

Comptes Rendus Biologies

Amelia Franke-Radowiecka, Nina Zmijewska, Tomasz Zubkiewicz, Michal Zalecki, Magdalena Klimczuk, Żaneta Listowska and Jerzy Kaleczyc

Nerve structures of the heart and their immunohistochemical characterization in 10-week-old porcine foetuses

Volume 343, issue 1 (2020), p. 53-62

Published online: 5 June 2020

https://doi.org/10.5802/crbiol.4

This article is licensed under the Creative Commons Attribution 4.0 International License. http://creativecommons.org/licenses/by/4.0/



Les Comptes Rendus. Biologies sont membres du Centre Mersenne pour l'édition scientifique ouverte www.centre-mersenne.org e-ISSN: 1768-3238 **2020**, 343, nº 1, p. 53-62 https://doi.org/10.5802/crbiol.4



News and events / Actualités

Nerve structures of the heart and their immunohistochemical characterization in 10-week-old porcine foetuses

Les structures nerveuses du cœur et leur caractéristique immunohistochimique chez des fœtus de porc de 10 semaines

Amelia Franke-Radowiecka*, ^a, Nina Zmijewska^a, Tomasz Zubkiewicz^a, Michal Zalecki^a, Magdalena Klimczuk^a, Żaneta Listowska^a and Jerzy Kaleczyc^a

^a Department of Animal Anatomy, University of Warmia and Mazury, Faculty of Veterinary Medicine, Oczapowskiego 13, 10-689 Olsztyn, Poland. E-mail: ameliaf@uwm.edu.pl (A. Franke-Radowiecka).

Abstract. The aim of the present study was to investigate the distribution of neuronal perikarya and nerve fibres and their chemical coding in the heart of 10-week-old porcine foetuses. The foetuses, obtained from a local slaughterhouse, were fixed by immersion in 4% buffered (pH 7.4) paraformaldehyde. Cryostat sections of the hearts were processed for single- or double-labelling immunofluorescence methods using antibodies against protein gene product (PGP) dopamine beta-hydroxylase (DβH), acetylcholine vesicular transporter (VAChT) and calcitonin gene-related peptide (CGRP). Numerous clusters of the PGP-positive nerve cells were observed throughout the heart wall. The majority of the clusters were found beneath the epicardium around the root of the aorta, pulmonary trunk and main veins. Some single PGP-positive neurons or small clusters of nerve cells were observed in the epicardium of heart atria and ventricles. The richest network of PGP-positive nerve fibres was observed in the base of the heart near the main cardiac blood vessel openings. Many bundles of PGP-positive nerve fibres were observed throughout the four cardiac chambers. Double-immunohistochemical staining revealed that the majority of the neurons contained immunoreactivity to VAChT, some of them stained for CGRP and the single neuronal somata were D β H-positive. The nerve fibres supplying the heart expressed immunoreactivity to D β H, VAChT or CGRP. The distribution and neurochemical coding of the nerve structures in ten-week-old foetuses are different from those observed in the hearts of juvenile

Résumé. L'objectif du présent étude était d'examiner la disposition des cellules et des fibres nerveuses, ainsi que de leur codage, aux cœurs de fœtus de porc de 10 semaines. Les fœtus, obtenus dans un abattoir locale, ont été fixés par immersion dans la solution de paraformaldéhyde 4% (pH

^{*} Corresponding author.

7,4). Des coupes de cœurs, obtenues à l'aide du cryostat, ont subi une coloration immunohistochimique simple ou double afin de dépister, dans des structures nerveuses, la présence du produit génétique protéiné (PGP), de la dopamine bêta-hydroxylase (D\(\beta\)H), du transporteur vésiculaire de l'acétylcholine (VAChT) et du peptide relié au gène calcitonine (CGRP). De nombreuses concentrations de cellules nerveuses PGP-positives ont été observées dans la paroi cardiaque. La plupart des concentrations ont été retrouvées en dessous de l'épicarde, autour de la racine de l'aorte, du tronc pulmonaire et des veines principales. Des neurones singulaires PGP-positifs ou des petites concentrations de cellules nerveuses ont été observées dans l'épicarde des oreillettes et des ventricules du cœur. Le plus riche réseau de fibres nerveuses PGP-positives a été observé dans la base du cœur, près de l'acheminement des vaisseaux principaux. Dans les quatre chambres du cœur, de nombreux faisceaux de fibres nerveuses PGP-positives ont été observées. Des colorations immunohistochimiques doubles ont démontré que la plupart de neurones présentaient une immunoréactivité par rapport à VAChT, certains par rapport à CGRP, et des corps singulaires de cellules nerveuses étaient positives par rapport à D β H. Des fibres nerveuses alimentant le cœur présentaient une immunoréactivité par rapport à D\(\beta H \), VAChT ou CGRP. La distribution et le codage neurochimique des structures nerveuses chez des fœtus de 10 semaines diffèrent de ceux observés dans les cœurs de jeunes porcs.

