

December 2022

## Therapies for Mitochondrial Disorders

Kayli Sousa Smyth

*Technological University of the Shannon: Midlands Midwest, kaylisousasmyth@gmail.com*

Anne Mulvihill

*Technological University of the Shannon: Midlands Midwest, ammulvihill@ait.ie*

Follow this and additional works at: [https://arrow.tudublin.ie/sure\\_j](https://arrow.tudublin.ie/sure_j)

 Part of the [Alternative and Complementary Medicine Commons](#), [Animal Structures Commons](#), [Biochemistry Commons](#), [Biological Factors Commons](#), [Biology Commons](#), [Biotechnology Commons](#), [Cell Anatomy Commons](#), [Cell Biology Commons](#), [Cells Commons](#), [Developmental Biology Commons](#), [Embryonic Structures Commons](#), [Medical Education Commons](#), [Medical Sciences Commons](#), [Molecular Biology Commons](#), [Nucleic Acids, Nucleotides, and Nucleosides Commons](#), [Other Analytical, Diagnostic and Therapeutic Techniques and Equipment Commons](#), [Other Biochemistry, Biophysics, and Structural Biology Commons](#), [Other Cell and Developmental Biology Commons](#), [Physical Sciences and Mathematics Commons](#), [Surgical Procedures, Operative Commons](#), and the [Therapeutics Commons](#)

### Recommended Citation

Smyth, Kayli Sousa and Mulvihill, Anne (2022) "Therapies for Mitochondrial Disorders," *SURE\_J: Science Undergraduate Research Journal*: Vol. 4: Iss. 1, Article 3.

Available at: [https://arrow.tudublin.ie/sure\\_j/vol4/iss1/3](https://arrow.tudublin.ie/sure_j/vol4/iss1/3)

This Article is brought to you for free and open access by the Current Publications at ARROW@TU Dublin. It has been accepted for inclusion in SURE\_J: Science Undergraduate Research Journal by an authorized administrator of ARROW@TU Dublin. For more information, please contact [arrow.admin@tudublin.ie](mailto:arrow.admin@tudublin.ie), [aisling.coyne@tudublin.ie](mailto:aisling.coyne@tudublin.ie), [gerard.connolly@tudublin.ie](mailto:gerard.connolly@tudublin.ie).



This work is licensed under a [Creative Commons Attribution-NonCommercial-Share Alike 4.0 License](#)

---

## Therapies for Mitochondrial Disorders

### Cover Page Footnote

Firstly, I would like to thank my incredibly supportive, kind and knowledgeable supervisor Dr Anne Mulvihill, for all the encouragement and motivation that facilitated me in writing this article. Her unconditional praise and positive reassurance made me believe in myself and in my ability to complete this project in a timely manner. I am eternally grateful and privileged to have had Dr Anne Mulvihill as my supervisor, and for her overall guidance throughout my entire academic journey. This includes her encouragement towards my masters application, to study my passion for genetic and genomic counselling. In addition, I would like to thank my parents for their continuous support and review of this article. In particular, my father 'Roger Smyth' who helped me think of better ways of phrasing sentences, as well as teaching me how to avoid grammatical error. Their unconditional support and positive regard meant a lot to me along the way. Lastly, I would like to thank Dr Anne Friel for inviting me to participate in the SUREJ article writing. This has been a wonderful opportunity that I am very grateful for, and my scientific writing has improved 100 fold throughout the journey.

# Therapies for Mitochondrial Disorders

Kayli Sousa Smyth<sup>1\*</sup>, Anne Mulvihill<sup>1</sup>

<sup>1</sup> Department of Pharmaceutical Sciences & Biotechnology, Technological University of the Shannon: Midlands  
Midwest, Athlone, Co. Westmeath, Ireland

\*Corresponding Author e-mail: kaylisousasmyth@gmail.com>

Received 24<sup>th</sup> August 2022, Accepted for publication 14<sup>th</sup> November, Published 22<sup>nd</sup> December 2022

## Abstract

Mitochondria are cytoplasmic, double-membrane organelles that synthesise adenosine triphosphate (ATP). Mitochondria contain their own genome, mitochondrial DNA (mtDNA), which is maternally inherited from the oocyte. Mitochondrial proteins are encoded by either nuclear DNA (nDNA) or mtDNA, and both code for proteins forming the mitochondrial oxidative phosphorylation (OXPHOS) complexes of the respiratory chain. These complexes form a chain that allows the passage of electrons down the electron transport chain (ETC) through a proton motive force, creating ATP from adenosine diphosphate (ADP). This study aims to explore current and prospective therapies for mitochondrial disorders (MTDS). MTDS are clinical syndromes coupled with abnormalities of the ETC and OXPHOS, caused by pathogenic variants in mtDNA or nDNA. Many MTDS emerge from either homoplasmic or heteroplasmic mutations of the DNA, and include mitochondrial encephalomyopathy, lactic acidosis and stroke-like episodes. Current therapies include increasing mitochondrial biogenesis, the use of antioxidants, dietary approaches, and exercise. However, these are mostly symptomatic and supportive therapies. Future therapies comprise of personalised and precision medicine approaches which include gene, mitochondrial, cell, and *in utero*-based therapies. Obstacles towards discovering effective therapies include the rarity of MTDS, its pathogenic complexity and lack of clinical trials. Despite the lack of current curative therapies for MTDS, emerging therapies promise exciting and clinically meaningful therapies in the future.

Keywords: Mitochondrial diseases; Mitochondria; mtDNA; Oxidative phosphorylation; Therapies; ROS

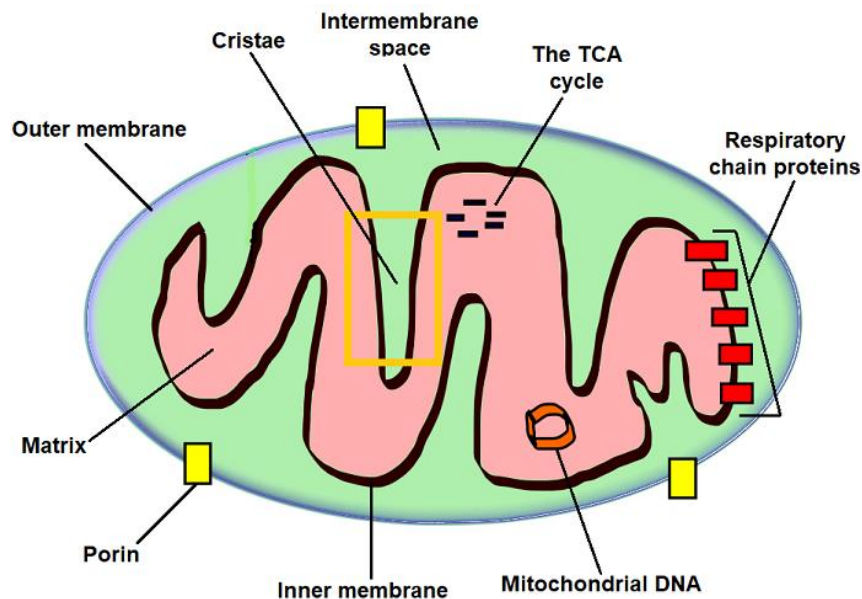
## 1. Introduction

Mitochondria are cytoplasmic, double-membrane organelles in eukaryotic cells, derived originally from bacteria by endosymbiosis (Zeviani and Donato, 2004). Their main purpose is to synthesise adenosine triphosphate (ATP) which is crucial for eukaryotic life. They are also unique as organelles, because they contain their own genome (Zeviani and Donato, 2004; Annesley and Fisher, 2019).

### 1.1 Structure

The structure of mitochondria, as well as its mitochondrial protein and lipid arrangement, determines the capability of mitochondrial function in a cell. The efficiency of mitochondrial function is limited by the number of mitochondria in a cell, its inner elements, and its capability to obtain inputs and supply required products (Glancy, 2020).

Mitochondria consist of an inner lumen/matrix surrounded by a double membrane, as shown in **Figure 1**. This double membrane consists of an outer mitochondrial membrane (OMM) and an inner mitochondrial membrane (IMM), which separate the lumen of the mitochondria from the cytoplasm (Kühlbrandt, 2015).



**Figure 1. General structure of the mitochondria.** The outer and inner membrane split the mitochondria into two sections, the matrix and the intermembrane space (Kühlbrandt, 2015). Porins in the outer membrane allow exchange of small metabolites between the cytosol and mitochondria (Endo and Sakaue, 2019). The tricarboxylic acid (TCA) cycle components and mitochondrial DNA are embedded within the matrix. Within the cristae folds are the machinery for oxidative phosphorylation (OXPHOS), the main mitochondrial function. The respiratory chain proteins are found on the edge of the matrix and are discussed in section 1.2.2.

### 1.1.1 The outer membrane (TOM)

TOM is porous to ions and small uncharged molecules  $< 5,000$  Da, that pass through the pore-forming membrane proteins (porins), as shown in **Figure 1**. Due to TOM's porosity, no membrane potential exists across this membrane (Kühlbrandt, 2015; Supinski *et al.*, 2020).

