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Meat quality of buffalo young bulls fed faba bean as protein source

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ABSTRACT

Sixteen Italian Mediterranean Buffalo young bulls were divided into two groups fed isoprotein and isoenergy diets and only differing for protein source of concentrate: faba bean (FB) vs soybean (SB). Animals were slaughtered at 350 kg BW. Meat from FB group showed significantly lower fat, protein, cholesterol and saturated fatty acids than SB group. Significant differences were also found between the three muscles analysed [*Longissimus thoracis* (LT), *Semitendinosus* (ST) and *Iliopsoas* plus *Psoas minor* (IP)]. ST showed the most favourable fatty acids profile: lower SFA, higher PUFA, MUFA, ω -3, ω -6, CLA and, consequently, lower values for both atherogenic and thrombogenic indexes. Results showed that faba bean can be used as a protein source alternative to soybean in the diet of young buffalo bulls for the production of high quality meat.

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1. Introduction

The Italian Mediterranean Buffalo (Bubalus bubalis) amounts to about 380000 heads and it is mainly bred for milk production in order to produce Mozzarella, the well known fresh cheese which is highly appreciated both in national and international markets. In recent years, an increasing interest for buffalo meat was seen in Italy. Up to few years ago, buffalo meat was little appreciated since only low quality meat from females with productive or reproductive problems was marketed for human consumption (Borghese et al., 2010). More recently, it has been demonstrated that using adequate feeding systems, favourable nutritional characteristics can be achieved (Infascelli, Gigli, & Campanile, 2004). The fatty acids profile and the cholesterol content of buffalo meat appears particularly interesting (Cutrignelli, Calabrò, Laudadio, Grasso, & Di Lella, 1996; Di Luccia et al., 2003; Infascelli et al., 2003, 2005; Spanghero, Gracco, Valusso, & Piasentier, 2004). In a recent cross-sectional study, Giordano et al. (2010), demonstrated that buffalo meat can confer significant benefits in terms of cardiovascular risk profile, including lower carotid atherosclerotic burden and susceptibility to oxidative stress.

In recent years in order to reduce feeding costs, several farmers attempt protein sources alternative to soybean in buffalo diet (Cutrignelli, Calabrò, Tudisco, Infascelli, & Piccolo, 2011) and tried to increase the domestic production of GMO-free plant proteins. Indeed, large part of the available soybean is genetically modified thus still determining public concerns due its health potential risks (Tudisco, Ma stellone, et al., 2010).

Faba bean (Vicia faba var. minor) is relatively cheap in spite of its high nutritional value (Larralde & Martinez, 1991) traditionally attributed to the high protein content (25-35%) and to the high lysine content despite the imbalance in sulphur aminoacids. Additionally, Faba bean as grain legume increases the sustainability of crop-livestock systems through the safeguarding of soil fertility. The occurrence of some anti-nutritional factors (ANF) has hampered a wider nutritional utilization of faba bean (Calabrò et al., 2009). However, the considerable progress in plant breeding notably decreased the presence of secondary plant metabolites (Duc, Marget, Esnault, Le Guen, & Bastianelli, 1999). Recent studies performed in other species (Cutrignelli, Calabrò, et al., 2008; Cutrignelli, Piccolo, et al., 2008; Lanza et al., 2011; Scerra et al., 2011) showed that the use of faba bean as the main protein source did not negatively affect animal performance and meat fatty acids profile and cholesterol content. Because few data are available on buffalo species, the aim of this study was to evaluate the influence of a diet containing faba bean on the nutritional characteristics of meat from young buffalo bulls.

2. Material and methods

2.1. Animals and diets

The trial was carried outin a farm situated at 47 m a.s.l. in Southern Italy (Cassino, FR), where about 200 Italian Mediterranean buffaloes are bred. All experiments were authorized by the Animal Welfare Commission of the Department of Veterinary Medicine and Animal Production, University of Naples Federico II.

Sixteen eight days buffalo males were equally divided into two groups (FB, faba bean and SB, soybean) and received 6.0 L/head/d of





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acidified milk replacer (180 g/L of water) until 56 days of age. Therefore, the replacer amount was gradually decreased, whilst administering the same volume, in order to obtain a complete weaning at the age of 80 days. Starting from the 70th day, corn silage was administered, where-as roughly chopped mixed hay and weaning concentrate were added from the fifth week. After weaning until the beginning of trial, the animals were fed mixed hay and corn silage *ad libitum*, whereas concentrate was fed in the amount of 2.0 kg/d

At 84 days, each animal was placed in individual box up to the slaughtering weight and fed (2.7% BW) isoprotein (CP: 15.2% DM) and isoenergy (0.91 veal forage unit_VFU/kg DM) diets, characterized by 39% of NDF and differing in protein source of concentrate: faba bean tannin-free variety (*Vicia faba minor* L.) vs. whole seed soybean (*Soja hispida*). As forage, a mixed hay and a corn silage produced by the farm were administered to both groups. Every two months, samples of each feed were collected to determine the chemical composition (AOAC, 2000; Van Soest, Robertson, & Lewis, 1991) and to estimate the nutritive value as net energy for meat production (VFU/kg DM; INRA, 1988). In Table 1, percentages (%DM of diet), crude protein and nutritive values of diet ingredients are depicted.

Faba bean, soybean and the other ingredients of concentrates were also analysed for the fatty acid profile. For this purpose, the total fat was extracted from 0.5 g of each samples into 20×150 mm test tubes with Teflonlined screw caps (Folch, Lees, & Sloane Stanley, 1957). Fatty acids were methylated using HCl in methanol prepared by adding 20 mL of acetyl chloride to 100 mL of methanol (Christie, 1993). Fatty acid methyl esters were analysed by a gas-chromatography (GC) apparatus (Focus ThermoQuest equped a computer integration system Chrome-card.), equipped with a CP-SIL 88 fused silica capillary column $[30 \text{ m} \times 0.25 \text{ mm} \text{ (internal diameter) with } 0.2\text{-}1 \,\mu\text{m} \text{ film thickness;}$ Varian, Walnut Creek, CA, USA]. The GC conditions were the following: the oven temperature was programmed at 160 °C and held for 1 min; then increased up to 230 °C at a rate of 1 °C/min and held for 3 min. The injector and detector temperatures were set up at 220 °C and 250 °C, respectively. Helium was used as carrier gas with flow of 1.5 mL/min. Fatty acids were identified using a mixture of standard fatty acids (Larodan Fine Chemical AB, Malmo, Sweden) and quantified using the peak areas, and were expressed as mg fatty acid/g dry matter of feed sample (Antongiovanni, Lercker, & Serchiari, 2007; D'Urso et al., 2008; Tudisco, Cutrignelli, et al., 2010).

