We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



167,000





Our authors are among the

TOP 1% most cited scientists





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Chapter

A Short Communication: Non-acid Nucleic Blood Multi-Factors Panels for Primary Breast Cancer Detection - A Systematic Review and Network Meta-Analysis

Vahid Raja, Ziba Farajzadegan, Marjan Mansourian, Khojaste Ghasemi, Mohammad Sadegh Aboutalebi, Rasool Nouri and Fariborz Mokarian

Abstract

This study aimed to compare the non-acid nucleic blood multi-factor panels together and with mammography in terms of sensitivity, specificity, and accuracy in primary breast cancer detection (I, II, III, and IV). We systematically reviewed studies assessing non-acid nucleic blood tumor markers panels' diagnostic value in both healthy women and patients (before any anticancer treatment) for the detection of primary breast cancer. Out of the 2358 titles initially identified, 12 studies and 9 panels were included in the network meta-analysis. Panels I (MSA + B2m) and J (GATA3 + E-cadherin) had the highest sensitivity in all stages of primary breast cancer but had no significant difference with mammography. Panels L (MSA + CA15–3) and B (M-CSF+ CA15–3) had the highest specificity in all stages compared to other panels but no remarkable difference with mammography. Panels J (GATA3 + E-cadherin) and I (MSA + B2m) respectively had the highest accuracy in primary breast cancer detection but no considerable difference with mammography in terms of accuracy. Panel J, including GATA3 + E-cadherin, demonstrated a higher diagnostic value for primary breast cancer detection (I, II, III, and IV) than the rest of the panels.

Keywords: primary breast cancer, blood tumor markers, timely diagnosis, sensitivity and specificity, multi-factor panels, network meta-analysis

1. Introduction

Based on our previous study [1], the necessity of a noninvasive, accessible, costeffective, and reliable method for breast cancer detection based on blood factors was proved. Furthermore, blood multi-factor panels can be the best choice for such a method thanks to improving the sensitivity and specificity of cancer detection considerably compared to the individual state. In that study [1], we had determined the best non-acid nucleic blood multi-factor panels for breast cancer detection in early stages and locoregional breast cancer (I, II, and III). In this brief study, however, we compared the best non-acid nucleic blood multi-factor panels in primary breast cancer detection (I, II, III, and IV) by conducting a network meta-analysis. In fact, this study aimed to offer new insight into the diagnostic value of the best panels of non-acid nucleic blood tumor markers to detect primary breast cancer along all stages not only in early stages. The breast malignancy that emerges and can be diagnosed for the first time is named primary breast cancer, and if it recurs after primary treatment including surgery, chemotherapy, hormone therapy, and radiotherapy individually or collectively, it will be named secondary breast cancer [2]. Primary breast cancer comprises locoregional (I, II, III) and metastatic stages (IV) [3].

2. Materials & method

The systematic reviews of the observational studies were conducted based on PRISMA guidelines (Preferred Reporting Items for Systematic Reviews and Metaanalysis) [4]. Eligibility criteria, search strategy (supplementary material 1 B), databases, study selection, data extraction, and statistical analysis conformed to our former study [1]. The difference is that, in this brief study, we systematically reviewed the studies that have simultaneously assessed several tumor markers in the form of a panel to diagnose and detect breast cancer in all stages of primary breast cancer (I, II, III, and IV).

The included panels were **B**: M-CSF + CA15–3, **C**: VEGF + CA15–3, **D**: VEGF + M-CSF + CA 15–3, **E**: VEGF + M-CSF, **F**: p16+ c-MYC+ P53, **G**: CA15–3 + CEA, **I**: MSA + B2m, **J**: GATA3 + E-cadherin and **L**: MSA + CA15–3.

All these panels were made based on simultaneous measurement of two or three blood tumor markers in patients and healthy people using a compatible linear combination method [5]. Panels (B, C, D, E, F, G) were assessed in more than one study (multiple studies), and panels (I, J, L) were only assessed in one study (single study). We conducted direct and indirect paired comparisons of the sensitivity, specificity, and accuracy of the included blood tumor markers panels for diagnosing primary breast cancer in all stages. All the investigations were conducted in comparison to mammography (M) as the gold standard [6–8], like our previous study (**Figure 1**) [1].

