



Carcinogenicity evaluation for the application of carbon nanotubes as biomaterials in rasH2 mice

SUBJECT AREAS:
BIOMATERIALS
CARBON NANOTUBES AND FULLERENES
TOXICOLOGY
ONCOGENESIS

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The application of carbon nanotubes (CNTs) as biomaterials is of wide interest, and studies examining their application in medicine have had considerable significance. Biological safety is the most important factor when considering the clinical application of CNTs as biomaterials, and various toxicity evaluations are required. Among these evaluations, carcinogenicity should be examined with the highest priority; however, no report using transgenic mice to evaluate the carcinogenicity of CNTs has been published to date. Here, we performed a carcinogenicity test by implanting multi-walled CNTs (MWCNTs) into the subcutaneous tissue of rasH2 mice, using the carbon black present in black tattoo ink as a reference material for safety. The rasH2 mice did not develop neoplasms after being injected with MWCNTs; instead, MWCNTs showed lower carcinogenicity than carbon black. Such evaluations should facilitate the clinical application and development of CNTs for use in important medical fields.

Carbon nanotubes (CNTs) are fibrous particles with nanoscale diameter and microscale length. CNTs have unique physical and chemical characteristics, and the study and development of these particles is ongoing in various fields¹⁻⁶. For example, CNTs have already been used in the electrodes of lithium batteries and displays because of their superior electrical properties⁷⁻⁹, as well as in tennis rackets and golf clubs, to take advantage of their good mechanical properties. Furthermore, applications in airplanes and motor vehicle parts are expected in the future^{10,11}. In the medical field, the use of CNTs as biomaterials has been extensively studied in recent years¹²⁻¹⁴. CNTs have been reported to be an excellent scaffold material for bone and nerve regeneration¹⁵⁻¹⁷, and studies examining the efficacy of combining CNTs with existing implants as a reinforcement to improve their mechanical strength and durability are ongoing¹⁸. CNTs also have applications in cancer therapy, including in drug delivery systems and cancer cell imaging, and the ability of CNTs to absorb electromagnetic waves make them suitable for use in hyperthermia in cancer^{19,20}.

Because of their expected applications in various fields, such as those described above, the biological safety of CNTs has been studied worldwide. However, most previous studies have examined the inhalation of CNTs, as this provides the most convenient method of internalization, and studies on the safety of the inhaled particles are urgently required²¹⁻²³. Because CNTs have a long, thin, fibrous structure similar to that of asbestos, which can cause malignant mesothelioma when inhaled into the lungs, CNTs have been suspected to have carcinogenic properties²⁴. Various studies examining this possibility have been performed²⁵⁻²⁸; because the results have shown that CNTs and asbestos have different safety characteristics, a different standard value for the safe inhalation of CNTs has been proposed^{29,30}.

As opposed to inhalation, the application of CNTs as a biomaterial requires completely different evaluations of safety. Such evaluations include implantation tests, cytotoxicity tests, carcinogenicity tests, and in vivo pharmacokinetic studies³¹. Although some evaluations of the safety of CNTs when used as a biomaterial have been



published in the last several years, the number of these reports is quite small compared to that for inhalation³². The carcinogenicity of CNTs is the most important issue to be addressed when considering them as a biomaterial, and tests using experimental animals are important for studying carcinogenicity in humans. However, to the best of our knowledge, no evaluations examining the implantation of CNTs in transgenic animals with increased susceptibility to carcinogenic substances have been performed to date.

The most representative transgenic mouse models with increased sensitivity to carcinogens are the p53^{+/-} and rasH2 mouse models³³. While rasH2 mice have been used in previous carcinogenicity studies on biomaterials, these tests examined solid materials, and no study involving particles has been performed to date. The positive and negative controls used for the evaluation of solid materials are *N*-methyl-*N*-nitrosourea (MNU) and high-density polyethylene³⁴. However, while MNU can be used as a positive control when evaluating the carcinogenicity of particles, a reference standard of particles that have been established to have no carcinogenicity should be used as the reference negative control²⁶.

Asbestos is often used as a control material in carcinogenicity tests of CNTs. However, asbestos is only an appropriate control in inhalation tests, and is not appropriate as a control for the carcinogenicity of implantable materials because it has never been used as a biomaterial. As toxicity due to inhalation and implantation are completely different, it is not possible to evaluate the carcinogenicity of CNTs using asbestos as a reference. We therefore applied carbon black, which is used as a black tattoo ink and has been histologically proven to have no carcinogenicity to the human body, as the most appropriate reference for evaluating the carcinogenicity of CNTs used as biomaterials³⁵ in this subcutaneous administration study using rasH2 mice. Our results have important implications on further studies examining the applicability of CNTs as a biomaterial.

Results

CNTs, reference material and positive control. The CNTs used for evaluation were multi-walled CNTs (MWCNTs) (VGCF-S; Showa Denko, Tokyo, Japan) with a mean diameter of 100 nm, mean length of 10 μm, and a >99.9% carbon content; they were manufactured by chemical vapor deposition. Sterilization was performed by autoclaving at 121°C for 15 min. Tween 80 (Polysorbate 80; NOF, Tokyo, Japan) was used as a surfactant to disperse the MWCNTs uniformly in solution.

