



UvA-DARE (Digital Academic Repository)

Pediatric connective tissue diseases

Looking through the skin and beyond

Schonenberg-Meinema, D.

Publication date

2022

Document Version

Final published version

[Link to publication](#)

Citation for published version (APA):

Schonenberg-Meinema, D. (2022). *Pediatric connective tissue diseases: Looking through the skin and beyond*. [Thesis, fully internal, Universiteit van Amsterdam].

General rights

It is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), other than for strictly personal, individual use, unless the work is under an open content license (like Creative Commons).

Disclaimer/Complaints regulations

If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please Ask the Library: <https://uba.uva.nl/en/contact>, or a letter to: Library of the University of Amsterdam, Secretariat, Singel 425, 1012 WP Amsterdam, The Netherlands. You will be contacted as soon as possible.



PEDIATRIC Looking
CONNECTIVE through
TISSUE the skin and
DISEASES beyond

DIENEKE SCHONENBERG-MEINEMA

**Pediatric connective tissue diseases:
looking through the skin and beyond**

Dieneke Schonenberg-Meinema

The work in this thesis was performed at the Department of Pediatric Immunology, Rheumatology and Infectious Diseases at Amsterdam University Medical Centers, at the location of AMC/Emma Children's Hospital from the University of Amsterdam, The Netherlands. This thesis contains collaborative projects with Department of Rheumatology from Ghent University in Belgium and Department of Pediatric Rheumatology from Sophia Children's Hospital, Erasmus Medical Center, Rotterdam, The Netherlands.

The research described in this thesis was financially supported by "Stichting Steun Emma" and "Zeldzame Ziektenfonds". Both had no involvement in the study design, data collection, data analysis, interpretation of data or decision to publish the study results.

Printing of this thesis was supported by a non-commercial grant from Stichting NVLE fonds:



Voor onderzoek naar lupus,
APS, sclerodermie en MCTD

Cover: Gineke Schonenberg

Lay-out and printing by: Optima Grafische Communicatie (www.ogc.nl)

**Pediatric connective tissue diseases:
looking through the skin and beyond**

ACADEMISCH PROEFSCHRIFT

ter verkrijging van de graad van doctor aan de Universiteit van Amsterdam op gezag
van de Rector Magnificus
prof. dr. ir. P.P.C.C. Verbeek
ten overstaan van een door het College voor Promoties ingestelde commissie, in het
openbaar te verdedigen in de Aula der Universiteit
op vrijdag 9 december 2022, te 11.00 uur

door

Dieneke Schonenberg geboren te Rijnsburg

PROMOTIECOMMISSIE

Promotor:	prof. dr. T.W. Kuijpers	AMC-UvA
Copromotores:	dr. J.M. van den Berg dr. V. Smith	AMC-UvA UZ Gent
Overige leden:	prof. dr. R.F. van Vollenhoven prof. dr. R. ten Cate prof. dr. A.E. Voskuyl prof. dr. M. Maas dr. A. van Royen-Kerkhof dr. S.W. Tas	AMC-UvA Universiteit Leiden Vrije Universiteit Amsterdam AMC-UvA UMC Utrecht AMC-UvA

Faculteit der Geneeskunde

A decorative graphic consisting of a red dotted line that forms a large, irregular, looping shape. The line starts at the top center, curves down and left, then loops back up and right, then down and left, then up and right, and finally down and left, ending at the bottom center. The overall shape is reminiscent of a stylized 'S' or a calligraphic flourish.

Voor Zenani

en Minou

TABLE OF CONTENTS

Chapter 1	General introduction and outline of thesis	9
Part I.	Nailfold videocapillaroscopy in cSLE	33
Chapter 2	Capillaroscopy in childhood-onset systemic lupus erythematosus: a first systemic review	35
Chapter 3	Nailfold capillary abnormalities in childhood-onset systemic lupus erythematosus: a cross-sectional study compared with healthy controls	55
Chapter 4	Standardized nailfold capillaroscopy in children with rheumatic diseases: a worldwide study	75
Chapter 5	A capillary scleroderma pattern in childhood-onset may be associated with disease damage: important lessons from longitudinal follow-up	99
Part II.	New biomarkers in (systemic) connective tissue diseases	121
Chapter 6	Gene signature fingerprints divide SLE patients in subgroups with similar biological disease profiles: a multicenter longitudinal study	123
Chapter 7	Rarities in rare: illuminating the microvascular and dermal status in juvenile localized scleroderma. a case series	151
Chapter 8	Consider the wrist: a retrospective study on pediatric connective tissue disease with MRI	167
Part III.	Discussion, summary and appendix	179

1

**General introduction,
aims and outline of thesis**

GENERAL INTRODUCTION AND AIMS

Autoimmunity in children

Autoimmunity is a pathological phenomenon in the body when the immune system reacts with an inflammatory reaction to its own cells or organs, as if it were foreign. The word 'auto-immune' is a combined word from two ancient Greek and Latin words, meaning immunity (Latin) to itself (auto from Greek). Autoimmune diseases in children are fortunately rare but, when present, these diseases are often chronic and a lifelong burden. They may give limitations in normal growth and development during childhood, leading to invalidating disabilities in adult life. Moreover, these diseases often give limitations in normal daily activities such as school, sports and social activities.

In these last decades there has been an enormous growth in international collaborations between physicians and researchers in the field of pediatric rheumatology. Existing networks are expanding and cross-linking, leading to larger cohort studies. These collaborations are important in rare rheumatic diseases in children, for which new scientific insights and therapeutic developments are only implemented until long investigating and approving procedures are completed in adult patients. Much improvement has been made during the last decades in terms of early diagnoses and treatment options for (systemic) autoimmune diseases in children. This has led to a higher quality of life for patients with childhood-onset of chronic autoimmune diseases and led to better outcomes for these children in adulthood. Still, there are a lot of unmet needs as will be described below.

The autoimmune diseases that are studied in this thesis are childhood-onset systemic lupus erythematosus (cSLE), juvenile mixed connective tissue disease (JMCTD) and localized scleroderma (LS). The focus of this thesis is the identification of novel disease biomarkers, mainly in imaging, which may help to correctly classify these (systemic) autoimmune disease at an early stage of disease. The value of these different prosperous biomarkers are examined in terms of monitoring disease activity and risk for damage. Novel biomarkers have to lead to adapted personalized treatment choices and better outcomes for individual pediatric patients.

Systemic Lupus Erythematosus (SLE)

Incidence, prevalence and demographics

SLE is a systemic autoimmune disease which is chronic and thus lifelong. It often leads to severe inflammation with multi-organ involvement but there is a huge variety of clinical presentations. These presentations vary not only in type of organ involvement but also in severity of the inflammation from mild to very severe, and are sometimes even

life-threatening (see figure 1). SLE is characterized by the presence of one or more auto-antibodies against nuclear antigens of cells, such as anti-nuclear antibodies (ANA) and anti-double stranded DNA (anti-ds-DNA), or against cell-surface antigens, for instance to red blood cells leading to haemolytic anaemia. The auto-antibodies in SLE lead to formation of immune complexes and a cascade of inflammatory responses leading to low complement levels of C3 and C4.

The incidence of adult-onset SLE (aSLE) is estimated up to 25 per 100,000 and the prevalence is 20-150 per 100,000 (1-5). SLE with a childhood onset (before the age of 18 years) is called childhood-onset SLE (cSLE). Fortunately, cSLE has a lower incidence than aSLE, of 1-2 per 100,000 and a prevalence of 2-25 per 100,000 children worldwide (5-7). Approximately 10-20% of all SLE cases start during childhood, and those childhood patients have a mean age of onset at 11-12 years (8). SLE occurs predominantly in women, the female:male ratio is approximately 7-15: 1 (1, 7).

The prevalence of SLE, in both adults and children, is higher *and* more severe in people with Asian, African-American, African-Caribbean and Hispanic background compared to white individuals (1, 9). In a recent epidemiologic study, black females had the highest prevalence of SLE (230 per 100,000) (3). It has also been shown that non-white patients are diagnosed at a significant younger median age (10, 11).

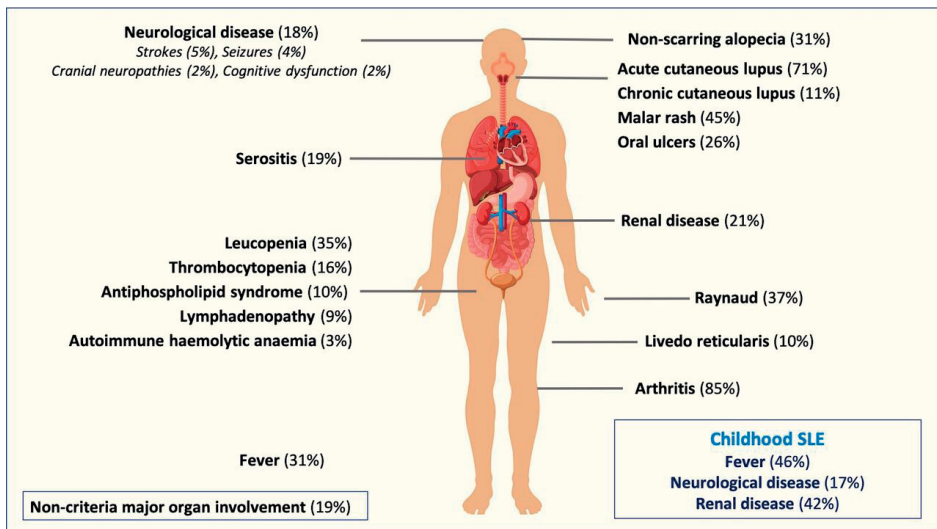


Figure 1. Multi-organ involvement In SLE (with consent from Fanouriakis et al. (12))

Classification criteria and disease activity scores

As mentioned above, SLE is a heterogeneous disease with many different presentations of (multi-)organ involvement. For high quality studies, a homogeneous study population is needed, or homogeneous subgroups. Over the years classification criteria for SLE have evolved and been adapted (figure 2). The Systemic Lupus International Collaborating Clinics (SLICC) 2012 criteria were proposed because of some limitations in the earlier ACR 1997 criteria (13). The SLICC 2012 criteria defined 17 criteria of which a minimum of 4 should be present with a minimum of 1 (of 11) clinical and 1 (of 6) immunological criteria (14). Presence of lupus nephritis with presence of ANA or anti-double-stranded DNA (anti-ds-DNA) antibodies without other criteria was also qualified for SLE classification. The SLICC 2012 criteria were validated and showed a higher sensitivity than ACR 1997 criteria (97 versus 83%) but a lower specificity (84 versus 96%) (14). In 2019, the new weighted EULAR/ACR criteria for SLE were published (15). These EULAR/ACR classification criteria start with the presence of ANA and thereafter other criteria (7 clinical and 3 immunological) must be met with each weighted points between 2 and 10. Patients are classified with SLE if they have a score of ≥ 10 . Classification criteria were developed to improve detection of new/early SLE in order to prevent delay in diagnosis. In a validation cohort, these EULAR/ACR 2019 criteria had a sensitivity of 96 percent and a specificity of 93% (15).

Recently, all classification criteria were compared for their performance, including the SLICC 2012 criteria, adapted with weighing factors for clinical manifestations with scores between 1-26. There was overall agreement for the diagnosis of aSLE by physician-rated patient scenarios without any statistically significant differences between the ACR 1997, SLICC 2012 and EULAR 2019 criteria (17). It was concluded that both (unweighted) SLICC 2012 or the more complex (weighted) EULAR/ACR 2019 can be used in the research field of SLE, although performance is always dependent on the type of patient population.

In a subsequent study, the performance of these three different classification criteria were also assessed in cSLE. SLICC 2012 criteria showed highest sensitivity (95%) and ACR 1997 the highest specificity (95%) (18). These high numbers for sensitivity and specificity show that all current available classification criteria seem sufficient for studies in cSLE-patients. In this thesis, the SLICC 2012 criteria were used.

Multiple SLE disease activity scoring systems have been developed, validated and improved over the last decades. Two of the most used disease activity scores are Systemic Lupus Erythematosus Disease Activity Index (SLEDAI) and British Isles Lupus Assessment Group (BILAG) (19, 20). These disease activity scores are mostly used in clinical trials to evaluate patient outcomes and treatment responses but can also be used in

daily clinical practice for follow-up. In clinical setting the SLEDAI is easy to use in daily practice, although this score needs some blood results and therefore it often takes a week after the patient was seen. The scoring of BILAG is more time consuming because to scoring list is longer than SLEDAI and scoring items need to be compared with the last visit (in terms of: improving, same or worse). Comparing these two scoring systems it was concluded by Yee et al. that SLEDAI is reliable when compared to BILAG but some patients, requiring increased treatment, were less likely to be picked up by SLEDAI (21).

Suspicion of SLE		
ACR	SLICC	EULAR/ACR
any 4 of 11	Histology compatible with lupus nephritis and ANA or anti-dsDNA OR any 4 of 17 (at least one immunological)	ANA positive 10 points weighted items (highest in each domain counted only)

Figure 2. Comparison of different classification criteria for SLE over the last decades (with consent from Aringer et al. (16))

Childhood-onset versus adult-onset SLE: disease damage and mortality

In general, cSLE patients suffer from more severe disease than aSLE patients. Longitudinal studies show that this is reflected in higher disease activity, not only at disease onset but also during follow-up and by higher disease damage scores (22, 23). These same studies show that cSLE-patients are more prone for renal involvement, which can cause chronic kidney insufficiency leading to dialysis or kidney transplantation (10, 22, 23). Neuropsychiatric involvement is also higher in cSLE compared to aSLE (10). Large studies show that especially renal and neuropsychiatric involvement (in cSLE) is associated with a higher risk for irreversible disease damage (25, 26). In a long-term study in adults with cSLE (with a median disease duration of 20 years), 62% of patients had disease damage, predominantly in musculoskeletal, neuropsychiatric and renal systems. Specific disease damage occurred at a low median age of 20 years (cerebrovascular accidents), 24 years (renal transplants), 34 years (replacement arthroplasties) and 39

years (myocardial infarctions) (24). All of these data show that the damage in cSLE patients already occurs at a relatively early age when their peers are in the bloom of their life. Children with SLE differ from the aSLE patients in other types of damage (such as growth failure and puberty delay) but also in terms of disease burden in daily life, with different indicators for quality of life and educational goals. Health related quality of life (HRQoL) in adults with a childhood-onset of SLE is impaired in most domains, compared to the general population. The overall HRQoL score was irrespective of disease related organ damage by SLICC damage index (SDI) (24). After the educational phase of their life, SLE has a great impact on employment as an adult independent individual. Groot et al. showed that in adults with SLE, with a childhood-onset of disease, 50% of patients had adjusted their vocational choice, 44% of patients did not have a paid job and 51% had been declared work disabled. Disease related organ damage was equally prevalent in patients with and without paid employment (27).

Over the last decades, mortality rates decreased significantly from a 5-year survival rate of 50% in 1955 to >95% in the early 2000s (28, 29). The mean age of death has risen from 42 years in 1970s to 58 years in 2013. Nevertheless, the age-specific standard mortality *ratio* (compared to healthy age-matched peers) was still found to be the highest for SLE-patients in the age group of 25-34 years when compared to older age groups (28). In a more recent study was shown that mortality in patients with cSLE still is still 4.8% only 5 years after diagnosis, and in all age groups comorbidities attributed to 16.3% all-cause mortality (30).

It is necessary to lower the mortality rate in SLE further but also to lower risks of irreversible disease damage. Besides that, drug toxicity also needs to be considered constantly because of lifelong dependence on immunosuppressive medication, which is needed to halt disease progression and prevent disease flares. In particular for adolescent patients, which will have to live their whole life with this disease, it is needed to improve the daily quality of life in all domains, and to support their ability to pursuit their goals in life. Considering all of the above, it means that although a lot of progress has been made in the last decades, there are still many unmet needs for more disease biomarkers in SLE to improve treatment decisions and to predict risk for damage.

New techniques in the assessment of novel SLE disease biomarkers

Nailfold videocapillaroscopy

Nailfold capillaroscopy is an attractive imaging tool to use in daily clinical practice because it is non-invasive and easy to use with direct imaging results during the nailfold investigation of the patient. Over the last years, the quality of imaging of the nailfold microvascular bed has greatly improved with x200 magnification videocapillaroscopy. The main reason for us to further investigate capillary abnormalities in cSLE was because of the observation that many of our cSLE patients showed nailfold capillary hemorrhages (figure 3). Initially, Raynaud's phenomenon or acro-cyanotic complaints were an indication in these patients to look at their nailfold capillaries. Because SLE is a disease with intravascular inflammation, some studies had already used this technique to observe the capillary bed in the nailfolds of SLE patients, also without outspoken acro-cyanotic complaints. Recently, a systematic literature review showed that *adult* SLE patients have significantly more abnormal capillary shapes and -hemorrhages, which were associated with disease activity (by SLEDAI) (31). This systemic review also indicated that most data were cross-sectional of nature and less is known about longitudinal follow-up of these capillary abnormalities. Data on nailfold abnormalities in patients with cSLE were not investigated in this review.

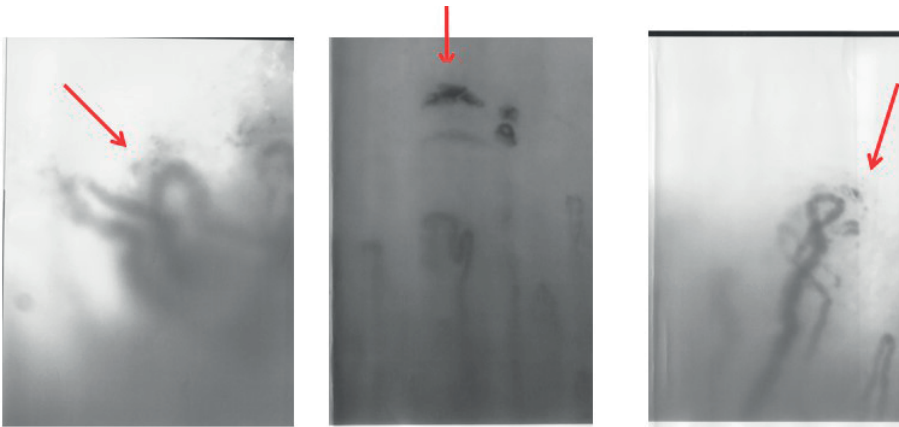


Figure 3. Our first observations of capillary hemorrhages (red arrows) in cSLE patients by light microscopy

A possible relation of these nailfold capillary hemorrhages with the observed vascular problems in SLE seems logical. Hypothetically, these capillary hemorrhages might have diagnostic and/or prognostic value or might be a biomarker to monitor disease activity. It is already known that one of the negative long-term effects in SLE is a higher risk for cardiovascular disease. Studies in adults with SLE have shown that the overall risk of cardiovascular disease is increased up to 17-fold, but even >50-fold in female patients

between 35-44 years (32). In the earlier mentioned study of Groot et al., it was shown that cardio- and cerebrovascular complications occur in 5-10% of adults with cSLE, with the majority of events occurring before the age of 40 (24). Hersh et al. found that, although the percentage of cardiovascular complications was similar between adult- and childhood-onset patient groups, the cSLE-patients were (much) younger (mean age of 32 compared to 48 years) at presentation of the myocardial infarction (22). Clearly, in these patients premature coronary-artery atherosclerosis is prevalent and occurs more frequent and at a much lower age than in healthy age-matched controls (33). This is worrisome, specifically since these studies only included survivors of cardiovascular events and most likely the numbers found on coronary artery disease is an underestimation of the real prevalence and incidence. Traditional risk factors for cardiovascular disease (such as diabetes, obesity, hypertension) do not fully explain this increased risk for cardiovascular disease in SLE-patients: non-traditional and more SLE-specific risk factors such as chronic/cumulative corticosteroid use, vascular injury, high levels of pro-inflammatory immune complexes and decreased levels of endothelial progenitor cells (EPC) for vascular repair and/or remodeling, also seem to play an important role (34). The pathophysiology of premature atherosclerosis in SLE is not yet completely

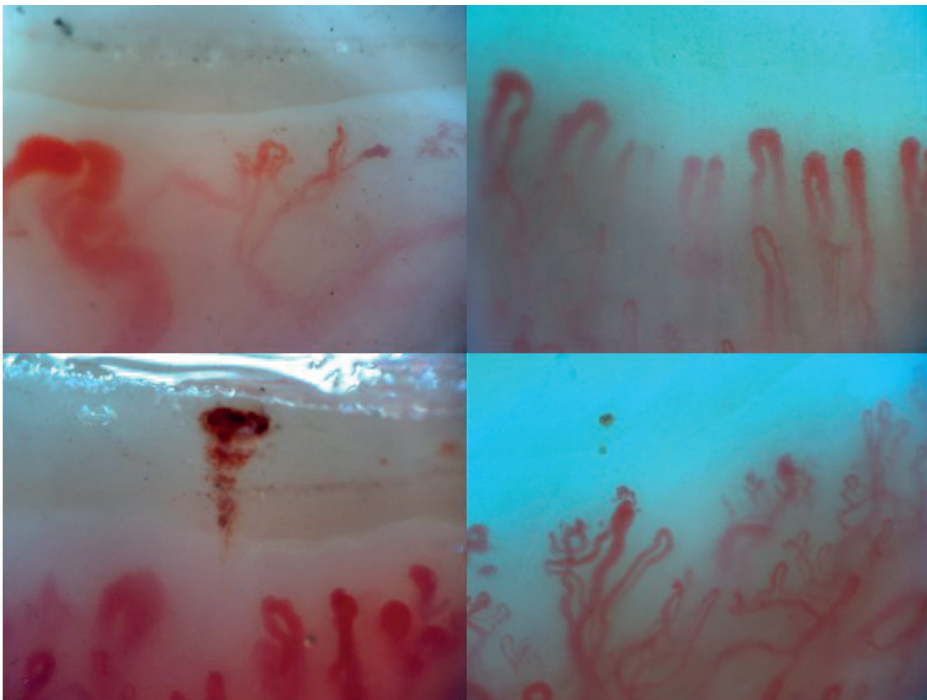


Figure 4. Nailfold capillary scleroderma pattern with: giant capillaries (upper left/right and lower left), capillary hemorrhages (lower left), avascular areas (upper left) and ramifications (lower right)

understood, but endothelial dysregulation due to endothelial cell damage combined with lower endothelial progenitor cells and continuous inflammation seem to play an important role (35, 36). Capillary abnormalities in SLE might be a mirror for the sick endothelium and may be important to detect in follow-up to monitor vascular health in SLE over time.

Besides capillary hemorrhages and abnormal capillary shapes, a so-called capillary scleroderma pattern (figure 4) has been observed in a subgroup of SLE patients (37, 38). In general, a capillary scleroderma pattern in SLE has been suggested to be a sign that these patients are at risk for of possible overlap disease and these patients should be monitored for clinical features of SSc (39). Although it was suggested to be a useful diagnostic marker, the actual value of the observation of this scleroderma pattern in SLE has not yet been thoroughly investigated longitudinally.

All of these above observations on the nailfold capillary bed of SLE patients leads to the question whether these pathological capillary changes reflect overall disease severity in SLE, are related to specific organ involvement, or show different subgroups of patients correlated with a specific type of capillary abnormalities. In short, if capillary abnormalities prove to be a biomarker of SLE, reflecting disease activity or damage, they will be important for use in clinical practice.

Interferon gene signatures and phenotyping of patients

Interferon (IFN) is produced by plasmacytoid dendritic cells, induced by (viral) infections, but also in response to immune complexes in rheumatic diseases (40). IFNs, especially type 1 IFN, play an important role in the pathogenesis of SLE (41). An IFN gene signature is a description of a group with the same upregulated genes. In SLE, a chronic and high 'type 1 IFN signature' has been associated with more severe SLE disease activity and with specific type of organ involvement such as nephritis and (muco)cutaneous involvement (42). It has been shown in cSLE that the type of organs that are involved mostly do not change after the first two years after diagnosis (24). This could mean that treatment regimens might be chosen or rapidly adapted to specific phenotypes in those first years after diagnosis.

It has also been suggested that high levels of IFN type 1 have a modulating effect on the endothelium via plaque-residing macrophages, potentiating foam cell and extracellular trap formation, inducing endothelial dysfunction and lead to exacerbated atherosclerosis outcomes in patients suffering from inflammatory diseases and risk for atherosclerosis (36). This links to the paragraph above on premature coronary-artery atherosclerosis in SLE whose pathophysiology is important to unravel further.

There is a high complexity in different types of IFN signatures in (c)SLE, but new insights point to some stratification in different disease subgroups (43, 44). Possibly, these new stratification of IFN signatures can predict (future) severity and treatment responses better than the known disease activity scores such as SLEDAI and BILAG, by defining more specific SLE subgroups. This might be very helpful in new treatment strategies to become more steroid-sparing and lower toxic side effects. This is extra important because damage in SLE is also partially related to steroid toxicity.

New biomarkers and importance for implementation in treatment choices

Newer treatments for children with SLE, such as belimumab, are now more widely available, and other new treatments are being developed. Other new treatments for this disease targeting IFN, JAK, IL-12 and IL-23 are being investigated, as well as the combined use of rituximab and belimumab as anti-B cell therapy (45). Nevertheless, it is still unknown *which patients need which therapy at which time point* to prevent irreversible disease damage. It is necessary to further define SLE subgroups that need different personalized treatment approaches. This thesis aims to provide some of many steps to the new insights that are still needed to reach this goal.

A broader spectrum of connective tissue diseases (CTD's)

Overlap disease in systemic connective tissue diseases

There is a group of patients (children and adults) that show clinical *overlap* features between the diseases SLE, systemic sclerosis (SSc) and/or dermatomyositis (DM) (see figure 5). These patients often have anti-ribonucleoprotein (RNP) antibodies combined with Raynaud's phenomenon, arthritis and/or myositis (46, 47). In the past, there has been a lot of debate whether to call this phenomenon of symptoms an 'overlap disease' or a disease entity of its own (48, 49). This overlapping group of symptoms in systemic autoimmunity has been defined as mixed connective tissue disease (MCTD), or as undifferentiated CTD (UCTD) when classification criteria are not (yet) met. For MCTD, different sets of classification criteria have been proposed, such as Sharp, Alarcon-Segovia, Kasukawa, and Kahn, of which most have good specificity but sometimes lower sensitivity (50). The existence of four sets of diagnostic criteria shows that there is still lot of debate on the best classification of this disease. UCTD is considered to be a diagnosis of exclusion when a patient with an autoimmune disease does not fulfil the criteria for other specific systemic autoimmune diseases such as SLE, SSc, DM and Sjögren's syndrome (or MCTD). Although juvenile dermatomyositis (JDM) is not considered as a lifelong disease, MCTD and UCTD are considered to be chronic and lifelong, just like SLE and SSc. Approximately one quarter of MCTD patients have a juvenile-onset (jMCTD) of the disease, i.e. onset of disease before the age of 18 (51). Treatments for MCTD or UCTD are chosen according the most severe symptoms. In contrast with SLE, MCTD patients

are in general not at risk for the development of nephritis but they do seem to be at risk for pulmonary involvement with severe vascular complications such as pulmonary hypertension (52). Recently, it was highlighted that there are still no clinical practice guidelines for MCTD patients, not only in terms of standard of care but also in consensus of classification criteria (53).

Literature on jMCTD is scarce, but a recent longitudinal study showed that during follow-up of jMCTD, SLE-like and myositis-like symptoms decreased over time while SSc-like symptoms increased. This study was a cohort of 55 patients with a mean disease duration of 16 years, and presence of rheumatoid factor (RF) was a predictor for ongoing active disease (54). These findings are in line with a study (in adults) that showed that 58% of MCTD-patients did not change their initial clinical presentation and 17% of patients evolved into SSc (48). In a large cohort of adult- and jSSc patients, it was shown that the juvenile group showed more overlap disease and relatively more musculoskeletal involvement. Looking at organ-specific damage, another longitudinal study (median 15 years after disease onset) showed that patients with jMCTD showed impaired right- and left-ventricle cardiac function compared to healthy controls, although no patients had signs of pulmonary hypertension (55). In terms of survival, children with MCTD did better than adults (55, 56).

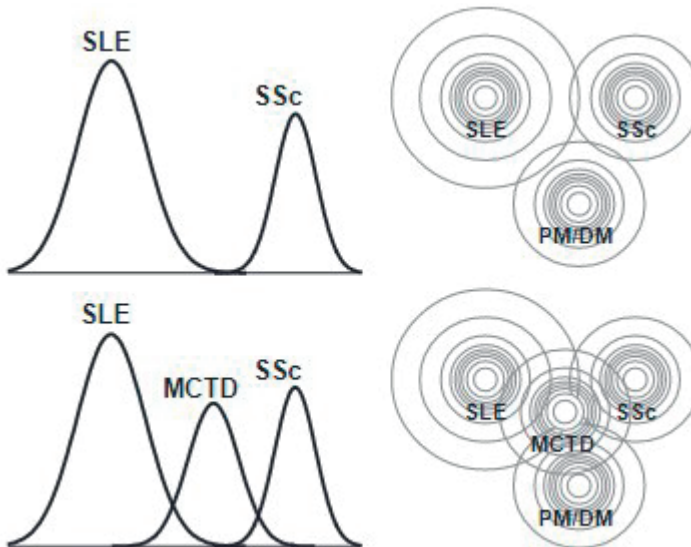


Figure 5. Visualization of overlapping disease in the CTD-spectrum (with consent from Aringer et al.(49))

It is important to have more predictive markers to know which patients are at risk for cardiac, pulmonary and kidney inflammation because of screening protocols in daily

clinical practice. The optimal frequency for urine-analysis, pulmonary function tests (for detection of restriction and interstitial lung disease) and cardiac ultrasound (to screen for pulmonary hypertension) is still unknown. In (j)MCTD/UCTD, capillary abnormalities such as a scleroderma pattern might be helpful in detecting patients with risk for SSc-like symptoms.

Furthermore, if children with MCTD, with or without RF-positivity, are more prone for musculoskeletal disease, this is important to realize in screening and treatment of these children. Their young bodies are growing with risks for growth deformities and bone or cartilage erosions. In JIA, it has been established that RF-positive patients have higher risk for erosive damage than RF-negative patients (57). It seems that MRI, as imaging modality without the radiation side effects, has a better performance of detecting bone erosions than X-ray (58). There is scarcity in studies that investigate musculoskeletal disease activity or -damage by MRI in patients with systemic autoimmune diseases.

Localized scleroderma (LS)

Whereas SSc in the young is ultra-rare, the entity of localized scleroderma (LS) , also called morphea, is a slightly more common autoimmune disease, mostly limited to the skin. LS is predominantly diagnosed in childhood with a mean age of onset at 6-8 years (59, 60). Although LS is a part of the scleroderma spectrum, it is a pathological process of *local* and chronic inflammation of a part of the skin, in contrast to the systemic involvement in SSc. LS leads to localized fibrosis and/or loss of subcutaneous tissue. The pathogenesis of this rare disease is probably multifactorial with a possible genetic predisposition combined with environmental factors leading to microvascular injury. An inflammatory cascade of pro-fibrotic cytokines is present, combined with activation T-lymphocytes and expression of vascular adhesion molecules (61-63). Subsequently, this chronic inflammation of the affected skin can lead to severe damage with growth deformities, functional disabilities and cosmetic mutilation. The incidence of LS is rare and estimated at 3.4 cases per million children per year (64, 65).

Defined subtypes in LS are linear scleroderma (majority), circumscribed morphea, generalised morphea, pan-sclerotic morphea and a mixed subtype (65, 66). Progressive hemofacial atrophy, deep morphea and bullous morphea are other subtypes (61). The affected skin areas can be anywhere on the body including the scalp and face, which is a subgroup that is called localized scleroderma 'en coup de sabre' (figure 6). New insights in the disease show that patients with LS also can have extra-cutaneous manifestations such as arthritis, uveitis, dental abnormalities and, although extremely rare, even neurological involvement in patients with LS en coup de sabre (67, 68).

LS can be mild and mono-phasic for which only topical anti-inflammatory treatment is needed, mostly prescribed by a dermatologist. Nevertheless, many patients need early and aggressive systemic treatment. This is supported by long-term studies which showed that 25-44% of the patients developed significant disability, 56% of patients had permanent damage, and 31-89% of patients had ongoing active disease in their adult life (69, 70). In this latter, more severe, subgroup of patients with LS the disease was chronic or had a course with exacerbations and remissions.



Figure 6. Localized Scleroderma en coup de sabre (with consent from Kreuter et al. The Lancet Rheum May 2022; 4 (5); E374)

Due to its rarity, there is often a diagnostic delay in LS, and even after diagnosis not all physicians are aware of the possibilities of effective *systemic* treatment options. Patients in need of systemic anti-inflammatory treatment regimens are frequently treated jointly by paediatric rheumatologists and dermatologists with experience in prescribing those treatments (71-74). According to a recent international consensus guideline, given the rarity of the disease, it is advised that children with suspected LS are referred to a specialised paediatric rheumatology centre for clinical assessment and treatment (71).

Disease activity in LS can be measured clinically by the Localized Scleroderma Cutaneous Assessment Tool (LoSCAT). LoSCAT gives measurements for severity of disease activity with Skin Severity Index (LoSSI) and provides a Skin Damage Index (LoSDI) (75, 76). LoSSI describes body surface area involvement, degree of erythema, skin thickness and appearance of new lesion or old lesion extension, each graded from 0 to 3, at 18 anatomic sites. LoSDI measures damage by a similar scoring system describing three domains: skin atrophy, subcutaneous tissue loss and hypo-hyperpigmentation, also graded from 0 to 3 (75, 76). The outcome of LoSCAT is a quantitative parameter and it describes the most severe part of the lesion. In practice, it is an adequate measurement when used by the same person and it is correlated with measurements of physician

global assessments (PGA's) (77). In LoSCAT, skin scores need to be given for the scoring items dermal atrophy, subcutaneous atrophy and skin thickness. There is risk for some variability in the inter-observer scoring of these items, especially for scores 1 and 2 in the range of 0-3 (78).

As LS can be a slowly smouldering disease, it is highly recommended to use frequent photography in longitudinal follow-up, because over the years subcutaneous fat atrophy can be slowly progressive, even in absence of skin redness. This means that disease progression can be missed with consequence risk for damage. An unmet need is the lack of objective but also accessible biomarkers that more quantitatively reflect disease activity. Currently, it is difficult to decide when to increase treatment or when it is possible and safe to taper treatment and stop. In this thesis, a pilot study will evaluate multiple imaging techniques in the search of new biomarkers in LS.

Outline of thesis

In **chapter 1** the background and aspects in diagnosis, follow-up and treatment of the autoimmune diseases cSLE, jMCTD and LS are highlighted with the current problems in treating these patients in the daily practice a pediatric rheumatologist.

Part I. Nailfold capillaroscopy in cSLE

The focus in part I is on detailed microvascular descriptions of the nailfold capillaries in cSLE and in healthy controls.

In **chapter 2** the results of a systematic literature review on nailfold capillary abnormalities in cSLE are reported and discussed.

Next, in **chapter 3** the results are shown from our cross-sectional study on capillary abnormalities in cSLE, compared to matched healthy controls.

In **chapter 4** a world-wide multicenter study compares of the capillary findings in pediatric patients with different rheumatic diseases and healthy controls.

In **chapter 5** the longitudinal data are described from our cSLE-cohort with follow-up nailfold videocapillaroscopy, specifically looking at capillary scleroderma patterns and disease damage as primary outcome.

Part II. New biomarkers in (systemic) connective tissue diseases

In part II other disease biomarkers are discussed which can be used to improve the clinical care of patients with autoimmune diseases such as cSLE, jMCTD/UCTD and LS.

In **chapter 6** subtypes of different interferon signatures are discussed and their role with neutrophil- and plasmablast signatures in stratifying cSLE patients to specific fingerprints with different (milder or severe) disease phenotypes.

In **chapter 7** unique quantitative data are described investigating skin thickness (by high frequency ultrasonography), skin hardness (by durometer) and dynamics of microcirculation (by laser speckle contrast analysis (LASCA)), in skin lesions of patients with LS, compared to contra-lateral healthy skin.

In **chapter 8** various MRI abnormalities in children with CTD are reported in relation to the clinical severity of arthritis according to their treating physicians.

Part III. Discussion, summary and appendix

In **chapter 9** the results from this thesis are discussed in a broader perspective. This leads to a recommendations on the future needs for research projects and needs in patient care for cSLE, jMCTD and LS.

In **chapter 10**, a summary is given in English and Dutch of chapter 2 until 8.

REFERENCES

1. Chakravarty EF, Bush TM, Manzi S, Clarke AE, Ward MM. Prevalence of adult systemic lupus erythematosus in California and Pennsylvania in 2000: estimates obtained using hospitalization data. *Arthritis and rheumatism*. 2007;56(6):2092-4.
2. Pons-Estel GJ, Alarcon GS, Scofield L, Reinlib L, Cooper GS. Understanding the epidemiology and progression of systemic lupus erythematosus. *Seminars in arthritis and rheumatism*. 2010;39(4):257-68.
3. Izmirlly PM, Ferucci ED, Somers EC, Wang L, Lim SS, Drenkard C, et al. Incidence rates of systemic lupus erythematosus in the USA: estimates from a meta-analysis of the Centers for Disease Control and Prevention national lupus registries. *Lupus Sci Med*. 2021;8(1).
4. Stojan G, Petri M. Epidemiology of systemic lupus erythematosus: an update. *Curr Opin Rheumatol*. 2018;30(2):144-50.
5. Pons-Estel GJ, Ugarte-Gil MF, Alarcon GS. Epidemiology of systemic lupus erythematosus. *Expert Rev Clin Immunol*. 2017;13(8):799-814.
6. Hiraki LT, Feldman CH, Liu J, Alarcon GS, Fischer MA, Winkelmayr WC, et al. Prevalence, incidence, and demographics of systemic lupus erythematosus and lupus nephritis from 2000 to 2004 among children in the US Medicaid beneficiary population. *Arthritis and rheumatism*. 2012;64(8):2669-76.
7. Mina R, Brunner HI. Update on differences between childhood-onset and adult-onset systemic lupus erythematosus. *Arthritis Res Ther*. 2013;15(4):218.
8. Martinez-Barrio J, Ovalles-Bonilla JG, Lopez-Longo FJ, Gonzalez CM, Montoro M, Valor L, et al. Juvenile, adult and late-onset systemic lupus erythematosus: a long term follow-up study from a geographic and ethnically homogeneous population. *Clinical and experimental rheumatology*. 2015;33(6):788-94.
9. Lewis MJ, Jawad AS. The effect of ethnicity and genetic ancestry on the epidemiology, clinical features and outcome of systemic lupus erythematosus. *Rheumatology (Oxford)*. 2017;56(suppl_1):i67-i77.
10. Ambrose N, Morgan TA, Galloway J, Ionnoau Y, Beresford MW, Isenberg DA, et al. Differences in disease phenotype and severity in SLE across age groups. *Lupus*. 2016;25(14):1542-50.
11. Somers EC, Marder W, Cagnoli P, Lewis EE, DeGuire P, Gordon C, et al. Population-based incidence and prevalence of systemic lupus erythematosus: the Michigan Lupus Epidemiology and Surveillance program. *Arthritis Rheumatol*. 2014;66(2):369-78.
12. Fanouriakis A, Tziolos N, Bertsias G, Boumpas DT. Update on the diagnosis and management of systemic lupus erythematosus. *Ann Rheum Dis*. 2021;80(1):14-25.
13. Hochberg MC. Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus. *Arthritis and rheumatism*. 1997;40(9):1725.
14. Petri M, Orbai AM, Alarcon GS, Gordon C, Merrill JT, Fortin PR, et al. Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis and rheumatism*. 2012;64(8):2677-86.
15. Aringer M, Costenbader K, Daikh D, Brinks R, Mosca M, Ramsey-Goldman R, et al. 2019 European League Against Rheumatism/American College of Rheumatology classification criteria for systemic lupus erythematosus. *Ann Rheum Dis*. 2019;78(9):1151-9.
16. Aringer M, Leuchten N, Johnson SR. New Criteria for Lupus. *Curr Rheumatol Rep*. 2020;22(6):18.
17. Petri M, Goldman DW, Alarcon GS, Gordon C, Merrill JT, Fortin PR, et al. Comparison of the 2019 European Alliance of Associations for Rheumatology/American College of Rheumatology

- Systemic Lupus Erythematosus Classification Criteria With Two Sets of Earlier Systemic Lupus Erythematosus Classification Criteria. *Arthritis care & research*. 2021;73(9):1231-5.
18. Batu ED, Akca UK, Kisaarslan AP, Sag E, Demir F, Demir S, et al. The Performances of the ACR 1997, SLICC 2012, and EULAR/ACR 2019 Classification Criteria in Pediatric Systemic Lupus Erythematosus. *The Journal of rheumatology*. 2021;48(6):907-14.
 19. Bombardier C, Gladman DD, Urowitz MB, Caron D, Chang CH. Derivation of the SLEDAI. A disease activity index for lupus patients. The Committee on Prognosis Studies in SLE. *Arthritis and rheumatism*. 1992;35(6):630-40.
 20. Yee CS, Farewell V, Isenberg DA, Rahman A, Teh LS, Griffiths B, et al. British Isles Lupus Assessment Group 2004 index is valid for assessment of disease activity in systemic lupus erythematosus. *Arthritis and rheumatism*. 2007;56(12):4113-9.
 21. Yee CS, Isenberg DA, Prabu A, Sokoll K, Teh LS, Rahman A, et al. BILAG-2004 index captures systemic lupus erythematosus disease activity better than SLEDAI-2000. *Ann Rheum Dis*. 2008;67(6):873-6.
 22. Hersh AO, von Scheven E, Yazdany J, Panopalis P, Trupin L, Julian L, et al. Differences in long-term disease activity and treatment of adult patients with childhood- and adult-onset systemic lupus erythematosus. *Arthritis and rheumatism*. 2009;61(1):13-20.
 23. Brunner HI, Gladman DD, Ibanez D, Urowitz MD, Silverman ED. Difference in disease features between childhood-onset and adult-onset systemic lupus erythematosus. *Arthritis and rheumatism*. 2008;58(2):556-62.
 24. Groot N, Shaikhani D, Teng YKO, de Leeuw K, Bijl M, Dolhain R, et al. Long-Term Clinical Outcomes in a Cohort of Adults With Childhood-Onset Systemic Lupus Erythematosus. *Arthritis Rheumatol*. 2019;71(2):290-301.
 25. Insfran CE, Aikawa NE, Pasoto SG, Filho DMN, Formiga FFC, Pitta AC, et al. 2019-EULAR/ACR classification criteria domains at diagnosis: predictive factors of long-term damage in systemic lupus erythematosus. *Clinical rheumatology*. 2021.
 26. Holland MJ, Beresford MW, Feldman BM, Huggins J, Norambuena X, Silva CA, et al. Measuring Disease Damage and Its Severity in Childhood-Onset Systemic Lupus Erythematosus. *Arthritis care & research*. 2018;70(11):1621-9.
 27. Groot N, Kardolus A, Bijl M, Dolhain R, Teng YKO, Zirkzee E, et al. Effects of Childhood-onset Systemic Lupus Erythematosus on Academic Achievements and Employment in Adult Life. *The Journal of rheumatology*. 2021;48(6):915-23.
 28. Tselios K, Gladman DD, Sheane BJ, Su J, Urowitz M. All-cause, cause-specific and age-specific standardised mortality ratios of patients with systemic lupus erythematosus in Ontario, Canada over 43 years (1971-2013). *Ann Rheum Dis*. 2019;78(6):802-6.
 29. Nieves CEF, Izmirly PM. Mortality in Systemic Lupus Erythematosus: an Updated Review. *Current Rheumatology Reports*. 2016;18(4).
 30. Yu CY, Kuo CF, Chou IJ, Chen JS, Lu HY, Wu CY, et al. Comorbidities of systemic lupus erythematosus prior to and following diagnosis in different age-at-onset groups. *Lupus*. 2022;9612033221100908.
 31. Cutolo M, Melsens K, Wijnant S, Ingegnoli F, Thevissen K, De Keyser F, et al. Nailfold capillaroscopy in systemic lupus erythematosus: A systematic review and critical appraisal. *Autoimmun Rev*. 2018;17(4):344-52.
 32. Manzi S, Meilahn EN, Rairie JE, Conte CG, Medsger TA, Jr., Jansen-McWilliams L, et al. Age-specific incidence rates of myocardial infarction and angina in women with systemic lupus erythematosus: comparison with the Framingham Study. *Am J Epidemiol*. 1997;145(5):408-15.
 33. Asanuma Y, Oeser A, Shintani AK, Turner E, Olsen N, Fazio S, et al. Premature coronary-artery atherosclerosis in systemic lupus erythematosus. *N Engl J Med*. 2003;349(25):2407-15.

34. Mohan S, Barsalou J, Bradley TJ, Slorach C, Reynolds JA, Hasni S, et al. Endothelial progenitor cell phenotype and function are impaired in childhood-onset systemic lupus erythematosus. *Arthritis Rheumatol.* 2015;67(8):2257-62.
35. Westerweel PE, Luyten RK, Koomans HA, Derksen RH, Verhaar MC. Premature atherosclerotic cardiovascular disease in systemic lupus erythematosus. *Arthritis and rheumatism.* 2007;56(5):1384-96.
36. Chen HJ, Tas SW, de Winther MPJ. Type-I interferons in atherosclerosis. *J Exp Med.* 2020;217(1).
37. Pavlov-Dolijanovic S, Damjanov NS, Vujasinovic Stupar NZ, Marcetic DR, Sefik-Bukilica MN, Petrovic RR. Is there a difference in systemic lupus erythematosus with and without Raynaud's phenomenon? *Rheumatol Int.* 2013;33(4):859-65.
38. Ingegnoli F. Capillaroscopy abnormalities in relation to disease activity in juvenile systemic lupus erythematosus. *Microvascular research.* 2013;87:92-4.
39. Ingegnoli F, Zeni S, Meani L, Soldi A, Lurati AF, Fantini F. Evaluation of nailfold videocapillaroscopic abnormalities in patients with systemic lupus erythematosus. *Jcr-J Clin Rheumatol.* 2005;11(6):295-8.
40. Eloranta ML, Alm GV, Ronnblom L. Disease mechanisms in rheumatology--tools and pathways: plasmacytoid dendritic cells and their role in autoimmune rheumatic diseases. *Arthritis and rheumatism.* 2013;65(4):853-63.
41. Postal M, Vivaldo JF, Fernandez-Ruiz R, Paredes JL, Appenzeller S, Niewold TB. Type I interferon in the pathogenesis of systemic lupus erythematosus. *Curr Opin Immunol.* 2020;67:87-94.
42. Oke V, Gunnarsson I, Dorschner J, Eketjall S, Zickert A, Niewold TB, et al. High levels of circulating interferons type I, type II and type III associate with distinct clinical features of active systemic lupus erythematosus. *Arthritis Res Ther.* 2019;21(1):107.
43. Wahadat MJ, Bodewes ILA, Maria NI, van Helden-Meeuwsen CG, van Dijk-Hummelman A, Steenwijk EC, et al. Type I IFN signature in childhood-onset systemic lupus erythematosus: a conspiracy of DNA- and RNA-sensing receptors? *Arthritis Res Ther.* 2018;20(1):4.
44. Chiche L, Jourde-Chiche N, Whalen E, Presnell S, Gersuk V, Dang K, et al. Modular transcriptional repertoire analyses of adults with systemic lupus erythematosus reveal distinct type I and type II interferon signatures. *Arthritis Rheumatol.* 2014;66(6):1583-95.
45. Trentin F, Zucchi D, Signorini V, Elefante E, Bortoluzzi A, Tani C. One year in review 2021: systemic lupus erythematosus. *Clinical and experimental rheumatology.* 2021;39(2):231-41.
46. Sharp GC, Irvin WS, Tan EM, Gould RG, Holman HR. Mixed connective tissue disease--an apparently distinct rheumatic disease syndrome associated with a specific antibody to an extractable nuclear antigen (ENA). *Am J Med.* 1972;52(2):148-59.
47. Kasukawa R. Mixed connective tissue disease. *Intern Med.* 1999;38(5):386-93.
48. Cappelli S, Bellando Randone S, Martinovic D, Tamas MM, Pasalic K, Allanore Y, et al. "To be or not to be," ten years after: evidence for mixed connective tissue disease as a distinct entity. *Seminars in arthritis and rheumatism.* 2012;41(4):589-98.
49. Aringer M, Steiner G, Smolen JS. Does mixed connective tissue disease exist? Yes. *Rheum Dis Clin North Am.* 2005;31(3):411-20, v.
50. John KJ, Sadiq M, George T, Gunasekaran K, Francis N, Rajadurai E, et al. Clinical and Immunological Profile of Mixed Connective Tissue Disease and a Comparison of Four Diagnostic Criteria. *Int J Rheumatol.* 2020;2020:9692030.
51. Burdt MA, Hoffman RW, Deutscher SL, Wang GS, Johnson JC, Sharp GC. Long-term outcome in mixed connective tissue disease: longitudinal clinical and serologic findings. *Arthritis and rheumatism.* 1999;42(5):899-909.

52. Sharp GC. MCTD: a concept which stood the test of time. *Lupus*. 2002;11(6):333-9.
53. Chaigne B, Scire CA, Talarico R, Alexander T, Amoura Z, Avcin T, et al. Mixed connective tissue disease: state of the art on clinical practice guidelines. *RMD Open*. 2018;4(Suppl 1):e000783.
54. Hetlevik SO, Flato B, Rygg M, Nordal EB, Brunborg C, Hetland H, et al. Long-term outcome in juvenile-onset mixed connective tissue disease: a nationwide Norwegian study. *Ann Rheum Dis*. 2017;76(1):159-65.
55. Witzczak BN, Hetlevik SO, Sanner H, Barth Z, Schwartz T, Flato B, et al. Effect on Cardiac Function of Longstanding Juvenile-onset Mixed Connective Tissue Disease: A Controlled Study. *The Journal of rheumatology*. 2019;46(7):739-47.
56. Scalapino K, Arkachaisri T, Lucas M, Fertig N, Helfrich DJ, Londino AV, Jr., et al. Childhood onset systemic sclerosis: classification, clinical and serologic features, and survival in comparison with adult onset disease. *The Journal of rheumatology*. 2006;33(5):1004-13.
57. van Rossum MA, Zwinderman AH, Boers M, Dijkmans BA, van Soesbergen RM, Fiselier TJ, et al. Radiologic features in juvenile idiopathic arthritis: a first step in the development of a standardized assessment method. *Arthritis and rheumatism*. 2003;48(2):507-15.
58. Hu L, Huang Z, Zhang X, Chan Q, Xu Y, Wang G, et al. The performance of MRI in detecting subarticular bone erosion of sacroiliac joint in patients with spondyloarthritis: a comparison with X-ray and CT. *Eur J Radiol*. 2014;83(11):2058-64.
59. Zulian F, Athreya BH, Laxer R, Nelson AM, Feitosa de Oliveira SK, Punaro MG, et al. Juvenile localized scleroderma: clinical and epidemiological features in 750 children. An international study. *Rheumatology (Oxford)*. 2006;45(5):614-20.
60. Li SC. Scleroderma in Children and Adolescents: Localized Scleroderma and Systemic Sclerosis. *Pediatr Clin North Am*. 2018;65(4):757-81.
61. Kaushik A, Mahajan R, De D, Handa S. Paediatric morphea: a holistic review. Part 1: epidemiology, aetiopathogenesis and clinical classification. *Clin Exp Dermatol*. 2020;45(6):673-8.
62. Fett N, Werth VP. Update on morphea: part I. Epidemiology, clinical presentation, and pathogenesis. *J Am Acad Dermatol*. 2011;64(2):217-28; quiz 29-30.
63. Yamane K, Ihn H, Kubo M, Yazawa N, Kikuchi K, Soma Y, et al. Increased serum levels of soluble vascular cell adhesion molecule 1 and E-selectin in patients with localized scleroderma. *J Am Acad Dermatol*. 2000;42(1 Pt 1):64-9.
64. Herrick AL, Ennis H, Bhushan M, Silman AJ, Baildam EM. Incidence of childhood linear scleroderma and systemic sclerosis in the UK and Ireland. *Arthritis care & research*. 2010;62(2):213-8.
65. Zulian F. Scleroderma in children. *Best Pract Res Clin Rheumatol*. 2017;31(4):576-95.
66. Laxer RM, Zulian F. Localized scleroderma. *Curr Opin Rheumatol*. 2006;18(6):606-13.
67. Zulian F, Vallongo C, Woo P, Russo R, Ruperto N, Harper J, et al. Localized scleroderma in childhood is not just a skin disease. *Arthritis and rheumatism*. 2005;52(9):2873-81.
68. Chiu YE, Vora S, Kwon EK, Maheshwari M. A significant proportion of children with morphea en coup de sabre and Parry-Romberg syndrome have neuroimaging findings. *Pediatric dermatology*. 2012;29(6):738-48.
69. Saxton-Daniels S, Jacobe HT. An evaluation of long-term outcomes in adults with pediatric-onset morphea. *Arch Dermatol*. 2010;146(9):1044-5.
70. Piram M, McCuaig CC, Saint-Cyr C, Marcoux D, Hatami A, Haddad E, et al. Short- and long-term outcome of linear morphea in children. *Br J Dermatol*. 2013;169(6):1265-71.
71. Zulian F, Culp R, Sperotto F, Anton J, Avcin T, Baildam EM, et al. Consensus-based recommendations for the management of juvenile localised scleroderma. *Ann Rheum Dis*. 2019;78(8):1019-24.
72. Zulian F, Tirelli F. Treatment in Juvenile Scleroderma. *Curr Rheumatol Rep*. 2020;22(8):45.

73. Li SC, O'Neil KM, Higgins GC. Morbidity and Disability in Juvenile Localized Scleroderma: The Case for Early Recognition and Systemic Immunosuppressive Treatment. *The Journal of pediatrics*. 2021.
74. Li SC, Zheng RJ. Overview of Juvenile localized scleroderma and its management. *World J Pediatr*. 2020;16(1):5-18.
75. Arkachaisri T, Vilaiyuk S, Torok KS, Medsger TA, Jr. Development and initial validation of the localized scleroderma skin damage index and physician global assessment of disease damage: a proof-of-concept study. *Rheumatology (Oxford)*. 2010;49(2):373-81.
76. Arkachaisri T, Vilaiyuk S, Li S, O'Neil KM, Pope E, Higgins GC, et al. The localized scleroderma skin severity index and physician global assessment of disease activity: a work in progress toward development of localized scleroderma outcome measures. *The Journal of rheumatology*. 2009;36(12):2819-29.
77. Kelsey CE, Torok KS. The Localized Scleroderma Cutaneous Assessment Tool: responsiveness to change In a pediatric clinical population. *J Am Acad Dermatol*. 2013;69(2):214-20.
78. Agazzi A, Fadanelli G, Vittadello F, Zulian F, Martini G. Reliability of LoSCAT score for activity and tissue damage assessment in a large cohort of patients with Juvenile Localized Scleroderma. *Pediatric rheumatology online journal*. 2018;16(1):37.



Part I



Nailfold videocapillaroscopy in cSLE

2

Capillaroscopy in childhood-onset Systemic Lupus Erythematosus (cSLE): a first systematic review

Dieneke Schonenberg-Meinema, Karin Melsens, Amara Nassar-Sheikh Rashid,
Maurizio Cutolo, Taco W. Kuijpers, J. Merlijn van den Berg, Vanessa Smith

Published in Clinical and Experimental Rheumatology. 2020 Mar-Apr;38(2):350-354

ABSTRACT

BACKGROUND: Recently, a systematic review indicated that, compared to healthy controls, adult patients with systemic lupus erythematosus (SLE) show significant more abnormal capillary morphology and hemorrhages in nailfold capillaroscopy and that these capillary changes are associated with disease activity. As yet, no systematic literature evaluation of capillaroscopy in childhood-onset SLE (cSLE) has been performed.

OBJECTIVE: To systematically review the literature on nailfold capillary characteristics in cSLE.

METHODS: Search terms “SLE or Lupus”, “Capillaroscopy” and “Juvenile or Childhood or Pediatric or Child” were used in PubMed, Embase and Web of Science. Capillary findings were evaluated according to the current international consensus-based definitions for analysis of capillaroscopic characteristics from the European League against Rheumatism (EULAR) Study Group on Microcirculation in Rheumatic Diseases (SG MC/RD).

RESULTS: After screening eighty search hits, six articles were retained, of which two case-control studies and four case series. For capillary density, no difference was found between cSLE and healthy controls (one study). Differences in capillary diameter, capillary morphology, hemorrhages and semi-quantitative score were inconclusive or non-interpretable. A scleroderma pattern was not detected in case control studies but was reported in a minority of cSLE patients in three out of four case series.

CONCLUSIONS: Literature on nailfold capillary findings in cSLE is scarce and inconclusive. To evaluate capillary characteristics in cSLE, prospective longitudinal studies are needed. Future studies should use uniform definitions for capillary characteristics and findings should be compared with healthy controls, matched for age and ethnicity. The EULAR SG MC/RD is stepping forward to this need.

INTRODUCTION

Nailfold capillaroscopy is a noninvasive magnification method to visualize the capillaries in the fingertips. Capillary changes in adult and pediatric patients with Raynaud's phenomenon (RP) are characterized by scleroderma patterns when associated with a scleroderma spectrum disease [1-5]. Notably, a scleroderma pattern is one of the 2013 classification criteria for systemic sclerosis (SSc) developed by American College of Rheumatology (ACR) and the European League Against Rheumatism (EULAR). The severity of this scleroderma pattern seems associated with (the risk of) severe organ inflammation and fibrosis in SSc [6, 7].

Recently, Cutolo et al. concluded in a systematic review on capillaroscopic characteristics in adults with systemic lupus erythematosus (SLE) that a significantly higher number of tortuous capillaries, abnormal capillary morphology and hemorrhages are detected in SLE-patients when compared to healthy controls [8]. The "semi-quantitative nailfold capillaroscopic score (NFC)", rating capillary changes from zero to three or as "normal/minor/major/severe pattern", is also higher in adult SLE-patients than in healthy controls and seems to correlate with disease activity [8]. In his review, Cutolo focused only on adult patients with SLE. He used standardized definitions for uniformity and global interpretability, from the EULAR Study Group on Microcirculation in Rheumatic Diseases (SG MC/RD), to summarize capillaroscopic literature data [7].

As capillaries from healthy children quantitatively differ from adults in terms of capillary density, -diameter, morphology and presence of microhemorrhages, capillary changes should be separately described in adult and pediatric cohorts [9-11]. Moreover, patients with childhood-onset SLE (cSLE) have been found to show a more severe disease presentation and disease course than adult-onset SLE patients [12-15]. These reports in children suggest that SLE-patients should be studied in separate cohorts for children and adults.

For that reason, the aim of our study was to systematically review the literature on nailfold capillary changes in cSLE with the standard terminology of the EULAR SG MC/RD.

METHODS

Search strategy and process

To identify all original research articles that reported on the nailfold capillaroscopic assessment of cSLE, literature was systematically searched, with the latest update on

the 2nd of April 2019, with search terms “SLE or Lupus”, “Capillaroscopy” and “Juvenile or Childhood or Pediatric or Child” in PubMed, Embase and Web of Science. The search was not restricted in publication date.

Two reviewers (DS and AN) screened the search hits for relevance by title and abstract. An overview of the search process is shown in figure 1. SLE should be diagnosed according to the international classification criteria from American College of Rheumatology (ACR) or Systemic Lupus International Collaborating Clinics (SLICC). Articles with a different topic, objective or patient population (for instance Raynaud’s phenomenon) mentioning capillaroscopy as a secondary outcome were included if they described capillaroscopic details from a sub-cohort of cSLE patients. There was no minimum count for study population. Reference lists of the retained manuscripts were checked for additional relevant articles.

Quality appraisal and level of evidence

The articles underwent quality appraisal by authors DS and MvdB, using a standardized scoring sheet from the National Institutes of Health (NIH) Quality Assessment tool for Observational Cohort and Cross Sectional Studies [16] which contains 15 questions, resulting in a score ranging between 0-15 (see supplementary file 1). In a second phase, DS and AN reached consensus on the scores. No articles were excluded based on their quality, as the aim of this review was to provide a description of literature. As such, case-control studies as well as case series were considered.

Evaluation of capillaroscopic variables

The capillaroscopic findings were evaluated by quantitative, semi-quantitative and qualitative assessment according to the parameters and definitions from the EULAR SG MC/RD [1, 17, 18].

The following quantitative capillary variables were extracted from the included studies: density (as number of capillaries per linear mm), dimension of a capillary limb (apical, afferent/efferent width or loop length in micrometer (μm), normal morphology as hairpin (stereotype hairpin shape), crossing (once or twice) and tortuous (limbs bend but do not cross) and abnormal morphology as all other shapes and lastly hemorrhages (over the top or around the whole length of the capillary) [17-19].

The reported quantitative variables that defined semi-quantitative assessments in the reviewed articles were translated by the current definitions from EULAR SG MC/RD as described above.

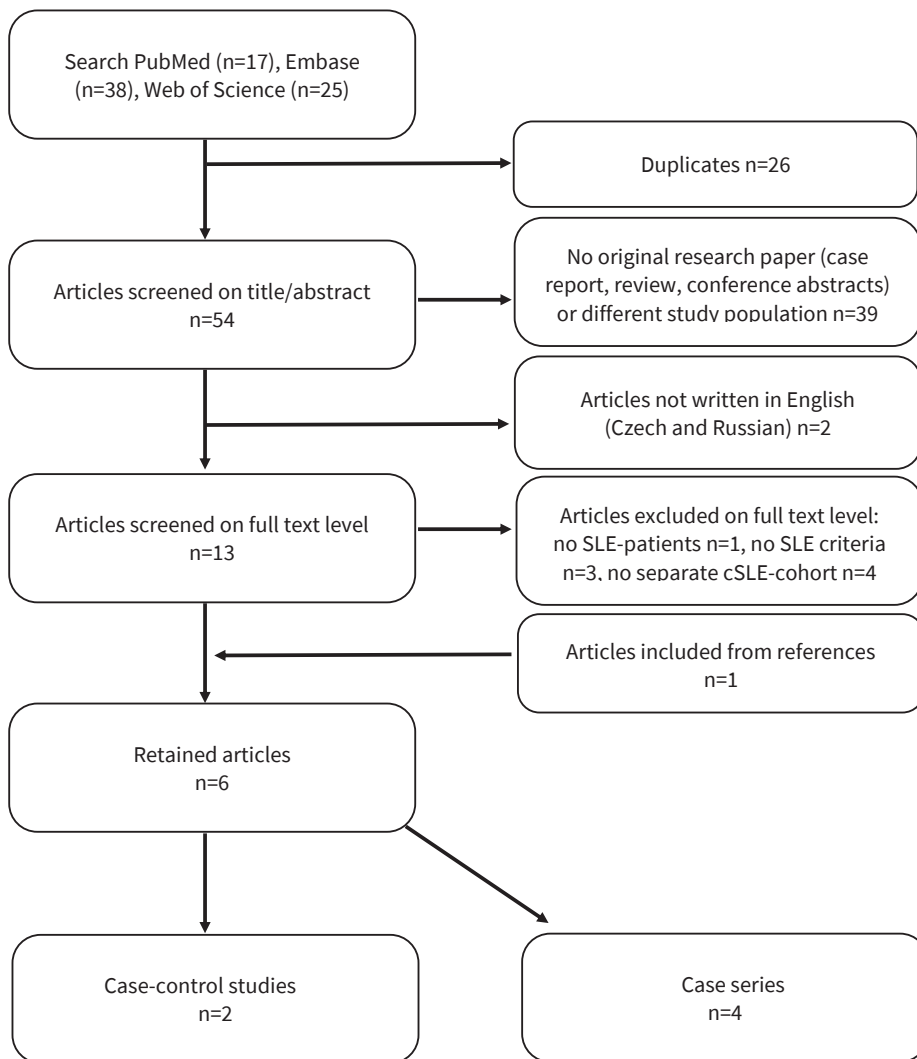


Figure 1. Flow chart search process and selection

Qualitative assessment was reported as detection of the scleroderma pattern. Scleroderma pattern was defined as by Cutolo et al. (2000) or Maricq et al. (1981) with “enlarged/giant capillaries, hemorrhages and a-vascularity” as main observations with possible detection of “ramified/bushy capillaries” as a sign of neo-angiogenesis [20, 21].

RESULTS

The literature search provided a total of eighty hits (see figure 1). Duplicates (n=26) and articles that were not an original research paper (i.e. case report, review or conference meeting, n=39) were excluded. Articles written in other languages than English, French, German, Dutch or Italian were also excluded (n=2). One extra article was found by screening references.

Subsequently, fourteen original articles remained for review on a full text level by DS, AN, KM and VS. One article, with left ventricular function as primary outcome, was included as it described a well-defined cohort of cSLE and capillary abnormalities. Another eight articles were excluded for the following reasons: no capillaroscopy performed in patients with SLE (n=1), no separate pediatric cohort of SLE patients (n=3) or SLE was not diagnosed (or this was not reported) according to the ACR or SLICC criteria (n=4).

Finally, six articles were retained for this review. An overview of the retained studies with their sample sizes, demographics, (different) used capillaroscopic techniques and magnification scales is shown in table 1.

Quality appraisal and level of evidence

Supplementary file A shows the quality appraisal of the included articles. The retained articles consisted of two case-control studies and four case series. Two articles have an NIH score $\leq 7/15$ (low to moderate quality), which concerned the two case-control studies. Reasons for these scores are the limitations of the study design (cross-sectional cohort descriptions), small sample sizes with no/limited follow-up period or the absence of details on the disease duration.

