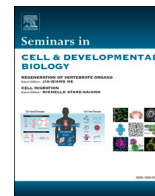




Contents lists available at ScienceDirect

Seminars in Cell and Developmental Biology

journal homepage: www.elsevier.com/locate/semcdb

Maternal selection of human embryos in early gestation: Insights from recurrent miscarriage

Jan J. Brosens^{a,b,*}, Phillip R. Bennett^c, Vikki M. Abrahams^d, Rosanna Ramhorst^e, Arri Coomarasamy^f, Siobhan Quenby^{a,b}, Emma S. Lucas^a, Rajiv C. McCoy^g^a Division of Biomedical Sciences, Warwick Medical School, University of Warwick, Coventry, UK^b Tommy's National Centre for Miscarriage Research, University Hospitals Coventry & Warwickshire NHS Trust, Coventry CV2 2DX, UK^c Tommy's National Centre for Miscarriage Research, Imperial College London, UK^d Department of Obstetrics, Gynecology & Reproductive Sciences, Yale University, New Haven, CT, USA^e CONICET, Universidad de Buenos Aires, Instituto de Química Biológica de la Facultad de Ciencias Exactas y Naturales IQUBICEN, Buenos Aires, Argentina^f Tommy's National Centre for Miscarriage Research, Institute of Metabolism and Systems Research, University of Birmingham, UK^g Department of Biology, Johns Hopkins University, Baltimore, Maryland 21218, USA

ARTICLE INFO

Keywords:

Miscarriage
Embryo
Aneuploidy
Endometrium
Decidualization
Placenta
Selection

ABSTRACT

Compared to most mammals, human pregnancy is unusual in that it involves chromosomally diverse embryos, cyclical breakdown and regeneration of the uterine mucosa, and intimate integration of fetal and maternal cells at the uteroplacental interface. Not surprisingly, pregnancy often falters in early gestation. Whether these losses result in clinical miscarriages depends on the origins and impacts of chromosomal errors on fetal development and the ability of the decidualizing endometrium to engage in embryo biosensing and selection. Aneuploidy originating in oocytes during meiosis drives the age-related risk of miscarriage. By contrast, the frequency of endometrial cycles with an impaired decidual response may account for the stepwise increase in miscarriage rates with each pregnancy loss independently of maternal age. Additional physiological mechanisms operate in early gestation to ensure that most failing pregnancies are lost before vascular maternal-fetal connections are established by the end of the first trimester. Here, we summarise how investigations into the mechanisms that cause miscarriage led to new insights into the processes that govern maternal selection of human embryos in early gestation.

1. Introduction

Pregnancy begins when the implanting embryo breaches the uterine mucosa. Once the conceptus embeds in the endometrial stroma, maternal serum and urine levels of embryo-derived human chorionic gonadotrophin (hCG) rise, typically around 7–9 days after ovulation [1]. Based on sensitive urine hCG measurements, several prospective studies reported miscarriage rates of approximately 30% in young, healthy women trying to conceive [1–3]. Many losses, however, occur soon after implantation and escape detection. The population prevalence of women with one, two or three or more self-reported miscarriages is 10.8%, 1.9%, and 0.7%, respectively [4]. More than 92% of recognised

miscarriages occur before 12 weeks of pregnancy [5].

Two independent risk factors, maternal age and the number of previous pregnancy losses, have disproportionate effects on the miscarriage rate [5,6]. The age-specific risk of miscarriage follows a J-shaped curve [5,7]. The risk is significantly higher in adolescents, flattens between the ages of 20 and 34 years, and rises again sharply in women over 34 years old (Fig. 1 A). Fetal chromosomal errors, or aneuploidies, are the primary driver of age-specific miscarriage risk. The J-shaped curve of age-related miscarriages mirrors the incidences of meiotic chromosome errors in oocytes [8], preimplantation embryos [9,10], and fetal tissues [11,12] (Fig. 1A). The impact of previous pregnancy losses on miscarriage rates is also well documented but poorly understood. Several large

Abbreviations: aPL, anti-phospholipid antibodies; APS, antiphospholipid syndrome; BMPC, bone marrow-derived progenitor cells; hCG, human chorionic gonadotrophin; KIR, killer immunoglobulin-like receptors; HLA-C, human leukocyte antigen-C; PGT-A, pre-implantation genetic testing for aneuploidy; PSSC, precocious separation of sister chromatids; uNK cells, uterine natural killer cells.

* Correspondence to: Division of Biomedical Sciences, Warwick Medical School, University Hospitals Coventry and Warwickshire, Coventry CV2 2DX, United Kingdom.

E-mail address: J.J.Brosens@warwick.ac.uk (J.J. Brosens).

<https://doi.org/10.1016/j.semcb.2022.01.007>

Received 15 November 2021; Received in revised form 21 January 2022; Accepted 21 January 2022

Available online 31 January 2022

1084-9521/© 2022 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

epidemiological studies have documented a stepwise increase in the recurrence risk of miscarriage by approximately 10% with each additional loss (Fig. 1B), independently of maternal age and other co-variables [5,6].

All clinical definitions of recurrent miscarriage are based on an arbitrary number of consecutive or non-consecutive clinical pregnancy losses, usually two or three [13]. Consequently, many studies take a binary approach that does not account for the marked differences in prognoses between individual patients. Numerous pathological mechanisms are invoked to explain recurrent miscarriage [13,14], although none provide a robust explanation for the stepwise increase in recurrence rate with each additional miscarriage. It is important to emphasize that multiple miscarriages do not preclude a successful pregnancy. Even after four consecutive losses, it remains more likely that a subsequent pregnancy will succeed than fail for women under the age of 36 years (Fig. 1B). Thus, a pivotal challenge is to define the mechanisms that account for recurrence of miscarriage without precluding the possibility of a successful pregnancy.

This review starts with an overview of different paradigms of early pregnancy loss. We then summarise the mechanisms accounting for fetal aneuploidies, beginning with gametes and pre-implantation embryos, and assess their contributions to the recurrence risk of miscarriage. Next, we discuss how recent insights into the dynamic nature of embryo-endometrial interactions at implantation have highlighted the importance of maternal checkpoint failures in determining the recurrence risk of miscarriage. Finally, we focus on the pathogenic pathways that converge on the uteroplacental interface in early pregnancy and explore the role of local immune cells.

2. Miscarriage paradigms

The clinical approach to miscarriage is based on the general principle that all physiological processes in the body depend on cooperation between different cell populations in tissues and organs. Early pregnancy loss, discerned or not, is therefore attributed to either inborn errors (i.e., embryonic aneuploidies) or extrinsic pathological processes purported to interfere with the cooperative interactions between embryonic and maternal cells. Consequently, it is common practice to test recurrent miscarriage patients for a host of subclinical disorders, including endocrine perturbations, structural uterine anomalies, thrombophilia, immune disorders, pathogenic mutations, and lifestyle factors [13,14]. The test results then guide treatment. While this approach appears sensible, two vexing problems remain unresolved. First, no modifiable risk factors are identified in over 50% of couples affected by recurrent miscarriage [13,14]. Second, there is limited or no evidence that treatments targeting perceived risk factors of miscarriage improve the prognosis for patients, despite numerous clinical trials [14]. Even in the presence of overt chronic disease, including autoimmune disorders, the

incidence of miscarriage is largely unaffected [15].