The shape of precipitated solid contents obtained by drying commercially available black tattoo ink (Sumi-Black; Unique Tattoos, Subiaco, Australia) was nearly uniform, with a diameter of 30–50 nm. The tattoo ink particles contained 99.57 wt% C and 0.43 wt% Na, with no other element being detected. Comparative Raman and transmission electron microscope studies showed that the black tattoo inks were composed of nano-sized carbon black particles. Therefore, the carbon black particles from the black tattoo inks were used as the reference material³⁵. The CNT mixture was created by sonicating a physiological saline solution containing 0.1 vol% Tween 80 and 3 mg/mL MWCNTs for 30 min by ultrasonic dispersion; the reference mixture was created from carbon black (3 mg/mL) using the same procedure. A positive control solution with known tissue toxicity was prepared by mixing MNU (Wako, Osaka, Japan) with Tween 80 to obtain a 3 mg/mL solution.

Carcinogenicity study using transgenic rasH2 mice. CNT safety was examined by analyses involving subcutaneous implantation in rasH2 mice (CLEA Japan, Tokyo, Japan), which express the *c-Ha-ras* proto-oncogene derived from humans. Test substances were implanted when the mice were 6 weeks old, and tumor onset was evaluated after 26 weeks (at 32 weeks of age) to predict the carcinogenicity of these substances in humans. This test period

was chosen because rasH2 mice seldom develop spontaneous tumors until they are 34 weeks old²⁷.

Six-week-old male rasH2 mice were anesthetized using diethyl ether, and 550 μL of CNT mixture (approximately 75 mg/kg) was injected into the subcutaneous tissue of the back of each mouse using a 1-mL syringe and a 26 G needle (CNT group). The carbon black, solvent, and MNU groups were injected with the corresponding substances by using the same procedure. Each group consisted of 10 rasH2 mice (total 40), and all the mice were weighed once every week during the survival period, except at 10 and 20 weeks, when no and incorrect measurement was performed, respectively. Mice were euthanized after 26 weeks and dissected. Subjects that died or were euthanized because of extreme weakness before 26 weeks were dissected at the time of death.

All the mice in the CNT group and solvent group were alive at 26 weeks after receiving the injection. In the carbon black group, 1 mouse died at 22 weeks, while the 9 remaining mice survived until 26 weeks. In the MNU group, 1 mouse each died at 12, 14, 17, and 21 weeks following injection, while the remaining 6 mice were alive at 26 weeks (Fig. 1a). Statistically significant differences in survival were observed between the MNU and CNT groups, and between the MNU and control groups ($p = 0.029406$ for both comparisons). When weight change during the study period was examined, the CNT group, carbon black group, and solvent group showed similar changes, while the 4 mice that died prior to the end of the study in the MNU group showed rapid decrease in weight from several weeks before the death. (Fig. 1b).

Histological examination showed a mass in the spleen of 1 mouse from the CNT group; however, it was histologically shown to be an inflammatory pseudotumor (Fig. 2a). The single mouse that died prior to 26 weeks in the carbon black group did not show any neoplasms, and the cause of death was unknown. Of the 9 surviving mice, 1 had a hemangioma in the spleen, while 1 other mouse was found to have a neoplasm in the lung, which was histologically judged to be an adenoma (Fig. 2b). None of the mice in the solvent group were found to have onset of neoplasm in any organ. In contrast, all 10 mice in the MNU group developed neoplasms, including forestomach tumors, papillomas of the skin (face, thigh, and injection site), and perineal tumors in 10, 6, and 5 mice, respectively. In addition, a lung adenoma was observed in 1 mouse, and 2 mice were found to have malignant lymphoma (Fig. 2c). Abnormal growth of squamous cells was histologically shown in all the mice in the MNU group.

Spleen tumors from 1 mouse from each of the CB and CNT groups were also examined by immunostaining for factor VIII, and were diagnosed as a hemangioma and inflammatory pseudotumor, respectively (Fig. 3). The type and frequency of neoplasms in each group are shown in Table 1. A significantly larger number of papillomas were observed in the forestomach and perineal in the MNU group compared to the other groups (significant differences at $p=0.00000541250$ and $p=.016254$, respectively).

No neoplasms were detected at the injection sites in the CNT group and macrophage-phagocytosed CNTs were observed; furthermore, inflammatory cells such as neutrophils and lymphocytes were not detected in the surrounding tissues (Fig. 4a). Similarly, neoplasms were not observed at the injection sites in the carbon black group, and images of macrophages taken after they had phagocytosed carbon black particles were similar to those for the CNT group (Fig. 4b). The injection site did not show any change in the solvent group, although a skin tumor was observed at the injection site of 2 mice from the MNU group, which was determined to be a squamous cell tumor upon analysis of a tissue image (Fig. 4c).

Discussion

Although some studies examining the safety of CNTs as a biomaterial have been published, the evaluation of these results is difficult, and