Most mitochondrial proteins are made in the cytoplasm and are imported into the organelle. These larger proteins require special translocases to enter the mitochondria. Translocases consist of proteins in both the OMM and IMM (Hewitt *et al.*, 2013; Kühlbrandt, 2015).

The composition of the IMS is analogous to the cytosol in terms of ions and small molecules. As such, it acts as a functional barrier towards the passage of small molecules between the cytosol and matrix, whilst maintaining the protein gradient that drives OXPHOS. This is what allows for normal mitochondrial function (Cooper, 2000; Kühlbrandt, 2015; Suspinsky *et al.*, 2020).

### 1.1.2 The inner membrane (TIM)

TIM has many folds named cristae that extend into the matrix, shown in **Figure 1**. It contains the enzymes involved in energy conversion and reactive oxygen species (ROS) formation processes (Glancy, 2020). In addition, it acts as an anchor for elements of the electron transport chain (ETC) (Supinski *et al.*, 2020). It also contains ATP synthase, a channel that uses a proton motive force to drive ATP synthesis (Musante *et al.*, 2007).

The inner compartment, enclosed by TIM, is called the mitochondrial matrix, shown in **Figure 1**. The matrix has a large protein mass of up to 5000mg/ml and a pH of 7.9-8, creating the transmembrane electrochemical gradient that drives ATP synthesis (Kühlbrandt, 2015).

### 1.1.3 Mitochondrial genome

Mitochondria have their own genome comprised of circular deoxyribonucleic acid (DNA) molecules (Nass and Nass, 1963, cited by Kauppila *et al.*, 2017, p. 57). The mammalian mitochondrial proteome comprises approximately 1,200 proteins, nearly all encoded by nuclear DNA (nDNA), which are translated in the cytosol and introduced into mitochondria (Kauppila *et al.*, 2017). The performance of mitochondria relies on coordinated synthesis of nDNA and mitochondrial DNA (mtDNA) encoded proteins, for proper structure and function (Glancy, 2020; Supinski *et al.*, 2020).

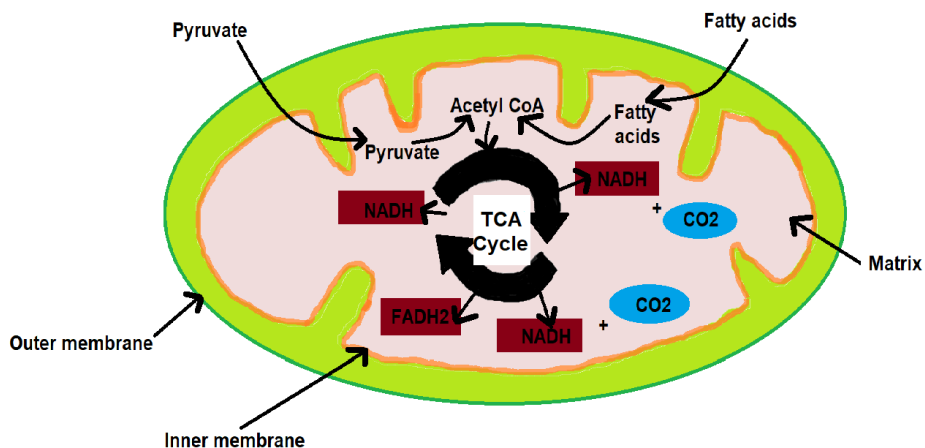
Mitochondria are semi-autonomous, as the mtDNA encodes around 1% of the mitochondrial proteome, and these thirteen proteins are crucial for function. The mitochondrial genome is inherited from the mother who transmits her oocyte mtDNA to her offspring, and so forth (Zeviani and Donato, 2004).

Mutations in the nuclear genome can lead to mitochondrial disorders (MTDS), including the diminishing of mtDNA *e.g.*, mutations in the gene for the mtDNA polymerase  $\gamma$  (POLG) (Guerra *et al.*, 2017). POLG is involved in a subunit of DNA polymerase gamma, the only DNA polymerase associated with replication and repair of mtDNA (Kanungo *et al.*, 2018). Mutations in mtDNA can have serious consequences for the mitochondria and its function. Heteroplasmy is where two or more differing mtDNA variants coexist inside a cell and is a frequent cause of mtDNA disorders. Homoplasmy is where all mtDNA within a cell are identical, either normal or mutant (Shalem and Riikka, 2020).

## 1.2 Functions of mitochondria

Mitochondria play a crucial role in the manufacture of metabolic energy in eukaryotic cells and are accountable for most of the valuable energy obtained from the degradation of carbohydrates and fatty acids (Cooper, 2000). This energy is in the form of ATP, an energy-rich compound that drives the main cellular functions including force generation, folding/breaking down of proteins and manufacturing/maintaining membrane potentials (Kühlbrandt, 2015).

Aside from ATP synthesis, the mitochondria have other functional roles including the manufacture of nicotinamide adenine dinucleotide (NADH) and guanosine triphosphate (GTP) in the TCA cycle (**Figure 2**), biosynthesis of amino acids, cellular signalling hubs and the production of phospholipids for membrane biogenesis (Kühlbrandt, 2015).



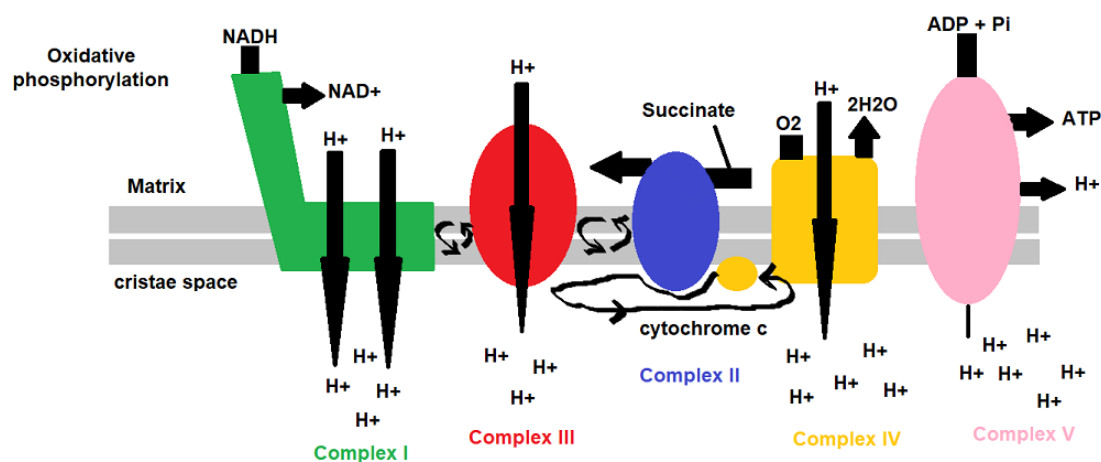
**Figure 2. The TCA cycle.** This occurs in the mitochondrial matrix and creates the reducing power used in OXPHOS, as well as metabolites used in other pathways. Macromolecules, such as fatty acids, must be broken down into acetyl coenzyme A (CoA) before entering the TCA cycle (Cooper, 2000).

### 1.2.1 Oxidative metabolism

The primary stages of glucose metabolism (glycolysis) occur in the cytosol, converting glucose to pyruvate, which is carried into mitochondria to complete oxidation to carbon dioxide ( $\text{CO}_2$ ) (Blanco and Blanco, 2017). Primary oxidation of pyruvate to CoA, shown in **Figure 2**, is degraded to  $\text{CO}_2$  through the TCA cycle (Cooper, 2000). The pyruvate dehydrogenase complex catalyses the conversion of pyruvate to acetyl CoA, making NADH and  $\text{CO}_2$  when in the presence of CoA, nicotinamide adenine dinucleotide and triphenylphosphonium (Park *et al.*, 2018).

### 1.2.2 Respiratory chain complexes

Mitochondrial OXPHOS complexes of the respiratory chain, known as NADH, include ubiquinone oxidoreductase (complex I), succinate dehydrogenase (complex II), ubiquinone-cytochrome c oxidoreductase (complex III), cytochrome c oxidase (complex IV) and ATP synthase (complex V), shown in **Figure 3** (Sharma *et al.*, 2009).



**Figure 3. The five components of the mitochondrial ETC.** Complex I oxidises NADH from the TCA cycle, then oxidised ubiquinone reduces ubiquinol via the acceptance of two electrons. In complex II, ubiquinol is re-oxidised, then transfers electrons to reduce oxygen to water in

complex IV. This releases redox energy, which delivers protons from the mitochondrial matrix to the periplasmic space, generating a proton-motive force across the IMM. Complex V uses this force (created by hydrogen ions) to generate ATP from adenosine diphosphate (ADP) and inorganic phosphate, constituting OXPHOS (Sharma *et al.*, 2009).

