


# Gene Expression Profiles of the Cochlea and Vestibular Endorgans: Localization and Function of Genes Causing Deafness

Annals of Otolaryngology, Rhinology & Laryngology  
1–43  
© The Author(s) 2015  
Reprints and permissions:  
sagepub.com/journalsPermissions.nav  
DOI: 10.1177/0003489415575549  
aor.sagepub.com  


Shin-ya Nishio, PhD<sup>1,2</sup>, Mitsuru Hattori, PhD<sup>1</sup>, Hideaki Moteki, MD, PhD<sup>1</sup>, Keita Tsukada, MD, PhD<sup>1</sup>, Maiko Miyagawa, MD, PhD<sup>1,2</sup>, Takehiko Naito, MD, PhD<sup>1</sup>, Hidekane Yoshimura, MD, PhD<sup>1</sup>, Yoh-ichiro Iwasa, MD, PhD<sup>1</sup>, Kentaro Mori, MD<sup>1</sup>, Yutaka Shima, MD<sup>1</sup>, Naoko Sakuma, MD<sup>1,3</sup>, and Shin-ichi Usami, MD, PhD<sup>1,2</sup>

## Abstract

**Objectives:** We sought to elucidate the gene expression profiles of the causative genes as well as the localization of the encoded proteins involved in hereditary hearing loss.

**Methods:** Relevant articles (as of September 2014) were searched in PubMed databases, and the gene symbols of the genes reported to be associated with deafness were located on the Hereditary Hearing Loss Homepage using *localization*, *expression*, and *distribution* as keywords.

**Results:** Our review of the literature allowed us to systematize the gene expression profiles for genetic deafness in the inner ear, clarifying the unique functions and specific expression patterns of these genes in the cochlea and vestibular endorgans.

**Conclusions:** The coordinated actions of various encoded molecules are essential for the normal development and maintenance of auditory and vestibular function.

## Keywords

deafness, gene expression, immunocytochemistry, in situ hybridization, localization

## Introduction

Hearing loss is one of the most common and frequently diagnosed sensory disorders worldwide, with 50% to 70% of cases attributable to genetic causes.<sup>1</sup> Hereditary hearing loss demonstrates great heterogeneity. To date, over 80 genes have been identified as causing nonsyndromic hearing loss, and approximately 100 genes are presumed to be involved in hearing loss.<sup>2</sup>

The coordinated actions of various encoded molecules are essential for the normal development and maintenance of auditory processing in the cochlea. The identification of deafness-associated genes has been the most influential factor in the recent extensive advances in our knowledge of the biology of hearing.

In terms of clinical applications, the most remarkable aspect of these advances is that ENT clinicians can now make highly accurate molecular diagnoses through the use of genetic testing, enabling a clearer understanding of the mechanisms involved, more appropriate and precise treatment selection, and greatly improved genetic counseling.

Recent advances in genetic analysis technology using massively parallel DNA sequencing have not only accelerated the exploration of novel genes involved in genetic

hearing loss but have also allowed the identification of mutations in rare causative genes.

However, such rapid progress in gene/mutation identification has, at times, disrupted our understanding of their precise function. It is now necessary to systematize the huge amounts of data available on the expression and localization profiles of causative genes.

The present review highlights the gene expression profiles of the causative genes as well as the localization of the encoded proteins involved in hereditary hearing loss. We conducted a literature search for studies describing in situ hybridization, reporter expression, and immunocytochemistry. In addition, we also reported functional predictions using

<sup>1</sup>Department of Otorhinolaryngology, Shinshu University School of Medicine, Matsumoto, Japan

<sup>2</sup>Department of Hearing Implant Sciences, Shinshu University School of Medicine, Matsumoto, Japan

<sup>3</sup>Department of Otorhinolaryngology and Head and Neck Surgery, Yokohama City University School of Medicine, Yokohama, Japan

## Corresponding Author:

Shin-ichi Usami, Department of Otorhinolaryngology, Shinshu University School of Medicine, 3-1-1 Asahi, Matsumoto 390-8621, Japan.  
Email: usami@shinshu-u.ac.jp

gene ontology analysis and summarized the cochlea and vestibular distribution of functional genes.

## Subjects and Methods

### Literature Search

PubMed database was searched as of September 2014. The gene symbols of the genes reported to be associated with deafness were located on the Hereditary Hearing Loss Homepage,<sup>2</sup> with *localization*, *expression*, and *distribution* used as keywords. After the database search, papers reporting on immunocytochemistry, reporter expression, or in situ hybridization were selected. For each gene linked to deafness, cell-specific localization as well as the methods utilized in the study were annotated and summarized (Tables 1 and 2).

### Computer Analysis and Database Survey

The gene ontology was analyzed using the PANTHER website<sup>3,4</sup> with the symbols for the genes expressed in each type of cochlea cell as well as in the vestibular endorgans (Table 3). We also used the GeneCard website.<sup>5</sup> Gene expression profiles for each human body part were surveyed using the RefEX website.<sup>6</sup> The results of the RefEX search produced by DBCLS are licensed under a Creative Commons Attribution 2.1 Japan License as (CC) BY. The domain structure information was cited from the UniProt website.<sup>7</sup>

## Results and Discussions

A review of the gene expression profiles for hearing loss-associated genes in the inner ear revealed their unique functions and specific expression patterns of the genes in the cochlear and vestibular endorgans (Figures 1, 2, and 3). Each cell-specific gene expression profile is discussed in the following.

### Inner Hair Cells Are Crucial for Sound Transduction

Inner hair cells (IHCs), the actual sensory receptors, play a crucial role in the conversion of mechanical movements to electric signals.<sup>8-14</sup> Hearing ability relies on the rapid gating of the mechano-electrical transduction (MET) channels believed to be located in the tip of the hair cell stereocilia (Figure 2A).<sup>15,16</sup> The genes encoding the MET channel have not yet been identified. Transmembrane channel 1 and 2 genes (*TMC1* and 2) are 2 candidates encoding the MET channels in the stereocilia.<sup>17,18</sup> Furthermore, the MET channels are believed to bind directly to the tip link between the adjacent cilia consisting of the cadherin 23 (*CDH23*) and protocadherin 15 (*PCDH15*). Movement of the stereocilia

mechanically pulls the MET channels and potassium ion ( $K^+$ ) included in the endolymph flow into the IHCs (Figure 3).<sup>9,11,16</sup> This potassium ion incorporation results in depolarization of the IHCs, and the subsequent opening of the voltage-dependent calcium channels.<sup>9,19</sup> Calcium ions ( $Ca^{2+}$ ) act as triggers for the exocytosis of vesicles, which contain the neurotransmitter glutamate.<sup>11,16,19</sup>

The majority (95%) of the fibers of the auditory nerve, which conveys electric stimulation to the brain, arise from IHCs. At the bottom of the IHCs is another unique component known as the ribbon synapse. The ribbon synapse is a type of neuronal synapse consisting of thousands of vesicles. This multivesicular component acts as a large reservoir of the glutamate neurotransmitter. The broad dynamic range of IHC sound sensing is ensured by the rapid, multiple vesicular exocytosis of the ribbon synapse at a level corresponding to the loudness of the sound.<sup>19,20</sup> Otoferlin (encoded by *OTOF*) is known to play an important role in this calcium-dependent exocytosis process, with *otof* knockout mice displaying deafness due to the absence of exocytosis in IHCs.<sup>21</sup>

With regard to gene expression, various genes linked to deafness are expressed in IHCs and in outer hair cells (OHCs; Figure 1). This reflects the unique and important characteristics of IHCs in the sound transduction mechanisms. To examine the gene expression linked to deafness in IHCs, we analyzed the gene ontology and searched a number of databases. As a result, gene expression in the IHCs and OHCs was characterized into 4 groups of genes.

The first group of genes is associated with vesicle transport, neuronal transmission, and calcium-binding functions. This group includes *CABP2* (calcium-binding protein 2), membrane traffic protein *OTOF* (otoferlin), *SLC17A8* (vesicular glutamate transporter 3), and *TBC1D24* (TBC domain-containing RAB-specific GTPase-activating protein 24). Mutations in these genes are known to cause DFNB93, DFNB48, DFNB9, DFNA25, and DFNA65/DFNB86.<sup>22-26</sup> The limited expression of these genes in the sensor cells and neurons reflects the importance of calcium ion ( $Ca^{2+}$ ) and vesicle transport for effective neuronal transmission. *TMPRSS3* (transmembrane protease serine 3) is a serine protease required for epithelial sodium channel (ENaC) maturation, and *TMPRSS3* expression is also limited in IHCs, OHCs, and spiral ganglions.<sup>27</sup>

The second group of genes consists of components of the stereocilia and includes *ACTG1* (actin gamma 1), *CDH23* (cadherin 23), *CIB2* (calcium- and integrin-binding family member 2), *ESPN* (espin), *MYO3A* (myosin IIIa), *MYO7A* (unconventional myosin VIIa), *MYO15A* (unconventional myosin XV), *PCDH15* (Protocadherin 15), *PDZD7* (PDZ domain-containing 7), *RDX* (radixin), *STRC* (stereocilin), *TMC1* (transmembrane channel-like protein 1), *TPRN* (taperin) *TRIOBP* (TRIO and F-actin-binding protein), *USH1C* (harmonin), *USH2A* (usherin), and *WHRN* (whirlin) (Figures 2A, 2B). Mutations in these genes also cause hearing

**Table 1. Summary of Gene Expression Profiles of the Causative Genes and Localization of the Encoded Proteins Involved in Hereditary Hearing Loss in the Cochlea.**

Gene Symbol	Gene Name	Locus	Inner			Outer			External					Inner			Method	Reference	Suspected Function (Description Survey Results)			
			Hair Cell	Hair Cell	Hair Cell	Spiral Ganglion	Pillar Cell	Supporting Cell	Hensen's Cell	Claudius' Cell	External Sulcus Cell	Spiral Prominence	Spiral Ligament	Sria Vasculans	Reissner's Membrane	Inner Dental Cell				Tectorial Membrane	Spiral Limbus	Inner Sulcus Cell
<i>ACTG1</i>	Actin gamma 1	DFNA20/ 26	+	+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	Mouse, guinea pig	IF, EM	28, 100	Cytoskeletal non-muscle actin protein gamma. Localized in F-actin gap region of stereocilia. ESPN also observed in this region.
<i>ADCY1</i>	Adenylylate cyclase type 1	DFNB44	+	+	-	-	-	+	+	+	+	+	+	+	+	+	+	+	Rat	IHC	101, 102	cAMP synthetase required for functional AMPA receptor and synaptic plasticity regulation. ESPN-ADCY1 complex is involved in microvillar elongation.
<i>BDP1</i>	B double prime 1, subunit of RNA polymerase III transcription initiation factor IIIB	DFNB49	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Mouse	IHC	103	Component of U6 RNA transcriptional initiation complex. BDP1 binds to U6 RNA polymerase III promoter.
<i>CABP2</i>	Calcium binding protein 2	DFNB93	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Mouse	IF	104	Calcium-binding protein required for the regulation of $Ca_v1.3$ $Ca^{2+}$ channels.
<i>CCDC50</i>	Coiled-coil domain-containing 50	DFNA44	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Mouse	IF	105	Effector of EGF-mediated cell signaling and required for microtubule-based cytoskeleton organization in pillar cells and the stria vascularis.
<i>CDH23</i>	Cadherin-related 23	DFNB12/ USH1D	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Mouse, guinea pig	ISH, IF, EM	106, 107	Component of tip-link and transient lateral links of stereocilia. Component of pre-synaptic region of IHCs and OHCs.
<i>CEACAM16</i>	Carcinoembryonic antigen-related cell adhesion molecule 16	AD-NSHL	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Mouse	IHC, IF, ISH, X-gal	108, 109	Glycoprotein that interacts with TECTA. CEACAM16 may have a role in connecting stereocilia with the tectorial membrane.
<i>CHD7</i>	Chromodomain helicase DNA-binding protein 7	CHARGE syndrome	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Mouse	X-gal	110	Transcriptional regulator that binds to enhancer elements in the nucleoplasm. CHD7 also functions as a positive regulator of ribosomal RNA biogenesis.
<i>CIB2</i>	Calcium- and integrin-binding family member 2	DFNB48/ USH1J	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Mouse	GFP	23	Component of the tip-link region of stereocilia and interacts with WHRN and MYO7A. CIB2 also functions as an inhibitor of calcium responses.
<i>CLDN14</i>	Claudin 14	DFNB29	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Mouse	IF, X-gal, ISH	60, 111	Component of tight junction complexes at the apex of the cuticular plate.
<i>CLCS</i>	Chloride intracellular channel 5	DFNB02	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Mouse, rat	IF, GFP, EM	112, 113	Component of the chloride channels involved in chloride ion transport. CLCS and RDX may stabilize the filamentous actin core of stereocilia.
<i>CLPP</i>	Caseinolytic mitochondrial matrix peptidase proteolytic subunit	PRLT53	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Mouse	IF	114	Component of a mitochondrial ATP-dependent proteolytic complex, which is required for the stress response signaling pathway.
<i>CLRN1</i>	Clarin 1	USH3	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Mouse	ISH, IF, EM	115-119	Expressed in photoreceptors and hair cell synapses and may be a component of pre-synaptic complexes in sensor cells.
<i>COCH</i>	Cochlin	DFNA9	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Mouse, human	ISH, IHC, IF	119, 120	Extracellular matrix protein. COCH may contribute to the innate immune response against bacterial infection.
<i>COL2A1</i>	Collagen, type II, alpha 1	STL1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Mouse	IF, EM	121	Structural components of the extracellular matrix of chondrocytes and the tectorial membrane.

(continued)

Gene Symbol	Gene Name	Gene Locus	Inner Hair Cell	Outer Hair Cell	Spiral Ganglion Cell	Pillar Cell	Supporting Cell	Hensen's Cell	Claudius' Cell	External Sulcus Cell	Spiral Prominence	Spiral Ligament	Stria Vascularis	Reissner's Membrane	Inter-Dental Cell	Tectorial Membrane	Spiral Limbus	Inner Sulcus Cell	Other/Detail	Model	Method	Reference	Suspected Function (Description Survey Results) <sup>a</sup>
COL4A3	Collagen, type IV, alpha 3	Alport syndrome	-	-	-	-	-	-	-	-	-	+	-	+	-	-	+	-	Basilar membrane, Schwann cells, spiral lamina	Human	IHC	86	Structural components of the extracellular matrix of the basilar membrane and spiral ligament.
COL4A4	Collagen, type IV, alpha 4	Alport syndrome	-	-	-	-	-	-	-	+	+	+	-	-	-	-	+	+	Basilar membrane	Mouse	IHC	122	Structural components of the extracellular matrix of the basilar membrane and spiral ligament.
COL4A5	Collagen, type IV, alpha 5	Alport syndrome	-	-	-	-	-	-	-	-	-	+	-	+	-	-	+	-	Basilar lamina	Human	IHC	86	Structural components of the extracellular matrix of the basilar membrane and stria vascularis portion of the spiral ligament.
COL4A6	Collagen, type IV, alpha 6	X linked-NSHL	NA	NA	+	NA	NA	NA	NA	NA	NA	+	+	NA	NA	NA	NA	NA	Basilar lamina membrane	Mouse	IHC	123	Structural components of the extracellular matrix of the basilar membrane and stria vascularis portion of the spiral ligament.
COL9A1	Collagen, type IX, alpha 1	STL4	-	-	-	-	-	-	-	-	-	+	-	-	-	+	+	-	Basilar membrane, spiral lamina	Rat, mouse	IF, EM	82, 124	Structural components of the extracellular matrix of the tectorial membrane, covalently cross-linked to type II collagen.
COL9A2	Collagen, type IX, alpha 2	STL5	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Basilar membrane	Mouse	IHC	NA	Cartilage-specific fibril-associated collagen. The function of this protein in the inner ear is not well known.
COL9A3	Collagen, type IX, alpha 3	AR-NSHL	-	-	-	-	-	-	-	-	-	+	-	-	-	+	+	-	Basilar membrane, spiral lamina	Rat, mouse	IF, EM	82, 124	Structural components of the extracellular matrix of the tectorial membrane, covalently cross-linked to type II collagen.
COL11A1	Collagen, type XI, alpha 1	STL2	-	-	-	-	-	+	+	-	-	+	+	-	-	+	-	-	Greater epithelial ridge, lateral wall	Mouse	ISH	83	Structural components of the extracellular matrix of the tectorial membrane.
COL11A2	Collagen, type XI, alpha 2	DFNA13/DFNB33/STL3	-	-	-	-	-	+	+	-	-	+	+	-	-	-	+	-	Spiral limbus region	Mouse	ISH	84	Structural components of the extracellular matrix of the tectorial membrane.
CRYM	Crystallin, mu	AD-NSHL	-	-	-	NA	NA	NA	NA	NA	NA	+	-	-	-	-	-	NA	Mouse	IF	125	Directly binds to thyroid hormone (T3) with high affinity in the presence of NADPH, CRYM and NADPH complex transport T3 into the nucleus and activate T3-dependent transcription.	
DFNA5	Deafness, autosomal dominant 5	DFNA5	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Inversely correlated with estrogen receptor gene expression, p53 induces DFNA5 expression in response to DNA damage.
DFNB59	Pejvakit	DFNB59	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	Cochlea nuclei, kinocillum	Mouse	IF, ISH	126, 127	DFNB59 is markedly similar to DFNA5 and is important to the functioning of both hair cells and neurons. Pjvk mutation causes ANSD.
DIAPH1	Diaphanous, drosophila, homolog of, 1	DFNA1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Actin polymerization protein in hair cells of the inner ear. <i>MITF</i> regulates <i>DIAPH1</i> expression.
DIAPH3	Diaphanous, drosophila, homolog of, 3	AD-ANSD1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Remodelling of cytoskeleton and may act in the elongation process of the non-branched actin filaments. <i>DIAPH3</i> directly binds to and stabilizes microtubules.

(continued)

**Table 1. (continued)**

Gene Symbol	Gene Name	Locus	Inner			Outer			Internal			External			Inter			Inner			Reference	Method	Suspected Function (Description Survey Results)	
			Hair Cell	Hair Cell	Hair Cell	Spiral Ganglion	Pillar Cell	Supporting Cell	Hensen's Cell	Claudius' Cell	External Sulcus Cell	Spiral Prominence	Spiral Ligament	Spiria Vascularis	Reissner's Membrane	Dental Cell	Tectorial Membrane	Spiral Limbus	Sulcus Cell	Other/Detail				Model
DSPP	Desitin sialoprophosphoprotein	DFNA39	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Small integrin-binding ligand N-linked glycoprotein family of secreted phosphoproteins, which are involved in bone mineralization.	
EDN3	Endothelin 3	WS4	+	+	-	-	+	+	-	-	-	-	-	-	-	-	-	-	+	Boettcher's cells	Guinea pig	IF	128	EDN3 directly binds to EDNRB. EDN3-EDNRB complex acts in the normal differentiation and development of melanocytes. EDNRB required for normal differentiation and development of melanocytes. SOX10 enhances the expression of EDNRB. EDNRB inhibits the Na-K transporter and activates G protein-coupled inwardly rectifying potassium channels (Kir-3).
EDNRB	Endothelin receptor type B	WS4	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	EDNRB has a role as a GTPase-activating protein for small GTPases.
ELMOD3	ELMO domain-containing protein 3	DFNB88	+	+	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	Rat	IF	129	ELMO domain-containing protein. ELMOD3 has a role as a GTPase-activating protein for small GTPases.
EP58	Epidermal growth factor receptor kinase substrate 8	AR-NSHL	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Mouse	IF	130	MYO15A-WHRN-EP58 complex has an essential role in stereocilia elongation.
ESPN	Espin	DFNB36	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Mouse	IHC	131	Directly binds to actin filaments. Actin-bundling activities of ESPN are not inhibited by calcium ions.	
ESRR8	Estrogen-related receptor beta	DFNB35	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	Basilar membrane	Mouse, rat	ISH, IF	132	Estrogen-related receptor acts in epigenetic transcriptional induction at pluripotency loci during somatic cell reprogramming.
EYA1	Eyes absent homolog 1	BOR1	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	Eye	Mouse	ISH	133	Involved in the PAX-EYA-SIX regulatory pathway. EYA1 act as a phosphorylation-dependent transcription factor or transcriptional modulator.
EYA4	Eyes absent homolog 4	DFNA10	NA	NA	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	Bony cochlea capsule	Rat	ISH	134	Involved in innate immune response regulation by modulating the phosphorylation of signal transducers for intracellular pathogens.
FOXL1	Forkhead box L1	PDS	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Transcriptional enhancer acts in the SLC26A4 promoter. Digenic mutations of FOXL1 and SLC26A4 cause Pendred syndrome.
GIPC3	GIPC PDZ domain-containing family, member 3	DFNB15/72/95	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Stereocilia	Mouse	IF	135	Required for postnatal maturation of hair bundles and also has potassium channel activity. GIPC3 is required for long-term survival of hair cells and spiral ganglion.
GJB2	Gap junction protein, beta 2	DFNA3A/DFNB1A	-	-	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	Rat	IF	136	Gap junction protein for potassium recirculation and the transport of other metabolites.
GJB3	Gap junction protein, beta 3	DFNA2B/DFNB91	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Auditory nerve fiber	Mouse	ISH	137	Gap junction protein gated by voltage. GJB3 closes at low pH when exposed to long-chain alkanols.
GJB6	Gap junction protein, beta 6	DFNA3B/DFNB1B	-	-	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	-	Rat	IF	136	Involved in the generation of the endocochlear potential (EP). GJB6 is also required for auditory hair cell survival after the onset of hearing.
GPR98	G protein-coupled receptor 98	USH2C	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Ankle-link	Mouse	IF, EM	138	Large calcium-binding protein expressed in the ankle link of stereocilia. VGLR1 directly binds to PDZD7. VGLR1 also expressed in the synaptic region of hair cells.

