

ISSN: (Print) (Online) Journal homepage: https://www.tandfonline.com/loi/tneo20

Diversity of the Ganoderma species in Uruguay

G. Morera, S. Lupo, S. Alaniz & G. Robledo

To cite this article: G. Morera, S. Lupo, S. Alaniz & G. Robledo (2021) Diversity of the Ganoderma species in Uruguay, Neotropical Biodiversity, 7:1, 570-585, DOI: 10.1080/23766808.2021.1986329

To link to this article: <u>https://doi.org/10.1080/23766808.2021.1986329</u>

0

© 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.



View supplementary material



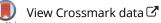
Published online: 04 Jan 2022.

Submity	/our	article	to	this	iournal	

лII	Article views: 897
	Article views: 897



View related articles



OPEN ACCESS Check for updates

Diversity of the Ganoderma species in Uruguay

G. Morera ^[], S. Lupo ^[], S. Alaniz ^[] and G. Robledo ^[], ^{c,d,e}

^aSección Micología, Facultad de Ciencias, Universidad de la República, Montevideo, Uruguay; ^bDepartamneto de Protección Vegetal, Facultad de Agronomía, Universidad de la República, Montevideo, Uruguay; ^cBioTecA3 – Centro de Biotecnología Aplicada al Agro y Alimento – Ing. Agr, Universidad Nacional de Córdoba, Córdoba, Argentina; ^dCONICET, Consejo Nacional de Investigaciones Científicas y Técnicas, Córdoba, Argentina; ^eFundación Fungicosmos, www.fungicosmos.org, Córdoba, Argentina

ABSTRACT

Ganoderma is a cosmopolitan genus that includes a great diversity of species. Many of them have been historically described based only on morphological characteristics; however, due to their morphological plasticity, there is no complete understanding about their relationship and taxonomic status. Commonly applied names, particularly in the southern Neotropics, come from species of North Hemisphere distribution (e.g. *G. lucidum, G. resinaceum* and *G. applanatum*). The objective of the present work was to perform a survey of *Ganoderma* species thriving in Uruguay. We aimed to identify and characterize them through molecular, morphological and ecological analysis. The results confirm the presence of four reddish laccate species first registered for Uruguay (*G. dorsale, G. platense, G. martinicense* and *G. mexicanum*), and one non-laccate species (*G. australe s.l.*) composed of two clades. The species are morphologically differentiated mainly by its stipe, pilear surface, context, pores, basidiospores and cutis cells. Regarding the ecological data, the species present differences in substrate preferences. In addition, a taxonomic discussion regarding phylogenetic relationships and taxonomic status of Uruguayan *Ganoderma* species is presented.

ARTICLE HISTORY

Received 21 October 2020 Accepted 16 September 2021

KEYWORDS ITS; mycogeography; phylogeny; systematics; South American polypores

Introduction

Ganoderma (P. Karst.) harbors at least 220 species, being the most diverse genus of Ganodermataceae [1–4]. However, due to their high diversity and phenotypic plasticity, the phylogenetic relationship and taxonomic status of many species remain unclear until now [5,6]. In the last 20 years, the use of molecular tools, mainly through the amplification and sequencing of Internal Transcribed Spacer (ITS), has been incorporated into systematic studies and into circumscription of Ganoderma species around the world. In this sense, through phylogenies based on molecular characters, the tendency in recent years has been to reinterpret the variations in characters of morphologically defined species, reinterpret the ecological relationships (relationship with hosts), determine their distribution and arrive at an understanding of the biogeographic processes that shape it [5,7–11].

Ganoderma species are morphologically characterized by the formation of sessile to stipitate basidiomata, with a glossy reddish laccate to opaque non-laccate cover, ellipsoid to ovoid double-walled basidiospores with truncated apex and endosporium with columnar ornamentations [12]. This cosmopolitan genus is comprised of parasitic and saprophytic species that decay the wood of plants from temperate and tropical areas around the world [6,13,14]. These fungi are described as white rot decayers that play a critical role in the dynamics of wood decomposition in tropical forests [15]. Moreover, they are the main cause of tree deterioration in public ornamental and commercial plantations [10,15–18].

In the last decades, some researchers have tried to elucidate the diversity of *Ganoderma* genus in the Neotropics, particularly Bazzalo and Wright [19], Gilbertson and Ryvarden [20], Gottlieb and Wright [21,22], Gottlieb et al. [7], Ryvarden [12,23], Torres-Torres and Dávalos [24], Torres-Torres et al. [25,26], and more recently Cabarroi-Hernández et al. [6] and Loyd et al. [11] for the northern limit of Neotropical distribution. Uruguay in particular harbors a great biodiversity due to its transitional condition and ecoregions diversity [27]. Some *Ganoderma* species have been historically recorded in Uruguay (Table 1), while Gazzano [28,29] and Martinez [30] have made more recent contributions to the diversity of the genus in Uruguay.

Some Uruguayan cited species as *G. applanatum* (Pers.) Pat. (= *G. lipsiense* (Batsch) G.F. Atk.), *G. sessile* Murrill, *G. lucidum* P. Karst. and *G. resinaceum* Boud were distributed out of the southern Neotropics [14,35,36], while others as *G. lorenzianum* (Kalchbr.)

MS. Guillermo Morera. Main author of the work, he collected the specimens, analyzed the data and wrote the manuscript.Dra. Sandra Lupo Rizzo. Participated in field campaigns, molecular analysis and reviewed the manuscript.Dra. Sandra Alaniz Ferro. Participated in the revision and correction of the manuscript.Dr. Gerardo Robledo. Contributed in morphological and phylogenetic analyses, and reviewed the manuscript.

© 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

CONTACT G. Morera 🖾 mguillemorera@gmail.com

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Pat., *G. nitens* (Fr.) Pat. lack of phylogenetic studies and their taxonomic status is unclear. Until now, there is no complete understanding on the systematic of the *Ganoderma* genus in Uruguay and its species diversity remains unknown.

The objective of the present work was to perform an updated survey of the *Ganoderma* species thriving in Uruguay and characterize them through molecular, morphological and ecological analysis. We also aimed to discuss the taxonomic status of the species of Uruguay and contrast with previous reports. We hypothesized that the *Ganoderma* specimens of Uruguay correspond to native species that have not been previously reported for the country. Those species are different from those registered to date (with names of species described from the Northern Hemisphere).

Materials and methods

Fungal specimens, basidiomata description and host characterization

Fresh Ganoderma basidiomata were collected from indigenous and urban ecosystems of Uruguay, during 2017 and 2018 field expeditions. Geographic location, host species and substrate condition (living tree, dead trunk, roots and stump) data were taken at the collection site following Urcelay and Robledo [37]. For fresh basidiomata, small pieces were aseptically taken from the context, placed into 2% malt extract agar (MEA) and incubated in darkness at 25°C. Pure cultures and basidiomata were deposited in MVHC. In addition, specimens from national herbaria (MVHC and MVM) were examined. Herbarium acronyms follow Thiers [38] (continuously updated, http://sweetgum.nybg.org/).

For morphological analyses and basidiomata identification, macroscopic and microscopic observations were made on basidiomata following the terminology and methodology according to authors [6,7,19,22–26]. Macroscopic features of basidiomata were analyzed and measured, particularly: pileus dimension, colour, texture, shape and appearance of surface, margin and stipe. The colour, presence of melanoid deposits and texture of context were also inspected and pores were described and

measured (pores/mm). Microscopic features (basidiospores, chlamydospores in context, generative and somatic hyphae and cuticular cells) were analyzed with an optical microscope by mounting small sections of basidiomata with 5% KOH or Melzer's Reagent (to test for dextrinoid or amyloid reaction). In particular, for the analysis of the hyphal system, sections of basidiomata were treated for 24-48 h in 3% NaOH at 50-60°C [39]. Thirty basidiospores were measured from each specimen (length and width) and values were expressed as rank of mean values. Width was measured in the widest part of the spore and the length considered from the base to the truncated apex of the basidiospore. Then, Q ratio was calculated as the relation: length/width. Measures of cuticular cells were made from the middle part of the basidiomes.

Host relationships of each *Ganoderma* species in Uruguay were characterized by host range and native/exotic status. The preference of substrate condition was analyzed through the relative frequency of each *Ganoderma* species in each substrate condition (LT = stem of living tree, DT = dead trunks, S = stumps and R = soil, arising from roots of living or dead trees) following Urcelay and Robledo [37]. Then, substrate preference was determined by transforming this relative frequency into a percentage.