### 1.2.3 Mitochondrial fission and fusion

Mitochondria are dynamic organelles that are continually undergoing fusion and fission. Fusion combines elements between the mitochondrial contents of a cell, avoiding the complete loss of important elements (Youle and van der Bliek, 2012). It results in the creation of an individual mitochondrion from previous independent structures (Scott and Youle, 2010). Fusion is facilitated by mitofusion 1 and mitofusion 2 in the OMM and optic atrophy 1 (localised in the IMM), which functions to fuse internal/external membranes (Supinski *et al.*, 2020). Mitofusion 1 and 2 are dynamin-like GTP hydrolase enzymes, critical to mitochondrial function, due to their ability to stabilise fission/fusion dynamics (Sidarala *et al.*, 2022).

Fission creates new organelles and helps quality control within the mitochondria. It is facilitated by the formation of a multimeric complex comprising of dynamic-related protein 1, which exerts a mechanical force around the OMM, cutting it into two pieces (Supinski *et al.*, 2020). It is crucial that cytochrome c is not released during mitochondrial fission/fusion, as it assists with ATP synthesis. If apoptosis occurs, cytochrome c can disperse into the cytosol and trigger apoptosis of the cell (Halestrap *et al.*, 1997; Ow *et al.*, 2008).

### 1.2.4 Mitochondrial biogenesis (MB)

MB is the creation of new mitochondria in cells. The main controller of MB is a transcriptional coactivator, peroxisome proliferator-initiated receptor gamma coactivator 1- $\alpha$  (PGC1 $\alpha$ ). Disease mechanisms hinder MB, damaging the upkeep of appropriate levels of proteins *e.g.*, a mutated TOM complex seen in patients with Alzheimer's disease (Goyal and Chaturvedi, 2021). Treatments for MTDS include therapies that initiate MB and mitochondrial restoration, such as administration of resveratrol (sirtuin 1 activator) or human recombinant transcription factor a (Supinski *et al.*, 2020).

## 2. Mitochondrial disorders (MTDS)

MTDS refer to clinical syndromes linked with abnormalities of the ETC and OXPHOS. Clinical manifestations include mtDNA/nDNA depletion, muscle weakness and pigmentary retinopathy. The most common MTDS can be seen below in Table 1 (Zeviani and Donato, 2004). Primary mitochondrial diseases (PMD) are a heterogeneous clinical cluster of disorders that occur due to dysfunction of the mitochondrial respiratory chain (MRC) (Chinnery, 2000). Deficiencies of almost 400 genes across the two genomes have been associated with PMD (Pitceathly *et al.*, 2020).

**Table 1. Common MTDS (Zeviani and Donato, 2004).**

<b>Types of MTDS</b>	<b>Phenotype</b>	<b>mtDNA mutation</b>
Kearns-Sayre syndrome (KSS)	Ataxia, neuropathy and short stature.	Singular deletions or copies (commonly sporadic).
Pearson's syndrome	Death in infancy is common and anaemia.	Singular deletions or copies (commonly sporadic).
Mitochondrial Encephalopathy, Lactic Acidosis, and Stroke-like episodes (MELAS)	Stroke like episodes and lactic acidosis.	Heteroplasmic point mutations (inherited maternally).
Myoclonic epilepsy with ragged red fibres (MERRF)	Epilepsy, myoclonus and deafness.	Heteroplasmic point mutations (inherited maternally).
Neuropathy, ataxia and retinitis pigmentosa (NARP)	Weakness, ataxia and pigmentary retinopathy.	Heteroplasmic point mutations (inherited maternally).
Leber's hereditary optic neuropathy (LHON)	Loss of vision.	Homoplasmic point mutations (Inherited maternally)
Sensorineural hearing loss (SNHL)	Hearing loss.	Homoplasmic point mutations (Inherited maternally)

## 2.1. Heteroplasmic mutations

Heteroplasmic mutations, where only some copies of mtDNA contain the mutation, progress to various clinical phenotypes *e.g.*, Leigh syndrome (LS), MERRF, MELAS and NARP. The heterogenous symptoms of MTDS range from organ specific to multisystemic dysfunction with varying clinical classes (Bottani *et al.*, 2020).

For example, heteroplasmic mutations in LS are attributable to mtDNA mutations, however some defects are X-linked or sporadic. LS symptoms usually occur in early childhood and include developmental delay, cardiomyopathy, hypotonia and movement disorder (Kanungo *et al.*, 2018).

## 2.2. Mitochondrial myopathies (MitM)

MitM are developing muscle disorders caused by damage to OXPHOS in the mitochondria. Myopathy is disease of muscle tissue. It is a common symptom of adult-onset MTDS due to the elevated energy required by the skeletal muscle. It can result in proximal myopathy and weakness of limbs. Mutations in either mtDNA or nDNA can cause MitM (Ahmed *et al.*, 2018).

## 3. Therapies for mitochondrial disorders (MTDS)

Current treatments for MTDS are mostly symptomatic. One of the main symptomatic treatments used is the 'one-size-fits-all' approach, which is designed to treat all types of MTDS. This 'one-size-fits-all' approach includes diet, exercise, increasing MB with resveratrol, antioxidants, and pharmacological therapy (Bottani *et al.*, 2020). Dietary approaches can include the use of vitamin B2, vitamin B3 and a ketogenic diet. In terms of exercise, a combination of aerobic and endurance training has proven to be a safe and beneficial treatment for patients with mtDNA mutations (Tarnopolsky, 2014; Voet *et al.*, 2019). Many potential treatments have been suggested for MTDS in the future. These are usually more specific



therapies that can be broadly categorised into precision medicine approaches, personalised therapies, cell replacement therapies and gene therapies (Bottani *et al.*, 2020).

Precision medicine strategies may be able to treat MTDS with specific mutations or unusual metabolic hallmarks (Bottani *et al.*, 2020). Personalised therapy will soon allow patients to obtain earlier diagnoses, risk assessments and better treatments with lower costs. Both have the potential to customize therapy for individual patients and to produce the best response with the highest safety ensured. Cell replacement therapies and gene therapy also have great potential for the future (Vogenberg *et al.*, 2010).

### **3.1 Precision medicine approaches for treating patients with MTDS**

#### **3.1.1 Gene therapies**

Gene therapy is a current strategy to combat MTDS, which introduces modified gene products into the mitochondria via protein import machinery and inhibition of mutant mtDNA (Friedmann and Roblin, 1972, cited by Zeviani and Donato, 2004, p. 2167). This is done via sequence antigenomic peptide-nucleic acids (Zeviani and Donato, 2004). It can treat diseases of a single recessive genetic defect (Bottani *et al.*, 2020). Similarly, it can target many organs at the same time, rendering it a desirable treatment method (Pitceathly *et al.*, 2020). Two gene therapies for treating spinal muscular atrophy and retinal dystrophy have been approved by the FDA (Mendell *et al.*, 2017; Russell *et al.*, 2017).

#### **Adeno-associated virus (AAV)**

AAV's are non-enveloped viruses belonging to the parvovirus family that can be manipulated to transport DNA to target cells. It is one of the safest forms of gene therapy using recombinant AAV particles with the DNA sequence of interest, but without viral genes (Rose *et al.*, 1966, cited by Naso *et al.*, 2017; Pitceathly *et al.*, 2020). AAV-mediated gene therapies for MTDS have undergone clinical trials for neurometabolic disorders *e.g.*, Hunter syndrome (Naso *et al.*, 2017; Pitceathly *et al.*, 2020). At present, there is an AAV-vector-based gene replacement clinical trial running in patients with LHON (Hanaford *et al.*, 2022).

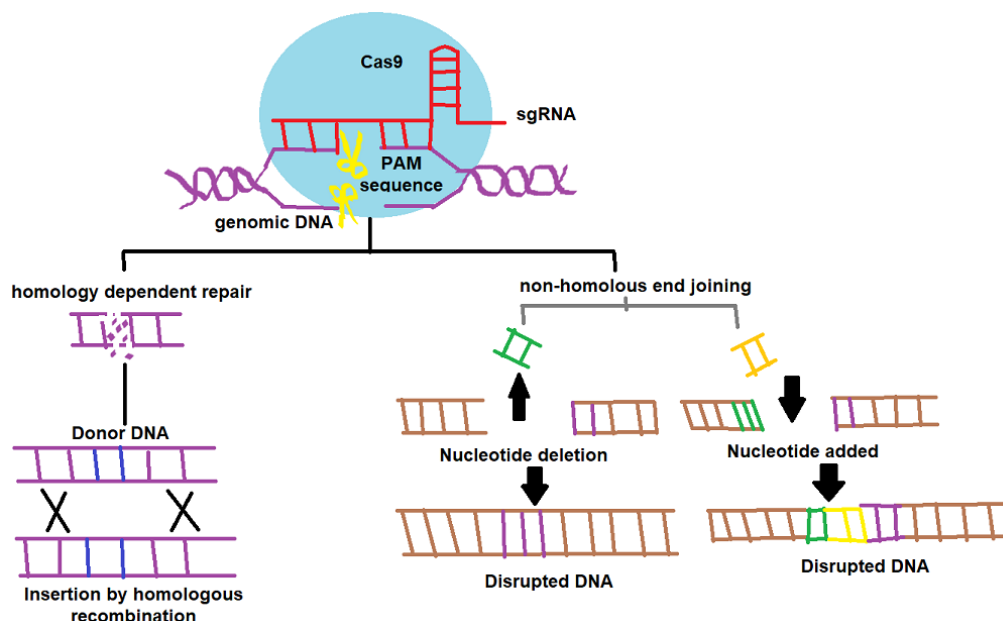
AAV's have been utilised to transport molecular scissors such as Zinc finger nucleases (ZFNs) and Transcription activator-like effector nucleases (TALENs) *in vivo*, to remove mutated mtDNA. ZFNs and TALENS are chimeric nucleases that have engineerable, sequence-specific DNA attaching modules, bound to a non-specific DNA cleavage domain. They can undergo genetic modification by triggering DNA-double-strand breaks, which initiate error-prone non-homologous end joining or homologous directed repair at genomic locations (Bottani *et al.*, 2020). TALENS and mitochondrially targeted ZFNs (mtZFNs) lowered mtDNA heteroplasmy and restored molecular and biochemical phenotypes in a mouse model of heteroplasmic MTDS (Bacman *et al.*, 2018; Gammage *et al.*, 2018).