(continued)

Table 1. (continued)

Gene Symbol	Gene Name	Locus	Inner Hair Cell	Outer Hair Cell	Spiral Ganglion	Pillar Cell	Supporting Cell	Hensen's Cell	Claudius' Cell	External Sulcus Cell	Spiral Prominence	Spiral Ligament	Spiral Vascubris	Reissner's Membrane	Inter-Dental Cell	Tectorial Membrane	Spiral Limbus	Inner Sulcus Cell	Other/Detail	Model	Method	Reference	Suspected Function (Description Survey Results)
GF5M2	G-protein signaling modulator 2	DFNB82	+	+	-	+	+	+	-	NA	NA	NA	NA	NA	NA	NA	NA	NA	Apical surface of hair and supporting cells	Mouse	IF	52	G protein activation modulator, involved in embryonic neuroblast asymmetric cell division.
GRHL2	Grainyhead-like 2	DFNA28	-	-	-	-	-	-	-	-	-	-	+	+	-	NA	NA	-	Basilar membrane	Mouse	ISH	139	Grainyhead-like transcription factor family protein. GRHL2 has an essential role in epithelial morphogenesis and epidermal development.
GRXCR1	Glutaredoxin, cysteine-rich 1	DFNB25	+	+	NA	NA	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Stereocilia	Mouse	IF, GFP	140	The glutaredoxin protein and the catalytic domain of this protein are predicted to be involved in the reversible S-glutathionylation.
GRXCR2	Glutaredoxin, cysteine-rich 2	DFNB101	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Stereocilia bundle	NA	IF	141	Expressed in the stereocilia and has an essential role in maintaining stereocilia bundles. GRXCR2 has a glutaredoxin domain but lacks catalytic activity.
HARS2	Histidyl-tRNA synthetase 2, mitochondrial	PRLTS2	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Histidyl tRNA synthetase, a highly conserved tRNA synthetase for protein synthesis in mitochondria
HGF	Hepatocyte growth factor	DFNB39	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	HGF/MET signaling induces rearrangement of the actin cytoskeleton. Mutations in HGF cause hearing loss due to OHC degeneration and cochlea amplification loss.
HSD17B4	Hydroxysteroid (17-beta) dehydrogenase 4	PRLTS1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Enzyme involved in peroxisomal fatty acid beta-oxidation.
ILDR1	Immunoglobulin-like domain-containing receptor 1	DFNB42	+	+	-	+	+	+	-	-	-	-	-	-	-	-	-	-	-	Mouse	ISH	142	Involved in lipoprotein transport
KARS	Lysyl-tRNA synthetase	DFNB89	+	+	+	+	+	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	Type II ganglion	Mouse	IF	143	Lysyl-tRNA synthetase, which acts in the aminoacylation of tRNA-Lys in the cytoplasm and mitochondria.
KCNE1	Potassium voltage-gated channel, KQT-related family, member 1	Jervell and Lange-Nielsen syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	+	+	+	+	+	+	Heart, kidney	Mouse	IF	144	Voltage-gated potassium channel. KQT-like subfamily protein, KCNE1 is expressed on the apical surface of the marginal cells in the stria vascularis and is involved in EP generation by the endolymph, particularly in its high potassium ion concentration.
KCNJ10	Potassium inwardly rectifying channel, subfamily J, member 10	PDS	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	+	+	+	+	+	+	Mouse	ISH, IF	145	Inwardly rectifying potassium channel. The sensitivities of KCNJ10 are controlled by intracellular pH but not extracellular pH.	
KCNQ1	Potassium voltage-gated channel, KQT-like subfamily, member 1	Jervell and Lange-Nielsen syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	+	+	+	+	+	+	Heart, kidney	Mouse	IF	144	Voltage-gated potassium channel, KQT-like subfamily protein. KCNQ1 has an important role in ion homeostasis of the endolymph, particularly in its high potassium ion concentration.
KCNQ4	Potassium voltage-gated channel, KQT-like subfamily, member 4	DFNA2A	-	+	NA	NA	-	NA	NA	NA	-	NA	-	NA	NA	NA	-	-	Mouse	ISH, IF	146, 147	Member of the voltage-gated potassium channel gene family for OHC. Alternative transcriptional variants without Ca <sup>2+</sup> binding domain for auto-regulation are expressed at the basal turn of the mouse cochlea.	

(continued)

**Table 1. (continued)**

Gene Symbol	Gene Name	Locus	Inner Hair Cell	Outer Hair Cell	Spiral Ganglion	Pillar Cell	Supporting Cell	Hensen's Cell	Claudius' Cell	External Sulcus Cell	Spiral Prominence	Spiral Ligament	Spiral Vasculis	Reissner's Membrane	Inner Dental Cell	Tectorial Membrane	Spiral Limbus	Inner Sulcus Cell	Other/Detail	Model	Method	Reference	Suspected Function (Description Survey Results) <sup>3</sup>
LARS2	Leucyl-tRNA synthetase 2, mitochondrial	PRLT54	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Leucyl aminoacyl-tRNA synthetases containing a mitochondrial-targeting sequence.
LHFPLS	Lipoma HMGIC fusion partner-like 5	DFNB66/67	+	+	NA	+	+	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	Tip of stereocilia	Mouse	IF	148	Tetraspan membrane protein of hair cell stereocilia and involved in the tip link complex.
LOXHD1	Lipoxygenase homology domains 1	DFNB77	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Stereocilia	Mouse	ISH, IF	43	Involved in the regulation of stereocilia elongation. Mutation of <i>LOXHD1</i> causes "fused stereocilia" and "membrane ruffling" at the apical surface of hair cells.
LRTOMT1/COMT2	Leucine-rich transmembrane and O-methyltransferase domain-containing	DFNB63	+	+	NA	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Mouse	IF	149	<i>LRTOMT</i> encodes 2 different reading frame proteins. Its expression is restricted to hair cells of the cochlea and vestibule, and may be involved in the maintenance of stereocilia.	
MARVELD2	Tricellulin	DFNB49	+	+	NA	NA	NA	NA	NA	NA	NA	NA	+	NA	NA	NA	NA	NA	Mouse	IF	150	Tight junction protein that contributes to the structure and function of tricellular contacts between neighboring cells.	
MIR96	Micro-RNA 96	DFNA50	+	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Mouse	ISH	151	Non-coding microRNA that down-regulates <i>MITF</i> , <i>SLC26A5</i> , and <i>PTRFQ</i> gene expression. <i>MIR96</i> expression is restricted to IHCs, OHCs, and spiral ganglions.	
MITE	Microphthalmia-associated transcription factor	WS2A	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Important for the development of various types of neural crest-derived melanocytes. <i>SOX10</i> and <i>PAX3</i> directly regulate <i>MITF</i> gene expression.
MSRB3	Methionine sulfoxide reductase B3	DFNB74	+	+	+	NA	+	NA	NA	NA	NA	NA	-	NA	NA	NA	NA	NA	Mouse	IF	152	Involved in the oxo-reduction of oxidized methionine residues. <i>MSRB3</i> is required for the repair of oxidatively damaged proteins.	
MYH9	Myosin, heavy chain 9, non-muscle	DFNA17	-	+	-	+	+	+	+	+	+	+	-	+	-	-	+	-	Rat	IHC	153	<i>MYH9</i> encodes a non-muscle myosin and may be involved in actin degeneration and reorganization of the actomyosin network.	
MYH14	Myosin, heavy chain 14, non-muscle	DFNA4	+	+	-	+	+	+	+	+	+	+	+	-	-	-	-	-	Mouse	IHC	154	ATP-dependent molecular motors that interact with cytoskeletal actin. <i>MYH14</i> is involved in the regulation of cytokinesis, cell motility, and cell polarity.	
MYO3A	Unconventional myosin IIIA	DFNB30	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Rat	IF	155	Myosin IIIA expression is restricted to the retina and IHCs and to OHCs of the cochlea. <i>MYO3A</i> is localized in the tip density region of stereocilia and acts in the maintenance of stereocilia morphology.	
MYO6	Unconventional myosin VI	DFNA22/DFNB37	+	+	-	NA	NA	NA	NA	-	-	-	-	-	-	-	-	-	Mouse, rabbit	IF	156	<i>MYO6</i> is expressed in the cuticular plate region of IHCs and OHCs. <i>MYO6</i> is involved in stereocilia formation and may have an important role in anchoring stereocilia.	
MYO7A	Unconventional myosin VIIA	DFNA11/DFNB2/USH1B	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Mouse	IF	157	<i>MYO7A</i> encodes a component of the USH complex (including <i>CDH23</i> , <i>SANS</i> , <i>USH1C</i> , and <i>MYO7A</i> ) in the tip links of stereocilia.	

(continued)

Table I. (continued)

Gene Symbol	Gene Name	Locus	Inner Hair Cell	Outer Hair Cell	Spiral Ganglion	Pillar Cell	Supporting Cell	Hensen's Cell	Claudius' Cell	External Sulcus Cell	Spiral Prominence	Spiral Ligament	Spiral Vascularis	Reissner's Membrane	Inter-Dental Cell	Tectorial Membrane	Spiral Limbus	Inner Sulcus Cell	Other/Detail	Model	Method	Reference	Suspected Function (Description Survey Results) <sup>a</sup>
<i>MYO15A</i>	Unconventional myosin XVa	DFNB3	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Mouse, rat, IF guinea pig	IF	158	<i>MYO15A</i> directly binds to <i>WHRN</i> to form the <i>MYO15A-WHRN</i> complex. This complex is essential for stereocilia elongation.
<i>NDP</i>	Norrie disease	Norrie disease	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	Capillary plexus between the Corti and the spiral ganglion	Mouse	AP	159	Cystine knot growth factor family protein (norm). <i>NDP</i> induces the <i>FZD4</i> - and <i>LRP</i> -dependent activation of the classic Wnt signaling pathway.
<i>OTOA</i>	Otoancorin	DFNB22	+	-	-	-	-	-	-	-	-	-	-	-	+	-	+	Apical surface of the spiral limbus	Mouse	IF	160	Non-collagenous glycoproteins of the acellular gels of the inner ear. <i>OTOA</i> stabilizes the conformation of tectorial members.	
<i>OTOF</i>	Otoferlin	DFNB9	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	Ribbon synapse	Mouse	IF	21	Correlated with afferent synaptogenesis and involved in the late steps of synaptic vesicle exocytosis. <i>OTOF</i> may act as the major $Ca^{2+}$ sensor for IHC ribbon synapses.	
<i>OTOG</i>	Otogelin	AR-NSHL	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	Basilar membrane, otoconial membrane	Mouse	X-gal, IF	161	N-glycosylated protein comprising the acellular membranes of the tectorial membrane.	
<i>OTOG</i>	Otogelin-like protein	DFNB84	+	+	+	+	-	-	+	-	+	-	-	-	-	+	-	-	Mouse	IF	88	Acellular membranes of the cochlea and vestibular system.	
<i>P2RX2</i>	Purinergic receptor P2X <sub>2</sub> , ligand-gated ion channel, 2	DFNA41	+	+	+	+	-	-	-	-	+	-	-	+	-	-	+	-	Mouse	IF	162	ATP activates <i>P2RX2</i> channels to modify OHC electromotility. Extracellular $Ca^{2+}$ is required for the effective gating.	
<i>PAX3</i>	Paired box 3	WS1/WS3	-	-	+	-	-	-	-	-	-	-	+	-	-	-	-	-	Glial cells	Mouse	X-gal	163	<i>SOX10</i> and <i>PAX3</i> strongly activate <i>MITF</i> gene expression, which is required for the differentiation and development of melanocytes.
<i>PCDH15</i>	Protocadherin 15	DFNB23/USH1F	+	+	NA	NA	-	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Tip-link	Mouse	IF	164	Component of the tip links and transient lateral links of stereocilia. <i>PCDH15</i> and <i>CDH23</i> directly bind to form the tip link.	
<i>PDZD7</i>	PDZ domain-containing 7	USH2C	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Ankle-link	Mouse	IF	165	<i>PDZD7</i> encodes a component of the USH complex and is expressed in stereocilia. <i>PDZD7</i> interacts with <i>SWAS</i> ( <i>USH1G</i> ), <i>GRR98</i> , and <i>USH2A</i> .	
<i>PNPT1</i>	Polyribonucleotide nucleotidyltransferase 1, mitochondrial	DFNB70	+	+	+	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	-	Mouse	IF	166	Subunit of the exosome complex. <i>PNPT1</i> has 3-prime-to-5-prime exonuclease activity and is involved in the import of RNAs to the mitochondria.	
<i>POLR1C</i>	Polymerase I, RNA, subunit C	TCS3	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	-	NA	NA	NA	NA	16-kDa subunit of mouse RNA polymerase I complex.
<i>POLR1D</i>	Polymerase I, RNA, subunit D	TCS2	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	-	Mouse	ISH	167	Transcription factor restrictedly expressed in the spiral ligament fibrocytes. <i>POLR1D</i> may have a role in potassium ion homeostasis.	
<i>POU3F4</i>	POU domain, class 3, transcription factor 4	DFNX2 (DFN3)	-	-	-	-	-	-	-	-	+	-	-	+	-	-	-	-	-	Mouse	IF	168	POU family of transcription factors and is involved in the maintenance of inner ear hair cells. <i>POU3F4</i> activates <i>MYO7A</i> gene expression.

(continued)



**Table 1. (continued)**

Gene Symbol	Gene Name	Locus	Inner			Outer			External				Internal				Method	Reference	Suspected Function (Description Survey Results) <sup>a</sup>				
			Hair Cell	Hair Cell	Supporting Cell	Hensen's Cell	Claudius' Cell	External Sulcus Cell	Spiral Prominence	Spiral Ligament	Sria Vascularis	Reissner's Membrane	Dental Cell	Tectorial Membrane	Spiral Limbus	Sulcus Cell				Inner Cell	Other/Detail	Model	
<i>PP5I</i>	Phosphoribosyl pyrophosphatase I	DRX1 (DFN2)	+	+	-	-	+	-	-	-	-	-	-	-	-	-	-	Greater epithelial ridge	Mouse	ISH	169	Phosphoribosylpyrophosphate synthetase that catalyzes the phosphorylation of ribose 5-phosphate to 5-phosphoribosyl-1-pyrophosphate. <i>PP5I</i> is necessary for the salvage pathways of purine and pyrimidine biosynthesis.	
<i>PTPRQ</i>	Protein tyrosine phosphatase receptor type, Q	DFNB84	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	Basal region of stereocilia	Rat, chick, mouse, frog	IF, EM	170, 171	Protein tyrosine phosphatase receptor protein and has an important role in shaft connector formation in hair bundles. <i>PTPRQ</i> is necessary for the long-term survival of high-frequency auditory hair cells.	
<i>RDX</i>	Radixin	DFNB24	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	Stereocilia	Mouse	IF	36	Cytoskeletal protein that may be involved in anchoring actin to the plasma membrane of cochlear stereocilia.	
<i>SANS</i>	Scaffold-containing inlyrin repeats and SAM domain	USH1G	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Stereocilia	Rat, guinea pig	IF, GFP	172	<i>SANS</i> interacts with <i>USH1C</i> and <i>MYO7A</i> directly. This protein is involved in hair bundle cohesion.		
<i>SEMA3E</i>	Sema domain, immunoglobulin domain, short basic domain, secreted, 3E	CHARGE syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Encoding high-affinity proteins to receptor plexin D1. <i>SEMA3E</i> is a critical determinant of synaptic specificity in sensory motor circuits in mice.	
<i>SERPIN6</i>	Serpin peptidase inhibitor, clade B, member 6	ARNSHL	+	+	-	-	+	+	+	+	+	+	+	+	+	+	+	Lateral wall	Mouse	IF, GFP	173	<i>SERPIN6</i> encodes serpin peptidase inhibitor protein. <i>SERPIN6</i> has no effect on cochlear development but is required for cochlea homeostasis.	
<i>SIX1</i>	SIX homeobox 1	DFNA23/BOS3	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	Greater epithelial ridge	Mouse	X-gal	174	<i>PAX-EYA-SIX</i> regulatory pathway ( <i>PAX3</i> and <i>EYA2</i> are involved in cochlea hair cells). <i>SIX1</i> has phosphorylation-dependent transcription modulation activity.	
<i>SIX5</i>	SIX homeobox 5	BOS2	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	<i>SIX5</i> encodes an activator of <i>IGFBP5</i> expression, and a mutation in this gene causes Myotonic Dystrophy or BOK syndrome.
<i>SLC17A8</i>	Vesicular glutamate transporter 3	DFNA25	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Mouse, rat	IF	175, 176	Glutamate transporter protein acts in glutamate release from hair cells at the first synapse of the auditory pathway.	
<i>SLC26A4</i>	Pendrin	DFNB4/PDS	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Spindle-shaped cells	Mouse	ISH, IF	177, 178	Acts as a chloride, bicarbonate, and iodide ion transporter in the spiral prominence. Pendrin also contributes to pH homeostasis and mineralization in the organ of Corti and vestibular organs.	
<i>SLC26A5</i>	Prestin	DFNB61	-	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Rat	IF	179	Prestin is the motor protein of cochlea OHCs and is involved in the sound amplification process.	
<i>SMACD/ABLO</i>	Second mitochondrial-derived activator of caspase	DFNA64	+	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Greater epithelial ridge	Mouse	IF	180	Regulates programmed cell death (apoptosis) in specific situations or tissues.	
<i>SMPX</i>	Small muscle protein, X-linked	DFNX4 (DFN6)	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	Boettcher's cells, root cells	Mouse	IF	181	<i>SMPX</i> encodes small muscle proteins that may protect the hearing organs from mechanical stress.	
<i>SNAIL2</i>	Snail family zinc finger 2	WS2D	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	Small family of zinc finger transcription factor 2 triggers epithelial-mesenchymal transitions and developmental processes.

(continued)



**Table 1. (continued)**

Gene Symbol	Gene Name	Locus	Inner Hair Cell		Outer Hair Cell		Spiral Ganglion	Pillar Cell	Supporting Cell	Hensen's Cell	Claudius' Cell	External Sulcus Cell		Spiral Prominence	Spiral Ligament	Stria Vascularis	Reissner's Membrane	Inter-Dental Cell		Tectorial Membrane	Spiral Limbus	Inner Sulcus Cell	Other/Detail	Model	Method	Reference	Suspected Function (Description Survey Results) <sup>a</sup>
			+	-	+	-						+	-					+	-								
TRIOBP	TRIO and F-actin-binding protein	DFNB28	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Mouse	IF, EM	192	Encodes the actin-binding protein and is essential for the development of rootlets that provide durable flexibility and mechanical rigidity to the stereocilia bundles.	
TSPEAR	Thrombospondin-type laminin G domain and EAR repeats	DFNB98	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Mouse	IF	193	Thrombospondin-type laminin G domain and EAR repeats-containing protein. TSPEAR expressed in the base of the hair bundles of inner and outer hair cells.	
USH1C	Harmonin	DFNB18/ USH1C	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	Mouse	IF	194, 195	Scaffold protein for CDH23, SANS, and MYO7A complexes in the tip link of stereocilia.	
USH2A	Usherin	USH2A	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Mouse	IF	196	Scaffold protein and forms a complex with USH1C and VLGR1. Usherin is present in the ankle links in stereocilia.	
WFS1	Wolfgramin	DFNA6/ 14/38	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Mouse	IF, BH	197	Encodes the endoplasmic reticulum protein and may act as an ER calcium channel or regulator of ER calcium channel activity. WFS1 may be involved in ER stress responses.	
WHRN	Whirlin	DFNB31/ USH2D	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	Mouse	IF	198	Scaffolding protein that facilitates synaptic transmission in the central nervous system. SANS, EPS8, and MYO15A are colocalized with WHRN in the tip of stereocilia.	

Abbreviations: BOR, branchiootorenal syndrome; EM, electron microscopy; IF, immunofluorescence; IHC, immunohistochemistry; ISH, in situ hybridization; NSHL, nonsyndromic hearing loss; PRLTS, Perrault syndrome; STL, Stickler syndrome; TCS: Treacher Collins syndrome; USH, Usher syndrome; WS, Waardenburg Syndrome; +, expressed; -, not expressed; NA, not applicable.

<sup>a</sup>Suspected functions were summarized from the description in the OMIM database (<http://omim.org>) and each reference.

**Table 2.** Summary of Gene Expression Profiles of the Causative Genes and Localization of the Encoded Proteins Involved in Hereditary Hearing Loss in the Vestibular Endorgans.