DNA extraction, PCR and sequencing

DNA extractions from pure cultures were performed, using the CTAB protocol of Doyle and Doyle [40] with modifications [41]. The PCR reaction of the ITS region (including ITS1, 5.8s and ITS2) was performed. The following PCR primers were alternatively used: ITS1/ITS4 [42], ITS1-F/ITS4-B [43], and ITS4/ITS5 [42]. The PCR mixture was prepared in a 25 μ l final volume, with 2 μ l of genomic DNA solution (10 ng), 16 μ l of mQ water, 0.25 μ l of Taq polymerase (1 U), 0.5 μ l of each primer (10 mM), 0.7 μ l of 50 mM MgCl2, 2.5 μ l of dntps (2.5 mM) and 2.5 μ l (10X) of buffer. PCR reactions were performed in a MultiGene Optimax thermocycler (Labnet International Inc) with the cycling conditions as follows: 3 min at 94°C, followed by 35 cycles, each

 Table 1. Ganoderma species recorded in Uruguay.

Taxon	Author
G. australe (Fr.) Pat.	Felippone [6667]
G. applanatum (Pers.) Pat. (= G. lipsiense (Batsch) G.F. Atk.)	Felippone [66], Herter [31], Gazzano [28,29,32] Martínez [30]
G. fornicatum (Fr.) Pat.	Felippone [66]
G. lorenzianum (Kalchbr.) Pat.	Patouillard [65], Felippone [66]
G. lucidum (Curtis) P. Karst.	Spegazzini [33], Felippone [66], Wright & Blumenfeld [34], Gazzano [28,29,32], Martínez [30]
G. nitens (Fr.) Pat.	Patouillard [65], Felippone [66]
G. resinaceum Boud.	Felippone [66], Gazzano [32], Martínez [30]
G. sessile Murrill	Gazzano [29]

572 🕒 G. MORERA ET AL.

Table 2. Ganoderma species, specimens, location, Gen Bank accession numbers for ITS sequences and reference source.

pecies/Voucher/Culture reference	Locality	ITS GenBank accession number	References
anoderma adspersum BS351.74	Belgium	X78742/X78763	Moncalvo et al. [13]
aTO00	Italy	AM906057	Guglielmo et al. [44]
anoderma annulare			
CTC 16803	Brazil	JQ520160	GenBank
noderma applanatum		10 - 20 - 21	
	Japan	JQ520161	GenBank
noderma australe s.l. VHC 5620	Uruguay	MN191569	This work
VHC 5564	Uruguay	MN191568	This work
VHC 5582	Uruguay	MN191570	This work
VHC 5659	Uruguay	MN191554	This work
VHC 5660	Uruguay	MN191553	This work
VHC 5668	Uruguay	MN191555	This work
VHC 5601	Uruguay	MN191556	This work
VHC 5605	Uruguay	MN191557	This work
VHC 5661	Uruguay	MN191558	This work
VHC 5646	Uruguay	MN191552	This work This work
VHC 5640 VHC 5641	Uruguay	MN191551 MN191550	This work
VHC 5680	Uruguay Uruguay	MN 191550 MN 191547	This work
VHC 5647	Uruguay	MN191549	This work
VHC 5587	Uruguay	MN191548	This work
VHC 5568	Uruguay	MN191559	This work
VHC 5645	Uruguay	MN191560	This work
VHC 5697	Uruguay	MN191561	This work
VHC 5717	Uruguay	MN191572	This work
VHC 5711	Uruguay	MN191562	This work
VHC 5722.2 VHC 5724	Uruguay	MN191563 MN191565	This work This work
VHC 5725	Uruguay Uruguay	MN 191565 MN191564	This work
VHC 5746	Uruguay	MN191573	This work
VHC 5688	Uruguay	MN191566	This work
VHC 5708	Uruguay	MN191567	This work
VHC 5579	Uruguay	MN191546	This work
VHC 5732	Uruguay	MN191545	This work
C 7177	Uruguay	MN191571	This work
705	Taiwan	X78750.1	Moncalvo and Buchanan [5]
noderma concinnum	D - U. J.	141077500	Conta Dagan da et al. [52]
obledo 3192 (FCOS) obledo 3235 (FCOS)	Bolivia Bolivia	MN077522 MN077523	Costa-Rezende et al. [53] Costa-Rezende et al. [53]
anoderma dorsale	DOIMA	1011077525	costa-nezende et al. [55]
IVHC 5588	Uruguay	MN191582	This work
VHC 5701	Uruguay	MN191581	This work
VHC 5653	Uruguay	MN191578	This work
VHC 5648	Uruguay	MN191579	This work
VHC 5654	Uruguay	MN191580	This work
noderma ecuadoriense		1/11/2020 1	
51799	Ecuador	KU128524	Crous et al. [45]
AC126	Ecuador Ecuador	KU128525	Crous et al. [45]
ly-2-4 noderma gibbosum	ECUdUUI	KU128526	Crous et al. [45]
5D34	China	EU273513	GenBank
noderma lipsiense	IU	20273313	Genbuik
AFC2424	Argentina	AF169977/8	Gottlieb et al. [7]
noderma lobatum	-		
AFC2411	Argentina	AF169989/90	Gottlieb et al. [7]
35222.48	USA	X78740/X78761	Moncalvo et al. [13]
noderma lucidum	A	A F 4 70007 / A F 4 70000	
AFC2419	Argentina	AF170007/AF170008	Gottlieb et al. [7]
AFC2493 MNUT1	Argentina USA	AF170009/A1470010 MG654070	Gottlieb et al. [7] Loyd et al. [11]
175217	England	KJ143911	Zhou et al. [46]
(V 33217 T	Norway	Z37096/ Z37073	Moncalvo et al. [13]
35 270.81	France	Z37049/Z37099	Moncalvo et al. [13]
ai 2272	Sweden	JQ781851	Cao et al. [14]
ai11593	Finland	JQ781852	Cao et al. [14]
CRC 37043	China	EU021460	Wang et al. [9]
noderma martinicense			
VHC 5635	Uruguay	MN191574	This work
VHC 5583	Uruguay	MN191575	This work
IVHC 5637 IVHC 5684	Uruguay	MN191577 MN191576	This work This work
46TX	Uruguay USA	MG654185	Loyd et al. [11]
art08_55 T	Martinique	KF963256	GenBank
· · · · · · · · · · · · · · · · ·	· ······ · · · · · · · · · · · · · · ·		· · · · ·
anoderma mexicanum			

(Continued)

Table 2. (Continued).

