Histomorphometric Effcets of Oral use of Tobacco in Testes of Offsprings of Swiss Albino Mice

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> Author`s Contribution

¹Conception, Synthesis and Planning of the research ^{4.6.5}Active participation in active methodology ^{2.3}Interpretation, analysis and discussion

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ABSTRACT

Objective: To study the gross micro structural changes in the testis of the offspring of Swiss albino mice exposed to oral use of tobacco during pregnancy.

Study Design: Experimental

Duration and Setting of Study: This study was conducted from July 2017 to December 2017 at Isra University Hyderabad

Methodology: 20 female albino mice were mated. After confirmation of pregnancy by vaginal plug they were divided into two groups; experimental A and control B of 10 each. Experimental mice of group A were given 5% tobacco in their chow diet andOwater ad libitum; however, control group B were given only normal chowOdiet andOwaterOad libitum. After 15 days of birth the 10 male offsprings each of control and experimental mice were randomly selected. Their initial and final body weight was recorded. They were sacrificedOby cervical dislocations andOtheir testes were taken away for further gross & histologicalOanalysis.

Results: Noticeable changes were observed in the body weight and weight of testes. The mean initial weight of experimental male offspring was found to be 1.76 ± 0.33 g, while in control group it was 2.60 ± 0.45 g. The final weight in experimental offspring was 9.38 ± 0.59 g and in control group it was 12.75 ± 0.96 g. Statistically the difference of body weight in offspring was found to be significant (p value <0.05). The testes weight was markedly decreased in experimental group as compared to control group. The mean testes weight in experiment group was recorded as 0.03 ± 0.004 g however in control group it was recorded as 0.07 ± 0.004 g.

Simultaneously, 5 micro structural variations were also observed in the testes of offsprings of mice. InOtheOexperimental groupOof off-springs, very fewOlayers and decreased number of spermatocytes were noticed in seminiferous tubules of 9 testes. Sperm degenerative changes, cellular inflammation and mild to massive hyalinization were noticed in 9, 6, and 9 testes of experimental group respectively. Loss of architecture of seminiferous tubules in 8 testes as well as destruction of the basement membrane in 7 testes were also observed in experiment group testes. However, the decreased number of spermatocytes in only 1 testes and mild hyalinization in 2 testes of control group were observed.

Conclusion: The consumption of smokeless tobacco has vital effects on the body weight, micro structure and weight of testes of offsprings of mice given with cellular injury of seminiferous tubules especially decreased sperm count, cellular inflammation, destruction of basement membrane as well as massive hyalinization.

Key Words: smokeless tobacco, offspring, testes.

Introduction

Infertility is commonest between couples of childbearing ages and around 50% of the known factors of main infertility are accredited to male factor.¹

Though, the etiology for male factor sterility is not clearly recognized. Whereas, some people can possibly be genetically susceptible for being sub-fertile.² There remain chief epigenetic factors that are concerned as main factors of male sterility. The male reproductive system is thought to be responsive to several chemical and medicines that are found to cause adversative impacts on male's reproductive function under bound settings.³

The use of tobacco products and the health consequences of smoking and smokeless tobacco (SLT) are eminent. Crucial reasons exist for enhanced mortality and morbidity within developed nations and therefore, primacy is growing in underdeveloped countries too.⁴ In spite of expanding data on harmful reproductive results of tobacco, it is comparatively vague whether, the smoking and smokeless tobacco has similar consequences on male's reproductive events particularly among evidently reproductively active individuals.²

Two chief kinds of smokeless tobacco (SLT) used in United States are chewing tobacco and snuff.⁵ Several significant, harmful individual's health consequences have been associated to its consumption. These comprise: periodontitis oral leukoplakia and submucous fibrosis;⁶ astrointestinal abnormalities, oropharyngeal, esophageal and ancreatic cancers; stomach cancer as well.^{7,8}

Other likely unfavorable health consequences of SLT consist of toxicity of the cardiovascular, immune, and/or reproductive systems.⁹