A different perspective on miscarriage emerged from the parent-offspring conflict hypothesis [16], which posits that the interactions between genetically distinct fetal and maternal cells at the mammalian uteroplacental interface are driven by conflict as much as cooperation [17,18]. Conflicts arise because embryonic genes are evolutionarily selected to maximise the chance of implantation whereas maternal genes evolve to minimise the risk of continued investment of resources in a failing pregnancy or in supporting a fetus with low fitness [18]. This hidden evolutionary tug-of-war between maternal and embryonic genomes is credited for not only the remarkable diversity of placental structures amongst mammals but also the emergence of unique reproductive features [19]. A good example is the vast array of complex chromosomal imbalances in human embryos, many of which do not preclude successful pregnancy [20,21]. A peculiar maternal reproductive feature, confined mainly to humans and other simians, is the cyclical shedding of the upper endometrial layer at menstruation [22]. Further, comparative studies of placental mammals showed that human trophoblast cells penetrate the uterine wall very deeply, remodel uterine arteries extensively, and induce a marked maternal cardiovascular response [22,23]. Uterine perfusion increases from 45 mL/minute during the menstrual cycle to 750 mL/minute at term [24]. Maternal energy expenditure over the duration of pregnancy is close to physiological limits and comparable to that of endurance athletes [25].

Thus, human pregnancy uniquely starts with the implantation of a genetically distinct embryo in a freshly regenerated endometrium and leads to the formation of a deeply invading placenta capable of radically altering maternal physiology. At a glance, this reproductive strategy seems reckless. However, several physiological ‘checkpoints’ in early gestation ensure that most pregnancies involving a low-fitness conceptus or an inadequately prepared endometrium fail before the onset of placental perfusion at around 12 weeks (Fig. 2A) [26]. The earlier the pregnancy loss, the smaller the impact on maternal fitness [18,27]. Thus, the parent-offspring conflict paradigm posits that the high rate of pregnancy loss soon after conception reflects, at least partly, a robust maternal implantation checkpoint. It predicts that failure of this initial checkpoint will lead to more clinically recognised pregnancies but also more miscarriages.

3. Genetic instability in human embryos

Many human embryos perish during early development, especially prior to the blastocyst stage [28]. Given that early embryonic mortality often coincides with major waves of embryonic genome activation prior to blastocyst formation, genetic causes have been suspected. While direct evidence remains elusive, recent statistical modelling suggests the occurrence of ~0.3–0.4 lethal or nearly lethal *de novo* point mutations per potential human zygote [29], while others have proposed that the

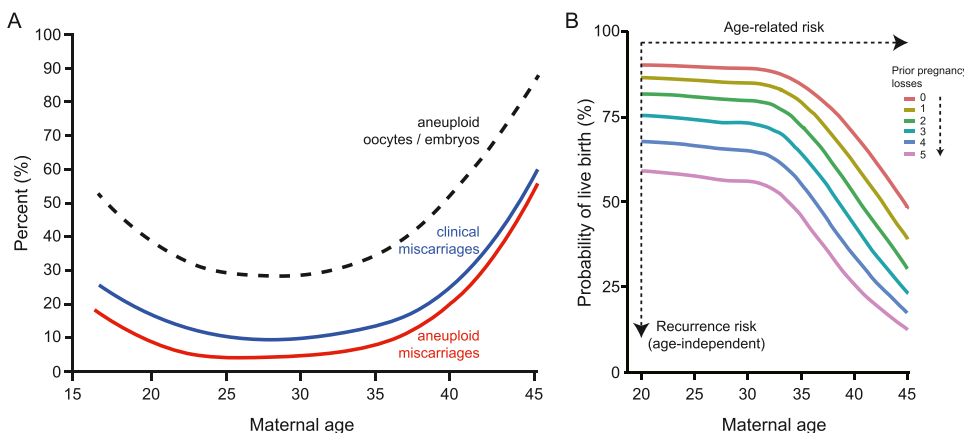


Fig. 1. Risks of miscarriage. (A) Composite graph showing risk of miscarriage according to maternal age. Superimposed are the age-dependent incidences of oocyte/embryo aneuploidy and aneuploid miscarriage. The composite graph is based on data extracted from several studies [5, 7–9]. (B) Miscarriage rates increase with maternal age and with each additional miscarriage. The term ‘recurrence risk’ of miscarriage denotes the stepwise increase in miscarriage rates, independently of maternal age, interpregnancy interval, or a previous live birth. Adapted from Kolte and colleagues [6], with permission.