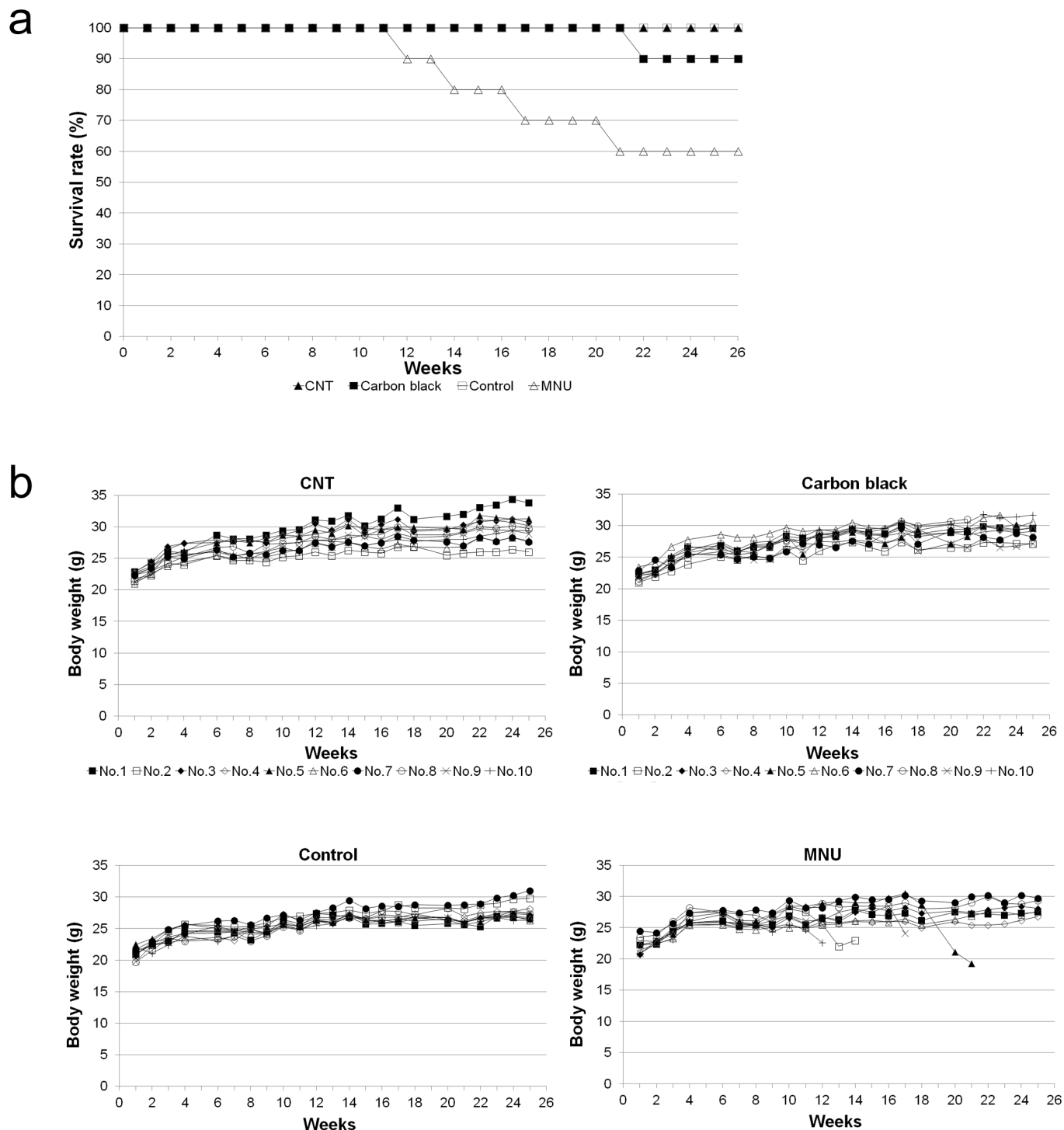


Figure 1 | Survival rate and change in the weights of the transgenic rasH2 mice. (a) Survival rate: All the mice in the CNT group and solvent group were alive at 26 weeks after injection. In the carbon black group, 1 mouse died at 22 weeks, and the remaining 9 mice were alive at 26 weeks. In the MNU group, 1 mouse died at each of 12, 14, 17, and 21 weeks following the injection, and the remaining 6 mice were alive at 26 weeks. The survival rate was statistically different between the MNU and CNT groups, and between the MNU and control groups ($p = 0.029406$ for both comparisons). (b) Weight change in mice: The CNT group, carbon black group, and solvent group showed similar weight change, while the 4 mice in MNU group that died before the end of the study period showed rapid decrease in weight from several weeks before death.

different results have been reported for the same type of safety test^{37–40}. The biggest reason for such discrepancy is the lack of a nanoparticle that has established biological safety and is suitable for use as a reference material²⁶. Similar to CNTs, which are nanosized carbon particles, tattoo ink is made up of nanosized carbon black particles, and is long known to be safe for use in the human body. In 2011, we

therefore reported that black tattoo ink (carbon black) could be an optimal reference material for evaluating the safety of CNTs³⁵. While some studies that used carbon black particles as a negative control for CNTs had been published before our report, the researchers in those studies chose carbon black particles arbitrarily^{41–45}. As there are numerous types and grades of carbon black available, a standard for

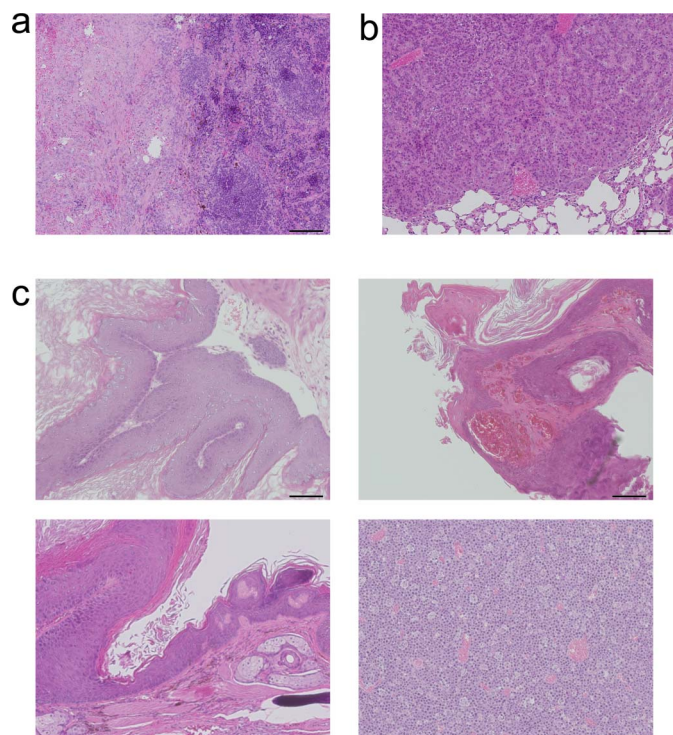


Figure 2 | Tissue images of masses that developed in the rash2 mice.

