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**Transcriptome characterization and expression profiling in chestnut cultivars resistant
or susceptible to the gall wasp *Dryocosmus kuriphilus***

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44 **Abstract** – The oriental gall wasp *Dryocosmus kuriphilus* represents a limiting pest for the European Chestnut
45 (*Castanea sativa*, Fagaceae) as it creates severe yield losses. The European Chestnut is a deciduous tree, having
46 major social, economic and environmental importance in Southern Europe, covering an area of 2.53 million
47 hectares, including 75,000 ha devoted to fruit production. Cultivars show different susceptibility and very few are
48 resistant to gall wasp. To deeply investigate the plant response and understand which factors can lead the plant to
49 develop or not the gall, the study of transcriptome is basic (fundamental). To date little transcriptomic information
50 are available for *C. sativa* species. Hence, we present a *de novo* assembly of the chestnut transcriptome of the
51 resistant Euro-Japanese hybrid ‘Bouche de Bétizac’ (BB) and the susceptible cultivar ‘Madonna’ (M), collecting
52 RNA from buds at different stages of budburst. The two transcriptomes were assembled into 34,081 (BB) and
53 30,605 (M) unigenes, respectively. The former was used as a reference sequence for further characterization
54 analyses, highlighting the presence of 1,444 putative Resistance Gene Analogues (RGAs) and about 1,135
55 unigenes, as putative MiRNA targets. A global quantitative transcriptome profiling comparing the resistant and
56 the susceptible cultivars, in the presence or not of the gall wasp, revealed some GO enrichments as “response to
57 stimulus” (GO:0050896), and “developmental processes” (e.g.: post embryonic development, GO:0009791).
58 Many up-regulated genes appeared to be transcription factors (e.g.: RAV1, AP2/ERF, WRKY33) or protein
59 regulators (e.g.: RAPTOR1B) and storage proteins (e.g.: LEA D29) involved in “post-embryonic development”.
60 Our analysis was able to provide a large amount of information, including 7k simple sequence repeat (SSR) and
61 335k single-nucleotide polymorphism (SNP)/INDEL markers, and generated the first reference unigene catalogue
62 for the European Chestnut. The transcriptome data for *C. sativa* will contribute to understand the genetic basis of
63 the resistance to gall wasp and will provide useful information for next molecular genetic studies of this species
64 and its relatives.

65 **Keywords:** RNA-Seq, *Castanea*, resistance, assembly, Gene Ontology

66 Introduction

67

68 The oriental gall wasp *Dryocosmus kuriphilus* Yasumatsu (Hymenoptera: Cynipidae) is considered the most
69 invasive alien pest for the European chestnut (*Castanea sativa* Miller) currently reported in almost the whole
70 Europe. The purpose of this study was a deeply investigation of the chestnut transcriptome to understand the
71 genetic basis of the plant response to gall wasp infestation and to provide a large amount of information for
72 molecular genetic studies on this species.

73 The European sweet chestnut (*Castanea sativa* Miller) is a multipurpose tree species mostly distributed across
74 Southern Europe, from Turkey to Portugal, but found also in Northern countries such as UK. Its wide distribution
75 and presence in mountain and high hill areas makes the tree an important resource as provider of ecosystemic
76 services in these sites. Besides its value as a forest species and its importance for the landscape and environment,
77 *Castanea sativa* still represents a relevant economic resource for the nut production, being Italy, Turkey and
78 Portugal the major producing countries in Europe. Despite the progressive decline of the last 50 years of
79 production in Europe due to a series of factors including diseases and pest, climatic change, aging and urbanization
80 of mountain people, recently *C. sativa* production is showing a slow production recovery (FAOSTAT 2018).

81 The diffusion of the gall wasp *D. kuriphilus* in Europe, represented a major threat for chestnut; the pest, native of
82 China, was reported for the first time in Piemonte (NW Italy) in 2002 (Brussino et al. 2002) and now spread in
83 most of the European Countries where chestnut is present. The pest lays eggs into the buds in early summer of the
84 first year; larvae and eggs are found in the buds at the end of winter of the following year, but there is no outer
85 symptoms of the presence of the wasp until after budburst. The evidence of the infestation is the formation of
86 galls, round green and reddish structures that develop on the young shoots in spring, due to the reaction of the
87 plant to the presence of the feeding larvae. Following pupation, adults fly out of the gall in early summer and lay
88 eggs into the new formed dormant buds of the chestnut tree. The thelytokous parthenogenetic reproduction system
89 of the wasp causes an exponential population increase in a short time, while dispersal through propagation material
90 is favoured by the absence of external symptoms in buds during winter.

91 The damage that galls can cause involve directly leaves, shoots and inflorescences, and indirectly the whole
92 biomass; leaf surface is reduced, and the amount of vegetative buds is decreased, year-by-year (Kato and Hijii
93 1997). Although plant death is rare and usually associated with other factors such as diseases, the interruption of
94 plant growth and the reduction of fruiting results in yield losses of up to 50-70% in the Chinese chestnut (*C.*
95 *mollissima*) and Japanese chestnut (*C. crenata*) (Dixon et al. 1986). The assessment of yield loss in *Castanea*
96 *sativa* (Sartor et al. 2015) showed similar data indicating that infestation values, determined as No. galls/bud,
97 above 0.6 cause a drastic decrease of productivity (60% on average).

98 In Japan, after the accidental introduction of the gall wasp in 1941, breeding programs were carried out to obtain
99 resistant cultivars starting from resistant genotypes found in *C. crenata*. More recently, the trait was found to be
100 present in other *Castanea* species (*C. mollissima*, *C. pumila*) and in *C. sativa* (Sartor et al. 2015). Studies on the
101 genetic bases of resistance, agree on the hypothesis that more mechanisms may be responsible of the trait in the
102 different chestnut genotypes (Shimura 1972b; Anagnostakis et al. 2009).

103 Following the introduction of the wasp, in Italy several strategies of control were tested, the most successful being
104 the biological control by *Torymus sinensis* (Kamijio) (Quacchia et al. 2008; Picciau et al. 2017; Ferracini et al.,
105 2018). In parallel, studies were conducted on the susceptibility and resistance to the pest in the cultivated and wild

106 *C. sativa* germplasm (Sartor et al. 2015). In fact, there were reports of resistance in *C. crenata* and a large variation
107 in susceptibility observed across genotypes (Shimura 1972a). Among cultivars, the interspecific hybrid ‘Bouche
108 de Bétizac’ (*C. sativa* ‘Bouche Rouge’ x *C. crenata* ‘CA04’) was found to be asymptomatic in spring (no galls),
109 although buds were oviposited and contained larvae in winter (Sartor et al. 2009; Dini et al. 2012). In this case,
110 the occurrence of a hypersensitive response at budburst was postulated to explain larvae death and regular shoot
111 development (Dini et al. 2012). Following these advancements, a segregating progeny accounting 250 individuals
112 was obtained from ‘Bouche de Bétizac’ X ‘Madonna’ (*C. sativa*, highly susceptible cultivar) in order to map the
113 trait (Torello Marinoni et al. 2017) and a transcriptome analysis, described in this paper, was carried out. The
114 purpose was to create a catalogue of *C. sativa* unigenes, likely including genes involved in plant-insect interaction,
115 and to isolate molecular markers for the mapping of traits of interest.