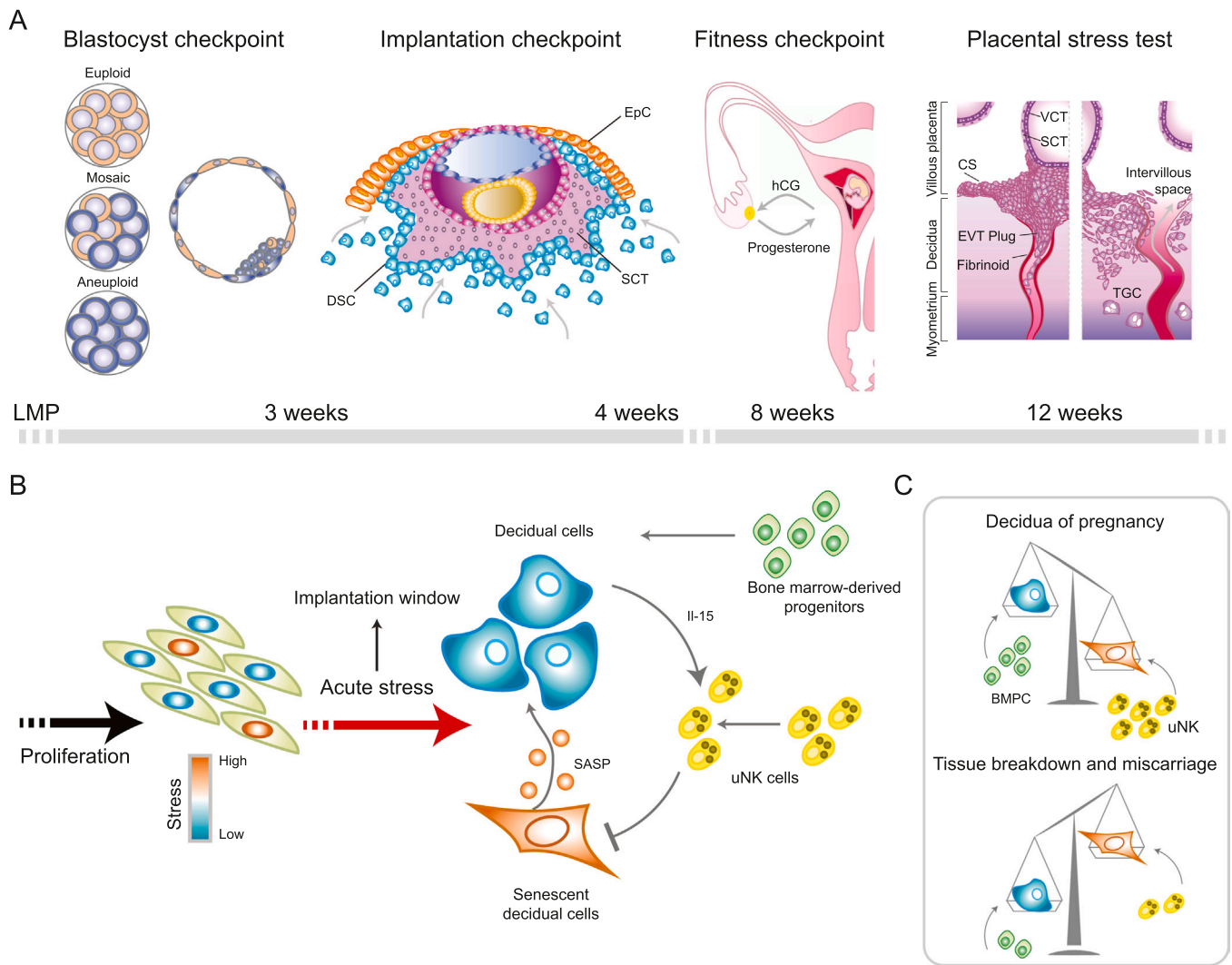


Fig. 2. Tests for success. (A) Several physiological mechanisms limit the risk of prolonged maternal investment in a failing pregnancy. The ‘blastocyst checkpoint’ refers to embryo-intrinsic mechanisms that balances developmental arrest with self-correction in response to different levels of mosaicism. The ‘implantation checkpoint’ involves biosensing of the conceptus by encapsulating maternal decidual cells, whereas the ‘fitness checkpoint’ refers to the maintenance of ovarian progesterone production in response to embryonic fitness signals, such as hCG. Marked vascular changes impose a further stress-test on the decidual-placental interface at the end of the first trimester. EpC, epithelial cells; DSC, decidualized stromal cells; SCT, syncytiotrophoblast; VCT, villous cytotrophoblast; CS, cytotrophoblast shell; EVT, extravillous cytotrophoblast; TGC, trophoblast giant cells; LMP, last menstrual period. (B) Transition of the cycling endometrium into the decidua of pregnancy requires cooperation between decidual cells and uNK cells to eliminate senescent decidual cells. SASP, senescence-associated secretory phenotype; IL-15, interleukin 15. (C) Endometrial fate decisions at implantation pivot on the balance between decidual subsets. Lack of BMPC and/or loss of uterine natural killer (uNK) cell activity drive a pro-senescent decidual response that renders uteroplacental interface vulnerable to tissue breakdown.

genome-wide burden of weakly deleterious mutations influences embryonic survival [30]. Meanwhile, it has long been recognized that lethal whole chromosome abnormalities are very common in human embryos, with aneuploid miscarriages representing only the tip of the iceberg. In this section, we explore the origins and mechanisms of age-dependent embryonic aneuploidy, the functional and fitness consequences of aneuploidy, and the contribution of aneuploidy to miscarriage.

3.1. Origins of human aneuploidy

The fidelity of chromosome segregation during human gamete formation and early embryonic development is strikingly low. It is estimated that 40–60% of human embryos are lost between fertilization and birth, primarily due to aneuploidy, i.e. extra or missing chromosomes compared to the euploid 46-chromosome set [28]. Although a rigorous phylogenetic perspective on the evolution of embryonic aneuploidy is

lacking, the rates in humans are much higher when compared to mice [31], but perhaps comparable to those reported in non-human primates, such as rhesus macaque (~75%) [32]. We note, however, that the practical challenge of controlling for maternal age, ovarian stimulation medications, IVF culture conditions, and other relevant environmental factors complicates such interspecific comparisons.

Preimplantation genetic testing for aneuploidy (PGT-A) seeks to improve the success of in vitro fertilization (IVF) by selecting euploid embryos for transfer [33]. It also offers an exceptional resource for studying human chromosome abnormalities. PGT-A data consistently reveal substantial karyotypic diversity arising from a variety of mechanisms of chromosome mis-segregation [34]. These include maternal meiotic errors, such as classical nondisjunction, precocious separation of sister chromatids (PSSC), and reverse segregation [35]. Nondisjunction is elevated during adolescence and declines through the early twenties, while PSSC and reverse segregation increase exponentially with advancing maternal age, starting around the mid-thirties [8,36]. The

mechanisms of these age associations remain obscure, though several non-mutually exclusive hypotheses have been proposed that generally involve deterioration of chromosome cohesion in oocytes during decades-long meiotic arrest [37]. Paternal meiotic errors are comparatively rare, despite the prevalence of male factor infertility. Aneuploidies affect only 1–5% of sperm and exhibit no discernible age association, though statistical power is limited given this infrequency [10].

While meiotic errors affect all embryonic cells, mitotic errors result in mosaicism, with two or more karyotypically distinct cell lineages (Fig. 2 A). Mosaic aneuploidies are thought to arise through mechanisms including mitotic nondisjunction, anaphase lag, or endoreplication [36]. Current estimates of the incidence of mosaicism range from 4% to 90% [38], further fuelling a long-standing debate over the clinical usefulness of PGT-A [39]. While these wide-ranging estimates reflect both technical and biological variability, one crucial limitation of PGT-A is the reliance on biopsies of one or few cells. The resulting sampling variability is further compounded by the unknown spatial distribution of aneuploid cells within the embryo.

One notable observation from aggregated PGT-A data is the large variation in aneuploidy rates among embryos from different women, even after controlling for maternal age. Such patterns of over-dispersion imply that aneuploidy rates are influenced by environmental or parental genetic risk factors [40]. Suspected environmental risk factors include exposure to endocrine-disrupting chemicals, such as bisphenol A and its analogues, which disrupt meiosis in model organisms and cultured human tissue [41]. Genetic risk factors for human aneuploidy have proven largely elusive, though recent studies are providing initial headway (Box 1).

3.2. Functional and fitness consequences of aneuploidy

Why certain chromosome abnormalities cause preimplantation embryonic arrest, but others remain viable into later development, is poorly understood. Phenotypic impacts of deletions and duplications, including aneuploidy and sub-chromosomal structural variation, are mediated by effects on gene expression [42]. These include primary dosage effects of chromosome gains and losses, secondary effects of dysregulated transcription factors that propagate to other chromosomes, as well as tertiary effects such as cellular stress responses, all of which have been documented in human embryos [43,44]. These effects may propagate to the level of translation and elicit the build-up of misfolded proteins, which can aggregate and exert proteotoxic stress on cells [42]. Yet the relationship between gene dosage and activity at protein level is not necessarily linear due to buffering mechanisms at the translational and post-translational stages [45,46]. Understanding these relationships using quantitative methods is a major goal of genomic research into the functional impacts of aneuploidy.

The fitness consequences of aneuploidy are linked to the number and identity of aneuploid cells and chromosomes. Meiotic aneuploidies arise during egg formation, impact all cells of the embryos, and are unambiguously harmful. Mitotic aneuploidies are restricted to the descendant cells of the erroneous mitotic division. While severe mosaicism may result in early embryonic arrest, lower levels of mosaicism are compatible with normal development and healthy live birth [20,47,48]. This observation implies that certain mosaic aneuploidies are subject to negative selection and actively or passively eliminated from the conceptus throughout development. Studies using mouse models and human embryonic stem cells indicate that aneuploid cells are purged from mosaic fetal lineages by autophagy-mediated apoptosis but tolerated within the trophoctoderm, which goes on to form the placenta [49]. Analysis of single-cell sequencing data from human embryos indicates that this selection process may intensify during post-implantation development [44]. Additional mechanisms have been identified by which aneuploid cells are extruded from human and non-human primate embryos during blastocyst formation [32].

Recently, extended in vitro embryo culture systems have been used to examine the developmental consequences of specific aneuploidies during and after implantation. Popovic et al. [50], for example, reported that while trisomies of chromosomes 16, 21, and 22 remain viable through day 12 postfertilization, autosomal monosomies tend to arrest at the time of implantation or shortly thereafter. These findings were echoed by Shahbazi et al. [51], who also demonstrated that embryos with trisomy 16 exhibit hypo-proliferation of the trophoblast, potentially driven by cell adhesion defects induced by over-expression of the chromosome 16 gene *E-cadherin (ECAD)*. Future work combining such systems with functional genomic profiling will help identify additional causal genes and mechanisms that drive mortality of aneuploid embryos.

