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Author(s)	Kobayashi, Sumitaka; Sata, Fumihiro; Ikeda-Araki, Atsuko; Miyashita, Chihiro; Minatoya, Machiko; Ikeno, Tamiko; Kato, Shizue; Fujikura, Kaori; Mizutani, Futoshi; Chisaki, Yoichi; Kishi, Reiko
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1	Relationships between maternal perfluoroalkyl substance levels, polymorphisms of receptor
2	genes, and adverse birth outcomes in the Hokkaido birth cohort study, Japan
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4	Sumitaka Kobayashi ¹ , Fumihiro Sata ^{1,2} , Atsuko Ikeda-Araki ^{1,3} , Chihiro Miyashita ¹ , Goudarzi
5	Houman ^{1,4} , Yusuke Iwasaki ⁵ , Tamie Nakajima ⁶ , Reiko Kishi ^{1,*}
6	
7	1. Center for Environmental and Health Sciences, Hokkaido University, North-12, West-7, Kita-
8	ku, Sapporo 060-0812, Japan
9	2. Health Center, Chuo University, 42-8, Ichigaya-Hommura-cho, Shinjuku-ku, Tokyo 162-8473,
10	Japan
11	3. Faculty of Health Sciences, Hokkaido University, North-12, West-5, Kita-ku, Sapporo 060-
12	0812, Japan
13	4. Department of Respiratory Medicine, Faculty of Medicine, Hokkaido University, North-15,
14	West-7, Kita-ku, Sapporo 060-8638, Japan
15	5. Department of Biopharmaceutics and Analytical Science, Hoshi University, 2-4-41, Ebara,
16	Shinagawa-ku, Tokyo 142-8501, Japan
17	6. College of Life and Health Sciences, Chubu University, 1200, Matsumoto-cho, Kasugai 487-

2	* Corresponding author
3	Reiko Kishi, PhD, MD, MPH
4	Center for Environmental and Health Sciences, Hokkaido University
5	North-12, West-7, Kita-ku, Sapporo 060-0812, Japan
6	E-mail: rkishi@med.hokudai.ac.jp
7	Tel: +81-(0)11-706-4746
8	Fax: +81-(0)11-706-4725
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10	Highlights
11	• Maternal perfluorooctanesulfonate levels were associated with birth weight.
12	• Gene-perfluorooctanesulfonate interaction were related to birth outcomes.
13	• Only female infants show gene-perfluorooctanesulfonate interaction.
14	• <i>LXRB</i> (rs1405655) affects birth weight, chest circumference, and ponderal index.
15	
16	Abstract
17	We assessed the associations between perfluorooctanesulfonate (PFOS) and perfluorooctanoate
18	(PFOA) levels in third trimester maternal serum, the maternal genotypes of genes encoding

1	nuclear receptors, and birth outcomes. We studied a prospective birth cohort of healthy pregnant
2	Japanese women ($n = 372$) recruited in Sapporo between July 2002 and October 2005. We
3	analyzed PFOS and PFOA levels using liquid chromatography-tandem mass spectrometry and
4	analyzed 13 single nucleotide polymorphisms (SNPs) of proliferator-activated receptor alpha,
5	gamma, gamma coactivator 1A, delta, constitutive androstane receptor, liver X receptor alpha,
6	and beta (LXRB) using real-time polymerase reaction (PCR). We employed multiple linear
7	regression models to establish the influences of log10-transformed PFOS and PFOA levels and
8	maternal genotypes on birth size. In female infants, we identified interactions between PFOS
9	levels, the maternal genotype of LXRB (rs1405655), and birth weight. The estimated mean
10	changes in birth weight in response to PFOS levels, the maternal genotype LXRB (rs1405655)-
11	TC/CC (compared to TT), and their interactions were -502.9 g (95% confidence interval $[CI] =$
12	-247.3, -758.5 g), -526.3 g (95% CI = -200.7, -852.0 g), and 662.1 g (95% CI = 221.0, 1,103.2
13	g; $p_{int} = 0.003$), respectively. Interactions between PFOS levels and the maternal genotype of
14	LXRB (rs1405655) also significantly affected birth chest circumference and the Ponderal index
15	($p_{int} = 0.037$ and 0.005, respectively). Thus, interactions between PFOS levels and the maternal
16	genotype of LXRB (rs1405655) affects birth sizes in female infants. We found that certain SNPs
17	modify the effects of PFOS levels on birth size.

1 Keywords

2 Perfluoroalkyl substance; Pregnancy; Polymorphism; Liver X receptor; Birth size; Ponderal index

3

4 Abbreviations

- 5 BCC, birth chest circumference
- 6 BHC, birth head circumference
- 7 BL, birth length
- 8 BMI, body mass index
- 9 BPI, birth Ponderal index

10 BW, birth weight

- 11 CAR, constitutive androstane receptor
- 12 CI, confidence interval
- 13 LOD, limit of detection
- 14 LXR, liver X receptor
- 15 LXRA, liver X receptor alpha
- 16 LXRB, liver X receptor beta
- 17 PFAS, perfluoroalkyl substance
- 18 PFOA, perfluorooctanoate

1	PFOS,	perfluorooctanesulfonate
_		

- 2 PPAR, peroxisome proliferator-activated receptor
- 3 PPARA, peroxisome proliferator-activated receptor alpha
- 4 PPARD, peroxisome proliferator-activated receptor delta
- 5 PPARG, peroxisome proliferator-activated receptor gamma
- 6 PPARGC1A, peroxisome proliferator-activated receptor gamma coactivator 1 alpha
- 7 SNP, single nucleotide polymorphism
- 8
- 9

10 **1. Introduction**

- Perfluoroalkyl substances (PFASs) have been manufactured for more than 60 years, and have
 been used in a wide range of industrial and consumer products; specifically, they are used as
- 13 processing aids in impregnation agents during the production of paper and carpets. Humans are
- 14 commonly exposed to PFASs through food. The trans-placental transfer rates of
- 15 perfluorooctanoate (PFOA) and perfluorooctanesulfonate (PFOS), which are representative
- 16 PFASs, have been determined to be 0.73 and 0.42 %, respectively [1]. It has therefore been
- 17 suggested that PFOS and PFOA undergo maternal-to-fetal transfer. PFOS and PFOA have
- 18 recently been found to lead to a reduction in birth size [2-19]. Currently, however, the biological

2	restriction are not clearly understood.
3	Genetic factors may be key to elucidating the underlying biological mechanisms between
4	these relationships. The heritability of birth weight (BW) is moderate, with estimates of 38% from
5	studies on twins. This suggests that both genetic and environmental factors affect the etiology of
6	birth weight [20]. The "Developmental origins of health and disease (DOHaD)" theory suggests
7	that fetal conditions not only affect birth characteristics such as BW, birth length (BL), birth chest
8	circumference (BCC), birth head circumference (BHC), and birth Ponderal index (BPI), but also
9	have lifelong health implications [21,22]. Despite widespread interest in this hypothesis, only
10	limited advances have been made regarding fetal gene-environment interactions with respect to
11	chemical exposure during pregnancy and maternal genetic factors. Therefore, the gene-
12	environment interaction studies are necessary to improve the current understanding of fetal
13	conditions.
14	Future adverse health effects have recently been partially revealed by five investigations
15	into interactions between the genotypes of specific biologically relevant genes and PFOS and
16	PFOA levels [23-27]. Of these five studies, four investigated the interactions between specific
17	genetic susceptibility-conferring genes and prenatal PFOS and PFOA levels in relation to changes
18	in maternal lipid levels [23], cord sex hormone levels [24], reductions in BW [25], and the risk of

mechanism underlying the association between PFOS and PFOA levels and fetal growth

1	atopic dermatitis at 2 years of age [27]. Maternal genotypes of the receptor genes peroxisome
2	proliferator-activated receptor (PPAR) gamma coactivator 1 alpha (PPARGC1A) and PPAR delta
3	(PPARD) have been revealed to crosstalk with PFOS levels during pregnancy. Specifically, they
4	were found to reduce maternal lipid levels [23]. Furthermore, the fetal genotypes of cytochrome
5	P450 17A1 (CYP17A1) have been shown to crosstalk with PFOS levels during pregnancy by
6	changing cord sex hormone levels [24]. In addition, the maternal genotypes of the xenobiotic
7	metabolizing genes cytochrome P450 1A1 (CYP1A1) and glutathione S-transferase mu 1
8	(GSTM1) have been revealed to interact with PFOS and PFOA during pregnancy, augmenting the
9	reduction in BW [25] and the risk of dermatitis at 2 years of age [27]. Previous studies have also
10	suggested that PFOS and PFOA during pregnancy act on the developing fetus, thus affecting fetal
11	growth and child diseases after birth, and that differences in maternal genetic susceptibility can
12	affect fetal their growth and child diseases.
13	PFOS and PFOA are known to change the induction levels of PPAR alpha (PPARA), PPAR
14	gamma (PPARG), PPAR delta (PPARD), constitutive androstane receptor (CAR), liver X receptor
15	(LXR) alpha (LXRA), and LXR beta (LXRB) [28-30]. However, to date no interactions between
16	maternal PFOS and PFOA and the genetic polymorphisms of their receptors have been reported
17	with respect to birth size. As PFOS and PFOA bind to receptors and elicit biological responses,
18	we hypothesize that the maternal genotypes of receptor genes may modify the associations

1	between PFOS and PFOA levels during pregnancy and decreased birth size. Sex-specific
2	associations between PFOS and PFOA levels during pregnancy and reductions in birth size have
3	been reported [17], as have the associations between PFOS levels during pregnancy and the
4	maternal genotypes of PPARGC1A and PPARD with respect to maternal lipid levels [23]. The
5	present study aimed to expand these studies to examine the associations between PFOS and PFOA
6	levels during pregnancy and the maternal genotypes of the receptor genes of PPARA, PPARG,
7	PPARGC1A, PPARD, CAR, LXRA, and LXRB regarding their effects on birth outcomes (BW, BL,
8	BCC, BHC, and BPI). This study also aimed to investigate the sex differences with respect to the
9	effects of associations on birth outcomes.
10	
11	2. Methods

12 2.1. Study participants

This prospective birth cohort study was based on the Hokkaido Study on Environment and Children's Health (Sapporo cohort). The study protocol has been described previously [31,32]. Briefly, from July 2002 to October 2005, pregnant Japanese women (n = 514) were recruited from a local obstetrics and gynecology hospital in Sapporo City. Of these, ten participants withdrew from the study. Of the remaining 504, after excluding mothers with pregnancy-induced hypertension and gestational diabetes mellitus as exclusion criteria, 372 participants were included in the current investigation; complete data on the levels of PFOS, PFOA, and fatty acids,
 maternal genotypes, and birth size were available for each participant.

3

4 2.2. Data collection

Each participant completed a self-administered questionnaire upon enrollment regarding their maternal age, height before pregnancy, weight before pregnancy, annual household income, maternal educational level, maternal smoking in the third trimester, and maternal alcohol consumption during pregnancy. Maternal records were also obtained to collect information on parity, gestational age, infant sex, BW, BL, BCC, and BHC. The pre-pregnancy body mass index (BMI) was defined as maternal weight before pregnancy over the square of maternal height before pregnancy (kg/m²). BPI was defined as BW divided by the cube of BL (kg/m³).

12

13 **2.3.** Measurement of PFOS and PFOA in maternal serum

For each participant, a 40 mL blood sample was taken from the maternal peripheral vein in the 3rd trimester. All samples were stored at -80 °C until analysis. In 447 maternal blood samples, PFOS and PFOA levels were measured using liquid chromatography-tandem mass spectrometry (LC/MS/MS). The measurement protocol has been described previously [23,24,33]. Samples from other participants were not analyzed, either because they were not available or because the

1	sample volume was insufficient. Of the 447 blood samples, 228 were collected during pregnancy
2	and 159 were obtained after delivery (owing to anemia during pregnancy). For all the samples,
3	the limit of detection (LOD; 0.50 ng/mL) for PFOS was excessed. However, 16 (5.9%) samples
4	had PFOA levels below the LOD (0.50 ng/mL); these were assigned a value of 0.25 ng/mL (50%
5	of LOD).

7 2.4. Maternal genotype analysis

We analyzed the genotypes of the 494 maternal blood samples. The remaining samples were not 8 9 analyzed because they were not available or because there was insufficient blood volume for 10 measurement. Maternal blood samples were collected when participants gave birth; 400 µL of each sample was used to isolate and purify genomic DNA using a QIAamp DNA Blood Mini Kit 11 (Qiagen GmbH, Hilden, Germany) or a Maxwell 16 DNA Purification Kit (Promega, Madison, 1213WI, USA), according to the manufacturer's instructions [34]. We evaluated 13 genetic polymorphisms, namely those in PPARA (rs1800234 and rs135561), PPARG (rs3856806), 14PPARGC1A (rs2970847 and rs8192678), PPARD (rs1053049 and rs2267668), CAR (rs2307424 1516 and rs2501873), LXRA (rs2279238), and LXRB (rs1405655, rs2303044, and rs4802703). 17When selecting genetic polymorphisms, we examined three genes, i.e., PPARs, CAR, and

18 LXRs, from the orphan receptors that were expected to be activated by PFASs and affected by

1	fatty acid levels. Next, using the single nucleotide polymorphism (SNP) database (dbSNP), we
2	selected the following 13 genetic polymorphisms, which have been reported to be located in
3	potentially functional regions (mainly promotor and coding regions), and to be associated with
4	disease susceptibilities to cancer, non-alcohol fatty acid disease, type 2 diabetes mellitus, obesity,
5	and other diseases: <i>PPARA</i> (T>C, Val227Ala; dbSNP ID: rs1800234; G>A, dbSNP ID: rs135561),
6	PPARD (T>C, dbSNP ID: rs1053049; A>G, dbSNP ID: rs2267668), PPARG (C>T, His449His;
7	dbSNP ID: rs3856806), <i>PPARGC1A</i> (C>T, Thr394Thr; dbSNP ID: rs2970847; G>A, dbSNP ID:
8	rs8192678), <i>CAR</i> (T>C, Pro180Pro; dbSNP ID: rs2307424; A>G, dbSNP ID: rs2501873), <i>LXRA</i>
9	(C>T, Ser99Ser; dbSNP ID: rs2279238), and LXRB (T>C, dbSNP ID: rs1405655; G>A, dbSNP
10	ID: rs2303044; G>A; dbSNP ID: rs4802703) [35-45]. Approximately 5 % or more of the minor
11	alleles among pregnant Japanese women are necessary to secure statistical power when examining
12	adverse health outcomes. All 13 genetic polymorphisms satisfied a minor allele frequency of \geq
13	5 %. These polymorphisms were analyzed using high-throughput gene expression of pre-
14	amplification, real-time polymerase chain reaction (PCR) with dynamic chips, and TaqMan gene
15	expression measurements. The assessment protocol has been described previously [23].
16	

17 2.5. Fatty acid measurement in maternal serum

18 For each participant, a 40 mL blood sample was taken from the maternal peripheral vein in the

1	third trimester. All samples were stored at -80 °C until analysis. In 491 maternal blood samples,
2	fatty acid levels were measured using gas chromatography-mass spectrometry (GC/MS) as
3	described previously [23,46]. Samples for other participants were not analyzed either because
4	they were not available or because the sample volume was insufficient. Of the 491 blood samples
5	analyzed, 307 were collected during pregnancy and 184 were obtained after delivery (owing to
6	anemia during pregnancy). The nine fatty acids targeted for measurement were palmitic acid,
7	palmitoleic acid, stearic acid, oleic acid, linoleic acid, α -linolenic acid, arachidonic acid,
8	eicosapentaenoic acid, and docosahexaenoic acid. The detection limits were 2.4 $\mu\text{g/mL}$ for
9	palmitic acid, 0.069 μ g/mL for palmitoleic acid, 1.3 μ g/mL for stearic acid, 3.6 μ g/mL for oleic
10	acid, and 2.0 μ g/mL for the other fatty acids. The detection rates for all fatty acids were more than
11	99.0 % except for eicosapentaenoic acid (97.8 %). Non-fasting blood triglyceride levels were
12	measured using triglyceride E-Test Wako Kits (Wako, Osaka, Japan) after lipid extraction,
13	according to the methods described by Folch et al. [47].

