

Accepted Manuscript

---

This document is the Accepted Manuscript version of a Published Work that appeared in final form in *Environmental Science and Technology*, copyright © American Chemical Society after peer review and technical editing by the publisher.

To access the final edited and published work see  
<https://doi.org/10.1021/acs.est.1c04957>

Mengyu Bai, Yan Lin, Rachel R. Hurley, Lixin Zhu, and Daoji Li. Controlling Factors of Microplastic Riverine Flux and Implications for Reliable Monitoring Strategy. *Environmental Science & Technology*. 2021. 56(1), 48-61.

It is recommended to use the published version for citation.

---

# Controlling factors of microplastic riverine flux and implications for reliable monitoring strategy

Mengyu Bai<sup>1,2,3</sup>, Yan Lin<sup>4</sup>, Rachel Hurley<sup>4</sup>, Lixin Zhu<sup>1,2,3</sup>, Daoji Li<sup>1,2,3\*</sup>

<sup>1</sup> State Key Laboratory of Estuarine and Coastal Research, East China Normal University, 500 Dongchuan Road, Shanghai 200241, China

<sup>2</sup> Plastic Marine Debris Research Center, East China Normal University, 500 Dongchuan Road, Shanghai 200241, China

<sup>3</sup> Regional Training and Research Center on Plastic Marine Debris and Microplastics, IOC-UNESCO, 500 Dongchuan Road, Shanghai 200241, China

<sup>4</sup> Norwegian Institute for Water Research (NIVA), Gaustadelléen 21, 0349 Oslo, Norway

**Abstract:** A significant proportion of marine plastic debris and microplastics is assumed to be derived from river systems. In order to effectively manage plastic contamination of the marine environment, an accurate quantification of riverine flux of land-based plastics and microplastics is imperative. Rivers not only represent pathways to the ocean, but are also complex ecosystems that support many life processes and ecosystem services. Yet riverine microplastics research is still in its infancy, and many uncertainties still remain. Major barriers exist in two aspects. First, nonharmonized sampling methodologies make it problematic for compiling data across studies to better estimate riverine fluxes of microplastics globally; Second, the significant spatiotemporal variation of microplastics in rivers which was affected by the river characteristics, MPs properties, etc. also have important influence on the estimation of riverine MPs fluxes. In this study, we made a comprehensive review from the above two aspects based on published peer-reviewed studies and provide recommendations and suggestions for a reliable monitoring strategy of riverine MPs, which is beneficial to the further establish sampling methods for rivers in different geographical locations. Besides, methods for achieving a high level of comparability across studies in different geographical

28 contexts are highlighted. Riverine microplastic flux monitoring is another important part of  
29 this manuscript. The influential factors and calculation methods of microplastic flux in rivers  
30 are also discussed in this paper.

31 **Key words:** River water; Microplastic; Monitoring strategy; Sampling methodologies; Flux

## 32 1. Introduction

33 Microplastics (MPs), defined as plastic debris smaller than 5 mm<sup>1</sup>, are derived from the  
34 breakdown of larger plastic items (e.g., fibrous or fragmented MPs) or are deliberately  
35 produced (e.g., microbeads or glitter). The mismanagement of plastic litter worldwide has led  
36 to the widespread occurrence of MPs<sup>2</sup>. MPs have similar shapes and sizes to plankton and can  
37 be unintentionally ingested by fish, filter feeders and planktivores<sup>3,4</sup>. After entering organisms,  
38 MPs can be transferred through the food web, which has been detected in both invertebrates  
39 and zooplankton<sup>5,6</sup>; although, the true risk associated with this is still yet to be fully articulated.  
40 MPs can be transported over long distances to remote islands<sup>7</sup>, polar regions<sup>8</sup>, Arctic Sea ice  
41<sup>9</sup>, and mountain lakes in Mongolia<sup>10</sup> by physical factors (such as wind, ocean currents and  
42 river flow)<sup>11-13</sup>. The ocean is viewed as a major MP sink, whereas rivers are viewed as  
43 important pathways for plastic litter transport into the oceans. Meijer et al.<sup>14</sup> revealed that  
44 1000 rivers globally transport 0.8-2.7 million metric tonnes of plastic litter each year to the  
45 ocean, accounting for 80% of annual global emissions. The Danube River was estimated to  
46 transport 1553 tonnes of plastic debris to the Black Sea annually at a rate of 7.5 mg/m<sup>3</sup>•s<sup>15</sup>.  
47 River basins are the main contributors of plastics to estuaries, where transportation and  
48 accumulation patterns are determined by the fluctuation of flow regimes<sup>16</sup>. Other studies have  
49 also confirmed river basins as the major sources of inland MPs to estuaries, as well as exporters  
50 of MPs to the oceans<sup>17-20</sup>.

51 Yet, river environments represent a crucial component of the global hydrosphere and  
52 biosphere, supporting important ecological diversity. Rivers are often described in terms of

53 conduits of plastic to the ocean, but they are complex and dynamic systems that can  
54 accumulate, store, and remobilize plastic particles over different spatial and temporal scales.  
55 The majority of plastics are produced, consumed, and disposed of on land <sup>21</sup>. Due to this  
56 quantity and extent of MP sources, rivers often exhibit elevated MP concentrations compared  
57 to the marine environment <sup>22</sup>. High MP concentrations have been found in both river sediment  
58 <sup>23-25</sup> and surface waters <sup>26, 27</sup>. Many riverine ecosystems globally are expected to be exposed  
59 to MP in water and sediments, and these organisms also need protection from the potential  
60 adverse effects of contamination. Especially given the high degree of complexity in river  
61 systems, the fate and transport of MP remains relatively under-researched. The behavior of  
62 plastic litter in freshwater, especially in rivers, varies greatly from that in the marine  
63 environment, and freshwater-specific studies are required to investigate relevant processes of  
64 MP movement and accumulation in rivers. Inconsistent sampling methods hinder the  
65 possibility to compare between results, and different sample types reveal different snapshots  
66 of riverine MP contamination <sup>28</sup>. Hermsen et al. <sup>29</sup> established a means for assessing  
67 methodological and data quality based on 10 criteria, and found that the information integrity  
68 of most studies needs to be further improved. Further publications have since proposed  
69 reporting guidelines for MP studies, covering topics from field sampling to quantification from  
70 laboratory data <sup>30</sup>.

71 River stratification and hydrodynamic action have a great influence on the distribution  
72 and transport characteristics of plastic litter <sup>31-33</sup>. Artificial structures, such as dams <sup>34-36</sup>,  
73 bridges <sup>37</sup> and human-created tributaries <sup>38</sup>, hydropower station <sup>39</sup>, as well as natural  
74 characteristics, such as riparian vegetation <sup>40</sup> and river curvature <sup>41</sup>, geographical factors  
75 determined by the shape of the river <sup>42</sup> will affect river flow and may lead to the accumulation  
76 of plastic litter and MP. Hydrological conditions are also an important control on MP fate and  
77 transport, affecting the partitioning between the water and sediment phase <sup>28, 43, 44</sup>, influencing

78 the movement of MPs with channel bed sediments<sup>43,45,46</sup>, and acting as a control on the export  
79 of MPs downstream<sup>47</sup>.

80 Currently, the reported values from literatures utilize an array of sampling methods, size  
81 limits, processing methods, and different instrumentation under different conditions. There is  
82 also variability in the definition of MPs (such as the size categories analyzed) and the degree  
83 of quality control implemented into methodologies. Several studies have now reviewed the  
84 different sampling methodologies for assessment of MP occurrence in water and sediments<sup>42,</sup>  
85<sup>48-50</sup>. This article instead specifically focuses on the emission of MPs from rivers to the ocean  
86 on a certain time scale (riverine MP flux) and reviews the approaches to monitoring and  
87 establishing flux estimates within this context. This review intends to address the following  
88 key issues related to MP monitoring: (I) comparative analysis of the different sampling  
89 methods; and (II) review of the the calculation and influential factors of river MP flux, and  
90 requirements for improvement of the methods for both short-term field sampling and long-  
91 term monitoring to achieve more accurate river MP flux calculations.

## 92 **2. Materials and methods**

93 An extensive and systematic literature search was conducted in ISI Web of Science  
94 (WOS), Scopus, Google Scholar and Elsevier Science Direct for this review. The following  
95 key words were used in literature collecting: “microplastic(s)”, “river”, “fresh water”,  
96 “riverine”, “plastic flux” and “stream”. A total of 83 scientific articles and reviews about  
97 riverine MPs fluxes published before June, 2021 were included and reviewed. Data analysis  
98 and visualization were based on Python 3.8.5 and IBM SPSS Statistics 26.

99 This review includes global (locate in 5 Asian countries, 8 European countries, 3  
100 countries in the Americas, 1 African country and 2 Oceanian countries) rivers with different  
101 discharge, and covers multiple types of rivers locating in different climatic zones. The physical  
102 geography classifications of rivers include tributaries (e.g., Ottawa River and its tributaries,

103 Keelung River-the tributary of Tamsui River), river estuaries (e.g., Tamar estuary, Yangtze  
 104 River estuary), and the main stream of larger river catchments (e.g., Beijiang River, Pearl  
 105 River).

106 **3. Sampling methodologies for MPs in rivers**

107 To link the sources and fate of MPs in freshwater environments, it is necessary to  
 108 establish a unified approach to enhance data comparability. This review will introduce  
 109 sampling methodologies for MPs in rivers based on the different equipment and deployment  
 110 locations.

111 *3.1 Sampling equipment*

112 By reviewing studies about MPs in river water, sampling equipment can be summarized  
 113 as the following three types: i) Direct sampling-using precleaned stainless buckets, water  
 114 samplers, glass jars or other containers to directly collect surface water. In some studies, the  
 115 direct sampling process is accompanied by volume-reduction operations to decrease the  
 116 volume of water; ii) Pumps - where various types of submersible pumps are used to draw a  
 117 larger volume of water; and iii) Nets - which include Manta, Neuston and other surface nets  
 118 that are widely used in water sampling. Some types of nets (e.g., Bongo net) can also acquire  
 119 the samples from lower layers of the water column. A detailed comparison of the sampling  
 120 approaches is shown in Table 1.

121  
 122

Table 1 Comparison of the specific sampling methodologies of MP in river water

Commonly used	Types	Sampling volume	Sampling layer	Water volume calculating method	Advantages	Disadvantages	
Direct sampling	Stainless bucket; Water sampler; Glass jar; Niskin bottles; Other containers	Laboratory filtration; Volume reduced (stainless steel sieves / bolting-silk)	Min (less than 100 liters)	Surface water layer; Surface microlayer; Limited water column	Use containers for volume determination	Cheap, easy to operate and quick; Smaller size and fibrous MPs can be captured; Accurate and controllable volume of filtered water; Boat is not necessary	Small sampling volume; Sample is easily contaminated; Possible to transport bulky samples to the lab; Hard to sample water column in deep water; Need to bring bulky samples to the lab (without volume reduced process)

Pump	Teflon pump; Peristaltic pump; Submersible pump; Plankton pump	Laboratory filtration; Volume reduced (stainless steel sieves / bolting- silk)	Medium (several cubic meter)	Full water layer	Flowmeter; Rated power multiply by running time	Relatively cheap, easy to operate and quick; Relatively smaller size and fibrous MPs can be captured; Accurate and controllable volume of filtered water; Possible to sample the water column	Relatively small sampling volume; Need power supply and boat; Relatively higher cost; Possible to transport bulky samples to the lab; Need to bring bulky samples to the lab (without volume reduced process)
Net	Manta net; Neuston net; Plankton net; Bongo net and other nets	Volume reduced	Max (up to a few hundred cubic meters)	Full water layer	Flowmeter; Towing length multiply by net opening size	Large volume sampling; Net aperture can be changed according to the actual situation; Volume reduced on site; Possible to sample the water column(plankton net)	Net and manual sample transfer may bring into contamination; Risk of clogging; Expensive and not easy operation; Mesh sizes impose a relatively large lower size limit, potentially underestimating MP concentrations

123

124

The different approaches to sampling MPs in river water can be methodologically divided

125

into bulk water sampling and volume-reduced sampling<sup>51</sup>. The primary difference is that the

126

former method collects the entire volume of the sample without performing volume reduction

127

(e.g., use of filters or sieves) in situ. Bulk sampling has advantages including: (i) non-in-situ

128

filtration, which may reduce potential contamination; (ii) samples with low concentrations or

129

small particle sizes can be obtained by collecting large volume samples; and (iii) reducing

130

subjective errors by quantitative sample collection. Direct sampling and pumping can be all

131

categorized together as bulk sampling.