(a) A mass was observed in the spleen of 1 mouse from the CNT group, but it was found to be an inflammatory pseudotumor and not a neoplasm. (b) One mouse from the carbon black group was alive at the 26th week and was found to have developed a neoplasm in the lung, which was diagnosed as an adenoma. (c) All 10 mice in the MNU group developed forestomach tumors, with abnormal growth of squamous cells (left upper panel). Papillomas of skin were observed in 6 mice (right upper panel). Perineal tumors were found to have developed in 5 mice (left lower panel), while 2 mice developed malignant lymphoma (right lower panel). Hematoxylin-eosin staining. Scale bar: 100 μ m.

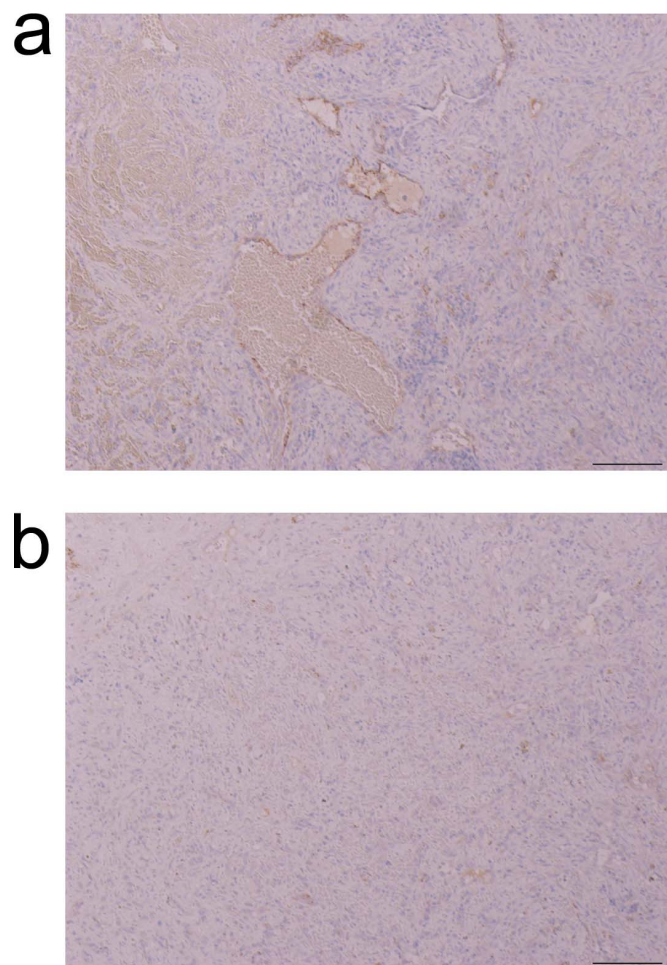


Figure 3 | Immunostaining of the spleen tissue for Factor VIII.

(a) Hemangioma was diagnosed in the spleen of a mouse from the carbon black group. Immunostaining revealed vascular endothelial-like cells stained in dark brown. Atypical nucleus was not observed. (b) Inflammatory pseudotumor was diagnosed in the spleen of a mouse from the CNT group. Few cells were stained. Scale bar: 100 μ m.

Table 1 | Incidence of neoplastic changes in resH2 mice following injection with a CNT, carbon black, solvent, or MNU test substance

Organ	Diagnosis/total number	control	carbon black	CNT	MNU
		10	10	10	10
Skin (back area)	Papilloma	0	0	0	2
	Keratoacanthoma	0	0	0	0
Skin (face)	Papilloma	0	0	0	3
	Keratoacanthoma	0	0	0	0
Skin (thigh)	Papilloma	0	0	0	1
	Keratoacanthoma	0	0	0	0
Spleen	Inflammatory pseudotumor	0	0	1	0
	Hemangioma	0	1	0	0
Hematopoietic system	Malignant lymphoma	0	0	0	2
	Epithelial thymoma	0	0	0	0
Kidneys	Hemangioma	0	0	0	0
Pancreas	Hemangioma	0	0	0	0
Lungs	Adenocarcinoma	0	0	0	0
	Adenoma	0	1	0	1
Forestomach	Hemangioma	0	0	0	0
	Papilloma	0	0	0	10*
	Basal cell tumor	0	0	0	0
	Squamous cell carcinoma	0	0	0	0
Perineal	Papilloma	0	0	0	5**

*, **: significant differences at $p=0.0000054125$ and $p=0.016254$, respectively (Fisher's direct method).