3. Results and discussion

3.1 Study selection

Study selection conformed to our former study [1]. However, in this brief study, among the 54 studies relevant to our research question which contained 86 unique blood tumor markers panels (supplementary material 2) conforming to our eligibility criteria, only 12 studies and 9 panels presented enough data for estimating sensitivity and specificity in all stages (I, II, III, and IV) of primary breast cancer and could be included in the systematic review and network meta-analysis. These 12 studies were similar in terms of pre-analytical procedures and analytical methods (**Table 1**).

All the included and excluded studies are presented in Figure 2.

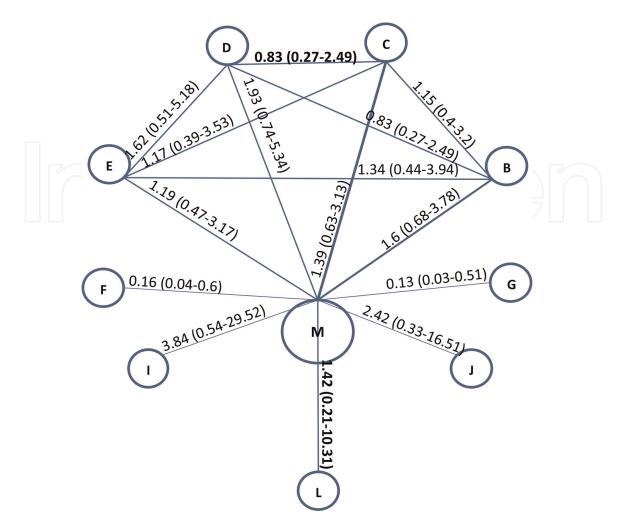


Figure 1.

Multiple comparison of different panels for sensitivity. B: M-CSF + CA15–3, C: VEGF + CA15–3, D: VEGF + M-CSF + CA 15–3, E: VEGF + M-CSF, F: p16+ c-MYC+ P53, G: CA15–3 + CEA, I: MSA + B2m, J: GATA3 + E-cadherin. L: MSA + CA15–3 M = mammography.

Association between diagnosis of primary breast cancer and blood tumor markers panels:

Panels I (MSA + B2m) and J (GATA3 + E-cadherin) had the highest sensitivity in primary breast cancer but did not have noticeable differences with mammography. Panels G (CA15–3 + CEA) and F (p16+ c-MYC+ P53) had the lowest sensitivity than the rest of the panels and mammography as mammography exhibited a remarkably better function than them, with OR = 0.13 and 95% CL (0.04–0.46) and OR = 0.15 and 95% CL (0.04–0.52) (Figure 3a, Table 2). In diagnostic tests, sensitivity had a vital role in screening diseases [21]. As a result, we can claim that the panels which had the highest sensitivity can be promising diagnostic tests in primary breast cancer screening, which included panels I and J in all stages of primary breast cancer. Panels L (MSA + CA15–3) and B (M-CSF + CA15–3) had the highest specificity but did not have remarkable differences with mammography. Panels G (CA15-3 + CEA) and D (VEGF + M-CSF + CA 15–3) had the lowest specificity as mammography demonstrated a superior function in specificity, with OR = 0.06 and 95% CL (0.01–0.39) and OR = 0.06 and 95% CL (0.02–0.19) (Figure 3b, Table 3). Mammography had a better function in specificity than a large number of panels, since it exhibited the highest specificity after panel L with OR = 2.54 and 95% CL (0.1–177.46) in diagnosing

