

**UCC Library and UCC researchers have made this item openly available.  
Please [let us know](#) how this has helped you. Thanks!**

|                                    |  |
|------------------------------------|--|
| <b>Title</b>                       | Interactions between medications and the gut microbiome in inflammatory bowel disease  |
| <b>Author(s)</b>                   | Eckenberger, Julia; Butler, James C.; Bernstein Charles N.; Shanahan, Fergus; Claesson, Marcus J.  |
| <b>Publication date</b>            | 2022-10  |
| <b>Original citation</b>           | Eckenberger, J., Butler, J. C., Bernstein C. N., Shanahan, F. and Claesson, M. J. (2022) 'Interactions between medications and the gut microbiome in inflammatory bowel disease', <i>Microorganisms</i> , 10 (10), 1963, (18pp). doi: 10.3390/microorganisms10101963   |
| <b>Type of publication</b>         | Article (peer-reviewed)  |
| <b>Link to publisher's version</b> | <a href="http://dx.doi.org/10.3390/microorganisms10101963">http://dx.doi.org/10.3390/microorganisms10101963</a><br>Access to the full text of the published version may require a subscription.  |
| <b>Rights</b>                      | © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license ( <a href="https://creativecommons.org/licenses/by/4.0/">https://creativecommons.org/licenses/by/4.0/</a> ). |
| <b>Item downloaded from</b>        | <a href="http://hdl.handle.net/10468/13841">http://hdl.handle.net/10468/13841</a>  |

Downloaded on 2022-12-08T08:39:11Z



**UCC**

University College Cork, Ireland  
Coláiste na hOllscoile Corcaigh



## Article

# Interactions between Medications and the Gut Microbiome in Inflammatory Bowel Disease

Julia Eckenberger <sup>1,2</sup> , James C. Butler <sup>1,2,3</sup>, Charles N. Bernstein <sup>4,5</sup>, Fergus Shanahan <sup>1,6</sup> and Marcus J. Claesson <sup>1,2,\*</sup>

- <sup>1</sup> APC Microbiome Ireland, University College Cork, T12 YT20 Cork, Ireland  
<sup>2</sup> School of Microbiology, University College Cork, T12 TP07 Cork, Ireland  
<sup>3</sup> The SFI Centre for Research Training in Genomics Data Science, H91 TK33 Galway, Ireland  
<sup>4</sup> Inflammatory Bowel Disease Clinical and Research Centre, University of Manitoba, Winnipeg, MB R3A 1R9 Canada  
<sup>5</sup> Section of Gastroenterology, Department of Internal Medicine, University of Manitoba, Winnipeg, MB R3A 1R9, Canada  
<sup>6</sup> Department of Medicine, University College Cork, T12 AK54 Cork, Ireland  
\* Correspondence: m.claesson@ucc.ie

**Abstract:** In view of the increasing evidence that commonly prescribed, non-antibiotic drugs interact with the gut microbiome, we re-examined the microbiota variance in inflammatory bowel disease (IBD) to determine the degree to which medication and supplement intake might account for compositional differences between disease subtypes and geographic location. We assessed the confounding effects of various treatments on the faecal microbiota composition (16S rRNA gene sequencing) in persons with Crohn's disease (CD; n = 188) or ulcerative colitis (UC; n = 161) from either Cork (Ireland) or Manitoba (Canada) sampled at three time points. The medication profiles between persons with UC and CD and from different countries varied in number and type of drugs taken. Among Canadian participants with CD, surgical resection and overall medication and supplement usage is significantly more common than for their Irish counterparts. Treatments explained more microbiota variance (3.5%) than all other factors combined (2.4%) and 40 of the 78 tested medications and supplements showed significant associations with at least one taxon in the gut microbiota. However, while treatments accounted for a relatively small proportion of the geographic contribution to microbiome variance between Irish and Canadian participants, additive effects from multiple medications contributed significantly to microbiome differences between UC and CD.

**Keywords:** inflammatory bowel disease; gut microbiota; drugs



**Citation:** Eckenberger, J.; Butler, J.C.; Bernstein, C.N.; Shanahan, F.; Claesson, M.J. Interactions between Medications and the Gut Microbiome in Inflammatory Bowel Disease. *Microorganisms* **2022**, *10*, 1963. <https://doi.org/10.3390/microorganisms10101963>

Academic Editor: Konstantinos Triantafyllou

Received: 28 August 2022

Accepted: 30 September 2022

Published: 4 October 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Inflammatory bowel disease (IBD) which comprises Crohn's disease (CD) and ulcerative colitis (UC) have been linked with changes in the gut microbiome [1,2], although cause and consequence have not been disentangled [3,4]. Moreover, no uniform microbiome pattern or signature has been identified consistently across multiple studies [5]. This may be due to the heterogeneity of IBD and/or variations in study design and patient populations [3]. Medications have also been shown to alter the gut microbiome and add an additional layer of confounding factors. Not only antibiotics have a well-described, cumulative long-lasting effect on the intestinal microbiome [6], but also commonly prescribed non-antibiotic medications have been shown to impact the gut microbiome [7,8]. Standard therapies for IBD include anti-inflammatory drugs, such as 5-aminosalicylic acids (5-ASA) and corticosteroids, and immunosuppressants, such as thiopurines, methotrexate and biologics (TNF- $\alpha$  inhibitors and integrin or interleukin receptor antagonists). Several studies have shown that these drugs can impact the microbial composition [9–13], but vice versa, gut microbiota can modulate their pharmacological activity via drug (in)activation

and biotransformation [14]. For instance, bacterial azoreductases release 5-ASA from its prodrugs, while bacterial N-acetyltransferases are responsible for the inactivation of 5-ASA [9]. Aminosalicylates in turn can increase the levels of some Firmicutes while reducing the levels of Bacteroidetes and Proteobacteria [15].

Moreover, commonly used non-IBD drugs can also affect the composition and metabolic function of gut microbiota [16]. In a major *in vitro* study, Maier and colleagues [8] tested the effect of more than 1000 drugs against 40 representative gut bacterial strains and found that approximately a quarter of human targeted drugs inhibited the growth of at least one test strain, with butyrate and propionate producing bacteria being more sensitive and  $\gamma$ -Proteobacteria being more drug-resistant. Pathogens and commensals alike have been shown to be able to metabolize and/or bio-transform a variety of drugs leading to an altered bacterial metabolism and changes in microbial composition due to metabolic cross-feeding and changes in the intestinal microenvironment [17,18]. To this end, a variety of studies have elucidated the microbiota-altering effects of commonly prescribed medications (proton pump inhibitors [19,20], lipid lowering statins [21], laxatives [22], metformin [23], beta blockers [24], ACE inhibitors [25], and SSRI antidepressants [26]). In some cases, the drugs explained more of the microbiota variability than the disease itself [27,28], which bears the question as to what degree this variability is secondary to the disease, or to the drugs treating it.

We recently showed in a large intercontinental twin city study of the microbiome in IBD that geographic location (Ireland vs. Canada) had a major influence on microbiota variance almost equivalent to that of a diagnosis of Crohn's disease itself [29]. While the influence of geography may, in part, be due to cultural and ethnic influences, differences in treatment on either side of the Atlantic may also have a contribution. Therefore, the purpose of the present study was to re-examine and disentangle microbiota variance in IBD to determine the degree to which differences in treatment at different locations might account for the apparent geographic influence. The results confirm that the overall trends of microbiota composition and diversity, as previously reported by us, remain different across IBD-subtypes and geographic location. Only a small part of the effect of geographic location is explained by the differences in medication and supplement intake. However, a large proportion of the disease-associated shift in microbial composition between persons with UC and CD can be explained by additive interaction effects from multiple medications.

## 2. Materials and Methods

The V3-V4 16S rRNA amplicon sequences from our previous study, which had been processed in a single laboratory in Cork utilizing the same protocols, were downloaded and pre-processed as previously described [29], with the exception of the taxonomic classification, which was here performed against the SILVA database (v132) [30] within the mothur suite (v1.39.3) [31] utilizing the `classify.seqs` function with a bootstrap cut-off of 80%. OTUs that fell below that cut-off were assigned as unclassified at that particular rank. Species-level resolution was provided by SPINGO [32] using a similarity score of 0.5 and bootstrap cut-off of 0.8 against the SILVA database (v132). Conflicts between the two methods were resolved by means of BLASTn (v 2.8.1; 10 May 2021) [33]. The raw medication data was classified based on the anatomical therapeutic chemical (ATC) classification system, which hierarchically classifies medications and dietary supplements based on the therapeutic use of their main active ingredient (1st level: anatomical main group; 2nd level: therapeutic subgroup; 3rd level: pharmacological subgroup; 4th level: chemical subgroup; 5th level: chemical substance) [34] and recorded as a qualitative variable. While surgical resection is neither medication nor supplement, it is a common treatment for IBD as 80% of patients with CD require surgery during their lifetime [35]. It also has been shown to significantly alter the microbiome of patients [36] and was therefore included in the analysis as treatment. Long-term dietary habits were captured through frequencies of medium food servings of 157 items via Food Frequency Questionnaires (FFQ) and were summarized into

one factor, the Healthy Food Diversity (HFD) Index, as previously described [29,37]. An active state of IBD was defined as a faecal calprotectin measurement of  $\geq 250$   $\mu\text{g/g}$  [38].