116 With a similar approach, a highly informative genetic map of Chinese chestnut was constructed to extend genomic
117 studies in the Fagaceae and to aid the introgression of Chinese chestnut blight resistance genes into American
118 chestnut (Kubisiak et al. 2013). The transcriptome-based genetic map was created with 329 simple sequence repeat
119 and 1,064 single nucleotide polymorphism markers all derived from expressed sequence tag sequences. Genetic
120 maps for each parent were developed and combined to establish 12 consensus linkage groups spanning 742 cM.
121 Another paper compared the root transcriptome of the susceptible species *C. sativa* and the resistant species *C.*
122 *crenata* after *P. cinnamomi* inoculation to elucidate chestnut defense mechanisms to ink disease (Serrazina et al.
123 2015); results of RNA-seq enabled the selection of candidate genes for ink disease resistance in *Castanea*.

124 In this paper, we sequenced, assembled and functionally characterized the transcriptome of two chestnut cultivars
125 (a cynipid-resistant and a cynipid-susceptible), during the early stages of the interaction plant-pest, generating an
126 extraordinary amount of information. A set of genes regulated in both the susceptible and the resistant cultivars
127 was highlighted. The functional annotation of RGAs and miRNA target genes was attempted and SSR and SNP
128 markers were identified/classified to populate a catalogue suitable for genetic trait dissection. The chestnut
129 transcriptome assembly will open the possibility to deeply study the plant response and understand which factors
130 can lead the plant to develop or not the gall.

131

132 **Materials and Methods**

133

134 **Chestnut Material**

135 Buds from cultivar ‘Madonna’ (*C. sativa*) and the Eurojapanese hybrid ‘Bouche de Bétizac’ (*C. sativa* ‘Bouche
136 Rouge’ X *C. crenata* ‘CA04’), were harvested from single plants at different times of budburst from April 21st to
137 May 12th. The cultivar ‘Madonna’ buds were harvested in areas highly infested by the cynipid, while the ‘Bouche
138 de Bétizac’ buds were collected from plants infested by the cynipid, maintained in screenhouses set up in the
139 forest nursery of Chiusa Pesio (CN, Piedmont, Italy) as described in Sartor et al. 2015. The collection was carried
140 out once a week in order to gather material representative of four different stages of bud sprouting (1-closed bud;
141 2-bud that initiates to swell; 3-end of bud swelling; scales separated; 4-brown scales fallen, bud enclosed by green
142 scales), and be able to sample tissues during the defensive response. Buds were immediately frozen in liquid
143 nitrogen and stored at -80°C until use.

144 The identity of the cultivars was checked by SSR analysis (CsCAT1, CsCAT3, CsCAT6, CsCAT16, CsCAT17,
145 QpZAG110; Steinkellner et al. 1997; Marinoni et al. 2003) according to the protocol by Torello Marinoni et al.
146 (2013).

147

148 **RNA and DNA extraction**

149 Buds were disrupted in liquid nitrogen using a baked mortar and pestle treated with DEPC water. Nucleic acids
150 were extracted using a buffer containing: 2% CTAB, 2% polyvinylpyrrolidone (PVP) K-30 (soluble), 100 mM
151 Tris HCl (pH 8.0), 25 mM EDTA, 2.0 M NaCl, 0.5 g/L spermidine, 2% b-mercaptoethanol. After two buffered
152 chloroform extractions, the upper phase containing the nucleic acids was divided in two parts for RNA and DNA
153 extraction. DNA was precipitated with 0.7 volumes of isopropanol and washed in 70% ethanol; it was dried and
154 resuspended in 50 ml of sterile water.

155 Total RNA was precipitated overnight with 8 M LiCl at 4°C. The next day RNA was added of an SSTE buffer
156 (5.0 M NaCl, 0.5% SDS, 10 mM Tris HCl, pH 8.0, 1 mM EDTA), and treated at 65°C for 10 min. Following two
157 chloroform purifications, RNA was precipitated and then washed in ethanol 100% and 70%. Total RNA was
158 purified with RNeasy Mini Kit (Qiagen). RNA yield and quality were evaluated using spectrophotometric
159 determinations (Dini et al. 2012).

160 DNA samples were used to check the cynipid presence/absence by diagnostic PCR following the protocol by
161 Sartor et al. (2012), choosing the 320bp amplicon of 28S Ribosomal DNA sequence as marker to detect the larva
162 presence.

163

164 **RNA sequencing**

165 For RNA sequencing we considered 4 thesis: ‘Bouche de Bétizac’ infested, ‘Bouche de Bétizac’ not infested,
166 ‘Madonna’ infested, ‘Madonna’ not infested. To have a representative sample, the total RNA extracted from single
167 buds, belonging to the same thesis, but collected at four different stages of sprouting, was pooled together..

168 From these materials, in collaboration with Evrogen (Moscow, Russia), 4 tagged cDNA libraries were obtained:
169 ‘Bouche de Bétizac’ infested (BI), ‘Bouche de Bétizac’ uninfested (BNI), ‘Madonna’ infested (MI), ‘Madonna’
170 uninfested (MNI). The sequencing was commissioned to BMR Genomics (Padova, Italy), preparing a unique pool
171 of the four tagged samples in equimolar concentration. For the sequencing a single Hiseq 1000 protocol (2PE x
172 100bp) was used, following the TruSeq DNA protocol (Illumina).

173

174 **Transcriptome assembly and completeness**

175 The Illumina reads were adapter trimmed (Scythe, <https://github.com/vsbuffalo/scythe>) and quality filtered
176 (Sickle, <https://github.com/najoshi/sickle>). Reads were then separated according to their sequence tag using a
177 custom Python script. Reads sizing less than 15 bases were deleted. Quality check of the raw/filtered reads was
178 carried out using FastQC (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>). Filtered reads were
179 assembled using ABySS v.2.0 (Simpson et al. 2009) with default parameters; a preliminary assembly optimization
180 was performed by varying *k* values in the range 25-60 and the best assembly was picked using some metrics
181 adopted in Assemblathon2 (<https://github.com/ucdavis-bioinformatics/assemblathon2-analysis>). A final filtering
182 cutoff criterium (500 bp) was applied and shorter contigs were filtered out.