Quantitative assessment of capillaroscopy in cSLE

In supplementary file B (tables B.1-B.4), capillary characteristics are described in detail according to the quantitative assessment. An overview of significant capillary characteristics in cSLE patients compared to healthy controls is shown in table 2. Details on the quantitative assessment of nailfold capillaries were reported in only one case control study. For density per millimeter, no significant difference was observed between cSLE patients and healthy children [22]. The same research group did not find significant differences in capillary length between cSLE patients and healthy controls [22]. For capillary morphology, Ingegnoli (2005) observed that cSLE patients show significantly increased 'elongated, tortuous loops' compared to healthy controls ($p=0.002$), although the term 'tortuous' was not defined in this study [22]. The presence of microhemorrhages was not observed in the cSLE-patients from this case-control study.

Table 1. Overview of included studies (n=6): sample size and used techniques

Author	Sample size cSLE patients	Sex M/F	Average age (years)	Average disease duration at inclusion (years)	Disease specific serologic abnormalities (%)	Concomitant therapies (%)	Raynaud's phenomenon (%)	Technique	Magnification	Number of fingers
Chung et al. 2015 [23]	92	13/79	12.4	3.6	Anti-ds-DNA (75) Anti-Sm (6.5) Anti-RNP (25)	Steroids (98.9) Cyclophosphamide (51.1) Hydroxychloroquine (17.4) Azathioprine (37) Mycophenolate mofetil (3.3)	23.9	NR	200x	10
Ingegnoli et al. 2013 [25]	35	2/33	13.2	2.3	NR	NR	34.3	NVC	200x	10
Piotto et al. 2012 [27]	30	5/25	14.4	4.4	NR	NR	36.6	Optical microscopy	10x-16x	8 ^a
Ingegnoli et al. 2005 [24]	123 ^b	4/119	29.9, range 5-69	7.9	Anti-ds-DNA (77.2) Anti-Sm (20.3) Anti-RNP (26.8)	NR	54	NVC	100x-200x	10
Ingegnoli et al. 2005 [22]	34	3/31	14.5	NR	NR	NR	NR	NVC	200x	10
Spencer-Green et al. 1983 [26]	7	0/7	15.3	NR	NR	NR	NR	Stereo microscopy	25-40x	4 ^c

^a thumbs excluded ^b including adults, separate size of cSLE sub-cohort is not reported, childhood-onset SLE data are reported in percentages of capillary characteristics ^c 2nd-3rd finger bilaterally

NR=not reported, NVC = nailfold video capillaroscopy, Anti-ds-DNA = anti-double-stranded DNA antibodies, Anti-Sm = anti-Smith antibody, Anti-RNP = anti-ribonucleoprotein antibody

Table 2. Significance of capillaroscopic changes in cSLE compared to healthy age-matched controls (data from 2 case control studies)

	Density	Mean density	Significant	Non-significant	Conclusion
Quantitative evaluation	Density		0 studies	1 study [22]	No significant difference in density between cSLE and healthy controls
	Dimensions	Diameter (mean width)	0 studies	0 studies	No data
		Length	0 studies	1 study [22]	Inconclusive
	Morphology	Normal morphology	1 study [22] ^a	0 studies	Suggestive that cSLE show more 'tortuous capillaries' compared to healthy controls
		Abnormal morphology	0 studies	0 studies	No data
Hemorrhages		0 studies	0 studies	No data	
Qualitative evaluation	Semi-quantitative score		0 studies	0 studies	No data
	Other patterns		0 studies	2 studies [22, 26] ^b	"Scleroderma pattern" is not significantly more detected in cSLE patients compared to healthy controls

^a 'tortuous' on itself was not defined, though rather included in the author's definition of 'major abnormalities', defined as: "normal or decreased density", "a high presence of alterations in dimensions (>10% of longer loops or presence of enlarged loops)", presence of normal variations according to the EULAR SG MCRD (">50% tortuous") or presence of abnormal shapes ("meandering or branched") or "presence of hemorrhages."^b according to Maricq (1981) or Cutolo (2000)

Semi-quantitative assessment of capillaroscopy in cSLE

Compared to healthy controls, a significantly increased detection of ‘major morphologic loop abnormalities’ was reported by Ingegnoli. It remained unclear if the authors assessed this capillary characteristic semi-quantitatively or quantitatively, since they specified the finding further as the presence of ‘elongated and tortuous capillaries’ (see paragraph 3.2) [22].

In case series of (c)SLE patients, the reported percentages of ‘major capillary abnormalities’ varied from 23 to 36% (see supplementary file C). These percentages are interpreted with caution as in these studies the term ‘tortuous’ was part of the definition for ‘major capillary abnormalities’, but was not further defined [23-25].

Qualitative assessment of capillaroscopy in cSLE

Both Spencer-Green (1983) and Ingegnoli (2005) did not detect a scleroderma pattern in cSLE (including follow-up) nor in healthy controls (see supplementary file D) [22, 26]. A scleroderma pattern was detected in 2.7% (n= 5/188) of the patients from three case series (see supplementary file D) [24, 25, 27].

DISCUSSION

This systematic review shows that there is a major lack of studies of capillaroscopy in cSLE, especially in the current light of the standardized definition for abnormal capillary morphology from the consensus of the EULAR SG MC/RD [17, 18]. After screening, only two-case control studies and four case series were retained. The main reasons for excluding articles were the lack of reporting the definition for diagnosing SLE or the fact that the pediatric patient cohort was not separately described.

Based on the results of our systematic review, we cannot make any solid conclusions on the nailfold capillary changes that might be present in cSLE-patients. As in adults with SLE, the capillary density of cSLE patients seems to be preserved. Unfortunately, because of the limited number of studies, we cannot draw any conclusions on the dimension of capillaries and the presence of hemorrhages. The observation from Ingegnoli that cSLE-patients have significantly more ‘major morphologic loop abnormalities’ compared to healthy controls could not be adopted in our conclusion because it was unknown if the authors meant ‘tortuous’ as being a variant of normal morphology, such as described by Andrade and adopted with the EULAR SG MC/RD or ‘tortuous’ as abnormal morphology (15,16). The latter being capillaries that also have other shapes, such as multiple crossing, branching or a non-convex tip. Using the same standardized definition for abnormal

capillary morphology, as proposed by EULAR SG MC/RD, will simplify the comparison and interpretation of data between future studies that are ongoing in this field worldwide. From literature, it seems that tortuous capillary morphology, as described by the EULAR SG MC/RD, is a frequent capillaroscopic finding, also in the healthy pediatric/adolescent population and as such considered a-specific and as a normal morphological variation of the nailfold capillary [10]. On the other hand, if tortuous loops form the majority of the capillaries (>50% of the capillaries) this seems to be more characteristic for SLE in adults [8, 22].

Compared to healthy controls, adult SLE-patients show significantly more abnormal capillary morphology as well as a higher semi-quantitative score, as was shown by Cutolo in his review from 40 studies on adults with SLE [8]. From our systematic review of the pediatric literature, it is still unknown if these findings could also apply to cSLE-patients.

Whereas it was not detected in case-control studies, a scleroderma pattern was detected in a minority of cSLE-patients from case series [24, 25, 27]. Although definitions for a scleroderma pattern differed in these case series, the described definitions seem to match our pre-defined criteria for a scleroderma pattern according to Maricq and Cutolo [20, 21] (see supplementary file D). A scleroderma pattern in a minor percentage of SLE patients was also reported in the review of capillaroscopy findings in adults with SLE [8]. Future studies, explicitly longitudinal data, will further elucidate if the (pathological) detection of a scleroderma pattern in (c)SLE-patients might be correlated with a specific clinical (overlap) phenotype.

Noteworthy, one case control study indicates that capillaries from healthy children show differences according to age. Density increased progressively in relation to age although this trend was not significant ($r=0.226$, $p=0.15$) but capillary length differed significantly according to age ($r=0.485$, $p=0.001$) [22]. These findings emphasize that capillaroscopic abnormalities in SLE-patients should be studied in separate cohorts for adults and children. Even the transition period from child to adult should be considered as a separate age category in studies. Besides age, ethnicity should also be investigated as a confounder in capillary characteristics, definitely in SLE, as it is very well known that ethnicity has a major impact on severity of the disease [10, 28].

CONCLUSION

Studies on nailfold capillary findings in cSLE are scarce, have small sample sizes and are inconclusive. In order to better evaluate capillary findings in (c)SLE, prospective, longitudinal studies are needed that use uniform definitions. Results from these (larger) cohort-studies should be compared with healthy controls, matched for age and ethnicity.

Supplementary file A: Quality appraisal (n=6)

	Chung et al. 2015 [23]	Ingegnoli et al. 2013 [25]	Piotto et al. 2012 [27]	Ingegnoli et al. 2005 [24]	Ingegnoli et al. 2005 [22]	Spencer-Green et al. 1983 [26]
1. Was the research question or objective in this paper clearly stated?	Yes	Yes	Yes	Yes	Yes	Yes
2. Was the study population clearly specified and defined?	Yes	Yes	Yes	Yes***	Yes	Yes
3. Was the participation rate of eligible persons at least 50%?	NR	NR	NR	Yes	NR	NR
4a. Were all the subjects selected or recruited from the same or similar populations (including the same time period)? ^d	Yes	Yes	Yes	Yes	Yes	Yes
4b. Were inclusion and exclusion criteria for being in the study pre-specified and applied uniformly to all participants? ^d	Yes	Yes	Yes	Yes	Yes	Yes
5. Was a sample size justification, power description, or variance and effect estimates provided?	NR	NR	NR	NR	NR	NR
6. For the analyses in this paper, were the exposure(s) of interest measured prior to the outcome(s) being measured?	Yes	Yes	Yes	Yes	No	No
7. Was the timeframe sufficient so that one could reasonably expect to see an association between exposure and outcome if it existed?	Yes	NR	NA	NR	Yes	No
8. For exposures that can vary in amount or level, did the study examine different levels of the exposure as related to the outcome (e.g., categories of exposure, or exposure measured as continuous variable)?	Yes	Yes	Yes	Yes	No	No
9. Were the exposure measures (independent variables) clearly defined, valid, reliable, and implemented consistently across all study participants?	Yes	NR	Yes	Yes	NR	No
10. Was the exposure(s) assessed more than once over time?	NR	Yes	No	NR	NR	No
11. Were the outcome measures (dependent variables) clearly defined, valid, reliable, and implemented consistently across all study participants?	No	Yes	Yes	Yes	Yes	Yes
12. Were the outcome assessors blinded to the exposure status of participants?	NR	NR	NR	Yes	Yes	Yes
13. Was loss to follow-up after baseline 20% or less?	NR	NR	NR	NR	NR	NR
14. Were key potential confounding variables measured and adjusted statistically for their impact on the relationship between exposure(s) and outcome(s)?	Yes	NR	NR	Yes	No	No
Total score **	9	8	8	11	7	6

<http://www.nhlbi.nih.gov/health-pro/guidelines/in-develop/cardiovascular-risk-reduction/tools/cohort>.

NR = not reported; this item was not reported in the manuscript. NA = not applicable. No = item was reported, answer was No. ** NR, NA, No's were scored as zero. Yes equals 1 point. *** including adults, separate size of cSLE sub-cohort is not reported

Supplementary file B: Summary quantitative capillary characteristics**Table B.1** Capillary density

In all tables, the results from case-control studies and case series without healthy controls are separated by a bolder black line.

Author	Study type	Sample size	Description of capillaroscopic characteristic	Result in SLE patients	Result in healthy subjects	Significance
Ingegnoli et al. 2005 [22]	Cross-sectional and prospective ^a	34 cSLE 50 healthy children 20 healthy adults	Mean density per mm (range) ^b	No exact results given	Healthy children 6.1 (5-8) Healthy adults 7.3 (6-8)	Not significant (p-value not mentioned)

^a one follow-up in a selected sub-cohort. ^b normal: 6-8 capillaries per mm.

Table B.2 Capillary dimensions

Author	Study type	Sample size	Description of capillaroscopic characteristic	Result in SLE patients	Result in healthy subjects	Significance
Ingegnoli et al. 2005 [22]	Cross-sectional and prospective ^a	34 cSLE 50 healthy children 20 healthy adults	Length in μm (range) ^b	No exact results given	Healthy children 341.04 (159-500) Healthy adults 442.77 (359-500)	No significant difference in capillary length between HC and cSLE (p-value not mentioned)
Piotto et al. 2012 [27]	Cross-sectional	30 cSLE	Capillary elongation and dilatation ^c	No elongated capillaries No exact results given on dilatations	Not applicable	Not applicable

^a one follow-up in a selected sub-cohort. ^b normal length 200-500 μm , elongated defined as >10% longer than normal. ^c according to Andrade 1990 [29]: dilated when widened in afferent, transitional and efferent branches with calibers ranging from 4-9 times normal dimension. Giant loops if calibers >10 greater than those of normal adjacent loops.

Table B.3 Capillary morphology

Author	Study type	Sample size	Description of capillaroscopic characteristic	Result in SLE patients	Result in healthy subjects	Significance
Ingegnoli et al. 2005 [22]	Cross-sectional and prospective ^a	34 cSLE 50 healthy children 20 healthy adults	Tortuous, meandering and branched loops ^b	No exact results given	No exact results given	Significantly increased elongated, tortuous loops in SLE (p=0.0002)
Protto et al. 2012 [27]	Cross-sectional	30 cSLE	Crossed, bushy and bizarre capillaries ^c	No tortuous and crossed capillaries	Not applicable	Not applicable

^a one follow-up in a selected sub-cohort. ^b "tortuous" is not defined. ^c defined according to Andrade 1990 [29]: "atypical capillaries, such as crossed, bushy and bizarre capillaries".

Table B.4 Capillary hemorrhages

Author	Study type	Sample size	Description of capillaroscopic characteristic	Result in SLE patients	Result in healthy subjects	Significance
Ingegnoli et al. 2005 [22]	Cross-sectional and prospective ^a	34 cSLE 50 healthy children 20 healthy adults	Presence of pathologic hemorrhages ^b	No exact results given	10% (7/70 total healthy controls including adults) ^c	Not stated

^a one follow-up in a selected sub-cohort. ^b defined as: "over the top or around the whole length of capillary". ^c extra note reported in text: "microhemorrhages grouped within limited areas", though 'microhemorrhages is not defined.

Supplementary file C: Summary semi-quantitative analysis

Author	Study type	Sample size	Description of capillaroscopic characteristic	Result in SLE patients	Result in healthy subjects	Significance
Ingegnoli et al. 2005 [22]	Cross-sectional and prospective ^a	34 cSLE 50 healthy children 20 healthy adults	Normal Minor abnormalities Major abnormalities ^b	Approximately 30% Approximately 30-40% Approximately 25% ^c	28/70 (incl adults) 39/70 3/70	Significantly increased major morphologic loop abnormalities (p=0.0002) ^d
Chung et al. 2015 [23]	Cross-sectional capillaroscopy	92 cSLE 50 control (for echocardiography only)	Normal Major abnormalities ^b	No exact results given 32/92 (34.8%)	Not stated	Not applicable
Ingegnoli et al. 2013 [25]	Cross-sectional with minimal 6 months follow-up ^e	35 cSLE 27 adult onset SLE	Normal Minor abnormalities ^f Major abnormalities ^g	12/35 (adult-onset 7/27) 14/35 (adult-onset 10/27) 8/35 (adult-onset 9/27)	Not applicable	No significant differences in pattern between childhood-onset and adult-onset SLE patients (p-value not mentioned)
Ingegnoli et al. 2005 [24]	Retrospective	123 SLE ^h	Normal Minor abnormalities Major abnormalities ^b	35/123 41/123 44/123	Not applicable	Not applicable

^a one follow-up in a selected sub-cohort. ^b according to Ingegnoli 2013 [25] defined as: "normal or decreased density", "a high presence of alterations in dimensions (>10% of longer loops or presence of enlarged loops)", presence of normal variations according to the EULAR SG MCRD (">50% tortuous") or presence of abnormal shapes ("meandering or branched") or "presence of hemorrhages"; ^c detailed numbers not mentioned; estimated data from figure. ^d defined as: "characterized by elongated, tortuous loops, in SLE"; ^e no details on follow-up period. ^f defined as: "density 6-8/mm, elongated <10%, tortuous <50%, parallel rows, no pathologic hemorrhages".

^g defined as: "density ≤6-8/mm, elongated >10%, tortuous > 50%, enlarged, meandering, branched, disarrangement, +/- pathological hemorrhages"; ^h size of cSLE sub-cohort not reported.

Nailfold videocapillaroscopy in cSLE

Supplementary file D: Summary qualitative scoring patterns

Author	Study type	Sample size	Description of capillaroscopic characteristic	Result in SLE patients	Result in healthy subjects	Significance
Ingegnoli et al. 2005 [22]	Cross-sectional and prospective ^a	34 cSLE 50 healthy children 20 healthy adults	Scleroderma pattern ^b	0/34 ^c	0/70	No difference
Spencer-Green et al. 1983 [26]	Cross-sectional	7 cSLE 34 healthy children	Normal ^d Abnormal pattern ^e	7/7 0/7	34/34 0/34	No difference
Ingegnoli et al. 2013 [25]	Cross-sectional with minimal 6 months follow-up ^f	35 cSLE 27 adult onset SLE	Scleroderma pattern ^b	1/35 1/27	Not applicable	No significant differences in pattern between childhood-onset and adult-onset SLE patients ^g
Piotto et al. 2012 [27]	Cross-sectional	30 cSLE	Normal ^g Unspecific microangiopathy ⁱ Scleroderma pattern ^j	28/30 1/30 1/30	Not applicable	Not applicable
Ingegnoli et al. 2005 [24]	Retrospective	123 SLE ^k	Scleroderma pattern ^b	3/123	Not applicable	Not applicable

^a one follow-up in a selected sub-cohort. ^b defined as: "density <6-8/mm, elongated >10%, tortuous, enlarged, meandering, branched, disarrangement with or without vascular areas, ++ pathological hemorrhages"; ^c none of the cSLE patients developed a scleroderma pattern during follow-up period. ^d defined as: "uniform, homogeneous distribution and appearance of loops"; ^e defined as: "large, dilated loops with capillary a-vascularity and non-uniform appearance"; ^f no details on follow-up period ^g p-value not mentioned. ^h defined as: "presence of parallel non-dysmorphic capillaries and lack of deletion areas"; ⁱ defined as: "dilated or giant capillaries and other morphological changes in absence of deletion areas"; Deletion areas defined as: "absence of two or more successive capillaries"; ^j defined as: "dilated or giant capillary loops and vascular deletion areas"; Deletion areas defined as: "absence of two or more successive capillaries"; ^k including adults, the size of cSLE sub-cohort is not reported.

REFERENCES

1. Cutolo, M., Atlas of capillaroscopy in rheumatic diseases. 2010, Milan: Elsevier.
2. Maricq, H.R., et al., Diagnostic potential of in vivo capillary microscopy in scleroderma and related disorders. *Arthritis Rheum*, 1980. 23(2): p. 183-9.
3. Maricq, H.R., et al., Microvascular abnormalities as possible predictors of disease subsets in Raynaud phenomenon and early connective tissue disease. *Clin Exp Rheumatol*, 1983. 1(3): p. 195-205.
4. Koenig, M., et al., Autoantibodies and microvascular damage are independent predictive factors for the progression of Raynaud's phenomenon to systemic sclerosis: a twenty-year prospective study of 586 patients, with validation of proposed criteria for early systemic sclerosis. *Arthritis Rheum*, 2008. 58(12): p. 3902-12.
5. Pain, C.E., et al., Raynaud's syndrome in children: systematic review and development of recommendations for assessment and monitoring. *Clin Exp Rheumatol*, 2016. 34 Suppl 100(5): p. 200-206.
6. Smith, V., et al., Nailfold capillaroscopy for prediction of novel future severe organ involvement in systemic sclerosis. *J Rheumatol*, 2013. 40(12): p. 2023-8.
7. Pizzorni, C., et al., Nailfold capillaroscopic parameters and skin telangiectasia patterns in patients with systemic sclerosis. *Microvasc Res*, 2016. 111: p. 20-24.
8. Cutolo, M., et al., Nailfold capillaroscopy in systemic lupus erythematosus: A systematic review and critical appraisal. *Autoimmun Rev*, 2018.
9. Piotto, D.P., et al., Nailfold videocapillaroscopy in healthy children and adolescents: description of normal patterns. *Clin Exp Rheumatol*, 2016. 34 Suppl 100(5): p. 193-199.
10. Terreri, M.T., et al., Nail fold capillaroscopy: normal findings in children and adolescents. *Semin Arthritis Rheum*, 1999. 29(1): p. 36-42.
11. Gerhold, K. and M.O. Becker, Nailfold capillaroscopy in juvenile rheumatic diseases: known measures, patterns and indications. *Clin Exp Rheumatol*, 2014. 32(6 Suppl 86): p. S-183-8.
12. Ambrose, N., et al., Differences in disease phenotype and severity in SLE across age groups. *Lupus*, 2016. 25(14): p. 1542-1550.
13. Choi, J.H., et al., Comparison of clinical and serological differences among juvenile-, adult-, and late-onset systemic lupus erythematosus in Korean patients. *Lupus*, 2015. 24(12): p. 1342-9.
14. Martinez-Barrio, J., et al., Juvenile, adult and late-onset systemic lupus erythematosus: a long term follow-up study from a geographic and ethnically homogeneous population. *Clin Exp Rheumatol*, 2015. 33(6): p. 788-94.
15. Fatemi, A., M. Matinfar, and A. Smiley, Childhood versus adult-onset systemic lupus erythematosus: long-term outcome and predictors of mortality. *Clin Rheumatol*, 2017. 36(2): p. 343-350.
16. Institute, H.N. Quality Assessment Tool for Observational Cohort and Cross-Sectional Studies. [cited 2018 12th march 2018]; Available from: <https://www.nhlbi.nih.gov/health-pro/guidelines/in-develop/cardiovascular-risk-reduction/tools/cohort>.
17. Smith, V., et al., An EULAR study group pilot study on reliability of simple capillaroscopic definitions to describe capillary morphology in rheumatic diseases. *Rheumatology (Oxford)*, 2016. 55(5): p. 883-90.
18. Cutolo, M., et al., Reliability of simple capillaroscopic definitions in describing capillary morphology in rheumatic diseases. *Rheumatology (Oxford)*, 2018. 57(4): p. 757-759.

19. Smith, V., et al., Reliability of the qualitative and semiquantitative nailfold videocapillaroscopy assessment in a systemic sclerosis cohort: a two-centre study. *Ann Rheum Dis*, 2010. 69(6): p. 1092-6.
20. Cutolo, M., et al., Nailfold videocapillaroscopy assessment of microvascular damage in systemic sclerosis. *J Rheumatol*, 2000. 27(1): p. 155-60.
21. Maricq, H.R., Wide-field capillary microscopy. *Arthritis Rheum*, 1981. 24(9): p. 1159-65.
22. Ingegnoli, F., et al., Capillaroscopic observations in childhood rheumatic diseases and healthy controls. *Clin Exp Rheumatol*, 2005. 23(6): p. 905-11.
23. Chung, H.T., et al., Subclinical deterioration of left ventricular function in patients with juvenile-onset systemic lupus erythematosus. *Lupus*, 2015. 24(3): p. 263-72.
24. Ingegnoli, F., et al., Evaluation of nailfold videocapillaroscopic abnormalities in patients with systemic lupus erythematosus. *Jcr-Journal of Clinical Rheumatology*, 2005. 11(6): p. 295-298.
25. Ingegnoli, F., Capillaroscopy abnormalities in relation to disease activity in juvenile systemic lupus erythematosus. *Microvasc Res*, 2013. 87: p. 92-4.
26. Spencer-Green, G., et al., Nailfold capillary abnormalities in childhood rheumatic diseases. *J Pediatr*, 1983. 102(3): p. 341-6.
27. Piotto, D.G., et al., Nailfold capillaroscopy in children and adolescents with rheumatic diseases. *Rev Bras Reumatol*, 2012. 52(5): p. 722-32.
28. Pons-Estel, G.J., et al., Understanding the epidemiology and progression of systemic lupus erythematosus. *Semin Arthritis Rheum*, 2010. 39(4): p. 257-68.
29. Andrade, L.E., et al., Panoramic nailfold capillaroscopy: a new reading method and normal range. *Semin Arthritis Rheum*, 1990. 20(1): p. 21-31.



3

Nailfold capillary abnormalities in childhood-onset systemic lupus erythematosus: a cross-sectional study compared with healthy controls

Dieneke Schonenberg-Meinema, Sandy C. Bergkamp, Amara Nassar-Sheikh Rashid, Leontien B. van der Aa, Godelieve J. de Bree, Rebecca ten Cate, Maurizio Cutolo, A. Elisabeth Hak, Petra C.E. Hissink Muller, Marieke van Onna, Taco W. Kuijpers, Vanessa Smith, J. Merlijn van den Berg

Published in Lupus. 2021 Apr;30(5):818-827

ABSTRACT

OBJECTIVES: For selection of high-risk systemic lupus erythematosus (SLE) patients it is necessary to obtain indicators of disease severity that predict disease damage. As in systemic sclerosis, nailfold capillary abnormalities could be such a biomarker in SLE. The primary objective of this cross-sectional study is to describe capillary abnormalities in childhood-onset SLE (cSLE) cohort (onset < 18 years) and compare them with matched healthy controls. The secondary objective is to correlate the observed capillary abnormalities with demographical variables in both cohorts and with disease-specific variables in cSLE patients.

METHODS: Healthy controls were matched for ethnic background, age and gender. Videocapillaroscopy was performed in eight fingers with 2-4 images per finger. Quantitative and qualitative assessments of nailfold capillaroscopy images were performed according to the definitions of the EULAR study group on microcirculation in Rheumatic Diseases.

RESULTS: Both groups (n=41 cSLE-patients and n=41 healthy controls) were comparable for ethnic background (p=0.317). Counted per mm, cSLE-patients showed significantly more 'giants' (p=0.032), 'abnormal capillary shapes' (p=0.003), 'large capillary hemorrhages' (p<0.001) and 'pericapillary extravasations' (p<0.001). Combined 'abnormal capillary shapes and pericapillary extravasations' (in the same finger) were detected in 78% (32/41 patients). By qualitative analysis, 'microangiopathy' was detected in 68.3% (28/41) and a 'scleroderma pattern' in 17.1% (7/41) of the cSLE-patients (without scleroderma symptoms). The difference of percentage positive anti-RNP antibodies in the group with or without a scleroderma pattern was not significant (p=0.089). The number of 'abnormal capillary shapes per mm' was significantly correlated with treatment-naivety. The number of 'large pathological hemorrhages per mm' was significantly correlated with SLEDAI score and presence of nephritis. Compared to healthy controls, 'pericapillary extravasations' were found in significantly higher numbers per mm (p<0.001) as well as in percentage of patients (p<0.001).

CONCLUSIONS: Our observations confirm that giants, abnormal capillary morphology and capillary hemorrhages are also observed in cSLE, as was already known for adults with SLE. Number of capillary hemorrhages in cSLE was significantly correlated with disease activity. A high frequency and total amount of "pericapillary extravasations" was observed in cSLE patients, possibly revealing a new subtype of capillary hemorrhage that might reflect endothelial damage in these pediatric patients.

INTRODUCTION

Nailfold capillaroscopy (NFC), a non-invasive magnification method, is used to visualize the capillaries of the fingertips. NFC is a diagnostic instrument, used in patients with Raynaud's phenomenon: a capillary scleroderma pattern is associated with systemic sclerosis (SSc) [1-3].

Systemic Lupus Erythematosus (SLE) patients can also show capillary abnormalities in NFC. As concluded in a recent review, adults with SLE show a significantly higher number of tortuous capillaries, abnormal capillary morphology, hemorrhages and "semi-quantitative NFC score", when compared to healthy controls [4]. Additionally, the NFC-score (by rating severity of capillary changes) also seems to correlate with disease activity [4]. Studies on nailfold capillary findings in children with SLE are scarce and inconclusive. In our recently published systematic review, data from six published studies on this topic were not comparable as different definitions for abnormal morphology were used [5]. Moreover, the definition for abnormal capillary morphology was recently further specified and revised by the European League Against Rheumatism (EULAR) Study Group on Microcirculation in Rheumatic Diseases (SG MCRD) [6, 7].

The diagnosis of childhood-onset (c)SLE is often delayed due to heterogeneity of presenting symptoms, and is dependent on recognition by and experience of the clinical physician. To prevent organ damage, it is important to prevent delay in diagnosis. Delay in diagnosis is specifically mentioned as one of the patients' unmet needs in a recent publication of 'state of the art on clinical guidelines' [8]. Prevention of delay in diagnosis is especially important for cSLE-patients, because it was shown that they have more severe symptoms at presentation and a more severe disease course compared to patients with adult onset SLE [9-13]. Heterogeneity is not only applicable for disease symptoms but also for disease severity with mild to severely affected patients and, depending on type of organ manifestations, a higher risk of mortality [12]. SLE is associated with progressive (irreversible) organ damage, which has shown to be a predictor of additional morbidity and early mortality [14]. A recent international recommendation for treatment in SLE is based on the treat-to-target principle: 'since damage predicts subsequent damage and death, prevention of damage accrual should be a major therapeutic goal in SLE' [15]. Steroid-related damage is an important factor in SLE and has become an outcome parameter for damage in long-term SLE follow-up studies [16, 17]. Selection of patients who need aggressive and steroid-sparing treatment in early phases of the disease will lead to less organ damage and lower cumulative steroid-use. For selection of high-risk patients it is necessary to obtain indicators of disease severity that predict (severe) future disease damage. Nailfold capillary abnormalities could be such an in-

indicator or biomarker in SLE. For systemic sclerosis (SSc), multiple studies have shown that capillary abnormalities (by qualitative description) can be of use as a prognostic biomarker [18-22].

This study was conducted by the EULAR SG MCRD. The primary objective of this cross-sectional study is to describe possible capillary abnormalities in cSLE patients and compare them with healthy controls, matched for skin pigmentation, age and gender. These demographic variables have been described as confounding factors in healthy controls in interpreting capillary characteristics, such as density [23, 24]. The secondary objective is to correlate the observed capillary abnormalities with demographical variables in both cohorts and with disease-specific variables in cSLE patients.

METHODS

Patients and controls

Consecutive patients with (suspected) cSLE were cross-sectional included during a visit at the (outpatient) clinic. Criteria for inclusion were SLE diagnosis according to the 2012 Systemic Lupus International Collaborating Clinics (SLICC) classification criteria [25] and age of disease onset < 18 years old. Patients were excluded if they did not fulfil a minimum of four SLICC criteria, if they declined capillaroscopy examination/analysis of their capillaroscopy images, if it was impossible to collect images with good quality (due to nailfold skin thickness) or when a patient was too sick to undergo capillaroscopy examination. Demographical and clinical data were collected from patient charts. For cSLE-patients with one-time cross-sectional capillaroscopy, informed consent was waived by our ethical committee. Nevertheless, most cSLE-patients were part of a longitudinal cohort study for which an informed consent by patients (from 12 years of age) and/or both parents (for patients below 16 years) was signed. If capillaroscopy was performed longitudinally, images from the first capillaroscopy were used.

For healthy controls, children and adolescents from schools around the Amsterdam University Medical Centers (AUMC) and via personal contacts of the authors were approached for one-time capillaroscopy. This project was approved, combined with our longitudinal cohort study (Dutch trial register registration no. NL60885.018.17) by the ethical committee from the AUMC. Inclusion followed if they did not suffer from a chronic disease and had signed informed consent (child from 12 years of age and/or both parents for children below 16 years old). Age, gender, ethnic background, Raynaud symptoms and periungual trauma were noted. Disease activity was measured by Sys-

temic Lupus Erythematosus Activity Index (SLEDAI) score. Patients and healthy controls were coded with a unique study number.

Nailfold capillaroscopy technique and image collection

NVC was performed with a x200 magnification lens from Optilia. All images were collected by one investigator (DS). The patients/healthy children stayed in a room of 20-22°C for a minimum of 15-20 minutes. During capillaroscopy they were in sitting position with the hand on a table at the level of their heart. A drop of oil was applied to the fingers before examination. In total, eight fingers per cSLE-patient (excluding the thumbs) were examined. Per finger, four images were stored. From November 2017 until June 2018, a cohort of healthy children were included. In healthy children, eight fingers were examined and two images per finger were stored (according to the EULAR SG MRCD study protocol). From this larger healthy pediatric study cohort, healthy controls were matched with our cohort of cSLE-patients according to ethnic background, age and gender (in that order).

Image analysis

Post-examination, the following quantitative capillaroscopy characteristics were evaluated by primary investigator (DS) with a grid per millimeter: density (number of capillaries in distal row per mm), number of abnormal shapes (as defined by EULAR SG MCRD as all other shapes than hairpin (stereotype hairpin shape), crossing (once or twice) and tortuous (limbs bend but do not cross)) [6, 7], number of giant capillaries (if apical diameter >50 µm), maximum apical diameter (in µm, by Optipix software version 1.7.6), and number of capillary hemorrhages [3]. Hemorrhages were defined in two subtypes: 'large pathological hemorrhages' as large deposit of hemosiderin with a cap-like appearance [1] and 'pericapillary extravasations' as small point-shaped hemorrhages surrounding the capillary loop (image 1). Examined subjects were asked for finger trauma and manicure treatment in the 2-3 weeks prior to examination.

Qualitatively, three capillary patterns were described. A scleroderma pattern was defined by presence of giant capillaries, possibly combined with large pathological capillary hemorrhages, loss of capillaries and abnormal capillary shapes, according to the 'Fast Track Algorithm' [26]. If the observed capillary pattern showed abnormal capillary morphology or hemorrhages, but did not match the criteria for scleroderma, it was called 'microangiopathy', referring to non-specific abnormalities. A normal pattern showed no capillary abnormalities.

Capillaries from images of low visible quality were excluded and not analyzed.

Statistical analysis

Statistical analysis was performed with IBM SPSS Statistics type 26. Descriptive statistics were reported in terms of percentages, means and standard deviations or medians and inter-quartile ranges depending on distribution of outcome data. Demographical differences between both study groups were calculated with a paired t-test (in case of normal data distribution), McNemar test (for binary and nominal outcome variables) and Wilcoxon signed rank test (in case of no normal data distribution). Linear regression by ANOVA and logistic regression were used for respectively numerical and categorical outcome data. Demographic and clinical variables (only for the cSLE-cohort) were tested as co-variate factors for the amount (per mm) of ‘abnormal capillary shapes’, ‘large hemorrhages’ or ‘pericapillary extravasations’. Type of ethnic background was analyzed as an ordinal variable for three types of skin pigmentation: white/white-mixed, Asian/North-African/Middle-eastern and African/Afro-Caribbean. P-values <0.05 were considered as statistically significant.

RESULTS

Inclusion and demographics

Fifty-two patients with (suspected) cSLE were eligible for inclusion between April 2016 until September 2019. After revising SLICC-criteria, seven patients did not fulfil a minimum of four criteria and were excluded. Two patients were excluded because it was not possible to obtain clear capillaroscopy images due to skin thickness around their nailfold. One patient, with circulatory insufficiency admitted on intensive care unit, was too sick to undergo capillaroscopy examination. Therefore, forty-one patients were included for analysis.

The same number of healthy controls (n=41) were matched from a cohort of healthy children (n=140) with capillaroscopy images, first by matching for ethnic background (p=0.317). The cSLE-cohort had significantly more female patients (36 (87.8%) versus 29 (70.7%), p=0.039) and higher median age (median 17 versus 12 years, p<0.001) compared to healthy controls (see table 1).

In total, 8055 capillaries from 1147 images could be analyzed from 41 cSLE-patients. From healthy controls (n=41), 4253 capillaries were analyzed from 656 images. Disease characteristics of the cSLE-cohort are shown in table 1. Fifty six percent (56.1%) of patients were treatment naive and investigated at time of diagnosis.

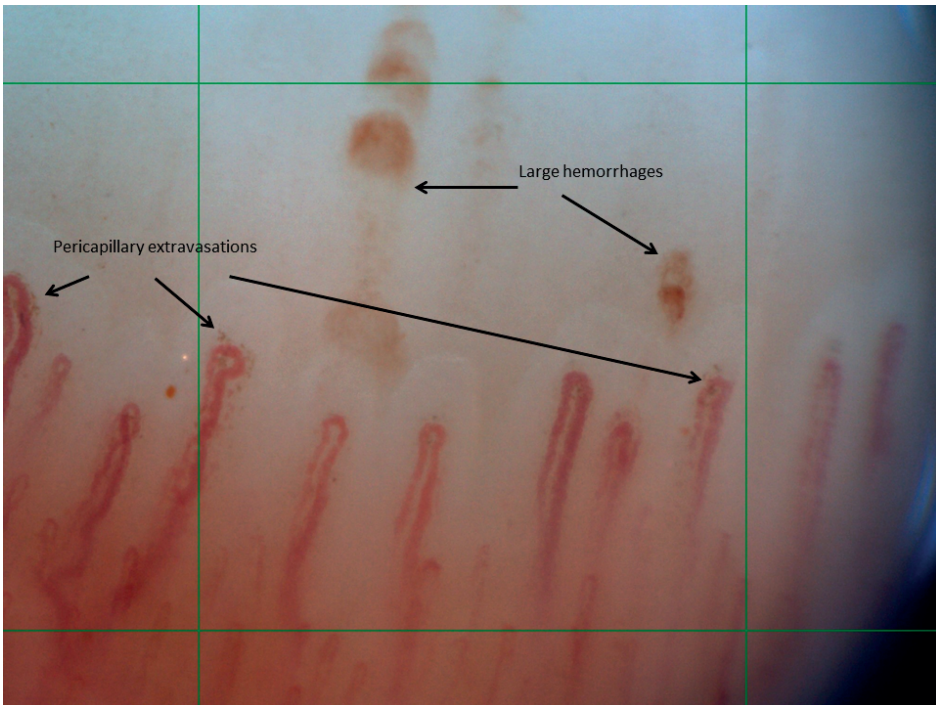


Image 1. ‘Large hemorrhages’ versus ‘pericapillary extravasations’.
Magnification 200x; green lines show 1 mm grid

Quantitative capillary variables

Compared to healthy controls and counted per millimeter, cSLE-patients showed significantly more giant capillaries ($p=0.032$), abnormal capillary shapes ($p=0.03$), more large pathological hemorrhages ($p<0.001$) and more pericapillary extravasations ($p<0.001$) (table 2). In total, large pathological hemorrhages and pericapillary extravasations were significantly more observed in respectively 75.6% (31/41) and 87.8% (36/41) of cSLE-patients compared to healthy controls (resp. in 17.1% (7/41) and in 36.6% (15/41), McNemar test; resp. $p<0.001$ and $p<0.001$)).

Qualitative capillary patterns

Compared to healthy controls, cSLE-patients showed significantly more abnormal capillary patterns ($Z=-5.291$, $p<0.001$) (table 2). In total, thirty-two patients (32/41, 78%) showed a specific combination of ‘pericapillary extravasations’ and ‘abnormal capillary shapes’ combined in the same finger (as shown in image 2), including all patients ($n=7$) with a scleroderma pattern. Looking at frequency and localization of these two combined capillary abnormalities, three patients (3/32, 9.4%) showed this combination of capillary abnormalities in all eight examined fingers, 59.4% (19/32) in four or more

fingers and 78.1% (25/32) in three or more fingers. Three other patients with high number of 'pericapillary extravasations' (with a total count of 96, 71 and 128 extravasations) were also qualitatively analyzed as 'microangiopathy', but these three patients did not show the specific combination with 'abnormal capillary shapes'.

Clinical details of cSLE-patients with capillary scleroderma pattern (n=7/41, 17.1%) are shown in supplementary file 1. Five out of seven patients (71.4%) with a scleroderma pattern had positive anti-RNP antibodies versus 32.4% (n=11/34) of patients without a scleroderma pattern, this difference was not significant (p=0.089). None of these patients showed any signs of sclerodactyly nor other classification criteria for SSc. This was also not detected at follow-up (range 1-9 years). Two healthy controls showed one giant capillary (per person) with diameters of 54.7 and 62.4 μm . As no other capillary abnormalities were found in these healthy individuals, this was not scored as a scleroderma pattern in these two healthy controls. Both giants were observed in the second fingers (with more frequent use and risk for trauma) while the giants in cSLE-patients were observed in the fourth/fifth fingers.

Correlations with demographic and clinical variables

Capillary morphology

In cSLE-patients, the amount of 'abnormal shapes per mm' was significantly correlated with periungual trauma (p=0.049) and treatment-naivety (p=0.022). In healthy controls, no correlations were found for the amount of abnormal shapes per mm (supplementary file 2).

Apical diameter

cSLE patients showed no significant correlation between the presence of giants and Raynaud's phenomenon (OR 2.3, 95% CI 0.48 – 11.08, p=0.299). There was also no significant correlation between the amount of 'giants per mm' and presence of anti-RNP antibodies (supplementary file 3).

Large pathological hemorrhages

In cSLE-patients, 'large pathological hemorrhages per mm' showed a significant correlation with SLEDAI scores (at diagnosis (p=0.009) and at moment of capillaroscopy (p=0.002)) and nephritis (p=0.012). In healthy controls, the amount of 'large pathological hemorrhages per mm' was significantly correlated with periungual trauma (p=0.004) (see table 3).

Table 1. Demographical variables and clinical characteristics of study groups

	cSLE-patients, n=41	Healthy controls, n=41	p-value
Female, n (%)	36 (87.8)	29 (70.7)	0.039^a
Ethnicity, n (%)			0.317 ^b
African/Afro-Caribbean	18 (43.9)	15 (36.6)	
White	15 (36.6)	14 (36.6)	
North-African/Middle-Eastern	3 (7.3)	4 (9.8)	
Asian	3 (7.3)	5 (12.2)	
Mixed/other	2 (4.9)	2 (4.9)	
Age at capillaroscopy in years, median (IQR)	17 (14-18)	12 (11-16.5)	<0.001^c
Raynaud's phenomenon / acro-cyanotic symptoms, n (%)	14 (34.1)	2 (4.9)	0.002^a
Age at onset in years, median (IQR 25-75)	14 (12.5-16)		
Disease duration in months, median (IQR)	12.9 (0.1-44.5)		
Prednisone naive, n (%)	23 (56.1)		
ANA at diagnosis, n (%)	41 (100)		
ANA + anti-ds-DNA	26 (63.4)		
ANA + anti-RNP	16 (39)		
ANA + anti-Sm	14 (34.1)		
Cutaneous involvement, n (%)	27 (65.9)		
Nephritis, n (%)	13 (31.7)		
Neuropsychiatric involvement, n (%)	6 (14.6)		
Antiphospholipid antibodies, n (%)	5 (12.2)		
SLEDAI score at diagnosis, median (IQR)	12 (8-16)		
SLEDAI score at capillaroscopy, median (IQR)	5 (3-10.5)		

Bold indicates statistically significant p values (<0.05). ^a McNemar test. ^b Wilcoxon signed rank test: ordinal variables (3 groups: white/mixed/other, Asian/North-African/Middle-Eastern and African/Afro-Caribbean). ^c paired t-test. ANA= Anti-Nuclear Antibodies, anti-ds-DNA= anti-double stranded DNA antibodies, anti-RNP= anti-Ribonucleoprotein, anti-Sm= anti-Smith antibodies, SLEDAI= Systemic Lupus Erythematosus Disease Activity Index, IQR= interquartile range.

Table 2. Capillary characteristics

Quantitative parameters	cSLE-patients, n=41	Healthy controls, n=41	p-value
Density per mm, mean (SD)	6.83 (1.06)	6.53 (0.86)	0.117 ^a
Max apical diameter in μm , median (IQR)	37.7 (35.2-45.9)	38.6 (32.8-41.2)	0.206 ^b
Giant capillaries per mm, mean (SD)	0.04 (0.13) ^c	0.003 (0.013)	0.032^b
Abnormal shapes per mm, median (IQR)	0.31 (0.13-0.73) ^c	0.21 (0.06-0.38)	0.003^b
Hemorrhages per mm, median (IQR)	1.1 (0.39-2.34) ^c	0 (0-0.16)	
Large pathological hemorrhages per mm	0.07 (0-0.24)	0 (0-0)	<0.001
Pericapillary extravasations per mm	1.11 (0.28-2.15)	0 (0-0.13)	<0.001
Qualitative patterns			
Normal capillary pattern, n (%)	6 (14.6)	37 (90.2)	<0.001^b
Microangiopathy, n (%)	28 (68.3) ^c	4 (9.8)	
Scleroderma pattern, n (%)	7 (17.1) ^c	0 (0)	

Bold indicates statistically significant p values (<0.05). Mm= millimeter, SD= standard deviation, μm =micrometer. ^a paired t-test. ^b Wilcoxon signed rank test. ^cnaifold capillary abnormalities which are also described in adult-onset SLE (4).

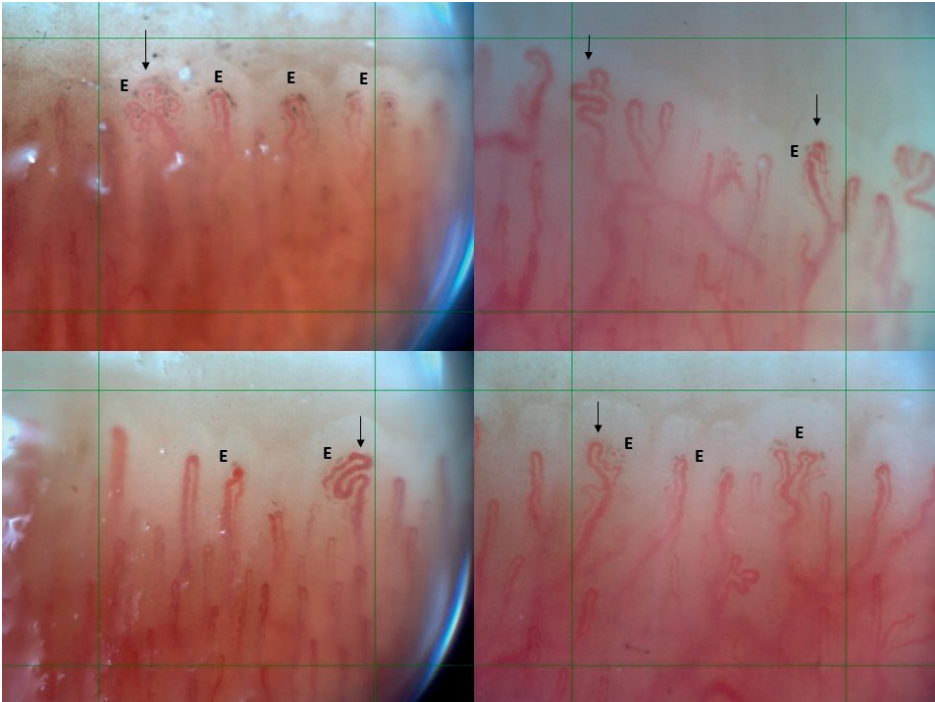


Image 2. Combination of pericapillary extravasations (E) and abnormal shapes (black arrows). Magnification 200x; green lines show 1 mm grid; images from four different patients

Pericapillary extravasations

In both cSLE-patients ($p < 0.001$) and in healthy controls ($p = 0.001$), the amount of 'pericapillary extravasations per mm', was significant positively correlated with darker skin pigmentation (see table 4). In healthy controls, the presence of 'pericapillary extravasations' (observed or not) was significantly correlated with darker skin pigmentation (logistic regression, OR 14.33, 95% CI 3.30-62.32, $p < 0.001$).

Table 3. Correlations between clinical and demographical variables and amount of “large hemorrhages per mm”

Variable	Regression coefficient β (95% CI) cSLE-patients	p-value	Regression coefficient B (95% CI) healthy controls	p-value
Skin pigmentation (ordinal)	0.103 (-0.001 – 0.207)	0.052	0.000 (-0.051 – 0.052)	0.990
Trauma	-0.068 (-0.376 – 0.239)	0.656	0.163 (0.054 – 0.271)	0.004
Raynaud/acrocyanosis	-0.087 (-0.298 – 0.124)	0.410	-0.037 (-0.247 – 0.174)	0.726
Treatment-naivety	-0.061 (-0.264 – 0.141)	0.543		
Disease duration	-0.002 (-0.006 – 0.001)	0.241		
SLEDAI at diagnosis	0.019 (0.005 – 0.032)	0.009		
SLEDAI at capillaroscopy	0.021 (0.008 – 0.035)	0.002		
Anti-RNP	-0.017 (-0.224 – 0.189)	0.866		
Cutaneous involvement	-0.128 (-0.337 – 0.081)	0.222		
Neuropsychiatric involvement	0.121 (-0.162 – 0.404)	0.392		
Nephritis	0.260 (0.06 – 0.460)	0.012		
Antiphospholipid antibodies	0.000 (-0.071 – 0.072)	0.989		

Table 4. Correlations between clinical, demographical variables and amount of “pericapillary extravasations per mm”

Variable	Regression coefficient β (95% CI) cSLE-patients	p-value	Regression coefficient B (95% CI) healthy controls	p-value
Skin pigmentation (ordinal)	0.846 (0.536 – 1.157)	<0.001	0.146 (0.063 – 0.230)	0.001
Trauma	-0.587 (-1.740 – 0.567)	0.310	-0.028 (-0.253 – 0.197)	0.803
Raynaud/acrocyanosis	0.193 (-0.612 – 0.997)	0.631	-0.129 (-0.521 – 0.262)	0.507
Treatment-naivety	0.265 (-0.501 – 1.031)	0.489		
Disease duration	0.011 (-0.002 – 0.024)	0.109		
SLEDAI at diagnosis	0.034 (-0.021 – 0.090)	0.216		
SLEDAI at capillaroscopy	0.025 (-0.31 – 0.081)	0.372		
Anti-RNP	0.518 (-0.249 – 1.284)	0.180		
Cutaneous involvement	-0.423 (-1.218 – 0.373)	0.289		
Neuropsychiatric involvement	-0.855 (-1.901 – 0.192)	0.106		
Nephritis	0.564 (-0.237 – 1.366)	0.163		
Antiphospholipid antibodies	-0.011 (-0.283 – 0.261)	0.934		

Bold indicates statistically significant p values (<0.05)

DISCUSSION

Our observations confirm that giants, abnormal capillary morphology and capillary hemorrhages are also observed in cSLE, as was already known for adults with SLE [4]. The uniqueness of our cohort is that more than half of the patients (23/41, 56.1%) were treatment-naïve at the moment of capillaroscopy examination. This is the first study to describe abnormal capillary morphology in cSLE since the new published definitions for abnormal capillary shapes from EULAR SG MCRD in 2016 [6]. In this cross-sectional study and compared to healthy controls, cSLE-patients show significantly more giant capillaries, abnormal capillary morphology and capillary hemorrhages, both in absolute numbers (per mm) as well as in percentage of patients. The high number (median 1.1 per mm) of capillary hemorrhages in cSLE patients and the observation of two different subtypes of capillary hemorrhages are the other prominent findings of our study. Large hemorrhages were also observed in healthy controls but these were significantly correlated with trauma, which seems a logical explanation.

We found a significant correlation between the amount of large hemorrhages and SLEDAI score (at diagnosis and at capillaroscopy). Significantly higher SLEDAI scores in adult SLE with major capillary changes (defined by abnormal shapes and capillary hemorrhages) have been described before, further specified by a correlation between more capillary hemorrhages in the patient group with a SLEDAI score of >12 [27]. Ingegnoli also showed a linear correlation with SLEDAI score and severity of capillary abnormalities, by semi-quantitatively scoring patterns between 0-2 [28]. Approximately half of patients (56%) in our cohort were analyzed at the moment of diagnosis (treatment naïve). Improvement of abnormal capillary changes due to therapeutic intervention has been described in SSc [29, 30]. In our cohort, the median SLEDAI score of 5 at the moment of capillaroscopy is interpreted as a low disease activity score which may underestimate our results. Our significant correlation between the amount of abnormal capillary shapes and treatment-naïvety confirms this. Presence of nephritis was significantly correlated with large pathological hemorrhages, while no other disease manifestations showed correlations with capillary abnormalities. An explanation could be that the found capillary abnormalities are representative for SLE in general and not specific for certain clinical symptoms of this severe disease.

A novel finding in this study was the observation of ‘pericapillary extravasations’: small point-shaped hemorrhages surrounding the capillary apex. These extravasations were observed in significantly higher frequency and count per mm in cSLE-patients, as compared to healthy controls. The ‘pericapillary extravasations’ seem a distinct subtype of capillary hemorrhage and were six times more often observed than ‘large pathological

hemorrhages', when analyzed per mm. Interestingly, pericapillary extravasations were not correlated with periungual trauma (table 4), suggesting a pathophysiological origin such as endothelial wall damage. To our knowledge, such extravasations have been sporadically described in adult SLE-patients (and never in children), as "pearl necklaces of extravasates" or "extravasations of red blood cells, with the impression of punched out windows" [31, 32]. A possible explanation for this new observation could be that the quality and resolution of images from NVC have significantly improved in the last years. Hypothetically, this subtype of capillary hemorrhages might be small extravasations from a vulnerable capillary possibly due to endothelial activation and damage. It is possible that these 'pericapillary extravasations' are a reflection of endothelial dysregulation, as has been demonstrated in SLE-patients [33-35], leading to vasculopathy, which may be related to the pathogenesis of SLE. Pericapillary extravasations do not show migration towards the peripheral area (along with nail growth) as large hemorrhages do in a scleroderma pattern. Possibly, smaller hemosiderin deposits are cleared faster by phagocytic cells. The endothelial activation and damage, as described in SSc [36], does also seem to play a role in SLE [33-35]. SLE occurs 2 to 4 times more frequently among non-white populations [37] and this non-white population also seems to have a more severe disease course [11, 38]. It might be that the significant higher amount of extravasations, found in our non-white cSLE-patients, reflects this.

The combination of 'pericapillary extravasations and abnormal capillary shapes' were mostly observed in the fourth and fifth digits. These digits are less used in daily activities, suggesting that these capillary changes are less likely to be caused by trauma and further supporting a possible origin from a pathophysiological damage of endothelium. Multivariate analysis to determine correlations between detection of such 'specific microangiopathy pattern' with disease characteristics could not be performed due to small sample size of the group (< 10 subjects) that did not show this 'specific microangiopathy pattern' (n=9/41) [39]. All seven cSLE-patients with a scleroderma pattern also showed this specific combination of 'capillary abnormal shapes and pericapillary extravasations'. However, these patients did not have any other clinical criteria for SSc and positive anti-RNP antibodies were not significantly more detectable in these patients. Longitudinal studies are needed for clinical follow-up of these cSLE-patients with a capillary scleroderma pattern for a correct interpretation of this finding. The hypothesis for pathogenesis of a scleroderma pattern is that the capillary first typically enlarges due to endothelial damage forming a micro-aneurysmatic giant capillary, which might subsequently lead to a capillary microhemorrhage. These microhemorrhages are closely associated with the enlarged loops and have an obvious apical capillary genesis [1]. In our cohort we did not observe a correlation between the amount of large hemorrhages and the number of giant capillaries per mm (regression coefficient B 0.71, 95% CI -0.06

– 1.49, $p=0.07$). This observation also leads to the question if the pathogenesis of large capillary hemorrhages is different in SLE than in scleroderma, both distinct systemic autoimmune diseases with incidentally clinical overlap with other connective tissue diseases.

The limitations of this study include the relatively small sample size of this cSLE-cohort with 41 patients, due to the rarity of this disease. Our male/female ratio was 1/7, representing the general known male/female ratio of 1/8-10 in adults with SLE [37] and 1/5-6 in cSLE (14). Secondly, it is known that SLE occurs 2 to 4 times more frequently among non-white populations [37] which is also shown in our data (63.4% non-white). In cSLE, a median onset of 12.6 years (IQR 10.4-14.5) at diagnosis is described in the literature (14) which corresponds with our cohort with a median age of 14 years (IQR 12.5-16) at diagnosis. Although median age was significantly lower in the healthy cohort (12 versus 17 years), this difference will probably not make a difference for our outcome data, as it still concerned a pediatric (teenage) population. The same argument applies to matching of gender which was significantly different but with percentages of 87 versus 70% a majority of females in both cohorts.

CONCLUSION

This study confirms that children with SLE, like adult SLE-patients, also show significantly more giants, abnormal capillary morphology and capillary hemorrhages, when compared to healthy controls. In our study, these abnormal capillary findings were significantly correlated with SLEDAI scores, treatment-naivety and nephritis, thus making nailfold capillary abnormalities potentially interesting as disease biomarker(s). A prominent finding was the observation of a newly described subtype of capillary hemorrhage which we called “pericapillary extravasations”. By assessment of intra- and inter-observer variability we need to determine if these pericapillary extravasations are reproducible, to confirm if they are a distinct finding from large capillary hemorrhages.

ACKNOWLEDGEMENTS

We would like to thank to Dr. M.D.J. Wolvers for her advice on the statistical analyses of our data.

We would like to thank all healthy children, their parents and primary schools “Knotwilg”, “Piet Hein” and “Burgemeester Amersfoordtschool” for their participation in undertaking a nailfold capillaroscopy.

Supplementary file 1. Clinical characteristics of cSLE-cases with capillary scleroderma pattern, n=7.

	Case 1	Case 2	Case 3	Case 4	Case 5	Case 6	Case 7	
Demographics (age at presentation)	Girl, Caucasian, 12 years old	Girl, Caucasian, 13 years old	Boy, Asian, 13 years old	Girl, Afro-Caribbean, 15 years old	Girl, Afro-Caribbean, 17 years old	Girl, Afro-Caribbean	Girl, Caucasian, 17 years old	
Clinical symptoms	Butterfly rash of erosive ulcerative skin disease, extensive oral aphthous ulcers, chilblains, auto-immune hepatitis, splenomegaly with calcifications, leukopenia, thrombocytopenia	Butterfly rash and discoid skin lesions trunk, arms after sun exposure, aphthous ulcers, leukopenia	Pleuritis, pericarditis, ascites, myositis, polyarthritits, fever, lymphadenopathy, deep venous thrombosis, hepatosplenomegaly, aphthous ulcers, leukopenia	Pleuritis, pericarditis, myositis, polyarthritits, fever, aphthous ulcers, leukopenia, thrombocytopenia	Aphthous ulcers, lymphadenopathy, skin rash (resulting in hyperpigmentation)	Vasculitis, arthritis, nephritis, mood disorder (intracranial hypertension/resorption)		Aphthous ulcers, leukopenia, thrombocytopenia
Skin biopsy	Vacuolar degeneration of the basal layer with abundant nuclear dust, also localized around the superficial blood vessels. Immunofluorescence: positive lupus band with (granular) staining of IgG, IgM, C1q and C3	The epidermis shows a hyperkeratotic basket-weave stratum corneum, with some vacuolar degeneration of the basal layer. The epidermis is atrophic with follicular plugging. Below the epidermis a mild perivascular and perifollicular, predominantly lymphocytic infiltrate	-	-	A-specific inflammation lymph node, lip biopsy inconclusive	Around superficial vessels, subcutaneous tissue and deep dermal plexus inflammation with lymphocytes, histiocytes, neutrophilic granulocytes en abundant nuclear dust. Also eosinophilic granulocytes around vessels and focal interstitial degranulation. Vessel wall is swollen with focal fibroid change. Immunofluorescence: depositions of complement factors (C1q>>C3c), combined with IgM (and some IgG)	-	
ANA / anti-ds-DNA	positive / positive	positive / positive	positive / positive	positive / positive	positive / negative	positive / positive	positive / positive	
Other auto-antibodies	anti-RNP, anti-Ro52, anti-SS-A, anti-Sm	anti-RNP, anti-SS-A, anti-Sm	anti-RNP, anti-SS-A, anti-Sm, rheumatoid factor	anti-RNP, anti-Sm, anti-Ro52, anti-ds-DNA, rheumatoid factor	none	anti-C1q antibodies	Anti-RNP, anti-Sm	

Nailfold videocapillaroscopy in cSLE

Supplementary file 1. Clinical characteristics of cSLE-cases with capillary scleroderma pattern, n=7. (continued)

	Case 1	Case 2	Case 3	Case 4	Case 5	Case 6	Case 7
Anti-phospholipid antibodies	negative	negative	positive	negative	positive	negative	negative
C3/C4	low	low	low	low	normal	low	normal
Coombs test	positive	positive	positive	positive	positive	positive	negative
SLEDAI at presentation	17	10	10	29	4	35	4
SLEDAI at capillaroscopy	17	10	8	6	4	35	4
Disease duration at capillaroscopy	at diagnosis	at diagnosis	4 years	5 years	at diagnosis	at diagnosis	at diagnosis
Discoloration of fingers	acrocyanosis in winter	no	biphasic Raynaud's phenomenon	acrocyanosis during whole year	no	no	Raynaud
Sclerodactyly	no	no	no	no	no	no	no
Pulmonary disease	no	no	restrictive pulmonary function	no	no	no	no
Nephritis	no	no	proteinuria: biopsy refused by parents/patient	nephritis class V	no	nephritis class IV	no
Medication at capillaroscopy (and ever used)	None (prednisolone, hydroxychloroquine, rituximab, mycophenolate mofetil, belimumab)	None (prednisolone, hydroxychloroquine, azathioprine)	prednisolone, hydroxychloroquine, methotrexate (rituximab, mycophenolate mofetil)	prednisolone, hydroxychloroquine, mycophenolate mofetil, cyclophosphamide (azathioprine, methotrexate, rituximab, belimumab)	Prednisolone (hydroxychloroquine)	None (prednisolone, hydroxychloroquine, mycophenolate mofetil)	(hydroxychloroquine, nifedipine, prednisolone)
Follow-up period	4 years	5 years	8 years	9 years	Lost to follow-up	2 years	1 year

Supplementary file 2. Correlations between clinical and demographical variables and amount of “abnormal shapes per mm”

Variable	Regression coefficient β (95% CI) cSLE	p-value	Regression coefficient B (95% CI) healthy controls	p-value
Skin pigmentation (ordinal)	0.037 (-0.1 – 0.173)	0.587	0.021 (-0.035 – 0.078)	0.454
Trauma	0.369 (0.001 – 0.737)	0.049	-0.012 (-0.146 – 0.121)	0.854
Raynaud/acrocyanosis	0.184 (-0.076 – 0.444)	0.160	-0.066 (-0.298 – 0.167)	0.572
Treatment-naivety	0.281 (0.042 – 0.519)	0.022		
Disease duration	0.005 (0.001 – 0.01)	0.01		
SLEDAI at diagnosis	0.009 (-0.009 – 0.028)	0.324		
SLEDAI at capillaroscopy	-0.01 (-0.029 – 0.008)	0.257		
Anti-RNP	0.226 (-0.023 – 0.475)	0.074		
Cutaneous involvement	-0.145 (-0.408 – 0.118)	0.271		
Neuropsychiatric involvement	0.065 (-0.293 – 0.422)	0.716		
Nephritis	-0.137 (-0.405 – 0.132)	0.309		
Antiphospholipid antibodies	-0.029 (-0.118 – 0.061)	0.518		

Bold indicates statistically significant p values (<0.05)

Supplementary file 3. Correlations between clinical and demographical variables and amount of “giant capillaries per mm”

Variable	Regression coefficient β (95% CI) cSLE	p-value
Skin pigmentation (ordinal)	-0.013 (-0.057 – 0.030)	0.543
Trauma	-0.026 (-0.149 – 0.098)	0.678
Raynaud/acro-cyanosis	0.054 (-0.030 – 0.137)	0.200
Treatment-naivety	0.013 (-0.068 – 0.095)	0.740
Disease duration	-0.001 (-0.002 – 0.000)	0.166
SLEDAI at diagnosis	-0.002 (-0.008 – 0.004)	0.487
SLEDAI at capillaroscopy	0.001 (-0.005 – 0.007)	0.629
Anti-RNP	0.052 (-0.029 – 0.134)	0.200
Cutaneous involvement	-0.022 (-0.107 – 0.063)	0.598
Neuropsychiatric involvement	-0.028 (-0.142 – 0.086)	0.619
Nephritis	-0.034 (-0.120 – 0.052)	0.430
Antiphospholipid antibodies	-0.002 (-0.031 – 0.026)	0.874

Bold indicates statistically significant p values (<0.05)

REFERENCES

1. Cutolo, M., Atlas of capillaroscopy in rheumatic diseases. 2010, Milan: Elsevier.
2. Maricq, H.R., et al., Diagnostic potential of in vivo capillary microscopy in scleroderma and related disorders. *Arthritis Rheum*, 1980. 23(2): p. 183-9.
3. Smith, V., et al., Standardisation of nailfold capillaroscopy for the assessment of patients with Raynaud's phenomenon and systemic sclerosis. *Autoimmun Rev*, 2020. 19(3): p. 102458.
4. Cutolo, M., et al., Nailfold capillaroscopy in systemic lupus erythematosus: A systematic review and critical appraisal. *Autoimmun Rev*, 2018. 17(4): p. 344-352.
5. Schonenberg-Meinema, D., et al., Capillaroscopy in childhood-onset systemic lupus erythematosus: a first systematic review. *Clin Exp Rheumatol*, 2020. 38(2): p. 350-354.
6. Smith, V., et al., An EULAR study group pilot study on reliability of simple capillaroscopic definitions to describe capillary morphology in rheumatic diseases. *Rheumatology (Oxford)*, 2016. 55(5): p. 883-90.
7. Cutolo, M., et al., Reliability of simple capillaroscopic definitions in describing capillary morphology in rheumatic diseases. *Rheumatology (Oxford)*, 2018. 57(4): p. 757-759.
8. Tamirou, F., et al., Systemic lupus erythematosus: state of the art on clinical practice guidelines. *RMD Open*, 2018. 4(2): p. e000793.
9. Ambrose, N., et al., Differences in disease phenotype and severity in SLE across age groups. *Lupus*, 2016. 25(14): p. 1542-1550.
10. Choi, J.H., et al., Comparison of clinical and serological differences among juvenile-, adult-, and late-onset systemic lupus erythematosus in Korean patients. *Lupus*, 2015. 24(12): p. 1342-9.
11. Martinez-Barrio, J., et al., Juvenile, adult and late-onset systemic lupus erythematosus: a long term follow-up study from a geographic and ethnically homogeneous population. *Clin Exp Rheumatol*, 2015. 33(6): p. 788-94.
12. Fatemi, A., M. Matinfar, and A. Smiley, Childhood versus adult-onset systemic lupus erythematosus: long-term outcome and predictors of mortality. *Clin Rheumatol*, 2017. 36(2): p. 343-350.
13. Sousa, S., et al., Clinical features and long-term outcomes of systemic lupus erythematosus: comparative data of childhood, adult and late-onset disease in a national register. *Rheumatol Int*, 2016. 36(7): p. 955-60.
14. Nived, O., et al., High predictive value of the Systemic Lupus International Collaborating Clinics/American College of Rheumatology damage index for survival in systemic lupus erythematosus. *J Rheumatol*, 2002. 29(7): p. 1398-400.
15. van Vollenhoven, R.F., et al., Treat-to-target in systemic lupus erythematosus: recommendations from an international task force. *Ann Rheum Dis*, 2014. 73(6): p. 958-67.
16. Davidson, J.E., et al., Quantifying the burden of steroid-related damage in SLE in the Hopkins Lupus Cohort. *Lupus Sci Med*, 2018. 5(1): p. e000237.
17. Heshin-Bekenstein, M., et al., Longitudinal disease- and steroid-related damage among adults with childhood-onset systemic lupus erythematosus. *Semin Arthritis Rheum*, 2019. 49(2): p. 267-272.
18. Smith, V., et al., Nailfold capillaroscopy for prediction of novel future severe organ involvement in systemic sclerosis. *J Rheumatol*, 2013. 40(12): p. 2023-8.
19. Smith, V., et al., Do worsening scleroderma capillaroscopic patterns predict future severe organ involvement? a pilot study. *Ann Rheum Dis*, 2012. 71(10): p. 1636-9.
20. Caramaschi, P., et al., Scleroderma patients nailfold videocapillaroscopic patterns are associated with disease subset and disease severity. *Rheumatology (Oxford)*, 2007. 46(10): p. 1566-9.

