



REVIEW

REVISED Circadian stabilization loop: the regulatory hub and therapeutic target promoting circadian resilience and physiological health [version 2; peer review: 2 approved]

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Abstract

The circadian clock is a fundamental biological mechanism that orchestrates essential cellular and physiological processes to optimize fitness and health. The basic functional unit is the cell-autonomous oscillator, consisting of intersecting negative feedback loops. Whereas the core loop is primarily responsible for rhythm generation, auxiliary loops, most notably the secondary or stabilization loop, play pivotal roles to confer temporal precision and molecular robustness. The stabilization loop contains opposing nuclear receptor subfamilies REV-ERBs and retinoic acid receptor-related orphan receptors (RORs), competing to modulate rhythmic expression of the basic helix-loop-helix ARNT like 1 (*Bmal1*) genes in the core loop as well as other clock-controlled genes. Therefore, REV-ERBs and RORs are strategically located to interface the oscillator and the global transcriptomic network, promoting cellular homeostasis and physiological fitness throughout lifespan. Disruption of REV-ERB and ROR functions has been linked with diseases and aging, and pharmacological manipulation of these factors has shown promise in various mouse disease models. Nobiletin is a natural compound that directly binds to and activates ROR α/γ , modulating circadian rhythms, and shows robust *in vivo* efficacies to combat clock-associated pathophysiologies and age-related decline. Results from several studies demonstrate an inverse relation between nobiletin efficacy and clock functional state, where nobiletin elicits little effect in young and healthy mice with growing efficacy as the clock is perturbed by environmental and genetic challenges. This mode of action is consistent with the function of the stabilization loop to promote circadian and physiological resilience. Future studies should further investigate the function and mechanism of REV-ERBs and RORs, and test strategies targeting these factors against disease and aging.

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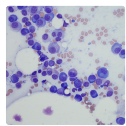
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Any reports and responses or comments on the article can be found at the end of the article.

Keywords

Circadian oscillator, core loop and stabilization/secondary loop, REV-ERBs and RORs, ligands and drugs, circadian amplitude and resilience, physiological health, healthy aging



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REVISED Amendments from Version 1

This new version of the article does not include any major changes in the shape and contents of the original one. However, some minor changes have been made. We have responded to reviewer comments and expanded on a statement in some parts including “The circadian timing system and health implications”, “The stabilization loop”, and “Nobiletin (NOB): A natural ROR agonist”. We have revised references to address the content better.

Any further responses from the reviewers can be found at the end of the article

The circadian timing system and health implications

Circadian rhythms are daily cycles of intrinsic processes in living organisms. While light/dark cycles of our environment are the predominant input (or zeitgeber, time giver) to reset our internal rhythms, it is now clear that other factors including feeding-fasting state, nutrients, physical activity, and temperature are all capable of manipulating the circadian cycle.^{1–3} Fundamentally, the circadian timing system is a molecular circuit governing cellular and physiological homeostasis throughout lifespan. Alterations to this clock machinery, by either environmental stresses or genetic defects, have been shown to cause or correlate with dysfunction of diverse physiological processes and increased risks for various diseases involving both peripheral organs and the brain.^{4–6}

At the pinnacle of the circadian timing system is the master pacemaker located in the suprachiasmatic nucleus (SCN) of the hypothalamus. The SCN clock synchronizes semi-autonomous cellular oscillators in other brain regions and peripheral organs through neuronal and hormonal signals.^{7–9} The ubiquitous cellular oscillator, present in the SCN and throughout the body, contains interlocked transcriptional and translational feedback loops controlling the expression of downstream target genes.¹⁰ The core clock genes functioning in the oscillator include circadian locomotor output cycles kaput (*Clock*)/neuronal PAS domain-containing protein 2 (*Npas2*), basic helix-loop-helix ARNT like 1 (*Bmal1*), period 1 (*Per1*)/period 2 (*Per2*)/period 3 (*Per3*), cryptochrome 1 (*Cry1*)/cryptochrome 2 (*Cry2*), *Rev-erba/Rev-erbb* (nuclear receptor subfamily 1 group D member 1/2 (*Nr1d1/2*)), and retinoic acid receptor-related orphan receptor alpha (*Rora*)/retinoic acid receptor-related orphan receptor beta (*Rorb*)/Retinoic acid receptor-related orphan receptor gamma (*Rorc*) (*Nr1f1/2/3*), D-box binding protein (*Dbp*), and nuclear factor, interleukin-3 regulated protein (*Nfil3*). By acting on consensus promoter elements or directing the expression of secondary regulators of gene expression, the encoded core clock proteins play a prevalent role in the global gene expression landscape where more than 80% of genes have been shown to oscillate in at least one location in the body.^{11,12}

Perhaps one of the most important physiological functions of the clock is to safeguard energy homeostasis.^{13,14} It has been postulated that an evolutionary origin of the circadian system is energy partitioning: photosynthesis using oxygen during the day and anaerobic metabolism including nitrogen fixation at night.¹⁵ In mammals, central and peripheral clocks coordinately drive rhythmic expressions of metabolic-related genes in organs with high metabolic activity including liver, muscle, and adipose tissue.^{3,16–18} Over the past 15 years or so, a growing body of evidence has established that the clock gene machinery influences energy homeostasis directly and genetic mutations in clock genes lead to metabolic dysfunctions, including deficient insulin resistance, glucose intolerance, leptin resistance, and abnormal glucocorticoid and melatonin levels.^{17,19,20} In accordance, human subjects who were exposed to a controlled circadian misalignment condition displayed glucose intolerance, insulin resistance, and other comorbidities.^{21–23} In addition, our lifestyle choices that affect circadian rhythms may also evoke adverse metabolic consequences. For example, external stimuli including abnormal light exposure,²⁴ jet-lag,^{20,25} and high-fat diet induced-obesity^{26,27} can trigger desynchronization of the internal clock accompanied by many tissue disorders. Furthermore, sleep deprivation, a common occurrence in modern lifestyle, is associated with increased body mass index and type 2 diabetes incidence and has been identified as an independent risk factor for hypertension, obesity, and coronary heart disease.^{28,29} In addition, sleep and feeding alterations and shift work are highly correlated with elevated metabolic syndrome markers such as triglycerides, and lower high-density lipoprotein (HDL)-cholesterol levels.^{29–31}

Dysregulated clocks are also involved in brain dysfunction and diseases.^{32–34} Sleep is well known to be regulated by the clock, and elegant studies combining human genetics and mechanistic investigation have revealed molecular links between several mutations in clock genes, including *PER2* and casein kinase I isoform delta (*CSNK1D*), and sleep disorders.³⁵ An emerging area of interest is the crosstalk between the clock and neurodegenerative diseases.^{36–38} Circadian clocks have been shown to control several aspects of brain functions linked to neurodegeneration including dopamine synthesis, inflammatory response, oxidative stress, and cellular metabolism.^{33,39} Consistently, circadian and sleep disruptions are closely associated with neurodegenerative diseases including Alzheimer’s disease and Parkinson’s disease,⁴⁰ as evidenced by amyloid-beta (A β) oscillation,⁴¹ sundowning behaviors,⁴² and neuronal inflammation in mouse genetic mutants.⁴⁰

Given the fundamental role of the clock in cellular and physiological homeostasis and the myriads of chronic diseases associated with circadian dysregulation, it is not surprising that age-related decline over time is strongly correlated with and likely exacerbated by dysfunction in the clock system.⁴³ It is well known that a number of physiological parameters display blunted circadian rhythms during aging, including sleep, temperature, and hormone secretion.^{43,44} More recently, global transcriptomic profiling revealed profound rewiring in the clock network, notably dampening of oscillatory gene expression in accordance with the physiological decline.⁴⁵ A key role of the circadian rhythms in aging is further highlighted by two large-scale gene profiling studies where circadian gene expression changes emerged from unbiased analyses as a top underlying pathway during aging.^{46,47} For example, a comparative multi-tissue gene profiling approach was undertaken to search for pathways correlated with maximum lifespan in 26 species, and identified the circadian system as a pillar that governs metabolic and inflammatory pathways for longevity regulation.⁴⁷ Furthermore, interventional fasting paradigms designed to incorporate circadian timing were recently reported to markedly prolong lifespan in *Drosophila* and mice,^{48,49} including 35% lifespan extension in male mice. The convergent spotlight on circadian remodeling during aging provides compelling evidence for the notion that a robust circadian system is key to health and healthspan.⁵⁰

The stabilization loop

The core loop of the oscillator is primarily responsible for generating the near-24hr rhythm *via* the negative feedback between CLOCK/BMAL1 and CRY/PER. Through binding to E-box elements, the CLOCK/BMAL1 heterodimer activates the expression of many Clock-Controlled Genes (CCGs). As PER/CRY proteins accumulate and reach critical levels in the cytoplasm, they translocate to the nucleus to inhibit the activity of CLOCK/BMAL1, thereby inhibiting their own transcription.¹⁰ On the other hand, the secondary loop, mainly involving the opposing transcription factors REV-ERBs and RORs,^{51,52} confers stability and robustness for the core loop, and is also strategically located at the interface between the core oscillator and many downstream clock-controlled genes (Figure 1). Growing evidence suggests a regulatory and therapeutic potential of the stabilization loop in physiology, disease, and aging.⁵³

REV-ERBs and RORs, the main components of the stabilization loop, are multi-functional nuclear receptors to repress and activate target gene expression, respectively.^{52,54,55} REV-ERBs and RORs bind as monomers to the same consensus

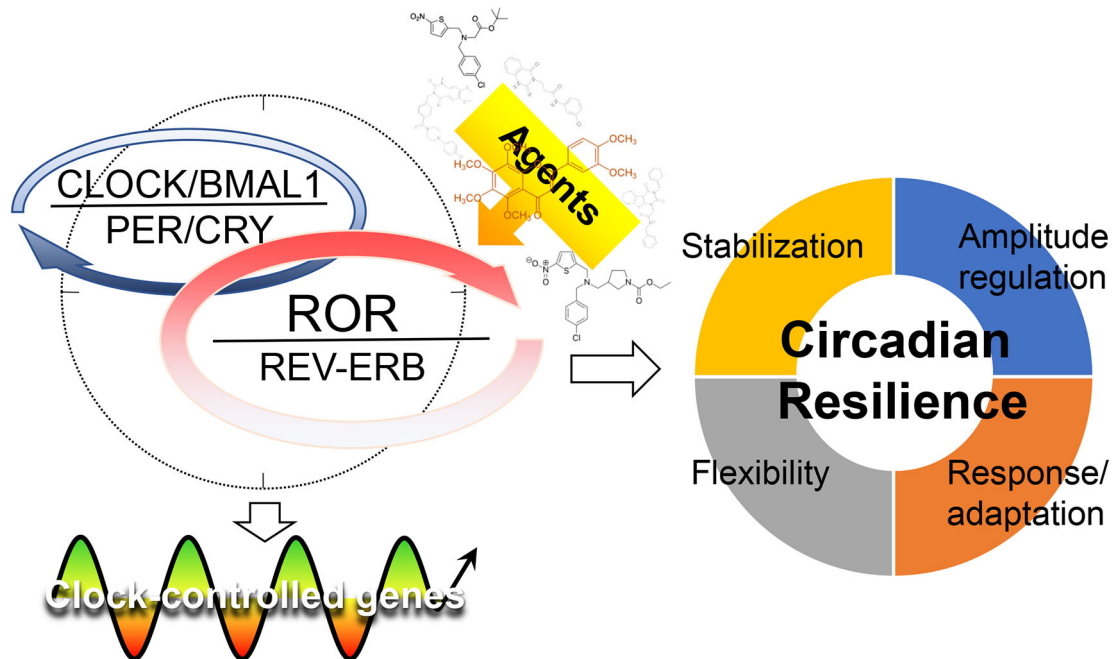


Figure 1. The stabilization loop of the circadian oscillator is strategically situated at the interface of the rhythm-generating core loop and the circadian output network. The core oscillator mainly consists of core and stabilization loops as indicated by the dotted circle. The REV-ERB and retinoic acid receptor-related orphan receptors (RORs) compete at the consensus ROR response elements (ROREs) of target gene promoters, including basic helix-loop-helix ARNT like 1 (*Bmal1*) and many clock-controlled genes (CCGs), to regulate circadian transcription in a tissue-specific manner. They may interact *via* other mechanisms and in clock-independent processes – see text for details. REV-ERBs and RORs play regulatory roles in many tissue and organismal functions and targeting these receptors by small-molecule agents may strengthen circadian resilience, ultimately conferring beneficial effects to promote health and healthspan. CLOCK, circadian locomotor output cycles kaput; CRY, cryptochrome; PER, period.