15 2.6. Statistical analyses

16 Of the 372 participants, one (0.3%), three (0.8%), and six (1.6%) had missing data on parity, 17 educational level, and annual household income, respectively. Using simple imputation, these 18 participants were assigned to the multiparous parity group, to an educational level group with

1	more than high school graduation, and to an annual household income group of more than 5
2	million Japanese yen (the most frequent group). First, we examined the characteristics of the
3	participants. A chi-squared test, independent <i>t</i> -test, and a Mann-Whitney's U-test were employed
4	to test whether there were differences between the male and female groups. Then, a chi-squared
5	test was used to determine whether the frequency of genotype distribution conformed to the
6	Hardy-Weinberg equilibrium (HWE). Third, PFOS, PFOA and fatty acid levels were log10-
7	transformed before the following analyses, because of their non-normal distribution. Birth
8	outcomes were defined using BW, BL, BCC, BHC, and BPI. Multiple linear regression analyses
9	were used to evaluate the associations between PFAS levels and birth outcomes in both the crude
10	and adjusted models. Infant sex was also subjected to stratified analysis. Maternal age
11	(continuous), pre-pregnancy BMI (continuous), maternal smoking in the third trimester (yes/no),
12	maternal alcohol consumption during pregnancy (yes/no), parity (primiparous/multiparous),
13	educational level (less than or equal to high school graduation/more than high school graduation),
14	annual household income (less than 5 million Japanese yen/more than or equal to 5 million
15	Japanese yen), cesarean section (yes/no), maternal blood sampling periods (during pregnancy or
16	after delivery), gestational age (continuous), and infant sex (male/female; total infants only) were
17	adjusted in the multiple linear regression models (except for in the crude model). Multiple linear
18	regression analyses were then performed to evaluate the interactions between infant sex and PFAS

1	levels regarding birth outcomes, for both the crude and adjusted models. The covariates were the
2	same as those used in the fourth analysis, except for the infant sex. Multiple linear regression
3	analyses were then conducted again to evaluate the association between PFOS or PFOA levels
4	and maternal genotypes, and the associations between birth outcomes and maternal genotypes
5	(for both the crude and adjusted models); infant sex was also subjected to a stratified analysis.
6	The covariates were the same as those used in the fourth analysis. Multiple linear regression
7	analyses were also used to evaluate the interactions between PFOS or PFOA levels and maternal
8	genotypes on birth outcomes, in both the crude and adjusted models; infant sex was subjected to
9	a stratified analysis and the covariates were the same as those used in the fourth analysis. Multiple
10	linear regression analyses were then used to evaluate the association between PFAS levels and
11	birth outcomes in both the crude and adjusted models after stratification, based on the maternal
12	genotypes of LXRB (rs1405655). This was done because only the effects of the interactions
13	between PFOS levels and the maternal genotype of LXRB (rs1405655) on BW, BCC, and BPI in
14	female infants were statistically significant. Infant sex was also subjected to a stratified analysis.
15	The covariates were the same as those used in the fourth analysis. Multiple linear regression
16	models were then used to evaluate the interactions between PFOS levels and the maternal
17	genotype of LXRB (rs1405655) regarding fatty acid levels. Infant sex was also subjected to a
18	stratified analysis. The covariates, except for cesarean section, gestational age, and infant sex,

1	were the same as those used in the fourth analysis. In addition, PFOS levels were divided into
2	quartile levels before the following analyses, because of their non-normal distribution. Multiple
3	linear regression analyses were used to evaluate the dose-dependent associations between PFOS
4	levels and BW, BCC, and BPI, in both the crude and adjusted models. This was done after
5	stratification based on the maternal genotype of LXRB (rs1405655) because only the effects of
6	the interactions between PFOS levels and the maternal genotype of LXRB (rs1405655) on BW,
7	BCC, and BPI were statistically significant. Infant sex was also subjected to a stratified analysis;
8	the covariates were the same as those used in the fourth analysis.
9	The PFAS-sex interaction term was defined as "log10-transformed PFOS or PFOA levels
10	(continuous) \times sex (0 = males and 1 = females)." The PFOS-gene or PFOA-gene interaction term
11	was defined as "log ₁₀ -transformed PFOS or PFOA levels (continuous) \times genotype (0 = referent
12	genotype and 1 = genotype to be compared)." The interaction term was included in the multiple
13	linear regression models except in stratified analysis. Furthermore, we refuted or confirmed the
14	validity of the results for all participants using sensitivity analyses. We achieved this by restricting
15	participants to those whose maternal blood samples were collected during pregnancy (i.e., before
16	delivery). In the sensitivity analyses, multiple linear regressions were performed using the same
17	covariates for all participants.



- 1 performed using SPSS software version 26 (IBM Corp., Armonk, NY, USA).
- $\mathbf{2}$

3	2.7. Ethics
4	Written informed consent was obtained from all participants. All procedures were conducted in
5	accordance with the principles of the Declaration of Helsinki. The study protocol was approved
6	by the Institutional Ethical Board for Human Gene and Genome Studies and the Epidemiological
7	Studies Programs of the Hokkaido University Center for Environmental and Health Sciences
8	(approval number: 21-137; approval date: September 3 rd , 2021).
9	
10	3. Results
11	The characteristics of this study are summarized in Table 1. The mean maternal age (30.8 vs. 29.7
12	years; $p = 0.020$), median PFOA levels in maternal blood (1.4 vs. 1.2 ng/mL; $p = 0.041$), mean
13	BW (3,098.3 vs 2,997.8 g; $p = 0.012$), mean BL (48.3 vs. 47.7 cm; $p = 0.004$), and BHC (33.6 vs.
14	32.8 cm; $p < 0.001$) were all significantly lower in females than in males, respectively.
15	The maternal genotype frequencies are summarized in Table 2. All genotypes conformed to
16	Hardy-Weinberg equilibrium ($p > 0.05$, for all).
17	The associations between PFOS and PFOA levels in maternal blood and birth outcomes are
18	shown in Table 3. Multiple linear regression analysis showed that maternal PFOS levels during

1	pregnancy were associated with lower BW (mean reduction = 182.3 g [95% confidence interval
2	(CI) = 28.2, 336.5]) for all infants, and lower BW (mean reduction = 292.1 g [95% CI = 79.8,
3	504.3]) and lower BL (mean reduction = 1.384 cm [95% CI = 0.297 , 2.472]) in female infants.
4	Furthermore, maternal PFOA levels during pregnancy were associated with lower BW (mean
5	reduction = 183.0 g [95% CI = 4.1, 361.9]) in female infants, after adjusting for the covariates.
6	These results were almost the same as the results of the sensitivity analysis (Supplementary Table
7	1). However, no interactions were detected between PFOS levels in maternal blood and fetal sex
8	on birth outcomes, and between PFOA levels in maternal blood and fetal sex on birth outcomes
9	(Supplementary Table 2).
10	The interactions between PFOS and PFOA levels in maternal blood and the maternal genotype
11	of LXRB (rs1405655) on birth size in female infants are summarized in Table 4 (also see Fig. 1.).
12	Regarding the interaction between PFOS levels in maternal blood and the maternal genotype of
10	
13	LXRB (rs1405655) on BW (after adjusting for the covariates), the mean estimated decrease (95%
13 14	<i>LXRB</i> (rs1405655) on BW (after adjusting for the covariates), the mean estimated decrease (95% CI) in BW for a one-unit increase in PFOS levels in maternal blood was 502.9 g (95 % confidence
13 14 15	<i>LXRB</i> (rs1405655) on BW (after adjusting for the covariates), the mean estimated decrease (95% CI) in BW for a one-unit increase in PFOS levels in maternal blood was 502.9 g (95 % confidence interval [CI] = 247.3, 758.5 g). The corresponding mean estimated decrease in BW for the <i>LXRB</i>
13 14 15 16	<i>LXRB</i> (rs1405655) on BW (after adjusting for the covariates), the mean estimated decrease (95% CI) in BW for a one-unit increase in PFOS levels in maternal blood was 502.9 g (95 % confidence interval [CI] = 247.3, 758.5 g). The corresponding mean estimated decrease in BW for the <i>LXRB</i> (rs1405655)-TC/CC genotype was 526.3 g (95 % CI = 200.7, 852.0 g), compared with the <i>LXRB</i> -
13 14 15 16 17	<i>LXRB</i> (rs1405655) on BW (after adjusting for the covariates), the mean estimated decrease (95% CI) in BW for a one-unit increase in PFOS levels in maternal blood was 502.9 g (95 % confidence interval [CI] = 247.3, 758.5 g). The corresponding mean estimated decrease in BW for the <i>LXRB</i> (rs1405655)-TC/CC genotype was 526.3 g (95 % CI = 200.7, 852.0 g), compared with the <i>LXRB</i> -TT genotype. The estimated increase in BW related to interactions between PFOS levels in

1	for interaction $[p_{int}] = 0.003$). Moreover, after adjusting for the covariates, the effects of the
2	interactions between PFOS levels in maternal blood and the maternal genotype of LXRB
3	(rs1405655) on the BCC ($p_{int} = 0.037$) and BPI ($p_{int} = 0.005$) were also significant. These results
4	were almost the same as those obtained from the sensitivity analysis (Supplementary Table 3).
5	However, no interactions were observed between PFOS and fatty acid levels in maternal blood
6	(Supplementary Table 4). In male infants, there were no interactions between PFOS levels in
7	maternal blood and the maternal genotype of LXRB (rs1405655) regarding BW, BL, BCC, BHC,
8	or BPI (data not shown). Across all infants, there were also no interactions between PFOS levels
9	in maternal blood and the SNPs of PPARA, PPARG, PPARGCIA, PPARD, CAR, or LXRA
10	regarding BW, BL, BCC, BHC, or BPI (data not shown). Moreover, there were no interactions
11	between PFOA levels in maternal blood and all the SNPs of PPARA, PPARG, PPARGCIA, PPARD
12	CAR, LXRA, or LXRB regarding BW, BL, BCC, BHC, or BPI (data not shown).
13	The associations between PFOS and PFOA levels in maternal blood and birth outcomes,
14	stratified by the maternal genotype of <i>LXRB</i> (rs1405655) in female infants, are shown in Table 5.
15	After adjusting for the covariates, for the LXRB-TT genotype the estimated decreases in BW, BCC
16	and BPI for a one-unit increase in PFOS levels in maternal blood were 538.5 g (95 $\%$ CI = 278.1,
17	799.0 g), 1.368 cm (95 % CI = 0.159, 2.576 cm), and 0.242 kg/m ³ (95 % CI = 0.056, 0.429 kg/m ³),
18	respectively. For the LXRB-TC/CC genotype, the estimated increase in BPI for a one-unit increase

1	in PFOS levels in maternal blood was 0.321 kg/m^3 (95 % CI = 0.038, 0.605 kg/m ³). However, the
2	associations between PFOS levels in maternal blood and BW and BCC were not significant. These
3	results were almost the same as those obtained in the sensitivity analysis (Supplementary Table
4	5).
5	The dose-dependent associations between PFOS quartile levels in maternal blood and
6	BW, BCC, and BPI, stratified by the maternal genotype of LXRB (rs1405655) in female infants,
7	are illustrated in Fig. 2 (also see Supplementary Table 6). PFOS dose-dependent associations
8	with BP (<i>p</i> for trend $[p_{trend}] = 0.001$), BCC ($p_{trend} = 0.004$), and BPI ($p_{trend} = 0.018$) were
9	observed among female infants born to mothers with the LXRB (rs1405655) TT genotype, after
10	adjusting for the covariates. Female infants born to mothers with the rs1405655 TT genotype
11	had the highest quartiles of PFOS levels (\geq 7.2 ng/mL) weighed 306.1 g less (95% CI = 136.2,
12	475.9 g) in BW, were 1.242 cm shorter (95% CI = 0.431, 1.271 cm) in BCC, and were 0.144
13	kg/m ³ smaller (95% CI = 0.023, 0.265 kg/m ³) in BPI, compared with female infants born to
14	mothers with levels in the lowest quartile of PFOS (<3.7 ng/mL) and who had the rs1405655 TT
15	genotype (after adjusting for the covariates).
16	

4. Discussion

1	Here, we found that female infants born to mothers with LXRB (rs1405655) major homozygotes
2	(TT genotype) appeared to be susceptible to the adverse health effects of maternal PFOS levels.
3	This was manifested in increased risks of reductions in BW, BCC, and BPI. In contrast, female
4	infants born to mothers with the LXRB (rs1405655) heterozygote and minor homozygotes
5	(TC/CC genotype) appeared to be resistant to the adverse health effects of maternal PFOS
6	levels. This suggests that there was a gene-environment interaction between a specific LXRB TT
7	genotype of receptor genes and PFOS exposure during pregnancy, with relation to decreasing
8	birth size.
9	We found that increased PFOS levels in maternal blood were associated with lighter BW
10	and shorter BL, but only in female infants. Furthermore, we discovered that increased PFOA
11	levels in maternal blood were associated with lighter BW, but again only in female infants. Sex
12	differences in fetal growth related to PFOS and PFOA levels have also been reported in previous
13	18 studies [2-19]. Four of these studies reported that PFOS levels were only associated with
14	reduced birth outcomes in male infants [8,12,14,16]. These studies reported median PFOS levels
15	in maternal blood of 13.8 [12], 25.7 [14], and 27.2 ng/mL [16]. A previous study also reported a
16	median PFOS level in cord blood of 3.0 ng/mL [8], which is equal to approximately 7.1 ng/mL in
17	maternal blood, when converted using data from Cai et al. [1]. Conversely, three studies have
18	reported PFOS levels to be associated only with reduced birth outcomes in female infants

1	[10,13,19]. These studies reported the following median PFOS levels in maternal blood: 5.4 [19],
2	19.6 [10], and 30.1 ng/mL [13]. The PFOS levels obtained in our study (median = 5.2 ng/mL)
3	were comparable to these values.
4	Of the 18 abovementioned studies, one reported that PFOA levels were only associated
5	with reduced birth outcomes in male infants [7]; it reported a median PFOA level in maternal
6	blood of 2.3 ng/mL [7]. Conversely, six of these studies reported that PFOA levels were only
7	associated with reduced birth outcomes in female infants [5,8,10,13,18,19]; they reported the
8	following median PFOA levels in maternal blood: 1.6 [19], 3.7 [10], and 4.6 ng/mL [13]. These
9	studies also reported the following median PFOA levels in cord blood: 1.3 [8] and 2.0 ng/mL [18];
10	these are equal to approximately 1.7 and 2.8 ng/mL ng/mL in maternal blood, when converted
11	using data from Cai et al. [1]. The PFOA levels in our study (median = 1.3 ng/mL) were lower
12	than these values.
13	Regarding the sex-specific influence on the associations between PFOS and PFOA levels
14	during pregnancy and fetal growth, one possible mechanism may explain the observed
15	associations between PFOS and hepatic nuclear factor 4 alpha (HNF4 α) via LXRB. Cell studies
16	have revealed that the exposure of primary human hepatocytes to PFOS results in decreased
17	HNF4 α protein expression [48]. Furthermore, the overexpression of HNF4 α has been shown to
18	increase the promoter expression of LXRA [49]. LXRA shares 78 % of its amino acid sequence