21. Marino Claverie, L., et al., Organ involvement in Argentinian systemic sclerosis patients with “late” pattern as compared to patients with “early/active” pattern by nailfold capillaroscopy. *Clin Rheumatol*, 2013. 32(6): p. 839-43.
22. Boulon, C., et al., Correlation between capillaroscopic classifications and severity in systemic sclerosis: results from SCLEROCAP study at inclusion. *Clin Exp Rheumatol*, 2019. 37 Suppl 119(4): p. 63-68.
23. Terreri, M.T., et al., Nail fold capillaroscopy: normal findings in children and adolescents. *Semin Arthritis Rheum*, 1999. 29(1): p. 36-42.
24. Andrade, L.E., et al., Panoramic nailfold capillaroscopy: a new reading method and normal range. *Semin Arthritis Rheum*, 1990. 20(1): p. 21-31.
25. Petri, M., et al., Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis Rheum*, 2012. 64(8): p. 2677-86.
26. Smith, V., et al., Fast track algorithm: How to differentiate a “scleroderma pattern” from a “non-scleroderma pattern”. *Autoimmun Rev*, 2019. 18(11): p. 102394.
27. Shenavandeh, S. and S. Habibi, Nailfold capillaroscopic changes in patients with systemic lupus erythematosus: correlations with disease activity, skin manifestation and nephritis. *Lupus*, 2017: p. 961203316686702.
28. Ingegno, F., et al., Evaluation of nailfold videocapillaroscopic abnormalities in patients with systemic lupus erythematosus. *Jcr-Journal of Clinical Rheumatology*, 2005. 11(6): p. 295-298.
29. Sulli, A., et al., Scoring the nailfold microvascular changes during the capillaroscopic analysis in systemic sclerosis patients. *Ann Rheum Dis*, 2008. 67(6): p. 885-7.
30. Miniati, I., et al., Autologous stem cell transplantation improves microcirculation in systemic sclerosis. *Ann Rheum Dis*, 2009. 68(1): p. 94-8.
31. Maricq, H.R. and E.C. LeRoy, Patterns of finger capillary abnormalities in connective tissue disease by “wide-field” microscopy. *Arthritis Rheum*, 1973. 16(5): p. 619-28.
32. Groen, H., et al., Pulmonary function in systemic lupus erythematosus is related to distinct clinical, serologic, and nailfold capillary patterns. *Am J Med*, 1992. 93(6): p. 619-27.
33. Mak, A. and N.Y. Kow, Imbalance between endothelial damage and repair: a gateway to cardiovascular disease in systemic lupus erythematosus. *Biomed Res Int*, 2014. 2014: p. 178721.
34. Tyden, H., et al., Endothelial dysfunction is associated with activation of the type I interferon system and platelets in patients with systemic lupus erythematosus. *RMD Open*, 2017. 3(2): p. e000508.
35. Lee, W.F., et al., Biomarkers associating endothelial Dysregulation in pediatric-onset systemic lupus erythematosus. *Pediatr Rheumatol Online J*, 2019. 17(1): p. 69.
36. Mostmans, Y., et al., The role of endothelial cells in the vasculopathy of systemic sclerosis: A systematic review. *Autoimmun Rev*, 2017. 16(8): p. 774-786.
37. Pons-Estel, G.J., et al., Understanding the epidemiology and progression of systemic lupus erythematosus. *Semin Arthritis Rheum*, 2010. 39(4): p. 257-68.
38. Lewis, M.J. and A.S. Jawad, The effect of ethnicity and genetic ancestry on the epidemiology, clinical features and outcome of systemic lupus erythematosus. *Rheumatology (Oxford)*, 2017. 56(suppl_1): p. i67-i77.
39. Peduzzi, P., et al., A simulation study of the number of events per variable in logistic regression analysis. *J Clin Epidemiol*, 1996. 49(12): p. 1373-9.

4

Standardized nailfold capillaroscopy in children with rheumatic diseases: a worldwide study

Karin Melsens*, Maurizio Cutolo*, **Dieneke Schonenberg-Meinema**, Ivan Foeldvari,
Maria C. Leone, Yora Mostmans, Valerie Badot, Rolando Cimaz, Joke Dehoorne,
Ellen Deschepper, Tracy Frech, Johanna Hernandez-Zapata, Francesca Ingegnoli,
Archana Khan, Dorota Krasowksa, Hartwig Lehman, Ashima Makol,
Miguel A. Mesa-Navas, Malgorzata Michalska-Jakubus, Ulf Müller-Ladner,
Laura Nuño-Nuño, Rebecca Overbury, Carmen Pizzorni, Mislav Radic, Divya Ramadoss,
Angelo Ravelli, Silvia Rosina, Clara Udaondo, J Merlijn van den Berg, Ariane L. Herrick,
Alberto Sulli, Vanessa Smith

On behalf of EULAR Study Group on Microcirculation in Rheumatic Diseases

*Shared first authors

Published in Rheumatology (Oxford): 2022 Aug 25 Online ahead of print

ABSTRACT

OBJECTIVE: To standardly assess and describe nailfold videocapillaroscopy (NVC) assessment in children and adolescents with juvenile rheumatic and musculoskeletal diseases (jRMD) versus healthy controls (HC).

METHODS: In consecutive jRMD children and matched HC from 13 centres worldwide, 16 NVC images per patient were acquired locally and read centrally for standard evaluation by international consensus definitions from the EULAR Study Group on Microcirculation in Rheumatic Diseases. Ninety-five patients with juvenile idiopathic arthritis (JIA), 22 with dermatomyositis (JDM), 20 with systemic lupus erythematosus (cSLE), 13 with systemic sclerosis (jSSc), 21 with localized scleroderma (lSc), 18 with mixed connective tissue disease (MCTD) and 20 with primary Raynaud's phenomenon (PRP) were included. NVC differences between juvenile subgroups and HC were calculated through multivariable regression analysis.

RESULTS: A total number of 6474 images were assessed from 413 subjects (mean age 12.1-years, 70.9% female). The quantitative NVC-characteristics were significantly lower (↓) or higher (↑) in the following subgroups compared to HC: For density: ↓ in jSSc, JDM, MCTD, cSLE and lSc; For dilations: ↑ in jSSc, MCTD and JDM; For abnormal shapes: ↑ JDM and MCTD; For haemorrhages: ↑ in jSSc, MCTD, JDM and cSLE. The qualitative NVC-assessment of JIA, lSc and PRP did not differ from HC, whereas the cSLE and jSSc, MCTD, JDM, cSLE subgroups showed more non-specific and scleroderma patterns respectively.

CONCLUSION: This analysis resulted from a pioneering registry of NVC in jRMD. The NVC-assessment in jRMD differed significantly from HC. Future prospective follow up will further elucidate the role of NVC in jRMD.

INTRODUCTION

Nailfold capillaroscopic examination has proven its worth in the field of rheumatology [1, 2]. In adults, together with the identification of autoantibodies, it allows discrimination between primary and secondary Raynaud's phenomenon (RP). The latter is mostly related to an underlying connective tissue disease (CTD) [3]. Moreover, its role was formalized, through the incorporation of "an abnormal nailfold capillaroscopy" as criterion (referring to the scleroderma pattern) in the systemic sclerosis (SSc) 2013 classification criteria [4]. Ever since its increased use, assembled efforts have led to the standardisation of the capillaroscopic technique and reading in adults [5-8]. In children and adolescents however, nailfold capillaroscopy is less used and studies are scarce and usually small, using non-standardised methods [9]. The European League Against Rheumatism (EULAR) Study Group of Microcirculation in Rheumatic Diseases found it thus timely to start an international collaborative to collect nailfold capillaroscopic data from children and adolescents with and without juvenile Rheumatic and Musculoskeletal Diseases (jRMD). We hypothesise that through nailfold videocapillaroscopy (NVC), as in adult disease, we may detect microvascular abnormalities in jRMD, presumably caused by systemic inflammatory immune responses.

This article presents the first data from a collaborative effort, in which the NVC-features of a large international cohort of children and adolescents are standardly described with international consensus definitions from the EULAR Study Group of Microcirculation in Rheumatic Diseases. The results could serve as a framework for paediatricians to interpret NVC-assessments in their rheumatology practice. For researchers, it could provide the prerequisites to further investigate NVC-characteristics and clinical correlations in disease specific studies.

PATIENTS AND METHODS

Study patients and healthy controls

Children and adolescents from 13 different centres worldwide were examined using NVC. All participating centres obtained approval from the local ethics committee (for Belgium: B670201627545) and written informed consents from all participants or representatives were obtained. Details on centre contributions and their approval numbers are found in the Supplementary file, table S1 [7].

Co-investigators were asked to include consecutive patients with RP and/or a definite diagnosis of a jRMD according to the physician's opinion and fulfilling the established

classification criteria, irrespective of the disease duration or disease activity status [10-16]. RP was defined as the observation of at least a biphasic colour change after cold exposure [17]. Patients with an indefinite diagnosis or overlap (other than mixed connective tissue disease) were excluded. Additionally, the presence of antinuclear antibodies (ANA) and, if available, the specifications on extractable nuclear antigen antibodies, were documented, as per discretion of the local investigator. No records were taken on therapies, with the assumption that all patients were receiving a variety of therapies and the unawareness of a specific therapy that is clearly correlated with the presence of NVC-abnormalities.

Each jRMD patient with evaluable NVC images was manually matched to a healthy control child (HC) from the same gender and age group (groups with the following age ranges: < 5 years, 5-7 years, 8-10 years, 11-14 years and 15-18 years) [18]. If no exact age and gender matched HC was available, the authors selected an as close as possible available match, where priority was given to similar age over same gender. The HC were provided by different centres. They were mainly recruited at schools, some were siblings of patients or family members from the investigators. No reliable records were taken on the presence of RP in HC, recruited in circumstances where serological or clinical assessment was unavailable (e.g. in schools).

Capillaroscopic technique and reading method

Each child was examined with a standardised NVC-technique [7]. All fingers, except for the thumbs, were assessed with a x200-magnification contact lens (Videocap/Optilia/Inspectis/Dino-Lite microscope, depending on local equipment). Two adjacent central images per nailfold were captured, coded, and saved (set of 16 images per child) and the investigators were asked to place a grid on all images, corresponding to one-millimetre (mm) nailfold in real life, using centre-dependent image analysis software (DS Medica-Videocap, Italy; Optilia-Mediscopes, Sweden; Inspectis, Sweden; Dino-Capture, Taiwan). The time to acquire the 16 images per child took about ten minutes. No specific training was given to the investigators, who were all operating in a capillaroscopic expert centre. The NVC-images were collected digitally through Web Share and read at the Ghent University by a trained observer, who was blind for healthy or disease status (KM) [19]. The graphic viewer "IrfanView" (Version 4.51) was used to correct for image sizes, which varied among centres, and to measure the dimension of the capillaries, using the one-mm grid as a reference. The reading and reporting method followed the capillaroscopic protocol from the EULAR Study Group of Microcirculation in Rheumatic Diseases (figure 1) [7, 20].

The quantitative NVC-assessment in the one-mm grid consisted of the following NVC-parameters:

- the “capillary density” (the number of capillaries in the distal row);
- the “capillary dimension” (the number of dilated capillaries (dilations) having an apical limb diameter between 20-50um and the number of giant capillaries (giants), having an apical limb diameter of >50um);
- the “capillary morphology” (the number of capillaries with a normal morphology -capillaries with a hairpin shape; once or twice-crossing shape; or tortuous shape i.e. limbs bend but do not cross; on the condition that the tip is convex- and abnormal morphology -all capillaries whose shape does not correspond to the definition of a normal shape) [21, 22] and;
- the presence of microhemorrhages (red or brown amorphous structures in the pericapillary/periungual region).

Nailfold Video Capillaroscopic Protocol	Left Hand				Right Hand			
	2 nd	3 rd	4 th	5 th	2 nd	3 rd	4 th	5 th
QUANTITATIVE ASSESSMENT <i>image level per linear mm</i>								
Capillary density	-	-	-	-	-	-	-	-
Number of dilations	-	-	-	-	-	-	-	-
Number of giants	-	-	-	-	-	-	-	-
Number of normal shapes	-	-	-	-	-	-	-	-
Number of abnormal shapes	-	-	-	-	-	-	-	-
Presence of microhaemorrhages	-	-	-	-	-	-	-	-
QUALITATIVE ASSESSMENT <i>subject level</i>								
Non-scleroderma pattern								
Normal pattern				_____				
Non-specific pattern				_____				
Scleroderma pattern								
_____				_____				

Figure 1. Standardized nailfold videocapillaroscopy (NVC) protocol.

Two images of the second, third, fourth and fifth digit of each hand are assessed according to the standard format to report on capillaroscopic characteristics, including the quantitative (per linear mm) and qualitative (the capillary pattern) assessment (image level). To report at subject level, the capillaroscopic parameters are deduced from all the obtained NVC-images from an individual subject:

- by calculating the mean of the capillary density, the mean of the number of dilations and the mean of the number of abnormal shapes (the sum of each NVC-parameter at image level divided by the number of assessed images per subject);
- by describing the presence or absence of the parameters ‘giants’ and ‘microhemorrhages’ in a dichotomous way and;
- by deriving the overall qualitative assessment (details in section 2.2).

To obtain the quantitative NVC-parameters at subject level, the means were calculated, except for the two NVC-parameters “giants” and “microhaemorrhages”, which were both reported in a dichotomous way at subject level, as being present or not.

The qualitative NVC-assessment consisted of categorizing the overall capillary pattern at subject level in a “scleroderma pattern” (a pattern with the presence of giants or the combination of abnormal shapes with an extremely lowered number of capillaries) or a “non-scleroderma pattern”. The latter included a “normal pattern” (capillaries of the distal row are normally shaped, homogenous in dimension and their density is ≥ 7 / linear mm) and a “non-specific pattern” [8]. To depict the overall capillary pattern at subject level, the following rules were applied: - as soon as one of the images was categorized as a “scleroderma pattern” the overall capillary pattern was a “scleroderma pattern”; -when no “scleroderma pattern” was present (none of the 16 images), the most dominant “non-scleroderma pattern” depicted the overall capillary pattern; - if both the “normal” and “non-specific” patterns were equally represented (eight “normal” patterns and eight “non-specific” patterns), the “non-specific pattern” was assigned as overruling capillary pattern. Examples of the quantitative and qualitative NVC-assessments at image level are given in figure 2.

Capillaroscopic characteristics	Non-scleroderma pattern		Scleroderma pattern		
	Normal	Non-specific abnormalities	Early	Active	Late
Density (mm)	≥ 7	↓	≥ 7	4-6	≤ 3
Dimension (μm)	Normal	20-50	>50 (giant)	>50 (giant)	-
Abnormal morphology	-	+	-	+	++

Figure 2. Standardized assessment of NVC-images according to the internationally consensus EULAR Study Group on Microcirculation in Rheumatic Diseases definitions.

A) An example of a stereotype “normal” pattern. Density: 8 capillaries per linear mm (↓). Dimension: no dilations, no giants. Morphology: no abnormal shapes. Microhemorrhages: absent. Interpretation: normal pattern (non-scleroderma pattern)

B) An example of a “non-specific” pattern. Density: 8 capillaries per linear mm (↓). Dimension: presence of 3 dilations per linear mm, no giants. Morphology: presence of 2 abnormal shapes (S). Microhemorrhages: present. Interpretation: non-specific abnormalities (non-scleroderma pattern)

C) An example of a “scleroderma” pattern. Density: 5 capillaries per linear mm (↓). Dimension: presence of a giant (↓). Morphology: no abnormal shapes. Microhemorrhages: present. Interpretation: (active) scleroderma pattern.

Adapted from Smith V et al. *Standardization of nailfold capillaroscopy for the assessment of patients with Raynaud’s phenomenon and systemic sclerosis*. Autoimmunity reviews. 2020 [7].

Evaluability and handling of missing data

The evaluability was assessed per image based on the difficulty of the capillaroscopic reading. If all NVC-parameters were evaluable, the visibility of that image was considered as “good”. On the other hand, an image was scored as “bad” if any NVC-parameter was not evaluable. A “bad” image was still included for analysis and the concerning unevaluable parameter was then encoded as a missing. A subject was excluded from further analysis if less than four out of the 16 images were scored as “good”.

Statistical analysis

Analyses have been performed by IBM SPSS version 27 and R version 4.1.1. To assess the matched sample quality (jRMD vs HC), an Optimal Pair Propensity Score Matching method was used, that minimizes the overall distance (age and gender) between matched pairs [23]. Descriptive statistics were used to represent demographic features and to describe the NVC-parameters per subgroup, with means and standard deviations (SDs) for continuous variables and proportions for categorical variables. The following six NVC-parameters were analysed at subject level: the mean capillary density, the mean number of dilations, the presence of giants, the mean number of abnormal shapes, the presence of microhemorrhages and the capillary pattern. For comparison of these six NVC-parameters between disease subgroups and the overall HC-group, unmatched multivariable regression analysis was performed, adjusted for age and gender. Raw p-values were reported, and a Bonferroni correction was applied (α -level set at 0.007, as seven subgroups were compared to the overall HC-group) and corresponding 99.3% confidence intervals (CI) are reported. The mean differences (for continuous variables) and the odds ratios (for categorical variables) were presented, in comparison with the HC-subgroup. Additionally, exploratory subgroup analysis in the HC-group was done by calculating the Pearson correlation coefficient to quantify the linear relationship between age and the quantitative NVC-parameters density, dimension, and morphology.

RESULTS

A total number of 6474 NVC images were assessed from 413 subjects, consisting of 120 boys and 293 girls, with a mean age of 12.1 years (± 3.72 SD). In some subjects, less than 16 NVC-images were assessed, due to erroneous storage of the images or because adjacent NVC-images were overlapping. Demographics per subgroup are shown in table 1 and details on ANA are found in the Supplementary file (table S2).

Nailfold videocapillaroscopy in cSLE

Table 1. Demographics (n=413)

	HC n=204	PRP n=20	cSLE n=20	JDM n=22	jSSc n=13	ISc n=21	MCTD n=18	OA n=22	PA n=23	ERA n=20	PSA n=15	JIAS n=15
Age (years), mean ±SD	12.0 ±3.6	14.6 ±2.1	13.6±4.0	11.4±3.9	13.1±3.6	11.2±3.5	13.5±3.2	10.8±4.1	11.4±4.0	13.6±2.8	12.1±4.4	9.4±3.9
Female gender, n (%)	145 (71.1)	16 (80.0)	14 (70.0)	15 (68.2)	12 (92.3)	14 (66.7)	15 (83.3)	18 (81.8)	15 (65.2)	11 (55.0)	9 (60.0)	9 (60.0)
Race, n (%)												
White	161 (78.9)	17 (85.0)	8 (40.0)	17 (77.3)	5 (38.5)	15 (71.4)	12 (66.7)	19 (86.4)	20 (87.0)	17 (85.0)	13 (86.7)	8 (53.3)
Asian	6 (2.9)	0 (0.0)	3 (15.0)	0 (0.0)	3 (23.1)	1 (4.8)	1 (5.6)	2 (9.1)	0 (0.0)	0 (0.0)	2 (13.3)	6 (40.0)
Black	11 (5.4)	0 (0.0)	5 (25.0)	2 (9.1)	1 (7.7)	1 (4.8)	0 (0.0)	1 (4.5)	0 (0.0)	0 (0.0)	0 (0.0)	1 (6.7)
Other	26 (12.7)	3 (15.0)	4 (20.0)	3 (13.6)	4 (30.8)	4 (19.0)	5 (27.8)	0 (0.0)	3 (13.0)	3 (15.0)	0 (0.0)	0 (0.0)
Disease duration (years), mean ±SD	-	1.3±1.9	2.4±2.7	3.3±4.2	2.5±2.5	3.8±3.2	2.1±2.8	6.2±4.5	4.5±3.6	3.5±3.7	4.7±5.1	4.2±4.1
Raynaud's phenomenon, n (%)	4* (2.0)	20 (100)	4 (20.0)	0 (0.0)	13 (100)	2 (9.5)	10 (55.6)	2 (9.1)	1 (4.3)	2 (10.0)	0 (0.0)	1 (6.7)
ANA, n/total valid (%)	-	5/20 (25.0)	17/18 (94.4)	13/16 (81.2)	13/13 (100.0)	2/11 (18.2)	14/16 (87.5)	9/17 (40.9)	13/21 (59.1)	6/15 (40.0)	5/10 (50.0)	1/15 (6.7)

*4/204 (2.0%) of HC had RP, in which the cause remained undefined (primary or secondary), as no rheumatological work-out had been performed in volunteers at schools. ANA, antinuclear antibodies detected by indirect immunofluorescence-screening on Hep-2 cell substrates; cSLE, childhood onset systemic lupus erythematosus; ERA, enthesitis related arthritis; HC, healthy controls; JDM, juvenile dermatomyositis; JIAS(S), juvenile idiopathic arthritis (systemic); jSSc, juvenile systemic sclerosis; ISc, localized scleroderma; MCTD, mixed connective tissue disease; OA, oligoarthritis; PA, polyarthritis; PRP, primary Raynaud's phenomenon; PSA, psoriatic arthritis.

Evaluability and matching

In 5912 NVC-images (91.3%) all NVC-parameters could be assessed. Most subjects had more than 12 images of “good” visibility (Supplementary file, table S3). From the 209 studied patients, five subjects had a bad general visibility (<4/16 “good” images): in four subjects (three with the systemic form of juvenile idiopathic arthritis (JIAS) and one with psoriatic arthritis), it was related to a technical problem with the focus of the lens, and in one patient with juvenile dermatomyositis (JDM) it was inherent to the capillaroscopic abnormalities, in which diffuse microhemorrhages made it impossible to assess other NVC-parameters. Those five subjects were excluded for further analysis (and were not matched).

As such, statistical analyses are performed on an age and gender balanced sample of 204 patients with jRMD and 204 HC. An Optimal Pair Propensity Score Matching method showed no imbalances in gender and a standardized mean difference in age below 0.1, which corresponds to a maximum age difference of three years within matched pairs.

Quantitative NVC-assessment per disease subset

Table 2 shows the NVC-parameters per subgroup, together with the mean differences and odds ratios per disease subset versus HC.

Capillary density

The mean capillary density in HC was 8.5 capillaries/linear mm (± 1.2 SD), which was similar to the mean capillary density from patients with PRP and JIA. On the other hand, in patients with mixed connective tissue disease (MCTD), JDM and juvenile SSC (jSSc), a significantly lower capillary density was seen (7.1 ± 1.3 SD, 6.3 ± 2.0 SD and 5.2 ± 1.9 SD respectively, $p < 0.001$).

The density in childhood onset systemic lupus erythematosus (cSLE) (7.6 ± 1.3 SD) and localized scleroderma (lSc) (7.7 ± 1.2 SD) was significantly lower compared to HC as well ($p = 0.006$ and $p = 0.005$ respectively). However, the mean difference remained small (less than one capillary per linear mm). The results are visualized in figure 3A.

Capillary dimension

In HC, a mean of $0.5 (\pm 0.6)$ SD capillary dilations per linear mm was observed. In lSc and JIA patients, the capillary dimension did not differ from HC. An increased number of dilated capillaries was seen in PRP (1.0 ± 0.9 SD, $p = 0.008$) and in cSLE (0.9 ± 1.1 SD, $p = 0.035$), but was only statistically significantly different from HC in JDM, MCTD and in jSSc, in which a mean of respectively $1.5 (\pm 1.2)$ SD, $1.8 (\pm 1.0)$ SD and $1.8 (\pm 0.6)$ SD capillary dilations per linear mm were observed ($p < 0.001$) (table 2, figure 3B).

Table 2. Comparison of the quantitative and qualitative NVC-assessment in the different subgroups compared to HC (n=408).

	HC n=204	PRP n=20	cSLE n=20	JDM n=21	JSSc n=13	ISc n=21	MCTD n=18	JIA n=91
QUANTITATIVE ASSESSMENT								
Density								
Mean capillary density/linear mm, \pm SD MD, (99.3CI)	8.5 \pm 1.2	8.7 \pm 0.8 0.2 (-0.6;1.0) p=0.467	7.6 \pm 1.3 -0.8 (-1.6;0.0) p=0.006	6.3 \pm 2.0 -2.1 (-2.9;-1.4) p<0.001	5.2 \pm 1.9 -3.2 (-4.2;-2.2) p<0.001	7.7 \pm 1.2 -0.8 (-1.6;0.0) p=0.005	7.1 \pm 1.3 -1.4 (-2.3;-0.5) p<0.001	8.7 \pm 1.2 0.2 (-0.2;0.6) p=0.229
Dimensions								
Mean number of dilations/linear mm, \pm SD MD, (99.3CI)	0.5 \pm 0.6	1.0 \pm 0.9 0.4 (0.0;0.9) p=0.008	0.9 \pm 1.1 0.3 (-0.1;0.8) p=0.035	1.5 \pm 1.2 1.1 (0.6;1.5) p<0.001	1.8 \pm 0.6 1.3 (0.7;1.8) p<0.001	0.4 \pm 0.4 -0.1 (-0.5;0.4) p=0.675	1.8 \pm 1.0 1.2 (0.8;1.7) p<0.001	0.4 \pm 0.6 -0.1 (-0.3;0.2) p=0.441
Subjects with giants, n (%) OR (99.3CI)	3 (1.5) 4.5 (0.1;63.1) p=0.185	1 (5.0) 4.4 (0.1;58.5) p=0.188	1 (5.0) 4.4 (0.1;58.5) p=0.188	12 (57.1) 71.8 (13.2;627.7) p<0.001	11 (84.6) 283.7 (33.2;5229.1) p<0.001	0 (0.0) 1.3 (0.0;31.0) p=0.869	10 (55.6) 73.3 (12.0;701.3) p<0.001	1 (1.1) 0.9 (0.0;11.7) p=0.943
Morphology								
Mean number of abnormal shapes/linear mm, \pm SD, MD, (99.3CI)	0.3 \pm 0.3	0.2 \pm 0.3 0.0 (-0.3;0.2) p=0.675	0.3 \pm 0.4 0.1 (-0.2;0.3) p=0.452	0.9 \pm 1.0 0.7 (0.4;0.9) p<0.001	0.5 \pm 0.4 0.2 (-0.1;0.6) p=0.043	0.4 \pm 0.6 0.2 (-0.1;0.4) p=0.059	0.6 \pm 0.4 0.3 (0.1;0.6) p=0.001	0.2 \pm 0.3 0.0 (-0.2;0.1) p=0.719
Microhemorrhages								
Subjects with microhemorrhages, n (%) OR (99.3CI)	80 (39.2) 1.6 (0.4;5.9) p=0.317	11 (55.0) 1.6 (0.4;5.9) p=0.317	16 (80.0) 5.2 (1.3;29.8) p=0.001	17 (85.3) 6.2 (1.6;35.4) p<0.001	11 (84.6) 6.8 (1.2;80.8) p=0.002	8 (38.1) 1.0 (0.3;3.5) p=0.964	14 (77.8) 4.6 (1.1;26.8) p=0.003	27 (29.7) 0.7 (0.3;1.4) p=0.134

Table 2. Comparison of the quantitative and qualitative NVC-assessment in the different subgroups compared to HC (n=408). (continued)

	HC n=204	PRP n=20	cSLE n=20	JDM n=21	jSSc n=13	ISc n=21	MCTD n=18	JIA n=91
QUALITATIVE ASSESSMENT								
Scleroderma pattern								
Subjects with a scleroderma pattern, n (%), OR (99.3CI)	3 (1.5)	1 (5.0) 4.4 (0.1;61.3) <i>p=0.189</i>	3 (15.0) 11.4 (1.1;114.8) <i>p=0.005</i>	13 (61.9) 85.3 (15.8;748.4) <i>p<0.001</i>	12 (92.3) 465.9 (45.3;18977.3) <i>p<0.001</i>	0 (0.0) 1.3 (0.0;31.3) <i>p=0.861</i>	11 (61.1) 85.7 (14.3;812.4) <i>p<0.001</i>	1 (1.1) 1.0 (0.0;12.0) <i>p=0.959</i>
Non-scleroderma patterns								
Subjects with a non-specific pattern, n (%), OR (99.3CI)	73 (35.8)	12 (60.0) 2.2 (0.6;8.5) <i>p=0.096</i>	15 (75.0) 4.5 (1.2;22.2) <i>p=0.002</i>	4 (19.0) 0.4 (0.1;1.8) <i>p=0.131</i>	1 (7.7) 0.2 (0.0;1.4) <i>p=0.031</i>	7 (33.3) 1.0 (0.2;4.0) <i>p=0.957</i>	7 (38.9) 1.1 (0.2;4.0) <i>p=0.901</i>	24 (26.4) 0.6 (0.3;1.4) <i>p=0.118</i>
Subjects with a normal pattern, n (%)	128 (62.7)	7 (33.3)	2 (10.0)	4 (19.0)	0 (0.0)	14 (66.7)	0 (0.0)	66 (72.5)

CI: confidence intervals; cSLE, childhood onset systemic lupus erythematosus; HC, healthy controls; JDM, juvenile dermatomyositis; JIA, juvenile idiopathic arthritis; jSSc, juvenile systemic sclerosis; ISc, localized scleroderma; MCTD, mixed connective tissue disease; MD: mean difference (compared to HC); OR: odds ratio compared to HC; PRP, primary Raynaud's phenomenon; SD, standard deviation.

The mean differences (MD) (by multivariable regression analysis, adjusted for the matching factors age and gender) and odd's ratios per subgroup are compared to HC, showing the raw *p*-values and the Bonferroni corrected CI. When a significant difference (*p*<0.007, α -level set at 0.007, as seven subgroups were compared) between the subgroup and the HC group existed, these values are indicated in bold.

Nailfold videocapillaroscopy in cSLE

In jSSc, 37.09% of the capillaries were dilated and 15.96% of the capillaries were giants versus 5.60% dilations and 0.03% giants in HC ($p < 0.001$). The same trends were observed in MCTD and JDM with respectively 26.33%, and 28.89% of dilated capillaries and 5.03% and 4.76% of giants ($p < 0.001$).

Capillary morphology

The mean number of abnormal capillary shapes per linear mm in HC was 0.3 (± 0.3 SD) and about the same values were observed in PRP, cSLE, lSc and JIA. JDM and MCTD subjects exhibited significantly more abnormal shapes compared to HC, being 0.9 (± 1.0 SD) per linear mm and 0.6 (± 0.4 SD) per linear mm ($p < 0.001$) (table 2, figure 3C).

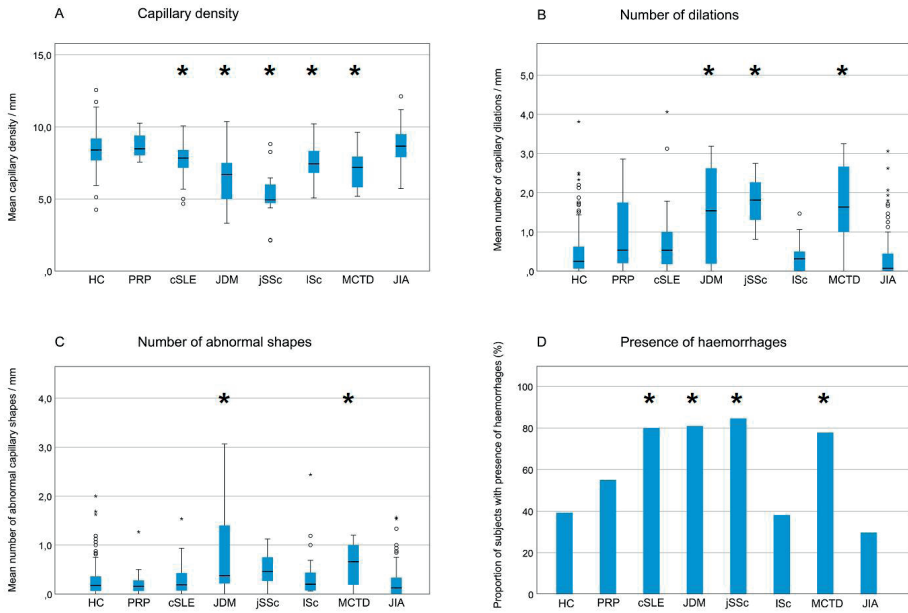


Figure 3. Quantitative NVC-assessment per subgroup, compared to the healthy subset (HC). cSLE, childhood onset systemic lupus erythematosus; HC, healthy controls; JDM, juvenile dermatomyositis; JIA, juvenile idiopathic arthritis; jSSc, juvenile systemic sclerosis; lSc, localized scleroderma; MCTD, mixed connective tissue disease; PRP, primary Raynaud’s phenomenon. A) Box plot of the capillary density per linear mm; B) Box plot of the number of dilations per linear mm; C) Box plot of the number of abnormal shapes per linear mm. Variations in the subgroups are evidenced by the wide blue boxes and by the presence of outliers ($\leq 1.5 \times$ interquartile range) and extreme values ($\geq 3 \times$ interquartile range); D) Bar graph of the proportion of subjects with presence of nailfold microhemorrhages. The statistical significant differences between each subgroup and the overall HC subgroup are indicate with an asterisk (*).

Of note, although the mean number of abnormal shapes in jSSc, being 0.5 (± 0.4 SD) per linear mm, was not significantly different from HC (after Bonferroni correction), we found in a post-hoc analysis that the proportion of the mean number of abnormal shapes on the mean capillary density (number of abnormal shapes/total number of capillaries), did reveal a significant difference (12.15% in jSSc versus 3.25% in HC, $p < 0.001$).

Presence of microhemorrhages

Figure 3D shows the proportions of subjects with microhemorrhages per subgroup. In 39.2% of the HC, microhemorrhages were found. The proportion increased in PRP to 55.0% ($p = 0.317$) and was lower in JIA (29.7%, $p = 0.134$). Significantly more microhemorrhages were observed in patients with CTD: cSLE 80.0%, JDM 85.3%, jSSc 84.6%, and MCTD 77.8%. The OR are represented in table 2.

Qualitative assessment

A “normal pattern” was observed in 62.7% of HC and was present in similar proportions in JIA and lSc. The “non-specific pattern” was the most dominant pattern in PRP (60.0%) and cSLE (75.0%), however, only in cSLE was it statistically significantly different from the proportion in HC (OR 4.5, CI 1.2-22.2, $p = 0.002$). The odds to exhibit a “scleroderma pattern” was higher in the CTD-subsets compared to the odds in HC (table 2).

A “scleroderma pattern” was observed in three out of 204 HC (1.5%), in one out of 20 PRP (5.0%) and in one out of 91 JIA-patients (1.0%). In all five of them, clinical signs of an underlying CTD were absent. One of the HC with a “scleroderma pattern” had RP (a 16-year-old white girl). Another HC had a local trauma at the same finger in which giants were observed (a 6-year-old black boy). In the last “healthy” child, no explanation for this observed abnormality was at hand (a 14-year-old boy with mixed ethnicities (white/black)). The boy with PRP, whom had a “scleroderma pattern”, had no antinuclear antibodies at multiple occasions and no jRMD-related symptoms, but reported a compulsive habit of nail biting. The boy with JIA, whom had a “scleroderma pattern”, reported on nail biting as well.

Exploratory analysis of age-related nailfold capillaroscopic findings in HC

No age-related increase in capillary density was observed ($R = -0.20$, $p = 0.773$) (figure 4A). On the contrary, the youngest age group (less than five-year-old children) exhibited a significantly higher capillary density compared to the other age groups. The Pearson’s correlation coefficient between the capillary density and age, by leaving out the youngest age group, resulted in a significant, though very weak positive correlation ($R = 0.14$, $p = 0.046$) (figure 4B).

There was a mild association between age and the number of capillary dilations, as reflected in a positive, but also small, correlation coefficient ($R=0.18$, $p=0.011$) (figure 4C). And there was no association with the number of abnormal shapes ($R=-0.02$, $p=0.758$) (figure 4D). No age-influence was found for the presence of microhemorrhages (data not shown).

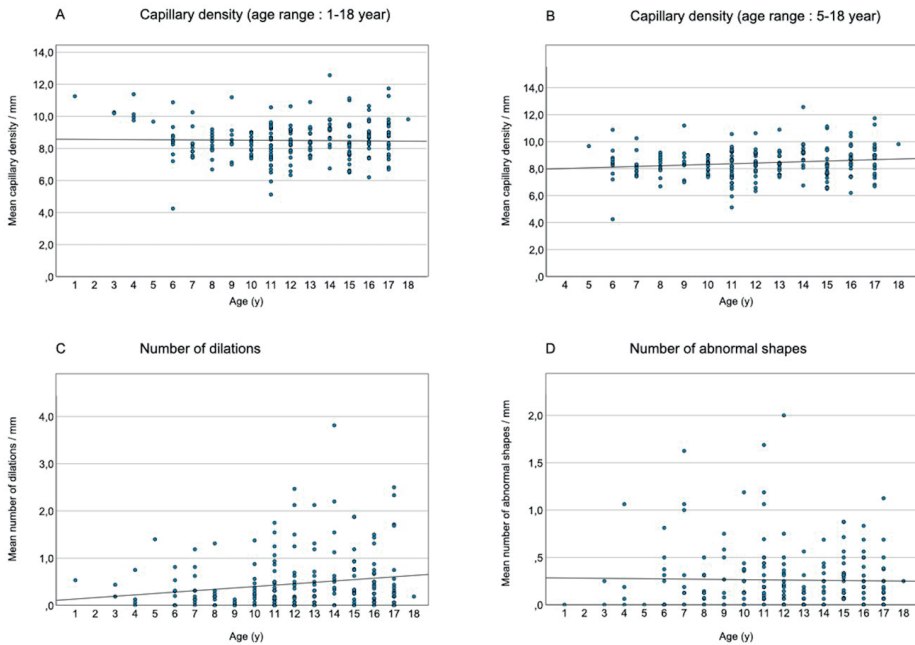


Figure 4. Exploratory analysis of the quantitative NVC-assessment in relation to age in the healthy subset. Scatter plot and fitted regression lines of: A) the capillary density in total healthy subset ($n=204$); B) the capillary density in healthy subset without the youngest group ($n=197$); C) the mean number of capillary dilations per linear mm ($n=204$); D) the mean number of abnormal capillary shapes per linear mm ($n=204$).

DISCUSSION

This study is a first multi-centre analysis of data from the international paediatric NVC-registry, set up by the EULAR Study Group of Microcirculation in Rheumatic Diseases. The NVC-characteristics of patients with varying jRMD and PRP are for the first time described in a standardised way and compared to a large group of HCs. The study reveals that the NVC-assessment in CTD such as jSSc, MCTD, JDM and cSLE differs significantly from HC. The NVC-characteristics from a large sample of JIA-patients are comparable with those from HC.

This study is one of the first of its kind, in that the NVC-images were assessed quantitatively and qualitatively, according to the standardised international consensus definitions from the EULAR Study Group on Microcirculation in Rheumatic Diseases [7, 20, 39]. The results are in accordance with the few previous publications on this topic in jRMD, in which a lowered capillary density is attested in jSSc, MCTD and JDM [24-29]. Additionally, we observed a lower capillary density in cSLE and lSc [28-30]. Interestingly, the mean capillary density in HC was 8.5/linear mm (± 1.2 SD), which is in the normal range for density in adults (where the cut-off value is 7/linear mm) and may indicate that the same cut-off values for normality can be used in children [8].

Although Piotto *et al.* had previously found a strong correlation between density and age in 100 healthy children (older than 5 years) ($R=0.796$, $p<0.001$), we can only at best report that there is a very weak association in our healthy subgroup ($R=0.14$, $p=0.046$) [18]. Other reports on this topic from Herrick *et al.* and Ingegnoli *et al.* are more in line with our results [28, 31].

It is novel in the capillaroscopic research of jRMD to report on capillaroscopic dimension as the mean number of dilations per mm [18, 29, 32]. We observed a significantly higher number of dilations, and also a higher proportion of subjects with giants in jSSc, MCTD and JDM compared to HC, consistent with observations in adults [33-35]. In keeping with the findings of Herrick *et al.*, our subgroup analysis in HC revealed a trend for the number of dilations to rise with age [31].

So far, only one study in children reports on the capillary morphology by using the standardised “simple” definitions from the EULAR Study Group on Microcirculation in Rheumatic Diseases [5, 7, 18, 32, 34, 39]. Concerning the presence of microhemorrhages, we observed higher prevalence rates than previously reported in HC (39% compared to prevalence rates ranging from 10% till 20% in literature) [28, 32]. Keeping in mind that as it stands, no reliable distinction between pathological and non-pathological types of microhemorrhages can be made, especially in children who are more frequently exposed to (micro)traumatic events [32, 36].

Not unexpectedly, we observed “scleroderma patterns” in CTDs and “non-specific abnormalities” as the predominant pattern in cSLE [24, 25, 27, 28, 37-39]. Remarkably, in three out of five “unexpected” cases with a “scleroderma pattern”, trauma or the compulsive habit of nail biting was noted, which underlines the need of repeated assessments in these cases and careful follow-up.

The consecutive input from centres dispersed across Europe, the American continent and South-Asia is a major advantage of our study. Despite the dominance of the white ethnicity within the study population, we obtained images from a relatively larger proportion of JIAS-patients from the South-Asian subcontinent (40%) and from a relatively larger proportion of black patients amongst those suffering from cSLE, which reflect data from a real-life world study population. Both diseases are known to have a higher prevalence in these ethnicities [40, 41].

Another advantage of our study is the standardised acquisition of NVC images from different centres with similar magnification (200X) and the centralised reading method. By using the videocapillaroscope, which is considered the gold standard device to obtain reliable nailfold images, and by applying a standardised methodology, we believe that our results add value for the implementation of the NVC-examination in clinical practice [7]. While instructions on the NVC-technique and capturing method were provided to the operators only by a written study protocol, the high reported general evaluability (97.6% of the jRMD-subjects) attests the applicability of this technique [18, 34]. However, it needs to be kept in mind that, while thoroughly validated in adults, the used NVC- technique and methodology needs further validation in children and adolescents.

This study has some limitations, linked to its exploratory nature. Firstly, the subgroups of patients with jRMD remain relatively small and no pairwise comparison with age- and sex matched healthy controls was performed. Likewise, for the interpretation of the age-related NVC-characteristics in the healthy subset, it is important to note that only seven children were below five years old. Secondly, inherent to the study design, we did not obtain reliable clinical information from our healthy controls. Children were mostly recruited at schools, in the absence of their parents. An underlying diagnosis of RP or rheumatic disease might as such have been missed Thirdly, our analysis did not correct for centre contribution because we were not able to match all jRMD patients with HC from the same centre and because samples per subgroup were very small for some centres.

To conclude, this study pioneers in the standardised NVC-assessment of children and adolescents with jRMD, recruited across the world and had the aim of describing their NVC-assessment in a detailed and comprehensive manner. Our results support the regular use of NVC in children with PRP, CTD and even lSc. A further step of this international project is to follow children and adolescents prospectively in order to shed light on clinical associations with NVC-abnormalities in larger disease specific samples. The EULAR Study Group of Microcirculation in Rheumatic Diseases advocates the use of

standardised NVC-assessments and terminology to improve the accuracy of NVC-studies and to facilitate comparisons in the future.

FUNDING STATEMENT

Vanessa Smith is a Senior Clinical Investigator of the Research Foundation – Flanders (Belgium) (FWO) [1.8.029.20N]. The FWO was not involved in study design, collection, analysis and interpretation of data, writing of the report, nor in the decision to submit the manuscript for publication. Vanessa Smith is supported by an unrestricted educational chair on systemic sclerosis of Janssen-Cilag NV. Janssen-Cilag NV was not involved in study design, collection, analysis and interpretation of data, writing of the report, nor in the decision to submit the manuscript for publication. Maria C Leone was a research fellow, who received an EULAR scientific training bursary, during the data acquisition period of the study.

ACKNOWLEDGEMENTS

The study was conducted on behalf of the EULAR Study Group on Microcirculation in Rheumatic Diseases. The Ghent University Hospital and Laboratory of Experimental Rheumatology of the University of Genova and IRCCS San Martino Hospital are member of the European Reference Network on Rare and Complex Connective Tissue and Musculoskeletal Diseases (ERN ReCONNET) and the Ghent University Hospital also of the Flemish Network on rare connective tissue diseases. We acknowledge the support given by the Fund for Scientific Research in Rheumatology and the Fund Aline, managed by the King Baudouin Foundation to Vanessa Smith. We are grateful to the schools which have allowed a substantial recruitment for healthy controls: “De Wassenaar” in Bruges-Belgium, “Sint Barbara College” and “Sint-Paulus” in Ghent-Belgium, “Sint-Pieterscollege” in Brussels-Belgium, and “Knotwilg”, “Piet Hein” in Amsterdam-the Netherlands and “Burgemeester Amersfoordtschool” in Badhoevedorp-the Netherlands. We thank the study-coordinators and investigators at each participating site for their logistical support. We acknowledge The EULAR Study Group on Microcirculation in Rheumatic Diseases-collaborators/authors: Liselotte Deroo, Amber Vanhaecke, Sandy Bergkamp, Amara Nassar-Sheikh Rashid, Carmen Pizzorni, Patrone Elisa, Walter Hermann, Francesco Bica, Francesco Cattelan, Maurizio Gattinara, Zeynep Özcan, Sara Murias, Rosa Alcobendas, Augustin Remesal, Pallavi Pimpale, Raju Khubchandani and Dorota Krasowska.

SUPPLEMENTARY FILE 1: Centre contributions and EC approval numbers

Centre name (EC approval number)	HC N=204	PRP n=20	jRMD n=189
Ghent University Hospital, Ghent-Belgium (B670201627545)	62*	0	29
Brugmann University Hospital, Brussels-Belgium (B670201627545)	0	0	13
Amsterdam UMC, Amsterdam-The Netherlands (NL60885.018.17)	88*	7	33
University of Genoa and "Giannina Gaslini" Children Hospital, Genoa-Italy (392REG2017)	13	2	26
Gaetano Pini Hospital, Milan-Italy (752REG2017)	0	0	7
An der Schön Klinik Hamburg-Eilbek, Hamburg-Germany (PV5373)	3	10	35
Klinik für Allgemeine Pädiatrie und Neonatologie, Giessen-Germany (AZ 66/17)	10	0	14
Medical University of Lublin, Lublin-Poland (KE-0254/44/2017)	0	0	3
University Hospital La Paz, Madrid-Spain (PI-2635)	5	0	12
Mayo Clinic College of Medicine, Rochester-USA (18-003557)	21	0	3
University of Utah, Utah-USA (00118171)	1	1	2
Clinica Universitaria Bolivariana, Medellin-Colombia **	0	0	0
SRCC Children's Hospital, Mumbai-India (R-201901)	1	0	12

Table S1. Contributions per participating centre for healthy controls (HC), primary Raynaud's phenomenon (PRP), and juvenile diseases (jRMD) (n=413)

* Data obtained from schools

** Data transfer contract had not been finished at the moment of data analysis of this study.

SUPPLEMENTARY FILE 2: Extractable nuclear antigen specifications per subgroup

	PRP n=20	cSLE n=20	JDM n=22	jSSc n=13	lSc n=21	MCTD n=18	JIA n=95
ANA, n/total valid (%)	5/20 (25.0)	17/18 (94.4)	13/16 (81.2)	13/13 (100.0)	2/11 (18.2)	14/16 (87.5)	35/59 (89.8)
dsDNA, n (%)		6 (33.3)					
RNP, n (%)		8 (44.4)		1 (7.7)		11 (68.7)	
Sm, n (%)		4 (22.2)				3 (18.7)	
CenP, n (%)				1 (7.7)	1 (9.1)		1 (1.7)
PM/Scl, n (%)			1 (6.2)	3 (23.1)			
Scl70, n (%)				3 (23.1)			
Th/To, n (%)				2 (15.4)			
SSA and/or SSB, n (%)		6 (33.3)	1 (6.2)	1 (7.7)		4 (25.0)	
Ku, n (%)						1 (6.2)	
RibP, n (%)		2 (11.1)					
NXP2, n (%)			1 (4.5)				

Table S2. Specifications on the extractable nuclear antigen per disease subgroup (n=209)

ANA, anti-nuclear antibodies detected by indirect immunofluorescence-screening on Hep-2 cell substrates; CenP, anti-centromere protein antibody; cSLE, childhood onset systemic lupus erythematosus; JDM, juvenile dermatomyositis; JIA, juvenile idiopathic arthritis; jSSc, juvenile systemic sclerosis; lSc, localized scleroderma; MCTD, mixed connective tissue disease; NXP2, nuclear matrix protein antibodies; PM/Scl, anti-exome antibodies; PRP, primary Raynaud's phenomenon; RibP, anti-ribosomal P antibodies; RNP, ribonucleoprotein antibodies; Sm: anti-Smith antibodies.

SUPPLEMENTARY FILE 3: Evaluability and Matching

	HC n=204	PRP n=20	cSLE n=20	JDM n=22	jSSc n=13	lSc n=21	MCTD n=18	JIA n=95				
								OA n=22	PA n=23	ERA n=20	PsA n=15	JIAS n=15
Subjects with <4/16 "good" images, n (%)	0 (0.0)	0 (0.0)	0 (0.0)	1 (4.5)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (6.7)	3 (20.0)
Subjects with 4-8/16 "good" images, n (%)	8 (3.4)	1 (5.0)	2 (10.0)	2 (9.1)	0 (0.0)	0 (0.0)	0 (0.0)	1 (4.5)	2 (8.7)	0 (0.0)	1 (6.7)	2 (13.3)
Subjects with 9-12/16 "good" images, n (%)	19 (9.3)	0 (0.0)	2 (10.0)	0 (0.0)	3 (23.1)	0 (0.0)	1 (5.6)	6 (27.3)	3 (13.0)	5 (25.0)	2 (13.3)	2 (13.3)
Subjects with 13-16/16 "good" images, n (%)	177 (86.8)	19 (95.0)	16 (80.0)	19 (86.4)	10 (76.9)	21 (100.0)	17 (94.4)	15 (68.2)	18 (78.3)	15 (75.0)	11 (78.6)	8 (53.3)

Table S3. Visibility of NVC-images per subgroup (n=413)

An image was considered as a "good" image when all parameters were evaluable. On the other hand, an image was scored as "bad" if any capillaroscopic parameter was not evaluable. A "bad" image was still included for analysis, but the concerning parameter was then encoded as a missing. Standardly, 16 images were taken at subject level. Subjects with less than four "good" images were excluded for further analysis (n=5).

REFERENCES

1. Cutolo M. Atlas of capillaroscopy in rheumatic diseases: Elsevier srl, 2015.
2. Cutolo M, Pizzorni C, Sulli A, Smith V. Early diagnostic and predictive value of capillaroscopy in systemic sclerosis. *Curr Rheumatol Rev.* 2013; 9(4):249-253.
3. Koenig M, Joyal F, Fritzler MJ, Roussin A, Abrahamowicz M, Boire G, et al. Autoantibodies and microvascular damage are independent predictive factors for the progression of Raynaud's phenomenon to systemic sclerosis: a twenty-year prospective study of 586 patients, with validation of proposed criteria for early systemic sclerosis. *Arthritis Rheum.* 2008 Dec; 58(12):3902-3912.
4. van den Hoogen F, Khanna D, Fransen J, Johnson SR, Baron M, Tyndall A, et al. 2013 classification criteria for systemic sclerosis: an American college of rheumatology/European league against rheumatism collaborative initiative. *Ann Rheum Dis.* 2013; 72(11):1747-1755.
5. Cutolo M, Melsens K, Herrick AL, Foeldvari I, Deschepper E, De Keyser F, et al. Reliability of simple capillaroscopic definitions in describing capillary morphology in rheumatic diseases. *Rheumatology (Oxford).* 2018 Apr 1; 57(4):757-759.
6. Dinsdale G, Moore T, O'Leary N, Tresadern P, Berks M, Roberts C, et al. Intra-and inter-observer reliability of nailfold videocapillaroscopy - A possible outcome measure for systemic sclerosis-related microangiopathy. *Microvascular research.* 2017 Jul; 112:1-6.
7. Smith V, Herrick AL, Ingegnoli F, Damjanov N, Angelis, Denton CP, et al. Standardisation of nailfold capillaroscopy for the assessment of patients with Raynaud's phenomenon and systemic sclerosis. *Autoimmunity reviews.* 2020 Jan 9:102458.
8. Smith V, Vanhaecke A, Herrick AL, Distler O, Guerra MG, Denton CP, et al. Fast track algorithm: How to differentiate a "scleroderma pattern" from a "non-scleroderma pattern". *Autoimmunity reviews.* 2019 Sep 11:102394.
9. Ingegnoli F, Herrick AL. Nailfold capillaroscopy in pediatrics. *Arthritis care & research.* 2013 Sep; 65(9):1393-1400.
10. Alarcon-Segovia D, Cardiel MH. Comparison between 3 diagnostic criteria for mixed connective tissue disease. Study of 593 patients. *The Journal of rheumatology.* 1989 Mar; 16(3):328-334.
11. Bohan A, Peter JB. Polymyositis and dermatomyositis (first of two parts). *N Engl J Med.* 1975 Feb 13; 292(7):344-347.
12. Hochberg MC. Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus. *Arthritis and rheumatism.* 1997 Sep; 40(9):1725.
13. Kasukawa R, Tojo T, Miyawaki S. Preliminary diagnostic criteria for classification of mixed connective tissue disease. In: Kasukawa R, Sharp G, eds. *Mixed Connective Tissue Disease and Antinuclear Antibodies.* Amsterdam: Elsevier:41.
14. Petri M, Orbai AM, Alarcon GS, Gordon C, Merrill JT, Fortin PR, et al. Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis and rheumatism.* 2012 Aug; 64(8):2677-2686.
15. Petty RE, Southwood TR, Baum J, Bhattay E, Glass DN, Manners P, et al. Revision of the proposed classification criteria for juvenile idiopathic arthritis: Durban, 1997. *J Rheumatol.* 1998 Oct; 25(10):1991-1994.
16. Zulian F, Woo P, Athreya BH, Laxer RM, Medsger TA, Jr., Lehman TJ, et al. The Pediatric Rheumatology European Society/American College of Rheumatology/European League against Rheumatism provisional classification criteria for juvenile systemic sclerosis. *Arthritis and rheumatism.* 2007 Mar 15; 57(2):203-212.

17. Herrick AL. The pathogenesis, diagnosis and treatment of Raynaud phenomenon. *Nature reviews Rheumatology*. 2012 Aug; 8(8):469-479.
18. Piotto DP, Sekiyama J, Kayser C, Yamada M, Len CA, Terreri MT. Nailfold videocapillaroscopy in healthy children and adolescents: description of normal patterns. *Clin Exp Rheumatol*. 2016 Sep-Oct; 34 Suppl 100(5):193-199.
19. University Ghent. MCRD6 - webshare [Internet]. 2016. Available from: www.mcrd6capped.ugent.be
20. Ingegnoli F, Herrick AL, Schioppo T, Bartoli F, Ughi N, Pauling JD, et al. Reporting items for capillaroscopy in clinical research on musculoskeletal diseases: a systematic review and international Delphi consensus. *Rheumatology (Oxford, England)*. 2021 Mar 2; 60(3):1410-1418.
21. Cutolo M, Melsens K, Wijnant S, Ingegnoli F, Thevissen K, De Keyser F, et al. Nailfold capillaroscopy in systemic lupus erythematosus: A systematic review and critical appraisal. *Autoimmunity reviews*. 2018 Apr; 17(4):344-352.
22. Smith V, Beeckman S, Herrick AL, Decuman S, Deschepper E, De Keyser F, et al. An EULAR study group pilot study on reliability of simple capillaroscopic definitions to describe capillary morphology in rheumatic diseases. *Rheumatology (Oxford, England)*. 2016 May; 55(5):883-890.
23. Greifer N. Matching methods [Internet]. [updated 2022, March 7; cited 2022 Mar 20]. Available from: <https://cran.r-project.org/web/packages/MatchIt/vignettes/matching-methods.html#caliper-matching-caliper>,
24. Spencer-Green G, Schlesinger M, Bove KE, Levinson JE, Schaller JG, Hanson V, et al. Nailfold capillary abnormalities in childhood rheumatic diseases. *The Journal of pediatrics*. 1983 Mar; 102(3):341-346.
25. Silver RM, Maricq HR. Childhood dermatomyositis: serial microvascular studies. *Pediatrics*. 1989 Feb; 83(2):278-283.
26. Scheja A, Elborgh R, Wildt M. Decreased capillary density in juvenile dermatomyositis and in mixed connective tissue disease. *The Journal of rheumatology*. 1999 Jun; 26(6):1377-1381.
27. Dolezalova P, Young SP, Bacon PA, Southwood TR. Nailfold capillary microscopy in healthy children and in childhood rheumatic diseases: a prospective single blind observational study. *Annals of the rheumatic diseases*. 2003 May; 62(5):444-449.
28. Ingegnoli F, Zeni S, Gerloni V, Fantini F. Capillaroscopic observations in childhood rheumatic diseases and healthy controls. *Clin Exp Rheumatol*. 2005 Nov-Dec; 23(6):905-911.
29. Piotto DG, Len CA, Hilário MO, Terreri MT. Nailfold capillaroscopy in children and adolescents with rheumatic diseases. *Rev Bras Reumatol*. 2012; 52(5):722-732.
30. Schonenberg-Meinema D, Melsens K, Nassar-Sheikh Rashid A, Cutolo M, Kuijpers TW, van den Berg JM, et al. Capillaroscopy in childhood-onset systemic lupus erythematosus: a first systematic review. *Clinical and experimental rheumatology*. 2020 Mar-Apr; 38(2):350-354.
31. Herrick AL, Moore T, Hollis S, Jayson MI. The influence of age on nailfold capillary dimensions in childhood. *The Journal of rheumatology*. 2000 Mar; 27(3):797-800.
32. Terreri MT, Andrade LE, Puccinelli ML, Hilario MO, Goldenberg J. Nail fold capillaroscopy: normal findings in children and adolescents. *Semin Arthritis Rheum*. 1999 Aug; 29(1):36-42.
33. Hajas A, Szodoray P, Nakken B, Gaal J, Zold E, Laczik R, et al. Clinical course, prognosis, and causes of death in mixed connective tissue disease. *The Journal of rheumatology*. 2013 Jul; 40(7):1134-1142.
34. Ingegnoli F, Ardoino I, Boracchi P, Cutolo M, co-authors E. Nailfold capillaroscopy in systemic sclerosis: data from the EULAR scleroderma trials and research (EUSTAR) database. *Microvasc Res*. 2013 Sep; 89:122-128.

35. Manfredi A, Sebastiani M, Campomori F, Pipitone N, Giuggioli D, Colaci M, et al. Nailfold Videocapillaroscopy Alterations in Dermatomyositis and Systemic Sclerosis: Toward Identification of a Specific Pattern. *The Journal of rheumatology*. 2016 Aug; 43(8):1575-1580.
36. Bergkamp SC, Schonenberg-Meinema D, Nassar-Sheikh Rashid A, Melsens K, Vanhaecke A, Boumans MJH, et al. Reliable detection of subtypes of nailfold capillary haemorrhages. *Clin Exp Rheumatol*. 2021.
37. Cony M, Klene-Boudard C, Fontan I, Sanciaume C, Sarrat P, Taieb A, et al. [Periungual capillaroscopy patterns in normal children]. *Archives francaises de pediatrie*. 1992 Mar; 49(3):171-174.
38. Russo RA, Katsicas MM. Clinical characteristics of children with Juvenile Systemic Sclerosis: follow-up of 23 patients in a single tertiary center. *Pediatric rheumatology online journal*. 2007 May 1; 5:6.
39. Schonenberg-Meinema D, Bergkamp SC, Nassar-Sheikh Rashid A, van der Aa LB, de Bree GJ, Ten Cate R, et al. Nailfold capillary abnormalities in childhood-onset systemic lupus erythematosus: a cross-sectional study compared with healthy controls. *Lupus*. 2021 Mar 3:961203321998750.
40. McCarty DJ, Manzi S, Medsger TA, Jr., Ramsey-Goldman R, LaPorte RE, Kwok CK. Incidence of systemic lupus erythematosus. Race and gender differences. *Arthritis and rheumatism*. 1995 Sep; 38(9):1260-1270.
41. Saurenmann RK, Rose JB, Tyrrell P, Feldman BM, Laxer RM, Schneider R, et al. Epidemiology of juvenile idiopathic arthritis in a multiethnic cohort: ethnicity as a risk factor. *Arthritis and rheumatism*. 2007 Jun; 56(6):1974-1984.

5

A nailfold capillary scleroderma pattern may be associated with disease damage in childhood-onset systemic lupus erythematosus: important lessons from longitudinal follow-up

Dieneke Schonenberg-Meinema, Sandy C. Bergkamp, Amara Nassar-Sheikh Rashid, Mariken P. Gruppen, Maritza A. Middelkamp, Wineke Armbrust, Koert M. Dolman, A. Elisabeth Hak, Petra C.E. Hissink Muller, Marieke van Onna, Joost F. Swart, Taco W. Kuijpers, Sylvia Kamphuis, Vanessa Smith, J. Merlijn van den Berg

Published in Lupus Science & Medicine. 2022 Feb;9(1):e000572

ABSTRACT

OBJECTIVES: To observe if capillary patterns in childhood-onset SLE (cSLE) change over time and find associations between a capillary scleroderma pattern with disease activity, damage or scleroderma-like features.

METHODS: Clinical and (yearly) capillaroscopy data from a longitudinal cohort of cSLE-patients (minimum of 4 SLICC criteria, onset < 18 years) were analyzed. Disease activity was measured by SLEDAI and disease damage by SDI. A scleroderma pattern was defined according to the 'Fast track algorithm' from the EULAR Study Group on Microcirculation in Rheumatic Diseases. An abnormal capillary pattern, not matching a scleroderma pattern, was defined as 'microangiopathy'.

RESULTS: Our cohort consisted of 53 cSLE patients with a median disease onset of 14 years (IQR 12.5-15.5 years), median SLEDAI score at diagnosis was 11 (IQR 8-16), median SLEDAI at follow up was 2 (IQR 1-6). A scleroderma pattern (ever) was seen in 18.9%, while only 13.2% of patients had a normal capillary pattern. Thirty-three patients had follow-up capillaroscopy of which 21.2% showed changes in type of capillary pattern over time. Type of capillary pattern was not associated with disease activity. Raynaud's phenomenon (ever) was equally distributed among patients with different capillaroscopy patterns ($p=0.26$). Anti-RNP antibodies (ever) were significantly more detected (Chi square, $p=0.016$) in the scleroderma pattern subgroup ($n=7/10$, 70%). Already 5 years after disease onset more than 50% of patients with a scleroderma pattern had SLE-related disease damage (HR 4.5, 95%CI 1.1-18.8, $p=0.034$), but they did not develop clinical features of systemic sclerosis at follow-up. Number of detected fingers with a scleroderma pattern was similar between cSLE, jSSc and jUCTD.

CONCLUSION: This longitudinal study shows that the majority of capillary patterns in cSLE are abnormal and they can change over time. Irrespective of disease activity, a capillary scleroderma pattern in cSLE may be associated with higher risk for SLE-related disease damage.