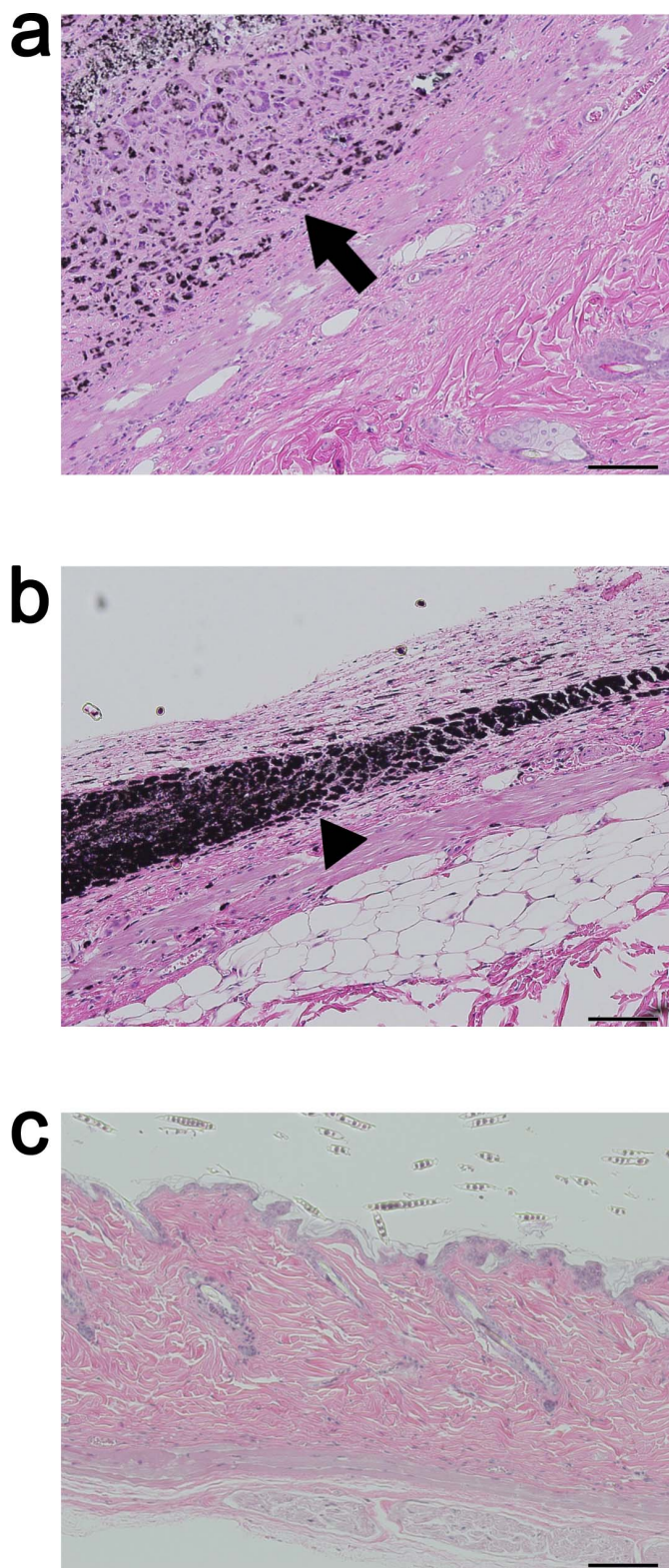


Figure 4 | Images of the skin and subcutaneous tissue at the injection site of rasH2 mice, at 26 weeks after injection with a CNT, carbon black, or MNU test substance. (a) None of the mice in the CNT group developed neoplasms, and macrophages accumulated after phagocytosing CNT particles. Inflammatory cells such as neutrophils and lymphocytes were not observed in the surrounding tissues. (b) In the carbon black group, macrophages were similarly found to have phagocytosed carbon black particles, and no neoplasms were observed. (c) In the MNU group, a skin tumor was detected at the injection site of 2 mice, which was found to be a squamous cell tumor. Arrow: CNTs, Arrowhead: carbon black. Hematoxylin-eosin staining. Scale bar: 100 μm .

its use as a control should be established. We therefore proposed that the optimal standard carbon black should have a carbon purity of $\geq 99.5\%$ and a particle size of 30–50 nm, similar to that used in tattoo ink³⁵. This carbon black and CNTs are both made of highly pure carbon, which allows comparisons at similar weights and enables the quantitative evaluation of CNT safety for use as a biomaterial. While our previous study found that the toxicity of CNTs is comparable to that of the carbon black used in tattoos in an *in vivo* implantation study, cytotoxicity study, and cytokine production assay of CNTs³⁵, this is the first study to use carbon black as a reference material to examine the subcutaneous implantation of CNTs in test animals susceptible to cancer.

The rasH2 mice used in this study express the human-derived c-H-ras proto-oncogene, which causes them to spontaneously develop multiorgan tumors after approximately 35 weeks of age, leading to death. However, these mice seldom develop tumors before 34 weeks of age, and carcinogenicity tests can be performed until then. As this particular characteristic can reduce the test time, work load, and cost of such studies, and the sensitivity of evaluation is high, rasH2 mice are mainly used to evaluate the carcinogenicity of chemical substances that are used in humans⁴⁶. This rasH2 mouse model is approved as a test animal for new drug applications by regulatory authorities such as the Food and Drug Administration in the USA, the Committee for Proprietary Medicinal Products in the EU, and the Ministry of Health, Labour and Welfare in Japan. The company that developed these mice is maintaining this strain in order to preserve study accuracy, so that highly reproducible evaluations can be achieved^{47,48}. Although their application in carcinogenesis tests for solid implants of biomaterials has only begun in recent years, such as in tests for the potential carcinogenicity of IC tags^{36,48–50}, the implantation test of CNTs in rasH2 mice reported here is the first to evaluate nanosized particles.

All 10 positive control mice injected with MNU developed neoplasms in 26 weeks, with 4 mice dying prior to the end of the study. In contrast, none of the mice in the CNTs group died; instead, tissue examination of the CNT group mice after dissection showed an inflammatory pseudotumor in only 1 mouse, and neoplasm was not observed in any mouse. The survival rate was significantly different between the MNU and CNT groups. Because the amount of test substance used in this study, i.e., 75 mg/kg, was remarkably higher than that used in conventional mouse implantation tests^{37–40}, and the mice used have higher sensitivity to carcinogenicity, we judged that it was not necessary to perform the test with a higher density when neoplasm was not observed. In the carbon black group, a hemangioma in the spleen and neoplasm in the lung of 1 mouse each were also observed. The results from this initial subcutaneous implantation test of carcinogenicity involving transgenic mice with higher susceptibility to cancer have therefore shown that CNTs do not induce the formation of neoplasms, as the carcinogenicity was lower than that of the carbon black used in tattoo ink.

During the majority of carcinogenesis, there is a complex relationship between factors such as the accumulation of mutations, DNA disorders, and chemical changes in nucleotides. Because the occurrence of tumors in transgenic mice such as rasH2 mice is influenced by both mutated oncogenes and mutated antioncogenes, studies using rasH2 mice can evaluate only the result of the complex process of carcinogenesis³⁵. In contrast, genotoxicity tests can evaluate the carcinogenicity of a material for individual parameters, thereby allowing evaluation to be performed relatively easily and the ISO10993 series requires that multiple types of tests be performed³¹. Typical genotoxicity tests include bacterial reverse mutation assays (Ames test), micronucleus assays, and Comet assays. CNTs have already been tested using such methods to examine carcinogenicity following inhalation^{51–53}. These genotoxicity tests are also important for the evaluation of carcinogenicity with regard to the application of CNTs as biomaterials, and while many previous reports have found



that CNTs have no genotoxicity, some have reported differing results, and no consensus has been achieved^{38,54}. This discrepancy may result from differences in test conditions, sample size, and evaluation method, although the most important cause is the absence of a suitable reference material²⁶. As shown in this study, carbon black—similar to that used for tattoo ink—is an appropriate reference material for future studies. The carcinogenicity of CNTs when used as a biomaterial can be further clarified by comprehensive evaluations of the results of genotoxicity tests and implantation tests such as the one performed in this study.

This study is the first to evaluate the carcinogenicity of CNTs as a biomaterial using transgenic rasH2 mice that overexpress the rasH2 oncogene and are highly sensitive to carcinogenesis. Although CNTs showed no carcinogenicity in this study and had lower carcinogenicity than the reference material, it cannot be completely ruled out on the basis of these results alone. A larger number of carcinogenicity studies comparing the results for CNTs to those for reference materials with established biological safety, such as the carbon black used in this study, should be performed. Results from various genotoxicity studies and studies using test animals susceptible to cancer should also be comprehensively evaluated to judge the carcinogenicity of CNTs. The completion of studies that use CNTs as a biomaterial is expected to result in progress in various important medical fields, including cancer treatment, regenerative medicine, and the development of drug delivery systems.

Methods

Reference material. The shape of precipitated solid contents obtained by drying commercially available black tattoo ink (Sumi-Black; Unique Tattoos, Subiaco, Australia) was determined by scanning electron microscopy (JSM-7000F and JED-2300, JEOL, Tokyo, Japan) to be nearly uniform, with a diameter of 30–50 nm (Fig. S1a). Elemental and compositional analyses performed using energy dispersive x-ray spectroscopy showed that the tattoo ink particles contained 99.57 wt% C and 0.43 wt% Na, with no other element being detected. Comparative Raman studies (R-XN1-532, Kaiser, Ann Arbor, MI) on the tattoo ink particles were performed using the industrial general carbon black (Vulcan XC 72, Cabot, Boston, MA), which is used as a filler for rubber products, as a control. The tattoo ink particles showed a Raman feature similar to that of the general carbon black, where the G band at 1582 cm^{-1} originated from the E_{2g2} graphite mode and the D band at 1360 cm^{-1} was a defect-induced mode (Fig. S1b). The R values (defined as the integrated intensity of the D band divided by the intensity of the G band) for the tattoo ink particles and general carbon black were comparable. Observation by transmission electron microscope (JEM-2010, JEOL) revealed that the morphologies of the tattoo ink particles were almost the same as that for general carbon black (Fig. S1c).

Carcinogenicity study. Visual inspection of tumor onset in transgenic rasH2 mice was performed during dissection. Each organ was fixed separately, embedded, and cut into thin sections before being stained with hematoxylin and eosin; subsequently, they were placed on a slide and observed using an optical microscope.

As the diagnosis of hemangioma by hematoxylin and eosin staining was difficult in the spleen tissue, vascular endothelial cells were also immunostained for Factor VIII. Two toxicologic pathology specialists diagnosed the tumor in accordance with a globally recognized text of pathological tumor diagnosis in mice (International classification of rodent tumors; the mouse. Editor: Ulrich Mohr. WHO International Agency for Research on Cancer. Springer-Verlag Berlin Heidelberg 2001).

All animal experimentation procedures were performed in compliance with the guidelines of the institutional animal care committee of Shinshu University and the experiments were approved by the committee.

Statistical analysis. Survival rates were calculated using the Kaplan-Meier method and tested by the log-rank test. The tumor occurrence was tested by Fisher's direct method. All statistical analyses were performed using statistical software (Statcel, OMS, Tokorozawa, Japan), and p values of less than 0.05 were considered significant.