1	with <i>LXRB</i> and binds to similar ligands with similar affinities [50]. Therefore, the decreased
2	activation of <i>LXRB</i> could be affected by PFOS exposure via the decreased activation of HNF4 α .
3	Studies in rabbits and mice have shown that $HNF4\alpha$ synergistically enhances the signal
4	transducer and activator of transcription 5b (STAT5b) [51] to regulate the sex-dependent
5	expression of liver cytochrome P450 (CYP) genes [52,53]. Hence, we speculate that the sex
6	differences observed here in the associations between PFOS levels in maternal blood and birth
7	outcomes might have been modified by the maternal genotype of LXRB (rs1405655). This may
8	have resulted in the obtained reductions in BW, BCC, and BPI, only in female infants.
9	The LXR agonist has been shown to selectively increase interleukin-1beta (IL-1 β)
10	messenger ribonucleic acid (mRNA) levels in human macrophages in vitro [54]; increased
11	levels of IL-1 β have also been associated with a risk of preterm birth [55]. As a possible
12	biological mechanism, the activation of LXRB may increase IL-1 β levels, leading to reductions
13	in BW, BCC, and BPI. Compared with the LXRB (rs1405655) minor homozygous (CC)
14	genotype, the LXRB (rs1405655) TT genotype of major homozygotes has been associated with
15	higher risks of tuberculosis [36], decreased insulin secretion in type 2 diabetes mellitus [56],
16	and Alzheimer's disease [57]. It has previously been shown that the linkage disequilibrium over
17	the genomic region containing LXRB underlies the intronic associations among rs1405655,
18	rs2303044, and rs4802703 [23]. Thus, we consider that the LXRB (rs1405655) TT genotype was

1	associated with higher activation of LXR and IL-1 β than the TC/CC genotype, and with
2	subsequent decreases in BW, BCC, and BPI. Compared with the LXRB (rs1405655) TT
3	genotype without minor C allele, there may have been a protective effect regarding the
4	increased risk of birth size reduction in the minor allele heterozygote (TC genotype) and
5	homozygotes (CC genotype) for the increasing PFOS levels during pregnancy. Future
6	investigations into the adverse birth health effects of various alleles at these positions are
7	needed, in relation to the LXRB promotor and transcription factor binding.
8	The maternal genotype LXRA (rs2279238) is located at an exon-splicing-enhancer region,
9	but the nucleotide variation is synonymous, thus resulting in a protein sequence of the mutant
10	genotype that is identical to that of the wild genotype [58]. Therefore, we observed no
11	interactions between PFOS and PFOA levels during pregnancy and the maternal genotype
12	LXRA (rs2279238) regarding birth outcomes.
13	One of the direct LXR target genes is <i>fatty acid transporter protein</i> [59]. In addition, LXR
14	regulates the long chain acyl-CoA synthase 3 (ACSL3) and sterol regulatory element binding
15	protein (SREBP) genes during lipogenesis [60-62]. However, no changes in fatty acid levels
16	were observed regarding interactions between PFOS levels during pregnancy and the maternal
17	genotype LXRB (rs1405655, rs2303044, and rs4802703) in a previous study [23]. Here, no
18	changes in fatty acid levels were obtained regarding interactions between these factors, when

1	stratified by infant sex. Our data suggest that the decreases BW, BCC, and BPI resulting from
2	the interactions between PFOS levels during pregnancy and the maternal genotype LXRB
3	(rs1405655) were not affected by maternal fatty acid levels. Further studies are needed to
4	elucidate the biological mechanism underlying the effects of the interactions between PFOS
5	levels during pregnancy and the maternal genotype LXRB (rs1405655) regarding the observed
6	decreases in BW, BCC, and BPI.
7	A previous study found that the maternal genotypes <i>PPARGC1A</i> (rs8192679) and <i>PPARD</i>
8	(rs1053049 and rs2267668) modified the association between PFOS and fatty acid levels during
9	pregnancy [23]. However, here we found that the maternal genotypes PPARA (rs1800234 and
10	rs135561), PPARG (rs3856806), PPARGC1A (rs2970847 and rs8192678), PPARD (rs1053049
11	and rs2267668), and CAR (rs2307424 and rs2501873) did not modify the association between
12	PFOS and PFOA levels during pregnancy and birth outcomes. Moreover, no associations
13	between PFOS and PFOA levels in maternal blood and birth outcomes, stratified by the
14	maternal genotypes PPARA (rs1800234 and rs135561), PPARG (rs3856806), PPARGC1A
15	(rs2970847 and rs8192678), PPARD (rs1053049 and rs2267668), and CAR (rs2307424 and
16	rs2501873), were observed (data not shown). Overall, the results of these studies suggest that
17	these genotypes in mothers did not modify the associations between PFOS and PFOA levels
18	during pregnancy and birth outcomes via maternal fatty acid levels.

1	One advantage of this study is that the risk of maternal exposure misclassification was
2	minimal because the PFOS and PFOA levels were determined based on measured levels in
3	blood samples using LC/MS/MS with a reliable, well-established, and accurate method.
4	However, this study has some limitations. First, no statistical corrections were performed for
5	multiple comparisons. As multiple hypothesis tests were performed in this study, this may have
6	increased the possibility of false positive errors. One limitation of the adopted no multiple
7	correction approach is that it does not consider type I errors. Second, although potential study
8	limitations include the small sample size used, a significant interaction was observed between
9	PFOS levels during pregnancy and the maternal genotype LXRB (rs1405655) regarding BW,
10	BCC, and BPI in female infants. Third, fetal genotypes were not considered in this study
11	because we focused on the association between maternal PFOS and PFOA exposure and
12	maternal receptor genotypes on birth size in this study. As the placenta is a fetal origin
13	tissue and is likely a significant target organ for PFOS and PFOA, and a key mediator of
14	sex differences in developmental outcomes, we need to examine the association
15	between maternal PFOS and PFOA exposure and fetal receptor genotypes on birth size
16	in a future study. Finally, as the study population considered mostly of infants with BWs
17	greater than 2,500 g, our findings may not be representative of infants with low birth weights.

1	Extensive efforts should be made to examine other polymorphisms that affect further
2	child growth trajectory regarding interactions between PFOS and PFOA levels during
3	pregnancy and maternal and child genotypes. Here, we only examined maternal genotypes; the
4	roles that child genotypes play in modifying the adverse effects of PFOS and PFOA levels
5	during pregnancy and maternal-child gene interactions still need to be elucidated.
6	
7	5. Conclusion
8	Here, we have demonstrated that PFOS levels during pregnancy were associated with decreased
9	BW, BCC, and BPI, and that those associations were significantly modified by the maternal
10	genotype LXRB (rs1405655). This study provides evidence of gene-environment interactions
11	between PFOS levels during pregnancy and the maternal genotype LXRB (rs1405655). For the
12	first time, we reported the existence of the LXRB (rs1405655) TC/CC genotype, which protects
13	against decreases in birth size regarding increased PFOS levels during pregnancy. This suggests
14	the importance of further assessing the role of genetic susceptibility when evaluating mother-
15	child relationships and environmental health. Further studies with gene-environment interaction
16	approaches may help identify more preventive genetic groups that can guide precision public
17	health in the future [63,64].

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4

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15	

16 **Conflicts of interest**

17 The authors declare no conflicts of interest.

18

1 Data statement

2	The data and materials used to derive our conclusions are unsuitable for public deposition due to
3	ethical restrictions and the specific legal framework in Japan. It is prohibited by the Act on the
4	Protection of Personal Information (Act No. 57 of May 30, 2003, amended on September 9,
5	2015) to publicly deposit data containing personal information. The Ethical Guidelines for
6	Epidemiological Research enforced by the Japan Ministry of Education, Culture, Sports,
7	Science and Technology and the Ministry of Health, Labour and Welfare also restrict the open
8	sharing of the epidemiologic data. All inquiries about access to data should be sent to
9	rkishi@med.hokudai.ac.jp. The person responsible for handling inquiries sent to this e-mail
10	address is Professor Reiko Kishi, Principal Investigator of the Hokkaido Study on Environment
11	and Children's Health, Center for Environmental and Health Sciences, Hokkaido University.
12	
13	
14	References
15	[1] D. Cai, Q.Q. Li, C. Chu, S.Z. Wang, Y.T. Tang, A.A. Appleton, R.L. Qiu, B.Y. Yang, L.W.
16	Hu, G.H. Dong, X.W. Zeng, High trans-placental transfer of perfluoroalkyl substances
17	alternatives in the matched maternal-cord blood serum: evidence from a birth cohort study, Sci.
18	Total Environ. 705 (2020) 135885. https://doi.org/10.1016/j.scitotenv.2019.135885.

2	[2] J. Ashley-Martin, L. Dodds, T.E. Arbuckle, M.F. Bouchard, M. Fisher, A.S. Morriset, P.
3	Monnier, G.D. Shapiro, A.S. Ettinger, R. Dallaire, S. Taback, W. Fraser, R.W. Platt, Maternal
4	concentrations of perfluoroalkyl substances and fetal markers of metabolic function and birth
5	weight, Am. J. Epidemiol. 185 (3) (2017) 185-193. https://doi.org/10.1093/aje/kww213.
6	
7	[3] C.C. Bach, B.H. Bech, E.A. Nohr, J. Olsen, N.B. Matthiesen, E.C. Bonefeld-Jørgensen, R.
8	Bossi, T.B. Henriksen, Perfluoroalkyl acids in maternal serum and indices of fetal growth: the
9	Aarhus birth cohort, Environ. Health Perspect. 124 (6) (2016) 848-854.
10	https://doi.org/10.1289/ehp.1510046.
11	
12	[4] A.C. Callan, A. Rotander, K. Thompson, J. Heyworth, J.F. Mueller, J.Ø. Odland, A.L.
13	Hinwood, Maternal exposure to perfluoroalkyl acids measured in whole blood and birth
14	outcomes in offspring, Sci. Total Environ. 569-570 (2016) 1107-1113.
15	https://doi.org/10.1016/j.scitotenv.2016.06.177.
16	
17	[5] W. Cao, X. Liu, X. Liu, Y. Zhou, X. Zhang, H. Tian, J. Wang, S. Feng, Y. Wu, P. Bhatti, S.
18	Wen, X. Sun, Perfluoroalkyl substances in umbilical cord serum and gestational and postnatal

1	growth in a Chinese birth cohort, Environ. Int. 116 (2018) 197-205.
2	https://doi.org/10.1016/j.envint.2018.04.015.
3	
4	[6] M.H. Chen, S. Ng, C.J. Hsieh, C.C. Lin, W.S. Hsieh, P.C. Chen, The impact of prenatal
5	perfluoroalkyl substances exposure on neonatal and child growth, Sci. Total Environ. 607-608
6	(2017) 669-675. https://doi.org/10.1016/j.scitotenv.2017.06.273.
7	
8	[7] H.B. Lauritzen, T.L. Larose, T. Øien, T.M. Sandanger, J.Ø. Odland, M. van de Bor, G.W.
9	Jacobsen, Maternal serum levels of perfluoroalkyl substances and organochlorines and indices
10	of fetal growth: a Scandinavian case-cohort study, Pediatr. Res. 81 (1-1) (2017) 33-42.
11	https://doi.org/10.1038/pr.2016.187.
12	
13	[8] M. Li, X.W. Zeng, Z.M. Qian, M.G. Vaughn, S. Sauvé, G. Paul, S. Lin, L. Lu, L.W. Hu, B.Y.
14	Yang, Y. Zhou, X.D. Qin, S.L. Xu, W.W. Bao, Y.Z. Zhang, P. Yuan, J. Wang, C. Zhang, Y.P.
15	Tian, M. Nian, X. Xiao, C. Fu, G.H. Dong, Isomers of perfluorooctanesulfonate (PFOS) in cord
16	serum and birth outcomes in China: Guangzhou birth cohort study, Environ. Int. 102 (2017) 1-8.
17	https://doi.org/10.1016/j.envint.2017.03.006.
18	

1	[9] D.V. Lind, L. Priskorn, T.H. Lassen, F. Nielsen, H.B. Kyhl, D.M. Kristensen, H.T.
2	Christesen, J.S. Jørgensen, P. Grandjean, T.K. Jensen, Prenatal exposure to perfluoroalkyl
3	substances and anogenital distance at 3 months of age in a Danish mother-child cohort, Reprod.
4	Toxicol. 68 (2017) 200-206. https://doi.org/10.1016/j.reprotox.2016.08.019.
5	
6	[10] M. Maisonet, M.L. Terrell, M.A. McGeehin, K.Y. Christensen, A. Holmes, A.M. Calafat,
7	M. Marcus, Maternal concentrations of polyfluoroalkyl compounds during pregnancy and fetal
8	and postnatal growth in British girls, Environ. Health Perspect. 120 (10) (2012) 1432-1437.
9	https://doi.org/10.1289/ehp.1003096.
10	
11	[11] C.B. Manzano-Salgado, M. Casas, M.J. Lopez-Espinosa, F. Ballester, C. Iñiguez, D.
12	Martinez, O. Costa, L. Santa-Marina, E. Pereda-Pereda, T. Schettgen, J. Sunyer, M. Vrijheid,
13	Prenatal exposure to perfluoroalkyl substances and birth outcomes in a Spanish birth cohort,
1 /	
14	Environ. Int. 108 (2017) 278-284. https://doi.org/10.1016/j.envint.2017.09.006.
14	Environ. Int. 108 (2017) 278-284. https://doi.org/10.1016/j.envint.2017.09.006.
14 15 16	Environ. Int. 108 (2017) 278-284. https://doi.org/10.1016/j.envint.2017.09.006. [12] K.J. Marks, A.J. Cutler, Z. Jeddy, K. Northstone, K. Kato, T.J. Hartman, Maternal serum
14 15 16 17	Environ. Int. 108 (2017) 278-284. https://doi.org/10.1016/j.envint.2017.09.006. [12] K.J. Marks, A.J. Cutler, Z. Jeddy, K. Northstone, K. Kato, T.J. Hartman, Maternal serum concentrations of perfluoroalkyl substances and birth size in British boys, Int. J. Hyg. Environ.
14 15 16 17 18	Environ. Int. 108 (2017) 278-284. https://doi.org/10.1016/j.envint.2017.09.006. [12] K.J. Marks, A.J. Cutler, Z. Jeddy, K. Northstone, K. Kato, T.J. Hartman, Maternal serum concentrations of perfluoroalkyl substances and birth size in British boys, Int. J. Hyg. Environ. Health. 222 (5) (2019) 889-895. https://doi.org/10.1016/j.ijheh.2019.03.008.

2	[13] Q. Meng, K. Inoue, B. Ritz, J. Olsen, Z. Liew, Prenatal exposure to perfluoroalkyl
3	substances and birth outcomes; an updated analysis from the Danish national birth cohort, Int. J.
4	Environ. Res. Public Health. 15 (9) (2018) 1832. https://doi.org/10.3390/ijerph15091832.
5	
6	[14] S.K. Sagiv, S.L. Rifas-Shiman, A.F. Fleisch, T.F. Webster, A.M. Calafat, X. Ye, M.W.
7	Gillman, E. Oken, Early-pregnancy plasma concentrations of perfluoroalkyl substances and birth
8	outcomes in project viva: confounded by pregnancy hemodynamics? Am. J. Epidemiol. 187 (4)
9	(2018) 793-802. https://doi.org/10.1093/aje/kwx332.
10	
11	[15] Y. Shi, L. Yang, J. Li, J. Lai, Y. Wang, Y. Zhao, Y. Wu, Occurrence of perfluoroalkyl
12	substances in cord serum and association with growth indicators in newborns from Beijing,
13	Chemosphere. 169 (2017) 396-402. https://doi.org/10.1016/j.chemosphere.2016.11.050.
14	
15	[16] D. Valvi, Y. Oulhote, P. Weihe, C. Dalgård, K.S. Bjerve, U. Steuerwald, P. Grandjean,
16	Gestational diabetes and offspring birth size at elevated environmental pollutant exposures,
17	Environ. Int. 107 (2017) 205-215. https://doi.org/10.1016/j.envint.2017.07.016.
18	

1	[17] N. Washino, Y. Saijo, S. Sasaki, S. Kato, S. Ban, K. Konishi, R. Ito, A. Nakata, Y. Iwasaki,
2	K. Saito, H. Nakazawa, R. Kishi, Correlations between prenatal exposure to perfluorinated
3	chemicals and reduced fetal growth, Environ. Health Perspect. 117 (4) (2009) 660-667.
4	https://doi.org/10.1289/ehp.11681.
5	
6	[18] H. Wang, H. Du, J. Yang, H. Jiang, K. O, L. Xu, S. Liu, J. Yi, X. Qian, Y. Chen, Q. Jiang, G.
7	He, PFOS, PFOA, estrogen homeostasis, and birth size in Chinese infants, Chemosphere. 221
8	(2019) 349-355. https://doi.org/10.1016/j.chemosphere.2019.01.061.
9	
10	[19] S. Wikström, P.I. Lin, C.H. Lindh, H. Shu, C.G. Bornehag, Maternal serum levels of
11	perfluoroalkyl substances in early pregnancy and offspring birth weight, Pediatr. Res. 87 (6)
12	(2020) 1093-1099. https://doi.org/10.1038/s41390-019-0720-1.
13	
14	[20] L. Grunnet, S. Vielwerth, A. Vaag, P. Poulsen, Birth weight is nongenetically associated
15	with glucose intolerance in elderly twins, independent of adult obesity, J. Intern. Med. 262 (1)
16	(2007) 96-103. https://doi.org/10.1111/j.1365-2796.2007.01793.x.
17	

1	[21] D.J. Barker, The origins of the developmental origins theory, J. Intern. Med. 261 (5):
2	(2007) 412-417. https://doi.org/10.1111/j.1365-2796.2007.01809.x.
3	
4	[22] T.P. Fleming, M.A. Velazquez, J.J. Eckert, Embryos, DOHaD and David Barker, J. Dev.
5	Orig. Health Dis. 6 (5) (2015) 377-383. https://doi.org/10.1017/S2040174415001105.
6	
7	[23] S. Kobayashi, F. Sata, H. Goudarzi, A. Araki, C. Miyashita, S. Sasaki, E. Okada, Y.
8	Iwasaki, T. Nakajima, R. Kishi, Associations among perfluorooctanesulfonic/perfluorooctanoic
9	acid levels, nuclear receptor gene polymorphisms, and lipid levels in pregnant women in the
10	Hokkaido study, Sci. Rep. 11 (1) (2021) 9994. https://doi.org/10.1038/s41598-021-89285-2.
11	
12	[24] S. Kobayashi, F. Sata, A. Ikeda-Araki, C. Miyashita, S. Itoh, H. Goudarzi, Y. Iwasaki, T.
13	Mitsui, K. Moriya, N. Shinohara, K. Cho, R. Kishi, Associations among maternal perfluoroalkyl
14	substance levels, fetal sex-hormone enzymatic gene polymorphisms, and fetal sex hormone
15	levels in the Hokkaido study, Reprod. Toxicol. 105 (2021) 221-231.
16	https://doi.org/10.1016/j.reprotox.2021.09.003.
17	

1	[25] E.J. Kwon, J.S. Shin, B.M. Kim, S. Shah-Kulkarni, H. Park, Y.L. Kho, E.A. Park, Y.J. Kim,
2	E.H. Ha, Prenatal Exposure to perfluorinated compounds affects birth weight through GSTM1
3	polymorphism, J. Occup. Environ. Med. 58 (6) (2016) e198-e205.
4	https://doi.org/10.1097/JOM.000000000000739.
5	
6	[26] M. Ghisari, H. Eiberg, M. Long, E.C. Bonefeld-Jørgensen, Polymorphisms in phase I and
7	phase II genes and breast cancer risk and relations to persistent organic pollutant exposure: a
8	case-control study in Inuit women, Environ. Health. 13 (1) (2014) 19.
9	https://doi.org/10.1186/1476-069X-13-19.
10	
11	[27] H.J. Wen, S.L. Wang, P.C. Chen, Y.L. Guo, Prenatal perfluorooctanoic acid exposure and
12	glutathione s-transferase T1/M1 genotypes and their association with atopic dermatitis at 2 years
13	of age, PLoS One. 14 (1) (2019) e0210708. https://doi.org/10.1371/journal.pone.0210708.
14	
15	[28] A.C. Behr, C. Plinsch, A. Braeuning, T. Buhrke, Activation of human nuclear receptors by
16	perfluoroalkylated substances (PFAS), Toxicol. In Vitro. 62 (2020) 104700.
17	https://doi.org/10.1016/j.tiv.2019.104700.
18	

1	[29] J.A. Bjork, J.L. Butenhoff, K.B. Wallace, Multiplicity of nuclear receptor activation by
2	PFOA and PFOS in primary human and rodent hepatocytes, Toxicology. 288 (1-3) (2011) 8-17.
3	https://doi.org/10.1016/j.tox.2011.06.012.
4	
5	[30] J.P. Vanden Heuvel, J.T. Thompson, S.R. Frame, P.J. Gillies, Differential activation of
6	nuclear receptors by perfluorinated fatty acid analogs and natural fatty acids: a comparison of
7	human, mouse, and rat peroxisome proliferator-activated receptor-alpha, -beta, and -gamma, liver
8	X receptor-beta, and retinoid X receptor-alpha, Toxicol. Sci. 92 (2) (2006) 476-489.
9	https://doi.org/10.1093/toxsci/kfl014.
10	
11	[31] R. Kishi, A. Araki, M. Minatoya, T. Hanaoka, C. Miyashita, S. Itoh, S. Kobayashi, Y. Ait
12	Bamai, K. Yamazaki, R. Miura, N. Tamura, K. Ito, H. Goudarzi, members of the Hokkaido
13	study on environment and children's health, The Hokkaido birth cohort study on environment
14	and children's health: cohort profile-updated 2017, Environ. Health Prev. Med. 22 (1) (2017) 46.
15	https://doi.org/10.1186/s12199-017-0654-3.
16	
17	[32] R. Kishi, A. Ikeda-Araki, C. Miyashita, S. Itoh, S. Kobayashi, Y. Ait Bamai, K. Yamazaki,
18	N. Tamura, M. Minatoya, R.M. Ketema, K. Poudel, R. Miura, H. Masuda, M. Itoh, T.

1	Yamaguchi, H. Fukunaga, K. Ito, H. Goudarzi, members of the Hokkaido study on environment
2	and children's health, Hokkaido birth cohort study on environment and children's health: cohort
3	profile 2021, Environ. Health Prev. Med. 26 (1) (2021) 59. https://doi.org/10.1186/s12199-021-
4	00980-у.
5	
6	[33] K. Inoue, F. Okada, R. Ito, S. Kato, S. Sasaki, S. Nakajima, A. Uno, Y. Saijo, F. Sata, Y.
7	Yoshimura, R. Kishi, H. Nakazawa, Perfluorooctane sulfonate (PFOS) and related
8	perfluorinated compounds in human maternal and cord blood samples: assessment of PFOS
9	exposure in a susceptible population during pregnancy, Environ. Health Perspect. 112 (11)
10	(2004) 1204-1207. https://doi.org/10.1289/ehp.6864.
11	
12	[34] S. Kobayashi, F. Sata, S. Sasaki, S. Ban, C. Miyashita, E. Okada, M. Limpar, E. Yoshioka,
13	J. Kajiwara, T. Todaka, Y. Saijo, R. Kishi, Genetic association of aromatic hydrocarbon receptor
14	(AHR) and cytochrome P450, family 1, subfamily A, polypeptide 1 (CYP1A1) polymorphisms
15	with dioxin blood concentrations among pregnant Japanese women, Toxicol. Lett. 219 (3)
16	(2013) 269-278. https://doi.org/10.1016/j.toxlet.2013.03.013.
17	

1	[35] S. Cresci, J.M. Huss, A.L. Beitelshees, P.G. Jones, M.R. Minton, G.W. Dorn, D.P. Kelly,
2	J.A. Spertus, H.L. McLeod, A PPARa promoter variant impairs ERR-dependent transactivation
3	and decreases mortality after acute coronary ischemia in patients with diabetes, PLoS One. 5 (9)
4	(2010) e12584. https://doi.org/10.1371/journal.pone.0012584.
5	
6	[36] M. Han, L. Liang, L.R. Liu, J. Yue, Y.L. Zhao, H.P. Xiao, Liver X receptor gene
7	polymorphisms in tuberculosis: effect on susceptibility, PLoS One. 2014; 9 (5) (2014) e95954.
8	https://doi.org/10.1371/journal.pone.0095954.
9	
10	[37] L.C. Kaupert, S.H.V. Lemos-Marini, M.P. De Mello, R.P. Moreira, V.N. Brito, A.A.L.
11	Jorge, C.A. Longui, G. Guerra Jr, B.B. Mendonca, T.A. Bachega, The effect of fetal androgen
12	metabolism-related gene variants on external genitalia virilization in congenital adrenal
13	hyperplasia, Clin. Genet. 84 (5) (2013) 482-488. https://doi.org/10.1111/cge.12016.
14	
15	[38] A. Leońska-Duniec, P. Cieszczyk, Z. Jastrzębski, A. Jażdżewska, E. Lulińska-Kuklik, W.
16	Moska, K. Ficek, M. Niewczas, A. Maciejewska-Skrendo, The polymorphisms of the PPARD
17	gene modify post-training body mass and biochemical parameter changes in women, PLoS One.
18	13 (8) (2018) e0202557. https://doi.org/10.1371/journal.pone.0202557.

1	
т	

2	[39] L.O. Lima, S. Almeida, M.H. Hutz, M. Fiegenbaum, PPARA, RXRA, NR112 and NR113
3	gene polymorphisms and lipid and lipoprotein levels in a Southern Brazilian population, Mol.
4	Biol. Rep. 40 (2) (2013) 1241-1247. https://doi.org/10.1007/s11033-012-2166-y.
5	
6	[40] Y.C. Lin, P.F. Chang, M.H. Chang, Y.H. Ni, A common variant in the peroxisome proliferator-
7	activated receptor- γ coactivator-1 α gene is associated with nonalcoholic fatty liver disease in
8	obese children, Am. J. Clin. Nutr. 97 (2) (2013) 326-331. https://doi.org/10.3945/ajcn.112.046417.
9	
10	[41] J. Lin, Y. Chen, W.F. Tang, C. Liu, S. Zhang, Z.Q. Guo, G. Chen, X.W. Zheng, PPARG
11	rs3856806 C>T polymorphism increased the risk of colorectal cancer: a case-control study in
12	eastern Chinese Han population, Front. Oncol. 9 (2019) 63.
13	https://doi.org/10.3389/fonc.2019.00063.
14	
15	[42] H. Naito, M. Kamijima, O. Yamanoshita, A. Nakahara, T. Katoh, N. Tanaka, T. Aoyama,
16	F.J. Gonzalez, T. Nakajima, Differential effects of aging, drinking and exercise on serum
17	cholesterol levels dependent on the PPARA-V227A polymorphism, J. Occup. Health. 49 (5)
18	(2007) 353-362. https://doi.org/10.1539/joh.49.353.

2	[43] K. Solaas, V. Legry, K. Retterstol, P.R. Berg, K.B. Holven, J. Ferrières, P. Amouyel, S.
3	Lien, J. Romeo, J. Valtueña, K. Widhalm, J.R. Ruiz, J. Dallongeville, S. Tonstad, H. Rootwelt,
4	B. Halvorsen, M.S. Nenseter, K.I. Birkeland, P.M. Thorsby, A. Meirhaeghe, H.I. Nebb,
5	Suggestive evidence of associations between liver X receptor β polymorphisms with type 2
6	diabetes mellitus and obesity in three cohort studies: HUNT2 (Norway), MONICA (France) and
7	HELENA (Europe), BMC Med. Genet. 11 (2010) 144. https://doi.org/10.1186/1471-2350-11-
8	144.
9	
10	[44] K.S. Vimaleswaran, V. Radha, M. Anjana, R. Deepa, S. Ghosh, P.P. Majumder, M.R. Rao,
11	V. Mohan, Effect of polymorphisms in the PPARGC1A gene on body fat in Asian Indians, Int. J.
12	Obes. (Lond). 30 (6) (2006) 884-891. https://doi.org/10.1038/sj.ijo.0803228.
13	
14	[45] Z. Wang, A. Dessa Sadovnick, A.L. Traboulsee, J.P. Ross, C.Q. Bernales, M. Encarnacion,
15	I.M. Yee, M. de Lemos, T. Greenwood, J.D. Lee, G. Wright, C.J. Ross, S. Zhang, W. Song, C.
16	Vilariño-Güell, Nuclear receptor NR1H3 in familial multiple sclerosis, Neuron. 90 (5) (2016)
17	948-954. https://doi.org/10.1016/j.neuron.2016.04.039.
10	

1	[46] X. Jia, M. Tagawa, H. Yatsuya, H. Naito, Y. Hayashi, H. Yetti, S. Sasaki, A. Araki, C.
2	Miyashita, T. Ikeno, R. Kishi, T. Nakajima, Association of maternal whole blood fatty acid
3	status during the prenatal period with term birth dimensions: a cross-sectional study, J. Perinat.
4	Med. 43 (5) (2015) 565-575. https://doi.org/10.1515/jpm-2014-0277.
5	
6	[47] J. Folch, M. Lees, G.H. Sloane Stanley, A simple method for the isolation and purification
7	of total lipides from animal tissues, J. Biol. Chem. 226 (1) (1957) 497-509.
8	
9	[48] K.M. Beggs, S.R. McGreal, A. McCarthy, S. Gunewardena, J.N. Lampe, C. Lau, U. Apte,
10	The role of hepatocyte nuclear factor 4-alpha in perfluorooctanoic acid- and
11	perfluorooctanesulfonic acid-induced hepatocellular dysfunction, Toxicol. Appl. Pharmacol. 304
12	(2016) 18-29. https://doi.org/10.1016/j.taap.2016.05.001.
13	
14	[49] D. Theofilatos, A. Anestis, K. Hashimoto, D. Kardassis, Transcriptional regulation of the
15	human Liver X Receptor α gene by hepatocyte nuclear factor 4α , Biochem. Biophys. Res.
16	Commun. 469 (3) (2016) 573-579. https://doi.org/10.1016/j.bbrc.2015.12.031.
17	

