



AperTO - Archivio Istituzionale Open Access dell'Università di Torino

Nanomedicine formulations for the delivery of antiviral drugs: A promising solution for the treatment of viral infections

This is the author's manuscript Original Citation: Availability: This version is available http://hdl.handle.net/2318/1662691 since 2018-03-17T18:10:28Z Published version: DOI:10.1080/17425247.2017.1360863 Terms of use: Open Access Anyone can freely access the full text of works made available as "Open Access". Works made available

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)





This is the author's final version of the contribution published as:

Lembo D, Donalisio M, Civra A, Argenziano M, Cavalli R. Nanomedicine formulations for the delivery of antiviral drugs: a promising solution for the treatment of viral infections, Expert Opin Drug Deliv., 15(1), 2018, 93-114, doi: 10.1080/17425247.2017.1360863.

The publisher's version is available at:

www.tandfonline.com/loi/iedd20

When citing, please refer to the published version.

Link to this full text: http://hdl.handle.net/314205 This full text was downloaded from iris-Aperto: https://iris.unito.it/

iris-AperTO

University of Turin's Institutional Research Information System and Open Access Institutional Repository

Nanomedicine formulations for the delivery of antiviral drugs: A promising solution for the treatment of viral infections

3

4 Abstract

Introduction. Viral infections represent a public health problem and one of the leading causes of global mortality. Nanomedicine strategies can be considered a powerful tool to enhance the effectiveness of antiviral drugs, often associated with solubility and bioavailability issues. Consequently, high doses and frequent administrations are required, resulting in adverse side effects. To overcome these limitations, various nanomedicine platforms have been designed.

10 Area covered. This review focuses on the state of the art of organic-based nanoparticles for the 11 delivery of approved antivirals. A brief description of the main characteristics of nanocarriers is 12 followed by an overview of the most promising research addressing the treatment of most important 13 viral infections.

Expert Opinion. The activity of antiviral drugs could be improved with nanomedicine formulations. Indeed, nanoparticles can affect the fate of the encapsulated drugs, allowing controlled release kinetics, enhanced bioavailability, modified pharmacokinetics, and reduced side effects. In addition, the physicochemical properties of nanocarriers can enable their capability to target specific sites and to interact with virus structures. In this regard, nanomedicines can be considered an opportunity to enhance the therapeutic index of antivirals. Efficacy, safety, and manufacturing issues need to be carefully assessed to bring this promising approach to the clinic.

Keywords: antiviral drugs, nanomedicines, viral infections, nanoparticles, nanotherapeutics, targeted
 delivery.

- 23
- 24
- 1

25 Article highlight box

- Nanocarrier incorporation of antiviral drugs can modify their pharmacokinetics and pharmacodynamics properties and reduce the side effects.
- Long-acting nanoformulations can be applied in the treatment of infectious diseases to promote
 compliance and patient adherence.
- Nanomedicines formulations can overcome biological barriers and achieve effective drug concentrations in viral reservoirs.
- The peculiar physico-chemical properties of nanocarriers can enable their capability to interact
 with virus structures.
- Nanomedicine can provide an effective platform for treating viral diseases by small interfering
 RNA (siRNA) delivery.
- Nanomedicine strategies can be considered a powerful tool to enhance the potential of currently
 approved antiviral drugs.

39 Index

- 40 1. Introduction
- 41 2. Nanodelivery systems for antiviral drugs
- 42 3. Nanoformulations for approved drugs against specific human viral infections
- 43 4. Future challenges
- 44 5. Conclusion
- 45 6. Expert Opinion

47 **1. Introduction**

Viral infections pose significant global health challenges, particularly because resistant viral strains and adverse side effects associated with the prolonged use of antiviral drugs can reduce the effectiveness of antiviral therapies. In the past several decades, a new paradigm for enhancing the efficacy of bioactive molecules – the use of nanotechnology in medicine (i.e., nanotherapeutics and nanodelivery systems) – has attracted increasing attention and has been largely exploited in many medical fields, both in basic research and in the drug development pipeline [1].

The use of nanodelivery systems offers some advantages that can improve therapeutic treatments. Indeed, the biopharmaceutical properties, and those of absorption, distribution, metabolism and excretion (ADME), of conventional drugs depend upon their physicochemical characteristics.

57 Nanomaterials can affect and govern the fate of the encapsulated drug [2]. Interestingly, nanocarrier 58 formulations can modify the physico-chemical properties of the incorporated molecules, thereby 59 allowing sustained/controlled release, modified pharmacokinetics, and targeting specific sites of action. 50 This can lead to an increase in the effectiveness of the drug and a decrease in the associated side effects 51 [3].

62 In this context, the nanomedicine rationale can be exploited to enhance the therapy of viral infections, taking into consideration the narrow therapeutic indices, high doses, and frequent administration 63 needed for a number of antivirals drugs due to their limited aqueous solubility, short half-life, and/or 64 65 slow uptake by the body tissues [4]. In addition to the physico-chemical limitations of drugs, prolonged anti-viral therapy can result in the development of drug-resistant virus strains necessitating daily 66 repeated administrations, which can lead to a demanding drug-regimen. This is especially the case of 67 chronic diseases, such as the acquired human immunodeficiency syndrome (AIDS) due to human 68 immunodeficiency virus (HIV) infection, where no suspension of treatment is allowed in order to avoid 69 viral resistance [5]. Thus, it is clear that the treatment of viral diseases is a challenging problem. 70

However, nanomedicine-based strategies may be useful for both the prevention and therapy of these diseases. The design of a nanodelivery systems can be approached from different points of view, although it generally takes into account the type of nanocarrier/nanomaterial used and the type of viral disease, leading to approaches that are respectively technologically and biologically driven.

Viruses act as intracellular parasites that use the replication system of the infected cells because they 75 76 lack the genetic information necessary for the generation of metabolic energy and macromolecular 77 synthesis. This behavior leads to an intricate relationship between viruses and infected cells that should be considered in the development of efficient, safe and targeted therapies. Current antiviral drugs 78 79 interfere with specific phases of the virus life cycle by targeting viral structures and enzymes that are 80 essential for virus replication. The major mechanisms of antivirals actions against DNA and RNA 81 viruses are reported in Figure 1. Most RNA viruses replicate in the cytoplasm, whereas DNA viruses, 82 Influenza virus and retroviruses (HIV) enter into the nucleus for genome replication. Notably, HIV reverse transcriptase acts before the entry of the genome into the cellular nucleus; by contrast, HBV 83 84 reverse transcriptase acts on a viral RNA genome of new viral progeny.

85 Intriguingly, nanomedicine formulations could represent a new avenue for controlling the amount, dosage frequency, and delivery site of antivirals, as well as for targeting the virus life cycle. Indeed, the 86 features of nanoparticles, such as sizes, morphology and surface charge, can be modulated to promote 87 88 the targeting of the drugs. Moreover, nanocarriers can be engineered to enhance their ability to reach 89 specific extracellular or intracellular targets and to compete with viruses for attachment to cell surface 90 receptors, both of which are key factors for controlling viral diseases and overcoming drug resistance. Consequently, passive and active drug targeting are possible with nanomedicine approaches, as 91 92 previously described for anticancer drugs [6]. Passive targeting depends on the composition and 93 physicochemical characteristics of the nanocarriers; active targeting is based on the presence of a 94 specific ligand on the nanocarrier surface, which acts as a recognition device for viral nanostructures.

95 These drug-targeting strategies might facilitate the local and specific release of antiviral drugs while
96 minimizing damage to healthy cells and tissues, thereby preventing side effects.

In addition, these approaches can be exploited for the design of broad-spectrum antiviral-based 97 nanodelivery systems that target the viral membrane or cell receptor to compete with attachment of the 98 virus to the target cell, thereby preventing virus internalization [7]. Recently, suitable engineered 99 100 nanoparticles (NPs) were developed that interact with cell receptors (i.e., glycan receptors) and 101 competitively bind to cells to prevent viral infection [8]. Interestingly, this rationale has been proven by using liposomal decoys to capture influenza virus and delay the disease [9]. Alternatively, 102 103 nanoassemblies have been generated to mimic glycan receptors, thereby neutralizing heparin sulfate 104 dependent viruses [10]. In addition, the site-specific drug release potential, as well as the ability to overcome biological barriers of nanomedicine, can offer the advantage of decreasing the administered 105 dose regimen of antivirals. To date, a number of nanocarriers, either organic (e.g., liposomes, NPs, 106 micelles) or inorganic (e.g., silver and gold NPs), have been studied to improve the delivery and 107 therapeutic efficacy of antiviral medicines. Consequently, various potential nanostructures might be 108 fine-tuned and employed to meet the different needs of viral disease therapy [11]. 109

This review highlights the state of the art of organic NPs used for the delivery of approved antiviral drugs. A brief classification of the main types of nanomedicines is given, and a detailed overview of the most common viral infections is provided. Given the large number of papers and reviews on this topic, it has not been possible to reference all the literature in the field, but some examples are reported which indicate how the formulation of antivirals as nanomedicines could represent a promising solution for the prevention and treatment of viral infections.

117 **2.** Nanodelivery systems for antiviral drugs

Various organic-based nanodelivery systems for antiviral agents, including liposomes, polymeric NPs, 118 119 solid lipid NPs, hybrid NPs, dendrimers, nanoemulsions, micellar systems and self-assembled 120 nanostructures, have been proposed [12]. To develop efficient nanocarrier platforms, various criteria 121 should be considered, including safety, biocompatibility, biodegradability and compatibility with the drug. It is worth noting that the main parameters that determine NP functionality are particle size, size 122 distribution, shape and surface characteristics (e.g., chemistry, charge) [13]. Additionally, NP features 123 124 should be fine-tuned to optimize the pharmacokinetics profile, in vivo biodistribution and viral interactions. Figure 2 reports an overview of the main drawbacks of antivirals and nanotechnological 125 strategies to overcome the limits of antiviral therapy. For example, nanocarriers for oral 126 127 nanoformulations should be resistant to the acidic pH of stomach and intestinal enzymes, and be able to penetrate the mucus secretion that limits the intestinal presence of these drugs. Drug release kinetics is 128 another important parameter that influences the therapeutic effectiveness of nanodelivery systems. An 129 130 ideal system should incorporate a good amount of drug and release its payload once it has arrived at the 131 site of action. Smart nanocarriers that are able to control and self-regulate drug delivery have been 132 investigated, particularly for cancer therapy and also for infectious diseases [14]. Finally, to enhance the effectiveness of the drug, it is necessary to take the route of administration into account. The 133 majority of antiviral drugs are administered orally, although some are delivered via intravenous or 134 135 subcutaneous routes (Fig. 3). With regard to the toxicity of organic NPs based on biocompatible components, many papers have reported no or slight toxicity related to the surface charge or solvent 136 residues [15]. The general characteristics of current nanoformulations for antivirals are described 137 below. 138

139

140 **2.1 Liposomes**

Liposomes are nanocarriers comprised of phospholipid bilayers encapsulating an aqueous core [12]. 141 These spherical lipid vesicles improve the solubility of compounds by compartmentalizing, 142 solubilizing, and delivering a wide range of hydrophilic and hydrophobic molecules. These unique 143 capabilities, coupled with their biocompatibility, biodegradability and low toxicity make liposomes 144 very attractive as drug delivery vehicles. They have consequently been widely investigated as one of 145 146 the most extensively used nanocarriers, and in 1995 the first liposomal product, consisting of doxorubicin-loaded pegylated liposomes was approved by the US Food and Drug Administration 147 148 (FDA) for cancer treatment. Liposomes range in size from 20 nm in the case of small unilamellar vesicles, to more than 100 nm for large and multilamellar vesicles. In basic research, the most common 149 150 preparation method is the dehydration-rehydration technique, which has relatively high loading efficiency compared with other methods, such as reverse-phase evaporation or detergent depletion, all 151 of which have lower encapsulation efficiencies. Liposomes have a natural tendency for hepatic 152 accumulation, and thus represent an optimal drug delivery system for the treatment of liver diseases, 153 154 including chronic hepatitis C virus infections [16]. However, as drug carriers, liposomes have certain drawbacks. Besides their low in vitro-in vivo stability, they have low encapsulation efficiency and have 155 a high production cost. In addition, their rapid removal from the blood and capture by cells of the 156 157 reticuloendothelial system restrict their therapeutic applications.

158

159 **2.2 Niosomes and ethosomes**

160 Niosomes are vesicular delivery systems that result from the self-assembly of hydrated surfactant 161 monomers. They may have some advantages over liposomes with respect to chemical stability, lower 162 cost of chemicals, and the large number of surfactant classes available for the design of this vesicular 163 system [17]. Niosomes can encapsulate both hydrophilic and lipophilic molecules, by entrapping

hydrophilic ones in vesicular aqueous core or adsorbed on the bilayer surfaces, while lipophilic drugs are encapsulated by their partitioning into the lipophilic domain of the bilayers. Because of their nonionic nature and high biodegradability, niosomes have shown excellent biocompatibility and low toxicity. However, segregation of non-ionic surfactants may cause toxicity [18].

Ethosomes are lipid multilamellar vesicles composed of phospholipids, ethanol and water. Because of 168 their ability to fuse with skin lipids, ethosomes deliver topical agents to the skin more efficiently than 169 170 liposomes. On the basis of their composition, they can be classified into classical ethosomes, binary ethosomes, and transethosomes. Classical ethosomes are composed of phospholipids, a high 171 172 concentration of ethanol up to 45% w/w, and water. They are a modification of classical liposomes 173 showing better skin permeation and stability profiles. Binary ethosomes were developed by adding another type of alcohol (i.e. propylene glycol and isopropyl alcohol) to the classical ethosomes. 174 Transethosomes are the new generation of ethosomal systems developed in order to combine the 175 advantages of classical ethosomes and deformable liposomes (transfersomes). They contain the basic 176 177 components of classical ethosomes and an additional compound, such as a penetration enhancer or an 178 edge activator (surfactant) in their formula [19].

179

180 2.3 Polymeric nanoparticles

Polymeric nanoparticles (NPs) were developed later than liposomes to improve their stability and drug payload. They are solid colloidal particles, generally below 500 nm in size, and are composed of a biocompatible polymeric matrix that can be made of synthetic or natural polymers [12]. The therapeutic molecule is either entrapped, adsorbed or covalently attached. Because of their polymeric composition, polymeric NPs may have greater stability than liposomes in biological fluids and under storage conditions. Polymeric NPs are prepared by several methods, including solvent evaporation, spontaneous emulsification, solvent diffusion and polymerization. The preparation conditions have important effects on the characteristics of the NPs obtained, such as size and release rate. NPs can be
loaded with lipophilic and hydrophilic drugs, and different chemical approaches have been proposed,
including covalent chemistry, hydrophobic interactions and entrapment.