132

Volume-reduced sampling usually employs nets or sieves<sup>52</sup> to reduce the sample volume

133

and concentrate the particulate load for MP analysis. Net sampling represents a commonly

134

used volume-reduced method. Manta and Neuston nets are often used to sample surface waters

135

<sup>53</sup>, whilst stationary conical driftnets and hand nets are also used in some studies<sup>15, 32, 54</sup>. Nets

136

can be deployed at the river surface to capture the floating MP components, as well as in the

137

middle and lowermost sections of the water column with the use of Bongo nets, for example

138

<sup>50</sup>. All the nets should be equipped with flow metres whilst in operation to calculate the volume

139

of water that has been sampled. Nash et al.<sup>55</sup> advised using two flowmeters simultaneously,

140

with one equipped inside the net and another equipped outside the net to better understand

141

how the clogging of the net may affect filtration efficiency. Figure 1 depicts three typical nets

142

used in studies: (a) Manta net, (b) Bongo net and (c) Plankton net.

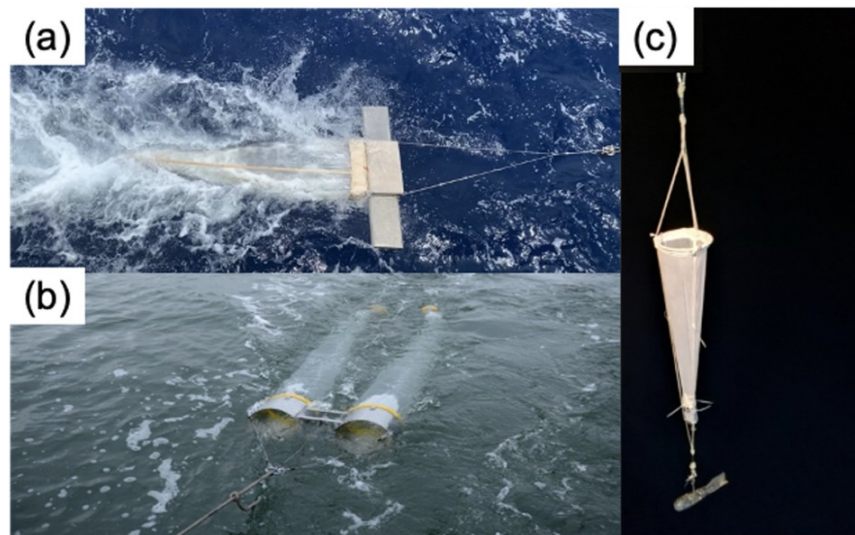


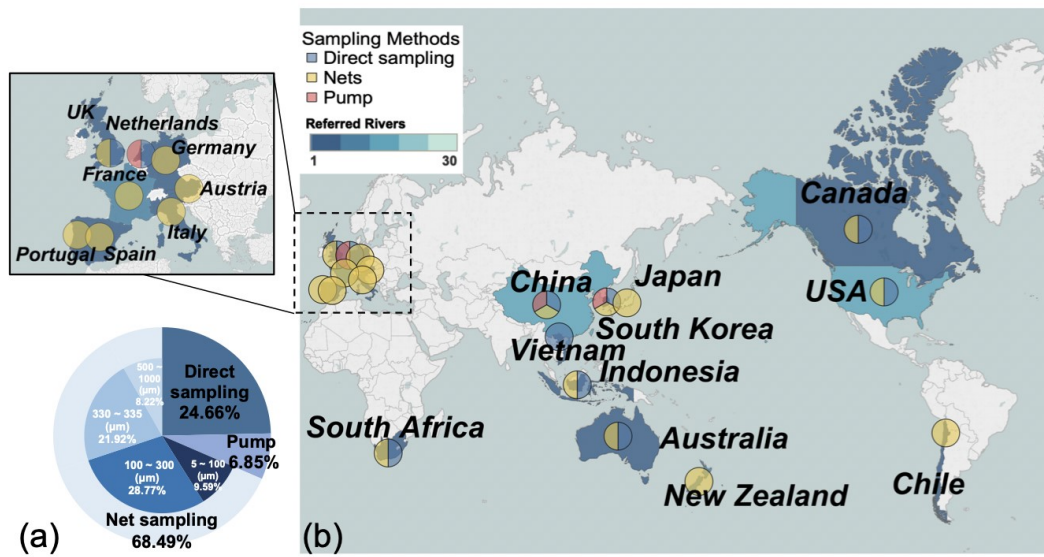
Figure 1 Three normal nets in practical application (a)Manta net,(b)Bongo net,(c)Plankton net.

144  
145  
146  
147

By reviewing the sampling equipment for MPs in river water, it was found that net sampling with mesh sizes between 100-335  $\mu\text{m}$  accounted for more than 50% of studies, which is represented in Figure 2(a). Clogging risks (which may lead to the sample loss or backflow) may occur when the mesh sizes are smaller than the given range, while more particles will escape from the net if the mesh sizes are too large. It also indicates that net sampling is more widely used and therefore offers greater potential for data comparability. Figure 2(b) shows the geographical distribution of river MPs studies, which refers to 5 Asian countries, 8 European countries, 3 countries in the Americas, 1 African country and 2 Oceanian countries) rivers with different flow regimes (monthly discharge ranges between  $4.15 \times 10^5$  to  $7.41 \times 10^{10}$   $\text{m}^3$ ). Kataoka et al.<sup>56</sup> conducted surveys in 29 rivers in Japan, which is the country with the most rivers involved in sampling. The U.S.A has the second largest number of sampled rivers. The third largest is China, which has 13 investigated rivers in this review.

158





160 Figure 2 (a) The proportions of three sampling methods in reviewing studies. (b) Global rivers discussed in this  
 161 review.  
 162  
 163

164 Small samples volumes and use of different (coarser) sieve or mesh sizes can lead to  
 165 inaccurate determination of MPs concentrations<sup>57-60</sup>. Several studies<sup>61, 62</sup> used small volume  
 166 sampling methods (e.g., 1 L surface water in rivers) and the MPs were found to be fibrous or  
 167 mostly fibrous (80%). By comparing the results from a 1 L surface grab sampling method with  
 168 a 335  $\mu\text{m}$  Neuston tow, Barrows et al.<sup>57</sup> found that the concentration of MPs (n/L) of the  
 169 former is 3 orders of magnitude higher than the latter. Furthermore, Norén et al.<sup>63</sup> reported  
 170 that plastic fibers filtered using an 80  $\mu\text{m}$  mesh size net were up to 5 orders of magnitude more  
 171 abundant than those obtained using a 450  $\mu\text{m}$  mesh size net. Dris et al.<sup>58</sup> reported that a  
 172 plankton net with 100  $\mu\text{m}$  mesh size collected 100 times the amount of MPs compared with  
 173 sampling by 330  $\mu\text{m}$  Manta net. Studies using nets for sampling must consider the minimum  
 174 mesh size that can be feasibly used in order to sample small microplastic particles and report  
 175 more accurate MP concentrations, whilst balancing trade offs related to clogging and the flow  
 176 conditions of the river.

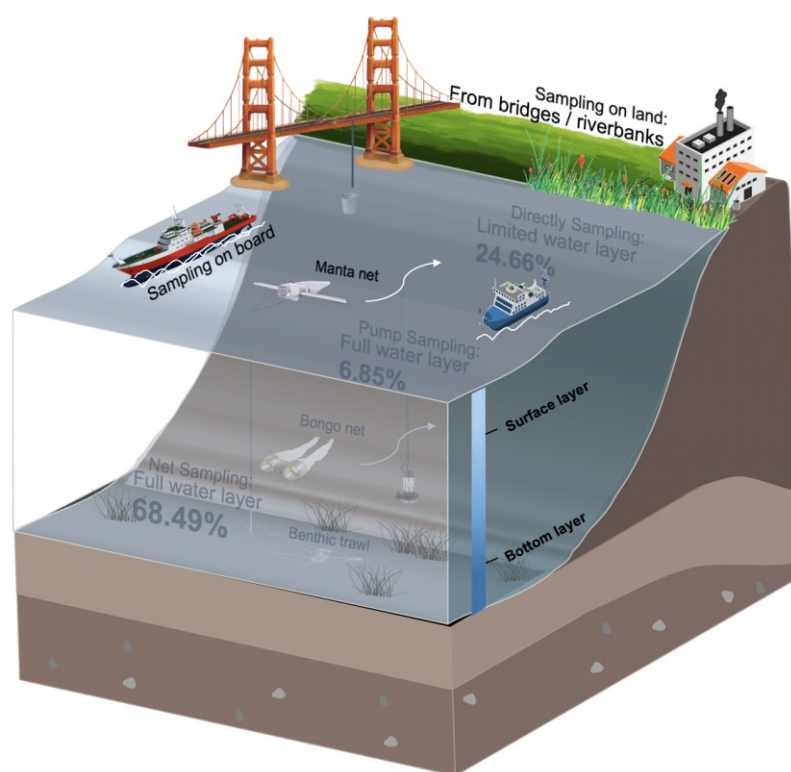
177 MP fibers are typically long and thin and so the portion captured by the net may depend  
 178 on several conditions such as the orientation and curvature of the particle and the flow

179 conditions. This represents an additional important consideration when using nets to sample  
180 river water: fibers observed in net samples may represent an unknown proportion of the total  
181 fiber contamination at that sampling location. A field survey conducted with 100 L water  
182 pumped and a 60  $\mu\text{m}$  sieve, the overall mean MP concentrations in the Yangtze River and East  
183 China Sea were respectively  $157.2 \pm 75.8 \text{ n/m}^3$  and  $112.8 \pm 51.1 \text{ n/m}^3$ , with fibers accounting  
184 for more than 80% of the total <sup>64</sup>. Zhao et al.<sup>65</sup> showed that a 32  $\mu\text{m}$  sieve can retain abundant  
185 fibers by filtering the surface water in the Yangtze River Estuary and East China Sea, which  
186 contained 79.1% and 83.2% fibrous MPs, respectively. Studies using sieves or nets with small  
187 mesh sizes will collect large quantities of fibrous MPs, although data comparability is poor  
188 compared with using Manta trawls with mesh sizes between 100-335  $\mu\text{m}$  as they are less often  
189 used in sampling activities.

### 190 *3.2 Sampling locations*

191 Flux estimates normally utilize measured plastic data and extrapolate to the total river  
192 channel or river catchment based on factors of time or discharge <sup>64,66</sup>. However, several factors  
193 may influence the movement of plastic particles from initial input to eventual release to the  
194 ocean, such as deposition, trapping, and remobilisation. Besides, the spatial context of the  
195 sampling locations also introduces additional uncertainty in the flux estimates. Exploring  
196 sampling locations is conducive to establishing scientific monitoring methods according to the  
197 specific conditions of river. It is also important to consider the deployment location for  
198 different sampling methods, as this may also determine appropriate sampling locations. This  
199 section will review aspects related to sampling locations.

200 Firstly, from the perspective of deployment location, sampling techniques can be  
201 deployed from bridges or from the riverbanks, for example with buckets or nets attached to  
202 retractable rods. Also, sampling activities can be conducted from the research vessels in the  
203 rivers. Figure 3 presents a schematic diagram showing specific sampling measures.



205 Figure 2 Schematic diagram of sampling measures with the proportion of each method in the reviewed papers  
 206  
 207

208 Research vessels may hinder the flow of water and interfere with the stability of the water  
 209 body, which may influence the effectiveness or representativeness of sampling. Sampling from  
 210 bridges or riverbanks is less heavily affected by this factor, although introducing the net into  
 211 the river may still interrupt flows and generate additional water turbulence. The mode of  
 212 deployment will depend on the geographical context of the site. For example, deploying from  
 213 bridges necessitates the occurrence of this form of infrastructure that also offers safe access.  
 214 The use of research vessels requires that rivers are over a certain depth, and that vessels are  
 215 available in the vicinity or can be entered into the water. Sampling from river banks requires  
 216 access and stability, and is only recommend for narrower channels where it is possible to take  
 217 representative samples from within reach of the bank.

218 When selecting riverine MP monitoring sites, the following aspects should be considered:  
 219 representativeness, accessibility, hydrology, and stability (e.g., potential for long-term  
 220 monitoring) <sup>67</sup>. Replicate samples, or simultaneous multiple samples from different points

221 across the river cross-section can help to account for potential variability in MP loads. For  
222 example, small scale spatial variability across a river cross-section or temporal variability  
223 across a short (e.g., 1 hour) sampling period. Wong et al.<sup>68</sup> emphasized the importance of  
224 sampling locations to the MPs flux estimation. Complex hydrodynamic processes in estuaries  
225 may alter the transport and distribution of MPs (e.g., sedimentation, aggregation, resuspension,  
226 hyporheic exchange and biological effects including biofouling, ingestion and excretion<sup>46, 69,</sup>  
227 <sup>70</sup>). Xiong et al.<sup>71</sup> concluded that the plastic debris might accumulate in the river estuary area  
228 due to tidal activities, and emphasized that MPs flux calculations that are only based on the  
229 data obtained from the river estuary will lead to an overestimation. González et al.<sup>67</sup>  
230 recommended to sampling upstream for reference to avoid the influence of estuaries, and the  
231 selection of exact monitoring sites depending on the actual situation on site (e.g., population  
232 density, waste discharge source, possibility of sampling location implementation). To  
233 minimize these effects, multimedial, multi-layer sampling, and long-time scale monitoring  
234 are effective methods. It is recommended to collect samples from multiple reaches of the river  
235 (e.g., covering the upper reaches of the river, river outlets and the place where the river meets  
236 the sea). Cowger et al.<sup>72</sup> also advised for the adoption of depth-integrated sampling.