Health professionals have a well-thought-out exposure to smokeless tobacco noxious to reproductive process, influencing facets from pregnancy and fertility results in development of fetus and child. Tobacco comprises of hundreds of mixes that have harmful impact on reproductive well-being, like metals, nicotine, and carbon monoxide. Annually, together with over 4,000,000 births within the US, 10% to 20% of pregnancies lead to miscarriage or abortion prior to delivery, other 10% of pairs face childlessness or decreased fertility.¹

Nicotine is used up in numerous ways varying from smokeless tobacco products like snuff and chewing tobacco to additional used up method: smoked tobacco. Cigarette tobacco holds numerous substances.¹² Proteins and carbohydrates are the foremost representative parts however alkaloids are considerably present too. Nicotine, specifically, represents 95% of the full alkaloids. Nicotine is extremely cyanogenic element and is absorbed quickly via the tract, oral membrane and skin. Around, 80%- 90% of nicotine is broken down liver. however kidney, lungs and by gonads are involved too.² The usage of nicotine during pregnancy is very harmful because of it is going to directly affect fetal organs development. Nicotine can easily pass through membrane barriers because of its lipophilic nature and activate nicotinic acetylcholine receptors (nAChRs).13

Endogenous agonists like acetylcholine usually tie nAChRs to modulate downstream physiological and cellular responses; although, exogenous agents; for instance nicotine, can contend for the holding sites and exert different, and possibly pathological, effects.¹⁴ Nicotine delivery is fast in whole body, taking place in seconds to minutes as of exposure, by maximum affinities within brain, kidney, lung, liver, spleen, and skeletal muscle.¹⁵ Nicotine too amasses in breast milk, amniotic fluid, placental tissue, and fetal blood¹⁶, directed towards important fetal and neonatal exposure. Animal models researches evidently proved that fetal and neonatal exposure to nicotine ends up in a varied choice of short and long-time health consequences for offspring, shortfalls together with in postnatal reproductive function.17, 18

Previous researches have revealed that tobacco brings apoptosis and has degenerative effects on testicular tissues that are linked to metabolize oxidative stress.¹⁹

Most of the research consideration on Smokeless Tobacco has focalized on malignancy from a clinical or epidemiological stance, together with practice²⁰, harm-reduction potential²¹, and risk perceptions.²² Partial researches via animal models have shown that long term SLT exposure may ends up in reproductive harmfulness together with low sperm production and count among male mice [23]. Longstanding contact of mice with SLT reduced antioxidant defense leading to longstanding inflammation within lung and liver²⁴ and can cause testicular cancer. Short term contact research on various SLT kinds in animal models are deficient, mainly their effects on structure of offspring's testes. Having increased frequency and worldwide practice of culturally-specific tobacco products, this research aims at investigating the effects of smokeless tobacco on microstructure of testes in offsprings of mice.

Methodology

This experimental study was conducted from July 2017 to December 2017 at Isra University Hyderabad. Twenty non-pregnant Swiss albino mice with normal weight of 34 grams and 10–12 weeks old were chosen which were separated into 2 groups of ten mice each. Female mice were kept alone in polycarbonate cages in a temperature (22 °C) and humidity-regulated room (55%). Water and food were given ad libitum with dark and light periods kept on 12-hour cycles.

All female mice were coupled with adult male mice, controlled from copulating for 7 day to raise their sexual impulse for opposite sex. Pregnancy was verified by existence of mucus vaginal plug amid 1- 10 days of pairing. When pregnancy confirmed, male and female mice were separated.

All female mice were divided in two groups according to block randomization:

- 10 pregnant mice from Group A (Experimental).
- 10 pregnant mice from Group B (Control).

From day 1 (gestation) till parturition, Experimental Group A females (Per cage 2) were made contact to 5% tobacco combined with food ad libitum together with fresh water and Control Group B were made contact to normal forage ad libitum along with fresh water. 1 to 2 days before birthing (GD 18), barriers were made and kept separately. While birth, every mother/offspring set were upheld in hygienic filtered air, besides the quantity of litter by means of feasible offspring with estimated period of pregnancy and size were ascertained. 10 male off-springs were haphazardly chosen from Group A (Experimental) and 10 male off-spring were haphazardly chosen from Group B (Control).