The synthetic polymers most widely used are polyesters, such as polylactides (PLAs), and polylactide– polyglycolide copolymers, poly(lactic-co-glycolic acid) (PLGAs), polycaprolactones (PCLs) and polyacrylates (PCAs). The most common polymer used is PLGA, and several PLGA drug delivery systems have been approved by the FDA. Natural polymers, such as alginate and chitosan, have also been used, and they generally exhibit immunogenicity lower than that of synthetic ones.

196

197 **2.4 Polymeric micelles**

Polymeric micelles are colloidal structures of block copolymers, such as poly(butyl methacrylate) 198 (PBMA), polystyrene (PLS), PLA or poly(ethyleneoxide)-poly(propylene oxide) (PEO-PPO). The 199 200 polymers spontaneously form micellar structures above the critical micelle concentration and temperature, causing the polymer chains to become hydrophobic and resulting in aggregation [12]. On 201 202 the other hand, the polymers become insoluble in aqueous solution, and they act like inert materials below the aforementioned critical conditions. The hydrophobic fragments form the spherical inner core, 203 in which poorly-water soluble-drugs can be encapsulated, and hydrophilic fragments form the outer 204 205 shell. The outer hydrophilic shell can be functionalized with different moieties, such as folate, 206 monoclonal antibodies and monosaccharides (i.e. mannose, glucose, fructose), to achieve active targeting and/or pH/temperature responsive nanocarriers. They have raised particular interest as nano-207 sized drug delivery systems, not only because they provide increased solubility and stability of 208 hydrophobic drugs, but also due to their in vivo advantages versus the free drug, such as increased 209 210 circulation half-life, enhanced bioavailability and improved cellular uptake [20]. Polymeric micelles suitable for drug delivery have the appropriate charge density and polymer chain length, and the 211

212 particle diameter is usually within 5–100 nm. Size control depends on the chemical structure of the 213 polymers and is not dependent on the preparation process, which is an advantage that other types of NPs lack. Similar to other organic NPs, polymeric micelles have low toxicity because they disassemble 214 215 into single polymer chains that can be easily excreted and exhibit no toxicity. Micellar stability mainly depends on the copolymer self-aggregation tendency. Since polymeric micelles are dynamic systems, 216 they are liable to dissociate, especially upon administration when they are diluted to a concentration 217 218 below the CMC. Moreover, blood components can alter the kinetic stability of micelles and cause 219 dissociation [21].

220

221 **2.5 Solid lipid nanoparticles**

Solid lipid nanoparticles (SLNs) are made of lipids that are solid at body temperature, such as fatty 222 acids and triglycerides [17]. The method of preparation usually involves heating to liquefy the lipids. 223 Upon cooling, SLNs separate and can be readily dried. They are stabilized with emulsifiers and 224 co-emulsifiers, such as polysorbates, poloxamers, fatty acid co-esters, lecithin and bile salts. The 225 226 amount of stabilizer used must be enough to prevent aggregation without reducing drug uptake. The 227 proper selection of lipids and surfactants can affect the particle size, long-term stability during storage, drug loading and behaviors of release. SLNs are less toxic and easier to scale up than synthetic polymer 228 229 NPs. Furthermore, the state of the lipids depends upon the characteristics of the medium (e.g., temperature, pH), which allows for better controlled release, safety and efficacy. Common 230 disadvantages of SLNs are unpredictable gelation tendency and inherent low incorporation rates 231 resulting from the crystalline structure of the solid lipid [22]. 232

- 233
- 234
- 235

236 **2.6 Dendrimers**

237 Dendrimers are nanosized, radially symmetric molecules with homogenous, well-defined and monodisperse structures, which typically have a symmetric core, an inner shell and an outer shell. 238 These polymers are shaped like the head of a tree and have two unique characteristics, namely, a 239 globular structure and polyvalency, which is found in many naturally occurring systems. Tomalia et al. 240 reported the synthesis of the first family of dendrimers, known as poly(amido amine) (PAMAM), 241 242 which is one of the most successfully used dendrimers [23]. One of the dendrimer-based formulations was designed specifically with HIV and HSV antiviral activity. Vivagel[®], a carbomer gel containing a 243 dendrimer (SPL7013), has demonstrated efficacy against human immunodeficiency virus and herpes 244 245 simplex virus in *in vitro* and animal models. Phase I clinical trials have been conducted, and the results concerning its safety were generally favorable [24-25]. 246

247

248 2.7 Cyclodextrin derivatives

Cyclodextrins are nanometric biomaterials consisting of a-1,4-linked cyclic glucopyranose oligomers, 249 and they are synthesized by enzymatic action on hydrolyzed starch. The most common (so-called 250 "native") forms of cyclodextrins are α -, β -, and γ -cyclodextrin, consisting of six, seven or eight 251 252 glucopyranose units, respectively. They have a characteristic toroidal shape formed by linkages of 253 multiple glucose units, thereby creating a hydrophilic exterior and a hydrophobic cavity. This 254 distinctive structure can form inclusion complexes with compounds whose geometry and polarity are 255 compatible with that of their cavity. The production of drug/cyclodextrin complexes has become one of 256 the most extensively investigated approaches to improve the stability, solubility, dissolution rate and bioavailability of drugs. Indeed, cyclodextrins have been employed to overcome the main 257 258 biopharmaceutical drawbacks of antiviral agents [26]. Cyclodextrin-based nanosponges are hypercross-linked polymers that have cyclodextrin units as their building blocks, and they are characterizedby their marked capacity to encapsulate a great variety of substances [27].

261

262 **2.8 Nanoemulsions**

Nanoemulsions are biphasic systems consisting of oil nanodroplets in the order of 100 nm. A typical 263 264 nanoemulsion contains oil, water and an emulsifier. The addition of an emulsifier is critical for the 265 creation of small droplets as it decreases the interfacial tension (i.e., the surface energy per unit area) 266 between the oil and water phases of the emulsion [28]. Nanoemulsions are potential tools for improving 267 the oral bioavailability of poorly aqueous soluble drugs. Self-nanoemulsifying drug delivery systems 268 (SNEDDS) are anhydrous homogenous liquid mixtures consisting of oil, surfactant, drug and coemulsifier or solubilizer, which spontaneously form oil-in-water nanoemulsions of approximately 200 269 270 nm or less upon dilution with water under gentle stirring [29]. These systems offer a number of advantages including decreased dose and dosing frequency (due to improved bioavailability), high 271 drug-loading efficiency and high stability. However, questions regarding the physical and chemical 272 stability of drugs solubilized in lipid and excipients mixture have not yet been adequately addressed 273 [30]. Successful large-scale production of the SNEDDS exemplified for two HIV protease inhibitors 274 ritonavir (Norvir[®], Abbott Laboratories, Abbott Park, IL, USA) and saquinavir (Fortovase[®], Roche 275 276 Pharmaceuticals, Nutley, NJ, USA), has generated considerable interest.

277

278 2.9 Nanosuspensions

Nanosuspensions or nanocrystals are nanoformulations comprising 100% pure drug nanoparticles with sizes in the nanoscale range, generally stabilized by surfactants or polymers. These drug nanodelivery systems are derived from traditional pharmaceutical technology. Nanosuspensions are usually obtained in liquid media by bottom-up or top-down methods or a combination of both techniques. They have been designed to enhance the solubility, dissolution rate and bioavailability of drugs via various administration routes. Due to their small sizes, nanosuspensions can also be considered nanotechnology-based drug delivery systems for nanomedicine products. Many nanocrystal formulations have been developed to increase the bioavailability of drugs and to decrease adverse side effects [31]. Currently, 9 nanocrystal-based drug products are on the market.

288

289 2.10 Hydrogel based nanocarriers

Hydrogels are swellable polymer networks, cross linked together either by physical or chemical cross-290 291 linking, with high fluid absorption capacity. The use of these versatile, highly biocompatible structures 292 for drug delivery has been extensively studied. Some hydrogels exhibit a high drug loading efficiency and display prolonged drug release. Moreover, drug release from hydrogel nanoparticles can be 293 294 controlled physically or chemically. Hydrogels at the nanoscale have a large surface area for multivalent conjugations and subsequent surface modifications. The targeting capacity can be enhanced 295 by coupling targeting ligands to the nanoparticle surface [32]. Efforts are being made to functionalize 296 hydrogel-based nanoformulations for brain delivery to eradicate neuro-AIDS. Owing to their 297 biodegradability and biocompatibility, bio-polymeric nanogels have shown excellent potential as 298 targeted drug carriers for the central nervous system. Nanogels might be modified using hydrophobic 299 300 moieties or many such compounds with the ability to enhance the permeability of BBB by crossing the tight junctions between endothelial cells. Moreover, nanogel surface charge and hydrophobicity have a 301 great influence on plasma protein adsorption, affecting the uptake via transcytosis [33]. 302

Hydrogels have been also explored for intravaginal drug delivery to prevent sexual transmission of HIV and other vaginal infections, providing excellent convenience for frequent usage and prolonged drug release. They could be topically applied over the vulvovaginal area to achieve a high bioavailability at the targeted tissues and potentially reduce the side effects associated with oral

formulations [34]. Thermosensitive hydrogels provide a method for convenient administration, showing the ability to spread along the vaginal canal after injection and coat the vaginal epithelium by hydrogel formation. Furthermore, the formed hydrogel acts as a drug reservoir releasing the drug that would be absorbed by epithelia or directly penetrated into virus to prohibit virus propagation [35].

Nanolipogels are an emerging platform being investigated as an alternative to the more widely used liposomes or polymer nanoparticles. These dual structure nanoparticles have a distinct lipid bilayer encompassing a polymer core, and they have been synthesized by UV-induced gelation of a hydrogel network within liposomes using various processes. Nanolipogels have been used to incorporate physico-chemically diverse agents within the lipid bilayer and in the hydrogel core, showing high encapsulation efficiency and controlled release kinetics. Studies have demonstrated the biological utility of nanolipogels for use in HIV infection chemoprophylaxis as topical microbicides [36].

318

319 2.11 Stimuli responsive drug delivery systems

On-demand drug delivery is becoming feasible through the design of stimuli-responsive systems that recognize specific triggers and react in a dynamic way. Nanoscale stimuli-responsive systems are able to control drug release in response to specific stimuli, either exogenous (variations in temperature, magnetic field, ultrasound intensity, light or electric pulses) or endogenous (changes in pH, enzyme concentration or redox gradients). They can take advantage of specific microenvironmental changes associated with pathological situations such as neoplastic diseases, ischemia, inflammatory diseases or infections [37].

327 Smart nanoparticles with stimuli responsive release are proposed for delivery of anti-retroviral agents, 328 exploiting as triggers semen, enzymes, endosomal escape, temperature and magnetic field [38]. Novel 329 stimuli-responsive strategies are provided especially in the development of vaginal nanocarriers for the 330 prevention of sexually transmitted infections. Among them are Eudragit[®] S100-based vaginal delivery

systems for Tenofovir. The presence of Eudragit[®], a pH-sensitive methacrylic acid/methyl methacrylate 331 copolymer (soluble at pH 7.6 but insoluble at pH 4.5), enabled the release of drug depending on 332 environment pH changes as occurring upon intravaginal ejaculation and, therefore, increased the 333 release of Tenofovir from nanoparticles in simulated seminal fluid [39]. Furthermore, Huang et al. 334 developed pH-sensitive cellulose acetate phthalate (CAP) nanofibers to deliver tenofovir disoproxil 335 fumarate- or etravirine, and fibers dissolved quickly in the presence of human semen releasing the drug 336 337 payload [40]. Moreover, many enzymes in the semen, like hyaluronidase and prostate specific antigen, can act as triggers for the delivery of drugs on contact with semen. 338

339

340 **2.12 Toxicology aspects**

To date, the toxicity of nanomaterials delivering antiviral drugs has been studied predominantly in vitro 341 or to assess the component biocompatibility [41]. Generally, in vivo biodistribution studies of injected 342 nanoformulations have revealed high accumulation of drugs in various organs without investigating the 343 toxicity profile after prolonged administration. Furthermore, the effects of oral nanoformulations on gut 344 microbiota is also an important aspect to take into account when analyzing possible modulation on the 345 metabolic activity of enteric bacteria after treatment. Studies of immunogenicity, hypersensitivity 346 reactions or other adverse immune effects of orally or intravenously administered drug delivery 347 348 systems used to cure infections are poor and require more attention [42]. Increasing evidence suggests a potent immune modulatory role of nanoformulations, especially lipid nanoparticles and engineered 349 350 liposomes used as vaccine adjuvants, in order to boost antigen-specific immune responses [43-45]. In the past decade, nanoformulated vaccine candidates have been developed and tested in vivo against 351 viral infections, such as respiratory syncytial virus and HIV. Interestingly, nanoparticles efficiently 352 enter the lymphatic system, and through classic receptor-mediated endocytosis particles smaller than 353 150 nm are endocytosed by macrophages and dendritic cells and antigens are presented for following 354

activation of T cells. A different effect on the immune response has been observed according to the nanoparticle size: a stronger CD4 or CD8 T cell response has been promoted by a particle size major or minor than 50nm, respectively [46].

Cellular uptake and interaction of nanocarrier are affected by their composition, size, shape, and surface properties (area, porosity, charge, surface modifications, coating) [47,48]. The biological activity and therapeutic efficacy achieved by drug loaded in nanosystems depend on these different physico-chemical parameters. They are likely to modify biological responses, such as translocation across epithelia to other organs, induction of oxidative stress, binding to proteins and receptors, and localization in cellular organelles as mitochondria.

In regard to the safety of human exposure to nanomaterials, genotoxicity studies, including the Ames test (the Salmonella typhimurium reverse mutation assay), mouse lymphoma assay (*in vitro* mammalian assay) and chromosome aberration assay (in vivo rodent assay), are crucial. These studies have often revealed a high genotoxicity especially by inorganic nanoparticles [49].

Furthermore, nanomaterials should be tested on primary cells and cell lines to test their genotoxic potential to induce strand breaks and oxidized DNA lesions and to affect the cell cycle progression

370

371 3. Nanoformulations for approved drugs against specific human viral infections

372 **3.1 HIV infection**

The human immunodeficiency virus (HIV) is the cause of acquired immunodeficiency syndrome (AIDS) and represents a major public health issue and cause of mortality globally [50]. At present, there is no effective vaccine for HIV or a way to eradicate the established infection due to its peculiar pathogenesis: 1) HIV has a high mutation rate that promotes immune escape and antiviral resistance; 2) the HIV replication cycle requires integration of the viral genome (provirus) in cellular DNA; and 3) cellular HIV reservoirs (i.e., latently infected CD4+ memory T cells, macrophages and hematopoietic cells) and anatomical HIV reservoirs (i.e., mucosa, brain, lymph nodes) are created where low levels of
viral replication are maintained even in the presence of treatment [51].