#### **Clustered regularly interspaced palindromic repeats (CRISPR) /CRISPR-like protein 9 (Cas9) technology**

CRISPR/Cas9 is another gene-editing tool that can correct errors in the genome, as well as switch on/off genes in cells and organisms. It is a cheaper/less time-consuming alternative to TALENS. Its mechanism can be seen below in **Figure 4** (Redman *et al.*, 2016).

Editing the mitochondrial genome has proven difficult due to sub-efficient delivery of a single guide ribonucleic acid (sgRNA) and Cas9 enzyme complexes into the mitochondria (Bottani *et al.*, 2020). In 2015 Jo *et al.*, effectively engineered the mtDNA with CRISPR/Cas9 technology

and showed that the flagged Cas9 localised inside the mitochondria, whilst the sgRNAs permitted depletion of directed elements of mtDNA (Bottani *et al.*, 2020).



**Figure 4. CRISPR/Cas9 mechanism.** Cas9 cuts both strands of DNA and is directed to its target by a sgRNA, which attaches to genomic DNA, allowing for modification of the genome. For Cas9 to cut, the sequence of DNA must be at the 3' end of the sgRNA, this is called the protospacer adjacent motif (PAM). The DNA can be repaired via non-homologous end joining (resulting in random deletion/insertion of DNA) or homology dependent repair (where the homologous section of DNA is utilised as a repair template). Homology dependent repair allows for precise genome repair, as it can make alterations down to a single base pair (Redman *et al.*, 2016).

### 3.1.2 Cell replacement therapies

Cell replacement therapies utilise cell-based products to replace dead cells to re-establish the role of the impacted tissue (Lindvall *et al.*, 2004). It may be a possible cure to various MTDS, such as mitochondrial neurogastrointestinal encephalopathy (MNGIE). MNGIE is an autosomal recessive disorder caused by mutations in the thymidine phosphorylase (TP) gene (Hirano *et al.*, 2004). Autologous haematopoietic stem cell transplantation (AHSCT) and white blood cell-encapsulated thymidine phosphorylase are being investigated as a treatment. Following AHSCT, clinical and biochemical enhancements were effective in repairing TP activity and lowering thymidine concentrations to regular circulating levels in patients with MNGIE (Hirano *et al.*, 2006; Filosto *et al.*, 2012; Bottani *et al.*, 2020; Pitceathly *et al.*, 2020).

### 3.1.3 Mitochondrial therapies

#### Mitochondria based gene editing tools

Hashimoto *et al.*, (2015) used a new molecular technique called mitochondrial-targeted TALENS (mitoTALENS), which show great promise for the future. mitoTALENS can be engineered to identify specific DNA sequences, to initiate double-stranded breaks to breakdown DNA (Pitceathly *et al.*, 2020). They can cause positive heteroplasmic shifts in cells lines with different pathogenic mtDNA mutations and have worked well in cultured mammalian cells (Ahmed *et al.*, 2018; Kazama *et al.*, 2019).

## Mitochondrial transplantation (MT)

This is a current ‘one-size-fits-all’ method of restoring mitochondrial function, through immediate transplantation of good-quality mitochondria into target tissues *e.g.*, to repair function to diseased heart and liver (Cowan *et al.*, 2017; Supinski *et al.*, 2020). Supinski *et al.*, (2020) described how MT into ischemic cardiac tissue increases cardiac function, myocardial contractility and may rescue other organs. Other MT transplants, such as into liver, may be a possible treatment for certain MTDS, particularly for disorders that impact this single organ (McCully *et al.*, 2017, cited by Parikh *et al.*, 2016, p. 181).

### 3.1.4 *In utero*

#### Molecular bypass (MBP) therapy for treating disorders with mtDNA instability

MBP therapy is an experimental treatment, which works by replenishing the regular number of deoxynucleotides (dNTPs) in the mitochondria of cells in patients with mtDNA degeneration syndromes (Marks, 2021). nDNA and mtDNA both rely on individual balanced pools of dNTPs to restore normal function of DNA replication and fix any DNA damage. By importing dNTPs from the cytosol to the mitochondria, it has the potential to restore mitochondrial dNTP discrepancies (Desler *et al.*, 2006).

Diminished thymidine kinase 2 (TK2) activity has shown to cause imbalance of mitochondrial dNTPs and cause mtDNA mutations in non-dividing cells (Desler *et al.*, 2006). MBP therapy with deoxypyrimidine monophosphates and deoxynucleoside were used in an open-labelled study, conducted on 16 early-onset TK2-patients. Results showed amelioration of motor abilities, discontinuation of feeding tubes and prolonged survival (Bottani *et al.*, 2020).

#### Pre-implantation therapies

Pre-implantation genetic diagnosis is a current preventative method, which facilitates families with a known history of mtDNA mutations (Rai *et al.*, 2018). It is an *in vitro* fertilization (IVF) technique, where the fertilized egg with the pathogenic mtDNA mutation is cultured until the blastocyst stage, then it is biopsied for genetic analysis, prior to implantation (Treff *et al.*, 2012; Sallevelt *et al.*, 2017). However, it is limited, as it only helps women with small concentrations of mtDNA mutations in their oocytes (Bottani *et al.*, 2020).

## 3.2 Personalised therapies for MTDS

### 3.2.1 Gene therapies

Allotropic gene expression is a technique used to override mtDNA mutations by re-expressing the absent mtDNA-encoded protein from the nucleus. In this instance, an engineered nuclear form of a mitochondrial gene encodes a protein that may be imported into the mitochondria. Nine patients with LHON syndrome, who had a G11778A mutation causing optic neuropathy, underwent a clinical trial with a single dose of a recombinant AAV2 carrying the normal ND4 gene, with the goal to improve their vision. 66% of the nine patients’ visual acuity improved, resulting in amelioration of their visual field (Bottani *et al.*, 2020).

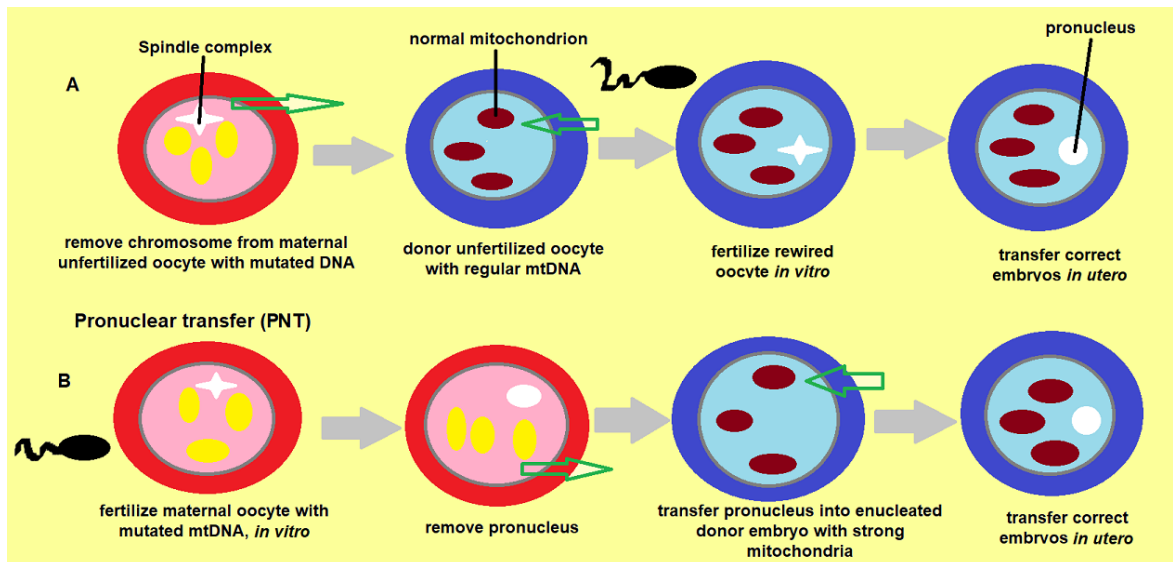
### 3.2.2 Mitochondrial therapies

#### Mitochondrial donation (MD)

Mitochondrial spindle transfer (MST) is a new IVF treatment which includes transferring the mothers nDNA from an affected egg, into an unnucleated egg from an unaffected donor, leading

to an embryo bearing nDNA from both parents (Tachibana *et al.*, 2009, cited by Bottani *et al.*, 2020, p. 25).