3.3. The role of aneuploidy in miscarriage

Chromosome abnormalities are a common finding in sporadic miscarriage, with reported rates of 40–60% [52,53]. Some recent studies reported even higher rates, reflecting the demographic trends in delaying pregnancy [53], and the application of increasingly sensitive platforms for aneuploidy detection [54]. The age-related risk of miscarriage is largely accounted for by increased frequency of meiotic trisomy [7]. The incidence of chromosomal errors in sporadic miscarriage also varies by gestational age, peaking between 9 and 14 weeks of gestation [53]. Counterintuitively, anembryonic miscarriages and clinically recognized losses before 6 weeks of gestation are disproportionately euploid [55–57]. There is no evidence of geographic or ancestry-related variation in the incidence or pattern of chromosomal abnormalities [53], suggesting that the mechanisms causing aneuploid miscarriages are an inherent feature of human reproduction.

Box 1

The search for genetic risk factors for human aneuploidy.

Identification of parental genetic risk factors of embryonic aneuploidy could potentially lead to screening of couples at risk of infertility and miscarriage. The largest genome-wide association study (GWAS) of aneuploidy risk to date reported one significant quantitative trait locus (QTL)—a common set of linked polymorphisms spanning the centrosome regulator *PLK4* (polo like kinase 4). The embryos of women carrying the risk haplotype had higher rates of mitotic aneuploidy and were less likely to progress to the blastocyst stage [109]. Tripolar mitosis of diploid cells is a candidate mechanism driving this maternal effect [42] and exemplifies a broader class of complex mosaicism that is compatible with cleavage-stage development but arrests upon embryonic genome activation prior to blastocyst formation [10]. A further study of trisomy 21 reported putative associations at several meiosis-related genes, albeit below genome-wide significance thresholds [45]. Both studies were constrained to the discovery of common genetic variation because of small sample sizes (hundreds to thousands of patients). Population genetic theory predicts that heritability of traits closely tied to fitness will have an outside contribution from rare and *de novo* mutations [46]. Characterization of such rare variants will require alternative approaches, such as whole-genome sequencing and meticulous validation in animal models or human cell lines.

Alternatively, maternal age may be such a powerful driver of aneuploidy that it masks the impact of ancestry and local environmental factors.

In the context of recurrent miscarriage, there are three pertinent outstanding questions. First, is the underlying rate of embryonic aneuploidy altered in recurrent miscarriage? One PGT-A study, involving 46,439 preimplantation human embryos, observed a small but significant elevation of maternal meiotic (but not mitotic) aneuploidy in blastocyst-stage embryos from recurrent miscarriage patients (age-adjusted odds ratio: 1.14, 95% confidence interval: 1.01–1.27) [10]. While intriguing, this modest effect size implies that patient-specific risk of aneuploid conception may be a minor contributor to recurrent miscarriage. Second, is the ratio of euploid versus aneuploid pregnancy losses different in recurrent miscarriage? The literature is replete with contradictory studies reporting mostly reduced, sometimes unchanged, and occasionally elevated rates of chromosome abnormalities in recurrent versus sporadic miscarriage [58–61]. A conspicuous problem with many studies is the failure to account for confounding variables, such as gestational age and the number of previous miscarriages. A final question, with important implications for clinical management, is whether embryonic aneuploidy offers a plausible explanation for the recurrence risk of miscarriage? As illustrated in **Box 2**, this seems doubtful.

4. Embryo selection at implantation

The intrinsic genetic instability of human embryos imposes a major challenge onto the endometrium: how to eliminate embryos of low fitness without compromising implantation of high-quality embryos? Here, we describe the cellular events that enable the endometrium to recognise and respond to embryos of different qualities and discuss how pathogenic mechanisms that compromise this endometrial function cause miscarriage, irrespective of the ploidy status of the conceptus.

4.1. Endometrial cyclicity and decidualization

Between menarche and menopause, the endometrium undergoes hundreds of cycles of tissue breakdown, menstrual shedding, and scar-free regeneration in response to the rise and fall of ovarian hormone production. In each cycle, rapid oestrogen-dependent endometrial growth is followed by progesterone-dependent differentiation of glands and stroma. During the midluteal phase, the endometrial stroma remodels intensively, heralding the start of a short implantation window. This process of tissue remodelling, termed decidualization, is driven foremost by the transformation of stromal fibroblasts into epithelial-like decidual cells and accumulation of uterine natural killer (uNK) cells [62].

Decidualization of endometrial stromal cells is a multistep differentiation process, which starts with an acute cellular stress response and

release of proinflammatory mediators (Fig. 2B) [63–66]. This initial inflammatory phase coincides with the implantation window [62,66,67]. After approximately four days of inflammatory reprogramming, specialized anti-inflammatory decidual cells emerge, highly resistant to metabolic and oxidative stressors, and exquisitely responsive to embryonic signals [62, 68–70]. However, some stromal cells already burdened by replication stress fail to differentiate and are earmarked for cellular senescence [63,65]. Senescent decidual cells are progesterone-resistant and abundantly secrete a complex mixture of extracellular matrix proteins and proteinases, proinflammatory cytokines, and chemokines (termed senescence associated secretory phenotype), which cause sterile inflammation and induce secondary senescence in neighbouring decidual cells [65]. If left unchecked, spatiotemporal propagation of this senescence phenotype renders the decidua vulnerable to breakdown. Progesterone-dependent decidual cells can escape this default pathway by co-opting uNK cells to eliminate their senescent counterparts through perforin- and granzyme-containing granule exocytosis (Fig. 2B) [63,65]. This process rejuvenates the endometrium at implantation and enables transformation of the stroma into a tightly adherent, immune-protective decidual matrix that coordinates trophoblast invasion and accommodates the placenta throughout pregnancy. Expansion of the decidua in early pregnancy relies on recruitment and differentiation of circulating bone marrow-derived progenitor cells (BMPC) into decidual cells [71]. In a non-conception cycle, however, progesterone production by the ovarian corpus luteum declines in the late-luteal phase, leading to a preponderance of senescent decidual cells, influx of neutrophils and macrophages, tissue breakdown and menstrual shedding [72].

4.2. Implantation checkpoints

Decidual transformation of the endometrium occurs in all mammals with an invading placenta. Only in menstruating species is this differentiation process initiated in each cycle, instead of triggered by the implanting embryo [22]. Maternal control of the decidual process imposes multiple checkpoints on the conceptus (Fig. 2A). First, the embryo must implant at the right time in the cycle to direct the decidual reaction away from tissue destruction. Clinically, a delayed rise in hCG levels beyond the putative implantation window is strongly associated with miscarriage in the first two weeks of pregnancy [1]. Once embedded, the conceptus is rapidly encapsulated by migratory decidual cells [73–75]. These cells serve as biosensors of embryo quality, engaging in both negative and positive selection. In response to signals from low-quality embryos, as defined by morphological criteria, decidual cells mount an endoplasmic reticulum stress response, which inhibits secretion of crucial implantation factors and hinders embryo encapsulation [73,75,76]. By contrast, secreted factors from successful embryos enhance the

Box 2

Can aneuploid embryos account for the recurrence risk of miscarriage?.