2.2. Meat nutritional characteristics

When animals reached about 350 kg BW (429.7 \pm 57.6 days of age) they were slaughtered in an authorized slaughterhouse (1.5 km from the farm) according to EU legislation (EU Regulation 882/2004). After 15 days of ageing at 4 \pm 1 °C, the right side of each animal was dissected. The pH (Hanna pH-meter, mod. HI 9025, equipped with an electrode FC 230C) was measured in *Longissimus thoracis* (LT), *Semitendinosus* (ST) and *lliopsoas* plus *Psoas minor* (IP) muscles within 1 h from death and at the end of ageing.

Samples of LT, ST and IP muscles were collected and rapidly transported, upon refrigeration temperature, to the laboratories for the chemical analysis. The muscles were homogenised and divided into three different aliquots: two were vacuum-packed and frozen (-20 °C)

Table 1

Diet ingredients: percentage (% of DM diet), crude protein and nutritive value.

	Corn silage	Mixed hay	FB concentrate ^a	SB concentrate ^b
% of DM diet	30.0	20.0	50.0	50.0
Crude protein, % DM	7.60	7.80	22.8	22.8
Energy, VFU/kg DM	0.80	0.60	1.10	1.12

DM: dry matter; VFU: veal forage unit.

FB: faba bean; SB: soybean.

^a corn (10%), barley (11%), wheat bran (23%) faba bean 29% CP (56%).

 $^{\rm b}~$ corn (15%), barley (15%), wheat bran (26%) and soybean 35% CP (44%).

until the analysis (cholesterol content and fatty acid composition). The last one was analysed for moisture, crude fat, ash and protein by using a food analyser (FoodScan Lab, FOSS Electric, Denmark). The cholesterol extraction was made according to Naeemi, Ahmad, Al-Sharrah, and Behbahani (1995) and for its quantification a GC was utilized. The GC (Focus ThermoQuest) was equipped with a split/splitless injector, a flame ionization detector, a fused silica capillary column (SACTM-5, Supelco, 30 m × 0.25 mn × 0.25 µm film thickness) and computer integration system Chrome-card. The operative conditions were the following: oven temperature isothermal 285 °C; injector and detector temperature 300 ° C; injection volume 1.0 µL; carrier gas: helium (25 cm/s); split ratio: 1:100. The identification of cholesterol was made by comparing the peak retention time of samples with an external standard (C-8667, SIGMA). The concentration was expressed as g/100 g of edible part, each chromatographic analysis was replicated three times.

To determine the fatty acids profile, total fat was previously extracted (Folch et al., 1957) and subsequently turned into methyl esters (FAMEs) by direct transesterification (Christie, 1993) according to Chiofalo et al. (2010) and Chiofalo et al. (2011). The oil extracted from each sample was suspended in a mixture of sulphuric acid/methanol (1:9, mL/mL) and heated for three h. The FAMEs were isolated by adding 1.0 mL of n-hexane. The mixture was shaken and, after two min, the formed toplayer with n-hexane was transferred into the vial for the GC injection. The FAMEs were analyzed by GC-FID (Agilent Technologies 6890 N, Palo Alto, CA, U.S.A.) with a split/splitless injector, a flame ionization detector and fused silica capillary column Omegawax 250 (Supelco, Bellefonte, PA, U.S.A.; 30 m \times 0.25 mm I.D., 0.25 μ m film thickness). Column temperature was programmed: initial isotherm of 160 °C (6 min.), increment of 3 °C/min and final isotherm of 250 °C (30 min.). Temperature of the injector and detector: 250 °C. Injection volume: 1.0 µL. Carrier gas: helium (1 mL/min). Split ratio: 1:50. Identification of fatty acids was made by comparing the relative retention times of FAME peaks from samples with standards from Supelco (Bellefonte, PA, U.S.A.). Chromatogram peak areas were acquired and calculated by Chemstation software (Agilent, Palo Alto, CA, U.S.A.). The concentration of each fatty acid was expressed as g/100 g, considering 100 g the summation of the areas of all fatty acid methyl esters identified. For each sample the chromatographic analysis was replicated three times.

On the basis of the identified fatty acids, the Atherogenic Index (AI) and the Thrombogenic Index (TI) were estimated using the following equations proposed by Ulbricht and Southgate (1991):

$$AI = \frac{C12:0 + (4 \cdot C14:0) + C16:0}{n-6 \text{ PUFA} + n-3 \text{ PUFA} + \text{MUFA}}$$
$$TI = \frac{C14:0 + C16:0 + C18:0}{(0.5 \cdot \text{MUFA}) + (0.5 \cdot n-6 \text{ PUFA}) + (3 \cdot n-3 \text{ PUFA}) + (n-3 \text{ PUFA}/n-6 \text{ PUFA})}$$

2.3. Statistical analysis

The statistical analysis was performed (Proc GLM, SAS, 2000) with the following model: $y_{ijk} = \mu + D_i + M_j + (D^*M)_{ij} + \epsilon_{ijk}$ where: y = dependent variable, $\mu =$ mean; D = dietary effect (i = FB, SB), M = muscle effect (j = LT, ST, IP), D*M = interaction diet \times muscle and $\epsilon =$ error. The mean was statistically compared using the Least Significant Differences (LSD) test.

3. Results

3.1. Fatty acid profile of the diet ingredient

The fatty acid profile of faba bean, soybean and concentrates is depicted in Table 2. Soybean showed higher concentration of palmitic (C16:0), oleic (C18:1 n-9) and linoleic (C18:2 n-6) acids than faba bean. Conversely, the levels of stearic (C18:0) and linolenic (C18:3 n-3) acids were higher in faba bean than soybean. These results

reflected the fatty acid profile of FB and SB concentrates despite a lower difference was recorded in that case.

