

Mycosphere 11(1): 1514–1526 (2020) www.mycosphere.org

Article

Doi 10.5943/mycosphere/11/1/11

ISSN 2077 7019

A dynamic portal for a community-driven, continuously updated classification of *Fungi* and fungus-like organisms: outlineoffungi.org

Wijayawardene NN¹, Hyde KD², Dai DQ¹,*, Tang LZ¹,*, Aptroot A³, Castañeda-Ruiz RF⁴, Druzhinina IS⁵, Cai F⁵, Ekanayaka AH², Erdoğdu M⁶, Fiuza PO⁻, Gentekaki E², Goto BT⁶, Haelewaters D⁵,¹0,¹1¹,¹², Hongsanan S¹³, Jeewon R¹⁴, Kirk PM¹⁵, Jayalal RGU¹⁶, Karunarathna SC¹⁻,¹8,¹9, Wanasinghe DN¹⁻,¹18,¹9, Lumbsch HT²⁰, Madrid H²¹,²², Maharachchikumbura SSN²³, Monteiro JS²⁴, Shivaprakash N²⁵, Pfliegler WP²⁶, Phillips AJL²¬, Saxena RK²⁶, Stadler M²⁵,³0, Tian Q², Tokarev YS³¹, Tsurykau A³²,³3, Ertz D³⁴,³⁵, Lee HB³⁶, Etayo J³¬, Vizzini A³⁶, Jones EBG³ց, Lin CG², Li WJ², Dai YC⁴⁰, Fan XL⁴¹, McKenzie EHC⁴², Shivas RG⁴³, Hustad V⁴⁴, Leontyev DV⁴⁵, de Hoog GS⁴⁶, Niskanen T⁴¬, Boekhout T⁴⁶,⁴9, Gaya E⁵⁰, Thines M⁵¹,⁵²

¹ Center for Yunnan Plateau Biological Resources Protection and Utilization, College of Biological Resource and Food Engineering, Qujing Normal University, Qujing, Yunnan 655011, P.R. China.

² Center of Excellence in Fungal Research, Mae Fah Luang University, Chiang Rai 57100, Thailand

³ Laboratório de Botânica / Liquenologia, Instituto de Biociências, Universidade Federal de Mato Grosso do Sul, Avenida Costa e Silva s/n, Bairro Universitário, CEP 79070-900, Campo Grande, Mato Grosso do Sul, Brazil

⁴ Instituto de Investigaciones Fundamentales en AgriculturaTropical, 'Alejandro de Humboldt' (INIFAT), Santiago deLas Vegas, Habana, Cuba

⁵ Fungal Genomics Group, College of Resources and Environmental Sciences, Nanjing Agricultural University, Nanjing, Jiangsu, P.R. China.

⁶ Department of Landscape Architects, Faculty of Agriculture, Kırşehir Ahi Evran University, Kırşehir, Turkey

⁷ Universidade Federal do Rio Grande do Norte (UFRN), Programa de Pós-graduação em Sistemática e Evolução, Centro de Biociências, Campus Universitário, Av. Senador Salgado Filho, 3000, Lagoa Nova, 59078-970, Natal-RN, Brazil

⁸ Departamento de Botânica e Zoologia, Universidade Federal do Rio Grande do Norte, Campus Universitário, 59072–970, Natal, RN, Brazil

⁹ Faculty of Science, University of South Bohemia, Branišovská 31, 370 05 České Budějovice, Czech Republic

¹⁰ Department of Botany and Plant Pathology, Purdue University, 915 W. State Street, West Lafayette, Indiana 47907,

¹¹ Herbario UCH, Universidad Autónoma de Chiriquí, Apartado Postal 0427, David, Panama

¹² Department of Biology, Research Group Mycology, Ghent University, K.L. Ledeganckstraat 35, 9000 Ghent, Belgium

¹³ Shenzhen Key Laboratory of Laser Engineering, College of Optoelectronic Engineering, Shenzhen University, Shenzhen, P.R. China

¹⁴ Department of Health Sciences, Faculty of Science, University of Mauritius, Reduit, Mauritius

¹⁵ Biodiversity Informatics and Spatial Analysis, Royal Botanic Gardens, Kew, Richmond, Surrey TW9 3DS, UK

¹⁶ Department of Natural Resources, Sabaragamuwa University of Sri Lanka, Belihuloya, 70140, Sri Lanka

¹⁷ CAS key Laboratory for Plant Diversity and Biogeography of East Asia, Kunming Institute of Botany, Chinese Academy of Science, Kunming 650201, Yunnan, P.R. China

¹⁸ World Agroforestry Centre, East and Central Asia, Kunming 650201, Yunnan, P.R. China

¹⁹ Centre for Mountain Futures (CMF), Kunming Institute of Botany, Kunming, Yunnan, 650201, P.R. China

²⁰ Science & Education, The Field Museum, 1400 S. Lake Shore Drive, Chicago, IL 60605, USA

²¹ Escuela de Tecnología Médica, Facultad de Salud, Universidad Santo Tomás, Los Carreras 753, Osorno, Chile

²² Centro de Genómica y Bioinformática, Facultad de Ciencias, Universidad Mayor, Camino La Pirámide 5750, Huechuraba, Santiago, Chile