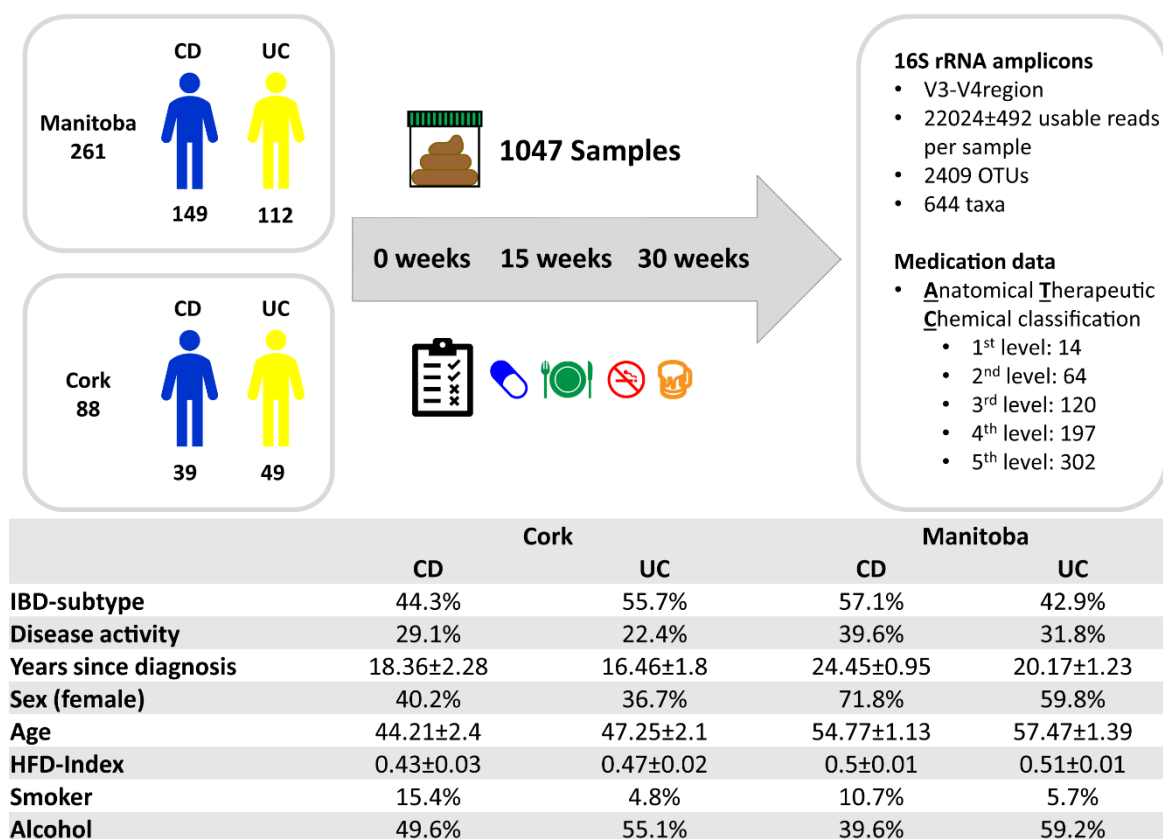
All statistical analyses were performed in the R environment version 4.1.0 and visualizations were produced with the `ggplot2_3.3.5` package [39]. In order to alleviate the constant sum constraint of compositional data [40], zeros were removed from the raw counts via the count zero multiplicative method within the `zCompositions_1.3.4` package [41] and subsequently subjected to a centered log ratio (CLR) transform, i.e.; the data were expressed as logarithms of ratios with the geometric mean as denominator using the `propr_4.2.6` package [42]. Beta diversity (i.e.; between sample diversity) of the microbiome was evaluated via principal component analysis on Aitchison distances [43] within `phyloseq_1.36.0` [44], while `vegan_2.5-7` [45] was used for permutational multivariate analysis of variance between groups. Alpha diversity (i.e.; within sample diversity) was calculated with `iNEXT_2.0.20` [46]. Differential taxa abundance and effect sizes without adjusting for confounding factors was computed via `ALDEx2_1.24.0` [47] using 1000 Monte Carlo samples. Differences in the usage of medications and supplements between the different IBD-subtypes and participants from the two different geographic locations were tested for significance with Fischer's Exact tests. Distance-based redundancy analysis (dbRDA) was performed with the `capscale` function in `vegan_2.5.7` [45] to evaluate the effect of environmental factors on the medication profiles (Jaccard distances) and gut microbial composition (Aitchison distances). Here, medication profiles and microbial abundances constitute the set of response variables, while the environmental factors represent the predictive variables. The proportion of explained, compared to the total fitted variance indicates how much of the variation between samples is due to differences in environmental factors. Selection of the most relevant species and features in the dbRDA was implemented with the `ordiselect` function in `goeveg_0.5.1` [48]. Differences in numbers of used medications as well as the number of changed medications and intra-personal differences in alpha and beta diversity measures were assessed via Wilcoxon rank sum tests.

Explained variance of single covariates in a multivariate data set was computed with the `VpThemAll_0.0.0.9` wrapper [49], which chooses a model via the `ordistep` function that explains most variance in microbiota composition and then looks at each included metadata variable separately, conditioning out the effect of all other included metadata variables using the `varpart` function. The difference between the naïve (shared) effect and the unique effects amounts to the interaction effect. The variation in community data with respect to explanatory tables was calculated with the `varpart` function in `vegan_2.5-7` [45] allowing only permutations within the samples of the same patient to adjust for multiple measurements. Significant associations and effect sizes of covariates with single taxa were computed with the `metadeconfoundR_0.2.8` package [50], specifying "patient" as random variable; *p*-values were adjusted for multiple testing where appropriate, using the Benjamini and Hochberg method [51].

### 3. Results

After extracting from the Clooney et al. [29] data set samples of persons with IBD with all three time points and no missing data, 349 persons were included for further analysis, of which 188 were diagnosed with CD and 161 with UC. The sampling time points were on average  $15.18 \pm 0.42$  weeks apart. Usage information on medication and dietary supplements had been collected for all of the 1047 samples. Of the original 3148 operational taxonomic units (OTUs) that were clustered at  $\geq 97\%$  identity, 2409 remained after filtering, leaving on average  $22,024 \pm 492$  usable reads per sample. Applying the ATC classification system [34] to the raw medication data yielded 302 different chemical substances (5th ATC level). A number of dietary supplements ( $n = 74$ ) had no ATC classifier and were aggregated under "Other supplements". For the analysis, the medications and supplements were then further combined into 120 pharmacological subgroups (3rd ATC level), and drug usage was recorded as a qualitative yes–no variable. Long-term dietary habits were summarized into one factor, the Healthy Food Diversity (HFD) Index, as previously described [29,37].

An active state of IBD was defined as a faecal calprotectin measurement of  $\geq 250$   $\mu\text{g/g}$  [38]. (Figure 1, Supplementary Tables S1 and S2).

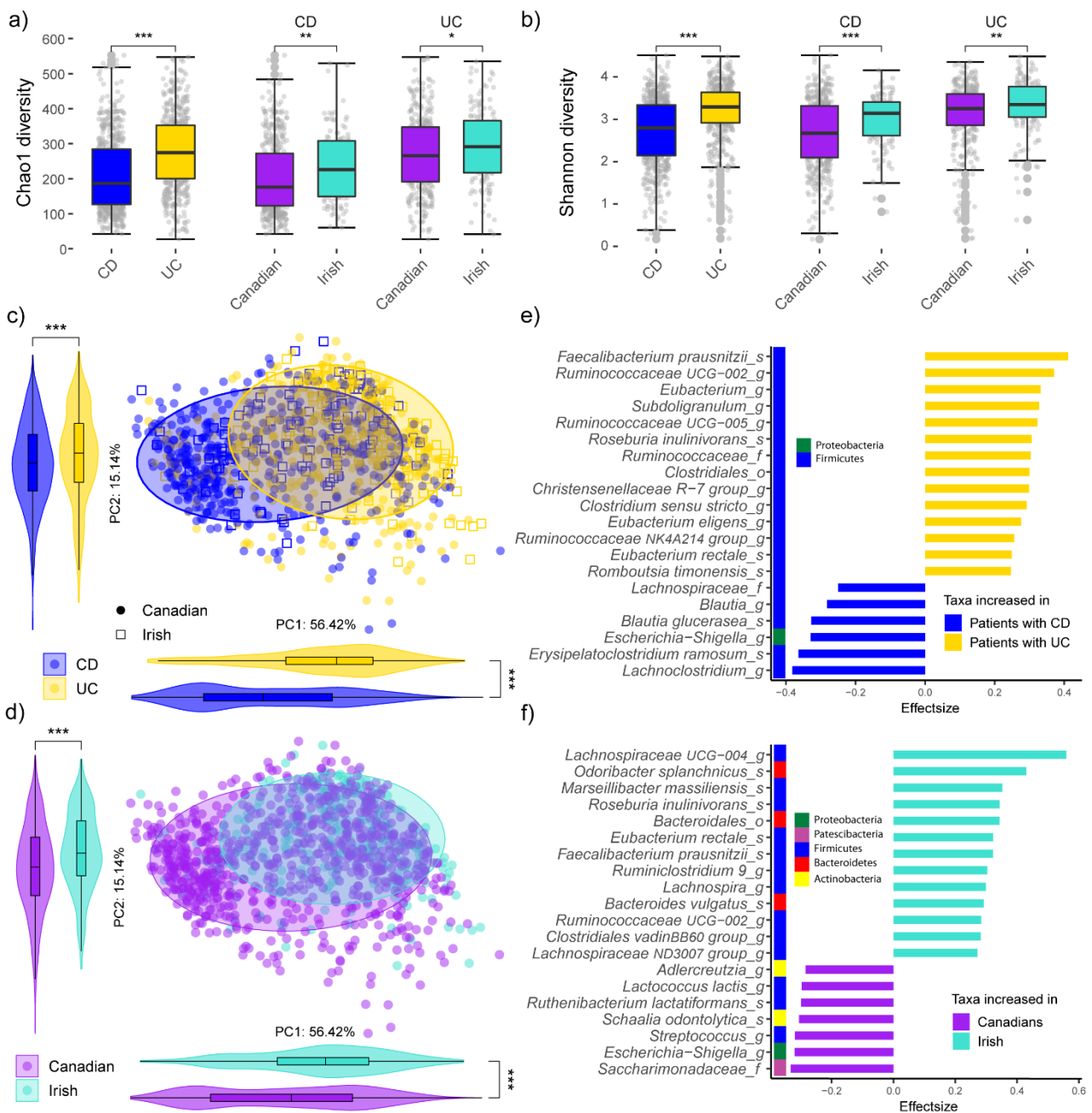


**Figure 1.** Subject characteristics and sample sizes of the study cohort. The ATC classification system classifies medications hierarchically based on the therapeutic use of the main active ingredient: 1st level: anatomical main group; 2nd level: therapeutic subgroup; 3rd level: pharmacological subgroup; 4th level: chemical subgroup; 5th level: chemical substance.