First author and year	Country	Study design	Sample size Clinical and stages population	Panel	Number of panel components	Sensitivity	Specificity	Accuracy	Method of chemical evaluation	Type of sample	Score
S	Poland	Case-	240 I:29	VEGF + CA	2	96.25	65	80.6	ELISA	plasma	11
Zajkowska,		control	Bc:120 II:30	15–3	2	91.25	67.5	79.3	CMIA		
М			B:60 III:31	M-CSF + CA	3	96.25	57.5	76.8			
2016			H:60 IV:30	15–3	2	90	76	83			
[9]			Median age	VEGF + M-	VEGF	76.25	85				
			(range) 54	CSF + CA 15-	M-CSF	60	90				
			(34–72)	3 VEGF+ M- CSF*	CA 15–3	83.75	75				
Sacks, N. P.	Australia.	Case-	131 I/ II:34	MSA + CA15–	2	84	100	89.2	ELISA	serum	11
1987		control	Bc:72 III/ IV:38	3							
[10]			B:13								
			H:46								
Liu, Y.	China	Case-	248 I/ II:57	*p16+ c-	3	30	90	65.3	ELISA	serum	10
2017		control	Bc102 III/ IV:45	MYC + TP53	p16	27.5	90				
[11]			H146		c-MYC	11.8	90				
			$\textbf{50.88} \pm \textbf{7.12}$		TP53	24.5	90				
Molina,	Spain	Case-	292 I/ II:118	Ca15.3	2	29.2	90	51.2	ELISA	serum	9.5
Rafael		control	Bc186 III/ IV:68	+	Ca15.3	15.6					
1998[12]			B56	CEA	CEA	18.3					
			H50						70		
Ławicki,	Poland	Case-	200 I/ II/ III:77	VEGF	2	84	90	87	ELISA	plasma	11
Sławomir		control	Bc100 IV:23(with	+CA15-3	VEGF	61	96				
2016			B50 metastases)		CA 15–3	65	96				
[13]			H50								
			48(20–78)						S P		
Lawicki, S.	Poland	Case-	200 I/ II/ III:77	VEGF+ CA	2	83	90	86.5	ELISA	plasma	11.5
2017		control	Bc100 IV:23	15–3	VEGF	60	95				
[14]			B50		CA 15–3	64	95				
			H50								
			48(20–78)								

4

First author and year	Country	Study design	Sample size Clinical and stages population	Panel	Number of panel components	Sensitivity	Specificity	Accuracy	Method of chemical evaluation	Type of sample	
Tjandra, JJ 1988 [15]	Australia	Case– control	161 I:32 Bc109 II:24 B31 IIII/ IV:53 H21 III	MSA + B2m	2 MSA B2m	93 88 39	90 95 90	90.3	Radioimmunoassay + ELISA	serum	12
Ławicki, S 2013 [16]	Poland	Case– control	190 I:25 Bc110 II:35 B40 III:25 H40 IV:25(with 44 (30–78) metastases)	M- CSF + CA15– 3	2 M-CSF CA 15–3	85 60 53	90 95 95	87.1	ELISA	plasma	11
Ławicki, Sławomir 2013 [17]	Poland	Case- control	200 I/ II/ III/75 Bc100 IV:25(with B50 metastases) H50 51 (40–70)	VEGF+ CA 15–3 M-CSF+ CA 15–3 VEGF+ M- CSF VEGF+ M- CSF+ CA 15– 3	2 2 3 VEGF M-CSF CA 15-3	61 67 63 75 44 53 36	86 86 78 92 94 92	73.5 76.5 74.5 76.5	ELISA	plasma	11
Luo, M. 2019 [18]	China	Case– control	200 I/ II:47 Bc120 III/ IV:73 H80 59.88 ± 9.05 (32-70)	GATA3 + E- cadherin GATA3 E-cadherin	2	90 87.5 82.5	91.7 73.3 87.5	90.6	ELISA	serum	11.5
Looi, Koksun 2006 [19]	China	Case- control	123 Bc41 H82 Multi cancer	p16 + c-MYC + P53	3	43.9	97.6	79.6	ELISA	serum	8

ъ

First author and year	Country	Study design	Sample size and population	Clinical stages	Panel	Number of panel components	Sensitivity	Specificity	Accuracy	Method of chemical evaluation	Type of sample	Score
Guadagni,	Italy	Case-	2191	I:392	CEA	2	39	85	54	RIA kit	serum	9.5
Fiorella		control	BC 1453	II:562	+	CA 15.3	33					
2001			B738	III:153	CA 15.3	CEA	16.7					
[20]			mean age,	IV:48								
			57 years	Metastatic:240								
			(range 25–	Local								
			97 years	recurrence:58								

B: benign; H: Healthy; ELISA: the enzyme-linked immunosorbent assay; CMIA: luminescent microparticle immunoassay; RIA: radioimmunoassay. The scoring system based on the CASP checklist (specified for diagnostic studies) was applied to all studies. The sensitivity, specificity, and accuracy of all studies were evaluated in all stages of primary breast cancer (I, II, III, and IV). *Based on linear combination (5).