183 The ‘Bouche de Bétizac’ unigenes set was selected for transcriptome fine characterization procedures. Its initial
184 gene set was further filtered, for contaminants deriving from pests and fungi, using Blastn against three databases:
185 1) *Biorhiza pallida* transcriptome (gently provided by Prof. Graham Stone, Univ. of Edinburgh); 2) *Synergus*
186 *japonicus* draft genome (gently provided by Prof. Graham Stone, Univ. of Edinburgh); 3) *Aureobasidium*
187 *pullulans*. High ranked contigs having high identity with insects and the fungus sequences were removed.
188 Transcriptome completeness was then assessed by means of the CEGMA pipeline (Parra et al. 2007) measuring
189 the percentages of 248 different Core Eukaryotic Genes (CEGs) mapped in the chestnut assembly.

190

191 **Transcriptome functional annotation**

192 Functional annotation of the unigenes was locally performed with Blast2GO (Conesa et al. 2005) and gene
193 ontology (GO) terms were predicted by assigning functional classifications (Gene Ontology Consortium, 2000)
194 as well as potential properties of gene products. The blast cut-off E-value was 10^{-5} . The GO terms were assigned
195 to the representative transcripts for each sample through an enrichment analysis using Fisher's exact test (p-value
196 <0.01), with a false discovery rate (FDR) correction in terms of biological processes and molecular functions. The
197 unigenes were submitted to ORF predictor (Min et al. 2005;
198 <http://bioinformatics.yzu.edu/tools/OrfPredictor.html>) and compared with data from the *C. mollissima* genome
199 (www.hardwoodgenomics.org/organism/Castanea/mollissima).

200 *Resistance genes analogs (RGA) analysis* - candidates genes were identified by means of a Blastp analysis against
201 the Plant Resistance Genes database (<http://prgdb.crg.eu>; Sanseverino et al. 2012). Positive hits were validated
202 via HMMERv3 (hmmer.janelia.org/software) software, searching against PFAM hidden Markov models for NB-
203 ARC, TIR and several leucine-rich repeat motifs (Finn et al. 2016).

204 *Mirna target analysis* - Transcribed sequences were subjected to psRNATarget (Dai et al. 2011;
205 <http://plantgrn.noble.org/psRNATarget/>) analysis against miRBase Release 21 (Griffiths-Jones et al. 2008). A
206 maximum expectation of 2.5 was adopted, allowing a maximum energy to unpair the target site of 25, and
207 considering 17 bp upstream and 13 bp downstream of the target site sequence. Inhibition of translation was
208 considered for mismatches in the 9th to 11th mature miRNA nucleotides. Any enrichment of GO terms was verified
209 by comparing the putative miRNA targets against the whole transcript dataset by means of the Gossip package
210 implemented in the Blast2Go suite: a Fisher's exact test was applied collecting terms with P values $<e^{-4}$ and false
211 discovery rate <0.01 .

212

213 **Differentially expressed genes (DEGs)**

214 The produced clean reads were mapped to the reference transcriptome (B) using BWA software with default
215 parameters. Only the reads that could be uniquely mapped to the transcriptome were used for subsequent
216 processing. The retained reads were quantified using the “count” function implemented in GFOLD algorithm
217 (Feng et al. 2012; <https://bitbucket.org/feeldead/gfold>), which considered the expression level of each gene by
218 normalizing to the read per kilobase of exon per million mapped reads (RPKM) value. Differentially expressed
219 genes were identified using the “diff” function of GFOLD algorithm, which was biologically meaningful for single
220 replicate experiments. Genes with four fold change (GFOLD > 1 or < -1) were considered differentially expressed
221 between two samples. The transcriptome profile of the two varieties was analysed considering four pairwise
222 comparisons (Figure 2A; BI vs BNI; BI vs MI; MI vs MNI; BNI vs MNI).

223

224 **SSR identification and primer design**

225 SSR motifs were identified with the suite SciRoKo (Kofler et al. 2007; <http://kofler.or.at/bioinformatics>). Perfect
226 and imperfect mono, di-, tri-, tetra-, penta- and hexanucleotide motifs were targeted with default parameters.
227 Primer pairs were designed from the flanking sequences using Primer3 software (Rozen and Skaletsky 2000) in
228 batch mode, as implemented in the SciRoKo package. The target amplicon size range was set as 125-450 bp. The
229 optimal annealing temperature was 60° C, and the optimal primer length 20 bp.

230

231 **SNP mining**

232 Two transcriptomic read sets were constructed by pooling filtered reads from infested and not infested ‘Bouche
233 de Bétizac’ buds (BI + BNI), and reads from infested and not infested ‘Madonna’ buds (MI + MNI). The two
234 pools were independently back-aligned to the reference transcriptome (‘Bouche de Bétizac’) with the Burrows-
235 Wheeler Aligner (BWA, <http://bio-bwa.sourceforge.net>) using *mem* as algorithm with default parameters.
236 Automated SNP calling on was carried out on sorted bam alignment files by SAMtools mpileup and vcftools in a
237 multi-sample call pipeline. To populate the starting SNP table, a minimum mapping quality of 25 was required,
238 with a minimum SNP quality of 20. SNP characterization (test-cross/intercross; homozygous/heterozygous) was
239 addressed through custom bash scripts. The full SNP data set was organized into a relational database, available
240 upon request.

241

242 **Results**

243

244 **Transcriptome sequencing and assembly**

245 The whole Illumina sequencing experiment resulted in 361M raw pair ended reads with an average length of 100
246 bp. Filtering/trimming operations reduced the reads to 298M (83%, Table 1). The total amount of high quality
247 sequence was 59.7 Gb (‘Bouche de Bétizac’, 28 Gb; ‘Madonna’, 31.7 Gb).

248 *‘Bouche de Bétizac’ assembly metrics* - The best draft assembly was established using a k value of 47 and the
249 initial reference resulted in about 1 million contigs. By applying a cutoff of 500 bp, the assembled transcriptome
250 was reduced to 39,365 scaffolds of average length of 1,032 bp. The 50% of the *de novo* assembly (N_{50}) was
251 included in 11,440 scaffolds of 1,142 bp or larger, with 41.76% G+C bases (Table 2). About 40.6 Mb represented
252 the final assembly span. The longest scaffold was 13.7 kb. Contigs in the 500-1000 bp range were 24,880 (63.2%,
253 Fig. 1). The contigs exceeding 1kb were 14,481 (36.8%), of these 13,876 (35.3%) were in the length range 1000-
254 3000 bp.