- 10 male offspring from Group A (Experimental).
- 10 male offspring from Group B (Control).

15 days after birth, their initial and final body weight was recorded. The male offsprings of both groups experimental and control were sacrificed, and gonads were detached. For the morphological and histological measurements, the weight of gonads was recorded though weighing scale machine. For the testicular tissues of offsprings in individual groups, hematoxylin and Eosin (H&E) stained segments were used. Slides of four-micron tissue sections were set and investigated for the histological study to see the changes under the light microscope.

All the data was recorded in the proforma and analyzed by SPSS version 20. Mean and standard deviation were calculated for numerical variables. Frequency and percentage calculated for categorical variables. t-test was applied to compare the body weight, testes weight and histological changes of offsprings among both groups, p-Value <0.05 was considered as significant.

Results

In this study significant decrease in the body weight of the offsprings in the experimental group as compared to control group was observed. The mean initial body weight of experimental offsprings was found to be 1.76 ± 0.33 g, which was significantly lower as compare to control group as 2.60 ± 0.45 g, p-value 0.001. The final body weight in experimental offsprings was also decreased as 9.38 ± 0.59 g in contrast to control group as 12.75 ± 0.96 g, p-value 0.0001. (Table.1)

Table. 1. Body weight comparison among offsprings					
Groups	Experiment	Control	p- Value		
Initial body	176+0.33	2 60+0 45	0.001		
weight	grams	grams	0.001		
Final body	9.38± 0.59	12.75±0.96	0.0001		
weight	grams	grams			

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No mortality was observed in the offsprings during the experiment period and the appearance of all control animals was indicative of good health however the experiment animals were observed as lethargic and slothful. In comparison of weight of testes of 2 weeks age of offspring's, 0.03 \pm 004 g was observed in experiment group however the 0.07 \pm 004 g was observed in control group (Table. 2). Micro structural variations were found in testes of offsprings of mice. In experimental group of offspring's, insufficient layers and reduced number of spermatocytes were detected in seminiferous tubules of 9 testes (Table. 3). Formation of spermatogenic cell lines showed numerous spermatocytes in control group as linked to experiment group. Sperms degenerative changes, cellular inflammation and mild to massive hyalinization were noticed in 9, 6, and 9 testes of experimental group respectively. Loss of architecture of seminiferous tubules in 8 testes as well as destruction of basement membrane in ST of 7 testes was also observed in experiment group animals. (Table. 3) Figures 2, 3, 4 and 5.

Table. 2 Testes weight comparison among offsprings				
Groups	Experiment	Control	p-Value	
Testes mean	0.03 ± 0.004	0.07 ±	0.01	
weight /g		0.004		

Spermatozoa in the seminiferous tubule lumen of the control group were seen to be more abundant than in the experimental group, Figure 1. However, the decreased number of spermatocytes in only 1 ST and mild hyalinization in 2 testes of control group were observed, (Table. 3) that might have occurred due to the environmental threats on the animals.

Table. 3 Histological comparisons of testes of offsprings					
Histological changes	Experiment	Control	p-		
	N=10	N=10	Value		
Decreased	9	1			
spermatocyte layers &					
Sperm degenerative			0.001		
changes					
Cellular inflammation	6	0			
Hyalinization	9	2			
Loss of seminiferous	8	0			
architecture					
Basement membrane	7	0			
destruction					



Figure 1. Photomicrograph of testicular tissue isolated from control group showing uniform seminiferous tubules lined by normal layers of spermatogenic cells and basement membrane of normal thickness (H&E- X 100).