sequence, namely the ROR response elements (ROREs, or RREs) consisting of an (A/G) GGTCA flanked by an A/T-rich 5' in the regulatory regions of the target genes.^{56–58} In the core oscillator, perhaps the most important function of REV-ERB/ROR is to maintain and fine-tune the oscillatory expression of *Bmal1*, encoding the chief circadian transcription activator in the core loop, thereby serving as a stabilizing mechanism.^{52,55,59} In addition, several other core clock genes have been shown to possess the RORE elements in their promoter region,⁶⁰ suggesting an important stabilizing function by these opposing regulators. More broadly, gene expression and profiling studies have illustrated a prominent role of REV-ERBs and RORs in transcriptomic landscape in various tissues/cells.^{61–65} For example, studies of liver-specific ROR α /ROR γ double knockout mice revealed a key role of RORs in lipogenesis, *via* direct transcriptional regulation of the insulin-induced gene 2 (*Insig2*)-sterol regulatory element-binding protein 1 (*SREBP*) cascade.⁶⁵

Traditionally REV-ERBs and RORs are considered to interact and compete as repressors and activators on shared target genes. While *Rora* expressions display relatively moderate circadian oscillatory amplitude, *Rory* is more rhythmic, exhibiting a strong oscillatory pattern of expression in several tissues such as liver, brown adipose tissue (BAT), kidney, and small intestines with peak expression around ZT16, but not in the hypothalamus, muscle, or brainstem.^{66–68} Notably, *Rev-erbs* are among the most oscillatory genes (highest amplitude) in both protein and mRNA expression.^{64,69} It is believed that their opposing functions and circadian patterns (expression and promoter recruitment) play a key role in amplitude regulation of clock-controlled genes. In a pioneering study, the mRNA oscillation of *Bmal1*, *Clock*, and *Cry1* genes show strong amplitude in wild-type animals but is diminished in *Rev-erba* deficient mice.⁵⁵ Although dispensable for *Bmal1* oscillation *per se*, RORs were found to contribute to its amplitude.^{52,70,71} More recently, correlation analysis of publicly available mouse circadian transcriptomic datasets, including both microarray and RNA-sequencing, identified a strong correlation between *Rorc* expression level and amplitude with the percentage of cycling transcripts in respective tissues, consistent with a regulatory role of ROR γ in circadian oscillation.⁷² In addition to jointly regulating *Bmal1* transcription, additional modes of genetic and molecular interplay exist between REV-ERBs and RORs. *Rorc* itself contains a functional RORE element in its promoter, therefore subject to transcriptional regulation by REV-ERBs/RORs.⁶⁰ Moreover, molecular studies have demonstrated a facilitated recruitment mechanism where REV-ERBs are recruited to the target gene promoter by RORs, in a process that requires chromatin remodeling by SWI/SNF factors.⁷³ These observations suggest an interconnectivity, rather than a simple competition, relationship between these master regulators. As discussed below, our studies of an ROR agonist, nobiletin (NOB), provide further evidence that RORs, and likely REV-ERBs, regulate circadian gene expression levels and amplitude in a context-dependent manner, potentially dictated by an inherent requirement to maintain circadian and physiological resilience.

Consistent with the broad gene regulatory roles of REV-ERBs and RORs, mouse genetic mutants exhibit various circadian and physiological phenotypes. *Rev-erba* (*Nr1d1*)-deficient mice showed disrupted circadian rhythms including a shorter period length (0.5 h) and exaggerated light-induced phase shifts compared to wild-type (WT) mice.^{55,74} *Rev-erbb* (*Nr1d2*) knockout (KO) mice also displayed a strong diurnal change of gene expression including inhibition of *Bmal1* transcription.⁵¹ While individual KO mice retained circadian rhythmicity, *Nr1d1/2* double knockout led to severe disruption of overt rhythms, consistent with functional redundancy between these two subtypes.^{70,75,76} *Rora*- and *Rorb*-deficient mice were reported to display altered circadian behavior such as circadian locomotor activity and shortened period length, while no significant alteration in wheel activity is apparent in *Rorc*-deficient mice.^{52,70,71,77,78} These results indicated that REV-ERBs and RORs are required for the maintenance of normal circadian behavior and period length.

With respect to tissue physiology, REV-ERBs and RORs show overlapping but distinct expression patterns and their deficiency led to a wide range of other physiological deficits. REV-ERB α and REV-ERB β are expressed in skeletal muscle, adipose tissue, liver, and brain with tissue-specific patterns. Whereas REV-ERB α is broadly expressed in a rhythmic manner in many tissue types with robust amplitude, REV-ERB β is highly expressed in fewer tissues including certain brain regions (pineal and prefrontal cortex), thyroid, uterus, and pituitary. Deficiency of both *Rev-erbs* causes liver steatosis, in contrast to relatively minor changes upon loss of each subtype alone.^{75,79}

Like REV-ERBs, the three members of the ROR subfamily, ROR α , ROR β , and ROR γ , display significant sequence similarities. ROR α is expressed broadly, notably in skeletal muscle, liver, kidney, lungs, adipose tissue, skin, and brain.⁶¹ *Rora* KO (*Rora*^{-/-}) and *staggerer* mutant (*Rora*^{sg/sg}) mice displayed debilitating cerebellar ataxia and are mostly infertile.^{52,71,80,81} ROR α -deficient mice also showed a multitude of other defects including thin long bones⁸² and abnormal retinal development,^{83,84} the latter corresponding to high expression levels of ROR α in the ganglion cell layer, the inner nuclear layer, and cone photoreceptors in the outer layer. ROR β expression is more limited, mainly in the nerve system. *Rorb*^{-/-} mice exhibited reproductive abnormality and serious degeneration of postnatal retina.⁵⁹ ROR γ is expressed in several peripheral tissues including skeletal muscle, liver, kidney, adipose tissue, and particularly thymus.⁶¹ In accordance, ROR γ KO led to reduced levels of thymocytes and abnormal lymphoid organ development.⁸⁵ This and many other studies have since established a key role of ROR γ as the master transcription factor for Th17 cell

development, although circadian clock involvement in this function is not fully understood. Several studies have also examined double disruption of RORs, providing evidence for their overlapping functions. As mentioned above, in the liver where both ROR α and ROR γ are expressed, double *Rora/c* KO led to strong disruption of lipogenesis *via* a direct regulation of the *Insig2* gene.⁶⁵

Therapeutic relevance of REV-ERBs and RORs

REV-ERB α and RORs have been implicated in various diseases including metabolic diseases, immune diseases, and cancers.^{53,61,86,87} REV-ERB α and RORs show altered expressions and disrupted rhythms during disease development.^{88–90} Furthermore, alteration of REV-ERB α and RORs affects the organism susceptibility to diseases in both humans and mice and is involved in many pathways associated with pathological processes and diseases.^{61,87,90,91–93}

Metabolic disorders

Myriad studies have illustrated a regulatory role of REV-ERBs and RORs in energy metabolism. REV-ERB α was found to regulate *de novo* glucose synthesis.^{94–96} In accordance, REV-ERB α -deficient mice showed a higher plasma glucose level,^{75,97} whereas activation of REV-ERB α diminished plasma glucose levels, improving disease phenotypes.^{94–97} RORs are also involved in glucose metabolism. Single nucleotide polymorphism in *RORA* (rs7164773) has been shown to correlate with increased risk of type 2 diabetes in the Mexico Mestizo population, providing human genetic evidence for a role of ROR α in insulin sensitivity.⁹⁸ In addition, it was reported that ROR α was required for the secretion of FGF21, a hormone associated with glucose tolerance and hepatic lipid metabolism.^{99–101} In another study, ROR γ was found to regulate transcription of various genes involved in glucose metabolism including glucose-6-phosphatase (*G6p*), phosphoenolpyruvate carboxykinase 1 (*Pck*), and glucose transporter 2 (*Glut2*), and *Rorc*-deficient mice in fact displayed a significantly higher insulin sensitivity and glucose tolerance than WT mice, particularly at ZT (zeitgeber time, with ZT0 and ZT12 corresponding to light on and off respectively) 4–6.¹⁰² In pharmacological studies, SR1078, an agonist of ROR α and ROR γ , was able to improve insulin sensitivity, blood glucose, and triglyceride levels in diabetic rodents.¹⁰³ In comparison, mice treated with SR3335, an ROR α inverse agonist, showed dramatically decreased glucose levels in the plasma compared with the control mice, by inhibiting *Pck* expression and gluconeogenesis.¹⁰⁴

With respect to lipid metabolism, *Rev-erba*^{-/-} mice showed increased very low-density lipoprotein (VLDL) and triglyceride levels, consistent with the observed upregulation of apolipoprotein C-III (*ApoC3*), a critical regulator in triglyceride metabolism.^{105,106} Depletion of both *Rev-erbs* in the liver synergistically de-repressed several metabolic genes as well as genes that control the positive limb of the molecular clock.¹⁰⁷ Consistent with these genetic results, administration of the REV-ERB agonist SR9009 decreased cholesterol levels in the plasma in both wild-type and low-density lipoprotein receptor (*Ldlr*) null mice through downregulating cholesterol biosynthesis gene expression.¹⁰⁸ Extensive mouse studies also point to a key role of RORs in lipid metabolism. In *Rora*^{sg/sg} *staggerer* mice, expression levels of hepatic sterol regulatory element-binding protein 1, isoform c (*Srebp-1c*), and fatty acid synthase (*Fas*) were decreased, whereas expression of PGC-1 α and β , coactivators involved in oxidative metabolism and gluconeogenesis, were elevated.^{63,109} At the molecular level, gene expression and cistrome analysis showed that ROR α and/or ROR γ broadly regulate genes involved in lipid metabolism in both liver and muscle tissues.^{65,102,110,111} Furthermore, structural and biochemical studies identified lipid moieties, mainly cholesterol metabolic intermediates, as possible endogenous ligands for ROR α/γ , consistent with the notion that RORs may function as a lipid sensor in the regulation of lipid metabolism.^{63,109,110,112–115}

Immune diseases

Mounting evidence indicates circadian rhythms in immunity and inflammation. For example, rheumatoid arthritis patients exhibit diurnal variations in functional disability such as joint pain and stiffness in morning time.^{116,117} REV-ERB α deletion abolished the diurnal rhythms of various inflammatory factors and aggravates inflammation in diseases including autoimmune encephalomyelitis,^{118,119} fulminant hepatitis,⁹¹ neuroinflammation,^{120,121} heart failure,^{122,123} myocardial infarction,¹²⁴ and ulcerative colitis.^{118,125} At the molecular level, REV-ERB α regulates rhythmic transcription of inflammation-related genes involved in macrophage polarization, immune cell differentiation, and NF- κ B signaling.^{90,125} For example, REV-ERB α was found to obstruct NF- κ B signaling in human endometrial stroma cells and macrophages/microglia cells in mouse models, suppressing expression of inflammatory genes such as *IL-1 β* , *IL-6*, *IL-18*, tumor necrosis factor alpha (*Tnf α*), NACHT, LRR and PYD domains-containing protein 3 (*Nlrp3*), and C-C motif chemokine 2 (*Ccl2*).^{90,120,121,126} Activation of REV-ERB α by SR6472 inhibits NF- κ B signaling and NLRP3 inflammatory activity to prevent cytokines and chemokines productions, consistent with an anti-inflammatory role of REV-ERB α .^{2,90,121}