- Oberlin, A., Endo, M. & Koyama, T. Filamentous growth of carbon through benzene decomposition. *J. Syst. Growth* **32**, 335–349 (1976).
- Iijima, S. Helical microtubules of graphitic carbon. *Nature* **354**, 56–58 (1991).
- Qu, L., Dai, L., Stone, M., Xia, Z. & Wang, Z. L. Carbon nanotube arrays with strong shear binding-on and easy normal lifting-off. *Science* **322**, 238–242 (2008).
- Shannon, M. A. *et al.* Science and technology for water purification in the coming decades. *Nature* **452**, 301–310 (2008).
- Xiong, F., Liao, A. D., Estrada, D. & Pop, E. Low-power switching of phase-change materials with carbon nanotube electrodes. *Science* **332**, 68–570 (2011)
- Van Noorden, R. Chemistry: the trials of new carbon. *Nature* **469**, 14–16 (2011).
- Endo, M. *et al.* Vapor-grown carbon fibers (VGCFs) basic properties and their battery applications. *Carbon* **39**, 1287–1297 (2001).
- Kang, S. J. *et al.* High-performance electronics using dense, perfectly aligned arrays of single-walled carbon nanotubes. *Nat. Nanotechnol.* **2**, 230–236 (2007).
- Scrosati, B. Nanomaterials: paper powers battery breakthrough. *Nat. Nanotechnol.* **2**, 598–599 (2007).
- Pugno, N. M., Bosia, F. & Carpinteri, A. Multiscale stochastic simulations for tensile testing of nanotube-based macroscopic cables. *Small* **4**, 1044–1052 (2008).
- Byrne, M. T. & Gun'ko, Y. K. Recent advances in research on carbon nanotube-polymer composites. *Adv. Mater.* **22**, 1672–1688 (2010).
- Pagona, G. & Tagmatarchis, N. Carbon nanotubes: materials for medicinal chemistry and biotechnological applications. *Curr. Med. Chem.* **13**, 1789–1798 (2006).
- Saito, N. *et al.* Carbon nanotubes for biomaterials in contact with bone. *Curr. Med. Chem.* **15**, 523–527 (2008).
- Saito, N. *et al.* Carbon nanotubes: biomaterial applications. *Chem. Soc. Rev.* **38**, 1897–1903 (2009).
- Harrison, B. S. & Atala, A. Carbon nanotube applications for tissue engineering. *Biomaterials* **28**, 344–353 (2007).
- Dubin, R. A., Callegari, G., Kohn, J. & Neimark, A. Carbon nanotube fibers are compatible with Mammalian cells and neurons. *IEEE Trans Nanobioscience* **7**, 11–14 (2008).
- Usui, Y. *et al.* Carbon nanotubes with high bone-tissue compatibility and bone-formation acceleration effects. *Small* **4**, 240–246 (2008).
- Lee, H. H., Sang Shin, U., Lee, J. H. & Kim, H. W. Biomedical nanocomposites of poly(lactic acid) and calcium phosphate hybridized with modified carbon nanotubes for hard tissue implants. *J. Biomed. Mater. Res. B Appl. Biomater.* **98B**, 246–254 (2011).
- Shi Kam, N. W., Jessop, T. C., Wender, P. A. & Dai, H. Nanotube molecular transporters: internalization of carbon nanotube-protein conjugates into mammalian cells. *J. Am. Chem. Soc.* **126**, 6850–6851 (2004).
- Yang, F. *et al.* Magnetic functionalised carbon nanotubes as drug vehicles for cancer lymph node metastasis treatment. *Eur. J. Cancer.* **47**, 1873–1882 (2011).
- Muller, J. *et al.* Respiratory toxicity of multi-wall carbon nanotubes. *Toxicol. Appl. Pharmacol.* **207**, 221–231 (2005).
- Lam, C. W., James, J. T., McCluskey, R., Arepalli, S. & Hunter, R. L. A review of carbon nanotube toxicity and assessment of potential occupational and environmental health risks. *Crit. Rev. Toxicol.* **36**, 189–217 (2006).
- Carrero-Sanchez, J. C. Biocompatibility and toxicological studies of carbon nanotubes doped with nitrogen. *Nano Lett.* **6**, 1609–1616 (2006).
- Poland, C. A. Carbon nanotubes introduced into the abdominal cavity of mice show asbestos-like pathogenicity in a pilot study. *Nat. Nanotechnol.* **3**, 423–428 (2008).
- Ma-Hock, L. *et al.* Inhalation toxicity of multiwall carbon nanotubes in rats exposed for 3 months. *Toxicol. Sci.* **112**, 468–481 (2009).
- Auffan, M. *et al.* Towards a definition of inorganic nanoparticles from an environmental, health and safety perspective. *Nat. Nanotechnol.* **4**, 634–641 (2009).
- Oyabu, T. *et al.* Biopersistence of inhaled MWCNT in rat lungs in a 4-week well-characterized exposure. *Inhal. Toxicol.* **23**, 784–791 (2011).
- Nagai, H. *et al.* Diameter and rigidity of multiwalled carbon nanotubes are critical factors in mesothelial injury and carcinogenesis. *Proc. Natl. Acad. Sci. USA* **108**, E1330–E1338 (2011).
- Porter, D. W. *et al.* Mouse pulmonary dose- and time course-responses induced by exposure to multi-walled carbon nanotubes. *Toxicology* **269**, 136–147 (2010).
- Kobayashi, N. *et al.* Biological response and morphological assessment of individually dispersed multi-wall carbon nanotubes in the lung after intratracheal instillation in rats. *Toxicology* **276**, 143–153 (2010).
- ISO 10993-1:2009 Biological evaluation of medical devices Part 1: Evaluation and testing.
- van der Zande, M., Junker, R., Walboomers, X. F. & Jansen, J. A. Carbon nanotubes in animal models: a systematic review on toxic potential. *Tissue Eng. Part B Rev.* **17**, 57–69 (2011).
- Long, G. G., Morton, D., Peters, T., Short, B. & Skydsgaard, M. Alternative mouse models for carcinogenicity assessment: industry use and issues with pathology interpretation. *Toxicol. Pathol.* **38**, 43–50 (2010).
- Palazzi, X. & Kergozien-Framery, S. Use of rasH2 transgenic mice for carcinogenesis testing of medical implants. *Exp. Toxicol. Pathol.* **61**, 433–441 (2009).
- Hara, K. *et al.* Evaluation of CNT toxicity in comparison to tattoo ink nanoparticles for use as a biomaterial. *Mater. Today* **14**, 434–440 (2011).
- Urano, K. *et al.* Examination of percutaneous application in a 26-week carcinogenicity test in CB6F1-TG rasH2 mice. *J. Toxicol. Sci.* **32**, 367–375 (2007).
- Firme, 3rd, C. P. & Bandaru, P. R. Toxicity issues in the application of carbon nanotubes to biological systems. *Nanomedicine* **6**, 245–256 (2010).
- Beg, S. *et al.* Advancement in carbon nanotubes: basics, biomedical applications and toxicity. *J. Pharm. Pharmacol.* **63**, 141–163 (2011).
- Shvedova, A. A. *et al.* Mechanisms of pulmonary toxicity and medical applications of carbon nanotubes: Two faces of Janus? *Pharmacol. Ther.* **121**, 192–204 (2009).
- Yang, K. & Liu, Z. In vivo biodistribution, pharmacokinetics, and toxicology of carbon nanotubes. *Curr. Drug Metab.* (2012) [Epub ahead of print]