3.2. Meat pH, chemical composition, cholesterol and collagen

The pH of *Longissimus thoracis, Semitendinosus* and *lliopsoas* plus *Psoas minor*, despite significant differences were recorded between muscles, generally indicated a correct glycolitic process in muscle (Fig. 1): indeed, pH values at slaughter were always lower than 7.0 and at dissection always higher than 5.5.

Dietary treatments affected some qualitative parameters of meat (Table 3). In particular, FB group showed significantly (P < 0.05) lower value for fat, protein and cholesterol if compared with SB group.

The muscle type highly affected all the parameters (P < 0.001) except cholesterol content. In particular, ST showed the lowest fat (P < 0.01) and IP the highest protein content (P < 0.01).

3.3. Meat fatty acids profile

The dietary treatment significantly influenced the proportion of C17:1-cis10, C20:0, C22:0, C20:4 n-6, C24:0, C24:1-cis15, C22:5 n-3, C22:6 n-3 (P < 0.05), C20:3 n-6 (P < 0.01) and C1:0-anteiso, C15:0, C17:0, C23:0 (P < 0.001). The large part of fatty acids was significantly (P < 0.01) affected by muscle type, except C18:1-trans10 and C18:1-trans11, trans10-trans12 CLA, C24:1-cis15 and C22:6 n-3 (Table 4).

The great number of analysed fatty acids was divided in classes according to a particular nutritional interest (Table 5). It can be observed that saturated fatty acids (SFA) were significantly (P > 0.05) higher in SB than FB group (50.3 vs. 49.9 g/100 g). By contrast, monoinsaturated (MUFA), polyunsaturated (PUFA), ω -6, ω -3, CLA were not affected by dietary treatment, as well as PUFA/SFA and both ω -6/ ω -3 ratios and quality indexes (AI, TI).

Significant (P < 0.01) differences were found comparing the three muscles, and ST showed the most favourable fatty acids profile: lower SFA, higher PUFA, MUFA, ω -3, ω -6, CLA and, consequently, lower values for both quality indexes.

The interactions dietary treatment × muscle was never significant.

4. Discussion

4.1. Fatty acid profile of the diets ingredients

The results concerning fatty acids profile of faba bean and soybean are in agreement with those reported by Lanza et al. (2011) and in contrast with those of Grela and Günther (1995) who found lower linoleic (C18:2) acid content in faba bean. As reported by Lanza et al. (2011) legume seeds are characterized by a high content of linoleic and palmitic acids (C16:0), while level of linolenic acid (C18:3) is low and variable among varieties (Grela & Günther, 1995). Some authors (Priolo, Lanza, Galofaro, Fasone, & Bella, 2003; Ryan, Galvin, O'Connor, Maguire, & O'Brien, 2007; Yoshida, Saiki, Yoshida, Tomiyama, & Mizushina, 2008; Yoshida, Tomiyama, Saiki, & Mizushina, 2007) reported low and similar

Table 2				
Fatty acid	profile of faba	bean, sov	/bean and	concentrates

mg FA/g DM	Faba bean	Soybean	FB concentrate	SB concentrate
C16:0	2.874	3.987	3.256	3.720
C18:0	0.933	0.642	0.655	0.444
C18:1 n-9	2.376	4.754	2.773	3.881
C18:2 n-6	12.33	14.57	12.49	13.13
C18:3 n-3	2.145	1.628	1.928	1.569

FA: fatty acids.

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FB: faba bean; SB: soybean.



Fig. 1. pH values at slaughtering and dissection in the soybean and faba bean groups and in the three tested muscles. FB: faba bean group; SB: soybean group. LT: *Longissimus thoracis*; ST: *Semitendinosus*; IP: *Iliopsoas* plus *Psoas minor* muscles. a,b,c: P < 0.05.

linolenic content in peas (3.8 - 9.3% total FA), faba bean (3.8 - 5.1% total FA) and chickpeas (2.4 - 3.2% total FA) legume seeds. However, the favourable level of C18:3 n-3 in faba bean was lower when compared with linseed or algae (Cooper et al., 2004), but higher when compared with chickpeas (Priolo et al., 2003).

4.2. Meat quality

In Italy, the consumers' growing interest for buffalo meat is due to its nutritional characteristics (Cutrignelli et al., 1996; Di Luccia et al., 2003; Infascelli et al., 2003, 2004, 2005; Spanghero et al., 2004). Particularly, buffalo meat seems to be highly suitable for patients who need dietetical foods. Therefore, the significant lower fat and cholesterol content found in meat from animals fed diet including faba bean come to have a great appeal. In each case, the average level of cholesterol found in this study (33.3 mg/100 g), although sensibly higher than that reported for buffalo by Samoggia, Lo Fiego, De Grossi, and Bergonzoni (1993) on gracilis muscle (27.7 mg/100 g) and on the diaphragm muscle (31.4 mg/100 g) is lower than that obtained by Cutrignelli et al. (1996) in Longissimus dorsi muscle of young buffalo bulls (41.1 mg/100 g). In the literature, the content of cholesterol in buffalo meat is rather disomogeneous: Kesava Rao, Kowale, Murthy, and Sharma (1992) found 83 mg/l00 g in the Longissimus dorsi muscle, Yadav and Singh (1974) indicated a sensibly lower content (32 mg/100 g), Sinclair, Slattery, and O'Dea (1982) found 46 mg/100 g in Australian buffalo meat. Some authors found no differences in cholesterol content among muscles (Bohac & Rhee, 1988; Cifuni, Napolitano, Riviezzi, Braghieri, & Girolami, 2004) but, again, in contrast with others (Browning, Huffman, Egbert, & Jungst, 1990; Eichhorn et al., 1986; Rusman, Soeparno, Setiyono, & Suzuki, 2003). This contradiction is probably due to the different muscles analysed in each

Table 3
Chemical composition and cholesterol content.