INTRODUCTION

Abnormalities in nailfold capillaries of systemic lupus erythematosus (SLE) patients, visualized by capillaroscopy, have been described in literature [1]. Two systematic literature reviews showed that these capillary abnormalities mainly consist of capillary hemorrhages and abnormal capillary shapes in adults with SLE but literature in childhood-onset SLE (cSLE) is scarce and mainly inconclusive [1, 2]. By qualitative analysis, capillary patterns in SLE are mainly described as ‘non-specific changes’ but a scleroderma pattern has also been described in SLE with percentages varying from 3 to 26% [3-9]. A scleroderma pattern in nailfold capillaries was first described in patients with systemic sclerosis (SSc) and is characterized by giant capillaries, sometimes in combination with large pathological capillary hemorrhages, loss of capillaries and abnormal capillary shapes with a deterioration of capillary architecture [10]. Studies have suggested that SLE-patients with a scleroderma pattern might be patients with (subclinical) overlapping features of other connective tissue diseases (CTD), such as SSc and dermatomyositis (DM) [8]. Additionally, the finding of a scleroderma pattern in SLE patients have been associated with the occurrence of Raynaud’s phenomenon as well as with the presence of anti-ribonucleic protein (RNP) antibodies. Even more, these patients seem at risk for the development of pulmonary arterial hypertension or pulmonary fibrosis [3, 7, 8, 11]. In a cross-sectional study we previously showed that a scleroderma pattern in nailfold capillaries can be observed in patients with cSLE, characterized by typical SLE symptoms like lupus nephritis, malar rash, serositis, aphthous ulcers, leukopenia, thrombocytopenia, but without any clinical signs of SSc [9].

Vasculopathy is an important feature of SSc and this is a dynamic process in this disease changing over time. Changes of the nailfold capillary pattern, both deterioration, improvement as well as complete normalization have been described during follow-up of SSc-patients [12-16]. In one of these longitudinal studies, the appearance of digital ulcers correlated with the type of capillary patterns identified over time [15]. Only one study with longitudinal follow-up of nailfold capillaroscopy in SLE patients is available, describing changes in capillary pattern in cSLE as well as adult onset SLE-patients. This study suggested that a scleroderma pattern might be considered a red flag for the potential development of scleroderma spectrum disorders such as SSc and mixed/undifferentiated connective tissue disease [11]. More longitudinal follow-up data are obviously needed for better understanding of the meaning of a scleroderma capillary pattern in SLE.

The primary aims of this longitudinal prospective study are to observe if (ab)normal capillary patterns in individual cSLE patients change over time and if so, if these (changes

in) capillary patterns associate with disease activity and disease damage. Additionally it is studied if a capillary scleroderma pattern in cSLE-patients indicates an increased risk for development of clinical symptoms for SSc over time. Secondary objective is to compare total finger counts with a scleroderma pattern in cSLE to patients diagnosed with juvenile systemic sclerosis (jSSc), juvenile dermatomyositis (JDM) and juvenile undifferentiated connective tissue disease (jUCTD) to give more value to this abnormal observation.

METHODS

Study design and patients

Between April 2016 and April 2021, patients with systemic auto-immune diseases who visited the (out)patient clinics of the Amsterdam UMC and Leiden UMC were included. Prospective capillaroscopy data were obtained in a cSLE cohort and three disease control cohorts (jSSc, JDM and jUCTD) visiting the (outpatient) clinics. Inclusion criteria for cSLE-patients were diagnosis according to the 2012 Systemic Lupus International Collaborating Clinics (SLICC) classification criteria [17] and age of disease onset < 18 years old. Patients were diagnosed with JDM and jSSc according to their respective EULAR/ACR criteria [18, 19]. Juvenile UCTD patients were defined as patients with Raynaud's phenomenon and anti-nuclear auto-immune antibodies, but without fulfilling the classification criteria for cSLE, JDM or jSSc. Capillaroscopy was part of routine clinical follow-up, demographic and clinical data were recorded from the patient charts. For cSLE patients that already diagnosed in the past (before capillaroscopy), retrospective clinical and demographical data were used from the time of diagnosis (autoimmune serology, SLEDAI at diagnosis and type of organ involvement).

All cSLE-patients visiting the (outpatient) clinic in Amsterdam UMC were asked to participate in this longitudinal cohort study, patients from Leiden UMC were asked to participate during two pre-planned visits using the same videocapillaroscope from Amsterdam UMC. Patients were excluded if it was impossible to collect images with good quality (due to nailfold skin thickness) or when a patient was too sick to undergo capillaroscopy examination. Demographic and clinical characteristics and disease activity were collected at the study visit. Age, gender, ethnic background, Raynaud symptoms (ever in time) and periungual trauma were noted. Disease onset was defined at the date of first SLE-symptom. Disease activity was measured using the Systemic Lupus Erythematosus Activity Index (SLEDAI)²⁰; disease damage by the SLICC damage index (SDI) [21]. Disease activity was also graded by severity: inactive or mild for SLEDAI <3, moderate for SLEDAI between 3-6 and severe for SLEDAI >6.

This study was approved by the ethical committee from the Amsterdam University Medical Centers (Dutch trial register registration no. NL60885.018.17). All patients were coded with a unique study number. For follow-up, according to the Dutch Medical Research Involving Human Subjects Act, an informed consent was signed by children from 12 years of age, and/or both parents (if alive and authorized) for children below 16 years old.

Nailfold capillaroscopy technique and image analysis

Nailfold videocapillaroscopy (NVC) was performed with a x200 magnification lens from Optilia. Images were collected by two investigators (DS or SB, respectively six and three years of experience in capillaroscopy examination). Before start of examination, the patients stayed in a room of 20-22 °C for a minimum of 15-20 minutes. Patients were in sitting position with the hand on a table at the level of their heart during capillaroscopy examination. A drop of oil was applied to the fingers before examination. In total, eight fingers per cSLE-patient (excluding the thumbs) were examined. Per finger, four images were stored.

Qualitatively, three capillary patterns were described. First, a scleroderma pattern was defined as extremely lowered density (≤ 3 capillaries / mm) with abnormal shapes or the presence of giants by 'fast track algorithm' according to the 'EULAR Study Group on Microcirculation in Rheumatic Diseases standardized capillaroscopy evaluation chart' [10, 22]. Second, a capillary pattern was designated as 'microangiopathy' (also described as 'non-specific changes') if the observed capillary pattern showed abnormal capillary morphology (according to the EULAR study group criteria) [22, 23] and/or capillary hemorrhages, but did not match the criteria for a scleroderma pattern. Third, a normal capillary pattern was defined if a patient did not have any capillary abnormalities. If the type of capillary pattern in a patient changed over time, the worst pattern was used in analyses. In case of microangiopathy or scleroderma pattern, the number of fingers demonstrating this abnormal pattern was counted.

Statistical analysis

Statistical analysis was performed with IBM SPSS Statistics type 26. Descriptive statistics were reported in terms of percentages, means and standard deviations or medians and inter-quartile ranges. For differences of variables between groups, Chi-square, Mann-Whitney U and Kruskal Wallis test analyses were used depending on (distribution, groups and number of categories of) outcome data. Logistic regression was used for occurrence of disease damage and scleroderma pattern as binary outcome data with odds ratio (OR) reported with 95% confidence interval (CI). Variables for univariate logistic regression were demographic (age and gender), auto-antibodies anti-RNP and anti-Sm, organ

manifestations and SLEDAI at diagnosis. Significant variables from univariate analyses and anti-RNP antibodies, nephritis and neuropsychiatric organ involvement (known to give risk for respectively overlap disease or damage from literature [7, 24]) were chosen as co-variables for multivariate logistic regression analysis. Longitudinal analysis (with significant variable(s) from univariate analysis) for comparison between different capillary patterns and disease damage as endpoint was performed by Cox regression analysis. P-values <0.05 were considered as statistically significant.

RESULTS

Of 56 eligible patients, capillary images could be analyzed in 53 cSLE-patients at the first study visit (Figure 1). Thirty-three of these 53 patients (62.3%) had a minimum of one follow-up visit for NVC with a range of 8-60 months after first capillaroscopy. N=14 and n=8 patients respectively had two and three follow-up visits for NVC. The reasons for no follow-up NVC (n=20) were: no consent (n=2), lost to follow-up (n=4), transition to adult care (n=6), <1 year of diagnosis (n=4), no time/logistic reason (n=3) and no evaluable images (n=1) (Figure 1). Clinical follow-up data in our longitudinal cSLE-cohort ranged from 0.5-16 years after disease onset. The baseline characteristics are listed in table 1. The patient groups with or without follow-up NVC were comparable for all baseline characteristics.

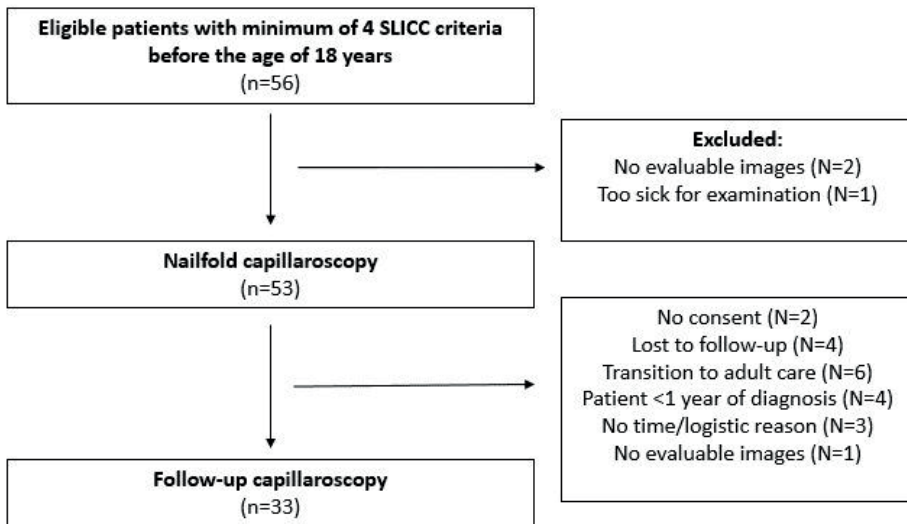


Figure 1. Flow chart longitudinal study cohort of cSLE-patients

Type of capillary patterns and change in time

At baseline, 13.2% (7/53) showed a capillary scleroderma pattern. In total in our cSLE cohort, 10/53 (18.9%) of all patients showed a nailfold scleroderma pattern (ever) at minimal one examination, 36/53 (67.9%) showed microangiopathy and 7/53 (13.2%) a normal capillary pattern. Range for NVC follow up time was 1-5 years. Figure 2 shows examples of nailfold capillary images with a scleroderma pattern in our cSLE-patients. Observed microangiopathy pattern consisted of abnormal capillary shapes and capillary hemorrhages (without the typical giant capillaries or extremely lowered density $\leq 3/\text{mm}$ as in a scleroderma pattern).

At longitudinal follow-up with NVC assessment, most patients (26/33, 78.8%) showed the same capillary pattern as baseline (see supplementary file 1). Of the 7 patients with a baseline capillary scleroderma pattern, in 5/7 patients NVC was repeated, which showed that 4/5 patients (80%) had a persistent scleroderma pattern and one patient (n=1/5, 20%) changed to a microangiopathy pattern (with severe active disease). Range of NVC follow-up in these patients with a scleroderma pattern was 14-50 months.

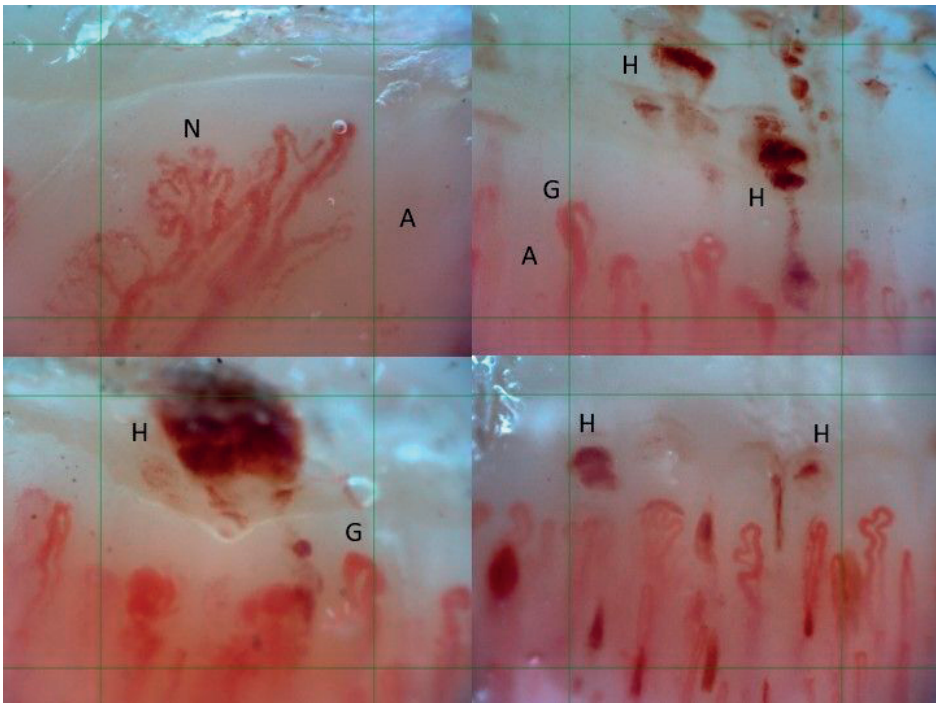


Figure 2. Capillary scleroderma patterns in patients with cSLE showing giant capillaries (G), hemorrhages (H) and neovascularization (= abnormal shapes) (N) with avascular areas (A).
Green grid: 1 mm.

Of the 39 patients with a baseline microangiopathy pattern, in 24/39 patients NVC was repeated. Images from one patient could not be interpreted because of indistinct visualization of capillaries (poor quality images). Eighteen of these patients (n=18/24, 75%) showed persistent microangiopathy and three patients (3/24, 12.5%) changed to a scleroderma pattern (two patients with inactive/mild and one with severe disease activity). In three other patients (3/24, 12.5%) microangiopathy changed to a normal pattern (two patients with inactive/mild disease and one with moderate disease activity). Range of NVC follow-up in these patients was 9-42 months.

Of the 7 patients with a normal capillary pattern, in 4/7 patients NVC was repeated. In all patients (n=4/4, 100%) this normal pattern persisted (two patients with inactive/mild disease and two with moderate disease activity). Range of NVC follow-up in these patients was 8-22 months.

Mean time between NVC in patients with a change in capillary pattern was 21.1 months (range 9-50 months). Mean time between NVC in patients without any change in pattern was comparable with 15.8 months (range 6-52 months) (see supplementary file 1). Almost all patients but one (with a worsening of capillary pattern to scleroderma pattern), with and without change in capillary pattern, were on treatment for SLE.

Two patients with a scleroderma pattern had digital lesions at presentation of their disease and thus at time of first capillaroscopy. At follow-up their digital lesions improved but the capillary scleroderma pattern persisted in both patients.

Capillary patterns and association disease activity/-damage

There was no significant difference in 'SLEDAI at diagnosis' between patients with different capillary patterns (sub-analysis of n=31 treatment-naive patients, Kruskal-Wallis, p=0.18). Overall, patients in follow-up had lower disease activity with median SLEDAI 2 (IQR 1.5-6) than patients at diagnosis with a median SLEDAI of 12 (IQR 8-17). Raynaud's phenomenon (ever) was equally distributed among patients with different capillaroscopy patterns (Chi square, p=0.26). Anti-RNP antibodies (all subtypes, ever in time) were significantly more detected (Chi square, p=0.016) in the scleroderma pattern subgroup than in the other groups, (in 42.8%, 22.2% and 70% of patients with a normal, microangiopathy and scleroderma pattern respectively). Of the anti-RNP positive patients with a scleroderma pattern, 85.7% (n=6/7) also showed positive anti-ds-DNA antibodies and 71.4% (n=5/7) also showed anti-Sm antibodies. The anti-RNP antibodies were directed against 70kD-protein in 7/18 patients, in the other patients the anti-RNP antibodies were directed against A- or C-protein. The more severe disease variables nephritis and neuropsychiatric involvement were equally distributed at diagnosis among different capillary

patterns (resp. $p=0.29$ and $p=0.44$). By univariate regression analysis, discoid rash (OR 9.3, 95% CI 2.0-42.8, $p=0.004$) and anti-RNP antibodies (OR 6.8, 95% CI 1.5-30.9, $p=0.013$) were significantly associated with the occurrence of a capillary scleroderma pattern. By multivariate regression analysis this was only seen for discoid rash (OR 5.9, 95% CI 1.2-30.2, $p=0.033$) (see supplementary file 2).

Table 1. Demographical variables and clinical characteristics of all cSLE patients (total and per subgroup with/without follow-up capillaroscopy).

	Total, n=53	Capillaroscopy follow-up, n=33	No capillaroscopy follow-up, n=20	p-value ^o
Female, n (%)	47 (88.7)	29 (87.9)	18 (90)	0.81
Ethnicity, n (%)				
African/Afro-Caribbean	21 (39.6)	14 (42.4)	7 (35)	0.23
White	21 (36.6)	13 (39.4)	8 (40)	
North-African/Middle-Eastern	4 (7.5)	3 (9.1)	1 (5)	
Asian	4 (7.5)	3 (9.1)	1 (5)	
Mixed/other	3 (5.7)	0	3 (15)	
Age at first capillaroscopy in years, median (IQR)	17 (14-17)	16 (14-17.5)	17 (14.3-17)	0.14
Raynaud's phenomenon / acro-cyanotic symptoms, n (%)	17 (32.1)	10 (30.3)	7 (35)	0.72
Age at onset in years, median (IQR 25-75)	14 (12.5-15.5)	15 (13.5-15.5)	13.5 (11.3-15.8)	0.22
Prednisone naïve, n (%)	31 (58.5)			
ANA at diagnosis, n (%)	52 (98.1)			
ANA + anti-ds-DNA	37 (69.8)	23 (69.7)	14 (70)	0.99
ANA + anti-RNP	18 (34)	14 (42.4)	4 (20)	0.10
ANA + anti-Sm	18 (34)	11 (33.3)	7 (35)	0.90
Cutaneous involvement, n (%)	38 (71.7)	22 (66.7)	17 (85)	0.14
Nephritis, n (%)	17 (32.1)	11 (33.3)	6 (30)	0.80
Neuropsychiatric involvement, n (%)	8 (15.1)	5 (15.2)	3 (15)	0.99
Antiphospholipid antibodies, n (%)	6 (11.3)	3 (9.1)	3 (15)	0.33
SLEDAI at diagnosis, median (IQR)	11 (8-16)	12 (8-17)	10 (6.5-14.8)	0.38
SLEDAI at first capillaroscopy, median (IQR)	6 (3.5-12)			
SLEDAI at second capillaroscopy, median (IQR)		2 (1-6)		
Capillary pattern: normal / microangiopathy / scleroderma pattern, n (%)	7 / 36 / 10 (13.2 / 67.9 / 18.9)	4 / 21 / 8 (12.1 / 63.6 / 24.2)	3 / 15 / 2 (15 / 75 / 10)	0.44
Disease damage present, n (%)	10 (18.9)	6 (18.2)	4 (20)	0.87

^o Chi square/Fisher exact analysis between the two subgroups

At final follow-up, disease damage (as measured by SDI), by univariate regression analysis, was significantly associated with detection of a capillary scleroderma pattern (ever) (OR 7.6 95% CI 1.6-35.9, $p=0.01$). In this univariate analysis neuropsychiatric involvement was also significantly associated with the risk of development of disease damage (OR 6.5 95%CI 1.3-33.2, $p=0.024$) as was discoid lupus (OR 5.1, 95% CI 1.2-22.6, $p=0.003$). By multivariate analysis, only neuropsychiatric involvement was a significant variable for occurrence of disease damage (OR 7.8, 95% CI 1.2-15.4, $p=0.032$) (see supplementary file 3). Cox regression analysis for occurrence of disease damage between patients with versus without a scleroderma pattern showed a significantly higher hazard for developing disease damage (Figure 3, Hazard Ratio (HR) 4.6 (95%CI 1.1-18.8, $p=0.034$). In this longitudinal regression model for disease damage, neuropsychiatric involvement was again a significant co-variable (HR 8.1, 95% CI 1.9-35.8, $p=0.005$), whereas discoid rash was not ($p=0.158$). Specific details on disease damage were end-stage renal disease ($n=1$), growth failure ($n=1$), extensive scarring in face ($n=2$), avascular skeletal necrosis ($n=2$), cognitive impairment/major psychosis ($n=1$), history of cerebrovascular accident ($n=3$), loss of digits ($n=1$), cardiac valve disease ($n=1$) and seizures requiring therapy for > 6 months ($n=1$). In figure 3 is seen that, between 4-5 years after disease onset, half of the cSLE-patients with a capillary scleroderma pattern had irreversible disease damage compared to <10% of cSLE-patients without a capillary scleroderma pattern.

Patients with a capillary scleroderma pattern did not show criteria to diagnose SSc (puffy hands, sclerodactyly or skin thickening, telangiectasia, interstitial lung disease or digital tip ulcers) or overlap disease during follow-up (table 2).

Capillary patterns in cSLE compared to disease controls

At time of first capillaroscopy, the median number of affected fingers was 5 (IQR 3-7) with microangiopathy and 4 (IQR 2-6) with a scleroderma pattern. At second capillaroscopy, the median number of affected fingers was 6.5 (IQR 2.3-8) with microangiopathy and with a scleroderma pattern 6 (IQR 4.5-7). Table 3 shows the number of fingers with a capillary scleroderma pattern in cSLE compared with patient cohorts of JDM ($n=12$), jUCTD ($n=13$) and jSSc ($n=7$). At first NVC and compared with cSLE, the number of fingers with a scleroderma pattern was significantly higher in patients with JDM, but this was not significantly different for patients with jSSc or jUCTD versus cSLE, nor was it different at follow-up.

Table 2. Clinical characteristics of cSLE-patients (n=10/53) with a capillary scleroderma pattern

SLE- characteristics	Patient 1	Patient 2	Patient 3	Patient 4	Patient 5	Patient 6	Patient 7	Patient 8	Patient 9	Patient 10
Butterfly rash	+	+	-	-	-	-	-	+	-	+
Photosensitive rash	+	+	+	+	+	+	-	+	+	+
Lupus nephritis	-	-	+	+	-	+	-	-	+	+
Autoimmune cytopenia	+	+	+	-	-	+	+	+	+	+
Positive coombs	+	+	+	+	+	+	-	-	+	+
Low C3/C4	+	+	+	+	-	+	-	-	+	+
Serositis	-	-	+	+	-	+	-	-	+	-
Damage	+	+	+	+	-	-	-	-	+	-
Type of auto-antibodies	ANA, Anti-ds-DNA, anti-Sm, anti-RNP, anti-Ro52, anti-SS-A	ANA, Anti-ds-DNA, anti-Sm, anti-RNP, anti-Ro52, anti-SS-A	ANA, Anti-ds-DNA, anti-Sm, anti-RNP, anti-SS-A	ANA, Anti-ds-DNA, anti-Sm, anti-RNP, anti-Ro52, anti-SS-A	ANA	ANA, Anti-ds-DNA, anti-C1q	ANA, Anti-ds-DNA, anti-Sm, anti-RNP	ANA	ANA, Anti-ds-DNA, anti-Sm, anti-RNP, anti-Ro52, anti-SS-A	ANA, Anti-ds-DNA, anti-Sm, anti-RNP, anti-Ro52, anti-SS-A

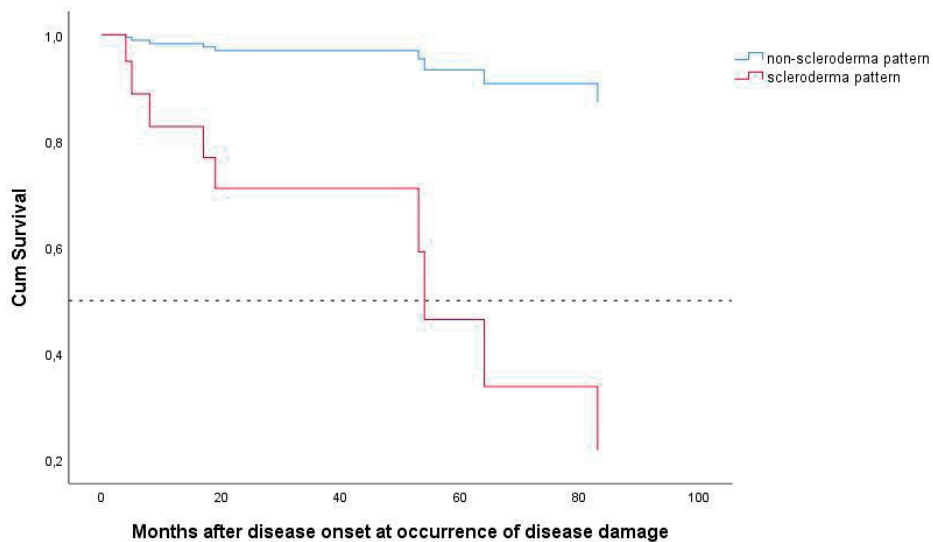


Figure 3. Cox regression analysis for occurrence of disease damage in cSLE patients with nailfold capillary non-scleroderma pattern (n=23) versus a capillary scleroderma pattern (n=10): Hazard Ratio (HR) 4.6 (95% CI 1.1-18.8) for scleroderma pattern group, p=0.034

Table 3. Different patients groups with a capillary scleroderma pattern and follow-up data.

Patient group	JDM n=11	jUCTD n=13	jSSc n=7	cSLE n=53	p-value
Scleroderma pattern at first capillaroscopy, n (%)	7 (63.6)	9 (69.2)	7 (100)	7 (13.2)	0.001/<0.001/<0.001 [°]
No fingers with scleroderma pattern, median (IQR)	8 (7-8)	3 (2.5-8)	8 (6-8)	4 (2-6)	0.03 / 0.76 / 0.10 ^b
FU patients, n (%)	7 (77.8)	3 (23.1)	5 (71.4)	33 (62.3)	
Scleroderma pattern at FU, n/total FU patients (%)	3/7 (42.9)	2/3 (66.7)	5/5 (100)	8/33 (24.2)	0.208/0.181/ 0.003 [°]
No fingers with scleroderma pattern in FU, median (IQR)	7 (7)	5 (5)	8 (5-8)	5.5 (2.5-7)	0.09 / 0.89 / 0.07 ^b

FU = follow-up. Bold indicates statistically significant p values (<0.05). [°] Chi square/Fisher exact analysis between the two subgroups. ^b Mann-Whitney U test between cSLE and resp. JDM/jUCTD/jSSc.

DISCUSSION

This is the first study reporting longitudinal follow-up data of nailfold capillaroscopy combined with clinical data in cSLE. We showed that nailfold capillary patterns in cSLE can change over time, but these changes were irrespective of disease activity. Strikingly, cSLE patients with a capillary scleroderma pattern had a higher risk for SLE-related disease damage, although SLEDAI (at baseline nor at follow-up) did not significantly differ between different capillary pattern groups. Subsequently, as we have shown that capillary patterns can worsen over time, it seems necessary to repeat capillaroscopy during clinical follow-up. Furthermore, we are also the first to show that in cSLE both microangiopathy and a scleroderma pattern, if detected, were observed in the majority of eight examined fingers. At baseline and follow-up, the total number of fingers with a scleroderma pattern, if present in cSLE, was comparable to patients with either jSSc/jUCTD (baseline) and JDM/jSSc/jUCTD (follow-up). This suggests that this finding is not coincidental in cSLE, and in juvenile patients might not only be a specific finding for jSSc.

Anti-RNP antibodies were significantly more detected in cSLE-patients with a capillary scleroderma pattern, but Raynaud's phenomenon was not, also not during follow-up. A capillary scleroderma pattern has been linked to overlap disease or mixed connective tissue disease [25-27]. In our study with up to 10 years of clinical follow up after diagnosis however, cSLE patients with a capillary scleroderma pattern did not evolve into a scleroderma overlap disease. Clinical follow-up of these patients included routine medical history, physical examination, laboratory biomarkers and pulmonary function tests (minimal every 2-3 years), cardiac ultrasound (on indication) and pulmonary CT scan for signs of fibrosis (on indication). The mean follow-up was 7 years after diagnosis,

one patient was lost to follow-up. Looking at the available anti-RNP titers (in only one-third) in our patients, these titers were quite low and this might subsequently mean that this subgroup of cSLE-patients are not at risk for overlap disease. Interestingly, our data do suggest that cSLE patients with anti-RNP seem to be a more severe subgroup with more SLE-related disease damage, which was also stated by Dayal et al. [28]. On the other hand, a recent study in SSc showed that the presence of anti-RNP antibodies fits with a subgroup of SSc with a better prognostic outcome in terms of survival [29]. Thus, the detection of anti-RNP antibodies might have a different prognostic meaning in these different systemic autoimmune diseases or scleroderma spectrum disorders. Most patients with anti-RNP antibodies in the scleroderma pattern group also had multiple, more SLE-specific, auto-antibodies detected, besides other clinical SLICC criteria. This shows that these patients are different than UCTD-patients with only anti-RNP and Raynaud symptoms.

Our finding that a capillary scleroderma pattern is a significant risk factor for irreversible SLE-related disease damage in cSLE is new. Longitudinally, we found that half of the cSLE-patients with a nailfold capillary scleroderma pattern had already irreversible disease damage within only 5 years of diagnosis, meaning that this disease damage develops at a young age, probably around their twenties. We have also shown that this damage could not be predicted by SLEDAI at diagnosis nor by SLEDAI at follow-up, and most of these patients had low disease activity over time. It has been described that SLE-patients with neuropsychiatric involvement and/or nephritis reflect a severe subgroup²⁴. In our study this was confirmed for neuropsychiatric involvement, but not for nephritis. Previously, in our cross-sectional study in cSLE, we did find a significant correlation between nephritis and the number of capillary hemorrhages. We have also described that the number of capillary hemorrhages significantly correlated with SLEDAI [9]. In this follow-up study, the capillary abnormalities remained visible although patients had low global disease activity or even inactive disease. In addition, patients with low disease activity and a scleroderma pattern at NVC, were significantly more at risk for developing disease damage, despite their low disease activity over time. This might imply an ongoing vasculopathy in SLE leading to this disease damage, irrespective of disease activity, as measured by SLEDAI. The disease damage in our cSLE-cohort was typical SLE-induced disease damage such as cerebrovascular incidents, seizures (>6 months of treatment), major psychosis, end-stage renal disease and avascular skeletal necrosis and extensive (facial) scarring. Only 7/53 patients never used prednisolone as treatment, all others used prednisone at some time in different dosages. As was shown in the specific details of disease damage, two patients suffered from skeletal necrosis and one from growth failure, probably related to chronic prednisolone use. In univariate regression analysis, prednisolone use (ever) was not associated with occurrence of

disease damage, but cumulative steroid dose was not calculated in this study. All other patients had specific SLE-related disease damage, which was not related to medication. Also, most disease damage already occurred in the first years after diagnosis.

As in our cross-sectional study [9], microangiopathy with abnormal capillary shapes and high number of capillary hemorrhages, was again a predominant and longitudinal a persistent finding in this current longitudinal study. As we have shown as well, two types of hemorrhages can be reliably differentiated and reproduced by different raters [30]. The same two types of capillary hemorrhages were observed over time in our longitudinal cohort. Other studies have described the capillary abnormalities in (c)SLE as non-specific abnormalities [1, 31, 32]. We hypothesize that the predominant finding of ‘microangiopathy’ in (c)SLE might be capillary leakage and revascularization and might be due to endothelial dysregulation in SLE, and may not be so non-specific at all. Dysregulation of endothelial cells in SLE has been linked to an increased risk of cardiovascular disease in these patients [33-35], which is one of the most important prognostic factors for mortality in SLE, especially in cSLE patients [36]. Thus, this microangiopathy should be further analyzed in this severe and chronic disease and might be a possible new biomarker or a “lupus-pattern” reflecting (early) vasculopathy in SLE which warrants additional therapy to prevent future damage.

A limitation of our study is the relatively low number of patients that was included. Although almost all eligible patients from the outpatient clinic could be included, the prevalence of these systemic autoimmune diseases at pediatric age is rare. Another limitation of our study is that not all patients had follow-up visits with capillaroscopy due to several reasons. The typical age of patients at cSLE onset is in their teens which means that patients are relatively soon transferred to adult care after diagnosis, roughly around the age of 18 years. In the Amsterdam UMC, these patients were included in our longitudinal cohort but not all longitudinal capillaroscopy data could be completed. Patients with transition to other hospitals were lost to follow-up.

CONCLUSION

We conclude that a capillary scleroderma pattern in cSLE did not reflect a SLE-subgroup at risk for developing SSc-like symptoms but we suggest that a capillary scleroderma pattern in SLE may be associated with a higher risk of developing SLE-related disease damage.

ACKNOWLEDGEMENTS

We would like to thank all patients for (repeated) participation of undergoing capillaroscopy examination. We also thank prof. dr. R. ten Cate, dr. L.B. van der Aa and G.E. Legger for their help in including patients. We would also like to thank M.D.J. Wolvers for her advice on statistics for the longitudinal data-analyses.

Supplementary table 1. Follow-up of capillary patterns in time (in months)

Subject	Visit 1	Visit 2	Visit 3	Visit 4	Visit 5	Visit 6	Visit 7	Time 1-2	Time 2-3	Time 3-4	Time 4-5	Time 5-6	Time 6-7
1	N	N						12					
2	N	N						22					
3	N	N						12					
4	N	N						8					
5	MA	MA	MA	Poor quality images				20	5	18			
6	MA	MA	MA	MA				15	20	16			
7	MA	MA	MA					15	20				
8	MA	MA	MA	Scl				16	14	20			
9	MA	Scl	Scl	Scl	Scl			13	13	13	9		
10	MA	MA	MA					15	15				
11	MA	MA						16					
12	MA	MA	MA					18	29				
13	MA	MA						52					
14	MA	MA						25					
15	MA	Scl						50					
16	MA	MA	MA					12	13				
17	MA	MA						36					
18	MA	MA	MA					10	26				
19	MA	MA						16					
20	MA	MA	MA					15	19				
21	MA	MA	MA					13	17				
22	MA	MA						23					
23	MA	N						15					
24	MA	N						9					
25	MA	MA						13					
26	MA	N						21					
27	MA	Poor quality images						20					
28	MA	MA						19					
29	Scl	Scl	Scl	Scl	Scl	Scl	Scl	9	8	5	6	10	9
30	Scl	Scl	Scl	Scl				10	14	14			
31	Scl	Scl	Scl	Scl				16	15	18			
32	Scl	MA						20					
33	Scl	Scl						14					

N= Normal pattern, MA=MicroAngiopathy, Scl=Scleroderma pattern. Bold: change in capillary pattern

Supplementary table 2. Logistic regression analyses for occurrence of scleroderma pattern (ever)

Univariate analysis	OR (95% CI)	p-value
SLEDAI at diagnosis	1.05 (0.96-1.15)	0.277
Discoid lupus	9.25 (2.0-42.77)	0.004
Nephritis	0.47 (0.09-2.48)	0.372
Neuropsychiatric involvement	1.54 (0.26-9.08)	0.632
Anti-phospholipid antibodies	0.71 (0.15-3.37)	0.669
Anti-RNP antibodies	6.79 (1.49-30.92)	0.013
Anti-Sm antibodies	2.31 (0.57-9.36)	0.242
Multivariate analysis		
Discoid lupus	5.89 (1.15-30.19)	0.033
Anti-RNP antibodies	4.11 (0.79-21.25)	0.092

OR= odds ratio, CI=confidence interval. NA=not available. Bold=statistical significant

Supplementary table 3. Logistic regression analyses for occurrence of SLE disease damage (by SDI)

Univariate analysis	OR (95% CI)	p-value
Age at diagnosis	1.19 (0.89-1.61)	0.243
Gender	1.18 (0.12-11.42)	0.884
SLEDAI at diagnosis	1.09 (0.99-1.19)	0.063
Discoid lupus	5.1 (1.17-22.61)	0.003
Nephritis	0.47 (0.09-2.48)	0.372
Neuropsychiatric involvement	6.5 (1.27-33.20)	0.024
Anti-phospholipid antibodies	0.92 (0.46-1.85)	0.819
Anti-RNP antibodies	3.88 (0.93-16.19)	0.063
Anti-Sm antibodies	1.38 (0.34-5.70)	0.655
Prednisolon use (ever)	NA	0.999
Scleroderma pattern (ever)	7.6 (1.61-35.85)	0.01
Multivariate analysis		
Discoid lupus	2.24 (0.33-15.44)	0.411
Neuropsychiatric involvement	7.75 (1.19-15.44)	0.032
Nephritis	0.50 (0.07-3.66)	0.496
Anti-RNP antibodies	2.01 (0.32-12.63)	0.455
Scleroderma pattern (ever)	4.75 (0.73-31.11)	0.104

OR= odds ratio, CI=confidence interval. NA=not available. Bold=statistical significant

REFERENCES

1. Cutolo M, Melsens K, Wijnant S, et al. Nailfold capillaroscopy in systemic lupus erythematosus: A systematic review and critical appraisal. *Autoimmun Rev* 2018;17(4):344-52.
2. Schonenberg-Meinema D, Melsens K, Nassar-Sheikh Rashid A, et al. Capillaroscopy in childhood-onset systemic lupus erythematosus: a first systematic review. *Clinical and experimental rheumatology* 2020;38(2):350-54.
3. Donnarumma JFS, Ferreira EVM, Ota-Arakaki J, et al. Nailfold capillaroscopy as a risk factor for pulmonary arterial hypertension in systemic lupus erythematosus patients. *Adv Rheumatol* 2019;59(1):1.
4. Lambova SN, Muller-Ladner U. Capillaroscopic pattern in systemic lupus erythematosus and undifferentiated connective tissue disease: what we still have to learn? *Rheumatol Int* 2013;33(3):689-95.
5. Pavlov-Dolijanovic S, Damjanov N, Ostojic P, et al. The prognostic value of nailfold capillary changes for the development of connective tissue disease in children and adolescents with primary raynaud phenomenon: a follow-up study of 250 patients. *Pediatric dermatology* 2006;23(5):437-42.
6. Shenavandeh S, Habibi S. Nailfold capillaroscopic changes in patients with systemic lupus erythematosus: correlations with disease activity, skin manifestation and nephritis. *Lupus* 2017;961203316686702.
7. Furtado RN, Pucinelli ML, Cristo VV, et al. Scleroderma-like nailfold capillaroscopic abnormalities are associated with anti-U1-RNP antibodies and Raynaud's phenomenon in SLE patients. *Lupus* 2002;11(1):35-41.
8. Pavlov-Dolijanovic S, Damjanov NS, Vujasinovic Stupar NZ, et al. Is there a difference in systemic lupus erythematosus with and without Raynaud's phenomenon? *Rheumatol Int* 2013;33(4):859-65.
9. Schonenberg-Meinema D, Bergkamp SC, Nassar-Sheikh Rashid A, et al. Nailfold capillary abnormalities in childhood-onset systemic lupus erythematosus: a cross-sectional study compared with healthy controls. *Lupus* 2021;30(5):818-27.
10. Smith V, Vanhaecke A, Herrick AL, et al. Fast track algorithm: How to differentiate a "scleroderma pattern" from a "non-scleroderma pattern". *Autoimmun Rev* 2019;18(11):102394.
11. Ingegnoli F. Capillaroscopy abnormalities in relation to disease activity in juvenile systemic lupus erythematosus. *Microvascular research* 2013;87:92-4.
12. Avouac J, Lepri G, Smith V, et al. Sequential nailfold videocapillaroscopy examinations have responsiveness to detect organ progression in systemic sclerosis. *Seminars in arthritis and rheumatism* 2017;47(1):86-94.
13. Smith V, Decuman S, Sulli A, et al. Do worsening scleroderma capillaroscopic patterns predict future severe organ involvement? a pilot study. *Ann Rheum Dis* 2012;71(10):1636-9.
14. Paolino S, Ferrari G, Pizzorni C, et al. Long-term follow-up of nailfold videocapillaroscopic microvascular parameters in mixed connective tissue disease versus systemic sclerosis patients: a retrospective cohort study. *Clinical and experimental rheumatology* 2019;37 Suppl 119(4):102-07.
15. Ghizzoni C, Sebastiani M, Manfredi A, et al. Prevalence and evolution of scleroderma pattern at nailfold videocapillaroscopy in systemic sclerosis patients: Clinical and prognostic implications. *Microvascular research* 2015;99:92-5.

16. Sulli A, Pizzorni C, Smith V, et al. Timing of transition between capillaroscopic patterns in systemic sclerosis. *Arthritis and rheumatism* 2012;64(3):821-5.
17. Petri M, Orbai AM, Alarcon GS, et al. Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis and rheumatism* 2012;64(8):2677-86.
18. Zulian F, Woo P, Athreya BH, et al. The Pediatric Rheumatology European Society/American College of Rheumatology/European League against Rheumatism provisional classification criteria for juvenile systemic sclerosis. *Arthritis and rheumatism* 2007;57(2):203-12.
19. Bottai M, Tjarnlund A, Santoni G, et al. EULAR/ACR classification criteria for adult and juvenile idiopathic inflammatory myopathies and their major subgroups: a methodology report. *RMD Open* 2017;3(2):e000507.
20. Bombardier C, Gladman DD, Urowitz MB, et al. Derivation of the SLEDAI. A disease activity index for lupus patients. The Committee on Prognosis Studies in SLE. *Arthritis and rheumatism* 1992;35(6):630-40.
21. Stoll T, Stucki G, Malik J, et al. Association of the Systemic Lupus International Collaborating Clinics/American College of Rheumatology Damage Index with measures of disease activity and health status in patients with systemic lupus erythematosus. *The Journal of rheumatology* 1997;24(2):309-13.
22. Smith V, Herrick AL, Ingegnoli F, et al. Standardisation of nailfold capillaroscopy for the assessment of patients with Raynaud's phenomenon and systemic sclerosis. *Autoimmun Rev* 2020;19(3):102458.
23. Cutolo M, Melsens K, Herrick AL, et al. Reliability of simple capillaroscopic definitions in describing capillary morphology in rheumatic diseases. *Rheumatology (Oxford)* 2018;57(4):757-59.
24. Fatemi A, Matinfar M, Smiley A. Childhood versus adult-onset systemic lupus erythematosus: long-term outcome and predictors of mortality. *Clinical rheumatology* 2017;36(2):343-50.
25. Alarcon-Segovia D, Cardiel MH. Comparison between 3 diagnostic criteria for mixed connective tissue disease. Study of 593 patients. *The Journal of rheumatology* 1989;16(3):328-34.
26. Aringer M, Steiner G, Smolen JS. Does mixed connective tissue disease exist? Yes. *Rheum Dis Clin North Am* 2005;31(3):411-20.
27. Cappelli S, Bellando Randone S, Martinovic D, et al. "To be or not to be," ten years after: evidence for mixed connective tissue disease as a distinct entity. *Seminars in arthritis and rheumatism* 2012;41(4):589-98.
28. Dayal NA, Isenberg DA. SLE/myositis overlap: are the manifestations of SLE different in overlap disease? *Lupus* 2002;11(5):293-8.
29. Fairley JL, Hansen D, Proudman S, et al. Clinical characteristics and survival in systemic sclerosis-mixed connective tissue disease and systemic sclerosis-overlap syndrome. *Arthritis care & research* 2020 doi: 10.1002/acr.24167 [published Online First: 2020/02/15]
30. Bergkamp SC, Schonenberg-Meinema D, Nassar-Sheikh Rashid A, et al. Reliable detection of subtypes of nailfold capillary haemorrhages in childhood-onset systemic lupus erythematosus. *Clinical and experimental rheumatology* 2021 Sep-Oct;39(5):1126-1131.
31. Ingegnoli F, Zeni S, Meani L, et al. Evaluation of nailfold videocapillaroscopic abnormalities in patients with systemic lupus erythematosus. *Clin Rheumatol* 2005;11(6):295-98.
32. Ingegnoli F, Zeni S, Gerloni V, et al. Capillaroscopic observations in childhood rheumatic diseases and healthy controls. *Clinical and experimental rheumatology* 2005;23(6):905-11.
33. Westerweel PE, Luyten RK, Koomans HA, et al. Premature atherosclerotic cardiovascular disease in systemic lupus erythematosus. *Arthritis and rheumatism* 2007;56(5):1384-96.

Nailfold videocapillaroscopy in cSLE


34. Sciatti E, Cavazzana I, Vizzardi E, et al. Systemic Lupus Erythematosus and Endothelial Dysfunction: A Close Relationship. *Curr Rheumatol Rev* 2019;15(3):177-88.
35. Mauro D, Nerviani A. Endothelial Dysfunction in Systemic Lupus Erythematosus: Pathogenesis, Assessment and Therapeutic Opportunities. *Rev Recent Clin Trials* 2018;13(3):192-98.
36. Hersh AO, Trupin L, Yazdany J, et al. Childhood-onset disease as a predictor of mortality in an adult cohort of patients with systemic lupus erythematosus. *Arthritis care & research* 2010;62(8):1152-9.



Part II

A large, abstract red splatter graphic that starts on the left side of the page and extends towards the right, partially overlapping the text. The splatter consists of numerous small red dots and larger, irregular red shapes, creating a dynamic, energetic feel. The background is white.

**New biomarkers in (systemic) connective
tissue diseases**

A solid red square graphic located on the right edge of the page, partially overlapping the red splatter graphic.

6

Gene signature fingerprints divide SLE patients in subgroups with similar biological disease profiles: a multicenter longitudinal study

M. Javad Wahadat, **Dieneke Schonenberg-Meinema**, van Helden-Meeuwsen CG, van Tilburg S, Groot N, Ellen JH Schatorjé, Esther PAH Hoppenreijns, Petra CE Hissink Muller, Danielle MC Brinkman, Dvorak D, Maaïke Verkaaik, Bouchalova K, J Merlijn van den Berg, Sylvia Kamphuis, Marjan A. Versnel

Published in Rheumatology. 2022 Feb 10: online ahead of print

ABSTRACT

OBJECTIVES: Clinical phenotyping and predicting treatment responses in Systemic Lupus Erythematosus (SLE) patients is challenging. Extensive blood transcriptional profiling has identified various gene modules that are promising for stratification of SLE patients. We aimed to translate existing transcriptomic data into simpler gene signatures suitable for daily clinical practice.

METHODS: RT-PCR of multiple genes from the Interferon M1.2, Interferon M5.12, neutrophil (NPh) and plasma cell (PLC) modules followed by a principle component analysis, was used to identify indicator genes per gene signature. Gene signatures were measured in longitudinal samples from two childhood onset SLE cohorts (n=101 and n=34, respectively) and associated with clinical features. Disease activity was measured using SELENA-SLEDAI. Cluster analysis subdivided patients into three mutually exclusive fingerprint-groups termed 1) all-signatures-low, 2) only IFN high (M1.2 and/or M5.12) and 3) high NPh and/or PLC.

RESULTS: All gene signatures were significantly associated with disease activity in cross-sectionally collected samples. The PLC-signature showed the highest association with disease activity. Interestingly in longitudinally collected samples, the PLC-signature was associated with disease activity and showed a decrease over time. When patients were divided into fingerprints, the highest disease activity was observed in the high NPh and/or PLC group. The lowest disease activity was observed in the all-signatures-low group. The same distribution was reproduced in samples from an independent SLE cohort.

CONCLUSIONS: The identified gene signatures are associated with disease activity and suitable tools to stratify SLE patients into groups with similar activated immune pathways that may guide future treatment choices.

INTRODUCTION

Systemic Lupus Erythematosus (SLE) is an autoimmune disease characterized by its heterogeneity at the clinical, cellular and molecular level [1]. This often poses a challenge for clinicians to reliably divide patients into homogeneous disease subgroups. Additionally, patients with distinct clinical disease phenotypes respond to the same medication and vice versa, underlining that solely using clinical phenotype to decide which treatment to start is not enough. Identification of tools to cluster patients in homogeneous groups with similar underlying aberrantly activated immune pathways to guide treatment blocking these pathways, is an important research topic. Transcriptional profiling resulted in the identification of so-called “gene signatures”. A gene signature is a group of simultaneously upregulated genes caused by a change in the cell’s biological processes. In SLE multiple gene signatures with correlations to unique clinical features have been identified. However, application into clinical practice is challenging, due to a lack of consensus regarding the genes representing a signature and the feasibility to implement transcriptional profiling on individual patients in the clinical setting.

The most well-known gene signature in SLE is the type I Interferon (IFN-I) gene signature, which is present in more than 50% of the patients and correlates to disease activity in several cross-sectional studies [2-6]. Transcriptomic data of SLE blood has revealed three different upregulated IFN-annotated modules, respectively called M1.2, M3.4 and M5.12 [7]. The M1.2 module is induced by IFN-I, while both the M3.4 and M5.12 modules are induced by a combination of IFN-I and type II IFN (IFN-II). When studied over time in SLE patients, each module displayed a different dynamic pattern, with the highest variation in the M5.12 module. These fluctuations indicated that the IFN signature could be used as biomarker for disease activity. However, the few studies that have investigated the parallel change of disease activity with IFN gene signatures over time, have shown a lack of association [8, 9]. This implicates the involvement of other immune pathways than the IFN route.

When focusing on pathways which have already been shown to correlate with SLE disease manifestations and/or disease activity, two other gene signatures stand out: the neutrophil (NPh) and plasma cell (PLC) signature [8-12]. Neutrophils and plasma cells are increased in SLE patients and play a role in disease pathogenesis [13, 14]. Neutrophils of SLE patients are more active, have lower phagocytic capacity and are prone to spontaneously release Neutrophil Extracellular Traps [15]. Plasma cells are the source of pathogenic auto-antibodies in SLE. The NPh signature was associated with lupus nephritis and vascular inflammation while the PLC signature correlated with disease activity [8-10, 16]. Moreover, in studies with extensive transcriptional profiling, these

and other gene signatures were used to divide patients into subgroups with similar biological disease profiles [12, 17].

Here, we investigated whether we could translate complex transcriptomic data, reflecting multiple different immune pathways, into simple gene signatures suitable for introduction into clinical practice. Additionally, we studied their association with disease activity and clinical outcome using prospectively collected clinical data and blood samples over time from two childhood-onset SLE (cSLE) cohorts. cSLE is an excellent disease model to study, as cSLE represents the more severe clinical phenotype, has a higher genetic component and children with SLE lack the comorbidities common in adult-onset SLE and that may confound translational studies.

METHODS

Patient recruitment

Patients fulfilled the Systemic Lupus International Collaborating Clinics (SLICC) classification criteria or the 2019 EULAR/ACR [18, 19]. Blood specimens, demographics and clinical characteristics were prospectively collected. Disease activity was assessed by the Safety of Estrogens in Lupus National Assessment-Systemic Lupus Erythematosus Disease Activity Index (SELENA-SLEDAI) at each visit [20]. Disease flares were indicated by an increase of >3 or >12 points from the previous visit for a mild/moderate or severe flare respectively. Disease domains were derived from the SELENA-SLEDAI items.

Additionally, 51 healthy controls (HC), without symptoms of underlying viral infections or the use of any medications, were included. Written informed consent was obtained from all participants in compliance with the Declaration of Helsinki. The study was approved by the medical ethics review committee of the Erasmus Medical Center, Rotterdam, the Netherlands (MEC-2019-0412).

Clustering strategy

A semi-manual and an automated clustering strategy was performed (supplementary figure S1). For the semi-manual clustering strategy, the combination of a positive or negative score per gene signature was used to identify 16 clusters. These clusters were subsequently divided into 3 so-called fingerprints consisting of patients with mutually exclusive combinations of positive and negative gene signatures. For the automated clustering strategy, an unsupervised hierarchical clustering method was used to identify clusters that were enriched in the cSLE cohort.

Supplementary methods

Details on blood collection and real-time PCR, gene selection and signature definitions, ultrasensitive IFN- α Simoa and statistics are described here.

RESULTS

Cohort description

Between March 2013 and January 2021, 101 Childhood-onset SLE (cSLE) patients with median disease duration of 0.5 (0-8.2) years at enrolment (Cohort-I, Table 1) were prospectively recruited at the outpatient clinic of three academic hospitals in the Netherlands and one in the Czech Republic. Fifty-one HC were included in the study. For 73/101 patients, blood samples from 2-4 longitudinal time points with a median follow up time of 344 (29-1542) days were available. As a replication cohort, 34 adults with cSLE with median disease duration of 15.8 (3.8-40) years were included (Cohort-II, Table 1).

Four dynamic gene signatures are present in SLE patients

Four gene signatures were assessed based on indicator genes in 51 HC and 101 cSLE patients (supplementary- Figure S1, table T1). All four gene signatures were significantly higher expressed in patients compared to HC (Figure 1A). High positive associations were present between the genes representing each gene signature, while poor associations were found between genes of different gene modules (Figure 1B). The M1.2 and M5.12 IFN signatures had the highest association, followed by the M5.12 and NPh signature, while poor associations were observed between the other signatures (Figure 1C).

In 73 cSLE patients, each gene signature was determined at a second time-point and in 42 cSLE patients at a third and/or fourth time point (Figure 1D). In 51 out of 73 patients (70%), at least one or more signatures changed from positive to negative or vice versa (Figure 1D). The M5.12 IFN signature showed the highest variability within individual patients (coefficient of variation [CoV] 0.66 ± 2.74), followed by the PLC- (CoV 0.54 ± 4.39), NPh- (CoV -0.35 ± 4.23) and M1.2 IFN (CoV 0.05 ± 1.14) signatures (Figure 1E). These findings imply that gene signatures are driven by different pathways and have a dynamic character over time, which makes them potential biomarkers for changes in disease activity.

Gene signatures are associated with disease activity

To investigate associations with disease characteristics we stratified patients into groups based on a low or high gene signature using the mean + $2xSD_{HC}$ per score as a threshold. A high M1.2 IFN, M5.12 IFN, NPh or PLC signature was more prevalent in patients with

a higher SELENA-SLEDAI (supplementary figure S2A-D). The highest association was found for the PLC signature ($p < 0.0001$, $r = 0.473$) (Figure 2A).

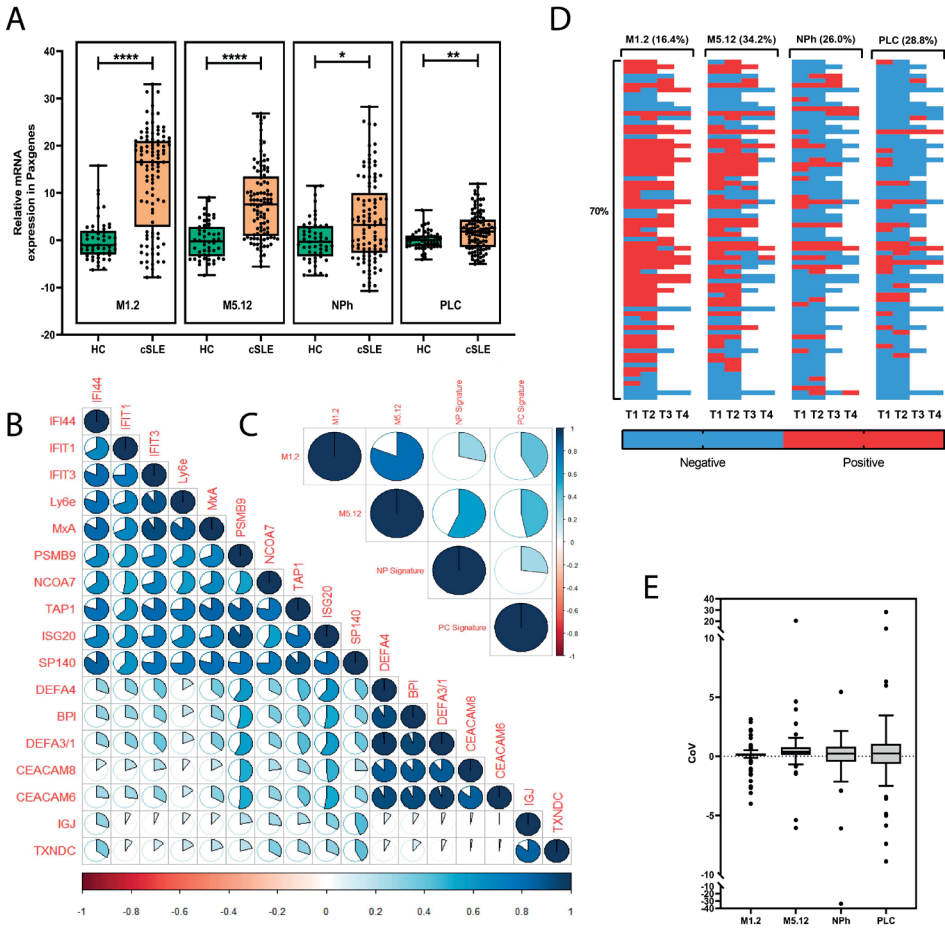


Figure 1. Four dynamic gene signatures are present in cSLE patients
 A). Relative expression of four gene signature scores in HC (N=51) vs cSLE patients (N=101). B). Correlation matrix between gene signature associated genes based on relative expression. C). Correlation matrix between each individual gene signature. D). Heatmap indicating a positive or negative gene signature score over time in 73 cSLE patients. Each row represents the same patient. Horizontal percentages indicate the number of patients that showed a dynamic signature over time. Vertical percentage indicates the number of patients with at least one dynamic gene signature. E). Coefficient of variation (CoV) per gene signature indicating the intraindividual difference per gene signature. Each dot represents the CoV of one patient. Lines indicate the mean \pm SD.
 Mann-Whitney-U test was used to compare two groups; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; **** $p < 0.0001$. For correlations Spearman's rho was used. A full circle represents a rho of 1. HC, healthy control. T, time point. NPh, Neutrophil signature. PLC, Plasma cell signature.

Furthermore, we investigated the association of disease domains, derived from the SELENA-SLEDAI, with the different gene signatures (Figure 2B). Univariate analysis showed skin-, hematological-, and immunological domain involvement to be associated

Table 1. Patients and healthy control characteristics

Category	Total cSLE cohort (n=101)	COHORT-I				COHORT-II	
		EMC cohort (n=49)	AUMC cohort (n=35)	TCH cohort (n=12)	RUMC cohort (n=5)	Adult cSLE cohort (n=34)	HC (n=51)
Demographics							
Gender	Female	84 (83.2%)	30 (85.7%)	8 (66.7%)	5 (100%)	32 (94.1%)	40 (78.4%)
	Male	17 (16.8%)	5 (14.3%)	4 (33.3%)	0 (0%)	2 (5.9%)	11 (21.6%)
Ethnicity	White	43 (42.6%)	15 (42.9%)	1 (8.3%)	2 (40%)	26 (76.5%)	45 (88.2%)
	Non-white	58 (57.4%)	24 (49.0%)	11 (91.7%)	3 (60%)	8 (23.5%)	6 (11.8%)
Age at enrolment (years)		15.6 (5.1-23)	15.2 (5.2-18.1)	16.7 (11.8-23)	14.4 (5.1-17.2)	32.4 (18.5-56.4)	29 (20-65)
Disease duration at enrolment (years)		0.51 (0-8.2)	0.18 (0-6.8)	1.11 (0-8.2)	0.04 (0-1.8)	15.8 (3.8-40)	.
SELENA-SLEDAI at enrolment		4 (0-27)	4 (0-18)	4 (0-27)	4 (0-15)	4 (0-14)	.
SELENA-SLEDAI at enrolment	≤4	53 (52.5%)	25 (51.0%)	18 (51.4%)	7 (58.3%)	23 (67.6%)	.
	5 to 7	12 (11.9%)	9 (18.4%)	2 (5.7%)	0 (0%)	8 (23.5%)	.
	≥8	36 (35.6%)	15 (30.6%)	15 (42.9%)	5 (41.7%)	3 (8.9%)	.
Longitudinal samples							
Number of visits (median)		73 (72.2%)	43 (87.8%)	18 (51.4%)	9 (75%)	0	0
		2 (1-4)	2 (1-4)	2 (1-4)	2 (1-4)	.	.

Table 1. Patients and healthy control characteristics (continued)

Category	Total cSLE cohort (n=101)	COHORT-I			COHORT-II		
		EMC cohort (n=49)	AUMC cohort (n=35)	TCH cohort (n=12)	RUMC cohort (n=5)	Adult cSLE cohort (n=34)	HC (n=51)
Follow up time (days)	344 (29-1542)	267 (51-1542)	511 (98-1059)	91 (29-182)	120.5 (105-182)	.	.
Flare	14 (19.2%)	7 (14.2%)	6 (17.1%)	1 (8.3%)	0 (0%)	.	.
Severe	7 (9.6%)	1 (2.0%)	6 (17.1%)	0 (0%)	0 (0%)	.	.
Treatment at enrolment							
None	27 (26.7%)	15 (30.6%)	8 (22.9%)	3 (25%)	0 (0%)	0 (0%)	51 (100%)
Hydroxychloroquine	67 (66.3%)	33 (67.3%)	24 (68.6%)	6 (50%)	4 (80%)	31 (91.2%)	.
Mycophenolate Mofetil	28 (27.7%)	16 (32.7%)	9 (25.7%)	0 (0%)	3 (60%)	14 (41.2%)	.
MTX	3 (3.0%)	2 (4.1%)	0 (0%)	0 (0%)	1 (20%)	2 (5.9%)	.
Azathioprine	8 (7.9%)	2 (4.1%)	5 (14.3%)	1 (8.3%)	0 (0%)	16 (47.1%)	.
Cyclophosphamide	0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)	8 (23.5%)	.
Prednisone	29 (28.7%)	15 (30.6%)	8 (22.9%)	4 (33.3%)	2 (40%)	31 (91.2%)	.
Rituximab	4 (4.0%)	0 (0%)	4 (11.4%)	0 (0%)	0 (0%)	1 (2.9%)	.
Belumimab	1 (1.0%)	0 (0%)	1 (2.9%)	0 (0%)	0 (0%)	0 (0%)	.

Data are presented as median (range) or as number (% of total). Non-white ethnicity = Hindu, Suriname, Hispanic, Asian, African-American, Mixed. MTX = Methotrexate. EMC (Erasmus Medical Center); AUMC (Amsterdam University Medical Center); TCH (Czech Republic Palacky University Olomouc); RUMC (Radboud University Medical Center); cSLE = childhood onset SLE; HC = healthy controls.

with a high M1.2 IFN signature. No domain was associated with a high M5.12 IFN signature. Constitutional and musculoskeletal domain involvement showed an association with a high NPh signature. Additionally, all domains except for the skin, renal and CNS were associated with a high PLC signature. In the multivariate model, skin involvement was associated with a high M1.2 IFN signature, musculoskeletal domain associated with a high NPh and PLC signature, and the constitutional domain was associated with a high PLC signature (Figure 2B).

Next, we determined whether changes over time in disease activity are accompanied by changes in gene signatures. For this purpose, 20 treatment naïve ($\text{Tx}_{\text{naïve}}$) children with at least one subsequent sample after start of treatment (median time between samples: 62.5 days) were chosen as the optimal patient group to investigate this. The decrease in disease activity over time was accompanied by a significant reduction of the PLC signature but was not mirrored by changes in M1.2 IFN, M5.12 IFN and NPh signatures (Figure 2C-D). Interestingly, when investigating the effect of medication, the additional use of Mycophenolate Mofetil (MMF) and/or Prednisone to Hydroxychloroquine treatment also led to a decrease in the PLC signature ($p=0.0005$), whereas the addition of Prednisone increased the NPh signature ($p=0.0195$) (Figure 2D). In addition, the IFN α 2 levels, neutrophil counts and anti-dsDNA levels were measured in the same samples to study other factors than medication use, that may influence the signatures. Neutrophil counts and anti-dsDNA levels showed the same trend as the NPh and PLC signatures, while the IFN α 2 levels in general decreased in contrast to the varying activation of the M1.2 and M5.12 IFN signature (supplementary figure S3A-C). Together, these data indicate that gene signatures are associated with disease activity and are influenced by medication use and cell compositions. However, the correlation coefficients (figure 2A) were low indicating that testing individual gene signatures will not be sufficient to identify homogeneous subgroups of patients.

Gene fingerprints identify SLE patients with similar disease activity

In our search to find homogenous subgroups of patients, we first used a semi-manual clustering strategy. Based on the four described gene signatures, patients were allocated into 16 unique clusters (Figure 3A). A cluster represents a combination of either a positive or negative gene signature score. To reduce data complexity the clusters were distributed over 3 mutually exclusive groups with matching underlying activated immune pathways forming a so-called fingerprint. Fingerprint-1 is described as “all-signature-low” which indicates patients with a low score in all four gene signatures. Fingerprint-1 consists of patients in cluster 16 ($n=25$; 24.8%). Fingerprint-2 represents patients with high IFNs, meaning a high M1.2 and/or high M5.12 score and consists of patients in clusters 1, 2 and 6 ($n=34$; 33.7%). Lastly, fingerprint-3 defines patients with

high NPh and/or PLC signature independent of the IFN signatures and includes patients in clusters 3, 4, 5, 7 to 15 (n=42; 41.5%). Notably, the majority of these patients have a positive M1.2 and/or M512 gene signature (36/42) (Figure 3A-B).