41. Wang, L. *et al.* Dispersion of single-walled carbon nanotubes by a natural lung surfactant for pulmonary in vitro and in vivo toxicity studies. *Part. Fibre Toxicol.* **7**, 31 (2010).
42. Palomäki, J. *et al.* Long, needle-like carbon nanotubes and asbestos activate the NLRP3 inflammasome through a similar mechanism. *ACS Nano* **27**, 6861–6870 (2011).
43. Patlolla, A. K., Berry, A. & Tchounwou, P. B. Study of hepatotoxicity and oxidative stress in male Swiss-Webster mice exposed to functionalized multi-walled carbon nanotubes. *Mol. Cell Biochem.* **358**, 189–199 (2011).
44. Sanchez, V. C., Weston, P., Yan, A., Hurt, R. H. & Kane, A. B. A 3-dimensional in vitro model of epithelioid granulomas induced by high aspect ratio nanomaterials. *Part. Fibre Toxicol.* **8**, 17 (2011).
45. Teeguarden, J. G. *et al.* Comparative proteomics and pulmonary toxicity of instilled single-walled carbon nanotubes, crocidolite asbestos, and ultrafine carbon black in mice. *Toxicol. Sci.* **120**, 123–135 (2011).
46. Ando, K. *et al.* Chemically induced forestomach papillomas in transgenic mice carry mutant human c-Ha-ras transgenes. *Cancer Res.* **52**, 978–982 (1992).
47. Boverhof, D. R. *et al.* Transgenic animal models in toxicology: historical perspectives and future outlook. *Toxicol. Sci.* **121**, 207–233 (2011).
48. Urano, K., Tamaoki, N. & Nomura, T. Establishing a laboratory animal model from a transgenic animal: rasH2 mice as a model for carcinogenicity studies in regulatory science. *Vet. Pathol.* **49**, 16–23 (2012).
49. Urano, K. *et al.* Use of IC tags in short-term carcinogenicity study on CB6F1 TGrasH2 mice. *J. Toxicol. Sci.* **31**, 407–418 (2006).
50. Palazzi, X. & Kergozien-Framery, S. Use of rasH2 transgenic mice for carcinogenesis testing of medical implants. *Exp. Toxicol. Pathol.* **61**, 433–441 (2009).
51. Di Sotto, A., Chiaretti, M., Carru, G. A., Bellucci, S. & Mazzanti, G. Multi-walled carbon nanotubes: Lack of mutagenic activity in the bacterial reverse mutation assay. *Toxicol. Lett.* **184**, 192–197 (2009).
52. Thurnherr, T. *et al.* A comparison of acute and long-term effects of industrial multiwalled carbon nanotubes on human lung and immune cells in vitro. *Toxicol. Lett.* **200**, 176–186 (2011).
53. Naya, M. *et al.* Evaluation of the genotoxic potential of single-wall carbon nanotubes by using a battery of in vitro and in vivo genotoxicity assays. *Regul. Toxicol. Pharmacol.* **61**, 192–198 (2011).
54. Sargent, L. M., Reynolds, S. H. & Castranova, V. Potential pulmonary effects of engineered carbon nanotubes: in vitro genotoxic effects. *Nanotoxicology* **4**, 396–408 (2010).

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Author contributions

S.T., K.H., K.A., H.T., K.M. and N.S. designed the study, analysed the data and wrote the manuscript. S.T., K.H., K.A., Y.U., M.S., H.H., N.O., K.N., S.K., K.S. and N.N. performed experiments. N.I. and M.O. contributed to discussions. H.K. supervised the project. All Authors discussed the results and commented on the manuscript.

Additional information

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The authors have noticed that Figure 4C was published incorrectly in the original version of this Article. The correct Figure 4C appears below in Figure 1.

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