Species/Voucher/Culture reference	Locality	ITS GenBank accession number	References
XAL D.Jarvio 143	Mexico	MK531823	Cabarroi-Hernández et al. [6]
BAFC2580	Brazil	AH008108	Gottlieb et al. [7]
Ganoderma multipileum			
DAI 9447	China	KJ143914	Zhou et al. [46]
Ganoderma multiplicatum			
SPC9	Brazil	KU569553	Bolaños et al. [60]
	Brazil	KU569515	Bolaños et al. [60]
URM 83346 Ganoderma oerstedii	Brazil	JX310823	Bolaños et al. [60]
ATCC 52409	Argentina	Z37058/Z37083	Moncalvo et al. [13]
ATCC 52410	Argentina	X78739/X78760	Moncalvo et al. [13]
ATCC 52411	Argentina	Z37059/Z37084	Moncalvo et al. [13]
Ganoderma parvulum	5		
URM 83344	Brazil	JX310819	Correia de Lima Júnior et al. [57]
URM 2948	Brazil	JX310821	Correia de Lima Júnior et al. [57]
Ganoderma platense		101404505	
MVHC 5586	Uruguay	MN191585	This work
MVHC 5565 MVHC 5721	Uruguay Uruguay	MN191584 MN191591	This work This work
MVHC 5732.2	Uruguay	MN191592	This work
MVHC 5686	Uruguay	MN191593	This work
MVHC 5692	Uruguay	MN191596	This work
MVHC 5687.2	Uruguay	MN191597	This work
MVHC 5687	Uruguay	MN191594	This work
MVHC 5690	Uruguay	MN191595	This work
NC 5332	Uruguay	MN191587	This work
NC 7187 NC 5104	Uruguay Uruguay	MN191588 MN191589	This work This work
MVHC 5694	Uruguay	MN191389 MN191590	This work
MVHC 5691	Uruguay	MN191586	This work
BAFC384	Argentina	AH008109	Gottlieb et al. [7]
Ganoderma polychromum	5		
330OR	USA	MG654196	Loyd et al. [11]
BJ280CA	USA	MG910492	Loyd et al. [11]
Ganoderma resinaceum			
DP2	Italy	AM906060	Guglielmo et al. [44]
CIRM BRFM 753 HMAS86599	France	FJ805250 AY884177	GenBank
BR 4150	England France	KJ143915	Wang et al. [9] Zhou et al. [46]
GR-101	India	GU451246	Mohanty et al. [63]
CBS 22036	The Netherlands	JQ520201	Park et al. [60]
BCRC 36147	The Netherlands	KJ143916	Zhou et al. [46]
Ganoderma sessile			
111TX	USA	MG654306	Loyd et al. [11]
103SC	USA	MG654304	Loyd et al. [11]
113FL	USA	MG654307	Loyd et al. [11]
228DC 117TX	USA USA	MG654319 MG654309	Loyd et al. [11] Loyd et al. [11]
BAFC2373	Argentina	AH008111	Gottlieb et al. [7]
Ganoderma sp.	Aigentina	Anotorri	
MUCL27886	India	AF255190	Moncalvo and Buchanan [5]
LXT.8	Vietnam	AF255188	Moncalvo and Buchanan [5]
PKB96/330	Japan	AF255105	Moncalvo and Buchanan [5]
TAI-05	Taiwan	AF255193/4	Moncalvo and Buchanan [5]
CP331	Papua New Guinea	AF255125	Moncalvo and Buchanan [5]
JM97/31	USA	AF255098	Moncalvo and Buchanan [5]
JM98/132	China South Africa	AF255115 AF255149	Moncalvo and Buchanan [5]
JM98/2 ME-GAN-24	South Africa USA	AF255149 AF255131/2	Moncalvo and Buchanan [5] Moncalvo and Buchanan [5]
RV-PR10	Puerto Rico	AF255131/2 AF255133	Moncalvo et al. [13]
JMCR.132	Costa Rica	AF255138	Moncalvo and Buchanan [5]
MUCL40406	Ecuador	AF255139	Moncalvo and Buchanan [5]
MUCL40324	French Guiana	AF255141	Moncalvo and Buchanan [5]
NIAST824	South Korea	AF255114	Moncalvo and Buchanan [5]
HMAS60686	China	AF255191/2	Moncalvo and Buchanan [5]
BAFC2531	Chile	AF255176	Moncalvo and Buchanan [5]
BAFC2449	Argentina	AF255187	Moncalvo and Buchanan [5]
Ganoderma subamboinense var. laevisporum	Argontina	V70706	Cottligh et al [7]
ATCC 52419 ATCC 52420	Argentina Argentina	X78736 JQ520205	Gottlieb et al. [7] Park et al. [60]
UMNFL 100	USA	MG654373	Loyd et al. [11]
Ganoderma tornatum		11005 1575	
BAFC2764	Argentina	AF169993/4	Gottlieb et al. [7]
BAFC2582	Brazil	AF169985/6	Gottlieb et al. [7]
BAFC1139	Argentina	AF169979/80	Gottlieb et al. [7]
Ganoderma tuberculosum			
BAFC2488	Argentina	AH008114	Gottlieb and Wright [22]

(Continued)

Table 2. (Continued).

Species/Voucher/Culture reference	Locality	ITS GenBank accession number	References
PLM684	USA	MG654369	Loyd et al. [11]
UMNFL160	USA	MG654364	Loyd et al. [11]
Ganoderma zonatum			·
FL-02	USA	KJ143921	Zhou et al. [46]
179NC	USA	MG654417	Loyd et al. [11]
UMNFL105	USA	MG654408	Loyd et al. [11]
UMNSC4	USA	MG654415	Loyd et al. [11]
123FL	USA	MG654416	Loyd et al. [11]
BAFC2374	Argentina	AH008110	Gottlieb and Wright [22]
Cristataspora coffeata			
FLOR 50933	Brazil	KU315204	Costa-Rezende et al. [53]
Foraminispora rugosa			
HUEFS_DHCR560	Brazil	MF409963	Costa-Rezende et al. [53]
FLOR 52191	Brazil	KU315200	Costa-Rezende et al. [4]

consisting of 60 s at 94°C, 45 s at 50°C, 60 s at 72°C, and a final extension step at 72°C for 5 min. PCR products were verified by electrophoresis in 1.0% agarose gels in TBE buffer, stained with EZ vision®One (Amresco®) and visualized under UV light transillumination. GeneRuler DNA Ladder Mix marker (Thermo) was used as molecular size marker. PCR products were purified and sequenced by Macrogen (Seoul, Korea). Sequences were submitted to GenBank (Table 2).

Phylogenetic analyses

The sequences obtained were manually edited (visual inspection of sequences and chromatograms, resolution of conflicts and pair the extremes) with Bioedit V.7.0.5.3 [47] and incorporated into alignments with sequences of specimens from other parts of the world obtained from GenBank (Table 2). Multiple alignment was made using ProbCons 1.12 from the CIPRES Science Gateway [48]. Subsequently, the best evolutionary model for each region (ITS1, 5.8S and ITS2) was estimated using the Corrected Akaike Informational Criteria (AICc), implemented by the jModelTest2 v.1.6 software [49]. The phylogenetic analysis was conducted in two independent ways: Bayesian Inference (BI) and Maximum Likelihood (ML), performed with MrBayes 3.2.7 [50] and RAxML 8.2.12 [51], respectively, in CIPRES Science Gateway [52]. Cristataspora coffeata (Murrill) Robledo, Costa-Rezende and de Madrignac Bonzi (FLOR 50933) and Foraminispora rugosa (Berk.) Costa-Rezende, Drechsler-Santos and Robledo (FLOR 52191 and HUEFS DHCR560) were used as outgroup [4,53]. For the BI, two independent runs were performed, starting with random trees, with four independent and simultaneous chains, 10,000,000 MCMC generations, and maintaining 1 tree every 1000 generations. Burn in discarded values was indicated as 0.25. The estimated models for each partition were incorporated, as indicated below (see Results). The average standard deviation of split frequencies was limited to

below 0.01. Convergence of the Markov chains to a stationary distribution was visually inspected using the Tracer v.1. 7. 1 program [54]. A GTRGAMMA nucleotide model and 1000 bootstrap iterations were indicated for the ML analysis. The rest of the parameter values were set by default. Since the topologies of trees obtained in each analysis were convergent, only the consensus BI tree is shown with values of Bayesian posterior probability (BPP) and ML bootstrap (ML) separated by cross bars (BPP/BS). A clade was considered strongly supported if it showed a 0.95 BPP and/or 80% BS [55].

Results

Fungal specimens, basidiomata description and host characterization

A total of 163 *Ganoderma* specimens from collections generated in this work (n = 90), MVHC (n = 57) and MVM (n = 16) were morphologically and ecologically analyzed (Tables 3 and 4). *Ganoderma australe* was the most commonly collected species, with 101 specimens. The 62 remaining specimens belong to four reddish laccate species: *G. mexicanum* (n = 3), *G. martinicense* (n = 10), *G. platense* (n = 22) and *G. dorsale* (n = 27).

Morphologically assigned species are presented in Figures 1 and 2. *Ganoderma* species of Uruguay were primarily differentiated by their pilear surface into two groups: reddish laccate and non-laccate (Table 3 and Figure 1).

The first group is composed of two species with great and robust basidiomes (*G. martinicense* and *G. platense*) and two species with smaller basidiomes (*G. dorsale* and *G. mexicanum*).