RORs also play important roles in immunity.⁸⁷ Extensive research has established ROR γ t, a subtype of ROR γ , as a master regulator of Th17 cell differentiation and therefore highly involved in autoimmune diseases.¹²⁷ In Th17 cells, ROR γ t is expressed at dramatically higher levels during daytime than at nighttime.¹²⁸ This diurnal expression pattern in turn

up-regulates BMAL1-dependent *Rev-erb* expression during daytime and conversely represses *NFIL3* transcription. Given the central role of *ROR γ t* in Th17 cells, several compounds targeting *ROR γ t* have been tested in autoimmune disease models. For example, SR1001, an *ROR α* and *ROR γ* inverse agonist, inhibited Th17 cell differentiation under autoimmune disease conditions.¹²⁹ Moreover, this effect is associated with decreased expression of several cytokines such as IL17A, IL17F, IL21, and IL22 by specially targeting TH17.^{130,131} Likewise, SR2211, an *ROR γ* inverse agonist, suppressed Th17 cell differentiation and reduced IL17a and IL23R expression levels as well as intracellular IL17 protein level.¹³²

Brain diseases

Circadian disruption can adversely impact brain development and function, potentially leading to various mood and neurological disorders.^{33,38} Previously, *Rev-erbb* knockout mice were found to exhibit enhanced anxiety, and treatment of an REV-ERB agonist showed anxiolytic effects.¹³³ On the other hand, acute administration of SR8278, a REV-ERB antagonist, improves anxiety symptom and maniac-like behavior.^{134,135} Furthermore, REV-ERB α was shown to diminish fatty acid-binding protein 7 (*Fabp7*) expression, thereby impairing neuronal differentiation and depleting neuronal progenitor cells.^{136,137} Relatedly, deficiency of REV-ERB α adversely affected hippocampal neurogenesis, which contributes to altered mood behaviors.¹³⁸

ROR α is highly expressed in several brain regions such as cerebellar Purkinje cells (PC) and thalamus, and functions to regulate brain development.^{61,139,140} The classical *ROR α* -deficient *staggerer* mice have been shown to present severe ataxia because of cerebellar neurodegeneration^{80,81,141} and abnormal PC development.^{80,141,142} Likewise, *Rora* KO mice exhibit reduced numbers and sizes of PC in the cerebellar region reminiscent of clinical observations from patients with autism-spectrum disorder (ASD).^{143,144} *ROR α* also showed neuroprotective effects in astrocytes and neurons during hypoxia.^{112,145} *ROR β* is highly expressed in the retina, pineal gland, and suprachiasmatic nucleus, and has been implicated in visual function, motor ability, and circadian rhythms.^{61,78,146} For example, *ROR β* -deficient mice showed abnormal motor and olfactory functions, anxiety control, and alterations in circadian behavior.⁷⁷ The noteworthy question regarding a potential functional overlap in the neuronal system between *ROR α* and *ROR β* remains to be investigated.

Muscle pathologies

REV-ERBs (α and β) and RORs (α and γ) are highly expressed in the skeletal muscle where they modulate myofiber types and energy metabolism^{61,147} and may be targeted against myopathies.¹⁴⁸ In an early study, REV-ERB α -deficient mice showed a marked increase in the relative amount of the slow (type I) myosin heavy chain (MyHC) isoform compared to WT controls.¹⁴⁷ Extensive research since has further revealed the regulatory roles of REV-ERBs in skeletal muscle function.^{86,149,150} For example, REV-ERB β has been implicated in skeletal muscle lipid metabolism since ectopic expression of its dominant-negative form upregulated expression of genes associated with fatty acid uptake in skeletal muscle.¹⁵¹ Consistently, SR8278, an antagonist of REV-ERBs, was found to activate expression of myogenesis genes including Myogenic determination 1 (*Myod*), Myogenin (*Myog*), and Major histocompatibility complex 3 (*Mhc3*), suggesting a role of REV-ERBs in myogenesis.¹⁵²

Loss-of-function studies also suggest an important role of *ROR α* in skeletal muscle metabolism.¹⁵³ For example, ectopic expression of a dominant-negative *ROR α* in C2C12 cells or mouse skeletal muscle broadly alters the expression of genes associated with lipid metabolism, lipogenesis, and energy expenditure, including carnitine palmitoyltransferase-1 (Cpt1), caveolin 3 (Cav3), and Srebp1c and its downstream targets.^{110,154}

Cancer

REV-ERB α has been implicated in the progression and development of various cancers.¹⁵⁵ Activation of REV-ERB α by SR9009 and SR9011 was found to confer cancer cell-selective cytotoxicity as well as *in vivo* efficacy against glioma, and autophagy and lipogenesis were identified as cellular hallmarks closely associated with this anti-cancer activity.¹⁵⁶ In a recent study investigating lung adenocarcinoma-associated cachexia,¹⁵⁷ REV-ERB α functions as a key effector whose exaggerated turnover contributes to gluconeogenesis gene induction and glucose production in mice.

A number of studies have shown that *ROR α* expression is significantly decreased during tumor development and progression, and exogenous *ROR α* expression repressed cell proliferation and tumor growth.^{158–163} For example, down-regulated *ROR α* expression has been observed in colorectal cancer and mammary cancer, and is associated with poor prognosis in patients with hepatocellular and breast carcinoma.^{158,159,161,162,164,165} Conversely, restoring *ROR α* expression suppressed cell migration and tumor growth of breast cancer cells as well as metastasis in nude mice, which was accompanied by up-regulated expression of semaphorin 3F (SEMA3F), a tumor suppressor that reduces tumor growth and invasion.¹⁶¹ In colon cancer HCT116 cells treated with DNA-damage agents, a p53-*ROR α* crosstalk was required for

apoptosis, where *Rora* gene transcription was dependent on p53 and ROR α in turn rendered greater p53 protein stability.¹⁶⁶ In ROR γ deficient mice, there was an aggravated development of T-cell lymphomas within the first months after birth, which rapidly metastasized to other organs including liver and spleen.¹⁶⁷

Nobiletin (NOB): A natural ROR agonist

NOB is a natural bioactive polymethylated flavonoid.^{168,169} Many studies have provided functional evidence both *in vitro* and *in vivo* for its biological efficacy in diverse disease models,^{168,170,171} including metabolic diseases and inflammation. In our previous unbiased chemical screen, we identified NOB, along with its close analog tangeretin, as a clock-enhancing small molecule in cell-based circadian reporter assays.¹⁷² Focusing on NOB, we demonstrated a circadian clock-dependent efficacy to blunt obesity and metabolic dysfunction in mouse models, and importantly identified ROR α and ROR γ as its direct targets *via* radioactive ligand binding assays. NOB shows robust binding to the LBDs of ROR α and ROR γ , with somewhat higher affinity for ROR γ . Currently there is no functional evidence to suggest subtype selectivity analogous to CRY-selective compounds.¹⁷³ Subsequent published studies, from our group and others, have provided further evidence that NOB plays a beneficial role in strengthening circadian physiologies in various mouse models, including aging, metabolic disorders, cardiovascular disease, and Alzheimer's disease (AD).^{171,174–182} In further support of NOB as an anti-inflammatory agent, recent studies demonstrated a potent role of NOB against neuroinflammation and astrogliosis, accompanied by mitigation of A β plaque deposition, in an amyloid AD mouse model.¹⁸³ Given the increasing appreciation of circadian rhythms in aging, recent studies have also tested its effect in aging models. In naturally aged mice fed with either regular or high-fat diets (HFD), NOB was found to promote healthy aging at several levels, including metabolic homeostasis, inflammatory markers, tissue functions, and systemic behaviors.¹⁷⁵ An important target organ is skeletal muscle, where circadian gene reprogramming and metabolomic alteration support an improved mitochondrial function, accompanied by respiratory supercomplex formation. Notably, while NOB-mediated improvement in general healthy aging parameters seems more pronounced in metabolically challenged aged mice (HFD fed) than in those fed with regular diet, the latter group showed an extension of median lifespan, but not maximum lifespan.¹⁷⁵ In comparison, NOB was found to exhibit longevity effects in *C. elegans*, extending median lifespan by up to 21%.¹⁸⁴ Overall, these and many other studies underscore a strong health-promoting effect of NOB, at least in part via circadian mechanisms.

Mechanistic studies have begun to shed light on the circadian modulatory action of NOB. In addition to its clock amplitude-enhancing effects, NOB also alters the other two cardinal circadian parameters, period, and phase, at least *in vitro*.^{172,182} Following chronic treatment *in vivo* (10–12 weeks), NOB was able to strengthen oscillatory amplitude, as well as peak expression, of core clock components at both transcript and protein levels in HFD-fed mice, and wheel-running activity was also increased at nighttime.¹⁷² Given the extensive crosstalk between clocks and metabolism/physiology, these overt enhancements of circadian rhythms may result from both direct and indirect effects of NOB on the core oscillator/RORs and clock-regulated downstream functions, respectively. Acute *in vivo* effects on circadian rhythms remain to be investigated. Another important issue is related to the varying effects of NOB according to the clock functional state. There seems to be a general inverse correlation between NOB efficacy and clock health. For example, in young and healthy mice under normal husbandry conditions, NOB showed essentially no effects on circadian and metabolic functions, contrary to the profound improvements in obese or diabetic mouse models.^{172,178,179} Likewise, as mentioned above, aged mice further challenged with HFD known to dampen circadian rhythms showed a greater responsiveness to NOB in healthy aging compared with aged mice fed with normal diets.¹⁷⁵ A similar pattern was observed between WT and AD mice at old ages (>22 months) where NOB was found to mitigate neuroinflammation more markedly in the latter, correlating with a more severe circadian disruption in AD mice.¹⁸³ These *in vivo* results together suggest a role of NOB to enhance circadian resilience toward restoring normal circadian rhythms that may have evolved to operate within a physiological range. Either dampening or indiscriminately enhancing the normal circadian rhythms is likely detrimental to organismal health.

Further research should investigate the downstream cellular mechanisms intersecting with the clock machinery. In a recent study, an inhibitory function of NOB against triple-negative breast cancer (TNBC) was found after cell line screening.¹⁸⁵ Both *in vitro* and in xenografts, NOB was able to blunt TNBC cell growth, either alone or in combination with chemotherapeutic agents. The cellular mechanism entailed, at least in part, suppression of NF- κ B signaling, *via* a pathway where activation of RORs by NOB increased expression of its downstream target gene encoding I κ B α , and ChIP analysis showed that ROR recruitment to the I κ B α gene promoter was potentiated.¹⁸⁵ While this study illustrates a cellular pathway targeted by NOB in TNBC, it should be noted that the TNBC cells examined do not have a functional clock despite detectable clock gene expression, and NOB was not able to restore the core oscillator in these cells.^{185,186} Therefore, this is a scenario that NOB effects are mediated by ROR transcriptional regulation independent of oscillator function. However, since the host mice have circadian rhythms, whether NOB modulates host rhythms as part of the

effect against TNBC remains to be investigated. Finally, as a natural compound with an excellent safety profile, NOB is ideally suited for future trials in clinically relevant settings against clock-related disorders.