255 *‘Madonna’ assembly metrics* - The best draft assembly was established using a k value of 45 and consisted of
256 30,605 scaffolds of average length of 1,018 bp. The 50% of the *de novo* assembly (N_{50}) was included in 8,886
257 scaffolds of 1,120 bp or larger, with 41.44% G+C bases (Table 2). About 31.2 Mb represented the final assembly
258 span. The longest scaffold was 5.4 kb. Contigs in the 500-1000 bp range were 19,809 (64.7%, Fig. 1). The contigs
259 exceeding 1kb were 10,796 (35.3%), of these 10,362 (33.9%) were in the length range 1,000-3,000 bp.

260

261 **Transcriptome filtering and completeness**

262 The *de novo* assembled transcriptomes of both cultivars were produced and, through a reciprocal blast analysis,
263 20,950 common unigenes were detected (in addition, 18,415 were specific for ‘Bouche de Bétizac’, 9,655 for
264 Madonna). High ranked contigs having high identity with insects and the fungus sequences were removed,
265 Overall, 1,039 cynipid-like and 4,245 fungi-like contaminant sequences were filtered out. The resulting ‘Bouche
266 de Bétizac’ filtered transcriptome contained 34,081 contigs. The assembled transcriptomes of the two cultivars
267 are provided in Online Resource file 1 (‘Bouche de Bétizac’) and in Online Resource file 2 (‘Madonna’). The
268 CEGMA (Core Eukaryotic Genes Mapping Approach), pipeline was adopted to assess the completeness of the
269 transcriptome of ‘Bouche de Bétizac’ (both unfiltered and insect-filtered sets). The transcriptome draft (unfiltered)
270 was surveyed for the presence of 248 conserved eukaryotic genes (CEGs). More than 69% of the 248 full-length
271 CEGs were mapped (Online Resource file 3), some of the missing CEGs were present as partial matches and when
272 included the mapped CEGs rose to 90%. The same pipeline was used to analyse a filtered transcriptome (Online
273 Resource file1), where some insect contaminant transcripts were removed (about 1039 sequences). Results
274 resembled the unfiltered data in both complete and partial alignments. ORF predictor detected about 33,000
275 sequences with ORF, which were used for further analyses (RGA and miRNA target mining).

276

277 **Transcriptome functional annotation and differential gene expression**

278 Blast2GO analysis of the 34,081 contigs produced the following results: 21,926 contigs were fully annotated,
279 2,202 contigs received just a Blast annotation, 3,683 received a GO Mapping, 573 received an "InterPro Scan"
280 annotation (Online Resource Fig. 1a). About 7,000 sequences were not effectively annotated. Overall, the
281 Blast2GO annotation permitted the functional annotation of 27,345 (71.3%) unigenes. The annotation was mainly
282 referred to UniProtKB (UniProt Consortium) and TAIR 10 (www.arabidopsis.org/) databases (Online Resource
283 Fig. 1b); four species provided a total of 16,391 "useful" annotations. The higher number of homologies with the
284 chestnut transcriptome reported in Online Resource Fig. 1c was found in *Arabidopsis thaliana* (6,310 sequences),
285 *Vitis vinifera* (5,565 sequences), *Populus trichocarpa* (2,339 sequences), and *Ricinus communis* (2,177
286 sequences).

287 The transcriptome profile of the two cultivars was analysed and four comparisons were carried out (Fig. 2a; BI vs
288 BNI; BI vs MI; MI vs MNI; BNI vs MNI). Infested ‘Bouche de Betizac’ buds (cynipid-resistant) compared to the
289 healthy buds (BI vs BNI) showed a relative low regulation (389 genes up and 168 down-regulated). Infested
290 ‘Madonna’ buds (cynipid-susceptible), compared to the healthy buds (MI vs MNI), showed a major down-
291 regulation (706 genes up and 2,108 down-regulated). Infested ‘Bouche de Betizac’ buds (cynipid-resistant)
292 compared to infested ‘Madonna’ buds (cynipid-susceptible; BI vs MI) showed a high level of regulation (2,488
293 genes up and 2,178 down-regulated). Indeed a very high variation in transcriptome was observed between the two
294 healthy cultivars (BNI vs MNI; 2,5k genes upregulated and 4,9k genes down-regulated, Fig. 2a). The genes
295 regulated in infested ‘Bouche de Bétizac’ and ‘Madonna transcriptomes are shown in Online Resource table 1 and
296 were further analyzed. Considering the biological processes (Fig. 3), almost 300 “response to stimulus”
297 (GO:0050896) related genes appeared up regulated. Up to 70 genes involved in “post embryo development”
298 (GO:0009791) appeared regulated and most of them were transcription factors involved in the plant development
299 (Fig. 3). Many up and down regulated proteins appeared associated to “death” (GO:0016265) processes and
300 “apoptosis” (GO:006915) and some of them were involved in the hyper-sensitive response. In the molecular
301 functions, the “transcriptional regulator activity” (GO:0030528) term was enriched only in the up-regulated genes.

302 In the cellular components, the “vacuolar part” (GO:0044437) term appeared significantly enriched only in up-
303 regulated genes (Fig. 3). Considering only the genes that were commonly regulated “BI vs BNI” and “BI vs MI”
304 (Venn diagram intersection, Fig. 2b) and not regulated in “BNI vs MNI” and “MI vs MNI”, ~100 genes were
305 highlighted (Fig. 2b; Online Resource table 2). Some GO enrichments were still highlighted for specific GO terms
306 (Fig. 3), such as some biological processes involved in response to stimulus (GO:0050896), and developmental
307 processes (e.g.: post embryonic development, GO:0009791). Many up-regulated genes appeared to be
308 transcription factors (e.g.: RAV1, AP2/ERF, WRKY33) or protein regulators (e.g.: RAPTOR1B) and storage
309 proteins (e.g.: LEA D29) involved in post-embryonic development.

310

311 **Resistance genes and miRNA target**

312 Protein sequences of 112 reference RGAs (from RGDB) were used to perform BLASTp searches against chestnut
313 unigenes. A total of 1,444 unigenes (Online Resource file 4), showing homology to 82 univoque proteins out of
314 the 112 encoded reference RGAs, were identified. Their putative functions (Fig. 4) were identified and 32 (39.0%)
315 belonged to the CC-NB-LRR type (CNL), 9 (10.9%) belonged to the TIR-NB-LRR type (TNL), 4 (4.9%)
316 belonged to the NB-LRR type (NL), 12 (14.6%) belonged to the receptor-like protein type (RLP), 10 (12.2%)
317 belonged to the receptor-like kinase class (RLK), and 1 (1.2%) belonged to the kinase-resistance related type.
318 Some of these proteins appeared as regulated by the presence of the cynipid (Fig. 2).