Gene Symbol	Gene Name	Locus	Semicircular Canal	Utricle	Sacculle	Hair Cell Type I	Hair Cell Type II	Support Cell	Transitional Epithelium	Model	Method	Reference
<i>ACTG1</i>	Actin gamma 1	DFNA20/26	NA	NA	NA	+	+	+	NA	Mouse, guinea pig	IF	199
<i>ADCY1</i>	Adenylate cyclase type 1	DFNB44	NA	+	NA	+	+	+	NA	Mouse	IF	102
<i>BDPI</i>	B double prime 1, subunit Of RNA polymerase III transcription initiation factor IIIB	DFNB49	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>CABP2</i>	Calcium-binding protein 2	DFNB93	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>CCDC50</i>	Coiled-coil domain-containing 50	DFNA44	NA	+	+	+	+	+	NA	Mouse	IF	105
<i>CDH23</i>	Cadherin-related 23	DFNB12/USH1D	+	+	+	+	+	+	+	Mouse	ISH	200, 201
<i>CEACAM16</i>	Carcinoembryonic antigen-related cell adhesion molecule 16	AD-NSHL	NA	NA	+	NA	NA	NA	NA	Mouse	ISH	202
<i>CHD7</i>	Chromodomain helicase DNA-binding protein 7	CHARGE syndrome	-	+	+	-	-	-	-	Mouse	IF	203
<i>CIB2</i>	Calcium- and integrin-binding family member 2	DFNB48/USH1J	NA	NA	NA	+	+	+	NA	Mouse	IF	204
<i>CLDN14</i>	Claudin 14	DFNB29	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>CLIC5</i>	Chloride intracellular channel 5	DFNB102	NA	NA	NA	+	+	NA	NA	Mouse	IF	113
<i>CLPP</i>	Caseolytic mitochondrial matrix peptidase proteolytic subunit	Perrault syndrome 3	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>CLRN1</i>	Clarin 1	USH3	NA	NA	+	+	+	NA	NA	Mouse	ISH	115
<i>COCH</i>	Cochlin	DFNA9	+	NA	NA	-	-	-	-	Mouse, human	IF, ISH, IHC	119, 205
<i>COL2A1</i>	Collagen, type II, alpha 1	Stickler syndrome	+	+	NA	NA	NA	NA	NA	Mouse	IF	81
<i>COL4A3</i>	Collagen, type IV, alpha 3 (Goodpasture antigen)	Alport syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>COL4A4</i>	Collagen, type IV, alpha 4	Alport syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>COL4A5</i>	Collagen, type IV, alpha 5	Alport syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>COL4A6</i>	Collagen, type IV, alpha 6	X linked-NSHL	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>COL9A1</i>	Collagen, type IX, alpha 1	Stickler syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>COL9A2</i>	Collagen, type IX, alpha 2	Stickler syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>COL11A1</i>	Collagen, type XI, alpha 1	Stickler syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>COL11A2</i>	Collagen, type XI, alpha 2	DFNA13/DFNB53/ Stickler syndrome	+	+	+	NA	NA	NA	NA	Mouse	ISH	84
<i>CRYM</i>	Crystallin, mu	AD-NSHL	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>DFNA5</i>	Deafness, autosomal dominant 5	DFNA5	+	NA	NA	NA	NA	NA	NA	Zebrafish	ISH	206
<i>DFNB59</i>	Pejvakian	DFNB59	+	+	+	NA	NA	NA	NA	Mouse	IF	126

(continued)

**Table 2. (continued)**

Gene Symbol	Gene Name	Locus	Semicircular Canal	Utricle	Sacculle	Hair Cell Type I	Hair Cell Type II	Support Cell	Transitional Epithelium	Model	Method	Reference
DIAPH1	Diaphanous, drosophila, homolog of, 1	DFNA1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
DIAPH3	Diaphanous, drosophila, homolog of, 3	AD-ANSD1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
DSPP	Dentin sialophosphoprotein	DFNA39	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EDN3	Endothelin 3	Waardenburg Syndrome type IV	+	+	+	-	-	-	-	Mouse	IHC	207
EDNRB	Endothelin receptor type B	Waardenburg Syndrome type IV	+	+	+	-	-	-	-	Mouse	IHC	207
ELMOD3	ELMO domain-containing protein 3	DFNB88	NA	+	+	+	+	NA	NA	Mouse	IF	129
EPS8	Epidermal growth factor receptor kinase substrate 8	AR-NSHL	NA	NA	NA	+	+	-	-	Rat, Mouse	IF	130
ESPN	Espn	DFNB36	+	+	+	+	+	NA	NA	Rat	IHC	131
ESRRB	Estrogen-related receptor beta	DFNB35	-	+	-	-	-	-	-	Mouse	ISH	132
EYA1	Eyes absent homolog 1	BOR syndrome 1	-	+	+	+	+	-	+	Mouse	ISH	208
EYA4	Eyes absent homolog 4	DFNA10	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
FOX11	Forkhead box 11	Pendred syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
GIPC3	GIPC PDZ domain-containing family, member 3	DFNB15/72/95	NA	NA	NA	+	+	NA	NA	Mouse	IF	135
GJB2	Gap junction protein, beta 2	DFNA3A/DFNB1A	NA	+	+	-	-	+	-	Mouse	IF	209
GJB3	Gap junction protein, beta 3	DFNA2B/DFNB91	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
GJB6	Gap junction protein, beta 6	DFNA3B/DFNB1B	NA	+	+	-	-	+	-	Mouse	IF	209
GPR98 (VLGR1)	G protein-coupled receptor 98	USH2C	NA	+	NA	+	+	NA	NA	Mouse	IF	138
GPSM2	G-protein signaling modulator 2	DFNB82	NA	+	+	+	+	+	-	Mouse	IF	52
GRHL2	Grainyhead-like 2	DFNA28	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
GRXCR1	Glutaredoxin, cysteine RIHC 1	DFNB25	NA	+	NA	+	+	-	+	Mouse	IHC	140
GRXCR2	Glutaredoxin, cysteine RIHC 2	DFNB101	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
HARS2	Histidyl-tRNA synthetase 2, mitochondrial	Perrault syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
HGF	Hepatocyte growth factor	DFNB39	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
HSD17B4	Hydroxysteroid (17-beta) dehydrogenase 4	Perrault syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
ILDR1	Immunoglobulin-like domain-containing receptor 1	DFNB42	NA	NA	NA	+	+	+	NA	Mouse	ISH	142
ILDRI	Immunoglobulin-like domain-containing receptor 1	DFNB42	NA	NA	+	NA	NA	NA	NA	Mouse	IHC	210
KARS	Lysyl-tRNA synthetase	DFNB89	-	-	-	+	+	+	-	Mouse	IF	143

(continued)

Table 2. (continued)

Gene Symbol	Gene Name	Locus	Semicircular Canal	Utricle	Sacculle	Hair Cell Type I	Hair Cell Type II	Support Cell	Transitional Epithelium	Model	Method	Reference
KCNE1	Potassium voltage-gated channel, Isk-related family, member 1	Jervell and Lange-Nielsen syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
KCNJ10	Potassium inwardly rectifying channel, subfamily J, member 10	Pendred syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
KCNQ1	Potassium voltage-gated channel, KQT-like subfamily, member 1	Jervell and Lange-Nielsen syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
KCNQ4	Potassium voltage-gated channel, KQT-like subfamily, member 4	DFNA2A	-	+	+	+	+	+	NA	Mouse	IHC	211
LARS2	Leucyl-tRNA synthetase 2, mitochondrial	Perrault syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
LHFPL5 (TMHS)	Lipoma HMGIC fusion partner-like 5	DFNB66/67	NA	+	+	+	+	+	NA	Mouse	IHC	14
LOXHD1	Lipoxygenase homology domains 1	DFNB77	NA	NA	+	+	+	-	NA	Mouse	ISH, IF	212
LRTOMT/COMT2	Leucine RIHC Transmembrane and O-methyltransferase domain-containing	DFNB63	NA	+	+	+	+	+	NA	Mouse	ISH, IHC	149
MARVELD2	Tricellulin	DFNB49	NA	+	NA	+	+	+	NA	Mouse	IF	53, 150
MIR96	Micro-RNA 96	DFNA50	NA	NA	NA	+	+	-	NA	Mouse	ISH	213
MITF	Microphthalmia-associated transcription factor	Waardenburg Syndrome type IIA	-	-	-	-	-	-	-	Mouse	ISH	214
MSRB3	Methionine sulfoxide reductase B3	DFNB74	NA	NA	NA	+	+	+	NA	Mouse	IF	152
MYH9	Myosin, heavy chain 9, non-muscle	DFNA17	-	-	-	-	-	-	-	Rat	ISH	153
MYH14	Myosin, heavy chain 14, non-muscle	DFNA4	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
MYO3A	Unconventional myosin IIIA	DFNB30	NA	+	+	+	+	-	NA	Rat	IHC	155
MYO6	Unconventional myosin VI	DFNA22/DFNB37	NA	+	+	+	+	-	NA	Lake sturgeon, zebrafish, oscaae, american shad, Xenopus	IHC	215
MYO6	Unconventional myosin VI	DFNA22/DFNB37	NA	+	NA	+	+	-	NA	Mouse	IHC	216
MYO7A	Unconventional myosin VIIA	DFNA11/DFNB2/USH1B	NA	NA	NA	+	+	-	NA	Mouse, rat, guinea pig	IHC	157
MYO15A	Unconventional myosin XVa	DFNB3	NA	+	+	+	+	-	NA	Mouse, rat, guinea pig	IF	158

(continued)

**Table 2. (continued)**

Gene Symbol	Gene Name	Locus	Semicircular Canal	Utricle	Sacculle	Hair Cell Type I	Hair Cell Type II	Support Cell	Transitional Epithelium	Model	Method	Reference
NDP	Norrie disease	Norrie disease	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
OTOA	Otoacortin	DFNB22	NA	+	+	+	+	-	NA	Mouse	IF	160
OTOF	Otoferlin	DFNB9	NA	+	NA	+	+	-	NA	Mouse	IF	217
OTOG	Otogelin	AR-NSHL	NA	+	+	-	-	-	-	Mouse	IF	218
OTOGL	Otogelin-like protein	DFNB84	NA	+	+	+	+	-	-	Mouse, zebrafish	ISH, IHC	88
P2RX2	Purinergic receptor P2X <sub>2</sub> ligand-gated ion channel, 2	DFNA41	NA	NA	NA	+	+	+	NA	Rat	GFP	219
PAX3	Paired box 3	Waardenburg syndrome type I/III	+	+	+	+	+	-	-	Mouse	X-gal	163
PCDH15	Protocadherin 15	DFNB23/USH1F	NA	+	+	+	+	-	NA	Mouse	IHC	220
PDZD7	PDZ domain-containing 7	Usher syndrome	NA	+	+	+	+	-	NA	Mouse, rat, chicken	IHC	165
PNPT1	Polyribonucleotide nucleotidyltransferase 1, mitochondrial	DFNB70	NA	NA	NA	+	NA	NA	NA	NA	NA	NA
POLR1C	Polymerase I, RNA, subunit C	Treacher Collins syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
POLR1D	Polymerase I, RNA, subunit D	Treacher Collins syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
POU3F4	POU domain, class 3, transcription factor 4	DFNX2 (DFN3)	NA	NA	NA	+	+	-	NA	Mouse	IF	221
POU4F3	POU domain, class 4, transcription factor 3	DFNA15	+	+	+	+	+	-	NA	Mouse	GFP	168
PRPS1	Phosphoribosyl pyrophosphate synthetase 1	DFNX1 (DFN2)	NA	+	NA	+	+	-	-	Murine	IHC	169
PTPRQ	Protein tyrosine phosphatase, receptor type, Q	DFNB84	NA	+	+	+	+	-	-	Mouse	IF	171
RDX	Radixin	DFNB24	NA	NA	NA	+	+	-	NA	Mouse	IF	222
SANS	Scaffold-containing ankyrin repeats and SAM domain	USH1G	NA	NA	NA	+	+	NA	NA	Guinea pig, rat	IHC	157
SEMA3E	Sema domain, immunoglobulin domain, short basic domain, secreted, 3E	CHARGE syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SERPINB6	Serpin peptidase inhibitor, clade B, member 6	AR-NSHL	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SIX1	SIX homeobox 1	DFNA23	+	+	+	+	+	-	NA	Mouse	X-gal	174
SIX5	SIX homeobox 5	Alport syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SLC17A8	Vesicular glutamate transporter 3	DFNA25	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SLC26A4	Pendrin	DFNB4/Pendred syndrome	-	+	+	-	-	-	+	Mouse	ISH, IHC	177, 178
SLC26A5	Prestin	DFNB61	NA	+	+	+	+	-	NA	Mouse	ISH, IHC	223

(continued)

**Table 2. (continued)**

Gene Symbol	Gene Name	Locus	Semicircular Canal	Utricle	Sacculle	Hair Cell Type I	Hair Cell Type II	Support Cell	Transitional Epithelium	Model	Method	Reference
SMAC1	Second mitochondrial-derived activator of caspase	DFNA64	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
DIABLO												
SMPX	Small muscle protein, X-linked	DFNX4 (DFN6)	+	+	NA	+	+	-	NA	Mouse	ISH	224
SNAI2	Snail family zinc finger 2	Waardenburg syndrome type IID	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
SOX10	SRY (sex determining region Y)-Box 10	Waardenburg syndrome type IV	+	+	+	-	-	-	NA	Mouse	ISH	214
STRC	Stereocilin	DFNB16	NA	+	+	+	+	-	NA	Mouse	IF	37
SYNE4	Nesprin-4	DFNB76	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
TBC1D24	TBC1 domain family, member 24	DFNA65/DFNB86	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
TCOF1	Treacher Collins-Franceschetti syndrome I	Treacher Collins syndrome	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
TECTA	Tectorin alpha	DFNA8/12/DFNB21	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
TJP2	Tight junction protein ZO 2	DFNA51	NA	+	+	-	-	+	NA	Mouse	IHC	54
TMCI	Transmembrane channel-like protein 1	DFNA36/DFNB7/11	+	+	NA	NA	NA	NA	NA	Mouse	ISH	225
TMCI	Transmembrane channel-like protein 1	DFNA36/DFNB7/11	NA	NA	NA	+	+	NA	NA	Mouse	ISH	38
TMIE	Transmembrane inner ear expressed protein	DFNB6	NA	+	+	+	+	-	-	Rat	IHC	187
TMPRSS3	Transmembrane protease, serine 3	DFNB8/10	NA	+	+	+	+	-	-	Mouse	ISH	189
TNC	Tenascin C	DFNA56	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
TPRN	Taperin	DFNB79	NA	NA	+	+	+	-	-	Mouse	GFP	191
TRIOBP	TRIO and F-actin-binding protein	DFNB28	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
TSPEAR	Thrombospondin-type laminin G domain and EAR REPEATS	DFNB98	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
USH1C	Harmonin	DFNB18/USH1C	+	+	+	+	+	-	-	Mouse	IF	194
USH2A	Usherin	USH2A	NA	NA	NA	+	+	-	NA	Mouse	IF	226
WFS1	Wolframin	DFNA6/14/38	NA	NA	NA	+	+	+	NA	Mouse	ISH, IHC	197
WHRN	Whirlin	DFNB31/USH2D	NA	+	NA	+	+	-	NA	Mouse	IF	227

Abbreviations: BOR, branchiootorenal syndrome; EM, electron microscopy; IF, immunofluorescence; IHC, immunohistochemistry; ISH, in situ hybridization; NSHL, nonsyndromic hearing loss; PRLTS, Perrault syndrome; STL, Stickler syndrome; TCS, Treacher Collins syndrome; USH, Usher syndrome; WS, Waardenburg Syndrome; +, expressed; -, not expressed; NA, not applicable.

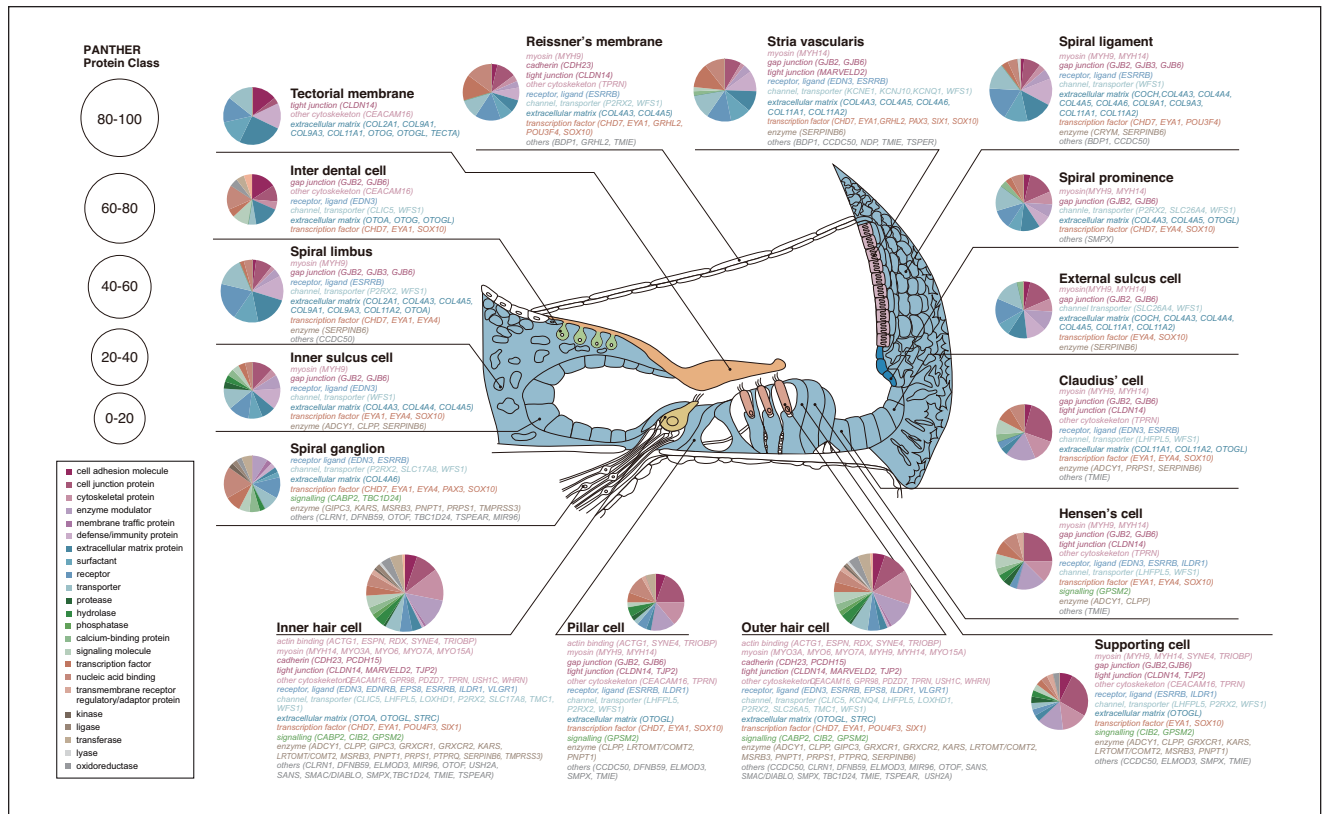




Table 3. (continued)

	Inner Hair Cell	Outer Hair Cell	Spiral Ganglion	Pillar Cell	Supporting Cell	Hensen's Cell	Claudius' Cell	External Sulcus	Spiral Prominence	Spiral Ligament	Stria Vascularis	Reissner's Membrane	Interdental Cell	Tectorial Membrane	Spiral Limbus	Inner Sulcus Cell	Vestibular Type I Hair Cell	Vestibular Type 2 Hair Cell	Supporting Cell	Transitional Cell	
Transporter	CLIC5 GPR98 LOXHD1 P2RX2 SLC17A8 TMCI	CLIC5 GPR98 LOXHD1 P2RX2 SLC26A5 TMCI KCNQ4	COL4A6 P2RX2 SLC17A8	P2RX2	P2RX2	COL11A1 COL11A2	COL11A1 COL11A2	COL4A3 COL4A4 COL4A5 COL11A1 COL11A2 SLC26A4	COL4A3 COL4A4 COL4A5 P2RX2 SLC26A4	COL4A3 COL4A4 COL4A5 COL4A6 COL11A1 COL11A2 COL11A1 COL11A2	COL4A3 COL4A4 COL4A5 COL11A1 COL11A2 KCNQ1	COL4A3 COL4A5 P2RX2	CLIC5 CLIC5	COL2A1 COL2A1 COL9A3 COL9A3 COL11A1	COL2A1 COL4A3 COL4A5 COL9A1 COL9A3 COL11A2 P2RX2	COL4A3 COL4A4 COL4A5	CLIC5 GPR98 LOXHD1 P2RX2 SLC26A5 TMCI KCNQ4 KCNQ4	CLIC5 GPR98 LOXHD1 P2RX2 P2RX2 P2RX2 P2RX2 KCNQ4	P2RX2 KCNQ4		
Protease	CLPP	CLPP		CLPP	CLPP	CLPP										CLPP					
Hydrolase	CIB2 CLPP PNPT1 PTPRQ	CIB2 CLPP PNPT1 PTPRQ	PNPT1	CLPP CLPP PNPT1	CLPP CLPP CIB2	CLPP										CLPP					
Phosphatase																					
Calcium-binding protein	CABP2 CIB2 TMPRSS3	CABP2 CIB2 TMPRSS3	CABP2 TMPRSS3		CIB2																
Signaling molecule	CLIC5 EDN3 ELMOD3 LOXHD1 TMPRSS3	CLIC5 EDN3 ELMOD3 LOXHD1 TMPRSS3	EDN3 TMPRSS3	ELMOD3	ELMOD3	EDN3 ELMOD3	EDN3 ELMOD3	EDN3 ELMOD3	EDN3 ELMOD3	EDN3	EDN3	EDN3	CLIC5 EDN3								
Transcription factor	ESRRB MARVELD2 POU4F3 SIX1	ESRRB MARVELD2 POU4F3 SIX1	ESRRB PAX3 SOX10	ESRRB SOX10	ESRRB SOX10	ESRRB SOX10	ESRRB SOX10	ESRRB SOX10	ESRRB SOX10	ESRRB POU3F4 POU3F4 PAX3 SIX1 SOX10	ESRRB MARVELD2 GRHL2 PAX3 SIX1 SOX10	ESRRB GRHL2 ESRRB GRHL2 SOX10	SOX10	ESRRB	ESRRB	SOX10	MARVELD2 PAX3 POU3F4 POU4F3 SIX1 SIX1	MARVELD2 MARVELD2 PAX3 POU3F4 POU4F3 SIX1 SIX1	MARVELD2 MARVELD2 PAX3 PAX3		
Nucleic acid binding	CHD7 CLIC5 ESRRB KARS PNPT1	CHD7 CLIC5 ESRRB KARS PNPT1	CHD7 ESRRB KARS PNPT1 PNPT1 SOX10	CHD7 ESRRB PNPT1 SOX10	ESRRB KARS SOX10	ESRRB SOX10	ESRRB SOX10	CHD7 SOX10	CHD7 SOX10	BDP1 CHD7 ESRRB	BDP1 CHD7 ESRRB PAX3 SOX10	CHD7 CHD7 SOX10	CHD7 CLIC5 CHD7 CLIC5 SOX10		CHD7 ESRRB	SOX10	CLIC5 KARS PAX3	CLIC5 KARS PAX3	KARS KARS KARS		
Transmembrane receptor/regulatory adaptor protein	EP58 GFSM2	EP58 GFSM2		GFSM2	GFSM2	GFSM2															
Kinase	PRPS1 KARS PRPS1	PRPS1 KARS PRPS1	PRPS1 KARS PRPS1																		
Ligase	ADCY1	ADCY1																			
Oxidoreductase	CLIC5 GRXCRI GRXCRI MSRB3	CLIC5 GRXCRI GRXCRI MSRB3	MSRB3 GRXCRI	MSRB3 GRXCRI	MSRB3 GRXCRI	MSRB3 GRXCRI	MSRB3 GRXCRI	MSRB3 GRXCRI	MSRB3 GRXCRI	MSRB3 GRXCRI	MSRB3 GRXCRI	MSRB3 GRXCRI	CLIC5 GRXCRI MSRB3	CLIC5 GRXCRI MSRB3	CLIC5 GRXCRI MSRB3	CLIC5 GRXCRI MSRB3	CLIC5 GRXCRI MSRB3	CLIC5 GRXCRI MSRB3	CLIC5 GRXCRI MSRB3	GRXCRI	GRXCRI
Transferase	CLIC5 GIPC3 LRTOMT PNPT1 PRPS1	CLIC5 GIPC3 LRTOMT PNPT1 PRPS1	PNPT1 PRPS1 GIPC3	LRTOMT PNPT1	LRTOMT PNPT1	LRTOMT PNPT1	LRTOMT PNPT1	LRTOMT PNPT1	LRTOMT PNPT1	LRTOMT PNPT1	LRTOMT PNPT1	LRTOMT PNPT1	CLIC5 GIPC3 LRTOMT PRPS1	CLIC5 GIPC3 LRTOMT PRPS1	CLIC5 GIPC3 LRTOMT PRPS1	CLIC5 GIPC3 LRTOMT PRPS1	CLIC5 GIPC3 LRTOMT PRPS1	CLIC5 GIPC3 LRTOMT PRPS1	CLIC5 GIPC3 LRTOMT PRPS1	LRTOMT PRPS1 CLIC5	LRTOMT PRPS1 CLIC5
Isomerase	CLIC5	CLIC5																			

<sup>a</sup>Gene ontology analyses were performed using PANTHER software as described previously.