Concluding remarks

Accumulating evidence from molecular, genetic and interventional studies highlight a critical role of the circadian secondary/stabilization loop, specifically the REV-ERB α/β and ROR $\alpha/\beta/\gamma$ nuclear receptors, in linking the core oscillator with physiology and behavior under both normal and pathological conditions. These are multi-functional transcription factors, playing important regulatory roles in circadian regulation as well as other processes not primarily tied to the clock (e.g., ROR γ t in Th17 differentiation and autoimmunity). It is therefore a challenge to dissect the underlying mechanisms and devise disease-specific interventions from the circadian perspective.¹⁴⁸ Whenever possible, detailed circadian characterization should be performed, especially at the tissue and organismal levels. As illustrated by pharmacological studies targeting these factors,^{53,187} including those on NOB, the concept of circadian resilience, or restoration of homeostatic clock function, should be an important consideration regarding intervention. Finally, given the tissue-specific nature of circadian regulation and the growing evidence for inter-organ communication with the clock system,¹⁸⁸ the functional effects, mechanistic pathways and interventional approaches should be interrogated accordingly in an integrative manner. In that regard, distribution and functional redundancy among the subtypes of these receptors should be considered. Despite the inherent complexity and practical challenges, targeting the circadian machinery, including the secondary loop, represents an exciting frontier in the 4th dimension for research and medicine.

Author contributions

Conceptualization: Z.C.; Original draft preparation: E.K. and Z.C.; Review and Editing: S.-H.Y. and Z.C.

Data availability

Underlying data

No data are associated with this article.

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References

- Bass J, Takahashi JS: **Circadian integration of metabolism and energetics.** *Science.* 2010; **330**: 1349–1354.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Güldür T, Otlı HG: **Circadian rhythm in mammals: time to eat & time to sleep.** *Biol. Rhythm. Res.* 2017; **48**: 243–261.
[Publisher Full Text](#)
- Panda S, et al.: **Coordinated transcription of key pathways in the mouse by the circadian clock.** *Cell.* 2002; **109**: 307–320.
[Publisher Full Text](#)
- Burris TP: **Nuclear hormone receptors for heme: REV-ERB α and REV-ERB β are ligand-regulated components of the mammalian clock.** *Mol. Endocrinol.* 2008; **22**: 1509–1520.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Bass J, Lazar MA: **Circadian time signatures of fitness and disease.** *Science.* 2016; **354**: 994–999.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Cederroth CR, et al.: **Medicine in the Fourth Dimension.** *Cell Metab.* 2019; **30**: 238–250.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Mohawk JA, Green CB, Takahashi JS: **Central and peripheral circadian clocks in mammals.** *Annu. Rev. Neurosci.* 2012; **35**: 445–462.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Liu AC, et al.: **Intercellular coupling confers robustness against mutations in the SCN circadian clock network.** *Cell.* 2007; **129**: 605–616.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Finger AM, et al.: **Intercellular coupling between peripheral circadian oscillators by TGF-beta signaling.** *Sci. Adv.* 2021; **7**:
[PubMed Abstract](#) | [Publisher Full Text](#)
- Takahashi JS: **Transcriptional architecture of the mammalian circadian clock.** *Nat. Rev. Genet.* 2017; **18**: 164–179.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Mure LS, et al.: **Diurnal transcriptome atlas of a primate across major neural and peripheral tissues.** *Science.* 2018; **359**.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Zhang R, Lahens NF, Ballance HI, et al.: **A circadian gene expression atlas in mammals: implications for biology and medicine.** *Proc. Natl. Acad. Sci. U. S. A.* 2014; **111**: 16219–16224.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Rutter J, Reick M, McKnight SL: **Metabolism and the control of circadian rhythms.** *Annu. Rev. Biochem.* 2002; **71**: 307–331.
[Publisher Full Text](#)
- Green CB, Takahashi JS, Bass J: **The meter of metabolism.** *Cell.* 2008; **134**: 728–742.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Rosbash M: **The implications of multiple circadian clock origins.** *LoS Biol.* 2009; **7**: e62.
- Ando H, et al.: **Rhythmic messenger ribonucleic acid expression of clock genes and adipocytokines in mouse visceral adipose tissue.** *Endocrinology.* 2005; **146**: 5631–5636.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Bass J: **Circadian topology of metabolism.** *Nature.* 2012; **491**: 348–356.
[Publisher Full Text](#)
- Harfmann BD, et al.: **Muscle-specific loss of Bmal1 leads to disrupted tissue glucose metabolism and systemic glucose homeostasis.** *Skelet. Muscle.* 2016; **6**: 12.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Stenvers DJ, Scheer FA, Schrauwen P, et al.: **Circadian clocks and insulin resistance.** *Nat. Rev. Endocrinol.* 2019; **15**: 75–89.
[Publisher Full Text](#)

20. Kettner NM, *et al.*: **Circadian Dysfunction Induces Leptin Resistance in Mice.** *Cell Metab.* 2015; **22**: 448–459.
[PubMed Abstract](#) | [Publisher Full Text](#)
21. Scheer FA, Hilton MF, Mantzoros CS, *et al.*: **Adverse metabolic and cardiovascular consequences of circadian misalignment.** *Proc. Natl. Acad. Sci. U. S. A.* 2009; **106**: 4453–4458.
[PubMed Abstract](#) | [Publisher Full Text](#)
22. Buxton OM, *et al.*: **Adverse metabolic consequences in humans of prolonged sleep restriction combined with circadian disruption.** *Sci. Transl. Med.* 2012; **4**: 129ra143.
23. McHill AW, *et al.*: **Impact of circadian misalignment on energy metabolism during simulated nightshift work.** *Proc. Natl. Acad. Sci. U. S. A.* 2014; **111**: 17302–17307.
[PubMed Abstract](#) | [Publisher Full Text](#)
24. Lucassen EA, *et al.*: **Environmental 24-hr cycles are essential for health.** *Curr. Biol.* 2016; **26**: 1843–1853.
[PubMed Abstract](#) | [Publisher Full Text](#)
25. Inokawa H, *et al.*: **Chronic circadian misalignment accelerates immune senescence and abbreviates lifespan in mice.** *Sci. Rep.* 2020; **10**: 2569.
[PubMed Abstract](#) | [Publisher Full Text](#)
26. Kohsaka A, *et al.*: **High-fat diet disrupts behavioral and molecular circadian rhythms in mice.** *Cell Metab.* 2007; **6**: 414–421.
[PubMed Abstract](#) | [Publisher Full Text](#)
27. Arble DM, Bass J, Laposky AD, *et al.*: **Circadian timing of food intake contributes to weight gain.** *Obesity (Silver Spring).* 2009; **17**: 2100–2102.
[PubMed Abstract](#) | [Publisher Full Text](#)
28. Young ME, Razezghi P, Cedars AM, *et al.*: **Intrinsic diurnal variations in cardiac metabolism and contractile function.** *Circ. Res.* 2001; **89**: 1199–1208.
[PubMed Abstract](#) | [Publisher Full Text](#)
29. Rudic RD, Curtis AM, Cheng Y, *et al.*: **Peripheral clocks and the regulation of cardiovascular and metabolic function.** *Methods Enzymol.* 2005; **393**: 524–539.
[Publisher Full Text](#)
30. Gangwisch JE, *et al.*: **Short sleep duration as a risk factor for hypertension: analyses of the first National Health and Nutrition Examination Survey.** *Hypertension.* 2006; **47**: 833–839.
[Publisher Full Text](#)
31. Gottlieb DJ, *et al.*: **Association of sleep time with diabetes mellitus and impaired glucose tolerance.** *Arch. Intern. Med.* 2005; **165**: 863–867.
[PubMed Abstract](#) | [Publisher Full Text](#)
32. Slat E, Freeman GM Jr, Herzog ED: **The clock in the brain: neurons, glia, and networks in daily rhythms.** *Handb. Exp. Pharmacol.* 2013;
[PubMed Abstract](#) | [Publisher Full Text](#)
33. Logan RW, McClung CA: **Rhythms of life: circadian disruption and brain disorders across the lifespan.** *Nat. Rev. Neurosci.* 2019; **20**: 49–65.
[PubMed Abstract](#) | [Publisher Full Text](#)
34. Burish MJ, Chen Z, Yoo SH: **Emerging relevance of circadian rhythms in headaches and neuropathic pain.** *Acta Physiol (Oxf).* 2019; **225**: e13161.
[PubMed Abstract](#) | [Publisher Full Text](#)
35. Jones CR, Huang AL, Ptacek LJ, *et al.*: **Genetic basis of human circadian rhythm disorders.** *Exp. Neurol.* 2013; **243**: 28–33.
[PubMed Abstract](#) | [Publisher Full Text](#)
36. Kondratova AA, Kondratov RV: **The circadian clock and pathology of the ageing brain.** *Nat. Rev. Neurosci.* 2012; **13**: 325–335.
[PubMed Abstract](#) | [Publisher Full Text](#)
37. Musiek ES, Holtzman DM: **Mechanisms linking circadian clocks, sleep, and neurodegeneration.** *Science.* 2016; **354**: 1004–1008.
[PubMed Abstract](#) | [Publisher Full Text](#)
38. Colwell CS: **Defining circadian disruption in neurodegenerative disorders.** *J. Clin. Invest.* 2021; **131**.
[PubMed Abstract](#) | [Publisher Full Text](#)
39. Kress GJ, *et al.*: **Regulation of amyloid-beta dynamics and pathology by the circadian clock.** *J. Exp. Med.* 2018; **215**: 1059–1068.
[PubMed Abstract](#) | [Publisher Full Text](#)
40. Homolák J, Mudrovčič M, Vukić B, *et al.*: **Circadian rhythm and Alzheimer's disease.** *Med. Sci.* 2018; **6**: 52.
41. Kang J-E, *et al.*: **Amyloid- β dynamics are regulated by orexin and the sleep-wake cycle.** *Science.* 2009; **326**: 1005–1007.
[PubMed Abstract](#) | [Publisher Full Text](#)
42. Volicer L, Harper DG, Manning BC, *et al.*: **Sundowning and circadian rhythms in Alzheimer's disease.** *Am. J. Psychiatr.* 2001; **158**: 704–711.
[Publisher Full Text](#)
43. Acosta-Rodriguez VA, Rijo-Ferreira F, Green CB, *et al.*: **Importance of circadian timing for aging and longevity.** *Nat. Commun.* 2021; **12**: 2862.
[PubMed Abstract](#) | [Publisher Full Text](#)
44. Gibson EM, Williams WP 3rd, Kriegsfeld LJ: **Aging in the circadian system: considerations for health, disease prevention and longevity.** *Exp. Gerontol.* 2009; **44**: 51–56.
[PubMed Abstract](#) | [Publisher Full Text](#)
45. Sato S, *et al.*: **Circadian Reprogramming in the Liver Identifies Metabolic Pathways of Aging.** *Cell.* 2017; **170**: 664–677.e11.
[PubMed Abstract](#) | [Publisher Full Text](#)
46. Lu JY, *et al.*: **Comparative transcriptomics reveals circadian and pluripotency networks as two pillars of longevity regulation.** *Cell Metab.* 2022; **34**: 836–856.e5.
[PubMed Abstract](#) | [Publisher Full Text](#)
47. Schaum N, *et al.*: **Ageing hallmarks exhibit organ-specific temporal signatures.** *Nature.* 2020; **583**: 596–602.
[PubMed Abstract](#) | [Publisher Full Text](#)
48. Acosta-Rodriguez V, *et al.*: **Circadian alignment of early onset caloric restriction promotes longevity in male C57BL/6j mice.** *Science.* 2022; **376**: 1192–1202.
[PubMed Abstract](#) | [Publisher Full Text](#)
49. Ulgherait M, *et al.*: **Circadian autophagy drives iTRF-mediated longevity.** *Nature.* 2021; **598**: 353–358.
[PubMed Abstract](#) | [Publisher Full Text](#)
50. Lopez-Otin C, Kroemer G: **Hallmarks of health.** *Cell.* 2021; **184**: 1929–1939.
[Publisher Full Text](#)
51. Guillaumond F, Dardente H, Giguère V, *et al.*: **Differential control of Bmal1 circadian transcription by REV-ERB and ROR nuclear receptors.** *J. Biol. Rhythm.* 2005; **20**: 391–403.
[PubMed Abstract](#) | [Publisher Full Text](#)
52. Sato TK, *et al.*: **A functional genomics strategy reveals Rora as a component of the mammalian circadian clock.** *Neuron.* 2004; **43**: 527–537.
[PubMed Abstract](#) | [Publisher Full Text](#)
53. Kojetin DJ, Burris TP: **REV-ERB and ROR nuclear receptors as drug targets.** *Nat. Rev. Drug Discov.* 2014; **13**: 197–216.
[PubMed Abstract](#) | [Publisher Full Text](#)
54. Harding HP, Lazar MA: **The orphan receptor Rev-ErbA alpha activates transcription via a novel response element.** *Mol. Cell. Biol.* 1993; **13**: 3113–3121.
[PubMed Abstract](#)
55. Preitner N, *et al.*: **The orphan nuclear receptor REV-ERB α controls circadian transcription within the positive limb of the mammalian circadian oscillator.** *Cell.* 2002; **110**: 251–260.
[PubMed Abstract](#) | [Publisher Full Text](#)
56. Giguère V, *et al.*: **Isoform-specific amino-terminal domains dictate DNA-binding properties of ROR alpha, a novel family of orphan hormone nuclear receptors.** *Genes Dev.* 1994; **8**: 538–553.
[PubMed Abstract](#) | [Publisher Full Text](#)
57. Carlberg C, Hooft van Huijsduijnen R, Staple JK, *et al.*: **RZR α , a new family of retinoid-related orphan receptors that function as both monomers and homodimers.** *Mol. Endocrinol.* 1994; **8**: 757–770.
[PubMed Abstract](#)
58. Hirose T, Smith RJ, Jetten AM: **ROR- γ : the third member of ROR/RZR orphan receptor subfamily that is highly expressed in skeletal muscle.** *Biochem. Biophys. Res. Commun.* 1994; **205**: 1976–1983.
[PubMed Abstract](#) | [Publisher Full Text](#)
59. André E, *et al.*: **Disruption of retinoid-related orphan receptor β changes circadian behavior, causes retinal degeneration and leads to vacillans phenotype in mice.** *EMBO J.* 1998; **17**: 3867–3877.
[PubMed Abstract](#) | [Publisher Full Text](#)
60. Ueda HR, *et al.*: **System-level identification of transcriptional circuits underlying mammalian circadian clocks.** *Nat. Genet.* 2005; **37**: 187–192.
[PubMed Abstract](#) | [Publisher Full Text](#)
61. Jetten AM: **Retinoid-related orphan receptors (RORs): critical roles in development, immunity, circadian rhythm, and cellular metabolism.** *Nucl. Recept. Signal.* 2009; **7**: e003.
62. Kang HS, *et al.*: **Transcriptional profiling reveals a role for RORalpha in regulating gene expression in obesity-associated inflammation and hepatic steatosis.** *Physiol. Genomics.* 2011; **43**: 818–828.
[PubMed Abstract](#) | [Publisher Full Text](#)
63. Lau P, *et al.*: **The orphan nuclear receptor, RORalpha, regulates gene expression that controls lipid metabolism: staggerer (SG/SG) mice are resistant to diet-induced obesity.** *J. Biol. Chem.* 2008; **283**: 18411–18421.
[PubMed Abstract](#) | [Publisher Full Text](#)