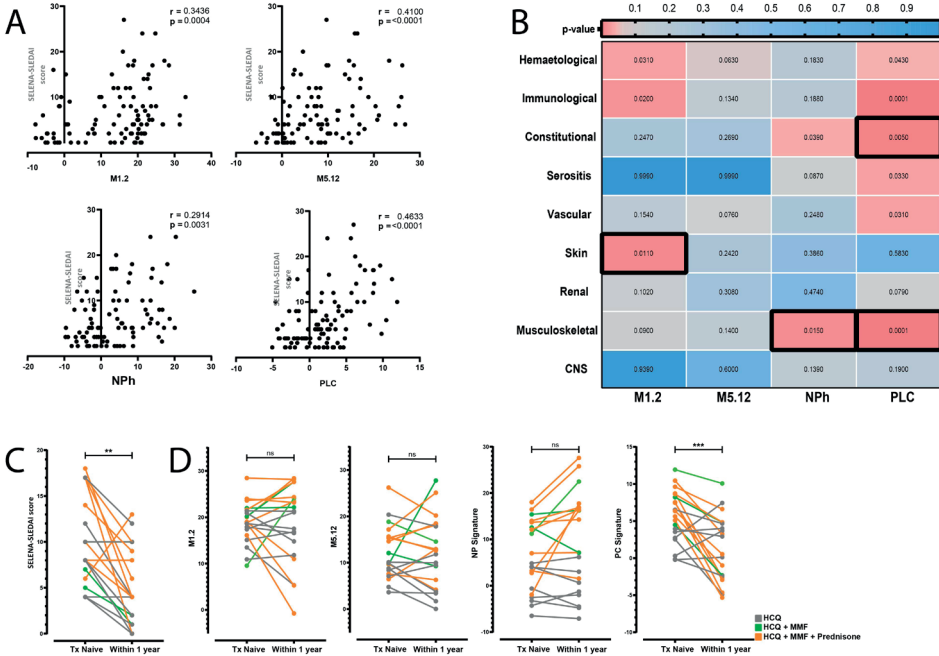


Figure 2. Gene signatures are associated with disease activity
 A). Correlation between the SELENA-SLEDAI and gene signature scores. B). Heatmap indicating the correlation between a high signature and a specific disease domain derived from the SELENA-SLEDAI. Numbers indicate the p-value based on univariate analysis. Black lined boxes indicate domains that were significant in the multivariate model. C) Longitudinal SELENA-SLEDAI. D) gene signature scores from 20 Tx_{naive} cSLE patients at the first and second time point (median time between two samples = 62.5 days).
 Mann-Whitney-U test was used to compare two groups; *p < 0.05; **p < 0.01; ***p < 0.001; ****p < 0.0001. For correlations Spearman's rho was used. Fisher's exact test was used to compare categorical data. NPh, Neutrophil signature. CNS, central nervous system. PLC, Plasma cell signature. Tx_{naive}, treatment naïve. HCQ, Hydroxychloroquine. MMF, Mycophenolate Mofetil.

As a first step we cross-sectionally analyzed data from samples taken at entry in the study (Figure 3C). Disease activity was significantly different between the three fingerprint groups. Patients with fingerprint-1 (median SELENA-SLEDAI = 2) had the lowest disease activity, while patients with fingerprint-3 had the highest disease activity (median SELENA-SLEDAI = 8). Interestingly, patients with fingerprint-2 formed an intermediate group, indicating that these patients had immunological activation and clinical disease activity but less prominent than patients with fingerprint-3 (Figure 3C). In the fingerprint-3 group, the highest number of patients was treatment naïve (Tx_{naive}; N=20/42) and recently diagnosed (median 10 days, supplementary figure S4A-B). Only 8 patients in this group had a disease duration of >1 year, of which 3 had a disease flare.

To filter out the component of high disease activity and lack of immunosuppressive medication in Tx_{naive} patients (n=27/101) at the first time point, we analyzed 73 samples taken at a subsequent second time point. This confirmed the observation from the first time point: patients within fingerprint-1 had the lowest disease activity, while patients with fingerprint-3 had the highest disease activity (Figure 3D).

To further address whether disease duration influenced our findings, the fingerprints were determined in a cohort of 34 adults with cSLE with a median disease duration of 15.8 years (cohort-II, Table 1). Distribution of patients within cohort-II based on fingerprints, showed an identical association with disease activity. This excludes disease duration from being a factor influencing the fingerprints (Figure 3E). Moreover, to investigate why a selection of patients with low disease activity (SELENA-SLEDAI ≤ 4) had fingerprint-3, we compared the use of medication between all patients with SELENA-SLEDAI ≤ 4 with fingerprint-1 and fingerprint-3 (Figure 3F). Patients in the fingerprint-3 group with SELENA-SLEDAI ≤ 4 were more often on prednisone ($p < 0.002$). This indicates that these patients, despite having low disease activity, still have activation of the underlying immune pathways leading to high NPh and/or PLC gene signature expression.

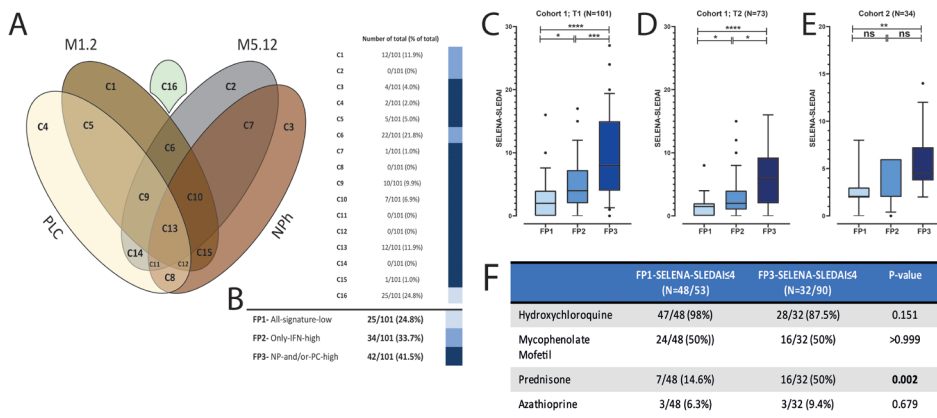


Figure 3. Gene fingerprints identify SLE patients with similar disease activity

A). Venn-diagram of gene clusters. Overlap between Venn's indicates a positive gene score of the involved gene signatures. B). Patient distribution over three fingerprint groups. Colored bars represent the clusters that are involved in each fingerprint. C). SELENA-SLEDAI distribution per fingerprint group in cSLE cohort-I; first time point (n=101). D). SELENA-SLEDAI distribution per fingerprint group in cSLE cohort-I; second time point (n=73). E). SELENA-SLEDAI distribution per fingerprint group in cSLE cohort-II; replication cohort (n=34). F) Medication use in patients with FP1 and FP3 with a SELENA-SLEDAI ≤ 4 . Dots represent individual patients.

Mann-Whitney-U test was used to compare the two groups; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; **** $p < 0.0001$. NPh, Neutrophil signature. PLC, Plasma cell signature. FP, fingerprint. ns, not significant. T, time point.

Gene fingerprints and clinical phenotype

We performed univariate and multivariate logistic regression analyses to test for specific organ domains involved in patients in the fingerprint-groups, fingerprint-1 was associated with significantly less skin involvement in the multivariate model and fingerprint-3 was associated with involvement of the musculoskeletal, constitutional and immunological organ domains (supplementary table T2).

Auto-antibody profiling of the patients revealed that patients with fingerprint-3 had higher anti-dsDNA levels than patients with fingerprints-1 and 2, reflecting the higher disease activity found in patients with fingerprint-3 (supplementary figure S4C-D). Interestingly, patients with fingerprint-3 were also more often anti-dsDNA positive than the other patients. Anti-SSA antibodies were primarily present in patients with fingerprint-2 and 3, while anti-SM and anti-RNP did not differ between the fingerprint groups (supplementary figure S4D).

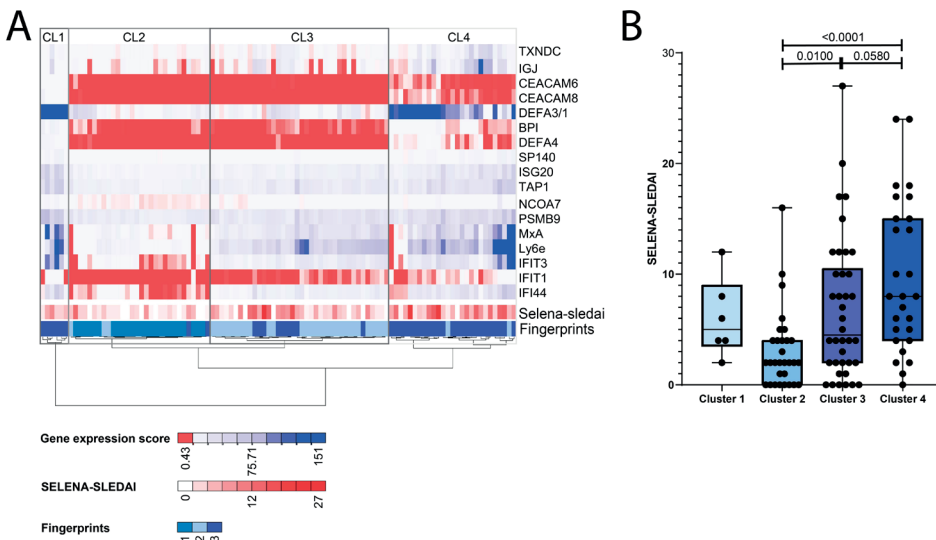


Figure 4. Hierarchical clustering parallels the identified gene fingerprints

A) Unsupervised hierarchical clustering using Ward's agglomerative method, passing the Euclidean distance between samples, identifying Cluster 1,2,3 and 4. Fingerprints 1,2,3 and SELENA-SLEDAI are depicted in the lowest two rows of the heatmap. Each column represents one patient. B). Association between SELENA-SLEDAI and clusters. Dots represent individual patients.

Mann-Whitney-U test was used to compare two groups (4B); * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; **** $p < 0.0001$. Red to white color indicates the magnitude of gene expression described as $2^{\wedge}-(dCT)$.

Hierarchical clustering identifies gene fingerprints

To investigate the robustness of the identified gene fingerprints, we additionally performed an automated clustering strategy to assess clusters that were enriched in our patient cohort. Unsupervised hierarchical clustering identified four major clusters that

paralleled the identified gene fingerprints (figure 4A). Cluster 1 represented a small group of patients, all with fingerprint-3. Cluster 2 represented patients with fingerprint-1. Cluster 3 represented patients with primarily fingerprint-2, while cluster 4 represented patients with primarily fingerprint-3. As in the respective fingerprint-groups, patients with cluster 2 had the lowest disease activity, while patients with cluster 4 had the highest disease activity (figure 4B). Interestingly, within cluster 3, patients that had a fingerprint-3 had a higher disease activity. These data indicate that gene fingerprints are robust tools that match with an automated clustering method and correctly identify patients with similar disease activity.

DISCUSSION

We studied four gene signatures in SLE derived from previously described transcriptomic data to develop a method that can easily be applied in clinical practice. These gene signatures were associated with disease activity in general and with disease domains derived from the SELENA-SLEDAI. Upon subgrouping of patients into fingerprints we found a significant difference in disease activity between the fingerprint groups. Fingerprint-1 was associated with low disease activity, while fingerprint-3 represented the patients with high disease activity. We replicated these findings in samples collected over time in the same patient cohort as well as in cross sectional samples of a replication cohort. Hierarchical clustering identified similar gene fingerprints indicating the robustness of our strategy.

Transcriptional profiling is elaborate and costly, resulting in signatures that consist of large gene sets that are simultaneously upregulated [12]. Translation of these data into signatures of a restricted number of genes would facilitate introduction into clinical practice. Therefore, we used a principal components analysis (PCA)- approach to identify genes that explain more than 95% of the total variance in the gene groups. Between the indicator genes that describe a specific gene signature, we observed a high association while genes from different gene signatures lacked this association. These results are in line with previous studies, where indicator genes from each individual gene signature are driven by their own unique pathway [12, 21]. Previously, Chiche et al. described the intra-individual variation of the M1.2 and M5.12 IFN gene signatures obtained from transcriptomics in longitudinal samples from 29 SLE patients [7]. We were able to reproduce these findings for the M1.2 and M5.12 IFN gene signatures obtained via the PCA approach, in our longitudinal cohort of 73 patients. Interestingly, demographic and clinical differences between the two cohorts didn't affect these observations. We further demonstrated for the first time, that the NPh and PLC signatures, showed a dynamic character over time. Importantly, the presence of the described signatures in our cohort

was in line with previous findings obtained by microarray analysis by Banchereau and colleagues [12]. This indicates that the signatures are a reliable approach that can substitute transcriptomic analysis.

Confirming previous data, disease activity was associated with each individual gene signature [5, 7, 10, 12]. In line with the findings of Banchereau et al, the PLC signature showed the best association with disease activity in a cross-sectional cohort [12]. Additionally, we now show in our longitudinal cohort of Tx naïve patients that the PLC signature aligns significantly with disease activity as well. Yet, the correlation coefficients are rather low. Disease activity is measured by scoring the involvement of various disease domains. Here we show that each gene signature is linked to specific disease domains. The M1.2 IFN signature was associated with the skin domain while the NPh and PLC signatures associated significantly with the musculoskeletal and constitutional domains. In contrast to previous findings the NPh signature was not associated with renal involvement [10, 12, 16]. As shown by Banchereau et al and Wither et al., the NPh signature is mostly increased during the active phase of lupus nephritis [10, 12]. The contrasting results between our study and previous published results might be due to the relative low number of patients that had active lupus nephritis at time of sample collection. Nevertheless, these data indicate that a low correlation of individual gene signatures with disease activity could be a consequence of the effect of these signatures on different disease domains.

Our longitudinal data indicate that there might be an association between prednisone use and a positive NPh signature. This observation is in line with previous data, showing that neutrophil numbers are increased in individuals using corticosteroids [12, 22, 23]. Our study is the first longitudinal study that confirms previous findings by Banchereau et al. showing that corticosteroids influence the NPh signature. Also, the finding that the PLC signature in longitudinal cSLE samples is sensitive to changes in disease activity and is affected by the use of prednisone and MMF is in line with previously described results [12]. Studies in the MRL/lpr mouse model for SLE showed that prednisone treatment was associated with a significant decrease of plasma cell numbers [24], that linked to a decrease in BLIMP-1, which regulates plasma cell formation [25]. Interestingly, BLIMP-1 correlated to increased plasma cell numbers and disease activity in SLE patients [25, 26]. Moreover, the neutrophil count and anti-dsDNA represented the NPh and PLC signatures, indicating that cell compositions are drivers of the gene signatures. Considering the IFN α 2 levels, we show that the IFN gene signatures don't have the ability to parallel changes in disease activity while ultra-sensitive analysis of IFN α 2 can. This is in line with previous findings [27]. This finding underscores that in contrast to the NPh and PLC signatures the IFN gene signatures are not influenced by the use of medication and

potentially have a biological role in disease manifestation. Future longitudinal studies in SLE patients are needed to confirm our observations.

We identified 3 fingerprints by cluster analysis of four different gene signatures. These fingerprints were able to discriminate between patients who are in remission (fingerprint-1- “all signature low”) or have high disease activity (fingerprint-3- “high NPh and/ or PLC”) in two different cohorts. This observation is in line with previous work showing that adult SLE patients with a low IFN-I signature had significantly lower disease activity compared to patients with a high IFN-I and high NPh signature [23]. Moreover, the demonstration that unsupervised hierarchical clustering analysis paralleled the identification of the gene fingerprint groups shows the robustness of this novel approach.

Our logistic regression model indicated that fingerprints were associated with different organ domains. The identified associations highly reflect the drivers of each fingerprint group. Fingerprint-1 is particularly driven by the M1.2 IFN gene signature, as this signature is associated with skin involvement. Fingerprint-2 seems to be driven by the M5.12 gene signature as this signature was not associated with any organ domain. Lastly, fingerprint-3 is driven by the NPh and PLC signatures, as these signatures are associated with the musculoskeletal and constitutional domains. Moreover, our results illustrate that autoantibody-profiles are different among patients within various fingerprint groups adding to the knowledge on the relation between autoantibody profiles, disease activity and disease phenotype [28, 29], yet also highlighting their gaps and underlining that mere autoantibody-profiles are not enough for subtyping SLE patients.

Previous data showed that the presence of an IFN signature is associated with an increased chance of disease flare in 5-years [30], supporting a role in disease pathogenesis. Also, higher baseline serum IFN-alpha levels measured by Simoa during SLE remission identified patients at risk for relapse [31]. Interestingly, in our cohorts the patients with only high IFN scores clustered together in fingerprint-2 and they had intermediate disease activity scores when compared to patients with fingerprint-1 and fingerprint-3. Further studies will have to show whether these patients may be specifically at risk to develop a disease flare. Recently, Northcott et al showed that high expression of IFN-I is associated with limited efficacy of glucocorticoids in SLE patients suggesting that IFN gene signatures can predict treatment efficacy and therefore are candidates for future individualized treatment choices [32].

This study has several strengths. This is a study measuring four different gene signatures in a longitudinal multicenter cohort of SLE patients with $\pm 25\%$ being Tx_{naive}. Moreover, the reproducibility of our observations in an independent replication cohort and

unsupervised clustering strategy with similar findings shows the robustness of this approach. Our study also has limitations. With the current cohort of 101 patients, we are underpowered to study specific disease phenotypes like lupus nephritis and neuropsychiatric lupus. Furthermore, disease activity was assessed by the SELENA-SLEDAI. A disadvantage of this scoring system is that it does not consider improvement or worsening of disease items. Therefore, the SELENA-SLEDAI is less sensitive to changes in disease activity compared to other measurement scales [33]. Lastly, it is important to mention that gene signatures and fingerprints, especially the NPh and PLC signatures, may be influenced by medication use. Therefore, medication use should always be considered as a confounding factor before results on fingerprints are interpreted.

CONCLUSION

In conclusion, this study shows that the approach using PCA to identify indicator genes for gene groups, is a successful method to translate existing transcriptomic data into a tool that can be applied in clinical practice. We confirmed the activation of four gene signatures previously identified by transcriptomics and reproduced these data in an independent replication cohort. Moreover, combining the gene signatures into so-called fingerprints enabled us to stratify patients into subgroups with similar activated immune pathways that were associated with disease activity over time in our longitudinal cohort study. The heterogeneity of SLE is reflected in the variability of drug responsiveness between patients. This is expected to become more given the current focus on the development of new biologicals that specifically target specific molecules or immune pathways. We have identified a molecular tool, stratifying patients into groups with similar biological disease profiles that has the potential to guide individualized treatment choices and improve the trial-and-error treatment approach of the present time. Our findings should be confirmed in large longitudinal studies to elucidate the applicability of this tool for prediction of responses on treatments interfering with the aberrantly activated immune pathways.

ACKNOWLEDGEMENTS

This work was made possible by the support of the Sophia Children's Hospital Fund, NVLE (Dutch patient organization for Lupus, APS, Scleroderma and MCTD) and Dutch Arthritis Society. The research was performed within the framework of the Erasmus Postgraduate School Molecular Medicine. The authors thank all the cSLE patients and HC for taking part in this study. The authors thank I. Bodewes, S. Bergkamp, D. Timmermans, K. van Rijswijk and L. van den Berg, for assistance with sample collection and data acquisition. Furthermore, the authors thank dr. J. Göpfert, head of Applied Biomarkers and Immunoassays at NMI for the measurement of the IFNa2 with the Simoa.

SUPPLEMENTARY METHODS

Blood collection and real-time PCR

Blood was collected in PAXgene RNA tubes and was stored in -80°C until use for whole blood RNA purification. RNA isolation, cDNA preparation and real-time PCR were performed according to manufacturer's protocol. In short, total RNA was isolated from PAXgene tubes and reverse-transcribed to cDNA. For calculation of relative gene expressions, samples were normalized to expression of the household gene Abl. Relative expression values were determined from normalized CT values using the $2^{-\Delta\text{CT}}$ method.

Gene selection and gene signature definitions

For each gene signature, genes were selected from the most frequently described genes from transcriptional studies published previously (supplementary table T1, figure S1). To identify correlated groups of genes and reduce data complexity, the expression of 10 IFN-I inducible genes from the M1.2 IFN module, 15 neutrophil (NPh)- and 7 plasma cell (PLC) associated genes were added to a principle component analysis (PCA). Kaiser-Meyer-Olkin measure of sampling adequacy were respectively 0.744, 0.796 and 0.792.

The gene scores for each signature were defined by the sum of the relative expression of 2-5 indicator genes. A \log_2 transformation was used as an intermediate step in the calculation process to generate both + (up) and - (downregulated) gene expression values. The gene signature formula is listed below. For the M1.2 IFN signature these were IFI44, IFIT1, IFIT3, Ly6e and MxA; for the NPh signature DEFA4, BPI, CEACAM6, CEAMCAM 8 and DEFA3/1; and for the PLC signature IGJ and TXNDC5. The gene score for the M5.12 IFN signature was defined by the relative expression of 5 indicator genes (PSMB9, ISG20, NCOA7, SP140 and TAP1) as previously described [21]. Mean and SD of each gene in HC were used to standardize expression levels. The gene scores per subject represent the sum of these standardized scores, calculated as previously described [4, 5]. Patients were divided in groups being positive and negative for a signature, using a threshold of $\text{mean} + 2 \times \text{SD}_{\text{HC}}$.

Gene signature formula

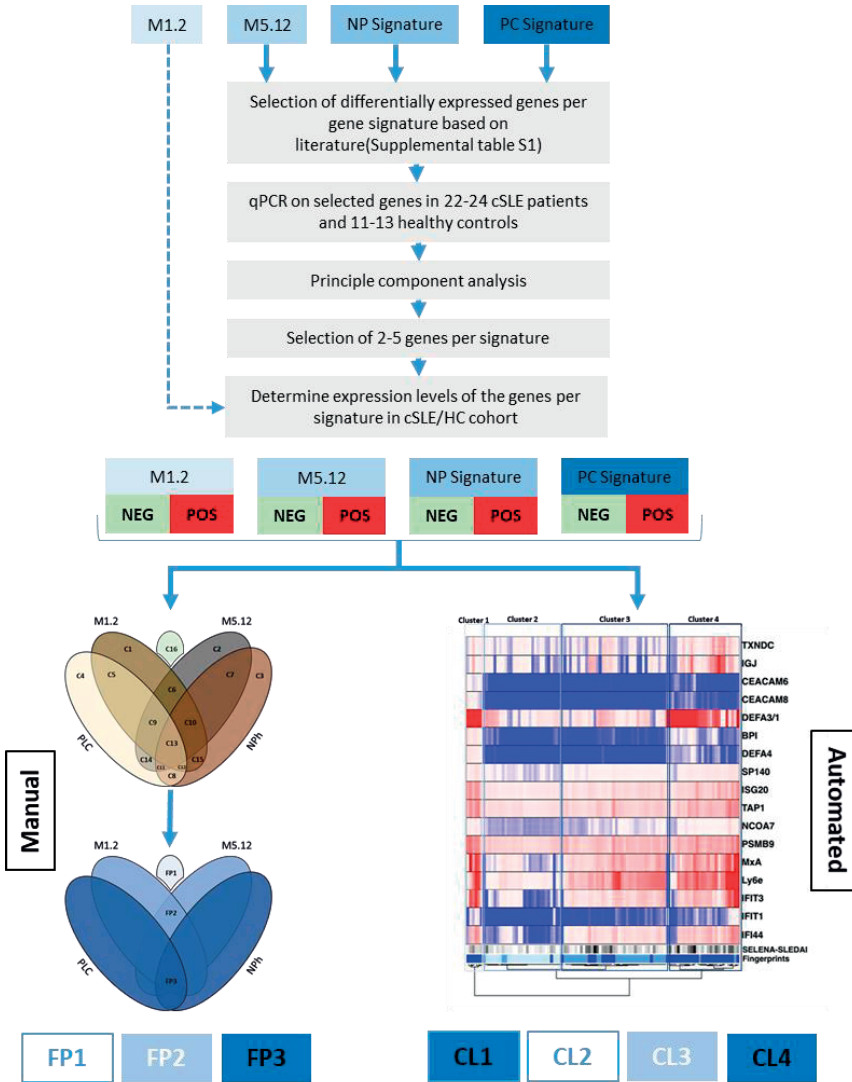
$\Sigma(\text{Log}_2^{-\Delta\text{Ct}}(\text{subjects}) - \text{mean Log}_2^{-\Delta\text{Ct}}(\text{healthy controls})) / \text{st.dev. Log}_2^{-\Delta\text{Ct}}(\text{healthy controls})$.

Ultrasensitive IFN- α Simoa

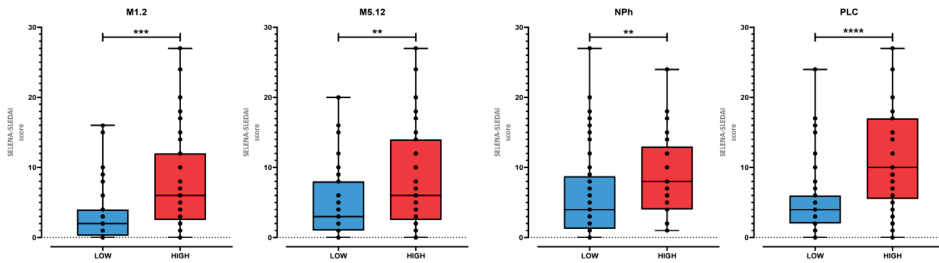
IFN- α 2 was measured in duplicates from serum samples (diluted two-fold in sample diluent) using the Simoa IFN- α Advantage Kit (no. 100860, Quanterix, Billerica, MA, USA) following the instructions of the supplied manual. Sample processing and analysis were done using an HD-X analyser (software version 1.6.1905.300; Quanterix). The lower limit of detection was 5 fg/ml.

Statistics

The non-parametric Mann-Whitney U (two groups) and Kruskal-Wallis (three groups) tests were used to analyze comparisons between medians. The paired *t* test was used to compare means of paired data. Chi squared test and Fisher's exact test were used to compare categorical data. Spearman's rho (r_s) coefficient or Pearson correlation coefficient was calculated to assess correlation. Regression analysis was performed to relate organ domains to gene signatures or fingerprints. All domains with an individual effect of $p < 0.1$ were used to build a multivariable model. Values of $p < 0.05$ were considered statistically significant. Unsupervised hierarchical clustering using Ward's agglomerative method, passing Euclidean distance between samples was used to identify clusters enriched in cSLE patients. Graphpad Prism 8.0 (Graphpad Software, La Jolla, CA, USA) and R statistical software were used for graph design and statistical analysis. Due to the exploratory nature of our study, we refrained from performing multiple testing correction.



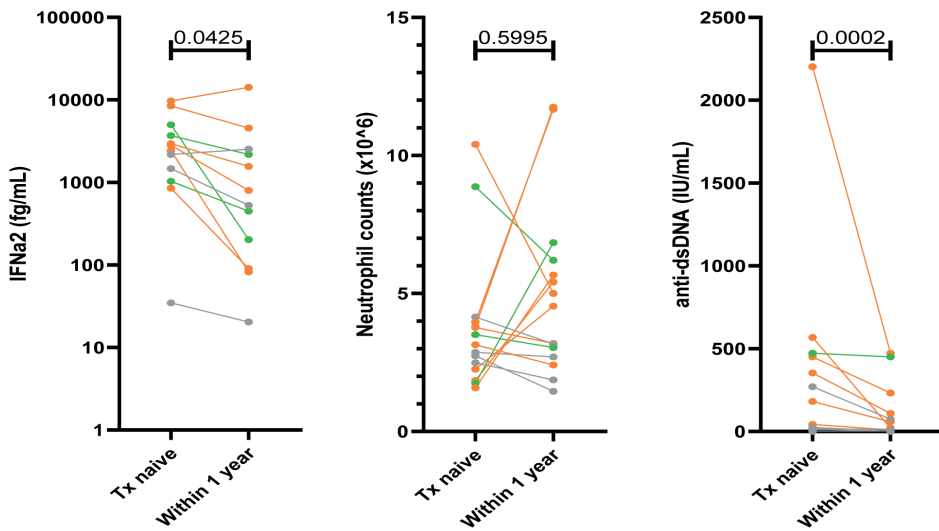
Supplementary figure S1. Flowchart illustrating the process of gene selection and clustering strategies for the M1.2, NP and PLC signature, genes were selected by choosing the most frequently described genes from transcriptional studies published previously. Gene expression was determined in 22-24 cSLE patients and 11-13 healthy controls by qPCR. Expression data was added into a principle component analysis. For each gene signature 2-5 indicator genes were selected. Expression of these indicator genes were further determined in the rest of the blood samples. Indicator genes of the M5.12 IFN signature were previously described by our group (21). A semi-manual and an automated clustering strategy were performed. The semi-manual approach identified 16 clusters based on the combination of a positive or negative score per gene signature. The gene signatures measured at the first study time point of each patient were used. These clusters were subsequently divided into 3 so-called fingerprints. As a starting point we chose a group of patients in which all gene signatures were low. We combined the two IFN gene signatures together due to their high inter-correlation (Figure 1A). We defined Fingerprint 2 by having either a positive M1.2 or a positive M5.12 gene signature but without having a positive NP or positive PLC gene signature. Fingerprint 3 was defined by having a positive NP and/or positive PLC gene signature, independent of the IFN gene scores. Notably, the fingerprint patient groups were mutually exclusive, each patient only fits in one fingerprint group. For the automated clustering strategy, an unsupervised hierarchical clustering method was used to identify clusters that were enriched in the cSLE cohort.
 IFN: Interferon, NP: Neutrophil, PLC: Plasma cell, FP: Fingerprint, CL: Cluster.



Supplementary figure S2. SELENA-SLEDAI score in patients with a low or high gene signature (N=101)

A). M1.2 IFN signature. B). M5.12 IFN signature. C). NPh signature. D). PLC signature. The mean + 2xSD_{HC} was used as a threshold for dividing patients into the low or high group.

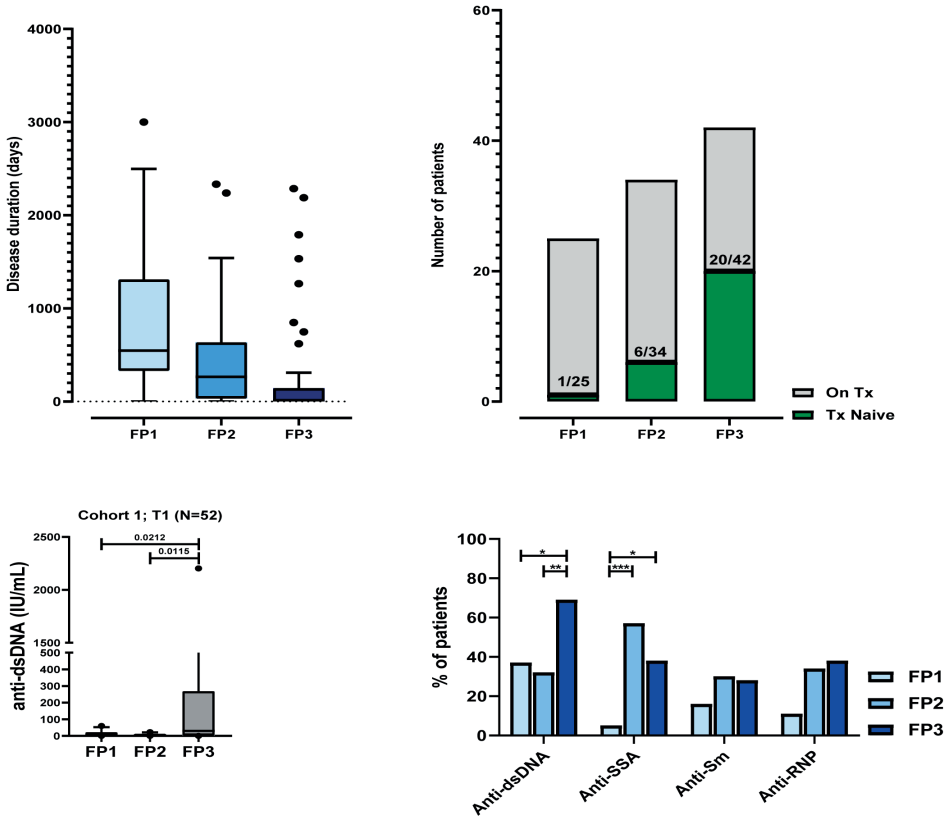
Mann-Whitney-U test was used to compare the two groups; *p<0.05; **p<0.01; ***p<0.001; ****p<0.0001. IFN: Interferon, NPh: Neutrophil, PLC: Plasma cell.



Supplementary figure S3. IFNa2, Neutrophil count and anti-dsDNA level in 20 Tx naive patients over time
A). IFNa2 (n=12). B). Neutrophil count (n=15). C). anti-dsDNA (n=16).

Mann-Whitney-U test was used to compare the two groups; *p<0.05; **p<0.01; ***p<0.001; ****p<0.0001. IFNa2, Interferon alpha 2.

New biomarkers in (systemic) connective tissue diseases



Supplementary figure S4. Disease duration, medication use and autoantibodies per fingerprint
 A). Distribution of disease duration between fingerprints. B). Number of treatment naïve patients per fingerprint. Numbers represent treatment naïve patients per group total. C). Anti-dsDNA level distribution per fingerprint in a selection of cSLE cohort 1; first time point (N=52). Levels are based on a fluorescent enzyme immune assay, reference value >10 IU/mL. D). Percentage of patients with a positive auto-antibody per fingerprint in cSLE cohort 1 (N=86).
 Mann-Whitney-U test was used to compare the two groups; chi-square test was used to compare categorical data
 *p < 0.05; **p < 0.01; ***p < 0.001; ****p < 0.0001. FP, Fingerprint. Tx, Treatment.
 On Tx indicates Hydroxychloroquine with or without Mycophenolate Mofetil and/or Prednisone.

Supplementary table T1. List of genes per signature

M1.2 IFN	M5.12 IFN				NPh	PLC			
IFI44	PSMB9				DEFA4	IGJ			
IFIT1	ISG20				BPI	TXNDC5			
IFIT3	NCOA7				DEFA3/DEFA1	CD38			
Ly6e	SP140				CEACAM8	GLDC			
MxA	TAP1				CEACAM6	TNFRSF17			
Serping					MPO	IGHA1			
IFI27					ELANE	IGKC/IGKV5-1			
IFI44L					AZU1				
XAF1					MMP8				
ISG15					LTF				
					CTSG				
					LCN2				
					MS4A3				
					OLFM4				
					RNS2				
M1.2									
<i>Bodewes, 2018</i>	IFI44	IFIT1	IFIT3	Ly6e	MxA	Serping	IFI27	IFI44L	XAF1
	ISG15								
M5.12									
<i>Bodewes, 2018</i>	PSMB9 ISG20 NCOA7 SP140 TAP1								
Neutrophil associated genes									
<i>Bennett, 2003</i>	RNS2	RNAse 2	MMP-9	RNS3	HUSI-1	ELASTASE 2	LCN2	CG	
	cathepsin G	MPO	NCF	DEF3	DEFA4	DEFA1	FALL-39	BPI	AZU
	EST	GCA	S100P	CD24A	CD66A	CD66B	TC1		
	CRISPR3	F2RPA							
<i>Villanueva, 2011</i>	MMP8	RNASE3	ELANE	LCN2	MPO	CTSG	DEFA4	BPI	AZU1
	CEACAM8	CEACAM1	LTF	CLU					
<i>Banchereau M5.15, 2016</i>	ARG1	AZU1	BPI	CAMP	CEACAM6	CEACAM8			
	COL17A1	CTSG	DEFA1	DEFA3	DEFA4	EIF1AY	ELA2	HLA-DRB1	HLA-DRB5
	HP	LOC653600	LTF	MMP8	MPO	MS4A3	OLR1	RETN	TCN1
<i>Carlucci, 2018</i>	MPO	ELANE	PRTN3	CTSG	AZU1	DEFA4			
<i>Wither, 2018</i>	MPO	DEFA4	DEFA3	DEFA1	MMP8	CEACAM6	CEACAM8		
	LTF	MS4A3	OLFM4	CRISP3	LCN2	BPI			
<i>Petri, 2019</i>	DEFA4	CEACAM8	CEACAM6	MMP8	LCN2	BPI		OLFM4	
	LTF								
Plasma cell associated genes									
<i>Streicher, 2014</i>	IGHA1	IGJ	IGKC	IGKV4-1	TNFRSF17				
<i>Banchereau M4.11, 2016</i>	CAMK1G	CD38	GLDC	IGJ	TNFRSF17	TXNDC5			
<i>Petri, 2019</i>	IGJ	TXNDC5							

*Grey color illustrates the indicator genes per gene signature.

Supplementary table T2. Binary logistic regression analysis of organ domains associated with different fingerprints as outcome measure

	Univariate								
	FP1			FP2			FP3		
	B	P	Exp	B	P	Exp	B	P	Exp
CNS	20.104	0.999	0	20.556	0.999	0	-1.34	0.281	0.262
Musculoskeletal	1.775	0.094	5.902	0.911	0.18	2.1488	-1.721	0.004	0.179
Renal	0.963	0.15	2.619	0.462	0.384	1.587	-1.197	0.015	0.302
Skin	1.281	0.02	3.6	-0.401	0.347	0.67	-0.683	0.109	0.505
Vascular	20.232	0.999	0	0.774	0.345	2.169	-1.658	0.022	0.19
Serositis	20.232	0.999	0	20.602	0.999	0	-21.991	0.999	0
Constitutional	1.402	0.191	4.062	1.869	0.08	6.482	-1.984	0.005	0.138
Immunological	0.912	0.061	2.489	0.816	0.061	2.261	-1.91	0.0001	0.148
Haematological	1.114	0.02	3.048	-0.147	0.73	0.863	-0.78	0.076	0.458
Multivariate									
CNS									
Musculoskeletal	1.495	0.173	4.461				-1.404	0.043	0.246
Renal							-1.038	0.075	0.354
Skin	1.118	0.05	3.06						
Vascular							-0.267	0.769	0.766
Serositis									
Constitutional							-2.062	0.012	0.127
Immunological	0.415	0.429	1.515	1.752	0.103	5.765	-1.449	0.008	0.235
Haematological	0.852	0.092	2.344	0.734	0.097	2.084	-0.217	0.689	0.805

A cut-off of $p < 0.10$ was set to select the variables for the multivariate logistic regression. In the analysis a value of 1 indicated the involvement of a fingerprint while 0 indicated no involvement of a fingerprint, a negative value for B therefore means more association with a certain domain, while a positive B indicated less association.

CNS: Central Nervous System, FP: Fingerprint.

REFERENCES

1. Mohan C, Putterman C. Genetics and pathogenesis of systemic lupus erythematosus and lupus nephritis. *Nat Rev Nephrol.* 2015;11(6):329-41.
2. Baechler EC, Batliwalla FM, Karypis G, Gaffney PM, Ortmann WA, Espe KJ, et al. Interferon-inducible gene expression signature in peripheral blood cells of patients with severe lupus. *Proc Natl Acad Sci U S A.* 2003;100(5):2610-5.
3. Bennett L, Palucka AK, Arce E, Cantrell V, Borvak J, Banchereau J, et al. Interferon and granulopoiesis signatures in systemic lupus erythematosus blood. *J Exp Med.* 2003;197(6):711-23.
4. Kirou KA, Lee C, George S, Louca K, Papagiannis IG, Peterson MG, et al. Coordinate overexpression of interferon-alpha-induced genes in systemic lupus erythematosus. *Arthritis Rheum.* 2004;50(12):3958-67.
5. Feng X, Wu H, Grossman JM, Hanvivadhanakul P, FitzGerald JD, Park GS, et al. Association of increased interferon-inducible gene expression with disease activity and lupus nephritis in patients with systemic lupus erythematosus. *Arthritis Rheum.* 2006;54(9):2951-62.
6. Chaussabel D, Quinn C, Shen J, Patel P, Glaser C, Baldwin N, et al. A modular analysis framework for blood genomics studies: application to systemic lupus erythematosus. *Immunity.* 2008;29(1):150-64.
7. Chiche L, Jourde-Chiche N, Whalen E, Presnell S, Gersuk V, Dang K, et al. Modular transcriptional repertoire analyses of adults with systemic lupus erythematosus reveal distinct type I and type II interferon signatures. *Arthritis Rheumatol.* 2014;66(6):1583-95.
8. Villanueva E, Yalavarthi S, Berthier CC, Hodgkin JB, Khandpur R, Lin AM, et al. Netting neutrophils induce endothelial damage, infiltrate tissues, and expose immunostimulatory molecules in systemic lupus erythematosus. *J Immunol.* 2011;187(1):538-52.
9. Streicher K, Morehouse CA, Groves CJ, Rajan B, Pilataxi F, Lehmann KP, et al. The plasma cell signature in autoimmune disease. *Arthritis Rheumatol.* 2014;66(1):173-84.
10. Wither JE, Prokopec SD, Noamani B, Chang NH, Bonilla D, Touma Z, et al. Identification of a neutrophil-related gene expression signature that is enriched in adult systemic lupus erythematosus patients with active nephritis: Clinical/pathologic associations and etiologic mechanisms. *PLoS One.* 2018;13(5):e0196117.
11. Carlucci PM, Purmalek MM, Dey AK. Neutrophil subsets and their gene signature associate with vascular inflammation and coronary atherosclerosis in lupus: ncbi.nlm.nih.gov; 2018.
12. Banchereau R, Hong S, Cantarel B, Baldwin N, Baisch J, Edens M, et al. Personalized Immunomonitoring Uncovers Molecular Networks that Stratify Lupus Patients. *Cell.* 2016;165(3):551-65.
13. Odendahl M, Jacobi A, Hansen A, Feist E, Hiepe F, Burmester GR, et al. Disturbed peripheral B lymphocyte homeostasis in systemic lupus erythematosus. *J Immunol.* 2000;165(10):5970-9.
14. Kaplan MJ. Neutrophils in the pathogenesis and manifestations of SLE. *Nat Rev Rheumatol.* 2011;7(12):691-9.
15. Wirestam L, Arve S, Linge P, Bengtsson AA. Neutrophils-Important Communicators in Systemic Lupus Erythematosus and Antiphospholipid Syndrome. *Front Immunol.* 2019;10:2734.
16. Jourde-Chiche N, Whalen E, Gondouin B, Speake C, Gersuk V, Dussol B, et al. Modular transcriptional repertoire analyses identify a blood neutrophil signature as a candidate biomarker for lupus nephritis. *Rheumatology (Oxford).* 2017;56(3):477-87.
17. Toro-Dominguez D, Martorell-Marugan J, Goldman D, Petri M, Carmona-Saez P, Alarcon-Riquelme ME. Stratification of Systemic Lupus Erythematosus Patients Into Three Groups of Disease Activity

- Progression According to Longitudinal Gene Expression. *Arthritis Rheumatol.* 2018;70(12):2025-35.
18. Petri M, Orbai AM, Alarcon GS, Gordon C, Merrill JT, Fortin PR, et al. Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis Rheum.* 2012;64(8):2677-86.
 19. Aringer M, Costenbader K, Daikh D, Brinks R, Mosca M, Ramsey-Goldman R, et al. 2019 European League Against Rheumatism/American College of Rheumatology Classification Criteria for Systemic Lupus Erythematosus. *Arthritis Rheumatol.* 2019;71(9):1400-12.
 20. Petri M, Kim MY, Kalunian KC, Grossman J, Hahn BH, Sammaritano LR, et al. Combined oral contraceptives in women with systemic lupus erythematosus. *N Engl J Med.* 2005;353(24):2550-8.
 21. Bodewes ILA, Al-Ali S, van Helden-Meeuwsen CG, Maria NI, Tarn J, Lendrem DW, et al. Systemic interferon type I and type II signatures in primary Sjogren's syndrome reveal differences in biological disease activity. *Rheumatology (Oxford).* 2018;57(5):921-30.
 22. Shoenfeld Y, Gurewich Y, Gallant LA, Pinkhas J. Prednisone-induced leukocytosis. Influence of dosage, method and duration of administration on the degree of leukocytosis. *Am J Med.* 1981;71(5):773-8.
 23. Chasset F, Ribl C, Trendelenburg M, Huynh-Do U, Roux-Lombard P, Courvoisier DS, et al. Identification of highly active systemic lupus erythematosus by combined type I interferon and neutrophil gene scores vs classical serologic markers. *Rheumatology (Oxford).* 2020;59(11):3468-78.
 24. Yan SX, Deng XM, Wang QT, Sun XJ, Wei W. Prednisone treatment inhibits the differentiation of B lymphocytes into plasma cells in MRL/MpSlac-lpr mice. *Acta Pharmacol Sin.* 2015;36(11):1367-76.
 25. Luo J, Niu X, Liu H, Zhang M, Chen M, Deng S. Up-regulation of transcription factor Blimp1 in systemic lupus erythematosus. *Mol Immunol.* 2013;56(4):574-82.
 26. Shapiro-Shelef M, Lin KI, McHeyzer-Williams LJ, Liao J, McHeyzer-Williams MG, Calame K. Blimp-1 is required for the formation of immunoglobulin secreting plasma cells and pre-plasma memory B cells. *Immunity.* 2003;19(4):607-20.
 27. Mathian A, Mouries-Martin S, Dorgham K, Devilliers H, Barnabei L, Ben Salah E, et al. Monitoring Disease Activity in Systemic Lupus Erythematosus With Single-Molecule Array Digital Enzyme-Linked Immunosorbent Assay Quantification of Serum Interferon-alpha. *Arthritis Rheumatol.* 2019;71(5):756-65.
 28. To CH, Petri M. Is antibody clustering predictive of clinical subsets and damage in systemic lupus erythematosus? *Arthritis Rheum.* 2005;52(12):4003-10.
 29. Jurencak R, Fritzler M, Tyrrell P, Hiraki L, Benseler S, Silverman E. Autoantibodies in pediatric systemic lupus erythematosus: ethnic grouping, cluster analysis, and clinical correlations. *J Rheumatol.* 2009;36(2):416-21.
 30. Mai L, Asaduzzaman A, Noamani B, Fortin PR, Gladman DD, Touma Z, et al. The baseline interferon signature predicts disease severity over the subsequent 5 years in systemic lupus erythematosus. *Arthritis Res Ther.* 2021;23(1):29.
 31. Mathian A, Mouries-Martin S, Dorgham K, Devilliers H, Yssel H, Garrido Castillo L, et al. Ultrasensitive serum interferon-alpha quantification during SLE remission identifies patients at risk for relapse. *Ann Rheum Dis.* 2019;78(12):1669-76.
 32. Northcott M. Glucocorticoid gene signatures in systemic lupus erythematosus and the effects of type I interferon: a cross-sectional and in-vitro study. *The Lancet Rheumatology.* 2021;Volume 3(issue 5):e357 - e70.

33. Mikdashi J, Nived O. Measuring disease activity in adults with systemic lupus erythematosus: the challenges of administrative burden and responsiveness to patient concerns in clinical research. *Arthritis Res Ther.* 2015;17:183.

7

Rarities in rare: illuminating the microvascular and dermal status in juvenile localized scleroderma. A case series

Amber Vanhaecke*, **Dieneke Schonenberg-Meinema***, Sofie de Schepper,
Sandy C. Bergkamp, Maria C. Leone, Maritza A. Middelkamp-Hup,
Amara Nassar-Sheikh Rashid, J Merlijn van den Berg, Taco W. Kuijpers,
Annamaria Iagnocco, Maurizio Cutolo, Vanessa Smith

*shared first authors

Published in Clinical and Experimental Rheumatology 2022; 40 (Suppl. 134): S12-S18

ABSTRACT

OBJECTIVE: To assess the (structural and functional) characteristics of the microvascular and dermal status in juvenile localised scleroderma (jLoS), using novel non-invasive standardised research tools commonly used in adult systemic sclerosis (SSc).

METHODS: Ten consecutive patients with a confirmed jLoS diagnosis were studied cross-sectional in this two-centre case series. For each patient, the most prominent lesion (i.e. “target lesion”) was chosen for further examination of the centre, edge and contralateral unaffected site. High-frequency ultrasonography was used to determine dermal thickness, durometer for skin hardness, and laser speckle contrast analysis (LASCA) for a dynamical evaluation of the microcirculation. The structure of the microcirculation was evaluated at the nailfolds of the 2nd-5th finger bilaterally, using nailfold videocapillaroscopy (NVC).

RESULTS: 6 linear and 4 plaque subtype jLoS lesions were included. Dermal thickness was thinner at the centre of the “target lesions” vs. the edges ($p<0.001$) and control sites ($p<0.001$). Skin hardness was harder at the centre of the “target lesions” vs. the edges ($p=0.012$) and control sites ($p=0.003$). A higher perfusion was found in the centre of the “target lesion” (124.87 ± 66.40 PU) vs. the edges (87.27 ± 46.40 PU; $p<0.001$) and control sites (67.85 ± 37.49 ; $p<0.001$). Of note, all patients had a “non-scleroderma” pattern on NVC.

CONCLUSION: This case series suggests the supportive value of both microcirculatory and dermal assessments of skin lesions using novel non-invasive research tools, adopted from adult SSc, for the identification, scoring and/or monitoring of jLoS.

INTRODUCTION

Scleroderma comprises a group of rare (incidence of ± 1 per 100.000/year) fibrosing disorders with similar histopathological findings [1-8]. Scleroderma can be broadly divided into systemic sclerosis (SSc, prevalence ranging between 7 and 44 per 100.000 individuals) and localised scleroderma (LoS, also called morphea, estimated prevalence ± 50 per 100.000 individuals). SSc is a multisystem connective tissue disease hallmarked by a triad of vasculopathy, autoimmunity and fibrosis of the skin and/or internal organs, LoS is pathological process of local and chronic inflammation mainly affecting part(s) of the skin and underlying tissues, which eventually leads to fibrosis and atrophic changes [1, 6-12]. In contrast to SSc, LoS is a very rare condition that usually presents in childhood (juvenile LoS [jLoS]) with a mean age of onset at 6-8 years [9, 12]. The estimated incidence of jLoS is estimated at 3.4 cases per million children per year [13, 14].

It is believed that SSc and (j)LoS share common pathophysiological pathways with an initial inflammatory phase accompanied by endothelial activation, followed by a fibrotic phase characterised by tissue collagenisation and appreciable skin thickness sometimes accompanied by atrophic changes [3, 7-9, 15-17]. The hypothesis of a common pathophysiological pathway is further supported by recent observations from our research group, as we found both in literature and in a pilot study that the coexistence of SSc and (j)LoS (2.4 to 7.4%) is higher than their individual prevalence in the healthy population [16].

Although it is not a lethal disease like SSc, (j)LoS can lead to severe physical damage with growth deformities, functional disabilities and cosmetic impairment, which may eventually lead to chronic psychological besides physical problems [18-20]. As there are no antifibrotic therapies yet that can cure the disease, early anti-inflammatory aggressive systemic treatment regimens are often required to halt disease progression and prevent poor outcomes [21-25].

In daily clinical practice the validated Localised Scleroderma Cutaneous Assessment Tool (LoSCAT) is being used combining assessment of disease activity using the modified Skin Severity Index (mLoSSI) and assessment of disease damage using the modified Skin Damage Index (mLoSDI) [26, 27]. Because the LoSCAT only provides a global impression of all affected (j)LoS localisations, there is a need for more validated (imaging) tools to objectively identify and monitor inflammation and tissue damage *per* (j)LoS lesion. Those tools are an unmet need in making treatment decisions, for instance when to increase, switch, taper or to stop treatment in young aged patients and should consequently predict and monitor treatment response. The lack of these tools not only hampers clinical daily treatment decisions, but also complicates the conduct of clinical

trials. Several non-invasive imaging modalities, such as thermography, laser doppler flowmetry and (doppler) ultrasound have been proposed, but are hampered by user dependency and lack of standardisation [28]. Magnetic resonance imaging (MRI) and cone beam computed tomography (CBCT) are other imaging possibilities with the disadvantages of high costs and invasiveness for young children, not only during examination (MRI) but also because of negative radiation risks (CBCT) [29-37].

Unlike in SSc, where the evaluation of the microcirculation (using non-invasive standardised tools such as nailfold videocapillaroscopy [NVC] and laser speckle contrast analysis [LASCA]) has earned a pivotal role and the evaluation of skin fibrosis using high-frequency ultrasonography (HFUS) and durometry is currently an area of significant research and standardisation efforts, exhaustive reports using these tools in jLoS are non-existent [1, 6-9, 38-54]. Against this background, we felt it was time to descriptively assess the (structural and functional) characteristics of the microvascular and dermal status in a case series of jLoS patients, using non-invasive standardised research tools (i.e. NVC, LASCA, HFUS and durometer) that are commonly used in adult SSc.

MATERIALS AND METHODS

Ethical Vote

This study was approved by the local institutional review boards and local ethics committees (Amsterdam University Medical Centre [2017-172], Ghent University Hospital [EC/2019/1639], and conducted in accordance with the Declaration of Helsinki and Helsinki and its amendments. All parents or legal guardians (for patients < 16 years old) and competent patients (over > 12 years old) signed written informed consent before inclusion.

Study population

This two-centre, observational study was conducted in 2019 at the tertiary Amsterdam University Medical Centre (The Netherlands) and in 2021 at the tertiary Ghent University Hospital (Belgium). Consecutive juvenile patients (≤ 18 years) with a confirmed diagnosis of jLoS, regardless of disease duration, who visited the paediatric rheumatology/immunology and dermatology department for an outpatient visit were recruited [6, 17]. The diagnosis of jLoS was made clinically and, if necessary in doubtful cases, confirmed by histopathological examination (6, 17). In addition, jLoS was classified according to Kreuter's guideline as limited, generalised, linear, deep or mixed type [6]. Patients without demonstrable skin involvement at the time of recruitment, or with the presence of other systemic diseases (e.g. juvenile systemic sclerosis, rheumatoid arthritis, lupus erythematosus) were deemed ineligible for the study.

Data collection

Demographic, anamnestic, clinical and serological data were collected cross-sectionally from electronic medical records. For the purpose of this study, all patients were subjected to a detailed skin and microvascular examination by a team of experienced (paediatric) dermatologists and rheumatologists on the same day.

Skin examination

First, a clinical skin evaluation was performed, including a detailed description of the jLoS lesions in terms of their number, anatomical locations and dimensions and an assessment of clinical signs of disease activity and damage by completing the mLoSSI and the mLoSDI (which are combined in the LoSCAT) [26, 27]. More specifically, disease activity was measured by assessing 3 separate items (new lesion/lesion extension, erythema and skin thickness) at 18 cutaneous anatomical sites (head, neck, chest, abdomen, upper back, lower back, upper arms, forearms, hands/fingers, buttocks/thighs, legs and feet), with a predefined score of 0-3. The scores for each anatomical site were based on the most severe (i.e. highest) score for each item [27]. Disease damage was measured by the comparable mLoSDI, scoring 0-3 on three items (i.e. dermal atrophy, subcutaneous atrophy and dyspigmentation) in the same 18 cutaneous anatomical areas as the mLoSSI [4]. Identically to the mLoSSI, the most severe score obtained from each item was used to calculate the mLoSDI [26].

When multiple lesions were present, the most prominent lesion (i.e. “target lesion”) was chosen for further examination, and in case of large lesions, the most affected site was designated by an expert dermatologist (M.M.-H.). Following the clinical skin evaluation, a bi-instrumental examination of the centre, edge and contralateral unaffected site of the “target lesion” was performed by an experienced investigator (A.V.) using HFUS to assess skin thickness, and a durometer to determine skin hardness. In case of presence of jLoS lesions on the contralateral site, the perilesional unaffected skin was examined.

HFUS images were taken by using a commercially available ultrasound system with a linear probe operating at 18 MHz in B-mode (Logiq S8, GE Healthcare, Chalfont St. Giles, Buckinghamshire, UK). Briefly, the probe was placed perpendicular to the skin by hand, without applying pressure, using a layer of ultrasound gel acting as a coupling agent between the skin surface and the probe. Images were obtained of the centre, edge and contralateral/perilesional unaffected site of the “target lesion”. The dermal thickness (DT), expressed in mm, was subsequently determined by measuring the distance between the epidermis-dermis interface and the dermis-subcutis interface three times and calculating an average DT value for each area [55, 56].

Durometer measurements were performed using a hand-held electronic durometer (RX-DD Digital Durometer, type OO), which has a calibrated continuous scale from 0 to 100 standard durometer units (DU). The durometer was placed perpendicular to the skin and left at rest by gravity of the durometer's weight. As during the HFUS examination, durometer readings were taken in the centre, edge and contralateral/perilesional unaffected site of the "target lesion" to obtain a durometer value for each of these areas [57, 58].

Microcirculation

The microcirculation was evaluated both structurally and dynamically, respectively by using nailfold videocapillaroscopy (NVC) and laser speckle contrast analysis (LASCA).

First, the structure of the microcirculation was evaluated by examining the nailfolds of the 2nd-5th finger bilaterally with an **NVC** probe equipped with a 200x magnification lens. Two adjacent fields in the middle of the nailfold, extending over 1mm and corresponding to the distal row of capillaries, were captured per finger, resulting in 16 images per patient. The NVC images, with a 1 mm grid, were coded and read centrally at the Ghent University Hospital. Quantitative and qualitative assessments of these images were performed according to the consented capillaroscopic definitions of the EULAR Study Group on Microcirculation in Rheumatic Diseases [38, 47, 59, 60].

For the dynamic evaluation of the microcirculation, **LASCA** was performed under standardised conditions as previously described, using a commercially available LASCA instrument (Pericam PSI, Perimed, Jarfalla, Sweden). During the measurements, the areas of interest, being the centre, edge and contralateral/perilesional unaffected site of the "target lesion", were illuminated perpendicularly with a laser beam for 30 seconds, at a fixed distance (20±0.5 cm). Then, the blood perfusion (BP) was evaluated by drawing a standardised circular region of interest (ROI) with a fixed diameter of 1cm in the middle of the area of interest, using LASCA software (PIMSoft 15.1, Perimed AB, Jarfalla, Sweden). Hence, a BP value, expressed in arbitrary perfusion units (PU), was recorded for each of these areas [49-53, 61].

Statistical analysis

Descriptive statistics were used to summarise the data. For nominal categorical variables, absolute numbers with percentages are shown, for ordinal categorical and skewed continuous variables, medians with interquartile ranges (IQR) are shown, and for symmetric continuous variables, means with standard deviation (SD) are shown. To compare the means between the "target lesions" and control sites, paired sample T-tests were used. Pearson's correlations examined the relationship between the mLoSSI/mLoSDI and LASCA, HFUS and durometer measurements of the "target lesions". Signifi-

cance was defined as $p < 0.05$. Statistical analysis are performed with SPSS, version 27 (IBM SPSS Inc., USA).

RESULTS

Study population

Ten patients with a confirmed jLoS diagnosis were included at the tertiary Amsterdam University Medical Centre ($n=9$) and the tertiary Ghent University Hospital ($n=1$). Their demographic, clinical and laboratory characteristics are summarised in table 1. The mean age was 14.6 years, and 60% were female patients. When categorised according to the jLoS subtype, there were 6 patients with linear (5 extremities, 1 ECDS) and 4 with plaque subtype. A total of 22 lesions were found, which were located on the trunk ($n=12$; with 4 on the chest, 4 on the abdomen and 4 on the back), lower extremities ($n=9$; with 6 on the upper legs, 2 on the lower legs, and 1 on the feet), and face ($n=1$).

NVC evaluation

By quantitative analysis, the mean capillary density was 7.4 (± 0.8) capillaries/linear mm. No giant capillaries were observed. The mean number of abnormal capillary shapes was 0.3 (± 0.3) capillaries/linear mm and 5 (50%) patients showed microhaemorrhages. By qualitative analysis, all patients had a “non-scleroderma pattern”. Of them, 5/10 (50%) were classified as having a “normal” NVC pattern, and 5/10 (50%) as having “non-specific abnormalities” (table 1, see end of manuscript).

Clinical and instrumental measurements

Table 2 (see end of manuscript) lists per patient the mLoSSI and mLoSDI measurements, as well as all instrumental measurements of the centre, edge and contralateral/perilesional unaffected sites of each “target lesion”. When examining skin thickness and hardness, significant differences were observed in the centre of the “target lesions” compared to both the edge of the “target lesions” and control sites (tables 3 and 4). More specifically, the dermal thickness was thinner in the centre of the “target lesions” than at the edge of the “target lesions” ($p < 0.001$, table 3) and the control sites ($p < 0.001$, table 4). The centre of the “target lesions” was harder than the edge of the “target lesions” ($p = 0.012$, table 3) and the control sites ($p = 0.003$, table 4). Furthermore, a significant higher BP was observed in the centre of the “target lesions” (124.87 ± 66.40 PU) than at the edge of the “target lesions” (87.27 ± 46.40 PU, $p = 0.001$, table 3) and the control sites (67.85 ± 37.49 PU, $p < 0.001$, table 4).

Table 1. Demographic, clinical and serological characteristics of jLoS patients.

n°	Age	Gender	Race	jLoS subtype	jLoS location	ANA	Age at diagnosis (years)	Disease duration (years)	Systemic treatment (ever)	NVC pattern	mLossI*	mLoSDI*
1	16	Male	African	Linear	Upper leg left	-	8	8	MTX, GCs	Normal	2	6
2	17	Female	Asian	Plaque	Thorax (left, abdomen left (x2), abdomen right , back, lower leg left)	+	10	7	MTX, GCs	Normal	2	3
3	14	Female	Caucasian	Plaque	Thorax left	-	5	9	MTX, tocilizumab	Non-specific	2	7
4	16	Male	Caucasian	Linear	Foot left	+	12	4	MTX, GCs	Non-specific	2	3
5	15	Male	Caucasian	Linear ECDS	Face left	+	12	3	MTX, GCs	Non-specific	3	5
6	18	Female	Mediterranean	Linear	Upper leg left	-	4	14	MTX	Normal	3	5
7	12	Female	African	Plaque	Upper leg right , abdomen right, back central	-	11	1	MTX, GCs	Non-specific	4	5
8	10	Female	Caucasian	Linear	Upper leg left	+	9	1	MTX, GCs	Non-specific	4	6
9	14	Male	Caucasian	Plaque	Back right	-	7	7	MTX, GCs	Normal	1	2
10	14	Female	Caucasian	Linear	Thorax left , upper leg left, upper leg right, thorax left, back middle	-	14	0	MTX, GCs	Normal	3	3

* Target lesion. ANA: antinuclear antibodies; ECDS: *en coup de sabre*; jLoS: juvenile localised scleroderma; mLossI: modified localised scleroderma severity index; mLoSDI: modified localised scleroderma severity index; NVC: nailfold video capillaroscopy; GCs: glucocorticosteroids.

Table 2. Detailed overview of measurements of the “target lesion” and the corresponding control site.

Patient n°	jLoS location	mLoSSI (0-9)			LASCA			mLoSDI (0-12)			HFUS			Durometer		
		Centre	Edge	Control site	Centre	Edge	Control site	Centre	Edge	Control site	Centre	Edge	Control site	Centre	Edge	Control site
1	Upper leg left	2	144.03	96.64	75.75	6	0.10	0.10	0.12	0.14	0.14	37.5	20.5	13.2		
2	Abdomen right	2	159.56	91.45	74.41	3	0.08	0.08	0.10	0.14	0.14	12.0	8.0	7.4		
3	Thorax left	2	144.17	122.14	98.40	7	0.08	0.08	0.09	0.12	0.12	25.6	16.6	7.7		
4	Foot left	2	34.83	24.42	22.16	3	0.08	0.08	0.10	0.15	0.15	33.4	29.9	19.1		
5	Face left	3	261.73	184.99	142.68	5	0.09	0.09	0.13	0.15	0.15	16.6	8.2	4.8		
6	Upper leg left	3	77.23	44.52	28.61	5	0.10	0.10	0.12	0.13	0.13	44.4	39.8	5.7		
7	Upper leg right	4	49.85	42.11	30.43	5	0.08	0.08	0.09	0.12	0.12	44.0	36.1	36.2		
8	Upper leg left	4	97.82	65.59	43.14	6	0.08	0.08	0.09	0.11	0.11	53.8	28.8	21.9		
9	Back right	1	109.47	96.06	81.35	2	0.07	0.07	0.08	0.12	0.12	16.2	19.6	15.4		
10	Thorax left	3	170.01	104.82	81.56	3	0.10	0.10	0.12	0.15	0.15	28.8	25.8	22.8		

jLoS: juvenile localised scleroderma; LASCA: laser speckle contrast analysis; mLoSSI: modified localised scleroderma severity index; mLoSDI: modified localised scleroderma damage index

Table 3. Comparison of measurements of centre of “target lesions” versus edge of “target lesions”.

Variable	Centre of “target lesions”	Edge of “target lesions”	p-value
HFUS, mean ± SD (mm)	0.086 ± 0.01	0.104 ± 0.02	< 0.001
Durometer, mean ± SD (DU)	31.23 ± 13.89	23.33 ± 10.77	0.012
LASCA, mean ± SD (PU)	124.87 ± 66.40	87.27 ± 46.60	0.001

Table 4. Comparison of measurements of jLoS “target lesions” versus control sites.

Variable	Centre of “target lesions”	Control site	p-value
HFUS, mean ± SD (mm)	0.086 ± 0.01	0.133 ± 0.01	< 0.001
Durometer, mean ± SD (DU)	31.23 ± 13.89	15.42 ± 9.88	0.003
LASCA, mean ± SD (PU)	124.87 ± 66.40	67.85 ± 37.49	< 0.001

Bold text: statistical significant finding ($p < 0.05$).

DU: durometer units; HFUS: high-frequency ultrasonography; LASCA: laser speckle contrast analysis; mm: millimetre; PU: perfusion units; SD: standard deviation.

DISCUSSION

This is the first case series in a jLoS population describing the use of non-invasive research tools to evaluate both microcirculatory and dermal properties, as commonly used in adult SSc. Our study provides evidence for future investigation of these tools in the detection and monitoring of (j)LoS lesions.