- ²³ School of Life Science and Technology, University of Electronic Science and Technology of China, Chengdu 611731, P.R. China
- ²⁴ Coordenação de Botânica, Museu Paraense Emílio Goeldi, Avenida Perimetral, 1901, Terra Firme, 66077-530, Belém, Pará, Brazil
- ²⁵ ICAR-Central Plantation Crops Research Institute (ICAR-CPCRI), Regional Station, Vittal 574243, Karnataka, India
- ²⁶ Department of Molecular Biotechnology and Microbiology, University of Debrecen, Debrecen, Egyetem tér 1., 4032, Hungary
- ²⁷ Universidade de Lisboa, Faculdade de Ciências, Biosystems and Integrative Sciences Institute (BioISI), Campo Grande, 1749-016 Lisbon, Portugal
- ²⁸ Birbal Sahni Institute of Palaeosciences, 53 University Road, Lucknow-226007, India
- ²⁹ Department of Microbial Drugs, Helmholtz-Zentrum für Infektionsforschung GmbH, Inhoffenstrasse 7, 38124, Braunschweig, Germany
- ³⁰ German Centre for Infection Research (DZIF), partner site Hannover-Braunschweig, 38124 Braunschweig, Germany
- ³¹ Laboratory of Microbiological Control, All-Russian Institute of Plant Protection, Shosse Podbelskogo 3, Pushkin, St. Petersburg, 196608, Russia
- ³² F. Skorina Gomel State University, Department of Biology, Sovetskaja Str. 104, Gomel 246019, Belarus
- ³³ Samara National Research University, Institute of Natural Sciences, Department of Ecology, Botany and Nature, Protection, Moskovskoye shosse 34, Samara 443086, Russia
- ³⁴ Meise Botanic Garden, Department of Research, Nieuwelaan 38, BE-1860 Meise, Belgium
- ³⁵ Fédération Wallonie-Bruxelles, Direction générale de l'Enseignement non obligatoire et de la Recherche scientifique, Rue A. Lavallée 1, BE-1080 Bruxelles, Belgium
- ³⁶ Environmental Microbiology Laboratory, Department of Agricultural Biological Chemistry, College of Agriculture & Life Sciences, Chonnam National University, Gwangju 61186, Korea
- ³⁷ Institute Zizur Mayor, Ronda San Cristóbal 196, 31180 Zizur Mayor, Navarra, Spain
- ³⁸ Department of Life Sciences and Systems Biology, University of Torino, Viale PA Mattioli 25, 10125 Torino, Italy
- ³⁹ Nantgaredig, 33 B St. Edwards Road Southsea Hants, Hampshire PO5 3DH, UK
- ⁴⁰ Institute of Microbiology, Beijing Forestry University, East Qinghua Road, 35, Haidian, Beijing 100083, P.R. China
- ⁴¹ The Key Laboratory for Silviculture and Conservation of Ministry of Education, Beijing Forestry University, Beijing 100083, P.R. China
- ⁴² Manaaki Whenua-Landcare Research, Private Bag No 92170, Auckland, New Zealand
- ⁴³ Centre for Crop Health, University of Southern Queensland, Toowoomba 4350, Australia
- ⁴⁴ Department of Natural Sciences, Northwest Missouri State University, Maryville, Missouri, 64468, USA
- ⁴⁵ Department of Botany, H.S. Skovoroda Kharkiv National Pedagogical University, Valentynivs'ka 2, Kharkiv 61168, Ukraine
- ⁴⁶ Center of Expertise in Mycology of Radboud University Medical Center / Canisius Wilhelmina Hospital, Nijmegen, The Netherlands
- ⁴⁷ The Jodrell Laboratory Royal Botanic Gardens, Kew, Surrey TW9 3AB, UK
- ⁴⁸ Westerdijk Fungal Biodiversity Institute, Utrecht, The Netherlands
- ⁴⁹ Institute of Biodiversity and Ecosystem Dynamics (IBED), University of Amsterdam, Amsterdam, the Netherlands
- ⁵⁰ Comparative Plant and Fungal Biology, Royal Botanic Gardens, Kew, TW9 3AE, UK
- ⁵¹ Goethe University, Department for Biological Sciences, Institute of Ecology, Evolution and Diversity, Max-vonLaue-Str. 13, D-60486 Frankfurt am Main, Germany
- ⁵² Senckenberg Biodiversity and Climate Research Centre, Senckenberganlage 25, D-60325 Frankfurt am Main, Germany

Wijayawardene NN, Hyde KD, Dai DQ, Tang LZ, Aptroot A, Castañeda-Ruiz RF, Druzhinina IS, Cai F, Ekanayaka AH, Erdoğdu M, Fiuza PO, Gentekaki E, Goto BT, Haelewaters D, Hongsanan S, Jeewon R, Kirk PM, Jayalal U, Karunarathna SC, Wanasinghe DN, Lumbsch HT, Madrid H, Maharachchikumbura SSN, Monteiro JS, Shivaprakash N, Pfliegler WP, Phillips AJL, Saxena RK, Stadler M, Tian Q, Tokarev YS, Tsurykau A, Ertz D, Lee HB, Etayo J, Vizzini A, Jones EGB, Lin CG, Li WJ, Dai YC, Fan XL, McKenzie EHC, Shivas RG, Hustad V, Leontyev DV, de Hoog GS, Niskanen T, Boekhout T, Gaya E, Thines M 2020 – A dynamic portal for a community-driven, continuously updated classification of *Fungi* and fungus-like organisms: outlineoffungi.org. Mycosphere 11(1), 1514–1526, Doi 10.5943/mycosphere/11/1/11

Abstract

The website http://outlineoffungi.org, is launched to provide a continuous up-to-date classification of the kingdom *Fungi* (including fossil fungi) and fungus-like taxa. This is based on

recent publications and on the outline of fungi and fungus-like taxa published recently (Mycosphere 11, 1060–1456, doi 10.5943/mycosphere/11/1/8). The website is continuously updated according to latest classification schemes, and will present an important platform for researchers, industries, government officials and other users. Users can provide input about missing genera, new genera, and new data. They will also have the opportunity to express their opinions on classifications with notes published in the 'Notes' section of the webpage following review and editing by the curators and independent experts. The website will provide a system to stay abreast of the continuous changes in fungal classification and provide a general consensus on the systematics of fungi.

Keywords – classification – community-driven – higher ranks – outline – portal – taxa

Introduction

Classification of fungi is a topic of key concern among mycologists and other researchers, such as plant pathologists and those in the applied sciences. Accurate species identification and classification are prerequisites for precise scientific communication and for future comparative studies. Morphology-based classification was widely used before DNA sequence data became available and opened up a more objective tool to infer natural relationships (White et al. 1990). In traditional taxonomy, asexual fungi have been placed in the Subdivision Deuteromycotina (Ainsworth 1966), which comprised three classes, Coelomycetes, Hyphomycetes and Agonomycetes (mycelia sterilia) (Sutton 1980). None of these taxa are used anymore, but the designations, coelomycetes and hyphomycetes, are still frequently used in an informal, descriptive manner. Kendrick (1989) stressed the importance of incorporating asexual fungi and sexual fungi into one classification system and Taylor (1995) discussed the possibility of using one classification system with the application of DNA sequences and phylogeny. In recent years, multi-locus phylogenetic analyses have come at the forefront of modern fungal taxonomy (e.g. Jiang et al. 2020, Samarakoon et al. 2020, Wibberg et al. 2020). Furthermore, DNA sequence analyses are used to link asexual morphs with sexual morphs. As a result of these developments, dual nomenclature that existed for asexual and sexual morphs was discontinued (Hawksworth et al. 2011), and a base for a global classification scheme for fungi developed.

Previous outlines of fungi and their limitations

Classification of sexual fungi has been presented as an outline by Eriksson (1982, 1999, 2001, 2005), Eriksson & Hawksworth (1986, 1998a, b), and Lumbsch & Huhndorf (2007, 2010). Hyde et al. (2011) published the first outline of asexual genera with links to their sexual genera, and this was updated by Wijayawardene et al. (2017b). Wijayawardene et al. (2018) incorporated both asexual and sexual genera in the most recent Outline of the *Ascomycota*. Wijayawardene et al. (2017a, b, 2018, 2020) and He et al. (2019) provided notes for all fungal (including fossil fungi) and fungus-like genera. Most recently, Wijayawardene et al. (2020) provided a classification of all fungi and fungus-like taxa as an outline that was authored by over 150 mycologists. These authors recognized two crucial limitations while compiling the outline.

- 1. As classification is not an exact science, there can be disagreement and dispute among different research groups on taxonomic boundaries of genera and higher taxa.
- 2. As phylogenetic data are still being collected at a very high pace and increasingly being used in taxonomy, there is the necessity of constantly updating classification schemes and incorporate new findings.

Overcoming limitations of static publication

Disagreements on the classification and identification of fungi can occur as a result of different types of DNA analyses performed, different gene regions analysed, the impact of taxonomic sampling, the significance of specific morphs used in species delineation, and the inclusion of type species and type specimens in analyses. Therefore, Wijayawardene et al. (2020)

included some alternative views on higher-level classification. It is important for readers to evaluate different classification proposals and make decisions as to which ones they will follow based on the available data and expert opinion. Collection of updated data was also a huge challenge while compiling the outline of Wijayawardene et al. (2020) as fungi and fungus-like organisms are extremely diverse and their classification and taxonomy is in a constant state of flux. As a consequence, a 'cut-off' date for gathering published data was set by Wijayawardene et al. (2020), as the authors were well-aware that continuous updating of the classification scheme would be necessary. As a result, the importance of a flexible, online platform has been discussed as a means to overcome static classification proposals.

Other online databases

Online databases are used as tools in current studies in mycology, including taxonomy, nomenclature and classification (Prakash et al. 2017, Větrovský et al. 2020). For example, Index (http://www.indexfungorum.org), MycoBank (https://www.mycobank.org) FungalNames (http://www.fungalinfo.net/) are important web tools that mainly deal with nomenclature. Several other important web based databases such as onestopshopfungi.org (https://onestopshopfungi.org/), dothideomycetes.org (https://dothideomycetes.org/), (https://fungalgenera.org/), fungalgenera.org facesoffungi.org (http://www.facesoffungi.org/), theyeasts.org (https://theyeasts.org/) provide information on pathogenic genera, Dothideomycetes genera, typification data, descriptions of species and other taxonomic ranks, and yeast genera, respectively (Jayasiri et al. 2015, Jayawardena et al. 2019, Monkai et al. 2019, Pem et al. 2019). Doctor Fungus (http://www.mycosesstudygroup.org/), Mycology (http://www.mycology.adelaide.edu.au/), and the Aspergillus and Aspergillosis Website (http://www.aspergillus.org.uk/) are dedicated websites for clinically important fungi. The website http://www.marinefungi.org deals with the latest taxonomy of marine fungi (Jones et al. 2019). None of these databases provides general changes in consensus classification and the reasons for changes. Hence, it is essential to have a database, which, based on new studies, is being continuously updated with balanced changes of a community-supported consensus classification.

Outline of Fungi web page

The Outline of *Fungi* web site is established to address the limitations recognized during the compilation of the published Outline of *Fungi* (Wijayawardene et al. 2020). The main objectives of the online resource, 'outlineoffungi.org' are to:

- 1. Present a continually updated consensus classification of fungi and fungus-like organisms based on recent literature and expert opinion.
- 2. Provide notes for important changes and additions to the outline.
- 3. Provide an opportunity for researchers to add missing data, suggestions to modify data, and provide critical comments on the consensus outline, based on their expert opinion.