237 Understanding river characteristics is essential for sampling. Sampling strategies should  
238 be adjusted accordingly, such as with respect to suspended sediment concentration, flow  
239 velocity/discharge, water depth, functional zoning of river basins and tidal influence. Studying  
240 the transport patterns at different time scales can help to better understand the fate of MPs, as  
241 rivers are globally highly diverse and have various features<sup>48</sup>. A sampling site with features  
242 that reduce turbulence complexity (e.g., homogeneous bed characteristics, gentle curve of the  
243 riverbank, stable with little interference) need to be identified and included in monitoring. Site  
244 metadata should also be recorded to allow for effective interpretation of observed MP  
245 concentrations, such as flow velocity, water level, suspended sediment concentrations,

246 meteorological conditions (including antecedent conditions), hydrogeomorphological context  
247 and bed substrate type. To investigate the impact of point and regional pollution sources, for  
248 example, artificial facilities, sewage treatment plants and population density (e.g., tourist areas,  
249 rural and densely populated areas), sampling activities should cover the upper and lower  
250 reaches of the sources<sup>22, 73</sup>.

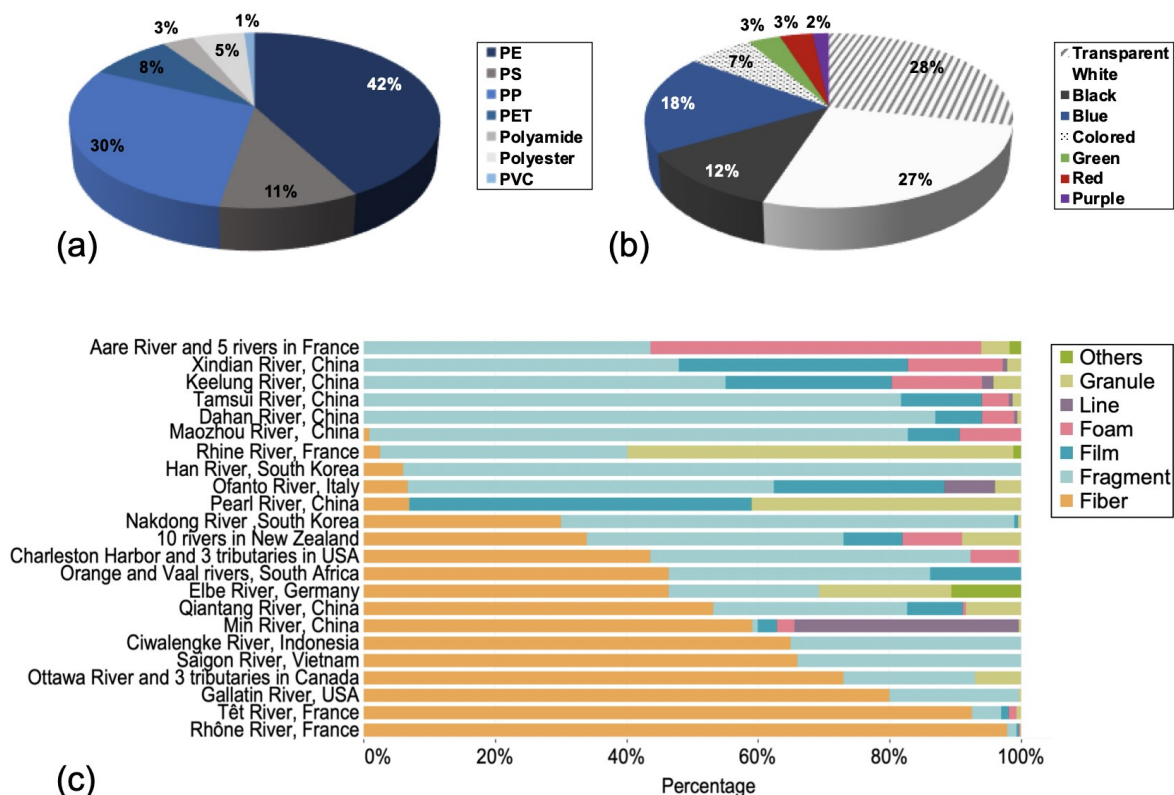
#### 251 4. Riverine MPs distribution, transport and flux

##### 252 4.1 Characteristics of MPs in rivers

253 The occurrence of chemical and colour compositions of MPs in river water according to  
254 reviewed papers were analyzed and sorted, and the proportion of occurrence frequency was  
255 calculated (Figure 4(a), (b)) (See Table S1 Microplastic composition in river water in  
256 Supporting Information). After analyzing, it was found that the most common polymer type  
257 detected in rivers is polyethylene (PE) (42%), followed by polypropylene (PP) (30%) and  
258 polystyrene (PS) (11%) (Figure 4a). Other common polymer types like PET, polyamide and  
259 polyester are also often observed. PE and PP comprised the majority of polymers in the studied  
260 rivers<sup>64, 66, 71, 74-80</sup>, which represent low-density plastic types. Besides, PE and PP are  
261 commonly used in disposable plastic products. While PS foam (normally used in food  
262 packaging and shockproof container) was the most abundant MP category by number in  
263 Hongkong waters<sup>81</sup>. In a study of river in Saigon River, Vietnam, the percentage of polyolefin  
264 and PS accounted for the most while PS foam food box scraps accounted for a very large  
265 proportion<sup>37</sup>.

266 Colourless MPs (including transparent and white were dominant in the articles included  
267 in this review (Figure 4 (b)). This phenomenon has been reported in many previous studies  
268 about MPs in river water<sup>24, 68, 82</sup>. The colourless aspect may be specifically engineered (e.g.,  
269 from white textiles) or caused by fading due to photodegradation. Wong et al.<sup>68</sup> sampled  
270 plastic particles in both river water and sediments from the beach, and found that the

271 colourless plastic particles in river water are less abundant than that in sediment. The higher  
 272 incidence of colourless particles found on river beaches could be interpreted as resulting  
 273 from greater exposure to ultraviolet light in this context, in comparison to particles moving  
 274 in the water.  
 275



276  
 277 Figure 4 (a) chemical, (b) colour and (c) shape compositions of riverine MPs referenced by the articles  
 278 discussed in this review. The ratios in (a) and (b) refer to the occurrence frequencies in all reviewed papers,  
 279 while the ratios in (c) are original data from each independent studies.  
 280

281 MPs from different release pathways partly have their own characteristics, for example,  
 282 fibrous and small size of MPs are typical features of MPs from sewage treatment plants, which  
 283 can be used to partly explain the origin of some of the numerous MPs in the shoreline and river  
 284 shore sediments with size ranges from 63 – 200  $\mu\text{m}$ <sup>83, 84</sup>. MPs derived from tire and road  
 285 marking found in river water can help to reveal contributions from runoff<sup>85, 86</sup>, while the MPs  
 286 formed from the fragmentation of fish lines and floating rafts also reveal the source of directly  
 287 discarded plastic waste<sup>79</sup>. It is helpful for the consideration of influencing factors and

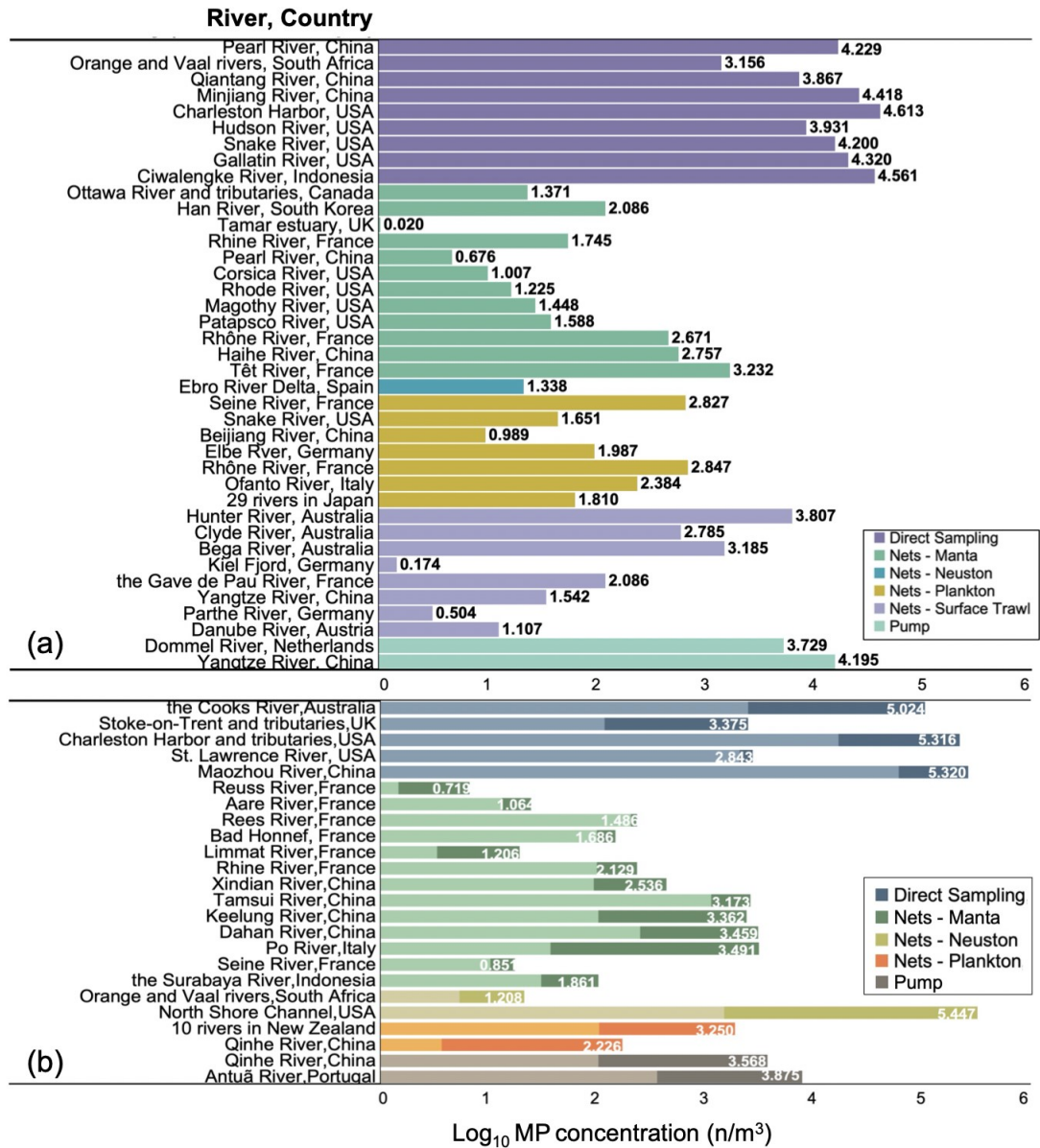
288 development of monitoring strategy of MPs flux in rivers by reviewing the major release  
289 pathways. Considering the shape information is important for the identification of source, fate  
290 and transport for riverine MPs, the original shape compositions data from independent  
291 researches was assessed (Figure 4 (c)). At present, the definition of MP shape has a unification  
292 of broad categories, but there are still differences in the subdivision process. The shapes were  
293 unified here and fall into the following categories: fiber, line, fragment (which includes hard  
294 fragment and film-type fragment <sup>87</sup>), film (which includes flake <sup>88</sup>), foam, granule (which  
295 includes sphere, microbead and pellet <sup>54, 62, 68, 88-90</sup>, opaque and transparent spherules <sup>87, 91</sup>),  
296 others (which includes combined <sup>87</sup>).

297 In general, fibers represent the dominant shape of MPs in rivers <sup>54, 62, 64, 65, 75-77, 79, 80, 92-94</sup>.  
298 Several studies emphasized the use of synthetic textiles as a source of riverine MPs pollution,  
299 also pointing towards sewage treatment plants as a potentially important pathway <sup>77, 95-97</sup>.  
300 Fibers are more easily entrained and maintained in suspension by river flows <sup>43</sup>, potentially  
301 leading to higher proportions observed in the water than in sediments. In some studies,  
302 fragments account for more than 50 % of the MP concentrations <sup>24, 68, 78, 82, 87, 88, 98, 99</sup>, which  
303 may be derived from the fragmentation of plastic products <sup>66, 100-102</sup>. Lahens et al. <sup>93</sup> also  
304 advised to investigate the role of in-situ macroplastic fragmentation as a source of MPs to  
305 rivers. Thompson et al.<sup>103</sup> found 9 types of MPs, including fibers and fragments typically  
306 derived from synthetic fabric, packages and rope in 23 out of 30 sedimentary samples around  
307 Plymouth, UK.