As shown in Table 1, there are currently 29 approved antiretroviral drugs to treat the infection [52], 381 belonging to five different classes based on the phase of replication affected: 1) nucleoside/nucleotide 382 reverse transcriptase inhibitors (NRTIs) [53]; 2) non-nucleoside reverse transcriptase inhibitors 383 (NNRTIs) [54]; 3) integrase inhibitors [55]; 4) protease inhibitors (PIs) [56]; 5) fusion/entry inhibitors 384 385 [57]. To reduce the incidence of viral resistance and increase drug efficacy, the gold standard of care for treatment of HIV infection involves the use of a combination of at least three antiretroviral therapy 386 (ART) drugs belonging to different classes, which are administered orally at least once a day. Only one 387 388 anti-HIV drug, the oligopeptide Enfuvirtide, requires subcutaneous administration [58]. Despite the potent activity of antiretroviral products, many challenges remain in eradicating HIV from infected 389 cells due to the toxicity and poor bioavailability of approved anti-HIV drugs, drug-drug interactions, 390 achieving efficacious drug concentrations in viral reservoirs, poor patient compliance, development of 391 drug resistance and systemic side effects. To overcome these problems, different ART nanodelivery 392 approaches have been developed in the last several decades, and they have been described in recent 393 reviews [59-62]. 394

Here, we highlight the most promising nanotechnology strategies for treatment of HIV infection, taking 395 396 into account the key limitations of the drugs (see Table 2). It is worth noting that the majority of the 397 marketed antiretroviral drugs are administered orally. Consequently, the new oral nanodelivery systems 398 should be able to overcome the various challenges raised either by the physico-chemical properties of 399 the drug or by the conditions of the gastrointestinal tract [63]. To address poor water solubility 400 problems, nanocrystal formulations could be used [31]. The formulation rationale is that nanoparticles produce a huge increase of the surface area, resulting in a significant increase of the drug dissolution 401 rate, consequently improving the oral bioavailability. Moreover, nanocrystals have mucoadhesion 402

403 properties, thus prolonging the retention time in the intestinal mucosa. Drug nanosuspensions, 404 consisting of 100 % pure drug nanocrystals, can be easily translated into clinical studies because they can be obtained with low amounts of excipients and exploiting current pharmaceutical manufacturing 405 processes. This type of nanoformulation can be considered a tool useful for traditional and innovative 406 product production. Liquid nanocrystal nanoformulation can permit the administration of antiretrovirals 407 to pediatric patients and to patients with swallowing problems. Nanocrystals are also able to mask the 408 409 drug's unpleasant taste, favoring the patient's compliance considering that HIV medicinal products are for chronic use. 410

411 Nanocrystals as innovative nanodelivery systems can be used to obtain sustained release [31]. For 412 example, paliperidone nanocrystals is a long acting injectable formulation on the market. Recently, two 413 antiretroviral drug nanocrystals have been considered for clinical phases, one developed by Janssen (rilpivirine LA) [64], and the other prepared by ViiV/Healthcare (cabotegravir LA) [65. Both are long-414 acting formulations produced by the nanotechnology top-down process of wet bead milling. The 415 versatility of nanocrystals can permit the delivery of different therapeutics for a synergistic therapy. 416 Intriguingly, an efficient targeted combination treatment might be obtained by the co-delivery of 417 different drug nanocrystals. 418

An interesting nanoART formulation consisting of a nanocrystal mixture of atazanavir, efavirenz, and ritonavir was prepared by high-pressure homogenization, followed by *in vitro* characterization. The results demonstrated that physical characteristics such as particle size, surfactant coating, surface charge and shape affected cell uptake and antiretroviral efficacy [66]. In addition nanocrystals can be used as such or surface-modified. Nevirapine nanocrystals were developed and their surfaces were modified with albumin, polysaccharides or polyethylene glycol to enhance drug targeting potential after intravenous administration to rats [67].

Polymers, either natural or synthetic, are key components of nanodelivery systems. They can be used as solid matrices, self-assembled aggregates, or coating materials (as reported for nanocrystals), according to their chemical structure and preparation process of nanomedicines. The interest of these materials is mainly related to their ability to protect the encapsulated drugs and to control the release kinetics.

430 Polymer-based nanosystems, such as nanoparticles and micelles, represent a versatile platform for431 improving HIV infection therapy.

432 The use of surfactant micelles is a technological strategy commonly used to increase the drug 433 solubility. Interestingly, polymer micelles have a high solubilization capability and an increased 434 stability with respect to surfactant micelles, as recently demonstrated with anticancer drugs.

The encapsulation in polymer micelles can govern the drug release kinetics and protect the dug from
the external environment. The application of polymer micelles for the delivery of anti-HIV drugs was
widely investigated.

Chiappetta et al. [68] studied polymeric micelles of different poly(ethylene oxide)-poly(propylene 438 oxide) (PEO-PPOs) block copolymers for the treatment of pediatric patients which encapsulated 439 efavirenz with increased aqueous solubility up to 34 mg/mL (drug solubility = $4\mu g/mL$). Moreover, 440 good oral bioavailability was obtained after the administration of the nanoformulation to rats. The 441 highest bioavailability was obtained with smaller micellar size [68]. Subsequently, the oral 442 443 pharmacokinetics of efavirenz-loaded micelles was evaluated in adult healthy volunteers in comparison to suspension and oil solution of the drugs. The micellar system showed a 3-fold increase in the oral 444 445 bioavailability [69].

To expand the application of polymeric micelles to mucosal administration routes, Sosnik *et al.* [70] generated mucoadhesive thermo-responsive chitosan-g-poly(N-isopropylacrylamide) polymeric micelle-encapsulated indinavir, and a significant 24-fold increase in aqueous solubility was obtained

449 [70]. Other architectures and compositions are currently under investigation to improve the residence450 time of drug-loaded micelles.

Various types of nanoparticles encapsulating antivirals, have been studied using polymers approved by 451 452 the regulation agencies, such as PLGA and PCL. Polymer nanoparticles can improve the performance of antiviral drugs. In particular, they can provide the sustained release of the encapsulated drug, thus 453 decreasing the administration frequency – which is a feature important for chronic treatments. 454 455 Interestingly, efavirenz-loaded poly(epsilon-caprolactone) (PCL) nanoparticles have been obtained by a spray-drying method and compared with efavirenz-loaded micelles and pure drug nanocrystals. The 456 457 encapsulation within polymer nanoparticles significantly increased the maximum concentration in 458 plasma and the oral bioavailability [71]. PLGA-based NPs containing didanosine have been developed to study their uptake by macrophages in vitro, and sustained drug release was demonstrated for up to 60 459 days [72]. 460

The co-delivery of two or more drugs in the same polymer nanocarrier is an attractive strategy for 461 obtaining a combination therapy. Combinations of ART drugs from different classes have been proved 462 463 to offer sustained efficacy, durability and long-term safety. Destache et al. [73] developed PLGA NPs to simultaneously encapsulate ritonavir, lopinavir and efavirenz, and evaluated their phagocytosis into 464 monocyte-derived macrophages. The sustained release of drugs from the PLGA nanocarrier showed 465 466 high levels of antiretrovirals in cells until day 28 without cytotoxicity. Furthermore, PLGA NPs were tested for efficacy *in vivo* after intraperitoneal injection in mice. The sustained release of the drugs was 467 confirmed, as the nanoformulated antiretrovirals were detected in blood and organs up to 35 days after 468 administration [73]. 469

Another interesting approach is the development of lipid nanoformulations such as SNEDDS for
improving oral drug delivery. This type of formulation was designed to overcome the stability problems
of emulsions. They can be reconstituted in water just before the administration or administered filled

in capsules, being an anhydrous system able to form o/w emulsions in the gastrointestinal environment [30]. In this regard, Patel *et al.* developed SNEDDS consisting of Maisine 35-1 as oil for improvement of oral bioavailability of nelfinavir mesylate [74]. The *ex vivo* drug release through rat intestinal membrane was much faster from both liquid (reconstituted) and solid SNEDDS nanoformulations in comparison with the administration pure drug suspension, suggesting the enhancement in oral permeability. Some products based on this nanotechnology are on the market [63].

Moreover, other kinds of lipid-based drug carriers have been proposed to enhance the oral bioavailability of antiretrovirals. Tenofovir loaded-cationic phosphatidylcholine proliposomes have been formulated with different stearylamine levels to improve the drug permeability for oral delivery. Proliposomes with 5% and 15% stearylamine presented enhancements of permeability by 16.5-fold and 5.2-fold, respectively [75].

Liposomes, due to their fluid lipid bilayer structure, have been successful in improving oral bioavailability of drugs. They can better adhere to biomembranes, form mixed-micelle structures with bile salts in the gut to increase the solubility of poorly-soluble drugs and are suitable candidates for lymphatic uptake [76]. To increase low bioavailability of saquinavir and increase delivery to mammalian cells, a novel liposome formulation was prepared and compared to PEGylated liposomes. PEGylated liposomes showed more sustained drug release, which persisted for 50 hours, and they were less cytotoxic than non-PEGylated liposomes or free drug [77].

Various lipid based formulations have been studied to enhance the intestinal lymphatic uptake after oral administration as an alternative pathway to reach blood circulation. The lymphatic absorption via Peyer's patches can protect the drug from the epatic first-pass metabolism [78]. Previously, Cavalli et al. [79,80] demonstrated that SLNs administered intraduodenally to rats were mainly targeted to the lymph. This strategy was exploted also with antivirals. SLNs loaded with lopinavir, which has very poor oral bioavailability, were formulated to target intestinal lymphatic vessels. The lipid formulation

increased the cumulative percentage dose into the lymph of rats, as well as the drug's bioavailability, most likely by crossing the gut via endothelial intercellular gaps [81,82]. Gaur *et al.* [83] developed efavirenz-loaded SLNs to enhance the oral bioavailability of this poorly water-soluble drug. The nanoformulation exhibited a 5.32-fold increase in peak plasma concentration and a 10.98-fold increase in the area under the curve compared with the drug aqueous suspension after oral administration [83].

Notably, one of the significant challenges in treating HIV infection is the abolishment of cellular and anatomical reservoirs of HIV, where viruses can persist and drug concentrations are suboptimal [60,84]. Different delivery strategies have been explored to improve targeting on the critical sites of HIV infection such as macrophages, lymph nodes and the brain [60,85-87]. Macrophage uptake of drugs depends on the different physical characteristics of nanocarriers such as particle size, surfactant coating, surface charge and shape. Polymeric NPs have been the principal nanoformulation investigated for drug uptake in target cells of HIV-1 infection.

To optimize macrophage targeting specificity, suitable ligands can be conjugated to nanocarrier surfaces, such as small molecules that are able to bind to target cell receptors [62,88]. The mannose receptor of macrophages may also serve as drug delivery cellular portals for nanocarriers. Thus, mannosylated poly(ethylene glycol)-conjugate nanocarriers have recently been explored in cellular uptake studies [89].

Atazanavir-encapsulated galactosylated liposomes were prepared to target the lectin receptors present on macrophages. The nanoformulations resulted in increased uptake by alveolar macrophages and better drug distribution in the lymph nodes, liver, spleen and lungs compared with free atazanavir (AZT). In addition, no toxicity was observed after intravenous injection in rats, confirming that the galactosylated liposomes might be a potential system for targeted drug delivery [90].

519 One of the main limitations in using AZT lies in their poor intracellular activation by cellular kinases 520 into their active triphosphorylated forms. The administration of tri-phosphorylated molecule (AZT-TP)

521 may bypass the metabolic triphosphorylation by cellular kinases in cell cytoplasm, but these derivatives 522 do not diffuse intracellularly because of their too hydrophilic feature. Several nanoformulations directly encapsulating the tri-phosphorylated molecule have been studied. Hillaireau et al. [91,92] prepared 523 524 aqueous-cored poly(isobutylcyanoacrylate) nanocapsules as carriers to overcome the cellular delivery of AZT-TP. To prevent leakage of the drug through the nanocapsule membrane, due to its relatively low 525 molecular weight, AZT-TP was complexed to cationic polymers, such as chitosan or polyethylenimine. 526 527 Nanocapsules efficiently delivered AZT-TP in vitro to macrophages, reaching relevant cellular concentrations for therapeutic purposes [91,92]. 528

Furthermore, chitosan nanoparticles based on ionic interactions between chitosan and AZT-TP were developed by Giacalone *et al.* for macrophage targeting [93]. They showed very high drug loading (up to 44% w/w) and increased *in vitro* delivery in murine macrophages compared with the free molecule [93].

It is known that, even at effective plasma drug concentrations, insufficient drug exposure to lymphoid
tissue may be one of the key factors in the inability to completely eliminate residual virus.

A number of nanoformulations to enhance drug accumulation in the lymph nodes are currently being
evaluated [94]. Among them, an attractive approach for the sustained and targeted delivery of anti-HIV
drugs is the use of surface-modified liposomes.

Zidovudine-loaded liposomes have been engineered by incorporating charges (positive or negative
using stearylamine and dicetyl phosphate, respectively) or the site-specific ligand mannose to enhance
uptake and localization of the liposomes in the lymph nodes and spleen, and increasing quantities of the
drug were detected in these organs [95].

A lipid-drug nanoparticle containing lopinavir, ritonavir, and tenofovir was developed and administered as a single subcutaneous dose in primates, resulting in a 50-fold higher intracellular drug concentration in the lymph nodes compared with free drug. After 7 days, persistent levels of the drugs were detected

545 in the blood, and therefore an improvement on what is achieved using current oral therapies [96]. 546 Endsley *et al.* [97] designed novel peptide-coated lipid NPs for targeting IDV on CD4-expressing cells. Four candidate peptides showed marked binding specificity to CD4 and were used to anchorate 547 on lipid NPs to be targeted in the lymph nodes [97]. Terminal cysteine containing candidate peptides 548 were conjugated to lipid nanoparticles through maleimide-linked phopholipids for targeting to CD4 549 cells. Interestingly, two peptides bounded on lipid nanoparticles showed a CD4 selectivity in a peptide 550 551 dose dependent manner. The results proved the key role played by pepetide-tagged nanoparticles for targeting delivery. 552

Roy *et al.* [98] developed an efavirenz-loaded polymer-based Pluronic nanocarrier, which was bioconjugated with anti-M-cell-specific antibodies to target the microfold cells in the gut-associated lymphoid tissue. A significantly improved sustained release and increased anti-HIV activity of the targeted loaded drug were observed compared with the free drug [98].

557 However, the treatment of virus reservoirs in the central nervous system (CNS) remains a challenge.

The brain is one of the main HIV reservoirs due to the inability of antiretroviral drugs to reach effectiveconcentrations in the CNS.