Fossil fungi

Fossil fungi are the ancestors of current fungi and represent extinct lineages, some of which evolved relatively early, probably more than 1500 million years ago. They were buried in sediments, silicified or trapped in amber, and have been preserved until now in sedimentary rocks (Samarakoon et al. 2020). They generally tend to be microscopic. However, a few large fungal bodies, such as mushrooms, have also been found as fossils. Fossil fungal remnants are found in the form of spores, mycelia, sporophores, symbiotic associations, and are commonly observed in macerated residues prepared for palynological studies. Although fungal remains are encountered in sediments of all ages, their frequency increases remarkably in the Tertiary Period (Dilcher 1965, Pirozynski & Weresub 1979, Kalgutkar & Jansonius 2000, Saxena & Tripathi 2011). Because of the fragmentary remains that make up the vast majority of fungal remains, fossil fungi usually lack characteristic features that are diagnostic, hampering their classification with extant fungi. Since DNA survival is limited, they are described on the basis of morphological characters only. For

example, spore taxa are characterised on their shape, size, symmetry, number and nature of apertures, septa and spore wall characters, which often allow up to genus-level classifications. In addition, fossil sporocarps without spores or hyphal details can mostly be assigned to order level or above. All fossil fungal taxa are registered with a recognized nomenclatural repository, e.g. Index Fungorum/MycoBank, to make their references accessible world-wide. This also helps ensuring that they are validly published, avoiding unnecessary introductions of later homonyms.

Fungus-like taxa

Fungal-like characteristics have evolved multiple times, both in prokaryotes (e.g. Myxobacteria) and eukaryotes (e.g. slime molds, Labyrinthulomycota, Oomycota). Myxobacteria are, however, almost exclusively studied by bacteriologists, and many members Labyrinthulomycota have been described under the zoological code in the corresponding phylum Labyrinthulida. In more recent classifications of myxomycetes, which have long been described under the *International Code of Nomenclature for algae, fungi and plants* (ICNafp), a classification entirely based on the *International Code of Zoological Nomenclature* (ICZN) is adopted (Adl et al. 2005). However, but this approach was criticized by Ronikier & Halamski (2018), who emphasized that the transfer of myxomycetes to zoological nomenclature would destabilize the nomenclature of the group, due to the existence of numerous homonyms. For this reason, the most recent published classification of Eumycetozoa was based on the botanical nomenclature (Leontyev et al. 2019). However, most of the non-Eumycetozoan slime molds, such as protosteloids, acrasids, copromyxids, fonticulids, or guttulinopsids, were described by protozoologists using zoological nomenclature. These groups, therefore, should be covered by ICZN. However, the Preamble 8 of the ICNafp states that the 'slime molds' are considered among the organisms for which the Code is applied, while ICZN does not mention these organisms at all, so coordination between ICZN and ICNafp is needed to resolve this issue.

The group most closely resembling opisthokont fungi are *Oomycota*, because of their hyphal growth, osmotrophic nutrition and formation of large numbers of asexual spores (Dick 2001, Beakes & Thines 2017). Because of these similarities, they have always been described under the ICNafp and its predecessors. Their early-diverging lineages mostly form only small, holocarpic thalli, i.e. the entire thallus converts into a sporangium. This is similar to the trophocytes found in some other members of the Straminipila-Alveolata-Rhizaria supergroup (Burki et al. 2007) groups, such as Perkinsozoa. However, the monophyletic branch starting from Miraculaceae (Buaya et al. 2017) until the highly diversified downy mildews (Thines & Choi 2016) is commonly recognised as the phylum *Oomycota* (Beakes & Thines 2017), and thus, treated under the ICNafp. Within Oomycota, the higher-level classification is still not fully resolved, owing to the difficulties in obtaining multiple genes for in-depth phylogenetic analyses from the often obligate biotrophic and holocarpic lineages that diverge before the main split into Saprolegniomycetes and Peronosporomycetes (Beakes & Thines 2017). Another poorly-known group related to Oomycota, Hyphochytriomycota, is usually treated under the ICNafp as well, but only few species of this group have been discovered and their higher-level relationships, as well as their relationships to some bacteriovorous protists and oomycetes are still poorly resolved. Thus, it is expected that for both oomycetes and hyphochytrids, there will be considerable efforts necessary to stabilise their nomenclature, which will also be reflected in the Outline of Fungi website.

Construction

As a starting point, all fungal genera in the database are listed according to Wijayawardene et al. (2020). The database will be updated based on new studies and observations by the users.

Database interface and visualization

The homepage comprises nine tabs. Viewers can follow them and use the underlying functions with a simple and user-friendly interface (Fig. 1).

Tools included in homepage

- 1. Home: The homepage provides an overview of the kingdom *Fungi*. Objectives of launching the website are also provided. (Fig. 1)
- 2. Outline: The 'Outline' provides a recent consensus taxonomic classification of the kingdom *Fungi* and fungus-like taxa of other kingdoms (e.g., slime moulds, oomycetes). (Fig. 2)
- 3. Archives: The 'Archives' provides recently published outlines.
- 4. Sexual-asexual links: This will be updated with all pleomorphic genera.
- 5. Curators: This section provides a list and contact details of the curators of the website (see Table 1).
- 6. History: This section provides a brief history of the classification of kingdom *Fungi* and fungus-like taxa.
- 7. References: This section provides reference list of citations used in the entries, history and related information.
- 8. Notes: The 'Notes' section publishes recent changes in outline since Wijayawardene et al. 2020 (see below for further details).
- 9. Contact: The 'Contact' section provides contact details for the website and allows users to address any comments and suggestions.