308 MPs concentration in river water is an important data source which can be used to  
309 extrapolate riverine MPs flux. Yet, an important problem arises in data extrapolation, related  
310 to MPs sizes. Several studies have shown that the lower limit of the mesh size is not equal to  
311 the detection limit of MP samples <sup>79, 93, 98, 104</sup>. Recently, Koelmans et al. <sup>105</sup> provide a method  
312 to mitigate size range differences: correction factors can be used to convert MPs sizes into

313 three default size ranges. Figure 5 shows the MPs density in global rivers in this review. To  
314 reduce the variation of different sampling size ranges, the data shown in Figure 5 has been  
315 standardized with the correction factors <sup>105</sup> (See Table S2 Correction factors used for  
316 mitigating size range differences in Supporting Information). MP densities in rivers show great  
317 differences, which may be attributable to the individual or combined effects of the influencing  
318 factors mentioned above, such as sampling methods, river morphology, watershed conditions,  
319 abnormal weather conditions, and so on. It is likely that many rivers exhibit a unique  
320 microplastic assemblage based on the quantity and diversity of sources and the  
321 hydrogeomorphological conditions of the river, which influence MP fate and the  
322 concentrations and particle types observed in a given sample.





324  
 325 Figure 5 MP concentration (n/m<sup>3</sup>) in global rivers (data shown in logarithmic form); (a) numbers represent the  
 326 mean values; (b) numbers represent the quantitative difference between the max and min values, lighter colored  
 327 columns represent the minimum MP concentration and darker colored columns represent the maximum MP  
 328 concentration.  
 329

330 *4.2 Fate and transport processes*

331 Hydrology (e.g., channel morphology, turbulence and tidal influence), spatiotemporal  
 332 variability in sources and environmental processes, artificial factors (e.g., basin population and  
 333 area, watershed function zoning), and the characteristics of MPs (small size thus easily widely  
 334 dispersed with currents and hydrodynamic processes) will influence the transport behaviors of

335 MPs in the freshwater environment <sup>42, 106-109</sup>. This is relevant for sampling MP in rivers and  
336 understanding what monitoring data conveys, which can also influence the accuracy of flux  
337 estimates. Currently, many fate and transport processes governing MP distributions in rivers  
338 are poorly understood and more research is needed to identify and unpick the dominant  
339 mechanisms. Sampled MP from rivers represent a snapshot of concentrations and information  
340 about possible fate and transport processes should be collected to aid in the interpretation of  
341 this data to evaluate what it means in the context of riverine flux.

342 In rivers, particles may partition between the water and sediment phase based upon  
343 numerous factors, such as particle density, flocculation and flow velocity. In the water column,  
344 the movement of particles is not uniform and MPs are not evenly distributed in the water layer.  
345 The density of a type of plastic imparts an important control over the depth of occurrence in  
346 the water <sup>110, 111</sup>, whilst the biodegradability, shape, oxidation resistance, flocculation /  
347 aggregation, surface properties and degree of biofilm formation are also influencing factors  
348 <sup>112</sup>. Low-density MPs mostly float close to the surface of rivers and denser MPs could be  
349 expected to accumulate at the bottom of rivers or buried in the sediments <sup>33, 35, 36, 113</sup>. A study  
350 conducted in the Nakdong River showed the MP concentration in surface river water was 3  
351 times higher than that in bottom water <sup>78</sup>. Yet, changes in MP particle density may occur as a  
352 result of biochemical processes, including surface biofilm generation <sup>114</sup>, ageing and leaching  
353 of additive chemicals <sup>115, 116</sup>, and lead to the change of settling rates <sup>117</sup>. Biofilms can be easily  
354 formed on the surface of plastics in the marine environment and attract adherence by algae  
355 and invertebrates, increasing the sinking speed <sup>118</sup>. MPs may sink to the bottom of the water  
356 layer through biofouling by organisms and accumulate in sediments <sup>118-120</sup>. Scherer et al. <sup>121</sup>  
357 found that the MP abundance in bottom sediment is  $6 \times 10^6$  times higher than that in the  
358 overlying water, and the studied results in the Nakdong River <sup>78</sup> showed that the MP content  
359 in sediment is 2827 times higher than that in the water column. However, several studies

360 identify high density polymer types in water samples and low density – and theoretically  
361 buoyant – polymer types in the sediments<sup>78, 93, 113, 122</sup>. This is partly due to turbulent flow,  
362 which is likely to lead to entrainment and mixing of particles within the water column. An  
363 uneven distribution of MPs in the vertical profile of rivers has been reported<sup>123</sup>, which was  
364 mainly affected by fluvial hydrodynamics. Water turbulence below the surface may mix  
365 particles with a density close to that of the surrounding water, and the density and shape of  
366 small items and particles will also affect their rising or sinking speed<sup>67</sup>. Drummond et al.<sup>46</sup>  
367 found that, for MPs smaller than 100 µm, retention in river sediments can be substantially  
368 increased with the influence of the hyporheic exchange. Besides, hyporheic abrasion may  
369 decrease particle size, thus influencing other variable such as surface area or propensity for  
370 biofilm formation, which could further influence fate and transport processes.

371 The morphological characteristic of a river is a key factor, and the morphology of riverbed  
372 forms may also impact the plastic debris travel distance<sup>124</sup>. For example, a sharp drop in MP  
373 concentrations was found by Mani et al.<sup>91</sup> in the section of the Rhine River with the lowest  
374 bed slope. Concentrations in the river decreased in the water column, which may be attributed  
375 to the lowest bed slope and low flow velocity in the river bed<sup>91</sup>. Also in the Rhine river, Klein  
376 et al.<sup>84</sup> found a dramatical increase in MPs concentration in sediments near to the confluence  
377 of tributaries and the main stream. Similarly, the concentration of MPs in the sediment of the  
378 Elbe River has been shown to decrease in the lower part of the river<sup>121</sup>.

379 Natural meteorological events, such as storms and heavy rainfall, are factors that  
380 influence the instantaneous concentrations and spatial accumulation of MPs. Barnes et al.<sup>11</sup>  
381 found that the wind action of a typhoon and heavy rainfall would increase the speed of MP  
382 migration from land into the aquatic environment. According to a study in the Yangtze River  
383 Estuary, typhoons are an influencing factor for MPs accumulation in the water environment<sup>80</sup>.  
384 Moore et al.,<sup>125</sup> showed that surface plastic debris on the California coast near the Los Angeles

385 stormwater conveying system increased from 10 n/m<sup>3</sup> to 60 n/m<sup>3</sup> after a heavy storm,  
386 indicating that the storm increased the export of MP from the catchment. A study in southern  
387 California coastal water also found that MPs accumulate in coastal areas from less than 1 n/m<sup>3</sup>  
388 to 18 n/m<sup>3</sup> after a typhoon and heavy rain event <sup>50</sup>. And the MP abundance in Venoge river  
389 water in Switzerland increased 150 times after a rainfall event <sup>126</sup>. Flooding can also lead to a  
390 flushing of MPs stored in river sediments and may export MP from catchments or redistribute  
391 particles, for example through overbank deposition <sup>43, 45</sup>. The thresholds for MP deposition,  
392 remobilization and entrainment have not yet been established for a representative range of MP  
393 particle types (sizes, shapes, polymer types), so the hydrological conditions under which  
394 sedimentation or transport of particles occurs is still poorly understood.

395         Some MPs may deposit in riverbank or floodplain sediments, due to overbank deposition  
396 during flood events. These sediments are subject to less erosion than channel bed sediments,  
397 and thus the river corridors can be seen as a possible storage and release component to MPs  
398 movement. Scheurer et al. <sup>127</sup> and Christensen et al. <sup>44</sup> all show MPs can accumulate in  
399 floodplains adjacent to rivers. The timescales over which particles will be remobilized from  
400 these sedimentary archives remains poorly understood, but they could constitute potential  
401 long-term legacy sources of MP contamination to the active channel into the future. The  
402 geomorphological context will present a dominant control in this case, where channel and  
403 floodplain morphology differ significantly, globally. Further research is needed to constrain  
404 this potential source and how it contributes to present and future MP fluxes in a variety of  
405 rivers.

406         Artificial facilities, such as dams, bridges and human-made tributaries, may introduce  
407 additional turbulence which is more likely to entrain MPs. Xiong et al. <sup>71</sup> found that the MP  
408 concentrations downstream of the Three Gorges Reservoir is an order of magnitude lower than  
409 that upstream. Lisa et al. <sup>35</sup> reported more MPs in the reservoir water and sediment than in the

410 upstream water and sediment. According to a study combining manual visual and static trawl  
411 sampling on the Thu Thiem Bridge in the Saigon River, a high concentration area of large  
412 plastic debris was observed near the bridge column, which may be caused by the eddy currents  
413 created by the bridge column that carry the plastic debris <sup>37</sup>. MPs may show a similar response,  
414 which should be further tested to better understand the fate and transport of MPs in the context  
415 of sampling and interpreting monitoring data.

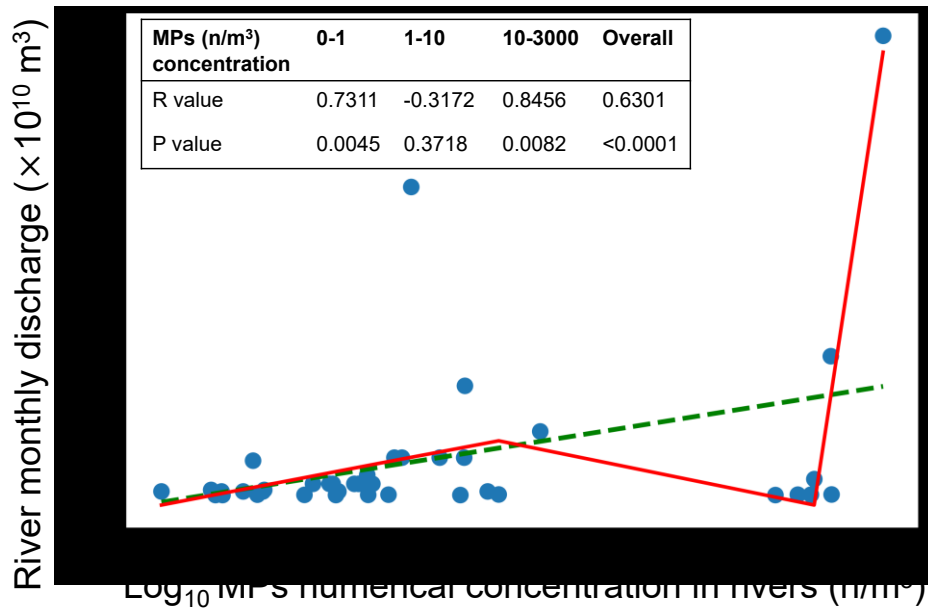
#### 416 *4.3 Spatiotemporal variability and relationship with discharge*

417 MPs are not evenly distributed in different river sections. The complex hydrological  
418 conditions near the estuary will affect the spatial distribution of MPs <sup>71</sup>. For example, MP  
419 abundance in the surface water of Qinhe River increased from upstream to downstream and  
420 reached its highest level in the estuary where MP accumulation zones had formed <sup>79</sup>. Human  
421 activities may lead to spatial variations in MPs. A high density of MPs was detected at  
422 stations near densely populated areas <sup>66, 80, 81, 83, 128, 129</sup>, and the lowest abundance was found  
423 at sites located far from urban centres <sup>77, 91</sup>. In a study conducted in the coastal water of  
424 South Korea <sup>32</sup>, the urban areas had a MP abundance of approximately two times that in rural  
425 areas, and there was a strong correlation between the population of the river and coastal  
426 basins and the mean MP abundances. In contrast, Kapp et al. <sup>130</sup> found high MP  
427 concentrations in a rural site impacted by agriculture where plastic film was widely used.  
428 These spatial patterns in potential MP sources can lead to a heterogeneous distribution of MP  
429 in the catchment.