Keywords. Porcine foetuses, Heart innervation, Peptides. **Mots-clés.** Cœur de fœtus, Innervation cardiaque, Peptides.

Manuscript received 24th September 2019, revised 5th December 2019, accepted 6th December 2019.

1. Introduction

An important aspect of the cardiac studies is the neuroanatomy of this organ [1-6]. Research on nerve structures has given new insights into the pathological aspects of the heart function. The acquired knowledge is already used for further investigations, i.e. clinical trials [2, 5]. The research has been mainly conducted on the human heart [3, 7, 8] as well as in several animal species including the mouse [5], rat [9], guinea pig [10], dog [11], rabbit [12], heifer [6] and pig [1, 2, 13, 14]. A very interesting field of science, which is being intensively developed is xenotransplantology [13–15]. Domestic pigs are currently thought to be the best candidates for organ donation. This fact encourages wider and more thorough research on this animal species. Current experiments in xenotransplantation most often use pigs as the donor and baboons as human models. Pigs, when it comes to their physiological parameters, are very similar species to humans [13-15], among others, they have a very similar cardiac output, close blood pressure or urine osmolarity [13-15]. Despite considerable knowledge of cardiac neuroanatomy [1–3, 13, 14], little is known about the innervation of the foetal heart in mammals, including the pig. The existing data mainly concern the human foetus heart [3, 16, 17]. These data indicate that the development of cardiac autonomic innervation takes place in several stages and continues to early postnatal life [17] well as autonomic neurons and their precursors display substantial plasticity [17, 18]. Disorders in the normal developmental path, even at early stages of development, may result in cardiovascular disease in childhood or adulthood. An example of this is sinus bradycardia in mice lacking Nrp1 (neuropilin 1) and Sema3a (semaphorin 3A), important axonal growth-cone protein. Their absence is caused a defective trunk of neural crest cells (NCC) migration and results in ectopic sympathetic ganglia and a malformed stellate ganglion as the main centre of neurons whose axons supply the heart tissue [17, 19–22]. Another example is hypertension and coronary heart disease, which early birth or low birth weight infants are predisposed to because the maturation of the autonomic nervous system has not yet been completed and the heart is under the dominant influence of sympathetic innervation. Due to the fact that the domestic pig is an important animal model in experimental neurocardiology [2, 13, 14] and xenotransplantology [13-15, 23], it is worth more closely investigating the proper development of heart innervation during prenatal development. Considering the rapid rate and high complexity of the development of the heart innervation, the delayed advancement of its the sympathetic component, as well as the first appearance of some biologically active substances (e.g. CGRP) in the nerve structures not earlier than in midpregnancy [17, 19–22], it was decided to examine the porcine foetus heart nerve supply in the second half of pregnancy. Therefore, the present study aimed at

investigating the distribution and chemical coding of nerve structures in the hearts of 10-week-old porcine foetuses (these foetuses, as a unique slaughterhouse research material, constituted the oldest foetal group obtained) and can be considered as a starting point for further investigations dealing with the developmental changes occurring in the earlier and later stages of pregnancy.

2. Materials and methods

Ten-week-old porcine foetuses (n = 5) were obtained from the "Warmia" slaughterhouse (Biskupiec, Poland). According to Polish law and an EU directive (No. 2010/63/EU) the experiments performed in the present study do not require the approval of the Ethics Committee. The age of foetuses was marked according to the crown-rump length (CRL) method. CRL sets the distance from the top of the head of the embryo or foetus entering the lower limit of the buttocks [24]. After removing the foetuses from the uterus, the thorax and abdominal skin were cut for better penetration of the fixative solution in all collected animals. The foetuses were fixed by immersion in 4% buffered paraformaldehyde (pH 7.4; 4 h), rinsed with phosphate buffer (pH 7.4; overnight, in the fridge) and transferred into an 18% buffered sucrose solution (pH 7.4) where they were stored until further processing. The hearts were dissected out, cut on 12 µm-thick cryostat sections and processed for single- or double-labelling immunofluorescence (as described earlier; [25, 26]) using primary antibodies against PGP, DBH, VAChT, CGRP and corresponding secondary antibodies (Table 1). After air-drying at room temperature (RT) for 30 min the sections were pre-incubated with a blocking mixture containing 10% normal horse serum, 1% bovine serum albumin and 0.05% Tween 20 in PBS (1 h, RT). They were then incubated with a mixture of appropriate secondary antibodies (listed in Table 1). After rinsing in PBS $(3 \times 10 \text{ min})$ the slides were cover-slipped with carbonate-buffered glycerol (pH 8.6).