The most utilised MD productive technologies include MST and Pronuclear Transfer (PNT), shown in **Figure 5**, which are both legally approved for use in the UK (Craven *et al.*, 2010; Craven *et al.*, 2017; Herbert and Turnbull, 2018).



**Figure 5. Mitochondrial donation.** MST includes the transfer of nuclear genetic material from a patient's egg with mutated mtDNA, to an enucleated donor's unfertilized metaphase II oocyte, with healthy mitochondria. It creates an oocyte with a patient's nDNA, but without the mutated mtDNA (Bottani *et al.*, 2020). PNT involves the transfer of a diploid nucleus into a donor zygote which has been enucleated. The zygote now contains healthy donor mtDNA, with both nDNA of the patient and the fertilising sperm (Craven *et al.*, 2010).

At the meeting of The American Society for Reproductive Medicine, in 2016, Dr Jogn Zhang discussed the results of the utilisation of MST in a woman bearing a mtDNA mutation causing LS. The outcome was the birth of a healthy baby, with fewer than 10% mutated mtDNA in tissues, when analysed 2 days after birth (Hudson *et al.*, 2019). PNT was utilised in 2005 to avoid transmission of a mtDNA disorder in a mito-mouse, a mouse model that normally builds-up large-scale mtDNA deletions (Sato *et al.*, 2005).

### Heteroplasmic shift

A method of correcting mtDNA mutations, focuses on the interference of mutant molecules, by utilising selective nucleases to move the heteroplasmy level under the maximum threshold (Bottani *et al.*, 2020). Many techniques have been developed based on this concept *e.g.*, oligonucleotides. However, it still needs to be determined if shifts in heteroplasmy are maintained over time (Pitceathly *et al.*, 2020).

### 3.2.3 In utero

Foetal gene therapy uses intracellular delivery of genetic material for the treatment of disease, as described in sections 3.1.1 and 3.2.1. By introducing gene therapy for foetal treatment, scientists can access organs during developmental changes (Tsukamoto *et al.*, 1995, cited by David and Peebles, 2008, p. 203). This may also be used a therapeutic method for inherited disorders, *e.g.*, LS, which would normally result in early death or life-long irreparable destruction (Rashnonejad *et al.*, 2019; Bottani *et al.*, 2020). *In Utero* Gene Therapy (IUGT)

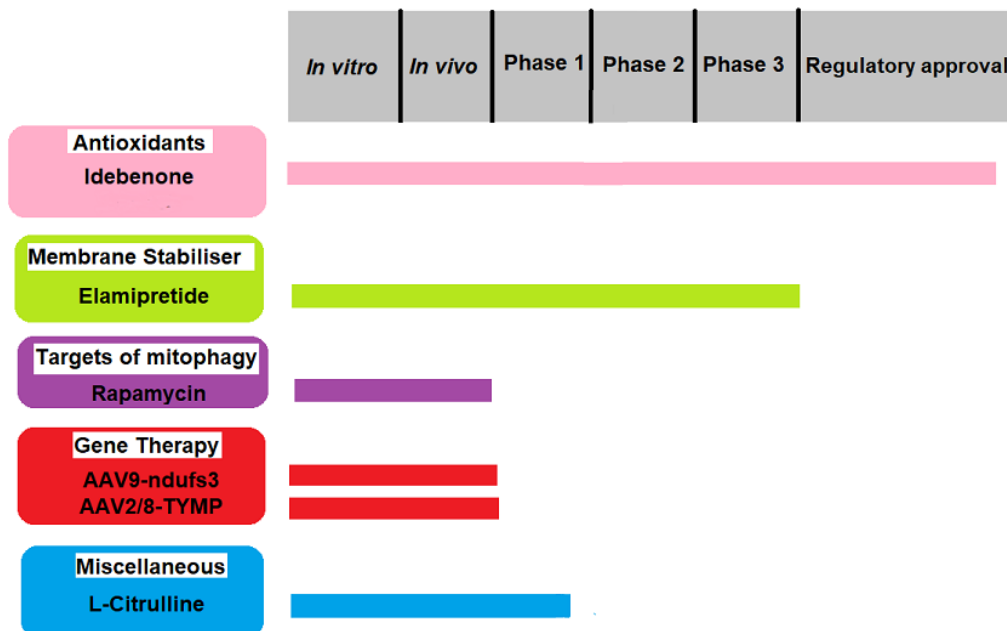
may overcome many of the disadvantages in postnatal gene therapy because of the tiny foetal size and the immature foetal immune system (Peranteau and Flake, 2020).

### 3.3 Genetic counselling (GC)

Most patients with MTDS are already symptomatic by the time they are diagnosed (Pitceathly *et al.*, 2020). As there is huge clinical variability and genetic heterogeneity involved with the complexity of MTDS, patients and families usually experience a long and complicated diagnosis process. Therefore, GC is crucial for individuals suffering from MTDS and should be made available to all patients due to delayed diagnosis, no clear prognosis, issues of family planning and genetic testing (Vento and Pappa, 2013).

### 3.4 Clinical trials for MTDS

A growing number of clinical trials, which are normally double blind and placebo regulated, have investigated the therapeutic effect of vitamins, co-factors, nutritional supplements, and gene therapy for MTDS treatment. Ongoing clinical trials are shown in **Figure 6**. However, these ongoing trials have not yet demonstrated beneficial primary and secondary results (Pitceathly *et al.*, 2020).



**Figure 6. Progress in clinical trial development for MTDS.** With idebenone (an antioxidant) reaching regulatory approval, elamipretide (a membrane stabiliser) reaching phase 3, rapamycin (a mitophagy target) and two gene therapies AAV9-ndufs3 and AAV2/8-TYMP reaching the *in vivo* stage, and L-citrulline (a miscellaneous compound) reaching phase 1 (Pitceathly *et al.*, 2020).

Failure of clinical trials in MTDS are mostly due to inadequacy of the compounds analysed. In addition, trial designs are hindered by the shortage of natural history data (due to their rarity), heterogeneity, validated biomarkers and result measures, to detect an effective treatment (Pitceathly *et al.*, 2020).

## 4. Discussion

From this review it is evident that mitochondria are an essential organelle, critical for synthesizing ATP for eukaryotic life. The mitochondria have protein coding genes which are crucial for function and are derived from its own genome and nDNA. The mitochondrial genome is inherited from the mother, who transmits her oocyte mtDNA to all her female offspring, and subsequently transmit their mtDNA to their offspring, and so on. Being such an important structure with many functions, anything inhibiting the mitochondria's role in the body can lead to severe and often fatal results to the cell/body if not corrected.

These effects result in genetically heterogenous clinical syndromes linked with abnormalities of OXPHOS, and most MTDS occur due to dysfunction of the MRC. Clinical manifestations affect the body's organs, cells, and systems in various debilitating ways. Heteroplasmic and homoplasmic mutations have led to many clinical phenotypes such as LS, LHON and MERRF. High levels of mitochondrial ROS can trigger ETC irregularities and destroy mitochondrial elements, potentially causing muscle disorders such as MitM.

Due to the severity of MTDS, research must continue to explore the defects, origins, and clinical manifestations of these disorders, in order to find more precision/personalised therapies for patients. This information is also crucial in order to advise the patients, treat symptoms/manifestations, with the ultimate goal of preventing the disorders before they emerge.

Current treatments are limited to 'one-size-fits-all' approaches *e.g.*, dietary approaches. However, future perspectives for treating MTDS are exciting and promising, with many treatments currently undergoing clinical trials, including those completed as shown in Table 2 below.

**Table 2. Completed clinical trials for MTDS.**

Current trials	Location	Date	References
<b>Dose escalating with KH176 to treat MELAS, LS and LHON.</b>	Drug Research Unit in Ghent.	Completed October 15th, 2021.	(Clicosm, 2022).
<b>EPI-743 for treating metabolism or MTDS.</b>	Bethesda, Maryland, National Institutes of Health.	Completed March 19 <sup>th</sup> , 2021.	(Clicosm, 2022).

Precision medicine approaches and personalised therapies are coming to the forefront, either of which may treat specific mutations at the genome level and prevent the disorders. Emerging precision medicine approaches include the use of gene therapy in AAV's and gene editing tools

such as CRISPR/Cas9. Personalised therapies include allotropic gene expression, MD with MST/PNT, and even foetal gene therapy for treating early-onset MTDS.