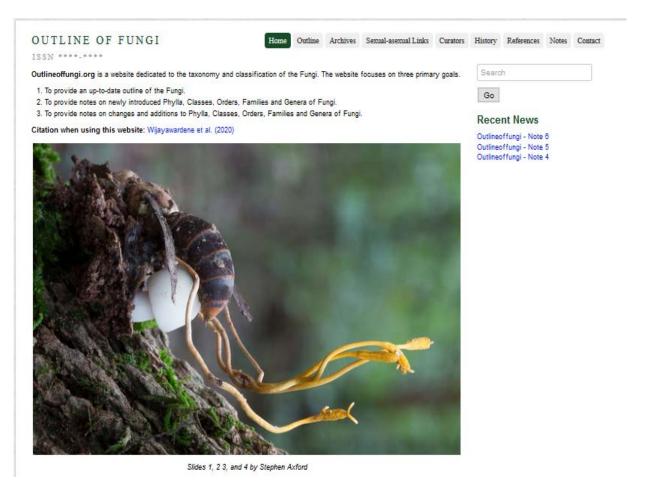


Figure 1 – The homepage view of outlineoffungi.org

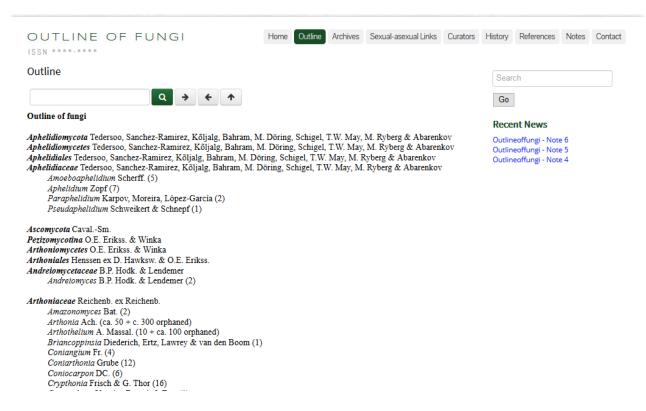


Figure 2 – Outline of fungi

Table 1 List of expert curators for Outline of *Fungi* webpage

Position	Name	Field of specialty	Contact details
Head curator	Nalin N. Wijayawardene	Asexual fungi, nomenclature	nalinwijayawardene@yahoo.com
Senior	Kevin D. Hyde	Dothideomycetes,	kdhyde3@gmail.com
Curator		Sordariomycetes	
Managing curators	Makbule Erdoğdu	Fungal plant pathogens	merdogdu@ahievran.edu.tr
	Andrei Tsurykau	Lichens, lichenicolous fungi	tsurykau@gmail.com
	Shiva Prakash Nedle	Basidiomycota, hyphomycetes	shivanedle@gmail.com
	Josiane S. Monteiro	Asexual fungi	kiotobelbio2003@yahoo.com.br
Curators	Alan J.L. Phillips	Botryosphaeriales, pathogens	alan.jl.phillips@gmail.com
	André Aptroot	Lichens	andreaptroot@gmail.com
	Bruno T. Goto	Glomeromycota	brunogoto@hotmail.com
	Danny Haelewaters	Leotiomycetes, Laboulbeniomycetes	danny.haelewaters@gmail.com
	Eleni Gentekaki	Basal fungi	gentekaki.ele@mfu.ac.th
	Damien Ertz	Lichens, lichenicolous fungi	damien.ertz@jardinbotaniquemeise.be
	Hugo Madrid	hyphomycetes, Eurotiomycetes	hugo.madrid@gmail.com
	Marc Stadler	<i>Xylariales</i> , secondary metabolites of fungi	marc.stadler@helmholtz-hzi.de
	Alfredo Vizzini	Basidiomycota, Agaricales, Boletales	alfredo.vizzini@unito.it
	Marco Thines	Oomycota, Ustilaginomycotina, nomenclature	marco.thines@senckenberg.de
	Chuan-Gen Lin	Hyphomycetes	chuangenlin@gmail.com
	Chain Och Em	Tijphomyceus	chamigemine ginan.com

Table 1 Continued.

Position	Name	Field of specialty	Contact details	
	Yu-Cheng Dai	Basidiomycota	yuchengd@yahoo.com	
	Patrícia O. Fiuza	Fresh water fungi, asexual fungi	patyfiuzabio@gmail.com	
	Paul M. Kirk	Nomenclature P.Kirk@kew.org		
	Don-Qin Dai	Dothideomycetes	cicidaidongqin@gmail.com	
	Ramesh K. Saxena	Fossil fungi	rksaxena2207@yahoo.com	
	Rafael F. Castañeda- Ruiz	hyphomycetes	rfcastanedaruiz@gmail.com	
	Irina S. Druzhinina	Asexual fungi	irina.s.druzhinina@mail.ru	
	Feng Cai	Asexual fungi	czfscf@hotmail.com	
	Anusha H. Ekanayaka	Leotiomycetes	hasinie88@gmail.com	
	Sajeewa S.N. Maharachchikumbura	Sordariomycetes	sajeewa83@yahoo.com	
	Sinang Hongsanan	Sooty moulds	sinang333@gmail.com	
	E.B. Gareth Jones	Aquatic fungi	torperadgj@gmail.com	
	Walter P. Pfliegler	Yeast (Ascomycota)	pfliegler.valter@science.unideb.hu	
	Samantha C. Karunarathna	Basidiomycota	samanthakarunarathna@gmail.com	
	Hyang Burm Lee	Basal fungi	hblee@jnu.ac.kr	
	Javier Etayo	Lichens, lichenicolous fungi	jetayosa@educacion.navarra.es	
	H. Thorsten Lumbsch	Lichens	tlumbsch@fieldmuseum.org	
	Rajesh Jeewon	Asexual fungi	r.jeewon@uom.ac.mu	
	R.G. Udeni Jayalal	Lichens	jayalal@appsc.sab.ac.lk	
	Qing Tian	Eurotiomycetes	tianqing124@gmail.com	
	Dhanushka N. Wanasinghe	Pleosporales	dnadeeshan@gmail.com	
	Xinlei Fan	Diaporthales, Botryosphaeriales, forest pathogenic fungi	xinleifan@bjfu.edu.cn	
	Eric H.C. McKenzie	Pucciniomycotina, Ustilaginomycotina, hyphomycetes	mckenziee@landcareresearch.co.nz	
	Teun Boekhout	Yeasts	t.boekhout@wi.knaw.nl	
	Vince Hustad	Geoglossales	vhustad@gmail.com	
	Dmitry Leontyev	Slime moulds	alwisiamorula@gmail.com	
	Sybren de Hoog	Clinical fungi	Sybren.deHoog@radboudumc.nl.	
	Tuula Niskanen	Agaricomycetes	t.niskanen@kew.org	
	Roger G. Shivas	Plant pathogens	roger.shivas@usq.edu.au	
	Gaya Ester	Lichens	e.gaya@kew.org	
	Wen-Jing Li	Asexual fungi	winnie20070653026@163.com	

Notes section

The publication of notes is recognized as an important part of outlineoffungi.org, which provides details of new additions, changes or opinions. This provides an opportunity for interested parties to better understand recent changes. It will also provide a platform to express opinions and judgements on fungal taxonomy with respect to the identification and placement of fungi in different classification schemes.