	Groups		Muscles			Prob.		MSE
	FB	SB	LT	ST	IP	Muscle	Group	
Fat, %	1.82 ^b	1.98 ^a	2.07 ^A	1.47 ^B	2.17 ^A	***	*	0.058
Moisture, %	75.9	75.7	75.5 ^B	75.5 ^B	76.5 ^A	***	NS	0.040
Protein, %	21.0 ^b	21.3 ^a	21.3 ^{Ab}	21.6 ^{Aa}	20.6 ^B	***	NS	0.130
Ash, %	0.76	0.57	0.72 ^A	0.94 ^A	0.33 ^B	***	NS	0.157
Cholesterol,	32.2 ^b	33.7 ^a	32.4	33.7	32.8	NS	*	4.549
mg/100 g								

FB: faba bean; SB: soybean.

LT: Longissimus thoracis; ST: Semitendinosus; IP: Iliopsoas plus Psoas minor. MSE: mean square error; along the column A,B: P < 0.01. a,b: P < 0.05. ****, *, NS: P < 0.001, P < 0.05, not significant, respectively.

Table 4

Fatty acids classes (g/100 g), ratios and quality indexes in the soybean and faba bean groups and in the three tested muscles.

	Group	s	Muscles	Muscles		Prob.		MSE
	FB	SB	LT	ST	IP	Muscle	Group	
SFA	49.9 ^b	50.3 ^a	52.5 ^A	44.2 ^B	53.5 ^A	***	*	9.40
MUFA	36.3	36.6	36.7 ^B	39.9 ^A	32.8 ^C	***	NS	10.29
PUFA	13.4	12.8	10.5 ^c	16.1 ^a	12.91 ^b	***	NS	6.38
ω-3	1.79	1.74	1.52 ^A	2.17 ^A	1.61 ^B	***	NS	4.36
ω-6	11.4	10.9	8.76 ^C	13.7 ^A	11.1 ^B	***	NS	0.0042
CLAs	0.21	0.23	0.21 ^b	0.25 ^{Aa}	0.20 ^{Bb}	***	NS	0.012
PUFA/SFA	0.26	0.28	0.20 ^{Bb}	0.36 ^A	0.24 ^a	***	NS	0.011
ω-6/ω-3	6.44	6.24	5.90 ^b	6.26 ^a	6.95 ^a	*	NS	1.53
AI	0.53	0.53	0.57 ^a	0.42 ^b	0.60 ^a	***	NS	0.0033
TI	1.34	1.35	1.51 ^a	1.02 ^b	1.50 ^a	***	NS	0.027

FB: faba bean; SB: soybean.

LT: Longissimus thoracis; ST: Semitendinosus; IP: Iliopsoas plus Psoas minor.

SFA: saturated fatty acids; MUFA: monounsaturated fatty acids; PUFA: polyunsaturated fatty acids; CLA: conjugated linoleic acids; AI: Atherogenic Index; TI: Thrombogenic index. MSE: mean square error. Along the row: A,B,C: P < 0.01 and a,b,c: P < 0.05. ****, *, NS: P < 0.001, P < 0.05, not significant, respectively.

experiment: indeed Bohac and Rhee (1988) and Cifuni et al. (2004) utilised *Longissimus thoracis, Semimembranous* and *Semitendinosus,* while Eichhorn et al. (1986) and Browning et al. (1990) analysed *Biceps femoris* and *Longissimus thoracis.* As theorised by Wheeler, Davis, Stoeker, and Hatmon (1987), the cholesterol content may be affected by the different physiological function of each muscle.

Table 5 Fatty acid profile in the soybean and faba bean groups and in the three tested muscles.

Concerning the protein content of meat, the differences between groups (21.0% vs 21.3%, for FB and SB respectively) even if significant, appear too little to have any importance from the nutritional point of view.

4.3. Fatty acid profile

Meat from buffaloes fed faba bean showed significantly lower levels of total saturated fatty acids (SFA). This result is very interesting and it is mainly due to the differences found for some long-chain SFA (C15:0, C17:0, C20:0 – C24:0) which were significantly higher in SB group. According to Hornstra and Lussenberg (1975) and Renaud et al. (1986) long chain SFA are involved in thrombus formation. Differences in fatty acid profile using faba bean as protein source have been already found by other authors. Cutrignelli, Calabrò, et al. (2008) reported a significantly higher value for stearic acid in young Marchigiana bulls fed faba bean respect to those fed soybean meal solvent extract. Similarly, Scerra et al. (2011) found a higher palmitic acid value in meat from lambs fed soybean respect to those fed faba bean.

In the current trial, the contents of myristic (C14:0), palmitic (C16:0) and stearic (C18:0) acids were always lower than those recorded by Poli et al. (1996) for Chianina young bulls, thus confirming the favourable characteristics of buffalo meat.

The dietary treatment did not affect the level of most unsaturated fatty acids which are believed to be important for human health. Interestingly, the arachidonic acid (C20:4 ω -6), derived from C18:2 ω -6 through the action of Δ 5 and Δ 6 desaturase enzymes and elongase