In summary, there is great hope for the future to treat MTDS, with these personalised and precision-based approaches. The more we learn about the rarity, complexity, and challenges of MTDS, the more precise methods of treatment we can develop. Current and future trials provide hope to overcome the extensive challenges of MTDS. We must not underestimate the importance of GC for patients suffering with MTDS. GC is crucial to explain to patients the complexities of the diseases and to support families/patients in their coping strategies, to discuss their options in terms of MTDS treatments, and to decide the best path to take so that MTDS can be better well managed in future generations.

## 5. Acknowledgements

Firstly, I would like to thank my incredibly supportive, kind and knowledgeable supervisor Dr Anne Mulvihill, for all the encouragement and motivation that facilitated me in writing this article. Her unconditional praise and positive reassurance made me believe in myself and in my ability to complete this project in a timely manner. I am eternally grateful and privileged to have had Dr Anne Mulvihill as my supervisor, and for her overall guidance throughout my entire academic journey. This includes her encouragement towards my masters application, to study my passion for genetic and genomic counselling.

In addition, I would like to thank my parents for their continuous support and review of this article. In particular, my father 'Roger Smyth' who helped me think of better ways of phrasing sentences, as well as teaching me how to avoid grammatical error. Their unconditional support and positive regard meant a lot to me along the way.

Lastly, I would like to thank Dr Anne Friel for inviting me to participate in the SUREJ article writing. This has been a wonderful opportunity that I am very grateful for, and my scientific writing has improved 100 fold throughout the journey.

## 6. References

- Ahmed, S. T., Craven, L., Russell, O. M., Turnbull, D. M., and Vincent, A. E. (2018) Diagnosis and treatment of mitochondrial myopathies. *Neurotherapeutics: the journal of the American Society for Experimental NeuroTherapeutics*, 15(4), pp. 943–953. Available at: <https://doi.org/10.1007/s13311-018-00674-4> [Accessed 29 December 2021].
- Annesley, S. and Fisher, R. P. (2019) Mitochondria in health and disease. *Cells*, 8(7), pp. 680. Available at: <https://doi.org/10.3390/cells8070680> [Accessed 31 October 2021].
- Bacman, S. R., Kauppila, J., Pereira, C. V., Nissanka, N., Miranda, M., Pinto, M., Williams, S. L., Larsson, N. G., Stewart, J. B. and Moraes, C. T. (2018) MitoTALEN reduces mutant mtDNA load and restores tRNA<sup>Ala</sup> levels in a mouse model of heteroplasmic mtDNA mutation. *Nature medicine*, 24(11), pp. 1696–1700. Available at: <https://doi.org/10.1038/s41591-018-0166-8> [Accessed 04 March 2022].
- Blanco, A. And Blanco, G. (2017) Chapter 14 - carbohydrate metabolism. *Medical Biochemistry*, Academic Press, pp. 283–323. Available at: <https://doi.org/10.1016/B978-0-12-803550-4.00014-8> [Accessed 02 October 2022].

Bottani, E., Lamperti, C., Prigione, A., Tiranti, V., Persico, N., and Brunetti, D. (2020) Therapeutic approaches to treat mitochondrial diseases: "one-size-fits-all" and "precision medicine" strategies. *Pharmaceutics*, 12(11), pp. 1083. Available at: <https://doi.org/10.3390/pharmaceutics12111083> [Accessed 29 December 2021].

Chinnery, P. F. (2000) Primary mitochondrial disorders overview. In M. P. Adam (Eds.) *et al.*, *GeneReviews*®. University of Washington, Seattle. Available at: <https://www.ncbi.nlm.nih.gov/abc/cardiff.ac.uk/books/NBK1224/> [Accessed 19 August 2022].

Clincosm (2022) *Mitochondrial disease clinical trials*. Available at: <https://www.clincosm.com/Mitochondrial-Disease> [Accessed 05 March 2022].

Cooper, M. G. (2000) *The cell: a molecular approach*. 2nd edn. Sunderland (MA): Sinauer Associates. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK9839/> [Accessed 05 March 2022].

Cowan, D. B., Yao, R., Thedsanamoorthy, J. K., Zurakowski, D., Del Nido, P. J. and McCully, J. D. (2017) Transit and integration of extracellular mitochondria in human heart cells. *Scientific reports*, 7(1), pp. 17450. Available at: <https://doi.org/10.1038/s41598-017-17813-0> [Accessed 03 March 2022].

Craven, L., Tuppen, H. A., Greggains, G. D., Harbottle, S. J., Murphy, J. L., Cree, L. M., Murdoch, A. P., Chinnery, P. F., Taylor, R. W., Lightowers, R. N., Herbert, M. and Turnbull, D. M. (2010) Pronuclear transfer in human embryos to prevent transmission of mitochondrial DNA disease. *Nature*, 465(7294), pp. 82–85. Available at: <https://doi.org/10.1038/nature08958> [Accessed 03 March 2022].

Craven, L., Alston, C. L., Taylor, R. W. and Turnbull, D. M. (2017) Recent advances in mitochondrial disease. *Annual review of genomics and human genetics*, 18, pp. 257–275. Available at: <https://doi.org/10.1146/annurev-genom-091416-035426> [Accessed 04 March 2022].

David, L. A. and Peebles, D. (2008) Gene therapy for the fetus: is there a future?. *best practice & research clinical obstetrics & gynaecology*, 22(1), pp. 203-218. Available at: <https://doi.org/10.1016/j.bpobgyn.2007.08.008> [Accessed 27 September 2022].

Desler, C., Petersen, M. B. and Rasmussen, J. L. (2006) The role of mitochondrial dNTP levels in cells with reduced TK2 activity. *Nucleosides, Nucleotides & Nucleic Acids*, 9-11(25), pp. 1171-1175. Available at: [10.1080/15257770600894501](https://doi.org/10.1080/15257770600894501) [Accessed 27 September 2022].

Endo, T. and Sakaue, H. (2019) Multifaceted roles of porin in mitochondrial protein and lipid transport. *Biochemical Society transactions*, 47(5), pp. 1269–1277. Available at: <https://doi.org/10.1042/BST20190153> [Accessed 01 November 2022].

Filosto, M., Scarpelli, M., Tonin, P., Lucchini, G., Pavan, F., Santus, F., Parini, R., Donati, M. A., Cotelli, M. S., Vielmi, V., Todeschini, A., Canonico, F., Tomelleri, G., Padovani, A. and Rovelli, A. (2012) Course and management of allogeneic stem cell transplantation in patients with mitochondrial neurogastrointestinal encephalomyopathy. *Journal of neurology*, 259(12), pp. 2699–2706. Available at: <https://doi.org/10.1007/s00415-012-6572-9> [Accessed 01 March 2022].



- Friedmann, T. and Roblin, R. (1972) Gene therapy for human genetic disease?. *Science (New York, N.Y.)*, 175(4025), pp. 949–955. Available at: <https://doi.org/10.1126/science.175.4025.949> [Accessed 04 November 2022].
- Gammage, P. A., Viscomi, C., Simard, M. L., Costa, A., Gaude, E., Powell, C. A., Van Haute, L., McCann, B. J., Rebelo-Guiomar, P., Cerutti, R., Zhang, L., Rebar, E. J., Zeviani, M., Frezza, C., Stewart, J. B. and Minczuk, M. (2018) Genome editing in mitochondria corrects a pathogenic mtDNA mutation in vivo. *Nature medicine*, 24(11), pp. 1691–1695. Available at: <https://doi.org/10.1038/s41591-018-0165-9> [Accessed 04 March 2022].
- Glancy, B. (2020). Visualizing mitochondrial form and function within the cell. *Trends in molecular medicine*, 26(1), pp. 58–70. Available at: <https://doi.org/10.1016/j.molmed.2019.09.009> [Accessed 10 February 2022].
- Glancy, B., Yuho, K., Prasanna, K. and Bradley, T. W. (2020) The Functional impact of mitochondrial structure across subcellular scales. *Frontiers in Physiology*, 11. Available at: <https://www.frontiersin.org/article/10.3389/fphys.2020.541040> [Accessed 10 February 2022].
- Goyal, S. and Chaturvedi, R. K. (2021) Mitochondrial protein import dysfunction in pathogenesis of neurodegenerative diseases. *Molecular Neurobiology*, 58, pp. 1418-1437. Available at: <https://doi.org/10.1007/s12035-020-02200-0> [Accessed 31 October 2022].
- Guerra, F., Arbini, A. A., and Moro, L. (2017) Mitochondria and cancer chemoresistance. *Biochimica et biophysica acta. Bioenergetics*, 1858(8), pp. 686–699. Available at: <https://doi.org/10.1016/j.bbabi.2017.01.012> [Accessed 29 December 2021].
- Halestrap, A. P., Woodfield, K. Y. and Connern, C. P. (1997) Oxidative stress, thiol reagents, and membrane potential modulate the mitochondrial permeability transition by affecting nucleotide binding to the adenine nucleotide translocase. *The Journal of biological chemistry*, 272(6), pp. 3346–3354. Available at: <https://doi.org/10.1074/jbc.272.6.3346> [Accessed 15 March 2022].
- Hanaford, A. R., Cho, Y. J. and Nakai, H. (2022) AAV-vector based gene therapy for mitochondrial disease: progress and future perspectives. *Orphanet journal of rare diseases*, 17(1), pp. 217. Available at: <https://doi.org/10.1186/s13023-022-02324-7> [Accessed 01 November 2022].
- Hashimoto, M., Bacman, R. S., Peralta, S., Falk, J. M., Chomyn, A., Chan, C. D., Williams, L. S. and Moraes, T. C. (2015) MitoTALEN: A general approach to reduce mutant mtDNA loads and restore oxidative phosphorylation function in mitochondrial diseases. *Molecular Therapy*, 23(10), pp. 1592-1599. Available at: <https://doi.org/10.1038/mt.2015.126> [Accessed 30 December 2021].
- Herbert, M. and Turnbull, D. (2018) Progress in mitochondrial replacement therapies. *Nature reviews. Molecular cell biology*, 19(2), pp. 71–72. Available at: <https://doi.org/10.1038/nrm.2018.3> [Accessed 03 March 2022].
- Hewitt, V., Lithgow, T. and Waller, R. F. (2013) Modifications and innovations in the evolution of mitochondrial protein import pathways. *Endosymbiosis*, pp. 19-35. Available at: <https://doi.org/10.1007/978-3-7091-1303-5-2> [Accessed 04 November 2022].
- Hirano, M., Nishigaki, Y. and Martí, R. (2004) Mitochondrial neurogastrointestinal encephalomyopathy (MNGIE): a disease of two genomes. *The neurologist*, 10(1), pp. 8–17. Available at: <https://doi.org/10.1097/01.nrl.0000106919.06469.04> [Accessed 28 October 2022].