430 As mentioned above, precipitation and storm events can cause large shifts in MPs  
431 concentration on short time scales. MPs abundance in rivers shows seasonal variation, which  
432 is manifested in the difference of MPs concentration in river water between rainy and dry  
433 season. The dominant shape of MPs also has differences between the dry and rainy seasons.  
434 In one study, fibers were most abundant in the dry season, while fragments were most abundant

435 in the rainy season <sup>81</sup>. There was a significant difference in MPs concentration in Hongkong  
436 waters near the Pearl River Estuary between the rainy (median = 2.657 n/m<sup>3</sup>, 0.227 mg/m<sup>3</sup>)  
437 and dry seasons (median = 0.183 n/m<sup>3</sup>, 0.023 mg/m<sup>3</sup>) <sup>81</sup>. According to Lebreton et al. <sup>18</sup>, 74.5%  
438 of the total plastic load emitted from rivers to the ocean occurs between May and October,  
439 with a peaks in August and minimal release in January. Soeun et al. <sup>78</sup> estimated that 70-80%  
440 of the annual MP load by to the ocean occurred in the wet season. A similar situation also  
441 occurred in the sediment of the Brisbane River; that is, the MP concentration in the wet season  
442 was higher than that in the dry season <sup>131</sup>. These findings may relate to the greater number of  
443 sources that are accessed by precipitation through the increase in connectivity between land  
444 and the river. However, in contrast, Fan et al. <sup>74</sup> has found the MP abundances in the river water  
445 were notably lower during the wet season, which they attributed to the dilution effect of the  
446 precipitation and subsequent increase in discharge. These effect was also reported in the  
447 Gallatin River <sup>62</sup> and the Yangtze Estuary <sup>71</sup>, where the abundance of MPs is inversely  
448 proportional to river discharge. To further discuss the relationship between the MPs  
449 concentration and river discharge, the correlation analysis was carried out with the database  
450 used in this article and collected river monthly discharge data.

451



452  
453  
454  
455  
456  
457  
Figure 6 Correlation between the MPs numerical concentration with river monthly discharge. The red lines represent the fitting curves after subsection regression, respectively for MPs concentration between 0-1, 1-10, 10-3000 n/m<sup>3</sup>; and the green line represents the overall regression analysis of all MPs concentration.

458 The correlation coefficients between MPs numerical concentration and river monthly  
459 discharge is shown in Figure 6. River monthly discharge data were obtained from original  
460 articles or U.S. Geological Survey (*usgs.gov*), and the monthly discharge of sampling time is  
461 selected. The average MP numerical concentration ranges (n/m<sup>3</sup>) were divided into 0-1; 1-10  
462 and 10-3000 (n/m<sup>3</sup>), that can be simply described as low, medium and high MPs numerical  
463 concentrations. It shows that both low and high MPs numerical concentration have a positive  
464 correlation with river monthly discharge (Pearson test, P<0.05), while medium concentration  
465 are not correlated with river discharge. In general, the MPs numerical concentrations are  
466 positively correlated with river discharge (Pearson test, P<0.0001). This finding suggests that  
467 the flux calculation should not simply multiply MP density by river discharge in a single  
468 snapshot in time and instead the measurements need to be integrated over a range of flows,  
469 while considering more practical situations (e.g., the influence of hyporheic exchange,  
470 biological effects, interception of plants). To obtain a more accurate river plastic flux model,  
it is necessary to couple the model with hydrodynamic simulations. When calculating for rivers

471 that lack actual measured data, it is advised to add a relationship coefficient between the river  
472 discharges and MPs concentration in riverine MPs flux equations according to specific river.

#### 473 *4.4 Riverine MPs flux*

474 There is no single method for assessing MP or total plastic flux in rivers, which reflects  
475 the different hydrogeomorphic settings of rivers globally, as well as the different data used to  
476 generate such estimates. As stated above, this relates to the difficulties associated with  
477 sampling rivers, especially those which are large, fast, or have a high suspended load. Different  
478 sampling methods are capable of capturing a different proportion of the total MPs load in a  
479 river, and the way that these data are interpreted will influence the accuracy of calculated flux  
480 estimates. The selection of sampling methods or geographical factors can lead to the  
481 differences in units or data expressions. For example, specific measurement of MPs and larger  
482 plastic flows with nets<sup>66</sup> and visual observation<sup>37</sup> reported either the numerical or mass flux.  
483 Conversely, some waste management infrastructures such as manual waste collection  
484 activities<sup>93</sup> and booms<sup>132</sup> were used in rivers, which are tend to report a total mass of plastics  
485 that is intercepted. Some estimates incorporate multiple measurements which have been  
486 conducted under different hydrological conditions or sampling over a longer time scale to  
487 consider the temporal variability in MPs and plastic flows<sup>66, 78, 133, 134</sup>.

488 For MPs flux calculation, a common method is to build a model and combine it with field  
489 data. Based on this, Moore et al.<sup>135</sup> estimated that two rivers in Los Angeles, the U.S.A, can  
490 transfer 2 billion pieces of MPs into Californian coastal waters in three days. Zhao et al.<sup>64</sup>  
491 adopted the mean MP concentration from field data to calculate the annual plastic flux, and  
492 the Yangtze River was estimated to have transported 16-20 trillion MPs through the top layer  
493 of water (approximately 30 cm depth), a total weight of 537.6-905.9 tonnes, into the East  
494 China Sea annually. The river discharge from the Nakdong River, South Korea, was calculated  
495 by dividing the river into two vertical portions at a downstream site: surface (from the surface



496 to 0.2 m) and subsurface (from 0.2 m to the bottom)<sup>78</sup>. The estimated total annual load in the  
497 Nakdong River reflecting the abundance of MPs in both surface and subsurface waters at the  
498 estuary across four seasons was 5.4 trillion particles, or 53.3 tonnes, in 2017. Mai et al.<sup>66</sup> used  
499 Manta trawls (330  $\mu\text{m}$ ) to sample the MPs in the surface water of the Pearl River Delta and  
500 calculated the riverine MP inputs by multiplying the concentrations of MPs and river discharge.  
501 The annual transport number of MPs in the Pearl River is 390 billion, which weighs 66 tonnes,  
502 and can be converted into an average plastic debris mass of 2900 tonnes year<sup>-1</sup>. By comparing  
503 the MP concentrations in the surface waters of 22 global rivers, the MP concentration in the  
504 Pearl River was at the lower middle level. Max et al.<sup>85</sup> modelled MPs loads in rivers. The  
505 model had three input factors: the density of the population connected to sewage systems, per  
506 capita input of MPs, and sewage treatment efficiency. Approximately 14400 tonnes of MPs  
507 from point sources were calculated to enter the North Sea, Baltic Sea, Black Sea,  
508 Mediterranean Sea and European River basins and then flow into the Atlantic Ocean in 2000.  
509 In addition, these numbers differed by sea. The MP load amount to the Mediterranean Sea was  
510 5600 tonnes, the load to the Black Sea was 4100 tonnes, the load to the European part of the  
511 Atlantic Ocean was 2700 tonnes, the load to the North Sea was 1100 tonnes, and the load to  
512 the Baltic Sea was 900 tonnes.

513 MP fluxes may be inaccurately estimated if the MPs data used are not comprehensive  
514 enough. Zhao et al.<sup>64</sup> has found an overestimation of more than 50% in the Yangtze River  
515 Estuary and East China Sea may occur if only use the data in July. Soeun et al.<sup>78</sup> reported the  
516 influence of small size MPs (< 300  $\mu\text{m}$ ), water layer transportation and seasonal variation to  
517 the estimation of riverine MPs load. Small sampling volume also may lead to the error  
518 estimation of riverine MPs flux which has been demonstrated in the study of Park et al.<sup>99</sup>.

## 519 5. **Perspective and remaining knowledge gaps**

520 The non-uniform sampling locations and methods of riverine MPs may lead to the  
521 underestimation or overestimation of the riverine MPs flux calculation, which has been  
522 demonstrated in 5.4. Considering that the mesh size is negatively correlated with the number  
523 of filtered MPs, a unified approach to minimizing the disparities must be identified. As a  
524 widely used method for sampling MPs in surface water, the net sampling with mesh size varies  
525 from 100-300  $\mu\text{m}$  has great data comparability (Figure 2). However, this only from the data  
526 comparability of the dimension of analysis, the selection of specific sampling methods should  
527 be targeted according to different rivers. To study the variation characteristics of riverine MPs  
528 and plastic debris loads, physical hydrological data along with monitored MP data are essential.  
529 Real-time data during sampling, including river flow, salinity, velocity, turbidity, sediment  
530 concentration and temperature, can be analysed with MP concentrations to research the  
531 correlations among data to gain further understanding.

532 A long term monitoring strategy of riverine MP should consider to establish the MP  
533 particle size distribution curves for the monitored river under different representative flow  
534 regimes. And the monitoring results are recommended to report a power law based distribution  
535 curve since plastic particles tend to break down over time to ever smaller pieces. Kooi et al.<sup>136</sup>  
536 also has suggested a universal equation for this purpose. A scientific monitoring strategy  
537 should also consider the seasonality which needs to set the sampling intervals according to the  
538 river flow regime. As indicated above, the river flow may greatly affect the trend of particle  
539 numbers in the river.

540 One can use the sampling equipment which are already available, however, monitoring  
541 results should be reported with necessary auxiliary information including the flow regime. The  
542 sampling methods described in detail in chapter 4. The monitoring results can then be  
543 comparable by extrapolating the monitoring results based on MP particle size distribution  
544 curve under the specific flow regime. The flow regime is primarily controlled by the climatic

545 conditions and may also be subject to considerable modification by natural impoundments,  
546 dams, or water storage. Flow characteristics may also be changed by water uses, such as  
547 withdrawal for irrigation. The discharge of a river (e.g., in m<sup>3</sup>/s) is the most important  
548 measurement that indicates the river's flow condition. When possible, people should report  
549 the hydrograph based on measurements of daily river discharges for the whole monitoring  
550 period, this is extremely important in determining the flow regimes of the microplastic  
551 sampling dates. A comprehensive monitoring strategy should cover both base flow regimes  
552 and high flow regimes.

553 MP abundance distribution in different particle size ranges can be derived based on the  
554 above steps, the riverine MPs flux can then be estimated by using the mass curves  
555 corresponding to different MP particle sizes <sup>105</sup>, rather than using a single reported value which  
556 may greatly sacrifice the accuracy of estimation.

557 Further research is required to evaluate the effect of sampling methodology on observed  
558 MPs concentrations and compositions, as different morphologies may dominate within  
559 different size classes of MPs <sup>137</sup>, and different sampling methodologies may be more effective  
560 at capturing different particle types <sup>79, 138</sup>. In order to make the monitoring results consistent  
561 and comparable, it is important to establish a monitoring strategy for riverine MPs flux  
562 considering spatial and temporal variations, one has also to acknowledge that the selection of  
563 sampling equipment in different locations depend on the availability and tradition, it is  
564 therefore not realistic to require people to use samplers of the same size. The following  
565 considerations are therefore recommended:

566 1) Determine how the estuarine processes affects the riverine MPs flux estimation, for  
567 example, the riverine MP flux in downstream sections may be unidirectional flows, but the  
568 influence mechanism of tidal current action is not clear enough;

- 569 2) The sampling methods have differences, it is important to calculate the proportion of the  
570 total MP load captured by each method, and how representative are different sampling  
571 campaigns in terms of the full flux across a given cross-section;
- 572 3) Normalize the riverine MP flux measurement in different section of the same catchment  
573 or different catchment for data comparability. Usually, MP sampler cannot cover the whole  
574 cross-section of a river, due to the velocity difference, the microplastic flux measured at  
575 different location along the cross section will be different. It is therefore suggested to  
576 develop a microplastic cross-sectional profile for the monitored location. For example, the  
577 profile can be developed by measurements at each quartile along the vertical and horizontal  
578 directions of the cross-section of a river.

#### 579 **Corresponding Author**

580 Daoji Li – State Key Laboratory of Estuarine and Coastal Research, East China Normal  
581 University, 500 Dongchuan Road, Shanghai 200241, China; orcid.org/0000-0002-3447-3485;  
582 E-mail: daojili@sklec.ecnu.edu.cn

#### 583 **Declaration of Competing Interest**

584 The authors declare that they have no known competing financial interests or personal  
585 relationships that could have appeared to influence the work reported in this paper.

#### 586 **Acknowledgments**

587 This study was funded by the National Key Research and Development Program of China  
588 (2016YFC1402205), National Natural Science Fund of China (41676190) and Sino-  
589 Norwegian cooperation project on capacity building for reducing plastic and microplastic  
590 pollution (SINOPLAST). We extend our thanks to the anonymous reviewers and dedicated  
591 editors for valuable suggestions to improve the quality of this manuscript.