### 3.1. Microbiota Composition Is Different Both between IBD Subtypes and Geographic Locations

In contrast to our original study, here we used a ‘compositionally aware’ analysis approach, which more accurately quantifies taxa without the confounding relative effect of total read count per sample [40]. As previously reported [29], alpha (i.e.; within sample) diversities were lower for all persons with CD compared to UC and also for Canadian participants in general (Wilcoxon  $p < 0.05$ ; Figure 2a,b, Supplementary Table S3). Abundances of OTUs were CLR-transformed, and beta (i.e.; between sample) diversity analysis was calculated from Aitchison distances of all OTUs present in at least 10% of the samples. The first two principal component axes captured a much higher proportion of the microbiota variation in the data set (71.6%), whilst showing the same significance (PERMANOVA  $p < 0.05$ ) for disease and location-associated shifts (Figure 2c,d). Differential taxa abundance analysis was carried out using ALDEx2, initially without accounting for any confounders. Here, the OTUs were aggregated based on their highest known taxonomic classification and filtered for an abundance in at least 10% of samples, resulting in 233 tested taxa. Of these, 108 OTUs were significantly different (Wilcoxon/Welch  $p < 0.05$ ) between persons with UC and CD (Figure 2e) but displayed only weak to moderate effect sizes, ranging from  $-0.38$  to  $0.41$ . *Faecalibacterium prausnitzii*, Ruminococcaceae UCG-002 and *Eubacterium* were the most increased taxa in patients with UC, while *Lachnoclostridium*, *Erysipelatoclostridium* and *Escherichia/Shigella* had the highest abundance in the microbiome of patients with CD (Supplementary Table S4). The effect sizes of the 100 taxa that were significantly different (Wilcoxon/Welch  $p < 0.05$ ) between subjects from Manitoba and Cork were slightly higher

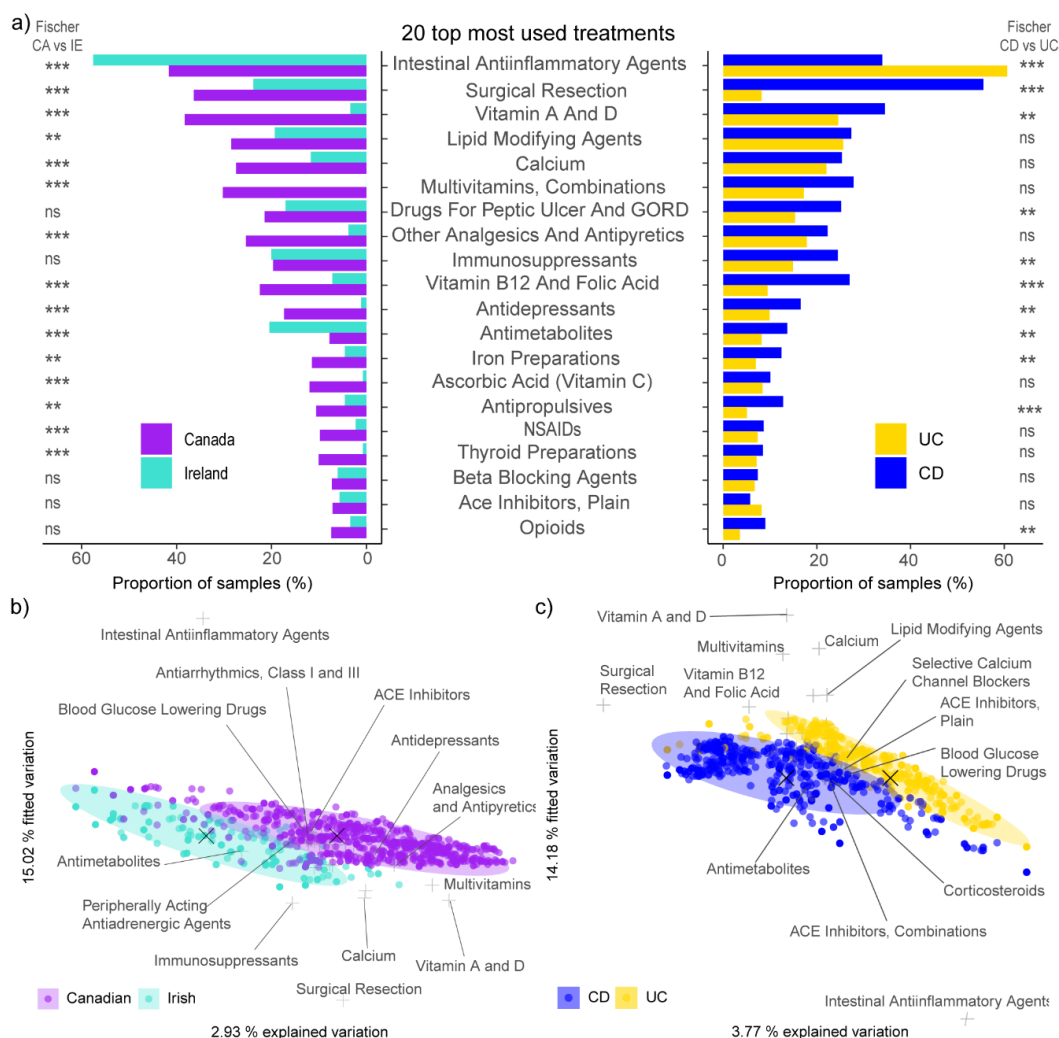
than between the IBD-subtypes (Figure 2f), ranging from  $-0.33$  to  $0.59$ . Members of Saccharimonadaceae, *Streptococcus* and *Ruthenibacterium lactatiformans* were the most enriched taxa in the Manitoba cohort, while Lachnospiraceae\_UCG-004, *Odoribacter splanchnicus* and Bacteroidales were the most increased taxa in the Irish cohort (Supplementary Table S5). Thus, with updated analysis, compared to our previous work [29], the overall trends of microbiota composition and diversity remain different across IBD-subtypes and geographic location.



**Figure 2.** Comparison of (a) Chao1 (species richness) and (b) Shannon diversity (species richness and evenness) between different IBD-subtypes and geographic location. Principal component analysis (PCA) based on Aitchison distances on all operational taxonomic units (OTUs) present in >10% of samples, with samples grouped by: (c) IBD-subtype; and (d) geographic location. Violin plots show projections of the PCA points onto PC1 and PC2. Stars show significant differences between the groups as determined by Wilcoxon test. The top 20 most differential OTU abundances between (e) IBD-subtypes; and (f) geographic location calculated with ALDEx2; \*  $p < 0.05$ ; \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ .

### 3.2. Higher Usage of Resection, Medications and Supplements among Canadian Participants

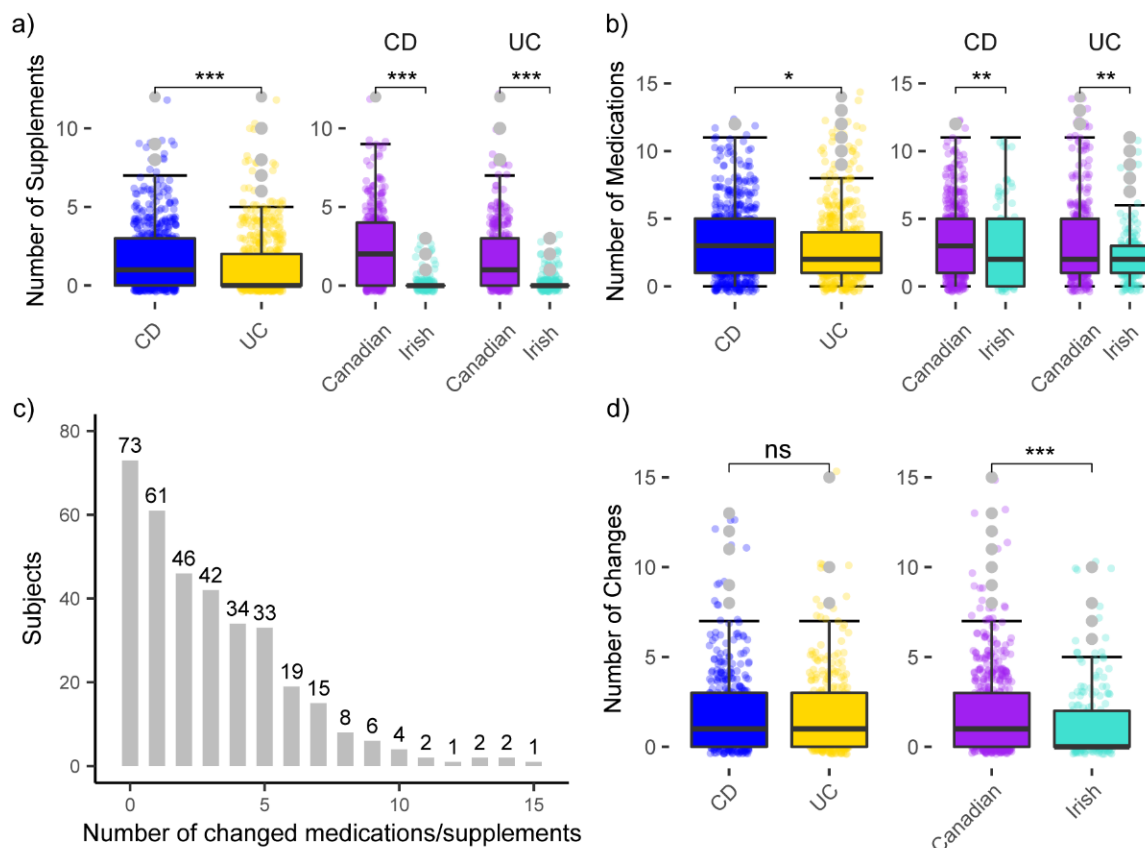
To disentangle the effect of medication on microbiota, we first quantified any significant differences in IBD treatments, both dependently and independently of geography. Of the 120 different pharmacological subgroups found in this study, 78 were recorded at least 5 times as “yes” in total and were included in the downstream analysis (default MetadeconfoundR requirement). We also included surgical resection as a treatment. Of these, 28 medications and supplements showed significantly (Fischer  $p < 0.05$ ) different usage between the Irish and the Canadian cohort (Figure 3a). Between persons with either CD or UC, 18 pharmacological subgroups were differentially (Fischer  $p < 0.05$ ) used. Surgical resection was significantly more present in the Manitoba cohort and in persons with CD. Intestinal anti-inflammatory agents, a subgroup which contains ‘locally acting corticosteroids’ and ‘aminosalicylic acid and similar agents’ were the most commonly used treatments, which were over-represented in persons with UC and Irish participants. The most taken supplements were vitamin A and D, which were over-represented in Canadians with CD (Figure 3a, Supplementary Table S6).