The omission of primary antisera and their replacement by normal, non-immune sera (rabbit, mouse or rat) were used to investigate the specificity of immunohistochemical labelling. No fluorescence was observed in any of these control stainings, which confirmed the specificity of the staining.

The labelled sections were viewed under a Zeiss Axiophot fluorescence microscope equipped with epifluorescence and an appropriate filter set for AlexaFluor 488 and AlexaFluor 555. The total number of cardiac nerve clusters or neurons in the large cardiac ganglia were counted in every fourth slice considering the size of the whole of cardiac neurons (neurons were not larger then 20 μ m). To determine the percentage (as described earlier, [25, 27]) of particular neuronal populations, at least 100 neuronal profiles for each combination of antisera were counted. The sections of sinoatrial and atrioventricular ganglia were collected from different, representative regions of the ganglia (from its upper, middle and lower one-third).

3. Results

3.1. Nerve cell bodies

Immunohistochemical staining for PGP showed that numerous clusters of nerve cells were observed throughout the foetus heart (Table 2). The majority of the clusters were found beneath the epicardium around the root of the aorta, pulmonary trunk and main veins. The sinoatrial ganglion was found to comprise the largest number (262 \pm 12) of the PGPpositive nerve cell bodies (Figure 1a). In the right atrial wall, a large PGP-positive nerve cell cluster was also observed - the atrioventricular ganglion (236 ± 13) . Neurons forming both of the ganglia were mainly oval with a large nucleus located in the cell centre. The size of these neurons was 16-20 µm. Around the cardiac segment of the ascending part of the aorta, numerous scattered spherical clusters consisting mostly of over a dozen nerve cells were observed. The others, containing from 10 to 30 PGP-positive neurons, were distributed near the pulmonary artery and smaller clusters (5-10 neurons) were observed near the venous vessels. Numerous smaller clusters consisting of 6-10 PGP-positive neurons were observed in the epicardium of both atria, in the vicinity of coronary arteries (Figure 1g). The lowest number of neurons was observed in the epicardium of the heart ventricles. They were single nerve cells or gathered into small clusters. The number of small nerve cell clusters in the whole heart area was 117 ± 21 . The size of neurons in small cell

Antigen	Species	Code	Dilution	Supplier					
Primary antibodies									
PGP 9.5	Mouse	7863-2004	1:400	Biorad					
DβH	Mouse	MAB308	1:500	Millipore					
DβH	Rabbit	BML-DZ1020-0050	1:500	Enzo					
VAChT	Rabbit	V5387	1:200	Sigma					
CGRP	Rabbit	C8198	1:600	Sigma					
Secondary reagents									
Goat AlexaFluor 488 anti-mouse		A11001	1:400	Invitrogen					
Goat AlexaFluor 488 anti-rat		A11006	1:400	Invitrogen					
Goat AlexaFluor 555 anti-rabbit		A11010	1:400	Invitrogen					

Table 1. List of primary antisera and secondary reagents used in the study

clusters was slightly smaller (11–17 μm) than those in the large ganglia.

Double immunohistochemical stainings revealed that the majority of the PGP-positive neurons contained immunoreactivity to VAChT (Figure 1a–c), CGRP (Figure 1g–i) and the single nerve cells were D β H-positive (Figure 1d–f). In the sinoatrial and atrioventricular ganglion in the heart of 10-week-old foetus pigs, a large number of PGP-positive neurons contained immunoreactivity to VAChT (53,8% and 51,7% respectively), some to CGRP (13,2% and 13,9% respectively) and a few were D β H-positive (1,6% and 1,2% respectively). Single neurons were DBH/VAChT-positive. In the smaller nerve body clusters, neurons mainly contained VAChT (41,22%), a few D β H (9,42%) and single nerve body cells were CGRP-positive (Figure 1g–i).