319 The assembled transcriptomes of both cultivars were scanned for the presence of recognition sites for known plant
320 miRNAs (miRNA targets) and the results are provided in Table 3 as well as in Online Resource file 5 (‘Bouche
321 de Bétizac’) and in Online Resource file 6 (‘Madonna’). The contigs/scaffolds of ‘Bouche de Bétizac’ showed, in
322 total, target annealing sites for 249 miRNAs, located in 1135 transcripts (Online Resource table 3). A total of 185
323 targets belonged to the ath-miR5021 family; a total of 146 targets belonged to the ath-miR5658; a total of 139
324 targets belonged to the ath-miR414 family. ReviGO analysis (Fig. 5) showed some GO enrichments for miRNA
325 targets transcripts (Online Resource table 4), particularly for the categories: nitrogen compound metabolic process
326 (GO:0006807), nucleobase-containing compound metabolic process (GO:0006139), developmental process
327 (GO:0032502), shoot system development (GO:0048367), phyllome development (GO:0048827) and response to
328 stimulus (GO:50896). They included 21 genes: three were involved in the trichome development, four were genes
329 involved in stress related phenomena and fourteen were miRNA target related to tissue development (leaf, shoot,
330 inflorescence). The latter group contained three structural proteins, five enzymes and six transcriptional factors.
331 Some of these proteins (Online Resource table 1) were significantly regulated in the resistant cultivar in the
332 presence of the cynipid.

333

334 **SSR identification and primer design**

335 A screening of the reference transcriptome resulted in the identification of 5,713 scaffolds containing 11,364
336 putative SSRs. About 14.9% of the unigenes contained an SSR (one SSR per 3.5 Kb) and the most abundant repeat
337 motifs were mono-nucleotides (4,519; 39.8%), followed by tri-nucleotides (2,296; 20.2%), di-nucleotides (1,908;
338 16.8%), hexanucleotides (1,282; 11.3%), penta-nucleotides (719; 6.3%) and tetra-nucleotides (640; 5.6%). The
339 most common di- and tri-nucleotide motifs were AG (1,449, 12.8%) and AAG (629; 5.5%). Frequencies and
340 repeat numbers for the 20 most present SSR motifs are reported in Figure 6 and complete statistics are presented

341 in Online Resource file 7). A batch analysis permitted the design of PCR primers for all the loci, leading to the
342 generation of 7,176 putative markers (Online Resource file 8).

343

344 **SNP mining**

345 Considering the two cultivars ‘Bouche de Bétizac’ (as reference) and ‘Madonna’, 335,468 reliable SNPs/Indels
346 (DP>10), across the two accessions, were detected. On the whole, 321,939 were SNPs and 13,529 were Indels
347 and among SNPs, 206,015 were transitions, 115,515 are transversions (ratio=1.78). Since the assembled
348 transcriptome is 36,094,445 bp long in 34,081 contigs, the average SNP frequency was calculated at 1/124 bp with
349 a mean of 8.4 SNP/INDEL per contig.

350 *SNP (inter/intra genotype)* - The number of SNPs/Indels between the two accessions was 335,468. The number
351 of SNPs in homozygous state between them was 25,154. The number of heterozygous loci was 232,578 in the
352 ‘Bouche de Bétizac’ genotype, and 159,742 in the ‘Madonna’ genotype.

353 *SNP markers* - Considering only SNP variants, loci were classified into those expected to segregate in a 1:1 ratio
354 ("testcross markers" AA x AB or AB x BB), and those in a 1:2:1 ratio ("intercross markers"; AB x AB). Testcross
355 markers were 76,764, considering the ‘Bouche de Bétizac’ genotype over the ‘Madonna’ one and 149,600
356 considering ‘Madonna’ genotype over ‘Bouche de Bétizac’ one (Table 4). Overall testcross markers were 226,364.
357 Intercross markers were 82,978 (24.7% of the total). Testcross and intercross SNPs were in all 309,343 (92.2% of
358 the total).

359

360 **Discussion**

361 The first fully resistant to gall wasp cultivar found was the hybrid ‘Bouche de Bétizac’ that showed no symptoms
362 both in orchard and under controlled conditions (Sartor et al., 2009). For this reason this cultivar was used for a
363 transcriptomic approach, starting from the hypothesis that resistance may be due to a hypersensitive reaction in
364 the bud tissues (Dini et al. 2012).

365 To conduct genetic dissection of traits involved in the insect-plant interaction and proceed to breeding practices,
366 there is a need for a transcriptome/genome reference sequence. High-throughput RNA sequencing is a useful
367 approach to obtaining a complete set of transcripts from species of interest. Because of the potential advantages
368 of these technologies (high-throughput vs low costs), many transcriptomes from model/non-model species have
369 been sequenced and assembled in the last years (over 2,100 papers in the period 2000-2018, ISI - Web of Science
370 survey). This consolidated approach was used, in the present study, to reconstruct the transcriptome of *C. sativa*
371 buds under biotic stress (i.e.: in the presence/absence of the chestnut gall wasp). In ‘Bouche de Bétizac’, a total of
372 34,081 unigenes were obtained by optimizing assembly procedures. The transcriptomes of both cultivars were
373 properly assembled, and while the ‘Bouche de Bétizac’ unigenes set (belonging to the resistant cultivar) was
374 selected for the functional characterization, the ‘Madonna’ one was just used for RNAseq data analysis, and
375 provided as supplementary materials. The ‘Bouche de Bétizac’ assembly was evaluated for its completeness with
376 CEGMA, Parra et al. 2007). This pipeline uses 248 Core Eukaryotic Genes (CEGs), which are highly conserved,
377 present in low copy numbers in higher eukaryotes, to describe the gene space. Based on the average degree of
378 conservation observed from each CEG, the CEGMA pipeline divides the CEGs into four groups (group 1 has the
379 least conserved CEGs while group 4 has the most conserved CEGs). The alignment trends (Online Resource file
380 1) using the new *C. sativa* reference transcriptome (‘Bouche de Bétizac’, filtered) showed how the lack of

381 complete alignments for less conserved ortholog groups (65%) is due to divergence, while partial alignments
382 confirm an even representation of different ortholog groups, indicating a gene space coverage of about 90% (range
383 86-95 %). Blast2GO analysis annotated 27,345 (71.3%) unigenes containing a wide range of biological, cellular
384 and molecular functions typical of a plant transcriptome resembling similar transcriptome assemblies (Garcia-
385 Seco et al. 2015; Cardoso-Silva et al. 2014), involving all the physiological processes (i.e.: Biological Process),
386 the majority of cellular compartments (i.e.: Cellular Component), and the functions of the proteins produced (i.e.:
387 Molecular Function).