**Figure 3.** Comparison of the 20 most common medications and supplements taken in the study cohort separated by IBD-subtype and geographic location (a). Ordination plot of Jaccard distance-based redundancy analysis (dbRDA) of used medications and supplements constrained by (b) geographic location and (c) IBD-subtype. The points represent samples, crosses represent medications and X represent the centroids of the depicted groups. The 5% of medications with the best axis fit are labelled; Fishers test: ns  $p > 0.05$ ; \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ .

Jaccard distance-based redundancy analysis (dbRDA), which is more suitable for quantifying distances between binary data (e.g.; medication A: TRUE vs. FALSE), was used to evaluate the effect of IBD-subtype and geographic location on medication usage patterns. Both the IBD-subtype ( $F = 33.71; p < 0.001$ ) and geographic location ( $F = 25.96; p < 0.001$ ) showed a significant effect, with a CD/UC diagnosis accounting for 26.6% and location for 19.5% of the explained variation in treatments (Figure 3b,c). Surgical resection and vitamin A and D supplementation were most prevalent in Canadian participants, while intestinal anti-inflammatories best distinguished Irish participants with UC from the other cohorts.

The participant groups from varying locations and with a different disease type not only diverged in their drug usage pattern, but also in the amount of ingested pharmaceutical compounds. Overall, persons with CD took a significantly higher number of supplements and medications (Wilcoxon  $p < 0.05$ ) than persons with UC. Similarly, Canadian participants used a significantly greater number of supplements and medications than their Irish counterparts (Figure 4a,b). Another source of variation between the groups was the frequency with which specific medications were changed over the course of the study. Only 73 of 349 participants did not change their medication regime during the 6 months of study, while another 93 persons changed five or more medications over the three time points (Figure 4c). Unsurprisingly, due to the higher usage of pharmaceutical products in the Canadian cohort, there were also significantly (Wilcoxon  $p < 0.05$ ) more changes of medications in this group, while there were no significant disparities between persons with UC and CD within their cohort (Figure 4d).

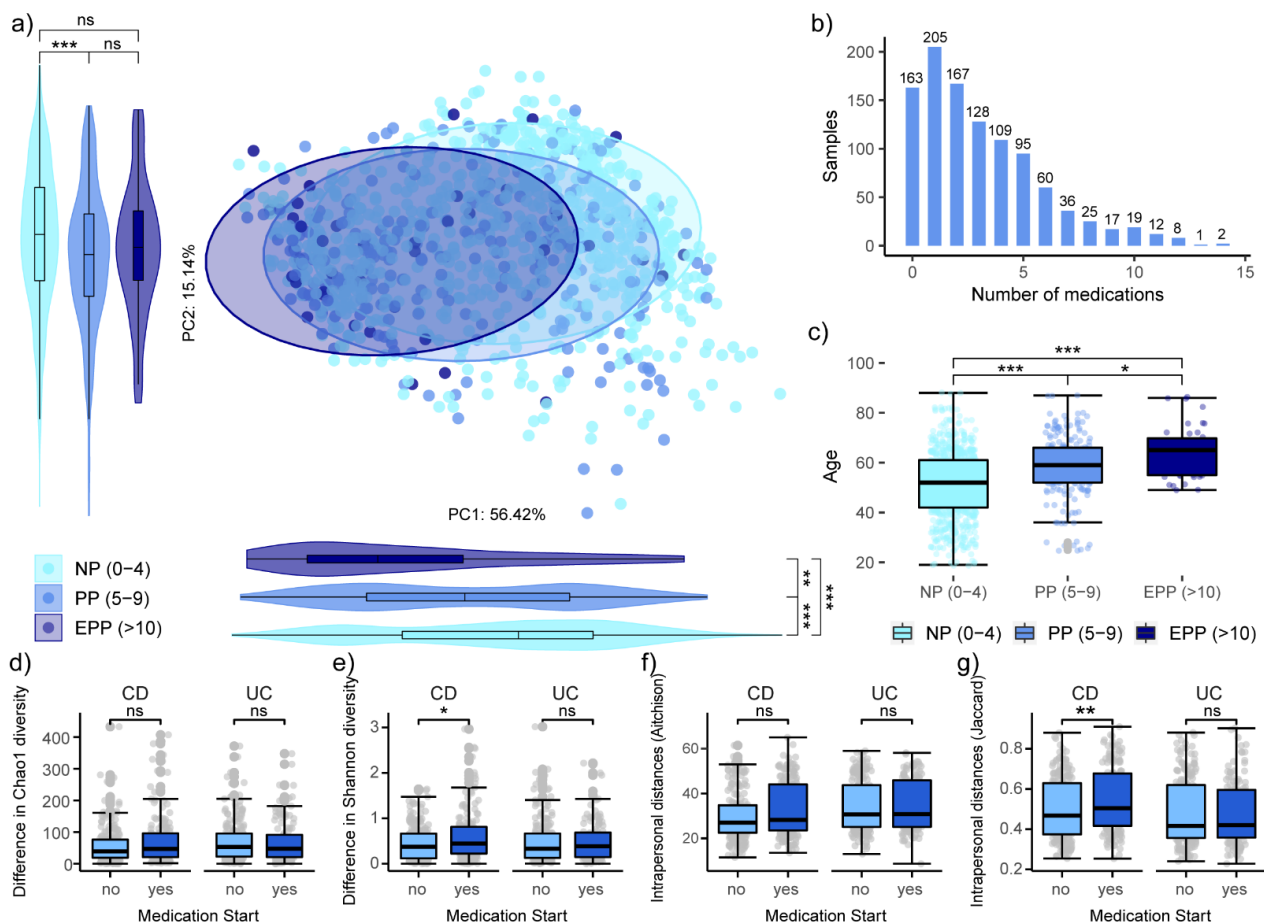


**Figure 4.** Comparison of the amount of (a) supplements and (b) medication taken per person and time point; (c) histogram of changes in medications and supplements with ATC classification per subject over the course of the study; (d) comparison of changes in medications and supplements between persons with differing IBD-subtype and from different geographic locations; Wilcoxon test: ns  $p > 0.05$ ; \*  $p < 0.05$ ; \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ .



### 3.3. Polypharmacy Associated with the Disease-Associated Microbiota Shift in Crohn's Disease

Polypharmacy (PP) is defined as the concurrent and regular use of at least 5 medications and excessive polypharmacy (EPP) as use of 10 or more different drugs. In our study, 120 persons at 233 time points fit in the former category, while 22 persons (42) fit the latter group (Figure 5b). Polypharmacy expectedly increased with age as persons in the EPP and PP group were significantly older than those that took less than five medications on a regular basis (Figure 5c). The microbial composition of persons with EPP, PP and no polypharmacy (NP) showed a gradual shift along the first PC axis (Figure 5a). This corresponded with the shift seen when grouping by IBD-subtype (Figure 2c), which is not surprising as persons with CD used significantly more medications than persons with UC (Figure 4b). Taking multiple medications at the same time can lead to increased side effects and drug interactions that amplify adverse complications [52].



**Figure 5.** (a) PCA based on Aitchison distances with CLR transformed OTUS with a prevalence of at least 10% grouped by polypharmacy. Violin plots show projections of the PCA points onto PC1 and PC2; (b) histogram of the number of medications used per sample; (c) comparison of the age of participants between the polypharmacy groups. Comparison of intra-personal: (d) Chao1 diversity; and (e) Shannon diversity differences, as well as (f) Aitchison; and (g) Jaccard distances. Stars show significant differences between the groups as determined by Wilcoxon test; ns  $p > 0.05$ ; \*  $p < 0.05$ ; \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ .

Since the intake of these pharmaceuticals might influence disease-associated microbiome disturbances, we investigated the influence of changes in the treatment regime on the microbiota. We compared intra-individual alpha and beta diversity differences between the first and second, and second and third time points of participants who did change their medications and those who did not (Supplementary Table S7). Persons with CD who started

new medications between time points displayed significantly higher Shannon diversity differences (Figure 5e) and intra-individual Jaccard distances (Figure 5g). This finding was not reproduced in persons with UC, which might be due to the higher microbial richness compared to persons with CD. Neither disease group showed significant differences in intra-individual species richness (Figure 5d) nor in Aitchison distances (Figure 5f) after starting a new medication. Thus, while presence or absence of certain taxa can be affected by a change in medication, this does not necessarily translate to overall differences in species richness. Additionally, it may have also been important to know when exactly a change in the medication regime has occurred. Unfortunately, this information was not available to us.