Delivering therapeutics across blood-brain barrier (BBB) have been deeply investigated. Nanoparticle 560 chemical modification or physical triggers can be used to enhance drug penetration to CNS [99]. 561 562 Nanoparticle features can be tuned to favour the BBB passage, particularly exploiting their surface functionalization. The binding of specific ligands such as peptides, glutathione, transferrin and 563 564 transferrin antibody, lectins and lactic acid, to the nanoparticle surface have been extensively explored to achieve drug delivery to the brain [100]. Interestingly, to increase the brain targeting efficacy 565 nanoparticles can be functionalized with apolipoprotein E modified peptide responsible for low-density 566 lipoprotein receptor (LDL) binding, highly expressed in the brain [101]. 567

Different strategies have been explored to increase the penetration of ART across the BBB, as summarized by Nair *et al.* [102]. To overcome the limitations of CNS delivery, a valid approach is to eradicate the treatment of neuroAIDS, as reported in a number of recent review articles [102-106]. The design of specific sized and structured nanocargo able to cross the BBB and reach the HIV brain reservoir is one of the goals of nanomedicine. For these purposes nanodelivery systems triggered by external physical stimuli might have great potential for future treatments, i.e. magnetic fields or ultrasound [107].

575 Polymeric NPs, properly modified to obtain an effective passive or active permeability across BBB,576 have been designed for drug delivery to the CNS.

577 The delivery of stavudine, zidovudine, and lamivudine across an in vitro BBB model has been explored using both methylmethacrylate-sulfopropylmethacrylate (MMA-SPM) and poly(butyl cyanoacryalate) 578 (PBCA)-based NPs [108]. More recently, Kuo et al. [109] have investigated the ability of MMA-579 SPM nanoparticles with grafted RMP-7, a synthetic linear pseudopeptide agonist of bradykinin type II 580 (B2) receptor, to deliver stavudine, delavirdine, and saquinavir across the BBB in a co-culture model 581 582 containing human brain-microvascular endothelial cells and human astrocytes [109]. Tat-peptideconjugated ritonavir-loaded PLGA/PLA based NPs, formulated using an emulsion-solvent evaporation 583 technique, delivered drug to neural cells, both with and without Tat-peptide conjugation [110]. 584

The transferrin receptor is highly expressed in the brain; therefore transferrin-conjugated nanocarriers are internalized by endocytosis. Transferrin-grafted poly(lactide-co-glycolide) nanoparticles have been developed to deliver the nevirapine across the BBB into human brain microvascular endothelial cells [107]. Lamivudine loaded on PAMAM dendrimer, and mannose-capped poly(propyleneimine)-PPI dendrimers, exerted stronger anti-HIV activity than free drug solution, with a 21-fold increase in cellular uptake [112].

Liposomes have been extensively investigated for delivery of anti-HIV drugs to the brain [113]. The physico-chemical features of lipid systems allow them to cross the BBB easily, and their ability to reach the CNS can be further increased by surface engineering with BBB targeting moieties or cellpenetrating peptides (CPPs) [114].

Interestingly, a saquinavir-loaded flaxseed oil-based nanoemulsion was developed for brain delivery.
When orally administered to mice, it had the ability to cross the BBB, probably due to the small size of
the lipid-based nanocarrier [115].

Nasal administration as a means to deliver therapeutic agents preferentially to the brain has gained significant recent interest. Intranasal delivery is a non-invasive approach that allows direct access of the therapeutic substances to the brain, bypassing the BBB. Animal and human studies have shown that drugs can be transported directly from the nasal cavity to the CNS via the olfactory epithelium and/or the trigeminal nerve system [116]. A nanoemulsion was prepared for the intranasal saquinavir mesylate delivery to the brain. A higher *in vivo* concentration of saquinavir in the brain was detected after intranasal administration of the nanoemulsion compared with intravenous administration [117].

More recently, Chiappetta *et al.* developed PEO–PPO polymeric micelles loaded with efavirenz for targeting on the CNS. After intranasal administration in rats, an increased accumulation of micelles in the brain was detected, and a 5-fold relative exposure index (the ratio between the area under the curve in the CNS and the area under the curve in the plasma) was calculated compared to intravenous administration [118].

Finally, the use of magnetic nanoformulations allows targeting on specific anatomic sites by applying an external magnetic field and determination of site specific dosing through magnetic resonance imaging [119]. Magnetoliposomes loaded with 3'-azido-3'-deoxythymidine-5'-triphosphate (AZTTP) were developed for targeted delivery across the BBB and consisted of magnetic nanoformulations encapsulated into liposomes, to escape the reticuloendothelial systems. Magnetoliposomes, packed in

615 monocytes/macrophage, increased the *in vitro* BBB transmigration of the drug and did not affect the 616 BBB integrity. Furthermore, they maintained antiviral activity and resulted in a sustained AZTTP release for two weeks [120]. Jayant et al. [121] engineered magnetic nanoparticles for sustained release 617 of Tenofovir by a layer-by-layer assembly of dextran sulfate and drug, exploiting the interaction 618 619 between the negative sulphate group of dextran and positive charged drug. These magnetic 620 nanoformulations showed a stronger antiviral activity in infected astrocytes and greater ability to cross 621 the BBB compared with the free drug [121]. HIV latency-breaking agents such as vorinostat can be packaged into ultrasmall magnetic nanoparticles in conjunction with tenofovir in order to 622 simultaneously reactivate and kill HIV in a sustained manner for 5-7 days across the BBB. 623 624 Nanoformulation showed a good BBB transmigration ability with marked in vitro antiviral efficacy in primary human astrocytes, with good cell viability after HIV infection [122]. 625

There is a critical requirement to select an efficient and biocompatible nanocarrier, which can be externally guided for on-demand delivery of cargo across the BBB. Magneto-electro carriers (MENCs) of BaTiO3@CoF2O4 (BTO@CFO) were explored for on-demand controlled release of anti-HIV drugs as potential therapy against NeuroAIDS. After administration MENCs were uniformly distributed inside the brain, and were non-toxic to the brain and other major organs and did not affect hepatic, kidney and neurobehavioral functioning [123].

633 **3.2 Herpes virus infections**

Herpes viridae is a family comprising several human pathogens divided into three different subfamilies 634 $(\alpha, \beta \text{ and } \gamma)$: herpes simplex virus type 1 (HSV-1) and type 2 (HSV-2) and Varicella Zoster virus 635 (VZV) belong to the α -herpes virus subfamily; human cytomegalovirus (HCMV), human herpes virus 636 6 (HHV-6) and human herpes virus 7 (HHV-7) belong to the β -herpes virus subfamily; while Epstein-637 Barr virus (EBV) and Kaposi's sarcoma-associated herpes virus (KSHV) belong to the γ -herpes virus 638 639 subfamily. Following primary infection all these viruses establish life-long latency in different cell types and tissues and they are all able to reactivate at a later point in time, leading to asymptomatic or 640 641 symptomatic infections. The approved drugs shown in Table 3 treat the acute symptomatic infections 642 but they are unable to eradicate the latent infections. Various nanomedicines are currently under 643 investigation for the treatment of herpetic infections (see Table 4).

Acyclovir (ACV) is the drug of choice for treating HSV infections. However, due to its short half-life and incomplete absorption it must be taken in its oral dosage form five times daily (up to 1200 mg/day), and the dosage interval for intravenous formulations is 8 h.

Infections caused by HSV are incurable, so that the main goal of antiviral therapy with ACV is to inhibit the replication of the virus and thereby prevent associated epithelial damage [124]. Currently, the treatments available for herpes simplex are conventional tablets and topical gel for application on outbreaks. The drugs that are commonly used for herpes simplex are acyclovir, valaclovir and famciclovir.

ACV was the first antiviral to be licensed for the treatment of HSV infections, and it is the drug of choice for the treatment of epidermal, ocular or systemic herpetic infections, despite the fact that most of its currently available dosage forms, that is, tablets, suspension, cream, fail to achieve suitable levels at target sites following oral, local, or parenteral administration [125]. The main reason is that Acyclovir is a class III drug according to the Biopharmaceutics Classification System [126]: it is

slightly soluble in water, has a short plasma half-life, its absorption from gastrointestinal tract is slow and incomplete, and oral bioavailability ranges from only 10% to 30%. As a consequence, higher doses are prescribed, resulting in systemic toxicity and adverse reactions [126]. Also in local therapy, ACV bioavailability is low and highly variable, associated with low retention at the vaginal mucosa and poor patient compliance, and requires frequent administrations [127-129].

For these reasons, the development of efficient nanoformulations of ACV is of increasing importance. Moreover, the increased incidence of sexually transmitted diseases (STDs) raises concern about the limitations of currently available therapies. Among various STDs, the infections with human *Herpes simplex* virus type-1 and type-2 remain among the most common [130].

666 Over the course of the past decade, different types of nanoparticles based on cyclodextrin have been 667 developed to improve the physico-chemical features of ACV [131-133]. In general, cyclodextrin-based 668 formulations allow high loading and the sustained release of ACV; in some cases, they also improve its 669 *in vitro* antiviral efficacy.

Recently, Eudragit RLPO[®], a copolymer of ethyl acrylate, methyl methacrylate and a low content of
methacrylic acid ester with quaternary ammonium groups, has been selected for preparation of ACV
loaded nanoparticles [134]. *In vitro* studies show that this formulation provides sustained drug release
over a period of 24 h.

Amany and colleagues [135] prepared stealth PLGA nanoparticles loaded with ACV with the aim of extending the drug plasma half-life after intravenous administration [135]. In another study, PLGA surface tailored nanoparticles were prepared for targeted hepatic delivery of ACV with the purpose of minimizing side effects [136]. Galactose, actively taken up by asialoglycoprotein receptors (ASGP-R), was conjugated to PLGA nanoparticles, to exploit the exclusive presence of ASGP-R on hepatocytes for liver targeting.

In vitro, the galactosylated nanoparticles resulted in less haemolysis and successfully exhibited a sustained release pattern. *In vivo* studies showed an improved bioavailability, increased residence time, and enhanced delivery of ACV to the liver upon galactosylation [136].

Nano-niosomes have also been used to encapsulate ACV and were found to increase ACV effectivenessand decrease its side effects [137].

In regard to ACV vaginal therapy of HSV-2 sexually transmitted infections, high drug concentrations in 685 686 genital tissues are desirable. Unfortunately, the incomplete coverage and short duration of action limit the effectiveness of vaginally administered drugs, including ACV. The use of mucoadhesive polymers 687 688 such as poly(acrylic acid) derivatives, in the preparation of delivery systems, may overcome these 689 limitations. For instance, mucoadhesive ACV-containing liposomes coated with mucoadhesive polymers (Carbopol® or chitosan) have been developed [138]. In recent years, efforts have been made 690 to develop new types of mucoadhesive polymers that form a covalent bonding with the mucous 691 membrane, producing a stronger bio-adhesion compared to conventional polymeric mucoadhesive 692 materials. In this regard, Yandrapu et al. [139] developed new thiolated dendrimers for mucoadhesive 693 drug delivery. The thiolated dendrimers were synthesized by conjugating PAMAM dendrimer with 694 cysteamine and further encapsulated with ACV [139]. The thiolated dendrimers showed sustained 695 release of ACV in addition to a high mucoadhesion capability. 696

697 The dendrimer SPL7013 has been formulated in a mucoadhesive carbopol gel (VivaGel[®]) for use as a 698 topical microbicide in the prevention of HIV and HSV infections. Potent antiviral activity against HIV-699 1 and HSV-2 was observed following vaginal administration of VivaGel[®] in human [25].

An interesting strategy to enhance the accumulation of nanoparticles is the design of mucus penetrating systems. Ensign prepared mucus-penetrating particles (MPPs) for mucosal drug delivery in order to enhance the distribution and increase the retention in the vagina *in vivo* [140]. MPPs provided uniform

distribution over the vaginal epithelium in mice, whereas conventional nanoparticles were aggregatedby the animals' vaginal mucus, resulting in poor distribution.

Solid lipid nanoparticles can play an important role as ocular delivery system [141]. In 2013, Seyfoddin and colleagues [142] developed two lipid formulations (namely solid lipid nanoparticles and nanostructured lipid carriers) to improve the ocular bioavailability of ACV [142]. The high encapsulation efficiency, superior physical properties and good release profile obtained from nanostructured lipid carriers indicated that this formulation could be used as a potential ocular drug delivery system for ACV.

711 Interestingly, solid lipid nanoparticles and nanoemulsions were studied for enhancing dermal delivery 712 of ACV by Jain [143]. Mechanism of topical permeation and dermal distribution studies suggested that 713 pilosebaceous route was followed by solid lipid nanoparticles for skin penetration, without major 714 morphological changes on rat skin surface.

Another class of nanocarriers which has been investigated for the topical dermal drug delivery of ACV are nanoemulsions. Schwarz and colleagues [144] optimized several water/oil/water nanoemulsions for the dermal application of ACV by using the phase inversion temperature method [144].

718

Nanoformulations have been also proposed for the two derivatives of Acyclovir, i.e. penciclovir and
valaciclovir. For example, SLNs have been prepared and evaluated as nanocarriers for the topical
delivery of penciclovir to prolong the drug release [145].

Ganciclovir (GCV) is the first choice antiviral treatment for human cytomegalovirus infections. However, the low bioavailability and short half-life of GCV necessitate the development of a carrier. In a recent study, guanosine-based GCV was used to synthetize hydrophobic GCV-poly(caprolactone) (GCV-PCL), which was then grafted with hydrophilic chitosan to form amphiphilic copolymers for the preparation of stable micelles [146]. Solid lipid nanoparticles were also investigated as drug delivery

727 systems for GCV. Intriguingly, borneol can be used as penetration enhancer. Recent studies 728 demonstrated that borneol increased the permeation of drugs across BBB and enhanced their 729 distribution in the brain tissue [147]. Ren prepared borneol-modified and non-borneol ganciclovir-730 loaded SLNs and showed that borneol could enhance the transport of ganciclovir (GCV) incorporated 731 in solid lipid nanoparticles to the brain in mice after their intravenous administration [148].

GCV plays an important role also in the treatment of ocular HCMV infections. Akhter [149] used the reverse-phase evaporation technique to prepare GCV mucoadhesive nanoemulsions, chitosan nanoparticles, and mucoadhesive niosomal dispersions [149]. All three formulations were successfully shown to be non-irritant and non-toxic. These results may therefore be useful for the development of GCV formulations for the treatment of ocular infections.

Then, Akhter and colleagues [150] prepared nano-niosomes in order to improve the oral bioavailability of GCV. *In vitro* release studies confirmed a sustained release profile of the niosomal dispersions; whereas *in vivo* studies in rats revealed a five-fold increment in GCV bioavailability after the oral niosome administration compared with the standard tablet formulation [150].

Foscarnet is a pyrophosphate analog mainly used for the treatment of GCV-resistant HCMV infections in patients with AIDS or in transplant recipients. A novel nanoparticulate system for foscarnet delivery was developed by Russo *et al.* [151]. Nanoparticles were obtained by ionotropic gelation of chitosan induced by foscarnet itself, acting as an ionotropic agent. Crosslinked nanoparticles showed a controlled drug release, and foscarnet released from nanoparticles maintained the antiviral activity of the free drug when tested *in vitro* against HCMV.