Changes to the outline/ classification could be due to three main reasons:

- 1. Changes to classification following recent publications.
- 2. Additions of taxa missing in Wijayawardene et al. (2020).
- 3. Correction or errors in Wijayawardene et al. (2020) (e.g. duplication of names, wrong placement of taxa).

Preparing notes

1. Addition of new taxa from new publications

The addition of new taxa will be coordinated with repositories such as Index Fungorum. The published materials/research articles that introduce new taxa will cross checked against repository lists by the four managing curators. Their main task will be to prepare entries for new additions. As the second step, the entries will be sent to curator/s for checking. Once the managing curator has edited the entry according to curators' comments, the senior curator and the head curator will check the entries. The head curator will cross check the validity of the taxa against repositories (e.g. Index Fungorum) and upload the entry.

A list of new taxa (genus level and above) will be gathered from Index Fungorum twice a year and cross checked against the entries provided in outlineoffungi.org. This will help to maintain a current outline.

Authors who publish new taxa are also encouraged to provide entries. This includes resurrection of synonymized genera in recent publications (e.g. Thiyagaraja et al. 2020).

2. Addition of missing taxa

Notes for missing taxa will be mainly expected from expert mycologists. They can prepare the entries and send these to the head curator. The curator/s will check and correct the entries which will be uploaded to the webpage.

3. Correcting mistakes in the existing outline

Notes which correct mistakes (such as duplication of names, incorrect author citations) will also be acceptable. However, the senior curator and the head curator will decide whether it is necessary to upload the note, or correct the web version of the outline.

OUTLINE OF FUNGI	Home Outline	e Archives Sexual-asexual Links	Curators	History References Notes Contact
Notes				Search
Outlineoffungi - Note 6				Go Recent News
Outlineoffungi - Note 5				Outlineoffungi - Note 6 Outlineoffungi - Note 5
Outlineoffungi - Note 4				Outlineoffungi - Note 4
Outlineoffungi - Note 3				
Outlineoffungi - Note 2				
Outlineoffungi - Note 1				

Figure 3 – Notes section

A platform to exchange different opinions

Different opinions on higher classification and synonymy of pleomorphic genera (Art. 59.1) are two important topics that can lead to confusion. For example, the higher-level classification of *Leotiomycetes* in Ekanayaka et al. (2019) is different from the classification in Johnston et al. (2019). Two main differences account for the alternative classification schemes: (i) the amount of sequence data [5-locus by Ekanayaka et al. (2019) vs 15-locus and genomic-scale by Johnston et al. (2019)] and (ii) the inclusion of type species and ex-type sequences in Johnston et al's (2019) analyses. However, the presentation of the alternative classifications provide perspective for

general users and mycologists interested in *Leotiomycetes* taxonomy. Further, if other authors have different opinions on existing classifications, we encourage them to provide them in the notes section.

Adopted or proposed names for pleomorphic genera are also controversial in some cases. For example, Wijayawardene et al. (2014) proposed to adopt *Anthracostroma* Petr. 1954 over *Camarosporula* Petr. 1954, but Rossman et al. (2015) did not agree and proposed to adopt the latter name over the former. This type of disagreement and controversial opinions might cause confusion among users if future authors referred to only one publication. Thus, it is essential to have a platform such as http://outlineoffungi.org to provide different opinions, which can eventually culminate in a consensus towards species concepts.

Links between sexual and asexual morphs

The new webpage also provides links between sexual and asexual morphs with the names that have been adopted for pleomorphic genera since 2011 (Art. 59.1). Wijayawardene et al. (2017b) provided the adopted names of pleomorphic genera based on previous studies (e.g. Wijayawardene et al. 2014, Rossman et al. 2015), but it is intended to include all pleomorphic genera according to recent developments and recommendations from the *International Commission on the Taxonomy of Fungi*.

A place to obtain data (from genus to higher rank) for phylogenetic analyses

Selecting taxa for phylogenetic studies is a challenge as it is important to include closely related genera in the analyses. Extracting data from static publications might cause problems due to outdated views on phylogenetic relationships. Thus, it is important to obtain data from a database that is kept updated. Since outlineoffungi.org will be continuously updated, with the supervision of senior taxonomists, users can obtain a list of related genera from it for their phylogenetic analyses and morphological comparisons.

Acknowledgements

This work was supported by the Key Laboratory of Yunnan Province Universities of the Diversity and Ecological Adaptive Evolution for Animals and Plants on the Yun-Gui Plateau, the National Natural Science Foundation of China (No. NSFC 31950410558, NSFC 31760013, 31260087, 31460561), and the Scientific Research Foundation of Yunnan Provincial Department of Education (2017ZZX186). Nalin N. Wijayawardene gratefully acknowledges grant number FAMP201906K provided by Guizhou Medical University. Kevin D. Hyde thanks the Foreign Experts Bureau of Yunnan Province, Foreign Talents Program (2018; grant no. YNZ2018002), Thailand Research grants entitled Biodiversity, phylogeny and role of fungal endophytes on above parts of *Rhizophora apiculata* and *Nypa fruticans* (grant no: RSA5980068), the future of specialist fungi in a changing climate: baseline data for generalist and specialist fungi associated with ants, *Rhododendron* species and *Dracaena* species (grant no: DBG6080013), Impact of climate change on fungal diversity and biogeography in the Greater Mekong Subregion (grant no: RDG6130001). Dong-Qin Dai would like to thank the Thousand Talents Plan, Youth Project of Yunnan Provinces for support. Alan JL Phillips acknowledges the support from UIDB/04046/2020 and UIDP/04046/2020 Centre grants from FCT, Portugal (to BioISI).