Our results demonstrate that both skin thickness (i.e. “atrophy”) and skin hardness (i.e. “fibrosis”) of a (j)LoS lesion can be quantitatively measured separately by HFUS and durometer, respectively. Another striking finding is that the centre of a (j)LoS lesion has a higher perfusion than the edge of the same affected skin lesion (scored by LASCA). This higher perfusion within (j)LoS plaques has also been demonstrated by other study groups that used thermography, although their control site was healthy skin instead of the edge of the “target lesions” [62].

Quantitative outcome measures are important for monitoring disease activity in (j)LoS, and the combined measurements of LoSCAT add up several different type of scoring items. To date, infrared thermography, MRI and ultrasonography have also been used to detect disease activity in (j)LoS but those tools may be limited in case of severe skin atrophy [28]. More biomarkers are needed for the treating physician in the chronic treatment of (j)LoS because it is still difficult to know when to increase, modify, taper or stop systemic treatment. Methotrexate, steroids and mycophenolate mofetil are known to be effective in systemic treatment of (j)LoS but not all patients respond to these drugs

and they are frequently not tolerated due to adverse side effects in their chronic use. Recently, biologic disease-modifying anti-rheumatic drugs like tocilizumab and abatacept, based on different molecular mechanisms, have been described as promising new treatment regimens for (j)LoS [63, 64]. This current pilot study explores and suggests the potential of novel, and non-invasive, research tools that appear to be easily applicable by using them in systemic treatment decisions in the daily clinical practice of (j)LoS. The separate quantitative outcome measurements obtained by HFUS (for dermal thickness), by durometer (for skin hardness) and by LASCA (for microcirculatory dynamics) make these tools particularly interesting as potential new disease biomarkers.

A limitation of this study is the relatively low number of patients, which can be explained by the high rarity of jLoS. Another limitation is the cross-sectional design since this was a first pilot study. Of note, LASCA is not very accessible to most hospitals, although HFUS and durometer are easier to acquire tools, making these interesting modalities for future longitudinal studies.

CONCLUSION

This case series suggests the supportive value of both microcirculatory and dermal assessment using novel non-invasive research tools, adopted from adult SSc, also for the identification, scoring and monitoring of (j)LoS lesions. Longitudinal studies should elaborate further the value of those non-invasive tools in this, potential severely invalidating, chronic skin disease.

ACKNOWLEDGEMENTS

We would like to thank Judith de Braaf for her administrative help during the examinations in Amsterdam UMC.

REFERENCES

1. Peterson LS, Nelson AM, Su WP, Mason T, O'Fallon WM, Gabriel SE. The epidemiology of morphea (localized scleroderma) in Olmsted County 1960-1993. *J Rheumatol*. 1997;24(1):73-80.
2. Murray KJ, Laxer RM. Scleroderma in children and adolescents. *Rheum Dis Clin North Am*. 2002;28(3):603-24.
3. Knobler R, Moinzadeh P, Hunzelmann N, Kreuter A, Cozzio A, Mouthon L, et al. European Dermatology Forum S1-guideline on the diagnosis and treatment of sclerosing diseases of the skin, Part 1: localized scleroderma, systemic sclerosis and overlap syndromes. *JEADV*. 2017;31(9):1401-24.
4. Torres JE, Sanchez JL. Histopathologic differentiation between localized and systemic scleroderma. *Am J Dermatopath*. 1998;20(3):242-5.
5. Kreuter A. Localized scleroderma. *Dermatol Ther*. 2012;25(2):135-47.
6. Kreuter A, Krieg T, Worm M, Wenzel J, Moinzadeh P, Kuhn A, et al. German guidelines for the diagnosis and therapy of localized scleroderma. *JDDG*. 2016;14(2):199-216.
7. Torok KS, Arkachaisri T. Methotrexate and corticosteroids in the treatment of localized scleroderma: a standardized prospective longitudinal single-center study. *J Rheumatol*. 2012;39(2):286-94.
8. Zulian F, Cuffaro G, Sperotto F. Scleroderma in children: an update. *Curr Opin Rheumatol*. 2013;25(5):643-50.
9. Zulian F, Athreya BH, Laxer R, Nelson AM, Feitosa de Oliveira SK, Punaro MG, et al. Juvenile localized scleroderma: clinical and epidemiological features in 750 children. An international study. *Rheumatology (Oxford, England)*. 2006;45(5):614-20.
10. Cutolo M, Soldano S, Smith V. Pathophysiology of systemic sclerosis: current understanding and new insights. *Expert Rev Clin Immunol*. 2019;15(7):753-64.
11. Bergamasco A, Hartmann N, Wallace L, Verpillat P. Epidemiology of systemic sclerosis and systemic sclerosis-associated interstitial lung disease. *Clin Epidemiol*. 2019;11:257-73.
12. Li SC. Scleroderma in Children and Adolescents: Localized Scleroderma and Systemic Sclerosis. *Pediatr Clin North Am*. 2018;65(4):757-81.
13. Herrick AL, Ennis H, Bhushan M, Silman AJ, Baildam EM. Incidence of childhood linear scleroderma and systemic sclerosis in the UK and Ireland. *Arthritis care & research*. 2010;62(2):213-8.
14. Zulian F. Scleroderma in children. *Best Pract Res Clin Rheumatol*. 2017;31(4):576-95.
15. Medsger TA, Jr., Bombardieri S, Czirjak L, Scorza R, Della Rossa A, Bencivelli W. Assessment of disease severity and prognosis. *Clinical and experimental rheumatology*. 2003;21(3 Suppl 29):S42-6.
16. Vanhaecke A, De Schepper S, Paolino S, Heeman L, Callens H, Gutermuth J, et al. Coexistence of systemic and localized scleroderma: a systematic literature review and observational cohort study. *Rheumatology (Oxford, England)*. 2020;59(10):2725-33.
17. Laxer RM, Zulian F. Localized scleroderma. *Curr Opin Rheumatol*. 2006;18(6):606-13.
18. Saxton-Daniels S, Jacobe HT. An evaluation of long-term outcomes in adults with pediatric-onset morphea. *Arch Dermatol*. 2010;146(9):1044-5.
19. Piram M, McCuaig CC, Saint-Cyr C, Marcoux D, Hatami A, Haddad E, et al. Short- and long-term outcome of linear morphoea in children. *Br J Dermatol*. 2013;169(6):1265-71.
20. Zulian F. Systemic sclerosis and localized scleroderma in childhood. *Rheum Dis Clin North Am*. 2008;34(1):239-55; ix.
21. Zulian F, Culpo R, Sperotto F, Anton J, Avcin T, Baildam EM, et al. Consensus-based recommendations for the management of juvenile localised scleroderma. *Ann Rheum Dis*. 2019;78(8):1019-24.
22. Zulian F, Tirelli F. Treatment in Juvenile Scleroderma. *Curr Rheumatol Rep*. 2020;22(8):45.

23. Li SC, O'Neil KM, Higgins GC. Morbidity and Disability in Juvenile Localized Scleroderma: The Case for Early Recognition and Systemic Immunosuppressive Treatment. *The Journal of pediatrics*. 2021.
24. Li SC, Zheng RJ. Overview of Juvenile localized scleroderma and its management. *World J Pediatr*. 2020;16(1):5-18.
25. Arkachaisri T, Pino S. Localized scleroderma severity index and global assessments: a pilot study of outcome instruments. *J Rheumatol*. 2008;35(4):650-7.
26. Arkachaisri T, Vilaiyuk S, Torok KS, Medsger TA, Jr. Development and initial validation of the localized scleroderma skin damage index and physician global assessment of disease damage: a proof-of-concept study. *Rheumatology (Oxford)*. 2010;49(2):373-81.
27. Arkachaisri T, Vilaiyuk S, Li S, O'Neil KM, Pope E, Higgins GC, et al. The localized scleroderma skin severity index and physician global assessment of disease activity: a work in progress toward development of localized scleroderma outcome measures. *The Journal of rheumatology*. 2009;36(12):2819-29.
28. Kaushik A, Mahajan R, De D, Handa S. Paediatric morphoea: a holistic review. Part 2: diagnosis, measures of disease activity, management and natural history. *Clin Exp Dermatol*. 2020;45(6):679-84.
29. Birdi N, Shore A, Rush P, Laxer RM, Silverman ED, Krafchik B. Childhood linear scleroderma: a possible role of thermography for evaluation. *J Rheumatol*. 1992;19(6):968-73.
30. Martini G, Murray KJ, Howell KJ, Harper J, Atherton D, Woo P, et al. Juvenile-onset localized scleroderma activity detection by infrared thermography. *Rheumatology (Oxford, England)*. 2002;41(10):1178-82.
31. Schanz S, Henes J, Ulmer A, Kotter I, Fierlbeck G, Claussen CD, et al. Response evaluation of musculoskeletal involvement in patients with deep morphea treated with methotrexate and prednisolone: a combined MRI and clinical approach. *AJR*. 2013;200(4):W376-82.
32. Horger M, Fierlbeck G, Kuemmerle-Deschner J, Tzaribachev N, Wehrmann M, Claussen CD, et al. MRI findings in deep and generalized morphea (localized scleroderma). *AJR*. 2008;190(1):32-9.
33. Weibel L, Howell KJ, Visentin MT, Rudiger A, Denton CP, Zulian F, et al. Laser Doppler flowmetry for assessing localized scleroderma in children. *Arthritis and rheumatism*. 2007;56(10):3489-95.
34. Hoffmann K, Gerbaulet U, el-Gammal S, Altmeyer P. 20-MHz B-mode ultrasound in monitoring the course of localized scleroderma (morphea). *ADV*. 1991;164:3-16.
35. Li SC, Liebling MS, Haines KA, Weiss JE, Prann A. Initial evaluation of an ultrasound measure for assessing the activity of skin lesions in juvenile localized scleroderma. *Arthritis care & research*. 2011;63(5):735-42.
36. Nezafati KA, Cayce RL, Susa JS, Setiawan AT, Tirkes T, Bendeck SE, et al. 14-MHz ultrasonography as an outcome measure in morphea (localized scleroderma). *Archives of dermatology*. 2011;147(9):1112-5.
37. Di Giovanni C, Puggina S, Meneghel A, Vittadello F, Martini G, Zulian F. Cone beam computed tomography for the assessment of linear scleroderma of the face. *Pediatr Rheumatol Onl J*. 2018;16(1):1.
38. Smith V, Vanhaecke A, Herrick AL, Distler O, Guerra MG, Denton CP, et al. Fast track algorithm: How to differentiate a "scleroderma pattern" from a "non-scleroderma pattern". *Autoimmun Rev*. 2019:102394.
39. Smith V, Pizzorni C, De Keyser F, Decuman S, Van Praet JT, Deschepper E, et al. Reliability of the qualitative and semiquantitative nailfold videocapillaroscopy assessment in a systemic sclerosis cohort: a two-centre study. *Ann Rheum Dis*. 2010;69(6):1092-6.

40. Cutolo M, Sulli A, Pizzorni C, Accardo S. Nailfold videocapillaroscopy assessment of microvascular damage in systemic sclerosis. *J Rheumatol.* 2000;27(1):155-60.
41. Mostmans Y, Richert B, Badot V, Nagant C, Smith V, Michel O. The importance of skin manifestations, serology and nailfold (video)capillaroscopy in morphea and systemic sclerosis: current understanding and new insights. *JEADV.* 2020.
42. Cutolo M, Smith V. State of the art on nailfold capillaroscopy: a reliable diagnostic tool and putative biomarker in rheumatology? *Rheumatology (Oxford, England).* 2013;52(11):1933-40.
43. Ingegnoli F, Ardoino I, Boracchi P, Cutolo M, co-authors E. Nailfold capillaroscopy in systemic sclerosis: data from the EULAR scleroderma trials and research (EUSTAR) database. *Microvascular research.* 2013;89:122-8.
44. Smith V, Riccieri V, Pizzorni C, Decuman S, Deschepper E, Bonroy C, et al. Nailfold capillaroscopy for prediction of novel future severe organ involvement in systemic sclerosis. *J Rheumatol.* 2013;40(12):2023-8.
45. Ingegnoli F, Boracchi P, Gualtierotti R, Smith V, Cutolo M, Foeldvari I, et al. A comparison between nailfold capillaroscopy patterns in adulthood in juvenile and adult-onset systemic sclerosis: A EUSTAR exploratory study. *Microvascular research.* 2015;102:19-24.
46. Ruaro B, Sulli A, Pizzorni C, Paolino S, Smith V, Cutolo M. Correlations between skin blood perfusion values and nailfold capillaroscopy scores in systemic sclerosis patients. *Microvascular research.* 2016;105:119-24.
47. Smith V, Herrick AL, Ingegnoli F, Damjanov N, De Angelis R, Denton CP, et al. Standardisation of nailfold capillaroscopy for the assessment of patients with Raynaud's phenomenon and systemic sclerosis. *Autoimmunity reviews.* 2020;19(3):102458.
48. Smith V, Vanhaecke A, Guerra MG, Melsens K, Vandecasteele E, Paolino S, et al. May capillaroscopy be a candidate tool in future algorithms for SSC-ILD: Are we looking for the holy grail? A systematic review. *Autoimmun Rev.* 2020;19(9):102619.
49. Ruaro B, Sulli A, Smith V, Paolino S, Pizzorni C, Cutolo M. Short-term follow-up of digital ulcers by laser speckle contrast analysis in systemic sclerosis patients. *Microvascular research.* 2015;101:82-5.
50. Ruaro B, Sulli A, Alessandri E, Pizzorni C, Ferrari G, Cutolo M. Laser speckle contrast analysis: a new method to evaluate peripheral blood perfusion in systemic sclerosis patients. *Ann Rheum Dis.* 2014;73(6):1181-5.
51. Cutolo M, Vanhaecke A, Ruaro B, Deschepper E, Ickinger C, Melsens K, et al. Is laser speckle contrast analysis (LASCA) the new kid on the block in systemic sclerosis? A systematic literature review and pilot study to evaluate reliability of LASCA to measure peripheral blood perfusion in scleroderma patients. *Autoimmun Rev.* 2018;17(8):775-80.
52. Ruaro B, Paolino S, Pizzorni C, Cutolo M, Sulli A. Assessment of treatment effects on digital ulcer and blood perfusion by laser speckle contrast analysis in a patient affected by systemic sclerosis. *Reumatismo.* 2017;69(3):134-6.
53. Lambrecht V, Cutolo M, De Keyser F, Decuman S, Ruaro B, Sulli A, et al. Reliability of the quantitative assessment of peripheral blood perfusion by laser speckle contrast analysis in a systemic sclerosis cohort. *Ann Rheum Dis.* 2016;75(6):1263-4.
54. van den Hoogen F, Khanna D, Fransen J, Johnson SR, Baron M, Tyndall A, et al. 2013 classification criteria for systemic sclerosis: an American college of rheumatology/European league against rheumatism collaborative initiative. *Ann Rheum Dis.* 2013;72(11):1747-55.

55. Sulli A, Ruaro B, Smith V, Paolino S, Pizzorni C, Pesce G, et al. Subclinical dermal involvement is detectable by high frequency ultrasound even in patients with limited cutaneous systemic sclerosis. *Arthritis research & therapy*. 2017;19(1):61.
56. Vanhaecke A, Cutolo M, Heeman L, Vilela V, Deschepper E, Melsens K, et al. HIGH FREQUENCY ULTRASONOGRAPHY: RELIABLE TOOL TO MEASURE SKIN FIBROSIS IN SSC? A systematic literature review and additional pilot study. *Rheumatology (Oxford, England)*. 2021.
57. Vanhaecke A, Verschuere S, Vilela V, Heeman L, Cutolo M, Smith V. Durometry in SSC: The hard facts. A systematic literature review and additional pilot study. *Rheumatology (Oxford, England)*. 2021;60(5):2099-108.
58. Poff S, Li SC, Kelsey CE, Foeldvari I, Torok KS. Durometry as an outcome measure in juvenile localized scleroderma. *Br J Dermatol*. 2016;174(1):228-30.
59. Smith V, Beeckman S, Herrick AL, Decuman S, Deschepper E, De Keyser F, et al. An EULAR study group pilot study on reliability of simple capillaroscopic definitions to describe capillary morphology in rheumatic diseases. *Rheumatology (Oxford, England)*. 2016;55(5):883-90.
60. Cutolo M, Melsens K, Herrick AL, Foeldvari I, Deschepper E, De Keyser F, et al. Reliability of simple capillaroscopic definitions in describing capillary morphology in rheumatic diseases. *Rheumatology (Oxford, England)*. 2018;57(4):757-9.
61. Sulli A, Ruaro B, Cutolo M. Evaluation of blood perfusion by laser speckle contrast analysis in different areas of hands and face in patients with systemic sclerosis. *Ann Rheum Dis*. 2014;73(11):2059-61.
62. Moore TL, Vij S, Murray AK, Bhushan M, Griffiths CE, Herrick AL. Pilot study of dual-wavelength (532 and 633 nm) laser Doppler imaging and infrared thermography of morphea. *Br J Dermatol*. 2009;160(4):864-7.
63. Foeldvari I. Update on the Systemic Treatment of Pediatric Localized Scleroderma. *Paediatr Drugs*. 2019;21(6):461-7.
64. Cutolo M, Soldano S, Montagna P, Trombetta AC, Contini P, Ruaro B, et al. Effects of CTLA4-Ig treatment on circulating fibrocytes and skin fibroblasts from the same systemic sclerosis patients: an in vitro assay. *Arthritis research & therapy*. 2018;20(1):157.

8

Consider the wrist: a retrospective study on pediatric connective tissue disease with MRI

Charlotte M. Nusman, J. Merlijn van den Berg, Amara Nassar-Sheikh Rashid,
Katerina Ntailiani, Apostolos Karantanas, Taco W. Kuijpers,
Mario Maas, **Dieneke Schonenberg-Meinema**

Published in Rheumatol Int. 2019 Dec;39(12):2095-2101

ABSTRACT

OBJECTIVE: The aim of this study is to describe the clinical characteristics and MRI findings of the wrist in a cohort of children suffering from connective tissue disease with musculoskeletal involvement.

METHODS: Ten patients with pediatric connective tissue disease [median age 14.7 years (IQR 12.7-16.6 years), 70% female] were identified from a large MRI database. Clinical findings during the disease course were retrospectively obtained from patient charts and findings at the time of MRI were prospectively registered in the MRI database. MRI wrist datasets were evaluated by three readers in consensus for synovitis, tenosynovitis, bone marrow changes, bone erosions and myositis.

RESULTS: Patients suffered from connective tissue disease with clinical overlap of subtypes systemic lupus erythematosus, Sjögren syndrome and dermatomyositis. Median onset of disease was at 12.3 years (IQR 7.8-14.8 years). Clinical arthritis activity was scored low (median visual analogue scale physician 19, IQR 7-31). Notwithstanding, extensive inflammatory abnormalities such as synovitis and tenosynovitis were found in the wrist of 7/10 patients. Osteochondral involvement was detected in 3/10 patients.

CONCLUSION: In a small cohort of children with connective tissue disease and musculoskeletal symptoms, severe inflammatory abnormalities of the involved wrist were present in the MRI, while clinical disease scores suggested mild disease activity. Therefore, clinicians should consider the wrist as vulnerable for joint damage and can add MRI as a helpful tool in the management of patients with pediatric connective tissue disease and musculoskeletal involvement.

INTRODUCTION

Connective tissue disease with potential musculoskeletal involvement in children comprises several disorders, such as systemic lupus erythematosus (SLE), mixed connective tissue disease (MCTD), systemic sclerosis (SSc) and juvenile polymyositis/dermatomyositis (PM/DM). Besides the variety of musculoskeletal symptoms, the above-mentioned disorders can manifest in many other organ systems [1]. For SLE, the recent international recommendation on treatment is based on the treat-to-target principle: since damage predicts subsequent damage and death, prevention of damage accrual should be a major therapeutic goal [2]. In general this also applies to the other mentioned connective tissue diseases.

In daily practice, the physician of children with connective tissue disease acquires information on the disease activity to make treatment decisions. Besides clinical and laboratory findings, imaging can be valuable in initiating and evaluating therapy and in the awareness and prevention of complications. MRI is the modality of choice for the imaging of inflammatory musculoskeletal changes due to its multiplanar capabilities and its capacity for visualizing both soft-tissue (e.g. synovium, tendons, muscle) and osteochondral structures [3].

In juvenile idiopathic arthritis, the wrist is known for its frequency of involvement, vulnerability to erosive damage and association to poor prognosis, and therefore warrants detailed examination in children with joint complaints [4-6].

Although several large cohort studies on the musculoskeletal symptoms of children with connective tissue disease have been published [7-10], no information on pediatric MRI of the joints in these diseases is available. For adult SLE [11, 12], MCTD [13] and SSc [14] a small number of studies has been published on the MRI appearance of the wrist. Based on these studies, wrist involvement in children with connective tissue disease is expected to show abnormalities indicating inflammatory arthritis as well as the common extra-articular involvement of skin, muscles and tendons visualized by MRI as soft tissue edema, indicating sclerosis, myositis and/or tenosynovitis [11-14].

The aim of this study is to describe the clinical characteristics and MRI findings of the wrist in a cohort of children suffering from connective tissue disease with musculoskeletal involvement.

METHODS

Pediatric patients with connective tissue disease that underwent MRI of the wrist in a tertiary pediatric rheumatology center between 2008 and 2018 were retrospectively included in this case series. MRI scans were performed at an open-bore 1.0T MRI scanner (Philips Panorama HFO) or at a 3.0T MRI scanner (Philips Achieva). No sedation was used. The clinically most affected wrist was imaged before and after intravenous (IV) contrast (Gadovist, Bayer Schering Pharma, Berlin, Germany, 1.0 mmol gadolinium/mL, dose 0.1 mmol/kg). Before IV contrast, coronal/axial T1- and T2-weighted images were acquired. After IV contrast, coronal and fat suppressed axial sequences were obtained. Approval by the regional ethics board and informed consent by patients and/or parents was waived for this study by the Medical Ethical Committee of the Amsterdam University Medical Center, location AMC (reference number W13_166, date August 6th 2013) as all examinations were in the context of regular patient care and data were completely anonymized.

Clinical parameters (clinical history, presentation/symptoms, laboratory values) were retrospectively acquired from the patient charts. MRI datasets were evaluated in a consensus reading session by a musculoskeletal radiologist (>25 years of experience), a pediatric rheumatologist (8 years of experience) and a resident in pediatrics with 5 years of experience in pediatric musculoskeletal radiology. The MRI's of the wrist were evaluated for presence and extent of synovial inflammation and tenosynovitis by means of validated scoring systems [15, 16]. The synovial inflammation score was based on a comprehensive evaluation of inflammatory features such as degree and extent of synovial enhancement and the presence of effusion [15]. Synovial inflammation was assessed on six locations: distal-radioulnar joint, radiocarpal joint, intercarpal joint, first carpometacarpal joint, carpometacarpal joints 2-5 and the pisotriquetral recess. A 0-2 (none, mild or moderate/severe inflammation) scale was used for every location, leading to a maximum total score of 12. Tenosynovitis was also scored on a 0-2 scale, with 0 reflecting no enhancement nor thickening of the synovial sheath, 1 reflecting enhancement and mild thickening of the synovial sheath, and 2 reflecting enhancement and moderate to significant thickening of the synovial sheath [16]. Tenosynovitis was evaluated over the entire trajectory of the tendon and separately in each of the six extensor compartments whereas the flexor tendons were scored as one group. Maximum total tenosynovitis score was 14. Other expected ancillary findings such as bone marrow edema, bone erosions and myositis were also evaluated and documented.

Descriptive statistics such as median, interquartile range (IQR) and frequencies were used to describe patient characteristics and clinical findings. SPSS software version 24

(IBM SPSS Statistics for Windows, Version 24.0. Armonk, NY: IBM Corp) was used for the evaluation of all data.

RESULTS

Clinical findings

Ten patients (7 female) with connective tissue disease and MRI of the wrist were identified. This concerns approximately 13% of the total population of pediatric patients with mixed connective tissue disease in the given time period (n=80). Median age at onset was 12.3 years with an IQR of 7.8 to 14.8 years. Median time from disease onset to MRI was 1.4 years with an IQR of 0.4 to 6.6 years. At time of MRI, median age was 14.7 years (IQR 12.7-16.6). Clinical arthritis activity as assessed on a Likert scale (0-5) was mild (score 3) in 6 of the patients (67%); none of the patients were considered to have 'severe' clinical arthritis activity on this Likert scale. Furthermore, clinical arthritis activity as assessed on a visual analogue scale (0-100) showed a median of 19 (IQR 7-31). At time of MRI, 4 patients used no medication and the other 6 patients used different types of medication, consisting of non-steroidal anti-inflammatory drugs, methotrexate, prednisone, Plaquenil and Cellcept. Clinical symptoms and treatment strategies during the disease course are shown in Table 1.

Imaging findings

MRI of the wrist showed extensive inflammatory findings in 7/10 patients. Severe synovial inflammation was seen in 7/10 patients, with total scores ranging between 7 and 12. Severe tenosynovitis as reflected by a score of 10 and higher was found in 6/10 patients. In those patients with severe tenosynovitis, flexor tendons were involved in 5/6 patients. An example is given in Figure 1.

The three patients with minimal inflammatory findings had total synovial inflammation scores of 2- 4 and total tenosynovitis scores of 0-3 respectively (which is less than 33% of the maximum scores in both items). Myositis, reflected by high signal intensity on the axial T2-weighted sequence as well as on the fat-suppressed T1-weighted sequence after administration of IV contrast, was found in 3/10 patients (33%). When present, myositis was always found in the volar thumb muscles (Figure 2)).

Furthermore, 1 patient showed extensive infiltration of the subcutaneous fat, clinically correlating active sclerosis of the skin (Figure 3). Osteochondral abnormalities, such as bone marrow edema and bone erosions, were found in 3/10 patients (Figure 4).

Table 1. Characteristics of 10 pediatric patients with connective tissue disease

	Ethnicity	Auto-antibody profile	Other symptoms (besides arthritis)	Treatment ever used
1	African / Afro-Caribbean	ANA, anti-RNP, anti-centromere, anti-Scl-70, anti-SS-A, anti-ds-DNA	Myositis, Raynaud, sclerodactyly, digital ulcerations, interstitial pulmonary involvement, nephritis	Hydroxychloroquine, MTX, MMF, prednisolone, RTX, etanercept, allogenic SCT
2	African/ Afro-Caribbean	ANA, anti-RNP, anti-Sm, anti-Ro52, anti-ds-DNA, RF	Myositis, Raynaud, pleuritis, pericarditis, haemolytic anaemia, nephritis, aphthous ulcers	Hydroxychloroquine, MTX, MMF, RTX, prednisolone, cyclophosphamide, belimumab
3	Asian	ANA, anti-RNP, anti-Sm, anti-Ro52, anti-ds-DNA, RF	Lymphadenopathy, haemolytic anaemia, parotitis, nephritis	Hydroxychloroquine, MTX, prednisolone, RTX
4	Caucasian	ANA, anti-SS-A, anti-SS-B, anti-Ro52, RF	Sicca, parotitis	Hydroxychloroquine
5	Caucasian	ANA, anti-RNP, anti-ds-DNA	Autoimmune mediated encephalopathy, leukopenia, aphthous ulcers	Hydroxychloroquine, MTX, MMF, RTX, prednisolone, IVIG, plasmapheresis
6	African / Afro-Caribbean	ANA, anti-RNP, anti-Sm, anti-ds-DNA	Leukopenia, thrombocytopenia, haemolytic anaemia	Hydroxychloroquine, MTX
7	North African / Middle east	ANA, anti-centromere A / anti-PM-Scl100	Dermatomyositis	MTX, IVIG, prednisolone, RTX
8	Caucasian	ANA, anti-RNP	Raynaud	MTX
9	Caucasian	ANA, anti-RNP	Raynaud	Hydroxychloroquine, MTX
10	Caucasian	ANA	Cutaneous LE, leukopenia, aphthous ulcers	MTX, plaquenil

anti-ds-DNA= anti-double stranded DNA antibodies, anti-RNP= anti-RiboNuclear Protein, anti-Sm= anti-Smith antibodies, RF = IgM rheumatoid factor; MTX=methotrexate, MMF=mycophenolate mofetil (cellcept), RTX=rituximab, IVIG=intravenous immunoglobulins, SCT = stem cell transplantation

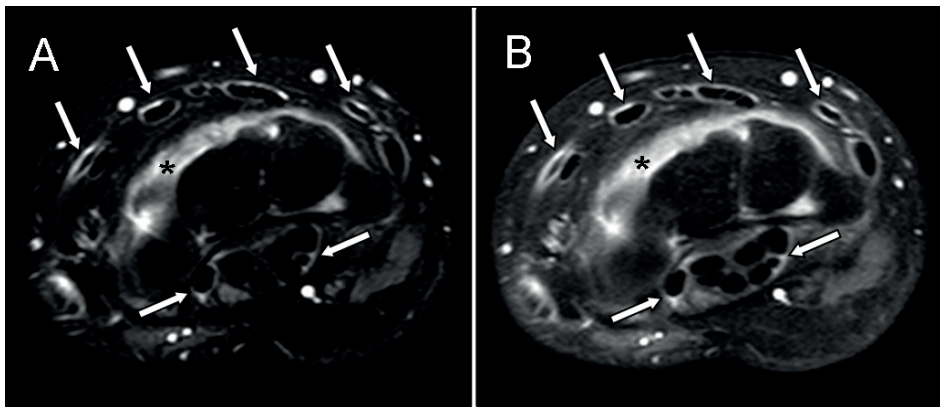


Figure 1. Multifocal inflammatory findings in the wrist from a 17 year old girl with mixed connective tissue disease.

The midcarpal region shows synovial inflammation (asterisk) and tenosynovitis (arrows) in the flexor and extensor tendons on axial T2-weighted images (A) and T1-weighted images with fat saturation after IV contrast (B).

Correlation clinical and imaging findings

In all patients, clinical arthritis activity scores were relatively low (i.e. VAS <50 (0-100) or Likert ≤ 4 (0-5)). On the other hand, in the majority of patients the total MRI scores for synovitis and tenosynovitis were clearly increased (i.e. total synovitis score >6 (0-12) and total tenosynovitis score >7 (0-14)). No correlation statistics were performed because of the small cohort.

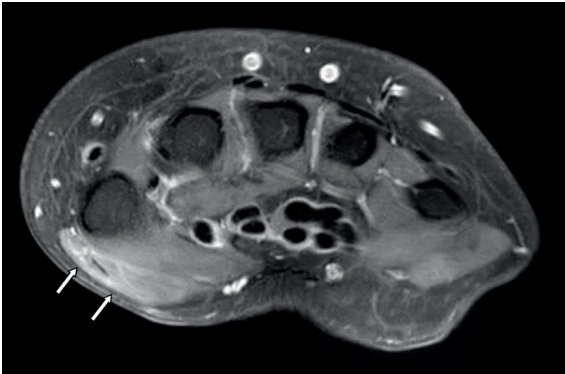


Figure 2. Osteochondral abnormalities in a 13 year old girl with mixed connective tissue disease. Coronal T1- (A) and T2-weighted images (B) with numerous foci of low (A) and high (B) signal intensity within the bone marrow, in keeping with bone marrow edema and/or bone erosions indicating extensive osteochondral damage.

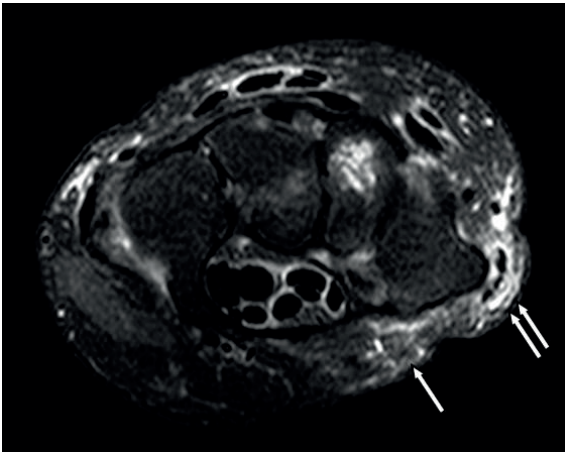


Figure 3. Soft tissue edema in mixed connective tissue disease in a 13 year old girl. Axial T2-weighted image (A) visualizing infiltration of the subcutaneous fat (high signal intensity), possibly indicative of active sclerosis. Also bone marrow edema in the trapezoid and tenosynovitis (enhancement on T1 after contrast administration, B) in both the flexor- and extensor-compartments are present

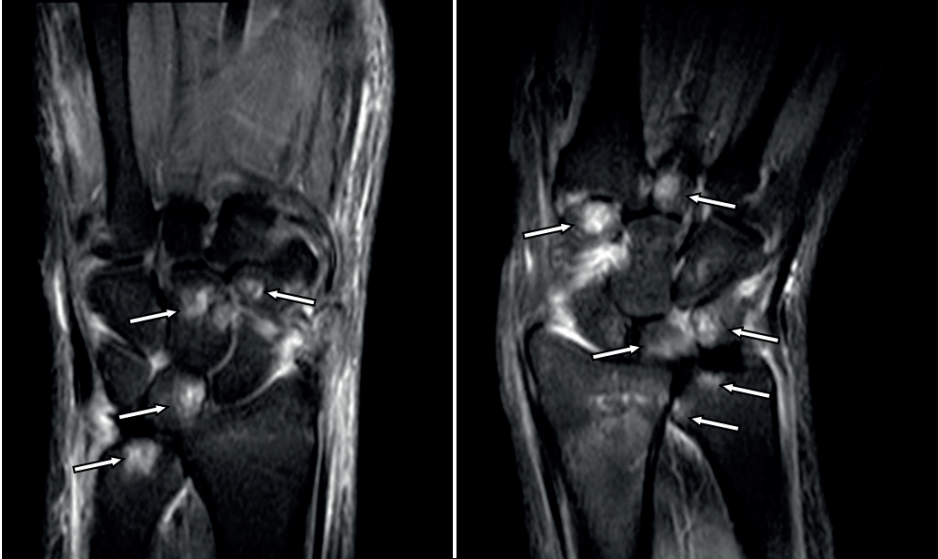


Figure 4. Myositis in an 8 year old boy with juvenile dermatomyositis. Axial fat-saturated T1-weighted MR image of the wrist after administration of IV contrast, visualizing high signal intensity (enhancement) in the thenar muscles (arrows), indicative of local myositis.

DISCUSSION

To the best of our knowledge, this cohort of pediatric patients with connective tissue disease with musculoskeletal involvement is unique in describing the imaging findings of the wrist. Extensive inflammatory and osteochondral abnormalities on MRI of the wrist were observed in these patients and included involvement of flexor tendons, myositis and infiltration of subcutaneous fat indicative of active sclerosis. Interestingly, the perceived/scored clinical arthritis activity of the wrist by the patients and their clinicians (VAS) was strikingly low, compared to the extent of the abnormalities detected with MRI. Based on these findings, the wrist deserves thorough evaluation in daily practice as a possible vulnerable joint in children with connective tissue disease. The pattern of inflammation and damage on the MRI of the wrist as observed in patients with connective tissue disease is much more extensive compared to other arthritic diseases of childhood, such as juvenile idiopathic arthritis or reactive arthritis [4]. Although a direct comparison has not been performed, findings of pediatric connective tissue disease upon imaging of the wrist show higher synovial inflammation scores, the presence of extensive tenosynovitis and preference for the flexor tendons compared to JIA patients [4]. The findings including the extensive tenosynovitis and extracapsular inflammation have also been described upon MRI of the wrist in adults with MCTD [13]. Additionally, high signal intensity on T2-weighted images in muscle and subcutaneous tissue, sugges-

tive for myositis and subcutaneous edema, have been described before in a recent study on pediatric mixed connective tissue disease [17].

The presence of bone erosions and bone marrow edema is commonly observed in adults with systemic autoimmune disease [11, 12]. However, Mosca et al. showed that bony depressions were equally frequent in the healthy control group and hence cast doubt on the clinical significance of these findings [11]. MRI studies in healthy children have shown that bony depressions in the wrist are very commonly present and may mimic erosive pathology [18, 19]. Nevertheless, erosions as found in our cohort can be considered 'pathologic', due to accompanying bone marrow edema and/or synovial inflammation.

The correlation between clinical and imaging findings could not be assessed due to the small cohort size. In general though, the clinical arthritis activity was scored low compared to the high MRI inflammation scores. Based on this finding, one could cautiously state that clinical arthritis activity underestimates the inflammation as assessed with MRI in pediatric connective tissue disease – a finding that needs confirmation in larger cohorts.

A limitation of the current study was the relatively low number of patients, resulting from the fact that connective tissue disease in children is a rare entity and MRI is not routinely performed in these affected children. Furthermore, the variety of different diagnoses within the connective disease decreased to the homogeneity of the cohort. On the other hand, homogeneity was achieved with the involvement and imaging of one specific joint, i.e. the wrist.

CONCLUSION

Children suffering from connective tissue disease with musculoskeletal symptoms show severe abnormalities on MRI of the involved wrist. Frequent findings include extensive (teno)synovitis, osteochondral damage, myositis and infiltration of the subcutaneous fat. Clinical arthritis activity was strikingly low compared to the extent of the MRI abnormalities. Considering the wrist as vulnerable for erosive disease, integration of MRI in the management of patients with pediatric connective tissue disease and musculoskeletal involvement deserves consideration to better interpret disease activity and subsequently damage.

REFERENCES

1. Berard RA, Laxer RM. Pediatric Mixed Connective Tissue Disease. *Curr Rheumatol Rep.* 2016;18(5):28.
2. van Vollenhoven RF, Mosca M, Bertsias G, Isenberg D, Kuhn A, Lerstrom K, et al. Treat-to-target in systemic lupus erythematosus: recommendations from an international task force. *Ann Rheum Dis.* 2014;73(6):958-67.
3. Cimmino MA, Bountis C, Silvestri E, Garlaschi G, Accardo S. An appraisal of magnetic resonance imaging of the wrist in rheumatoid arthritis. *SeminArthritis Rheum.* 2000;30(3):180-95.
4. Nusman CM, Hemke R, Schonenberg D, Dolman KM, Van Rossum MA, Van den Berg JM, et al. Distribution pattern of MRI abnormalities within the knee and wrist of juvenile idiopathic arthritis patients: signature of disease activity. *AJR Am J Roentgenol.* 2014;202(5):W439-46.
5. Al-Matar MJ, Petty RE, Tucker LB, Malleson PN, Schroeder ML, Cabral DA. The early pattern of joint involvement predicts disease progression in children with oligoarticular (pauciarticular) juvenile rheumatoid arthritis. *Arthritis Rheum.* 2002;46(10):2708-15.
6. Hemke R, Nusman CM, van der Heijde DM, Doria AS, Kuijpers TW, Maas M, et al. Frequency of joint involvement in juvenile idiopathic arthritis during a 5-year follow-up of newly diagnosed patients: implications for MR imaging as outcome measure. *Rheumatol Int.* 2014.
7. Sakamoto AP, Silva CA, Ferriani MP, Pereira RM, Bonfa E, Saad-Magalhaes C, et al. Characterization of chronic arthritis in a multicenter study of 852 childhood-onset systemic lupus erythematosus patients. *Rheumatol Int.* 2016;36(12):1641-8.
8. Hetlevik SO, Flato B, Rygg M, Nordal EB, Brunborg C, Hetland H, et al. Long-term outcome in juvenile-onset mixed connective tissue disease: a nationwide Norwegian study. *Ann Rheum Dis.* 2017;76(1):159-65.
9. Martini G, Foeldvari I, Russo R, Cuttica R, Eberhard A, Ravelli A, et al. Systemic sclerosis in childhood: clinical and immunologic features of 153 patients in an international database. *Arthritis and rheumatism.* 2006;54(12):3971-8.
10. Ramanan AV, Feldman BM. Clinical features and outcomes of juvenile dermatomyositis and other childhood onset myositis syndromes. *Rheum Dis Clin North Am.* 2002;28(4):833-57.
11. Mosca M, Tani C, Carli L, Vagnani S, Possemato N, Delle Sedie A, et al. The role of imaging in the evaluation of joint involvement in 102 consecutive patients with systemic lupus erythematosus. *Autoimmun Rev.* 2015;14(1):10-5.
12. Boutry N, Hachulla E, Flipo RM, Cortet B, Cotten A. MR imaging findings in hands in early rheumatoid arthritis: comparison with those in systemic lupus erythematosus and primary Sjogren syndrome. *Radiology.* 2005;236(2):593-600.
13. Cimmino MA, Iozzelli A, Garlaschi G, Silvestri E, Montecucco C. Magnetic resonance imaging of the hand in mixed connective tissue disease. *AnnRheumDis.* 2003;62(4):380-1.
14. Frerix M, Kroger K, Szalay G, Muller-Ladner U, Tarner IH. Is osteonecrosis of the lunate bone an underestimated feature of systemic sclerosis? A case series of nine patients and review of literature. *Semin Arthritis Rheum.* 2016;45(4):446-54.
15. Damasio MB, Malattia C, Tanturri de Horatio L, Mattiuz C, Pistorio A, Bracaglia C, et al. MRI of the wrist in juvenile idiopathic arthritis: proposal of a paediatric synovitis score by a consensus of an international working group. Results of a multicentre reliability study. *Pediatr Radiol.* 2012;42(9):1047-55.

16. Lambot K, Boavida P, Damasio MB, Tanturri de HL, Desgranges M, Malattia C, et al. MRI assessment of tenosynovitis in children with juvenile idiopathic arthritis: inter- and intra-observer variability. *Pediatr Radiol*. 2013;43(7):796-802.
17. Gorkem SB, Doria AS, Tse S. Imaging findings of mixed connective tissue disease in children and adolescents: a case series. *Jpn J Radiol*. 2019;37(5):371-9.
18. Müller LS, Avenarius D, Damasio B, Eldevik OP, Malattia C, Lambot-Juhan K, et al. The paediatric wrist revisited: redefining MR findings in healthy children. *Ann Rheum Dis*. 2011;70(4):605-10.
19. Avenarius DF, Ording Muller LS, Rosendahl K. Erosion or normal variant? 4-year MRI follow-up of the wrists in healthy children. *Pediatr Radiol*. 2016;46(3):322-30.



Part III

A large, abstract graphic of red splatters and dots, resembling a bloodstain or ink blot, originates from the left side of the page and extends towards the right, partially overlapping the text. The splatters are of various sizes and densities, creating a textured, organic shape.

Discussion, summary and appendix

A small, solid red square is located in the bottom right corner of the page.

9

Discussion

NAILFOLD CAPILLAROSCOPY IN cSLE

One of the main objectives of this thesis was describing the abnormalities of the nailfold capillaries in cSLE patients and correlate these to patient- and disease characteristics. Nailfold capillaroscopy is an attractive imaging tool for daily clinical practice. It is non-invasive and gives direct image results during the nailfold investigation of the patient in (outpatient) clinic. Most devices for capillaroscopy are quite easy to use in daily care with self-teaching practice, but for interpretation of images some training is needed (1). These trainings are provided at (international) conferences in our field. Reliability of new definitions for abnormal capillary morphology have been tested in a pilot study of participants in a one-day EULAR capillaroscopy course. The reliability for abnormal capillary morphology was excellent, also in attendees with low experience in this field, proving capillaroscopy to be suitable for direct applicability in practice after a short training course (2).

In adults with SLE it had already been observed that tortuous capillaries, abnormal capillary morphology and capillary hemorrhages were significantly more prevalent in SLE patients as compared to healthy controls (3). After showing, in **chapter 2**, that the literature on capillaroscopy in cSLE was scarce, we have examined our own cohort of cSLE patients cross-sectionally. In **chapter 3** is shown that the nailfold capillaries in these patients have many abnormalities when compared to matched healthy controls. These capillary abnormalities were also seen in cSLE patients *without* Raynaud's phenomenon or acro-cyanotic complaints. In the above mentioned review on adults with SLE, capillary abnormalities were also seen with (2 studies) but also without (7 studies) a correlation with Raynaud's phenomenon (3). This is an interesting finding because capillary abnormalities and Raynaud's phenomenon in SSc are always closely linked. It raises the question if the pathophysiology of these capillary abnormalities is quite different in these two systemic autoimmune diseases. It is already known that both diseases have long-term vascular complications, although quite different in type: SLE patients suffer from cerebral stroke, myocardial ischemia or thrombosis while pulmonary hypertension is a well-known complication in SSc.

We have been the first to describe the nailfold capillary abnormalities in cSLE in so much detail. The specific capillary abnormalities that we observed in cSLE patients were giant capillaries (apical diameter > 50 micrometer), abnormal capillary morphology and capillary hemorrhages. Capillary hemorrhages were seen in up to 88% of cSLE patients and the number of capillary hemorrhages was significantly correlated with disease activity at diagnosis as well as with disease activity at the moment of capillaroscopy (by SLEDAI). The high percentage of cSLE patients that shows capillary hemorrhages is

striking, as well as the high mean number of capillary hemorrhages per patient (at least one hemorrhage per mm, with a standard examination of eight mm in eight fingers per patient).

In SSc, the capillary hemorrhages are well known as part of the so-called ‘active stage’ capillary scleroderma pattern (4, 5). It has also been shown in SSc that the severity of this capillary scleroderma pattern (defined by early-, active- and late stage) is associated with the severity of organ involvement (6, 7). SSc is known to be a vascular disease with dysregulation of the endothelium (8). The endothelium in SLE is also dysregulated by intravascular pro-inflammatory cytokines due to auto-inflammation. This leads to oxidative stress, endothelial apoptosis and coagulopathy (9). As in SSc, the images from (video)capillaroscopy are visualizing (micro)vasculopathy in SLE. In **chapter 4**, some of the abnormal capillary findings in cSLE were confirmed in a multicenter cohort, although the number of investigated cSLE patients of this cohort was lower than in **chapter 3** and there was partial overlap of participating patients from **chapter 3 and 4**. In the international cohort of cSLE in **chapter 4**, it was confirmed that a large majority of 80% of cSLE patients showed capillary hemorrhages. cSLE patients also showed a significant lower capillary density when compared to healthy controls, although skin color was not taken into account. It has been shown that skin color can be of influence in capillary density (10, 11).

Chapter 3 and 4 show unique nailfold capillaroscopy data in cSLE because they describe, for the first time, observations of the abnormal capillary morphology by the new definition of EULAR study group on Microcirculation in Rheumatic Diseases from 2018 (2). In **chapter 3** we also described the *combination* of abnormal capillary morphology (shapes) and capillary hemorrhages, which were often seen in the same capillaries in our cSLE patient cohort. The significance of this finding was indicated by the percentage of observed patients ($n=32/41$, 78%) and number of fingers (78% in ≥ 3 fingers) affected, in which these abnormalities were combinable observed. The qualitative description of this finding was called ‘microangiopathy’ by us in **chapter 3**. In **chapter 4**, 75% of cSLE patients also showed the comparable ‘non-specific pattern’ versus 35% in healthy age-matched controls. In this international study, cSLE patients were the only disease subgroup with this finding of a ‘non-specific’ pattern. More recent studies show very similar results of SLE patients (children and adults) with abnormal ‘non-specific’ capillary patterns (12-14).

Another striking finding in **chapter 3** was the description of a new subtype of capillary hemorrhage, which we called ‘pericapillary extravasation’. To explore the reproducibility of this observation, inter- and intra-observer agreement had to be determined. As a first

step, we have already shown that there is a good agreement to detect different subtypes of capillary hemorrhages (15). At this moment, the meaning of this new ‘pericapillary extravasation’ is not yet exactly clear. We hypothesize that it might be capillary leakage that can be visually detected by the good quality imaging from videocapillaroscopy with 200x magnification. SLE is a vascular disease with a high degree of vascular inflammation and endothelial damage. This endothelial damage may be reflected by these different subtypes of nailfold capillary hemorrhages that we observe in (c)SLE.

As mentioned in the introduction of this thesis, SLE patients are known to be at risk for cardiovascular disease which can lead to a much lower life expectancy. In the past, the cardiovascular health of SLE patients has been studied by measuring carotid intima-media thickness (CIMT) and presence of carotid plaques in SLE patients, which were significantly different compared to controls (16). A recent study shows that more sensitive imaging measurements such as ‘total plaque area’ and increased ‘echogenicity of the plaque’ is significantly higher in patients taking corticosteroids (CS) (17). It is known that risk factors with significant influence on CIMT are traditional cardiovascular risk factors (age, HDL and triglyceride) and lupus related risk factors (disease duration, ESR, SLEDAI and use of CS) (16, 17). Endothelial dysregulation seems to play an important factor in the risk for this premature atherosclerosis (9). In a systematic review and meta-analysis we found that several dysregulated EC markers seem associated with disease activity scores but, more interestingly, other EC markers show dysregulated levels without a clear association to disease activity scores (*Bergkamp et al., under review*). This finding suggests dysregulation of the endothelium during low disease activity states or maybe even in disease remission. The identified EC markers that we have found in this systematic literature review are involved in many endothelial functions: EC activation, EC apoptosis, disturbed angiogenesis, defective vascular tone control, immune dysregulation and coagulopathy. Hypothetically this could mean that, even in a low disease activity state, the endothelium of SLE patients might still be at risk for premature atherosclerosis and EC markers might reflect this pre-atherosclerotic state before CIMT measurements can detect signs of a plaque.

Future needs and perspectives in capillaroscopy and endothelial cell markers

One of the steps to be studied in this field is the sensitivity of different devices for nailfold capillaroscopy and the number of fingers that needs to be investigated for optimal results. Sensitivity should especially be investigated in various devices to see if they also detect (smaller types of) capillary hemorrhages. It is important to know if these hemorrhages are also detectable with a handheld dermatoscope, or if they can be missed with lower magnifications than those in the videocapillaroscopy (with 200-300x magnifica-

tion). It would also be informative if less, and if so which, fingers can be investigated with the same sensitivity and quality of observations. If less fingers can be examined with good sensitivity of abnormal capillaries, this would be less time-consuming. For instance, capillary examination performed by dermatoscope in only 1-2 fingers takes only a few minutes. Hypothetically, digits 3 and 4 of the non-dominant hand, with less risk for traumatic abnormalities, might possibly be enough to detect capillary changes in systemic autoimmune diseases.

In chapter 4, a lower capillary density was found in cSLE patients. In future studies a correlation of lower capillary density in darker skin should be further examined and confirmed because this observation has been described in the past (10). This is important to know because people with different ethnic backgrounds have different incidence rates for SLE, as was discussed in the introduction of this thesis. In black patients a lower density might not be a pathological finding and this might be of importance in the interpretation of images from capillaroscopy in patients with different ethnic backgrounds.

The repeating observations of microangiopathy and non-specific pattern in cSLE lead to the hypothesis that a capillary ‘lupus pattern’ might exist as an entity. More (multi-center) studies with a detailed quantitative description of this abnormal capillary pattern in SLE (children and adults) should investigate this hypothesis further, according to international consensus definitions from the EULAR Study Group on Microcirculation in Rheumatic Diseases. Future longitudinal studies also have to show if the number of capillary hemorrhages/mm reflect treatment efficacy and microvascular health on the long-term. By combining EC marker analyses with capillaroscopy data it can also be investigated if the nailfold capillaries are a mirror of what happens to the protective internal lining of the blood vessel itself.

The reviewed studies in our systematic literature review on EC markers are mainly cross-sectional studies in adult SLE populations. To further unravel the pathophysiology of (cardio)vascular complications in SLE we are in progress to study most of these EC markers in our longitudinal cohort, with samples from pre-treatment and follow-up over time, during high and low disease activity and/or remission. Because the pediatric SLE population, in contrast to adults, does not have many other confounding risk factors for atherosclerosis, our study in cSLE patients will show results that are mostly determined by the sole effect of vascular inflammation in SLE, especially because pre-treatment samples will be studied. Next steps would be to compare these results with measurements from blood samples from adult patients with a childhood-onset and a long disease duration, again preferably obtained during high and low disease activity states. In the future we have to focus on prevention of (subclinical) atherosclerosis in the

subgroup of SLE patients at risk. The risk factors involved have to be defined in further detail for integration into standard clinical screening protocols regarding early detecting of premature atherosclerosis in SLE.

SLE-RELATED DAMAGE: (NEW) PREDICTIVE RISK FACTORS

In **chapter 5** was shown that a capillary scleroderma pattern has prognostic meaning in cSLE patients, as it predicts a higher risk for disease damage. cSLE patients with a capillary scleroderma pattern were at significant more risk to develop SLE-specific organ damage, which is irreversible. This finding was epochal because it was always thought that SLE patients with a capillary scleroderma pattern were patients at risk to develop overlapping symptoms with SSc. Nevertheless, in our cohort, these cSLE patients did not show any symptoms of overlap disease and the type of damage was purely lupus-related. Although the range of follow-up in these patients was up to 16 years after disease onset, our cohort consisted of relatively small patient numbers. More long-term follow-up studies with larger patient numbers should evaluate the risk for change into overlap disease of these (c)SLE patients with a capillary scleroderma pattern. If those patients are *not* at risk for overlap disease, this would mean that a capillary scleroderma pattern might not be a specific finding for SSc only.

The higher risk for (early) damage in cSLE versus aSLE is mostly reflected in ocular (cataract, retinal change or optic atrophy) and musculoskeletal damage (muscle atrophy, erosive arthritis, osteoporosis with fracture, avascular necrosis, osteomyelitis or ruptured tendon) (18). These data show that damage does not only result from inflammation but also from high and/or chronic (cumulative) CS treatment in SLE. Looking at the type of disease damage (by validated SDI scoring) in our cohort in **chapter 5**, this damage was partly *corticosteroid*-related (avascular skeletal necrosis and growth failure, n=3/53 patients) and partly *lupus*-related damage (skin scarring, end-stage renal disease, cerebrovascular accident, epilepsy, loss of digits, cardiac valve disease and cognitive impairment/major psychosis, n=10/53 patients). Two patients (n=2/10) showed both types of damage, lupus- and CS-related. Disease damage occurred already within five years after diagnosis and such early onset of damage has been shown in other studies as well (19, 20). One of these long-term studies showed that more than half of cSLE patients have damage after 10 years, with more increasing numbers in the years thereafter (19). Especially the presence of nephritis is a risk factor for mortality in young patients (21). This latter cohort consisted of 93% white patients. This might give an underestimation of damage because in general, black (female) patients have a higher incidence of SLE and are also at risk for more severe disease activity and -damage, as described in the in-

roduction. Looking at very recent (unpublished) data from an international multicenter project (n=1096 cSLE patients) from the Pediatric Rheumatology European Society (PReS) Lupus working party, one-third of cSLE patients already have damage after 6 years of disease duration. Mean age at diagnosis in this study was 12 years, meaning that many of these patients already suffer from damage in the adolescent phase of their life. More importantly, damage was more likely with an early disease onset (< 10 years of age) but not correlated with SLEDAI at diagnosis or SLEDAI at last visit (*PReS Lupus Working party unpublished data*). We showed this same finding in our data from **chapter 4**. So although we currently are doing better in terms of lowering mortality rates in SLE, this means that we should still focus more on early detection of the diagnosis (in order to prevent lupus-specific damage) and on better/earlier use of CS- sparing regimens (to prevent CS-related damage).

In the new treat-to-target (T2T) approach the lowest effective dose of CS is now state-of-the-art and explicitly recommended in the treatment of (c)SLE (22)). T2T strategy has recently been proposed for adults with SLE but is now also of interest for treatment of cSLE (23, 24). This strategy defines specific targets that should be reached and aimed for while treating patients with SLE. One of the suggested treatment targets in SLE is the Lupus Low Disease Activity State (LLDAS). LLDAS is defined as low disease activity score (SLEDAI ≤ 4) with zero scores for renal, central nervous system, serositis, vasculitis and constitutional components, no increase in any SLEDAI component since the previous visit, physician global assessment ≤ 1 , and prednisone dose ≤ 7.5 mg/day. LLDAS is a target which has been shown to significantly lower the risk for disease damage in adults with SLE (25-27). Recent studies show that LLDAS is also an achievable target in cSLE, even in the first year after diagnosis (28-30). This is a very important finding because disease damage in cSLE often occurs in the first years after diagnosis, and is often CS-related (20, 31). Another treatment target in T2T is disease remission for which different definitions are still being fine-tuned, for example remission “on” or “off treatment” (32). At this moment, an international cSLE T2T Lupus working party is establishing consensus definitions for different remission states, specifically for the cSLE patients.

Future needs and perspectives in SLE-related damage: prevention

Future studies have to show if T2T, by new treatment approaches, results in lower damage scores in cSLE on the long-term. The focus needs to be on lowering CS, but at the same time CS-sparing regimens need to be started early, preferably *as early as possible* after diagnosis. This means a high focus on the compliance of other (daily) oral medication (hydroxychloroquine, mycophenolate mofetil (MMF) and azathioprine (AZA)). Early (or earlier) starting of biologicals such as rituximab and belimumab (if available) should be considered when compliance of daily pills is a problem for the individual patient.

By defining subgroups with low and higher risk for damage in (c)SLE patients we can intervene early with different treatment protocols for different subgroups. A suggestion for stratifying SLE patients, by risk for cumulative CS use, is described in the next section on transcriptomics and personalized treatment.

The patients with high risk for long-term use of corticosteroids might profit from an early start with specific B cell depleting therapy early after diagnosis. A good example from current daily clinical practice is the difficult choice which patients should be treated with rituximab (RTX) and which patients should be treated with belimumab as first-choice of anti-B cell therapy. Both biologicals have shown efficacy in lowering disease activity and are therefore CS-sparing. Some RTX trials showed moderate efficacy, resulting in the opinion that RTX might not be the first choice in lupus nephritis, although this interpretation might not be correct (33). New insights in development of anti-drug-antibodies (ADA) against RTX might explain why some SLE patients fail in B cell depletion after RTX and this might also be the reason for different efficacy rates between SLE patients (34). It is important to find risk factors in patients that are prone to develop RTX-ADA in order to make the best treatment choices, especially because new treatment strategies also involve a combination of those two treatments with induction of RTX and maintenance therapy with belimumab. If RTX-ADA or anaphylactic reactions during infusion do not occur, treatment with RTX is an elegant therapy with 2 infusions every six months.

For SLE, CS are almost always needed for some time after diagnosis but it is important to pro-actively decrease the cumulative CS dose. For this reason, cumulative CS dose should be studied more often as co-variable or as outcome in longitudinal cohort studies in (c)SLE patients. Besides that, targeting anti-B cell therapy should be investigated as “top-down” strategy versus the currently used and slower “step-up” strategies, especially in high risk patients that are at risk for a high amount of cumulative CS use, as suggested in figure 1.

TRANSCRIPTOMICS IN cSLE: THE ROAD TO PERSONALIZED TREATMENT?

The heterogeneity of SLE patients makes it difficult to design standard treatment protocols for SLE. In the past decade, SHARE treatment recommendations were published to give guidance to (specialized) physicians in treatment choices for cSLE patients (35, 36). In clinical practice, as measured by SLEDAI and SDI, black SLE patients have more severe disease than patients with other ethnic backgrounds (37). This higher severity of disease was reflected by analysis of a large cohort of cSLE patients by increased plasmablast sig-

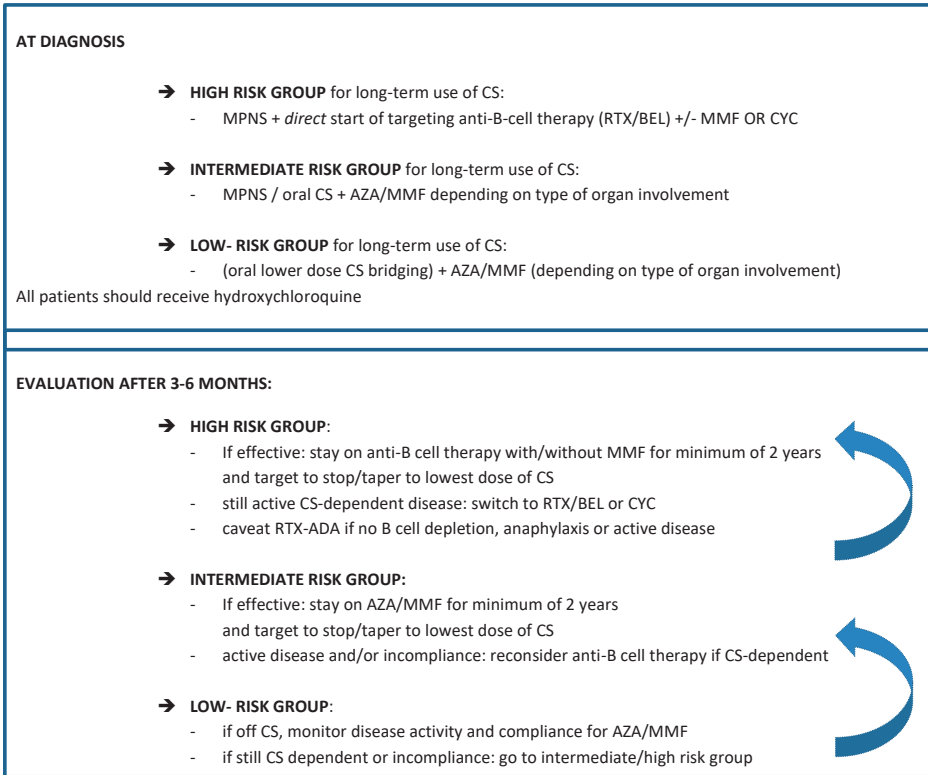


Figure 1. Example of personalized treatment regimens for SLE subgroups in the first year after diagnosis, stratified by risk for cumulative CS-usage
CS= corticosteroid(s), MPNS= methylprednisolon pulse, RTX= rituximab, BEL=belimumab, AZA= azathioprine, MMF= mycophenolate mofetil, ADA= anti-drug antibodies

natures in African-American patients, which correlated with higher SLEDAI (38). Those plasmablasts will form plasma cells and are the origin of the development of pathological auto-antibodies leading to systemic auto-inflammation. The ethnic background also influences IFN signatures (39). Recently, it was observed that SLE-patients can show different interferon signatures, differentiated in 'IFN high' and 'IFN low' subgroups but also in different modules by combining interferon type I and II expression profiles (38, 40-42). The results from **chapter 6** will help to stratify patients to make more personalized treatment choices. In our study, it was shown that by *combining* interferon signatures with neutrophil (NPh) and plasma cell (PLC) signatures, three different SLE-fingerprints can be seen in cSLE. These fingerprints showed that different types of IFN gene signatures were associated with different types of organ involvement and type of auto-antibodies (high titer of anti-ds-DNA and presence of anti-SSA, but not for presence of anti-Sm or -RNP). Our so-called 'fingerprint 3' with the highest disease activity and high NPh and/or PLC gene signatures consisted of patients who were more frequent on treatment with

CS. This could mean that this group is at the highest risk for disease damage. We need longitudinal studies to study these gene signatures in more detail over time, in search for associations with specific type of treatment and risk for damage, as also suggested by Northcott (41).

Current treatment choices are made depending on the severity of symptoms or clinical phenotype of the patient and they are re-evaluated on a regular basis, usually every 3-6 months,. Unfortunately many treatments (such as MMF, RTX or cyclophosphamide) often take up to 4-8 weeks to show some treatment effect. If after this period treatment efficacy is not enough, changes in treatment regimen have to be considered and this means delay *and* also more cumulative exposure to CS. Consequently, this leads to higher risk for CS-toxicity and higher risk for damage (due to longer period of higher disease activity *and* to longer CS use). There is need for more guidance leading to precision medicine to identify *which* patients need *which* therapy. This is not only necessary for patients with severe disease but also for patients with lower risk for high disease activity/-damage or flares. This latter group might not need (high dosage) CS, or only for a short period of time. There is need for more (predictive) disease biomarkers to identify patients that can taper faster to lower the risk for steroid-related damage.

Interferon signatures and ‘fingerprints’ as described in **chapter 6** will be helpful in stratifying patients into subgroups: patients with milder phenotype and patients with (potential) severe phenotypes that might need to start early with the currently available specific B-cell therapy or other novel therapy options (also see figure 1). However, it is necessary to measure IFN signatures directly after diagnosis in treatment-naïve patients because over time they do not always seem to correlate with disease activity. Northcott et al. showed that (initial) IFN high patients were more likely to be of East Asian ethnicity, younger at disease onset and more frequently showed anti-RNP, anti-Ro and anti-La auto-antibodies. IFN high patients also had significantly more active disease, lower attainment of LLDAS and higher rates of flare. In this study was concluded that IFN status, at diagnosis, had prognostic significance in the management of SLE (41). As was also shown in **chapter 6**, pre-treatment analysis for fingerprints in transcriptomics indeed seems to be helpful in identifying patients with higher disease activity but also seems to identify low-risk patients that might be able to taper steroids faster. By using this T2T strategy combined with transcriptomics, the risk for steroid-related damage may be reduced. The focus on lowering steroids as soon as possible is a reachable goal in developed countries like the Netherlands. A similar example is the shift in treatment of systemic juvenile idiopathic arthritis (sJIA) for which we start with anakinra (anti-IL-1) treatment up front, directly after diagnosis. As a consequence we see in our country that most of these sJIA patients do not have to use any CS at all during their disease course.

It is difficult to put this CS sparing strategy into financial numbers but in this example by starting with a more expensive medicine, it leads to faster remission rates and shorter treatment periods. This not only lowers the burden of chronic CS use but might also be cheaper in the long-term. Such insights are important to study further and can be implemented to other rheumatic diseases with high CS use, such as SLE.

