SC. Synthesis of the nextgeneration therapeutic antibodies that combine cell targeting and antibody-catalyzed prodrug activation. *Proc Natl Acad Sci USA* 2007; 104: 5584-5589 [PMID: 17372220 DOI: 10.1073/pnas.0700223104]
- 58 Chatterjee SK, Zetter BR. Cancer biomarkers: knowing the present and predicting the future. *Future* Oncol 2005; 1: 37-50 [PMID: 16555974 DOI: 10.1517/14796694.1.1.37]
- 59 de Graaf M, Boven E, Scheeren HW, Haisma HJ, Pinedo HM. Beta-glucuronidase-mediated drug release. Curr Pharm Des 2002; 8: 1391-1403 [PMID: 12052215 DOI: 10.2174/1381612023394485]
- Wu J, Liu Q, Lee RJ. A folate receptor-targeted liposomal formulation for paclitaxel. *Int J Pharm* 2006;
 316: 148-153 [PMID: 16564654 DOI: 10.1016/j.ijpharm.2006.02.027]

- 61 Lin YS, Tungpradit R, Sinchaikul S, An FM, Liu DZ, Phutrakul S, Chen ST. Targeting the delivery of glycan-based paclitaxel prodrugs to cancer cells via glucose transporters. *J Med Chem* 2008; **51**: 7428-7441 [PMID: 19053781 DOI: 10.1021/jm8006257]
- 62 Longley DB, Harkin DP, Johnston PG. 5-fluorouracil: mechanisms of action and clinical strategies. Nat Rev Cancer 2003; 3: 330-338 [PMID: 12724731 DOI: 10.1038/nrc1074]
- 63 Johnston PG, Kaye S. Capecitabine: a novel agent for the treatment of solid tumors. *Anticancer Drugs* 2001; 12: 639-646 [PMID: 11604550 DOI: 10.1097/00001813-200109000-00001]
- 64 Feng D, Shaikh AS, Wang F. Recent Advance in Tumor-associated Carbohydrate Antigens (TACAs)based Antitumor Vaccines. ACS Chem Biol 2016; 11: 850-863 [PMID: 26895482 DOI: 10.1021/acschembio.6b00084]
- 65 Hollingsworth RE, Jansen K. Turning the corner on therapeutic cancer vaccines. NPJ Vaccines 2019; 4: 7 [PMID: 30774998 DOI: 10.1038/s41541-019-0103-y]
- 66 Carrascal MA, Severino PF, Guadalupe Cabral M, Silva M, Ferreira JA, Calais F, Quinto H, Pen C, Ligeiro D, Santos LL, Dall'Olio F, Videira PA. Sialyl Tn-expressing bladder cancer cells induce a tolerogenic phenotype in innate and adaptive immune cells. *Mol Oncol* 2014; 8: 753-765 [PMID: 24656965 DOI: 10.1016/j.molonc.2014.02.008]
- 67 Pashov A, Monzavi-Karbassi B, Kieber-Emmons T. Immune surveillance and immunotherapy: lessons from carbohydrate mimotopes. *Vaccine* 2009; 27: 3405-3415 [PMID: 19200843 DOI: 10.1016/j.vaccine.2009.01.074]
- 68 Pashov A, Monzavi-Karbassi B, Raghava GP, Kieber-Emmons T. Bridging innate and adaptive antitumor immunity targeting glycans. *J Biomed Biotechnol* 2010; 2010: 354068 [PMID: 20617150 DOI: 10.1155/2010/354068]
- 69 Pinho SS, Reis CA. Glycosylation in cancer: mechanisms and clinical implications. *Nat Rev Cancer* 2015; 15: 540-555 [PMID: 26289314 DOI: 10.1038/nrc3982]
- 70 Ho WL, Hsu WM, Huang MC, Kadomatsu K, Nakagawara A. Protein glycosylation in cancers and its potential therapeutic applications in neuroblastoma. *J Hematol Oncol* 2016; 9: 100 [PMID: 27686492 DOI: 10.1186/s13045-016-0334-6]
- 71 Sørensen AL, Reis CA, Tarp MA, Mandel U, Ramachandran K, Sankaranarayanan V, Schwientek T, Graham R, Taylor-Papadimitriou J, Hollingsworth MA, Burchell J, Clausen H. Chemoenzymatically synthesized multimeric Tn/STn MUC1 glycopeptides elicit cancer-specific anti-MUC1 antibody responses and override tolerance. *Glycobiology* 2006; 16: 96-107 [PMID: 16207894 DOI: 10.1093/glycob/cwj044]
- 72 Tarp MA, Sørensen AL, Mandel U, Paulsen H, Burchell J, Taylor-Papadimitriou J, Clausen H. Identification of a novel cancer-specific immunodominant glycopeptide epitope in the MUC1 tandem repeat. *Glycobiology* 2007; 17: 197-209 [PMID: 17050588 DOI: 10.1093/glycob/cwl061]
- 73 Ryan SO, Vlad AM, Islam K, Gariépy J, Finn OJ. Tumor-associated MUC1 glycopeptide epitopes are not subject to self-tolerance and improve responses to MUC1 peptide epitopes in MUC1 transgenic mice. *Biol Chem* 2009; **390**: 611-618 [PMID: 19426130 DOI: 10.1515/BC.2009.070]
- 74 Madsen CB, Petersen C, Lavrsen K, Harndahl M, Buus S, Clausen H, Pedersen AE, Wandall HH. Cancer associated aberrant protein O-glycosylation can modify antigen processing and immune response. *PLoS One* 2012; 7: e50139 [PMID: 23189185 DOI: 10.1371/journal.pone.0050139]
- 75 Li M, Yu F, Yao C, Wang PG, Liu Y, Zhao W. Synthetic and immunological studies on trimeric MUC1 immunodominant motif antigen-based anti-cancer vaccine candidates. *Org Biomol Chem* 2018; 16: 993-999 [PMID: 29345713 DOI: 10.1039/c7ob02976d]
- 76 Lakshminarayanan V, Thompson P, Wolfert MA, Buskas T, Bradley JM, Pathangey LB, Madsen CS, Cohen PA, Gendler SJ, Boons GJ. Immune recognition of tumor-associated mucin MUC1 is achieved by a fully synthetic aberrantly glycosylated MUC1 tripartite vaccine. *Proc Natl Acad Sci USA* 2012; 109: 261-266 [PMID: 22171012 DOI: 10.1073/pnas.1115166109]
- 77 Miles D, Papazisis K. Rationale for the clinical development of STn-KLH (Theratope) and anti-MUC-1 vaccines in breast cancer. *Clin Breast Cancer* 2003; 3 Suppl 4: S134-S138 [PMID: 12620150 DOI: 10.3816/cbc.2003.s.002]
- 78 Rodríguez E, Schetters STT, van Kooyk Y. The tumour glyco-code as a novel immune checkpoint for immunotherapy. Nat Rev Immunol 2018; 18: 204-211 [PMID: 29398707 DOI: 10.1038/nri.2018.3]
- 79 Altenberg B, Greulich KO. Genes of glycolysis are ubiquitously overexpressed in 24 cancer classes. Genomics 2004; 84: 1014-1020 [PMID: 15533718 DOI: 10.1016/j.ygeno.2004.08.010]
- 80 Medina RA, Owen GI. Glucose transporters: expression, regulation and cancer. *Biol Res* 2002; **35**: 9-26 [PMID: 12125211 DOI: 10.4067/s0716-97602002000100004]