**Figure 1.** Gene expression profiles of the causative genes and localization of the encoded proteins involved in hereditary hearing loss in the cochlea. Pie charts indicate the results of gene ontology analysis of the gene expression profiles for each cell type.

loss.<sup>28-42</sup> Most of the Usher syndrome causative genes are included in this group and are expressed in the stereocilia (Figures 2A, 2B). *LOXHD1* and *GIPC3* are not components of the stereocilia; however, mutations in these genes in mice were shown to lead to degeneration of the stereocilia.<sup>43-45</sup> All of these genes are only expressed in the IHCs and OHCs.

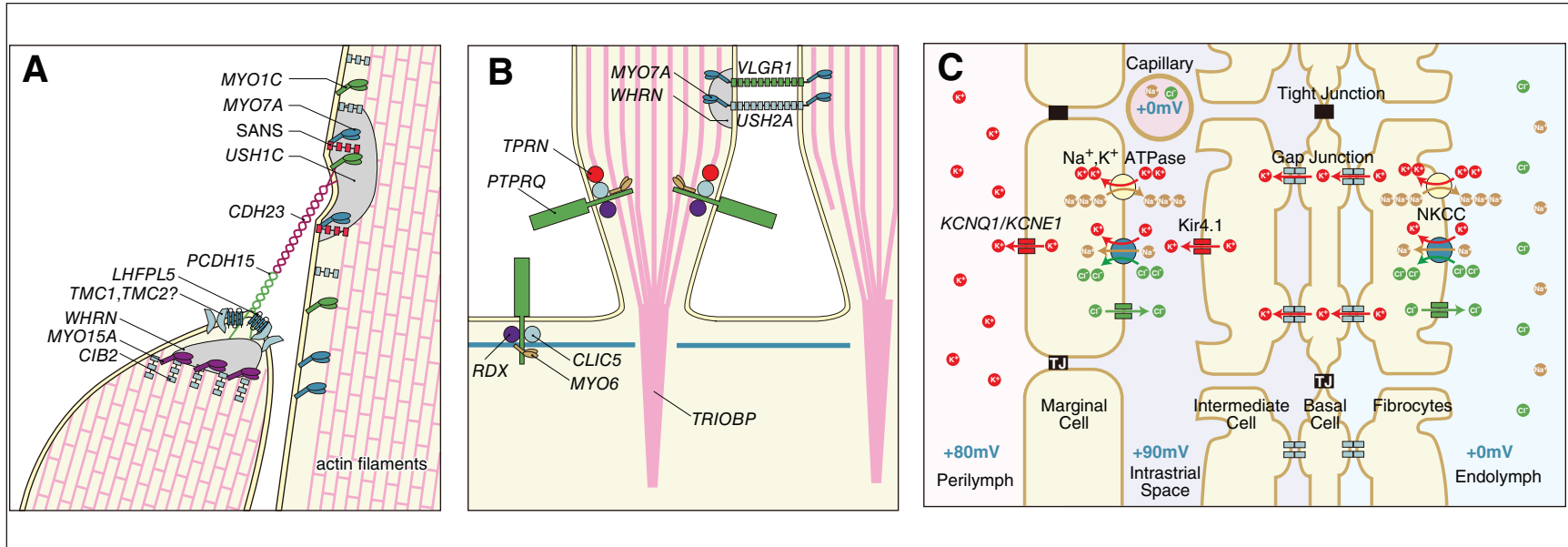
The third group comprises nuclear binding protein, transcription factor, receptor, and signaling molecule genes, including *SIX1* (Sine oculis homeobox drosophila homolog 1), *POU4F3* (POU domain class 4 transcription factor 3), *EDN3* (endothelin 3), *EDNRB* (endothelin receptor type B), *EPS8* (epidermal growth factor receptor kinase substrate 8), and *GPSM2* (G-protein signaling modulator 2). These genes may have an important role in the transcription signaling pathway in IHC and OHC differentiation (Figure 3A).<sup>46-52</sup>

The fourth group of genes comprises genes encoding tight junction proteins, including *MARVELD2* (tricellulin), *TJP2* (tight junction protein ZO 2), and *CLDN14* (claudin 14). These genes are components of the tight junctions. *MARVELD2* is expressed only in the IHCs and OHCs, while *TJP2* is expressed in the IHCs, OHCs, pillar cells, and adjacent supporting cells, and *CLDN14* is expressed in the IHCs, OHCs, supporting cells, and Reissner's membrane.

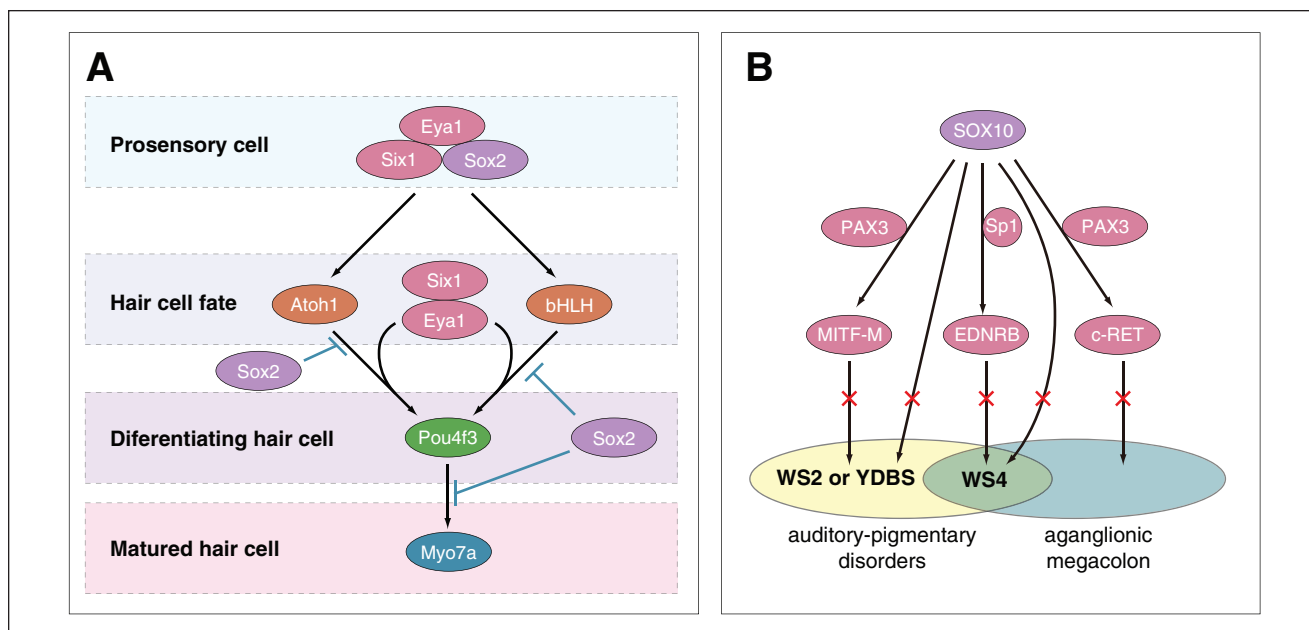
The tight junctions of these cells are believed to act as barriers that are required for normal hearing. Moreover, tricellulin proteins in the IHCs and OHCs form connections with adjacent cells and may prevent potassium ion (K<sup>+</sup>) leakage from the apical side of hair cells.<sup>53-55</sup>

### Outer Hair Cells Are the Center for Cochlea Amplification

OHCs in the organ of Corti are organized into 3 lines of cells that act as amplifiers of auditory signals. Similar to IHCs, OHCs also have stereocilia at the top of the cells. The OHC stereocilia are attached to the tectorial membrane; however, IHC stereocilia do not come into contact with the tectorial membrane. OHCs are electromotile, acting as an amplifier of basilar membrane vibrations. When OHCs are depolarized by K<sup>+</sup> ions from the stereocilia, motor proteins (prestin) located on the surface of the OHCs are shortened. As a result of the prestin movement, OHCs shrink in synchronization with basilar membrane vibrations.<sup>56,57</sup> As a result of this erector-evoked movement of the OHCs, basilar membrane movements are amplified, and the stereocilia of IHCs undergo more extensive swaying. This mechanism plays an important role in enhancing the dynamic range of



**Figure 2.** (A) Detailed structural components of the stereocilia tip link. Most of the Usher syndrome causative genes are expressed in the stereocilia and constructed tip link. (B) Detailed structural components of the stereocilia basal region. (C) Detailed structural components of the stria vascularis region. These figures are modified from previous reports.<sup>9,11,113</sup> TJ, tight junction.



**Figure 3.** (A) *EYA-SIX-SOX* transcriptional pathway for hair cell differentiation. Most of the transcriptional factors expressed in hair cells are involved in this pathway. (B) *SOX-PAX-MITF* transcriptional pathway for melanocyte differentiation. Most of the transcriptional factors expressed in melanocytes in the stria vascularis are involved in this pathway. These figures are modified from previous reports.<sup>98,99</sup> WS, Waardenburg syndrome; YDBS, Yemenite deaf-blind hypopigmentation syndrome.

sound sensors in the IHCs, particularly for low-intensity sounds.<sup>56,57</sup> The movement of prestin itself does not require ATP as an energy source. Rather, changes in voltage capacity cause conformational changes in the prestin on the OHC surface membrane, thereby acting as the motor to drive these movements. OHCs are also controlled by efferent neurons, which act as a feedback system and modulate these movements.<sup>58</sup>

Many genes are involved in the maintenance of this unique characteristic of OHCs (Figure 1), and mutations in these genes also cause hearing loss. The gene expression profiles of OHCs are quite similar to those of IHCs, and most genes are commonly expressed in both OHCs and IHCs; however, the expression of the mechanical amplification motor protein (prestine) gene *SLC26A5* is restricted to OHCs. Three myosin motor proteins encoded by *MYO3A*, *MYO7A*, and *MYO15A* are expressed only in IHCs and OHCs. These genes may contribute to effective mechanical transduction and may also be involved in effective sound amplification by OHCs.

### Pillar Cells Are Anchors for the Basilar Membrane and Hair Cells

Pillar cells act as the supporting cells located between IHCs and OHCs and are characterized by the presence of cross-linked actin filaments that ensure the necessary stiffness to support the hair cells on the basilar membrane

and synchronize the vibration of the basilar membrane and hair cells. Pillar cells express actin gamma encoded by *ACTG1* and tight junction proteins encoded by *CLDN14* and *TJP2*.<sup>54,59,60</sup> The gene expression profiles of pillar cells are similar to those of hair and other supporting cells; *ACTG1* expression is only observed in hair and pillar cells.<sup>28,52</sup> In contrast, many types of myosin and various stereocilia components are not expressed in pillar cells. The gap junction proteins are commonly observed among the cochlea supporting and lateral wall cells, and those encoded by *GJB2* and *GJB6* are also observed in these cells.<sup>61</sup>

### Inner Phalangeal Cells, Border Cells, and Deiters' Cells Are Neuroglial Cells in the Organ of Corti

Inner phalangeal cells, border cells, and Deiters' cells are all located adjacent to hair cells. Deiters' cells act as the supporting cells for OHCs and conform to the shape of the OHCs, while inner phalangeal cells and border cells act as the supporting cells for IHCs and conform to their shape. Furthermore, all of the aforementioned cells possess actin filaments and microtubules stretching from the basilar membrane to the reticular membrane in order to anchor the hair cells in the appropriate positions.

Another important role played by inner phalangeal and border cells is the glial cell-like uptake of the glutamate neurotransmitter, which is converted to glutamine.<sup>62-64</sup> After

this conversion, glutamine is transported to the hair cells, converted back to glutamate, and stored in the synaptic ribbon vesicles.<sup>62-64</sup>

The expression of tight junction proteins *CLDN14* and *TJP2* is expressed only in hair, pillar, inner phalangeal cell, Border cell, and Deiters' cells. Methionine-R-sulfoxide reductase B3 (*MSRB3*) acts as an oxidoreductase, and its expression is only observed in hair, spiral ganglion, and inner phalangeal cell and Border and Deiters' cells.<sup>65-72</sup> Mutations in *MSRB3* cause DFNB74,<sup>65-72</sup> revealing the importance of decreasing oxidative stress in the protection of hair and nerve cells. In contrast, many types of myosin and stereocilia components are not expressed in inner phalangeal, border, or Deiters' cells. However, gap junction proteins are commonly observed among the cochlea supporting and lateral wall cells, and those encoded by *GJB2* and *GJB6* are observed in this cell type. Mutations in gap junction genes are known to cause nonsyndromic hearing loss DFNA3A/DFNB1A and DFNB1B.<sup>66,67</sup>

### The Spiral Prominence Is the Center for Chloride, Bicarbonate, and Iodide Ion Transport

Hensen's and Claudius' cells are located adjacent to Deiters' cells. External sulcus cells and the spiral prominence are connected to the organ of Corti and stria vascularis. Most of these cells produce pendrin, which is encoded by *SLC26A4*. Pendrin acts as a chloride ( $\text{Cl}^-$ ), bicarbonate ( $\text{HCO}_3^-$ ), and iodide ( $\text{I}^-$ ) ion transporter and is expressed in the cochlea, kidney, and thyroid gland. Mutations in *SLC26A4* result in nonsyndromic hearing loss with an enlarged vestibular aqueduct (DFNB4) and Pendred syndrome (hearing loss and goiter are the main symptoms).<sup>68-70</sup> Among the supporting cells, the spiral prominence strongly expresses *SLC26A4* and is believed to be responsible for maintaining  $\text{Cl}^-$  and  $\text{HCO}_3^-$  ion concentrations.

### The Stria Vascularis and Spiral Ligament Are Batteries for Mechanical Transduction

The scala media of the cochlea is filled with endolymph, which has a high positive potential (+80 mV) due to the high concentration of  $\text{K}^+$  ions (150 mM).<sup>10,71,72</sup> The scala vestibuli and scala tympani are filled with perilymph, which has no potential (0 mV) and a low concentration of  $\text{K}^+$  ions (5 mM). The endocochlear potential (EP) and high concentration of  $\text{K}^+$  ions act as an energy source for efficient mechanoelectric transduction. This unique fluid is produced by the stria vascularis and spiral ligament. The stria vascularis is composed of 3 layers of cells (marginal, intermediate, and basal cells), with narrow intrastrial spaces (IS) between the marginal and

intermediate cell layers. Basal and intermediate cells are connected to each other by gap junctions and the  $\text{K}^+$  ions of the perilymph are transported into the basal cells by the  $\text{Na}^+/\text{K}^+$ ATPase and  $\text{Na}^+/\text{K}^+/\text{2Cl}^-$  co-transporter (NKCC) located on the basolateral side of the basal cells (Figure 2C). On the apical surface of the intermediate cells, *Kir4.1* encodes a potassium channel that transports  $\text{K}^+$  ions to the IS.<sup>10,71,72</sup> Marginal cells have NKCC and  $\text{Na}^+/\text{K}^+$ ATPase at the basolateral surfaces and voltage-gated  $\text{K}^+$  channels (KQT-like subfamily and ISK-related subfamily members) at the apical membranes; the genes for these channels are *KCNQ1* and *KCNE1*, respectively.<sup>10,72</sup> As a result of nonequivalent expression patterns on the basolateral and apical sides of the intermediate and marginal cells,  $\text{K}^+$  ions are efficiently pumped from the perilymph to the endolymph and  $\text{Na}^+$  ions are returned to the perilymph<sup>10,72</sup> (Figure 2C). This system requires ATP as an energy source, and blood vessels located in the stria vascularis supply oxygen and nutrients for ATP synthesis. The stria vascularis expresses many unique genes. *KCNQ1* and *KCNE1* are genes for the  $\text{K}^+$  channels expressed on the apical surface of the marginal cells of the stria vascularis. Mutations in these genes cause Jervell and Lange-Nielsen syndrome, which is characterized by congenital deafness, and long QT syndrome.<sup>73-75</sup> Paired box gene 3 (*PAX3*), SRY-Box 10 (*SOX10*), and Endothelin 3 (*EDN3*) are expressed in the stria vascularis. *PAX3*, *SOX10*, *MITF*, *EDNRB*, and *EDN3* are reported to be causative genes of Waardenburg syndrome.<sup>76-79</sup> *PAX3* and *SOX10* encode transcriptional activator proteins and directly bind to the promoter of the *MITF*, *EDNRB*, and *RET* genes (Figure 3B).<sup>78</sup> *MITF* codes for the protein that is presumed to be a transcriptional factor associated with melanocyte differentiation.<sup>79</sup> In the organ of Corti, melanocytes are observed only in the stria vascularis and may have an important role in oxidoreductase activity.

### The Tectorial Membrane Is a Sound Signal Enhancer

The tectorial membrane is a component of the organ of Corti and is comprised of collagens and non-collagenous glycoproteins; the membrane covers both the IHCs and OHCs. The longest OHCs stereocilia are connected to the tectorial membrane, and amplification of basilar membrane movement by the OHCs causes the endolymph to flow between the organ of Corti and tectorial membrane. As a result of this endolymph flow, IHCs more efficiently transduce sound signals.

Gene mutations associated with the tectorial membrane also causes hearing loss. Collagen, encoded by *COL2A1*, *COL9A1*, *COL9A3*, and *COL11A1*, is a

component of the tectorial membrane, and mutations in these genes appear to be related to nonsyndromic hearing loss or Stickler syndrome.<sup>80-84</sup> The collagen genes associated with hearing loss are not distributed in the IHCs, OHCs, or adjacent supporting cells, external sulcus cells, or the spiral prominence.<sup>80-86</sup> Further, no type IV collagen genes, including *COL4A3*, *COL4A5*, and *COL4A6*, are components of the tectorial membrane. Mutations in the type IV collagen genes cause Alport syndrome. This differential expression pattern among collagen genes might contribute to the differential roles of collagen in the hearing system. Inner ear-specific glycoproteins, coded by *TECTA*, *OTOG*, and *OTOGL*, are also expressed in the tectorial membrane. Mutations in *TECTA* cause DFNA8/12/DFNB21, those in *OTOG* cause nonsyndromic hearing loss, and those in *OTOGL* cause DFNB84.<sup>87-89</sup> Gene mutations in the tectorial membrane components can cause mild to moderate hearing loss (termed *cochlear conductive hearing loss*), and hearing aids are effective for such. This may indicate that gene mutations in the tectorial membrane components can cause a malformation of the membrane, which is required for enhancing sound signals.