Ganoderma martinicense is characterized by a large, commonly substipitate basidiomata with a tuberculous concentric zonated pilear surface and conspicuous melanoid deposits in the homogeneous context. Microscopically, it is characterized by the presence of a distinct smooth basidiospores ornamentation, formed by free pillars and cutis cells with weak

				Context			Pores		Bas	Basidiospores	Chlamydospores	ores		ט	Cutis cells
Taxon	Stipe	Surface	Margin	Color	Resinous/ melanoid deposits	Color	Shape	Pores/ mm	Size (µm)	Ornamentation	Description	Size (µm)	Shape	-	Melzer reaction Size (µm)
G. australe M1	Rarely present, mostly sessile	Tubercular, rugose, E concentric zonate, brownish to mate	Brownish, flat to slightly lobulated	Homogeneous, chocolate	Always present	Whitish to brownish	Circular to irregular	4-6 6	8.3- 10.3 × 5- 6.6	Thick, free to sub free pillars	In $10-13$ context, final $\times 9$ -or 11 intercalary, globose to slightly ellipsoid, $10-13$	10–13 × 9– 11	Branches of terminal skeletal hyphae embedded in melanoid substance	e de f	f Null 30- reaction 70 × 3- ed in 5 e
<i>G. australe</i> M2 Rarely pres mos sess	. Rarely present, mostly sessile	Concentric zonate, B brownish to greyish	Brownish, flat to slightly lobulated	Homogeneous, chocolate	Always present	Whitish to brownish	Circular to irregular	4-6	6.5- 10.1 × 4.2- 6.1	Thick, free to sub free pillars	uouute- walled. In context, very 11 × 9 rare, slightly ellipsoid, double- walled.	11 × 9	Branches of terminal skeletal hyphae embedded in melanoid	a f	f Null 5 reaction ed in
G. dorsale	Always present, lateral, vertical	Rough, semi-concentric F zonation, wine purplish to reddish	Reddish, flat to acute	Not fully homogeneous, first light brown becoming dark brown near the	Always present	Whitish to brownish	Circular to irregular	3-2 2		Distinct, thick pillars, sometimes with anastomosed	In context, rare, 5–13 × late or 4– intercalary, 7.5 double- walled.	,5-13 × 4- 7.5	substance Cylindrical to clavate, usually with apical protuberances (up to 2)	and to	substance lindrical to Strongly 20– clavate, usually amyloid 3; with apical × protuberances 3; (up to 2) 7,
G. martinkense	 G. martinicense Substipitate to sessile 	Tuberculous, rugose, V concentric colored, first purplish, then reddish and yellowish toward the margin the margin	Whitish, lobulated	ubes Not fully alternate dark and light brown	Always present	Whitish to yellowish	Circular to irregular	φ 4	10- 12.9 5.7- 7.7	appearance Distinct, free pillars	In culture, terminal or intercalary, double walled, variably smooth, sometimes with appendages, yellowish. In context, terminal or intercalary.	8- 12.5 × 8- 11 11 9-12 × 5-8	Spheroid pedunculated to clavate, occasionally with apical or basal branches	e, e, cal nnc	Weakly lated amyloid e, ally inches

					Resinous/									
					melanoid			Pores/				Size		Melzer
Taxon	Stipe	Surface	Margin	Color	deposits	Color	Shape	mm	(mn)	(μm) Ornamentation Description	Description	(mŋ)	Shape	reaction Size (µm)
G. mexicanum Present, lateral	Present, lateral	Rough, slightly semi- concentric zonation	Whitish, acute	Mostly light colored Slightly	Present, Whitish to Circular to 4–6 inconsciruous hrowoich irregular	Whitish to hrownish	Circular to irregular	4–6	7-9 × 5-6	$7-9 \times$ Inconspicuous, 5-6 mostly free	In context, 5	9-15.5 × 7_	Cylindrical to clinhtly	Strongly 30 × amvloid 4–
	horizontal	shiny, bordered to		heterogeneous,			in coard) 1	pillars	final or	1	clavate,	9
		reddish		being darker							intercalary,		generally	
				near the tubes							globose to		without	
											slightly		protuberances	ces
											ellipsoid,			
											double-			
											walled.			
											Dextrinoid.			
G. platense	Always	Rough to variably striated, Whitish,		Not fully	Present,	Whitish to Angular	Angular	3-4	7.5–9	7.5–9 Inconspicuous,	In context,	8.3–	Cylindrical to	Strongly 30–
	sessile	semi-orbicular	lobulated	homogeneous,	inconspicuous	brownish			× +	free pillars	infrequent,		slightly	amyloid 50
		zonation, shiny, wine	to acute	first light brown					6.2		terminal or		clavate,	×
		purplish on board		becoming dark							intercalary,	5.2-	sometimes	4-
				brown near the							brownish.		with knots	or 9
				tubes									apical	
													constrictions	SI

Table 3. (Continued).

576 🕒 G. MORERA ET AL.

Table 4. Number of specimens, distribution and host charac	terization (species, native/exotic status and substrate preferences) of
Ganoderma species collected and studied in this work.	
	Host characterization
Total	Substrate

Taxon	Total specimens	Distribution (Departments)	Native	Exotic	Substrate preferences
G. australe M1	13	Lavalleja, Montevideo, Maldonado, Paysandú, Rivera, Rocha, Florida, Colonia, Flores	Scutia buxifolia, Myrsine laetevirens, Alophyllus edullis, Lithraea sp.	Acacia melanoxylon, Gleditsia triachanthos, Tilia sp., Eucalyptus sp., Quercus sp., Salix sp., Grevillea sp.	LT (45%), DT (33%) and S (22%)
G. australe M2	88	Canelones, Lavalleja, Maldonado, Montevideo, Rivera, Rocha, Tacuarembó, Treinta y Tres, Florida, Cerro Largo, Río Negro, Artigas, Colonia, San José	Alophillus edulis, Lithraea sp., Myrcianthes sp., Scutia buxifolia, Myrsine laetevirens	Acacia longifolia, Acacia melanoxylon, Acacia sp., Eucalyptus globulus, Robinia pseudoacacia, Prunus persica, Pinus pseudostrobus, Quercus suber, Salix sp.	DT (46%), LT (39%), S (14%) and R (1%)
G. dorsale	27	Montevideo, Rocha, Treinta y Tres, Canelones, Rivera	Alophyllus edulis, Lithraea sp., Ruprechtia salicifolia, Scutia buxifolia, Celtis sp., Sebastiana sp.	Quercus suber	R (58%), LT (37%) and S (5%)
G. martinicense	10	Canelones, Montevideo, Paysandú, Rocha, Artigas	Tipuana tipu	Acacia sp.	R (87%) and S (13%)
G. mexicanum	3	Rocha, Paysandú	Unidentified wood	-	DT (100%)
G. platense	22	Canelones, Montevideo, Maldonado	-	Acacia dealbata, Fraxinus sp., Platanus acerifolia, Salix sp., Quercus sp., Araucaria sp.	LT (85%), DT (8%) S (7%)

LT = stem of living tree, DT = dead trunks, S = stumps, R = soil arising from roots of living or dead trees.

reaction and spheroid-pedunculated shape. On the other hand, *G. platense* basidiomata are smaller, semiorbicular zonate, homogeneous colored and always sessile. Its context is also different, presenting extremely inconspicuous melanoid lines and microscopically characterized by particular cutis elements with apical constrictions (Figure 2).

Ganoderma mexicanum produces stylized, flabelliform, laterally and horizontally stipitate basidiomata characterized by an almost light-colored context and basidiospores with fine ornamentation, whereas *G. dorsale* produces stylized, spatuliform, shellshaped and almost laterally and vertically stipitate basidiomata with no homogeneous distinctive context. Its basidiospores present notorious rough ornamentation and almost cylindrical cutis cells with a strong amyloid reaction (Figure 2).

Specimens with non-laccate surfaces are morphologically very similar, discernible however by their pilear surface, which is distinctly tubercular in *G. australe* M1 and distinctly zonate in *Ganoderma australe* M2.

Herbaria specimens showed a morphology consistent with the species recorded in this study. In that sense, *G. lucidum* is a name previously used for specimens corresponding to the *G. dorsale, G. martinicense, G. platense* and *G. mexicanum* species. The name *G. resinaceum* was previously used to name specimens corresponding to *G. platense* and *G. martinicense*. The names *G. lipsiense, G. applanatum* and *G. marmoratum* were used to refer to specimens of *G. australe* in the broad sense (from now on sensu lato or s.l.). A morphological key for *Ganoderma* species found in Uruguay is presented below.

Host relationships of each Ganoderma species in Uruguay and presence in distinct departments are presented in Table 4. Ganoderma martinicense and G. australe were found alternatively in several departments growing on native or exotic trees. On the other hand, G. platense was only recorded in the southeast (Montevideo, Canelones and Maldonado departments), growing preferentially on exotic trees, G. mexicanum was only found on dead stems of native forest, and G. dorsale, almost exclusively on native trees. Regarding their substrate preferences, G. mexicanum and G. australe M2 were found preferentially on dead stem wood (100% and 46%, respectively); G. martinicense and G. dorsale, preferentially on roots (87% and 58%, respectively); and G. australe M1 and G. platense, preferentially on live stems (45% and 85%, respectively).