3.2. Nerve fibres

Immunohistochemical staining for PGP showed that many bundles of nerve fibres have been observed throughout all cardiac chambers of the porcine foetal heart (Table 3). The richest network of PGP-positive nerve fibres was observed in the base of the heart in the vicinity of the main cardiac blood vessel openings (Figure 2a). The nerve bundles were sometimes associated with single neurons or small clusters of nerve cells. Most of the nerve bundles were observed in the epicardium, with less in the myocardium and endocardium. Generally, many of them were found in

the left and right atrial epicardium and were less numerous observed in the left and right ventricular epicardium. Many PGP-positive nerve fibres were distributed in the endocardium, in the interatrial septum and some in the interventricular septum. Thinner bundles or single nerve fibres were observed in the myocardium of the atria and ventricles (Figure 2g) of the heart.

The majority of epicardial, myocardial and endocardial nerve fibres were D\(\beta\)H-positive (Table 2), (Figure 2a–c). A low number of fibres were VAChT-positive. DBH/VAChT-positive nerves were not observed (Figure 2d–f). Few axons observed in the epicardium or myocardium contained immunoreactivity to CGRP and single nerves were found in the endocardium of four heart chambers (Figure 2g–i). There was no co-localization of CGRP with other studied substances in the nerve fibres supplying the foetus heart.

4. Discussion

As mentioned earlier, the development of the autonomic innervation of the heart is a multi-stage process [17,18]. During the fifth week of human prenatal development, the NCC starts to migrate to the dorsal aorta and then differentiate into neurons. After this process, the neurons migrate to form a paravertebral sympathetic chain or parasympathetic cardiac ganglia and finally the axons of nerve body cells elongate and penetrate the cardiac tissue. This process continues until the first weeks after birth [17, 18, 28, 29].

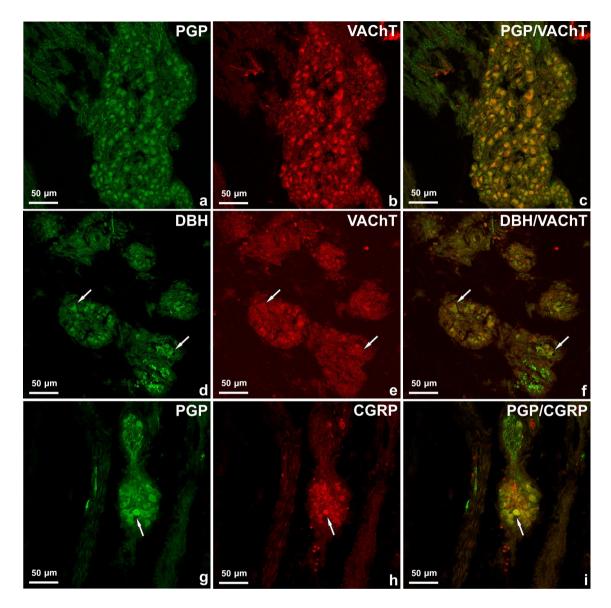


Figure 1. Longitudinal-section of sinoatrial ganglion in the heart of 10-week-old foetus pig (a–c) labelled for PGP (a) and VAChT (b). The large number of PGP-positive neurons contained immunoreactivity to VAChT. Merged images 1a and 1b (c). Nerve cell cluster located around the cardiac segment of the ascending part of the aorta (d–f). Some neurons contained immunoreactivity to D β H (d) but were VAChT-negative (e, arrows). In the small nerve body cluster (g–i) which was observed in the epicardium of left atria, in the vicinity of coronary arteries, single PGP-positive neurons (g) contained CGRP (h). Merged images 1g and 1h (i).

According to the current study, in the heart of the 10-weeks-old porcine foetus, there are typical cardiac nerve structures. However, analysing the obtained results, it is clear that the process of maturation, migra-

tion and proliferation of the nervous system structures in the heart area has not yet been completed. This can be concluded by the number of neurons in the sino-atrial, atrio-ventricular ganglion, as well as

Table 2. Total number of PGP-positive neurons counted in different areas of the heart and percentages of particular neuronal subpopulations containing substances investigated in cardiac PGP-positive neurons

Ganglia	Total number of neurons	Percentage of particular neuronal subpopulations		
	PGP	PGP/VAChT	PGP/CGRP	PGP/DβH
Sinoatrial	262 ± 12	53,8%	13,2%	1,6%
Atrioventricular	236 ± 13	51,7%	13,9%	1,2%
Smaller clusters of neurons	117 ± 21	41,22%	Single	9,42%

Table 3. Relative frequency of PGP-IR, D β H-IR, VAChT-IR and CGRP-IR nerve fibres in the 10-week-old porcine foetus heart

Nerve fibres		Right atrium	Right ventricle	Left atrium	Left ventricle
Epicardium	PGP	+++++	++++	+++++	++++
	PGP/DβH	++++	+++	++++	+++
	PGP/VAChT	++	+	++	+
	PGP/CGRP	++	+	++	+
Myocardium	PGP	+++	+++	+++	+++
	PGP/DβH	++	++	++	++
	PGP/VAChT	+	+	+	+
	PGP/CGRP	++	++	++	++
Endocardium	PGP	+++	+++	+++	+++
	PGP/DβH	++	++	++	++
	PGP/VAChT	+	+	+	+
	PGP/CGRP	++	++	++	++

Many fibres +++++, moderate number of fibres ++++, low number of fibres ++++, few fibres ++, single fibres +.

by the number and distribution of smaller nerve cell clusters compared to juvenile pigs [1, 2, 13, 14]. Previous studies have shown that in the hearts of fewweek-old piglets there were an average of 359 ± 178 neurons in the large ganglia [2] while in hearts of 10-week-old pig foetuses there were fewer (249 \pm 10) nerve cell bodies observed. The number of small ganglia in piglets [2] were more numerous than in the heart of porcine foetus (361 ± 52 and 117 ± 21 , respectively) and the most of them were observed in the left atrial and in the left and right ventricle area of the piglet hearts [2]. The small cardiac ganglia in the foetus were mainly observed on the base of the heart and in the vicinity of coronary arteries while the least were observed in the epicardium of the heart ventricles. There were single nerve cells or gathered into

very small clusters. In comparing the results, the differences in the distribution of a small clusters of neurons in the hearts of piglets [2, 13, 14] and 10-weekold foetuses are visible, which strongly suggests that they are still in the process of migration. Moreover, it may seem that the innervation of the porcine heart matures more slowly than in humans. It has been observed that in 24 week-old human foetuses (roughly, proportionately similar pregnancy time in a pig) the distribution pattern of heart innervation has been observed in adults [16]. This may be associated with a more extensive cardiac intrinsic nervous system in the pig than in humans, which is a characteristic feature of the porcine heart but also makes a large difference in the morphology of the heart of both species and may today pose difficulties for the function of the

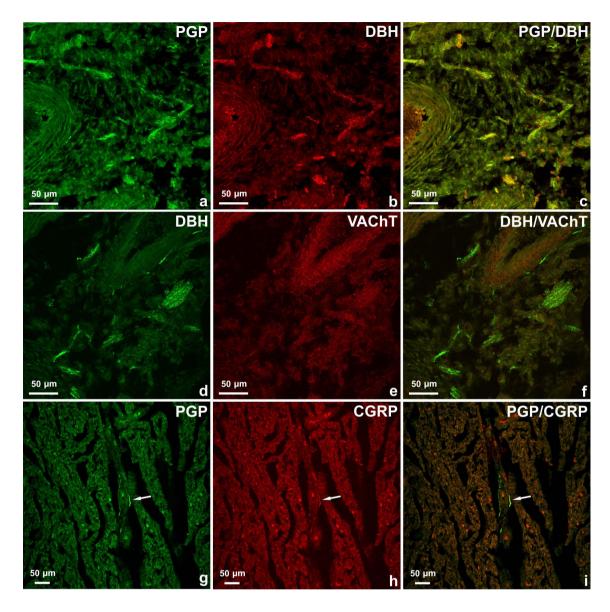


Figure 2. The richest network of nerve fibres was observed in the base of the heart in the vicinity of the main cardiac blood vessel openings (a–f). The majority of PGP-positive nerve fibres (a) were D β H-positive (b). D β H-positive nerve fibres (d) were VAChT-negative (e). PGP-positive nerve fibres (g, arrow) were observed in right ventricle myocardium. Some of them contained immunoreactivity to CGRP (h). Merged images 2g and 2h (i).

heart donor by the transgenic pig [13–15].