#### 592 **Supporting Information Available**

593 This information is available free of charge via the Internet at <http://pubs.acs.org>.

594

#### 595 **References:**

- 596 1. Kershaw, P.; Rochman, C. *Sources, fate and effects of microplastics in the marine environment:*  
597 *part 2 of a global assessment*; 2015; pp [http://safety4sea.com/wp-content/uploads/2017/01/GESAMP-](http://safety4sea.com/wp-content/uploads/2017/01/GESAMP-Report-Microplastics-in-the-Marine-Environment-2017_01.pdf)  
598 [Report-Microplastics-in-the-Marine-Environment-2017\\_01.pdf](http://safety4sea.com/wp-content/uploads/2017/01/GESAMP-Report-Microplastics-in-the-Marine-Environment-2017_01.pdf).
- 599 2. Thompson, R. C., *Microplastics in the Marine Environment: Sources, Consequences and Solutions.*  
600 *Marine Anthropogenic Litter* **2015**, 185-200.
- 601 3. de Sá, L. C.; Luís, L. G.; Guilhermino, L., Effects of microplastics on juveniles of the common  
602 goby (*Pomatoschistus microps*): Confusion with prey, reduction of the predatory performance and  
603 efficiency, and possible influence of developmental conditions. *Environmental Pollution* **2015**, *196*,  
604 359-362.
- 605 4. Besseling, E.; Foekema, E. M.; Van Franeker, J. A.; Leopold, M. F.; Kühn, S.; Bravo Rebolledo,  
606 E. L.; Heße, E.; Mielke, L.; Ijzer, J.; Kamminga, P.; Koelmans, A. A., Microplastic in a macro filter  
607 feeder: Humpback whale *Megaptera novaeangliae*. *Marine Pollution Bulletin* **2015**, *95*, (1), 248-252.
- 608 5. Farrell, P.; Nelson, K., Trophic level transfer of microplastic: *Mytilus edulis* (L.) to *Carcinus*  
609 *maenas* (L.). *Environmental Pollution* **2013**, *177*, 1-3.

- 610 6. Setälä, O.; Fleming-Lehtinen, V.; Lehtiniemi, M., Ingestion and transfer of microplastics in the  
611 planktonic food web. *Environmental Pollution* **2014**, *185*, 77-83.
- 612 7. Sul, J.; Spengler, A.; Costa, M., Here, there and everywhere. Small plastic fragments and pellets  
613 on beaches of Fernando de Noronha (Equatorial Western Atlantic). *Marine pollution bulletin* **2009**, *58*,  
614 1236-8.
- 615 8. Barnes, D. K. A.; Walters, A.; Gon Alves, L., Macroplastics at sea around Antarctica. *Marine*  
616 *Environmental Research* **2010**, *70*, (2), 250-252.
- 617 9. Obbard, R. W.; Sadri, S.; Wong, Y. Q.; Khitun, A. A.; Baker, I.; Thompson, R. C., Global warming  
618 releases microplastic legacy frozen in Arctic Sea ice. *Earth's Future* **2014**, *2*, (6), 315-320.
- 619 10. Free, C. M.; Jensen, O. P.; Mason, S. A.; Eriksen, M.; Williamson, N. J.; Boldgiv, B., High-levels  
620 of microplastic pollution in a large, remote, mountain lake. *Marine Pollution Bulletin* **2014**, *85*, (1),  
621 156-163.
- 622 11. Barnes, D.; Galgani, F.; Thompson, R.; Barlaz, M., Accumulation and fragmentation of plastic  
623 debris in global environments. *Philosophical transactions of the Royal Society of London. Series B,*  
624 *Biological sciences* **2009**, *364*, 1985-98.
- 625 12. Ng, K. L.; Obbard, J. P., Prevalence of microplastics in Singapore's coastal marine environment.  
626 *Marine Pollution Bulletin* **2006**, *52*, (7), 761-767.
- 627 13. Martinez, E.; Maamaatuaiahutapu, K.; Taillandier, V., Floating marine debris surface drift:  
628 Convergence and accumulation toward the South Pacific subtropical gyre. *Marine Pollution Bulletin*  
629 **2009**, *58*, (9), 1347-1355.
- 630 14. Meijer, L.; van Emmerik, T.; van der Ent, R.; Schmidt, C.; Lebreton, L., Over 1000 rivers  
631 accountable for 80% of global riverine plastic emissions into the ocean. *Science Advances* **2021**, *7*,  
632 (18), eaaz5803.
- 633 15. Lechner, A.; Keckeis, H.; Lumesberger-Loisl, F.; Zens, B.; Krusch, R.; Tritthart, M.; Glas, M.;  
634 Schludermann, E., The Danube so colourful: A potpourri of plastic litter outnumbers fish larvae in  
635 Europe's second largest river. *Environmental Pollution* **2014**, *188*, 177-181.
- 636 16. Barletta, M.; Lima, A. R. A.; Costa, M. F., Distribution, sources and consequences of nutrients,  
637 persistent organic pollutants, metals and microplastics in South American estuaries. *Science of The*  
638 *Total Environment* **2019**, *651*, 1199-1218.
- 639 17. Fok, L.; Cheung, P. K., Hong Kong at the Pearl River Estuary: A hotspot of microplastic pollution.  
640 *Marine Pollution Bulletin* **2015**, *99*, (1-2), 112-118.
- 641 18. Lebreton, L. C. M.; van der Zwet, J.; Damsteeg, J.-W.; Slat, B.; Andrady, A.; Reisser, J., River  
642 plastic emissions to the world's oceans. *Nature Communications* **2017**, *8*, (1), 15611.
- 643 19. SC, J. S.; Silva, J. D. B.; França, E. J. d.; Araújo, M. C. B. d.; Gusmão, F., Microplastics ingestion  
644 by a common tropical freshwater fishing resource. *Environmental Pollution* **2017**, *221*, 218-226.
- 645 20. Pazos, R. S.; Bauer, D. E.; Gómez, N., Microplastics integrating the coastal planktonic community  
646 in the inner zone of the Río de la Plata estuary (South America). *Environmental Pollution* **2018**, *243*,  
647 134-142.
- 648 21. Hurley, R.; Horton, A.; Lusher, A.; Nizzetto, L., Plastic waste in the terrestrial environment. In  
649 *Plastic Waste and Recycling*, Elsevier: 2020; pp 163-193.
- 650 22. McCormick, A. R.; Hoellein, T. J.; London, M. G.; Hittie, J.; Scott, J. W.; Kelly, J. J., Microplastic  
651 in surface waters of urban rivers: concentration, sources, and associated bacterial assemblages.  
652 *Ecosphere (Washington, D.C)* **2016**, *7*, (11), e01556.
- 653 23. Castaneda, R.; Avlijas, S.; Simard, A.; Ricciardi, A., Microplastic pollution in St. Lawrence River  
654 sediments. *Canadian Journal of Fisheries and Aquatic Sciences* **2014**.
- 655 24. Wu, P.; Tang, Y.; Dang, M.; Wang, S.; Jin, H.; Liu, Y.; Jing, H.; Zheng, C.; Yi, S.; Cai, Z., Spatial-  
656 temporal distribution of microplastics in surface water and sediments of Maozhou River within  
657 Guangdong-Hong Kong-Macao Greater Bay Area. *Science of The Total Environment* **2020**, *717*,  
658 135187.
- 659 25. Rodrigues, M. O.; Abrantes, N.; Gonçalves, F. J. M.; Nogueira, H.; Marques, J. C.; Gonçalves, A.  
660 M. M., Spatial and temporal distribution of microplastics in water and sediments of a freshwater system  
661 (Antuã River, Portugal). *Science of The Total Environment* **2018**, *633*, 1549-1559.
- 662 26. McCormick, A.; Hoellein, T. J.; Mason, S. A.; Schluep, J.; Kelly, J. J., Microplastic is an abundant  
663 and distinct microbial habitat in an urban river. *Environ Sci Technol* **2014**, *48*, (20), 11863-71.

664 27. Mintenig, S. M.; Kooi, M.; Erich, M. W.; Primpke, S.; Redondo- Hasselerharm, P. E.; Dekker, S.  
665 C.; Koelmans, A. A.; van Wezel, A. P., A systems approach to understand microplastic occurrence and  
666 variability in Dutch riverine surface waters. *Water Research* **2020**, *176*, 115723.  
667 28. Woodward, J.; Li, J.; Rothwell, J.; Hurley, R., Acute riverine microplastic contamination due to  
668 avoidable releases of untreated wastewater. *Nature Sustainability* **2021**.  
669 29. Hermsen, E.; Mintenig, S. M.; Besseling, E.; Koelmans, A. A., Quality criteria for the analysis of  
670 microplastic in biota samples: a critical review. *Environmental Science & technology* **2018**, *52*, (18),  
671 10230-10240.  
672 30. Cowger, W.; Booth, A. M.; Hamilton, B. M.; Thaysen, C.; Primpke, S.; Munno, K.; Lusher, A. L.;  
673 Dehaut, A.; Vaz, V. P.; Liboiron, M., Reporting guidelines to increase the reproducibility and  
674 comparability of research on microplastics. *Applied spectroscopy* **2020**, *74*, (9), 1066-1077.  
675 31. Zheng, Y.; Li, J.; Cao, W.; Jiang, F.; Zhao, C.; Ding, H.; Wang, M.; Gao, F.; Sun, C., Vertical  
676 distribution of microplastics in bay sediment reflecting effects of sedimentation dynamics and  
677 anthropogenic activities. *Marine Pollution Bulletin* **2020**, *152*, 110885.  
678 32. Song, Y. K.; Hong, S. H.; Eo, S.; Jang, M.; Han, G. M.; Isobe, A.; Shim, W. J., Horizontal and  
679 Vertical Distribution of Microplastics in Korean Coastal Waters. *Environmental Science & Technology*  
680 **2018**, *52*, (21), 12188-12197.  
681 33. Lenaker, P.; Baldwin, A.; Corsi, S.; Mason, S.; Reneau, P.; Scott, J., Vertical Distribution of  
682 Microplastics in the Water Column and Surficial Sediment from the Milwaukee River Basin to Lake  
683 Michigan. *Environmental Science & Technology* **2019**, *53*, (21), 12227-12237.  
684 34. Weideman, E. A.; Perold, V.; Ryan, P. G., Little evidence that dams in the Orange–Vaal River  
685 system trap floating microplastics or microfibrils. *Marine Pollution Bulletin* **2019**, *149*, 110664.  
686 35. Lisa, W.; Susan, M.; Patrick, J., The effect of dams on river transport of microplastic pollution.  
687 *Science of the Total Environment* **2019**, *664*, 834-840.  
688 36. Zhang, K.; Gong, W.; Lv, J.; Xiong, X.; Wu, C., Accumulation of floating microplastics behind  
689 the Three Gorges Dam. *Environmental Pollution* **2015**, *204*, 117-123.  
690 37. van Emmerik, T.; Kieu-Le, T.-C.; Loozen, M.; van Oeveren, K.; Strady, E.; Bui, X.-T.; Egger, M.;  
691 Gasperi, J.; Lebreton, L.; Nguyen, P.-D.; Schwarz, A.; Slat, B.; Tassin, B., A Methodology to  
692 Characterize Riverine Macroplastic Emission Into the Ocean. *Frontiers in Marine Science* **2018**, *5*.  
693 38. Baldwin, A. K.; Corsi, S. R.; Mason, S. A., Plastic Debris in 29 Great Lakes Tributaries: Relations  
694 to Watershed Attributes and Hydrology. *Environmental Science & Technology* **2016**, *50*, (19), 10377-  
695 10385.  
696 39. Song, J.; Hou, C.; Zhou, Y.; Liu, Q.; Wu, X.; Wang, Y.; Yi, Y., The flowing of microplastics was  
697 accelerated under the influence of artificial flood generated by hydropower station. *Journal of Cleaner*  
698 *Production* **2020**, *255*, 120174.  
699 40. Balas, C.; Williams, A.; Simmons, S.; Ergin, A., A statistical riverine litter propagation model.  
700 *Marine Pollution Bulletin* **2001**, *42*, (11), 1169-1176.  
701 41. Williams, A.; Simmons, S., Movement patterns of riverine litter. *Water, Air, and Soil Pollution*  
702 **1997**, *98*, (1), 119-139.  
703 42. Campanale, C.; Savino, I.; Pojar, I.; Massarelli, C.; Uricchio, V. F., A Practical Overview of  
704 Methodologies for Sampling and Analysis of Microplastics in Riverine Environments. *Sustainability*  
705 **2020**, *12*, (17), 6755.  
706 43. Hurley, R.; Woodward, J.; Rothwell, J. J., Microplastic contamination of river beds significantly  
707 reduced by catchment-wide flooding. *Nature geoscience* **2018**, *11*, (4), 251-257.  
708 44. Christensen, N. D.; Wisinger, C. E.; Maynard, L. A.; Chauhan, N.; Schubert, J. T.; Czuba, J. A.;  
709 Barone, J. R., Transport and characterization of microplastics in inland waterways. *Journal of Water*  
710 *Process Engineering* **2020**, *38*, 101640.  
711 45. Ockelford, A.; Cundy, A.; Ebdon, J. E., Storm response of fluvial sedimentary microplastics.  
712 *Scientific reports* **2020**, *10*, (1), 1-10.  
713 46. Drummond, J. D.; Nel, H. A.; Packman, A. I.; Krause, S., Significance of Hyporheic Exchange  
714 for Predicting Microplastic Fate in Rivers. *Environmental Science & Technology Letters* **2020**, *7*, (10),  
715 727-732.