$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		Groups		Muscles			Prob.		MSE
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	g/100 g FA	FB	SB	LT	ST	IP	Muscle	Group	
C15:0-iso 0.15 ^b 0.17 ^a 0.18 ^h 0.18 ^b 0.19 ^h *** NS 0.0081 C15:0-anteiso 0.20 ^h 0.25 ^A 0.22 ^{Abb} 0.18 ^B 0.27 ^A *** 0.081 C14:1-cts9 0.48 0.46 0.35 ^B 0.73 ^A 0.32 ^B *** NS 0.0076 C15:0 0.29 ^B 0.36 ^A 0.31 ^{ABb} 0.27 ^A 0.36 ^A *** 0.0003 C16:0 21.0 20.8 21.1 ^{AB} 20.4 ^B 21.6 ^A *** 0.0003 C17:0-iso 0.50 0.53 0.47 ^a 0.57 ^a 0.50 ^b ** NS 0.192 C17:0-iso 1.10 ^B 1.24 ^A 0.36 ^B *** 0.0011 C17:1-cist10 0.40 ^b 0.45 ^s 0.41 ^{Abb} 0.48 ^A 0.36 ^B *** 0.0029 C18:1-rans10 0.56 0.64 0.61 0.63 0.52 NS NS 0.652 C18:1-rans11 1.30 1.31 1.28 1.24 ^A 1.41 NS 0.023 0.165 0.64 0.651 0.56	C14:0	1.18	1.23	1.31 ^A	0.91 ^B	1.40 ^A	***	NS	0.53
C15:0-anteiso 0.20 ⁸ 0.25 ^A 0.23 ^{Bb} 0.18 ^B 0.27 ^{Aa} *** *** NS 0.081 C14:1-cisS 0.48 ^B 0.46 ^A 0.35 ^B 0.73 ^A 0.32 ^B *** NS 0.0276 C15:0 0.29 ^B 0.36 ^A 0.17 ^{AB} 0.02 ^{BA} *** NS 0.0276 C17:0-iso 0.50 0.53 0.47 ^A 0.57 ^A 0.50 ^B ** NS 0.122 C16:1n-7 1.18 1.14 1.10 ^B 1.38 ^A 1.00 ^B *** 0.0011 C17:0-is10 0.40 ^A 0.45 ^A 0.41 ^{ABD} 0.48 ^{AA} 0.36 ^B *** NS 0.0629 C18:1-trans10 0.56 0.64 0.61 0.63 0.52 NS NS 0.029 C18:1-trans11 1.30 1.31 1.28 1.24 1.41 NS 0.024 C18:1-trans10 0.56 0.64 0.61 0.33 0.52 NS NS 0.024 C18:1-trans10 1.30 3.1 1.28 1.24 1.41 NS	C15:0-iso	0.15 ^b	0.17 ^a	0.18 ^A	0.12 ^B	0.19 ^A	***	NS	0.00096
Cl4:1-cis9 0.48 0.46 0.35 ^k 0.73 ^A 0.32 ^k *** NS 0.0276 C15:0 0.20 ^g 0.36 ^A 0.31 ^{ABD} 0.27 ^{BC} 0.36 ^A *** NS 0.243 C17:0-iso 0.50 0.53 0.47 ^A 0.57 ^A 0.50 ^b * NS 0.192 C16:1-7 1.18 1.14 1.10 ^B 1.32 ^A 0.97 ^B 1.24 ^A *** NS 0.614 C17:0 1.10 ^B 1.22 ^A 1.23 ^A 0.97 ^B 1.24 ^A *** 0.0011 C18:1-trans10 0.46 ^D 0.45 ^A 0.41 ^{ABb} 0.48 ^{Aa} 0.36 ^B *** NS 0.662 C18:1-trans11 1.30 1.31 1.28 1.24 1.41 NS NS 0.024 C18:1-trans11 1.30 1.31 1.28 1.24 1.41 NS NS 0.024 C18:1-trans11 1.29 1.26 1.17 ^B 1.49 ^A 1.17 ^B NS 0.004 <td>C15:0-anteiso</td> <td>0.20^B</td> <td>0.25^A</td> <td>0.22^{ABb}</td> <td>0.18^B</td> <td>0.27^{Aa}</td> <td>***</td> <td>***</td> <td>0.081</td>	C15:0-anteiso	0.20 ^B	0.25 ^A	0.22 ^{ABb}	0.18 ^B	0.27 ^{Aa}	***	***	0.081
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C14:1-cis9	0.48	0.46	0.35 ^B	0.73 ^A	0.32 ^B	***	NS	0.0276
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C15:0	0.29 ^B	0.36 ^A	0.31 ^{ABb}	0.27 ^{Bc}	0.36 ^{Aa}	***	***	0.00036
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C16:0	21.0	20.8	21.1 ^{AB}	20.4 ^B	21.6 ^A	***	NS	2.43
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C17:0-iso	0.50	0.53	0.47 ^a	0.57 ^a	0.50^{b}	*	NS	0.192
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C16:1n-7	1.18	1.14	1.10 ^B	1.38 ^A	1.00 ^B	***	NS	0.614
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C17:0	1.10 ^B	1.22 ^A	1.23 ^A	0.97 ^B	1.24 ^A	***	***	0.0011
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C17:1-cis10	0.40 ^b	0.45 ^a	0.41 ^{ABb}	0.48 ^{Aa}	0.36 ^B	***	*	0.0089
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C18:0	24.6	25.0	27.2 ^A	19.8 ^B	27.6 ^A	***	NS	6.062
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C18:1-trans10	0.56	0.64	0.61	0.63	0.52	NS	NS	0.039
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C18:1-trans11	1.30	1.31	1.28	1.24	1.41	NS	NS	0.165
C18:1-cis111.291.26 1.17^B 1.49^A 1.17^B ***NS 0.024 C18:2n-67.917.87 6.49^B 9.13^{Aa} 8.10^{Ab} ***NS 2.77 C20:0 0.19^b 0.21^a 0.21^A 0.17^B 0.21^A **** 0.0023 C18:3n-6 0.11 0.10 0.09^B 0.14^A 0.09^B ***NS 0.0023 C20:1-cis11 0.10 0.11 0.10^{ABb} 0.13^{Aa} 0.08^B ***NS 0.0013 C18:3n-3 0.57 0.59 0.51^b 0.65^a 0.56^b *NS 0.007 c18:3n-3 0.57 0.59 0.51^b 0.66^a 0.56^b *NS 0.007 c18:3n-3 0.57 0.05 0.04^b 0.06^a 0.5^b *NS 0.007 c18:3n-3 0.57 0.05 0.04^b 0.06^a 0.5^b *NS 0.007 c18:3n-3 0.57 0.05 0.04^b 0.06^a 0.5^b *NS 0.0032 c20:2-cis11,14 0.21 0.20 0.18^B 0.26^A 0.18^B **NS 0.0026 C22:0 0.17^b 0.21^a 0.17^b 0.21^a 0.17^b * 0.0026 C20:3n-6 0.63^A 0.53^B 0.42^C 0.80^A 0.56^B **NS 0.0078 C20:3n-6 0.63^A 0.53^B 0.42^C 0.80^A 0.56^B **NS 0.0078	C18:1n-9	31.0	30.8	31.2 ^{Ab}	33.2 ^{Aa}	27.8 ^B	***	NS	8.70
C18:2n-67.917.87 649^B 9.13^{Aa} 8.10^{Ab} ***NS 2.77 C20:0 0.19^b 0.21^a 0.21^A 0.17^B 0.21^A **** 0.0023 C18:3n-6 0.11 0.10 0.09^B 0.14^A 0.09^B ***NS 0.0028 C20:1-cis11 0.10 0.11 0.10^{Abb} 0.13^{Aa} 0.08^B ***NS 0.0013 C18:3n-3 0.57 0.59 0.51^b 0.65^a 0.56^b *NS 0.015 cis9-trans 11 CLA 0.05 0.05 0.04^b 0.06^a 0.05^b *NS 0.0022 cis9-trans 12 CLA 0.17 0.18 0.17^{ab} 0.19^a 0.15^b *NS 0.0022 C20:2-cis11,14 0.21 0.20 0.18^B 0.26^A 0.18^B ***NS 0.0026 C20:3n-6 0.63^A 0.53^B 0.42^C 0.80^A 0.56^B ***NS 0.0026 C20:3n-3 0.29 0.28 0.23^{Bb} 0.33^{Aa} 0.28^b **NS 0.0078 C20:4n-6 0.47^a 0.21^A 0.16^B 0.22^A 0.18^B *** 0.0026 C23:0 0.17^B 0.21^A 0.16^B 0.29^A 0.18^B *** 0.0024 C23:0 0.17^B 0.21^A 0.16^B *** $*$ 0.0024 C24:1-cis15 0.39^b 0.49^A 0.16^B 0.21^A 0.18^B *** $*$	C18:1-cis11	1.29	1.26	1.17 ^B	1.49 ^A	1.17 ^B	***	NS	0.024
C20:0 0.19^{b} 0.21^{a} 0.21^{A} 0.17^{B} 0.21^{A} **** 0.0023 C18:3n-6 0.11 0.10 0.09^{B} 0.14^{A} 0.09^{B} ***NS 0.0088 C20:1-cis11 0.10 0.11 0.10^{ABb} 0.13^{Aa} 0.08^{B} ***NS 0.0013 C18:3n-3 0.57 0.59 0.51^{b} 0.65^{a^3} 0.56^{b} *NS 0.0015 cis9-trans11 CLA 0.05 0.05 0.04^{b} 0.06^{a^3} 0.05^{b} *NS 0.0027 trans10-trans12 CLA 0.17 0.18 0.17^{ab} 0.13^{a} 0.15^{b} *NS 0.0032 C20:2-cis11,14 0.21 0.20 0.18^{B} 0.26^{A} 0.18^{B} ***NS 0.0026 C20:3n-6 0.63^{A} 0.53^{B} 0.42^{C} 0.80^{A} 0.56^{B} ***NS 0.0078 C20:3n-3 0.29 0.28 0.23^{Bb} 0.33^{Aa} 0.28^{b} ***NS 0.0078 C20:4n-6 2.43^{a} 2.05^{b} 1.56^{B} 3.19^{Ab} 2.05^{Ba} ****** 0.024 C20:5n-3 0.22^{A} 0.16^{B} 0.21^{A} 0.16^{B} *** 0.0024 0.024^{A} 0.026^{A} C24:0 0.16^{b} 0.20^{a} 0.16^{B} 0.21^{A} 0.15^{B} *** 0.024^{A} 0.025^{A} C20:3n-3 0.29^{A} 0.16^{B} 0.21^{A} 0.16^{B} 0.20^{A} <	C18:2n-6	7.91	7.87	6.49 ^B	9.13 ^{Aa}	8.10 ^{Ab}	***	NS	2.77
C18:3n-60.110.100.09 0.14^{A} 0.09***NS0.0088C20:1-cis110.100.11 0.10^{ABb} 0.13^{Aa} 0.08^{B} ***NS0.0013C18:3n-30.570.59 0.51^{b} 0.65^{a} 0.56^{b} *NS0.015cis9-trans11 CLA0.050.05 0.04^{b} 0.06^{a} 0.05^{b} *NS0.0007trans10-trans12 CLA0.170.18 0.17^{ab} 0.19^{a} 0.15^{b} *NS0.0032C20:2-cis11,140.210.20 0.18^{B} 0.26^{A} 0.18^{B} ***NS0.0026C20:2-cis1,140.210.20 0.18^{B} 0.26^{A} 0.18^{B} ***NS0.032C20:2-cis1,140.210.20 0.18^{B} 0.26^{A} 0.18^{B} ***NS0.0026C20:3 n-60.63^{A}0.53^{B} 0.42^{C} 0.80^{A} 0.56^{B} ***NS0.0078C20:3 n-30.290.28 0.23^{Bb} 0.33^{Aa} 0.28^{b} **NS0.0078C20:4 n-62.43^{a}2.05^{b} 1.56^{B} 3.19^{Ab} 2.05^{Ba} ***NS0.024C20:5 n-30.22^{B} 0.19^{A} 0.16^{B} 0.22^{A} 0.18^{B} *** 0.024 0.034C24:0 0.16^{b} 0.20^{a} 0.16^{B} 0.21^{A} 0.15^{B} ****0.0034C24:1-cis15 0.39^{b} 0.49^{a} <	C20:0	0.19 ^b	0.21 ^a	0.21 ^A	0.17 ^B	0.21 ^A	***	*	0.0023
C20:1-cis110.100.110.10^{ABb}0.13^{Aa}0.08^B***NS0.0013C18:3n-30.570.590.51^b0.65^a0.56^b*NS0.015cis9-trans 11 CLA0.050.050.04^b0.06^a0.05^b*NS0.0007trans10-trans12 CLA0.170.180.17^{ab}0.19^a0.15^b*NS0.0032C20:2-cis11,140.210.200.18^B0.26^A0.18^B***NS0.0026C20:3 n-60.63^A0.53^B0.42^C0.80^A0.56^B*****0.015C20:3 n-30.290.280.23^{Bb}0.33^{Aa}0.28^b***NS0.0078C20:4 n-62.43^a2.05^b1.56^B3.19^{Ab}2.05^{Ba}******0.024C20:5 n-30.22^B0.19^A0.17^B0.22^A0.18^B***0.024C24:00.17^B0.21^A0.16^B0.22^A0.18^B***0.024C20:4 n-62.43^a2.05^b1.56^B3.19^{Ab}2.05^{Ba}***%0.024C20:5 n-30.22^B0.19^A0.17^B0.22^A0.18^B***%0.024C24:00.16^b0.20a0.16^B0.21^A0.15^B***%0.0034C24:1-cis150.39^b0.49^30.440.410.43NS\$0.0015C24:n-60.370.32_c0.26^B_a0.46^A0.33^B_a	C18:3n-6	0.11	0.10	0.09 ^B	0.14 ^A	0.09 ^B	***	NS	0.00088
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C20:1-cis11	0.10	0.11	0.10 ^{ABb}	0.13 ^{Aa}	0.08 ^B	***	NS	0.0013
cis9-trans 11 CLA0.050.050.04b0.06a0.05b*NS0.0007trans10-trans12 CLA0.170.180.17ab0.19a0.15b*NS0.0032C20:2-cis11,140.210.200.18B0.26A0.18B****NS0.0039C22:00.17b0.21a0.17b0.21a0.17b****NS0.0066C20:3n-60.63A0.53B0.42C0.80A0.56B******0.017C20:3n-30.290.280.23 ^{Bb} 0.33 ^{Aa} 0.28b***NS0.0078C20:4n-62.43a2.05b1.56B3.19Ab2.05 ^{Ba} *****0.555C23:00.17B0.21A0.16B0.22A0.18B******0.024C20:5n-30.22B0.19A0.16B0.22A0.18B******0.0034C24:00.16b0.20a0.16B0.21A0.15B******0.0017C24:1-cis150.39b0.49a0.440.410.43NS*0.0021C22:4n-60.370.320.26 ^{Ba} 0.46 ^A 0.33 ^{Ba} ***NS0.021	C18:3n-3	0.57	0.59	0.51 ^b	0.65 ^a	0.56 ^b	*	NS	0.015
trans10-trans12 CLA0.170.180.17^{ab}0.19^{a}0.15^{b}*NS0.0032C20:2-cis11,140.210.200.18^B0.26^A0.18^B***NS0.0039C22:00.17^{b}0.21^{a}0.17^{b}0.21^{a}0.17^{b}***NS0.0032C20:3n-60.63^A0.53^B0.42^C0.80^A0.56^B*****0.0026C20:3n-30.290.280.23^{Bb}0.33^{Aa}0.28^{b}**NS0.0078C20:4n-62.43^a2.05^b1.56^B3.19^{Ab}2.05^{Ba}*****0.555C23:00.17^B0.21^A0.16^B0.22^A0.18^B***0.024C20:5n-30.22^B0.19^A0.16^B0.22^A0.16^B******0.0017C24:1-cis150.39^b0.49^a0.440.410.43NS***0.0021C22:4n-60.370.320.26^{Ba}0.46^A0.33^{Ba}***NS0.021	cis9-trans 11 CLA	0.05	0.05	$0.04^{\rm b}$	0.06^{a}	0.05 ^b	*	NS	0.0007
$ \begin{array}{cccccc} C20:2-cis11,14 & 0.21 & 0.20 & 0.18^B & 0.26^A & 0.18^B & $^{***} & NS & 0.039 \\ C22:0 & 0.17^b & 0.21^a & 0.17^b & 0.21^a & 0.17^b & $^{***} & 0.0026 \\ C20:3n-6 & 0.63^A & 0.53^B & 0.42^C & 0.80^A & 0.56^B & $^{***} & NS & 0.0078 \\ C20:3n-3 & 0.29 & 0.28 & 0.23^{Bb} & 0.33^{Aa} & 0.28^b & $^{**} & NS & 0.0078 \\ C20:4n-6 & 2.43^a & 2.05^b & 1.56^B & 3.19^{Ab} & 2.05^{Ba} & $^{***} & $^{**} & 0.555 \\ C23:0 & 0.17^B & 0.21^A & 0.16^B & 0.22^A & 0.18^B & $^{***} & $^{***} & 0.0245 \\ C20:5n-3 & 0.22^B & 0.19^A & 0.16^B & 0.22^A & 0.16^B & $^{***} & $^{***} & 0.0017 \\ C24:0 & 0.16^b & 0.20^a & 0.16^B & 0.21^A & 0.15^B & $^{**} & $^{**} & 0.0017 \\ C24:1-cis15 & 0.39^b & 0.49^a & 0.44 & 0.41 & 0.43 & NS & $^{**} & 0.0055 \\ C22:4n-6 & 0.37 & 0.32 & 0.26^{Ba} & 0.46^A & 0.33^{Ba} & $^{***} & NS & 0.021 \\ \end{array}$	trans10-trans12 CLA	0.17	0.18	0.17 ^{ab}	0.19 ^a	0.15 ^b	*	NS	0.0032
C22:0 0.17^{b} 0.21^{a} 0.17^{b} 0.21^{a} 0.17^{b} *** 0.0026 C20:3n-6 0.63^{A} 0.53^{B} 0.42^{C} 0.80^{A} 0.56^{B} **** 0.015 C20:3n-3 0.29 0.28 0.23^{Bb} 0.33^{Aa} 0.28^{b} **NS 0.0078 C20:4n-6 2.43^{a} 2.05^{b} 1.56^{B} 3.19^{Ab} 2.05^{Ba} **** 0.555 C23:0 0.17^{B} 0.21^{A} 0.16^{B} 0.22^{A} 0.18^{B} *** 0.0245 C20:5n-3 0.22^{B} 0.19^{A} 0.17^{B} 0.29^{A} 0.16^{B} *** 0.0017 C24:0 0.16^{b} 0.20^{a} 0.16^{B} 0.21^{A} 0.15^{B} *** 0.0017 C24:1-cis15 0.39^{b} 0.49^{a} 0.44 0.41 0.43 NS* 0.0021 C22:4n-6 0.37 0.32 0.26^{Ba} 0.46^{A} 0.33^{Ba} ***NS 0.021	C20:2-cis11,14	0.21	0.20	0.18 ^B	0.26 ^A	0.18 ^B	***	NS	0.0039
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	C22:0	0.17 ^b	0.21 ^a	0.17 ^b	0.21 ^a	0.17 ^b	*	*	0.0026
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	C20:3n-6	0.63 ^A	0.53 ^B	0.42 ^C	0.80 ^A	0.56 ^B	***	**	0.015
C20:4n-62.43a2.05b 1.56^{B} 3.19^{Ab} 2.05^{Ba} **** 0.55 C23:0 0.17^{B} 0.21^{A} 0.16^{B} 0.22^{A} 0.18^{B} ****** 0.0245 C20:5n-3 0.22^{B} 0.19^{A} 0.17^{B} 0.29^{A} 0.16^{B} ****** 0.0034 C24:0 0.16^{b} 0.20^{a} 0.16^{B} 0.21^{A} 0.15^{B} *** 0.0017 C24:1-cis15 0.39^{b} 0.49^{a} 0.44 0.41 0.43 NS* 0.0055 C22:4n-6 0.37 0.32 0.26^{Ba} 0.46^{A} 0.33^{Ba} ***NS 0.021	C20:3n-3	0.29	0.28	0.23 ^{Bb}	0.33 ^{Aa}	0.28 ^b	**	NS	0.0078
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	C20:4n-6	2.43 ^a	2.05 ^b	1.56 ^B	3.19 ^{Ab}	2.05 ^{Ba}	***	*	0.555
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	C23:0	0.17 ^B	0.21 ^A	0.16 ^B	0.22 ^A	0.18 ^B	***	***	0.0245
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	C20:5n-3	0.22 ^B	0.19 ^A	0.17 ^B	0.29 ^A	0.16 ^B	***	***	0.0034
C24:1-cis15 0.39 ^b 0.49 ^a 0.44 0.41 0.43 NS * 0.0065 C22:4n-6 0.37 0.32 0.26 ^{Ba} 0.46 ^A 0.33 ^{Ba} *** NS 0.021	C24:0	0.16 ^b	0.20 ^a	0.16 ^B	0.21 ^A	0.15 ^B	**	*	0.0017
C22:4n-6 0.37 0.32 0.26 ^{Ba} 0.46 ^A 0.33 ^{Ba} *** NS 0.021	C24:1-cis15	0.39 ^b	0.49 ^a	0.44	0.41	0.43	NS	*	0.0065
	C22:4n-6	0.37	0.32	0.26 ^{Ba}	0.46 ^A	0.33 ^{Ba}	***	NS	0.021
C22:5n-3 0.54^{a} 0.45^{b} 0.38^{B} 0.69^{A} 0.44^{B} *** * 0.039	C22:5n-3	0.54 ^a	0.45 ^b	0.38 ^B	0.69 ^A	0.44 ^B	***	*	0.039
C22:6n-3 0.20 ^b 0.24 ^a 0.22 0.22 0.21 NS * 0.010	C22:6n-3	0.20 ^b	0.24 ^a	0.22	0.22	0.21	NS	*	0.010