388 A differential gene expression analysis was conducted to highlight regulated genes emerging in two genetic
389 contexts (cultivars susceptible/resistant to the cynipid) following the infestation with the wasp. The RNAseq
390 analyses were conducted using bulks of buds to catch macroscopic variations between resistant and sensitive
391 varieties in the presence of the pest. To the scope, genes were analyzed with the GFOLD suite, specifically
392 implemented as a tool for studies with few or no replicates (Feng et al. 2012), as it generalizes the fold change by
393 considering the posterior distribution of log fold change, such that each gene is assigned a reliable fold change.
394 Considering a 1-fold variation, up to 557 genes showed to be regulated in the infested ‘Bouche de Bétizac’
395 transcriptome, while 2814 in the infested cultivar ‘Madonna’, most of which (75%) were down-regulated genes.
396 This high difference in number of regulated genes was mostly expected since the phenotype of the sensitive buds
397 in respect of the resistant ones appeared very different (Dini et al. 2012). As reviewed elsewhere (Schuman et al.
398 2016) plant respond to pests with a multilayer approach. Here, we observed, as expected, many and different
399 “response to stimulus” up-regulated genes. Among them, some were involved in the likely recognition of specific
400 elicitors and patterns of damage. Indeed, as many as sixty LRR proteins were observed to be regulated during the
401 interaction between chestnut bud and cynipid; some other genes were implied with a “transcriptional regulator
402 activity” role. Intriguingly, 16 WRKY and 6 ERF/AP2 genes were here observed as up regulated. Recently, those
403 categories of transcription factors are reported to be involved in response to both aphid attack and *P. syringae*
404 infection regulation, but it is known that some of them are up-regulated in an insect-specific manner (e.g.: WRKY-
405 33, Barah et al. 2013). Dini et al. (2012) highlighted the occurrence of an HR in the resistant cultivar ‘Bouche de
406 Bétizac’ as response to the cynipid infestation, resulting in cell and larvae death. This fact was here confirmed,
407 since more than 100 genes appeared associated to “death” and “apoptosis” processes, including genes for HR
408 response. Also the “vacuolar part” term appeared significantly enriched in up-regulated genes; they were
409 prominently positively regulated membrane proteins coding for ionic channels and pumps, consistently with the
410 role attributed to vacuolar proteins in plant immunity (Hatsugai 2015; Zhang et al. 2010). Some other genes
411 involved in “post embryo development” were over-represented. For example, Contig_32550 is a putative
412 homologue to RAV1 transcription factor, which has been suggested to be a negative regulator of growth and
413 development. The regulation of RAV1 (and RAV2) may serve not only for immediate physiological responses,
414 but also for developmental adaptation in response to the environmental stimuli, such as response to touch stimulus,
415 happening during the cynipid growth in the gall infection.

416 During gall formation, many biochemical, physiological, and molecular changes occur in plant tissues requiring
417 continuous activity of unknown stimuli (Harper et al. 2004). Studies on the insect-plant interaction carried out on
418 cynipids of oak and rose showed three stages of gall formation: initiation, growth and maturation (Harper et al.
419 2004). The growth stage starts with cell proliferation, differentiation and hypertrophy due to stimuli from the
420 cynipid gall wasps that are able to redirect host-plant development to form novel structures to protect and nourish

421 the developing larvae. Comparisons between inner-gall and non-gall tissue protein signatures by Schönrogge et
422 al. (2000) have identified a number of inner-gall proteins, such as a NAD-dependent formate dehydrogenase
423 (NFD) and a biotin carboxyl carrier protein (BCCP), the latter being a subunit of a class II acetyl CoA-carboxylase
424 (ACCase), involved in the production of triacylglycerol lipids. Further studies (Harper et al. 2004) showed that
425 genes for this inner-gall putative BCCP reveal differential expression throughout gall development. Fluorescent
426 in situ hybridization demonstrated many of the inner-gall cells to be polytenized. The expression of putative BCCP
427 and the polytenization of the nuclei in gall cells are typical of nutritive, secretory cells, found in seeds and also in
428 tapetal cells in pollen. Recent studies (Pawłowski et al. 2017) on protein patterns in healthy and gall tissues showed
429 changes in abundance of 21 proteins. Interestingly, some functions (Online Resource Table 1) appeared here to
430 be up-regulated in infested ‘Bouche de Bétizac’, such as some subunit of ATP synthase and many HSPs, while
431 others showed to be down-regulated in Madonna infested galls, such as ascorbate peroxidase, actin and many
432 stress-related and pathogenesis-related proteins (PRP).

433 Each contig was scanned for the presence of recognition sites for known plant miRNAs (miRNA targets). The
434 latter analysis highlighted some findings, which deserves to be deeply discussed. microRNAs (miRNAs) were
435 discovered in ‘90s and are now recognized as one of the major regulatory gene families in plants and in eukaryotes
436 in general. They play important roles in a variety of biological phenomena, such as development and responses to
437 abiotic and biotic stresses, by regulating complementary target transcripts. In particular, it was primarily
438 recognized that miRNA activity results in gene expression repression impairing mRNA stability, by guiding
439 mRNA degradation, at a protein synthesis initiation level, by its inhibition, or through degradation of a protein,
440 via the binding of the 3’UTR of a target transcript. We highlighted target annealing sites for 249 miRNAs located
441 in 1,135 transcripts and, interestingly, we observed enrichments for certain specific GO categories. The more
442 intriguing were the ones involved in some developmental processes (e.g.: shoot/phyllome development), as well
443 as the ones involved to the response to stimulus (Online Resource Table 3). The gFOLD analysis showed a trend
444 of up-regulation in infested buds belonging to the susceptible cultivar (‘Madonna’) and thus, as expected, we
445 spotted some enriched GO terms likely related to plant/insect interaction, which deserve to be discussed. A first
446 example of regulated genes is referred to some miRNA targets (in contig_4038, contig_37578, contig_2740 and
447 contig_15619), regulated by a unique miRNA (miR5658), which are involved in the energetic
448 regulation/reprogramming of the cell in condition of stress and involved in defense responses, as well as in growth
449 and development (Online Resource Table 4). Some other miRNA target genes were observed (Online Resource
450 Table 4) and most of them were transcription factors (GRF7, ZFP8, DOT2, TCP3, ATHB-15) or generally
451 involved in gene regulation (LHP1, YUC4, ARF) playing a role in root, shoot and flower development as factors
452 most likely involved in the reshaping of the tissue towards the formation of a suitable gall. These enrichments are
453 intriguing and will be the target for future analyses aimed at understanding the functions of the identified genes
454 and thus providing useful tools to molecular breeders. Indeed, RNA interference technology involving siRNA and
455 miRNA have emerged as an attractive tool used by plant biologists not only to decipher the plant function, but
456 also to develop plants with improved and novel traits by the manipulation of both desirable and undesirable genes.
457 One of our target was the identification of SNP markers; however, transcriptomic data from NGS sequencing
458 made it also possible the mining of microsatellite motifs. SSR markers are multi-allelic and are widely applied for
459 genetic analyses, regardless of their cost for development and for implementation in throughput facilities. During
460 the last years the exploitation of publicly available EST database, at first, and the explosion of NGS data