Photo 2. Photomicrograph of offspring of mice testicular tissue isolated from experiment showing loss of architecture and destruction of the basement membrane.X 100



Photo 3. Photomicrograph of offspring of mice testicular tissue isolated from experiment showing variable sized and shaped seminiferous tubules lined by few layers of spermatogenic cells. Marked with hyalinization in the seminiferous tubules. X 400



 $\label{eq:photo-4} Photo 4. Photomicrograph of offspring of mice testicular tissue isolated from experiment showing cellular swelling and fatty infiltration with variable size and number of spermatocytes in the ST. <math display="inline">\times$ 400



Photo 3. Photomicrograph of left rat testicular tissue isolated from experiment showing massive hyalinization with edematous changes X 400

Discussion

The outcomes of this study showed that smokeless tobacco has lethal effects on reproductive roles of male offsprings mice that is adequate reason to cause sterility. Mice used within current study have been revealed to possess a clear reproductive system. This study specifies that smokeless tobacco (SLT) treatment in male offsprings mice impacts the weight of testes also brings hazardous effects on testes structure.

Numerous results from scientific studies found on inferences of smokeless tobacco usage on the organs of animals and human both and their offsprings as well.^{25, 26, and 27} But, there is shortage of scientific literature on the consequences of smoke extracts of *smokeless tobacco* on testes of offsprings of male mice as a marker of toxicity.

The results of this study showed a significant decrease in the body weight of the offsprings of

experimental mice as compared to the controls. This finding is in line with the study of Essien & Akpan³³ showing similar results in the offsprings of Albino rats consuming nitrosamine.

Another study by EL-Meligy et al³⁴ that highly supports our findings, as reported that the fetuses of nicotine treated mothers are lighter than those of control mothers and this difference in fetal body weight when compared to those of control group were recorded to be highly significant.

The organs weight of the experiment group presented a noteworthy reduction in weight of testes. These finding agree with the Oyeyipo IP²⁸ who has observed same changes in adult rats administered orally different doses of smokeless tobacco.

Smokeless tobacco and its major constituent Nicotine is engrossed into buccal mucosa 3-4 times quicker in chewing cases as compare to smoking. Degree of Nicotine grows slowly with intensive chewing practice and residual chemicals endure for a longtime as compare to among smoking cases.²⁹ However, threshold concentration distressing male reproductive system is so far not defined.

Smokeless tobacco is identified to interrupt the hypothalamus-pituitary axis by disturbing the testicular microcirculation. As per testosterone acts on seminiferous tubules to start and uphold spermatogenesis, decrease of this sex hormone level by impaired Leydig cell functions or trouble in the estrogen/androgen proportion might be a reason of reduced sperm counts.²⁹

Most prominently, we found evidence of few layers and decreased number of spermatocytes observed in seminiferous tubules. This research evidently proves that chewing tobacco causes statistically large lessening in sperm concentration. These explanations are in step with results of others like Said et al.³⁰ and Aprioku JS et al.³¹

The significant findings of this study were the reformed morphology and histopathological variation in testes of offsprings of the tobacco processed by mothers. The chief results were the fatty infiltration of seminiferous tubules. Besides, we also observed the cell swellings, mild to massive hyalinization. Loss of architecture of seminiferous tubules as well as destruction of basement membrane. These results are in accordance with similar results of other researches who have found comparable variations in the testes of adult experimental animals cured with nicotine or nitrosamines.^{31, 32}

The present investigation shows that smokeless tobacco usage of local kind, during pregnancy, has important upshots on body weight, weight and microstructure of testes of offsprings of mice, especially decrease the sperm count as well as results in mild to massive hyalinization, inflammation and destruction of seminiferous tubules. Although, more study is bucked up to affirm the part and mechanics of other brands and mixes of tobacco.

CONCLUSION:

The Consumption of smokeless tobacco has vital effects on the body weight, weight and micro structure of testes of offsprings of mice given with cellular injury of seminiferous tubules parenchyma, especially decreased sperm count, cellular inflammation, destruction of basement membrane as well as massive hyalinization.