### 3.4. Treatment Explains More Variation in Microbial Composition Than IBD-Subtype

Having established differences in both microbiota and treatment between locations, as well as microbiota–polypharmacy associations, we further assessed the overall effect of these treatments on the gut microbiota in more detail. Thus, we performed multivariate regression analysis of the explained microbiota variance (Aitchison distances) of different variables such as medications, patient demographics, environmental factors, and disease status and activity. The full explanatory regression models contained 16 of those variables, which together explained 7.88% of the total variation in microbial composition ( $R^2$ : 0.788; Figure 6a, Supplementary Table S8). This is slightly less than in our original study (9.7%) [29] and is likely due to the fact that here we considered a slightly different number of participants and no control subjects. A conservative estimate of the unique effect of each variable present in the full model was obtained by evaluating all the variables separately, while at the same time partitioning out the effects of all other included covariates. Thus, the difference in explained variation between the naive effect and the unique effect are the interaction effects between the variables. When taken together, all medications explained more microbiota variation than IBD-subtype alone (1.47% vs. 0.25%) but less than surgical resection (1.68%) or geographic location (1.23%). It was also notable that the interaction effects associated with the proportion of variance explained by IBD-subtype were much higher (1.57%) than the interaction effects explained by geographic location (0.50%) (Figure 6a, Supplementary Table S8). Treatment (surgical resection and medications combined) explained more variation (3.48%) than all other clinical and environmental factors together (2.40%) (Figure 6b). The effects of disease remission and intestinal anti-inflammatory agents were positively correlated with UC, while surgical resection, drugs for peptic ulcers and gastro-oesophageal reflux disease (GORD) and antacids as well as EPP and insulins and analogues were positively associated with the effect of CD on the gut microbiome (Figure 6c). In concordance to the previous differential taxa abundance analysis (Figure 2e), *Escherichia/Shigella*, *Klebsiella* and *Blautia* were most associated with CD, whilst a high abundance of several *F. prausnitzii* species characterized persons with UC. Irish participants exhibited a higher abundance of Lachnospiraceae UCG-004 than Canadians, who in turn showed a higher proportion of *Escherichia/Shigella* and *Ruthenibacterium lactatiformans* (Figure 6d).



When clustering the taxa and metadata associations by their effect sizes (Figure 6e), we noticed that the majority of treatments seem to amplify the effect of IBD on the microbiota in so far that many of the associations between treatments and microbiota had the same direction (i.e.; positive or negative) as the associations between IBD and microbiota. As detailed further below, many of the taxa that we found to be increased or decreased in persons with IBD were also correlated with commonly used treatments of these participants. The health-associated taxa *F. prausnitzii*, Lachnospiraceae NK4AI36-group and *Subdoligranulum* [4,5,53,54] were not only depleted in CD compared to UC, but also had the highest number of (negative) correlations with other covariates (21, 20, 17 and 17 associations, respectively). Equally, many taxa that are positively correlated with IBD, such as *Escherichia/Shigella* (n = 18), *Streptococcus* (n = 22), *Klebsiella* (n = 19) and Veillonellaceae (n = 19), as well as bacteria from the oral cavity such as *Rothia dentocariosa* (n = 22), *Fusobacterium nucleatum* (n = 21) or *Oribacterium sinus* (n = 19), were among those that showed the highest number of positive correlations with medication and supplements (Figure 6e). At higher taxonomic ranks, Verrucomicrobiae, Bacteroidia and  $\gamma$ -Proteobacteria were significantly (Fischer  $p < 0.05$ ) less associated with medications than Actinobacteria, Bacilli and Clostridia, among others (Figure 7a, Supplementary Table S10). It was also notable that the Clostridia taxa Ruminococcaceae, Clostridiales vadinBB60 group and unclassified Clostridiales had the highest proportion of negative associations. While Lachnospiraceae were relatively unaffected by medications (Figure 7c), they also showed a relatively high proportion of negative associations (Figure 7d, Supplementary Table S11).

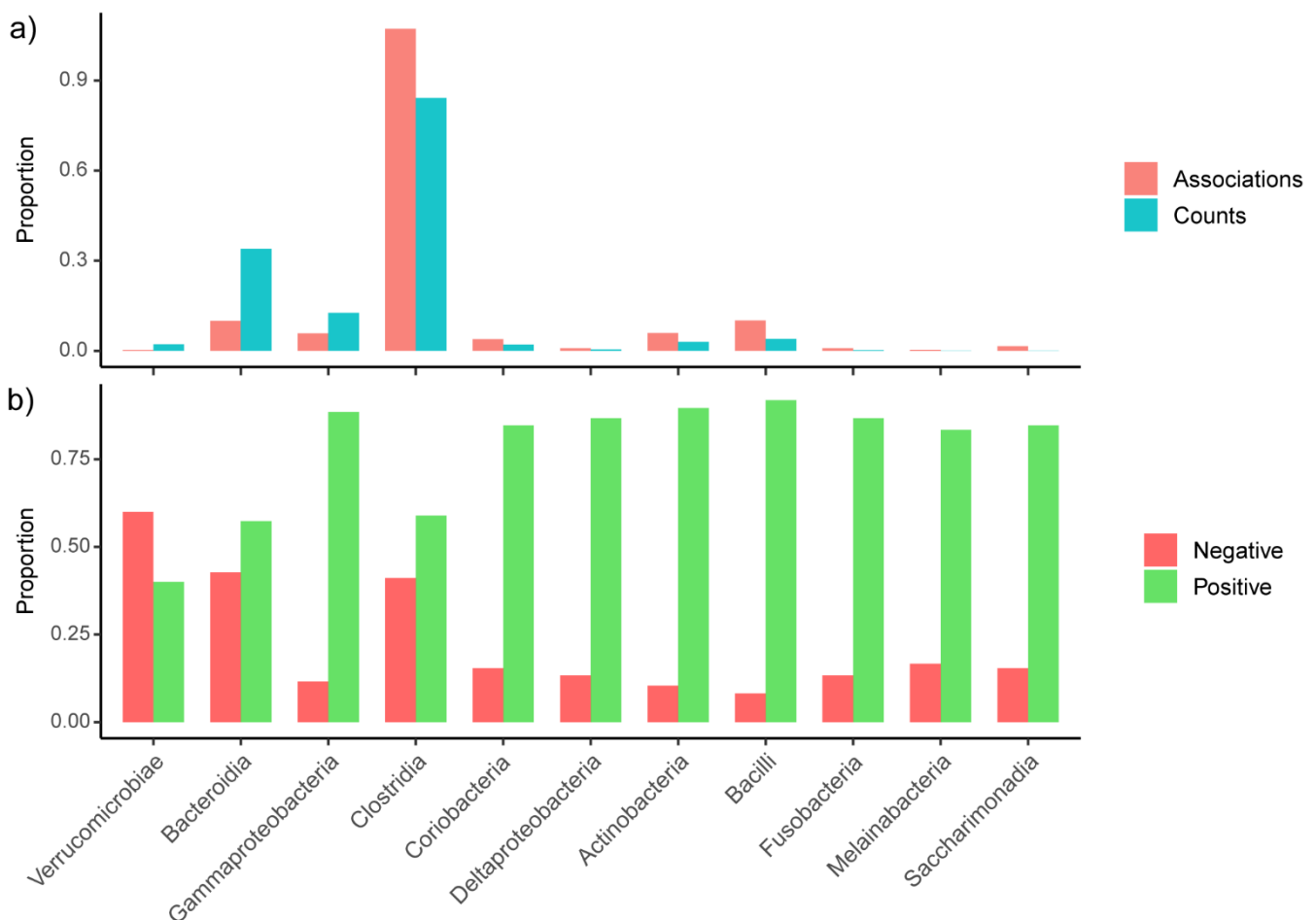
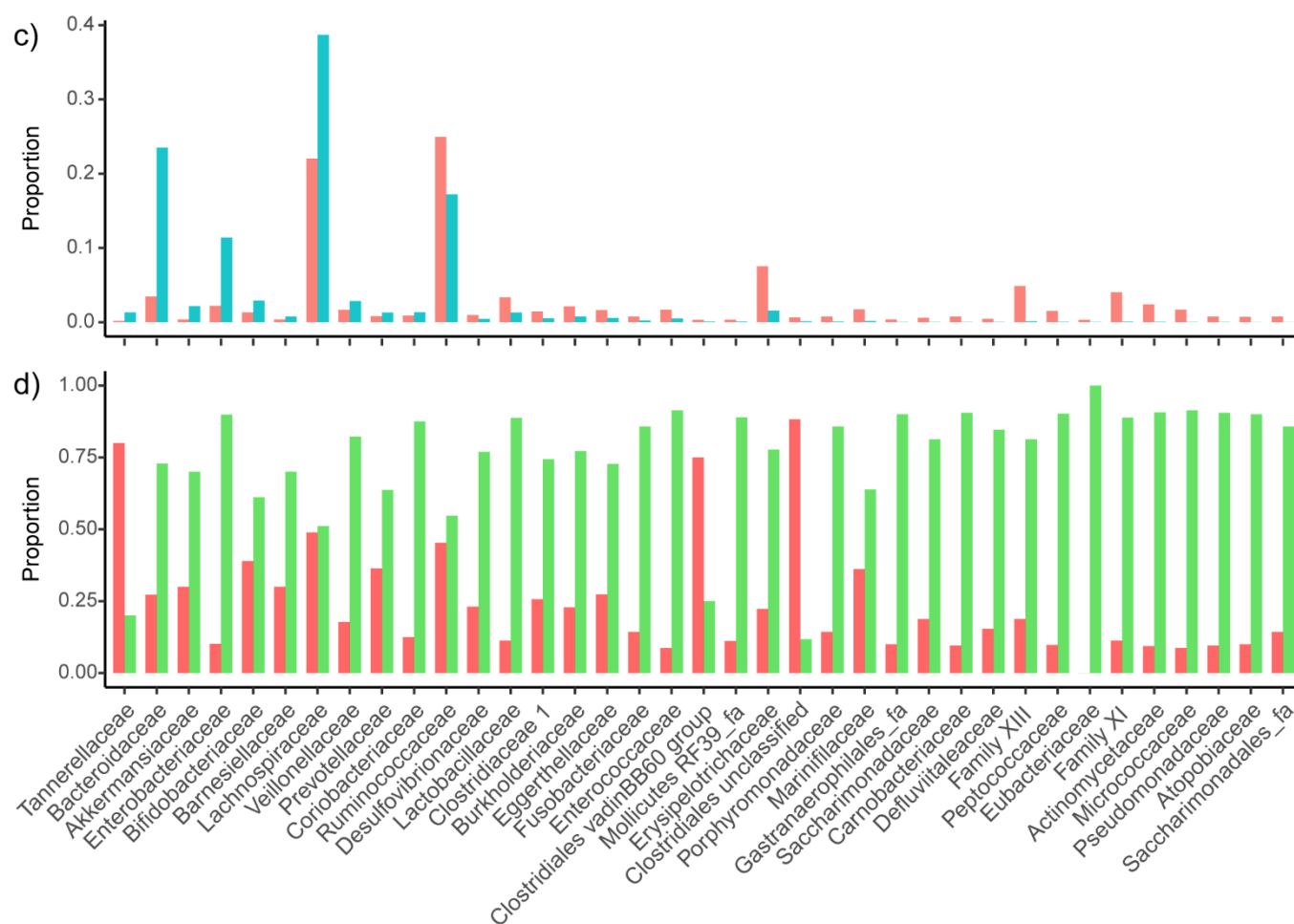


Figure 7. Cont.