### Reissner's Membrane Separates the Perilymph and Endolymph

Reissner's membrane is a component of the cochlea, separating the scala vestibuli and the scala media. Reissner's membrane comprises 2 cell layers and tight junction proteins that prevent the leakage of the endolymph into the perilymph. *CLDN14*, which encodes tight junction proteins and *CDH23* are expressed in Reissner's membrane.<sup>55,90</sup>

### Gene Expression Profiles of the Vestibular Endorgans

Our review also focused on the vestibular system. Most of the genes associated with hereditary hearing loss are also expressed in the vestibular endorgans. However, only a limited number of genes, namely, *COCH* and *SLC26A4*, have been reported to be associated with vestibular dysfunctions and/or vertigo,<sup>91-94</sup> and the involvement of these genes remains unclear. In the vestibular system, mechanical transduction occurs by linear and rotatory acceleration. This mechanoelectric transduction is conducted by the hair cell stereocilia of the utricle and saccule together with the crista-ampullaris of the semicircular canals. Hair cells of the vestibular endorgans are slightly different from those of the organ of Corti. Vestibular stereocilia are linked to the kinocilium, and

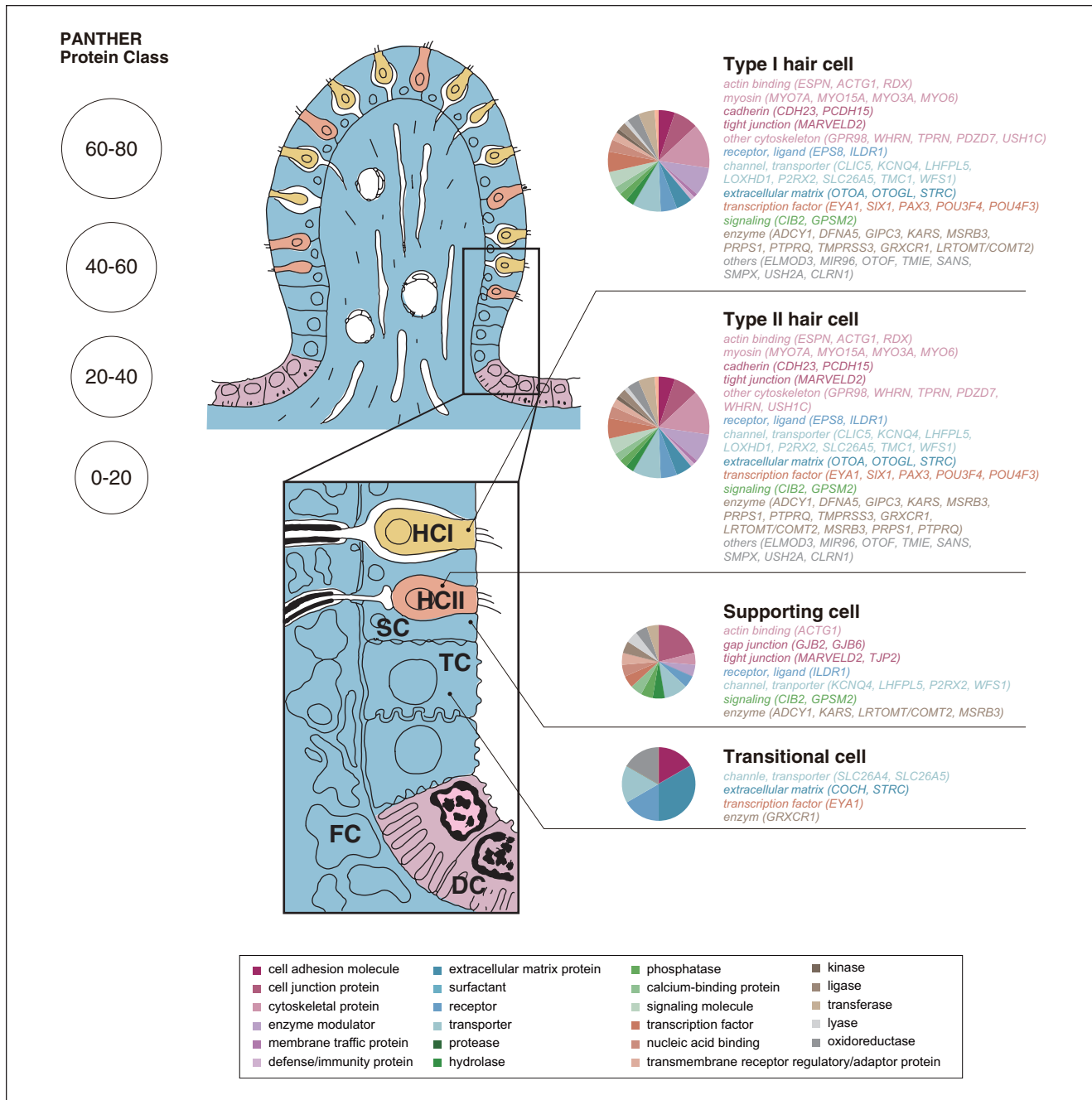
stereocilia movement toward the kinocilium depolarizes the hair cells.

Unlike the cochlea, only a limited number of reports are available for deafness-related causative genes expressed in the vestibular endorgans. A large proportion of the causative genes for deafness are also expressed in the vestibular endorgans, with the currently available information indicating that only 2 out of 72 genes (*MITF* and *MYH9*) are restricted to the cochlea. This may reflect differences between the cochlea and vestibular endorgans (Figure 4).

The remaining genes are expressed in both the cochlea and vestibular endorgans; thus, it is difficult to explain the absence of vertigo in most patients. Possible explanations for this inconsistency are as follows: (1) other molecules compensate for the functional loss of sensory activity in the vestibular endorgans, (2) congenital vestibular dysfunction is compensated by visual and somatosensory input, and (3) the vestibular sensory system requires a lower degree of sensitivity than does the audio system. It is noteworthy that a missense mutation in *CDH23* causes DFNB12, and vertigo is not associated with the mutation. Nonsense or frameshift mutations, which have more deleterious effects on protein function, cause Usher syndrome type 1D, which is characterized by retinitis pigmentosa and vertigo. Some of these inconsistencies can be explained by the third point; however, further studies are required to elucidate the effects of mutations linked to deafness on vestibular functions.<sup>29,95,96</sup>

### Gene Expression Profiles of Genes Causing Deafness in Other Body Parts

Some mutations in the deafness causative genes only cause hearing loss (nonsyndromic hearing loss); however, others cause syndromic hearing loss with various associated symptoms. The presence of these associated symptoms may be related to the expression profiles of the genes correlated with deafness in other parts of human body. To elucidate the gene expression profiles of genes previously reported to cause deafness, we conducted the database search summarized in Table 4. We also added information on the domain structure and protein interactions. Most of the genes known to cause deafness, including *GJB2*, *CDH23*, and *TECTA*, are expressed mainly in the cochlea; only a small percentage are expressed in other body parts. In contrast, most genes causing syndromic hearing loss, including *COL4A3*, *COL4A4*, *COL4A5* (Alport syndrome); *COL2A1*, *COL9A1*, *COL11A1*, *COL11A2* (Stickler syndrome); *PAX3* (Waardenburg syndrome); and *EYAI* (BOR syndrome), are expressed in other parts of the human body. These



**Figure 4.** Gene expression profiles of the causative genes and localization of the encoded proteins involved in hereditary hearing loss in the vestibular endorgan. Pie charts indicate the results of gene ontology analysis of the gene expression profiles for each cell type.

results can satisfactorily explain the presence of various symptoms in addition to hearing loss. However, many of the genes associated with nonsyndromic hearing loss are also expressed in other body parts. Further investigation is needed to elucidate the mechanisms underlying nonsyndromic hearing loss. One possible explanation of the

forementioned inconsistencies is the presence of alternative splicing variants as *OTOF* transcription variants NM\_001100393 and NM\_001144074 are expressed in the brain and kidney to some extent, while NM\_004802 and NM\_194248 are not observed in other body parts and are only expressed in the cochlea.<sup>97</sup>



**Table 4.** Summary of Gene Expression Profiles of Each Deafness Gene in Organs.

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<i>ACTG1</i> Actin, cytoplasmic 2		EST: High expression in muscle and heart. GeneChip: High expression in muscle and heart. CAGE: High expression in muscle and heart. RNA-seq: High expression in muscle and heart.		Itself ACTB CFL2 MLHI
<i>ADCY1</i> Adenylate cyclase type I		EST: High expression in brain and lung. GeneChip: High expression in brain and lung. CAGE: High expression in brain and lung. RNA-seq: High expression in brain and lung.	493 – 520 Interaction with calmodulin region 1024 – 1047 Interaction with calmodulin region	
<i>BSND</i> Barttin		EST: High expression in kidney and urinary organs. GeneChip: nodata CAGE: nodata RNA-seq: High expression in kidney and urinary organs.		
<i>BDP1</i> Transcription factor TFIIB component B homolog		EST: High expression in brain and lung. GeneChip: High expression in brain and lung. CAGE: High expression in brain and lung. RNA-seq: High expression in brain and lung.	295 – 345 Myb-like domain, 823 – 877 approximate repeat 878 – 932 repeat 2 933 – 987 3 repeat 988 – 1040 4 repeat 1041 – 1094 5 repeat 1095 – 1148 6 repeat 1149 – 1203 7 repeat 1204 – 1257 8; approximate repeat 1258 – 1327 9; approximate repeat 1 – 299 Interaction with ZBTB43 region 355 – 470 Required for phosphorylation by CSNK2A1 region 823 – 1327 9 X 55 AA repeats region 144 – 177 coiled coil 1078 – 1103 coiled coil 1223 – 1284 coiled coil	
<i>CABP2</i> Calcium-binding protein 2		EST: High expression in brain and lung. GeneChip: High expression in brain and lung. CAGE: High expression in brain and lung. RNA-seq: High expression in brain and lung.	78 – 113 EF-hand 1 domain 111 – 146 EF-hand 2 domain 152 – 187 EF-hand 3 domain 189 – 220 EF-hand 4 domain	
<i>CCDC50</i> Coiled-coil domain-containing protein 50		EST: High expression in kidney and urinary organs. GeneChip: nodata CAGE: High expression in kidney and urinary organs. RNA-seq: High expression in kidney and urinary organs.	63 – 130 coiled coil	OTUD7B, RIPK1, UBB
<i>CDH23</i> Cadherin 23		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: High expression in kidney and urinary organs.	34 – 132 Cadherin 1 domain 133 – 236 Cadherin 2 domain 237 – 348 Cadherin 3 domain 349 – 460 Cadherin 4 domain 461 – 561 Cadherin 5 domain 562 – 671 Cadherin 6 domain 672 – 784 Cadherin 7 domain 779 – 890 Cadherin 8 domain 891 – 995 Cadherin 9 domain 996 – 1102 Cadherin 10 domain 1103 – 1208 Cadherin 11 domain 1210 – 1313 Cadherin 12 domain 1314 – 1418 Cadherin 13 domain	


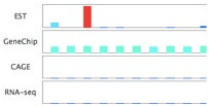

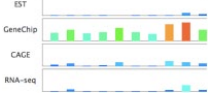

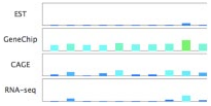




(continued)

Table 4. (continued)

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<i>CEACAM16</i> Carcinoembryonic antigen-related cell adhesion molecule 16		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata	1420 – 1527 Cadherin 14 domain 1529 – 1634 Cadherin 15 domain 1635 – 1744 Cadherin 16 domain 1745 – 1851 Cadherin 17 domain 1852 – 1959 Cadherin 18 domain 1960 – 2069 Cadherin 19 domain 2070 – 2174 Cadherin 20 domain 2175 – 2293 Cadherin 21 domain 2297 – 2402 Cadherin 22 domain 2403 – 2509 Cadherin 23 domain 2510 – 2611 Cadherin 24 domain 2614 – 2722 Cadherin 25 domain 2729 – 2846 Cadherin 26 domain 2847 – 2975 Cadherin 27 domain 133 – 218 Ig-like C2-type 1 domain 223 – 309 Ig-like C2-type 2 domain	
<i>CIB2</i> Calcium- and integrin-binding family member 2		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata	66 – 101 EF-hand 1 domain 103 – 138 EF-hand 2 domain 144 – 179 EF-hand 3 domain	
<i>CLDN14</i> Claudin 14		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata		
<i>CLIC5</i> Chloride intracellular channel protein 5		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata	260 – 400 GST C-terminal domain	SRC
<i>COCH</i> Cochlin		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata	28 – 121 LCCL domain 165 – 346 VWFA 1 domain 367 – 537 VWFA 2 domain	
<i>COL1A2</i> (DFNA13/DFNB53/ Stickler syndrome) Collagen alpha 2(XI) chain		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata	57 – 228 Laminin G-like domain 399 – 447 Collagen-like 1 domain 487 – 545 Collagen-like 2 domain 546 – 590 Collagen-like 3 domain 805 – 862 Collagen-like 4 domain 863 – 899 Collagen-like 5 domain 1099 – 1156 Collagen-like 6 domain 1157 – 1172 Collagen-like 7 domain 1441 – 1499 Collagen-like 8 domain 1541 – 1735 Fibrillar collagen NCI domain 215 – 486 Nonhelical region 487 – 1500 Triple-helical region	DDR2
<i>COL4A6</i> Collagen alpha 6(IV) chain		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata	1467 – 1691 Collagen IV NCI domain 23 – 46 7S domain region 47 – 1463 Triple-helical region 515 – 517 Cell attachment site motif 560 – 562 Cell attachment site motif 986 – 988 Cell attachment site motif	

(continued)

Table 4. (continued)

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<i>COL2A1</i> (Stickler syndrome) Collagen, type II, alpha-1			32 – 90 VWF domain 1253 – 1487 Fibrillar collagen NCI domain 201 – 1214 Triple-helical region 1215 – 1241 Nonhelical region (C-terminal)	
<i>COL4A3</i> (Alport) Collagen, type IV, alpha-3			1445 – 1669 Collagen IV NCI domain 29 – 42 7S domain 43 – 1438 Triple-helical region 1427 – 1444 Epitope recognized by Goodpasture antibodies region 1479 – 1557 Required for the anti-angiogenic activity of tumstatin region 1610 – 1628 Required for the anti-tumor cell activity of tumstatin region 791 – 793 Cell attachment site motif 996 – 998 Cell attachment site motif 1154 – 1156 Cell attachment site motif 1306 – 1308 Cell attachment site motif 1345 – 1347 Cell attachment site motif 1432 – 1434 Cell attachment site motif	
<i>COL4A4</i> (Alport) Collagen, type IV, alpha-4			1465 – 1690 Collagen IV NCI domain 39 – 64 7S domain 65 – 1459 Triple-helical region 94 – 96 Cell attachment site motif 145 – 147 Cell attachment site motif 189 – 191 Cell attachment site motif 310 – 312 Cell attachment site motif 724 – 726 Cell attachment site motif 785 – 787 Cell attachment site motif 989 – 991 Cell attachment site motif 1212 – 1214 Cell attachment site motif	
<i>COL4A5</i> (Alport) Collagen, type IV, alpha-5			1461 – 1685 Collagen IV NCI domain 27 – 41 Nonhelical region (NC2) region 42 – 1456 Triple-helical region	
<i>COL11A1</i> (Stickler syndrome) Collagen, type XI, alpha-1			71 – 243 Laminin G-like domain 442 – 490 Collagen-like 1 domain 532 – 586 Collagen-like 2 domain 583 – 641 Collagen-like 3 domain 616 – 674 Collagen-like 4 domain 643 – 699 Collagen-like 5 domain 1393 – 1450 Collagen-like 6 domain 1429 – 1487 Collagen-like 7 domain 1483 – 1541 Collagen-like 8 domain 1577 – 1805 Fibrillar collagen NCI domain 230 – 419 Nonhelical region 420 – 508 Triple-helical region (interrupted) 509 – 511 Short nonhelical segment region 512 – 528 Telopeptide region 529 – 1542 Triple-helical region 1543 – 1563 Nonhelical region (C-terminal)	

(continued)

Table 4. (continued)

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<i>COL9A1</i> (Stickler syndrome) Collagen, type IX, alpha-1			50 – 244 Laminin G-like domain 269 – 324 Collagen-like 1 domain 325 – 356 Collagen-like 2 domain 358 – 403 Collagen-like 3 domain 416 – 472 Collagen-like 4 domain 473 – 516 Collagen-like 5 domain 587 – 643 Collagen-like 6 domain 655 – 712 Collagen-like 7 domain 713 – 755 Collagen-like 8 domain 790 – 847 Collagen-like 9 domain 848 – 899 Collagen-like 10 domain 24 – 268 Nonhelical region (NC4) 269 – 405 Triple-helical region (COL3) 406 – 417 Nonhelical region (NC3) 418 – 756 Triple-helical region (COL2) 757 – 786 Nonhelical region (NC2) 787 – 901 Triple-helical region (COL1) 902 – 921 Nonhelical region (NC1)	
<i>COL9A2</i> (Stickler syndrome) Collagen, type IX, alpha-2			27 – 163 Triple-helical region 4 (COL4) 164 – 180 Nonhelical region 4 (NC4) 181 – 519 Triple-helical region 3 (COL3) 520 – 549 Nonhelical region 3 (NC3) 550 – 632 Triple-helical region 2 (COL2) 633 – 634 Nonhelical region 2 (NC2) 635 – 664 Triple-helical region 1 (COL1) 665 – 689 Nonhelical region 1 (NC1)	
<i>CRYM</i> Thiomorpholine-carboxylate dehydrogenase				
<i>DFNA5</i> Non-syndromic hearing impairment protein				
<i>DIAPH1</i> Protein diaphanous homolog 1			84 – 449 GBD/FH3 domain 583 – 764 FH1 domain 769 – 1171 FH2 domain 1194 – 1222 DAD domain	ORF, PPM1F, RHOA
<i>DSPP</i> Dentin sialophosphoprotein			488 – 490 Cell attachment site motif	
<i>ELMOD3</i> ELMO domain-containing protein 3			170 – 324 ELMO domain	
<i>EPS8</i> Epidermal growth factor receptor kinase substrate 8			327 – 488 Helicase ATP-binding domain 542 – 702 Helicase C-terminal domain 6 – 18 Nuclear localization signal motif 441 – 444 DEVH box motif	PSMC5, RAD23B, XPC

(continued)

**Table 4. (continued)**

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<i>ESPN</i> Espin		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: [blue line]	651 – 668 WH2 domain 756 – 830 coiled coil	
<i>ESRRB</i> Steroid hormone receptor		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: [blue line]	103 – 123 NR C4-type zinc finger 139 – 163 NR C4-type zinc finger	
<i>EYA1</i> Eyes absent homolog 1		EST: [blue line] GeneChip: [green bars] CAGE: [blue line] RNA-seq: [blue line]		
<i>EYA4</i> Eyes absent homolog 4		EST: [blue line] GeneChip: [green bars] CAGE: [blue line] RNA-seq: [blue line]		
<i>GIPC3</i> PDZ domain-containing protein GIPC3		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: [blue line]	112 – 192 PDZ domain	
<i>GJB2</i> Gap junction beta 2 protein		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: [blue line]		
<i>GJB3</i>		EST: [blue line] GeneChip: [green bars] CAGE: [blue line] RNA-seq: [blue line]		
<i>GJB6</i> Gap junction beta 6 protein		EST: [blue line] GeneChip: nodata CAGE: [blue line] RNA-seq: [blue line]		
<i>GPSM2</i> G-protein signaling modulator 2		EST: [blue line] GeneChip: [green bars] CAGE: [blue line] RNA-seq: [blue line]	24 – 57 TPR 1 repeat 62 – 95 TPR 2 repeat 102 – 135 TPR 3 repeat 142 – 184 TPR 4 repeat 202 – 235 TPR 5 repeat 242 – 275 TPR 6 repeat 282 – 315 TPR 7 repeat 322 – 355 TPR 8 repeat 489 – 511 GoLoco 1 domain 544 – 566 GoLoco 2 domain 594 – 616 GoLoco 3 domain 628 – 650 GoLoco 4 domain	itself, GNAI1, NUMA1
<i>GRHL2</i>		EST: nodata GeneChip: [green bars] CAGE: [blue line] RNA-seq: [blue line]		

(continued)

**Table 4. (continued)**

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<i>GRXCR1</i> Glutaredoxin domain-containing cysteine-rich protein 1		NA	127 – 234 Glutaredoxin domain	
<i>GRXCR2</i> Glutaredoxin domain-containing cysteine-rich protein 2		NA		
<i>HGF</i> Hepatocyte growth factor		NA	27 – 515 Sema domain 563 – 655 IPT/TIG 1 domain 657 – 739 IPT/TIG 2 domain 742 – 836 IPT/TIG 3 domain 1078 – 1345 Protein kinase domain 1212 – 1390 Interaction with RANBP9 region 1320 – 1359 Interaction with MUC20 region	CBL, DNAJA3, EGFR, FGR, HGF, HGF, <i>inlB</i> (from a different organism), INPPL1, KDR, Kdr (from a different organism), LCK, LYN, MUC1, NCK1, NCK2, PIK3R1, PIK3R2, PIK3R3, PLCG1, PLXNB1, PLXNB2, PLXNB3, PTK2 (from a different organism), PTPNI, PTPNI1, PTPRB, PTPRJ, SH2B3, SH2D1A, SH2D1B, SH2D2A, SH2D3C, SHB, SHC1, SHC2, SHC4, SHD, SLA2, SOCS5, SOCS6, SRC, STAP1, SYK, TEC, TENC1, TNSI, TNS, VAV3, YES1, ZAP70
<i>ILDR1</i> Immunoglobulin-like domain-containing receptor 1			24 – 162 Ig-like V-type domain	
<i>KARS</i> Lysine-tRNA ligase				
<i>KCNQ4</i> Potassium voltage-gated channel subfamily KQT member 4			546 – 650 A-domain (Tetramerization) region 610 – 645 coiled coil 283 – 288 Selectivity filter motif	
<i>LHFPL5</i> Tetraspan membrane protein of hair cell stereocilia				
<i>LOXHD1</i> Lipoxygenase homology domain containing protein 1			18 – 134 PLAT 1 domain 147 – 262 PLAT 2 domain 275 – 395 PLAT 3 domain 406 – 525 PLAT 4 domain 536 – 650 PLAT 5 domain	