References

- Kumar N, Singh AK. Trends of male factor infertility, an important cause of infertility: A review of literature. Journal of human reproductive sciences. 2015 Oct;8(4):191.
- Oyeyipo IP, Raji Y, Emikpe BO, Bolarinwa AF. Effects of nicotine on sperm characteristics and fertility profile in adult male rats: a possible role of cessation. Journal of reproduction & infertility. 2011 Jul;12(3):201.
- Skakkebaek NE, Rajpert-De Meyts E, Buck Louis GM, Toppari J, Andersson AM, Eisenberg ML, Jensen TK, Jørgensen N, Swan SH, Sapra KJ, Ziebe S. Male reproductive disorders and fertility trends: influences of environment and genetic susceptibility. Physiological reviews. 2015 Nov 18;96(1):55-97.
- 4. US Department of Health and Human Services. The health consequences of smoking—50 years of progress: a report of the Surgeon General. Atlanta, GA: US Department of Health and Human Services, Centers for Disease Control and Prevention, National Center for Chronic Disease Prevention and Health Promotion, Office on Smoking and Health. 2014 Nov 27;17.
- 5. Gupta PC, Ray CS. Smokeless tobacco and health in India and South Asia. Respirology. 2003 Dec;8(4):419-31.
- Johnson GK, Slach NA. Impact of tobacco use on periodontal status. Journal of Dental Education. 2001 Apr 1;65(4):313-21.
- Aro P, Ronkainen J, Storskrubb T, Vieth M, Engstrand L, Johansson SE, Bolling-Sternevald E, Bolinder G, Alving K, Talley NJ, Agréus L. Use of tobacco products and gastrointestinal morbidity: an endoscopic population-based

study (the Kalixanda study). European journal of epidemiology. 2010 Oct 1;25(10):741-50.

- Shankaran K, Kandarkar SV, Contractor QQ, Kalro RH, Desai HG. Electron microscopic observations in gastric mucosa of habitual tobacco chewers. The Indian journal of medical research. 1994 Jun;99:267-71.
- Willis D, Popovech M, Gany F, Zelikoff J. Toxicology of smokeless tobacco: Implications for immune, reproductive, and cardiovascular systems. Journal of Toxicology and Environmental Health, Part B. 2012 Jul 1;15(5):317-31.
- World Health Organization, Research for International Tobacco Control. WHO report on the global tobacco epidemic, 2008: the MPOWER package. World Health Organization; 2008 Feb 11.
- 11. Samet JM, Yoon SY, World Health Organization. Gender, women, and the tobacco epidemic. 2010.
- O'connor RJ. Non-cigarette tobacco products: what have we learnt and where are we headed?. Tobacco control. 2012 Mar 1;21(2):181-90.
- Wickstrom R. Effects of nicotine during pregnancy: human and experimental evidence. Current neuropharmacology. 2007 Sep 1;5(3):213-22.
- Albuquerque EX, Pereira EF, Alkondon M, Rogers SW. Mammalian nicotinic acetylcholine receptors: from structure to function. Physiological reviews. 2009 Jan;89(1):73-120.
- 15. Benowitz NL, Hukkanen J, Jacob P. Nicotine chemistry, metabolism, kinetics and biomarkers. InNicotine psychopharmacology 2009 (pp. 29-60). Springer, Berlin, Heidelberg.
- Dahlström A, Lundell B, Curvall M, Thapper L. Nicotine and cotinine concentrations in the nursing mother and her infant. Acta Pædiatrica. 1990 Feb;79(2):142-7.
- Behl M, Rao D, Aagaard K, Davidson TL, Levin ED, Slotkin TA, Srinivasan S, Wallinga D, White MF, Walker VR, Thayer KA. Evaluation of the association between maternal smoking, childhood obesity, and metabolic disorders: a national toxicology program workshop review. Environmental health perspectives. 2013 Feb;121(2):170.
- Bruin JE, Gerstein HC, Holloway AC. Long-term consequences of fetal and neonatal nicotine exposure: a critical review. Toxicological sciences. 2010 Apr 2;116(2):364-74.
- B. A. Hanadi, A. M. Kelany, F. M. ElQudsi, H. A. Ameen, and S. A. El Karium, "The possible protective role of antioxidants (selenium, vitamin E) in reducing smoking effects on testes of albino rats," Assiut University Bulletin for Environmental Researches, vol. 14, no. 1, 2011.
- Timberlake DS, Huh J. Demographic profiles of smokeless tobacco users in the US. American journal of preventive medicine. 2009 Jul 31;37(1):29-34.
- Hatsukami DK, Lemmonds C, Tomar SL. Smokeless tobacco use: harm reduction or induction approach?. Preventive medicine. 2004 Mar 1;38(3):309-17.
- 22. O'Connor RJ, McNeill A, Borland R, Hammond D, King B, Boudreau C, Cummings KM. Smokers' beliefs about the relative safety of other tobacco products: findings from the