References

Adl SM, Simpson AG, Farmer MA, Andersen RA et al. 2005 – The new higher level classification of eukaryotes with emphasis on the taxonomy of protists. Journal of Eukaryotic Microbiology 52, 399–451. Doi: 10.1111/j.1550-7408.2005.00053.x

Ainsworth GC. 1966 – A general purpose classification of fungi. Bibliography of Systematic Mycology 1, 1–4.

- Beakes GW, Thines M. 2017 *Hyphochytriomycota* and *Oomycota*. In: Archibald J, Simpson A, Slamovits C (eds) Handbook of the Protists. Springer International Publishing, Cham, pp. 435–505.
- Buaya AT, Ploch S, Hanic L, Nam B et al. 2017 Phylogeny of *Miracula helgolandica* gen. et sp. nov. and *Olpidiopsis drebesii* sp. nov., two basal oomycete parasitoids of marine diatoms, with notes on the taxonomy of Ectrogella-like species. Mycological Progress 16, 1041–1050. Doi: 10.1007/s11557-017-1345-6
- Burki F, Shalchian-Tabrizi K, Minge M, Skjæveland Å et al. 2007 Phylogenomics reshuffles the eukaryotic supergroups. PloS ONE 2, e790. Doi: 10.1371/journal.pone.0000790
- Dick MW. 2001 Straminipilous Fungi. Springer, Dordrecht.
- Dilcher DL. 1965 Epiphyllous fungi from Eocene deposits in western Tennessee, U.S.A. Palaeontographica Abt. B 116, 1–54.
- Ekanayaka AH, Hyde KD, Gentekaki E, McKenzie EHC et al. 2019 Preliminary classification of Leotiomycetes. Mycosphere 10, 310–489. Doi: 10.5943/mycosphere/10/1/7
- Eriksson OE. 1982 Outline of the Ascomycetes 1982. Mycotaxon 15, 203–248.
- Eriksson OE. 1999 Outline of Ascomycota 1999. Myconet 3, 1–88.
- Eriksson OE. 2001 SSU rDNA sequences from Ascomycota. Myconet 6, 27–76.
- Eriksson OE. 2005 Outline of Ascomycota 2005. Myconet 11, 1–113.
- Eriksson OE, Hawksworth DL. 1986 Outline of the ascomycetes-1986. Systema Ascomycetum 5, 185–324.
- Eriksson OE, Hawksworth DL. 1998a Outline of the ascomycetes 1998. Systema Ascomycetum 16, 83–296.
- Eriksson OE, Hawksworth DL. 1998b Notes on ascomycete systematics. Nos 2256–2439. Systema Ascomycetum 16, 39–81.
- Hawksworth DL, Crous PW, Redhead SA, Reynolds DR et al. 2011 The Amsterdam declaration on fungal nomenclature. IMA Fungus 2: 105–112.
- He MQ, Zhao RL, Hyde KD, Begerow D et al. 2019 Notes, outline and divergence times of *Basidiomycota*. Fungal Diversity 99, 105–367. Doi: 10.1007/s13225-019-00435-4.
- Hyde KD, McKenzie EHC, KoKo TW. 2011 Towards incorporating anamorphic fungi in a natural classification –checklist and notes for 2010. Mycosphere 2, 1–88.
- Jiang S, Lücking R, Xavier-Leite AB, Marcela ES et al. 2020 Reallocation of foliicolous species of the genus *Strigula* into six genera (lichenized *Ascomycota*, *Dothideomycetes*, *Strigulaceae*). Fungal Diversity. Doi: 10.1007/s13225-020-00445-7.
- Jayawardena RS, McKenzie EHC, Chen YJ, Phillips AJL et al. 2019 https://onestopshopfungi.org/, a database to enhance identification of phytopathogenic genera. Asian Journal of Mycology 2, 281–286. Doi: 10.5943/ajom/2/1/18
- Jayasiri SC, Hyde KD, Ariyawansa HA, Bhat DJ et al. 2015 The Faces of Fungi database: fungal names linked with morphology, phylogeny and human impacts. Fungal Diversity 74, 3–18. Doi: 10.1007/s13225-015-0351-8
- Jones EBG, Pang KL, Abdel-Wahab MA, Scholz B, et al. 2019 An online resource for marine fungi. Fungal Diversity 96, 347–433.
- Johnston PR, Quijada L, Smith CA, Baral HO et al. 2019 A multigene phylogeny toward a new phylogenetic classification for the *Leotiomycetes*. IMA Fungus 10, 1. Doi: 10.1186/s43008-019-0002-x
- Kalgutkar RM, Jansonius J. 2000 Synopsis of fungal spores, mycelia and fructifications. AASP Contribution Series 39, 1–423.
- Kendrick WB. 1989 Subdivision *Deuteromycotina* a fungal chimera. Sydowia 41, 6–14.
- Leontyev DV, Schnittler M, Stephenson S, NovozhilovYK et al. 2019 Towards a phylogenetic classification of Myxomycetes. Phytotaxa 399, 209–238.
- Lumbsch HT, Huhndorf SM. 2007 Outline of Ascomycota-2017. Myconet 13, 1–99.