Table 1.

6

Characteristics of articled included in network meta-analysis.

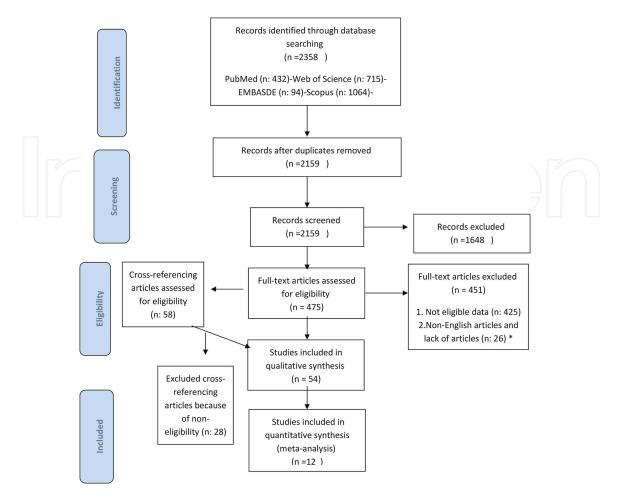


Figure 2.

Flow diagram of included and excluded articles. *Although we sent emails to articles' authors to get their full texts, we did not receive any answers.

primary breast cancer. Panels J (GATA3 + E-cadherin) and I (MSA + B2m) possessed the highest accuracy in primary breast cancer but did not show significant differences with mammography. Panel L (MSA + CA15–3) did not demonstrate considerable differences with panel I; therefore, we could consider them approximately similar regarding accuracy. Panels G (CA15–3 + CEA) and F (p16+ c-MYC+ P53) possessed the lowest accuracy in primary breast cancer as mammography exhibited a considerably superior function in accuracy, with OR = 0.15 and 95% CL (0.07–0.3) and OR = 0.37 and 95% CL (0.17–0.74) (**Figure 3c, Table 4**).

The best panels based on total function: J: GATA3 + E-cadherin, I: MSA + B2m.

In diagnosing primary breast cancer, panels J and I exhibited the highest accuracy and total function compared to other panels. Overall, we recommend panel J because it had an even better function in accuracy than panel I, despite being minor (**Table 1**) and its study had a larger sample size (200). Panel J was made of GATA3 and E-cadherin. GATA3 is a transcription factor that plays a crucial role in the development and progression of breast cancer and can reverse the epithelial-mesenchymal transition. It also regulates the proliferation, differentiation, and development of cells. E-cadherin is a member of the cadherin family mainly expressed in epithelial cells. E-cadherin mediates the adhesion of allogeneic epithelial cells and plays a key role in epithelial cell aggregation and adhesion. Studies have demonstrated that the expression of cadherin is closely related to the invasion of breast cancer [18].