Analysing the distribution of nerve fibres in the heart area of the porcine foetus and the piglet, differences in the distribution and quantity of fibres are also noticeable. As described by Crick *et al.* [14], a

gradient of immunoreactivity for PGP was observed from dense plexus of nerve fibres in the epicardium and endocardium, to a lesser extent in the myocardial tissue. Considering the distribution of innervation in different areas of the piglet heart, the epicardial tissues displayed a ventricular-to-atrial gradient in density, the myocardial tissues displayed an atrialto-ventricular gradient and, for the endocardial tissues, a distinct right to left chamber gradient was observed, with the right ventricle possessing the highest and the left atrium possessing the lowest density of nerves [14]. Generally, significantly fewer nerve fibres were observed in the endocardium and myocardium in the pig foetus. Nerve plexuses and fibres were mostly arranged in the epicardium of both atria and were much less observed in the ventricles area, which is a clear difference between the foetal heart and this organ in the animal after birth. It is most probable that in the studied stage of prenatal development, the axons are in the process of elongation and penetrate the cardiac tissue.

Referring to the immunohistochemical characterization of nerve structures in the foetal heart, it was observed that the occurrence of VAChT, DBH or CGRP in nerve fibres is also slightly different than in juvenile pig in the studied stage of development.

It is known that the ontogenesis of parasympathetic and intrinsic cardiac innervation precedes sympathetic innervation [14, 16, 17, 30] and cholinergic nerves were presented in 12 weeks of gestation and adrenergic nerves were not identified in this organ up to 18 weeks of gestation in the human foetus heart [16,31-33] but there is no data in relation to the porcine foetus heart. Analysing the obtained results, it was observed that in the ten-week-old porcine foetus, the majority of cardiac neurons contain VAChT while the only few were DBH-positive compared to the piglet heart [14]. This observation may suggest a still-developing sympathetic part of the cardiac intrinsic nervous system. Another observation concerns the nerve fibres supplying the heart of the porcine foetus. Many of the fibres, especially those present in the epicardium and containing DBH, are mainly efferent post-ganglionic sympathetic nerves, while the intrinsic fibres are just forming. VACHTpositive nerve fibres were visible in the epicardium and single fibres were visible in the myocardium and endocardium. These nerves probably originated from the cardiac ganglia and were at the stage of penetration and elongation. CGRP-positive neurons and nerve fibres were the most abundant subpopulation in the piglet heart [14]. Most of these nerve structures originate from the intrinsic cardiac nervous system and simultaneously contain immunoreactivity

to AChE [14]. CGRP in these nerve fibres has probably been involved in coronary vasodilatation [34] and may also play a role in the neutrally-mediated negative inotropic effect on myocardial function observed during cholinergic coronary vasoconstriction in the pig [14, 35]. Other CGRP-positive nerve fibres are sensory and originate from vagal or dorsal root ganglia neurons [14,36]. In the ten-week-old porcine foetus, CGRP-positive neurons were mainly observed in large cardiac intrinsic ganglia (13.55%) and nerves were weakly visible in the entire heart area. These observations confirm the view that the autonomic innervation of the heart is still developing. In human CGRP/SP-positive nerve fibres (sensory) were not detected until 24 weeks of gestation [16]. It is known that sensory innervation appears later than autonomic innervation [13, 14, 16], which is also confirmed in the current study.

In summary, this study has described nerve structures of the heart and their immunohistochemical characterization in ten-week-old porcine foetuses. The distribution of nerve structures in ten-week-old foetuses represent a relatively high level of advancement, however, it is different from that observed in the heart of several-week-old pigs [2, 13, 14]. The neurochemical coding of the heart nerve structures at this stage of development is slightly different to that observed in the hearts of several-week-old pigs [13] and it is associated with a still developing the cardiac nervous system. The present results will constitute the basis for further observations of the developing heart in the porcine foetus and it is worth highlighting these studies, which provide details on the development and functioning of the heart in a pig as a potential donor of this organ for humans.

Conflicts of interest

The authors have no conflict of interest to declare.

Acknowledgements

The authors would like to thank Anna Przyborowska and Aleksander Penkowski for their contributions in preparing this paper.

This publication was supported by RID (Project financially co-supported by Minister of Science and Higher Education in the range of the program entitled "Regional Initiative of Excellence" for the years 2019-2022). Project No. 010/RID/2018/19, amount of funding 12.000.000 PLN.