64. Zhang Y, *et al.*: **GENE REGULATION. Discrete functions of nuclear receptor Rev-erbalpha couple metabolism to the clock.** *Science*. 2015; **348**: 1488–1492.
[PubMed Abstract](#) | [Publisher Full Text](#)
65. Zhang Y, *et al.*: **The hepatic circadian clock fine-tunes the lipogenic response to feeding through RORalpha/gamma.** *Genes Dev*. 2017; **31**: 1202–1211.
[PubMed Abstract](#) | [Publisher Full Text](#)
66. Littleton ES, Childress ML, Gosting ML, *et al.*: **Genome-wide correlation analysis to identify amplitude regulators of circadian transcriptome output.** *Sci. Rep.* 2020 Dec 14; **10**(1): 21839.
[PubMed Abstract](#) | [Publisher Full Text](#)
67. Takeda Y, Jothi R, Birault V, *et al.*: **ROR γ directly regulates the circadian expression of clock genes and downstream targets in vivo.** *Nucleic Acids Res.* 2012 Sep 1; **40**(17): 8519–8535.
[PubMed Abstract](#) | [Publisher Full Text](#)
68. Ikeda R, Tsuchiya Y, Koike N, *et al.*: **REV-ERB α and REV-ERB β function as key factors regulating Mammalian Circadian Output.** *Sci. Rep.* 2019 Jul 15; **9**(1): 10171.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
69. Zhao X, *et al.*: **Circadian amplitude regulation via FBXW7-targeted REV-ERB α degradation.** *Cell*. 2016; **165**: 1644–1657.
[PubMed Abstract](#) | [Publisher Full Text](#)
70. Liu AC, *et al.*: **Redundant function of REV-ERB α and β and non-essential role for Bmal1 cycling in transcriptional regulation of intracellular circadian rhythms.** *PLoS Genet.* 2008; **4**: e1000023.
[PubMed Abstract](#) | [Publisher Full Text](#)
71. Akashi M, Takumi T: **The orphan nuclear receptor ROR α regulates circadian transcription of the mammalian core-clock Bmal1.** *Nat. Struct. Mol. Biol.* 2005; **12**: 441–448.
[PubMed Abstract](#) | [Publisher Full Text](#)
72. Littleton ES, Childress ML, Gosting ML, *et al.*: **Genome-wide correlation analysis to identify amplitude regulators of circadian transcriptome output.** *Sci. Rep.* 2020; **10**: 21839.
[PubMed Abstract](#) | [Publisher Full Text](#)
73. Zhu B, *et al.*: **Coactivator-Dependent Oscillation of Chromatin Accessibility Dictates Circadian Gene Amplitude via REV-ERB Loading.** *Mol. Cell.* 2015; **60**: 769–783.
[PubMed Abstract](#) | [Publisher Full Text](#)
74. Duez H, Staels B: **Rev-erb- α : an integrator of circadian rhythms and metabolism.** *J. Appl. Physiol.* 2009; **107**(107): 1972–1980.
[Publisher Full Text](#)
75. Cho H, *et al.*: **Regulation of circadian behaviour and metabolism by REV-ERB-alpha and REV-ERB-beta.** *Nature*. 2012; **485**: 123–127.
[PubMed Abstract](#) | [Publisher Full Text](#)
76. Ikeda R, *et al.*: **REV-ERBalpha and REV-ERBbeta function as key factors regulating Mammalian Circadian Output.** *Sci. Rep.* 2019; **9**: 10171.
[PubMed Abstract](#) | [Publisher Full Text](#)
77. Masana MI, Sumaya IC, Becker-Andre M, *et al.*: **Behavioral characterization and modulation of circadian rhythms by light and melatonin in C3H/HeN mice homozygous for the ROR β knockout.** *Am. J. Phys. Regul. Integr. Comp. Phys.* 2007; **292**: R2357–R2367.
[PubMed Abstract](#) | [Publisher Full Text](#)
78. Schaeren-Wierners N, André E, Kapfhammer JP, *et al.*: **The ExDression pattern of the orphan nuclear receptor ROR β in the developing and adult rat nervous system suggests a role in the processing of sensory information and in circadian rhythm.** *Eur. J. Neurosci.* 1997; **9**: 2687–2701.
[Publisher Full Text](#)
79. Lazar MA: **Rev-erbs: Integrating Metabolism Around the Clock.** *A Time for Metabolism and Hormones.* Sassone-Corsi P, Christen Y, editors. Cham: 2016; pp. 63–70.
[PubMed Abstract](#) | [Publisher Full Text](#)
80. Hamilton BA, *et al.*: **Disruption of the nuclear hormone receptor ROR α in staggerer mice.** *Nature*. 1996; **379**: 736–739.
[PubMed Abstract](#) | [Publisher Full Text](#)
81. Steinmayr M, *et al.*: **Staggerer phenotype in retinoid-related orphan receptor α -deficient mice.** *Proc. Natl. Acad. Sci.* 1998; **95**: 3960–3965.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
82. Meyer T, Kneissel M, Mariani J, *et al.*: **In vitro and in vivo evidence for orphan nuclear receptor ROR α function in bone metabolism.** *Proc. Natl. Acad. Sci.* 2000; **97**: 9197–9202.
[PubMed Abstract](#) | [Publisher Full Text](#)
83. Fujieda H, Bremner R, Mears AJ, *et al.*: **Retinoic acid receptor-related orphan receptor α regulates a subset of cone genes during mouse retinal development.** *J. Neurochem.* 2009; **108**: 91–101.
[PubMed Abstract](#) | [Publisher Full Text](#)
84. Ino H: **Immunohistochemical characterization of the orphan nuclear receptor ROR α in the mouse nervous system.** *J. Histochem. Cytochem.* 2004; **52**: 311–323.
[PubMed Abstract](#) | [Publisher Full Text](#)
85. Sun Z, *et al.*: **Requirement for ROR γ in thymocyte survival and lymphoid organ development.** *Science*. 2000; **288**: 2369–2373.
[PubMed Abstract](#) | [Publisher Full Text](#)
86. Everett LJ, Lazar MA: **Nuclear receptor Rev-erb α : up, down, and all around.** *Trends Endocrinol. Metab.* 2014; **25**: 586–592.
[PubMed Abstract](#) | [Publisher Full Text](#)
87. Cook DN, Kang HS, Jetten AM: **Retinoic Acid-Related Orphan Receptors (RORs): Regulatory Functions in Immunity, Development, Circadian Rhythm, and Metabolism.** *Nucl Receptor Res.* 2015; **2**.
[PubMed Abstract](#) | [Publisher Full Text](#)
88. Duez H, Staels B: **The nuclear receptors Rev-erbs and RORs integrate circadian rhythms and metabolism.** *Diab. Vasc. Dis. Res.* 2008; **5**: 82–88.
[PubMed Abstract](#) | [Publisher Full Text](#)
89. Pivovarovova O, *et al.*: **Regulation of the clock gene expression in human adipose tissue by weight loss.** *Int. J. Obes.* 2016; **40**: 899–906.
[PubMed Abstract](#) | [Publisher Full Text](#)
90. Wang S, *et al.*: **REV-ERB α integrates colon clock with experimental colitis through regulation of NF- κ B/NLRP3 axis.** *Nat. Commun.* 2018; **9**: 1–12.
91. Pourcet B, *et al.*: **Nuclear receptor subfamily 1 group D member 1 regulates circadian activity of NLRP3 inflammasome to reduce the severity of fulminant hepatitis in mice.** *Gastroenterology*. 2018; **154**: 1449–1464. e1420.
[PubMed Abstract](#) | [Publisher Full Text](#)
92. Wang S, Li F, Lin Y, *et al.*: **Targeting REV-ERB α for therapeutic purposes: promises and challenges.** *Theranostics*. 2020; **10**: 4168–4182.
[PubMed Abstract](#) | [Publisher Full Text](#)
93. Goumidi L, *et al.*: **Impact of REV-ERB alpha gene polymorphisms on obesity phenotypes in adult and adolescent samples.** *Int. J. Obes.* 2013; **37**: 666–672.
[PubMed Abstract](#) | [Publisher Full Text](#)
94. Solt LA, *et al.*: **Regulation of circadian behaviour and metabolism by synthetic REV-ERB agonists.** *Nature*. 2012; **485**: 62–68.
[PubMed Abstract](#) | [Publisher Full Text](#)
95. Yin L, *et al.*: **Rev-erb α , a heme sensor that coordinates metabolic and circadian pathways.** *Science*. 2007; **318**: 1786–1789.
[PubMed Abstract](#) | [Publisher Full Text](#)
96. Yuan X, Dong D, Li Z, *et al.*: **Rev-erb α activation down-regulates hepatic Pck1 enzyme to lower plasma glucose in mice.** *Pharmacol. Res.* 2019; **141**: 310–318.
[PubMed Abstract](#) | [Publisher Full Text](#)
97. Delezie J, *et al.*: **The nuclear receptor REV-ERB α is required for the daily balance of carbohydrate and lipid metabolism.** *FASEB J.* 2012; **26**: 3321–3335.
[PubMed Abstract](#) | [Publisher Full Text](#)
98. Gamboa-Meléndez MA, *et al.*: **Contribution of common genetic variation to the risk of type 2 diabetes in the Mexican Mestizo population.** *Diabetes*. 2012; **61**: 3314–3321.
[PubMed Abstract](#) | [Publisher Full Text](#)
99. Kharitononkov A, *et al.*: **FGF-21 as a novel metabolic regulator.** *J. Clin. Invest.* 2005; **115**: 1627–1635.
[PubMed Abstract](#) | [Publisher Full Text](#)
100. Reitman ML: **FGF21: a missing link in the biology of fasting.** *Cell Metab.* 2007; **5**: 405–407.
[PubMed Abstract](#) | [Publisher Full Text](#)
101. Inagaki T, *et al.*: **Endocrine regulation of the fasting response by PPAR α -mediated induction of fibroblast growth factor 21.** *Cell Metab.* 2007; **5**: 415–425.
[PubMed Abstract](#) | [Publisher Full Text](#)
102. Takeda Y, *et al.*: **Retinoic acid-related orphan receptor γ (ROR γ): a novel participant in the diurnal regulation of hepatic gluconeogenesis and insulin sensitivity.** *PLoS Genet.* 2014; **10**: e1004331.
[PubMed Abstract](#) | [Publisher Full Text](#)
103. Wang Y, *et al.*: **Identification of SR1078, a synthetic agonist for the orphan nuclear receptors ROR α and ROR γ .** *ACS Chem. Biol.* 2010; **5**: 1029–1034.
[PubMed Abstract](#) | [Publisher Full Text](#)
104. Kumar N, *et al.*: **Identification of SR3335 (ML-176): a synthetic ROR α selective inverse agonist.** *ACS Chem. Biol.* 2011; **6**: 218–222.
[PubMed Abstract](#) | [Publisher Full Text](#)
105. Chomez P, *et al.*: **Increased cell death and delayed development in the cerebellum of mice lacking the rev-erbA (alpha) orphan**

- receptor. *Development*. 2000; **127**: 1489–1498.
[PubMed Abstract](#) | [Publisher Full Text](#)
106. Raspé E, et al.: **Identification of Rev-erb α as a physiological repressor of apoC-III gene transcription**¹. *J. Lipid Res.* 2002; **43**: 2172–2179.
[PubMed Abstract](#) | [Publisher Full Text](#)
107. Bugge A, et al.: **Rev-erb α and Rev-erb β coordinately protect the circadian clock and normal metabolic function.** *Genes Dev.* 2012; **26**: 657–667.
[PubMed Abstract](#) | [Publisher Full Text](#)
108. Sitaula S, Zhang J, Ruiz F, et al.: **Rev-erb regulation of cholesterologenesis.** *Biochem. Pharmacol.* 2017; **131**: 68–77.
[PubMed Abstract](#) | [Publisher Full Text](#)
109. Wada T, et al.: **Identification of oxysterol 7 α -hydroxylase (Cyp7b1) as a novel retinoid-related orphan receptor α (ROR α)(NR1F1) target gene and a functional cross-talk between ROR α and liver X receptor (NR1H3).** *Mol. Pharmacol.* 2008; **73**: 891–899.
[PubMed Abstract](#) | [Publisher Full Text](#)
110. Lau P, Nixon SJ, Parton RG, et al.: **ROR α regulates the expression of genes involved in lipid homeostasis in skeletal muscle cells: caveolin-3 and CPT-1 are direct targets of ROR.** *J. Biol. Chem.* 2004; **279**: 36828–36840.
[PubMed Abstract](#) | [Publisher Full Text](#)
111. Kim K, et al.: **ROR α controls hepatic lipid homeostasis via negative regulation of PPAR γ transcriptional network.** *Nat. Commun.* 2017; **8**: 162.
[PubMed Abstract](#) | [Publisher Full Text](#)
112. Boukhtouche F, et al.: **Human retinoic acid receptor-related orphan receptor α 1 overexpression protects neurones against oxidative stress-induced apoptosis.** *J. Neurochem.* 2006; **96**: 1778–1789.
[PubMed Abstract](#) | [Publisher Full Text](#)
113. Kallen JA, et al.: **X-ray structure of the hROR α LBD at 1.63 Å: structural and functional data that cholesterol or a cholesterol derivative is the natural ligand of ROR α .** *Structure.* 2002; **10**: 1697–1707.
[PubMed Abstract](#) | [Publisher Full Text](#)
114. Lind U, et al.: **Identification of the human ApoAV gene as a novel ROR α target gene.** *Biochem. Biophys. Res. Commun.* 2005; **330**: 233–241.
[PubMed Abstract](#) | [Publisher Full Text](#)
115. Mamontova A, et al.: **Severe atherosclerosis and hypoalphalipoproteinemia in the staggerer mouse, a mutant of the nuclear receptor ROR α .** *Circulation.* 1998; **98**: 2738–2743.
[PubMed Abstract](#) | [Publisher Full Text](#)
116. Straub RH, Cutolo M: **Circadian rhythms in rheumatoid arthritis: implications for pathophysiology and therapeutic management.** *Arthritis Rheum.* 2007; **56**: 399–408.
[PubMed Abstract](#) | [Publisher Full Text](#)
117. Bechtold DA, Gibbs JE, Loudon AS: **Circadian dysfunction in disease.** *Trends Pharmacol. Sci.* 2010; **31**: 191–198.
[Publisher Full Text](#)
118. Amir M, et al.: **REV-ERB α regulates TH17 cell development and autoimmunity.** *Cell Rep.* 2018; **25**: 3733–3749.e8.
[PubMed Abstract](#) | [Publisher Full Text](#)
119. Chang C, et al.: **The nuclear receptor REV-ERB α modulates Th17 cell-mediated autoimmune disease.** *Proc. Natl. Acad. Sci.* 2019; **116**: 18528–18536.
[PubMed Abstract](#) | [Publisher Full Text](#)
120. Griffin P, et al.: **Circadian clock protein Rev-erb α regulates neuroinflammation.** *Proc. Natl. Acad. Sci.* 2019; **116**: 5102–5107.
[PubMed Abstract](#) | [Publisher Full Text](#)
121. Guo D-K, et al.: **Pharmacological activation of REV-ERB α represses LPS-induced microglial activation through the NF- κ B pathway.** *Acta Pharmacol. Sin.* 2019; **40**: 26–34.
[PubMed Abstract](#) | [Publisher Full Text](#)
122. Reitz CJ, et al.: **SR9009 administered for one day after myocardial ischemia-reperfusion prevents heart failure in mice by targeting the cardiac inflammasome.** *Commun Biol.* 2019; **2**: 353.
[PubMed Abstract](#) | [Publisher Full Text](#)
123. Zhang L, et al.: **REV-ERB α ameliorates heart failure through transcription repression.** *JCI Insight.* 2017; **2**.
[PubMed Abstract](#) | [Publisher Full Text](#)
124. Stujanna EN, et al.: **Rev-erb agonist improves adverse cardiac remodeling and survival in myocardial infarction through an anti-inflammatory mechanism.** *PLoS One.* 2017; **12**: e0189330.
[Publisher Full Text](#)
125. Zhou Z, et al.: **Circadian pharmacological effects of berberine on chronic colitis in mice: role of the clock component Rev-erb α .** *Biochem. Pharmacol.* 2020; **172**: 113773.
[PubMed Abstract](#) | [Publisher Full Text](#)
126. Zhao W, et al.: **Activation of Rev-erb α attenuates lipopolysaccharide-induced inflammatory reactions in human endometrial stroma cells via suppressing TLR4-regulated NF- κ B activation.** *Acta Biochim. Biophys. Sin.* 2019; **51**: 908–914.
[Publisher Full Text](#)
127. Jetten AM, Cook DN: **(Inverse) Agonists of Retinoic Acid-Related Orphan Receptor gamma: Regulation of Immune Responses, Inflammation, and Autoimmune Disease.** *Annu. Rev. Pharmacol. Toxicol.* 2020; **60**: 371–390.
[PubMed Abstract](#) | [Publisher Full Text](#)
128. Yu X, et al.: **TH17 cell differentiation is regulated by the circadian clock.** *Science.* 2013; **342**: 727–730.
[PubMed Abstract](#) | [Publisher Full Text](#)
129. Solt LA, et al.: **Suppression of TH17 differentiation and autoimmunity by a synthetic ROR ligand.** *Nature.* 2011; **472**: 491–494.
[PubMed Abstract](#) | [Publisher Full Text](#)
130. Ivanov II, et al.: **The orphan nuclear receptor ROR γ t directs the differentiation program of proinflammatory IL-17+ T helper cells.** *Cell.* 2006; **126**: 1121–1133.
[PubMed Abstract](#) | [Publisher Full Text](#)
131. Yang XO, et al.: **T helper 17 lineage differentiation is programmed by orphan nuclear receptors ROR α and ROR γ .** *Immunity.* 2008; **28**: 29–39.
[PubMed Abstract](#) | [Publisher Full Text](#)
132. Solt LA, et al.: **Identification of a selective ROR γ ligand that suppresses Th17 cells and stimulates T regulatory cells.** *ACS Chem. Biol.* 2012; **7**: 1515–1519.
[PubMed Abstract](#) | [Publisher Full Text](#)
133. Banerjee S, et al.: **Pharmacological targeting of the mammalian clock regulates sleep architecture and emotional behaviour.** *Nat. Commun.* 2014; **5**: 5759.
[PubMed Abstract](#) | [Publisher Full Text](#)
134. Chung S, et al.: **Impact of circadian nuclear receptor REV-ERB α on midbrain dopamine production and mood regulation.** *Cell.* 2014; **157**: 858–868.
[PubMed Abstract](#) | [Publisher Full Text](#)
135. Son GH, Chung S, Ramirez VD, et al.: **Pharmacological Modulators of Molecular Clock and their Therapeutic Potentials in Circadian Rhythm-Related Diseases.** *Med. Chem. (Los Angeles).* 2016; **6**: 12.
[Publisher Full Text](#)
136. Young JK, Heinbockel T, Gondré-Lewis MC: **Astrocyte fatty acid binding protein-7 is a marker for neurogenic niches in the rat hippocampus.** *Hippocampus.* 2013; **23**: 1476–1483.
[PubMed Abstract](#) | [Publisher Full Text](#)
137. Giachino C, et al.: **Molecular diversity subdivides the adult forebrain neural stem cell population.** *Stem Cells.* 2014; **32**: 70–84.
[PubMed Abstract](#) | [Publisher Full Text](#)
138. Schnell A, et al.: **The nuclear receptor REV-ERB α regulates Fbp7 and modulates adult hippocampal neurogenesis.** *PLoS One.* 2014; **9**: e99883.
[PubMed Abstract](#) | [Publisher Full Text](#)
139. Sashihara S, Felts PA, Waxman SG, et al.: **Orphan nuclear receptor ROR α gene: isoform-specific spatiotemporal expression during postnatal development of brain.** *Mol. Brain Res.* 1996; **42**: 109–117.
[PubMed Abstract](#) | [Publisher Full Text](#)
140. Matsui T, Sashihara S, Oh Y, et al.: **An orphan nuclear receptor, mROR α , and its spatial expression in adult mouse brain.** *Mol. Brain Res.* 1995; **33**: 217–226.
[PubMed Abstract](#) | [Publisher Full Text](#)
141. Dussault I, Fawcett D, Matthyssen A, et al.: **Orphan nuclear receptor ROR α -deficient mice display the cerebellar defects of staggerer.** *Mech. Dev.* 1998; **70**: 147–153.
[PubMed Abstract](#) | [Publisher Full Text](#)
142. Chen XR, et al.: **Mature Purkinje cells require the retinoic acid-related orphan receptor- α (ROR α) to maintain climbing fiber mono-innervation and other adult characteristics.** *J. Neurosci.* 2013; **33**: 9546–9562.
[PubMed Abstract](#) | [Publisher Full Text](#)
143. Doulazmi M, et al.: **A comparative study of Purkinje cells in two ROR α gene mutant mice: staggerer and ROR α -/-.** *Dev. Brain Res.* 2001; **127**: 165–174.
[PubMed Abstract](#) | [Publisher Full Text](#)
144. Janmaat S, et al.: **Age-related Purkinje cell death is steroid dependent: ROR α haplo-insufficiency impairs plasma and cerebellar steroids and Purkinje cell survival.** *Age.* 2011; **33**:

- 565–578.
[PubMed Abstract](#) | [Publisher Full Text](#)
145. Jolly S, *et al.*: **Cell-autonomous and non-cell-autonomous neuroprotective functions of ROR α in neurons and astrocytes during hypoxia.** *J. Neurosci.* 2011; **31**: 14314–14323.
[PubMed Abstract](#) | [Publisher Full Text](#)
146. Srinivas M, Ng L, Liu H, *et al.*: **Activation of the blue opsin gene in cone photoreceptor development by retinoid-related orphan receptor β .** *Mol. Endocrinol.* 2006; **20**: 1728–1741.
[PubMed Abstract](#) | [Publisher Full Text](#)
147. Pircher P, Chomez P, Yu F, *et al.*: **Aberrant expression of myosin isoforms in skeletal muscles from mice lacking the rev-erbA α orphan receptor gene.** *Am. J. Phys. Regul. Integr. Comp. Phys.* 2005; **288**: R482–R490.
[PubMed Abstract](#) | [Publisher Full Text](#)
148. Welch RD, Flaveny CA: **REV-ERB and ROR: therapeutic targets for treating myopathies.** *Phys. Biol.* 2017; **14**: 045002.
[PubMed Abstract](#) | [Publisher Full Text](#)
149. Xiong X, Gao H, Lin Y, *et al.*: **Inhibition of Rev-erbalpha ameliorates muscular dystrophy.** *Exp. Cell Res.* 2021; **406**: 112766.
[PubMed Abstract](#) | [Publisher Full Text](#)
150. Boulanguiez A, *et al.*: **NR1D1 controls skeletal muscle calcium homeostasis through myoregulin repression.** *JCI Insight.* 2022; **7**.
[PubMed Abstract](#) | [Publisher Full Text](#)
151. Ramakrishnan SN, Lau P, Burke LJ, *et al.*: **Rev-erb β regulates the expression of genes involved in lipid absorption in skeletal muscle cells: evidence for cross-talk between orphan nuclear receptors and myokines.** *J. Biol. Chem.* 2005; **280**: 8651–8659.
[Publisher Full Text](#)
152. Welch RD, *et al.*: **Rev-Erb co-regulates muscle regeneration via tethered interaction with the NF- κ B cytosome.** *Mol. Metab.* 2017; **6**: 703–714.
[PubMed Abstract](#) | [Publisher Full Text](#)
153. Fitzsimmons RL, Lau P, Muscat GE: **Retinoid-related orphan receptor alpha and the regulation of lipid homeostasis.** *J. Steroid Biochem. Mol. Biol.* 2012; **130**: 159–168.
[PubMed Abstract](#) | [Publisher Full Text](#)
154. Raichur S, *et al.*: **Identification and validation of the pathways and functions regulated by the orphan nuclear receptor, ROR alpha1, in skeletal muscle.** *Nucleic Acids Res.* 2010; **38**: 4296–4312.
[PubMed Abstract](#) | [Publisher Full Text](#)
155. Ercolani L, *et al.*: **Circadian clock: Time for novel anticancer strategies?** *Pharmacol. Res.* 2015; **100**: 288–295.
[Publisher Full Text](#)
156. Sulli G, Manoojian ENC, Taub PR, *et al.*: **Training the Circadian Clock, Clocking the Drugs, and Drugging the Clock to Prevent, Manage, and Treat Chronic Diseases.** *Trends Pharmacol. Sci.* 2018; **39**: 812–827.
[PubMed Abstract](#) | [Publisher Full Text](#)
157. Verlande A, *et al.*: **Glucagon regulates the stability of REV-ERBalpha to modulate hepatic glucose production in a model of lung cancer-associated cachexia.** *Sci. Adv.* 2021; **7**.
[PubMed Abstract](#) | [Publisher Full Text](#)
158. Zhu Y, McAvoy S, Kuhn R, *et al.*: **RORA, a large common fragile site gene, is involved in cellular stress response.** *Oncogene.* 2006; **25**: 2901–2908.
[PubMed Abstract](#) | [Publisher Full Text](#)
159. Kottorou AE, *et al.*: **Altered expression of NFY-C and RORA in colorectal adenocarcinomas.** *Acta Histochem.* 2012; **114**: 553–561.
[PubMed Abstract](#) | [Publisher Full Text](#)
160. Moretti RM, Montagnani Marelli M, Sala A, *et al.*: **Activation of the orphan nuclear receptor ROR α counteracts the proliferative effect of fatty acids on prostate cancer cells: Crucial role of 5-lipoxygenase.** *Int. J. Cancer.* 2004; **112**: 87–93.
[PubMed Abstract](#) | [Publisher Full Text](#)
161. Xiong G, Wang C, Evers BM, *et al.*: **ROR α suppresses breast tumor invasion by inducing SEMA3F expression.** *Cancer Res.* 2012; **72**: 1728–1739.
[PubMed Abstract](#) | [Publisher Full Text](#)
162. Du J, Xu R: **ROR α , a potential tumor suppressor and therapeutic target of breast cancer.** *Int. J. Mol. Sci.* 2012; **13**: 15755–15766.
[Publisher Full Text](#)
163. Ye Y, *et al.*: **The Genomic Landscape and Pharmacogenomic Interactions of Clock Genes in Cancer Chronotherapy.** *Cell systems.* 2018; **6**: 314–328.e2.
[PubMed Abstract](#) | [Publisher Full Text](#)
164. Fu R-D, Qiu C-H, Chen H-A, *et al.*: **Retinoic acid receptor-related receptor alpha (RORalpha) is a prognostic marker for hepatocellular carcinoma.** *Tumor Biol.* 2014; **35**: 7603–7610.
[PubMed Abstract](#) | [Publisher Full Text](#)
165. Ou Z, *et al.*: **Regulation of the human hydroxysteroid sulfotransferase (SULT2A1) by ROR α and ROR γ and its potential relevance to human liver diseases.** *Mol. Endocrinol.* 2013; **27**: 106–115.
[Publisher Full Text](#)
166. Kim H, *et al.*: **DNA damage-induced ROR α is crucial for p53 stabilization and increased apoptosis.** *Mol. Cell.* 2011; **44**: 797–810.
[PubMed Abstract](#) | [Publisher Full Text](#)
167. Ueda E, *et al.*: **High incidence of T-cell lymphomas in mice deficient in the retinoid-related orphan receptor ROR γ .** *Cancer Res.* 2002; **62**: 901–909.
[PubMed Abstract](#)
168. Huang H, *et al.*: **The Multifunctional Effects of Nobiletin and Its Metabolites in vivo and In Vitro.** *Evid. Based Complement. Alternat. Med.* 2016; **2016**: 2918796.
[PubMed Abstract](#)
169. Evans M, Sharma P, Guthrie N: **Bioavailability of Citrus Polymethoxylated Flavones and Their Biological Role in Metabolic Syndrome and Hyperlipidemia.** InTech; 2012; 1–19.
[Publisher Full Text](#)
170. Mulvihill EE, Burke AC, Huff MW: **Citrus Flavonoids as Regulators of Lipoprotein Metabolism and Atherosclerosis.** *Annu. Rev. Nutr.* 2016; **36**: 275–299.
[PubMed Abstract](#) | [Publisher Full Text](#)
171. Mileykovskaya E, Yoo SH, Dowhan W, *et al.*: **Nobiletin: Targeting the Circadian Network to Promote Bioenergetics and Healthy Aging.** *Biochemistry, Biokhimiia.* 2020; **85**: 1554–1559.
[PubMed Abstract](#) | [Publisher Full Text](#)
172. He B, *et al.*: **The small molecule nobiletin targets the molecular oscillator to enhance circadian rhythms and protect against metabolic syndrome.** *Cell Metab.* 2016; **23**: 610–621.
[PubMed Abstract](#) | [Publisher Full Text](#)
173. Miller S, Son YL, Aikawa Y, *et al.*: **Isoform-selective regulation of mammalian cryptochromes.** *Nat. Chem. Biol.* 2020 Jun; **16**(6): 676–685.
[PubMed Abstract](#) | [Publisher Full Text](#)
174. Nohara K, *et al.*: **Ammonia-lowering activities and carbamoyl phosphate synthetase 1 (Cps1) induction mechanism of a natural flavonoid.** *Nutr. Metab. (Lond.)* 2015; **12**: 23.
[PubMed Abstract](#) | [Publisher Full Text](#)
175. Nohara K, *et al.*: **Nobiletin fortifies mitochondrial respiration in skeletal muscle to promote healthy aging against metabolic challenge.** *Nat. Commun.* 2019; **10**: 3923.
[PubMed Abstract](#) | [Publisher Full Text](#)
176. Nohara K, *et al.*: **Cardiolipin Synthesis in Skeletal Muscle Is Rhythmic and Modifiable by Age and Diet.** *Oxidative Med. Cell. Longev.* 2020; **2020**: 5304768.
177. Kim E, *et al.*: **Effects of the Clock Modulator Nobiletin on Circadian Rhythms and Pathophysiology in Female Mice of an Alzheimer's Disease Model.** *Biomolecules.* 2021; **11**: 1004.
[PubMed Abstract](#) | [Publisher Full Text](#)
178. Rakshit K, Matveyenko AV: **Induction of Core Circadian Clock Transcription Factor Bmal1 Enhances Beta-Cell Function and Protects Against Obesity-Induced Glucose Intolerance.** *Diabetes.* 2021; **70**: 143–154.
[PubMed Abstract](#) | [Publisher Full Text](#)
179. Petrenko V, *et al.*: **In pancreatic islets from type 2 diabetes patients, the dampened circadian oscillators lead to reduced insulin and glucagon exocytosis.** *Proc. Natl. Acad. Sci. U. S. A.* 2020; **117**: 2484–2495.
[PubMed Abstract](#) | [Publisher Full Text](#)
180. Nohara K, Nemkov T, D'Alessandro A, *et al.*: **Coordinate Regulation of Cholesterol and Bile Acid Metabolism by the Clock Modifier Nobiletin in Metabolically Challenged Old Mice.** *Int. J. Mol. Sci.* 2019; **20**.
[PubMed Abstract](#) | [Publisher Full Text](#)
181. Oyama Y, Bartman CM, Gile J, *et al.*: **The Circadian PER2 Enhancer Nobiletin Reverses the Deleterious Effects of Midazolam in Myocardial Ischemia and Reperfusion Injury.** *Curr. Pharm. Des.* 2018; **24**: 3376–3383.
[PubMed Abstract](#) | [Publisher Full Text](#)
182. Shinozaki A, *et al.*: **Potent Effects of Flavonoid Nobiletin on Amplitude, Period, and Phase of the Circadian Clock Rhythm in PER2::LUCIFERASE Mouse Embryonic Fibroblasts.** *PLoS One.* 2017; **12**: e0170904.
[PubMed Abstract](#) | [Publisher Full Text](#)
183. Wirianto M, *et al.*: **The clock modulator Nobiletin mitigates astrogliosis-associated neuroinflammation and disease hallmarks in an Alzheimer's disease model.** *FASEB J.* 2022; **36**: e22186.