Future needs and perspectives in transcriptomics

It will be informative to combine all of the hopeful (and novel) SLE disease biomarkers, that are described in this thesis. We are planning to analyse if (the number of) nailfold capillary hemorrhages, the presence of a capillary scleroderma pattern and/or dysregulated EC markers are correlated to the different IFN signatures. We need to know if specific transcriptomic ‘fingerprints’ or other biomarkers can predict the risk for damage better than SLEDAI, preferably at the moment of diagnosis. Correlating IFN-signatures with specific patterns in B cell subsets, especially in pre-treatment samples, is another interesting option to profile these (c)SLE patients. This could also lead to insights in further stratification to different SLE phenotypes with different risk profiles, and concomitant treatment strategies. Once optimal profiling phenotypes in cSLE are established, choosing personalized treatment regimens for different (c)SLE phenotypes is the next step forward. In the future this can also be implemented in drug trials. These trials are often set up by dividing patients with/without nephritis but it might also be interesting to look at responses between patients with different types of gene signature profiles.

OVERLAP DISEASE IN SYSTEMIC AUTOIMMUNITY

It is important to know in MCTD patients with systemic disease and overlapping symptoms which patients will develop to SLE- or SSc-like disease, especially in children that will probably suffer from this systemic autoimmunity for decades to come. Musculoskeletal involvement seems more prevalent in JMCTD than in adults with MCTD (43). Some studies describe arthritis in MCTD as evolvement into rheumatoid arthritis (RA) because of the combination of arthritis with a positive rheumatoid factor (RF) or anti-CCP antibodies. These specific, arthritis-linked, autoantibodies are probably not reflecting a different disease, such as RA, but may reflect disease severity and risk for bone erosions. The MRI abnormalities in **chapter 8** confirmed this severity of inflammatory and osteochondral abnormalities in a small cohort of CTD patients (3/10 RF-positive). Only some of these patients were JMCTD patients, others fulfilled the criteria for SLE and Sjögren's disease. Most important finding was that the clinical arthritis activity scores

with visual analogue scale (VAS), by experienced clinicians were relatively low. Because of this discrepancy, caution should be given to a possible underestimation of the severity of musculoskeletal involvement of children with CTD, but follow-up studies in these patients are needed. Instead of an indication for RA, the relevance of RF in these MCTD patients may be related to more severe musculoskeletal disease.

The presence of a capillary scleroderma pattern does not seem to be specific for patients with SSc. This pattern has been observed in different CTD's including SLE and (J)DM (44). A capillary scleroderma pattern in (J)DM has also been described in many studies (45). A 'scleroderma-like DM-pattern' was even proposed by some, with the observation of specific giant-ramified capillaries, almost exclusively seen in the DM patients (46). In our capillaroscopy studies in **chapter 4 and 5** it is shown that the majority of MCTD/UCTD, but also JDM patients, show a capillary scleroderma pattern. In those chapters it was also shown that a scleroderma pattern can be seen in up to 15% of cSLE patients, without any overlapping symptoms with SSc. Further investigation of the specific details of the capillary abnormalities in SLE- and (J)DM-patients with a scleroderma pattern will have to elucidate if this is (or is not) different from the capillary scleroderma pattern in (j)SSc. In Sjögren's disease, capillary abnormalities are not that common or only mild (47). In SLE patients the capillaries show more pathological findings as has been described in **chapter 2-5**. Schematically a spectrum of mild to severe vasculopathy, as seen by nailfold capillaroscopy, can be visualized as in figure 2. For jMCTD/UCTD, with overlapping clinical symptoms, the capillary pattern seems to be more severe with a high percentage of scleroderma patterns as was also shown in our patient cohorts from **chapter 4 and 5** (69 and 61%, respectively), although the cohort numbers were small (n=13 and n=18, respectively).

If the type of capillary pattern does not stratify MCTD patients, EC markers or IFN signatures may indicate whether an MCTD patient is likely to have (or will evolve into) SSc-like or more SLE-like disease. In SSc, endothelial dysregulation is prominent in many endothelial functions (8). In a recent study on EC markers in jMCTD, dysregulated levels of sICAM, IL-6 and vWF were observed in patients after a median disease duration of 17 years, compared to controls. By regression analysis with adjustment for cardiovascular risk factors, only sICAM-1 remained significant as sign for endothelial dysregulation from underlying auto-inflammation (48). In JDM, endothelial markers, such as endoglin and ICAM-1 seemed suitable for measurement of severe vasculopathy for identifying patients with capillary low end-row loop (ERL) score (<4 capillaries/mm). Nevertheless, combined with other biomarkers such as galectin-9 and CXCL-10 and clinical data, ICAM-1 and VEGF did not aid in identification of subgroups for disease severity (49). However, it is questionable if the variable ERL is enough to reflect the severity of vasculopathy in

JDM. The pathological capillaries in JDM also include many giants and hemorrhages in the active phase of this disease and a lower density can also be seen during lower disease activity states, without giants and hemorrhages. As mentioned earlier, it is important to take the ethnic background in the interpretation of capillary density into account. The complexity of endothelial dysregulation in these different CTDs is large and future studies have to show if EC markers and/or nailfold capillary changes will be helpful in differentiating MCTD from other CTD's.

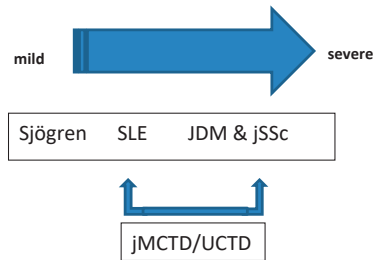


Figure 2. Spectrum of nailfold capillary abnormalities in different systemic autoimmune diseases
 SLE=systemic lupus erythematosus, JDM=juvenile dermatomyositis, jSSc=juvenile systemic sclerosis, jMCTD=juvenile mixed connective tissue disease, UCTD=undifferentiated connective tissue disease

Future needs and perspectives in overlap disease of systemic autoimmunity

Some of the MCTD/UCTD patients might have this disease as a separate entity but they often show overlap symptoms and in time evolve into SSc or SLE. It is not yet known how to predict the clinical course of these patients. During follow-up of MCTD/UCTD patients, clinicians need to know which patients have to be screened for SSc-like symptoms (with pulmonary function tests and CT scans of the thorax) and which for more SLE-like symptoms (such as regular urine analysis). These organs (lungs and kidney) can be inflamed without the patient experiencing symptoms, especially in the early stages of organ involvement. We need large longitudinal databases, with a multicenter set-up due to the rarity of these diseases, in the pediatric *and* adult population. Longitudinal follow-up of clinical variables is essential but we also need to know what happens to the capillary abnormalities. Further detailed specification of the capillary pathological patterns is necessary, also in a longitudinal setting. Again, the key in understanding the pathophysiology of these overlapping diseases might be to combine clinical variables with EC markers, transcriptomics and capillaroscopy data to define better subgroups with prognostic meaning.

LOCALIZED SCLERODERMA

In **chapter 7** pediatric patients with an auto-inflammatory skin disease, defined as LS, have been described. LS is a very rare disease and is often not recognized, or diagnosed with a delay of many years after onset. Ongoing disease activity is difficult to objectify because there are different types of smoldering symptoms that reflect disease activity and can lead to invalidating damage. The surface area of the affected skin often shows slowly increasing hyper- or hypopigmentation, skin atrophy, slight redness of edges and progressive skin hardness. LS can slowly smolder for years, with ongoing disease activity leading to (preventable) skin damage but LS can also diminish spontaneously and stay inactive. It is important to realize that after many years of disease remission, LS can flare up again. Another problem besides recognition and progression of the disease is the fact that many physicians are not aware of the systemic treatment opportunities for LS. Many dermatologists have little experience with these systemic immunosuppressant's in children. Because of its low prevalence, we believe that all pediatric patients with (severe) LS should be treated in academic hospitals in multidisciplinary expertise teams with pediatric rheumatologists and dermatologists for the best quality of care.

The Localized Scleroderma Assessment Tool (LoSCAT) has been validated for assessment and follow-up by measuring disease activity (modified Localized Skin Severity Index (mLoSSI)) and -damage (modified Localized Skin Damage Index (mLoSDSI)), but it has some limitations as described in the introduction of this thesis with variability in inter-observer scoring (50). General aspects of the skin in children are different from adults allowing for adaptations of the pediatric skin thickness scoring in LS and jSSc (51). Lastly, only the most severe part of the lesion is being scored in LoSCAT which is most often the center of the lesion. Although surface area is also a scoring item, there is still a risk when using LOSCAT that slowly deteriorating skin (smoldering disease) is not well detected or documented.

In **chapter 7** we have shown that the skin durometer, HFUS and LASCA are promising non-invasive imaging techniques that can measure skin abnormalities in LS. Of these, the durometer is likely to be the most preferred in terms of costs and handiness. The durometer was able to produce measurements of skin hardness, not only with significant differences between affected and unaffected skin, but also between edge and center of the affected lesion. If this technique is used in follow-up of patients it is very important that the measurements will be performed at the exact same skin sites as before. This should be documented carefully in patient charts with photographic images. The durometer as imaging technique in LS (and/or in (j)SSc) needs to be further validated but it might be a disease activity biomarker with higher sensitivity and inter-observer

rates than LoSCAT. High-frequency ultrasonography (HFUS) and laser speckle contrast analysis (LASCA) also showed potential to be used as imaging techniques in LS, but they have more practical limitations because of training, expertise and costs.

Some physicians that treat patients with LS are not always fully aware of the available systemic treatments for this disease, other than topical anti-inflammatory crèmes which are often not effective enough. Methotrexate (MTX) can be used as a systemic therapy, with or without prednisolone bridging. These two medicines frequently show side effects and MTX is often not well tolerated for long-term treatment, which is necessary in LS. Another systemic treatment option is MMF, which is mostly well-tolerated by patients. Nevertheless, for MMF compliance is often a problem.

Future needs and perspectives in treatment of localized scleroderma

In conclusion, there is an urgent need for new systemic treatment options in LS. Biologicals, such as tocilizumab and abatacept, have been described to be promising in treatment of LS but are still not officially approved. This gives an extra important reason for more longitudinal and multicenter studies in LS because these (new) treatments have only been described in case reports or case series, mainly because of the rarity of this disease (52-56). In future studies, new imaging techniques as described in **chapter 7** might be helpful in better monitoring (new) treatment effects, and especially the durometer seems an imaging technique that can be used in the context of multicenter trials.

CONCLUSIONS

This thesis aimed to provide direction on novel disease biomarkers from, mainly non-invasive, imaging tools in different (systemic) autoimmune diseases in children, with a specific emphasis on cSLE:

- By detection of nailfold capillaroscopy, capillary hemorrhages (in presence, type and per mm) are correlated with disease activity in (c)SLE.
- We hypothesize that a specific capillary ‘lupus pattern’, consisting of abnormal capillary morphology combined with hemorrhages, seems to exist and this observation needs to be studied further.
- A capillary scleroderma pattern is present in up to 15% of SLE patients and is associated with lupus-related damage and *not* with SSc-like overlap disease.
- Profiling of cSLE patients with ‘fingerprints’ from IFN-signatures and NPh/PLC modelling is a promising measurement for stratification at diagnosis (in a pre-treatment stage) for the implementation of personalized and CS-sparing treatment strategies.

- The severity of musculoskeletal inflammation in pediatric CTD, as visualized by MRI, seems to be underestimated by clinical arthritis activity as assessed on a Likert scale (0-5).
- The durometer can give quantitative measurements of inflamed skin in LS: it should be validated to measure disease activity in this rare localized connective tissue disease.

As mentioned in the introduction, diagnosis and survival rates for (c)SLE have been greatly improved in the last decades. Nevertheless, patients with SLE still suffer daily from this chronic lifelong disease with high risk for organ damage. A high number of daily pills, fatigue, lower quality of life and limited social participation are examples of the daily burden for a SLE patient which is a very difficult perspective for the future of a teenager that receives this diagnosis at a young age. Still, even though many insights and improvements were made in last decades, more disease biomarkers are needed to predict, and subsequently to prevent, disease damage and improve quality of life of these patients. Translational- and international collaborative research is the indispensable key to success. Most importantly, those (novel) disease biomarkers should be easy to implement in daily clinical care of pediatric patients with (rare) rheumatic diseases. In the end, those children with a pediatric onset of rheumatic disease are growing up to be adults. We need to further improve and adapt clinical care of these patients constantly, because these (pediatric) patients have to live their whole life with the effects of a chronic inflammatory disease.

REFERENCES

1. Snow MH, Saketkoo LA, Frech TM, Stever JR, Lebedoff N, Herrick AL, et al. Results from an American pilot survey among Scleroderma Clinical Trials Consortium members on capillaroscopy use and how to best implement nailfold capillaroscopy training. *Clinical and experimental rheumatology*. 2019;37 Suppl 119(4):151.
2. Cutolo M, Melsens K, Herrick AL, Foeldvari I, Deschepper E, De Keyser F, et al. Reliability of simple capillaroscopic definitions in describing capillary morphology in rheumatic diseases. *Rheumatology (Oxford)*. 2018;57(4):757-9.
3. Cutolo M, Melsens K, Wijnant S, Ingegnoli F, Thevissen K, De Keyser F, et al. Nailfold capillaroscopy in systemic lupus erythematosus: A systematic review and critical appraisal. *Autoimmun Rev*. 2018;17(4):344-52.
4. Cutolo M. *Atlas of capillaroscopy in rheumatic diseases*. Milan: Elsevier; 2010.
5. Smith V, Vanhaecke A, Herrick AL, Distler O, Guerra MG, Denton CP, et al. Fast track algorithm: How to differentiate a “scleroderma pattern” from a “non-scleroderma pattern”. *Autoimmun Rev*. 2019;18(11):102394.
6. Smith V, Riccieri V, Pizzorni C, Decuman S, Deschepper E, Bonroy C, et al. Nailfold capillaroscopy for prediction of novel future severe organ involvement in systemic sclerosis. *The Journal of rheumatology*. 2013;40(12):2023-8.
7. Smith V, Decuman S, Sulli A, Bonroy C, Piette Y, Deschepper E, et al. Do worsening scleroderma capillaroscopic patterns predict future severe organ involvement? a pilot study. *Ann Rheum Dis*. 2012;71(10):1636-9.
8. Mostmans Y, Cutolo M, Giddelo C, Decuman S, Melsens K, Declercq H, et al. The role of endothelial cells in the vasculopathy of systemic sclerosis: A systematic review. *Autoimmun Rev*. 2017;16(8):774-86.
9. Westerweel PE, Luyten RK, Koomans HA, Derksen RH, Verhaar MC. Premature atherosclerotic cardiovascular disease in systemic lupus erythematosus. *Arthritis and rheumatism*. 2007;56(5):1384-96.
10. Terreri MT, Andrade LE, Puccinelli ML, Hilario MO, Goldenberg J. Nail fold capillaroscopy: normal findings in children and adolescents. *Seminars in arthritis and rheumatism*. 1999;29(1):36-42.
11. Piotto DP, Sekiyama J, Kayser C, Yamada M, Len CA, Terreri MT. Nailfold videocapillaroscopy in healthy children and adolescents: description of normal patterns. *Clinical and experimental rheumatology*. 2016;34 Suppl 100(5):193-9.
12. Zhao T, Lin FA, Chen HP. Pattern of Nailfold Capillaroscopy in Patients With Systemic Lupus Erythematosus. *Arch Rheumatol*. 2020;35(4):568-74.
13. Shenavandeh S, Habibi S. Nailfold capillaroscopic changes in patients with systemic lupus erythematosus: correlations with disease activity, skin manifestation and nephritis. *Lupus*. 2017;961203316686702.
14. Raeeskarami SR, Namazi N, Assari R, Najafizadeh SR, Hassannejad Z, Ziaee V. The Comparison of Nailfold Capillaroscopy between Juvenile Systemic Lupus Erythematosus and Healthy Controls: Correlation with Laboratory and Clinical Parameters. *Int J Vasc Med*. 2020;2020:7631958.
15. Bergkamp SC, Schonenberg-Meinema D, Nassar-Sheikh Rashid A, Melsens K, Vanhaecke A, Boumans MJH, et al. Reliable detection of subtypes of nailfold capillary haemorrhages in childhood-onset systemic lupus erythematosus. *Clinical and experimental rheumatology*. 2021.

16. Wu GC, Liu HR, Leng RX, Li XP, Li XM, Pan HF, et al. Subclinical atherosclerosis in patients with systemic lupus erythematosus: A systemic review and meta-analysis. *Autoimmun Rev*. 2016;15(1):22-37.
17. Croca SC, Griffin M, Farinha F, Isenberg DA, Nicolaidis A, Rahman A. Total plaque area and plaque echogenicity are novel measures of subclinical atherosclerosis in patients with systemic lupus erythematosus. *Rheumatology (Oxford)*. 2021;60(9):4185-98.
18. Brunner HI, Gladman DD, Ibanez D, Urowitz MD, Silverman ED. Difference in disease features between childhood-onset and adult-onset systemic lupus erythematosus. *Arthritis and rheumatism*. 2008;58(2):556-62.
19. Groot N, Shaikhani D, Teng YKO, de Leeuw K, Bijl M, Dolhain R, et al. Long-Term Clinical Outcomes in a Cohort of Adults With Childhood-Onset Systemic Lupus Erythematosus. *Arthritis Rheumatol*. 2019;71(2):290-301.
20. Holland MJ, Beresford MW, Feldman BM, Huggins J, Norambuena X, Silva CA, et al. Measuring Disease Damage and Its Severity in Childhood-Onset Systemic Lupus Erythematosus. *Arthritis care & research*. 2018;70(11):1621-9.
21. Torrente-Segarra V, Salman Monte TC, Rua-Figueroa I, De Una-Alvarez J, Balboa-Barreiro V, Lopez-Longo FJ, et al. Relationship between damage and mortality in juvenile-onset systemic lupus erythematosus: Cluster analyses in a large cohort from the Spanish Society of Rheumatology Lupus Registry (RELESSER). *Seminars in arthritis and rheumatism*. 2019;48(6):1025-9.
22. Abbas L, Joseph A, Kunzler E, Jacobe HT. Morphea: progress to date and the road ahead. *Ann Transl Med*. 2021;9(5):437.
23. van Vollenhoven RF, Mosca M, Bertsias G, Isenberg D, Kuhn A, Lerstrom K, et al. Treat-to-target in systemic lupus erythematosus: recommendations from an international task force. *Ann Rheum Dis*. 2014;73(6):958-67.
24. Smith EMD, Gorst SL, Al-Abadi E, Hawley DP, Leone V, Pilkington C, et al. 'It is good to have a target in mind': qualitative views of patients and parents informing a treat to target clinical trial in juvenile-onset systemic lupus erythematosus. *Rheumatology (Oxford)*. 2021;60(12):5630-41.
25. Petri M, Magder LS. Comparison of Remission and Lupus Low Disease Activity State in Damage Prevention in a United States Systemic Lupus Erythematosus Cohort. *Arthritis Rheumatol*. 2018;70(11):1790-5.
26. Tsang ASMW, Bultink IE, Heslinga M, Voskuyl AE. Both prolonged remission and Lupus Low Disease Activity State are associated with reduced damage accrual in systemic lupus erythematosus. *Rheumatology (Oxford)*. 2017;56(1):121-8.
27. Franklyn K, Lau CS, Navarra SV, Louthrenoo W, Lateef A, Hamijoyo L, et al. Definition and initial validation of a Lupus Low Disease Activity State (LLDAS). *Ann Rheum Dis*. 2016;75(9):1615-21.
28. Ozturk K, Caglayan S, Tanatar A, Baglan E, Yener Otar G, Kavrul Kayaalp G, et al. Low disease activity state in juvenile-onset systemic lupus erythematosus. *Lupus*. 2021;30(13):2144-50.
29. Smith EMD, Tharmaratnam K, Al-Abadi E, Armon K, Bailey K, Brennan M, et al. Attainment of Low Disease Activity and Remission Targets reduces the risk of severe flare and new damage in Childhood Lupus. *Rheumatology (Oxford)*. 2021.
30. Wahadat MJ, van den Berg L, Timmermans D, van Rijswijk K, van Dijk-Hummelman A, Bakx S, et al. LLDAS is an attainable treat-to-target goal in childhood-onset SLE. *Lupus Sci Med*. 2021;8(1).
31. Heshin-Bekenstein M, Trupin L, Yelin E, von Scheven E, Yazdany J, Lawson EF. Longitudinal disease- and steroid-related damage among adults with childhood-onset systemic lupus erythematosus. *Seminars in arthritis and rheumatism*. 2019;49(2):267-72.

32. van Vollenhoven RF, Bertsias G, Doria A, Isenberg D, Morand E, Petri MA, et al. 2021 DORIS definition of remission in SLE: final recommendations from an international task force. *Lupus Sci Med.* 2021;8(1).
33. Leandro M, Isenberg DA. Rituximab - The first twenty years. *Lupus.* 2021;30(3):371-7.
34. Oomen I, Nassar-Sheikh Rashid A, Bouts AHM, Gouw SC, Kuijpers TW, Rispens T, et al. Anti-rituximab antibodies affect pharmacokinetics and pharmacodynamics of rituximab in children with immune-mediated diseases. *Clinical and experimental rheumatology.* 2022;40(1):183-90.
35. Groot N, de Graeff N, Avcin T, Bader-Meunier B, Brogan P, Dolezalova P, et al. European evidence-based recommendations for diagnosis and treatment of childhood-onset systemic lupus erythematosus: the SHARE initiative. *Ann Rheum Dis.* 2017;76(11):1788-96.
36. Groot N, de Graeff N, Marks SD, Brogan P, Avcin T, Bader-Meunier B, et al. European evidence-based recommendations for the diagnosis and treatment of childhood-onset lupus nephritis: the SHARE initiative. *Ann Rheum Dis.* 2017;76(12):1965-73.
37. Lewis MJ, Jawad AS. The effect of ethnicity and genetic ancestry on the epidemiology, clinical features and outcome of systemic lupus erythematosus. *Rheumatology (Oxford).* 2017;56(suppl_1):i67-i77.
38. Banchereau R, Hong S, Cantarel B, Baldwin N, Baisch J, Edens M, et al. Personalized Immunomonitoring Uncovers Molecular Networks that Stratify Lupus Patients. *Cell.* 2016;165(3):551-65.
39. Oliveira JJ, Karrar S, Rainbow DB, Pinder CL, Clarke P, Rubio Garcia A, et al. The plasma biomarker soluble SIGLEC-1 is associated with the type I interferon transcriptional signature, ethnic background and renal disease in systemic lupus erythematosus. *Arthritis Res Ther.* 2018;20(1):152.
40. Chiche L, Jourde-Chiche N, Whalen E, Presnell S, Gersuk V, Dang K, et al. Modular transcriptional repertoire analyses of adults with systemic lupus erythematosus reveal distinct type I and type II interferon signatures. *Arthritis Rheumatol.* 2014;66(6):1583-95.
41. Northcott M, Jones S, Koelmeyer R, Bonin J, Vincent F, Kandane-Rathnayake R, et al. Type 1 interferon status in systemic lupus erythematosus: a longitudinal analysis. *Lupus Sci Med.* 2022;9(1).
42. Postal M, Vivaldo JF, Fernandez-Ruiz R, Paredes JL, Appenzeller S, Niewold TB. Type I interferon in the pathogenesis of systemic lupus erythematosus. *Curr Opin Immunol.* 2020;67:87-94.
43. Scalapino K, Arkachaisri T, Lucas M, Fertig N, Helfrich DJ, Londino AV, Jr., et al. Childhood onset systemic sclerosis: classification, clinical and serologic features, and survival in comparison with adult onset disease. *The Journal of rheumatology.* 2006;33(5):1004-13.
44. Nagy G, Czirjak L, Kumanovics G. Patients with Systemic Sclerosis with and without Overlap Syndrome Show Similar Microvascular Abnormalities. *Diagnostics (Basel).* 2021;11(9).
45. Piette Y, Reynaert V, Vanhaecke A, Bonroy C, Gutermuth J, Sulli A, et al. Standardised interpretation of capillaroscopy in autoimmune idiopathic inflammatory myopathies: A structured review on behalf of the EULAR study group on microcirculation in Rheumatic Diseases. *Autoimmun Rev.* 2022;21(6):103087.
46. Manfredi A, Sebastiani M, Campomori F, Pipitone N, Giuggioli D, Colaci M, et al. Nailfold Videocapillaroscopy Alterations in Dermatomyositis and Systemic Sclerosis: Toward Identification of a Specific Pattern. *The Journal of rheumatology.* 2016;43(8):1575-80.
47. Melsens K, Leone MC, Paolino S, Elewaut D, Gerli R, Vanhaecke A, et al. Nailfold capillaroscopy in Sjogren's syndrome: a systematic literature review and standardised interpretation. *Clinical and experimental rheumatology.* 2020;38 Suppl 126(4):150-7.
48. Skjelland M, Zamani M, Hetlevik S, Lilleby V, Halvorsen B, Skagen K. Increased Endothelial Activation in Patients with Mixed Connective Tissue Disease. *J Stroke Cerebrovasc Dis.* 2020;29(2):104563.

49. Wienke J, Pachman LM, Morgan GA, Yeo JG, Amoruso MC, Hans V, et al. Endothelial and Inflammation Biomarker Profiles at Diagnosis Reflecting Clinical Heterogeneity and Serving as a Prognostic Tool for Treatment Response in Two Independent Cohorts of Patients With Juvenile Dermatomyositis. *Arthritis Rheumatol.* 2020;72(7):1214-26.
50. Agazzi A, Fadanelli G, Vittadello F, Zulian F, Martini G. Reliability of LoSCAT score for activity and tissue damage assessment in a large cohort of patients with Juvenile Localized Scleroderma. *Pediatric rheumatology online journal.* 2018;16(1):37.
51. Foeldvari I, Torok KS. Review for best practice in clinical rheumatology juvenile systemic sclerosis - Updates and practice points. *Best Pract Res Clin Rheumatol.* 2021;35(3):101688.
52. Zulian F, Culpo R, Sperotto F, Anton J, Avcin T, Baildam EM, et al. Consensus-based recommendations for the management of juvenile localised scleroderma. *Ann Rheum Dis.* 2019;78(8):1019-24.
53. Foeldvari I. Update on the Systemic Treatment of Pediatric Localized Scleroderma. *Paediatr Drugs.* 2019;21(6):461-7.
54. Ulc E, Rudnicka L, Waskiel-Burnat A, Warszawik-Hendzel O, Niemczyk A, Olszewska M. Therapeutic and Reconstructive Management Options in Scleroderma (Morphea) en Coup de Sabre in Children and Adults. A Systematic Literature Review. *J Clin Med.* 2021;10(19).
55. Talia J, Bitar C, Wang Y, Whitfield ML, Khanna D. A Case of Recalcitrant Linear Morphea Responding to Subcutaneous Abatacept. *J Scleroderma Relat Disord.* 2021;6(2):194-8.
56. Wenzel D, Haddadi NS, Afshari K, Richmond JM, Rashighi M. Upcoming treatments for morphea. *Immun Inflamm Dis.* 2021;9(4):1101-45.

10

Summary

In the introduction of this thesis (chapter 1), background information is given about different autoimmune and so-called ‘connective tissue diseases’ (CTD) that are discussed in this thesis, such as Systemic Lupus Erythematosus (SLE), Mixed Connective Tissue Disease (MCTD) and Localized Scleroderma (LS). Although much progress has been made in the early diagnosis and treatment of these diseases in the last decades, still more biomarkers are needed that stratify patients by different (prognostic) disease severity grades. This stratification will lead to improve personalized treatment choices which will lower the risk for irreversible damage and improve their quality of life while living with a chronic disease.

PART I. NAILFOLD CAPILLAROSCOPY IN CSLE

This thesis consists of two parts. The first part is about nailfold capillaroscopy. The smallest blood vessels of the body (capillaries) can be visualized with this imaging device. Capillaroscopy is feasible and painless. Observations from capillaroscopy studies in children with CTD are being described with an emphasis on the severe autoimmune disease SLE. SLE with a childhood-onset is called cSLE.

In **chapter 2** a systematic literature review was provided on nailfold capillary abnormalities in cSLE. At the moment of literature search, in 2019, six articles were retained, of which two case-control studies and four case series. For capillary density, no difference was found between cSLE and healthy controls (one study). Differences in capillary diameter, capillary morphology, hemorrhages and semi-quantitative score were inconclusive or non-interpretable. A scleroderma pattern was reported in a minority of cSLE patients in three out of four case series. We concluded that literature on nailfold capillary findings in cSLE is scarce and inconclusive. To evaluate capillary characteristics in cSLE, prospective and longitudinal studies are needed with uniform definitions for capillary characteristics.

In **chapter 3** we described the capillary abnormalities from a cross-sectional study in cSLE (n=41), compared to capillary observations from matched healthy controls (n=41). The secondary objective was to correlate the observed capillary abnormalities with demographical variables and with disease-specific variables in cSLE patients. cSLE-patients showed significantly more ‘giants’ (p=0.032), ‘abnormal capillary shapes’ (p=0.003), ‘large capillary hemorrhages’ (p<0.001) and ‘pericapillary extravasations’ (p<0.001). By qualitative analysis, the pattern ‘microangiopathy’ was detected in 68.3% (28/41) and a ‘scleroderma pattern’ in 17.1% (7/41) of the cSLE-patients (but without scleroderma symptoms). ‘Microangiopathy’ consisted of the combination of capillary

hemorrhages and abnormal capillary shapes. The difference of percentage positive anti-RNP antibodies in the group with or without a scleroderma pattern was not significantly different ($p=0.089$). The number of 'abnormal capillary shapes per mm' was correlated with treatment-naivety. The number of 'large pathological hemorrhages per mm' was correlated with SLEDAI score and presence of nephritis. Compared to healthy controls, 'pericapillary extravasations' were found in significantly higher numbers per mm ($p<0.001$) as well as in percentage of patients ($p<0.001$) and that was never described before in such detail. These observations confirmed for the first time that giants, abnormal capillary morphology and capillary hemorrhages are also observed in cSLE, as was already known for adults with SLE. A high frequency and total amount of "pericapillary extravasations" was also seen in our cSLE patients, possibly revealing a new subtype of capillary hemorrhage that might reflect endothelial damage in these pediatric patients

Chapter 4 shows a world-wide multicenter study with comparison of capillary findings between pediatric patients with different rheumatic diseases compared to healthy controls. Patients with rheumatic diseases showed many significant differences compared to healthy controls; capillary density was lower in jSSc, JDM, MCTD en cSLE and in these diseases significantly more hemorrhages were observed as well. A scleroderma pattern was seen in jSSc, MCTD and JDM but also in 15% of cSLE. In cSLE, an abnormal 'non-specific' capillary pattern was also found. In healthy children, capillary density only showed a weak correlation with age ($R=0.14$, $p=0.046$), in contrary with findings from earlier studies.

Chapter 5 describes the longitudinal data from our cSLE-cohort ($n=53$) with a median disease onset of 14 years (IQR 12.5-15.5 years), median SLEDAI score at diagnosis was 11 (IQR 8-16) and median SLEDAI at follow up was 2 (IQR 1-6). A scleroderma pattern (ever) was seen in 18.9% and micro-angiopathy in 67.9%, while only 13.2% of patients had a normal capillary pattern. Thirty-three patients had follow-up capillaroscopy of which 21.2% showed changes in type of capillary pattern over time. Type of capillary pattern was not associated with disease activity. Raynaud's phenomenon (ever) was equally distributed among patients with different capillaroscopy patterns ($p=0.26$). Anti-RNP antibodies (ever) were significantly more detected (Chi square, $p=0.016$) in the scleroderma pattern subgroup ($n=7/10$, 70%). Already 5 years after disease onset more than 50% of patients with a scleroderma pattern had SLE-related disease damage (HR 4.5, 95%CI 1.1-18.8, $p=0.034$), but they did not develop clinical features of systemic sclerosis at follow-up. Number of detected fingers with a scleroderma pattern was similar between cSLE, jSSc and jUCTD. This longitudinal study shows that the majority of capillary patterns in cSLE are abnormal and they can change over time. Irrespective of

disease activity, a capillary scleroderma pattern in cSLE may be associated with higher risk for SLE-related disease damage.

PART II. NEW BIOMARKERS IN (SYSTEMIC) CONNECTIVE TISSUE DISEASES

The second part of this describes novel methods to might be used in treatment of cSLE patients and related CTD's. This concerned new techniques in blood analyses but also the 'older' technique MRI for imaging of the wrist. Also, results from new imaging techniques (durometer, high-frequency ultrasound and laser speckle contrast analysis) are described in children with the autoimmune disease localized scleroderma (LS) to quantitatively measure the inflammation in the skin.

Chapter 6 aimed to translate existing transcriptomic data into simpler gene signatures suitable for daily clinical practice in cSLE patients. RT-PCR of multiple genes from the Interferon M1.2, Interferon M5.12, neutrophil (NPh) and plasma cell (PLC) modules followed by a principle component analysis, was used to identify indicator genes per gene signature. Gene signatures were measured in longitudinal samples from two childhood onset SLE cohorts (n=101 and n=34, respectively) and associated with clinical features. Cluster analysis subdivided patients into three mutually exclusive fingerprint-groups termed 1) all-signatures-low, 2) only IFN high (M1.2 and/or M5.12) and 3) high NPh and/or PLC. All gene signatures were significantly associated with disease activity in cross-sectional collected samples. The PLC-signature showed the highest association with disease activity. Interestingly, in longitudinally collected samples, the PLC-signature was associated with disease activity and showed a decrease over time. When patients were divided into fingerprints, the highest disease activity was observed in the high NPh and/or PLC group. The lowest disease activity was observed in the all-signatures-low group. The same distribution was reproduced in samples from an independent SLE cohort. The identified gene signatures were associated with disease activity and are suitable tools to stratify SLE patients into groups with similar activated immune pathways that may guide future treatment choices.

In **chapter 7** a pilot study in LS patients. It describes imaging data from the most prominent lesion (i.e. "target lesion") with examination of the centre, edge and contralateral unaffected site from ten patients with LS. High-frequency ultrasonography was used to determine dermal thickness, durometer for skin hardness, and laser speckle contrast analysis (LASCA) for a dynamical evaluation of the microcirculation. Dermal thickness was thinner at the centre of the "target lesions" vs. the edges ($p < 0.001$) and control

sites ($p<0.001$). Skin hardness was harder at the centre of the “target lesions” vs. the edges ($p=0.012$) and control sites ($p=0.003$). A higher perfusion was found in the centre of the “target lesion” (124.87 ± 66.40 PU) vs. the edges (87.27 ± 46.40 PU; $p<0.001$) and control sites (67.85 ± 37.49 ; $p<0.001$). This case series suggests the supportive value of both microcirculatory and dermal assessments of skin lesions using novel non-invasive research tools, adopted from adult SSc, for the identification, scoring and/or monitoring of LS.

Chapter 8 describes ten patients with pediatric connective tissue disease (with clinical (overlap of) subtypes systemic lupus erythematosus, Sjögren syndrome and dermatomyositis) with a median age of 14.7 years (IQR 12.7-16.6 years) and 70% female. MRI wrist datasets were evaluated by three readers in consensus for synovitis, tenosynovitis, bone marrow changes, bone erosions and myositis. Clinical arthritis activity was scored low (median visual analogue scale physician 19, IQR 7-31). Notwithstanding, extensive inflammatory abnormalities (synovitis and tenosynovitis) were found in the wrist of 7/10 patients. Osteochondral involvement was detected in 3/10 patients. It was concluded that severe inflammatory abnormalities in the wrist could be present while clinical disease scores suggested relatively mild disease activity. These are important findings because the wrist is known to be vulnerable for joint damage.

PART III. DISCUSSION, SUMMARY AND APPENDIX

In **chapter 9** all results from this thesis are discussed in a broader perspective. This leads to a recommendations on the current and future needs in patient care and research topics, especially for cSLE, jMCTD and LS.

NEDERLANDSE SAMENVATTING

In de inleiding van dit proefschrift (**hoofdstuk 1**), wordt achtergrondinformatie gegeven over de verschillende auto-immuun- en zogenaamde ‘connective tissue’ ziekten (CTD’s) zoals Systemische Lupus Erythematosus (SLE), Mixed Connective Tissue Disease (MCTD) en geLokaliseerde Sclerodermie (LS). Hoewel er in de afgelopen jaren veel progressie is geweest in vroegtijdige diagnose en behandelingen van deze ziektebeelden, zijn er nog steeds nieuwe biomarkers nodig die patiënten beter verdelen in (prognostische) groepen met verschillende ernst van de ziekte. Deze verdeling in subgroepen zal leiden tot verbeterde gepersonaliseerde keuze in behandeling, en dit zal leiden tot lager risico op onherstelbare schade en de kwaliteit van leven verbeteren ondanks het leven met een chronische ziekte.

Deel 1. Nagelriem capillaroscopie in kinderen met SLE

Dit proefschrift bestaat uit twee delen. Het eerste deel gaat over nagelriem capillaroscopie. Met dit apparaat kunnen de kleinste bloedvaatjes (capillairen) van het lichaam bij de nagelriem in beeld worden gebracht. Dit onderzoek is makkelijk uitvoerbaar en pijnloos. De bevindingen van capillaroscopie onderzoek worden beschreven bij kinderen met CTD, met de nadruk op de ernstige auto-immuunziekte SLE. SLE op de kinderleeftijd wordt childhood-onset SLE (cSLE) genoemd.

In **hoofdstuk 2** werd een ‘systematisch literatuuroverzicht’ gegeven van alle gepubliceerde literatuur over capillaire afwijkingen bij cSLE. Op het moment van dit literatuuroverzicht in 2019 werden er zes artikelen gevonden waarvan twee casus-controle studies en vier casuïstiek series. In capillaire dichtheid werd geen verschil gevonden tussen cSLE en gezonde controles (1 studie). Verschillen in capillaire diameter, morfologie, bloedingen en semi-kwantitatieve score waren niet-conclusief of niet te interpreteren. Een sclerodermiepatroon werd gerapporteerd in een minderheid van de cSLE patiënten in drie van vier casuïstiek series. We concludeerden dat de literatuur over capillaire bevindingen van de nagelriem in cSLE beperkt is en niet-conclusief. Om capillaire karakteristieken in cSLE te evalueren zijn er prospectieve en longitudinale studies nodig met gebruik van uniforme definities voor capillaire afwijkingen.

In **hoofdstuk 3** beschrijven we de capillaire afwijkingen van een cross-sectionele studie in cSLE patiënten (n=41), in vergelijking met capillairen van vergelijkbare gezonde kinderen (n=41). Het tweede doel van deze studie was om de abnormale capillaire afwijkingen te correleren met demografische variabelen en met ziekte-specifieke factoren in cSLE patiënten. cSLE patiënten lieten significant meer ‘reuzencapillairen’ (p=0.032), ‘abnormale capillaire vormen’ (p=0.003), ‘grote capillaire bloedingen’ (p<0.001) en

'pericapillaire extravasaties' ($p < 0.001$) zien. Met behulp van kwalitatieve analyse werd het patroon van 'microangiopathy' gedetecteerd in 68.3% (28/41) en een 'sclerodermie patroon' in 17.1% (7/41) van de cSLE-patiënten (zonder sclerodermie symptomen). 'Microangiopathy' bestond uit de combinatie van capillaire bloedingen en abnormale capillaire vormen. Er was geen verschil tussen percentage positieve anti-RNP antistoffen in de groep met of zonder een sclerodermiepatroon ($p = 0.089$). Het aantal 'abnormale capillaire vormen per mm' was gecorreleerd met treatment-naïviteit. The aantal 'grote pathologische bloedingen per mm' was gecorreleerd met SLEDAI score en aanwezigheid van nefritis. Vergeleken met gezonde controles werden 'pericapillaire extravasaties' in significant hogere aantallen per mm ($p < 0.001$) gezien en ook in hoger percentage van patiënten ($p < 0.001$) en dat was nog nooit zo in detail beschreven. Deze observaties bevestigen dus voor het eerst dat reuzencapillairen, abnormale capillaire morfologie en capillaire bloedingen ook worden geobserveerd in cSLE, net als bij volwassenen met SLE. Een hoge frequentie en aantal van 'pericapillaire extravasaties' werd gezien in onze cSLE patiënten, dit lijkt een nieuw subtype van capillaire bloeding te zijn die mogelijk de endotheel-schade in deze patiënten reflecteert.

Hoofdstuk 4 beschrijft een internationale studie waarbij de capillaire afwijkingen bij kinderen met verschillende auto-immuunziekten werden beschreven, in vergelijking met gezonde controles. Patiënten met reumatische ziektebeelden lieten significant meer afwijkingen zien in vergelijking met gezonde controles; capillaire dichtheid was lager in jSSc, JDM, MCTD en cSLE en in deze ziektebeelden werden ook significant meer capillaire bloedingen gezien. Een sclerodermie patroon werd gezien in jSSc, MCTD en JDM maar ook in 15% van cSLE patiënten. In cSLE werd ook een afwijkend 'non-specifiek' patroon gevonden. In gezonde kinderen liet capillaire dichtheid slechts een zwakke correlatie zien met leeftijd ($R = 0.14$, $p = 0.046$), in tegenstelling tot bevindingen uit eerdere studies.

In **hoofdstuk 5** werden de resultaten beschreven van ons longitudinaal cSLE cohort ($n = 53$) met een mediane leeftijd van begin ziekte van 14 jaar (IQR 12.5-15.5 jaar), mediane SLEDAI score bij diagnose van 11 (IQR 8-16) en een mediane SLEDAI at follow-up van 2 (IQR 1-6). Een sclerodermie patroon (geobserveerd ooit tijdens follow-up) werd gezien in 18.9% van de patiënten en 'micro-angiopathie' bij 67.9%, terwijl slechts 13.2% van patiënten een normaal capillair patroon liet zien. $N = 33$ patiënten kregen follow-up capillaroscopie waarvan 21.2% veranderingen liet zien in soort capillaire patroon over de tijd heen. Het type capillaire patroon was niet geassocieerd met ziekteactiviteit. Het optreden van Raynaud fenomeen (ooit) was gelijkmatig verdeeld over de patiënten met verschillende capillaire patronen ($p = 0.26$). Anti-RNP antistoffen (ooit) werden significant meer gezien (Chi square, $p = 0.016$) in de subgroep met sclerodermie patroon ($n = 7/10$).

Meer dan 50% van de patiënten met een sclerodermie patroon had al SLE-gerelateerde schade door de ziekte opgelopen binnen 5 jaar na start van de klachten (HR 4.5, 95%CI 1.1-18.8, $p=0.034$), maar zij vertoonden geen klinische kenmerken van de ziekte SSc bij follow-up. Het aantal vingers met een sclerodermie patroon was hetzelfde bij patiënten met cSLE, jSSc en JUCTD. Deze longitudinale studie laat zien dat de meeste capillaire patronen in cSLE afwijkend zijn en ook kunnen veranderen over de tijd. Daarnaast lijkt, onafhankelijk van ziekteactiviteit, een sclerodermie patroon geassocieerd met een hoger risico op SLE-gerelateerde schade aan de organen.

Deel II. Nieuwe biomarkers in (systemische) ‘connective tissue’ ziektebeelden

Het tweede deel van dit proefschrift gaat over nieuwe onderzoeksmethoden die behulpzaam zouden kunnen zijn in de behandeling van kinderen met SLE en vergelijkbare “connective tissue” ziekten. Dit betreft nieuwe technieken om het bloed te analyseren maar ook over ‘oudere’ beeldvormende technieken zoals MRI van het polsgewricht. Daarnaast worden nieuwe beeldvormende technieken (durometer, hoogfrequentie echo en laser speckle contrast analyse) beschreven in kinderen met de auto-immuunziekte gelokaliseerde sclerodermie (LS) om op een kwantitatieve manier de inflammatie in de huid te meten.

Hoofdstuk 6 had als doel om bestaande ‘transcriptomic data’ te vertalen naar simpelere genetische signaturen die bruikbaar zijn in de dagelijkse praktijk bij cSLE patiënten. Meerdere genexpressies van de Interferon M1.2, Interferon M5.12, Neutrofielen (NPh) en Plasma Cel (PLC) modules, gevolgd door een 'principale componenten analyse', werden gebruikt om indicator genen per genetische signatuur te identificeren. Genetische signaturen werden gemeten in longitudinale samples van twee cSLE cohorten ($n=101$ en $n=34$) en geassocieerd met klinische verschijnselen. Cluster analyse verdeelde patiënten in drie exclusieve ‘fingerprints’ groepen genaamd 1) alle-signaturen-laag, 2) alleen IFN hoog (M1.2 en/of M5.12) en 3) hoog NPh en/of PLC. Alle genetische signaturen waren significant geassocieerd met ziekteactiviteit in de cross-sectionele samples. PLC-signatuur had de hoogste associatie met ziekteactiviteit. Een opvallende bevinding, in de longitudinale monsters, was dat de PLC-signatuur geassocieerd was met ziekteactiviteit en een lagere waarden liet zien over de tijd. Als patiënten werden verdeeld met 'fingerprints' dan werd de hoogste ziekteactiviteit geobserveerd in de hoog NPh en/of PLC groep. De laagste ziekteactiviteit werd geobserveerd in de alle-signaturen-laag groep. Dezelfde verdelingen in groepen werden gereproduceerd in een onafhankelijk cSLE cohort. De geïdentificeerde gen signaturen waren geassocieerd met ziekteactiviteit en zijn passende tools om SLE patiënten te verdelen in groepen met dezelfde type immuun-activatie die toekomstige behandelingskeuzes meer richting kunnen geven.

In **hoofdstuk 7** wordt een pilot studie beschreven bij patiënten met gelokaliseerde sclerodermie (LS). Hierin worden de data beschreven van beeldvormingstechnieken bij de meest prominente huidlaesies ("target laesie") met onderzoek van het centrum en rand van de huidlaesie en van de contralaterale niet-aangedane kant van tien patiënten met LS. Hoogfrequentie echo werd gebruikt om de huiddikte vast te stellen, een durometer voor het meten van de hardheid van de huid en Laser Speckle Contrast Analyse (LASCA) voor dynamische evaluatie van de kleine bloedvaatjes (microcirculatie). De huiddikte was significant minder in het centrum van de 'target laesies' ten opzichte van de randen ($p < 0.001$) en de niet-aangedane kant ($p < 0.012$). Een hogere bloedperfusie werd gevonden in het centrum van de "target lesion" (124.87 ± 66.40 PU) ten opzichte van de randen (87.27 ± 46.40 PU; $p < 0.001$) en de controle kant (67.85 ± 37.49 ; $p < 0.001$). Deze casuïstiek serie laat zien dat, overgenomen vanuit volwassen SSc, zowel metingen in microcirculatie als huiddikte/-hardheid van de aangedane huid waarde lijken te hebben als nieuwe non-invasieve instrumenten, in het vaststellen, scoren en/of monitoring van LS.

Hoofdstuk 8 beschrijft tien patiënten met een pediatrische "connective tissue" ziekte (met klinische overlap van de subtypes SLE, SSc, Sjögren syndroom en dermatomyositis) met een mediane leeftijd van 14.7 jaar (IQR 12.7-16.6 jaar) bestaand uit 70% vrouw. MRI beelden van de pols werden geëvalueerd door drie experts om consensus te krijgen voor het scoren van synovitis, tenosynovitis, beenmergveranderingen, boterosies en myositis. De klinische ziekteactiviteit werd als relatief laag gescoord (mediane visuele analoge school voor arts 19 (0-100), IQR 7-31). Desalniettemin werden er uitgebreide inflammatoire afwijkingen gevonden (synovitis en tenosynovitis) in de pols bij 7/10 patiënten. Kraakbeenbetrokkenheid werd gezien in 3/10 patiënten. Geconcludeerd werd dat er ernstige inflammatoire afwijkingen in de pols aanwezig kunnen zijn terwijl de klinische ziektescores als relatief mild werd beoordeeld. Dit zijn belangrijke bevindingen omdat specifiek de pols als gewricht kwetsbaar is voor gewrichtsschade.

Deel III. Discussie, samenvatting en appendix

In **hoofdstuk 9** worden alle resultaten van dit proefschrift bediscussieerd en in een breder perspectief geplaatst. Dit leidt tot aanbevelingen voor huidige en toekomstige behoeften in patiëntenzorg en voor onderwerpen van onderzoek, in het bijzonder voor cSLE, jMCTD en LS.

A

LIST OF ABBREVIATIONS

ACR: American College of Rheumatology
ADA: anti-drug antibodies
Anti-ds-DNA: anti-double-stranded DNA antibodies
Anti-RNP: anti-Ribonucleoprotein antibody
Anti-Sm: anti-Smith antibodies
aSLE: adult-onset Systemic Lupus Erythematosus
BEL: belimumab
BILAG: British Isles Lupus Assessment Group
CIMT: Carotid-Intima-Media-Thickness
CS: corticosteroids
cSLE: childhood-onset Systemic Lupus Erythematosus
CYC: cyclofosfamide
CTD: Connective Tissue Disease
EC: Endothelial Cell
EULAR: European League Against Rheumatism
HR: hazard ratio
HRQoL: health related quality of life
IFN: interferon
IQR: interquartile range
JDM: Juvenile Dermatomyositis
jMCTD: juvenile Mixed Connective Tissue Disease
jSSc: juvenile Systemic Sclerosis
LASCA: Laser Speckle Contrast Analysis (LASCA)
LLDAS: Lupus Low Disease Activity State
LoSCAT: Localized Scleroderma Assessment Tool
LS: Localized scleroderma
(M)CTD: (Mixed) Connective Tissue Disease
MMF: mycophenolate mofetil
MTX: methotrexate
NFC: nailfold capillaroscopy
NIH: National Institutes of Health
NPh: neutrophil
NVC: nailfold videocapillaroscopy
OR: odds ratio
PBP: peripheral blood perfusion
PLC: plasma cell
PReS: Pediatric Rheumatology European Society

RP: Raynaud's phenomenon

RTX: rituximab

SDI: SLICC Damage Index

SG MCRD: Study Group on Microcirculation in Rheumatic Diseases

SLE: Systemic Lupus Erythematosus

SLEDAI: Systemic Lupus Erythematosus Activity Index

SLICC: Systemic Lupus International Collaborating Clinics

SSc: Systemic Sclerosis

T2T: Treat to Target

UCTD: Undifferentiated Connective Tissue Disease

AUTHOR AFFILIATIONS (ALPHABETICALLY)

Author	Affiliation
Armbrust W (Wineke)	UMC Groningen, University of Groningen, Department of Pediatric Rheumatology and Immunology, Beatrix Children's Hospital, Groningen, The Netherlands
Badot V (Valerie)	Department of Rheumatology, CHU Brugmann, Université Libre de Bruxelles (ULB), Brussels, Belgium
Bergkamp SC (Sandy)	Department of Pediatric Immunology, Rheumatology and Infectious Diseases, Emma Children's Hospital Amsterdam University Medical Centres, University of Amsterdam, Amsterdam, The Netherlands
Bouchalova K (Katerina)	Paediatric Rheumatology, Department of Paediatrics, Faculty of Medicine and Dentistry, Palacky University Olomouc and University Hospital, Olomouc, Czech Republic
Brinkman DMC (Danielle)	Department of Pediatrics, Division of Pediatric Rheumatology, Willem Alexander Children's Hospital, Leiden University Medical Center, The Netherlands
Cimaz R (Rolando)	Pediatric Rheumatology Unit, Gaetano Pini Hospital, Department of Clinical Sciences and Community Health, Research Centre for Adult and Pediatric Rheumatic Diseases, Università degli Studi di Milano, Milan, Italy
Cutolo M (Maurizio)	Laboratory of Experimental Rheumatology and Academic Division of Clinical Rheumatology, Department of Internal Medicine, University of Genova; IRCCS San Martino Polyclinic, Genoa, Italy
Dolman KM (Koert)	Department of Pediatrics, Onze Lieve Vrouwe Gasthuis; Department of Pediatric Rheumatology, Reade, Amsterdam, The Netherlands
de Bree GJ (Godelieve)	Department of Infectious Diseases, Amsterdam University Medical Centers, University of Amsterdam, the Netherlands

Dehoorne J (Joke)	Department of Pediatric Rheumatology, Ghent University Hospital, Ghent, Belgium
Deschepper E (Ellen)	Department of Public Health and Primary Care, Biostatistics Unit, Ghent University, Ghent, Belgium
De Schepper S (Sophie)	Department of Dermatology, Ghent University Hospital, Ghent, Belgium
Dvorak D (Denis)	Paediatric Rheumatology, Department of Paediatrics, Faculty of Medicine and Dentistry, Palacky University Olomouc and University Hospital, Olomouc, Czech Republic
Foeldvari I (Ivan)	Centre for Paediatric and Adolescent Rheumatology, An der Schön Klinik, Hamburg, Germany
Frech T (Tracy)	University of Utah, Department of Internal Medicine, Division of Rheumatology; Salt Lake Veterans Affairs Medical Centre, Utah Vascular Research Laboratory, Salt Lake City, Utah, USA
Groot N (Noortje)	Department of Paediatric Rheumatology, Erasmus MC-Sophia Children's hospital, University Medical Center Rotterdam, Rotterdam, The Netherlands
Gruppen MP (Mariken)	Department of Pediatric Immunology, Rheumatology and Infectious Disease, Emma Children's Hospital, Amsterdam University Medical Centres, University of Amsterdam, Amsterdam, The Netherlands
Hak AE (Elisabeth)	Department of Rheumatology and Clinical Immunology, Amsterdam University Medical Centres, University of Amsterdam; Amsterdam Rheumatology and Immunology Center, Amsterdam, The Netherlands
Hernandez-Zapata J (Johanna)	Department of Pediatrics, Universidad de Antioquia, Medellín, Colombia
Herrick AL (Ariane)	Centre for Musculoskeletal Research, The University of Manchester; Salford Royal NHS Foundation Trust, Manchester Academic Health Science Centre, Manchester, UK

Hissink Muller PC (Petra)	Department of Pediatrics, Division of Pediatric Rheumatology, Willem Alexander Children's Hospital, Leiden University Medical Center, The Netherlands
Hoppenreijns EPAH (Esther)	Department of Paediatric Rheumatology, Amalia Children's Hospital, Radboud University Medical Center; Department of Paediatric Rheumatology, St. Maartenskliniek, Nijmegen, The Netherlands
Iagnocco A (Annamaria)	Academic Rheumatology Centre, Department of Clinical and Biological Science, University of Turin, Italy
Ingegnoli F (Francesca)	Clinical Rheumatology Unit, Gaetano Pini Hospital, Department of Clinical Sciences and Community Health, Research Centre for Adult and Pediatric Rheumatic Diseases, Università degli Studi di Milano, Milan, Italy
Kamphuis S (Sylvia)	Department of Paediatric Rheumatology, Erasmus MC-Sophia Children's hospital, University Medical Center Rotterdam, Rotterdam, The Netherlands.
Karantanas A (Apostolos)	Department of Medical Imaging, Heraklion University Hospital Crete, Greece
Khan A (Archana)	Department of Pediatric Rheumatology, SRCC Children's Hospital, Mumbai, India
Krasowska D (Dorota)	Department of Dermatology, Venereology and Pediatric Dermatology, Medical University of Lublin, Lublin, Poland
Kuijpers TW (Taco)	Department of Pediatric Immunology, Rheumatology and Infectious Diseases, Emma Children's Hospital, Amsterdam University Medical Centres, University of Amsterdam, Amsterdam, The Netherlands
Lehmann H (Hartwig)	Department of Pediatrics, University Medicine Gießen, Gießen, Germany
Leone MC (Maria)	Department of Rheumatology, Ghent University Hospital, Ghent, Belgium; Medical and Rheumatological Clinic, S. Maria Hospital, Terni, Italy
Maas M (Mario)	Department of Radiology, Amsterdam University Medical Centres, University of Amsterdam, Amsterdam, The Netherlands

Makol A (Ashima)	Division of Rheumatology, Mayo Clinic, Rochester, MN, USA
Melsens, K (Karin)	Department of Rheumatology, Ghent University Hospital; Department of Internal Medicine, Ghent, University, Ghent, Belgium
Mesa-Navas MA (Miguel)	Rheumatology Section, Clínica Universitaria Universidad Pontificia Bolivariana, Medellín, Colombia
Michalska-Jakubus M (Malgorzata)	Department of Dermatology, Venereology and Pediatric Dermatology, Medical University of Lublin, Lublin, Poland
Middelkamp-Hup MA (Maritza)	Department of Dermatology, Amsterdam University Medical Centres, University of Amsterdam, Amsterdam, The Netherlands
Mostmans Y (Yora)	Department of Immunology-Allergology, CHU Brugmann, Université Libre de Bruxelles (ULB); Dpt of Dermatology, CHU Brugmann, Université Libre de Bruxelles (ULB), Brussels, Belgium
Müller-Ladner U (Ulf)	Department of Rheumatology and Clinical Immunology, Campus Kerckhoff, Justus-Liebig University, Gießen; Kerckhoff-Klinik GmbH, Bad Nauheim, Germany
Nassar-Sheikh Rashid A (Amara)	Department of Pediatric Immunology, Rheumatology and Infectious diseases, Amsterdam University Medical Centres, University of Amsterdam, Amsterdam; Department of Pediatrics, Zaans Medisch Centrum, Zaandam, The Netherlands
Ntailiani K (Katerina)	Department of Medical Imaging, Heraklion University Hospital Crete, 71110, Voutes, Greece
Nusman CM (Charlotte)	Department of Radiology; Department of Pediatric Immunology, Rheumatology and Infectious Diseases, Emma Children's Hospital, Amsterdam University Medical Centres, University of Amsterdam, Amsterdam, The Netherlands
Nuño-Nuño L (Laura)	Division of Rheumatology, Hospital La Paz, Madrid, Spain

Overbury R (Rebecca)	University of Utah, Department of Internal Medicine, Division of Rheumatology; University of Utah, Department of Pediatrics, Division of Pediatric Rheumatology, Salt Lake City, Utah, USA
Pizzorni C (Carmen)	Laboratory of Experimental Rheumatology and Academic Division of Clinical Rheumatology, Department of Internal Medicine, University of Genova; IRCCS San Martino Polyclinic, Genoa, Italy
Radic M (Mislav)	University of Utah, Department of Internal Medicine, Division of Rheumatology, Salt Lake City, Utah, USA; University Hospital Split, Split, Croatia
Ramadoss D (Divya)	Department of Pediatric Rheumatology, SRCC Children's Hospital, Mumbai, India
Ravelli, A (Angelo)	Clinica Pediatrica e Reumatologia, IRCCS Istituto Giannina Gaslini; University of Genoa, Genoa, Italy; Sechenov First Moscow State Medical University, Moscow, Russian Federation
Rosina S (Silvia)	Clinica Pediatrica e Reumatologia, IRCCS Istituto Giannina Gaslini, Genoa, Italy
Schatorjé EJH (Ellen)	Department of Paediatric Rheumatology, Amalia Children's Hospital, Radboud University Medical Center; Department of Paediatric Rheumatology, St. Maartenskliniek, Nijmegen, The Netherlands
Schonenberg-Meinema D (Dieneke)	Department of Pediatric Immunology, Rheumatology and Infectious diseases, Amsterdam University Medical Centres, University of Amsterdam, Amsterdam, Netherlands
Smith V (Vanessa)	Department of Rheumatology, Ghent University Hospital; Department of Internal Medicine, Ghent University; Unit for Molecular Immunology and Inflammation, Inflammation Research Centre, VIB-Ghent University, Ghent, Belgium
Sulli A (Alberto)	Laboratory of Experimental Rheumatology and Academic Division of Clinical Rheumatology, Department of Internal Medicine, University of Genova; IRCCS San Martino Polyclinic, Genoa, Italy

Swart JF (Joost)	Department of Paediatric Immunology, Wilhelmina Children's Hospital, University of Utrecht, University Medical Center Utrecht, Utrecht, The Netherlands.
Ten Cate R (Rebecca)	Department of Pediatrics, Division of Pediatric Rheumatology, Willem Alexander Children's Hospital, Leiden University Medical Center, The Netherlands
Udaondo C (Clara)	Pediatric Rheumatology Department., La Paz Children's Hospital, Madrid, Spain
van den Berg JM (Merlijn)	Department of Pediatric Immunology, Rheumatology and Infectious diseases, Amsterdam University Medical Centres, University of Amsterdam, Amsterdam, the Netherlands
van der Aa LB (Leontien)	Department of Pediatrics, Division of Pediatric Rheumatology, Willem Alexander Children's Hospital, Leiden University Medical Center; Department of Pediatrics, Zaans Medisch Centrum, Zaandam, The Netherlands
Vanhaecke A (Amber)	Department of Internal Medicine, Ghent University, and Department of Rheumatology, Ghent University Hospital, Ghent, Belgium
van Helden-Meeuwssen CG (Cornelia)	Department of Immunology, Erasmus MC, University Medical Center Rotterdam, Rotterdam, The Netherlands
van Onna M (Marieke)	Department of Rheumatology and Clinical Immunology, Amsterdam University Medical Centres, University of Amsterdam; Amsterdam Rheumatology and Immunology Center, Amsterdam, The Netherlands
van Tilburg SJ (Sander)	Department of Immunology, Erasmus MC, University Medical Center Rotterdam, Rotterdam, The Netherlands
Verkaaik M (Maaike)	Department of Paediatric Rheumatology, Erasmus MC-Sophia Children's hospital, University Medical Center Rotterdam, Rotterdam, The Netherlands
Versnel MA (Marjan)	Department of Immunology, Erasmus MC, University Medical Center Rotterdam, Rotterdam, The Netherlands

Wahadat MJ (Javad)

Department of Paediatric Rheumatology and
department of Immunology Erasmus MC-Sophia
Children's hospital, University Medical Center
Rotterdam, Rotterdam, The Netherlands

AUTHOR CONTRIBUTIONS PER CHAPTER

Chapter 2: DSM and VS initiated this review, DS, ANSR and JMvdB screened the search hits and performed quality appraisal on retained articles. DSM, VS and KM were involved in data analysis and writing of the manuscript. TWK and MC were involved in editing and critical revision of the manuscript.

Chapter 3: DS, JMvdB, VS and TWK initiated this study. DS, SB, AN, LBvdAA, GJdB, RtC, AEH, MvO, TWK and JMvdB recruited patients and healthy controls. DS, SB and JMvdB were involved in data analysis and writing of the manuscript. All authors were involved in editing and critical revision of the manuscript.

Chapter 4: KM, MC, DSM, IF, ALH, AS and VS had substantial contributions to the ideation of the study, substantial contributions to the design of the study. KM, DSM, IF, MCL, YM, VB, RC, JDH, TF, JHZ, FI, AK, DK, HL, AM, MAMN, MMJ, UML, LNN, RO, CP, MR, RD, AR, SR, CU and JMvdB had substantial contribution to the acquisition of data. KM, DSM, MCL, EDS and VS had substantial contribution in analysis and interpretation of data. KM and VS were involved in drafting of the article. All authors were involved in editing and critical revision of the manuscript.

Chapter 5: DSM, JMvdB, VS and TK initiated this study. DSM, SB, ANSR, MG, MM-H, WA, KD, AEH, PHM, MvO, JS, TK and JMvdB recruited patients. DS-M, SB, VS, SSMK, JS and JMvdB were involved in data analysis and writing of the manuscript. All authors were involved in editing and critical revision of the manuscript.

Chapter 6: SK and MAV contributed to the study conception and design. MJW, CGvHM, ST and MAV contributed to the experimental work. MJW, DSM, CGvHM, SJvT, NG, EJHS, EPAHH, PCEHM, .DMCB, DD, MV, JMvdB, KB, SK and MAV contributed to the acquisition of data. MJW, DSM, SJvT, SK and MAV contributed to the analysis and interpretation of the data. MJW, SK and MAV contributed to the writing of the draft manuscript. MJW, DSM, CGvHM, SJvT, NG, EJHS, EPAHH, PCEHM, DMCB, DD, MV, JMvdB, KB, SK and MAV contributed to the rewriting of the manuscript.

Chapter 7: AvH, DSM, JMvdB and VS initiated this study. DSM, SdS, MAMH, ANSR, JMvdB and TWK recruited patients. AvH, DSM, SdS, SCB, MCL, ANSR, MAMH, JMvdB and VS were involved in acquisition of data. AvH, DSM, AI, MC and VS were involved in data analysis, interpretation of data and writing of manuscript. All authors were involved in editing and critical revision of the manuscript.

Chapter 8: CN, DSM and MM initiated this study. CN, JMvdB, ANSR, DSM, KN and MM were involved in acquisition of data. CN, KN, DSM and MM were involved in data analysis and writing of the manuscript. CN, KN, and DS. All authors were involved in editing and critical revision of the manuscript.

BIBLIOGRAPHY

This thesis:

Melsens K*, Cutolo M*, **Schonenberg-Meinema D**, Foeldvari I, Leone MC, Mostmans Y, Badot V, Cimaz R, Dehoorne J, Deschepper E, Frech T, Hernandez-Zapata J, Ingegnoli F, Khan A, Krasowksa, Hartwig D Lehman, Makol A, Mesa-Navas MA, Michalska-Jakubus M, Müller-Ladner M, Nuño-Nuño L, Overbury R, Pizzorni C, Radic M, Ramadoss D, Ravelli A, Rosina S, Udaondo C, van den Berg JM, Herrick AL, Sulli A, Smith V from EULAR Study Group on Microcirculation in Rheumatic Diseases.; *Standardized nailfold capillaroscopy in children with rheumatic diseases: a worldwide study*. *Shared first authors. *Rheumatology* (Oxford). 2022 Aug 25:keac487. doi: 10.1093/rheumatology/keac487. Online ahead of print.

Wahadat MJ, **Schonenberg-Meinema D**, van Helden-Meeuwsen CG, van Tilburg S, Groot N, Schatorjé EJH, Hoppenreijns EPAH, Hissink Muller PCE, Brinkman DMC, Dvorak D, Verkaaik M, Bouchalova K, van den Berg JM, Kamphuis S, Versnel MA. *Gene signature fingerprints divide SLE patients in subgroups with similar biological disease profiles: a multicenter longitudinal study*. *Rheumatology* (Oxford). 2022 Feb 10:keac083. doi: 10.1093/rheumatology/keac083. Online ahead of print.