461 production, secondarily, led to the identification of several thousands of new markers in virtually almost every
462 plant species (Portis et al. 2016). The most abundant repeat motifs were mononucleotides, followed by
463 trinucleotides, dinucleotides and hexanucleotides. This seems not consistent with the observations in Poplar and
464 Arabidopsis (Morgante et al. 2002), where tri-nucleotides are the most represented in transcriptomes; from our
465 analyses most of the mononucleotide motifs could be represented by terminal polyA (this happens for over 3,500
466 sequences). A more realistic number of mononucleotide repetition could be thus calculated to be about one
467 thousand (8.8%). The most common di- and tri-nucleotide motifs were AG (1,449, 12.8%) and AAG (629; 5.5%)
468 in accordance to the observation in a previous study, where AG and AAG were the predominant motifs (Stagel et
469 al. 2008; Zeng et al. 2010; Portis et al. 2007; Morgante et al. 2002). About 14.9% of the unigenes contained an
470 SSR (one SSR per 3.5 Kb), which is a value comparable to the success rate recorded from other fruit crops (Rai
471 et al. 2015).

472 SNP frequencies in the *Castanea sativa* transcriptome appear to be comparable to that found in the outbred, highly
473 heterozygous *Cynara cardunculus* transcriptome (Scaglione et al. 2012) and among *Citrus* species ESTs (Jiang et
474 al. 2010). Overall, 335k SNP/indels were identified and the number of loci in heterozygous state was very high
475 (232,578 in 'Bouche de Bétizac', and 159,742 in 'Madonna'). This is expected in *Castanea sativa* that being an
476 outbreeding crop, presents a high level of heterozygosity. Genetics of *C. sativa* has been limited studied so far, and
477 very few mapping populations are available. The testcross and intercross SNP markers identified (in all 309,343
478 markers) will be suitable information for mapping purposes using F₁ progeny in a 2 way pseudo-test cross
479 approach. The core set of SNPs will be pivotal to setup SNP arrays or to design custom SPET (Single Primer
480 Enrichment Technology, Nugen) assays, as example of targeted resequencing approach, through the selection of
481 known polymorphic regions in gene space.

482 Overall, the *de novo* assembly of the transcriptome chestnut, combined with extensive homology analyses, yielded
483 a number of contigs comparable to those found in literature for similar RNAseq experiments. The work is still at
484 a preliminary stage, but it was successful in classifying contigs, and once data will be readily available to the
485 international scientific community, they will guide a better understanding of the interaction chestnut-gall wasp.
486 Altogether, the corpus of produced information will lead to investigate the different mechanisms of resistance
487 against cynipid, and to address breeding strategies towards resistant cultivars. To the scope, the bioinformatics
488 pipeline adopted enabled identification of a large set of SSR/SNP markers for practical applications in breeding
489 programs and provenance/pedigree tracking. We believe that the availability of these transcriptome data for *C.*
490 *sativa* will contribute to understand the genetic basis of the resistance to gall wasp and meet the informational
491 needs for molecular genetic studies of this species and its relatives.

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496 **Data Archiving Statement**

497 Sequences have been submitted to NCBI's Short Read Archive (SRA).

498 SRA accession: PRJNA509688

499

500 **Compliance with Ethical Standards**

501 This article does not contain any studies with human participants or animals performed by any of the authors.

502

503 **Conflict of Interest**

504 The authors declare that they have no conflict of interest and in particular:

505 Author 1 (Alberto Acquadro) declare that he has no conflict of interest

506 Author 2 (Daniela Torello Marinoni) declare that she has no conflict of interest

507 Author 3 (Chiara Sartor) declare that she has no conflict of interest

508 Author 4 (Francesca Dini) declare that she has no conflict of interest

509 Author 5 (Matteo Macchio) declare that he has no conflict of interest

510 Author 6 (Roberto Botta) declare that he has no conflict of interest

511

512

513

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647 TABLES

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649 **Table 1 Sequencing results and statistics after polishing procedures.**

Cultivar	Raw reads (M)	filtered/trimmed reads (M)	Amount of sequence (Gb)
'Bouche de Bétizac', infested	59.77	49.02	9.8
'Bouche de Bétizac', not infested	110.15	90.93	18.2
'Bouche de Bétizac' (total)	169.92	139.95	28.0
'Madonna' infested	75.21	62.74	12.6
'Madonna' not infested	115.81	95.55	19.1
'Madonna' (total)	191.02	158.29	31.7
Total	360.94	298.24	59.7

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653 **Table 2 Statistics for the *de novo* assembled transcriptomes.** Values were calculated using
654 the Perl script `assemblathon.pl` (Bradnam et al. 2013).