ITC collaboration. Nicotine & Tobacco Research. 2007 Oct 1;9(10):1033-42.

- 23. Kumari A, Mojidra BN, Gautam AK, Verma Y, Kumar S. Reproductive toxic potential of panmasala in male Swiss albino mice. Toxicology and industrial health. 2011 Sep;27(8):683-90.
- Avti PK, Kumar S, Pathak CM, Vaiphei K, Khanduja KL. Smokeless tobacco impairs the antioxidant defense in liver, lung, and kidney of rats. Toxicological Sciences. 2005 Nov 9;89(2):547-53.
- Dar NA, Bhat GA, Shah IA, Iqbal B, Kakhdoomi MA, Nisar I, Rafiq R, Iqbal ST, Bhat AB, Nabi S, Shah SA. Hookah smoking, nass chewing, and oesophageal squamous cell carcinoma in Kashmir, India. British journal of cancer. 2012 Oct;107(9):1618.
- Avti PK, Kumar S, Pathak CM, Vaiphei K, Khanduja KL. Smokeless tobacco impairs the antioxidant defense in liver, lung, and kidney of rats. Toxicological Sciences. 2005 Nov 9;89(2):547-53.
- Spindel ER, McEvoy CT. The role of nicotine in the effects of maternal smoking during pregnancy on lung development and childhood respiratory disease. Implications for dangers of e-cigarettes. American journal of respiratory and critical care medicine. 2016 Mar 1;193(5):486-94.
- 28. Oyeyipo IP, Raji Y, Emikpe BO, Bolarinwa AF. Effects of oral administration of nicotine on organ weight, serum testosterone level and testicular histology in adult male

rats. Nigerian Journal of Physiological Sciences. 2010;25(1):81-6.

- 29. Sunanda P, Panda B, Dash C, Ray PK, Padhy RN, Routray P. Prevalence of abnormal spermatozoa in tobacco chewing sub-fertile males. Journal of human reproductive sciences. 2014 Apr;7(2):136.
- Said TM, Ranga G, Agarwal A. Relationship between semen quality and tobacco chewing in men undergoing infertility evaluation. Fertility and sterility. 2005 Sep 1;84(3):649-53.
- Aprioku JS, Ugwu TC. Comparative evaluation of the impact of subacute exposure of smokeless tobacco and tobacco smoke on rat testes. International journal of reproductive medicine. 2015;2015.
- Adedayo AD, Tijani AA, Musa AA, Adeniyi TD. Histological study of smoke extract of Tobacco nicotiana on the heart, liver, lungs, kidney, and testes of male Sprague-Dawley rats. Nigerian medical journal: journal of the Nigeria Medical Association. 2011 Oct;52(4):217.
- Essien A D. and Akpan J O. Altered morphology of liver and pancreas tissues of offsprings of albino rats by charred meat. N igerian Journal of Physiological Sciences 22 (1-2): 2007 49 53©Physiological Society of Nigeria.
- 34. EL-Meligy. Manal M.S, Randa H. Abdel Dady, Amal R. Abdel Smeaei and Heba M. Saad Eldien. Effect of nicotine administration and its witherawal on the reproductive organs, fertility, and pregnancy outcome in female rats MansouraJ. Forensic Med. Clin. Toxicol. Vol. XV, No. 1, Jan. 2007.