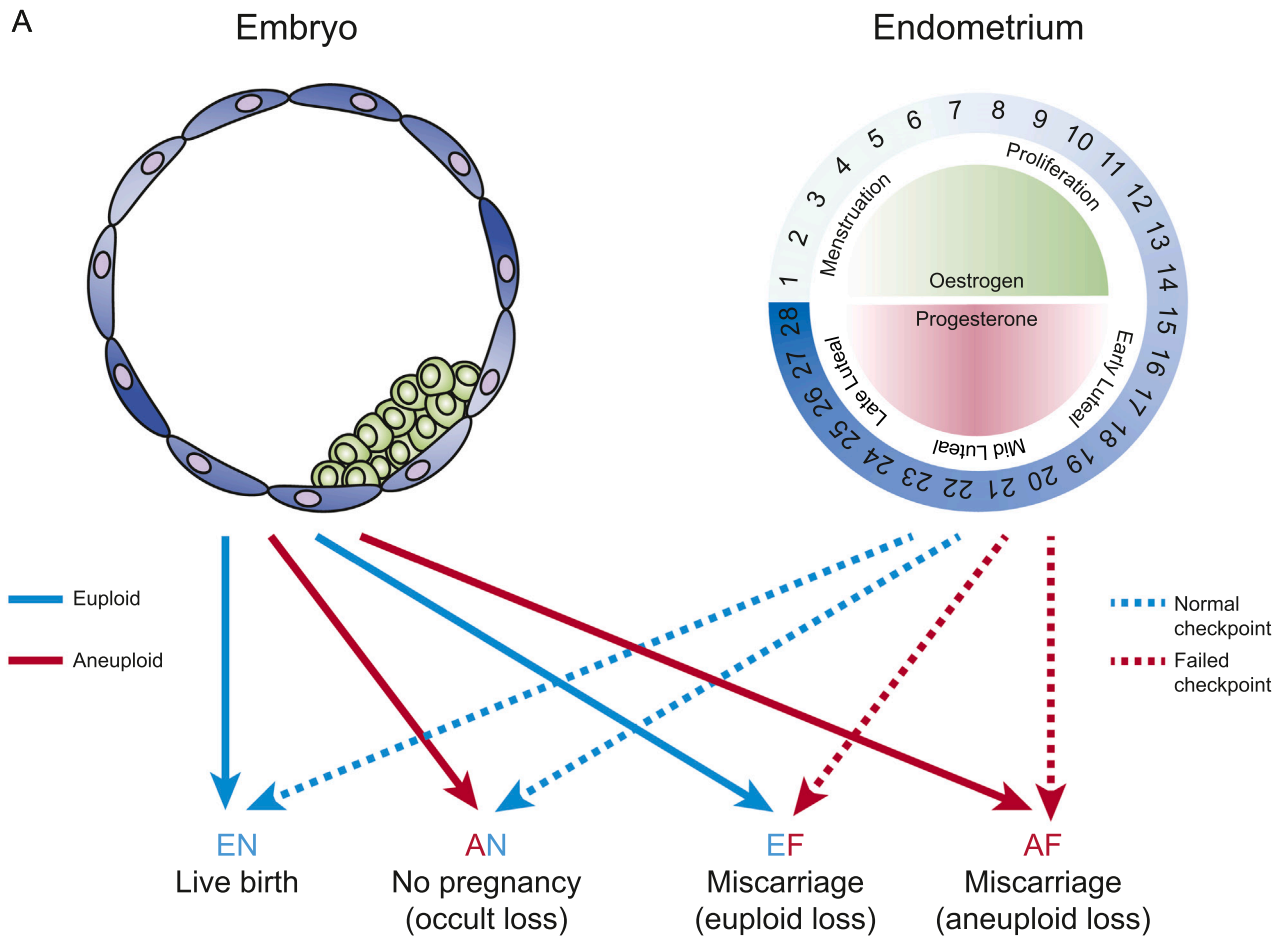
The diagnosis of an aneuploid pregnancy loss is often considered a compelling explanation for recurrent miscarriage. Some clinicians recommend no further investigations or treatments, whereas others advocate IVF treatment with PGT-A. However, can the recurrence risk of miscarriage be explained by embryonic aneuploidy alone? To answer this question, consider the following numbers:

At the age of 30, the proportion of embryos that are aneuploid is 30% (Fig. 1 A). In healthy women, the sporadic miscarriage rate is approximately 10% (Fig. 1B), half of which are reportedly aneuploid losses. As aneuploid pregnancies rarely result in live births, the aneuploid pregnancy rate will approximate the aneuploid miscarriage rate, that is, 5%. After three prior losses, however, the miscarriage rate rises to 40%. Some studies reported no change in the rate of aneuploid pregnancy losses between sporadic and recurrent miscarriage. If correct, the aneuploid pregnancy rate in recurrent miscarriage patients aged 30-years should approximate to 20%, i.e., half of the reported 40% miscarriage rate.

Is it possible to account for the relative increase in the aneuploid pregnancy rate from 5% to 20% between healthy women and recurrent miscarriage patients? This increase cannot be explained by a proportional increase in aneuploid embryos (since this would be from 30% to 120%). Thus, either the aneuploid miscarriage rate is overestimated or maternal selection at implantation is relaxed.

expression of maternal implantation and metabolic genes, thus actively promoting further implantation [68]. How human embryos signal their developmental potential remains unclear. However, it is striking that hsa-miR-320a and hyaluronidase 2, two putative embryonic fitness cues that act on decidual and uNK cells, respectively, are also implicated in

promoting pre-implantation development [77,78]. Next, it is incumbent on the embryo to secrete sufficient levels of hCG to rescue ovarian progesterone biosynthesis until the placenta takes over progesterone production around eight weeks of pregnancy. Human embryos are exquisitely adapted to meet this herculean task; the gene encoding the



B

Predicted miscarriage rates*		Reported miscarriage rates		
checkpoint failure	miscarriage rate	previous miscarriages	Magnus <i>et al.</i> 2019 [5]	Coomarasamy <i>et al.</i> , 2020 [112] (95 % CI)
1 in 13	10.6 %	0	11.6 %	11.3 % (7-17)
2 in 13	20.6 %	1	19.8 %	17.0 % (9-27)
3 in 13	30.0 %	2	27.7 %	28.0 % (20-36) ‡
4 in 13	38.8 %	3	41.9 % †	
5 in 13	47.2 %	4		39.6 % (35-44)
6 in 13	55.0 %	5		47.2 % (36-58)
7 in 13	62.5 %	6		63.9 % (54-73) †

Fig. 3. Predicted impact of implantation checkpoint failure on recurrence risk of miscarriage. (A) The implantation checkpoint hypothesis posits that the decidualizing endometrium mounts a tailored response to individual embryos, eliminating developmentally compromised embryos (leading to occult losses) but supporting competent embryos (leading to successful pregnancies). Checkpoint failure means that aneuploid embryos will escape detection (leading to aneuploid miscarriages), whilst euploid pregnancies will fail because the endometrium is unsupportive and prone to breakdown (leading to euploid miscarriages). Four scenarios can be considered in which the embryo is either euploid (E) or aneuploid (A) and the endometrium has either a normal (N) or failed (F) checkpoint, leading to three clinical outcomes: no pregnancy (occult loss), miscarriage, or live birth. (B) Predicted miscarriage rates with increasing frequency of cycles with implantation checkpoint failure. The predicted rates are compared to reported miscarriage rates associated with increasing number of pregnancy losses. *Based on 30% aneuploidy rate; †indicated number of previous miscarriages or more; ‡two and three previous miscarriages were combined in this study (Coomarasamy et al. [112]).

biologically active β -subunit hCG (*CGB*) is duplicated six times in the genome and the glycosylated protein evolved to have a much longer half-life compared to the ancestral luteinizing hormone [17,19].