FB: faba bean group; SB: soybean group.

LT: Longissimus thoracis; ST: Semitendinosus; IP: Iliopsoas plus Psoas minor.

MSE: mean square error. Along the row: A,B,C: P < 0.01 and a,b,c: P < 0.05.***, **, NS: P < 0.001, P < 0.01, P < 0.05, not significant, respectively.

(Gurr & Harvood, 1996), was significantly higher in FB group while its precursor was not found higher in meat from buffaloes fed faba bean. Similarly, C20:5 ω -3 was significantly higher in FB than in SB group, while its precursor C18:3 ω -3 wasn't different between groups. The ω -3 fatty acids play a very important physiological role, especially during fetal and infant growth, in the formation of the central nervous system and retina (Bourre, 2003; Bowen & Clandinin, 2005). Being antithrombotic, anti-inflammatory, antiarrhythmic and favoring plaque stabilization (Galli & Marangoni, 2006; Hu et al., 1999; Simon, Pong, Bernert, & Browner, 1995), they are also important for the prevention of cardiovascular diseases. Many significant differences were found between the three muscles studied concerning the FA profile, this was probably due to two main reasons: the sampling of the edible part of the muscle that also includes the infiltration fat, and the different type of muscle that, having a different role, favours the accumulation of different nutrients

These results are very interesting from the nutritional point of view. Muscles with a high percentage of unsaturated fatty acids (UFA) generally scored higher in taste panel evaluation and foods with high UFA, especially PUFA, are good for human health (Westerling & Hedrick, 1979). Conjugated linoleic acid (CLA) has been shown to have immunomodulating, anticarcinogenic and antiartheriosclerosis properties (Pastuschenko, Matthes, Hein, & Holzer, 2000; Whigham, Cook, & Atkinson, 2000). Additionally, high prominence is attributed to the MUFA, and particularly by the oleic acids that, reducing the oxidation of the cholesterol LDL, may slow the progression of atherosclerosis (Parthasarathy et al., 1990). Concerning the ω -6/ ω -3 ratio, if we compare the results obtained in this trial with those cited by Daley, Abbott, Doyle, Nader, and Larson (2010), we can notice that it is higher than the value reported for grass-fed beef, but lower than that reported for grain-fed beef. Nevertheless, the optimal nutritional value of the ω -6/ ω -3 ratio has still not been completely assessed both for human and animals. The studies on the relationship between ω -6/ ω -3 ratio and the pathogenesis of many diseases, including cancer, cardiovascular, inflammatory and autoimmune diseases, indicate that the optimal ratio may vary with the disease or condition under consideration. This is consistent with the fact that chronic diseases are multigenic and multifactorial. On the whole, the health authorities of many countries promote the intake of foods containing high levels of ω -6/ ω -3 ratio (West Suitor & Meyers, 2006) which seems to reduce the risk of many of the chronic diseases (Simopoulos, 2003).

According to Ulbricht and Southgate (1991), atherogenic and thrombogenic indexes might better characterize the health properties of vegetables or animal food respect to a simple approach based on total saturated fatty acids or on the ratio polyunsaturated/saturated fatty acids (Burr, 1989; Fehily, Pickering, Yarnell, & Elwood, 1994). Both the indexes were not affected by dietary treatment but were different between muscles. The AI for buffalo meat recorded in this trial was lower than those reported by Ulbricht and Southgate (1991) for raw minced beef and by Badiani et al. (2002) for cooked beef (0.72 and 0.77, respectively) while thrombogenic TI was lower if compared to that reported by these authors.

5. Conclusions

The results show that faba bean, that leads, by itself, to agronomical and economical advantages, can be used as a protein source alternative to soybean in the diet of young buffalo bulls for the production of high quality meat. In particular, fat percentage, cholesterol content and total saturated fatty acids decreased in meat from animals fed faba bean.

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