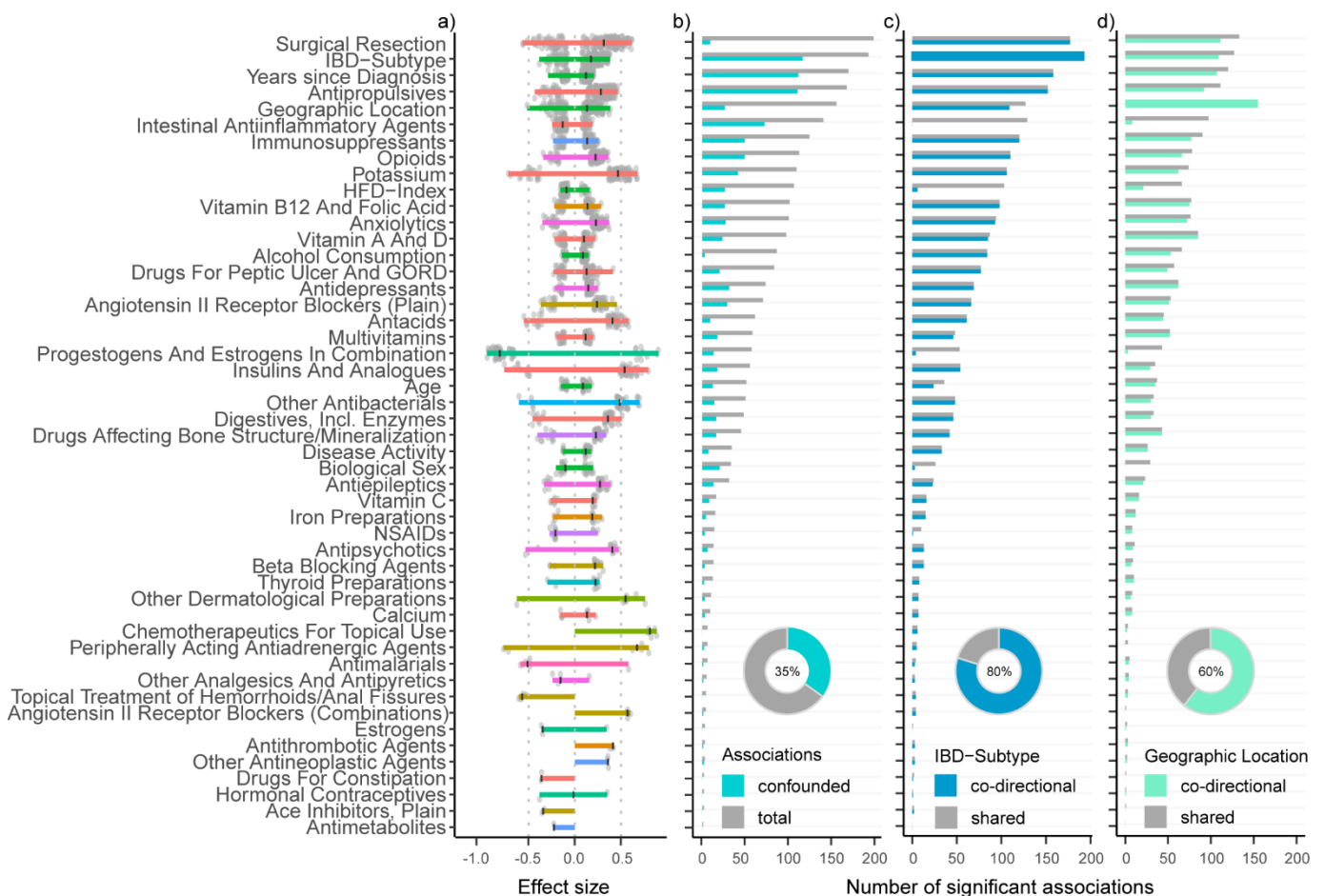


**Figure 7.** Taxa whose proportions of associations were significantly over- or under-represented compared to the overall distribution of OTU-counts at: (a) class; and (c) family level as determined by Fisher exact test; proportion of positive and negative associations for: (b) class level; and (d) family level. Bars are ordered by the ratio of associations to counts and negative to positive associations, respectively.

### 3.6. Additive Effects of Multiple Medications Amplify the Effect of IBD-Subtype on Gut Microbiota

In terms of specific treatment-association to taxa, surgical resection (198 associations), antipropulsives (167) and intestinal anti-inflammatory agents (140) showed the highest number of significant associations. A combination of estrogens and progesterones, chemotherapeutics for topical use, insulins and peripherally acting antiandrogenic agents showed the strongest associations with single taxa (Figure 8a). Of the overall 2754 significant associations between covariates and the taxonomic features, 35% were confounded, i.e.; the effect size value for a metadata/ taxonomic feature pair could be reduced to at least one other covariate while retaining its own significance (Figure 8b). Markedly, the proportion of confounded associations was lower for geographic location (16.8%) but much higher for IBD-subtype (60.4%). IBD-subtype showed interaction effects with either years since diagnosis, geographic location, biological sex or surgical resection, as well as five other treatments (A07E, B03A, A07D, N02A and A12B in Supplementary Table S2). Moreover, 2547 of all associations (92.5%) involved taxa that were also significant for the IBD-subtype, whereof 80% were in the same  $\pm$  direction, thus potentially amplifying the separation of the gut microbiome composition between persons with CD and persons with UC. For example, the immunosuppressant subgroup (L04A), which contains selective immunosuppressants such as mycophenolic acid but also tumor necrosis factor alpha (TNF $\alpha$ ) inhibitors and other immunosuppressants such as methotrexate and azathioprine showed significant

associations with 124 taxonomic features. Of these, 120 were not only also significant for IBD-subtype but also co-directional. A notable exemption to this exacerbating effect were intestinal anti-inflammatories, a pharmacological subgroup that contains locally acting corticosteroids and aminosalicyclic acid and similar agents and shared 129 of its 140 significant associations with IBD-subtype, but none of those were co-directional, suggesting that they counteracted the effect of IBD-subtype towards a healthier microbiome. Other variables that reduced the effect of IBD-subtype were a combination of progesterone and estrogen (54 shared, whereof only 4 were co-directional), HFD-index (106 shared, 6 co-directional) and biological sex (26 shared, 3 co-directional; Figure 8c). The amplifying effect for geographic location was much less pronounced, as 2037 associations (80%) were shared with other variables and only 60% of those were co-directional. Surgical resection, Vitamin A/D supplementation and antipropulsives had the most co-directional associations with geographic location (111, 92 and 85, respectively). Years since diagnosis and IBD-subtype also showed a high overlap with the effect of geographic location (107 and 109 co-directional; Figure 8d). While most of these effects were weak or moderate (Figure 8a), the summation of the many co-directional effects from different treatments might obscure which taxa were most depleted or increased due to a variable of interest in this case IBD-subtype and to a lesser degree geographic location.



**Figure 8.** Effect sizes (a) and number of significant associations by metadata variable that are (b) confounded: (c) shared/co-directional with disease; and (d) shared/co-directional with geographic location.

**4. Discussion**

In our previous study, we showed that geographic location accounted for the second highest explained variance in gut microbial composition after a diagnosis with CD [6]. The

present study extends the earlier observations by addressing differences in medication profiles in greater detail. We discovered that multiple medications and supplements were differentially used between persons with UC and CD as well as between Canadian and Irish participants. Indeed, Canadians were found to take significantly more medications and supplements than their Irish counterparts, and IBD-subtype accounted for only slightly more variation in drug usage patterns than geographic location. Despite this, when assessing the confounding effects of treatments on the microbiota, only a small part of the variation in microbial composition between participants from the different geographic locations was explained by the differences in medication and supplement intake.

In contrast, a major part of the disparity between the gut microbiomes of persons with UC versus CD seems to be due to, or amplified by, interaction effects with treatment. About half of the tested medications and supplements showed significant associations with at least one taxon from the gut microbiota, and together, treatments, including surgical resection, and medications and supplements, explained more variation in gut microbial composition than all other tested environmental factors.

Several taxa whose increase are generally reported with a shift away from healthy gut microbiome composition to an inflamed state, including *Escherichia/Shigella*, *Streptococcus*, *Klebsiella* and *Veillonellaceae* [3,4,55,56] were found here to be increased in the microbiome of persons with CD. These taxa also notably ranked amongst the highest number of positive associations with the tested covariates. Bacteria from the oral cavity such as *Rothia dentocariosa* and *Fusobacterium nucleatum*, which have been reported to be increased in PPI users [19,20] showed not only significant positive associations with drugs for peptic ulcers and GORD in the present study but also belong to the top 40 most affected bacteria. A depletion of *Faecalibacterium prausnitzii*, was in this and other studies associated with CD [5,29,57,58] and was also described in persons treated with the immunosuppressant azathioprine [59]. Whilst we were unable to confirm a negative association of *F. prausnitzii* with the intake of immunosuppressants, it was among the taxa which had the most negative associations with the tested drugs. The lack of diminution of this taxon by immunosuppressants might be explainable due to the fact that the ATC subgroup L04A not only includes thiopurines, but also TNF $\alpha$  inhibitors, the latter of which have been shown to increase SCFA producing bacteria [60]. It is notable though that immunosuppressants share nearly all their significant associations to taxonomic features with IBD-subtype, and all of those shared associations showed the same directionality and thus increase the disparity between the gut microbiota of persons with UC and CD. In contrast to that, intestinal anti-inflammatory agents are among the few drugs that did not follow this exacerbating trend. While this medication subgroup also shared most of its significant associations to taxa with IBD-subtype, none of them were co-directional. This observation is in agreement with earlier reports that 5-ASA drugs can partially recover the gut microbiome to a healthy status [29,61].