- 716 47. Taryono; Pe, E.; Wardiatno, Y.; Mashar, A. In *Macroplastic distribution, abundance, and*  
717 *composition which flows to Cimandiri estuary, West Java*, IOP Conference Series: Earth and  
718 Environmental Science, 2020; IOP Publishing: 2020; p 012031.
- 719 48. Skalska, K.; Ockelford, A.; Ebdon, J. E.; Cundy, A. B., Riverine microplastics: Behaviour, spatio-  
720 temporal variability, and recommendations for standardised sampling and monitoring. *Journal of*  
721 *Water Process Engineering* **2020**, *38*, 101600.
- 722 49. Dris, R.; Gasperi, J.; Rocher, V.; Tassin, B., Synthetic and non-synthetic anthropogenic fibers in  
723 a river under the impact of Paris Megacity: Sampling methodological aspects and flux estimations.  
724 *Science of The Total Environment* **2018**, *618*, 157-164.
- 725 50. Lattin, G. L.; Moore, C. J.; Zellers, A. F.; Moore, S. L.; Weisberg, S. B., A comparison of  
726 neustonic plastic and zooplankton at different depths near the southern California shore. *Marine*  
727 *Pollution Bulletin* **2004**, *49*, (4), 291-294.
- 728 51. Hidalgo-Ruz, V.; Gutow, L.; Thompson, R. C.; Thiel, M., Microplastics in the Marine  
729 Environment: A Review of the Methods Used for Identification and Quantification. *Environmental*  
730 *Science & Technology* **2012**, *46*, (6), 3060-3075.
- 731 52. Meng, Y.; Kelly, F. J.; Wright, S. L., Advances and challenges of microplastic pollution in  
732 freshwater ecosystems: A UK perspective. *Environmental Pollution* **2020**, *256*, 113445.
- 733 53. Faure, F.; Saini, C.; Potter, G.; Galgani, F.; de Alencastro, L. F.; Hagmann, P., An evaluation of  
734 surface micro- and mesoplastic pollution in pelagic ecosystems of the Western Mediterranean Sea.  
735 *Environmental Science and Pollution Research* **2015**, *22*, (16), 12190-12197.
- 736 54. Mel, C.; Wolfgang, L.; Philippe, K., Microplastic fluxes in a large and a small Mediterranean river  
737 catchments: The Têt and the Rhône, Northwestern Mediterranean Sea. *Science of the Total*  
738 *Environment* **2020**, *716*, 136984.
- 739 55. Nash, R. D. M.; Dickey-Collas, M.; Milligan, S. P., Descriptions of the Gulf VH/PRO-NET and  
740 MAFF/Guidline unencased high-speed plankton samplers. *Journal of Plankton Research* **1998**, *20*,  
741 (10), 1915-1926.
- 742 56. Kataoka, T.; Nihei, Y.; Kudou, K.; Hinata, H., Assessment of the sources and inflow processes of  
743 microplastics in the river environments of Japan. *Environmental Pollution* **2019**, *244*, 958-965.
- 744 57. Barrows, A. P. W.; Neumann, C. A.; Berger, M. L.; Shaw, S. D., Grab vs. neuston tow net: a  
745 microplastic sampling performance comparison and possible advances in the field. *Analytical Methods*  
746 **2017**, *9*, (9), 1446-1453.
- 747 58. Dris, R.; Gasperi, J.; Rocher, V.; Saad, M.; Renault, N.; Tassin, B., Microplastic contamination in  
748 an urban area: a case study in Greater Paris. *Environmental Chemistry* **2015**, *12*, (5), 592.
- 749 59. Koelmans, A. A.; Mohamed Nor, N. H.; Hermsen, E.; Kooi, M.; Mintenig, S. M.; De France, J.,  
750 Microplastics in freshwaters and drinking water: Critical review and assessment of data quality. *Water*  
751 *Research* **2019**, *155*, 410-422.
- 752 60. Karlsson, T. M.; Kärrman, A.; Rotander, A.; Hassellöv, M., Comparison between manta trawl and  
753 in situ pump filtration methods, and guidance for visual identification of microplastics in surface waters.  
754 *Environmental Science and Pollution Research* **2020**, *27*, (5), 5559-5571.
- 755 61. Miller, R. Z.; Watts, A. J. R.; Winslow, B. O.; Galloway, T. S.; Barrows, A. P. W., Mountains to  
756 the sea: River study of plastic and non-plastic microfiber pollution in the northeast USA. *Marine*  
757 *Pollution Bulletin* **2017**, *124*, (1), 245-251.
- 758 62. Barrows, A. P. W.; Christiansen, K. S.; Bode, E. T.; Hoellein, T. J., A watershed-scale, citizen  
759 science approach to quantifying microplastic concentration in a mixed land-use river. *Water Research*  
760 **2018**, *147*, 382-392.
- 761 63. Norén, F., Small plastic particles in Coastal Swedish waters. *KIMO Report* **2007**, 1-11.
- 762 64. Zhao, S.; Wang, T.; Zhu, L.; Xu, P.; Wang, X.; Gao, L.; Li, D., Analysis of suspended  
763 microplastics in the Changjiang Estuary: Implications for riverine plastic load to the ocean. *Water*  
764 *Research* **2019**, *161*, 560-569.
- 765 65. Zhao, S.; Zhu, L.; Wang, T.; Li, D., Suspended microplastics in the surface water of the Yangtze  
766 Estuary System, China: First observations on occurrence, distribution. *Marine Pollution Bulletin* **2014**,  
767 *86*, (1-2), 562-568.

- 768 66. Mai, L.; You, S.-N.; He, H.; Bao, L.-J.; Liu, L.-Y.; Zeng, E. Y., Riverine Microplastic Pollution  
769 in the Pearl River Delta, China: Are Modeled Estimates Accurate? *Environmental Science &*  
770 *Technology* **2019**, *53*, (20), 11810-11817.
- 771 67. González, D.; Hanke, G.; Tweehuysen, G.; Bellert, B.; Holzhauser, M.; Palatinus, A.; Hohenblum,  
772 P.; Oosterbaan, L. *Riverine litter monitoring-options and recommendations. MSFD GES TG marine*  
773 *litter thematic report*; JRC technical report: 2016.
- 774 68. Wong, G.; Löwemark, L.; Kunz, A., Microplastic pollution of the Tamsui River and its tributaries  
775 in northern Taiwan: Spatial heterogeneity and correlation with precipitation. *Environmental Pollution*  
776 **2020**, *260*, 113935.
- 777 69. Christoph, D.; Rummel, Annika; Jahnke, Elena; Gorokhova; Dana; Kühnel; Mechthild, Impacts  
778 of Biofilm Formation on the Fate and Potential Effects of Microplastic in the Aquatic Environment.  
779 *Environ.sci.technol.lett* **2017**.
- 780 70. Chubarenko, I.; Bagaev, A.; Zobkov, M.; Esiukova, E., On some physical and dynamical  
781 properties of microplastic particles in marine environment. *Marine Pollution Bulletin* **2016**, *108*, (1-2),  
782 105-112.
- 783 71. Xiong, X.; Wu, C.; James, J. E., Occurrence and fate of microplastic debris in middle and lower  
784 reaches of the Yangtze River – From inland to the sea. *Science of the Total Environment* **2019**, *659*,  
785 66-73.
- 786 72. Cowger, W.; Gray, A. B.; Guilinger, J. J.; Fong, B.; Waldschläger, K., Concentration Depth  
787 Profiles of Microplastic Particles in River Flow and Implications for Surface Sampling. *Environmental*  
788 *Science & Technology* **2021**, *55*, (9), 6032-6041.
- 789 73. Wagner, S.; Klöckner, P.; Stier, B.; Römer, M.; Seiwert, B.; Reemtsma, T.; Schmidt, C.,  
790 Relationship between Discharge and River Plastic Concentrations in a Rural and an Urban Catchment.  
791 *Environmental Science & Technology* **2019**, *53*, (17), 10082-10091.
- 792 74. Fan, Y.; Zheng, K.; Zhu, Z.; Chen, G.; Peng, X., Distribution, sedimentary record, and persistence  
793 of microplastics in the Pearl River catchment, China. *Environmental Pollution* **2019**, *251*, 862-870.
- 794 75. Jiang, Y.; Zhao, Y.; Wang, X.; Yang, F.; Chen, M.; Wang, J., Characterization of microplastics in  
795 the surface seawater of the South Yellow Sea as affected by season. *Science of The Total Environment*  
796 **2020**, *724*, 138375.
- 797 76. Li, L.; Geng, S.; Wu, C.; Song, K.; Sun, F.; Visvanathan, C.; Xie, F.; Wang, Q., Microplastics  
798 contamination in different trophic state lakes along the middle and lower reaches of Yangtze River  
799 Basin. *Environmental Pollution* **2019**, *254*, 112951.
- 800 77. Lin, L.; Zuo, L.; Peng, J.; Cai, L.; Fok, L.; Yan, Y.; Li, H.; Xu, X., Occurrence and distribution of  
801 microplastics in an urban river: A case study in the Pearl River along Guangzhou City, China. *Science*  
802 *of The Total Environment* **2018**, *644*, 375-381.
- 803 78. Soeun, E.; Sang, H.; Hong, Y., Spatiotemporal distribution and annual load of microplastics in the  
804 Nakdong River, South Korea. *Water Research* **2019**, *160*, 228-237.
- 805 79. Zhang, L.; Liu, J.; Xie, Y.; Zhong, S.; Yang, B.; Lu, D.; Zhong, Q., Distribution of microplastics  
806 in surface water and sediments of Qin river in Beibu Gulf, China. *Science of The Total Environment*  
807 **2020**, *708*, 135176.
- 808 80. Zhao, S.; Zhu, L.; Li, D., Microplastic in three urban estuaries, China. *Environmental Pollution*  
809 **2015**, *206*, 597-604.
- 810 81. Cheung, P. K.; Fok, L.; Hung, P. L.; Cheung, L. T. O., Spatio-temporal comparison of neustonic  
811 microplastic density in Hong Kong waters under the influence of the Pearl River Estuary. *Science of*  
812 *The Total Environment* **2018**, *628-629*, 731-739.
- 813 82. Yan, M.; Nie, H.; Xu, K.; He, Y.; Hu, Y.; Huang, Y.; Wang, J., Microplastic abundance,  
814 distribution and composition in the Pearl River along Guangzhou city and Pearl River estuary, China.  
815 *Chemosphere* **2019**, *217*, 879-886.
- 816 83. Browne, M. A.; Crump, P.; Niven, S. J.; Teuten, E.; Tonkin, A.; Galloway, T.; Thompson, R.,  
817 Accumulation of Microplastic on Shorelines Worldwide: Sources and Sinks. *Environmental Science &*  
818 *Technology* **2011**, *45*, (21), 9175-9179.
- 819 84. Klein, S.; Worch, E.; Knepper, T. P., Occurrence and Spatial Distribution of Microplastics in River  
820 Shore Sediments of the Rhine-Main Area in Germany. *Environmental Science & Technology* **2015**, *49*,  
821 (10), 6070-6076.