(continued)

**Table 4. (continued)**

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<i>LRTOMT</i> Transmembrane O-methyltransferase		NA	744 – 862 PLAT 6 domain 897 – 1015 PLAT 7 domain 1028 – 1153 PLAT 8 domain 1182 – 1300 PLAT 9 domain 1349 – 1467 PLAT 10 domain 1480 – 1595 PLAT 11 domain 1607 – 1725 PLAT 12 domain 1738 – 1859 PLAT 13 domain 1876 – 1947 PLAT 14 domain 139 – 140 S-adenosyl-L-methionine binding region	
<i>MARVELD2</i> MARVEL domain-containing protein 2		EST: nodata GeneChip: nodata CAGE: [blue line] RNA-seq: [blue line]	188 – 367 MARVEL domain 466 – 490 coiled coil 524 – 548 coiled coil	
<i>MSRB3</i> Methionine-R-sulfoxide reductase B3		EST: nodata GeneChip: nodata CAGE: [blue line] RNA-seq: [blue line]	189 – 192 Endoplasmic reticulum retention signal motif	
<i>MYH14</i> Myosin 14		EST: [orange bar] GeneChip: [orange bars] CAGE: [blue line] RNA-seq: [blue line]	105 – 800 Myosin motor domain 803 – 832 IQ domain 862 – 1947 coiled coil	
<i>MYH9</i> Myosin 9		EST: [red bars] GeneChip: [red bars] CAGE: [blue line] RNA-seq: [blue line]		CXCR4, GRB2, MEN1, NCL, SVIL
<i>MYO15A</i> Unconventional myosin XV		EST: [blue line] GeneChip: [green bars] CAGE: [blue line] RNA-seq: [blue line]	1222 – 1899 Myosin motor domain 1902 – 1924 IQ 1 domain 1925 – 1954 IQ 2 domain 1955 – 1976 IQ 3 domain 2065 – 2217 MyTH4 1 domain 2867 – 2953 SH domain 3050 – 3204 MyTH4 2 domain 3209 – 3530 FERM domain 1792 – 1799 Actin-binding region 1888 – 2029 Neck or regulatory domain region 2030 – 3530 Tail region 1323 – 1350 coiled coil	
<i>MYO3A</i> Myosin IIIa		EST: nodata GeneChip: [green bars] CAGE: [blue line] RNA-seq: [blue line]	21 – 287 Protein kinase domain 338 – 1053 Myosin motor domain 1055 – 1084 IQ 1 domain 1082 – 1111 IQ 2 domain 1346 – 1375 IQ 3 domain	
<i>MYO6</i> Unconventional myosin VI		EST: [red bar] GeneChip: [orange bars] CAGE: [blue line] RNA-seq: [blue line]	57 – 771 Myosin motor domain 814 – 834 IQ domain 273 – 317 Responsible for slow ATPase activity by similarity region 665 – 672 Actin-binding region	DAB2, Dab2

(continued)

**Table 4. (continued)**

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<p><i>MYO7A</i> Unconventional myosin VIIa</p>			<p>782 – 810 Required for binding calmodulin region 1116 – 1118 Interaction with OPTN region 864 – 1023 coiled coil 65 – 741 Myosin moto domain 745 – 765 IQ 1 domain 768 – 788 IQ 2 domain 791 – 811 IQ 3 domain 814 – 834 IQ 4 domain 837 – 857 IQ 5 domain 1017 – 1253 MyTH4 1 domain 1258 – 1602 FERM 1 domain 1603 – 1672 SH3 domain 1747 – 1896 MyTH4 2 domain 1902 – 2205 FERM 2 domain 632 – 639 Actin-binding region 858 – 935 coiled coil</p>	
<p><i>OTOA</i> Otoancorin</p>				
<p><i>OTOF</i> NM_001100393 NM_001144074 Otoferlin</p>			<p>241 – 338 C2 1 domain 404 – 514 C2 2 domain 947 – 1052 C2 3 domain 1479 – 1577 C2 4 domain 792 – 821 coiled coil</p>	
<p><i>OTOF</i> NM_004802 NM_194248 Otoferlin</p>			<p>241 – 338 C2 1 domain 404 – 514 C2 2 domain 947 – 1052 C2 3 domain 1479 – 1577 C2 4 domain 792 – 821 coiled coil</p>	
<p><i>OTOGL</i> Otogelin-like protein</p>		NA	<p>113 – 326 VWFD 1 domain 381 – 434 TIL 1 domain 473 – 683 VWFD 2 domain 736 – 791 TIL 2 domain 938 – 1141 VWFD 3 domain 1514 – 1734 VWFD 4 domain 2240 – 2332 CTCK domain</p>	
<p><i>P2RX2</i> P2X purinoceptor 2</p>			<p>320 – 333 Pore-forming motif region</p>	
<p><i>PCDH15</i> Protocadherin 15</p>			<p>40 – 147 Cadherin 1 domain 148 – 265 Cadherin 2 domain 278 – 395 Cadherin 3 domain 396 – 509 Cadherin 4 domain 510 – 616 Cadherin 5 domain 617 – 717 Cadherin 6 domain 719 – 819 Cadherin 7 domain 820 – 926 Cadherin 8 domain 927 – 1035 Cadherin 9 domain 1037 – 1144 Cadherin 10 domain 1145 – 1259 Cadherin 11 domain</p>	

(continued)



**Table 4. (continued)**

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<i>DFNB59</i> Pejvakin		EST GeneChip CAGE RNA-seq		
<i>PNPT1</i> Polyribonucleotide nucleotidyltransferase I, mitochondrial		EST GeneChip CAGE RNA-seq	605 – 664 KH domain 679 – 750 SI motif domain	
<i>POU3F4</i> POU domain, class 3, transcription factor 4		EST GeneChip CAGE RNA-seq	186 – 260 POU-specific domain	
<i>POU4F3</i> POU domain, class 4, transcription factor 3		EST GeneChip CAGE RNA-seq	179 – 256 POU-specific domain 56 – 65 POU-IV box motif	
<i>PRPS1</i> Ribose-phosphate pyrophosphokinase I		EST GeneChip CAGE RNA-seq	212 – 227 Binding of phosphoribosylpyrophosphate region	
<i>PTPRQ</i> Phosphatidylinositol phosphatase		EST GeneChip CAGE RNA-seq	36 – 99 Fibronectin type-III 1 domain 100 – 195 Fibronectin type-III 2 domain 199 – 294 Fibronectin type-III 3 domain 350 – 438 Fibronectin type-III 4 domain 441 – 539 Fibronectin type-III 5 domain 514 – 606 Fibronectin type-III 6 domain 610 – 705 Fibronectin type-III 7 domain 710 – 799 Fibronectin type-III 8 domain 804 – 894 Fibronectin type-III 9 domain 899 – 988 Fibronectin type-III 10 domain 993 – 1093 Fibronectin type-III 11 domain 1098 – 1190 Fibronectin type-III 12 domain 1192 – 1282 Fibronectin type-III 13 domain 1287 – 1380 Fibronectin type-III 14 domain 1384 – 1470 Fibronectin type-III 15 domain 1474 – 1578 Fibronectin type-III 16 domain 1583 – 1681 Fibronectin type-III 17 domain 1686 – 1787 Fibronectin type-III 18 domain 2036 – 2292 Tyrosine-protein phosphatase domain	
<i>RDX</i> Radixin		EST GeneChip CAGE RNA-seq	5 – 295 FERM domain 60 – 63 Phosphatidylinositol binding region	ITGB2
<i>SERPINE6</i> Serpin B6		EST GeneChip CAGE RNA-seq	Homeobox protein SIX1	MDF1

(continued)

Table 4. (continued)

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<i>SIX1</i> Homeobox protein SIX1, Sine oculis homeobox, drosophila, homolog of, 1		EST GeneChip CAGE RNA-seq		
<i>SLC17A8</i> Vesicular glutamate transporter 3		EST GeneChip CAGE RNA-seq		
<i>SLC26A4</i> Pendrin		EST GeneChip CAGE RNA-seq	535 – 729 STAS domain	
<i>SLC26A5</i> Prestin		EST GeneChip CAGE RNA-seq	525 – 713 STAS domain	
<i>SMAC</i>		EST GeneChip CAGE RNA-seq		
<i>SMPX</i> Small muscular protein		EST GeneChip CAGE RNA-seq		
<i>STRC</i> Stereocilin		EST GeneChip CAGE RNA-seq		
<i>TBC1D24</i> TBC1 domain family member 24		EST GeneChip CAGE RNA-seq	47 – 262 Rab-GAP TBC domain 368 – 554 TLD domain	
<i>TECTA</i> Alpha-tectorin		EST GeneChip CAGE RNA-seq	98 – 252 NIDO domain 260 – 314 VWFC domain 321 – 540 VWFD 1 domain 597 – 650 TIL 1 domain 712 – 929 VWFD 2 domain 984 – 1036 TIL 2 domain 1099 – 1317 VWFD 3 domain 1372 – 1425 TIL 3 domain 1486 – 1694 VWFD 4 domain 1805 – 2059 ZP domain	
<i>TJP2</i> Tight junction protein ZO 2		EST GeneChip CAGE RNA-seq	33 – 120 PDZ 1 domain 307 – 385 PDZ 2 domain 509 – 590 PDZ 3 domain 604 – 669 SH3 domain 678 – 876 Guanylate kinase-like domain 1188 – 1190 Interaction with SCRIB region	LASPI

(continued)

**Table 4. (continued)**

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<i>TMCI</i> Transmembrane channel-like protein 1		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: [blue line]		
<i>TMIE</i> Transmembrane inner ear-expressed protein		EST: [blue line] GeneChip: nodata CAGE: nodata RNA-seq: [blue line]		
<i>TMPRSS3</i> NM_024022 Transmembrane protease serine 3		EST: [blue line] GeneChip: [green bars] CAGE: [blue line] RNA-seq: [blue line]	72 – 108 LDL- receptor class A domain 109 – 205 SRCR domain 217 – 449 Peptidase S1 domain	
<i>TMPRSS3</i> NM_032405 Transmembrane protease serine 3		EST: [blue line] GeneChip: nodata CAGE: [blue line] RNA-seq: [blue line]	72 – 108 LDL- receptor class A domain 109 – 205 SRCR domain 217 – 449 Peptidase S1 domain	
<i>TNC</i> Tenascin		EST: [blue line] GeneChip: [green bars] CAGE: [blue line] RNA-seq: [blue line]	118 – 145 coiled coil 174 – 186 EGF-like 1 domain; incomplete 186 – 217 EGF-like 2 domain 217 – 248 EGF-like 3 domain 248 – 280 EGF-like 4 domain 280 – 311 EGF-like 5 domain 311 – 342 EGF-like 6 domain 342 – 373 EGF-like 7 domain 373 – 404 EGF-like 8 domain 404 – 435 EGF-like 9 domain 435 – 466 EGF-like 10 domain 466 – 497 EGF-like 11 domain 497 – 528 EGF-like 12 domain 528 – 559 EGF-like 13 domain 559 – 590 EGF-like 14 domain 590 – 621 EGF-like 15 domain 625 – 715 Fibronectin type-III 1 domain 716 – 804 Fibronectin type-III 2 domain 805 – 894 Fibronectin type-III 3 domain 895 – 990 Fibronectin type-III 4 domain 991 – 1075 Fibronectin type-III 5 domain 1076 – 1165 Fibronectin type-III 6 domain 1167 – 1256 Fibronectin type-III 7 domain 1258 – 1350 Fibronectin type-III 8 domain 1351 – 1439 Fibronectin type-III 9 domain 1440 – 1531 Fibronectin type-III 10 domain 1533 – 1621 Fibronectin type-III 11 domain 1622 – 1711 Fibronectin type-III 12 domain 1712 – 1801 Fibronectin type-III 13 domain 1802 – 1888 Fibronectin type-III 14 domain 1889 – 1977 Fibronectin type-III 15 domain 1975 – 2190 Fibrinogen C-terminal domain	
<i>TPRN</i> Taperin		EST: [blue line] GeneChip: [orange bars] CAGE: [blue line] RNA-seq: [blue line]		

(continued)

Table 4. (continued)

Gene Name	Image	Gene Expression Profiles (Brain, Blood, Connective Tissue, Reproductive Organs, Muscle, Digestive Organs, Liver, Lung, Kidney, Urinary Organs)	Domains	Interaction Proteins
<i>TSPEAR</i> Thrombospondin-type laminin G domain and EAR repeat-containing protein		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata	58 – 277 Laminin G-like domain 313 – 358 EAR 1 repeat 359 – 408 EAR 2 repeat 411 – 460 EAR 3 repeat 463 – 512 EAR 4 repeat 513 – 570 EAR 5 repeat 573 – 622 EAR 6 repeat 624 – 668 EAR 7 repeat	
<i>USH1C</i> Harmonin		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata	189 – 227 coiled coil 289 – 309 coiled coil 476 – 513 coiled coil 596 – 681 coiled coil	BET1, EXOC, NOC4L, SERTAD3
<i>USH2A</i> Usherin		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata	271 – 517 Laminin N-terminal domain 518 – 574 Laminin EGF-like 1 domain 5075 – 640 Laminin EGF-like 2 domain 641 – 693 Laminin EGF-like 3 domain 694 – 746 Laminin EGF-like 4 domain 747 – 794 Laminin EGF-like 5 domain 795 – 846 Laminin EGF-like 6 domain 847 – 899 Laminin EGF-like 7 domain 900 – 950 Laminin EGF-like 8 domain 951 – 1001 Laminin EGF-like 9 domain 1002 – 1052 Laminin EGF-like 10 domain 1058 – 1146 Fibronectin type-III 1 domain 1148 – 1244 Fibronectin type-III 2 domain 1245 – 1363 Fibronectin type-III 3 domain 1364 – 1468 Fibronectin type-III 4 domain 1517 – 1709 Laminin G-like 1 domain 1714 – 1891 Laminin G-like 2 domain 1869 – 1955 Fibronectin type-III 5 domain 1957 – 2054 Fibronectin type-III 6 domain 2055 – 2144 Fibronectin type-III 7 domain 2145 – 2239 Fibronectin type-III 8 domain 2243 – 2330 Fibronectin type-III 9 domain 2331 – 2433 Fibronectin type-III 10 domain 2437 – 2531 Fibronectin type-III 11 domain 2535 – 2622 Fibronectin type-III 12 domain 2624 – 2722 Fibronectin type-III 13 domain 2726 – 2819 Fibronectin type-III 14 domain 2820 – 2923 Fibronectin type-III 15 domain	
<i>WFS1</i> NM_006005 Wolframin		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata		
<i>WFS1</i> NM_001145805 Wolframin		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata		
<i>WHRN</i> Whirlin		EST: nodata GeneChip: nodata CAGE: nodata RNA-seq: nodata	140 – 223 PDZ 1 domain 279 – 361 PDZ 2 domain 816 – 899 PDZ 3 domain	

<sup>a</sup>Expression profile analysis for each human organ was performed using the RefEX database as described previously.

## Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

## Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This study was funded by a Health and Labour Sciences Research Grant for Research on Rare and Intractable Diseases and Comprehensive Research on Disability Health and Welfare from the Ministry of Health, Labour and Welfare of Japan (S.U.) and by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science and Culture of Japan (S.U.).

## References

- Morton CC, Nance WE. Newborn hearing screening: a silent revolution. *N Engl J Med*. 2006;354:2151-2164.
- Hereditary Hearing Loss homepage. <http://hereditaryhearingloss.org/>. Accessed February 25, 2015.
- Thomas PD, Campbell MJ, Kejariwal A, et al. PANTHER: a library of protein families and subfamilies indexed by function. *Genome Res*. 2003;13:2129-2141.
- Mi H, Lazareva-Ulitsky B, Loo R, et al. The PANTHER database of protein families, subfamilies, functions and pathways. *Nucleic Acids Res*. 2005;33:D284-D288.
- Rebhan M, Chalifa-Caspi V, Prilusky J, Lancet D. GeneCards: integrating information about genes, proteins and diseases. *Trends Genet*. 1997;13:163.
- Reference Expression Data Set. <http://refex.dbcls.jp/index.php?lang=en>. Accessed February 25, 2015.
- UniProt homepage. [www.uniprot.org](http://www.uniprot.org). Accessed February 25, 2015.
- Steel KP, Kros CJ. A genetic approach to understanding auditory function. *Nat Genet*. 2001;27(2):143-149.
- Gillespie PG, Müller U. Mechanotransduction by hair cells: models, molecules, and mechanisms. *Cell*. 2009;139:33-44.
- Zdebik AA, Wangemann P, Jentsch TJ. Potassium ion movement in the inner ear: insights from genetic disease and mouse models. *Physiology*. 2009;24:307-316.
- Kazmierczak P, Müller U. Sensing sound: molecules that orchestrate mechanotransduction by hair cells. *Trends Neurosci*. 2012;35:220-229.
- Marcotti W. Functional assembly of mammalian cochlear hair cells. *Exp Physiol*. 2012;97:438-451.
- Hudspeth AJ. Integrating the active process of hair cells with cochlear function. *Nat Rev Neurosci*. 2014;15:600-614.
- Feng X, Qin Z. Molecular analysis of hair cells in sensorineural hearing loss. *Audiol Neurootol*. 2014;19:267-274.
- Fettiplace R, Kim KX. The physiology of mechano-electrical transduction channels in hearing. *Physiol Rev*. 2014;94:951-986.
- Sakaguchi H, Tokita J, Müller U, Kachar B. Tip links in hair cells: molecular composition and role in hearing loss. *Curr Opin Otolaryngol Head Neck Surg*. 2009;17:388-393.
- Holt JR, Pan B, Koussa MA, Asai Y. TMC function in hair cell transduction. *Hear Res*. 2014;311:17-24.
- Pan B, Géléoc GS, Asai Y, et al. *TMC1* and *TMC2* are components of the mechanotransduction channel in hair cells of the mammalian inner ear. *Neuron*. 2013;79:504-515.
- Beutner D, Voets T, Neher E, Moser T. Calcium dependence of exocytosis and endocytosis at the cochlear inner hair cell afferent synapse. *Neuron*. 2001;29:681-690.
- Schmitz F. The making of synaptic ribbons: how they are built and what they do. *Neuroscientist*. 2009;15:611-622.
- Roux I, Safieddine S, Nouvian R, et al. Otoferlin, defective in a human deafness form, is essential for exocytosis at the auditory ribbon synapse. *Cell*. 2006;127:227-289.
- Schrauwen I, Helfmann S, Inagaki A, et al. A mutation in *CABP2*, expressed in cochlear hair cells, causes autosomal-recessive hearing impairment. *Am J Hum Genet*. 2012;91:636-645.
- Riazuddin S, Beltantseva IA, Giese AP, et al. Alterations of the *CIB2* calcium- and integrin-binding protein cause Usher syndrome type 1J and nonsyndromic deafness DFNB48. *Nat Genet*. 2012;44:1265-1271.
- Yasunaga S, Grati M, Cohen-Salmon M, et al. A mutation in *OTOF*, encoding otoferlin, a FER-1-like protein, causes DFNB9, a nonsyndromic form of deafness. *Nat Genet*. 1999;21:363-369.
- Robertson NG, Lu L, Heller S, et al. Mutations in a novel cochlear gene cause DFNA9, a human nonsyndromic deafness with vestibular dysfunction. *Nat Genet*. 1998;20:299-303.
- Azaiez H, Booth JT, Bu F, et al. *TBC1D24* mutation causes autosomal-dominant nonsyndromic hearing loss. *Hum Mutat*. 2014;35:819-823.
- Scott HS, Kudoh J, Wattenhofer M, et al. Insertion of beta-satellite repeats identifies a transmembrane protease causing both congenital and childhood onset autosomal recessive deafness. *Nat Genet*. 2001;27:59-63.
- Zhu M, Yang T, Wei S, et al. Mutations in the gamma-actin gene (*ACTG1*) are associated with dominant progressive deafness (DFNA20/26). *Am J Hum Genet*. 2003;73:1082-1091.
- Bork JM, Peters LM, Riazuddin S, et al. Usher syndrome 1D and nonsyndromic autosomal recessive deafness DFNB12 are caused by allelic mutations of the novel cadherin-like gene *CDH23*. *Am J Hum Genet*. 2001;68:26-37.
- Naz S, Griffith AJ, Riazuddin S, et al. Mutations of *ESPN* cause autosomal recessive deafness and vestibular dysfunction. *J Med Genet*. 2004;41:591-595.
- Walsh T, Walsh V, Vreugde S, et al. From flies' eyes to our ears: mutations in a human class III myosin cause progressive nonsyndromic hearing loss DNNB30. *Proc Natl Acad Sci USA*. 2002;99:7518-7523.
- Liu XZ, Walsh J, Mburu P, et al. Mutations in the myosin VIIA gene cause non-syndromic recessive deafness. *Nat Genet*. 1997;16:188-190.
- Wang A, Liang Y, Fridell RA, et al. Association of unconventional myosin *MYO15* mutations with human nonsyndromic deafness DFNB3. *Science*. 1998;280:1447-1551.
- Ahmed ZM, Riazuddin S, Ahmad J, et al. *PCDH15* is expressed in the neurosensory epithelium of the eye and ear and mutant alleles are responsible for both USH1F and DFNB23. *Hum Mol Genet*. 2003;12:3215-3223.