4.3. Implantation checkpoint failure and recurrent miscarriage

Recurrent miscarriage is associated with a pro-senescent decidual response, characterised foremost by a lack of anti-inflammatory decidual cells (Fig. 2B). This presents the embryo with a maternal environment that is easy to invade, devoid of biosensing properties, and prone to breakdown. The term ‘implantation checkpoint failure’ denotes the functional consequences of this pathological state, i.e., an endometrium that neither supports normal embryos nor eliminates abnormal embryos at implantation.

Decidualization is an iterative process, finetuned by cyclical recruitment of uNK cells and BMPC [63,79,80]. Hence, the frequency of cycles with an aberrant decidual response, leading to implantation checkpoint failure, is determined by the stringency of these opposing (homeostatic) regulatory mechanisms. For example, recurrent miscarriage is linked to loss of clonogenic BMPC during the implantation window, and the level of depletion correlates with the number of previous miscarriages [79,81], and thus, the recurrence risk. The abundance of uNK cells varies markedly throughout the luteal phase and between cycles [63], which on the one hand is in keeping with their homeostatic function but, on the other, also accounts for the inconsistent findings in different studies. Nevertheless, there is evidence that lower uNK cell activity in the endometrium and peripheral blood associates with higher miscarriage rates [65, 82–84]. Further, metabolic (e.g., obesity) and endocrine disorders (e.g., hypothyroidism) can perturb homeostatic regulation of the decidual response [4].

The implantation checkpoint failure hypothesis leads to several predictions. First, as implantation is the rate limiting step for pregnancy, lack of embryo selection at implantation should lead to more rapid conception or “superfertility”, which is consistent with clinical observations [85,86]. Second, the frequency of implantation checkpoint failure should determine the recurrence risk of miscarriage [65,81]. While this conjecture warrants further investigation, the predicted impact on miscarriage rates aligns with the reported stepwise increase in recurrence risk with each additional miscarriage (Fig. 3 and Supplementary file 1). Finally, checkpoint failure implies that the endometrium does not select efficiently against aneuploid embryos at implantation. Hence, the incidence of aneuploid pregnancy loss in recurrent miscarriage is predicted to mirror the incidence of meiotic errors in preimplantation embryos, i.e., a preponderance of losses should be euploid in younger but not older patients [61].

5. The uteroplacental interface

The development of the placenta in early pregnancy involves a sequence of events, starting with the emergence of different placental cell lineages and plugging of the uterine spiral arteries by invading trophoblast. Consequently, the early conceptus develops under hypoxic conditions, supported by endometrial gland secretions, until the onset of placental perfusion around 12 weeks of pregnancy. This section highlights how this sequence of events is disrupted in miscarriage.

5.1. Placental defects and miscarriage

Human placentation involves complex fetal-maternal interactions that are more extensive than in most other mammalian species [87]. Various placental cell lineages proliferate rapidly, and numerous villi cover the chorionic sac shortly after implantation. Each villus consists of a mesodermal core surrounded by an inner layer of progenitor cytotrophoblast cells and an outer layer of syncytiotrophoblast. At the tip of the villus, cytotrophoblast cells interrupt the syncytiotrophoblast and first form a columnar structure before spreading laterally to surround

the entire conceptus (Fig. 2 A) [88]. This cytotrophoblastic shell anchors the placenta but also plugs the terminal branches of the uterine spiral arteries [89]. Consequently, the conceptus is sealed off and develops under low oxygen tension, protected against a variety of stressors during the critical period of organogenesis [87]. During this phase, profuse glandular secretions, rich in growth factors, lipids, and carbohydrates, nourish the placenta and fetus [90,91]. Invasive trophoblast cells emerge from the decidual surface of the shell and migrate through the decidua and into the inner myometrium where they fuse into multinucleated trophoblast giant cells [88].

Towards the end of the first trimester, the trophoblast plugs in the spiral arteries dislocate progressively, allowing gradual perfusion of the intervillous space. During this period, oxygen tension rises steeply [89, 92]. The ensuing wave of reactive oxygen species stress-tests the resilience the placental-decidual interface [93]. Once the uteroplacental circulation is established around 12 weeks, the pregnancy has successfully negotiated the final checkpoint, and miscarriage rates drop sharply [5]. However, remodelling of the maternal spiral arteries is not yet complete. This process requires continued cooperation between interstitial and endovascular trophoblast cells to convert the decidual and inner myometrial portions of these arteries into large-bore vessels devoid of smooth muscle and elastin [94].

In 70% of first-trimester miscarriages, the cytotrophoblastic shell surrounding the conceptus is thin and fragmented across the placenta [95]. Deficient endovascular trophoblast invasion causes incomplete plugging of spiral arteries, precocious placental perfusion, and oxidative damage to immature villi [87,92,93,95]. These pathogenic events underpin both euploid and aneuploid miscarriages, suggesting orchestration by an aberrant decidual environment. Lesions confined to the placental edge may cause significant bleeding without necessarily imperilling the pregnancy and present clinically as a threatened miscarriage [87].

5.2. Decidual immune system and miscarriage

The first trimester decidua is rich in immune cells, principally uNK cells (~70%), macrophages (~20%), and T cells (~10%) [96]. Decidual cells control the influx and expansion of local immune cells [97,98]. Conversely, cooperation between different innate immune cell populations is essential for optimal decidualization and development of the uteroplacental interface (Box 3). Through this interdependency, impaired decidualization adversely impacts local immune populations and vice versa, hampering the distinction between cause and effect.

Because the semi-allogeneic conceptus expresses ‘foreign’ paternal proteins, miscarriage is frequently attributed to a breakdown in maternal T cell tolerance, akin to graft-versus-host disease in transplant immunology [99]. However, placental mammals arose at least 75 million years ago [100], and multiple adaptations have evolved to preclude placental trophoblast killing by decidual or systemic T cells [101]. uNK cells also engage in allo-recognition of fetal cells through binding of their killer immunoglobulin-like receptors (KIR) to human leukocyte antigen-C (HLA-C) molecules on invading trophoblast [101]. The genes encoding maternal KIR and fetal HLA-C are highly polymorphic, with different haplotype combinations stimulating or inhibiting uNK cell activity. Immunogenetic studies have consistently associated excessively inhibitory KIR and HLA-C combinations with adverse pregnancy outcome, including recurrent miscarriage [102,103]. Thus, uNK cell activity is essential for pregnancy. Nevertheless, the misconception that pregnancies fail because of an ‘overreactive’ immune response continues to resonate with many clinicians, fuelling a plethora of immune tests of uncertain value. Clinical trials have not demonstrated that immunosuppressive therapies (e.g., prednisolone, intralipids, intravenous immunoglobulin) increase live birth rates in recurrent miscarriage [13].

The uteroplacental interface is vulnerable to autoimmune disorders, most prominently antiphospholipid syndrome (APS), defined by the

Box 3

Examples of immune cell cooperation in early pregnancy.