747

748

749

750

751 3.3 Influenza virus infection

752 Seasonal orthomyxoviruses cause influenza in humans every year, affecting approximately 5% to 10% of the global adult population annually and associated with 250,000 to 500,000 deaths every year. To 753 date, three classes of anti-influenza drugs have been approved: inhibitors of the ion channel matrix 2 754 (M2) protein, known as adamantanes, inhibitors of neuraminidase, that are analogues of sialic acid, 755 756 blocking the release of progeny viruses from infected cells and, finally, a recent inhibitor of the viral 757 RNA polymerase (Table 5). Neuraminidase inhibitors, like oral Oseltamivir or inhaled Zanamivir, are the main drugs currently recommended for antiviral treatment. Zanamivir presents poor oral 758 bioavailability (approximately 2%), and it is therefore administered by inhalation. However, the 759 760 inhalation route is not recommended in patients with chronic respiratory conditions. Different 761 approaches to increasing the oral bioavailability of Zanamivir have been investigated.

Cao *et al.* prepared Zanamivir-loaded SLNs, which achieve relatively high entrapment efficiency, using a double emulsion solvent evaporation method [152]. Wang *et al.* developed dendrimer-cyclodextrin conjugates encapsulated with different guest molecules, including amantadine hydrochloride. The combination of PAMAM dendrimer and CDs can improve the drug loading ability of dendrimer and make amantadine hydrochloride suitable for dendrimer-based drug delivery systems. Indeed, they demonstrated that the drug is encapsulated within β -cyclodextrin cavity in the dendrimer conjugate [153].

769

770 **3.4 HBV infection**

Despite the availability of a prophylactic vaccine, Hepatitis B virus (HBV) causes 780,000 deaths every year and 240 million people were infected in 2014. The current approved therapy for chronic infection is based on immunomodulating peptides, such as interferon- α , and nucleos(t)ide analogues targeting the HBV polymerase. Orally administered polymerase inhibitors have certain drawbacks, including the poor oral bioavailability, as reported for Adefovir dipivoxil, and long-term treatment-induced viral resistance. Several drug delivery systems have been developed to improve the activity of the existing anti-HBV drugs (Table 5). While nanocarriers for interferon- α are reported in the paragraph relative to Hepatitis C virus, nanoformulations for the delivery of nucleos(t)ide analogues are described below.

To improve the bioavailability of adefovir dipivoxil, Dodiya and colleagues prepared and studied SLNs and nanosuspensions *in vivo* [154]. Both nanoformulations exhibited good bioavailability of 52% and 78%, for the nanosuspension and SLNs, respectively. Furthermore, higher accumulation of adefovir dipivoxil was detected in the liver and other organs after 24 hours after administration.

Du and colleagues [155] prepared nanoassemblies of an adefovir lipid derivative with cytochrome P450-triggered drug release with the purpose of improving hepatocyte targeting [155]. Cytochrome P450-induced cleavage thus provides a way to trigger the release of the active drug within hepatocytes. To ensure hepatocyte targeting, a galactose–ended polyoxyethylene fatty ether was added to assemblies. After its intravenous administration into infected mice, the resulting nanoassemblies showed long circulation times, liver targeting, and very high anti-HBV activity.

Stearic acid-g-chitosan oligosaccharide micelles, able to self-aggregate in aqueous medium, were prepared and loaded with Lamivudine stearate, the prodrug of Lamivudine. The resulting micelles were characterized by high entrapment efficiency, drug loading, and pH-sensitive drug release. Low cytotoxicity and high levels of antigen expression inhibition, and DNA replication inhibition were found compared with the free drug [156].

- 795
- 796
- 797
- 798

799 **3.5 HCV infection**

The approved treatment for chronic hepatitis C virus (HCV) infection is the combination of PEGylated 800 801 interferon- α (IFN), ribavirin and protease inhibitors, such as boceprevir, telaprevir or sofosbuvir (Table 5). A great deal of research has focused on the development of a carrier for ribavirin to improve its 802 therapeutic effect and safety [157]. Clinical use of ribavirin is often accompanied by severe side effects, 803 like hemolytic anemia, which occurs in approximately 50% of treated patients. Regarding liver 804 805 targeting of hepatitis treatments, nanotechnologies may provide important tools with which to promote the selective accumulation of the drug in the target cells; in turn, decreasing their toxic effects in other 806 tissues, improving drug efficacy and reducing administration frequency, with consequent increases in 807 808 patient compliance.

Craparo et al. developed polymer-based targeting micelles bearing galactose moieties on the surface for 809 the liver-targeted delivery of ribavirin [158-159]. Polymeric micelles were prepared starting with a 810 galactosylated polylactide-polyaminoacid conjugate (PHEA-EDA-PLA-GAL copolymer) able to self-811 assemble into micelles. In order to increase the entrapment within the system, ribavirin tripalmitate was 812 813 synthesized as a hydrophobic prodrug. No toxic effects on cell viability and good hemocompatibility were observed. The specificity of ribavirin tripalmitate-loaded PHEA-EDA-PLA-GAL micelles against 814 HepG2 was demonstrated *in vitro* by their ability to be internalized in HepG2 cells and the enhanced 815 816 accumulation of prodrug compared with non-galactosylated systems.

Biodegradable nanoparticles encapsulating ribavirin have also been developed with the aim of modulating the pharmacokinetics of the drug [160]. Ribavirin monophosphate-loaded nanoparticles were prepared from a mixture of poly(D,L-lactic acid) homopolymer and arabinogalactan (AG)–poly(llysine) conjugate. The stable retention of ribavirin by the nanoparticles within the systemic circulation can prevent accumulation of ribavirin in erythrocytes, and in turn reduce haemolytic anaemia. Ribavirin was shown to accumulate in the liver of mice following the intravenous administration of thedrug-loaded nanoparticles.

Hashim and colleagues [161] prepared ribavirin niosomes with the aim of investigating the influence of niosomal encapsulation upon the capacity to target the drug on the liver in rats. Niosomal formulations significantly increased the ribavirin liver concentration (6-fold) in comparison with ribavirin-free solution following intraperitoneal administration of a single dose of 30mg/kg.

Interferon- α (IFN α) has enormous potential as an anti-viral treatment for Hepatitis C, but its clinical success is hampered by its limited bioavailability, relatively narrow therapeutic index, and short halflife following parenteral administration. To achieve better tolerability, a PEGylated-IFN α formulation has been developed with better pharmacokinetics.

Liu and colleagues [162] investigated a method to obtain IFN α -loaded nanoparticles favourable for the preservation of IFN α -2b integrity and biological functionality [162]. The antiviral activity of the polysaccharide nanoparticles was shown to be highly preserved (above 97%) both *in vitro* and *in vivo*.

Li and co-workers encapsulated IFN- α into SLNs using the double emulsion solvent evaporation method. Antiviral assays demonstrated that SLNPs preserved the bioactivity of IFN- α after encapsulation; i.e. it maintained its antiviral activity [163].

838

839 4. Future challenges

Many strategies using nanomedicines have been proposed in order to improve the current treatment of viral infections. Although numerous efforts have been made in the field of nanotechnology-based drug delivery, and although pre-clinical and clinical results are encouraging, a number of problems remain that need to be addressed in the future (Figure 4). First, a crucial challenge for the clinical application of nanomedicines concerns their safety. Suitable long-term toxicity studies in animal models are necessary, especially in relation to their use in the treatment of chronic infections. Particular attention

846 should be paid to immune system compatibility, complement activation, and coagulation effect. The design of novel biocompatible and biodegradable nanomaterials might overcome some toxicity 847 848 drawbacks. However, some of the advantages associated with nanocarriers, such as the reduced drug 849 doses and the possible oral administration, may themselves limit the undesirable side effects. In addition, nanomedicine limitations can be related to poor drug loading capability and nanotoxicology 850 issues. In this regard, more research is needed to provide the robust preclinical data necessary to justify 851 852 clinical trials of biocompatible nanoformulations with good biodistribution profiles, sustained release dosages, and no significant side effects. Another challenge to address in the future is the design of 853 854 nanocarriers with high drug payload. High drug amounts within a nanosystem involve the use of less 855 amounts of excipients that may have safety problems. In addition, nanomedicine may be able to exploit 856 innovative administration routes, such as pulmonary and nasal, to overcome administration limitations. An important property of a nanocarrier should be its persistent stability for a sufficient length of time, 857 both *in vitro* and *in vivo*, to assure safety and its localization to the sites of interest and to minimize 858 toxicity of the liver and kidneys. In this regard, it is crucial to integrate nanotechnology with 859 physiological and pharmacological knowledge, and with an understanding of drug distribution and 860 metabolism in specific tissues. Pharmacokinetics studies play a key role. Stability should be also 861 862 assured during storage and administration.

Nanocarriers with modified drug release kinetics, such as sustained release systems, will be able to decrease the number of administrations required and doses. Currently, long-acting injectable delivery systems represent an important solution for problems caused by patient incompliance with infrequent dosing, thereby guaranteeing better long-term adherence especially in chronic diseases [164]. Another important challenge is related to the manufacturing of nanomedicines. Scaling-up their preparation method might not be easy and may prove to be expensive. Therefore, investments will need to be made in industrial processes in order to provide large batches of nanoproducts and render them cost-effective.

However, the potential for nanocarriers to reduce the required doses might compensate for the costincrease required for technological upgrades.

In addition, the regulatory aspects of nanomedicines should be taken into account to translate inindustrial products, suitable to clinical application.

Treatment of viral infections could be substantially improved in the future by the development of 874 "smart" delivery systems, such as stimuli sensitive nanocarriers. The design of triggered release 875 876 nanocarriers allows on-demand purchase of the drug. For this purpose, responsive polymers might be used as a key component, either in self-assembling copolymer aggregates or as a coating on the surface 877 of nanoparticles and nanocrystals, as reported previously. Their ability to undergo rapid structural 878 879 changes in their network in a triggered manner and to exhibit the desired drug release has been exploited to design smart nanosystems. The literature offers some initial evidence supporting the proof 880 of concept of such "smart" delivery systems. For example, Duan et al. [165] proposed pH-responsive 881 atazanavir or darunavir-loaded lipid nanoparticles as a long-acting targeted antiretroviral therapy [165]. 882 Additionally, purposely surface-modified nanocarriers might allow antiviral accumulation in infected 883 sites. Targeted delivery is a strategy to be exploited for addressing specific tissues, such as lymph 884 nodes. 885

Moreover, the future design of nanocarriers targeted on specific viral nanostructures is an important milestone, and it will be achieved by studying the mechanisms of interaction between viruses and nanoparticles. Deeper knowledge about the selective interactions can be exploited in strategies aimed at weakening the viral activity and in the therapy of infections. A possible challenge to targeting the delivery of drugs by nanomedicine is the possible induction of drug resistant mutations in not targeted anatomical sites where drugs reach suboptimal doses.

Nanodelivery of combinations of several drugs might be preferable, especially against viruses
characterized by a high degree of genetic variability, like HIV-1. Therefore, "combo" delivery systems
will be one of the future goals.

In the future, new antiviral drug candidates continue to be developed and subjected to in-depth 895 investigations. These candidates may be proteins, siRNA or antisense oligonucleotides; and purposely 896 tuned nanomedicines will be required for their protection and delivery. Nanoformulations are needed 897 898 for their administration. Although these studies are generally still in their early stages, encouraging results have been obtained. The development of microbicides against viral sexually transmitted 899 900 diseases has attracted much research [166]. In this context, RNA interference (RNAi) provides a highly 901 valid tool for downregulating the expression of viral and host mRNA targets required for viral infection and/or replication. The latest advances made in RNAi technology towards the treatment of HIV-1 902 infection have been recently summarized [167]. Currently, two RNAi products for HBV treatment from 903 Arbutus Biopharma are undergoing clinical trials: ARB-1467 (Phase II) and ARB-1598 (Phase I) [168]. 904 Finally, advanced nanomedicine will be considered. A novel approach now emerging is the cell-based 905 drug delivery system, which entails the use of cells as carriers of drug-encapsulated nanoformulations. 906 In particular, macrophages represent a proof of concept of this mechanism. They have been used as 907 cellular vehicles because of their ability to migrate to lymph nodes or travel across the BBB, ensuring 908 909 the sustained release of drugs over an extended period of time.

910

911 5. Conclusion

In this review the most common nanomedicine strategies for the delivery of approved antivirals have been described. Future nanomedicine design should address many challenges, such as high payload, target delivery, and toxicity issues, to overcome the limitations of current conventional formulations. A further understanding of the nanoparticle behavior either *in vitro* or *in vivo* might accelerate their

916	pharmaceutical development. A multidisciplinary approach of research with collaborative industrial
917	partners may permit the faster translation of nanoformulations to clinics and on the market for patients.

- 919
- 920
- 921

922 6. Expert Opinion

923 Considering the continual occurrence of viral infections, new drug delivery strategies are required to 924 treat and eradicate viral diseases. Nanomedicine formulations can play multiple roles in achieving this 925 goal by improving the biopharmaceutical properties of approved antiviral drugs. Moreover, 926 nanocarriers can enable the delivery of drugs to specific infection sites.

Moreover, it is the specific physico-chemical properties of nanocarriers that underlie their ability to overcome biological barriers or interact with virus nanostructures being in the same size range. In addition, nanoparticles with engineered surfaces can act as multifunctional systems able to interact with selective receptors able to inhibit viral infections.

Interestingly, the peculiar properties of nanomedicines, such as nanoscale size and surface charge, can 931 be exploited to improve the mucosal permeability of encapsulated drugs. Recently, numerous efforts 932 933 have been made in using nanotechnology to efficiently deliver and cross the mucosal barrier, mainly 934 thanks to the development of mucoadhesive and mucopenetrating nanoparticle systems [59,140,169]. 935 Mucoadhesive particles may protect encapsulated drugs from the low gastric pH and proteolytic 936 enzymes, improve the residence time in the gastrointestinal tract and, as a consequence, the bioavailability of orally administered drugs. In particular, polymer nanoparticles have been studied for 937 938 mucoadhesion, cell targeting and uptake, exploiting hydrophobic interactions, van der Waals interactions and polymer chain interpenetration with mucin. For these reasons, mucopenetrating 939

940 nanoparticles able to penetrate the adherent mucus layer and thus be retained longer seem more suitable 941 for oral and vaginal delivery than mucoadhesive nanosystems. However, it must be taken into account 942 that, during infection, the properties of mucus may be altered and nanoformulations could be 943 transported differently.

To overcome some limitations of antiviral drugs, the co-delivery of multimodal therapeutics for 944 "combo" therapy, previously investigated for cancer therapy, can be also proposed with nanomedicine. 945 946 Much research attention has been directed over the last few years to developing antiviral drug delivery systems for combinations of drugs or a minimum of at least two drugs. This issue is most relevant to 947 948 infections that require the simultaneous administration of multiple drugs to prevent the generation of 949 resistance and viral rebound. Instead of the co-encapsulation of several antiviral drugs in the same 950 nanoparticle, an alternative approach could consist of the administration of multiple single drug-loaded nanocarriers. The advantages of this approach are: better encapsulation efficiencies and drug loading 951 associated with the single drugs; greater ease in obtaining the correct ratio of drugs; and better patient 952 compliance. 953

Numerous efforts have also been made to optimize nanoformulations in order to improve their ability to cross the BBB by reducing particle size, by means of surface engineering and BBB transmigration studies. Nanomedicines can also provide the delivery proteins and biotech products, such as siRNA, by various administration routes, avoiding aggregation and chemical degradation, as well as stability during storage and administration [168].