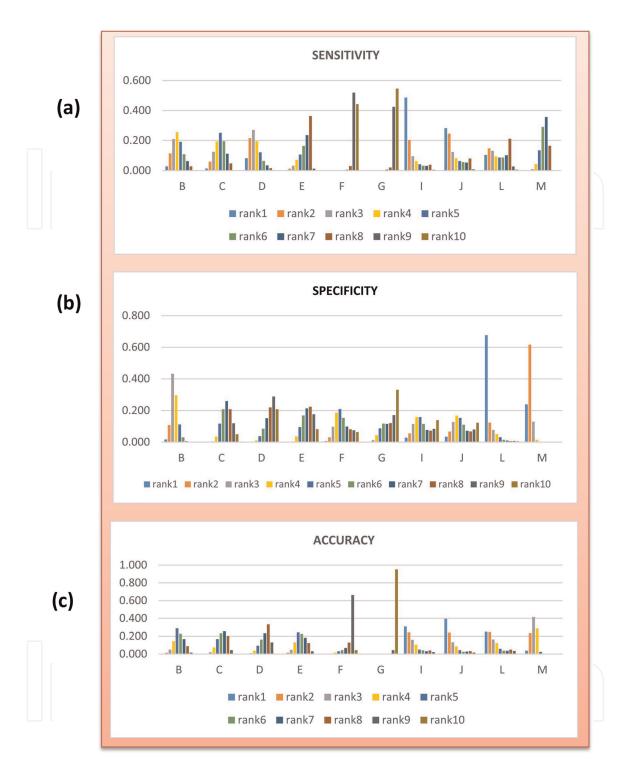


Figure 3.

Estimated rank probability of all panels' sensitivity, specificity, and accuracy. B: M-CSF + CA15–3, C: VEGF + CA15–3, D: VEGF + M-CSF + CA 15–3, E: VEGF + M-CSF, F: p16+ c-MYC+ P53, G: CA15–3 + CEA, I: MSA + B2m, J: GATA3 + E-cadherin. L: MSA + CA15–3 M = mammography.

4. Conclusion

In conclusion, panel J including GATA3 + E-cadherin with a sensitivity of 90 and specificity of 91.7 demonstrated a higher diagnostic value for primary breast cancer than the rest of the panels as it exhibited higher function in accuracy than mammography, with OR = 1.38 and 95% CL (0.42–4.41), although it was not remarkable. After

	В	С	D	Ε	F	G	Ι	J	L	Μ
В	1									
С	1.18	1								
	(0.42–2.96)									
D	0.83	0.71	1							
	(0.28–2.29)	(0.25–2)								
E	1.73	1.47	2.09	1						
	(0.62–4.83)	(0.56–4.24)	(0.72–							
			6.32)							
F	10.52	9.01	12.73	6.13	1					
	(2.42–	(2.16–	(2.85–	(1.34–						
	46.72)	39.41)	60.57)	27.66)						
G	11.94	10.21	14.57	6.93	1.12	1				
	(2.69–	(2.36–	(3.07–	(1.49–	(0.19–					
	56.3)	46.39)	70.46)	32.44)	6.78)					
Ι	0.44	0.37	0.52	0.25	0.04	0.04	1			
	(0.05-	(0.05–2.95)	(0.06–	(0.03-	(0.01–	(0.01–				
	3.44)		4.22)	2.01)	0.4)	0.37)				
J	0.64	0.54	0.77	0.37	0.06	0.05	1.46	1		
	(0.09–	(0.08–	(0.1–	(0.05–	(0.01–	(0.01–	(0.11–			
	4.78)	4.16)	6.26)	2.95)	0.57)	0.52)	22.19)			
L	1.13 (0.16–	0.98 (0.14–	1.37	0.65	0.11	0.09	2.6	1.79	1	
	7.97)	6.79)	(0.19–	(0.09–	(0.01–	(0.01–	(0.18–	(0.13–		
			10.07)	4.78)	1)	0.87)	36.26)	22.86)		
М	1.59 (0.69–	1.35 (0.65–	1.91	0.91	0.15	0.13	3.63	2.47	1.4	1
	3.53)	2.94)	(0.78–	(0.38–	(0.04–	(0.04–	(0.55–	(0.37–	(0.24–	
			4.88)	2.26)	0.52)	0.46)	25.11)	15.96)	8.3)	

B: M-CSF + CA15–3, C: VEGF + CA15–3, D: VEGF + M-CSF + CA 15–3, E: VEGF + M-CSF, F: p16+ c-MYC+ P53, G: CA15–3 + CEA, I: MSA + B2m, J: GATA3 + E-cadherin L: MSA + CA15–3 M = Mammography.

Table 2.

Relative effects and its 95% credible interval of all pairwise panels for sensitivity based on Bayesian network metaanalysis method.