35. Ebermann I, Phillips JB, Liebau et al. *PDZD7* is a modifier of retinal disease and a contributor digenic Usher syndrome. *J Clin Invest*. 2010;120:1812-1823.
36. Khan SY, Ahmed ZM, Shabbir MI, et al. Mutations of the *RDX* gene cause nonsyndromic hearing loss at the DFNB24 locus. *Hum Mutat*. 2007;28:417-423.
37. Verpy E, Masmoudi S, Zwaenepoel I, et al. Mutations in a new gene encoding a protein of the hair bundle cause non-syndromic deafness at the DFNB16 locus. *Nat Genet*. 2001;29:345-349.
38. Kurima K, Peters LM, Yang Y, et al. Dominant and recessive deafness caused by mutations of a novel gene, *TMC1*, required for cochlear hair-cell function. *Nat Genet*. 2002;30:277-284.
39. Mutations in a novel isoform of *TRIOBP* that encodes a filamentous-actin binding protein are responsible for DFNB28 recessive nonsyndromic hearing loss. *Am J Hum Genet*. 2006;78:144-152.
40. Ahmed ZM, Smith TN, Riazuddin S, et al. Nonsyndromic recessive deafness DFNB18 and Usher syndrome type 1C are allelic mutations of *USH1C*. *Hum Genet*. 2002;110:527-531.
41. Kimberling WJ, Weston MD, Möller C, et al. Localization of Usher syndrome II to chromosome 1q. *Genomics*. 1990;7:245-249.
42. Mburu P, Mustapha M, Varela A, et al. Defects in whirlin, a PDZ domain molecule involved in stereocilia elongation, cause deafness in the whirler mouse and families with DFNB31. *Nat Genet*. 2003;34:421-428.
43. Grillet N, Schwander M, Hildebrand MS, et al. Mutations in *LOXHD1*, an evolutionarily conserved stereociliary protein, disrupt hair cell function in mice and cause progressive hearing loss in humans. *Am J Hum Genet*. 2009;85:328-337.
44. Schraders M, Oostrik J, Huygen PL, et al. Mutations in *PTPRQ* are a cause of autosomal-recessive nonsyndromic hearing impairment DFNB84 and associated with vestibular dysfunction. *Am J Hum Genet*. 2010;86:604-610.
45. Ain Q, Nazil S, Riazuddin S, et al. The autosomal recessive nonsyndromic deafness locus DFNB72 is located on chromosome 19p13.3. *Hum Genet*. 2007;122:445-450.
46. Ruf RG, Berkman J, Wolf MT, et al. A gene locus for branchio-otic syndrome maps to chromosome 14q21.3-q24.3. *J Med Genet*. 2003;40:515-519.
47. Pingault V, Bondurand N, Kuhlbrodt K, et al. *SOX10* mutations in patients with Waardenburg-Hirschsprung disease. *Nat Genet*. 1998;18:171-173.
48. Vahava O, Morell R, Lynch ED, et al. Mutation in transcription factor *POU4F3* associated with inherited progressive hearing loss in humans. *Science*. 1998;279:1950-1954.
49. Edery P, Attié T, Amie J, et al. Mutation of the endothelin-3 gene in the Waardenburg-Hirschsprung disease (Shah-Waardenburg syndrome). *Nat Genet*. 1996;12:442-444.
50. Attié T, Till M, Pelet A, et al. Mutation of the endothelin-receptor B gene in Waardenburg-Hirschsprung disease. *Hum Mol Genet*. 1995;4:2407-2409.
51. Behlouli A, Bonnet C, Abdi S, et al. *EPS8*, encoding and actin-binding protein of cochlear hair cell stereocilia, is a new causal gene for autosomal recessive profound deafness. *Orphanet J Rare Dis*. 2014;9:55.
52. Walsh T, Shahin H, Elkan-Miller T, et al. Whole exome sequencing and homozygosity mapping identify mutation in the cell polarity protein *GPSM2* as the cause of nonsyndromic hearing loss DFNB82. *Am J Hum Genet*. 2010;87:90-94.
53. Riazuddin S, Ahmed ZM, Fanning AS, et al. Tricellulin is a tight-junction protein necessary for hearing. *Am J Hum Genet*. 2006;79:1040-1051.
54. Walsh T, Pierce SB, Lenz DR, et al. Genomic duplication and overexpression of *TJP2/ZO-2* leads to altered expression of apoptosis genes in progressive nonsyndromic hearing loss DFNA51. *Am J Hum Genet*. 2010;87:101-109.
55. Wilcox ER, Burton QL, Naz S, et al. Mutations in the gene encoding tight junction claudin-14 cause autosomal recessive deafness DFNB29. *Cell*. 2001;104:165-172.
56. Zheng J, Shen W, He DZ, Long KB, Madison LD, Dallos P. Prestin is the motor protein of cochlear outer hair cells. *Nature*. 2000;405:149-155.
57. Hudspeth AJ. Integrating the active process of hair cells with cochlear function. *Nat Rev Neurosci*. 2014;15:600-614.
58. Keller JP, Homma K, Duan C, Zheng J, Cheatham MA, Dallos P. Functional regulation of the SLC26-family protein prestin by calcium/calmodulin. *J Neurosci*. 2014;34:1325-1332.
59. Furness DN, Katori Y, Mahendrasingam S, Hackney CM. Differential distribution of beta- and gamma-actin in guinea-pig cochlear sensory and supporting cells. *Hear Res*. 2005;207:22-34.
60. Ben-Yosef T, Belyantseva IA, Saunders TL, et al. Claudin 14 knockout mice, a model for autosomal recessive deafness DFNB29, are deaf due to cochlear hair cell degeneration. *Hum Mol Genet*. 2003;12:2049-2061.
61. Forge A, Becker D, Casalotti S, Edwards J, Marziano N, Nevill G. Gap junctions in the inner ear: comparison of distribution patterns in different vertebrates and assessment of connexin composition in mammals. *J Comp Neurol*. 2003;467:207-231.
62. Oguchi T, Suzuki N, Hashimoto S, et al. Inner hair cells of mice express the glutamine transporter *SAT1*. *Hear Res*. 2012;292:59-63.
63. Furness DN, Lehre KP. Immunocytochemical localization of a high-affinity glutamate-aspartate transporter, GLAST, in the rat and guinea-pig cochlea. *Eur J Neurosci*. 1997;9:1961-1969.
64. Furness DN, Lawton DM. Comparative distribution of glutamate transporters and receptors in relation to afferent innervation density in the mammalian cochlea. *J Neurosci*. 2003;23:11296-11304.
65. Kwon TJ, Cho HJ, Kim UK, et al. Methionine sulfoxide reductase B3 deficiency causes hearing loss due to stereocilia degeneration and apoptotic cell death in cochlear hair cells. *Hum Mol Genet*. 2014;23:1591-1601.
66. Kelsell DP, Dunlop J, Stevens HP, et al. Connexin 26 mutations in hereditary non-syndromic sensorineural deafness. *Nature*. 1997;387:80-83.
67. del Castillo I, Villamar M, Moreno-Pelayo MA, et al. A deletion involving the connexin 30 gene in nonsyndromic hearing impairment. *N Engl J Med*. 2002;346:243-249.
68. Li XC, Everett LA, Lalwani AK, et al. A mutation in PDS causes non-syndromic recessive deafness. *Nat Genet*. 1998;18:215-217.

69. Usami S, Abe S, Weston MD, Shinkawa H, Van Camp G, Kimberling WJ. Non-syndromic hearing loss associated with enlarged vestibular aqueduct is caused by PDS mutations. *Hum Genet.* 1999;104:188-192.
70. Everett LA, Glaser B, Beck JC, et al. Pendred syndrome is caused by mutations in a putative sulphate transporter gene (PDS). *Nat Genet.* 1997;17:411-422.
71. Hibino H, Horio Y, Inanobe A, et al. An ATP-dependent inwardly rectifying potassium channel, KAB-2 (Kir4. 1), in cochlear stria vascularis of inner ear: its specific subcellular localization and correlation with the formation of endocochlear potential. *J Neurosci.* 1997;17:4711-4721.
72. Nin F, Hibino H, Doi K, Suzuki T, Hisa Y, Kurachi Y. The endocochlear potential depends on two K<sup>+</sup> diffusion potentials and an electrical barrier in the stria vascularis of the inner ear. *Proc Natl Acad Sci U S A.* 2008;105:1751-1756.
73. Neyroud N, Tesson F, Denjoy I, et al. A novel mutation in the potassium channel gene *KVLQT1* causes the Jervell and Lange-Nielsen cardioauditory syndrome. *Nat Genet.* 1997;15:186-189.
74. Tyson J, Tranebjaerg L, Bellman S, et al. *IsK* and *KvLQT1*: mutation in either of the two subunits of the slow component of the delayed rectifier potassium channel can cause Jervell and Lange-Nielsen syndrome. *Hum Mol Genet.* 1997;6:2179-2185.
75. Schulze-Bahr E, Wang Q, Wedekind H, et al. *KCNE1* mutations cause jervell and Lange-Nielsen syndrome. *Nat Genet.* 1997;17:267-268.
76. Tassabehji M, Read AP, Newton VE, et al. Waardenburg's syndrome patients have mutations in the human homologue of the Pax-3 paired box gene. *Nature.* 1992;355:635-636.
77. Tassabehji M, Newton VE, Read AP. Waardenburg syndrome type 2 caused by mutations in the human microphthalmia (*MITF*) gene. *Nat Genet.* 1994;8:251-255.
78. Watanabe A, Takeda K, Ploplis B, Tachibana M. Epistatic relationship between Waardenburg syndrome genes *MITF* and *PAX3*. *Nat Genet.* 1998;18:283-286.
79. Tachibana M, Takeda K, Nobukuni Y, et al. Ectopic expression of *MITF*, a gene for Waardenburg syndrome type 2, converts fibroblasts to cells with melanocyte characteristics. *Nat Genet.* 1996;14:50-54.
80. Cosgrove D, Samuelson G, Pinnt J. Immunohistochemical localization of basement membrane collagens and associated proteins in the murine cochlea. *Hear Res.* 1996;97:54-65.
81. Goodyear RJ, Richardson GP. Extracellular matrices associated with the apical surfaces of sensory epithelia in the inner ear: molecular and structural diversity. *J Neurobiol.* 2002;53:212-227.
82. Usami S, Takumi Y, Suzuki N, et al. The localization of proteins encoded by *CRYM*, *KIAA1199*, *UBA52*, *COL9A3*, and *COL9A1*, genes highly expressed in the cochlea. *Neuroscience.* 2008;154:22-28.
83. Shpargel KB, Makishima T, Griffith AJ. *Col11a1* and *Col11a2* mRNA expression in the developing mouse cochlea: implications for the correlation of hearing loss phenotype with mutant type XI collagen genotype. *Acta Otolaryngol.* 2004;124:242-248.
84. McGuirt WT, Prasad SD, Griffith AJ, et al. Mutations in *COL11A2* cause non-syndromic hearing loss (DFNA13). *Nat Genet.* 1999;23:413-419.
85. Cosgrove D, Kornak JM, Samuelson G. Expression of basement membrane type IV collagen chains during postnatal development in the murine cochlea. *Hear Res.* 1996;100:21-32.
86. Zehnder AF, Adams JC, Santi PA, et al. Distribution of type IV collagen in the cochlea in Alport syndrome. *Arch Otolaryngol Head Neck Surg.* 2005;131:1007-1013.
87. Verhoeven K, Van Laer L, Kirschhofer K, et al. Mutations in the human alpha-tectorin gene cause autosomal dominant non-syndromic hearing impairment. *Nat Genet.* 1998;19:60-62.
88. Yariz KO, Duman D, Seco CZ, et al. Mutations in *OTOGL*, encoding the inner ear protein otogelin-like, cause moderate sensorineural hearing loss. *Am J Hum Genet.* 2012;91:872-882.
89. Schraders M, Ruiz-Palmero L, Kalay E, et al. Mutations of the gene encoding otogelin are a cause of autosomal-recessive nonsyndromic moderate hearing impairment. *Am J Hum Genet.* 2012;91:883-889.
90. Wilson SM, Householder DB, Coppola V, et al. Mutations in *Cdh23* cause nonsyndromic hearing loss in waltzer mice. *Genomics.* 2001;74:228-233.
91. Fransen E, Van Camp G. The *COCH* gene: a frequent cause of hearing impairment and vestibular dysfunction? *Br J Audiol.* 1999;33:297-302.
92. Usami SI, Takahashi K, Yuge I, et al. Mutations in the *COCH* gene are a frequent cause of autosomal dominant progressive cochleo-vestibular dysfunction, but not of Meniere's disease. *Eur J Hum Genet.* 2003;11:744-748.
93. Suzuki H, Oshima A, Tsukamoto K, et al. Clinical characteristics and genotype-phenotype correlation of hearing loss patients with *SLC26A4* mutations. *Acta Oto Laryngologica.* 2007;127:1292-1297.
94. Miyagawa M, Nishio S, Usami SI, the Deafness Gene Study Consortium. Mutation spectrum and genotype-phenotype correlation of hearing loss patients caused by *SLC26A4* mutations in the Japanese: a large cohort study. *J Hum Gen.* 2014;59:262-268.
95. Miyagawa M, Nishio SY, Usami S. Prevalence and clinical features of hearing loss patients with *CDH23* mutations: a large cohort study. *PLoS One.* 2012;7:e40366.
96. Manji SS, Miller KA, Williams LH, et al. An ENU-induced mutation of *Cdh23* causes congenital hearing loss, but no vestibular dysfunction, in mice. *Am J Pathol.* 2011;179:903-914.
97. Yasunaga S, Grati M, Chardenoux S, et al. *OTOF* encodes multiple long and short isoforms: genetic evidence that the long ones underlie recessive deafness DFNB9. *Am J Hum Genet.* 2000;67:591-600.
98. Ahmed M, Wong EY, Sun J, et al. Eya1-Six1 interaction is sufficient to induce hair cell fate in the cochlea by activating *Atoh1* expression in cooperation with *Sox2*. *Dev Cell.* 2012;22:377-390.
99. Yokoyama S, Takeda K, Shibahara S. Functional difference of the *SOX10* mutant proteins responsible for the phenotypic variability in auditory-pigmentary disorders. *J Biochem.* 2006;140:491-499.
100. Belyantseva IA, Perrin BJ, Sonnemann KJ, et al. Gamma-actin is required for cytoskeletal maintenance but not development. *Proc Natl Acad Sci U S A.* 2009;106:9703-9708.

101. Drescher MJ, Khan KM, Beisel KW, et al. Expression of adenylyl cyclase type I in cochlear inner hair cells. *Brain Res Mol Brain Res*. 1997;45:325-330.
102. Santos-Cortez RL, Lee K, Giese AP, et al. Adenylate cyclase 1 (*ADCY1*) mutations cause recessive hearing impairment in humans and defects in hair cell function and hearing in zebrafish. *Hum Mol Genet*. 2014;23:3289-3298.
103. Giroto G, Abdulhadi K, Buniello A, et al. Linkage study and exome sequencing identify a *BDP1* mutation associated with hereditary hearing loss. *PLoS One*. 2013;8:e80323.
104. Cui G, Meyer AC, Calin-Jageman I, et al. Ca<sup>2+</sup>-binding proteins tune Ca<sup>2+</sup>-feedback to Cav1.3 channels in mouse auditory hair cells. *J Physiol*. 2007;585:791-803.
105. Modamio-Hoybjor S, Mencia A, Goodyear R, et al. A mutation in *CCDC50*, a gene encoding an effector of epidermal growth factor-mediated cell signaling, causes progressive hearing loss. *Am J Hum Genet*. 2007;80:1076-1089.
106. Wilson SM, Householder DB, Coppola V, et al. Mutations in *Cdh23* cause nonsyndromic hearing loss in waltzer mice. *Genomics*. 2001;74:228-233.
107. Kazmierczak P, Sakaguchi H, Tokita J, et al. Cadherin 23 and protocadherin 15 interact to form tip-link filaments in sensory hair cells. *Nature*. 2007;449:87-91.
108. Kammerer R, Rüttiger L, Riesenberger R, et al. Loss of mammal-specific tectorial membrane component carcinoembryonic antigen cell adhesion molecule 16 (*CEACAM16*) leads to hearing impairment at low and high frequencies. *J Biol Chem*. 2012;287:21584-21598.
109. Cheatham MA, Goodyear RJ, Homma K, et al. Loss of the tectorial membrane protein *CEACAM16* enhances spontaneous, stimulus-frequency, and transiently evoked otoacoustic emissions. *J Neurosci*. 2014;34:10325-10338.
110. Hurd EA, Adams ME, Layman WS, et al. Mature middle and inner ears express *Chd7* and exhibit distinctive pathologies in a mouse model of CHARGE syndrome. *Hear Res*. 2011;282:184-195.
111. Wilcox ER, Burton QL, Naz S, et al. Mutations in the gene encoding tight junction claudin-14 cause autosomal recessive deafness DFNB29. *Cell*. 2001;104:165-172.
112. Gagnon LH, Longo-Guess CM, Berryman M, et al. The chloride intracellular channel protein *CLIC5* is expressed at high levels in hair cell stereocilia and is essential for normal inner ear function. *J Neurosci*. 2006;26:10188-10198.
113. Salles FT, Andrade LR, Tanda S, et al. *CLIC5* stabilizes membrane-actin filament linkages at the base of hair cell stereocilia in a molecular complex with radixin, taperin, and myosin VI. *Cytoskeleton (Hoboken)*. 2014;71:61-78.
114. Jenkinson EM, Rehman AU, Walsh T, et al. Perrault syndrome is caused by recessive mutations in *CLPP*, encoding a mitochondrial ATP-dependent chambered protease. *Am J Hum Genet*. 2013;92:605-613.
115. Geng R, Geller SF, Hayashi T, et al. Usher syndrome IIIA gene *clarin-1* is essential for hair cell function and associated neural activation. *Hum Mol Genet*. 2009;18:2748-2760.
116. Adato A, Vreugde S, Joensuu T, et al. *USH3A* transcripts encode *clarin-1*, a four-transmembrane-domain protein with a possible role in sensory synapses. *Eur J Hum Genet*. 2002;10:339-350.
117. Zallocchi M, Meehan DT, Delimont D, et al. Localization and expression of *clarin-1*, the *Clrn1* gene product, in auditory hair cells and photoreceptors. *Hear Res*. 2009;255:109-120.
118. Zallocchi M, Meehan DT, Delimont D, et al. Role for a novel Usher protein complex in hair cell synaptic maturation. *PLoS One*. 2012;7:e30573.
119. Robertson NG, Resendes BL, Lin JS, et al. Inner ear localization of mRNA and protein products of *COCH*, mutated in the sensorineural deafness and vestibular disorder, DFNA9. *Hum Mol Genet*. 2001;10:2493-2500.
120. Robertson NG, Jones SM, Sivakumaran TA, et al. A targeted *Coch* missense mutation: a knock-in mouse model for DFNA9 late-onset hearing loss and vestibular dysfunction. *Hum Mol Genet*. 2008;17:3426-3434.
121. Goodyear RJ, Richardson GP. Extracellular matrices associated with the apical surfaces of sensory epithelia in the inner ear: molecular and structural diversity. *J Neurobiol*. 2002;53:212-227.
122. Cosgrove D, Samuelson G, Meehan DT, et al. Ultrastructural, physiological, and molecular defects in the inner ear of a gene-knockout mouse model for autosomal Alport syndrome. *Hear Res*. 1998;121:84-98.
123. Rost S, Bach E, Neuner C, et al. Novel form of X-linked nonsyndromic hearing loss with cochlear malformation caused by a mutation in the type IV collagen gene *COL4A6*. *Eur J Hum Genet*. 2014;22:208-215.
124. Asamura K, Abe S, Imamura Y, et al. Type IX collagen is crucial for normal hearing. *Neuroscience*. 2005;132:493-500.
125. Oshima A, Suzuki S, Takumi Y, Hashizume K, Abe S, Usami S. *CRYM* mutations cause deafness through thyroid hormone binding properties in the fibrocytes of the cochlea. *J Med Genet*. 2006;43:e25.
126. Delmaghani S, del Castillo FJ, Michel V, et al. Mutations in the gene encoding *pejvakin*, a newly identified protein of the afferent auditory pathway, cause DFNB59 auditory neuropathy. *Nat Genet*. 2006;38:770-778.
127. Schwander M, Sczaniecka A, Grillet N, et al. A forward genetics screen in mice identifies recessive deafness traits and reveals that *pejvakin* is essential for outer hair cell function. *J Neurosci*. 2007;27:2163-2175.
128. Franz P, Hauser-Kronberger C, Egerbacher M, et al. Localization of endothelin-1 and endothelin-3 in the cochlea. *Acta Otolaryngol*. 1997;117:358-362.
129. Jaworek TJ, Richard EM, Ivanova AA, et al. An alteration in *ELMOD3*, an Arl2 GTPase-activating protein, is associated with hearing impairment in humans. *PLoS Genet*. 2013;9:e1003774.
130. Furness DN, Johnson SL, Manor U, et al. Progressive hearing loss and gradual deterioration of sensory hair bundles in the ears of mice lacking the actin-binding protein *Eps8L2*. *Proc Natl Acad Sci U S A*. 2013;110:13898-13903.
131. Zheng L, Sekerková G, Vranich K, Tilney LG, Mugnaini E, Bartles JR. The deaf jerker mouse has a mutation in the gene encoding the *espin* actin-bundling proteins of hair cell stereocilia and lacks *espins*. *Cell*. 2000;102:377-385.
132. Collin RW, Kalay E, Tariq M, et al. Mutations of *ESRRB* encoding estrogen-related receptor beta cause autosomal-recessive nonsyndromic hearing impairment DFNB35. *Am J Hum Genet*. 2008;82:125-138.