Characteristics	'Bouche de Bétizac'	'Bouche de Bétizac' filtered	'Madonna'
Number of scaffolds	39,365	34,081	30,605
Total size of scaffolds	40,621,562 (39)	36,094,445	31,159,190
Longest scaffold	13,723	13,723	5,405
Shortest scaffold	500	500	500
Number of scaffolds > 1K nt	14,481 (36.8%)	13,112 (38.5%)	10,782 (35.2%)
Number of scaffolds > 10K nt	4 (0.0%)	4 (0.0%)	0
Number of scaffolds > 100K nt	0	0	0
Mean scaffold size	1,032	1,059	1,018
Median scaffold size	818	835	801
N50 scaffold length	1,142	1,191	1,12
L50 scaffold count	11,44	9,747	8,886
% GC	41.76	41.50	41.44

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Table 3 Abundance of putative miRNA annealing sites in the *Castanea sativa* transcriptome. miRNA families occurring fewer than four times were incorporated into the category labelled ‘other’.

miRNA family	No. of targets	miRNA family	No. of targets
ath-miR5021	185	ath-miR1886.1	5
ath-miR5658	146	ath-miR1886.2	5
ath-miR414	139	ath-miR4243	5
ath-miR838	15	ath-miR5641	5
ath-miR854a	13	ath-miR156i	5
ath-miR854b	13	ath-miR156a	4
ath-miR854c	13	ath-miR156b	4
ath-miR854d	13	ath-miR156c	4
ath-miR854e	13	ath-miR156d	4
ath-miR5653	13	ath-miR156e	4
ath-miR865-3p	11	ath-miR156f	4
ath-miR834	10	ath-miR171b	4
ath-miR400	9	ath-miR171c	4
ath-miR396a	8	ath-miR395a	4
ath-miR396b	8	ath-miR395d	4
ath-miR5648-5p	8	ath-miR395e	4
ath-miR4221	7	ath-miR397a	4
ath-miR157d	6	ath-miR397b	4
ath-miR163	6	ath-miR407	4
ath-miR837-5p	6	ath-miR156h	4
ath-miR156j	6	ath-miR415	4
ath-miR5654-3p	6	ath-miR447a.2-3p	4
ath-miR157a	5	ath-miR472	4
ath-miR157b	5	ath-miR831	4
ath-miR157c	5	ath-miR861-5p	4
ath-miR395b	5	ath-miR773b-3p	4
ath-miR395c	5	ath-miR5016	4
ath-miR395f	5	ath-miR5652	4
ath-miR773a	5	ath-miR5998a	4
ath-miR830-3p	5	ath-miR5998b	4
ath-miR835-5p	5	Others	305
ath-miR866-3p	5		

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Table 4 | Testcross and intercross markers evaluation. Data represents SNP sites having sequence information for each of the two samples analyzed.

SNP markers	‘B. de Bétizac’ vs ‘Madonna’	‘Madonna’ vs ‘B. de Bétizac’	In common
Putative testcross	76,764	149,600	-
Common intercross	-	-	82,978

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FIGURE CAPTIONS

Fig. 1 The distribution of the scaffold length in the chestnut transcriptome Assembled scaffold size: the length interval measured were set to 500 bp. Blue bars represent ‘Madonna’ scaffold; grey bars represent ‘Bouche de Bétizac’ scaffold.

Fig. 2 Differential gene expression in chestnut buds in the presence of the cynipid in resistant/susceptible cultivars. (a) Histogram representing variation of genes (> 1-fold) after inoculation of the cynipid in the two cultivars, considering four possible comparisons (BI vs BNI; BI vs MI; MI vs MNI; BNI vs MNI). (b) Venn diagram intersection of the four transcriptomes comparison. A white square highlights the genes commonly regulated in “BI vs BNI” and “BI vs MI” and not regulated in “BNI vs MNI” and “MI vs MNI”.

Fig. 3. Enriched GO terms in terms of biological processes, cellular components and molecular functions in up-regulated and down-regulated genes of ‘Bouche de Bétizac’ over ‘Madonna’ transcriptomes; X-axis and Y-axis are expressed as semantic space, using color (log₁₀ p-value) and size variation (log size).

Fig. 4 RGA analysis in the chestnut transcriptome (a) Representation of putative and univocal RGAs in the 34,081 analysed unigenes. (b) Analysis of the different categories of RGAs in the 82 univocal chestnut RGAs. The CNL class comprises resistance genes encoding proteins with at least a coiled-coil domain, a nucleotide binding site and a leucine-rich repeat (CC-NB-LRR); the TNL class includes those with a Toll-interleukin receptor-like domain, a nucleotide binding site and a leucine-rich repeat (TIR-NB-LRR); the RLP class, acronym for receptor-like protein, groups those with a receptor serine– threonine kinase-like domain, and an extracellular leucine- rich repeat (ser/thr-LRR); the RLK class contains those with a kinase domain, and an extracellular leucine-rich repeat (Kin-LRR); the ‘Others’ class includes all other genes which have been described as conferring resistance through different molecular mechanisms, e.g. Mlo and Asc-1; the kinase contain a kinase domain involved in resistance process.

Fig. 5 miRNA target enrichment analysis in the chestnut transcriptome (a) Representation GO category enriched in the miRNA target transcriptome subset. X axis is expressed in log₁₀ p-value, Y axis is expressed as semantic space scale.

Fig. 6 SSR most represented motifs

SUPPLEMENTARY MATERIAL CAPTIONS

ONLINE RESOURCE FILES

Online Resource file 1 The assembled transcriptome of the cultivar ‘Bouche de Bétizac’, filtered for contaminants deriving from pests and fungi

Online Resource file 2 The assembled transcriptome of the cultivar ‘Madonna’

Online Resource file 3 CEGMA pipeline results on the *C. sativa* transcriptome Prots = number of 248 ultra-conserved CEGs present in genome; %Completeness = percentage of 248 ultra-conserved CEGs present; Total = total number of CEGs present including putative orthologs; Average = average number of orthologs per CEG; %Ortho = percentage of detected CEGs that have more than 1 ortholog

Online Resource file 4 Unigenes, showing homology to 82 univoque proteins out of the 112 encoded reference RGAs, identified

Online Resources file 5 miRNA targets in cultivar ‘Bouche de Bétizac’

Online Resources file 6 miRNA targets in cultivar ‘Madonna’

Online Resource file 7 Complete statistics for the identified SSR loci

Online Resource file 8 primers designed for the selected SSR loci

ONLINE RESOURCE TABLE

Online Resource table 1 List of genes regulated in infested ‘Bouche de Bétizac’ and ‘Madonna transcriptomes

Online Resource table 2 List of the genes commonly regulated in “BI vs BNI” and “BI vs MI” (Venn intersection, Fig. 2C) and not regulated in “BNI vs MNI” and “MI vs MNI”

Online Resource table 3 Genes showing enriched GO-terms among the putative miRNA target transcripts

Online Resource table 4 The over-representation of GO-terms among the putative miRNA target transcripts. P-value (<0.01) was used to assess statistical significance. AgriGO was used to obtain GO terms from presumptive miRNA target and ReviGO was used to evaluate enriched GO terms

ONLINE RESOURCE FIGURE

Online Resource Figure 1 Annotation and categorization analysis of the chestnut transcriptome. (a) Blast2GO results. (b) Species more represented in the blast analysis and top blast hits. (c) Chart giving the distribution of the number of annotations (GO-terms) retrieved from the different source databases (e.g. UniProt, PDB, TAIR)