959 Children constitute the most challenging population for infection treatment, such as HVI or RSV.
960 Interestingly, nanomedicine could be an useful platform for the development of effective therapeutic
961 systems and appropriate dose adjustment for children and special patients with swallowing problems.

962 The advantages offered by long-acting injectables over oral formulations include eliminating the need
963 for daily oral tablet administration and therefore first-pass metabolism, lower antiviral doses with

964 consequently reduced hepatic and renal toxicities, and prevention of resistance. Long-acting injectable products have been successfully applied in the treatment of infectious diseases, most notably chronic 965 HIV infection, promoting compliance and patient adherence and reducing the burden of regimens 966 requiring daily treatment. Some long-acting injectable nanosuspensions of nonnucleoside reverse 967 transcriptase inhibitors, and integrase inhibitors are currently in clinical study and are highly 968 anticipated medications for the prevention of HIV infection [170]. Pharmacokinetic studies in humans 969 970 have demonstrated sustained drug concentrations after intramuscular administration at 4 weekly and 8 weekly intervals of rilpivirine long-acting injectable nanosuspension [171]. Preliminary in vivo studies 971 972 have shown that intramuscular cabotegravir can prevent simian/HIV acquisition from rectal, vaginal, 973 and intravenous challenge. Currently, there are two ongoing Phase II studies assessing cabotegravir as a preexposure prophylaxis agent in humans [172]. Crystalline nanoparticle formulations for the co-974 975 administration of rilpivirine and cabotegravir have progressed into phase II clinical trials as long-acting intramuscular maintenance therapy [173]. Nanomedicine production is generally more complex than 976 traditional small molecule based drugs. It is worth noting that complex formulations present unique 977 978 chemistry, manufacturing and control (CMC) properties, in particular when changes are needed for manufacturing scale-up. The development of nanotherapeutic products should be optimized with the 979 Quality by Design (QbD) approach to identify critical process parameters and material attributes. For 980 981 several nanotechnologies, the scaling up of manufacturing production processes using current 982 pharmaceutical technologies can be used.

Finally, the possibility of reducing the required number of antiviral doses might compensate for theincreased costs associated with technological upgrades.

985 Based on these premises, nanomedicines have the potential to become a revolutionary solution for the 986 treatment of viral diseases, although some issues still need to be addressed and solved for the 987 successful translation of nanotech products onto the market.

989 Acknowledgments

- MD was funded by Compagnia di San Paolo 2014 (project code: Torino_call2014_L2_207), while DL
- and RC were funded by University of Turin (ex-60 %) funds.

References

994	1.	Bamrungsap S, Zhao Z, Chen T, et al. Nanotechnology in therapeutics: a focus on nanoparticles								
995		as a drug delivery system. Nanomedicine 2012;7(8):1253-1271.								
996	2.	Hamidi M, Azadi A, Rafiei P, et al. A pharmacokinetic overview of nanotechnology-based drug								
997		delivery systems: an ADME-oriented approach. Crit Rev Ther Drug Carrier Syst.								
998		2013;30(5):435-67.								
999	3.	Kingsley JD, Dou H, Morehead J, et al. Nanotechnology: A Focus on Nanoparticles as a Drug								
1000		Delivery System. J Neuroimmune Pharmacol. 2006;1:340–350.								
1001	4.	Lembo D, Cavalli R. Nanoparticulate delivery systems for antiviral drugs. Antivir. Chem.								
1002		Chemother. 2010;21:53–70.								
1003	5.	Kim PS, Read SW. Nanotechnology and HIV: potential applications for treatment and								
1004		prevention. Wiley Interdiscip Rev Nanomed Nanobiotechnol. 2010;2(6):693-702.								
1005	6.	Sultana S, Khan MR, Kumar M, et al. Nanoparticles-mediated drug delivery approaches for								
1006		cancer targeting: a review. J Drug Target. 2013;21(2):107-25.								
1007	7.	Jackman JA, Lee J, Cho NJ. Nanomedicine for Infectious Disease Applications: Innovation								
1008		towards Broad-Spectrum Treatment of Viral Infections. Small 2016; 12(9):1133-1139.								
1009	8.	Raman R, Tharakaraman, K, Sasisekharan V, et al. Glycan-protein interactions in viral								
1010		pathogenesis. Curr Opin Struct Biol. 2016;40:153-162.								
1011	9.	Hendricks GL, Weirich KL, Viswanathan K, et al. Sialylneolacto-N-tetraose c (LSTc)-bearing								
1012		liposomal decoys capture influenza A virus. J Biol Chem. 2013;288(12):8061-8073.								
1013	10	. Lembo D, Donalisio M, Laine C, et al. Auto-associative heparin nanoassemblies: a biomimetic								
1014		platform against the heparan sulfate-dependent viruses HSV-1, HSV-2, HPV-16 and RSV. Eur J								
1015		Pharm Biopharm. 2014;88(1):275-82								

1016	11. Szunerits S, Barras A, Khanal M, et al. Nanostructures for the Inhibition of Viral Infections.
1017	Molecules 2015;20:14051-14081.
1018	12. Zazo H, Colino CI, Lanao JM. Current applications of nanoparticles in infectious diseases. J.
1019	Control. Release 2016;224:86–102.
1020	13. Dufort S, Sancey L, Coll JL. Physico-chemical parameters that govern nanoparticles fate also
1021	dictate rules for their molecular evolution. Adv Drug Deliv Rev 2012;64:179–189.
1022	••This article describes the main parameters affecting nanoparticle functionality and therapeutic
1023	efficacy.
1024	14. Liu D, Yang F, Xiong F, et al. The Smart Drug Delivery System and Its Clinical Potential.
1025	Theranostics 2016;6(9):1306-1323.
1026	15. Li J, Mao H, Kawazoe N, et al. Insight into the interactions between nanoparticles and cells.
1027	Biomater. Sci. 2017;5:173-189.
1028	16. Ripoli M, Angelico R, Sacco P, et al. Phytoliposome-Based Silibinin Delivery System as a
1029	Promising Strategy to Prevent Hepatitis C Virus Infection. J. Biomed. Nanotech 2016;12.4:770-
1030	780.
1031	17. Mehendale R, Joshi M, Patravale VB. Nanomedicines for treatment of viral diseases. Crit. Rev.
1032	Ther. Drug Carrier Syst. 2013;30:1–49.
1033	18. Moghassemi S, Hadjizadeh A. Nano-niosomes as nanoscale drug delivery systems: an
1034	illustrated review. J Control Release. 2014;185:22-36.

- 1035 19. Abdulbaqi IM, Darwis Y, Khan NA, et al. Ethosomal nanocarriers: the impact of constituents
 and formulation techniques on ethosomal properties, in vivo studies, and clinical trials. Int J
 1037 Nanomedicine. 2016;11:2279-304.
- 20. Cagel M, Tesan FC, Bernabeu E, et al. Polymeric mixed micelles as nanomedicines:
 Achievements and perspectives. Eur J Pharm Biopharm. 2017.pii: S0939-6411(16)30694-4.

1040	21. Owen SC, Chan DPY, Shoichet MS. Polymeric micelle stability. Nano Today 2012;7:53-65.
1041	22. Naseri N, Valizadeh H, Zakeri-Milani P. Solid Lipid Nanoparticles and Nanostructured Lipid
1042	Carriers: Structure, Preparation and Application. Adv Pharm Bull. 2015;5(3):305–313.
1043	23. Kannan RM, Nance E, Kannan S et al. Emerging concepts in dendrimer-based nanomedicine:
1044	from design principles to clinical applications. J Intern Med. 2014;276(6):579-617.
1045	24. Vacas-Córdoba E, Maly M, De la mata FJ, et al. Antiviral mechanism of polyanionic
1046	carbosilane dendrimers against HIV-I. Int J Nanomed. 2016;11:1281-1294.
1047	25. Roy U, Rodríguez J, Barber P, et al. The potential of HIV-1 nanotherapeutics: from in vitro
1048	studies to clinical trials. Nanomedicine (Lond). 2015;10(24):3597-3609.
1049	•This paper summarizes several HIV-1 nanotherapeutics at the preclinical level and/or Phase I/II
1050	clinical trials.
1051	26. Imperiale JC, Sosnik AD. Cyclodextrin complexes for treatment improvement in infectious
1052	diseases. Nanomedicine (Lond). 2015;10(10):1621-41.
1053	27. Trotta F, Zanetti M, Cavalli R. Cyclodextrin-based nanosponges as drug carriers. Beilstein J
1054	Org Chem. 2012;8:2091-9.
1055	28. Helgeson ME. Colloidal behavior of nanoemulsions: Interactions, structure and rheology.
1056	Current Opinion in Colloid Interface Science 2016;25:39-50.
1057	29. Date AA, Desai N, Dixit R, et al. Self-nanoemulsifying drug delivery systems: formulation
1058	insights, applications and advances. Nanomedicine 2010;5(10):1595-1616.
1059	30. Khan AW, Kotta S, Ansari SH, et al. Potentials and challenges in self-nanoemulsifying drug
1060	delivery systems. Exp. Opin. Drug Del. 2012;9:10:1305-1317.
1061	31. Leone F, Cavalli R. Drug nanosuspensions: a ZIP tool between traditional and innovative
1062	pharmaceutical formulations. Expert Opin Drug Deliv. 2015;12(10):1607-25.

- 32. Lai WF, He ZD. Design and fabrication of hydrogel-based nanoparticulate systems for in vivo
 drug delivery. J. Control. Release. 2016;243:269-282.
- 1065 33. Vashist A, Kaushik A, Vashist A, et al. Recent trends on hydrogel based drug delivery systems
 1066 for infectious diseases. Biomaterials Science. 2016;4(11):1535-1553.
- 1067 34. Lee AL, Ng VW, Poon GL, et al. Co-Delivery of Antiviral and Antifungal Therapeutics for the
 1068 Treatment of Sexually Transmitted Infections using a Moldable, Supramolecular Hydrogel.
 1069 Adv. Healthcare Mater. 2015;4(3):385–394.
- 1070 35. Li N, Yu M, Deng L. et al. Thermosensitive hydrogel of hydrophobically-modified
 1071 methylcellulose for intravaginal drug delivery. J Mater Sci: Mater Med 2012;23:1913–1919.
- 1072 36. Ramanathan R, Jiang Y, Read B, et al. Biophysical characterization of small molecule antiviral 1073 loaded nanolipogels for HIV-1 chemoprophylaxis and topical mucosal application Acta
 1074 Biomaterialia 2016;36:122–131.
- 1075 37. Mura S, Nicolas J, Couvreur P. Stimuli-responsive nanocarriers for drug delivery. Nature
 1076 materials. 2013;12(11):991-1003.
- ••Interesting article describing the recent advances in the design of nanoscale stimuli-responsive
 systems
- 1079 38. Adhikary RR, More P, Banerjee R. Smart nanoparticles as targeting platforms for HIV
 1080 infections. Nanoscale. 2015;7(17):7520-34.
- 39. Yoo JW, Giri N, Lee CH. pH-sensitive Eudragit nanoparticles for mucosal drug delivery. Int. J.
 Pharm. 2011;403:262–267.
- 40. Huang C, Soenen SJ, van Gulck E, et al. Electrospun cellulose acetate phthalate fibers for
 semen induced anti-HIV vaginal drug delivery. Biomaterials 2012;33:962–969.
- 41. Nyström AM, Fadeel B. Safety assessment of nanomaterials: Implications for nanomedicine. J.
 Control. Release 2012;161:403–408.

- **••**This article discusses some of the major challenges related to safety of nanomedicines.
- 1088 42. Örfi E, Szebeni J. The immune system of the gut and potential adverse effects of oral
 1089 nanocarriers on its function. Adv Drug Deliv Rev. 2016;106(Pt B):402-409.
- 43. Faulkner L, Buchan G, Slobbe L, et al. Influenza hemagglutinin peptides fused to interferon
 gamma and encapsulated in liposomes protects mice against influenza infection. Vaccine.
 2003;21(9-10):932-939.
- 44. Gao FS, Feng L, Zhang Q, et al. Immunogenicity of two FMDV nonameric peptides
 encapsulated in liposomes in mice and the protective efficacy in guinea pigs. PLoS One.
 2013;8(7):e68658.
- 45. Thoryk EA, Swaminathan G, Meschino S, et al. Co-Administration of Lipid Nanoparticles and
 Sub-Unit Vaccine Antigens Is Required for Increase in Antigen-Specific Immune Responses in
 Mice. Vaccines (Basel). 2016;4(4).
- 46. Jorquera PA, Tripp RA. Synthetic Biodegradable Microparticle and Nanoparticle Vaccines
 against the Respiratory Syncytial Virus. Vaccines (Basel). 2016;4(4):45-48.
- 47. Kumari A, Yadav SK. Cellular interactions of therapeutically delivered nanoparticles. Exp.
 Opin. Drug Deliv. 2011;8(2):141-151.
- 48. Yameen B, Choi W, Vilos C. Insight into nanoparticle cellular uptake and intracellular targeting.
 J. Control Release 2014;190:485–499.
- 49. Guo X, Li Y, Yan J, et al. Size- and coating-dependent cytotoxicity and genotoxicity of silver
 nanoparticles evaluated using in vitro standard assays. Nanotoxicology. 2016;10(9):1373-84.
- 50. Fauci AS, Folkers GK, Dieffenbach CW. HIV-AIDS: Much accomplished, much to do. Nat.
 Immunol. 2013;14(11):1104–1107.
- 1109 51. Sarmati L, D'Ettorre G, Parisi SG, et al. HIV replication at low copy number and its correlation
 1110 with HIV reservoir: a clinical perspective. Curr. HIV Res. 2015;13(3):250–257.