133. Abdelhak S, Kalatzis V, Heilig R, et al. A human homologue of the *Drosophila* eyes absent gene underlies branchio-otorenal (BOR) syndrome and identifies a novel gene family. *Nat Genet.* 1997;15:157-164.
134. Wayne S, Robertson NG, DeClau F, et al. Mutations in the transcriptional activator *EYA4* cause late-onset deafness at the DFNA10 locus. *Hum Mol Genet.* 2001;10:195-200.
135. Charizopoulou N, Lelli A, Schraders M, et al. Gipc3 mutations associated with audiogenic seizures and sensorineural hearing loss in mouse and human. *Nat Commun.* 2011;2:201.
136. Lautermann J, ten Cate WJ, Altenhoff P, et al. Expression of the gap-junction connexins 26 and 30 in the rat cochlea. *Cell Tissue Res.* 1998;294(3):415-420.
137. López-Bigas N, Olivé M, Rabionet R, et al. Connexin 31 (*GJB3*) is expressed in the peripheral and auditory nerves and causes neuropathy and hearing impairment. *Hum Mol Genet.* 2001;10:947-952.
138. McGee J, Goodyear RJ, McMillan DR, et al. The very large G-protein-coupled receptor *VLGRI*: a component of the ankle link complex required for the normal development of auditory hair bundles. *J Neurosci.* 2006;26:6543-6553.
139. Peters LM, Anderson DW, Griffith AJ, et al. Mutation of a transcription factor, *TFCP2L3*, causes progressive autosomal dominant hearing loss, DFNA28. *Hum Mol Genet.* 2002;11:2877-2885.
140. Odeh H, Hunker KL, Belyantseva IA, et al. Mutations in *Grxcr1* are the basis for inner ear dysfunction in the pirouette mouse. *Am J Hum Genet.* 2010;86:148-160.
141. Avenarius MR. The glutaredoxin-like cysteine-rich family of genes, *Grxcr1* and *Grxcr2*, in stereocilia development and function. Doctor thesis, University of Michigan; 2012.
142. Borck G, Ur Rehman A, Lee K, et al. Loss-of-function mutations of *ILDR1* cause autosomal-recessive hearing impairment DFNB42. *Am J Hum Genet.* 2011;88:127-137.
143. Santos-Cortez RL, Lee K, Azeem Z, et al. Mutations in *KARS*, encoding lysyl-tRNA synthetase, cause autosomal-recessive nonsyndromic hearing impairment DFNB89. *Am J Hum Genet.* 2013;93:132-140.
144. Knipper M, Claussen C, Rüttiger L, et al. Deafness in *LIMP2*-deficient mice due to early loss of the potassium channel *KCNQ1/KCNE1* in marginal cells of the stria vascularis. *J Physiol.* 2006;576:73-86.
145. Melnick M, Jaskoll T. An in vitro mouse model of congenital cytomegalovirus-induced pathogenesis of the inner ear cochlea. *Birth Defects Res A Clin Mol Teratol.* 2013;97:69-78.
146. Fang Q, Giordimaina AM, Dolan DF, Camper SA, Mustapha M. Genetic background of *Prop1(df)* mutants provides remarkable protection against hypothyroidism-induced hearing impairment. *J Assoc Res Otolaryngol.* 2012;13:173-184.
147. Kubisch C, Schroeder BC, Friedrich T, et al. *KCNQ4*, a novel potassium channel expressed in sensory outer hair cells, is mutated in dominant deafness. *Cell.* 1999;96:437-446.
148. Shabbir MI, Ahmed ZM, Khan SY, et al. Mutations of human *TMHS* cause recessively inherited non-syndromic hearing loss. *J Med Genet.* 2006;43:634-640.
149. Ahmed ZM, Masmoudi S, Kalay E, et al. Mutations of *LRTOMT*, a fusion gene with alternative reading frames, cause nonsyndromic deafness in humans. *Nat Genet.* 2008;40:1335-1340.
150. Nayak G, Lee SI, Yousaf R, Edelmann SE, et al. Tricellulin deficiency affects tight junction architecture and cochlear hair cells. *J Clin Invest.* 2013;123:4036-4049.
151. Weston MD, Pierce ML, Jensen-Smith HC, et al. MicroRNA-183 family expression in hair cell development and requirement of microRNAs for hair cell maintenance and survival. *Dev Dyn.* 2011;240:808-819.
152. Ahmed ZM, Yousaf R, Lee BC, et al. Functional null mutations of *MSRB3* encoding methionine sulfoxide reductase are associated with human deafness DFNB74. *Am J Hum Genet.* 2011;88:19-29.
153. Lalwani AK, Goldstein JA, Kelley MJ, Luxford W, Castelein CM, Mhatre AN. Human nonsyndromic hereditary deafness DFNA17 is due to a mutation in nonmuscle myosin *MYH9*. *Am J Hum Genet.* 2000;67:1121-1128.
154. Donaudy F, Snoeckx R, Pfister M, et al. Nonmuscle myosin heavy-chain gene *MYH14* is expressed in cochlea and mutated in patients affected by autosomal dominant hearing impairment (DFNA4). *Am J Hum Genet.* 2004;74:770-776.
155. Schneider ME, Dosé AC, Salles FT, et al. A new compartment at stereocilia tips defined by spatial and temporal patterns of myosin IIIa expression. *J Neurosci.* 2006;26:10243-10252.
156. Hertzano R, Shalit E, Rzedzinska AK, et al. A Myo6 mutation destroys coordination between the myosin heads, revealing new functions of myosin VI in the stereocilia of mammalian inner ear hair cells. *PLoS Genet.* 2008;4:e1000207.
157. Grati M, Kachar B. Myosin VIIa and sans localization at stereocilia upper tip-link density implicates these Usher syndrome proteins in mechanotransduction. *Proc Natl Acad Sci U S A.* 2011;108:11476-11481.
158. Belyantseva IA, Boger ET, Friedman TB. Myosin XVa localizes to the tips of inner ear sensory cell stereocilia and is essential for staircase formation of the hair bundle. *Proc Natl Acad Sci U S A.* 2003;100:13958-13963.
159. Ye X, Smallwood P, Nathans J. Expression of the Norrie disease gene (*Ndp*) in developing and adult mouse eye, ear, and brain. *Gene Expr Patterns.* 2011;11:151-155.
160. Zwaenepoel I, Mustapha M, Leibovici M, et al. Otoancorin, an inner ear protein restricted to the interface between the apical surface of sensory epithelia and their overlying acellular gels, is defective in autosomal recessive deafness DFNB22. *Proc Natl Acad Sci U S A.* 2002;99:6240-6245.
161. El-Amraoui A, Cohen-Salmon M, Petit C, Simmler MC. Spatiotemporal expression of otogelin in the developing and adult mouse inner ear. *Hear Res.* 2001;158:151-159.
162. Telang RS, Paramanathasivam V, Vlajkovic SM, et al. Reduced P2x(2) receptor-mediated regulation of endocochlear potential in the ageing mouse cochlea. *Purinergic Signal.* 2010;6:263-272.
163. Kim H, Ankamreddy H, Lee DJ, et al. Pax3 function is required specifically for inner ear structures with melanogenic fates. *Biochem Biophys Res Commun.* 2014;445:608-614.
164. Webb SW, Grillet N, Andrade LR, et al. Regulation of *PCDH15* function in mechanosensory hair cells by alternative splicing of the cytoplasmic domain. *Development.* 2011;138:1607-1617.
165. Grati M, Shin JB, Weston MD, et al. Localization of *PDZD7* to the stereocilia ankle-link associates this scaffolding

- protein with the Usher syndrome protein network. *J Neurosci*. 2012;32:14288-14293.
166. von Ameln S, Wang G, Boulouiz R, et al. A mutation in *PNPT1*, encoding mitochondrial-RNA-import protein PNPase, causes hereditary hearing loss. *Am J Hum Genet*. 2012;91:919-927.
  167. Minowa O, Ikeda K, Sugitani Y, et al. Altered cochlear fibrocytes in a mouse model of DFN3 nonsyndromic deafness. *Science*. 1999;285:1408-1411.
  168. Masuda M, Dulon D, Pak K, et al. Regulation of *POU4F3* gene expression in hair cells by 5' DNA in mice. *Neuroscience*. 2011;197:48-64.
  169. Liu X, Han D, Li J, et al. Loss-of-function mutations in the *PRPS1* gene cause a type of nonsyndromic X-linked sensorineural deafness, DFN2. *Am J Hum Genet*. 2010;86:65-71.
  170. Sakaguchi H, Tokita J, Naoz M, Bowen-Pope D, Gov NS, Kachar B. Dynamic compartmentalization of protein tyrosine phosphatase receptor Q at the proximal end of stereocilia: implication of myosin VI-based transport. *Cell Motil Cytoskeleton*. 2008;65:528-538.
  171. Goodyear RJ, Legan PK, Wright MB, et al. A receptor-like inositol lipid phosphatase is required for the maturation of developing cochlear hair bundles. *J Neurosci*. 2003;23:9208-9219.
  172. Grati M, Kachar B. Myosin VIIa and sans localization at stereocilia upper tip-link density implicates these Usher syndrome proteins in mechanotransduction. *Proc Natl Acad Sci U S A*. 2011;108:11476-11481.
  173. Tan J, Prakash MD, Kaiserman D, Bird PI. Absence of *SERPIN6A* causes sensorineural hearing loss with multiple histopathologies in the mouse inner ear. *Am J Pathol*. 2013;183:49-59.
  174. Zheng W, Huang L, Wei ZB, et al. The role of *Six1* in mammalian auditory system development. *Development*. 2003;130:3989-4000.
  175. Peng Z, Wang GP, Zeng R, Guo JY, Chen CF, Gong SS. Temporospatial expression and cellular localization of *VGLUT3* in the rat cochlea. *Brain Res*. 2013;1537:100-110.
  176. Seal RP, Akil O, Yi E, et al. Sensorineural deafness and seizures in mice lacking vesicular glutamate transporter 3. *Neuron*. 2008;57:263-275.
  177. Everett LA, Morsli H, Wu DK, Green ED. Expression pattern of the mouse ortholog of the Pendred's syndrome gene (*Pds*) suggests a key role for pendrin in the inner ear. *Proc Natl Acad Sci U S A*. 1999;96:9727-9732.
  178. Wangemann P, Itza EM, Albrecht B, et al. Loss of *KCNJ10* protein expression abolishes endocochlear potential and causes deafness in Pendred syndrome mouse model. *BMC Med*. 2004;2:30.
  179. Belyantseva IA, Adler HJ, Curi R, Frolenkov GI, Kachar B. Expression and localization of prestin and the sugar transporter GLUT-5 during development of electromotility in cochlear outer hair cells. *J Neurosci*. 2000;20:RC116.
  180. Cheng J, Zhu Y, He S, et al. Functional mutation of *SMAC/DIABLO*, encoding a mitochondrial proapoptotic protein, causes human progressive hearing loss DFNA64. *Am J Hum Genet*. 2011;89:56-66.
  181. Huebner AK, Gandia M, Frommolt P, et al. Nonsense mutations in *SMPX*, encoding a protein responsive to physical force, result in X-chromosomal hearing loss. *Am J Hum Genet*. 2011;88:621-627.
  182. Wakaoka T, Motohashi T, Hayashi H, et al. Tracing Sox10-expressing cells elucidates the dynamic development of the mouse inner ear. *Hear Res*. 2013;302:17-25.
  183. Horn HF, Brownstein Z, Lenz DR, et al. The LINC complex is essential for hearing. *J Clin Invest*. 2013;123:740-750.
  184. Rehman AU, Santos-Cortez RL, Morell RJ, et al. Mutations in *TBC1D24*, a gene associated with epilepsy, also cause nonsyndromic deafness DFN86. *Am J Hum Genet*. 2014;94:144-152.
  185. Zhang L, Hu L, Chai Y, Pang X, Yang T, Wu H. A dominant mutation in the stereocilia-expressing gene *TBC1D24* is a probable cause for nonsyndromic hearing impairment. *Hum Mutat*. 2014;35:814-818.
  186. Manji SS, Miller KA, Williams LH, Dahl HH. Identification of three novel hearing loss mouse strains with mutations in the *Tmc1* gene. *Am J Pathol*. 2012;180:1560-1569.
  187. Shin MJ, Lee JH, Yu DH, et al. Spatiotemporal expression of *tmc1* in the inner ear of rats during postnatal development. *Comp Med*. 2010;60:288-294.
  188. Guipponi M, Toh MY, Tan J, et al. An integrated genetic and functional analysis of the role of type II transmembrane serine proteases (TMPSs) in hearing loss. *Hum Mutat*. 2008;29:130-141.
  189. Fasquelle L, Scott HS, Lenoir M, et al. *Tmprss3*, a transmembrane serine protease deficient in human DFN8/10 deafness, is critical for cochlear hair cell survival at the onset of hearing. *J Biol Chem*. 2011;286:17383-17397.
  190. Ishiyama A, Mowry SE, Lopez IA, Ishiyama G. Immunohistochemical distribution of basement membrane proteins in the human inner ear from older subjects. *Hear Res*. 2009;254:1-14.
  191. Rehman AU, Morell RJ, Belyantseva IA, et al. Targeted capture and next-generation sequencing identifies *C9orf75*, encoding taperin, as the mutated gene in nonsyndromic deafness DFN79. *Am J Hum Genet*. 2010;86:378-388.
  192. Kitajiri S, Sakamoto T, Belyantseva IA, et al. Actin-bundling protein *TRIOBP* forms resilient rootlets of hair cell stereocilia essential for hearing. *Cell*. 2010;141:786-798.
  193. Delmaghani S, Aghaie A, Michalski N, Bonnet C, Weil D, Petit C. Defect in the gene encoding the EAR/EPTP domain-containing protein *TSPEAR* causes DFN98 profound deafness. *Hum Mol Genet*. 2012;21:3835-3844.
  194. Verpy E, Leibovici M, Zwaenepoel I, et al. A defect in harmonin, a PDZ domain-containing protein expressed in the inner ear sensory hair cells, underlies Usher syndrome type 1C. *Nat Genet*. 2000;26:51-55.
  195. Grillet N, Xiong W, Reynolds A, et al. Harmonin mutations cause mechanotransduction defects in cochlear hair cells. *Neuron*. 2009;62:375-387.
  196. Michalski N, Michel V, Bahloul A, et al. Molecular characterization of the ankle-link complex in cochlear hair cells and its role in the hair bundle functioning. *J Neurosci*. 2007;27:6478-6488.
  197. Cryns K, Thys S, Van Laer L, et al. The *WFS1* gene, responsible for low frequency sensorineural hearing loss and Wolfram syndrome, is expressed in a variety of inner ear cells. *Histochem Cell Biol*. 2003;119:247-256.

198. Wang L, Zou J, Shen Z, Song E, Yang J. Whirlin interacts with espin and modulates its actin-regulatory function: an insight into the mechanism of Usher syndrome type II. *Hum Mol Genet.* 2012;21:692-710.
199. Belyantseva IA, Perrin BJ, Sonnemann KJ, et al.  $\gamma$ -Actin is required for cytoskeletal maintenance but not development. *Proc Natl Acad Sci U S A.* 2009;106:9703-9708.
200. Wilson SM, Householder DB, Coppola V, et al. Mutations in *Cdh23* cause nonsyndromic hearing loss in waltzer mice. *Genomics.* 2001;74:228-233.
201. Johnson KR, Yu H, Ding D, Jiang H, Gagnon LH, Salvi RJ. Separate and combined effects of *Sod1* and *Cdh23* mutations on age-related hearing loss and cochlear pathology in C57BL/6J mice. *Hear Res.* 2010;268:85-92.
202. Kammerer R, Rüttiger L, Riesenberger R, et al. Loss of the mammal-specific tectorial membrane component cea cell adhesion molecule 16 (CEACAM16) leads to hearing impairment at low and high frequencies. *J Biol Chem.* 2012;287:21584-21598.
203. Hurd EA, Poucher HK, Cheng K, Raphael Y, Martin DM. The ATP-dependent chromatin remodeling enzyme CHD7 regulates pro-neural gene expression and neurogenesis in the inner ear. *Development.* 2010;137:3139-3150.
204. Riazuddin S, Belyantseva IA, Giese AP, et al. Alterations of the CIB2 calcium- and integrin-binding protein cause Usher syndrome type 1J and nonsyndromic deafness DFNB48. *Nat Genet.* 2012;44:1265-1271.
205. Robertson NG, Cremers CW, Huygen PL, et al. Cochlin immunostaining of inner ear pathologic deposits and proteomic analysis in DFNA9 deafness and vestibular dysfunction. *Hum Mol Genet.* 2006;15:1071-1085.
206. Busch-Nentwich E, Söllner C, Roehl H, Nicolson T. The deafness gene *dfna5* is crucial for *ugdh* expression and HA production in the developing ear in zebrafish. *Development.* 2004;131:943-951.
207. Stanchina L, Baral V, Robert F, et al. Interactions between *Sox10*, *Edn3* and *Ednrb* during enteric nervous system and melanocyte development. *Dev Biol.* 2006;295:232-249.
208. Kalatzis V, Sahly I, El-Amraoui A, Petit C. *Eya1* expression in the developing ear and kidney: towards the understanding of the pathogenesis of Branchio-Oto-Renal (BOR) syndrome. *Dev Dyn.* 1998;213:486-499.
209. Qu Y, Tang W, Dahlke I, et al. Analysis of connexin subunits required for the survival of vestibular hair cells. *J Comp Neurol.* 2007;504:499-507.
210. Morozko EL, Nishio A, Ingham NJ, et al. ILDR1 null mice, a model of human deafness DFNB42, show structural aberrations of tricellular tight junctions and degeneration of auditory hair cells. *Hum Mol Genet.* 2015;24(3):609-624.
211. Kharkovets T, Hardelin JP, Safieddine S, et al. KCNQ4, a K<sup>+</sup> channel mutated in a form of dominant deafness, is expressed in the inner ear and the central auditory pathway. *Proc Natl Acad Sci U S A.* 2000;97:4333-4338.
212. Grillet N, Schwander M, Hildebrand MS, et al. Mutations in LOXHD1, an evolutionarily conserved stereociliary protein, disrupt hair cell function in mice and cause progressive hearing loss in humans. *Am J Hum Genet.* 2009;85:328-337.
213. Sacheli R, Nguyen L, Borgs L, et al. Expression patterns of miR-96, miR-182 and miR-183 in the development inner ear. *Gene Expr Patterns.* 2009;9:364-370.
214. Watanabe K, Takeda K, Katori Y, et al. Expression of the *Sox10* gene during mouse inner ear development. *Brain Res Mol Brain Res.* 2000;84:141-145.
215. Coffin AB, Dabdoub A, Kelley MW, Popper AN. Myosin VI and VIIa distribution among inner ear epithelia in diverse fishes. *Hear Res.* 2007;224:15-26.
216. Mochizuki E, Okumura K, Ishikawa M, et al. Phenotypic and expression analysis of a novel spontaneous myosin VI null mutant mouse. *Exp Anim.* 2010;59:57-71.
217. Dulon D, Safieddine S, Jones SM, Petit C. Otoferlin is critical for a highly sensitive and linear calcium-dependent exocytosis at vestibular hair cell ribbon synapses. *J Neurosci.* 2009;29:10474-10487.
218. Simmler MC, Cohen-Salmon M, El-Amraoui A, et al. Targeted disruption of *otog* results in deafness and severe imbalance. *Nat Genet.* 2000;24:139-143.
219. Yan D, Zhu Y, Walsh T, et al. Mutation of the ATP-gated P2X<sub>2</sub> receptor leads to progressive hearing loss and increased susceptibility to noise. *Proc Natl Acad Sci U S A.* 2013;110:2228-2233.
220. Ahmed ZM, Riazuddin S, Ahmad J, et al. *PCDH15* is expressed in the neurosensory epithelium of the eye and ear and mutant alleles are responsible for both USH1F and DFNB23. *Hum Mol Genet.* 2003;12:3215-3223.
221. Xiang M, Gan L, Li D, et al. Essential role of POU-domain factor Brn-3c in auditory and vestibular hair cell development. *Proc Natl Acad Sci U S A.* 1997;94:9445-9450.
222. Kitajiri S, Fukumoto K, Hata M, et al. Radixin deficiency causes deafness associated with progressive degeneration of cochlear stereocilia. *J Cell Biol.* 2004;166:559-570.
223. Adler HJ, Belyantseva IA, Merritt RC Jr, Frolenkov GI, Dougherty GW, Kachar B. Expression of prestin, a membrane motor protein, in the mammalian auditory and vestibular periphery. *Hear Res.* 2003;184:27-40.
224. Wu L, Sagong B, Choi JY, Kim UK, Bok J. A systematic survey of carbonic anhydrase mRNA expression during mammalian inner ear development. *Dev Dyn.* 2013;242:269-280.
225. Kawashima Y, Géléoc GS, Kurima K, et al. Mechanotransduction in mouse inner ear hair cells requires transmembrane channel-like genes. *J Clin Invest.* 2011;121:4796-4809.
226. Adato A, Lefèvre G, Delprat B, et al. Usherin, the defective protein in Usher syndrome type IIA, is likely to be a component of interstereocilia ankle links in the inner ear sensory cells. *Hum Mol Genet.* 2005;14:3921-3932.
227. Kikkawa Y, Mburu P, Morse S, Kominami R, Townsend S, Brown SD. Mutant analysis reveals whirlin as a dynamic organizer in the growing hair cell stereocilium. *Hum Mol Genet.* 2005;14:391-400.