Phylogenetic analyses

A total of 53 new sequences were generated from Uruguayan specimens. The dataset alignment resulted in 140 DNA sequences comprising 639 bp. The best evolutionary models for each partition were as follows: K80 + G (ITS1), JC (5.8 S), K80 + G (ITS 2). The partition scheme was K80 + G (ITS1) with -lnL = 1373.0169, and equal base frequencies as follows: A = 0.25, C = 0.25, G = 0.25, T = 0.25, JC (5.8 S) with -lnL = 300.8947 with equal base frequencies, and K80 + G (ITS 2) with 1498.7406 and equal base frequencies. The Bayesian Inference consensus tree is presented in Figure 3.

A total of 19 supported clades were recovered through a phylogenetic analyses (Figure 3). Sequences corresponding to Uruguayan specimens

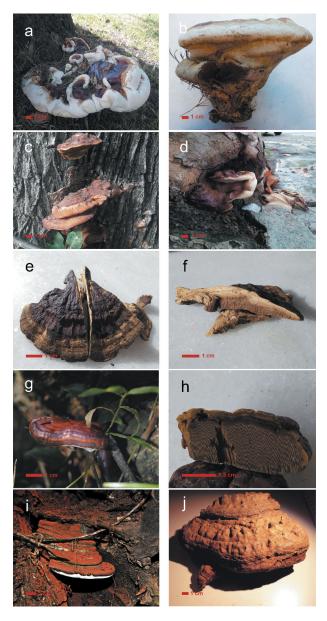


Figure 1. Basidiomata of *Ganoderma* species from Uruguay. *G. martinicense*: (a) (MVHC 5583) and (b) (MVHC 5635), *G. platense*: (c) (MVHC 5586) and (d) (MVHC 5687), *G. mexicanum* (MVHC 5652): (e) and (f), *G. dorsale*: (g) (MVHC 5648) and (h) (MVHC 5655), *G. australe* M2 (MVHC 5587): (i) *G. australe* M1 (MVHC 5582): (j).

were distributed in six terminal clades representing the following species (BPP/BS): Ganoderma platense Speg. (1/98), Ganoderma dorsale (Lloyd) Torrend (1/100), G. martinicense Welti & Courtec. (0.98/80),G. mexicanum Pat. (1/100) and Ganoderma australe s.l. composed of two terminal clades: one of them termed clade 1 (0.97/73), composed of specimens of G. australe M1 and the other one termed clade 2 (0.91/68) and composed of specimens of G. australe M2. The remaining clades represent next species: G. multipileum Hou. (0.96/97), G. multiplicatum (Mont.) Pat. (1/90), "G. lucidum" sensu authors [22] (0.76/94),G. tuberculosum Murrill (1/100), G. concinnum Ryvarden (1/100), G. lucidum (1/100), G. resinaceum

(0.9/59), *G. polychromum* Murrill (1/96), *G. sessile* (0.99/82), *G. zonatum* Murrill (1/100), *G. ecuadoriense* W.A. Salazar, C.W. Barnes & Ordoñez (1/100) and two unnamed taxa of the *G. australe/applanatum* complex from the Neotropic and Asia (1/89) and the Northern Hemisphere (1/88).

Key to Ganoderma species from Uruguay

- 3. Pores 4–6/mm, basidiospores with thick endosporic pillars...... *G. martinicense*
- 3'. Pores 3–4/mm, basidiospores with thin and tiny endosporic pillars...... *G. platense*
- 4[°]. Horizontally stipitate basidiomata, cream colored context and basidiospores with tiny endosporic pillar...... *G. mexicanum*

Discussion and integrative taxonomy

The taxonomic status of Ganoderma species in Uruguay was evaluated through ITS-based phylogenetic analyses in combination with morphological and ecological data. The topology recovered in our phylogenetic analyses is congruent with previous works [4-6]. Three main clades were recovered. One of them is composed of Ganoderma species with reddish laccate basidiomata, stipe and pilear surface, including traditional G. lucidum and G. resinaceum complexes (0.99/67). The second one is composed of almost-sessile Ganoderma species with non-laccate pilear surface, including G. australe and G. applanatum complexes (1/76). The third one is a small subclade of 2 reddish laccate species includina G. zonatum and G. ecuadoriense (0.96/66). Studied Uruguayan

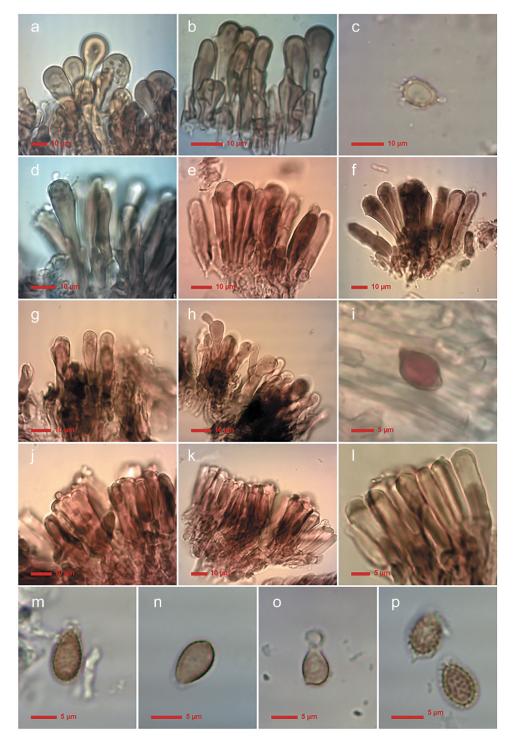


Figure 2. Microscopic features of laccate *Ganoderma* species from Uruguay. Pilear cells: *G. martinicense* (MVHC 5635): (a,b), *G. platense* (MVHC 5687): (d–f), *G. mexicanum* (MVHC 5652): (g–h), *G. dorsale* (MVHC 5655): (j–l). Chlamydospores from fresh *G. martinicense* (MVHC 5635) culture in MEA: (c) and dextrinoid characteristic chlamydospores from the context of *G. mexicanum* (MVHC 5652): (i). Basidiospores from *G. martinicense* (MVHC 5635): (m), *G. platense* (MVHC 5687): (n), *G. mexicanum* (MVHC 5652): (o) and *G. dorsale* (MVHC 5655): (p).

Ganoderma specimens are distributed in six clades representing species whose taxonomic resolutions are discussed below.

Ganoderma martinicense is characterized by developing short stipitate basidiomata of large dimensions (up to 30 cm upper view), with the pilear surface concentrically zonate, with melanoid incrustations in the context, pores 5–6/mm, basidiospores measuring 10– $12 \times 5-7$ µm and non-amyloid pear-shaped to shortly

cylindrical cutis cells. The species is distributed from SW North America through the Caribbean and the Neotropical Atlantic Rain Forests, [11,56], reaching Uruguay and NE Argentina. Ten of the studied specimens fit very well with that morphological description. Moreover, they are grouped with *G. martinicense* specimens, including the type in a strongly supported clade (1/88). Other names were used to label sequences of specimens that are conspecific with sequences of this

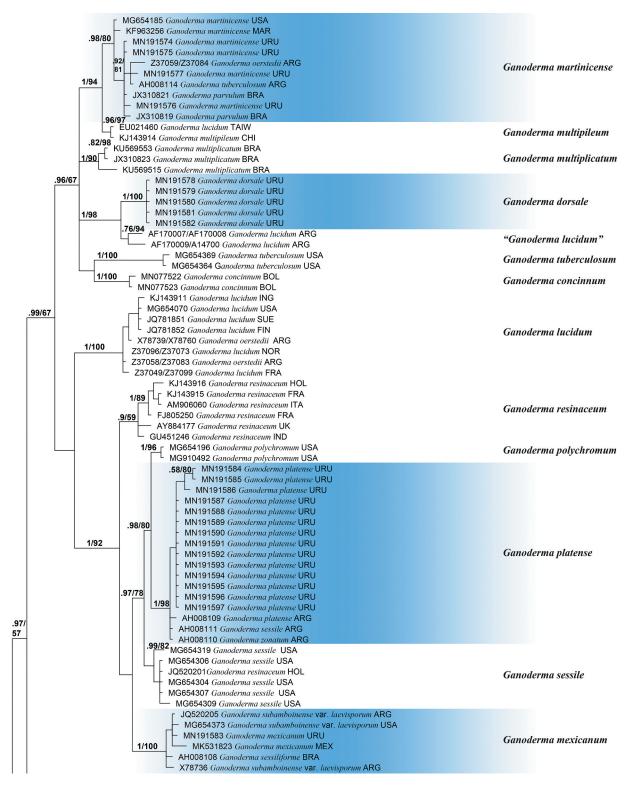


Figure 3. Consensus Bayesian Inference tree based on ITS sequence data used for positioning *Ganoderma* species found in Uruguay (highlighted in light blue). Values of Bayesian and Maximum likelihood are presented as PPB/BS. *Cristataspora coffeata* (FLOR 50933) and *Foraminispora rugosa* (FLOR 52191 and HUEFS DHCR560) were used as outgroup.

lineage (Figure 3), i.e. *G. tuberculosum* [7], *G. oerstedii* (Fr.) Murrill [13] and *G. parvulum* Murrill [57]. *Ganoderma tuberculosum* and *G. parvulum* form two different, distant and unrelated lineages [6,11]. *Ganoderma oerstedii* has been described as presenting a context lacking resinous lines, longer basidiospores

with semi-rough columnar ornamentations: 12– 15 \times 8–10 µm [12] and 9–14 \times 6–9 µm [19]. The pileipellis is formed by irregular, lobed and branched cells with up to seven short, wide protuberances [25]. Sequences of specimens from the type locality (San Juan de Puerto Rico) need to be included to determine

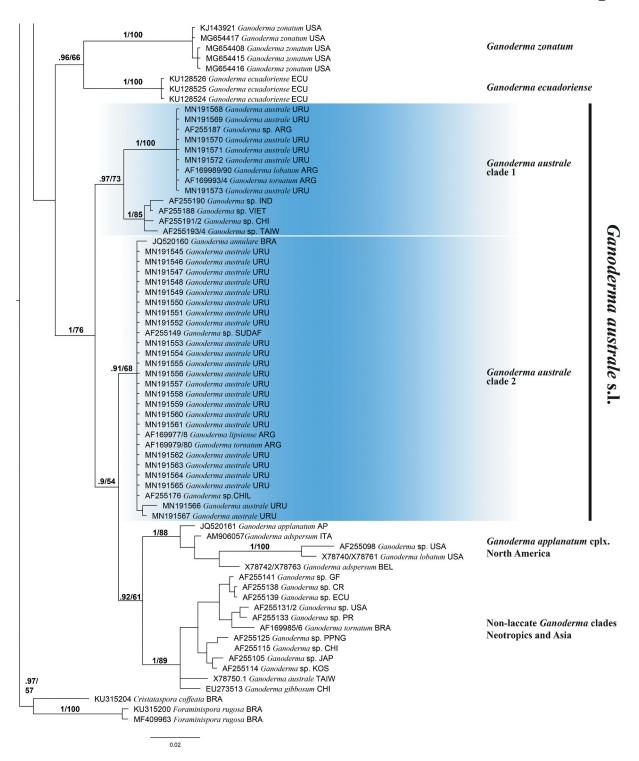


Figure 3. Continued.

the taxonomic status and the phylogenetic relationships of *G oerstedii*. The closest relative of *Ganoderma martinicense* is *G. multipileum* (1/94), an Asian species with small pores: 6–8/mm [9].

Five sequences of Uruguayan specimens are grouped together in a distinct, well-supported clade (1/100), unrelated to any available sequence. Specimens of this clade are characterized by a stylized basidiomata, laterally to centrally slender stipite, shiny reddish orange to violet colored radial and concentric zonate pilear surface, and melanoid bands in the context. Microscopically, the cutis is composed of clavate cells (Figure 2) and basidiospores with thick, often anastomosed endosporic ornamentations. The macro-morphological features suggest some Neotropical taxa including Ganoderma elegantum Ryvarden [12], G. concinnum [23] and G. dorsale [58]. Ganoderma elegantum is characterized by pores 6–7/mm, branched cutis cells (with up to six branches) and basidiospores with thin endosporic ornamentations [12]. Ganoderma concinnum. recorded in Colombia and Bolivia, is characterized by the formation of basidiomata with slender stipes of up to 20 cm long and it forms a different phylogenetic lineage [12,53]. Ganoderma dorsale (= G. lucidum var. dorsale in accordance with Torres-Torres et al. [25]) was described from Río Grande Do Sul in southern Brazil [59]. The macro and micro morphological characteristics of the specimens of this lineage agree with the description of the type specimen of G. dorsale [22,25] and fit well with the studied specimens: basidiomata shape and size, presence of melanoid lines in the context, basidiospores' shape and size, chemical reactions and shapes of pileipellis cells. Thus, considering morphology and distribution, Ganoderma dorsale is the most suitable name for the new clade of Uruguayan specimens. The closest relative is a phylogenetic clade named "G. lucidum" from NW Argentinean Yungas [7]. More sequences and morphological reassessments are necessary to evaluate the taxonomic status of the "G. lucidum" clade sensu Gottlieb et al. [7].

Ganoderma multiplicatum, also occurring in the Neotropics, is morphologically differentiated by its cutis cells with numerous protuberances [25,57,60,61]. The phylogenetic relationship between *G. multiplicatum*, *G. martinicense*, *G. multiplieum*, *G. dorsale*, "*G. lucidum*", *G. tuberculosum* and *G. concinnum* remains unsolved.

Ganoderma platense is characterized by developing sessile basidiomata, usually imbricated, with a semiorbicular zonate pilear surface context, usually with inconspicuous and discontinuous melanoid deposits. thin dissepiments and large pores (2-4/mm), pileipellis formed by cylindrical to slightly claviform elements, with apical constrictions and basidiospores measuring $9-13 \times 5-8$ µm, with thin, endosporic ornamentations [7,22,62]. The morphological characteristics of the studied specimens constituting this clade (Figures 1 and 2) agree with the description of G. platense and this name is hereinafter applied to the clade. Originally described from Buenos Aires (Argentina), it is currently known for growing on Platanus acerifolia trees of urban ecosystems of its type locality and on stumps of gallery forests of the Parana and Uruguay rivers [7,22,62]. Within the G. platense clade, two sequences correspond to specimens labelled as G. sessile and G. zonatum [7]; however, both species were phylogenetically circumscribed to distinct North American clades and are characterized by growing on hardwoods wood and monocots, respectively [11]. Ganoderma platense is grouped in the so called "resinaceum clade", together with G. resinaceum s.l. in Cabarroi-Hernández et al. [6], G. polychromum and G. sessile sensu Loyd et al. [11]. Ganoderma resinaceum encompasses North American, European and Asian populations, including more than one phylogenetic species. Nevertheless G. resinaceum sensu auctores from Eurasian descriptions present cutis cells with laterally diverticulate branches [22,25,63,64]. It contrasts with G. platense, which presents cylindrical to slightly clavate cells with apical constrictions. Ganoderma polychromum (Copel.) Murrill is distributed in North America, growing

on hardwoods and characterized by a context without melanoid deposits, smaller pores (4–5 mm) and larger basidiospores measuring $10.8-13.2 \times 6-7.5 \mu m$ [11].

Ganoderma mexicanum is known for its occurrence in Brazil, Colombia, Costa Rica, Cuba, French Guiana, Mexico, Nicaragua, South–eastern USA (Florida) [6], and now reported in Uruguay.

Morphologically, the studied specimen presents the diagnostic characters of light-colored context with variably abundant dextrinoid, smooth chlamydospores [6].

The sequences of non-laccate Ganoderma specimens collected in Uruguay were distributed in two clades: one of them with Southern Hemisphere specimens and the other one with specimens from South America and Asia [5]. Although the morphological analysis did not allow us to discriminate the specimens of each clade, the pilear surface generally appears tubercular and non-zonate in G. australe M1 and concentrically zonate G. australe M2. In addition, there seems to be ecological differences regarding host preferences: G. australe M1 specimens develop basidiomata on living hosts, whereas G. australe M2 on dead hosts. It was established that species of the G. australe/ applanatum complexes could have a recent origin (not earlier than 30 Ma), with a distribution pattern explained by a large-scale, episodic colonization model and subsequent distance isolation [5]. The recent origin, in addition to the remaining interfertility between specimens from both clades [5], may be the plausible explanation of the crypticity and lack of clear morphological differences for the specimens of the two clades of G. australe. Many names were proposed for sequence of specimens grouped in each clade: G. tornatum (Pers.) Bres., G. lobatum (Schwein.) G.F. Atk., G. annulare (Lloyd) Boedijn and G. lipsiense [7]. Ganoderma annulare and G. tornatum are largely considered synonyms of G. australe, and Ganoderma lipsiense was long considered a synonym of G. applanatum [21,25]. Ganoderma applanatum type locality is in the Northern Hemisphere and previous phylogenetic analyses suggested that it could be represented by a Northern Hemisphere clade [5.8]. Morphologically, G. applanatum lacks melanoid lines in the context [25], whereas all Uruguayan specimens present melanoid elements in the context. From the morphological, ecological and distribution data, M1 and M2 specimens should remain as G. australe s.l.

In this context, the previous records of *Ganoderma* in Uruguay (Table 1) should be questioned, and due to the absence of herbarium specimens, only speculations can be made in relation to published descriptions. Records of *G. lorenzianum* [65,66] have characteristics that resemble *G. mexicanum* due their morphological similarities regarding light colored context (picture in reference [67]), stipe and ovoid, smooth basidiospores (9–10 \times 6–7 µm). *G. lorenzianum* is an

earlier name than G. mexicanum but the description and picture offered in bibliography [67] and the low number of specimens of G. mexicanum analyzed in Uruguay is insufficient to assess the epitypification of these species. Focused studies on specimens corresponding to this species are urgently needed. In Uruguay, Ganoderma orbiforme (Fr.) Ryvarden (=G. fornicatum (Fr.) Pat.) was recorded by Felippone [66], but it currently forms a different phylogenetic clade [57] and no specimens or sequences corresponding to this species were recorded in the country. Ganoderma nitens was recorded by Patouillard [65] characterized by warty basidiospores, $10 \times 7 \ \mu$ m, the type is currently lost [68], so this record could be attributed to G. dorsale, a single reddish laccate Ganoderma species of Uruguay with warty spores.

Conclusions

An integrated comprehensive approach was carried out using morphological, molecular and ecological evidence to assess the diversity of *Ganoderma* in Uruguay. A total of five species (forming six supported clades) were found through the analysis of 163 basidiomata and 53 sequences. Among those species, *G. martinicense, G. mexicanum, G. platense* and *G. dorsale* were confirmed and identified for the first time in Uruguay. Particularly, *G. platense* and *G. dorsale* were first recovered in the phylogenetic analyses. On the other hand, non-laccate specimens were distributed in two clades and so far considered as *G. australe* s.l.

Acknowledgments

The authors kindly acknowledge Anaclara Cabrera Varela for her technical support in improving figures editions. Curators of CORD, MVM and MVHC herbaria are acknowledged for the loan of collections for this study. Daniel Newman, (ORCID: 0000-0002-5400-3691) is kindly acknowledged for discussions, comments and proofreading the English version of the manuscript. The assistance of Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) and Universidad Nacional de Córdoba, both of which supported the facilities used in this project, is also acknowledged. Authorities that granted permits to collect in Uruguay are kindly acknowledged.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This work was supported by Agencia Nacional de Investigación e Innovación [ANII, POS_NAC_2016_1_130911], Pedeciba, Universidad de la República, Uruguay, IDEA WILD, FONCYT [PICT 0830 to G. Robledo], and Fundación Fungicosmos.

ORCID

- G. Morera (D) http://orcid.org/0000-0001-9290-459X
- S. Lupo (p) http://orcid.org/0000-0003-2935-5545
- S. Alaniz (b) http://orcid.org/0000-0002-6530-7279
- G. Robledo (D) http://orcid.org/0000-0002-3840-1104

References

- Karsten PA. Enumeralio boletinearum et polyporearum fennicarum, systemate novo dispositarum. Rev Mycol. 1881;3:16–19.
- [2] Murrill WA. *Tomophagus* for dendrophagus. Torreya. 1905;5:197.
- [3] Moncalvo JM, Ryvarden L. A nomenclatural study of the Ganodermataceae. Oslo (Norway): Fungiflora; 1997.
- [4] Costa-Rezende DH, Robledo GL, Góes-Neto A, et al. Morphological reassessment and molecular phylogenetic analyses of *Amauroderma* s.lat. raised new perspectives in the generic classification of the Ganodermataceae family. Persoonia. 2017;39:254– 269.
- [5] Moncalvo JM, Buchanan PK. Molecular evidence for long distance dispersal across the Southern Hemisphere in the *Ganoderma applanatum-australe* species complex (Basidiomycota). Mycol Res. 2008;112(4):425–436.
- [6] Cabarroi-Hernández M, Villalobos-Arámbula AR, Torres-Torres MG, et al. The Ganoderma weberianum-resinaceum lineage: multilocus phylogenetic analysis and morphology confirm G. mexicanum and G. parvulum in the Neotropics. MycoKeys. 2019;59:95.
- [7] Gottlieb AM, Ferrer E, Wright JE. rDNA analyses as an aid to the taxonomy of species of *Ganoderma*. Mycol Res. 2000;104(9):1033–1045.
- [8] Moncalvo JM. Systematics of *Ganoderma*. *Ganoderma* diseases of perennial crops; 2000.
- [9] Wang DM, Wu SH, Su CH, et al. Ganoderma multipileum, the correct name for 'G. lucidum'in tropical Asia. Bot Stud. 2009;50:451–458.
- [10] Coetzee M, Marincowitz S, Muthelo VG, et al. Ganoderma species, including new taxa associated with root rot of the iconic Jacaranda mimosifolia in Pretoria, South Africa. IMA Fungus. 2015;6(1):249– 256.
- [11] Loyd AL, Barnes CW, Held BW, et al. Elucidating "lucidum": distinguishing the diverse laccate *Ganoderma* species of the United States. PloS One. 2018;13(7): e0199738.
- [12] Ryvarden L. Neotropical polypores: part 1: introduction, Ganodermataceae & Hymenochaetaceae. Oslo (Norway): Fungiflora; 2004.
- [13] Moncalvo JM, Wang HF, Hseu RS. Gene phylogeny of the *Ganoderma lucidum* complex based on ribosomal DNA sequences. Comparison with traditional taxonomic characters. Mycol Res. 1995;99(12):1489– 1499.
- [14] Cao Y, Wu SH, Dai YC. Species clarification of the prize medicinal *Ganoderma* mushroom "Lingzhi". Fungal Divers. 2012;56(1):49–62.
- [15] Papp V. Global diversity of the Genus Ganoderma. In: Sridhar KR, and Deshmukh SK, editors. Advances in Macrofungi: diversity, ecology and biotechnology. 10–33. CRC Press; 2019 Jan 30.

- [16] Glen M, Bougher NL, Francis AA, et al. *Ganoderma* and *Amauroderma* species associated with root-rot disease of *Acacia mangium* plantation trees in Indonesia and Malaysia. APP. 2009;38(4):345–356.
- [17] Rajchenberg M, Robledo G. Pathogenic polypores in Argentina. Forest Pathol. 2013;43(3):171–184.
- [18] Urcelay C, Robledo G, Heredia F, et al. Hongos de la madera en el arbolado urbano de Córdoba. Córdoba (Argentina): Instituto Multidisciplinario de Biología Vegetal (UNC-CONICET); 2012.
- [19] Bazzalo ME, Wright JE. Survey of the Argentine species of the *Ganoderma lucidum* complex. Mycotaxon. 1982;16:295–325.
- [20] Gilbertson RL, Ryvarden L. North American polypores 1–2. Oslo (Norway): Fungiflora; 1986–1987.
- [21] Gottlieb AM, Wright JE. Taxonomy of *Ganoderma* from Southern South America: subgenus Elfvingia. Mycol Res. 1999;103:1289–1298.
- [22] Gottlieb AM, Wright JE. Taxonomy of Ganoderma from Southern South America: subgenus Ganoderma. Mycol Res. 1999;103:661–673.
- [23] Ryvarden L. Studies in neotropical polypores 2: a preliminary key to neotropical species of *Ganoderma* with a laccate pileus. Mycologia. 2000;92 (1):180–191.
- [24] Torres-Torres MG, Guzmán-Dávalos L. The morphology of *Ganoderma* species with a laccate surface. Mycotaxon. 2012;119(1):201–216.
- [25] Torres-Torres MG, Guzmán-Dávalos L, de Mello Gugliotta A. *Ganoderma* in Brazil: known species and new records. Mycotaxon. 2013;121(1):93–132.
- [26] Torres-Torres MG, Ryvarden L, Guzmán-Dávalos L. Ganoderma subgénero Ganoderma en México. Rev Mex Micol. 2015;41:27–45.
- [27] Brazeiro A. Eco-regiones de Uruguay: biodiversidad, presiones y conservación: aportes a la Estrategia Nacional de Biodiversidad. Montevideo (Uruguay): Facultad de Ciencias, UDELAR; 2015.
- [28] Gazzano S. Notas sobre Basidiomycetes xilófilos del Uruguay. X. Hongos Aphyllophorales de la Región E y NE (Departamentos de Cerro Largo, Rivera, y Treinta y Tres). Comun Bot Mus Hist Nat Montevideo. 2001;6 (119):1–10.
- [29] Gazzano S. Notas sobre Basidiomycetes xilófilos del Uruguay. XIII. Aphyllophorales (Basidiomycota, Opistokonta) de la región Litoral Oesteeste y Noroeste de Uruguay. Comun Bot Mus Hist Nat Montevideo. 2010;138(7). 1–16.
- [30] Martínez Kopp S. Comunidades de Basidiomycetes lignícolas en bosques nativos de Uruguay y factores que condicionan su composición. Montevideo (Uruguay): Universidad Nacional de Córdoba; 2014.
- [31] Herter G. Florula Uruguayensis. Plantae Avasculares. Ostenia. Colección de Trabajos Botánicos dedicados a Don Cornelio Osten. 1933;7(84):1–13.
- [32] Gazzano S. Notas sobre Basidiomycetes xilófilos del Uruguay. VIII. Registro de Aphyllophorales y sus sustratos arbóreos. Comun Bot Mus Hist Nat Montevideo. 1998;6(109):1–12.
- [33] Spegazzini C. Fungi Argentini: novi v. critici. 1898;6.
- [34] Wright JE, Bolontrade MF. An undesirable immigrant. Mycologist. 1994;8(1):14–15.
- [35] Xing JH, Song J, Decock C, et al. Morphological characters and phylogenetic analysis reveal a new species within the *Ganoderma lucidum* complex from South Africa. Phytotaxa. 2016;266(2):115–124.

- [36] Richter CWK, Kirk PM, Stadler M. An assessment of the taxonomy and chemotaxonomy of *Ganoderma*. Fungal Diversity. 2015;71(1):1–15.
- [37] Urcelay C, Robledo G. Community structure of polypores (Basidiomycota) in Andean alder wood in Argentina: functional groups among wooddecay fungi? Austral Ecol. 2004;29(4):471–476.
- [38] Thiers B. Index Herbariorum: a global directory of public herbaria and associated staff. New York Botanical Garden's Virtual Herbarium; 2020 [cited 2020 Mar 31]. Available from: http://sweetgum. nybg.org/science/ih/
- [39] Gómez-Montoya N, Rajchenberg M, Robledo GL. Aegis boa (Polyporales, Basidiomycota) a new neotropical genus and species based on morphological data and phylogenetic evidences. Mycosphere. 2017;8(6):1261– 1269.
- [40] Doyle JJ, Doyle JL. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. Phytochemistry. 1987;19:11–15.
- [41] Morera G, Robledo G, Ferreira-Lopes V, et al. South American Fomitiporia (Hymenochaetaceae, Basidiomycota) 'jump on' exotic living trees revealed by multi-gene phylogenetic analysis. Phytotaxa. 2017;321(3):277–286.
- [42] White TJ, Bruns T, Lee SJWT, et al. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: PCR protocols: a guide to methods and applications. New York (NY): Academic Press, Inc.; 1990. p. 315–322.
- [43] Gardes M, Bruns TD. ITS primers with enhanced specificity for basidiomycetes-application to the identification of mycorrhizae and rusts. Mol Ecol. 1993;2(2):113– 118.
- [44] Guglielmo F, Gonthier P, Garbelotto M, et al. A PCR-based method for the identification of important wood rotting fungal taxa within *Ganoderma*, *Inonotus* sl and *Phellinus* sl. FEMS Microbiol Lett. 2008;282 (2):228–237.
- [45] Crous PW, Wingfield MJ, Richardson DM, et al. Fungal Planet description sheets. Persoonia. 2016;36 (316):400–468.
- [46] Zhou LW, Cao Y, Wu SH, et al. Global diversity of the *Ganoderma lucidum* complex (Ganodermataceae, Polyporales) inferred from morphology and multilocus phylogeny. Phytochemistry. 2015;114:7–15.
- [47] Hall TA. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. In: Nucleic acids symposium series. Vol. 41; London: Information Retrieval Ltd., c1979-c2000; 1999. p. 95–98.
- [48] Do CB, Mahabhashyam MS, Brudno M, et al. ProbCons: probabilistic consistency-based multiple sequence alignment. Genome Res. 2005;15 (2):330–340.
- [49] Darriba D, Taboada GL, Doallo R, et al. jModelTest 2: more models, new heuristics and parallel computing. Nat Methods. 2012;9(8):772.
- [50] Ronquist F, Huelsenbeck JP. MrBayes 3: bayesian phylogenetic inference under mixed models. Bioinformatics. 2003;19(12):1572–1574.
- [51] Stamatakis A. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. Bioinformatics. 2006;22(21):2688–2690.

- [52] Miller MA, Pfeiffer W, Schwartz T. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In: 2010 gateway computing environments workshop (GCE); IEEE; 2010 Nov 14. p. 1–8.
- [53] Costa-Rezende DH, Robledo GL, Drechsler-Santos ER, et al. Taxonomy and phylogeny of polypores with ganodermatoid basidiospores (Ganodermataceae). Mycol Prog. 2020;19(8):725 –41. DOI:10.1007/s11557-020-01589-1
- [54] Rambaut A, Drummond AJ, Xie D, et al. Posterior summarization in Bayesian phylogenetics using Tracer 1.7. Syst Biol. 2018;67(5):901.
- [55] Hyde KD, Udayanga D, Manamgoda DS, et al. Incorporating molecular data in fungal systematics: a guide for aspiring researchers. 2013; arXiv preprint arXiv:1302.3244.
- [56] Welti S, Courtecuisse R. The Ganodermataceae in the French West Indies (Guadeloupe and Martinique). Fungal Divers. 2010;43:103–126.
- [57] Correia de Lima Júnior N, Baptista Gibertoni T, Malosso E. Delimitation of some neotropical laccate *Ganoderma* (Ganodermataceae): molecular phylogeny and morphology. Rev Biol Trop. 2014;62(3):1197–1208.
- [58] Torrend C. Les Polyporacées du Brésil. I. Polyporacées stipités. Brotéria, Sér Bot. 1920;18:121–143.
- [59] Lloyd CG. Synopsis of the section Apus of the genus Polyporus. Lloyd Library; 1915. p. 4.

- [60] Bolaños AC, Bononi VLR, Gugliotta de Mello A. New records of *Ganoderma multiplicatum* (Mont.) Pat. (Polyporales, Basidiomycota) from Colombia and its geographic distribution in South America. Check List. 2016;12(4). DOI:10.15560/12.4.1948
- [61] Steyaert RL. Study of some *Ganoderma* species. Bull Jard Bot Belg. 1980;135–186.
- [62] Spegazzini C. Observations on and additions to Argentinian mycology. Bol Acad Nac Ci. 1926;28.
- [63] Mohanty PS, Harsh NSK, Pandey A. First report of Ganoderma resinaceum and G. weberianum from north India based on ITS sequence analysis and micromorphology. Mycosphere. 2011;2(4):469–474.
- [64] Ryvarden L, Gilbertson RL. European polypores: part 1: *abortiporus-Lindtneria*. Fungiflora A/S; 1993.
- [65] Patouillard N. Le genre *Ganoderma*. Bull Soc Mycol France. 1889;5:64–80.
- [66] Felippone F. Contribution á la flore mycologique de l Úruguay. Ann Cryptog Exot. 1928;1(4):338–348.
- [67] Kalchbrenner K. Szibériai és délamerikai gombák: fungi e Sibiria et America Australi [Értekezések a természettudományok köréből]. Budapest: Magyar Tudományos Akadémia Könyvkiadó Hivatala; 1878; 8. p. 16.
- [68] Ryvarden L. Type studies in Polyporaceae 36, species described by E. In: Fries Synopsys Fungorum. Vol. 39; 2019.