1	[50] K. Prüfer, J. Boudreaux, Nuclear localization of liver X receptor alpha and beta is
2	differentially regulated, J. Cell. Biochem. 100 (1) (2007) 69-85.
3	https://doi.org/10.1002/jcb.21006.
4	
5	[51] S.H. Park, C.A. Wiwi, D.J. Waxman, Signalling cross-talk between hepatocyte nuclear
6	factor 4alpha and growth-hormone-activated STAT5b, Biochem. J. 397 (1) (2006) 159-168.
7	https://doi.org/10.1042/BJ20060332.
8	
9	[52] C.A. Wiwi, M. Gupte, D.J. Waxman, Sexually dimorphic P450 gene expression in liver-
10	specific hepatocyte nuclear factor 4alpha-deficient mice, Mol. Endocrinol. 18 (8) (2004) 1975-
11	1987. https://doi.org/10.1210/me.2004-0129.
12	
13	[53] C.A. Wiwi, D.J. Waxman, Role of hepatocyte nuclear factors in transcriptional regulation of
14	male-specific CYP2A2, J. Biol. Chem. 280 (5) (2005) 3259-3268.
15	https://doi.org/10.1074/jbc.M409294200.
16	
17	[54] L. Ménégaut, C. Thomas, A. Jalil, J.B. Julla, C. Magnani, A. Ceroi, L. Basmaciyan, A.
18	Dumont, W. Le Goff, M.J. Mathew, C. Rébé, V. Dérangère, A. Laubriet, V. Crespy, J.P. Pais de

1	Barros, E. Steinmetz, N. Venteclef, P. Saas, L. Lagrost, M. Masson, Interplay between liver X
2	receptor and hypoxia inducible factor 1α potentiates interleukin- 1β production in human
3	macrophages, Cell. Rep. 31 (7) (2020) 107665. https://doi.org/10.1016/j.celrep.2020.107665.
4	
5	[55] I.M. Langmia, Y.D. Apalasamy, S.Z. Omar, Z. Mohamed, Impact of IL1B gene
6	polymorphisms and interleukin 1B levels on susceptibility to spontaneous preterm birth,
7	Pharmacogenet. Genomics. 26 (11) (2016) 505-509.
8	https://doi.org/10.1097/FPC.00000000000243.
9	
10	[56] C. Ketterer, K. Müssig, F. Machicao, N. Stefan, A. Fritsche, H.U. Häring, H. Staiger,
11	Genetic variation within the NR1H2 gene encoding liver X receptor β associates with insulin
12	secretion in subjects at increased risk for type 2 diabetes, J. Mol. Med. (Berl). 89 (1) (2011) 75-
13	81. https://doi.org/10.1007/s00109-010-0687-1.
14	
15	[57] J. Infante, E. Rodríguez-Rodríguez, I. Mateo, J. Llorca, J.L. Vázquez-Higuera, J. Berciano,
16	O. Combarros, Gene-gene interaction between heme oxygenase-1 and liver X receptor-beta and
17	Alzheimer's disease risk, Neurobiol. Aging. 31 (4) (2010) 710-714.
18	https://doi.org/10.1016/j.neurobiolaging.2008.05.025.

2	[58] E.T. Price, M.A. Pacanowski, M.A. Martin, R.M. Cooper-DeHoff, C.J. Pepine, I. Zineh,
3	J.A. Johnson, Liver X receptor alpha gene polymorphisms and variable cardiovascular
4	outcomes in patients treated with antihypertensive therapy: results from the invest-genes study,
5	Pharmacogenet. Genomics. 21 (6) (2011) 333-340.
6	https://doi.org/10.1097/FPC.0b013e3283452fec.
7	
8	[59] J. Zhou, M. Febbraio, T. Wada, Y. Zhai, R. Kuruba, J. He, J.H. Lee, S. Khadem, S. Ren, S.
9	Li, R.L. Silverstein, W. Xie, Hepatic fatty acid transporter Cd36 is a common target of LXR, PXR,
10	and PPARgamma in promoting steatosis, Gastroenterology. 134 (2) (2008) 556-567.
11	https://doi.org/10.1053/j.gastro.2007.11.037.
12	
13	[60] S.B. Joseph, B.A. Laffitte, P.H. Patel, M.A. Watson, K.E. Matsukuma, R. Walczak, J.L.
14	Collins, T.F. Osborne, P. Tontonoz, Direct and indirect mechanisms for regulation of fatty acid
15	synthase gene expression by liver X receptors, J. Biol. Chem. 277 (13) (2002) 11019-11025.
16	https://doi.org/10.1074/jbc.M111041200.
17	

18 [61] J.J. Repa, G. Liang, J. Ou, Y. Bashmakov, J.M. Lobaccaro, I. Shimomura, B. Shan, M.S.

1	Brown, J.L. Goldstein, D.J. Mangelsdorf, Regulation of mouse sterol regulatory element-binding			
2	protein-1c gene (SREBP-1c) by oxysterol receptors, LXRalpha and LXRbeta, Genes. Dev. 2000;			
3	14 (22) (2000) 2819-2830. https://doi.org/10.1101/gad.844900.			
4				
5	[62] M.S. Weedon-Fekjaer, K.T. Dalen, K. Solaas, A.C. Staff, A.K. Duttaroy, H.I. Nebb,			
6	Activation of LXR increases acyl-CoA synthetase activity through direct regulation of ACSL3 in			
7	human placental trophoblast cells, J. Lipid Res. 51 (7) (2010) 1886-1896.			
8	https://doi.org/10.1194/jlr.M004978.			
9				
10	[63] M.J. Khoury, M.F. Iademarco, W.T. Riley, Precision public health for the era of precision			
11	medicine, Am. J. Prev. Med. 50 (3) (2016) 398-401.			
12	https://doi.org/10.1016/j.amepre.2015.08.031.			
13				
14	[64] M.J. Khoury, M.S. Bowen, M. Clyne, W.D. Dotson, M.L. Gwinn, R.F. Green, K. Kolor,			
15	J.L. Rodriguez, A. Wulf, W. Yu, From public health genomics to precision public health: a 20-			
16	year journey, Genet. Med. 20 (6) (2018) 574-582. https://doi.org/10.1038/gim.2017.211.			
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1 Figure legends

2	Fig. 1. Interaction plots between PFOS levels and the maternal genotype <i>LXRB</i> (rs1405655) on
3	(a) birth weight, (b) birth chest circumference, and (c) birth Ponderal index among female
4	infants born to pregnant Japanese women in the Hokkaido study
5	
6	Fig. 2. Associations between PFOS quartile levels and (a) birth weight, (b) birth chest
7	circumference, and (c) birth Ponderal index stratified by the maternal genotype LXRB
8	(rs1405655) among female infants born to pregnant Japanese women in the Hokkaido study
9	Bars represent the changes (95% confidence interval [CI]) in birth weight (g), birth chest
10	circumference (cm), and birth Ponderal index (kg/m ³) compared to the reference group.
11	<i>ptrend</i> represents <i>p</i> -values for trends.
12	* <i>p</i> <0.05, ** <i>p</i> <0.01.

1 Fig. 1 (a)



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1 Fig. 1 (b)



1 Fig. 1 (c)



1 Fig. 2 (a)



1 Fig. 2 (b)



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1 Fig. 2 (c)



1 Table 1. Characteristics of the study participants (n = 372)

	Total	Male	Female	
Characteristics	(n = 372)	(n = 174)	(n = 198)	p value ^g
Mothers				
Age (years) ^{a,d}	30.2 ± 4.8	30.8 ± 4.7	29.7 ± 4.9	0.020
Pre-pregnancy body mass index (BMI) (kg/m ²) ^{a,d}	21.0 ± 2.9	20.7 ± 2.6	21.2 ± 3.1	0.106
Parity ^{b,e}				
Primiparous	173 (46.5)	79 (45.4)	94 (47.5)	0.727
Multiparous	198 (53.2)	94 (54.0)	104 (52.5)	
Missing data	1 (0.3)	1 (0.6)	0 (0.0)	
Smoking in the third trimester ^{b,e}				
No	303 (81.5)	147 (84.5)	156 (78.8)	0.159
Yes	69 (18.5)	27 (15.5)	42 (21.2)	
Alcohol consumption during pregnancy ^{b,e}				
No	260 (69.9)	121 (69.5)	139 (70.2)	0.890
Yes	112 (30.1)	53 (30.5)	59 (29.8)	
Educational level ^{b,e}				
Less than or equal to high school graduation	160 (43.0)	79 (45.4)	81 (40.9)	0.401
More than high school graduation	209 (56.2)	94 (54.0)	115 (58.1)	
Missing data	3 (0.8)	1 (0.6)	2 (1.0)	
Annual household income (million Japanese yen) ^{b,e}				
Less than 5	262 (70.4)	123 (70.7)	139 (70.2)	0.977
More than or equal to 5	104 (28.0)	49 (28.2)	55 (27.8)	
Missing data	6 (1.6)	2 (1.1)	4 (2.0)	
Cesarean section ^{b,e}				
No	296 (79.6)	138 (79.3)	158 (79.8)	0.907
Yes	76 (20.4)	36 (20.7)	40 (20.2)	
Blood sampling period ^{b,e}				
During pregnancy	235 (63.2)	106 (62.6)	126 (63.6)	0.843
After delivery	137 (36.8)	65 (37.4)	72 (36.4)	
Blood serum level ^{c,f}				
PFOS (ng/mL)	5.2 (3.7, 7.2)	5.3 (3.9, 7.0)	5.2 (3.4, 7.3)	0.491
PFOA (ng/mL)	1.3 (0.8, 1.8)	1.4 (0.9, 1.8)	1.2 (0.8, 1.7)	0.041
Triglyceride (mg/100-mL)	84.4 (61.7, 117.0)	82.8 (58.4, 123.3)	85.1 (64.5, 114.7)	0.686
Palmitic acid (µg/mL)	1,876.8 (1,557.3, 2,145.8)	1,785.5 (1,529.9, 2,412.1)	1,948.1 (1,564.8, 2,438.7)	0.203
Palmitoleic acid (µg/mL)	107.5 (77.5, 155.9)	104.4 (76.0, 150.6)	109.4 (80.1, 159.6)	0.380
Stearic acid (µg/mL)	526.4 (429.2, 635.5)	526.3 (425.5, 670.6)	526.7 (432.4, 623.8)	0.938
Oleic acid (µg/mL)	1,122.1 (891.4, 1,430.2)	1,112.1 (838.5, 1,435.8)	1,153.3 (927.0, 1,429.0)	0.372
Linoleic acid (µg/mL)	725.5 (532.1, 949.9)	746.2 (533.3, 951.2)	707.3 (527.3, 943.3)	0.719
Linolenic acid (µg/mL)	10.5 (5.5, 15.1)	10.4 (5.8, 15.4)	10.7 (5.1, 15.1)	0.925
Arachidonic acid (µg/mL)	65.0 (44.4, 93.8)	63.9 (47.2, 96.8)	67.2 (41.1, 92.0)	0.490
Eicosapentaenoic acid (µg/mL)	8.6 (4.8, 14.5)	8.5 (4.8, 14.1)	9.0 (4.8, 14.8)	0.971
Docosahexaenoic acid (µg/mL)	25.5 (13.5, 40.1)	24.1 (13.9, 40.8)	26.4 (13.1, 38.9)	0.690
Infants				

Sex ^{b,e}				
Male	174 (46.8)	174 (100.0)	0 (0.0)	(-)
Female	198 (53.2)	0 (0.0)	198 (100.0)	
Gestational age (weeks) ^{a,d}	38.9 ± 1.5	38.8 ± 1.5	39.0 ± 1.5	0.131
Birth weight $(BW)(g)^{a,d}$	$3,044.8 \pm 386.2$	$3,098.3 \pm 366.5$	$2,997.8 \pm 397.7$	0.012
Birth length (BL)(cm) ^{a,d}	48.0 ± 2.1	48.3 ± 2.1	47.7 ± 2.0	0.004
Birth chest circumference (BCC)(cm) ^{a,d}	31.4 ± 1.7	31.5 ± 1.5	31.3 ± 1.9	0.188
Birth head circumference (BHC)(cm) ^{a,d}	33.2 ± 1.4	33.6 ± 1.3	32.8 ± 1.4	< 0.001
Birth Ponderal index (BPI)(kg/m ³) ^{a,d}	2.754 ± 0.387	2.759 ± 0.507	2.749 ± 0.238	0.812

^a Mean \pm Standard deviation (SD). ^b n (%). 1

 $\mathbf{2}$

^a M (%).
^c Median (inter-quartile range; IQR).
^d Independent *t*-test.
^e Chi-squared test.
^f Mann-Whitney's U-test.
^g Male versus female. 3

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6

7

Abbreviations: PFOS, perfluorooctanesulfonate; PFOA, perfluorooctanoic acid.

 $\frac{8}{9}$

	Total	Male	Female			Total	Male	Female	
	(n = 3/2)	(n = 1/4)	(n = 198)			(n = 3/2)	(n = 1/4)	(n = 198)	
Gene name/genotype	n (%)	n (%)	n (%)	HWE ^a	Gene name/genotype	n (%)	n (%)	n (%)	HWE ^a
PPARA (rs1800234)					CAR (rs2307044)				
TT	334 (89.8)	158 (90.8)	176 (88.9)		TT	103 (27.7)	44 (25.3)	59 (29.8)	
TC	35 (9.4)	16 (9.2)	19 (9.6)	$\chi^2 = 3.468$	TC	195 (52.4)	92 (52.9)	103 (52.0)	$\chi^2 = 1.117$
CC	3 (0.8)	0 (0.0)	3 (1.5)	p = 0.062	CC	74 (19.9)	38 (21.8)	36 (18.2)	p = 0.291
PPARA (rs135561)					CAR (rs2501873)				
GG	331 (89.0)	159 (91.4)	172 (86.9)		AA	123 (33.1)	66 (37.9)	57 (28.8)	
GA	41 (11.0)	15 (8.6)	26 (13.1)	$\chi^2 = 1.265$	AG	194 (52.2)	80 (46.0)	114 (57.6)	$\chi^2 = 2.326$
AA	0 (0.0)	0 (0.0)	0 (0.0)	p = 0.261	GG	55 (14.8)	28 (16.1)	27 (13.6)	p = 0.127
PPARG (rs3856806)					LXRA (rs2278238)				
GG	263 (70.3)	129 (74.1)	134 (67.7)		CC	160 (43.0)	66 (37.9)	94 (47.5)	
GA	98 (26.3)	39 (22.4)	59 (29.8)	$\chi^2 = 0.257$	CT	168 (45.2)	92 (52.9)	76 (38.4)	$\chi^2 = 0.000$
AA	11 (3.0)	6 (3.4)	5 (2.5)	p = 0.612	TT	44 (11.8)	16 (9.2)	28 (14.1)	p = 0.992
PPARGC1A (rs2970847)	· · · ·	× /	× ,	1	LXRB (rs1405655)	~ /		× /	1
CC	235 (63.2)	101 (58.0)	134 (67.7)		TT	242 (65.1)	119 (68.4)	123 (62.1)	
СТ	120 (32.3)	62 (35.6)	58 (29.3)	$\gamma^2 = 0.112$	TC	111 (29.8)	47 (27.0)	64 (32.3)	$\gamma^2 = 1.744$
TT	17 (4.6)	11 (6.3)	6 (3.0)	p = 0.737	CC	19 (5.1)	8 (4.6)	11 (5.6)	p = 0.187
PPARGC1A (rs8192678)		()	- ()	F · · · · ·	LXRB (rs2303044)	- (-)	- (-)	()	r
GG	101 (27.2)	46 (26.4)	55 (27.8)		GG	252 (67.7)	121 (69.5)	131 (66.2)	
GA	182 (48.9)	93 (53.4)	89 (44.9)	$\gamma^2 = 0.156$	GA	106 (28.5)	46 (26.4)	60 (30.3)	$\gamma^2 = 0.461$
АА	89 (23.9)	35 (20.1)	54 (27.3)	n = 0.693	АА	14 (3.8)	7 (4.0)	7 (3.5)	n = 0.497
PPARD (rs1053049)	()		e (_,)	P	LXRB (rs4802703)	- (()	, ()	(212)	P
TT	226 (60.8)	108 (62.1)	118 (59.6)		GG	265 (71.2)	132 (75.9)	133 (67.2)	
TC	135(36.3)	58 (33.3)	77 (38.9)	$\gamma^2 = 3.004$	GA	95 (25.5)	36 (20.7)	59 (29.8)	$\gamma^2 = 0.918$
CC	11 (3.0)	8 (4.6)	3 (1.5)	n = 0.083	AA	12(3.2)	6 (3.4)	6 (3.0)	n = 0.338
<i>PPARD</i> (rs2267668)	11 (0.0)	0 (5 (1.5)	r 0.000		12 (0.2)	0 (0.1)	0 (0.0)	r 0.000
AA	242 (65.1)	118 (67.8)	124 (62.6)						
AG	120(323)	48 (27.6)	72 (36.4)	$\gamma^2 = 1.159$					
GG	10(2.7)	8 (4.6)	2(1.0)	n = 0.282					

1 Table 2. Maternal genotype frequencies (n = 372)

^a Chi-squared test was employed to test whether the frequency of genotype distribution conformed to the Hardy-Weinberg equilibrium (HWE).

 $\frac{2}{3}$

1 Table 3. Association between PFOS and PFOA levels and birth outcomes stratified by infant sex

		Total				Mal	es		Females			
	Crude		Adjusted		Crude		Adjusted		Crude		Adjusted	
Outcome	β (95% CI)	p value	β (95% CI)	p value	β (95% CI)	p value	β (95% CI)	p value	β (95% CI)	p value	β (95% CI)	p value
Exposure: PFOS (ng/mL)												
BW (g)	-77.0 (-256.0, 102.1)	0.398	-182.3 (-336.5, -28.2)	0.021	8.5 (-246.7, 263.6)	0.948	17.7 (-207.0, 242.5)	0.876	-165.3 (-414.4, 83.7)	0.192	-292.1 (-504.3, -79.8)	0.007
BL (cm)	0.023 (-0.945, 0.991)	0.963	-0.552 (-1.433, 0.328)	0.218	0.780 (-0.701, 2.261)	0.300	0.635 (-0.832, 2.102)	0.394	-0.709 (-1.965, 0.548)	0.267	-1.384 (-2.472, -0.297)	0.013
BCC (cm)	-0.158 (-0.960, 0.644)	0.698	-0.625 (-1.343, 0.093)	0.088	0.166 (-0.844, 1.177)	0.746	0.113 (-0.796, 1.022)	0.806	-0.467 (-1.683, 0.749)	0.450	-1.033 (-2.122, 0.056)	0.063
BHC (cm)	-0.225 (-0.869, 0.418)	0.492	-0.339 (-0.957, 0.280)	0.282	-0.226 (-1.120, 0.668)	0.618	-0.034 (-0.931, 0.864)	0.941	-0.335 (-1.228, 0.519)	0.424	-0.612 (-1.483, 0.258)	0.167
BPI (kg/m ³)	-0.054 (-0.234, 0.126)	0.556	-0.079 (-0.270, 0.111)	0.412	-0.077 (-0.430, 0.276)	0.668	-0.070 (-0.480, 0.321)	0.724	-0.037 (-0.187, 0.112)	0.625	-0.057 (-0.211, 0.097)	0.466
Exposure: PFOA (ng/mL)												
BW (g)	-69.0 (-206.9, 68.9)	0.326	-107.1 (-232.5, 18.4)	0.094	-29.9 (-223.9, 164.0)	0.761	-55.8 (-235.4, 123.8)	0.540	-130.3 (-325.1, 64.6)	0.189	-183.0 (-361.9, -4.1)	0.045
BL (cm)	-0.027 (-0.773, 0.719)	0.944	-0.408 (-1.122, 0.307)	0.262	0.047 (-1.083, 1.176)	0.935	-0.077 (-1.253, 1.099)	0.897	-0.254 (-1.239, 0.732)	0.612	-0.618 (-1.538, 0.302)	0.187
BCC (cm)	-0.234 (-0.851, 0.384)	0.458	-0.445 (-1.028, 0.138)	0.134	0.165 (-0.603, 0.933)	0.672	0.015 (-0.712, 0.742)	0.968	-0.644 (-1.593, 0.305)	0.182	-0.844 (-1.754, 0.067)	0.069
BHC (cm)	-0.169 (-0.065, 0.327)	0.502	-0.294 (-0.796, 0.207)	0.249	-0.283 (-0.962, 0.396)	0.412	-0.189 (-0.907, 0.528)	0.603	-0.262 (-0.946, 0.422)	0.450	-0.431 (-1.159, 0.298)	0.245
BPI (kg/m ³)	-0.054 (-0.193, 0.084)	0.442	-0.054 (-0.208, 0.101)	0.495	-0.016 (-0.284, 0.253)	0.908	-0.067 (-0.379, 0.245)	0.673	-0.091 (-0.207, 0.026)	0.126	-0.090 (-0.218, 0.038)	0.168
The associations between PFOS	or PFOA levels and birth out	comes were te	sted using a multiple linear	regression mo	odel.							
Crude: Non-adjusted.												

Adjusted for maternal age (continuous), pre-pregnancy BMI (continuous), maternal smoking in the third trimester (yes/no), maternal alcohol consumption during pregnancy (yes/no), parity (primiparous/multiparous), educational level (less than or equal to high school graduation/more than high school graduation), annual household income (less than 5 million Japanese yen/more than or equal to 5 million Japanese yen), cesarean section (yes/no), maternal blood sampling period (during pregnancy/after delivery), gestational age (continuous), and infant sex (male/female; total infants only). β (95% CI) represents the change in BW (g), BL (cm), BCC (cm), BHC (cm), and BPI (kg/m³) for each ten-fold increase in PFOS or PFOA levels (ng/mL).

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1 Table 4. Interactions between PFOS and PFOA levels and the maternal genotype *LXRB* (rs1405655) regarding birth outcomes in female infants

		Crude		Adjusted	
Outcome	Exposure/genotype	β (95% CI)	p value	β (95% CI)	p value
BW (g)	PFOS (ng/mL)	-365.4 (-664.1, -66.8)	0.017	-502.9 (-758.5, -247.3)	< 0.001
	LXRB-TC/CC (vs. TT)	-522.2 (-912.5, -131.8)	0.009	-526.3 (-852.0, -200.7)	0.002
	PFOS × LXRB-TC/CC (Interaction term)	664.2 (133.5, 1,194.8)	0.014	662.1 (221.0, 1,103.2)	0.003
BL (cm)	PFOS (ng/mL)	-0.754 (-2.284, 0.776)	0.332	-1.468 (-2.808, -0.129)	0.032
	LXRB-TC/CC (vs. TT)	-0.444 (-2.444, 1.556)	0.662	-0.594 (-2.301, 1.113)	0.493
	PFOS × LXRB-TC/CC (Interaction term)	0.287 (-2.433, 3.006)	0.836	0.420 (-1.892, 2.732)	0.720
BCC (cm)	PFOS (ng/mL)	-1.188 (-2.651, 0.276)	0.111	-1.767 (-3.089, -0.445)	0.009
	LXRB-TC/CC (vs. TT)	-2.157 (-4.070, -0.244)	0.027	-2.133 (-3.817, -0.449)	0.013
	PFOS × LXRB-TC/CC (Interaction term)	2.507 (-0.094, 5.108)	0.059	2.427 (0.146, 4.708)	0.037
BHC (cm)	PFOS (ng/mL)	-0.532 (-1.560, 0.495)	0.308	-0.786 (-1.819, 0.247)	0.135
	LXRB-TC/CC (vs. TT)	-1.437 (-2.780, -0.094)	0.036	-1.401 (-2.718, -0.084)	0.037
	PFOS × LXRB-TC/CC (Interaction term)	0.996 (-0.830, 2.822)	0.283	0.940 (-0.843, 2.723)	0.300
BPI (kg/m ³)	PFOS (ng/mL)	-0.189 (-0.368, -0.011)	0.037	-0.212 (-0.398, -0.025)	0.026
	LXRB-TC/CC (vs. TT)	-0.360 (-0.593, -0.127)	0.003	-0.340 (-0.578, -0.103)	0.005
	PFOS \times <i>LXRB</i> -TC/CC (Interaction term)	0.490 (0.173, 0.806)	0.003	0.468 (0.146, 0.789)	0.005
BW (g)	PFOA (ng/mL)	-136.6 (-382.3, 109.1)	0.274	-227.6 (-446.3, -8.9)	0.041
	LXRB-TC/CC (vs. TT)	-57.3 (-174.9, 60.3)	0.338	-67.6 (-166.1, 30.9)	0.177
	PFOA × LXRB-TC/CC (Interaction term)	38.1 (-369.5, 445.7)	0.854	157.8 (-185.5, 501.1)	0.366
BL (cm)	PFOA (ng/mL)	0.077 (-1.164, 1.318)	0.903	-0.488 (-1.614, 0.638)	0.394
	LXRB-TC/CC (vs. TT)	-0.213 (-0.807, 0.381)	0.480	-0.304 (-0.811, 0.204)	0.239
	PFOA × LXRB-TC/CC (Interaction term)	-0.813 (-2.871, 1.246)	0.437	-0.175 (-1.942, 1.593)	0.846
BCC (cm)	PFOA (ng/mL)	-0.207 (-1.397, 0.983)	0.732	-0.591 (-1.702, 0.519)	0.295
	LXRB-TC/CC (vs. TT)	-0.321 (-0.891, 0.248)	0.267	-0.379 (-0.880, 0.121)	0.137
	PFOA × LXRB-TC/CC (Interaction term)	-1.061 (-3.035, 0.914)	0.291	-0.457 (-2.201, 1.287)	0.606
BHC (cm)	PFOA (ng/mL)	0.268 (-0.561, 1.097)	0.525	0.002 (-0.857, 0.862)	0.995
	LXRB-TC/CC (vs. TT)	-0.671 (-1.068, -0.275)	0.001	-0.693 (-1.080, -0.306)	0.001
	PFOA × LXRB-TC/CC (Interaction term)	-1.182 (-2.558, 0.193)	0.092	-0.761 (-2.110, 0.588)	0.267
BPI (kg/m ³)	PFOA (ng/mL)	-0.136 (-0.283, 0.010)	0.068	-0.145 (-0.303, 0.012)	0.069
	LXRB-TC/CC (vs. TT)	-0.019 (-0.090, 0.051)	0.585	-0.016 (-0.086, 0.055)	0.666
	$PFOA \times LXRB$ -TC/CC (Interaction term)	0.130 (-0.113, 0.373)	0.293	0.157(-0.090, 0.404)	0.211

The associations between PFOS or PFOA levels and the maternal genotype LXRB (rs1405655) on birth outcomes were tested using a multiple linear regression model.

Crude: Non-adjusted.

Adjusted for maternal age (continuous), pre-pregnancy BMI (continuous), maternal smoking in the third trimester (yes/no), maternal alcohol consumption during pregnancy (yes/no), parity (primiparous/multiparous), educational level (less than or equal to high school graduation/more than high school graduation), annual household income (less than 5 million Japanese yen/more than or equal

to 5 million Japanese yen), cesarean section (yes/no), maternal blood sampling period (during pregnancy/after delivery), and gestational age (continuous).

 β (95% CI) represents the change in BW (g), BL (cm), BCC (cm), BHC (cm), and BPI (kg/m³).

8 PFOS and PFOA levels were log₁₀-transformed scales.

PFOS-LXRB or PFOA-LXRB interaction term was defined as " \log_{10} -transformed PFOS or PFOA levels (continuous) × genotype (0 = TT and 1 = TC/CC)."

		Crude		Adjusted	
Outcome	Genotype	β (95% CI)	p value	β (95% CI)	p value
Exposure: PF	OS (ng/mL)				
BW (g)	TT	-365.4 (-657.6, -73.3)	0.015	-538.5 (-799.0, -278.1)	< 0.001
	TC/CC	298.7 (-162.6, 760.1)	0.201	135.6 (-240.4, 511.6)	0.474
BL (cm)	TT	-0.754 (-2.037, 0.529)	0.247	-1.368 (-2.576, -0.159)	0.027
	TC/CC	-0.467 (-3.251, 2.317)	0.739	-1.616 (-3.894, 0.661)	0.161
BCC (cm)	TT	-1.188 (-2.487, 0.112)	0.073	-1.928 (-3.188, -0.668)	0.003
	TC/CC	1.319 (-1.215, 3.854)	0.303	0.391 (-1.750, 2.533)	0.716
BHC (cm)	TT	-0.532 (-1.402, 0.338)	0.228	-0.831 (-1.747, 0.086)	0.075
	TC/CC	0.464 (-1.391, 2.318)	0.464	-0.224 (-1.896, 1.448)	0.790
BPI (kg/m ³)	TT	-0.189 (-0.367, -0.012)	0.036	-0.242 (-0.429, -0.056)	0.011
	TC/CC	0.300 (0.032, 0.569)	0.029	0.321 (0.038, 0.605)	0.027
Exposure: PF	OA (ng/mL)				
BW (g)	TT	-136.6 (-378.0, 104.8)	0.265	-196.7 (-427.4, 34.0)	0.094
	TC/CC	-98.5 (-438.4, 241.4)	0.565	-108.0 (-420.6, 204.7)	0.493
BL (cm)	TT	0.077 (-0.968, 1.122)	0.884	-0.253 (-1.285, 0.799)	0.628
	TC/CC	-0.736 (-2.763, 1.291)	0.472	-1.044 (-2.949, 0.862)	0.278
BCC (cm)	TT	-0.207 (-1.273, 0.860)	0.702	-0.532 (-1.623, 0.560)	0.337
	TC/CC	-1.268 (-3.108, 0.573)	0.174	-0.870 (-2.639, 0.898)	0.329
BHC (cm)	TT	0.268 (-0.439, 0.975)	0.455	0.108 (-0.669, 0.886)	0.783
	TC/CC	-0.914 (-2.254, 0.425)	0.178	-0.674 (-2.054, 0.706)	0.333
BPI (kg/m ³)	TT	-0.136 (-0.280, 0.008)	0.064	-0.137 (-0.296, 0.021)	0.089
	TC/CC	-0.006(-0.209, 0.196)	0.951	0.049 (-0.196, 0.294)	0.692

1 Table 5. Associations between PFOS and PFOA levels and birth outcomes stratified by the maternal genotype *LXRB* (rs1405655) in female infants

The associations between PFOS or PFOA levels and birth outcomes were tested using a multiple linear regression model.

Crude: Non-adjusted.

Adjusted for maternal age (continuous), pre-pregnancy BMI (continuous), maternal smoking in the third trimester (yes/no), maternal alcohol consumption during pregnancy (yes/no), parity (primiparous/multiparous), educational level (less than or equal to high school graduation/more than high school graduation), annual household income (less than 5 million Japanese yen/more than or equal

to 5 million Japanese yen), cesarean section (yes/no), maternal blood sampling period (during pregnancy/after delivery), and gestational age (continuous).