Hirano, M., Martí, R., Casali, C., Tadesse, S., Uldrick, T., Fine, B., Escolar, D. M., Valentino, M. L., Nishino, I., Hesdorffer, C., Schwartz, J., Hawks, R. G., Martone, D. L., Cairo, M. S., DiMauro, S., Stanzani, M., Garvin, J. H., Jr. and Savage, D. G. (2006) Allogeneic stem cell transplantation corrects biochemical derangements in MNGIE. *Neurology*, 67(8), pp. 1458–1460. Available at: <https://doi.org/10.1212/01.wnl.0000240853.97716.24> [Accessed 18 October 2021].

Hudson, G., Takeda, Y. and Herbert, M. (2019) Reversion after replacement of mitochondrial DNA. *Nature*, 574(7778), pp. E8–E11. Available at: <https://doi.org/10.1038/s41586-019-1623-3> [Accessed 03 March 2022].

Jo, A., Ham, S., Lee, G. H., Lee, Y.-I., Kim, S., Lee, Y.-S. and Shin, J.-H. (2015) Efficient mitochondrial genome editing by CRISPR/cas9. *BioMed Research International*, pp. 1-10. Available at: <https://doi.org/10.1155/2015/305716> [Accessed 10 January 2022].

Kauppila, S. E. T., Kauppila, K. H. J., and Larsson, G. N. (2017) Mammalian mitochondria and aging: an update. *Cell Metabolism*, 25(1), pp. 57-71. Available at: <http://dx.doi.org/10.1016/j.cmet.2016.09.017> [Accessed 31 October 2021].

Kanungo, S., Morton, J., Neelakantan, M., Ching, K., Saeedian, J. and Goldstein, A. (2018) Mitochondrial disorders. *Annals of translational medicine*, 6(24), pp. 475. Available at: <https://doi.org/10.21037/atm.2018.12.13> [Accessed 17 February 2022].

Kazama, T., Okuno, M., Watari, Y., Yanase, S., Koizuka, C., Tsuruta, Y., Sugaya, H., Toyoda, A., Itok, T., Tsutsumi, N., Toriyama, K., Koizuka, N. and Arimura, S. (2019) Curing cytoplasmic male sterility via TALEN-mediated mitochondrial genome editing. *Nature Plants*, (5), pp. 722–730. Available at: <https://doi.org/10.1038/s41477-019-0459-z> [Accessed 04 March 2022].

Kühlbrandt, W. (2015) Structure and function of mitochondrial membrane protein complexes. *BMC Biology* 13, pp. 89. Available at: <https://doi.org/10.1186/s12915-015-0201-x> [Accessed 10 February 2022].

Lindvall, O., Kokaia, Z. and Martinez-Serrano, A. (2004) Stem cell therapy for human neurodegenerative disorders-how to make it work. *Nature medicine*, 10 Suppl, pp. S42–S50. Available at: <https://doi.org/10.1038/nm1064> [Accessed 01 March 2022].

Marks, W. J. (2021) *Medical definition of nucleoside bypass therapy*. Available at: [https://www.medicinenet.com/nucleoside\\_bypass\\_therapy/definition.htm](https://www.medicinenet.com/nucleoside_bypass_therapy/definition.htm) [Accessed 28 October 2022].

McCully, J. D., Cowan, D. B., Emani, S. M. and Del Nido, P. J. (2017) Mitochondrial transplantation: from animal models to clinical use in humans. *Mitochondrion*, 34, pp. 127–134. Available at: <https://doi.org/10.1016/j.mito.2017.03.004> [Accessed 01 March 2022].

Mendell, J. R., Al-Zaidy, S., Shell, R., Arnold, W. D., Rodino-Klapac, L. R., Prior, T. W., Lowes, L., Alfano, L., Berry, K., Church, K., Kissel, J. T., Nagendran, S., L'Italien, J., Sproule, D. M., Wells, C., Cardenas, J. A., Heitzer, M. D., Kaspar, A., Corcoran, S., Braun, L., Miranda, C., Meyer, K., Foust, D. K., Burghes, M. H. A., Likhite, S. and Kaspar, K. B. (2017) Single-dose gene-replacement therapy for spinal muscular atrophy. *The New England journal of medicine*, 377(18), pp. 1713–1722. Available at: <https://doi.org/10.1056/NEJMoa1706198> [Accessed 04 March 2022].

- Musante, L., Candiano, G., Bruschi, M., Santucci, L. And Ghiggeri, M. G. (2007) *Autoantibodies*. Elsevier. Available at <https://doi.org/10.1016/B978-044452763-9/50071-8> [Accessed 28 October 2022].
- Naso, M. F., Tomkowicz, B., Perry, W. L. and Strohl, W. R. (2017) Adeno-associated virus (AAV) as a vector for gene therapy. *BioDrugs: clinical immunotherapeutics, biopharmaceuticals and gene therapy*, 31(4), pp. 317–334. Available at: <https://doi.org/10.1007/s40259-017-0234-5> [Accessed 06 April 2022].
- Nass, K. M. M. and Nass, S. (1963) Intramitochondrial fibers with DNA characteristics: I. fixation and electron staining reactions. *The Journal of Cell Biology*, 19(3), pp. 593–611. Available at: <https://doi.org/10.1083/jcb.19.3.593> [Accessed 03 November 2022].
- Ow, Y.-L. P., Green, D. R., Hao, Z. and Mak, T. W. (2008) Cytochrome c: functions beyond respiration. *Nature Reviews Molecular Cell Biology*, 9(7), pp. 532–542. Available at: <https://doi-org.abc.cardiff.ac.uk/10.1038/nrm2434> [Accessed 02 October 2022].
- Parikh, S., Karaa, A., Goldstein, A., Ng, Y. S., Gorman, G., Feigenbaum, A., Christodoulou, J., Haas, R., Tarnopolsky, M., Cohen, B. K., Dimmock, D., Feyma, T., Koenig, M. K., Mundy, H., Niyazov, D., Saneto, R. P., Wainwright, M. S., Wusthoff, C., McFarland, R. and Scaglia, F. (2016) Solid organ transplantation in primary mitochondrial disease: proceed with caution. *Molecular genetics and metabolism*, 118(3), pp. 178–184. Available at: <https://doi.org/10.1016/j.ymgme.2016.04.009> [Accessed 01 March 2022].
- Park, S., Jeon, J. H., Min, B. K., Ha, C. M., Thoudam, T., Park, B. Y. and Lee, I. K. (2018) Role of the pyruvate dehydrogenase complex in metabolic remodeling: differential pyruvate dehydrogenase complex functions in metabolism. *Diabetes & metabolism journal*, 42(4), pp. 270–281. Available at: <https://doi.org/10.4093/dmj.2018.0101> [Accessed 27 September 2022].
- Peranteau, W. H. and Flake, A. W. (2020) The future of in utero gene therapy. *Molecular diagnosis and therapy*, 24(2), pp. 135–142. Available at: <https://doi.org/10.1007/s40291-020-00445-y> [Accessed 04 March 2022].
- Pitceathly, R., Keshavan, N., Rahman, J. and Rahman, S. (2020) Moving towards clinical trials for mitochondrial diseases. *Journal of inherited metabolic disease*, 44(1), pp. 22–41. Available at: <https://doi.org/10.1002/jimd.12281> [Accessed 29 December 2021].
- Rai, P. K., Craven, L., Hoogewijs, K., Russell, O. M. and Lightowlers, R. N. (2018) Advances in methods for reducing mitochondrial DNA disease by replacing or manipulating the mitochondrial genome. *Essays in biochemistry*, 62(3), pp. 455–465. Available at: <https://doi.org/10.1042/EBC20170113> [Accessed 03 March 2022].
- Rashnonejad, A., Amini Chermahini, G., Gündüz, C., Onay, H., Aykut, A., Durmaz, B., Baka, M., Su, Q., Gao, G. and Özkınay, F. (2019) Fetal gene therapy using a single injection of recombinant AAV9 rescued SMA phenotype in mice. *Molecular therapy: the journal of the American Society of Gene Therapy*, 27(12), pp. 2123–2133. Available at: <https://doi.org/10.1016/j.ymthe.2019.08.017> [Accessed 04 March 2022].
- Redman, M., King, A., Watson, C. and King, D. (2016) What is CRISPR/cas9?. *Archives of disease in childhood. Education and practice edition*, 101(4), pp. 213–215. <https://doi.org/10.1136/archdischild-2016-310459> [Accessed 27 March 2022].