184. Yang X, *et al.*: **Nobiletin Delays Aging and Enhances Stress Resistance of *Caenorhabditis elegans*.** *Int. J. Mol. Sci.* 2020; **21**.
[PubMed Abstract](#) | [Publisher Full Text](#)
185. Kim E, *et al.*: **ROR activation by Nobiletin enhances antitumor efficacy via suppression of I κ B/NF- κ B signaling in triple-negative breast cancer.** *Cell Death Dis.* 2022; **13**: 374.
[PubMed Abstract](#) | [Publisher Full Text](#)
186. Lellupitiyage Don SS, *et al.*: **Nobiletin affects circadian rhythms and oncogenic characteristics in a cell-dependent manner.** *PLoS One.* 2020; **15**: e0236315.
[PubMed Abstract](#) | [Publisher Full Text](#)
187. Chen Z, Yoo S-H, Takahashi JS: **Development and therapeutic potential of small-molecule modulators of circadian systems.** *Annu. Rev. Pharmacol. Toxicol.* 2018; **58**: 231–252.
[PubMed Abstract](#) | [Publisher Full Text](#)
188. Koronowski KB, Sassone-Corsi P: **Communicating clocks shape circadian homeostasis.** *Science.* 2021; **371**.
[PubMed Abstract](#) | [Publisher Full Text](#)

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Tsuyoshi Hirota 

Nagoya University, Nagoya, Japan

The authors addressed all my points in the revised manuscript, and I have no further comments.

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: circadian biology, small molecules

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Version 1

Reviewer Report 11 November 2022

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Tsuyoshi Hirota 

Nagoya University, Nagoya, Japan

In this manuscript, Kim, Yoo, and Chen beautifully and convincingly describe the relevance of REV-ERB and ROR nuclear hormone receptors as therapeutic targets of metabolic disorders, immune diseases, brain diseases, muscle pathologies, and cancer, as well as current challenges in the research, with special emphasis on natural compound nobiletin. This well-written review article

covers the translation of the molecular clock mechanism for health and healthy aging, which is an important topic in the field. I strongly support the publication of this article, and a brief description of the following points would be useful for further understanding:

- Are the therapeutic effects of nobiletin dependent on both ROR α and ROR γ ? As the authors described, isoforms of REV-ERB (α and β) and ROR (α , β , and γ) have different expression patterns and physiological functions. Therefore, it would be nice to discuss the possibility and potential of isoform-selective ligands as well.
- Because nobiletin is a natural compound and other REV-ERB/ROR ligands are synthetic compounds, it would be nice to mention the merit (and demerit) of natural compounds compared to synthetic compounds.

Minor points:

- Page 4, line 2: *Drosophila* to be italic.
- Page 5, "Therapeutic relevance of REV-ERBs and RORs" section, lines 2-3: "expression" is duplicated.
- Page 6, line 11: ZT may need an explanation.
- Page 6, the second paragraph, line 1: *Rev-erba* to be italic.
- Please check whether references 60 (Emery and Clayton, 2001) and 137 (Nagoshi et al., 2004) are proper literature in the context.

Is the topic of the review discussed comprehensively in the context of the current literature?

Yes

Are all factual statements correct and adequately supported by citations?

Yes

Is the review written in accessible language?

Yes

Are the conclusions drawn appropriate in the context of the current research literature?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: circadian biology, small molecules

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 28 Nov 2022

Eunju Kim, McGovern Medical School, The University of Texas Health Science Center at Houston (UTHealth Houston), Houston, USA

Point-by-Point Response

We thank both expert reviewers for their thorough and thoughtful comments. Our response is as follows.

Reviewer 2

In this manuscript, Kim, Yoo, and Chen beautifully and convincingly describe the relevance of REV-ERB and ROR nuclear hormone receptors as therapeutic targets of metabolic disorders, immune diseases, brain diseases, muscle pathologies, and cancer, as well as current challenges in the research, with special emphasis on natural compound nobiletin. This well-written review article covers the translation of the molecular clock mechanism for health and healthy aging, which is an important topic in the field. I strongly support the publication of this article, and a brief description of the following points would be useful for further understanding:

Thank you very much for the positive comments.

Are the therapeutic effects of nobiletin dependent on both ROR α and ROR γ ? As the authors described, isoforms of REV-ERB (α and β) and ROR (α , β , and γ) have different expression patterns and physiological functions. Therefore, it would be nice to discuss the possibility and potential of isoform-selective ligands as well.

Thank you for this valuable comment. NOB shows robust binding to the LBDs of ROR α and ROR γ , with somewhat higher affinity for ROR γ (PMID: 27076076). However, there is currently no functional evidence for a possible selectivity of NOB toward either ROR. We agree that potential isoform/subtype-selective ligands, such as those characterized for CRYs, would be valuable. We have added this discussion to the text on page 9.

Because nobiletin is a natural compound and other REV-ERB/ROR ligands are synthetic compounds, it would be nice to mention the merit (and demerit) of natural compounds compared to synthetic compounds.

Thank you for this excellent comment. NOB's excellent safety profile is indeed a significant advantage over other synthetic ligands which may require extensive medicinal chemistry efforts before in vivo and clinical applications. Without making a direct comparison, we have added a sentence on page 10 to highlight this point. Thank you.

Minor points:

Page 4, line 2: *Drosophila* to be italic.

As suggested, we have italicized the word.

Page 5, "Therapeutic relevance of REV-ERBs and RORs" section, lines 2-3: "expression" is duplicated.

As suggested, we corrected the sentence.

Page 6, line 11: ZT may need an explanation.

Apologies for this omission. We added the full name and explanation of ZT in the manuscript.

Page 6, the second paragraph, line 1: *Rev-erba-/-* to be italic.

As suggested, we have italicized the word.

Please check whether references 60 (Emery and Clayton, 2001) and 137 (Nagoshi *et al.*, 2004) are proper literature in the context.

As suggested, we have removed these references as they are not immediately relevant as the reviewer pointed out.

Thank you again.

Competing Interests: No competing interests were disclosed.

Reviewer Report 11 November 2022

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Shihoko Kojima 

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Evan S. Littleton

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Summary:

In the review article titled “Circadian stabilization loop: the regulatory hub and therapeutic target promoting circadian resilience and physiological health”, the authors discuss various studies on the role of *Rev-erbs* and *Rors* on health and aging with the main focus being their function within the circadian clock. The authors give a well-balanced and comprehensive approach to discussing the importance of these genes in many different physiological pathways, such as immune response, metabolic disorders, cancer, and more. The authors provide extensive citations to support their discussion as well. We only have minor comments, and believe the article is suitable for approval.

Minor Concerns:

1. In the second paragraph of the section titled “The circadian timing system and health implications”, *Dbp* and *Nfil3* are not mentioned as part of the circadian clock, despite their known regulation of multiple clock genes via D-boxes.
2. In the third paragraph of the section titled “The stabilization loop”, the authors state “While *Ror* expressions display relatively moderate circadian oscillatory amplitude, *Rev-erbs* are among the most oscillatory genes (highest amplitude) in both protein and mRNA expression”. However, *Rors* themselves differ in amplitude with *Rora* expression showing little to no rhythmicity in most tissues. Meanwhile, *Rorc* expression is rhythmic and displays similar amplitude to *Rev-erbs* in some tissues/cell types^{1,2,3}.

3. The nomenclature for mouse models should be superscripted (for example, *Rora^{sg/sg}* rather than *Rorasg/sg*)

References

1. Takeda Y, Jothi R, Birault V, Jetten AM: ROR γ directly regulates the circadian expression of clock genes and downstream targets in vivo. *Nucleic Acids Res.* 2012; **40** (17): 8519-35 [PubMed Abstract](#) | [Publisher Full Text](#)
2. Littleton E, Childress M, Gosting M, Jackson A, et al.: Genome-wide correlation analysis to identify amplitude regulators of circadian transcriptome output. *Scientific Reports.* 2020; **10** (1). [Publisher Full Text](#)
3. Ikeda R, Tsuchiya Y, Koike N, Umemura Y, et al.: REV-ERB α and REV-ERB β function as key factors regulating Mammalian Circadian Output. *Scientific Reports.* 2019; **9** (1). [Publisher Full Text](#)

Is the topic of the review discussed comprehensively in the context of the current literature?

Yes

Are all factual statements correct and adequately supported by citations?

Yes

Is the review written in accessible language?

Yes

Are the conclusions drawn appropriate in the context of the current research literature?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Circadian genomics, rhythmic gene expression, mouse, clock-controlled genes

We confirm that we have read this submission and believe that we have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 28 Nov 2022

Eunju Kim, McGovern Medical School, The University of Texas Health Science Center at Houston (UTHealth Houston), Houston, USA

Point-by-Point Response

We thank both expert reviewers for their thorough and thoughtful comments. Our response is as follows.

Reviewer 1

In the review article titled "Circadian stabilization loop: the regulatory hub and therapeutic target promoting circadian resilience and physiological health", the

authors discuss various studies on the role of Rev-erbs and Rors on health and aging with the main focus being their function within the circadian clock. The authors give a well-balanced and comprehensive approach to discussing the importance of these genes in many different physiological pathways, such as immune response, metabolic disorders, cancer, and more. The authors provide extensive citations to support their discussion as well. We only have minor comments, and believe the article is suitable for approval.

We thank the reviewer for the overall positive comments.

Minor Concerns:

1. In the second paragraph of the section titled “The circadian timing system and health implications”, Dbp and Nfil3 are not mentioned as part of the circadian clock, despite their known regulation of multiple clock genes via D-boxes.

Apologies for this omission. We have added Dbp and Nfil3 as additional core clock genes functioning in the oscillator on page 3.

2. In the third paragraph of the section titled “The stabilization loop”, the authors state “While Ror expressions display relatively moderate circadian oscillatory amplitude, Rev-erbs are among the most oscillatory genes (highest amplitude) in both protein and mRNA expression”. However, Rors themselves differ in amplitude with Rora expression showing little to no rhythmicity in most tissues. Meanwhile, Rorc expression is rhythmic and displays a similar amplitude to Rev-erbs in some tissues/cell types^{1,2,3}.

Thank you for this great comment. We have added a discussion text with these references to highlight the differences between the RORs (page 5, paragraph 2 in “the stabilization loop”). We thank the reviewer for this excellent suggestion.

3. The nomenclature for mouse models should be superscripted (for example, Rora^{sg/sg} rather than Rorasg/sg)

As suggested, we have superscripted the mouse model names.

Thank you again.

Competing Interests: No competing interests were disclosed.

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