Comparing the distribution of the number of reads and significant associations to medications for each taxon showed that some taxa were more resistant towards the effect of human targeted drugs, e.g.; Verrucomicrobiae, Bacteroidia and  $\gamma$ -Proteobacteria, while others, such as Actinobacteria, Bacilli and Clostridia, were more sensitive. These results concurred with a study from Maier et al. which found  $\gamma$ -Proteobacteria to be more drug resistant than highly abundant commensals such as *Roseburia intestinalis*, *Eubacterium rectale* and *Blautia obeum* [8].

While this study could not confirm treatment as a major factor explaining dissimilarities in gut microbiota of persons from different geographic locations, it shows that the highly variable medication profiles of persons with IBD and their effect on the faecal gut microbiota likely impede the discovery of a universally valid microbial signature distinguishing the IBD-subtypes and at least in part, explain the high disparity between different IBD microbiome studies. Furthermore, it highlights the need to include an exhaustive list of medication intakes (and ideally dosages) of study participants in the analysis that go beyond the common IBD therapeutics to improve reproducibility between IBD-studies.

There are several limitations to our study. Due to the highly variable drug usage patterns, not all medications could be assessed for univariate analysis. The evaluation of the confounding effects of medication on gut microbiota was further hampered by extreme polypharmacy and a multitude of changes in medication regimes over the course of the study. It is therefore possible that some of the reported effects of particular medications were under-estimated. However, while the exact effect sizes of single medications may not be robust, taken together they reveal a trend that treatments can exacerbate disease-associated shifts by additive effects of multiple medications on the microbiome as well as by affecting some groups of microbiota more than others. It is noteworthy though, that all described effects between medication and microbiome in this study are associative rather than causative, and thus intervention studies with treatment naïve persons with IBD and animal models will be needed to comprehensively disentangle the role of treatment and disease on the variation the gut microbiome of patients with IBD.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/microorganisms10101963/s1>, Table S1: Study cohort characteristics; Table S2: Medications in ATC3 classification; Table S3: Comparison of alpha diversity metrics between IBD-subtypes and geographic locations; Table S4: Differentially abundant taxa (highest available classification) between patients with UC and CD as determined with ALDEx2; Table S5: Differentially abundant taxa (highest available classification) between patients from Canada and Ireland as determined with ALDEx2; Table S6: Differentially used medications and supplements between patient from different geographic locations and IBD-subtype as determined with Fisher Exact tests; Table S7: Comparison of intra-personal alpha and beta diversity measures between patients that started a new drug and those who did not; Table S8: Multivariate regression analysis with vphemall. “naive” denotes the naïve effect of each factor on the gut microbial composition, “unique” denotes the conservative estimate after partitioning out the effect of all variables, “interaction” is the difference between “naive” and “unique”; Table S9: Number of significant, shared and co-directional associations between taxa and metadata as well as their minimum, maximum and median effect sizes; Table S10: Fisher Exact Test of significant associations between taxa and medications and taxa abundance on class level; Table S11: Fisher Exact Test of significant associations between taxa and medications and taxa abundance on family level.

**Author Contributions:** Conceptualization, J.E.; C.N.B.; F.S. and M.J.C.; methodology, J.E.; formal analysis, J.E. and J.C.B.; investigation, J.E. and M.J.C.; resources, C.N.B.; F.S. and M.J.C.; writing—original draft preparation, J.E. and M.J.C.; writing—review and editing, C.N.B. and F.S.; visualization, J.E.; supervision, M.J.C.; funding acquisition, F.S. and M.J.C. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by Science Foundation Ireland (grant numbers SFI/12/RC/2273\_P2 and 17/CDA/4765).

**Institutional Review Board Statement:** All subjects gave their informed consent for inclusion before they participated in the study. The study was conducted in accordance with the Declaration of Helsinki, and the protocol was approved by the Cork hospitals’ research ethics committee and the University of Manitoba Health Research Board (ECM 3 (gg) 06/01/09 & ECM 3 (e) 04/12/18) and (HS13164 (H2011:219)).

**Informed Consent Statement:** Informed consent was obtained from all subjects involved in the study.

**Data Availability Statement:** The data presented in this study are openly available in FigShare at [doi.org/10.6084/m9.figshare.21088885.v1](https://doi.org/10.6084/m9.figshare.21088885.v1) and [doi.org/10.6084/m9.figshare.20701291.v1](https://doi.org/10.6084/m9.figshare.20701291.v1). Sequence data are available at NCBI SRA PRJNA414072.

**Conflicts of Interest:** The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.



## References

1. Ryan, F.J.; Ahern, A.M.; Fitzgerald, R.S.; Laserna-Mendieta, E.J.; Power, E.M.; Clooney, A.G.; O'Donoghue, K.W.; McMurdie, P.J.; Iwai, S.; Crits-Christoph, A.; et al. Colonic microbiota is associated with inflammation and host epigenomic alterations in inflammatory bowel disease. *Nat. Commun.* **2020**, *11*, 1512. [[CrossRef](#)] [[PubMed](#)]
2. Alam, M.T.; Amos, G.C.A.; Murphy, A.R.J.; Murch, S.; Wellington, E.M.H.; Arasaradnam, R.P. Microbial imbalance in inflammatory bowel disease patients at different taxonomic levels. *Gut Pathog.* **2020**, *12*, 1. [[CrossRef](#)] [[PubMed](#)]
3. Khan, I.; Ullah, N.; Zha, L.; Bai, Y.; Khan, A.; Zhao, T.; Che, T.; Zhang, C. Alteration of Gut Microbiota in Inflammatory Bowel Disease (IBD): Cause or Consequence? IBD Treatment Targeting the Gut Microbiome. *Pathogens* **2019**, *8*, 126. [[CrossRef](#)] [[PubMed](#)]
4. Aldars-Garcia, L.; Chaparro, M.; Gisbert, J.P. Systematic Review: The Gut Microbiome and Its Potential Clinical Application in Inflammatory Bowel Disease. *Microorganisms* **2021**, *9*, 977. [[CrossRef](#)] [[PubMed](#)]
5. Pittayanon, R.; Lau, J.T.; Leontiadis, G.I.; Tse, F.; Yuan, Y.; Surette, M.; Moayyedi, P. Differences in Gut Microbiota in Patients With vs Without Inflammatory Bowel Diseases: A Systematic Review. *Gastroenterology* **2020**, *158*, 930–946.e1. [[CrossRef](#)] [[PubMed](#)]
6. Elvers, K.T.; Wilson, V.J.; Hammond, A.; Duncan, L.; Huntley, A.L.; Hay, A.D.; van der Werf, E.T. Antibiotic-induced changes in the human gut microbiota for the most commonly prescribed antibiotics in primary care in the UK: A systematic review. *BMJ Open* **2020**, *10*, e035677. [[CrossRef](#)]
7. Oh, B.; Boyle, F.; Pavlakakis, N.; Clarke, S.; Guminski, A.; Eade, T.; Lamoury, G.; Carroll, S.; Morgia, M.; Kneebone, A.; et al. Emerging Evidence of the Gut Microbiome in Chemotherapy: A Clinical Review. *Front. Oncol.* **2021**, *11*, 706331. [[CrossRef](#)]
8. Maier, L.; Pruteanu, M.; Kuhn, M.; Zeller, G.; Telzerow, A.; Anderson, E.E.; Brochado, A.R.; Fernandez, K.C.; Dose, H.; Mori, H.; et al. Extensive impact of non-antibiotic drugs on human gut bacteria. *Nature* **2018**, *555*, 623–628. [[CrossRef](#)]
9. Sargautiene, V.; Ligere, R.; Kalniņa, I.; Jākobsone, I.; Nikolajeva, V.; Derovs, A. The Effect of 5-Aminosalicylic Acid on Intestinal Microbiota. *Proc. Latv. Acad. Sci. Sect. B Nat. Exact Appl. Sci.* **2020**, *74*, 53–57. [[CrossRef](#)]
10. Huang, E.Y.; Inoue, T.; Leone, V.A.; Dalal, S.; Touw, K.; Wang, Y.W.; Musch, M.W.; Theriault, B.; Higuchi, K.; Donovan, S.; et al. Using Corticosteroids to Reshape the Gut Microbiome: Implications for Inflammatory Bowel Diseases. *Inflamm. Bowel Dis.* **2015**, *21*, 963–972. [[CrossRef](#)]
11. Swidsinski, A.; Loening-Baucke, V.; Bengmark, S.; Lochs, H.; Dorffel, Y. Azathioprine and mesalazine-induced effects on the mucosal flora in patients with IBD colitis. *Inflamm. Bowel Dis.* **2007**, *13*, 51–56. [[CrossRef](#)] [[PubMed](#)]
12. Letertre, M.P.M.; Munjoma, N.; Wolfer, K.; Pechlivanis, A.; McDonald, J.A.K.; Hardwick, R.N.; Cherrington, N.J.; Coen, M.; Nicholson, J.K.; Hoyles, L.; et al. A Two-Way Interaction between Methotrexate and the Gut Microbiota of Male Sprague-Dawley Rats. *J. Proteome Res.* **2020**, *19*, 3326–3339. [[CrossRef](#)] [[PubMed](#)]
13. Magnusson, M.K.; Strid, H.; Sapnara, M.; Lasso, A.; Bajor, A.; Ung, K.A.; Ohman, L. Anti-TNF Therapy Response in Patients with Ulcerative Colitis Is Associated with Colonic Antimicrobial Peptide Expression and Microbiota Composition. *J. Crohn's Colitis* **2016**, *10*, 943–952. [[CrossRef](#)] [[PubMed](#)]
14. Franzin, M.; Stefancic, K.; Lucafo, M.; Decorti, G.; Stocco, G. Microbiota and Drug Response in Inflammatory Bowel Disease. *Pathogens* **2021**, *10*, 211. [[CrossRef](#)] [[PubMed](#)]
15. Xu, J.; Chen, N.; Wu, Z.; Song, Y.; Zhang, Y.; Wu, N.; Zhang, F.; Ren, X.; Liu, Y. 5-Aminosalicylic Acid Alters the Gut Bacterial Microbiota in Patients With Ulcerative Colitis. *Front. Microbiol.* **2018**, *9*, 1274. [[CrossRef](#)] [[PubMed](#)]
16. Vich Vila, A.; Collij, V.; Sanna, S.; Sinha, T.; Imhann, F.; Bourgonje, A.R.; Mujagic, Z.; Jonkers, D.M.A.E.; Masclee, A.A.M.; Fu, J.; et al. Impact of commonly used drugs on the composition and metabolic function of the gut microbiota. *Nat. Commun.* **2020**, *11*, 362. [[CrossRef](#)]
17. Zimmermann, M.; Zimmermann-Kogadeeva, M.; Wegmann, R.; Goodman, A.L. Mapping human microbiome drug metabolism by gut bacteria and their genes. *Nature* **2019**, *570*, 462–467. [[CrossRef](#)]
18. Klunemann, M.; Andrejev, S.; Blasche, S.; Mateus, A.; Phapale, P.; Devendran, S.; Vappiani, J.; Simon, B.; Scott, T.A.; Kafkia, E.; et al. Bioaccumulation of therapeutic drugs by human gut bacteria. *Nature* **2021**, *597*, 533–538. [[CrossRef](#)]
19. Clooney, A.G.; Bernstein, C.N.; Leslie, W.D.; Vagianos, K.; Sargent, M.; Laserna-Mendieta, E.J.; Claesson, M.J.; Targownik, L.E. A comparison of the gut microbiome between long-term users and non-users of proton pump inhibitors. *Aliment. Pharmacol. Ther.* **2016**, *43*, 974–984. [[CrossRef](#)]
20. Weersma, R.K.; Zhernakova, A.; Fu, J. Interaction between drugs and the gut microbiome. *Gut* **2020**, *69*, 1510–1519. [[CrossRef](#)]
21. Vieira-Silva, S.; Falony, G.; Belda, E.; Nielsen, T.; Aron-Wisnewsky, J.; Chakaroun, R.; Forslund, S.K.; Assmann, K.; Valles-Colomer, M.; Nguyen, T.T.D.; et al. Statin therapy is associated with lower prevalence of gut microbiota dysbiosis. *Nature* **2020**, *581*, 310–315. [[CrossRef](#)] [[PubMed](#)]
22. Tropini, C.; Moss, E.L.; Merrill, B.D.; Ng, K.M.; Higginbottom, S.K.; Casavant, E.P.; Gonzalez, C.G.; Fremin, B.; Bouley, D.M.; Elias, J.E.; et al. Transient Osmotic Perturbation Causes Long-Term Alteration to the Gut Microbiota. *Cell* **2018**, *173*, 1742–1754.e17. [[CrossRef](#)] [[PubMed](#)]
23. Forslund, K.; Hildebrand, F.; Nielsen, T.; Falony, G.; Le Chatelier, E.; Sunagawa, S.; Prifti, E.; Vieira-Silva, S.; Gudmundsdottir, V.; Pedersen, H.K.; et al. Disentangling type 2 diabetes and metformin treatment signatures in the human gut microbiota. *Nature* **2015**, *528*, 262–266. [[CrossRef](#)] [[PubMed](#)]
24. Lin, Y.-T.; Lin, T.-Y.; Hung, S.-C.; Liu, P.-Y.; Hung, W.-C.; Tsai, W.-C.; Tsai, Y.-C.; Delicano, R.A.; Chuang, Y.-S.; Kuo, M.-C.; et al. Differences in the Microbial Composition of Hemodialysis Patients Treated with and without  $\beta$ -Blockers. *J. Pers. Med.* **2021**, *11*, 198. [[CrossRef](#)] [[PubMed](#)]