- 52. De Clercq E, Li G. Approved Antiviral Drugs over the Past 50 Years. Clin Microbiol Rev. 2016;
 29(3):695-747.
- 53. Das K, Arnold E. HIV-1 reverse transcriptase and antiviral drug resistance. Part 1. Curr. Opin.
 Virol. 2013;3:111–118.
- 54. Usach V, Melis J, Peris E. Non-nucleoside reverse transcriptase inhibitors: a re-view on
 pharmacokinetics, pharmacodynamics, safety and tolerability. J. Int. AIDS Soc. 2013;16 (Art.
 1117 18567).
- 1118 55. Métifiot M, Marchand C, Pommier Y. HIV integrase inhibitors: 20-year landmark and
 1119 challenges. Adv. Pharmacol. 2013;67:75–105.
- 56. Lv Z, Chu Y, Wang Y. HIV protease inhibitors: a review of molecular selectivity and toxicity.
 HIV AIDS (Auckl) 2015;7:95–104.
- 57. Bai Y, Xue H, Wang K, et al. Covalent fusion inhibitors targeting HIV-1 gp41 deep pocket.
 Amino Acids 2013;44:701–713.
- 58. Mcgowan I. An overview of antiretroviral pre-exposure prophylaxis of HIV infection. Am. J.
 Reprod. Immunol. 2014;71(6):624–630.
- 59. Nelson AG, Zhang X, Ganapathi U, et al. Drug delivery strategies and systems for HIV/AIDS
 pre-exposure prophylaxis and treatment. J Control Release 2015;219:669-80.
- 60. Giacalone G, Hillaireau H, Fattal E. Improving bioavailability and biodistribution of anti-HIV
 chemotherapy. Eur J Pharm Sci. 2015;75:40-53.
- 61. Kumar L, Verma S, Prasad DN, et al. Nanotechnology: a magic bullet for HIV AIDS treatment.
 Artif Cells Nanomed Biotechnol. 2015;43(2):71-86.
- 62. Ramana LN, Anand AR, Sethuraman S, et al. Targeting strategies for delivery of anti-HIV
 drugs. J Control Release. 2014;192:271-283.

1134	63. Sosnik A, Augustine R. Challenges in oral drug delivery of antiretrovirals and the innovative
1135	strategies to overcome them. Adv Drug Deliv Rev. 2016;103:105-120.
1136	••An interesting review on oral drug delivery of antiretrovirals.
1137	
1138	64. van 't Klooster G, Hoeben E, Borghys H, et al. Pharmacokinetics and disposition of rilpivirine
1139	(TMC278) nanosuspension as a long-acting injectable antiretroviral formulation. Antimicrob.
1140	Agents Chemother. 2010;54:2042–2050
1141	65. Andrews CD, Heneine W. Cabotegravir long-acting for HIV-1 prevention. Curr. Opin. HIV
1142	AIDS 2015;10:258–263.
1143	66. Nowacek AS, Balkundi S, McMillan J, et al. Analyses of nanoformulated antiretroviral drug
1144	charge, size, shape and content for uptake, drug release and antiviral activities in human

1145 monocyte-derived macrophages. J Control Release. 2011;150(2):204-11.

- 67. Shegokar R, Singh KK. Nevirapine nanosuspensions for HIV reservoir targeting. Pharmazie.
 2011;66(6):408-415.
- 68. Chiappetta DA, Hocht C, Taira C. Efavirenz-loaded polymeric micelles for pediatric anti-HIV
 pharmacotherapy with significantly higher oral bioavailability. Nanomedicine (London)
 2010;5:11–23.
- 69. Chiappetta DA, Hocht C, Taira C, et al. Oral pharmacokinetics of the anti-HIV efavirenz
 encapsulated within polymeric micelles. Biomaterials 2011;32:2379–2387.

1153 70. Sosnik A, Imperiale JC, Vazquez-Gonzalez B, et al. Mucoadhesive thermo-responsive chitosan-

- g-poly(NIPAAm) polymeric micelles via a one-pot gamma radiation-assisted pathway. Colloids
 Surf. B: Biointerfaces 2015;136:900–907.
- 1156 71. Tshweu L, Katata L, Kalombo L, et al. Enhanced oral bioavailability of the antiretroviral
 1157 efavirenz nano-encapsulated in poly(epsilon-caprolactone) nanoparticles by a spray-drying
 1158 method. Nanomedicine (London) 2014;9:1821–1833.

1159	72. Pattnaik G, Sinha B, Mukherjee B, et al. Submicron-size biodegradable polymer-based
1160	didanosine particles for treating HIV at early stage: an in vitro study. J. Microencapsul.
1161	2012;29(7):666–676.

- 1162 73. Destache CJ, Belgum T, Goede M, et al. Antiretroviral release from poly(DL-lactide-co1163 glycolide) nanoparticles in mice. J. Antimicrob. Chemother. 2010;65(10):2183–2187.
- 74. Patel A, Shelat P, Lalwani A. Development and optimization of solid self-nanoemulsifying drug
 delivery system (S-SNEDDS) using Scheffe's design for improvement of oral bioavailability of
 nelfinavir mesylate. Drug Deliv. Transl. Res. 2014;4:171–186.
- 75. Zidan AS, Spinks CB, Habib MJ, et al. Formulation and transport properties of tenofovir loaded
 liposomes through Caco-2 cell model. J Liposome Res. 2013;23(4):318-26.
- 76. Daeihamed M, Dadashzadeh S, Haeri A, et al. Potential of Liposomes for Enhancement of Oral
 Drug Absorption. Curr Drug Deliv. 2017;14(2):289-303.
- 1171 77. Ramana LN, Sharma S, Sethuraman S, et al. Investigation on the stability of saquinavir loaded
 1172 liposomes: implication on stealth, release characteristics and cytotoxicity. Int J Pharm.
 1173 2012;431(1-2):120-129
- 1174 78. Porter CJ, Pouton CW, Cuine JF, et al. Enhancing intestinal drug solubilisation using lipid1175 based delivery systems. Adv Drug Deliv Rev. 2008;60(6):673-691.
- 1176 79. Bargoni A, Cavalli R, Caputo O. Solid lipid nanoparticles in lymph and plasma after duodenal
 1177 administration to rats. Pharm. Res. 1998;15(5):745-750.
- 80. Cavalli R, Bargoni A, Podio V, et al. Duodenal administration of solid lipid nanoparticles
 loaded with different percentages of tobramycin. J Pharm Sci. 2003;92(5):1085-94.
- 1180 81. Aji AMR, Chacko AJ, Jose S, et al. Lopinavir loaded solid lipid nanoparticles (SLN) for
 1181 intestinal lymphatic targeting. Eur. J. Pharm. Sci. 2011;42(1–2):11–18.

1182	82. Negi JS, Chattopadhyay P, Sharma AK, et al. Development of solid lipid nanoparticles (SLNs)
1183	of lopinavir using hot self nano-emulsification (SNE) technique. Eur. J. Pharm. Sci. 2013;48 (1-
1184	2):231–239.

- 1185 83. Gaur PK, Mishra S, Bajpai M, et al. Enhanced oral bioavailability of efavirenz by solid lipid
 1186 nanoparticles: in vitro drug release and pharmacokinetics studies. Biomed Res Int.
 1187 2014;2014:363404.
- 1188 84. Cory TJ, Schacker TW, Stevenson M, et al. Overcoming pharmacologic sanctuaries. Curr. Opin.
 1189 HIV AIDS 2013;8(3):190–195.
- 1190 85. Gupta U, Jain NK. Non-polymeric nano-carriers in HIV/AIDS drug delivery and targeting. Adv.
 1191 Drug Deliv. Rev. 2010;62(4-5):478–490.
- 1192 86. Mamo T, Moseman EA, Kolishetti N, et al. Emerging nanotechnology approaches for
 1193 HIV/AIDS treatment and prevention. Nanomedicine (Lond.) 2010;5(2):269–285.
- 1194 87. Parboosing R, Maguire GE, Govender P, et al. Nanotechnology and the treatment of HIV
 1195 infection. Viruses 2012;4(4):488–520.
- 88. Gunaseelan S, Gunaseelan K, Deshmukh M, et al. Surface modifications of nanocarriers for
 effective intracellular delivery of anti-HIV drugs. Adv. Drug Deliv. Rev. 2010;62(4–5):518–
 531.
- 1199 89. Chen P, Zhang X, Jia L, et al. Optimal structural design of mannosylated nanocarriers for
 1200 macrophage targeting. J Control Release. 2014;194:341-9.
- 90. Garg M, Jain NK. Reduced hematopoietic toxicity, enhanced cellular uptake and altered
 pharmacokinetics of azidothymidine loaded galactosylated liposomes. J. Drug Target. 2006;14
 (1):1–11.

1204	91. Hillaireau H, Le Doan T, Appel M, et al. Hybrid polymer nanocapsules enhance in vitro
1205	delivery of azidothymidine-triphosphate to macrophages. J. Control. Release 2006;116(3):346-
1206	352.
1207	92. Hillaireau H, Le Doan T, Chacun H, et al. Encapsulation of mono- and oligo-nucleotides into
1208	aqueous-core nanocapsules in presence of various water-soluble polymers. Int. J. Pharm.
1209	2007;331(2):148–152.
1210	93. Giacalone G, Bochot A, Fattal E, et al. Drug-induced nanocarrier assembly as a strategy for the
1211	cellular delivery of nucleotides and nucleotide analogues. Biomacromolecules 2013;14(3):737-
1212	742.
1213	94. Shao J, Kraft JC, Li B, et al. Nanodrug formulations to enhance HIV drug exposure in
1214	lymphoid tissues and cells: clinical significance and potential impact on treatment and
1215	eradication of HIV/AIDS. Nanomedicine (Lond). 2016;11(5):545-64.
1216	95. Kaur CD, Nahar M, Jain NK. Lymphatic targeting of zidovudine using surface-engineered
1217	liposomes. J Drug Target. 2008;16(10):798-805.
1218	96. Freeling JP, Koehn J, Shu C, et al. Long-acting three-drug combination anti-HIV nanoparticles
1219	enhance drug exposure in primate plasma and cells within lymph nodes and blood. AIDS.
1220	2015;29(13):1727.
1221	97. Endsley AN, Ho RJY. Design and characterization of novel peptide-coated lipid nanoparticles
1222	for targeting anti-HIV drug to CD4 expressing cells. AAPS J. 2012;14(2):225–235.
1223	98. Roy U, Ding H, Pilakka-Kanthikeel S, et al. Preparation and characterization of anti-HIV
1224	nanodrug targeted to microfold cell of gut-associated lymphoid tissue. Int J Nanomedicine.
1225	2015;10:5819-5835.
1226	99. Guiot C, Zullino S, Priano L, et al. The physics of drug-delivery across the blood-brain barrier.
1227	Therapeutic Delivery 2016;7(3):153-156.
	54

1228	100.	Jain A, Jain SK. Ligand-appended BBB-targeted nanocarriers (LABTNs). Critical
1229	Review	ws [™] in Therapeutic Drug Carrier Systems 2015;32(2):149-180.
1230	101.	Portioli C, Bovi M, Benati D, et al. Novel functionalization strategies of polymeric
1231	nanopa	articles as carriers for brain medications. J Biomed Mater Res Part A. 2017;105:847-858.
1232	102.	Nair M, Jayant RD, Kaushik A, et al. Getting into the brain: Potential of nanotechnology
1233	in the	management of NeuroAIDS. Adv Drug Deliv Rev. 2016;103:202-217.
1234	•This a	article highlights the potential of nanocarriers in enhancing drug delivery to CNS.
1235	103.	Das MK, Sarma A, Chakraborty T. Nano-ART and NeuroAIDS. Drug Deliv Transl Res.
1236	2016;6	5(5):452-472.
1237	104.	Gomes MJ, Neves Jd, Sarmento B. Nanoparticle-based drug delivery to improve the
1238	efficac	ey of antiretroviral therapy in the central nervous system. Int J Nanomedicine
1239	2014;9	9:1757-69.
1240	105.	Rao KS, Ghorpade A, Labhasetwar V. Targeting anti-HIV drugs to the CNS. Expert
1241	Opin.	Drug Deliv. 2009;6(8):771–784.
1242	106.	Wong HL, Chattopadhyay N, Wu XY, et al. Nanotechnology applications for improved
1243	delive	ry of antiretroviral drugs to the brain. Adv. Drug Deliv. Rev. 2010;62(4-5):503–517.
1244	107.	Lakshmanan S, Gupta GK, Avci P, et al. Physical energy for drug delivery; poration,
1245	concer	ntration and activation. Adv Drug Deliv Rev. 2014;71:98-114.
1246	108.	Kuo YC, Chen HH. Effect of nanoparticulate polybutylcyanoacrylate and
1247	methy	Imethacrylate-sulfopropylmethacrylate on the permeability of zidovudine and lamivudine
1248	across	the in vitro blood-brain barrier. Int. J. Pharm. 2006;327:160-169.
1249	109.	Kuo YC, Lee CL. Methylmethacrylate-sulfopropylmethacrylate nanoparticles with
1250	surface	e RMP-7 for targeting delivery of antiretroviral drugs across the blood-brain barrier.
1251	Colloi	ds Surf. B: Biointerfaces 2012;90:75–82.

- 1252 110. Borgmann K, Rao KS, Labhasetwar V, et al. Efficacy of Tat-conjugated ritonavir-loaded
 1253 nanoparticles in reducing HIV-1 replication in monocyte-derived macrophages and
 1254 cytocompatibility with macrophages and human neurons. AIDS Res. Hum. Retrovir.
 1255 2011;27:853–862.
- 111. Kuo YC, Lin PI, Wang CC. Targeting nevirapine delivery across human brain
 microvascular endothelial cells using transferrin-grafted poly (lactide-co-glycolide)
 nanoparticles. Nanomedicine 2011;6:1011–1026.
- Huang RQ, Qu YH, Ke WL, et al. Efficient gene delivery targeted to the brain using a
 transferrin-conjugated polyethyleneglycol-modified polyamidoamine dendrimer. FASEB J.
 2007;21:1117–1125.
- 1262 113. Iannazzo D, Pistone A, Romeo R, et al. Nanotechnology approaches for antiretroviral
 1263 drugs delivery. J. AIDS HIV Infect. 2015;1:1–13.
- 1264 114. Fiandra L, Capetti A, Sorrentino L, et al. Nanoformulated Antiretrovirals for Penetration
 1265 of the Central Nervous System: State of the Art. J Neuroimmune Pharmacol. 2016:1-14.
- 1266 115. Vyas TK, Shahiwala A, Amiji MM. Improved oral bioavailability and brain transport of
 1267 saquinavir upon administration in novel nanoemulsion formulations. Int. J. Pharm.
 1268 2008;347:93–101.
- 1269 116. Md S, Mustafa G, Baboota S, et al. Nanoneurotherapeutics approach intended for direct
 1270 nose to brain delivery. Drug Dev Ind Pharm. 2015;41(12):1922-34.
- 1271 117. Mahajan HS, Mahajan MS, Nerkar PP, et al. Nanoemulsion-based intranasal drug
 1272 delivery system of saquinavir mesylate for brain targeting. Drug Deliv. 2014;21(2):148–154.
- 1273 118. Chiappetta DA, Hocht C, Opezzo JAW, et al. Intranasal administration of antiretroviral1274 loaded micelles for anatomical targeting to the brain in HIV. Nanomedicine 2013;8(2):223–237.

1275	119.	Sagar	V, At	tluri V	S, F	Pilakka-	Kanthikee	S,	et	al.	Magnetic	nanoth	erapeutics	for
1276	dysre	egulated s	synapti	ic plast	icity	during	neuroAID	S an	d dr	ug	abuse. Mo	l Brain.	2016;9(1)):57-
1277	66.													