References

- R. C. Arora, M. Waldmann, D. A. Hopkins, J. A. Armour, "Porcine intrinsic cardiac ganglia", *Anat. Rec.* 271 (2003), p. 249-258.
- [2] D. Batulevicius, V. Skripka, N. Pauziene, D. H. Pauza., "To-pography of the porcine epicardiac nerve plexus as revealed by histochemistry for acetylcholinesterase", *Auton. Neurosci.* 138 (2008), p. 64-75.
- [3] E. Gardner, R. O'Rahilly, "The nerve supplying and conducting system of the human heart at the end of the embryonic period proper", J. Anat. 121 (1976), p. 571-587.
- [4] D. H. Pauza, V. Skripka, N. Pauziene, "Morphology of the intrinsic cardiac nervous system in the dog: a wholemount study employing histochemical staining with acetylcholinesterase", Cells Tissues Organs 172 (2002), p. 297-320.
- [5] D. H. Pauza, K. Rysevaite, H. Inokaitis, M. Jokubauskas, A. G. Pauza, K. E. Brack, N. Pauziene, "Innervation of sinoatrial nodal cardiomyocytes in the mouse. A combined approach using immunofluorescent and electron microscopy", *J. Mol. Cell. Cardiol.* 75 (2014), p. 188-197.
- [6] N. Demus, "Organometry heart heifers depending on autonomous regulation heart rate. Органометрія серця теличок залежно від типу автономної регуляції серцевого ритму / Н.В. Демус // Науковий вісник Львівського національного університету ветеринарної медицини та біотехнологій імені С.З", Гжицького. Львів 17 (2015), р. 24-29.
- [7] D. H. Pauza, V. Skripka, N. Pauziene, R. Stropus, "Morphology, distribution and variability of the epicardiac neural ganglionated subplexuses in the human heart", *Anat. Rec.* 259 (2000), p. 353-382.
- [8] R. B. Smith, "Development of innervation of coronary arteries in human foetus up until 230 mm stage (mid-term)", Br. Heart J. 32 (1970), p. 108-113.
- [9] D. Batulevicius, N. Pauziene, D. H. Pauza, "Topographic morphology and age-related analysis of the neuronal number of the rat intracardiac nerve plexus", *Ann. Anat.* 185 (2003), p. 449-459.
- [10] D. Batulevicius, N. Pauziene, D. H. Pauza, "Architecture and age-related analysis of the neuronal number of the guinea pig intrinsic cardiac nerve plexus", *Ann. Anat.* 187 (2005), p. 225-243.
- [11] D. H. Pauza, V. Skripka, N. Pauziene, R. Stropus, "Anatomical study of the neural ganglionated plexus in the canine right atrium: implications for selective denervation and electrophysiology of the sinoatrial node in dog", *Anat. Rec.* 255 (1999), p. 271-294.
- [12] I. Saburkina, L. Gukauskiene, K. Rysevaite, K. E. Brack, A. G. Pauza, N. Pauziene, D. H. Pauza, "Morphological pattern of intrinsic nerve plexus distributed on the rabbit heart and interatrial septum", *J. Anat.* 224 (2014), p. 583-593.
- [13] S. J. Crick, R. H. Anderson, S. Yen Ho, M. N. Sheppard, "Localization and quantitation of autonomic innervation of