- Lumbsch HT, Huhndorf SM. 2010 Myconet Volume 14. Part one. Outline of Ascomycota 2009. Part Two. Notes on Ascomycete Systematics. Nos. 4751–5113. Fieldiana Life Earth Sci 1, 1–64. Doi: 10.3158/1557.1
- Monkai J, McKenzie EHC, Phillips AJL, Hongsanan S et al. 2019 https://fungalgenera.org/: a comprehensive database providing webbased information for all fungal genera. Asian Journal of Mycology 2, 298–305. Doi: 10.5943/ajom/2/1/20
- Pem D, Hongsanan S, Doilom M, Tibpromma S et al. 2019 https://www.dothideomycetes.org: An online taxonomic resource for the classification, identification, and nomenclature of *Dothideomycetes*. Asian Journal of Mycology 2, 287–297. Doi: 10.5943/ajom/2/1/19
- Pirozynski KA, Weresub LK. 1979 The classification and nomenclature of fossil fungi. in Kendrick, B. (ed.), The whole fungus, the sexual–asexual synthesis. Proceedings of the 2nd International Mycological Conference, University of Calgary, Kananaskis, Alberta (published by National Museum of Natural Sciences, National Museums of Canada and the Kananaskis Foundation) volume 2, 653–688.
- Prakash PY, Irinyi L, Halliday C, Chen S et al. 2017 Online Databases for Taxonomy and Identification of Pathogenic Fungi and Proposal for a Cloud-Based Dynamic Data Network Platform. Journal of Clinical Microbiology 55, 1011–1024. Doi: 10.1128/JCM.02084-16
- Quijada L, Tanney JB, Popov E, Johnston PR et al. 2020 Cones, needles and wood: *Micraspis* (*Micraspidaceae*, *Micraspidales* fam. et ord. nov.) speciation segregates by host plant tissues. Fungal Systematics and Evolution 5, 99–112. Doi: 10.3114/fuse.2020.05.05
- Ronikier A, Halamski AT. 2018 Is Myxomycetes (Amoebozoa) a truly ambiregnal group? A major issue in protist nomenclature. Protist 169, 484–493.
- Rossman AY, Crous PW, Hyde KD, Hawksworth DL et al. 2015 Recommended names for pleomorphic genera in *Dothideomycetes*. IMA fungus 6, 507–523. Doi: 10.5598/imafungus.2015.06.02.14
- Saccardo PA. 1904 De Diagnostica et nomenclatura mycologica, Admonita quaedam. Annales Mycologici 2, 195–198. [English translation by Clements FE (1904) J Mycol 10, 109–112]
- Samarakoon MC, Thongbai B, Hyde KD, Brnstrup M et al. 2020 Elucidation of the life cycle of the endophytic genus *Muscodor* and its transfer to *Induratia* in Induratiaceae fam. nov., based on a polyphasic taxonomic approach. Fungal Diversity 101, 177–210. Doi: 10.1007/s13225-020-00443-9
- Saxena RK, Tripathi SKM. 2011 Indian Fossil Fungi. Palaeobotanist 60, 1–208.
- Sutton BC. 1980 The Coelomycetes. Fungi imperfecti with pycnidia, acervuli and stromata. Commonwealth Mycological Institute, Kew.
- Taylor JW. 1995 Making the Deuteromycota redundant: a practical integration of mitosporic and meiosporic fungi. Canadian Journal of Botany 73 (Suppl.), s754–s759.
- Thines M, Choi YJ. 2016 Evolution, diversity, and taxonomy of the *Peronosporaceae*, with focus on the genus *Peronospora*. Phytopathology 106, 6–18. Doi: 10.1094/PHYTO-05-15-0127-RVW
- Thiyagaraja V, Lücking R, Ertz D, Wanasinghe DN et al. 2020 Evolution of non-lichenized, saprotrophic species of *Arthonia (Ascomycota, Arthoniales)* and resurrection of *Naevia*, with notes on *Mycoporum*. Fungal Diversity. Doi: 10.1007/s13225-020-00451-9
- Větrovský T, Morais D, Kohout P, Lepinay C et al. 2020 Global Fungi, a global database of fungal occurrences from high-throughput-sequencing metabarcoding studies. Scientific Data 7, 228. Doi: 10.1038/s41597-020-0567-7
- White TJ, Bruns T, Lee S, Taylor JW. 1990 Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ & White TJ (eds) PCR protocols: a guide to methods and applications, New York Academic Press, pp 315–322
- Wibberg D, Stadler M, Lambert C, Bunk B et al. 2020 High quality genome sequences of thirteen Hypoxylaceae (Ascomycota) strengthen the phylogenetic family backbone and enable the discovery of new taxa. Fungal Diversity. Doi: 10.1007/s13225-020-00447-5

- Wijayawardene NN, Crous PW, Kirk PM, Hawksworth DL et al. 2014 Naming and outline of *Dothideomycetes*–2014 including proposals for the protection or suppression of generic names. Fungal Diversity 69: 1–55. Doi: 10.1007/s13225-014-0309-2
- Wijayawardene NN, Hyde KD, Rajeshkumar KC, Hawksworth DL et al. 2017a Notes for genera: *Ascomycota*. Fungal Diversity 86, 1–594. Doi: 10.1007/s13225-017-0386-0
- Wijayawardene NN, Hyde KD, Tibpromma S, Wanasinghe DN et al. 2017b Towards incorporating asexual fungi in a natural classification: checklist and notes 2012–2016. Mycosphere 8:1457–1554. Doi: 10.5943/mycosphere/8/9/10
- Wijayawardene NN, Hyde KD, Lumbsch HT, Liu JK et al. 2018 Outline of Ascomycota: 2017. Fungal Diversity 88, 167–263. Doi: 10.1007/s13225-018-0394-8
- Wijayawardene NN, Hyde KD, Al-Ani LK, Tedersoo L et al. 2020 Outline of Fungi and fungus-like taxa. Mycosphere 11, 1060–1456. Doi: 10.5943/mycosphere/11/1/8