	В	С	D	E	F	G	Ŋ	I	J	L	1
В	1										
С	4.75 (1.41– 16.11)	1									
D	6.86 (1.97– 25.88)	1.46 (0.42– 5.27)	1	_							
E	5.19 (1.42– 18.95)	1.08 (0.31– 3.99)	0.75 (0.2– 2.74)	1							
F	2.8 (0.33– 24.19)	0.58 (0.07– 4.87)	0.4 (0.04– 3.57)	0.53 (0.06– 4.86)	1						

	В	С	D	Ε	F	G	Ι	J	L	Μ
G	6.82	1.43	1 (0.11–	1.33	2.62	1				
	(0.76–	(0.16–	9.29)	(0.14-	(0.17–					
	63.76)	12.82)		12.7)	35.24)					
Ι	2.74	0.57	0.39	0.53	1 (0.04–	0.39	1			
	(0.16–	(0.03-	(0.02-	(0.03-	30.25)	(0.02-				
	61.57)	12.81)	8.77)	12.6)		13.06)				
J	2.4	0.51	0.34	0.46	0.85	0.34	0.87	1		
	(0.14–	(0.03-	(0.02–	(0.03–	(0.03-	(0.01–	(0.02-			
	48.49)	9.19)	6.66)	9.64)	22.94)	9.86)	42.75)			
L	0.16	0.03	0.02	0.03	0.06	0.02	0.06	0.07	1	
	(0.01-	(0.01-	(0.01–	(0.01–	(0.01-	(0.01–	(0.01–	(0.01-		
	4.92)	1.04)	0.72)	0.89)	2.68)	0.96)	3.68)	4.26)		
М	0.42	0.09	0.06	0.08	0.15	0.06	0.16	0.18	2.54	1
	(0.14–	(0.03–	(0.02-	(0.02-	(0.02-	(0.01–	(0.01-	(0.01-	(0.1–	
	1.2)	0.24)	0.19)	0.25)	0.96)	0.39)	2.05)	2.3)	177.46)	

B: M-CSF + CA15–3, C: VEGF + CA15–3, D: VEGF + M-CSF + CA 15–3, E: VEGF + M-CSF, F: p16+ c-MYC+ P53, G: CA15–3 + CEA, I: MSA + B2m, J: GATA3 + E-cadherin L: MSA + CA15–3 M = Mammography.

Table 3.Relative effects and its 95% credible interval of all pairwise panels for specificity based on Bayesian network meta-
analysis method.

	В	С	D	Е	F	G	Ι	J	L	Μ
В	1									
С	1.11	1	_							
	(0.65-									
	1.9)									
D	1.24 (0.7–	1.11	1	_						
	2.18)	(0.65–								
		1.97)								
Е	1.03	0.92	0.83	1	_					
	(0.59–	(0.54–	(0.46–							
	1.81)	1.64)	1.46)							
F	1.8 (0.77–	1.61	1.45	1.78	1	-				
	4.42)	(0.7–4)	(0.61-	(0.73–						
	\Box		3.77)	4.23)						
G	4.42	3.98	3.58	4.32	2.45	1				
	(1.92–	(1.74–	(1.44–	(1.79–	(0.85–					
	10.23)	9.25)	8.49)	10.29)	6.76)					
Ι	0.54	0.48	0.44	0.53	0.3	0.12	1			
	(0.15–	(0.14–	(0.12-	(0.15-	(0.07–	(0.03–				
	1.81)	1.62)	1.48)	1.84)	1.15)	0.47)				
J	0.48	0.43	0.39	0.46	0.26	0.11	0.91	1	_	
	(0.13-	(0.12–	(0.1–	(0.13–	(0.06–	(0.03–	(0.17–			
	1.74)	1.55)	1.43)	1.84)	1.1)	0.43)	4.57)			
L	0.58	0.52	0.47	0.57	0.32	0.13	1.09	1.22	1	
L		1010	(0.13–	(0.16–	(0.08-	(0.03–	(0.22-	(0.25-		
Г	(0.17–	(0.16–	(0.13 -	(0.10-	(0.00-	(0.05	(0.22	(0.25		

	В	С	D	Е	F	G	Ι	J	L	Μ
М	(0.42–	(0.39–	(0.32–	(0.38–	(0.17–	0.15 (0.07– 0.3)	(0.4–	(0.42-	(0.34–	1

B: M-CSF + CA15–3, C: VEGF + CA15–3, D: VEGF + M-CSF + CA 15–3, E: VEGF + M-CSF, F: p16+ c-MYC+ P53, G: CA15–3 + CEA, I: MSA + B2m, J: GATA3 + E-cadherin L: MSA + CA15–3 M = Mammography.