- 1278 120. Saiyed ZM, Gandhi NH, Nair MP. Magnetic nanoformulation of azidothymidine 5'1279 triphosphate for targeted delivery across the blood-brain barrier. Int. J. Nanomedicine
 1280 2010;5:157-166.
- 1281 121. Jayant R. Layer-by-Layer (LbL) Assembly of Anti HIV Drug for Sustained Release to
 Brain Using Magnetic Nanoparticle. J. of Neuroimmune Pharmacol. 2014;9(1):25-25.
- 1283 122. Jayant RD, Atluri VS, Agudelo M, et al. Sustained-release nanoART formulation for the
 treatment of neuroAIDS. Int J Nanomedicine. 2015;10:1077–1093.
- 1285 123. Kaushik A, Jayant RD, Nikkhah-Moshaie R, et al. Magnetically guided central nervous
 1286 system delivery and toxicity evaluation of magneto-electric nanocarriers. Sci. Rep. 2016;6:
 1287 25309.
- 1288 124. Chayavichitsilp P, Buckwalter JV, Krakowski AC, et al. Herpes simplex. Pediatr. Rev.
 1289 2009;30(4):119–129.
- 1290 125. Perret F, Duffour M, Chevalier Y, et al. Design, synthesis, and in vitro evaluation of new
 1291 amphiphilic cyclodextrin-based nanoparticles for the incorporation and controlled release of
 1292 acyclovir . Eur J Pharm Biopharm. 2013;83(1):25–32.
- 1293 126. Lembo D, Swaminathan S, Donalisio M, et al. Encapsulation of Acyclovir in new
 1294 carboxylated cyclodextrin-based nanosponges improves the agent's antiviral efficacy. Int J
 1295 Pharm. 2013;443(1-2):262-72.
- 1296 127. Ruiz-Caro R, Gago-Guillan M, Otero-Espinar FJ, et al. Mucoadhesive tablets for
 1297 controlled release of acyclovir. Chem Pharm Bull. 2012;60(10):1249–1257.

- 1298 128. Pavelic Z, Skalko-Basnet N, Filipovic-Grcic J, et al. Development and in vitro
 1299 evaluation of a liposomal vaginal delivery system for acyclovir. J Control Release 2005;106(1–
 1300 2):34–43.
- 1301 129. De Clercq E. Antivirals for the treatment of herpes virus infections. J Antimicrob
 1302 Chemother. 1993;32(Suppl A):121–132.
- 1303 130. Centers for Disease Control and Prevention CDC. Incidence, prevalence, and cost of
 1304 sexually transmitted infections in the United States. http://www.cdc.gov/std">www.cdc.gov/std
 1305 2013.
- 1306 131. Ghera BB, Perret F, Chevalier Y, et al. Novel nanoparticles made from amphiphilic
 1307 perfluoroalkyl alpha-cyclodextrin derivatives: preparation, characterization and application to
 1308 the transport of acyclovir. Int J Pharm. 2009;375(1-2):155-62.
- 1309 132. Cavalli R, Donalisio M, Civra A, et al. Enhanced antiviral activity of Acyclovir loaded
 1310 into beta-cyclodextrin-poly(4-acryloylmorpholine) conjugate nanoparticles. J Control Release.
 1311 2009;137(2):116-22.
- 1312 133. Bencini M, Ranucci E, Ferruti P,et al. Preparation and in vitro evaluation of the antiviral
 1313 activity of the Acyclovir complex of a beta-cyclodextrin/poly(amidoamine) copolymer. J.
 1314 Controlled Release 2008;126(1):17-25.
- 1315 134. Gandhi A, Jana S, Sen KK. In-vitro release of acyclovir loaded Eudragit RLPO(®)
 1316 nanoparticles for sustained drug delivery. Int J Biol Macromol. 2014;67:478-82.
- 1317 135. Amany OK, Gehanne ASA, Ahmed SG, et al. Preparation of intravenous stealthy
 1318 acyclovir nanoparticles with increased mean residence time. AAPS PharmSciTech
 1319 2009;10:1427–36.
- 1320 136. Gupta S, Agarwal A, Gupta NK, et al. Galactose decorated PLGA nanoparticles for
 1321 hepatic delivery of acyclovir. Drug Dev Ind Pharm. 2013;39(12):1866-73.
- 1322 137. Azeem A, Anwer MKJ, et al. Niosomes in sustained and targeted drug delivery: recent
 1323 advances J. Drug Targeting. 2009;17:671-689.

- 1324 138. Naderkhani E, Erber A, Škalko-Basnet N, et al. Improved permeability of acyclovir:
 1325 optimization of mucoadhesive liposomes using the phospholipid vesicle-based permeation
 1326 assay. J Pharm Sci. 2014;103(2):661-8.
- 1327 139. Yandrapu SK, Kanujia P, Chalasani KB, et al. Development and optimization of
 1328 thiolated dendrimer as a viable mucoadhesive excipient for the controlled drug delivery: an
 1329 acyclovir model formulation. Nanomedicine. 2013;9(4):514-22.
- 1330 140. Ensign LM, Tang BC, Wang YY, et al. Mucus-penetrating nanoparticles for vaginal drug
 1331 delivery protect against herpes simplex virus. Sci Transl Med. 2012;4(138):138.
- 1332 141. Chetoni P, Burgalassi S, Monti D, et al. Solid lipid nanoparticles as promising tool for
 1333 intraocular tobramycin delivery: Pharmacokinetic studies on rabbits. Eur J Pharm Biopharm.
 1334 2016;109:214-223.
- 1335 142. Seyfoddin A, Al-Kassas R. Development of solid lipid nanoparticles and nanostructured
 1336 lipid carriers for improving ocular delivery of acyclovir. Drug Dev Ind Pharm. 2013;39(4):5081337 519.
- 1338 143. Jain S, Mistry MA, Swarnakar NK. Enhanced dermal delivery of acyclovir using solid
 1339 lipid nanoparticles. Drug Deliv Transl Res. 2011;1(5):395-406.
- 1340 144. Schwarz JC, Klang V, Karall S, et al. Optimisation of multiple W/O/W nanoemulsions
 1341 for dermal delivery of aciclovir. Int J Pharm. 2012;435(1):69-75.
- 1342 145. Lv Q, Yu A, Xi Y, et al. Development and evaluation of PCV-loaded solid lipid
 1343 nanoparticles for topical delivery. Int J Pharm. 2009;372(1-2):191-198.
- 1344 146. Sawdon AJ, Peng CA. Ring-opening polymerization of ε-caprolactone initiated by
 1345 ganciclovir (GCV) for the preparation of GCV-tagged polymeric micelles. Molecules.
 1346 2015;20(2):2857-2867.

- 1347 147. Yu B, Ruan M, Dong X, et al. The mechanism of the opening of the blood-brain barrier
 1348 by borneol: A pharmacodynamics and pharmacokinetics combination study. J Ethnopharmacol.
 1349 2013. pii: S0378-8741(13)00735-6.
- 1350 148. Ren J, Zou M, Gao P, et al. Tissue distribution of borneol-modified ganciclovir-loaded
 1351 solid lipid nanoparticles in mice after intravenous administration. Eur J Pharm Biopharm.
 1352 2013;83(2):141-148.
- 1353 149. Akhter S, Talegaonkar S, Khan ZI, et al. Assessment of ocular pharmacokinetics and
 1354 safety of Ganciclovir loaded nanoformulations. J Biomed Nanotechnol. 2011;7(1):144-5.
- 1355 150. Akhter S, Kushwaha S, Warsi MH, et al. Development and evaluation of nanosized
 1356 niosomal dispersion for oral delivery of Ganciclovir. Drug Dev Ind Pharm. 2012;38(1):84-92.
- 1357 151. Russo E, Gaglianone N, Baldassari S, et al. Preparation, characterization and in vitro
 1358 antiviral activity evaluation of foscarnet-chitosan nanoparticles. Colloids Surf B Biointerfaces.
 1359 2014:118:117-25.
- 1360 152. Cao Q, Wu H, Zhu L, et al. Preparation and evaluation of zanamivir-loaded solid
 1361 lipid nanoparticles. J Control Release. 2011;152 Suppl 1:e2-4.
- 1362 153. Wang H, Shao N, Qiao S, et al. Host-guest chemistry of dendrimer-cyclodextrin
 1363 conjugates: selective encapsulations of guests within dendrimer or cyclodextrin cavities
 1364 revealed by NOE NMR techniques. J Phys Chem B.2012; 116(36):11217-24.
- 1365 154. Dodiya S, Chavhan S, Korde A, et al. Solid lipid nanoparticles and nanosuspension of
 1366 adefovir dipivoxil for bioavailability improvement: formulation, characterization,
 1367 pharmacokinetic and biodistribution studies. Drug Dev Ind Pharm. 2013;39(5):733-43.
- 1368 155. Du L, Wu L, Jin Y, et al. Self-assembled drug delivery systems. Part7:hepatocyte1369 targeted nanoassemblies of an adefovir lipid derivative with cytochrome P450-triggered drug
 1370 release. Int J Pharm._2014;472(1-2):1-9.

- 1371 156. Li Q, Du YZ, Yuan H, et al. Synthesis of lamivudine stearate and antiviral activity of
 1372 stearic acid-g chitosan oligosaccharide polymeric micelles delivery system. Eur J Pharm
 1373 Sci.2010;41(3-4):498-507.
- 1374 157. Guo H, Sun S, Yang Z, et al. Strategies for ribavirin prodrugs and delivery systems for
 1375 reducing the side-effect hemolysis and enhancing their therapeutic effect. J. Control. Release
 1376 2015;209:27–36.
- 1377 158. Craparo EF, Triolo D, Pitarresi G, et al. Galactosylated micelles for a ribavirin prodrug
 1378 targeting to hepatocytes. Biomacromolecules 2013;4(6):1838–1849.
- 1379 159. Craparo EF, Teresi G, Licciardi M, et al. Novel composed galactosylated nanodevices
 1380 containing a ribavirin prodrug as hepatic cell targeted carriers for HCV treatment. J. Biomed.
 1381 Nanotechnol. 2013;9(6):1107–1122.
- 1382 160. Ishihara T, Kaneko K, Ishihara T, et al. Development of biodegradable nanoparticles for
 1383 liver-specific ribavirin delivery. J. Pharm. Sci. 2014;103(12):4005–4011.
- 1384 161. Hashim F, El-Ridy M, Nasr M, et al. Y. Preparation and characterization of niosomes
 1385 containing ribavirin for liver targeting. Drug Deliv. 2010;17(5):282–287.
- 1386 162. Liu G, Xu D, Jiang M, et al. Preparation of bioactive interferon alpha-loaded
 1387 polysaccharide nanoparticles using a new approach of temperature-induced water phase/water1388 phase emulsion. Int J Nanomedicine. 2012;7:4841-8.
- 1389 163. Li S, Zhao B, Wang F, et al. Yak interferon-alpha loaded solid lipid nanoparticles for
 1390 controlled release. Research in Veterinary Science 2010;88(1):148–153.
- 1391 164. Owen A, Rannard S. Strengths, weaknesses, opportunities and challenges for long acting
 1392 injectable therapies: Insights for applications in HIV therapy. Adv Drug Deliv Rev.
 1393 2016;103:144-56.
- •This paper describes long acting formulations for the treatment of chronic HIV infection.

1395	165.	Duan J, Freeling JP, Koehn J, et al. Evaluation of atazanavir and darunavir interactions
1396	with	lipids for developing pH-responsive anti-HIV drug combination nanoparticles. J Pharm
1397	Sci.	2014;103(8):2520-9.

- 1398 166. Steinbach JM. Protein and oligonucleotide delivery systems for vaginal microbicides
 1399 against viral STIs. Cell. Mol. Life Sci. 2015;72:469–503.
- 1400 167. Swamy MN, Wu H, Shankar P. Recent advances in RNAi-based strategies for therapy
 1401 and prevention of HIV-1/AIDS. Adv Drug Deliv Rev. 2016;103:174-86.
- This article presents the latest advances in RNAi technology towards prevention and treatment
 of HIV-1.
- 1404 168. Havel H, Finch G, Strode P, et al. Nanomedicines: From Bench to Bedside and Beyond.
 1405 The AAPS Journal 2016;18(6):1373-1378.
- 1406 169. Laffleur F, Bernkop-Schnürch A. Strategies for improving mucosal drug delivery.
 1407 Nanomedicine (Lond.) 2013;8:2061–2075.
- 1408 170. Hickey MB, Merisko-Liversidge E, Remenar JF et al. Delivery of long-acting injectable
 1409 antivirals: best approaches and recent advances. Current opinion in infectious diseases
 1410 2015,28(6):603-610.
- 1411 171. Williams PE, Crauwels HM, Basstanie ED. Formulation and pharmacology of long1412 acting rilpivirine. Curr Opin HIV AIDS. 2015;10(4):233-8.
- 1413 172. Whitfield T, Torkington A, van Halsema C. Profile of cabotegravir and its potential in
 1414 the treatment and prevention of HIV-1 infection: evidence to date. HIV AIDS (Auckl).
 1415 2016;8:157-164.
- 1416 173. Margolis DA, Boffito M. Long-acting antiviral agents for HIV treatment. Curr Opin
 1417 HIV AIDS. 2015;10(4):246-52.

1418	174.	Sathigari S, Chadha G, Lee YP, et al. Physicochemical characterization of efavirenz-
1419	cyclod	extrin inclusion complexes. AAPS PharmSciTech 2009;10:81-87.
1420	175.	Saiyed ZM, Gandhi NH, Nair MPN. AZT 50 -triphosphate nanoformulation suppresses
1421	humar	immunodeficiency virus type 1 replication in peripheral blood mononuclear cells. J.
1422	Neuro	virol. 2009;15(4):343–347.
1423	176.	Dutta T, Agashe HB, Garg M, et al. Poly (propyleneimine) dendrimer based
1424	nanoc	ontainers for targeting of efavirenz to human monocytes/ macrophages in vitro. J. Drug
1425	Target	. 2007;15(1):89–98.
1426	177.	Martinez-Skinner AL, Veerubhotla RS, Liu H, et al. Functional proteome of
1427	macro	phage carried nanoformulated antiretroviral therapy demonstrates enhanced particle
1428	carryii	ng capacity. J. Proteome Res. 2013; 12 (5):2282-2294.

1430

Figure	legend
	Figure

- 1433 **Figure 1**. Antiviral targeting strategies of drugs at different steps of the viral life cycle.
- 1434 Figure 2. Overview of main drawbacks of antivirals and nanotechnological strategies to overcome
- 1435 limits of antiviral therapy
- 1436 Figure 3. Administration routes of nanosystems studied in *in vivo* experiments and animal models used
- 1437 according to viral infections
- 1438 **Figure 4.** Future challenges of nanoformulations for antiviral drugs