- 822 85. Max, S.; Albert, A.; Koelmans, Export of microplastics from land to sea. A modelling approach.  
823 *Water Research* **2017**, *127*, 249-257.
- 824 86. Horton, A.; Svendsen, C.; Williams, R., Large microplastic particles in sediments of tributaries of  
825 the River Thames, UK – Abundance, sources and methods for effective quantification. *Marine*  
826 *pollution bulletin* **2017**, *114*, (1), 218-226.
- 827 87. Mani, T.; Burkhardt-Holm, P., Seasonal microplastics variation in nival and pluvial stretches of  
828 the Rhine River – From the Swiss catchment towards the North Sea. *Science of The Total Environment*  
829 **2020**, *707*, 135579.
- 830 88. Campanale, C.; Stock, F.; Massarelli, C.; Kochleus, C.; Bagnuolo, G.; Reifferscheid, G.; Uricchio,  
831 V. F., Microplastics and their possible sources: The example of Ofanto river in southeast Italy.  
832 *Environmental Pollution* **2020**, *258*, 113284.
- 833 89. Dikareva, N.; Simon, K. S., Microplastic pollution in streams spanning an urbanisation gradient.  
834 *Environmental Pollution* **2019**, *250*, 292-299.
- 835 90. Vermaire, J. C.; Pomeroy, C.; Herczegh, S. M.; Haggart, O.; Murphy, M., Microplastic abundance  
836 and distribution in the open water and sediment of the Ottawa River, Canada, and its tributaries.  
837 *FACETS* **2017**, *2*, (1), 301-314.
- 838 91. Mani, T.; Hauk, A.; Walter, U.; Burkhardt-Holm, P., Microplastics profile along the Rhine River.  
839 *Scientific Reports* **2015**, *5*, (1), 1-7.
- 840 92. Huang, Y.; Tian, M.; Jin, F.; Chen, M.; Liu, Z.; He, S.; Li, F.; Yang, L.; Fang, C.; Mu, J., Coupled  
841 effects of urbanization level and dam on microplastics in surface waters in a coastal watershed of  
842 Southeast China. *Marine Pollution Bulletin* **2020**, *154*, 111089.
- 843 93. Lahens, L.; Strady, E.; Kieu-Le, T.-C.; Dris, R.; Boukerma, K.; Rinnert, E.; Gasperi, J.; Tassin,  
844 B., Macroplastic and microplastic contamination assessment of a tropical river (Saigon River, Vietnam)  
845 transversed by a developing megacity. *Environmental Pollution* **2018**, *236*, 661-671.
- 846 94. Alam, F. C.; Sembiring, E.; Muntalif, B. S.; Suendo, V., Microplastic distribution in surface water  
847 and sediment river around slum and industrial area (case study: Ciwalengke River, Majalaya district,  
848 Indonesia). *Chemosphere* **2019**, *224*, 637-645.
- 849 95. Ding, L.; Mao, R. F.; Guo, X.; Yang, X.; Zhang, Q.; Yang, C., Microplastics in surface waters and  
850 sediments of the Wei River, in the northwest of China. *Science of The Total Environment* **2019**, *667*,  
851 427-434.
- 852 96. Han, M.; Niu, X.; Tang, M.; Zhang, B.-T.; Wang, G.; Yue, W.; Kong, X.; Zhu, J., Distribution of  
853 microplastics in surface water of the lower Yellow River near estuary. *Science of The Total*  
854 *Environment* **2020**, *707*, 135601.
- 855 97. Wang, G.; Lu, J.; Tong, Y.; Liu, Z.; Zhou, H.; Xiayihazi, N., Occurrence and pollution  
856 characteristics of microplastics in surface water of the Manas River Basin, China. *Science of The Total*  
857 *Environment* **2020**, *710*, 136099.
- 858 98. Gray, A. D.; Wertz, H.; Leads, R. R.; Weinstein, J. E., Microplastic in two South Carolina  
859 Estuaries: Occurrence, distribution, and composition. *Marine Pollution Bulletin* **2018**, *128*, 223-233.
- 860 99. Park, T.-J.; Lee, S.-H.; Lee, M.-S.; Lee, J.-K.; Lee, S.-H.; Zoh, K.-D., Occurrence of microplastics  
861 in the Han River and riverine fish in South Korea. *Science of The Total Environment* **2020**, *708*, 134535.
- 862 100. Lv, W.; Zhou, W.; Lu, S.; Huang, W.; Yuan, Q.; Tian, M.; Lv, W.; He, D., Microplastic pollution  
863 in rice-fish co-culture system: A report of three farmland stations in Shanghai, China. *Science of The*  
864 *Total Environment* **2019**, *652*, 1209-1218.
- 865 101. Liu, M.; Lu, S.; Song, Y.; Lei, L.; Hu, J.; Lv, W.; Zhou, W.; Cao, C.; Shi, H.; Yang, X.; He, D.,  
866 Microplastic and mesoplastic pollution in farmland soils in suburbs of Shanghai, China. *Environmental*  
867 *Pollution* **2018**, *242*, 855-862.
- 868 102. Cozar, A.; Echevarria, F.; Gonzalez-Gordillo, J. I.; Irigoien, X.; Ubeda, B.; Hernandez-Leon, S.;  
869 Palma, A. T.; Navarro, S.; Garcia-de-Lomas, J.; Ruiz, A.; Fernandez-de-Puelles, M. L.; Duarte, C. M.,  
870 Plastic debris in the open ocean. *Proceedings of the National Academy of Sciences* **2014**, *111*, (28),  
871 10239-10244.
- 872 103. Thompson, R.; Olsen, Y.; Mitchell, R., Lost at Sea: Where Is All the Plastic? *Science* **2004**, *304*,  
873 (5672), 838.
- 874 104. Hitchcock, J. N., Storm events as key moments of microplastic contamination in aquatic  
875 ecosystems. *Science of The Total Environment* **2020**, *734*, 139436.

- 876 105. Koelmans, A. A.; Redondo-Hasselerharm, P. E.; Mohamed Nor, N. H.; Kooi, M., Solving the  
877 Nonalignment of Methods and Approaches Used in Microplastic Research to Consistently Characterize  
878 Risk. *Environmental Science & Technology* **2020**, *54*, (19), 12307-12315.
- 879 106. González-Fernández, D.; Hanke, G.; Viejo Marin, J.; Cozar Cabañas, A. In *Modelling floating*  
880 *macro litter loads from rivers to the marine environment based on visual observations*, the European  
881 Geosciences Union (EGU) General Assembly Conference, 2019/04/1, 2019; 2019; p 18013.
- 882 107. Horton, A.; Walton, A.; Spurgeon, D., Microplastics in freshwater and terrestrial environments:  
883 Evaluating the current understanding to identify the knowledge gaps and future research priorities.  
884 *Science of The Total Environment* **2017**, *586*, 127-141.
- 885 108. Besseling, E.; Quik, J. T. K.; Sun, M.; Koelmans, A. A., Fate of nano- and microplastic in  
886 freshwater systems: A modeling study. *Environmental Pollution* **2017**, *220*, 540-548.
- 887 109. Lagarde, F.; Olivier, O.; Zanella, M.; Daniel, P.; Hiard, S.; Caruso, A., Microplastic interactions  
888 with freshwater microalgae: Hetero-aggregation and changes in plastic density appear strongly  
889 dependent on polymer type. *Environmental Pollution* **2016**, *215*, 331-339.
- 890 110. Cole, M.; Lindeque, P.; Halsband, C.; Galloway, T. S., Microplastics as contaminants in the  
891 marine environment: A review. *Marine Pollution Bulletin* **2011**, *62*, (12), 2588-2597.
- 892 111. Di, M.; Wang, J., Microplastics in surface waters and sediments of the Three Gorges Reservoir,  
893 China. *Science of the Total Environment* **2018**.
- 894 112. Andrady, A., The plastic in microplastics: A review. *Marine pollution bulletin* **2017**, *119*, (1), 12-  
895 22.
- 896 113. Tibbetts, J.; Krause, S.; Lynch, I.; Sambrook Smith, G., Abundance, Distribution, and Drivers of  
897 Microplastic Contamination in Urban River Environments. *Water* **2018**, *10*, 1597.
- 898 114. Long, M.; Moriceau, B.; Gallinari, M.; Lambert, C.; Huvet, A.; Raffray, J.; Soudant, P.,  
899 Interactions between microplastics and phytoplankton aggregates: Impact on their respective fates.  
900 *Marine Chemistry* **2015**, *175*, 39-46.
- 901 115. Waldschlger, K.; Lechthaler, S.; Stauch, G.; Schüttrumpf, H., The way of microplastic through  
902 the environment – Application of the source-pathway-receptor model (review). *Science of The Total*  
903 *Environment* **2020**, *713*, 136584.
- 904 116. Capolupo, M.; Sørensen, L.; Jayasena, K. D. R.; Booth, A. M.; Fabbri, E., Chemical composition  
905 and ecotoxicity of plastic and car tire rubber leachates to aquatic organisms. *Water Research* **2020**, *169*,  
906 115270.
- 907 117. Kowalski, N.; Reichardt, A. M.; Waniek, J. J., Sinking rates of microplastics and potential  
908 implications of their alteration by physical, biological, and chemical factors. *Marine pollution bulletin*  
909 **2016**, *109*, (1), 310-319.
- 910 118. Andrady, A., Microplastics in the marine environment. *Marine Pollution Bulletin* **2011**, *62*, (8),  
911 1596-1605.
- 912 119. Woodall, L. C.; Sanchez-Vidal, A.; Canals, M.; Paterson, G. L. J.; Coppock, R.; Sleight, V.;  
913 Calafat, A.; Rogers, A. D.; Narayanaswamy, B. E.; Thompson, R. C., The deep sea is a major sink for  
914 microplastic debris. *Royal Society Open Science* **2014**, *1*, (4), 140317-140317.
- 915 120. Alomar, C.; Estarellas, F.; Deudero, S., Microplastics in the Mediterranean Sea: Deposition in  
916 coastal shallow sediments, spatial variation and preferential grain size. *Marine Environmental*  
917 *Research* **2016**, *115*, 1-10.
- 918 121. Scherer, C.; Weber, A.; Stock, F.; Vurusic, S.; Egerci, H.; Kochleus, C.; Arendt, N.; Foeldi, C.;  
919 Dierkes, G.; Wagner, M.; Brennholt, N.; Reifferscheid, G., Comparative assessment of microplastics  
920 in water and sediment of a large European river. *Science of The Total Environment* **2020**, *738*, 139866.
- 921 122. Wang, J.; Peng, J.; Tan, Z.; Gao, Y.; Zhan, Z.; Chen, Q.; Cai, L., Microplastics in the surface  
922 sediments from the Beijiang River littoral zone: Composition, abundance, surface textures and  
923 interaction with heavy metals. *Chemosphere* **2017**, *171*, 248-258.
- 924 123. Haberstroh, C.; Arias, M.; Yin, Z.; Wang, M. C., Effects of hydrodynamics on the cross-sectional  
925 distribution and transport of plastic in an urban coastal river. *Water Environment Research* **2020**, *93*:  
926 186-200.
- 927 124. Horton, A.; Dixon, S., Microplastics: An introduction to environmental transport processes. *Wiley*  
928 *Interdisciplinary Reviews: Water* **2017**, *5*, e1268.

929 125. Moore, C. J.; Moore, S. L.; Weisberg, S. B.; Lattin, G. L.; Zellers, A. F., A comparison of  
930 neustonic plastic and zooplankton abundance in southern California's coastal waters. *Marine Pollution*  
931 *Bulletin* **2002**, *44*, (10), 1035-1038.

932 126. Faure, F.; Demars, C.; Wieser, O.; Kunz, M.; de Alencastro, L. F., Plastic pollution in Swiss  
933 surface waters: nature and concentrations, interaction with pollutants. *Environmental Chemistry* **2015**,  
934 *12*, (5), 582.

935 127. Scheurer, M.; Bigalke, M., Microplastics in Swiss floodplain soils. *Environmental science &*  
936 *technology* **2018**, *52*, (6), 3591-3598.

937 128. Desforges, J.-P. W.; Galbraith, M.; Dangerfield, N.; Ross, P. S., Widespread distribution of  
938 microplastics in subsurface seawater in the NE Pacific Ocean. *Marine Pollution Bulletin* **2014**, *79*, (1-  
939 2), 94-99.

940 129. Yonkos, L. T.; Friedel, E. A.; Perez-Reyes, A. C.; Ghosal, S.; Arthur, C. D., Microplastics in Four  
941 Estuarine Rivers in the Chesapeake Bay, U.S.A. *Environmental Science & Technology* **2014**, *48*, (24),  
942 14195-14202.

943 130. Kapp, K. J.; Yeatman, E., Microplastic hotspots in the Snake and Lower Columbia rivers: A  
944 journey from the Greater Yellowstone Ecosystem to the Pacific Ocean. *Environmental Pollution* **2018**,  
945 *241*, 1082-1090.

946 131. He, B.; Goonetilleke, A.; Ayoko, G. A.; Rintoul, L., Abundance, distribution patterns, and  
947 identification of microplastics in Brisbane River sediments, Australia. *Science of The Total*  
948 *Environment* **2020**, *700*, 134467.

949 132. Gasperi, J.; Dris, R.; Bonin, T.; Rocher, V.; Tassin, B., Assessment of floating plastic debris in  
950 surface water along the Seine River. *Environmental Pollution* **2014**, *195*, 163-166.

951 133. CJ, M.; GL, L.; AF, Z., Quantity and type of plastic debris flowing from two urban rivers to coastal  
952 waters and beaches of Southern California. *Journal of Integrated Coastal Zone Management* **2011**, *11*,  
953 (1), 65-73.

954 134. van Emmerik, T.; Strady, E.; Kieu-Le, T.-C.; Nguyen, L.; Gratiot, N., Seasonality of riverine  
955 macroplastic transport. *Scientific reports* **2019**, *9*, (1), 1-9.

956 135. CJ, M.; GL, L.; AF, Z., Working our way upstream: a snapshot of land-based contributions of  
957 plastic and other trash to coastal waters and beaches of Southern California. *in Proceedings of the*  
958 *Plastic Debris Rivers to Sea Conference* **2005**.

959 136. Kooi, M.; Koelmans, A. A., Simplifying Microplastic via Continuous Probability Distributions  
960 for Size, Shape, and Density. *Environmental Science & Technology Letters* **2019**, *6*, (9), 551-557.

961 137. Wang, Z.; Su, B.; Xu, X.; Di, D.; Huang, H.; Mei, K.; Dahlgren, R. A.; Zhang, M.; Shang, X.,  
962 Preferential accumulation of small (<300 µm) microplastics in the sediments of a coastal plain river  
963 network in eastern China. *Water Research* **2018**, *144*, 393-401.

964 138. Stanton, T.; Johnson, M.; Nathanail, P.; MacNaughtan, W.; Gomes, R. L., Freshwater microplastic  
965 concentrations vary through both space and time. *Environmental Pollution* **2020**, *263*, 114481.  
966