Rose, J. A., Hoggan, M. D. and Shatkin, A. J. (1966) Nucleic acid from an adeno-associated virus: chemical and physical studies. *Proceedings of the National Academy of Sciences of the United States of America*, 56(1), pp. 86–92. Available at: <https://doi.org/10.1073/pnas.56.1.86> [Accessed 04 November 2022].

Russell, S., Bennett, J., Wellman, J. A., Chung, D. C., Yu, Z. F., Tillman, A., Wittes, J., Pappas, J., Elci, O., McCague, S., Cross, D., Marshall, K. A., Walshire, J., Kehoe, T. L., Reichert, H., Davis, M., Raffini, L., George, L. A., Hudson, F. P., Dingfield, L., Zhu, X., Haller, A. J. Sohn, H. E., Mahajan, B. V., Pfeifer, W., Weckmann, M., Johnson, C. P., Gewaily, D., Drack, A., Stone, E., Wachtel, K., Simonelli, F., Leroy, P. B., Wright, F. J., High, A. K. and Maguire, A. M. (2017) Efficacy and safety of voretigene neparvovec (AAV2-hRPE65v2) in patients with RPE65-mediated inherited retinal dystrophy: a randomised, controlled, open-label, phase 3 trial. *Lancet (London, England)*, 390(10097), p. 849–860. Available at: [https://doi.org/10.1016/S0140-6736\(17\)31868-8](https://doi.org/10.1016/S0140-6736(17)31868-8) [Accessed 04 March 2022].

Sallevelt, S., Dreesen, J., Coonen, E., Paulussen, A., Hellebrekers, D., de Die-Smulders, C., Smeets, H. and Lindsey, P. (2017) Preimplantation genetic diagnosis for mitochondrial DNA mutations: analysis of one blastomere suffices. *Journal of medical genetics*, 54(10), pp. 693–697. Available at: <https://doi.org/10.1136/jmedgenet-2017-104633> [Accessed 03 March 2022].

Sato, A., Kono, T., Nakada, K., Ishikawa, K., Inoue, S., Yonekawa, H. and Hayashi, J. (2005) Gene therapy for progeny of mito-mice carrying pathogenic mtDNA by nuclear transplantation. *Proceedings of the National Academy of Sciences of the United States of America*, 102(46), pp. 16765–16770. Available at: <https://doi.org/10.1073/pnas.0506197102> [Accessed 03 March 2022].

Scott, I. and Youle, R. J. (2010) Mitochondrial fission and fusion. *Essays in biochemistry*, 47, pp. 85–98. Available at: <https://doi.org/10.1042/bse0470085> [Accessed 29 September 2022].

Shalem R. Modi and Riikka H. H. (2020) Recent advances in iPSC disease modeling, volume 1. In *Advances in Stem Cell Biology*, Academic Press, pp. 47–70. Available at: <https://doi.org/10.1016/B978-0-12-822227-0.00003-X> [Accessed 01 October 2022].

Sharma, L. K., Lu, J. and Bai, Y. (2009) Mitochondrial respiratory complex I: structure, function and implication in human diseases. *Current medicinal chemistry*, 16(10), pp. 1266–1277. Available at: <https://doi.org/10.2174/092986709787846578> [Accessed 12 March 2022].

Sidarala, V., Zhu, J., Levi-D'Ancona, E., Pearson, G. L., Reck, E. C., Walker, E. M., Kaufman, B. A., and Soleimanpour, S. A. (2022) Mitofusin 1 and 2 regulation of mitochondrial DNA content is a critical determinant of glucose homeostasis. *Nature communications*, 13(1), pp. 2340. Available at: <https://doi.org/10.1038/s41467-022-29945-7> [Accessed 01 October 2022].

Supinski, G. S., Murphy, M. P., and Callahan, L. A. (2009) MitoQ administration prevents endotoxin-induced cardiac dysfunction. *American journal of physiology. Regulatory, integrative and comparative physiology*, 297(4), pp. R1095–R1102. Available at: <https://doi.org/10.1152/ajpregu.90902.2008> [Accessed 14 February 2022].

Supinski, G. S., Schroder, E. A. and Callahan, L. A. (2020) Mitochondria and critical illness. *Chest*, 157(2), pp. 310–322. Available at: <https://doi.org/10.1016/j.chest.2019.08.2182> [Accessed 29 December 2021].

Tachibana M, Sparman M, Sritanaudomchai H, Ma H, Clepper L, Woodward J, Li Y, Ramsey C, Kolotushkina O. and Mitalipov S. (2009) Mitochondrial gene replacement in primate

offspring and embryonic stem cells. *Nature*, 461(7262), pp. 367-372. Available at: [doi:10.1038/nature08368](https://doi.org/10.1038/nature08368) [Accessed 04 November 2022].

Tarnopolsky M. A. (2014) Exercise as a therapeutic strategy for primary mitochondrial cytopathies. *Journal of child neurology*, 29(9), pp. 1225–1234. Available at: <https://doi.org/10.1177/0883073814538512> [Accessed 27 February 2022].

Treff, N. R., Campos, J., Tao, X., Levy, B., Ferry, K. M. and Scott, R. T., Jr (2012) Blastocyst preimplantation genetic diagnosis (PGD) of a mitochondrial DNA disorder. *Fertility and sterility*, 98(5), pp. 1236–1240. Available at: <https://doi.org/10.1016/j.fertnstert.2012.07.1119> [Accessed 04 March 2022].

Tsukamoto, M., Takahiro, O., Yoshida, S., Sugimura, T. and Terada, M. (1995) Gene transfer and expression in progeny after intravenous DNA injection into pregnant mice. *Nature Genetics*, 9(3), pp. 243-48. Available at: <https://doi.org/10.1038/ng0395-243> [Accessed 04 November 2022].

Vento, J. M. and Pappa, B. (2013) Genetic counseling in mitochondrial disease. *Neurotherapeutics: the journal of the American Society for Experimental NeuroTherapeutics*, 10(2), pp. 243–250. Available at: <https://doi.org/10.1007/s13311-012-0173-2> [Accessed 27 March 2022].

Voet, N. B., van der Kooi, E. L., van Engelen, B. G. and Geurts, A. C. (2019) Strength training and aerobic exercise training for muscle disease. *The Cochrane database of systematic reviews*, 12(12), pp. CD003907. Available at: <https://doi.org/10.1002/14651858.CD003907.pub5> [Accessed 27 February 2022].

Vogelberg, F. R., Isaacson Barash, C. and Pursel, M. (2010) Personalized medicine: part 1: evolution and development into theranostics. *P & T: a peer-reviewed journal for formulary management*, 35(10), pp. 560–576. Available at: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2957753/> [Accessed 27 March 2022].

Youle, R. J. and van der Bliek, A. M. (2012) Mitochondrial fission, fusion, and stress. *Science (New York, N.Y.)*, 337(6098), pp. 1062–1065. Available at: <https://doi.org/10.1126/science.1219855> [Accessed 02 October 2022].

Zeviani, M and Donato, D. S. (2004) Mitochondrial disorders. *Brain*, 127 (10) pp. 2153-2172. Available at: <https://doi.org/10.1093/brain/awh259> [Accessed 09 February 2022].

Zeviani, M and Donato, D. S. (2004) *Mitochondrial OXPHOS diseases due to mtDNA mutations.* [Table] *Brain*, 127 (10) pp. 2153-2172. Available at: <https://doi.org/10.1093/brain/awh259> [Accessed 09 February 2022].