25. Tuteja, S.; Ferguson, J.F. Gut Microbiome and Response to Cardiovascular Drugs. *Circ. Genom. Precis. Med.* **2019**, *12*, e002314. [[CrossRef](#)] [[PubMed](#)]
26. Munoz-Bellido, J.L.; Munoz-Criado, S.; Garia-Rodriguez, J.A. Antimicrobial activity of psychotropic drugs: Selective serotonin reuptake inhibitors. *Int. J. Antimicrob. Agents* **2000**, *14*, 177–180. [[CrossRef](#)]
27. Falony, G.; Joossens, M.; Vieira-Silva, S.; Wang, J.; Darzi, Y.; Faust, K.; Kurilshikov, A.; Bonder, M.J.; Valles-Colomer, M.; Vandeputte, D.; et al. Population-level analysis of gut microbiome variation. *Science* **2016**, *352*, 560–564. [[CrossRef](#)]
28. Forslund, S.K.; Chakaroun, R.; Zimmermann-Kogadeeva, M.; Marko, L.; Aron-Wisniewsky, J.; Nielsen, T.; Moitinho-Silva, L.; Schmidt, T.S.B.; Falony, G.; Vieira-Silva, S.; et al. Combinatorial, additive and dose-dependent drug-microbiome associations. *Nature* **2021**, *600*, 500–505. [[CrossRef](#)]
29. Clooney, A.G.; Eckenberger, J.; Laserna-Mendieta, E.; Sexton, K.A.; Bernstein, M.T.; Vagianos, K.; Sargent, M.; Ryan, F.J.; Moran, C.; Sheehan, D.; et al. Ranking microbiome variance in inflammatory bowel disease: A large longitudinal intercontinental study. *Gut* **2021**, *70*, 499. [[CrossRef](#)]
30. Quast, C.; Pruesse, E.; Yilmaz, P.; Gerken, J.; Schweer, T.; Yarza, P.; Peplies, J.; Glockner, F.O. The SILVA ribosomal RNA gene database project: Improved data processing and web-based tools. *Nucleic Acids Res.* **2013**, *41*, D590–D596. [[CrossRef](#)]
31. Schloss, P.D.; Westcott, S.L.; Ryabin, T.; Hall, J.R.; Hartmann, M.; Hollister, E.B.; Lesniewski, R.A.; Oakley, B.B.; Parks, D.H.; Robinson, C.J.; et al. Introducing mothur: Open-source, platform-independent, community-supported software for describing and comparing microbial communities. *Appl. Environ. Microbiol.* **2009**, *75*, 7537–7541. [[CrossRef](#)] [[PubMed](#)]
32. Allard, G.; Ryan, F.J.; Jeffery, I.B.; Claesson, M.J. SPINGO: A rapid species-classifier for microbial amplicon sequences. *BMC Bioinform.* **2015**, *16*, 324. [[CrossRef](#)] [[PubMed](#)]
33. Altschul, S.F.; Gish, W.; Miller, W.; Myers, E.W.; Lipman, D.J. Basic local alignment search tool. *J. Mol. Biol.* **1990**, *215*, 403–410. [[CrossRef](#)]
34. WHO Collaborating Centre for Drug Statistics Methodology. *Guidelines for ATC Classification and DDD Assignment*; Norwegian Institute of Public Health: Oslo, Norway, 2021.
35. Sica, G.S.; Biancone, L. Surgery for inflammatory bowel disease in the era of laparoscopy. *World J. Gastroenterol.* **2013**, *19*, 2445–2448. [[CrossRef](#)]
36. Fang, X.; Vazquez-Baeza, Y.; Elijah, E.; Vargas, F.; Ackermann, G.; Humphrey, G.; Lau, R.; Weldon, K.C.; Sanders, J.G.; Panitchpakdi, M.; et al. Gastrointestinal Surgery for Inflammatory Bowel Disease Persistently Lowers Microbiome and Metabolome Diversity (vol 27, pg 1368, 2021). *Inflamm. Bowel Dis.* **2021**, *27*, 1368.
37. Drescher, L.S.; Thiele, S.; Mensink, G.B. A new index to measure healthy food diversity better reflects a healthy diet than traditional measures. *J. Nutr.* **2007**, *137*, 647–651. [[CrossRef](#)]
38. Lin, J.F.; Chen, J.M.; Zuo, J.H.; Yu, A.; Xiao, Z.J.; Deng, F.H.; Nie, B.; Jiang, B. Meta-analysis: Fecal calprotectin for assessment of inflammatory bowel disease activity. *Inflamm. Bowel Dis.* **2014**, *20*, 1407–1415. [[CrossRef](#)]
39. Wickham, H. *ggplot2: Elegant Graphics for Data Analysis*; Springer: New York, NY, USA, 2016.
40. Gloor, G.B.; Macklaim, J.M.; Pawlowsky-Glahn, V.; Egozcue, J.J. Microbiome Datasets Are Compositional: And This Is Not Optional. *Front. Microbiol.* **2017**, *8*, 2224. [[CrossRef](#)]
41. Palarea-Albaladejo, J.; Martín-Fernández, J.A. zCompositions—R package for multivariate imputation of left-censored data under a compositional approach. *Chemom. Intell. Lab. Syst.* **2015**, *143*, 85–96. [[CrossRef](#)]
42. Quinn, T.P.; Erb, I.; Gloor, G.; Notredame, C.; Richardson, M.F.; Crowley, T.M. A field guide for the compositional analysis of any-omics data. *GigaScience* **2019**, *8*, giz107. [[CrossRef](#)]
43. Aitchison, J.; Barceló-Vidal, C.; