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Chapter

Retinal Disorders in Humans and Experimental ALS Models

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Abstract

Amyotrophic lateral sclerosis (ALS) is a rapidly progressive neurodegenerative disease that severely impairs the patient's mobility, as it mainly affects the upper and lower motor neurons in the spinal cord. In addition, alterations have also been demonstrated in different parts of the central nervous system (CNS), such as the brain and brainstem. The retina is a projection to the brain and is considered as a "window" to the CNS. Moreover, it is possible to use the retina as a biomarker in several neurodegenerative diseases, even in the absence of major visual impairment. Classically, it was thought that the eyes were not affected in ALS, with respect to extraocular muscles, whereas the remainder of the muscles of the body were distressed. Nevertheless, retinal changes have recently been found in this pathology and could help in diagnosis, follow-up, and even monitoring therapies in this disease.

Keywords: amyotrophic lateral sclerosis, ALS, retina, animal models, SOD1, microglia, protein aggregates, axon pathology, neurodegeneration, neuroinflammation

1. Introduction

Amyotrophic lateral sclerosis (ALS) is the most common progressive motor neuron disease, accounting for 80–90% of all motor neuron diseases cases [1]. Worldwide, the incidence per 100,000 people ranges from 0.3 to 2.5 cases per year [2–5]. Only 10% of the cases are familial [6], ranging from 2 to 15% depending on the population [7], whereas 90% of the cases are sporadic or seemingly sporadic. Overall, both the incidence [8] and the prevalence [9] of ALS vary according to location and race. ALS is more common in men than in women, with a ratio of 1.5:1 [5].

This neurodegenerative disease is rapidly progressive with a typical combination of symptoms of both upper motor neurons (UMNs) and lower motor neurons (LMNs) in different degrees, causing muscle fiber atrophy, which seriously affects the patient's mobility and quality of life [2, 4, 10]. ALS comprises overactive reflexes, as well as muscle weakness and stiffness, and it also involves the swallowing, speech, and respiratory muscles [10–12]. In fact, patients usually die within 2–3 years from diagnosis, frequently due to respiratory failure [5, 13]. The disease usually has a spinal onset, beginning in the extremities and spreading to the rest of the body; however, one in every four patients has a bulbar onset, which has a worse prognosis [4]. ALS is a heterogeneous disease with asymmetrical onset and spreading of UMN and LMN dysfunction, which makes its classification very complex [14]. In addition, no single specific test exists for ALS diagnosis; it is a diagnosis of exclusion based on the initial symptoms, the progression of the disease, and tests to eliminate overlapping conditions.

Although ALS has been considered an exclusively motor disease, over the last few years, several studies have focused on assessing the possible participation of nonmotor areas of the central nervous system (CNS) in this illness. Actually, neuroimaging tests have shown an overall reduction in brain volume, with a loss of focal gray matter and regional white-matter alterations [15–20]. The alteration of these areas leads to cognitive and behavioral changes [16, 18]. During the course of the disease, it has been found that 50% of ALS patients have some degree of cognitive impairment, mainly featuring executive dysfunction and mild memory loss [15, 21].

Classically, it was thought that the eyes were not affected in this disease, with respect to the eye motor muscles, whereas the remaining muscles of the body were affected [22]. However, some studies have found abnormal ocular movements in these patients [23–28]. Nevertheless, this classical concept did not refer to the retina or the optic nerve. Actually, these patients have demonstrated not only abnormal evoked potentials [29–33] but also astrogliosis in nonmotor areas, specifically in the occipital area [34]. Even a significant interocular difference of the P100 in ALS patients was demonstrated in a study of visual evoked potentials [33], similar to the existing asymmetry in the CNS of these patients [14]. Some researchers have also analyzed changes in the visual pathway (a nonmotor neuron area) using optical coherence tomography (OCT) in ALS patients [35–44], finding different changes in the retina and optic nerve, some with contradictory results, stressing the importance of classifying patients by both stage and type of ALS, given the high heterogeneity of the disease.

The retina is considered as an open window to the CNS, and it is possible to use it as a biomarker in multiple neurodegenerative diseases, whether or not there is visual impairment. In recent years, many studies have emphasized the importance of the retina in the diagnosis and monitoring of neurodegenerative diseases, with various pieces of evidence highlighting its value as a biomarker [45–57]. However, what was not so evident was the possible involvement of the retina in neuromuscular diseases, which are chronic progressive neurological diseases, such ALS, that predominantly affect the spinal cord, whereby the neurological involvement is far from the visual pathway.

The purpose of this review is to analyze the retinal changes that have been described in different animal models in this disease, to compare them with each other and to correlate them with the changes described in humans to highlight the possible role of the retina as a biomarker in this disease.

2. Retinal histopathological studies in amyotrophic lateral sclerosis patients and experimental models

ALS is a neurodegenerative disease, which shares some pathophysiological mechanisms common to other diseases of the CNS, such as vascular pathology, glutamate excitotoxicity, fragmentation, aggregation, and functional abnormalities of the mitochondria, impaired retrograde and anterograde axonal structure and transport, increased free-radical and oxidative stress, protein aggregation, and neuroinflammation [12, 58–61]. However, studies in the retina are scarce and have focused only on four such mechanisms, as described below.

2.1 Histopathological studies in ALS demonstrating intraretinal protein inclusions

(Table 1, Figure 1) Accumulated and altered proteins can interfere with neuronal traffic or can abduct proteins that are essential for proper neuronal functioning causing neurotoxicity [62]. The ubiquitin proteasome system plays an important role in ALS, with reactive ubiquitin inclusions being characteristic of this pathology [5, 63]. Among them, TDP-43 and p62 proteins are specifically indicative of ALS. These inclusions, which are positive for P62 and negative for TDP-43, have been demonstrated in the brain, hippocampus, and cerebellum in ALS patients [64, 65].

There are scarce studies that have focused on the histopathology of retinal tissue in both ALS patients and animal models of mammals with ALS. Actually, the first histopathological analysis in the retina was performed in 2014 on a patient with the C9orf72 mutation. In this study, protein intracytoplasmic p62-positive and pTDP43negative perinuclear aggregates, typical of ALS/frontotemporal dementia (FTD), were observed in the inner nuclear layer (INL) of the retina [66]. Both the poly-(GA) n dipeptide repeats and ubiquitin in the retina were positively stained for p62, similar to the perinuclear inclusions localized in the brains, specifically in the dentate gyrus, of patients with this mutation [66]. The authors suggested that most of the p62-positive inclusions found were likely placed within the cones of bipolar cells (OFF bipolar cells) and between amacrine and horizontal cells, because they were also stained with GLT-1 and recoverin; in addition, these retinal deposits could be related to the contrast sensitivity impairment manifested by the patient [66]. Moreover, Volpe et al. [67] analyzed two retinas from ALS patients with C9orf72 mutations and demonstrated (i) specific p62 inclusions mostly in the INL (94.9%) and in a smaller proportion in the retinal ganglion cell layer (GCL) (5.1%) in one patient, and (ii) ganglion cell axonal atrophy specifically in the papillomacular bundle in the second patient. On the other hand, abundant positive ubiquilin 2-positive inclusions were also shown in a transgenic mice experimental model with mutant UBQLN2, mostly in the inner plexiform layer (IPL), with a smaller amount in the outer plexiform layer (OPL) and a scarce amount in the GCL. This ubiquilin 2 aggregation in the layers of the retina with more synapses is associated to the ubiquilin 2 accumulation in the dendritic spines of the hippocampus, and it may also be related to the dementia observed in this experimental model. Furthermore, few ubiquilin 2-positive aggregates were detected between the neurosensorial retina and the retinal pigment epithelium, whose appearance was analogous to that of drusen [67]. Similarly, in patients with FTD and progranulin deficiency, lipofuscin deposits were found, sometimes associated with subretinal drusen-like aggregates [68]. Retinal thinning in these patients was detected by OCT before symptoms, suggesting that the eye is affected in programulin-deficient frontotemporal dementia disease [69].

Eye degeneration was reported in an ALS *Drosophila* model that expressed C9orf72 repeat expansion. The expansion of a noncoding GGGGCC hexanucleotide repeat of the C9orf72 gene on chromosome 9p21 is the most common point mutation in familiar ALS, which generates dipeptide repeat proteins that aggregate in the brain. It is note-worthy that some synthesized compounds revealed a significant biological effect by blocking the neurodegeneration of fly retina at different efficacy levels and upgrading

Mechanism	Author and year	Retinal tissue	Main retinal findings	Other comments
Protein inclusions	Fawzi et al. 2014	One patient with ALS secondary to a C9orf72 mutation	Protein intracytoplasmic p62 ⁺ /TDP43- perinuclear aggregates in the INL	Most of the p62-positive inclusions found were likely placed within OFF bipolar cells and between amacrine and horizontal cells; they may have been responsible for the contrast sensitivity impairment in this patient
	Volpe et al. 2015	UBQLN2P497H TG mice	Ubiquilin2 ⁺ inclusions mostly in the IPL, with a smaller amount in the OPL and in the GCL	Drusen-like ubiquilin 2-positive aggregates at the level of the sub-RPE space
		Two patients with ALS secondary to a C9orf72 mutation	First patient: Specific p62 ⁺ inclusions: 94.9% in the INL and 5.1% in the GCL	Second patient: ganglion cell axonal atrophy specifically in the papillomacular bundle
	Azoulay- Ginsburg et al. 2021	ALS fly <i>Drosophila</i> model expressing C9orf72 repeat expansion	Eye neurodegeneration	Compounds 9 and 4 of chemical chaperones blocked and upgraded the eye neurodegeneration
Neuroinflammation	Ringer et al. 2017	TG mouse model SOD1G93A	hSOD1 ⁺ vacuoles in the dendrites of excitatory retinal neurons in the IPL, with hardly any in the GCL and INL	No signs of activation of either the astroglia or the microglia of the retina
	Cho et al. 2019	Mouse model of ALS devoid of Ranbp2	↑ Amoeboid forms and microglial cells surrounding the RGCs	Hypertrophy in RGCs + ↑ metalloproteinases in RGCs + axonopathy in the optic nerve
Int	Rojas et al. 2021	TG mouse model SOD1G93A (late stage)	Microglial cells activation in retinal tissue	Loss of RGCs
			Cell thickening in the area occupied by each microglial cell	
			↑ Microglial arborization in the area with hyper- ramifications in the inferior sector of the OPL	M1 phenotype or proinflammatory state of microglia: neurotoxic
			Retractions of cells processes + migration and clustering of cells in some areas of the retina	

Mechanism	Author and year	Retinal tissue	Main retinal findings	Other comments
Retinal spheroids and axon pathology	Sharma et al. 2020	Retinal sections of 10 postmortem eyes from ALS patients	PAS⁺ spheroids (> 9.07 µm in diameter) in the RNFL	No significant correlation of retinal spheroids and axon pathology with clinical characteristics of the ALS patients (age at death, gender, disease duration, mode of disease onset, ALSFRS-R, and rate of disease progression)
			P-NF ⁺ spheroids (8 to 15 μm in diameter) in the peripheral and pRNFL	
			NP-NF ⁺ spheroids (7 to 10 μm in diameter) in the RNFL	
			↑ NP-NF signal in the RNFL and IPL	
Vasculopathy	Abdelhak et al. 2018	34 ALS patients with clinically diagnosed ALS who underwent an OCT	The outer wall thickness of retinal vessels was thicker in ALS patients than in controls	Thinning of the ONL, suggesting a possible impairment of rod and cone function
			There was also no correlation between the vessel measurements and clinical parameters	The whole retinal thickness was negatively correlated with the ALSFRS-R

ALS: amyotrophic lateral sclerosis; FTD: frontotemporal dementia; INL: inner nuclear layer; UBQLN2P497H: dysfunctional ubiquilin 2; TG: transgenic; IPL: inner plexiform layer; OPL: outer plexiform layer; ONL: outer nuclear layer; INL: inner nuclear layer; GCL: ganglion cell layer; sub-RPE: subretinal pigment epithelium; hSOD1: human superoxide dismutase 1; Ranbp2: RAN-binding protein 2; RGCs: retinal ganglion cells; PAS: periodic acid Schiff; P-NF: phosphorylated form of neurofilament; NP-NF: non-phosphorylated form of neurofilament; RNFL: retinal nerve fiber layer; OCT: optic coherence tomography; ALSFRS-R: ALS Functional Rating Scale—Revised.

Table 1.

Retinal findings in ALS animal models and patients.

this eye degeneration. The most active chemical chaperones were compound 9, which is a peptide derivative targeted to the endoplasmic reticulum, and compound 4, which is targeted to the lysosome. Consequently, both might be used as a new class of drug candidates to treat ALS and other protein misfolding disorders [70].

2.2 Histopathological studies in ALS and neuroinflammation in retinal tissue

(Table 1, Figure 1) Neuroinflammation is a pathophysiological mechanism, which involves the activation of astroglial and microglial cells, and it occurs in many neurodegenerative diseases, such as Parkinson's disease, Alzheimer's disease, ALS, and glaucoma [62]. Microglial cells are the macrophages of the CNS and have the ability to respond to injury by becoming activated; they can proliferate, migrate, and change shape, acquiring an amoeboid appearance in the most active state [62, 71]. On the one hand, in an attempt to protect against damage, microglial cells can secrete proinflammatory molecules, such as interferon γ or interleukin (IL)-1 β [62]. Nonetheless,



Figure 1.

Summary of retinal changes in amyotrophic lateral sclerosis. (A) Healthy retina. (B) Retinal changes in ALS. Most of retinal changes are mainly detected in the inner layers: (i) ganglion cell loss; (ii) activated microglia in outer plexiform layer (OPL) and inner layer complex (ILC) (constituting an inner plexiform layer and a nerve fiber-ganglion cell layer); (iii) p62-positive and TDP43-negative protein aggregates mostly in the inner nuclear layer with some in the ganglion cell layer (GCL); (iv) ubiquilin 2-positive inclusions mostly in the inner plexiform layer (IPL), with a smaller amount in the outer plexiform layer (OPL) and in the GCL; (v) periodic acid Schiff (PAS)-positive spheroids in the retinal nerve fiber layer (RNFL) and phosphorylated form of neurofilament (P-NF)-positive spheroids in the in the peripheral and peripapillary RNFL; (vi) non-phosphorylated form of neurofilament (NP-NF)-positive spheroids in the RNFL; (vii) hSOD1-positive vacuoles in in the IPL, with hardly any in the GCL and inner nuclear layer (INL).

uncontrolled activation of the M1 phenotype can lead to a state of chronic inflammation, which can induce neuronal death. On the other hand, microglial cells can also secrete anti-inflammatory molecules, such as IL-10 and the enzyme arginase 1 (Arg1), in order to control inflammation, repair tissue, and improve neuronal survival [62, 71–73]. Consequently, activated microglia can acquire two different activation phenotypes: an M1 or proinflammatory phenotype vs. an M2 or anti-inflammatory phenotype, both of which can be influenced by molecules derived from surrounding cells such as astrocytes [62, 74]. Astrocytes are glial cells of ectodermal origin that perform numerous functions for neuronal survival [75], such as maintenance of the volume and composition of the extracellular space, maintenance of the blood-brain barrier, and regulation of synaptic transmission [76], as well as metabolic maintenance and neuronal survival [77, 78]. When astrocytes are damaged and consequently activated, "astrogliosis" occurs [79]. If this astrogliosis is severe, a glial scar may form [75]. Reactive astrocytes can interact with microglia and neurons and can impair the function of neurons after an injury [80].

Astrocyte activation [81], microglial activation [82], and the appearance of lymphocytes [83] have been found in animal models of ALS (with SOD1 mutations) and in ALS patients. In ALS there are reactive microglia and astrocytes, which can result in motor neuron injury and subsequent death [73, 74, 81]. The SOD1G93A mouse model is one of the most suitable and widely used for preclinical studies in ALS, attributable to the animals having an analogous phenotype to patients. These animals develop limb paralysis due to the loss of motor neurons in the spinal cord, with a reduced lifetime of 150 days [84]. Microglial activation also occurs in ALS, as observed in SOD1-mutated mice and in spinal cord samples from ALS patients, which could exacerbate neuronal damage [73, 74, 81, 85]. In fact, it has been shown that exogenous extracellular mutation of SOD1G93A is not directly toxic to motor neurons, but requires microglial activation for toxicity in primary motor neuron and glia cultures [86]. Furthermore, in SOD1 transgenic mice, activated astrocytes and microglia have been shown to contribute to disease progression but not to disease

onset [87-89]. In ALS, microglial activation and proliferation have been observed in areas of significant motor neuron loss, such as the motor cortex, brainstem motor nucleus, corticospinal tract, and ventral horn of the spinal cord [90–94], as well as in areas with mild degeneration [95]. Precisely, in postmortem spinal cord analysis of patients with advanced stages of ALS, reactive astrocytes were found in the dorsal and ventral horn of the spinal cord [96] and in the gray [97] and white matter [34] of the cerebral cortex. Similarly, reactive microglia were found in the motor nucleus of the brainstem, motor cortex, corticospinal tract, and ventral horn of the spinal cord [91]. Reactive microglia were also observed in vivo using the PET imaging technique C-PK11195, finding a close relationship between microglial activation and upper motor neuron damage, but not lower motor neuron injury [98]. Moreover, in the SOD1 model, it was confirmed that overexpression of the SOD1 mutation in glial cells contributes to motor neuron damage, and that the degree of neuronal injury depends on the degree of glial cell pathology [99]. Microglial cells of SOD1-mutated mice suffer different degrees of morphological changes from resting to macrophagic amoeboid forms [91]. Lastly, symptomatic SOD1 transgenic mice also have increased numbers of microglial cells, mainly due to the proliferation of resident microglia [100].

Bearing in mind all of the above, both the microglia and the astrocytes play an important dual role in the progression of the ALS. Nevertheless, most studies about the involvement of microglia in ALS have been conducted in the motor cortex, brainstem motor nucleus, corticospinal tract, and ventral horn of the spinal cord. To our knowledge, there are only three studies that investigated the glial cells of the retina in relation to ALS [101–103].

In the first one, a mouse model of ALS devoid of RAN-binding protein 2 (Ranbp2), microglial activation was confirmed. Ranbp2 is a protein, which plays an important role in nucleocytoplasmic transport and whose regulation is affected in both sporadic and familiar ALS [104]. In this ALS mouse model, there was microglial activation with an increase in the number of microglial cells surrounding retinal ganglion cells (RGCs), as well as a noteworthy increase in amoeboid forms relative to controls. In addition, there was an increase in metalloproteinases in RGCs, and both hypertrophy in RGCs and axonopathy in the optic nerve were found [102].

In the second model, a TG mouse model of ALS SOD1 (SOD1G93A), there was a vacuolization, with hSOD1-positive vacuoles placed in the dendrites of excitatory retinal neurons, which were detected principally in the inner plexiform layer (IPL) and hardly in the GCL and INL; however, no signs of activation of either the astroglia or the microglia of the retina were shown compared with to the wild-type mice [101]. However, the authors did not rule out the possibility that the microglia were undergoing functional changes (in cytokines) related to the inflammatory process. Nevertheless, neuronal changes observed in this SOD1G93A ALS model in the brain at 50 days of age were followed by microglial morphological changes at 60 days [105–107]. Therefore, the authors concluded that, if there is an inflammatory process in the retina, microglia would be in a different, less reactive or even neuroprotective phenotype [101].

Lastly, the third transgenic murine SOD1G93A model of ALS in an advanced stage of the disease (120 days) showed a loss of the number of Brn3a⁺ RGCs and a microglial activation in retinal tissue [103]. Signs of microglial activation were found in different retinal sectors (superior, inferior, nasal, and temporal) of different retinal layers: outer plexiform layer (OPL) and inner layer complex (ILC) (constituted by an inner plexiform layer and a nerve fiber–ganglion cell layer). In addition, the microglial activation in this SOD1G93A model of ALS showed a cell thickening in the area occupied by each microglial cell, a significant increase in the area of microglial

arborization with hyper-ramifications in the inferior sector of the OPL, retractions of cell processes, and migration and clustering of cells in some areas of the retina, but no increase in the number of microglial cells [103]. Moreover, phenotypic analysis of the microglia showed an M1 phenotype or proinflammatory state of microglia, as the cells were intensely labeled with anti-IFN γ and anti-IL-1 β but did not stain with the characteristic M2 markers (anti-arginase 1 and anti-IL-10) [103]. The significant decrease in the total number of Brn3a⁺ RGCs at 120 days of illness would be consistent with the damage observed in the RGCs of the ALS models discussed above [37, 101, 102], as well as with the thinning of the peripapillary retinal nerve fiber layer (pRNFL), observed by OCT, in ALS patients compared with controls [36–44]. Consequently, these data would support that, in ALS, not only are motor neurons affected but also RGC loss occurs, considering this disease as a multisystemic disease [103].

In none of the abovementioned models were changes in the outer segments of the photoreceptors found. This could indicate that neither this layer of the retina nor the outer blood-retinal barrier (BRB) would be compromised in these animals. Because, when the outer BRB is disrupted, as in a glaucoma model of laser-induced ocular hyper-tension, there are morphological changes and an increase in the number of microglial cells in the photoreceptor outer segment layer [108–112]. Moreover, no changes in the number of microglial cells were found in either the OPL or the ILC [101, 103]; however, the group of Rojas et al. described signs of microglial activation [103]. This difference in results in the same experimental model could be due to the fact that Ringer et al. [101] used retinal sections, while Rojas et al. [103] used retinal whole mounts.

As mentioned above, microglial cells have two distinct phenotypic states that can exert neurotoxic or neuroprotective responses depending on the physiological conditions in which they are found. During ALS progression, activated microglia represent a continuum between the neuroprotective M2 phenotype and the neurotoxic M1 phenotype [113]. In SOD1 ALS animal models, in early stages of the illness, microglia in the lumbar spinal cord expressed markers related to the M2 neuroprotective phenotype (Ym1 and CD206); however, in the late stages of the disease, microglia in the lumbar spinal cord expressed markers related to the M1 neurotoxic phenotype (high levels of NADPH oxidase 2 (NOX2)) [74], suggesting that there is a polarization from a neuroprotective phenotype to a cytotoxic phenotype that induces motor neuron damage. In the retina, there is only one study that analyzed whether microglia are in an M1 or M2 activation phenotype [103]. The results of this study showed that, in 120-day-old SOD1G93A mice, microglia were strongly labeled with antibodies against M1 inflammatory cytokines (IFN γ and IL-1 β), but not with those against M2 anti-inflammatory cytokines (arginase-1 and IL-10), suggesting that at an advanced stage of the disease retinal microglial cells are in an M1 activation phenotype or in a pro-inflammatory state that could be neurotoxic to RGCs, as demonstrated by the loss of these neurons. These results are consistent with the findings in spinal cords of the same animal model, where microglia in an advanced stage of the disease showed a neurotoxic M1 phenotype, demonstrating the dual role (neuroprotective/neurodegenerative) of microglial cells during the ALS process [74]. Therapeutic approaches that target microglia polarization and result in the induction of the M2 phenotype are promising strategies to ameliorate local neurodegeneration and clinical outcome of the disease [114].

2.3 Histopathological studies in ALS and retinal spheroids and axon pathology

(**Table 1**, **Figure 1**) Alterations in axonal transport (retrograde and anterograde) are a hallmark of ALS, being impaired both in ALS patients and in mutant SOD1

mice. In the spinal motor neuron axons, an accumulation of altered mitochondria, neurofilaments, and autophagosomes [12, 58] was demonstrated. On the one hand, mutated dyneins in ALS mice cause this accumulation in the axons of mitochondria and autophagosomes [58]. On the other hand, altered autophagosomes do not eliminate either altered mitochondria or dilated endoplasmic reticules, which accumulate in the axons of motor neurons and cause them to malfunction [12].

There is only one study that focused on this important pathological mechanism in the retina [115]. This study analyzed retinal sections of postmortem eyes from ALS patients with periodic acid Schiff (PAS) and phosphorylated (P-NF) and nonphosphorylated (NP-NF) forms of neurofilament (NF), compared with age-matched controls. Three kinds of spheroids were revealed. First, PAS-positive spheroids with a diameter bigger than 9.07 μ m in the retinal nerve fiber layer (RNFL) were observed in most ALS patients (but only in half of controls), most commonly in the pRNFL and the peripheral RNFL, but rarely in the central RNFL in patients with ALS. The density of PAS-positive spheroids was significantly greater in the pRNFL. Second, P-NF-positive spheroids ranging from 8 to 15 µm in diameter were observed in the peripheral and pRNFL only in ALS patients. Additionally, ALS patients showed a stronger P-NF signal intensity in the RNFL in the peripheral, central, and peripapillary regions. Third, NP-NF spheroids ranging from 7 to 10 μ m in diameter were observed in the RNFL in some of ALS patients (but not in controls). In addition, in most of the ALS patients, the NP-NF signal was increased in the RNFL and IPL. Nevertheless, there was no significant correlation of these retinal spheroids and axon pathology with the clinical characteristics of the ALS patients (age at death, gender, disease duration, mode of disease onset, revised ALS functional rating scale, and rate of disease progression) [115].

Consequently, patients with ALS show not only hallmark findings in spinal cord motor neurons pointing to disrupted axon transport [116–121] but also retinal spheroids and axon pathology as a shared pathogenesis [115]. Transgenic mice with dysfunctional microtubule-associated motor proteins also display such findings [122–124].

2.4 Retinal vessel pathology and ALS

(**Table 1**, **Figure 1**) Retinal vessels are a reflection of small blood vessels in the brain [125]. Parallel vessel pathology in the retinal and cerebral small blood vessels has been demonstrated in many systemic diseases such as coronary heart disease [126] or stroke [127], as well as in some neurodegenerative diseases such as Alzheimer's disease [128, 129] (even in subjects at high genetic risk of developing Alzheimer's disease [130]) and Parkinson's disease [131].

Some ALS-induced changes have also been described in small blood vessels of the brain, which include a loss of pericytes, endothelial cell degeneration, capillary leakage, downregulation of tight junction proteins, and microhemorrhages in patients with ALS [132, 133]. Moreover, alterations of the structure of small blood vessels of the skin and muscles in ALS patients have been described [134, 135].

There was only one study that analyzed retinal vessel pathology in ALS patients with Spectralis OCT but not with angio-OCT. This study described a thicker outer wall of retinal vessels in ALS patients compared with controls, which may be related to the findings in small blood vessels in skin and muscle biopsies. There were neither significant differences in the vessel diameters between ALS patients with spinal onset and bulbar onset, nor a correlation between the vessel measurements and clinical parameters (disease duration and ALS Functional Rating Scale—Revised (ALSFRS-R)) [136].

3. Conclusions

Much research still remains to be conducted on the retina in both animal models and ALS patients. First, further research should aim to describe the different changes in the retina that occur in all pathogenic mechanisms of the disease. Second, there are several models with different genetic mutations that should also be analyzed. In addition, both the retinal and the choroid changes produced at different times in the evolution of the disease should be studied. It is known that ALS is a heterogeneous disease, with different forms of onset, development, and progression, which may potentially exhibit differences in the retina, as observed in the CNS.

The main findings found in the retina in ALS are summarized in **Figure 1** and **Table 1**. In conclusion, multiple studies have confirmed that the retina is affected in ALS,

mainly in the inner layers, and it could serve as a biomarker in this pathology. These retinal changes can be detected by noninvasive retinal imaging techniques to help in the diagnosis and monitoring of ALS disease. In addition, the retina could be used to evaluate the efficacy of different therapies in ALS in a noninvasive way.

Conflict of interest

"The authors declare no conflict of interest."



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References

[1] Román GC. Neuroepidemiology of amyotrophic lateral sclerosis: Clues to aetiology and pathogenesis. Journal of Neurology, Neurosurgery, and Psychiatry. 1996;**61**(2):131-137

[2] Rowland LP, Shneider NA.Amyotrophic lateral sclerosis. The New England Journal of Medicine.2001;344(22):1688-1700

[3] Sathasivam S. Motor neurone disease: Clinical features, diagnosis, diagnostic pitfalls and prognostic markers. Singapore Medical Journal. 2010;**51**(5):367-372

[4] Kiernan MC, Vucic S, Cheah BC, Turner MR, Eisen A, Hardiman O, et al. Amyotrophic lateral sclerosis. Lancet.
2011;377(9769):942-955

[5] Pratt AJ, Getzoff ED, Perry JJP. Amyotrophic lateral sclerosis: Update and new developments. Degenerative Neurological and Neuromuscular Disease. 2012;**2012**(2):1-14

[6] Byrne S, Walsh C, Lynch C, Bede P, Elamin M, Kenna K, et al. Rate of familial amyotrophic lateral sclerosis: A systematic review and meta-analysis. Journal of Neurology, Neurosurgery, and Psychiatry. 2011;**82**(6):623-627

[7] Conwit RA. Preventing familial ALS: A clinical trial may be feasible but is an efficacy trial warranted? Journal of the Neurological Sciences. 2006;**251**:1-2

[8] Haberlandt W. Genetic aspects of amyotrophic lateral sclerosis and progressive bulbar paralysis. Acta Geneticae Medicae et Gemellologiae. 1959;**8**:369-374

[9] Williams DB, Floate DA, Leicester J. Familial motor neuron disease: Differing penetrance in large pedigrees. Journal of the Neurological Sciences. 1988;**86**(2-3):215-230

[10] Yedavalli VS, Patil A, Shah P. Amyotrophic lateral sclerosis and its mimics/variants: A comprehensive review. Journal of Clinical Imaging Science. 2018;**8**:53

[11] Carra S, Crippa V, Rusmini P, Boncoraglio A, Minoia M, Giorgetti E, et al. Alteration of protein folding and degradation in motor neuron diseases: Implications and protective functions of small heat shock proteins. Progress in Neurobiology. 2012;**97**(2):83-100

[12] Zarei S, Carr K, Reiley L, Diaz K, Guerra O, Altamirano PF, et al. A comprehensive review of amyotrophic lateral sclerosis. Surgical Neurology International. 2015;**6**:171

[13] Wijesekera LC, Leigh PN. Amyotrophic lateral sclerosis. Orphanet Journal of Rare Diseases. 2009;**4**(1):3

[14] Zhang Q, Mao C, Jin J, Niu C,
Bai L, Dang J, et al. Side of limb-onset predicts laterality of gray matter
loss in amyotrophic lateral sclerosis.
BioMed Research International.
2014;2014:473250

[15] Ringholz GM, Appel SH, Bradshaw M, Cooke NA, Mosnik DM, Schulz PE. Prevalence and patterns of cognitive impairment in sporadic ALS. Neurology. 2005;**65**(4):586-590

[16] Abrahams S, Goldstein LH,
Suckling J, Ng V, Simmons A, Chitnis X, et al. Frontotemporal white matter changes in amyotrophic lateral sclerosis. Journal of Neurology.
2005;252(3):321-331

[17] Kassubek J, Unrath A, Huppertz HJ, Lulé D, Ethofer T, Sperfeld AD, et al. Global brain atrophy and corticospinal tract alterations in ALS, as investigated by voxel-based morphometry of 3-D MRI. Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders. 2005;**6**(4):213-220

[18] Mezzapesa D, Ceccarelli A, Dicuonzo F, Carella A, De CMF, Lopez M, et al. Whole-brain and regional brain atrophy in amyotrophic lateral sclerosis. AJNR. American Journal of Neuroradiology. 2007;**28**:255-259

[19] Ellis CM, Suckling J, Amaro E, Bullmore ET, Simmons A, Williams SC, et al. Volumetric analysis reveals corticospinal tract degeneration and extramotor involvement in ALS. Neurology. 2001;**57**(9):1571-1578

[20] Weis J, Katona I, Müller-Newen G, Sommer C, Necula G, Hendrich C, et al. Small-fiber neuropathy in patients with ALS. Neurology. 2011;**76**(23):2024-2029

[21] Meier SL, Charleston AJ, Tippett LJ. Cognitive and behavioural deficits associated with the orbitomedial prefrontal cortex in amyotrophic lateral sclerosis. Brain. 2010;**133**(11):3444-3457

[22] McLoon LK, Harandi VM, Brännström T, Andersen PM, Liu JX. Wnt and extraocular muscle sparing in amyotrophic lateral sclerosis. Investigative Ophthalmology and Visual Science. 2014;55(9):5482-5496

[23] Averbuch-Heller L, Helmchen C, Horn AK, Leigh RJ, Büttner-Ennerver JA. Slow vertical saccades in motor neuron disease: Correlation of structure and function. Annals of Neurology. 1998;**44**(4):641-648

[24] Gizzi M, DiRocco A, Sivak M, Cohen B. Ocular motor function in motor neuron disease. Neurology. 1992;**42**(5):1037-1046

[25] Puligheddu M, Congiu P, Aricò D, Rundo F, Borghero G, Marrosu F, et al. Isolated rapid eye movement sleep without atonia in amyotrophic lateral sclerosis. Sleep Medicine. 2016;**26**:16-22

[26] Sharma R, Hicks S, Berna CM, Kennard C, Talbot K, Turner MR.
Oculomotor dysfunction in amyotrophic lateral sclerosis: A comprehensive review. Archives of Neurology.
2011;68(7):857-861

[27] Shaunak S, Orrell RW, O'Sullivan E, Hawken MB, Lane RJ, Henderson L, et al. Oculomotor function in amyotrophic lateral sclerosis: Evidence for frontal impairment. Annals of Neurology. 1995;**38**(1):38-44

[28] Kang BH, Kim JI, Lim YMKK. Abnormal oculomotor functions in amyotrophic lateral sclerosis. Journal of Clinical Neurology. 2018;**14**(4):464-471

[29] Radtke RA, Erwin A, Erwin CW. Abnormal sensory evoked potentials in amyotrophic lateral sclerosis. Neurology. 1986;**36**(6):796-801

[30] Matheson JK, Harrington HJ, Hallett M. Abnormalities of multimodality evoked potentials in amyotrophic lateral sclerosis. Archives of Neurology. 1986;**43**(4):338-340

[31] Münte T, Tröger M, Nusser I, Wieringa B, Johannes S, Matzke M, et al. Alteration of early components of the visual evoked potential in amyotrophic lateral sclerosis. Journal of Neurology. 1998;**245**(4):206-210

[32] Subramaniam JS, Yiannikas C. Multimodality evoked potentials in motor neuron disease. Archives of Neurology. 1990;47(9):989-994 [33] González Díaz N, Escobar Barrios E, Escamilla Chávez C, Escobar RD. Potenciales evocados multimodales en pacientes con esclerosis lateral amiotrófica. Revista Mexicana de Medicina Física y Rehabilitación. 2004;**16**:5-11

[34] Kushner PD, Stephenson DT, Wright S. Reactive astrogliosis is widespread in the subcortical white matter of amyotrophic lateral sclerosis brain. Journal of Neuropathology and Experimental Neurology. 1991;**50**(3):263-277

[35] Roth NM, Saidha S, Zimmermann H, Brandt AU, Oberwahrenbrock T, Maragakis NJ, et al. Optical coherence tomography does not support optic nerve involvement in amyotrophic lateral sclerosis. European Journal of Neurology. 2013;**20**(8):1170-1176

[36] Ringelstein M, Albrecht P, Südmeyer M, Harmel J, Müller AK, Keser N, et al. Subtle retinal pathology in amyotrophic lateral sclerosis. Annals of Clinical Translational Neurology. 2014;**1**(4):290-297

[37] Volpe NJ, Simonett J, Fawzi AAST, Volpe NJ, Simonett J, Fawzi AA, et al. Ophthalmic manifestations of amyotrophic lateral sclerosis (an American ophthalmological society thesis). Transactions of the American Ophthalmological Society. 2015;**113**:1-15

[38] Hübers A, Müller HP, Dreyhaupt J, Böhm K, Lauda F, Tumani H, et al. Retinal involvement in amyotrophic lateral sclerosis: A study with optical coherence tomography and diffusion tensor imaging. Journal of Neural Transmission. 2016;**123**(3):281-287

[39] Simonett JM, Huang R, Siddique N, Farsiu S, Siddique T, Volpe NJ, et al. Macular sub-layer thinning and association with pulmonary function tests in amyotrophic lateral sclerosis. Scientific Reports. 2016;**6**(1):29187

[40] Mukherjee N, McBurney-Lin S, Kuo A, Bedlack R, Tseng H. Retinal thinning in amyotrophic lateral sclerosis patients without ophthalmic disease. Bhattacharya S, editor. PLoS One. 2017;**12**(9):e0185242

[41] Rohani M, Meysamie A, Zamani B, Sowlat MM, Akhoundi FH. Reduced retinal nerve fiber layer (RNFL) thickness in ALS patients: A window to disease progression. Journal of Neurology. 2018;**265**(7):1557-1562

[42] Liu Z, Wang H, Fan D, Wang W. Comparison of optical coherence tomography findings and visual field changes in patients with primary open-angle glaucoma and amyotrophic lateral sclerosis. Journal of Clinical Neuroscience. 2018;**48**:233-237

[43] Rojas P, De HR, Ramirez AI, Ferreras A, Salobrar-Garcia E, Muñoz-Blanco JL, et al. Changes in retinal OCT and their correlations with neurological disability in early ALS patients, a follow-up study. Brain Sciences. 2019;**9**(337):1-18

[44] Zhang Y, Liu X, Fu J, Zhang Y, Yang X, Zhang S, et al. Selective and inverse U-shaped curve alteration of the retinal nerve in amyotrophic lateral sclerosis: A potential Mirror of the disease. Frontiers in Aging Neuroscience | www.frontiersin.org. 2022;**13**(1):783431

[45] Bussel II, Wollstein G, Schuman JS. OCT for glaucoma diagnosis, screening and detection of glaucoma progression. The British Journal of Ophthalmology. 2014;**98**(Suppl. 2):ii15-ii19

[46] Ratchford JN, Quigg ME, Conger BA, Frohman BT, Frohman BE, Balcer LJ,

et al. Optical coherence tomography helps differentiate neuromyelitis optica and MS optic neuropathies. Neurology. 2009;**73**(4):302-308

[47] Dörr J, Wernecke KD, Bock M, Gaede G, Wuerfel JT, Pfueller CF, et al. Association of retinal and macular damage with brain atrophy in multiple sclerosis. Linden R, editor. PLoS One. 2011;**6**(4):e18132

[48] Gordon-Lipkin E, Chodkowski B, Reich DS, Smith SA, Pulicken M, Balcer LJ, et al. Retinal nerve fiber layer is associated with brain atrophy in multiple sclerosis. Neurology. 2007;**69**(16):1603-1609

[49] Siger M, Dziegielewski K, Jasek L, Bieniek M, Nicpan A, Nawrocki J, et al. Optical coherence tomography in multiple sclerosis: Thickness of the retinal nerve fiber layer as a potential measure of axonal loss and brain atrophy. Journal of Neurology. 2008;**255**(10):1555-1560

[50] Stricker S, Oberwahrenbrock T, Zimmermann H, Schroeter J, Endres M, Brandt AU, et al. Temporal retinal nerve fiber loss in patients with spinocerebellar ataxia type 1. Villoslada P, editor. PLoS One. 2011;**6**(7):e23024

[51] Albrecht P, Müller AK, Südmeyer M, Ferrea S, Ringelstein M, Cohn E, et al. Optical coherence tomography in parkinsonian syndromes. Paul F, editor. PLoS One. 2012;7(4):e34891

[52] La Morgia C, Barboni P, Rizzo G, Carbonelli M, Savini G, Scaglione C, et al. Loss of temporal retinal nerve fibers in Parkinson disease: A mitochondrial pattern? European Journal of Neurology. 2013;**20**(1):198-201

[53] Schneider E, Zimmermann H, Oberwahrenbrock T, Kaufhold F, Kadas EM, Petzold A, et al. Optical coherence tomography reveals distinct patterns of retinal damage in Neuromyelitis Optica and multiple sclerosis. PLoS One. 2013;8(6):e66151

[54] Kesler A, Vakhapova V, Korczyn AD, Naftaliev E, Neudorfer M. Retinal thickness in patients with mild cognitive impairment and Alzheimer's disease. Clinical Neurology and Neurosurgery. 2011;**113**(7):523-526

[55] Gordon-Lipkin E, Calabresi PA. Optical coherence tomography: A quantitative tool to measure neurodegeneration and facilitate testing of novel treatments for tissue protection in multiple sclerosis. Journal of Neuroimmunology. 2017;**304**:93-96

[56] Pfueller CF, Brandt AU, Schubert F, Bock M, Walaszek B, Waiczies H, et al. Metabolic changes in the visual cortex are linked to retinal nerve fiber layer thinning in multiple sclerosis. Kleinschnitz C, editor. PLoS One. 2011;**6**(4):e18019

[57] Bock M, Paul F, Dörr J. Diagnosis and monitoring of multiple sclerosis: The value of optical coherence tomography. Der Nervenarzt. 2013;**84**(4):483-492

[58] Ikenaka K, Katsuno M, Kawai K, Ishigaki S, Tanaka F, Sobue G. Disruption of axonal transport in motor neuron diseases. International Journal of Molecular Sciences. 2012;**13**:1225-1238

[59] Mejzini R, Flynn LL, Pitout IL, Fletcher S, Wilton SD, Akkari PA. ALS genetics, mechanisms, and therapeutics: Where are we now? Frontiers in Neuroscience. 2019;**13**:1-27

[60] Rojas P, Ramírez AI, Fernández-Albarral JA, López-Cuenca I, Salobrar-García E, Cadena M, et al. Amyotrophic lateral sclerosis: A neurodegenerative motor neuron disease with ocular involvement. Frontiers in Neuroscience. 2020;**14**:566858

[61] Soldatov VO, Kukharsky MS, Belykh AE, Sobolev AM, Deykin AV. Retinal damage in amyotrophic lateral sclerosis: Underlying mechanisms. Eye Brain. 2021;**13**:131-146

[62] Ramirez Sebastian JM. El ojo una ventana al cerebro. An Real Academic Doctors. 2017;**2**(3):357-381

[63] Han-Xiang D, Chen W, Seong-Tshool S, Boycott KM, Gorrie GH, Siddique N, et al. Mutations in UBQLN2 cause dominant X-linked juvenile and adult-onset ALS and ALS/dementia. Nature. 2011;477(7363):211-215

[64] Al-Sarraj S, King A, Troakes C, Smith B, Maekawa S, Bodi I, et al. p62 positive, TDP-43 negative, neuronal cytoplasmic and intranuclear inclusions in the cerebellum and hippocampus define the pathology of C9orf72linked FTLD and MND/ALS. Acta Neuropathologica. 2011;**122**(6):691-702

[65] Williams KL, Topp S, Yang S, Smith B, Fifita JA, Warraich ST, et al. CCNF mutations in amyotrophic lateral sclerosis and frontotemporal dementia. Nature Communications. 2016;7(1):11253

[66] Fawzi AA, Simonett JM, Purta P, Moss HE, Lowry JL, Deng HX, et al. Clinicopathologic report of ocular involvement in ALS patients with C9orf72 mutation. Amyotroph Lateral Scler Frontotemporal Degener. 2014;**15**(7-8):569-580

[67] Volpe NJ, Simonett J, Fawzi AA, Siddique T. Opthalmic manifestations of amyotrophic lateral sclerosis (an American ophthalmological society thesis). Transactions of the American Ophthalmological Society. 2015;**113**:1-15 [68] Ward ME, Chen R, Huang HY, Ludwig C, Telpoukhovskaia M, Taubes A, et al. Individuals with progranulin haploinsufficiency exhibit features of neuronal ceroid lipofuscinosis. Science Translational Medicine. 2017;**9**(385):eaah5642

[69] Ward ME, Taubes A, Chen R, Miller BL, Sephton CF, Gelfand JM, et al. Early retinal neurodegeneration and impaired ran-mediated nuclear import of TDP-43 in progranulin-deficient FTLD. The Journal of Experimental Medicine. 2014;**211**(10):1937-1945

[70] Azoulay-Ginsburg S, Di Salvio M, Weitman M, Afri M, Ribeiro S, Ebbinghaus S, et al. Chemical chaperones targeted to the endoplasmic reticulum (ER) and lysosome prevented neurodegeneration in a C9orf72 repeat expansion drosophila amyotrophic lateral sclerosis (ALS) model. Pharmacol Reports. 2021;**73**(2):536-550

[71] Ramirez AI, Rojas B, de Hoz R, et al. Microglia, Inflammation and Glaucoma. In: Glaucoma. SM Group Open Access eBooks. DE, USA: Dover; 2015. p. 1-16

[72] Munder M. Arginase: An emerging key player in the mammalian immune system: REVIEW. British Journal of Pharmacology. 2009;**158**(3):638-651

[73] Philips T, Robberecht W. Neuroinflammation in amyotrophic lateral sclerosis: Role of glial activation in motor neuron disease. Lancet Neurology. 2011;**10**(3):253-263

[74] Liao B, Zhao W, Beers DR, Henkel JS, Appel SH. Transformation from a neuroprotective to a neurotoxic microglial phenotype in a mouse model of ALS. Experimental Neurology. 2012;**237**(1):147-152

[75] De Hoz R, Rojas B, Ramírez AI, Salazar JJ, Gallego BI, Triviño A, et al.

Retinal macroglial responses in health and disease. BioMed Research International. 2016;**2016**:2954721

[76] Perea G, Navarrete M, Araque A. Tripartite synapses: Astrocytes process and control synaptic information. Trends in Neurosciences. 2009;**32**(8):421-431

[77] Bélanger M, Allaman I,
Magistretti PJ. Brain energy metabolism:
Focus on astrocyte-neuron metabolic
cooperation. Cell Metabolism.
2011;14(6):724-738

[78] Fernandez-Fernandez S, Almeida A, Bolaños JP. Antioxidant and bioenergetic coupling between neurons and astrocytes. The Biochemical Journal. 2012;**443**(1):3-12

[79] Sofroniew MV. Astrogliosis. Cold Spring Harbor Perspectives in Biology. 2015;7:2

[80] Burda JE, Sofroniew MV. Reactive gliosis and the multicellular response to CNS damage and disease. Neuron. 2014;**81**(2):229-248

[81] Vargas MR, Johnson JA. Astrogliosis in amyotrophic lateral sclerosis: Role and therapeutic potential of astrocytes. Neurotherapeutics. 2010;7(4):471-481

[82] Alexianu ME, Kozovska M, Appel SH. Immune reactivity in a mouse model of familial ALS correlates with disease progression. Neurology. 2001;**57**(7):1282-1289

[83] Engelhardt JI, Tajti J, Appel SH. Lymphocytic infiltrates in the spinal cord in amyotrophic lateral sclerosis. Archives of Neurology. 1993;**50**(1):30-36

[84] Gurney ME, Pu H, Chiu AY, Dal Canto MC, Polchow CY, Alexander DD, et al. Motor neuron degeneration in mice that express a human Cu,Zn superoxide dismutase mutation. Science (80-). 1994;**264**(5166):1772-1775

[85] Vinsant S, Mansfield C, Jimenez-Moreno R, Moore VDG, Yoshikawa M, Hampton TG, et al. Characterization of early pathogenesis in the SOD1G93A mouse model of ALS: Part I, background and methods. Brain and Behavior: A Cognitive Neuroscience Perspective. 2013;3(4):335-350

[86] Zhao W, Beers DR, Henkel JS, Zhang W, Urushitani M, Julien JP, et al. Extracellular mutant SOD1 induces microglial-mediated motoneuron injury. Glia. 2010;**58**(2):231-243

[87] Philips T, Rothstein JD. Rodent models of amyotrophic lateral sclerosis. Current Protocols in Pharmacology. 2015;**2015**:5.67.1-5.67.21

[88] Yamanaka K, Boillee S, Roberts EA, Garcia ML, McAlonis-Downes M, Mikse OR, et al. Mutant SOD1 in cell types other than motor neurons and oligodendrocytes accelerates onset of disease in ALS mice. Proceedings of the National Academy of Sciences of the United States of America. 2008;**105**(21):7594-7599

[89] Appel SH, Zhao W, Beers DR, Henkel JS. The microglial-motoneuron dialogue in ALS. Acta Myologica. 2011;**30**:4-8

[90] Lampson LA, PD K, Sobel RA. Strong expression of class II major histocompatibility complex (MHC) antigens in the absence of detectable T cell infiltration in amyotrophic lateral sclerosis (ALS) spinal cord. Journal of Neuropathology and Experimental Neurology. 1988;47:353

[91] Kawamata T, Akiyama H, Yamada T, Mcgeer PL. Immunologic reactions in amyotrophic lateral sclerosis brain and spinal cord tissue. The American Journal of Pathology. 1992;**140**(3):691-707

[92] Sargsyan SA, Monk PN, Shaw PJ. Microglia as potential contributors to motor neuron injury in amyotrophic lateral sclerosis. Glia. 2005;**51**(4):241-253

[93] Verine Boillé S, Vande VC, Cleveland DW. ALS: A disease of motor neurons and their nonneuronal neighbors. Neuron. 2006;**52**:39-59

[94] King AE, Dickson TC, Blizzard CA, Woodhouse A, Foster SS, Chung RS, et al. Neuron-glia interactions underlie ALS-like axonal cytoskeletal pathology. Neurobiology of Aging. 2011;**32**(3):459-469

[95] Ince PG, Shaw PJ, Slade JY, Jones C, Hudgson P. Familial amyotrophic lateral sclerosis with a mutation in exon 4 of the Cu/Zn superoxide dismutase gene: Pathological and immunocytochemical changes. Acta Neuropathologica. 1996;**92**(4):395-403

[96] Schiffer D, Cordera S, Cavalla P, Migheli A. Reactive astrogliosis of the spinal cord in amyotrophic lateral sclerosis. Journal of the Neurological Sciences. 1996;**139**(SUPPL):27-33

[97] Nagy D, Kato T, Kushner PD. Reactive astrocytes are widespread in the cortical gray matter of amyotrophic lateral sclerosis. Journal of Neuroscience Research. 1994;**38**(3):336-347

[98] Turner MR, Cagnin A, Turkheimer FE, Miller CCJ, Shaw CE, Brooks DJ, et al. Evidence of widespread cerebral microglial activation in amyotrophic lateral sclerosis: An [11C] (R)-PK11195 positron emission tomography study. Neurobiology of Disease. 2004;**15**(3):601-609

[99] Clement AM, Nguyen MD, Roberts EA, Garcia ML, Boillée S, Rule M, et al. Wild-type nonneuronal cells extend survival of SOD1 mutant motor neurons in ALS mice. Science (80-). 2003;**302**(5642):113-117

[100] Solomon JN, Lewis CAB, Ajami B, Corbel SY, Rossi FMV, Krieger C. Origin and distribution of bone marrow-derived cells in the central nervous system in a mouse model of amyotrophic lateral sclerosis. Glia. 2006;**53**(7):744-753

[101] Ringer C, Weihe E, Schütz B. SOD1 G93A mutant mice develop a Neuroinflammation-independent Dendropathy in excitatory neuronal subsets of the olfactory bulb and retina. Journal of Neuropathology and Experimental Neurology. 2017;**76**(9):769-778

[102] Cho K, in, Yoon D, Yu M, Peachey NS, Ferreira PA. Microglial activation in an amyotrophic lateral sclerosis-like model caused by Ranbp2 loss and nucleocytoplasmic transport impairment in retinal ganglion neurons. Cellular and Molecular Life Sciences. 2019;**76**:3407-3432

[103] Rojas P, Ramírez AI, Cadena M, Fernández-Albarral JA, Salobrar-García E, Santos-García I, et al. Retinal ganglion cell loss and microglial activation in a SOD1G93A mouse model of amyotrophic lateral sclerosis. International Journal of Molecular Sciences. 2021;**22**:1663

[104] Ferreira PA. The coming-of-age of nucleocytoplasmic transport in motor neuron disease and neurodegeneration. Cellular and Molecular Life Sciences. 2019;**76**(12):2247-2273

[105] Ringer C, Luisa-Sybille B, MaK S, Eiden LE, Weihe E, Schütz B. PACAP signaling exerts opposing effects on neuroprotection and neuroinflammation during disease progression in

the SOD1(G93A) mouse model of amyotrophic lateral sclerosis. Neurobiology of Disease. 2013;**54**:32-42

[106] Ringer C, Tune S, Mirjam BA, Schwarzbach H, Tsujikawa K, Weihe E, et al. Disruption of calcitonin generelated peptide signaling accelerates muscle denervation and dampens cytotoxic neuroinflammation in SOD1 mutant mice. Cellular and Molecular Life Sciences. 2017;74:339-358

[107] Ringer C, Weihe E, Schütz B. Presymptomatic alterations in subcellular β CGRP distribution in motor neurons precede astrogliosis in ALS mice. Neurobiology of Disease. 2009;**35**(2):286-295

[108] de Hoz R, Ramírez AI, González-Martín R, Ajoy D, Rojas B, Salobrar-Garcia E, et al. Bilateral early activation of retinal microglial cells in a mouse model of unilateral laser-induced experimental ocular hypertension. Experimental Eye Research. 2018;**171**:12-29

[109] Rojas B, Gallego BI, Ramírez AI, Salazar JJ, de Hoz R, Valiente-Soriano FJ, et al. Microglia in mouse retina contralateral to experimental glaucoma exhibit multiple signs of activation in all retinal layers. Journal of Neuroinflammation. 2014;**11**(1):1-24

[110] Fernández-Albarral JA, Ramírez AI, De Hoz R, López-Villarín N, Salobrar-García E, López-Cuenca I, et al. Neuroprotective and anti-inflammatory effects of a hydrophilic saffron extract in a model of glaucoma. International Journal of Molecular Sciences. 2019;**20**(17):1-22

[111] Ramírez AI, de Hoz R, Fernández-Albarral JA, Salobrar-Garcia E, Rojas B, Valiente-Soriano FJ, et al. Time course of bilateral microglial activation in a mouse model of laserinduced glaucoma. Scientific Reports. 2020;**10**(1):1-17

[112] Gallego BI, Salazar JJ, de Hoz R, Rojas B, Ramírez AI, Salinas-Navarro M, et al. IOP induces upregulation of GFAP and MHC-II and microglia reactivity in mice retina contralateral to experimental glaucoma. Journal of Neuroinflammation. 2012;**9**(92):1-18

[113] Gordon S, Martinez FO.Alternative activation of macrophages: Mechanism and functions. Immunity.2010;**32**:593-604

[114] Geloso MC, Corvino V, Marchese E, Serrano A, Michetti F, D'Ambrosi N. The dual role of microglia in ALS: Mechanisms and therapeutic approaches. Frontiers in Aging Neuroscience. 2017;**25**(9):242

[115] Sharma K, Amin MAM, Gupta N, Zinman L, Zhou X, Irving H, et al. Retinal spheroids and axon pathology identified in amyotrophic lateral sclerosis. Investigative Ophthalmology and Visual Science. 2020;**61**(13):8-10

[116] Delisle MB, Carpenter S. Neurofibrillary axonal swellings and amyotrophic lateral sclerosis. Journal of the Neurological Sciences. 1984;**63**(2):241-250

[117] Julien JP. A role for neurofilaments in the pathogenesis of amyotrophic lateral sclerosis. Biochemistry and Cell Biology. 1995;**73**(9-10):593-597

[118] Carpenter S. Proximal axonal enlargement in motor neuron disease. Neurology. 1968;**18**(9):841-851

[119] Munoz DG, Greene C, Perl DP, Selkoe DJ. Accumulation of phosphorylated neurofilaments in anterior horn motoneurons of amyotrophic lateral sclerosis patients. Journal of Neuropathology and Experimental Neurology. 1988;**47**(1):9-18

[120] Leigh PN, Dodson A, Swash M, Brion JP, Anderton BH. Cytoskeletal abnormalities in motor neuron disease. An immunocytochemical study. Brain. 1989;**112**(Pt 2(2)):521-535

[121] Murayama S, Bouldin TW, Suzuki K. Immunocytochemical and ultrastructural studies of upper motor neurons in amyotrophic lateral sclerosis. Acta Neuropathologica. 1992;**83**(5):518-524

[122] LaMonte BH, Wallace KE, Holloway BA, Shelly SS, Ascaño J, Tokito M, et al. Disruption of dynein/ dynactin inhibits axonal transport in motor neurons causing late-onset progressive degeneration. Neuron. 2002;**34**(5):715-727

[123] Laird FM, Farah MH, Ackerley S, Hoke A, Maragakis N, Rothstein JD, et al. Motor neuron disease occurring in a mutant dynactin mouse model is characterized by defects in vesicular trafficking. The Journal of Neuroscience. 2008;**28**(9):1997

[124] Xia CH, Roberts EA, Her LS, Liu X, Williams DS, Cleveland DW, et al. Abnormal neurofilament transport caused by targeted disruption of neuronal kinesin heavy chain KIF5A. The Journal of Cell Biology. 2003;**161**(1):55-66

[125] Patton N, Aslam T, Macgillivray T, Pattie A, Deary IJ, Dhillon B. Retinal vascular image analysis as a potential screening tool for cerebrovascular disease: A rationale based on homology between cerebral and retinal microvasculatures. Journal of Anatomy. 2005;**206**:319-348 [126] Liew G, Wang JJ. Retinal vascular signs: A window to the heart? Revista Española de Cardiología (English Edition). 2011;**64**(6):515-521

[127] Conijn MMA, Kloppenborg RP, Algra A, Mali WPTM, Kappelle LJ, Vincken KL, et al. Cerebral small vessel disease and risk of death, ischemic stroke, and cardiac complications in patients with atherosclerotic disease: The second manifestations of arterial diseasemagnetic resonance (SMART-MR) study. Stroke. 2011;42(11):3105-3109

[128] Lim JKH, Li Q-X, He Z, Vingrys AJ, Wong VHY, Currier N, et al. The eye as a biomarker for Alzheimer's disease. Frontiers in Neuroscience. 2016;**10**(536):1-14

[129] Salobrar-Garcia E, Méndez-Hernández C, de Hoz R, Ramírez AI, López-Cuenca I, Fernández-Albarral JA, et al. Ocular vascular changes in mild alzheimer's disease patients: Foveal avascular zone, choroidal thickness, and onh hemoglobin analysis. Journal of Personalized Medicine. 2020;**10**(4):1-13

[130] López-Cuenca I, Salobrar-García E, Sánchez-Puebla L, Espejel E, García del Arco L, Rojas P, et al. Retinal vascular study using OCTA in subjects at high genetic risk of developing Alzheimer's disease and cardiovascular risk factors. Journal of Clinical Medicine. 2022;**11**:3248

[131] Kromer R, Buhmann C, Hidding U, Keserü M, Keserü D, Hassenstein A, et al. Evaluation of retinal vessel morphology in patients with Parkinson's disease using optical coherence tomography. PLoS One. 2016;**11**(8):e0161136

[132] Garbuzova-Davis S, Sanberg PR.
Blood-CNS barrier impairment in
ALS patients versus an animal model.
Frontiers in Cellular Neuroscience.
2014;3(8):21

[133] Kovács-Öller T, Ivanova E, Szarka G, Tengölics ÁJ, Völgyi B, Sagdullaev BT. Imatinib sets Pericyte mosaic in the retina. International Journal of Molecular Sciences. 2020;**21**:2522

[134] Buckley AF, Bossen EH. Skeletal muscle microvasculature in the diagnosis of neuromuscular disease. Journal of Neuropathology and Experimental Neurology. 2013;72(10):906-918

[135] Kolde G, Bachus R, Ludolph AC. Skin involvement in amyotrophic lateral sclerosis. Lancet (London, England). 1996;**347**(9010):1226-1227

[136] Abdelhak A, Hübers A, Böhm K, Ludolph AC, Kassubek J, Pinkhardt EH. In vivo assessment of retinal vessel pathology in amyotrophic lateral sclerosis. Journal of Neurology. 2018;**265**(4):949-953