- the porcine heart II: endocardium, myocardium and pericardium", *J. Anat.* **195** (1999b), p. 359-373.
- [14] S. J. Crick, M. N. Sheppard, S. Yen Ho, R. H. Anderson, "Localization and quantitation of autonomic innervation of the porcine heart I: conduction system", J. Anat. 195 (1999a), p. 341-357.
- [15] M. Samiec, M. Skrzyszowska, "The possibilities of practical application of transgenic mammalian species generated by somatic cell cloning in pharmacology, veterinary medicine and xenotransplantology", Pol. J. Vet. Sci. 2 (2011), p. 329-340.
- [16] L. Gordon, J. M. Polak, G. J. Moscoso, A. Smith, D. M. Kuhn, J. Wharton, "Development of peptidergic innervation of the human heart", J. Anat. 183 (1993), p. 131-140.
- [17] W. Hasan, "Autonomic cardiac innervation", Organogenesis 9 (2013), p. 176-193.
- [18] S. R. Yiallourou, S. A. Sands, A. M. Walker, R. S. C. Horne, "Maturation of heart rate and blood pressure variability during sleep in term-born infants", *Sleep* 35 (2012), p. 177-186.
- [19] M. Ieda, H. Kanazawa, K. Kimura, K. Matsumura, Y. Tomita, T. Yagi, T. Onizuka, K. Shimoji, S. Ogawa, S. Makino, M. Sano, K. Fukuda, "Nerve growth factor is critical for cardiac sensory innervation and rescues neuropathy in diabetic hearts", *Circulation* 114 (2006), p. 2351-2363.
- [20] M. Ieda, H. Kanazawa, Y. Ieda, K. Kimura, F. Hartori, Y. Ieda, M. Taniguchi, J. K. Lee, K. Matsumura, Y. Tomita, S. Miyoshi, K. Shimoda, S. Makino, M. Sano, I. Kodama, S. Ogawa, K. Fukuda, "Sema3a maintains normal heart rhythm through sympathetic innervation patterning", *Nat. Med.* 13 (2007), p. 604-612.
- [21] A. Fantin, C. H. Madan, C. Ruhrberg, "Neuropilin ligands in vascular and neuronal patterning", *Biochem. Soc. Trans.* 37 (2009), p. 1228-1232.
- [22] Q. Schwarz, C. H. Maden, J. M. Vieira, C. Ruhrberg, "Neuropilin 1 signaling guides neural crest cells to coordinate pathway choice with cells specification", *Proc. Natl. Acad. Sci. USA* 106 (2009), p. 6164-6169.
- [23] M. M. Mohiuddin, B. Reichart, G. W. Byrne, C. G. A. McGregor, "Current status of pig heart xenotransplantation", *Int. J. Surg.* 23 (2015), p. 234-239.
- [24] H. E. Evans, W. O. Sack, "Prenatal development of domestic and laboratory mammals: growth curves, external features and selected references", *Anat. Histol. Embryol.* 2 (1973), p. 11-45.
- [25] A. Franke-Radowiecka, "Immunohistochemical characterisation of dorsal root ganglia neurons supplying the porcine mammary gland", *Histol. Histopathol.* 26 (2011), p. 1509-1517.
- [26] M. Zalecki, "Localization and neurochemical characteristics of the extrinsic sympathetic neurons projecting to the pylorus in the domestic pig", J. Chem. Neuroanat. 43 (2012), p. 1-13.
- [27] W. Sienkiewicz, A. Dudek, A. Zacharko-Siembida, M. Marszałek, "Immunohistochemical characterization of the jugular (superior vagal) ganglion in the pig", *Pol. J. Vet. Sci.* 20 (2017), p. 377-385.
- [28] J. A. Armour, "Intrinsic cardiac neurons involved in cardiac regulation possess alpha 1-, alpha 2-, beta 1- and beta 2adrenoceptors", Can. J. Cardiol. 13 (1997), p. 277-284.
- [29] D. C. Randall, D. R. Brown, A. S. McGuirt, G. W. Thompsom, J. A. Armour, J. L. Ardell, "Interactions within the intrinsic

- cardiac nervous system contribute to chronotropic regulation", *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **285** (2003), p. 1066-1075.
- [30] A. J. Pappano, "Ontogenetic development of autonomic neuroeffector transmission and transmitter reactivity in embryonic and fetal hearts", *Pharmacol. Rev.* 29 (1977), p. 3-33.
- [31] W. G. Dail, G. C. Palmer, "Localisation and correlation of catecholamine-containing cells with adenyl cyclase and phosphodiesterase activities in the human fetal heart", *Anat. Rec.* 177 (1973), p. 265-288.
- [32] L. Kanerva, A. Hervonen, H. Hervonen, "Morphological characteristics of the ontogenesis of the mammalian peripheral

- adrenergic nervous system with special remarks on the human fetus", *Med. Biol.* **52** (1994), p. 144-153.
- [33] S. Partanen, O. Korkala, "Catecholamines in human fetal heart", *Experientia* **30** (1974), p. 798-800.
- [34] A. Franco-Ceraceda, J. M. Lundberg, T. Hökfelt, "Somatostatin: an inhibitory parasympathetic transmitter in the human heart?", Eur. J. Pharmacol. 132 (1986), p. 101-102.
- [35] J. Cinca, A. Carreno, L. Moni, P. Blanch, J. Soler-Soler, "Neurally-mediated negative inototropic effect impairs myocardial function during cholinergic coronary vasoconstriction in pig", Circulation 94 (1996), p. 1101-1108.
- [36] S. Forsgren, "Distribution of calcitonin gene-related immunoreactivity in the bovine conduction system: correlation with substance", *Regul. Pept.* 52 (1994), p. 7-19.