

Agricultural viromics: from continental viromes to field level molecular diagnostics

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International Center for Agricultural Research in the Dry Areas (ICARDA)



International Livestock

Research Institute (ILRI)



International Food Policy Research Institute (IFPRI)

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International Maize and Wheat Improvement Center (CIMMYT)





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International Rice Research Institute (IRRI)



International Institute of Tropical Agriculture (IITA)



International Potato Center (CIP)



The Alliance of Bioversity International and the International Center for Tropical Agriculture (CIAT)





Plant Health Management under Changing Climates



Need for a Revitalized Strategy and Innovative Approaches



Next generation diagnostics

Better understanding of the major diseases contributing to seed degeneration, their epidemiology and cheap rapid accurate diagnostics



RNA sequencing and assembly for virus identification and discovery



RNA sequencing and assembly for virus identification and discovery



Plant infected with	siRNAs sequenced	Contigs identified	Contigs with Blastx hits 21–24 nt sRNA k = 15, cov = 30 ^a	Contig sizes $21-24$ nt sRNA $k = 15$, cov = 30^{a}	Contigs with Blastx hits 21–24 nt sRNA k=15, cov = 3 ^b	Contigs with Blastx hits only 22 nt sRNA k = 15, cov = 3 ^b
SPFMV	1,275,673	Total contigs	239		1633	431
		SPFMV	25	(≤949)	71	78
		SPCSV	0	-	0	0
		Badnavirus	38	(≤256)	62	55
		Mastrevirus	5	(≤210)	6	5
SPCSV	1,271,382	Total contigs	283		1675	285
		SPFMV	0	-	0	0
		SPCSV	10	(≤70)	64	12
		Badnavirus	44	(≤181)	63	44
		Mastrevirus	8	(≤260)	10	6
SPFMV + SPCSV	1,067,577	Total contigs	221		1363	581
		SPFMV	20	(≤1600)	43	51
		SPCSV	2	(≤81)	41	12
		Badnavirus	38	(≤266)	63	51
		Mastrevirus	5	(≤210)	8	5

Number of contigs assembled by Velvet using 21-24 nt sRNA, or only 22 nt sRNA sequences, with virus specific hits as identified using Translated Nucleotide Blast (Blastx).



SNP landscape highly similar between siRNA and RNA from virus particles



The African sweetpotato virome

~2000 samples and bioinformatics pipeline for virus identification using siRNA assembly and genome subtraction



Small RNA sequence analysis pipeline: VirusDetect_v1.6



Reference	Length	Coverage (%)	#contig	Depth	Depth (Norm)	%Identity	%lden Max	%lden Min	Genus	Descript	Description	
FJ150422	4806	4806 (100)	11	675.9	18.5	94.64	96.00	84.82	NA	Drosophila A virus isolate HD, complete genome.		
GQ342962	3260	3257 (99.9)	3	524.3	14.4	98.11	98.75	97.03	NA	Drosophila melanogaster birnavirus SW-2009a strain D segment A, complete sequen		
GQ342963	3014	3014 (100)	1	854.8	23.4	98.47	98.47	98.47	NA	Drosophila melanogaster birnavirus SW-2009a strain Di segment B, complete sequen		
KF947078	13534	1773 (13.1)	27	35.2	1.0	99.10	100	97.14	NA	Spodoptera frugiperda rhabdovirus isolate Sf, comple genome.		
GQ257737	12333	12333 (100)	6	661.2	18.1	96.87	99.29	91.67	NA	Nora virus isolate Umea 2007 complete genome.		
M32779	2225	2112 (94.9)	7	20.4	0.6	99.57	100	97.69	alphabaculovirus	Autographa californica nucleopolyhedrovirus insertio element IFP2.2 genomic sequence.		
EF690537	3107	3089 (99.4)	5	654.2	17.9	94.82	96.59	94.12	alphanodavirus	Flock house virus isolate TN0 segment RNA1 protein A mR complete cds.		
EF690538	1383	1383 (100)	1	378.8	10.4	94.44	94.44	94.44	alphanodavirus	Flock house virus isolate TNC segment RNA2 protein alpha mRNA, complete cds.		
AF014388	9264	9244 (99.8)	3	1096.2	30.1	98.03	98.21	96.88	cripavirus	Drosophila C virus strain EB, complete genome.		
↔ ok AF	014388	- 1k	- 2k	<u>-</u> 3k		4k 4k CONTIG220	++ +++ 5k	 6k		 8k		→ ■
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Order	Query ID	Que	ry Start	Quer	y End	Subjet St						
Order	Query ID CONTIG	Que	ry Start	Quer 1121	y End	8144	ent .	9264	1086/1121	(96%)	0.0	1
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Order 1 Alignme Query Sbjct	Query ID CONTIGS ent: : 1 : 8144	Que 50 1 tgaggttga iiiiiiiii tgaggttga	aatatgct	Quer 1121	y End	Subjet St 8144 caagtaatg	gggtga	agatgta agattta	1086/1121 agctattcaaag	60 8203	0.0	1

Sbjct: 8204 aaacgatgctcaacatggtgttcatccaatgactatagacactcataagatcgactcaaa 8263

http://bioinfo.bti.cornell.edu/virome/index



Results: the viruses 3193 viruses from 1168 samples



ssDNA satellite

Viroid 0.03%



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e-Xtra*

Badnaviruses of Sweet Potato: Symptomless Coinhabitants on a Global Scale

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FIGURE 1 Instatute suppression of SPPV-A and SPPV-B in leaves in coinfection with SPFMV, SPCSV, or both viruses (SPVD) relative to plants infected only with SPPV (only SPPV). Error bars indicate standard error of relative expression. "Significantly upregulated as compared to plants infected only by SPPV (only SPPV; p = 0.001).



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Can we make predictions about likely occurrence beyond evaluated areas?

Example for begomoviruses: Niche analysis using MaxEnt modeling and bioclimatic-ecosystem variables



Field LAMP kit









The Peruvian potato virome

• Sample collection: 994 geo-referenced potato leaf samples (Cajamarca, Huanuco, Junin, Huancavelica, Cusco)



Leaves



Weighting



General results



Are any of the new viruses of relevance?



Rugose stunting disease of potato: the history of the emergence and decline of a viral disease

- Observed since 1990 in Arequipa, Moquegua and Tacna and studied as SB26 and SB29
- Isometric virus, transmitted by transmitted by brown leafhoppers (*Russelliana solanicola*) and generated losses of between 35-85% depending on the variety
- Farmers changed their planting dates to manage the disease or quit potato al together and it is no longer considered a problem
- It was never determined what type of virus it was



Potato rugose stunting virus is a torradovirus and present in low frequencies in potatoes throughout Peru







The International Potato Center (known by its Spanish acronym CIP) is a research-for-development organization with a focus on potato, sweetpotato, and Andean roots and tubers. CIP is dedicated to delivering sustainable science-based solutions to the pressing world issues of hunger, poverty, gender equity, climate change and the preservation of our Earth's fragile biodiversity and natural resources. www.cipotato.org



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