β (95% CI) represents the change in BW (g), BL (cm), BCC (cm), BHC (cm), and BPI (kg/m³) for each 10-fold increase in PFOS or PFOA levels (ng/mL).

Supplementary Table 1. Sensitivity analysis of the associations between PFOS and PFOA levels and birth outcomes, stratified by infant sex, among participants whose maternal blood samples were collected during pregnancy (before delivery)
Total 1

	10ta					Males				Temates			
	Crude		Adjusted		Crude		Adjusted		Crude		Adjusted		
Outcome	β (95% CI)	p value	β (95% CI)	p value									
Exposure: PFOS (ng/mL)													
BW (g)	-137.2 (-350.7, 76.4)	0.207	-167.6 (-362.4, 27.2)	0.091	140.4 (-214.2, 495.1)	0.434	56.6 (-259.7, 372.8)	0.723	-317.7 (-586.4, -49.0)	0.021	-314.4 (-575.2, -53.6)	0.019	
BL (cm)	-0.329 (-1.546, 0.888)	0.595	-0.501 (-1.637, 0.634)	0.385	0.699 (-1.690, 3.089)	0.563	0.319 (-1.939, 2.577)	0.780	-1.117 (-2.337, 0.104)	0.073	-1.176 (-2.331, -0.020)	0.046	
BCC (cm)	-0.520 (-1.407, 0.366)	0.249	-0.623 (-1.431, 0.185)	0.130	0.342 (-1.084, 1.767)	0.635	-0.079 (-1.357, 1.199)	0.903	-1.024 (-2.181, 0.132)	0.082	-0.990 (-2.094, 0.115)	0.079	
BHC (cm)	-0.067 (-0.822, 0.688)	0.861	-0.125 (-0.871, 0.622)	0.743	0.302 (-0.988, 1.592)	0.644	0.061 (-1.212, 1.334)	0.924	-0.501 (-1.404, 0.402)	0.274	-0.440 (-1.376, 0.496)	0.354	
BPI (kg/m ³)	-0.026 (-0.311, 0.258)	0.857	-0.045 (-0.344, 0.253)	0.765	0.098 (-0.536, 0.732)	0.760	0.049 (-0.627, 0.725)	0.885	-0.097 (-0.289, 0.095)	0.321	-0.082 (-0.282, 0.118)	0.417	
Exposure: PFOA (ng/mL)													
BW (g)	-61.7 (-212.0, 88.6)	0.420	-50.7 (-197.9, 96.4)	0.497	98.5 (-144.2, 341.2)	0.423	-23.8 (-257.8, 210.2)	0.841	-174.8 (-368.4, 18.7)	0.076	-112.0 (-322.5, 98.5)	0.294	
BL (cm)	0.169 (-0.686, 1.024)	0.697	0.153 (-0.701, 1.007)	0.724	0.538 (-1.097, 2.172)	0.516	0.273 (-1.397, 1.943)	0.746	-0.176 (-1.058, 0.706)	0.694	0.082 (-0.849, 1.013)	0.862	
BCC (cm)	-0.227 (-0.851, 0.397)	0.474	-0.171 (-0.781, 0.439)	0.581	0.562 (-0.408, 1.533)	0.253	0.025 (-0.920, 0.971)	0.958	-0.735 (-1.560, 0.091)	0.081	-0.367 (-1.251, 0.517)	0.412	
BHC (cm)	-0.211 (-0.740, 0.319)	0.434	-0.213 (-0.773, 0.347)	0.454	-0.216 (-1.099, 0.667)	0.629	-0.600 (-1.534, 0.334)	0.205	-0.354 (-0.999, 0.290)	0.279	-0.222 (-0.965, 0.521)	0.555	
BPI (kg/m ³)	-0.078 (-0.278, 0.121)	0.440	-0.088 (-0.312, 0.136)	0.439	0.003 (-0.432, 0.437)	0.990	-0.151 (-0.650, 0.348)	0.550	-0.130 (-0.265, 0.006)	0.061	-0.117 (-0.274, 0.040)	0.143	

The associations between PFOS or PFOA levels and birth outcomes were tested using a multiple linear regression model.

Adjusted for maternal age (continuous), pre-pregnancy BMI (continuous), maternal smoking in the third trimester (yes/no), maternal alcohol consumption during pregnancy (yes/no), parity (primiparous/multiparous), educational level (less than or equal to high school graduation/more than high school graduation), annual household income (less than 5 million Japanese yen/more than or equal to 5 million Japanese yen), cesarean section (yes/no), maternal blood sampling pregnancy/after delivery), gestational age (continuous), and infant sex (male/female; total infants only). β (95% CI) represents the change in BW (g), BL (cm), BCC (cm), BHC (cm), and BPI (kg/m³) for each 10-fold increase in PFOS or PFOA levels (ng/mL).

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³ Crude: Non-adjusted. 4

		Crude		Adjusted	
Outcome	Exposure/sex	β (95% CI)	p value	β (95% CI)	p value
BW (g)	PFOS (ng/mL)	8.5 (-256.8, 273.7)	0.950	-85.7 (-313.2, 141.9)	0.460
	Female (vs. male)	19.9 (-243.2, 283.0)	0.882	-20.9 (-239.7, 197.9)	0.851
	PFOS × Sex (Interaction term)	-173.8 (-531.4, 183.8)	0.340	-171.4 (-468.4, 125.6)	0.257
BL (cm)	PFOS (ng/mL)	0.780 (-0.648, 2.208)	0.283	0.222 (-1.076, 1.520)	0.737
	Female (vs. male)	0.427 (-0.990, 1.843)	0.554	0.169 (-1.079, 1.417)	0.790
	PFOS × Sex (Interaction term)	-1.488 (-3.414, 0.437)	0.129	-1.372 (-3.066, 0.321)	0.112
BCC (cm)	PFOS (ng/mL)	0.166 (-1.030, 1.362)	0.785	-0.262 (-1.323, 0.799)	0.627
	Female (vs. male)	0.204 (-0.982, 1.391)	0.735	0.073 (-0.947, 1.093)	0.888
	PFOS × Sex (Interaction term)	-0.633 (-2.246, 0.980)	0.440	-0.643 (-2.028, 0.742)	0.362
BHC (cm)	PFOS (ng/mL)	-0.226 (-1.156, 0.704)	0.633	-0.205 (-1.119, 0.709)	0.660
	Female (vs. male)	-0.631 (-1.554, 0.291)	0.179	-0.625 (-1.504, 0.254)	0.163
	PFOS × Sex (Interaction term)	-0.128 (-1.382, 1.126)	0.841	-0.237 (-1.430, 0.956)	0.697
BPI (kg/m ³)	PFOS (ng/mL)	-0.077 (-0.346, 0.192)	0.575	-0.094 (-0.375, 0.187)	0.512
	Female (vs. male)	-0.039 (-0.305, 0.228)	0.776	-0.033 (-0.304, 0.237)	0.809
	PFOS × Sex (Interaction term)	0.040 (-0.323, 0.402)	0.830	0.025 (-0.342, 0.393)	0.892
BW (g)	PFOA (ng/mL)	-29.9 (-231.6, 171.7)	0.771	-115.2 (-293.1, 62.6)	0.203
	Female (vs. male)	-97.5 (-178.3, -16.6)	0.018	-143.0 (-211.1, -74.9)	< 0.001
	PFOA × Sex (Interaction term)	-100.3 (-375.8, 175.1)	0.474	15.0 (-215.8, 245.8)	0.898
BL (cm)	PFOA (ng/mL)	0.047 (-1.042, 1.135)	0.933	-0.493 (-1.506, 0.520)	0.339
	Female (vs. male)	-0.602 (-1.039, -0.166)	0.007	-0.813 (-1.200, -0.425)	< 0.001
	PFOA × Sex (Interaction term)	-0.300 (-1.788, 1.187)	0.692	0.157 (-1.158, 1.471)	0.815
BCC (cm)	PFOA (ng/mL)	0.165 (-0.743, 1.073)	0.721	-0.267 (-1.093, 0.559)	0.525
	Female (vs. male)	-0.194 (-0.558, 0.170)	0.295	-0.361 (-0.677, -0.045)	0.025
	PFOA × Sex (Interaction term)	-0.809 (-2.049, 0.431)	0.200	-0.326 (-1.398, 0.746)	0.551
BHC (cm)	PFOA (ng/mL)	-0.283 (-0.990, 0.424)	0.432	-0.414 (-1.124, 0.297)	0.253
	Female (vs. male)	-0.730 (-1.013, -0.446)	< 0.001	-0.813 (-1.085, -0.541)	< 0.001
	PFOA × Sex (Interaction term)	0.021 (-0.945, 0.986)	0.967	0.218 (-0.704, 1.140)	0.642
BPI (kg/m ³)	PFOA (ng/mL)	-0.016 (-0.220, 0.189)	0.880	-0.036 (-0.254, 0.183)	0.749
	Female (vs. male)	-0.007 (-0.089, 0.075)	0.866	-0.014 (-0.097, 0.070)	0.750
	PFOA × Sex (Interaction term)	-0.075 (-0.354, 0.204)	0.598	-0.033 (-0.317, 0.251)	0.820

Supplementary Table 2. Interactions between PFOS and PFOA levels and infant sex on birth outcomes 1

Abbreviations: BCC, birth chest circumference; BHC, birth head circumference; BL, birth length; BMI, body mass index; BPI, birth Ponderal index; BW, birth weight; CI, confidence interval; PFAS,

perfluoroalkyl substance; PFOS, perfluorooctanesulfonate; PFOA, perfluorooctanoic acid.

The associations between PFOS or PFOA levels and infant sex on birth outcomes were tested using a multiple linear regression model.

Crude: Non-adjusted.

Adjusted for maternal age (continuous), pre-pregnancy BMI (continuous), maternal smoking in the third trimester (yes/no), maternal alcohol consumption during pregnancy (yes/no), parity

(primiparous/multiparous), educational level (less than or equal to high school graduation/more than high school graduation), annual household income (less than 5 million Japanese yen/more than or equal

23456789to 5 million Japanese yen), cesarean section (yes/no), maternal blood sampling periods (during pregnancy/after delivery), and gestational age (continuous).

β (95% CI) represents the change in BW (g), BL (cm), BCC (cm), BHC (cm), and BPI (kg/m³).

10PFOS or PFOA levels were log₁₀-transformed scales.

The PFOS-sex or PFOA-sex interaction term was defined as "log₁₀-transformed PFOS or PFOA levels (continuous) \times sex (0 = male and 1 = female)."

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Supplementary Table 3. Sensitivity analysis of interactions between PFOS and PFOA levels and the maternal genotype LXRB (rs1405655) on birth 1

outcomes in female infants, among participants whose maternal blood samples were collected during pregnancy (before delivery) $\mathbf{2}$

		Crude		Adjusted	
Outcome	Exposure/genotype	β (95% CI)	p value	β (95% CI)	p value
BW (g)	PFOS (ng/mL)	-601.6 (-921.0, -282.1)	< 0.001	-592.1 (-906.1, -278.0)	< 0.001
	LXRB-TC/CC (vs. TT)	-655.5 (-1,077.7, -233.4)	0.003	-623.8 (-1,040.1, -207.5)	0.004
	PFOS × LXRB-TC/CC (Interaction term)	881.2 (322.3, 1,440.2)	0.002	836.0 (287.1, 1,384.9)	0.003
BL (cm)	PFOS (ng/mL)	-1.634 (-3.132, -0.135)	0.033	-1.694 (-3.131, -0.257)	0.021
	LXRB-TC/CC (vs. TT)	-0.910 (-2.890, 1.070)	0.365	-0.941 (-2.846, 0.964)	0.330
	PFOS × <i>LXRB</i> -TC/CC (Interaction term)	1.472 (-1.151, 4.094)	0.269	1.462 (-1.050, 3.974)	0.251
BCC (cm)	PFOS (ng/mL)	-2.166 (-3.540, -0.792)	0.002	-1.998 (-3.335, -0.661)	0.004
	LXRB-TC/CC (vs. TT)	-2.907 (-4.722, -1.092)	0.002	-2.522 (-4.294, -0.750)	0.006
	PFOS \times <i>LXRB</i> -TC/CC (Interaction term)	3.671 (1.268, 6.075)	0.003	3.148 (0.812, 5.485)	0.009
BHC (cm)	PFOS (ng/mL)	-0.732 (-1.812, 0.347)	0.182	-0.564 (-1.697, 0.570)	0.327
	LXRB-TC/CC (vs. TT)	-1.324 (-2.751, 0.102)	0.069	-1.091 (-2.593, 0.411)	0.153
	PFOS \times <i>LXRB</i> -TC/CC (Interaction term)	1.089 (-0.800, 2.979)	0.256	0.729 (-1.251, 2.710)	0.467
BPI (kg/m ³)	PFOS (ng/mL)	-0.264 (-0.494, -0.034)	0.025	-0.241 (-0.485, 0.003)	0.053
	LXRB-TC/CC (vs. TT)	-0.424 (-0.729, -0.120)	0.007	-0.383 (-0.707, -0.059)	0.021
	PFOS \times <i>LXRB</i> -TC/CC (Interaction term)	0.537 (0.135, 0.940)	0.009	0.490 (0.064, 0.917)	0.025
BW (g)	PFOA (ng/mL)	-219.1 (-467.7, 29.5)	0.084	-177.7 (-444.1, 88.7)	0.189
	LXRB-TC/CC (vs. TT)	-31.5 (-162.8, 99.8)	0.636	-35.9 (-163.6, 91.8)	0.579
	PFOA × <i>LXRB</i> -TC/CC (Interaction term)	127.0 (-277.9, 532.0)	0.536	177.3 (-218.6, 573.1)	0.377
BL (cm)	PFOA (ng/mL)	-0.076 (-1.210, 1.058)	0.895	0.006 (-1.176, 1.188)	0.992
	LXRB-TC/CC (vs. TT)	0.148 (-0.451, 0.747)	0.625	0.053 (-0.514, 0.619)	0.854
	PFOA × LXRB-TC/CC (Interaction term)	-0.325 (-2.172, 1.522)	0.728	0.142 (-1.614, 1.899)	0.873
BCC (cm)	PFOA (ng/mL)	-0.418 (-1.474, 0.638)	0.435	-0.118 (-1.235, 0.999)	0.835
	LXRB-TC/CC (vs. TT)	-0.169 (-0.726, 0.389)	0.551	-0.193 (-0.728, 0.343)	0.478
	PFOA \times <i>LXRB</i> -TC/CC (Interaction term)	-0.718 (-2.439, 1.002)	0.410	-0.453 (-2.112, 1.207)	0.590
BHC (cm)	PFOA (ng/mL)	0.015 (-0.789, 0.818)	0.971	0.131 (-0.781, 1.043)	0.776
	LXRB-TC/CC (vs. TT)	-0.463 (-0.888, -0.039)	0.033	-0.518 (-0.955, -0.081)	0.021
	PFOA \times <i>LXRB</i> -TC/CC (Interaction term)	-0.707 (-2.016, 0.602)	0.287	-0.477 (-1.832, 0.879)	0.488
BPI (kg/m ³)	PFOA (ng/mL)	-0.195 (-0.368, -0.022)	0.027	-0.173 (-0.372, 0.025)	0.087
	LXRB-TC/CC (vs. TT)	-0.048 (-0.140, 0.043)	0.297	-0.034 (-0.129, 0.061)	0.476
	$PFOA \times LXRB-TC/CC$ (Interaction term)	0.189 (-0.093, 0.471)	0.188	0.154 (-0.141, 0.449)	0.303

The associations between PFOS or PFOA levels and the maternal genotype LXRB (rs1405655) on birth outcomes were tested using a multiple linear regression model. 3

Crude: Non-adjusted.