Table 4.

Relative effects and its 95% credible interval of all pairwise panels for accuracy based on Bayesian network metaanalysis method.

panel J, panel I (MSA + B2m) with a sensitivity of 90 and specificity of 90.3 and panel L (MSA + CA15–3) with a sensitivity of 84 and specificity of 100 had the best function in primary breast cancer detection than the rest of the panels. However, more experimental studies are required with larger samples, on different populations, and using other chemical measurement methods to verify these results.

Acknowledgements

This study was supported by Research Institute and Cancer Prevention Research Center, Isfahan University of Medical Sciences.

Author contributions

Vahid Raja had the idea for the research. The literature search was performed by Vahid Raja, Mohammad Sadegh Aboutalebi, and Rasool Nouri. The data analysis was performed by Marjan Mansourian and Khojaste Ghasemi. The article was drafted by Vahid Raja and Ziba Farajzadegan. The article was critically revised by Vahid Raja, Ziba Farajzadegan, and Fariborz Mokarian.



Data availability

The data used to support the findings of this study are available from the corresponding author upon request.

Ethical statement

Our study did not require an ethical board approval because it did not contain human or animal trials.

Supplementary material

Including traditional meta-analysis of all panels, nod-splitting analysis of inconsistency for sensitivity, specificity and accuracy, ranking of different panels in sensitivity, specificity and accuracy, the search strategy for each data base, and 54 studies were identified relevant to our research question.

Author details

Vahid Raja^{1*}, Ziba Farajzadegan^{2*}, Marjan Mansourian³, Khojaste Ghasemi³, Mohammad Sadegh Aboutalebi⁴, Rasool Nouri⁵ and Fariborz Mokarian⁶

1 Clinical Laboratory Sciences, Amin Hospital, Isfahan university of Medical sciences, Isfahan, Iran

2 Community and Preventive Medicine, Isfahan University of Medical Sciences, Isfahan, Iran

3 Department of Biostatistics and Epidemiology, School of Public Health, Isfahan University of Medical Sciences, Isfahan, Iran

4 Faculty of Nursing and Midwifery, Isfahan University of Medical Sciences, Isfahan, Iran

5 Department of Medical Library and Information Sciences, Isfahan University of Medical Sciences, Isfahan, Iran

6 Hematology and Oncology Department of Internal Medicine, Isfahan University of Medical Sciences, Isfahan, Iran

*Address all correspondence to: www.mahantajhizs@gmail.com and farajzadegan@med.mui.ac.ir

IntechOpen

© 2022 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

References

[1] Raja V, Farajzadegan Z, Mansourian M, Ghasemi K, Aboutalebi MS, Nouri R, et al. Diagnostic value of nonacid nucleic blood tumor marker panels in early diagnosing breast cancer: A systematic review and network meta-analysis. Disease Markers. 2022;**2022**:15

[2] Sadler C, Goldfarb M. Comparison of primary and secondary breast cancers in adolescents and young adults. Cancer. 2015;**121**(8):1295-1302

[3] Zackrisson S, Cardoso F, Guidelines E. Clinical practice guidelines primary breast cancer: ESMO clinical practice Guidelines for diagnosis, treatment and follow-up† clinical practice guidelines. Annals of Oncology. 2015;**26**(5):8-30

[4] Moher D, Liberati A, Tetzlaff J, Altman DG, Group* P. Preferred reporting items for systematic reviews and meta-analyses: The PRISMA statement. Annals of internal medicine. 2009;**151**(4):264-269

[5] Pepe MS, Thompson ML. Combining diagnostic test results to increase accuracy. Biostatistics. 2000;1(2):123-140

[6] Yankaskas BC, Haneuse S, Kapp JM, Kerlikowske K, Geller B, Buist DS, et al. Performance of first mammography examination in women younger than 40 years. JNCI: Journal of the National Cancer Institute. 2010;**102**(10):692-701

[7] Sinclair N, Littenberg B, Geller B, Muss H. Accuracy of screening mammography in older women. American Journal of Roentgenology. 2011;**197**(5):1268-1273

[8] Ontario HQ. Screening mammography for women aged 40 to 49 years at average risk for breast cancer: An evidence-based analysis. Ont Health Technol Assess Ser. 2007;7(1):1-32

[9] Zajkowska M, Głażewska EK, Będkowska GE, Chorąży P, Szmitkowski M, Ławicki S. Diagnostic power of vascular endothelial growth factor and macrophage colonystimulating factor in breast cancer patients based on ROC analysis. Mediators of Inflammation. 2016;**2016**:8

[10] Sacks N, Stacker S, Thompson C, Collins J, Russell I, Sullivan J, et al. Comparison of mammary serum antigen (MSA) and CA15-3 levels in the serum of patients with breast cancer. British journal of cancer. 1987;**56**(6):820-824

[11] Liu Y, Liao Y, Xiang L, Jiang K, Li S, Huangfu M, et al. A panel of autoantibodies as potential early diagnostic serum biomarkers in patients with breast cancer. International Journal of Clinical Oncology. 2017;**22**(2):291-296

[12] Molina R, Jo J, Filella X, Zanon G, Pahisa J, Muñoz M, et al. c-erbB-2 oncoprotein, CEA, and CA 15.3 in patients with breast cancer: Prognostic value. Breast cancer research and treatment. 1998;**51**(2):109-119

[13] Ławicki S, Zajkowska M, Głażewska EK, Będkowska GE, Szmitkowski M. Plasma levels and diagnostic utility of VEGF, MMP-9, and TIMP-1 in the diagnosis of patients with breast cancer. Onco Targets and therapy. 2016;**9**:911

[14] Ławicki S, Zajkowska M, Głażewska EK, Będkowska GE, Szmitkowski M. Plasma levels and diagnostic utility of VEGF, MMP-2 and TIMP-2 in the diagnostics of breast cancer patients. Biomarkers. 2017;**22**(2): 157-164 [15] Tjandra J, McLaughlin P, Russell I, Collins J, McKenzie I. Comparison of mammary serum antigen (MSA) with β 2-microglobulin (β 2M) and carcinoembryonic antigen (CEA) assays in patients with breast cancer. European Journal of Cancer and Clinical Oncology. 1988;**24**(10):1633-1640

[16] Ławicki S, Będkowska G, Wojtukiewicz M, Szmitkowski M. Hematopoietic cytokines as tumor markers in breast malignancies. A multivariate analysis with ROC curve in breast cancer patients. Advances in Medical Sciences. 2013;**58**(2):207-215

[17] Ławicki S, Będkowska GE, Szmitkowski M. VEGF, M-CSF and CA 15-3 as a new tumor marker panel in breast malignancies: A multivariate analysis with ROC curve. Growth Factors. 2013;**31**(3):98-105

[18] Luo M, Huang Y, Huang J, Huang S, Wei L, Zhang Y, et al. Evaluation of the value of GATA3 combined with E-cadherin in the diagnosis of breast cancer. Journal of BU ON: Official Journal of the Balkan Union of Oncology. 2019;**24**(3):1038-1044

[19] Looi K, Megliorino R, Shi F-D, Peng X-X, Chen Y, Zhang J-Y. Humoral immune response to p16, a cyclindependent kinase inhibitor in human malignancies. Oncology Reports. 2006; **16**(5):1105-1110

[20] Guadagni F, Ferroni P, Carlini S, Mariotti S, Spila A, Aloe S, et al. A re-evaluation of carcinoembryonic antigen (CEA) as a serum marker for breast cancer: A prospective longitudinal study. Clinical Cancer Research. 2001; 7(8):2357-2362

[21] Stellman SD. Book review:Epidemiology. In: Gordis L, editor.Gordis Epidemiology, 6th ed.Philadelphia, PA, US: Saunders; 2009.

p. 2010. ISBN: 978-0-323-55229-5. Copyright © 2019 by Elsevier, Inc. All rights reserved