- Three distinct uNK cell subpopulations at the feto-maternal interface exert diverse roles in tissue homeostasis, spiral arteries remodelling through secretion of IFN γ and VEGF, and trophoblast invasion [96].
- In early pregnancy, macrophages acquire an anti-inflammatory (M2) phenotype, secrete TGF β , IL-10, and IDO (indoleamine 2,3-dioxygenase), and contribute to decidual angiogenesis, vascular transformation, and trophoblast invasion [110].
- Dendritic cells (DCs) are also directly involved in the decidualization process, promoting proliferation and differentiation [111]
- Activated regulatory T cells (Tregs) interact with DCs through CTLA4 (cytotoxic T-lymphocyte-associated protein 4), reducing the expression of co-stimulatory molecules (CD80 and CD86) and inhibiting effector T cell differentiation. Tregs also suppress cytotoxic uNK cell activity and regulate M2 macrophages and tissue DCs through release of heme oxygenase-1 [101].

presence of antiphospholipid antibodies (aPL), such as lupus anticoagulant, anticardiolipin or anti- β 2-glycoprotein 1 [104]. Since the main antigen of aPL, β 2-glycoprotein 1, is expressed constitutively on decidual and trophoblast cells, the maternal-fetal interface is a major target for these autoantibodies [105]. Unlike systemic APS, which is a thrombotic disorder, obstetric APS is primarily an inflammatory disorder [105], linked to fetal death and late pregnancy complications, such as preeclampsia, fetal growth restriction and preterm birth [106].

6. Perspective

Mammalian reproduction is characterised by remarkable diversification in pregnancy traits [19]. Based on comparative transcriptomics, hundreds of genes have been identified in pregnant endometrium that were gained or lost in primate and human lineages [107]. These genes not only underly the emergence of novel reproductive traits, such as spontaneous decidualization, menstruation, and deep haemochorial placentation, but also play a disproportionate role in reproductive disorders [107]. Despite these emerging insights, clinical practice remains firmly grounded in the misconception that early gestation represents a ‘vulnerable’ period, easily disrupted by a host of subclinical disorders [4, 14]. Here, we discussed how spontaneous decidualization enables active selection of embryos at implantation and elaborated the clinical consequences of implantation checkpoint failure. Endometrial fate decisions at implantation, that is, menstruation-like breakdown or transformation into a robust decidual matrix of pregnancy, pivot on balancing anti-inflammatory decidual cells and pro-inflammatory senescent decidual cell. While developmentally competent embryos promote cooperation between anti-inflammatory decidual cells and extra-uterine uNK cells and BMPC to form a robust decidua, low-fitness embryos subvert these interactions, thereby engineering their own demise [78]. However, the fitness consequences of aneuploidy depend on the identity of chromosomes; and embryos harbouring trisomies of smaller chromosomes appear particularly adept at evading maternal recognition at implantation, thus accounting for most age-related miscarriages. We posit that recurrence risk reflects the frequency of menstrual cycles resulting in implantation checkpoint failure, a paradigm that is already pointing towards novel therapeutic approaches. For example, a recent double-blind placebo-controlled feasibility trial demonstrated that sitagliptin, a dipeptidyl-peptidase IV (DPP4) inhibitor used in the management of diabetes, improves the decidual response at implantation by enhancing recruitment of BMPC [108]. Whether pre-pregnancy interventions, with sitagliptin or other drugs, lead to higher live birth rates in recurrent miscarriage patients will have to await further clinical trials.

Funding sources

The work was funded by Tommy’s Baby Charity, United Kingdom. RCM is supported by a National Institutes of Health (NIH) grant (R35GM133747), United States of America. JJB holds a Wellcome Trust

Investigator Award (212233/Z/18/Z), United Kingdom. The funders had no role in the writing of the article; or in the decision to submit the paper for publication.

Declaration of Competing Interest

None.

Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at [doi:10.1016/j.semcd.2022.01.007](https://doi.org/10.1016/j.semcd.2022.01.007).

References

- [1] A.J. Wilcox, C.R. Weinberg, J.F. O’Connor, D.D. Baird, J.P. Schlatterer, R. E. Canfield, E.G. Armstrong, B.C. Nisula, Incidence of early loss of pregnancy, *N. Engl. J. Med.* 319 (4) (1988) 189–194, <https://doi.org/10.1056/NEJM198807283190401>.
- [2] X. Wang, C. Chen, L. Wang, D. Chen, W. Guang, J. French, Conception, early pregnancy loss, and time to clinical pregnancy: a population-based prospective study, *Fertil. Steril.* 79 (3) (2003) 577–584, [https://doi.org/10.1016/s0015-0282\(02\)04694-0](https://doi.org/10.1016/s0015-0282(02)04694-0).
- [3] M.J. Zinaman, E.D. Clegg, C.C. Brown, J. O’Connor, S.G. Selevan, Estimates of human fertility and pregnancy loss, *Fertil. Steril.* 65 (3) (1996) 503–509.
- [4] S. Quenby, I.D. Gallos, R.K. Dhillon-Smith, M. Podsek, M.D. Stephenson, J. Fisher, J.J. Brosens, J. Brewin, R. Ramhorst, E.S. Lucas, R.C. McCoy, R. Anderson, S. Daher, L. Regan, M. Al-Memar, T. Bourne, D.A. MacIntyre, R. Rai, O.B. Christiansen, M. Sugiura-Ogasawara, J. Odendaal, A.J. Devall, P.R. Bennett, S. Petrou, A. Coomarasamy, Miscarriage matters: the epidemiological, physical, psychological, and economic costs of early pregnancy loss, *Lancet* 397 (10285) (2021) 1658–1667, [https://doi.org/10.1016/S0140-6736\(21\)00682-6](https://doi.org/10.1016/S0140-6736(21)00682-6).
- [5] M.C. Magnus, A.J. Wilcox, N.H. Morken, C.R. Weinberg, S.E. Haberg, Role of maternal age and pregnancy history in risk of miscarriage: prospective register based study, *Bmj* 364 (2019) 1869, <https://doi.org/10.1136/bmj.l869>.
- [6] A.M. Kolte, D. Westergaard, O. Lidegaard, S. Brunak, H.S. Nielsen, Chance of live birth: a nationwide, registry-based cohort study, *Hum. Reprod.* 36 (4) (2021) 1065–1073, <https://doi.org/10.1093/humrep/deaa326>.
- [7] T. Hassold, D. Chiu, Maternal age-specific rates of numerical chromosome abnormalities with special reference to trisomy, *Hum. Genet* 70 (1) (1985) 11–17, <https://doi.org/10.1007/bf00389450>.
- [8] J.R. Gruhn, A.P. Zielinska, V. Shukla, R. Blanshard, A. Capalbo, D. Cimadomo, D. Nikiforov, A.C. Chan, L.J. Newnham, I. Vogel, C. Scarica, M. Krapchev, D. Taylor, S.G. Kristensen, J. Cheng, E. Ernst, A.B. Bjorn, L.B. Colmorn, M. Blayney, K. Elder, J. Liss, G. Hartshorne, M.L. Grondahl, L. Rienzi, F. Ubaldi, R. McCoy, K. Lukaszuk, C.Y. Andersen, M. Schuh, E.R. Hoffmann, Chromosome errors in human eggs shape natural fertility over reproductive life span, *Science* 365 (6460) (2019) 1466–1469, <https://doi.org/10.1126/science.aav7321>.
- [9] J.M. Fransiak, E.J. Forman, K.H. Hong, M.D. Werner, K.M. Upham, N.R. Treff, R. T. Scott Jr., The nature of aneuploidy with increasing age of the female partner: a review of 15,169 consecutive trophectoderm biopsies evaluated with comprehensive chromosomal screening, *Fertil. Steril.* 101 (3) (2014) 656–663 e1, <https://doi.org/10.1016/j.fertnstert.2013.11.004>.
- [10] R.C. McCoy, Z.P. Demko, A. Ryan, M. Banjevic, M. Hill, S. Sigurjonsson, M. Rabinowitz, D.A. Petrov, Evidence of selection against complex mitotic-origin aneuploidy during preimplantation development, *PLoS Genet.* 11 (10) (2015), e1005601, <https://doi.org/10.1371/journal.pgen.1005601>.
- [11] T. Hassold, N. Chen, J. Funkhouser, T. Jooss, B. Manuel, J. Matsuura, A. Matsuyama, C. Wilson, J.A. Yamane, P.A. Jacobs, A cytogenetic study of 1000 spontaneous abortions, *Ann. Hum. Genet* 44 (2) (1980) 151–178.
- [12] S.I. Nagaoka, T.J. Hassold, P.A. Hunt, Human aneuploidy: mechanisms and new insights into an age-old problem, *Nat. Rev. Genet.* 13 (7) (2012) 493–504, <https://doi.org/10.1038/nrg3245>.

- [97] P. Nancy, E. Tagliani, C.S. Tay, P. Asp, D.E. Levy, A. Erlebacher, Chemokine gene silencing in decidual stromal cells limits T cell access to the maternal-fetal interface, *Science* 336 (6086) (2012) 1317–1321, <https://doi.org/10.1126/science.1220030>.
- [98] M. Silasi, Y. You, S. Simpson, J. Kaislasuo, L. Pal, S. Guller, G. Peng, R. Ramhorst, E. Grasso, S. Etemad, S. Durosier, P. Aldo, G. Mor, Human chorionic gonadotropin modulates CXCL10 expression through histone methylation in human decidua, *Sci. Rep.* 10 (1) (2020) 5785, <https://doi.org/10.1038/s41598-020-62593-9>.
- [99] G. Mor, P. Aldo, A.B. Alvero, The unique immunological and microbial aspects of pregnancy, *Nat. Rev. Immunol.* 17 (8) (2017) 469–482, <https://doi.org/10.1038/nri.2017.64>.
- [100] L. Liu, J. Zhang, F.E. Rheindt, F. Lei, Y. Qu, Y. Wang, Y. Zhang, C. Sullivan, W. Nie, J. Wang, F. Yang, J. Chen, S.V. Edwards, J. Meng, S. Wu, Genomic evidence reveals a radiation of placental mammals uninterrupted by the KPg boundary, *Proc. Natl. Acad. Sci. USA* 114 (35) (2017) E7282–E7290, <https://doi.org/10.1073/pnas.1616744114>.
- [101] A. Moffett, O. Chazara, F. Colucci, Maternal allo-recognition of the fetus, *Fertil. Steril.* 107 (6) (2017) 1269–1272, <https://doi.org/10.1016/j.fertnstert.2017.05.001>.
- [102] S.E. Hiby, R. Apps, A.M. Sharkey, L.E. Farrell, L. Gardner, A. Mulder, F.H. Claas, J. J. Walker, C.W. Redman, L. Morgan, C. Tower, L. Regan, G.E. Moore, M. Carrington, A. Moffett, Maternal activating KIRs protect against human reproductive failure mediated by fetal HLA-C2, *J. Clin. Investig.* 120 (11) (2010) 4102–4110, <https://doi.org/10.1172/JCI43998>.
- [103] S.E. Hiby, J.J. Walker, M. O’Shaughnessy K, C.W. Redman, M. Carrington, J. Trowsdale, A. Moffett, Combinations of maternal KIR and fetal HLA-C genes influence the risk of preeclampsia and reproductive success, *J. Exp. Med.* 200 (8) (2004) 957–965, <https://doi.org/10.1084/jem.20041214>.
- [104] G. Ruiz-Irastorza, M. Crowther, W. Branch, M.A. Khamashta, Antiphospholipid syndrome, *Lancet* 376 (9751) (2010) 1498–1509, [https://doi.org/10.1016/S0140-6736\(10\)60709-X](https://doi.org/10.1016/S0140-6736(10)60709-X).
- [105] V.M. Abrahams, L.W. Chamley, J.E. Salmon, Emerging treatment models in rheumatology: antiphospholipid syndrome and pregnancy: pathogenesis to translation, *Arthritis Rheuma* 69 (9) (2017) 1710–1721, <https://doi.org/10.1002/art.40136>.
- [106] K. Schreiber, S. Sciascia, P.G. de Groot, K. Devreese, S. Jacobsen, G. Ruiz-Irastorza, J.E. Salmon, Y. Shoenfeld, O. Shovman, B.J. Hunt, Antiphospholipid syndrome, *Nat. Rev. Dis. Prim.* 4 (2018) 18005, <https://doi.org/10.1038/nrdp.2018.5>.
- [107] K. Mika, M. Marinic, M. Singh, J. Muter, J.J. Brosens, V.J. Lynch, Evolutionary transcriptomics implicates new genes and pathways in human pregnancy and adverse pregnancy outcomes, *eLife* 10 (2021), <https://doi.org/10.7554/eLife.69584>.
- [108] S. Tewary, E.S. Lucas, R. Fujihara, P.K. Kimani, A. Polanco, P.J. Brighton, J. Muter, K.J. Fishwick, M. Da Costa, L.J. Ewington, L. Lacey, S. Takeda, J. J. Brosens, S. Quenby, Impact of sitagliptin on endometrial mesenchymal stem-like progenitor cells: a randomised, double-blind placebo-controlled feasibility trial, *EBioMedicine* 51 (2020), 102597, <https://doi.org/10.1016/j.ebiom.2019.102597>.
- [109] R.C. McCoy, Z. Demko, A. Ryan, M. Banjevic, M. Hill, S. Sigurjonsson, M. Rabinowitz, H.B. Fraser, D.A. Petrov, Common variants spanning PLK4 are associated with mitotic-origin aneuploidy in human embryos, *Science* 348 (6231) (2015) 235–238, <https://doi.org/10.1126/science.aaa3337>.
- [110] A. Schumacher, D.J. Sharkey, S.A. Robertson, A.C. Zenclussen, Immune cells at the fetomaternal interface: how the microenvironment modulates immune cells to foster fetal development, *J. Immunol.* 201 (2) (2018) 325–334, <https://doi.org/10.4049/jimmunol.1800058>.
- [111] V. Plaks, T. Birnberg, T. Berkutzki, S. Sela, A. BenYashar, V. Kalchenko, G. Mor, E. Keshet, N. Dekel, M. Neeman, S. Jung, Uterine DCs are crucial for decidua formation during embryo implantation in mice, *J. Clin. Investig.* 118 (12) (2008) 3954–3965, <https://doi.org/10.1172/JCI36682>.
- [112] A. Coomarasamy, A.J. Devall, J.J. Brosens, S. Quenby, M.D. Stephenson, S. Sierra, O.B. Christiansen, R. Small, J. Brewin, T.E. Roberts, R. Dhillion-Smith, H. Harb, H. Noordali, A. Papadopoulou, A. Eapen, M. Prior, G.C. Di Renzo, K. Hinshaw, B. W. Mol, M.A. Lumsden, Y. Khalaf, A. Shennan, M. Goddijn, M. van Wely, M. Al-Memar, P. Bennett, T. Bourne, R. Rai, L. Regan, I.D. Gallos, Micronized vaginal progesterone to prevent miscarriage: a critical evaluation of randomized evidence, *Am. J. Obstet. Gynecol.* (2020), <https://doi.org/10.1016/j.ajog.2019.12.006>.