Adjusted for maternal age (continuous), pre-pregnancy BMI (continuous), maternal smoking in the third trimester (yes/no), maternal alcohol consumption during pregnancy (yes/no), parity (primiparous/multiparous), educational level (less than or equal to high school graduation/more than high school graduation), annual household income (less than 5 million Japanese ven/more than or equal

to 5 million Japanese yen), cesarean section (yes/no), maternal blood sampling periods (during pregnancy/after delivery), and gestational age (continuous).

β (95% CI) represents the change in BW (g), BL (cm), BCC (cm), BHC (cm), and BPI (kg/m³).

456789PFOS or PFOA levels were log₁₀-transformed scales.

10 The PFOS-LXRB or PFOA-LXRB interaction term was defined as "log₁₀-transformed PFOS or PFOA levels (continuous) × genotype (0 = TT and 1 = TC/CC)."

1	Supplementary	y Table 4. Interactions between	PFOS levels and the maternal	genotype LXRB	(rs1405655) r	egarding fatty	acid levels in	female infants
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		Crude		Adjusted	
Outcome	Exposure/genotype	β (95% CI)	p value	β (95% CI)	p value
Triglyceride (mg/100-mL)	PFOS (ng/mL)	-0.187 (-0.327, -0.048)	0.009	-0.181 (-0.328, -0.033)	0.016
	LXRB-TC/CC (vs. TT)	0.140 (-0.042, 0.323)	0.131	0.120 (-0.068, 0.307)	0.210
	PFOS × LXRB-TC/CC (Interaction term)	-0.102 (-0.350, 0.146)	0.417	-0.085 (-0.339, 0.169)	0.512
Palmitic acid (µg/mL)	PFOS (ng/mL)	-0.114 (-0.230, 0.002)	0.053	-0.104 (-0.228, 0.020)	0.099
	LXRB-TC/CC (vs. TT)	0.054 (-0.097, 0.206)	0.478	0.058 (-0.100, 0.216)	0.472
	PFOS × LXRB-TC/CC (Interaction term)	-0.048 (-0.254, 0.157)	0.645	-0.053 (-0.267, 0.161)	0.625
Palmitoleic acid (µg/mL)	PFOS (ng/mL)	-0.192 (-0.361, -0.023)	0.026	-0.160 (-0.340, 0.019)	0.080
	LXRB-TC/CC (vs. TT)	0.049 (-0.171, 0.270)	0.659	0.058 (-0.170, 0.287)	0.616
	PFOS × LXRB-TC/CC (Interaction term)	0.005 (-0.295, 0.306)	0.971	-0.010 (-0.319, 0.300)	0.951
Stearic acid (µg/mL)	PFOS (ng/mL)	0.063 (-0.049, 0.175)	0.268	0.080 (-0.040, 0.199)	0.192
	LXRB-TC/CC (vs. TT)	0.028 (-0.119, 0.175)	0.705	0.030 (-0.123, 0.182)	0.703
	PFOS × LXRB-TC/CC (Interaction term)	-0.038 (-0.238, 0.161)	0.706	-0.042 (-0.249, 0.164)	0.687
Oleic acid (µg/mL)	PFOS (ng/mL)	-0.110 (-0.234, 0.014)	0.082	-0.089 (-0.222, 0.044)	0.190
	LXRB-TC/CC (vs. TT)	0.088 (-0.074, 0.251)	0.285	0.092 (-0.077, 0.262)	0.285
	PFOS × LXRB-TC/CC (Interaction term)	-0.110 (-0.331, 0.111)	0.328	-0.116 (-0.346, 0.113)	0.320
Linoleic acid (µg/mL)	PFOS (ng/mL)	-0.703 (-0.968, -0.437)	< 0.001	-0.681 (-0.964, -0.397)	< 0.001
	LXRB-TC/CC (vs. TT)	-0.116 (-0.463, 0.231)	0.509	-0.077 (-0.439, 0.284)	0.673
	PFOS × LXRB-TC/CC (Interaction term)	0.339 (-0.133, 0.810)	0.158	0.296 (-0.194, 0.785)	0.235
Linolenic acid (µg/mL)	PFOS (ng/mL)	-0.619 (-0.934, -0.303)	< 0.001	-0.594 (-0.930, -0.258)	0.001
	LXRB-TC/CC (vs. TT)	-0.081 (-0.492, 0.331)	0.700	-0.070 (-0.498, 0.357)	0.746
	PFOS × LXRB-TC/CC (Interaction term)	0.352 (-0.208, 0.913)	0.216	0.340 (-0.239, 0.920)	0.248
Arachidonic acid (µg/mL)	PFOS (ng/mL)	-0.438 (-0.711, -0.164)	0.002	-0.396 (-0.687, -0.105)	0.008
	LXRB-TC/CC (vs. TT)	-0.014 (-0.372, 0.343)	0.938	0.054 (-0.318, 0.425)	0.776
	PFOS × LXRB-TC/CC (Interaction term)	0.192 (-0.294, 0.678)	0.438	0.110 (-0.392, 0.613)	0.665
Eicosapentaenoic acid (µg/mL)	PFOS (ng/mL)	0.120 (-0.173, 0.413)	0.422	0.123 (-0.191, 0.436)	0.441
	LXRB-TC/CC (vs. TT)	0.130 (-0.253, 0.513)	0.503	0.086 (-0.313, 0.485)	0.671
	PFOS × LXRB-TC/CC (Interaction term)	-0.182 (-0.702, 0.339)	0.492	-0.143 (-0.684, 0.398)	0.602
Docosahexaenoic acid (µg/mL)	PFOS (ng/mL)	-0.189 (-0.491, 0.113)	0.220	-0.154 (-0.478, 0.170)	0.350
	LXRB-TC/CC (vs. TT)	0.022 (-0.373, 0.417)	0.913	0.053 (-0.361, 0.466)	0.801
	PFOS \times <i>LXRB</i> -TC/CC (Interaction term)	0.195 (-0.342, 0.732)	0.475	0.154 (-0.405, 0.714)	0.587

The association between PFOS levels and maternal genotype LXRB (rs1405655) on fatty acid levels were tested using a multiple linear regression model.

Crude: Non-adjusted.

Adjusted for maternal age (continuous), pre-pregnancy BMI (continuous), maternal smoking in the third trimester (yes/no), maternal alcohol consumption during pregnancy (yes/no), parity (primiparous/multiparous), educational level (less than or equal to high school graduation/more than high school graduation), annual household income (less than 5 million Japanese yen/more than or equal to 5 million Japanese yen), and maternal blood sampling periods (during pregnancy/after delivery).

 β (95% CI) represents the change in log₁₀-transformed fatty acid levels. PFOS and fatty acid levels were log₁₀-transformed scales. The PFOS-*LXRB* interaction term was defined as "log₁₀-transformed PFOS levels (continuous) × genotype (0 = TT and 1 = TC/CC)."

Supplementary Table 5. Sensitivity analysis of associations between PFOS and PFOA levels and birth outcomes stratified by the maternal genotype LXRB 1 (rs1405655) in female infants, among participants whose maternal blood samples were collected during pregnancy (before delivery) $\mathbf{2}$

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		Crude		Adjusted	
Outcome	Genotype	β (95% CI)	p value	β (95% CI)	p value
Exposure: PF	OS (ng/mL)				
BW (g)	TT	-601.6 (-947.8, -255.4)	0.001	-641.6 (-976.3, -306.9)	< 0.001
	TC/CC	279.7 (-112.1, 671.5)	0.157	259.4 (-164.0, 682.7)	0.222
BL (cm)	TT	-1.634 (-3.159, -0.108)	0.036	-1.774 (-3.293, -0.255)	0.023
	TC/CC	-0.162 (-2.303, 1.979)	0.880	-0.603 (-2.736, 1.529)	0.569
BCC (cm)	TT	-2.166 (-3.660, -0.672)	0.005	-2.348 (-3.845, -0.851)	0.003
	TC/CC	1.505 (-0.161, 3.171)	0.075	1.069 (-0.466, 2.605)	0.166
BHC (cm)	TT	-0.732 (-1.757, 0.292)	0.159	-0.841 (-1.964, 0.282)	0.140
	TC/CC	0.357 (-1.373, 2.087)	0.679	0.151 (-1.567, 1.869)	0.859
BPI (kg/m ³)	TT	-0.264 (-0.501, -0.028)	0.029	-0.267 (-0.525, -0.008)	0.044
	TC/CC	0.273 (-0.050, 0.596)	0.095	0.327 (-0.026, 0.680)	0.068
Exposure: PF	OA (ng/mL)				
BŴ(g)	TT	-219.1 (-490.9, 52.7)	0.113	-164.2 (-463.9, 135.5)	0.278
	TC/CC	-92.0 (-357.6, 173.6)	0.488	-56.6 (-367.1, 253.8)	0.713
BL (cm)	TT	-0.076 (-1.242, 1.091)	0.897	-0.067 (-1.228, 1.362)	0.918
	TC/CC	-0.401 (-1.822, 1.020)	0.572	0.013 (-1.527, 1.553)	0.986
BCC (cm)	TT	-0.418 (-1.582, 0.746)	0.477	-0.327 (-1.637, 0.983)	0.620
	TC/CC	-1.136 (-2.233, -0.039)	0.043	-0.357 (-1.486, 0.772)	0.525
BHC (cm)	TT	0.015 (-0.757, 0.786)	0.970	-0.032 (-0.969, 0.905)	0.946
	TC/CC	-0.692 (-1.827, 0.442)	0.225	-0.115 (-1.350, 1.119)	0.851
BPI (kg/m ³)	TT	-0.195 (-0.371, -0.019)	0.030	-0.166 (-0.381, 0.049)	0.128
	TC/CC	-0.006 (-0.229, 0.216)	0.953	0.050 (-0.316, 0.216)	0.706

The associations between PFOS or PFOA levels and birth outcomes were tested using a multiple linear regression model.

Crude: Non-adjusted.

Adjusted for maternal age (continuous), pre-pregnancy BMI (continuous), maternal smoking in the third trimester (yes/no), maternal alcohol consumption during pregnancy (yes/no), parity (primiparous/multiparous), educational level (less than or equal to high school graduation/more than high school graduation), annual household income (less than 5 million Japanese ven/more than or equal to 5 million Japanese yen), cesarean section (yes/no), maternal blood sampling periods (during pregnancy/after delivery), and gestational age (continuous).

β (95% CI) represents the change in BW (g), BL (cm), BCC (cm), BHC (cm), and BPI (kg/m³) for each 10-fold increase in PFOS or PFOA levels (ng/mL).

				Crude		Adjusted	
Outcome	Maternal genotype	PFOS level (ng/mL)	n _{Female}	β (95% CI)	p value	β (95% CI)	p value
BW (g)	TT	Quartile 1 (<3.7)	38	0.0 (Reference)		0.0 (Reference)	
		Quartile 2 (3.7 to <5.2)	27	-12.9 (-204.4, 178.5)	0.894	-58.0 (-219.9, 103.8)	0.479
		Quartile 3 (5.2 to <7.2)	26	-51.3 (-244.9, 142.3)	0.601	-115.9 (-279.1, 47.4)	0.162
		Quartile 4 (≥7.2)	32	-219.5 (-402.0, -37.0)	0.019	-306.1 (-475.9, -136.2)	0.001
		p for trend (p_{trend})			0.021		0.001
	TC/CC	Quartile 1 (<3.7)	16	0.0 (Reference)		0.0 (Reference)	
		Quartile 2 (3.7 to <5.2)	16	-0.3 (-287.4, 286.9)	0.999	22.0 (-213.6, 257.7)	0.852
		Quartile 3 (5.2 to <7.2)	22	68.1 (-198.8, 335.0)	0.613	84.5 (-135.7, 304.7)	0.446
		Quartile 4 (≥7.2)	21	225.1 (-44.4, 494.6)	0.100	119.7 (-104.5, 344.0)	0.290
		p for trend (p_{trend})			0.075		0.232
BCC (cm)	TT	Quartile 1 (<3.7)	38	0.000 (Reference)		0.000 (Reference)	
		Quartile 2 (3.7 to <5.2)	27	-0.134 (-0.983, 0.715)	0.755	-0.379 (-1.152, 0.394)	0.334
		Quartile 3 (5.2 to <7.2)	26	-0.200 (-1.058, 0.658)	0.645	-0.491 (-1.271, 0.289)	0.215
		Quartile 4 (≥7.2)	32	-0.839 (-1.648, -0.030)	0.042	-1.242 (-2.054, -0.431)	0.003
		p for trend (p_{trend})			0.050		0.004
	TC/CC	Quartile 1 (<3.7)	16	0.000 (Reference)		0.000 (Reference)	
		Quartile 2 (3.7 to <5.2)	16	-0.069 (-1.625, 1.487)	0.930	-0.094 (-1.428, 1.241)	0.889
		Quartile 3 (5.2 to <7.2)	22	-0.198 (-1.644, 1.248)	0.785	-0.138 (-1.385, 1.109)	0.826
		Quartile 4 (≥7.2)	21	1.209 (-0.252, 2.669)	0.103	0.571 (-0.699, 1.841)	0.372
		p for trend (p_{trend})			0.369		0.175
BPI (kg/m ³)	TT	Quartile 1 (<3.7)	38	0.000 (Reference)		0.000 (Reference)	
		Quartile 2 (3.7 to <5.2)	27	-0.063 (-0.180, 0.053)	0.284	-0.079 (-0.194, 0.036)	0.175
		Quartile 3 (5.2 to <7.2)	26	-0.085 (-0.203, 0.033)	0.157	-0.093 (-0.209, 0.023)	0.116
		Quartile 4 (≥7.2)	32	-0.110 (-0.222, 0.001)	0.051	-0.144 (-0.265, -0.023)	0.020
		p for trend (p_{trend})			0.045		0.018
	TC/CC	Quartile 1 (<3.7)	16	0.000 (Reference)		0.000 (Reference)	
		Quartile 2 (3.7 to <5.2)	16	0.104 (-0.063, 0.271)	0.218	0.130 (-0.045, 0.304)	0.142
		Quartile 3 (5.2 to <7.2)	22	0.179 (0.024, 0.334)	0.024	0.221 (0.058, 0.384)	0.009
		Quartile 4 (\geq 7.2)	21	0.188 (0.031, 0.345)	0.019	0.203 (0.037, 0.369)	0.017
		p for trend (p_{trend})			0.013		0.011

1 Supplementary Table 6. Associations of PFOS quartile levels with birth weight, birth chest circumference, and birth Ponderal index among female infants 2 stratified by the maternal genotype *LXRB* (rs1405655)

The associations between PFOS levels and birth outcomes were tested using a multiple linear regression model.

4 Crude: Non-adjusted.

Adjusted for maternal age (continuous), pre-pregnancy BMI (continuous), maternal smoking in the third trimester (yes/no), maternal alcohol consumption during pregnancy (yes/no), parity (primiparous/multiparous), educational level (less than or equal to high school graduation/more than high school graduation), annual household income (less than 5 million Japanese yen/more than or equal

to 5 million Japanese yen), cesarean section (yes/no), maternal blood sampling periods (during pregnancy/after delivery), and gestational age (continuous).

 β (95% CI) represents the change in BW (g), BCC (cm), and BPI (kg/m³) compared to the reference group.

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