Schonenberg-Meinema D, Bergkamp SC, Nassar-Sheikh Rashid A, Gruppen MP, Middelkamp-Hup MA, Armbrust W, Dolman KM, Hak AE, Hissink Muller PCE, van Onna M, Swart JF, Kuijpers TW, Kamphuis S, Smith V, van den Berg JM. *A capillary scleroderma pattern in childhood-onset may be associated with disease damage: important lessons from longitudinal follow-up*. *Lupus Sci Med*. 2022 Feb;9(1):e000572.

Vanhaecke A*, **Schonenberg-Meinema D***, de Schepper S, Bergkamp SC, Leone MC, Middelkamp-Hup MA, Nassar-Sheikh Rashid A, van den Berg JM, Kuijpers TW, Iagnocco A, Cutolo M, Smith V. *Rarities in rare: illuminating the microvascular and dermal status in juvenile localised scleroderma. A case series*. *Clin Exp Rheumatol* 2022; 40 (Suppl. 134): S12-S18. *shared first author

Schonenberg-Meinema D, Bergkamp SC, Nassar-Sheikh Rashid A, van der Aa LB, de Bree GJ, Ten Cate R, Cutolo M, Hak AE, Hissink Muller PC, van Onna M, Kuijpers TW, Smith V, van den Berg JM. *Nailfold capillary abnormalities in childhood-onset systemic lupus erythematosus: a cross-sectional study compared with healthy controls*. *Lupus*. 2021 Apr;30(5):818-827.

Schonenberg-Meinema D, Melsens K, Nassar-Sheikh Rashid A, Cutolo M, Kuijpers TW, van den Berg JM, Smith V. *Capillaroscopy in childhood-onset systemic lupus erythematosus: a first systemic review*. Clin Exp Rheumatol. 2020 Mar-Apr;38(2):350-354.

Nusman CM, van den Berg JM, Nassar-Sheikh Rashid A, Ntailiani K, Karantanas A, Kuijpers TW, Maas M, **Schonenberg-Meinema D**. *Consider the wrist: a retrospective study on pediatric connective tissue disease with MRI*. Rheumatol Int. 2019 Dec;39(12):2095-2101.

Other peer-reviewed publications:

Verkuil F, van den Berg JM, van Gulik EC, Barendregt AM, Nassar Sheikh-Rashid A, **Schonenberg-Meinema D**, Dolman KM, Kuijpers TW, Maas M, Hemke R. *Synovial signal intensity on static contrast-enhanced MRI for evaluation of disease activity in juvenile idiopathic arthritis - A look at the bright side of the knee*. Clin Imaging. 2022 Jun;86:53-60.

van der Krogt JMA, Verkuil F, van Gulik EC, Hemke R, van den Berg JM, **Schonenberg-Meinema D**, Kindermann A, Dolman KM, Benninga MA, Kuijpers TW, Maas M, Nusman CM. *Comparison of contrast-enhanced MRI features of the (teno)synovium in the wrist of patients with juvenile idiopathic arthritis and pediatric controls*. Rheumatol Int. 2022; 42: 1257–1264.

Verhoeven D, **Schonenberg-Meinema D**, Ebstein F, Papendorf JJ, Baars PA, van Leeuwen EMM, Jansen MH, Lankester AC, van der Burg M, Florquin S, Maas SM, van Koningsbruggen S, Krüger E, van den Berg JM, Kuijpers TW. *Hematopoietic stem cell transplantation in a patient with proteasome-associated autoinflammatory syndrome (PRAAS)*. J Allergy Clin Immunol. 2022 Mar;149(3):1120-1127.

Nassar-Sheikh Rashid A, **Schonenberg-Meinema D**, Berends SE, van den Berg JM, Mathôt RAA. *Population pharmacokinetics of infliximab in children with juvenile idiopathic arthritis*. Ther Drug Monit. 2022 April; 44(2):301-307.

Foeldvari I, Klotsche J, Hinrichs B, Helmus N, Kasapcopur O, Adrovic A, Sztajnbok F, Terrier MT, Anton J, Smith V, Katsicas M, Kostik M, Vasquez-Canizares N, Avcin T, Feldman B, Janarthanan M, Santos MJ, Sawhney S, **Schonenberg-Meinema D**, Sifuentes-Giraldo WA, Alexeeva E, Appenzeller S, Battagliotti C, Berntson L, Bica B, Costa Reis P, Eleftheriou D, Kallinich T, Lehman T, Marrani E, Minden K, Nielsen S, Nuruzzaman F, Patwardhan A, Khubchandani R, Stanevicha V, Uziel Y, Torok KS. *Under detection of interstitial lung disease in juvenile systemic sclerosis (jSSc)*. Arthritis Care Res (Hoboken). 2022 Mar;74(3):364-370.

Oomen I, Nassar-Sheikh Rashid A, Bouts AHM, Gouw SC, Kuijpers TW, Rispens T, de Vries A, Wolbink G, van den Berg JM, **Schonenberg-Meinema D**. *Anti-rituximab antibodies affect pharmacokinetics and pharmacodynamics of rituximab in children with immune-mediated diseases*. Clin Exp Rheumatol. 2022 Jan-Feb;40(1):183-190.

Chalhoub NE, Wenderfer SE, Levy DM, Rouster-Stevens K, Aggarwal A, Savani SI, Ruth NM, Arkachaisri T, Qiu T, Merritt A, Onel K, Goilav B, Khubchandani RP, Deng J, Fonseca AR, Ardoin SP, Ciurtin C, Kasapcopur O, Jelusic M, Huber AM, Ozen S, Klein-Gitelman MS, Appenzeller S, Cavalcanti A, Fotis L, Lim SC, Silva RM, Miramontes JR, Rosenwasser NL, Saad-Magalhaes C, **Schonenberg-Meinema D**, Scott C, Silva CA, Enciso S, Terreri MT, Torres-Jimenez AR, Trachana M, Al-Mayouf SM, Devarajan P, Huang B, Brunner HI; Childhood Arthritis and Rheumatology Research Alliance Lupus Nephritis Work Group and the Pediatric Rheumatology European Society Lupus Working Party. *International Consensus for the Dosing of Corticosteroids in Childhood-Onset Systemic Lupus Erythematosus With Proliferative Lupus Nephritis*. Arthritis Rheumatol. 2022 Feb;74(2):263-273.

Bergkamp SC, **Schonenberg-Meinema D**, Nassar-Sheikh Rashid A, Melsens K, Vanhaecke A, Boumans MJH, Hissink Muller PCE, Cutolo M, Kuijpers TW, van den Berg JM, Smith V. *Reliable detection of subtypes of nailfold capillary hemorrhages in childhood-onset systemic lupus erythematosus*. Clin Exp Rheumatol. 2021 Sep-Oct;39(5):1126-1131.

Maller J, Fox E, Park KT, Paul SS, Baszis K, Borocco C, Pahalad S, Quartier P, Reinhardt A, **Schonenberg-Meinema D**, Shipman L, Terreri MT, Simard J, Lavi I, Chalom E, Hsu J, Zisman D, Mellins ED; CARRA Legacy Registry Investigators. *Inflammatory Bowel Disease in Children with Systemic Juvenile Idiopathic Arthritis*. J Rheumatol 2021 Apr;48(4):567-574.

Nassar-Sheikh Rashid A, **Schonenberg-Meinema D**, Bergkamp SC, Bakhklakh S, de Vries A, Rispens T, Kuijpers TW, Wolbink G, van den Berg JM. *Therapeutic drug monitoring of anti-TNF drugs: an overview of applicability in daily clinical practice in the era of treatment with biologics in juvenile idiopathic arthritis (JIA)*. Pediatr Rheumatol Online J. 2021 Apr 29;19(1):59.

Ingegnoli F, Herrick AL, Schioppo T, Bartoli F, Ughi N, Pauling JD, Sulli A, Cutolo M, Smith V; European League against Rheumatism (EULAR) study group on microcirculation in rheumatic diseases and the Scleroderma Clinical Trials Consortium**. *Reporting items for capillaroscopy in clinical research on musculoskeletal diseases: a systematic review and international Delphi consensus*. Rheumatology (Oxford). 2021 Mar 2;60(3):1410-1418. ****Schonenberg-Meinema as collaborator in consortium**

Foeldvari I, Klotsche J, Kasapcopur O, Adrovic A, Terreri MT, Sakamoto AP, Stanevicha V, Sztajnbok F, Anton J, Feldman B, Alexeeva E, Katsicas M, Smith V, Avcin T, Marrani E, Kostik M, Lehman T, Sifuentes-Giraldo WA, Vasquez-Canizares N, Appenzeller S, Janarthanan M, Moll M, Nemcova D, Patwardhan A, Santos MJ, Sawhney S, **Schonenberg-Meinema D**, Battagliotti C, Berntson L, Bica B, Brunner J, Costa-Reis P, Eleftheriou D, Harel L, Horneff G, Kaiser D, Kallinich T, Lazarevic D, Minden K, Nielsen S, Nuruzzaman F, Uziel Y, Helmus N, Torok KS. *Differences sustained between diffuse and limited forms of juvenile systemic sclerosis in expanded international cohort. www.juvenile-scleroderma.com. Arthritis Care Res (Hoboken).* 2022 Oct;74(10):1575-1584

Barendregt AM, Veldkamp SR, Hissink Muller PCE, van de Geer A, Aarts C, van Gulik EC, Schilham MW, Kessel C, Keizer MP, Hemke R, Nassar-Sheikh Rashid A, Dolman KM, **Schonenberg-Meinema D**, Ten Cate R, van den Berg JM, Maas M, Kuijpers TW. *MRP8/14 and neutrophil elastase for predicting treatment response and occurrence of flare in patients with juvenile idiopathic arthritis. Rheumatology (Oxford).* 2020 Sep 1;59(9):2392-2401.

Verkuil F, van Gulik EC, Nusman CM, Barendregt AM, Nassar-Sheikh Rashid A, **Schonenberg-Meinema D**, Dolman KM, Maas M, Kuijpers TW, van den Berg JM, Hemke R. *Exploring contrast-enhanced MRI findings of the clinically non-inflamed symptomatic pediatric wrist. Pediatr Radiol.* 2020 Sep;50(10):1387-1396

Keuning MW, Kamp GA, **Schonenberg-Meinema D**, Dorigo-Zetsma JW, van Zuiden JM, Pajkrt D. *Congenital syphilis, the great imitator-case report and review. Lancet Infect Dis.* 2020 Jul;20(7):e173-e179.

Barendregt AM, Mazzoli V, van Gulik EC, **Schonenberg-Meinema D**, Nassar-Sheikh Al Rashid A, Nusman CM, Dolman KM, van den Berg JM, Kuijpers TW, Nederveen AJ, Maas M, Hemke R. *Juvenile Idiopathic Arthritis: Diffusion-weighted MRI in the Assessment of Arthritis in the Knee. Radiology.* 2020 May;295(2):373-380.

van Gulik EC, Verkuil F, Barendregt AM, **Schonenberg-Meinema D**, Rashid AN, Kuijpers TW, van den Berg JM, Hoving JL. *Experiences, perspectives and expectations of adolescents with juvenile idiopathic arthritis regarding future work participation; a qualitative study. Pediatr Rheumatol Online J.* 2020 Apr 15;18(1):33.

Luijten MAJ, Terwee CB, van Oers HA, Joosten MMH, van den Berg JM, **Schonenberg-Meinema D**, Dolman KM, Ten Cate R, Roorda LD, Grootenhuis MA, van Rossum MAJ, Haverman L. *Psychometric properties of the pediatric Patient-Reported-Outcomes Measurement Information System (PROMIS) item banks in a Dutch clinical sample of children with juvenile idiopathic arthritis*. *Arthritis Care Res (Hoboken)*. 2020 Dec;72(12):1780-1789.

Saper VE, Chen G, Deutsch GH, Guillerman RP, Birgmeier J, Jagadeesh K, Canna S, Schulert G, Deterding R, Xu J, Leung AN, Bouzoubaa L, Abulaban K, Baszis K, Behrens EM, Birmingham J, Casey A, Cidon M, Cron RQ, De A, De Benedetti F, Ferguson I, Fishman MP, Goodman SI, Graham TB, Grom AA, Haines K, Hazen M, Henderson LA, Ho A, Ibarra M, Inman CJ, Jerath R, Khawaja K, Kingsbury DJ, Klein-Gitelman M, Lai K, Lapidus S, Lin C, Lin J, Liptzin DR, Milojevic D, Mombourquette J, Onel K, Ozen S, Perez M, Phillippi K, Prahalad S, Radhakrishna S, Reinhardt A, Riskalla M, Rosenwasser N, Roth J, Schneider R, **Schonenberg-Meinema D**, Shenoi S, Smith JA, Sönmez HE, Stoll ML, Towe C, Vargas SO, Vehe RK, Young LR, Yang J, Desai T, Balise R, Lu Y, Tian L, Bejerano G, Davis MM, Khatri P, Mellins ED; Childhood Arthritis and Rheumatology Research Alliance Registry Investigators. *Emergent high fatality lung disease in systemic juvenile arthritis*. *Ann Rheum Dis*. 2019 Dec;78(12):1722-1731.

Hissink Muller PCE, van Braak WG, Schreurs D, Nusman CM, Bergstra SA, Hemke R, **Schonenberg-Meinema D**, van den Berg JM, Kuijpers TW, Koopman-Keemink Y, van Rossum MAJ, van Suijlekom-Smit LWA, Brinkman DMC, Allaart CF, Ten Cate R, Maas M. *No radiographic wrist damage after treatment to target in recent-onset juvenile idiopathic arthritis*. *Pediatr Rheumatol Online J*. 2019 Sep 4;17(1):62.

Brunner HI, Holland MJ, Beresford MW, Ardoin SP, Appenzeller S, Silva CA, Flores F, Goilav B, Avar Aydin PO, Wenderfer SE, Levy DM, Ravelli A, Khubchandani R, Avcin T, Klein-Gitelman MS, Ruperto N, Feldman BM, Ying J; Paediatric Rheumatology International Trial Organisation and Pediatric Rheumatology Collaborative Study Group***. *American College of Rheumatology Provisional Criteria for Clinically Relevant Improvement in Children and Adolescents With Childhood-Onset Systemic Lupus Erythematosus*. *Arthritis Care Res (Hoboken)*. 2019 May;71(5):579-590. *****Schonenberg-Meinema as co-author in collaborative study group**

Barendregt AM, van Gulik EC, Groot PFC, Dolman KM, van den Berg JM, Nassar-Sheikh Rashid A, **Schonenberg-Meinema D**, Lavini C, Rosendahl K, Hemke R, Kuijpers TW, Maas M, Nusman CM. *Prolonged time between intravenous contrast administration and image acquisition results in increased synovial thickness at magnetic resonance imaging in patients with juvenile idiopathic arthritis*. *Pediatr Radiol*. 2019 May;49(5):638-645.

Barendregt AM, van Gulik EC, Lavini C, Nusman CM, van den Berg JM, **Schonenberg-Meinema D**, Dolman KM, Kuijpers TW, Hemke R, Maas M. *Diffusion weight imaging for assessment of synovial inflammation in juvenile idiopathic arthritis: a promising imaging biomarker as alternative to gadolinium-based contrast agents*. *Eur Radiol*. 2018 Nov 30.

Hissink Muller P, Brinkman DMC, **Schonenberg-Meinema D**, van den Bosch WB, Koopman-Keemink Y, Brederije ICJ, Bekkering PW, Kuijpers TW, Van Rossum M, van Suijlekom-Smit LW, van den Berg JM, Boehringer S, Allaart CF, Ten Cate R. *Treat to target (drug-free) inactive disease in DMARD-naive juvenile idiopathic arthritis: 24-month clinical outcomes of a three-armed randomised trial*. *Ann Rheum Dis*. 2019 Jan;78(1):51-59.

van Gulik EC, Hemke R, Welsink-Karssies MM, **Schonenberg-Meinema D**, Dolman KM, Barendregt AM, Nusman CM, Maas M, Kuijpers TW, van den Berg JM. *Normal MRI findings of the knee in patients with clinically active juvenile idiopathic arthritis*. *Eur J Radiol*. 2018 May;102:36-40.

van Gulik EC, Welsink-Karssies MM, van den Berg JM, **Schonenberg-Meinema D**, Dolman KM, Barendregt AM, Nusman CM, Maas M, Kuijpers TW, Hemke R. *Juvenile idiopathic arthritis: magnetic resonance imaging of the clinically unaffected knee*. *Pediatr Radiol*. 2018 Mar;48(3):333-340

Hemke R, van den Berg JM, Nusman CM, van Gulik EC, Barendregt AM, **Schonenberg-Meinema D**, Dolman KM, Kuijpers TW, Maas M. *Contrast-enhanced MRI findings of the knee in healthy children: establishing normal values*. *Eur Radiol*. 2018 Mar;28(3):1167-1174.

Hemke R, Nusman CM, van den Berg JM, Lavini C, **Schonenberg-Meinema D**, Dolman KM, Kuijpers TW, Maas M. *Construct validity of pixel-by-pixel DCE-MRI: correlation with conventional MRI scores in juvenile idiopathic arthritis*. *Eur J Radiol*. 2017 Sep;94:1-5

van der Meulen PM, Barendregt AM, Cuadrado E, Magro-Checa C, Steup-Beekman GM, **Schonenberg-Meinema D**, Van den Berg JM, Li QZ, Baars PA, Wouters D, Voskuyl AE, Ten Berge IRJM, Huizinga TWJ, Kuijpers TW. *Protein array autoantibody profiles to determine diagnostic markers for neuropsychiatric systemic lupus erythematosus*. *Rheumatology (Oxford)*. 2017 Aug 1;56(8):1407-1416

Hissink Muller PC, de Meij TGJ, Westedt M, de Groot EFJ, Allaart CF, Brinkman DMC. **Schonenberg-Meinema D**, van den Berg JM, van Suijlekom-Smit LW, van Rossum MA, Budding AE, Ten Cate R. *Disturbance of microbial core species in new-onset juvenile idiopathic arthritis*. *J Pediatr Infect Dis* 2017;12:131-135

Nusman CM, Hemke R, Lavini C, **Schonenberg-Meinema D**, van Rossum MA, Dolman KM, van den Berg JM, Maas M, Kuijpers TW. *Dynamic contrast-enhanced magnetic resonance imaging can play a role in predicting flare in juvenile idiopathic arthritis*. *Eur J Radiol*. 2017 Mar;88:77-81.

Hissink Muller PC, Brinkman DM, **Schonenberg D**, Koopman-Keemink Y, Brederije IC, Bekkering WP, Kuijpers TW, van Rossum MA, van Suijlekom-Smit LW, van den Berg JM, Allaart CF, Ten Cate R. *A comparison of three treatment strategies in recent onset non-systemic Juvenile Idiopathic Arthritis: initial 3-months results of the BeSt for Kids-study*. *Pediatr Rheumatol Online J*. 2017 Feb 6;15(1):11.

Nusman CM, Lavini C, Hemke R, Caan MW, **Schonenberg-Meinema D**, Dolman KM, van Rossum MA, van den Berg JM, Kuijpers TW, Maas M. *Dynamic contrast-enhanced magnetic resonance imaging of the wrist in children with juvenile idiopathic arthritis*. *Pediatr Radiol*. 2017 Feb;47(2):205-213

Hoekman DR, Roelofs JJ, van Schuppen J, **Schonenberg-Meinema D**, D'Haens GR, Benninga MA. *Case report of cheilitis granulomatosa and joint complaints as presentation of Crohn's disease*. *Clin J Gastroenterol*. 2016 Apr;9(2):73-8.

Nusman CM, Hemke R, Benninga MA, **Schonenberg-Meinema D**, Kindermann A, van Rossum MA, van den Berg JM, Maas M, Kuijpers TW. *Contrast-enhanced MRI of the knee in children unaffected by clinical arthritis compared to clinically active juvenile idiopathic arthritis patients*. *Eur Radiol*. 2016 Apr;26(4):1141-8.

Hemke R, Kuijpers TW, Nusman CM, **Schonenberg-Meinema D**, van Rossum MA, Dolman KM, van den Berg JM, Maas M. *Contrast-enhanced MRI features in the early diagnosis of juvenile idiopathic arthritis*. *Eur Radiol*. 2015 Nov;25(11):3222-9.

Dontje B, van Santen HM, Niermeijer JM, **Schonenberg-Meinema D**, van Trotsenburg ASP. *Anti-TPO antibodies in encephalopathy: diagnostic marker or incidental finding? Auto-immune encephalopathy*. Journal of Pediatric Neurology. 2015 March 9.

Boerma RS, Wit FW, Orock SO, **Schonenberg-Meinema D**, Hartdorff CM, Bakia A, van Hensbroek MB. *Mortality risk factors among HIV-exposed infants in rural and urban Cameroon*. Trop Med Int Health. 2015 Feb;20(2):170-6.

Hemke R, Lavini C, Nusman CM, van den Berg JM, Dolman KM, **Schonenberg-Meinema D**, van Rossum MA, Kuijpers TW, Maas M. *Pixel-by-pixel analysis of DCE-MRI curve shape patterns in knees of active and inactive juvenile idiopathic arthritis patients*. Eur Radiol. 2014 Jul;24(7):1686-93.

Radke J, Pehl D, Aronica E, **Schonenberg-Meinema D**, Schneider U, Heppner FL, de Visser M, Goebel HH, Stenzel W. *The lymphoid follicle variant of dermatomyositis*. Neurol Neuroimmunol Neuroinflamm. 2014 Jul 28;1(2):e19.

Nusman CM, Hemke R, **Schonenberg D**, Dolman KM, van Rossum MA, van den Berg JM, Kuijpers TW, Maas M. *Distribution pattern of MRI abnormalities within the knee and wrist of juvenile idiopathic arthritis patients: signature of disease activity*. AJR Am J Roentgenol. 2014 May;202(5):W439-46.

Nusman CM, van den Berg JM, **Schonenberg-Meinema D**, Maas M, ten Cate R, Kuijpers TW. *Juvenile idiopathic arthritis: from biomarker to treatment*. Ned Tijdschr Geneesk. 2013;157(45): A6391.

Trip HF, **Schonenberg D**, Starreveld JS, Versteegh FG. *An enterovirus epidemic in infants in the summer and fall of 2006*. Eur J Clin Microbiol Infect Dis 2009 May;28(5):469-72.

Schonenberg D, Hartdorff CM, Hoekx JM, *A 4-year-old girl with photophobia and somnolence after a head trauma*. Acta Paediatr 2006 Apr;95(4):496.

Schonenberg D, van Meeteren M, Nelissen RG, van der Horst-Bruinsma IE, Poll RG, Nurmohamed MT. *Thrombosis prevention in orthopedic surgery: clinical practice in the Netherlands in 2002*. Ned Tijdschr Geneesk. 2003 Sep 20;147(38):1856-60.

D. Schonenberg-Meinema, A. van Royen-Kerkhof. Werkboek kinderreumatologie, derde druk, 2014. *Hoofdstuk 14 Juvenile dermatomyositis*.

Under review / accepted for publication

G. Biesbroek, B. Kapitein, I.M. Kuipers, M. P. Gruppen, D. van Stijn, T. E. Peros, M. van Veenendaal, M.H. Jansen, C. van der Zee, M. van der Kuip, E.G.J. von Asmuth, M.G. Mooij, M.E.J. den Boer, G.W. Landman, M.A van Houten, KIRI group****, K.E. van Meijgaarden, T.H.M. Ottenhoff, S.A. Joosten, N. Ketharanathan, M. Blink, C.L.H. Brackel, H. Zaaijer, P. Hombrink, J. M. van den Berg, E.P. Buddingh, T.W. Kuijpers. *Inflammatory responses in SARS-CoV-2 associated Multisystem Inflammatory Syndrome and Kawasaki Disease in children: an observational study*. Accepted in PLoS One. ******Schonenberg-Meinema as co-author KIRI group**

F. Verkuil, R. Hemke, E.C. van Gulik, A.M. Barendregt, A. Nassar-Sheikh Rashid, D. Schonenberg-Meinema, K.M. Dolman, C. den Harder, T.W. Kuijpers, J.M. van den Berg, M. Maas *Double inversion recovery MRI versus contrast-enhanced MRI for evaluation of knee synovitis in juvenile idiopathic arthritis*. Accepted in Insights into Imaging 2022.

S.C. Bergkamp, M. J. Wahadat, A. Salah, T.W. Kuijpers, V. Smith, S.W. Tas, J. M. van den Berg, S. S. M. Kamphuis, **D. Schonenberg-Meinema**. *Dysregulated Endothelial Markers in Systemic Lupus Erythematosus: a Systematic Review and Meta-Analysis*. Under review in Arthr Res Care.

S.C. Bergkamp, V. Smith, T.W. Kuijpers, M. Cutolo, J.M. van den Berg, **D. Schonenberg-Meinema**. *Correlations between capillary density and degree of skin pigmentation in healthy children analysed by nailfold video capillaroscopy* . Under review in Qualitative Imaging in Medicine and Surgery.

Smith EMD, Aggarwal A, Ainsworth J, Al-Abadi E, Avcin T, Bortey L, Burnham J, Ciurtin C, Hedrich CM, Kamphuis S, Levy DM, Lewandowski L, Maxwell N, Morand E, Özen S, Pain CE, Ravelli A, Saad Magalhaes C, Pilkington C, **Schonenberg-Meinema D**, Scott C, Tullus K, Beresford MW. *Treating childhood-onset Systemic Lupus Erythematosus to target: PReS-endorsed Principles and Recommendations from an International Task Force*. Under review in ARD

PORTFOLIO

1. PhD training

General courses

	Year	Workload
Computing in R	2021	2.5
Basis legislation in Science (BROK)	2020 / 2016	2.0
Scientific writing course	2018	1.5
Practical Biostatistics	2016	1.5

Seminars, workshops and master classes

JIR Winter School Management of CTD (virtual)	2022	1.5
JIR Winter School SLE	2018	1.5
Juvenile Systemic Sclerosis Symposium, Hamburg Germany	2016	1.0
Systemic JIA Preceptorship Genova, Germany	2015	1.5
Juvenile Systemic Sclerosis Symposium, Hamburg Germany	2014	1.0
Basis Immunology course Genova, Italy	2013	1.5

Presentations

Lupus-Cora – oral presentation (virtual)	2021	0.5
EULAR – poster (virtual)	2021	0.5
PReS – oral presentation (virtual)	2021	0.5
PReS – Lightening talk (virtual)	2020	0.5
PReS – poster (virtual)	2020	0.5
EULAR study group on Microcirculation	2019	0.5
NVCR assembly scientific meeting	2019	0.5
JIR SLE Winter School – oral presentation	2018	0.5
JIR SLE Winter School – poster	2018	0.5
KAISZ patient organisation – oral presentation	2018	0.5
Amsterdam Kindersymposium – oral presentation	2017	0.5
Young Investigator Meeting (YIM) PReS – 2 posters	2017	0.5
PReS – poster	2017	0.5
Amsterdam Kindersymposium – oral presentation	2015	0.5
Young Investigator Meeting (YIM) PReS – poster	2015	0.5

International conferences

Lupus-Cora (virtual)	2021	1.5
PReS (virtual)	2021	1.5

EULAR (virtual)	2021	1.5
PreS (virtual)	2020	1.5
EULAR Madrid, Spain	2019	1.5
Childhood Arthritis & Beyond Meeting Lake Louise, Canada	2019	1.5
PreS Lisbon, Portugal	2018	1.5
Therapeutic Drug Monitoring, Amsterdam NL	2018	1.5
EULAR Amsterdam, NL	2018	1.5
PreS/YIM Athens, Greece	2017	1.5
EULAR Madrid, Spain	2017	1.5
PreS Genova, Italy	2016	1.5
Autoimmunity Leipzig, Germany	2016	1.5
EULAR/YIM Rome, Italy	2015	1.5

Other

Writing METC-approved study protocol	2015-2016	4.0
Retro-/prospective data collection	2016-present	4.0

2. Teaching**Lecturing**

Verpleegkundigen nascholing SLE	2022	0.5
AIOS cursorisch onderwijs	2021 / 2018	4.0
Studentencollege met patiënt	2019	0.5
Refereeravond SLE	2017	2.0

Tutoring, mentoring

Bachelor student (Ishika)	2022	1.0
Research internship (Rosanne, Sandy, Ilja)	2017/2019/2020	6.0
PhD student (Amara, Sandy)	2018-2022	8.0

ECTS PhD period 2015-2022**Total 73**

3. Parameters of esteem	Year
Grants and Funds	
Zeldzame Ziekten Fonds	2021
Stichting Steun Emma	2016
Expertise centers	
Amsterdam UMC expertise center for juvenile idiopathic arthritis	2016 - present
Amsterdam UMC expertise center for juvenile SLE	2021- present
Amsterdam UMC expertise center for localized scleroderma	2021 - present
Networks and associations	
cSLE T2T International Task Force - member	2021-present
VSOP – national website for expertise network in cSLE	2021-2022
UCAN-CAN-DU study group	2019-present
PReS SLE working party - member	2018-present
EULAR Study Group Microcirculation in Rheumatic Diseases - member	2016-present
NVKR – member	2011-present
NVK – member	2004-present

DANKWOORD

Het overkoepelende thema is van dit proefschrift voor mij is **vrouwen**. De ziekten die zijn beschreven en die ik dagelijks behandel treffen voor het merendeel meisjes en jonge vrouwen. Er was een tijd dat dit juist reden was dat hier minder onderzoek naar werd gedaan, en op veel plekken op aarde is de toegang tot zorg voor zieke meisjes en vrouwen helaas nog steeds niet vanzelfsprekend. Ik heb in de huidige tijd de kans gekregen om mij te ontwikkelen tot kinderarts-reumatoloog en zo iets te kunnen bijdragen aan verbetering. Ik besef me heel goed dat ik dit in vorige generaties niet had kunnen doen als vrouw. Ik ben enorm dankbaar dat ik mijn ambities heb kunnen waarmaken en besef me goed dat ik dit grotendeels heb te danken aan waar mijn wieg stond.

Dit proefschrift is opgedragen ter nagedachtenis aan Zenani en Minou. Ze worden allebei verschrikkelijk gemist. Zenani en Minou waren allebei prachtige jonge tieners die door mij, en door zoveel andere collega's, nooit zullen worden vergeten. Ze namen zelf geen deel aan mijn onderzoek maar zijn wel de reden waarom dit onderzoek nodig is. Ik wil hen extra eren en daarom komen ze ook terug in lay-out door het hele boekje heen. Ik hoop dat ik iets heb kunnen bijdragen aan verbetering van de toekomstige zorg in systemische auto-immuunziekten, zodat niemand meer zal hoeven doormaken wat zij, en hun dierbaren, hebben moeten doorstaan.

Ik wil alle patiënten en hun ouders bedanken voor hun vertrouwen en deelname aan de studies. Dit proefschrift is voor jullie geschreven. Het is zo ontzettend moeilijk om te moeten leven met een chronische ziekte en alles wat daarbij komt kijken, zeker als kind of adolescent. Ik vind het belangrijk dat er oog is voor de mens achter de patiënt en ik hoop dat jullie dit ook zo voelen met regelmatig een lach maar soms ook een traan in de spreekkamer. Ik zal me altijd voor jullie blijven inzetten.

Stichting Emma, familie van Maasdam en het Zeldzame Ziekten Fonds; bedankt voor jullie vertrouwen in mijn onderzoek. Het was zonder jullie financiële steun niet gelukt. Ik ben zo dankbaar dat ik de kans kreeg in 2016 om een pilot-onderzoek te doen en nu jaren later ligt dit proefschrift hier en zijn er vele nieuwe lopende projecten, plannen en ideeën. Bedankt voor jullie steun!

Merlijn: toen ik begon aan mijn fellowship in 2011 zei ik dat ik geen wetenschapper was. Jij zei dat ik binnen 5 jaar gepromoveerd zou zijn. Nu ligt er toch een boekje, niet binnen die 5 jaar, maar die promotie komt er en dat had ik echt nooit gedacht. Ik ben er heel erg trots op en jij hebt me er doorheen gesleept. Af en toe een duwtje, dan weer een luisterend oor en veel advies maar vooral de gunfactor van tijd. Dankzij die maandag

research-dag is het mij gelukt. Je bent echt een maatje, inclusief soms wat gekibbel, maar bovenal iemand die mij door en door kent en begrijpt. Bedankt voor deze kans en route op mijn carrière, en wie weet is dit pas het begin. Ik heb nu zoveel nieuwe ideeën voor nieuwe onderzoeksprojecten, alleen helaas een gebrek aan tijd en vooral aan fondsen. Maar deze onderzoekslijn krijgt vorm en is in een stijgende lijn, we gaan ervoor. En daarna ga ik toch echt helpen om de plastic soep in zee op te ruimen.

Vanessa: de eerste keer dat ik je zag spreken was bij een workshop capillaroscopie tijdens een congres in San Diego in 2013. Ik had toen de eerste abnormale observaties bij capillaroscopie geanalyseerd in mijn SLE-patiënten en nam me voor contact te zoeken voor samenwerking met het idee: handig, want Gent is toch een beetje "om de hoek". Na 1 email was het contact snel gelegd en bezocht ik je in Gent, met mijn toenmalige onderzoeksgegevens. Alles kon direct de prullenbak in, dat was jouw advies: even slikken, maar je had wel gelijk. Als je het doet, dan moet het goed zijn. Hoe dat dan moest, dat heb jij mij geleerd. België bleek soms toch een afstand maar het lukte om samen projecten op te zetten en inmiddels is onze samenwerking niet meer uit ons leven weg te denken. Zonder jou was dit proefschrift niet gelukt, bedankt voor alles.

Taco: bij mijn sollicitatie was ik eerlijk en zei dat ik weinig onderzoek ambieerde. Dat was spannend en voelde een beetje naïef want ik was direct daarna bang om niet te worden aangenomen. Het tegendeel was waar. Je had juist iemand nodig voor patiëntenzorg en ik kon volop aan de slag. Toen kwamen opeens de onderzoeksvragen vanzelf. Vervolgens ligt er hier nu een proefschrift dat echt helemaal door mijzelf is opgebouwd en uitgevoerd en dat is zoveel waard. Je legde geen druk op mij in de snelheid van uitvoering en daar ben ik je dankbaar voor. Ik kon het op mijn eigen tempo doen, en op mijn eigen manier, ook al was dat soms anders dan hoe jij het zag. Bedankt voor steun en bovenal voor al die leerzame klinische lessen, je bent een geweldige dokter en een wandelende encyclopedie.

Rebecca: je stond aan de wieg van de ontwikkeling van kinderreumatologie in Nederland, en ook aan de wieg van mijn carrière in dit vakgebied. Aan het eind van mijn opleiding tot kinderarts kon ik een paar maanden met je meekijken en gaf je me een kans om het zwangerschapsverlof van Petra op te vangen. Het doel was verbeterde samenwerking tussen LUMC en AMC, en dat is ook precies gelukt. Inmiddels werken we nauw samen binnen KRANS en hebben we zelfs een gezamenlijke opleiding, wekelijkse patiëntenbesprekingen maar bovenal zeer korte lijntjes. Precies hoe jij het voor je zag en misschien nog wel meer dan dat. Bedankt voor al je hulp en steun. Je zei tegen me: vindt je eigen niche. Kijk nou, dat is gelukt!

Mario: mijn eerste ontmoeting met jou was toen ik ad hoc binnenliep in een radiologiebespreking om een formulier te laten ondertekenen voor mijn aanstelling in het AMC. Je reageerde hierop heel vriendelijk met een grap waardoor ik me niet meer opgelaten maar direct welkom voelde. Dat bleek een voorbode voor de goede sfeer en samenwerking van de jaren daarna. Altijd tijd nemen voor een spoedbeoordeling en de oprechte interesse in het verhaal van de patiënt achter de MRI-plaatjes. Ook al was de JIA-onderzoekslijn geheel anders dan die van de SLE, het is toch gelukt om die twee samen te brengen in dit proefschrift en jou in mijn promotiecommissie te krijgen. Bedankt voor je oprechte blijheid en interesse.

Sander: onze samenwerking begon tijdens mijn verplichte stage in volwassen reumatologie. Wat vond ik het moeilijk om weer volwassen patiënten te zien na al die jaren kindergeneeskunde. Doodsbang was ik om wat te missen. Jij was altijd een baken van rust, begrip en steun. Precies die kenmerken zie ik nu terug zie als je mijn arts-onderzoeker Sandy spreekt als mentor. Dat is voor jou vanzelfsprekend maar voor een ander zo waardevol in de academische hectiek. Ik ben dankbaar voor de samenwerking die de komende jaren zal groeien door onze gezamenlijke plannen. Want samen is beter; together everyone achieves more.

Ronald: het is een grote eer dat je in mijn commissie zit. Bedankt voor je uitnodiging in de ARC SLE research meetings, je oprechte interesse in waar ik mee bezig ben en navraag hoe het met mij gaat tijdens begeleiding van een levensbedreigend zieke patiënt. Het was even schrikken een paar maanden geleden. Gezondheid is het allerbelangrijkste en dat beseffen we als dokters elke dag. Bedankt voor je voorbeeld dat je hoogleraar kan zijn en nog steeds de mens achter de patiënt ziet. Dat inspireert mij enorm en is precies hoe geneeskunde ooit is bedoeld door Hippocrates.

Annet: dank je wel voor je enthousiasme om in mijn promotiecommissie deel te nemen, het is een eer. Je bent een expert in systemische auto-immuunziekten bij kinderen in Nederland, maar ook je internationale rol en betekenis is groot. Ik vind het spannend wat je van mijn proefschrift zal vinden. We hebben geprobeerd om de samenwerking op te zetten, de bureaucratische muren van UMCU bleken echter voor mij ondoordringbaar en vanwege tijdsgebrek gaf ik het op. Via een andere route met het landelijke CHILL-project hoop ik het alsnog te realiseren om capillaroscopie te kunnen uitvoeren bij de SLE patiënten in Utrecht. Geduld is een schone zaak, bedankt voor je steun.

Alexandre: jaren geleden vroeg ik je om mee te kijken bij een jonge patiënte waarbij ik twijfelde over een overlap-beeld. Dat was nog in de pre-fusie fase van AUMC. Je kwam speciaal hiervoor naar locatie AMC en dat waardeerde ik heel erg. De voor mij grappigste

herinnering is toen je zei dat mijn proefschrift bijna af was: die inleiding en discussie schrijf je in een weekend, die leest toch niemand. Je had helemaal gelijk, dat klopt. Behalve de commissie en daar zit je nu in. Ik hoop dat je het met interesse hebt gelezen en ik weet zeker dat we een mooie gedachten wisseling zullen hebben met al jouw ervaring in dit vakgebied. Ik kijk uit naar een goede samenwerking de komende jaren op locatie AMC. Ter info: er komen vele transitie patiënten aan.

Amara: je meer dan welkome komst (in een zeer moeilijke tijd) en helaas ook weer vertrek in onze vakgroep ging gepaard met tranen. Toen we definitief dat nieuws hoorden vielen we elkaar huilend in de armen. Een dierbaar tijdperk was voorbij. We waren jarenlang een team waarin we elkaar maar hoefden aan te kijken of we wisten wat de ander dacht. Vanzelfsprekend hadden we ook direct een band doordat onze kinderen precies dezelfde leeftijd hebben en het ploeteren was in die tropenjaren. Ik wil je dolgraag helpen om dit gevoel, van een geprint proefschrift dat voor je ligt, ook te ervaren. Je weet dat ik er altijd voor je ben, let's go for it! En in de tussentijd gaan we weer genieten van terrasjes tijdens congressen nu het eindelijk weer kan.

Sandy: wat ben ik blij dat je reageerde op het berichtje op StudentBoard jaren geleden, nota bene voor het onderzoek van Amara. Je bent nu niet meer weg te denken uit mijn SLE onderzoek. Ik zeg het te weinig maar je wordt zo enorm gewaardeerd en ik ben zo trots op je ontwikkeling. Je bleef vertrouwen hebben in ons als team, ook zelfs met een paar maanden zonder salaris. Je vertrouwen werd beloond, uiteindelijk kwamen er gelukkig echt (sporadische) fondsen binnen om je salaris te betalen. Ik heb midden in jouw promotietraject een moeilijke tijd gehad en je had alle begrip terwijl je het zelf ook moeilijk had, en hebt, met zieke dierbaren. Jouw promotie is de volgende en nogmaals, ik ben zo trots op je! Jij bent mijn eerste PhD student en ik hoop dat je snel de tweede mag inwerken voordat je aan de opleiding begint. Geloof in jezelf en wees trots, je kan het.

Ingrid: jij bent een onmisbare schakel in ons team. Het is zo fijn om jou als collega te hebben. Doortastend, menselijk en kundig. De laatste jaren klaar voor een nieuwe uitdaging die je met volle overtuiging hebt aangenomen en zoveel zal betekenen voor de resultaten van ons internationale research project voor jeugdreuma. Wie weet kan je voor mijn SLE project in de toekomst hetzelfde doen, eerst zal ik hopelijk fondsen werven en dan weet ik waar de expertise zit. Hou me scherp in de chaos!

Mariken: ik heb zoveel respect voor jou, je bent een alleskunner. Wat een dappere stap om later in je carrière voor een subspecialisatie te gaan, en ik ben zo blij dat je dat deed. Jouw visie op hoe patiëntenzorg hoort te zijn klopt precies met hoe Merlijn, Ingrid en ik

dat zien. Je nam een rol aan als fellow op een leeftijd die ongebruikelijk is met al jouw ervaring en expertise op andere vakgebieden waar wij ook weer van kunnen leren. Dit viel samen tot kinderreumatologische zorg zoals het moet zijn. Dus: don't ever leave us.

Marja Pannekoek: je weet het zelf niet maar jouw observaties van 'extravasaties' liggen ten grondslag aan dit proefschrift. Dit wekte namelijk mijn nieuwsgierigheid en heeft geleid tot dit boekje en tot inmiddels een volledige onderzoekslijn. Je werkt niet meer in het AMC maar ik zal je opsporen om je persoonlijk te kunnen bedanken.

Dear Maurizio: your atlas of capillaroscopy was the basis of my research. The findings of my studies in cSLE were sometimes surprising for you (no, these patients did not have overlap disease!). Thank you for being a part in my learning process and I hope that I can add some more knowledge to a future atlas.

Beste KIRI collega's Martijn, Dasja, Marceline, Michael en Henriette. We zijn 1 vakgroep maar werken ook veel gescheiden. Toch horen we echt bij elkaar. Bedankt voor jullie steun en feedback over de jaren waarin ik kon groeien als fellow naar volwaardig staflid met een eigen research lijn. Martijn: jou wil ik extra bedanken voor je steun voor onze kinderreumatologie-groep. Volledig dankzij jou heb ik een fonds gekregen van het zeldzame ziektefonds. Hoop nog lang tegen je aan te mogen zeuren als kamergenoot, en vice versa natuurlijk.

Ik wil vanzelfsprekend al mijn collega's bedanken in het Emma Kinderziekenhuis en dat zijn er teveel om op te noemen. Toch wil ik nog extra de kinder nefrologen benoemen. Tonny, Michiel, Rik, Arend en Jaap: wat hebben we een fijne samenwerking voor onze patiënten. We leveren topzorg op maat met onze combinatie-consulten en we zijn niet voor niets expertisecentrum geworden.

Pina (aka Jennifer): heldin. In je eentje de kinder dermatologie op zo'n hoog niveau uitvoeren. We gingen met Amara samen naar Gent en kwamen als garnalenvrouwtjes terug. Hopelijk lukt het om samen de zorg voor onze sclerodermie patiënten nog verder te verbeteren. Het is een groot voorrecht om met jou te werken en ik hoop dat nog heel lang kan doen. Blijf en hou vol!

Ook wil ik specifiek enkele oud-'verdiepers' noemen. Beste Koen, Daniel, Giske, Daan, Veronica, Cornelis, Laurens, Anne, Daria, Ilan en Marein: door jullie kon ik mijn wetenschaps-dagen ook echt efficiënt gebruiken. Het is echt mede door jullie dat dit proefschrift er daadwerkelijk ligt, ik ben jullie heel erg dankbaar. Het was ook erg gezell-

lig en leuk om te zien hoe jullie allemaal werden gegrepen door de kinderreumatologie, wat een fantastisch vak he?

Alle balie- en secretariaat-medewerkers, we werken door drukte zo vaak langs elkaar heen en daarom uit ik mijn waardering voor jullie onvoldoende, zeker in een fase waarin ik het erg zwaar had. Katenka, Simone W, Hester, Lilian, Anja, Madeleine, Simone H, Inez, Sabine, Mirthe en Monique: jullie zijn echt onmisbaar in het dagelijkse reilen en zeilen van onze afdeling, bedankt voor alles.

Hans van Goudoever: bedankt voor je persoonlijke bericht en aandacht in een zware periode. Dat heb ik echt enorm gewaardeerd.

Karin en Amber uit Gent: mijn ogen en mond vielen open op deze gespecialiseerde sclerodermie poli's waar ik altijd welkom was om mee te kijken. Ik heb er zoveel van geleerd, alle expertise in de uitvoering van capillaroscopie en Rodnan skin score heb ik van jullie geleerd. Wat fijn dat we konden samenwerken in projecten waarvan er 2 manuscripten in dit proefschrift staan. Bedankt voor al die leermomenten en steun.

Petra (Leiden) en Joost (Utrecht): ik wil jullie apart noemen want jullie hebben mij de richting van kinderreumatologie op gewezen die zo goed bij me past. Ik ben daar erg dankbaar voor want ik had het anders nooit overwogen. Het is nog extra leuk omdat we nog steeds zoveel kunnen samenwerken. Sylvia, ik herinner me een etentje op een koude winteravond bij Merlijn thuis waarin we hebben uitgesproken volledig samen te gaan werken met onze cohorten, en zo geschiedde. Bedankt dat je me meeneemt in je projecten, fondsaanvragen en netwerken. Hoe je alle ballen hoog houdt is mij een raadsel, ik ben zo blij met je steun. Alle overige collega's binnen de kinderreumatologie: Leontien, Wineke, Elizabeth, Esther, Ellen, Nico, Bas, Marc, Simone, Gijs, Danielle, Marleen en Amani: het is kostbaar dat we zo close samenwerken op landelijk niveau. Laten we dat goed vasthouden en verder uitbouwen voor al die zeldzame ziektebeelden die we behandelen.

Javad: je bent nu al professor in de dop. De ene na de andere onderzoeksprijs sleep je binnen, en terecht. Wat leuk en fijn dat we konden samenwerken, en nieuwe projecten zijn al gaande. Sterkte met afronding van ook jouw proefschrift binnenkort en succes met de opleiding tot kinderarts. Terecht dat je bent aangenomen en zo leuk dat je een collega wordt. We houden ongetwijfeld veel contact.

Iedereen van het 'radiologie-clubje' Robert, Charlotte N, Anouk, Charlotte vG en Floris. Bij jullie kon ik het een beetje afkijken, hoe het was om te promoveren. In het begin had

ik jullie tips hard nodig voor SPSS en statistiek. Het is extra leuk dat er een hoofdstuk over MRI in dit proefschrift kon worden opgenomen, zo is de cirkel weer rond.

Lieve Vu-ers: Jochim, Jikke, Kirsten, Evelien, Maartje, Esther, Eline en Brigitte. Velen van jullie gingen me voor in het promoveren, nu ben ik aan de beurt en ik had dat nooit gedacht. Bedankt voor jullie fijne en gezellige vriendschap tijdens onze studietijd.

Mijn carrière begon in Gouda en ik wil alle kinderartsen en verpleegkundigen van toen bedanken voor de leerzame tijd die een solide basis was voor mijn verdere carrière. De eerste keer reanimeren, opvang van een nieuwe diabeet, sepsis of ernstige uitdroging, met Ria als doorgewinterde super-verpleegkundige konden we alles aan. Die eerste stappen in de kindergeneeskunde waren nog op de oude Jozef locatie en ik had toen nooit kunnen dromen wat ik nu heb bereikt. Bedankt Cecil, Jurriaan, Ed, Dieneke 2 (ik was eerst!), Florens, Jeannette en Sander: zonder PORG was dit nooit gelukt, deze is ook voor jou! Lieve Judith, Esther, Helene, Minke, Hilde, Caroline, Hans en Michael wat waren we een leuk en goed team. Zo blij dat jullie mijn eerste collega's waren.

Alle collega's van de 'volwassen' reumatologie zoals wij dat noemen, bedankt voor de samenwerking, en een speciaal dankwoord voor Marieke en Liesbeth. Bij jullie weet ik zeker dat onze SLE-patiënten na de leeftijd van 18 jaar in goede handen zijn. Ik hoop nog lang samen te kunnen werken aan goede zorg voor deze kwetsbare jongeren.

Aan de trainsters van Mom in Balance, vooral aan Esther, Petra en Jorien: dankzij jullie bleef ik echt topfit. Het inspireerde me tot vaker sporten, halve marathons lopen en inmiddels een half fitness centrum in de tuin. Een fit lichaam doet veel voor een gezonde geest. Bedankt voor jullie enthousiaste en aanstekelijke trainingen!

Lieve vrienden "uit het dorp": Leonie, Barbara en Ivar, Maartje en Arjen, Marike en Dennis, Jacqueline en Rogier, Mirjam en Paul, Agnes en Marcel, Nicole en Sven, Manon en Duco, Joyce en Dave, Kim, Patrick, Diana en Jacob, Meike en Karel, Pauline en Onno, Ingrid en Michiel, allereerst bedankt aan iedereen die toestemming gaf om hun kinderen te onderzoeken als gezonde controle voor mijn capillaroscopie onderzoeken. Alle gegevens die daaruit kwamen bleken heel erg veel waard. Met schoolgaande kinderen in een nieuw dorp zijn er binnen no-time vele hele dierbare nieuwe vriendschappen geboren die inmiddels onmisbaar zijn. Op 9 december gaan we eindelijk weer eens een flink feestje vieren. Ik hoop dat er nog vele feestjes volgen met jullie.

Lieve jaarclub vriendinnen Hester, Kim, Meike, Suzan, Merle en Anna: al 25 jaar een trouwe vriendschap, door weer en wind, met meestal een schaterlach maar soms ook

met een traan. We hebben al vele levensfasen met elkaar doorlopen. We blijven trouw aan onze reisjes die inmiddels door geheel Europa gingen, misschien moeten we inmiddels meer intercontinentaal gaan denken. Ook jullie gaven toestemming om jullie eigen kinderen te onderzoeken. Terwijl jullie tijdens ons weekend weg lekker samen aan het borrelen waren, was ik helaas als een monnik aan de keukentafel hard aan het werk met mijn apparaat, maar het was het waard. Jullie zijn voor mij HET voorbeeld dat onze generatie vrouwen veel kansen kan pakken voor een goede carrière. We zijn allemaal hardwerkende vrouwen die deze kansen met beide handen hebben aangegrepen om expert te worden in hun eigen vakgebied. Dit zo'n groot verschil met de generaties voor ons. Nu is het tijd voor feest, en als er iemand weet hoe je moet feesten dan zijn jullie het wel. Let's go girls!

Kate en Berry: jullie vriendschap is alles waard. Jullie kunnen naar de andere kant van de wereld verhuizen maar onze band blijft toch ijzersterk. We konden leef en leed nog steeds intens met elkaar delen, was het niet via beeld dan toch wel met jaarlijkse vakanties "halverwege". Wat waren we blij dat jullie eindelijk echt naar Nederland terugkwamen, en het moment waarop kon ook niet beter. Laten we er een feestje van maken zoals in Shanghai, het dak mag eraf!

Lieve Caroline, Deborah en Hester: alle drie zo anders maar mij stuk voor stuk zo dierbaar met een hechte vriendschap al vanuit het begin van onze studententijd. Daarna alle mijlpalen van ons leven samen doorlopen. Door die tropenjaren met jonge kinderen zien we elkaar niet zoveel als daarvoor maar onze vriendschap is tijdloos en rotsvast. Jullie waren een grote reddingsboei toen ik jullie zo hard nodig had. Bedankt voor al die jaren vriendschap, jullie zijn voor mij onmisbaar.

Lieve schoonfamilie, opa en oma Politie, opa en oma Garage, Gladys en Riccardo: bedankt voor jullie steun en interesse. En voor die eindeloze jaren oppassen natuurlijk. Mede daardoor kon dit proefschrift tot stand komen. Wat een fijne familie kreeg ik erbij door met Aloys te trouwen. Ik ben heel dankbaar dat jullie erbij kunnen zijn op dit belangrijke moment voor mij op 9 december.

Ik wil graag ook dank uitspreken naar mijn oma's, die er niet meer zijn. Jullie kregen niet de kansen die ik wel kreeg. Ambitie ging verloren en bij een van jullie maakte dat plaats voor frustratie en depressie. Jullie moedigden me altijd aan om goed mijn best te doen op school. Ik besef me goed dat ik op de schouders sta van reuzinnen van de generaties voor mij. Ik keek bewonderend toe hoe mijn oudere nichten Heleen en Nicoline gingen studeren aan de universiteit, dat werden mijn voorbeelden, met dit als resultaat. Hopelijk kunnen jullie toch meekijken, dat is een troostende gedachte.

Lieve Mindy, sommige mensen zijn familie maar anderen *worden* familie. Je bent zo'n belangrijk onderdeel van ons gezin, niet alleen praktisch als reddende engel maar ook qua emotionele band. Onze kinderen houden zielsveel van je en jij van hen. Je gedachten over opvoeding sluiten volledig aan bij die van ons. Dankzij jou had ik de rust om dit proefschrift te kunnen maken. Ik kan me geen leven zonder jou voorstellen.

Lieve Maurits, 'Maus': ik hoop dat we je voldoende kunnen steunen. Wat fijn dat je volledig onderdeel bent van ons gezin. Het betekent heel veel voor ons allemaal. Ik verloor een zusje maar ik kreeg er een broer bij. Benieuwd of je nog rekenfoutjes gaat vinden in dit boek, het zou mij niets verbazen.

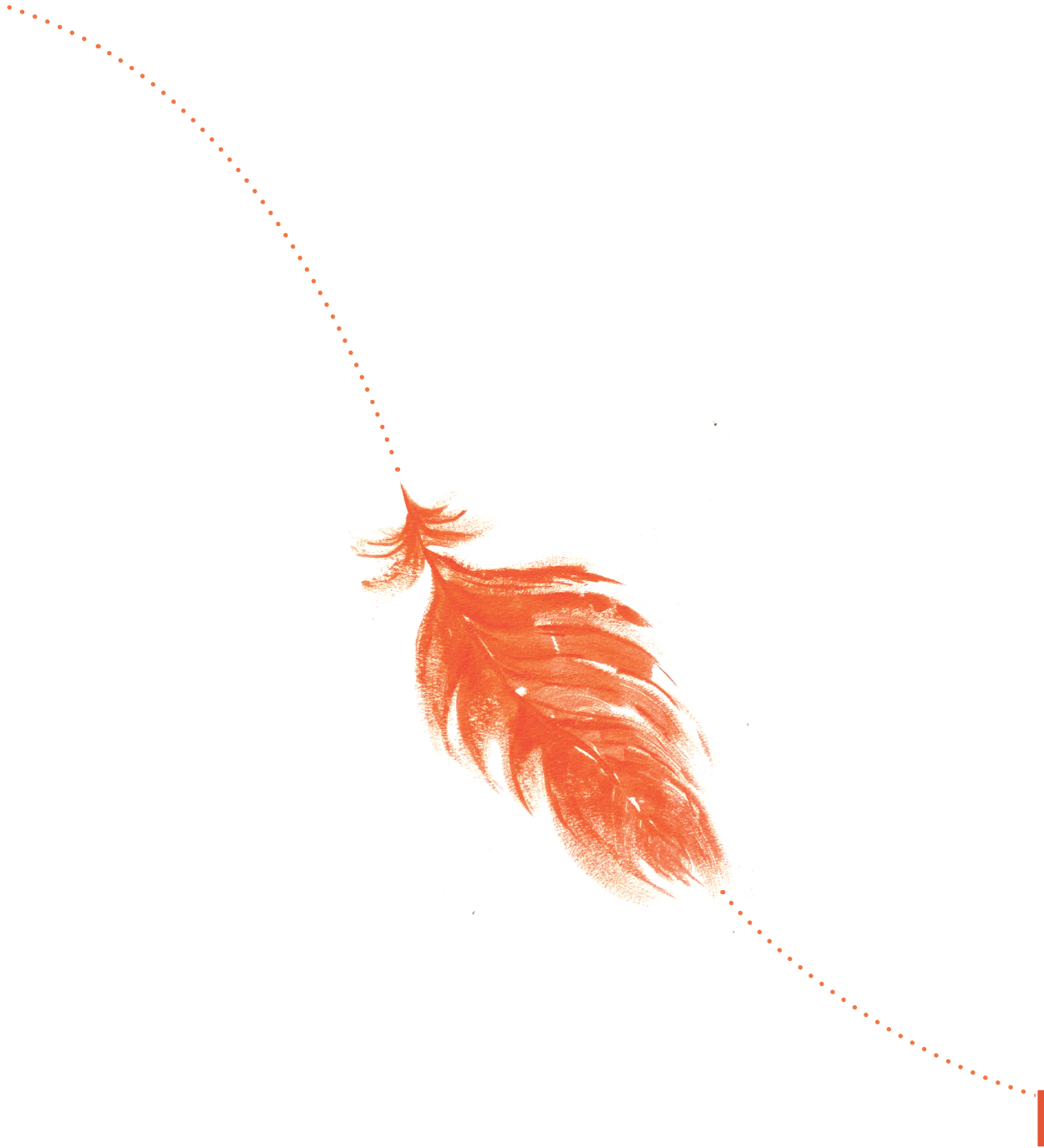
Lieve Marije en Bas! Mijn paranimfen, wat een woord he. Het voelt goed om daar straks te staan, als een blok, met zijn drieën. Ik zal daar zoveel steun aan hebben. Het is zoveel rijkdom om een grote familie te hebben. Door het leeftijdsverschil waren we soms met volledig andere levensfasen bezig, als tieners gingen we onze eigen weg maar nu is dat anders. Onze beroepskeuze is divers maar de gemeenschappelijke eigenschap is de serieuze en volhardende aanpak in ons werk. En Eva, als schoonzus pas je daar dus ook perfect tussen! We zien onze kinderen samen opgroeien en onze groepsapp wordt dagelijks druk gebruikt. Het meest ondenkbare gebeurde in Maart 2020 maar samen staan we sterk. Ik hou van jullie!

Lieve papa met je eindeloze oneliner quotes waarmee je ons hebt opgevoed: 'brutalen hebben de halve wereld'. 'Nee heb je, ja kan je krijgen'. 'Begrijp je het niet, verwonder je dan'. 'Niet klagen maar dragen' (het stukje bidden voor kracht liet je weg). 'Soms gaat het niet om wat je kan, maar wie je kent'. Je hebt me geleerd wat doorzettingsvermogen is en dat hard werken loont. Hier ligt het resultaat. Lieve mama, altijd met de vleugels beschermend om ons heen om het kwaad van de wereld buiten de deur proberen te houden. Zo moeilijk, met al die reislustige kinderen. Je staat altijd klaar voor ons, wil ieder pijntje of verdriet overnemen. Wat bijzonder dat je de omslag hebt kunnen ontwerpen. Je vond het heel spannend (want ik heb mijn perfectionisme niet van een vreemde) maar het is zo mooi en persoonlijk geworden. Ik ben trots en hopelijk ben jij dat ook! Jullie zijn zelf opgegroeid in een traditioneler gezin uit een hele andere tijd, eentje waar een vrouwelijke carrière niet werd nagestreefd. Jullie hebben ons wat anders meegegeven, en stimuleerden ons allemaal om ons eigen pad te zoeken. Al jaren is de vrijdag jullie oppasdag, dat begon 14 jaar geleden met jullie eerste kleinkind en is tot de dag van vandaag doorgegaan. Het is zo waardevol dat jullie zo'n sterke band opbouwen met alle kleinkinderen. Ik ben dankbaar voor mijn fijne jeugd en ook voor het feit dat jullie er altijd zijn voor ons, wat er ook gebeurt. Ik ben echt heel trots op jullie!

Allerliefste Willemijn: we missen je zo erg. Het blijft onwerkelijk en het zal nooit wennen. Met de veertjes zien we je terug, het is heel troostend om dat echt zo te voelen. Wat zou je trots op mij zijn geweest, je kon zo genieten voor een ander. Zelfs in de laatste periode had je altijd interesse voor anderen, in wat zij voelden of beleefden. Onze levensles is de enorme dankbaarheid die je uitstraalde waardoor we met gevoelens van geluk en rust het verdriet enigszins lijken te kunnen dragen. We zullen op dit soort dagen van mijlpalen de leegte weer extra voelen. Bij een concert van The Script waar we samen waren zong je huilend mee: "turn the pain into power". Ik heb het laten terugkomen in mijn stellingen en er naar geleefd de afgelopen jaren. Een van de vele laatste dingen die je tegen me zei was: wees lief voor jezelf. Wat kende me je goed. Ik zal genieten, en ik zal aan je denken en jou zo missen maar visualiseren dat je erbij bent met je stralende sprankelende lach.

Mijn lieve prachtige kids, ik hou zielsveel van jullie en vond het elke keer zo moeilijk om na zwangerschapsverlof weer te gaan werken, maar alles kwam goed. Ik ben zo benieuwd naar jullie toekomst maar ik geniet van elke fase. Ik heb de eer om jullie te zien opgroeien tot nu al zulke zelfstandige en sociale personen vol energie (net als papa). Ieder met uniek karakter en ook samen echt een team (met steeds minder ruzie). Het is uniek dat jullie bij mijn promotie kunnen zijn, dat is dan weer een groot voordeel dat ik er zo lang over heb gedaan. Mijn werk vergt soms veel energie en tijd maar jullie zijn zo lief en begripvol en snappen mijn ambitie. Siebe: alleskunner, doorzetter en zo lief en begripvol voor een ander. Het is altijd EN EN in plaats van OF OF, net als je moeder. Dat gaat je ver brengen, maar vergeet niet af en toe uit te rusten! Douwe: kleine grappige netwerker van me. Jij kent iedereen en iedereen kent jou. Ik probeer te leren van jouw relativiseringsvermogen. Je moet niet overal je spullen vergeten maar verander verder niet! Mijntje: blij pittige kletskaus. De creatieve wereld ligt aan je voeten, daar ga jij zeker je eigen weg in vinden. Blijf altijd vragen stellen zoals je nu doet. Je bent altijd bezig met creëren en hebt een talent voor presenteren en theater. Je kan niet wachten tot je groot bent maar blijf nu nog maar even ons kleine lieve meisje.

Aller- aller- allerliefste Aloys, jij bent alles voor mij. Mijn baken van rust, begrip en steun. En dat zeg ik je veel te weinig. Je weet altijd het juiste te zeggen in zware tijden. Je gunt mij alles en hebt in je carrière bepaalde keuzes gemaakt om te zorgen dat ik mijn ambities kon verwezenlijken. Dat was een generatie voor ons ondenkbaar geweest. Zonder jouw constante steun en vertrouwen had ik allang de handdoek in de ring gegooid om dit proefschrift af te ronden. Je bent nog trotser op mij dan ik op mezelf en dat is zo ontzettend lief. Ik prijs mezelf zo gelukkig dat je mijn levensmaatje wil zijn en dat nog heel lang! Want weet je nog: dat is de afspraak. Ik hou van je tot de maan en terug, en dan nog meer.



ABOUT THE AUTHOR

Dieneke Schonenberg-Meinema was born on 23 February 1977 in Rijnsburg where she grew up as the oldest of 4 siblings. In 1995 she completed high school at the Rijnlands Lyceum in Oegstgeest. From 1995-1997 she first studied 'Medical Biology' for two years at the 'Vrije Universiteit' in Amsterdam. In 1997 she started to study Medicine. In 2004, she graduated cum laude (with honor). She did the last internship in a rural hospital in Cameroon where she also started the foundation 'Give Milk Stop Aids' with one of her best friends Caroline. This foundation has helped many families with HIV-positive mothers during a decade. In 2004 she started her first job as a resident in pediatrics in the 'Groene Hart Ziekenhuis' in Gouda after which she specialized in pediatrics in the 'Leids Universitair Medisch Centrum'. In 2011, she started a fellowship Pediatric Rheumatology in Amsterdam UMC. This specialization was finished in 2016, from that time up till present she is a staff member of the Emma Children's Hospital from Amsterdam UMC.

In 2015 she observed abnormalities in her patients with nailfold capillaroscopy. She reached out for collaboration with experts in this field from Belgium and Italy. This subsequently led to the beginning of many research projects. In 2016 she received a research grant from the Emma Foundation for a pilot study. In 2017 her longitudinal cohort study proposal was approved by the medical ethical committee of Amsterdam UMC. The first results are written in this thesis, a second PhD student is working on the following ongoing projects. In 2018 Dieneke started a very fruitful collaboration with Sylvia Kamphuis, a pediatric rheumatologist from Erasmus MC Rotterdam. They decided to work closely together and work towards a national database of these patients which has been executed in 2022.

Dieneke is married to Aloys Meinema since 2007, together they have three children: Siebe (2008), Douwe (2010) and Mijntje (2013) and two cats Queen W (2019) and Winston Churchill (2021). In her free time she likes to spend time at the dinner table with friends and family or look at the sport activities from her children. She also likes to do running, fitness, tennis and skiing and loves to travel with her family, especially to Asia. Her youngest sister passed away in March 2020 due to metastatic breast cancer, at the young age of 33. She would say: "Vier het leven zonder spijt", which is the last quote of this PhD thesis, in her reminiscence.

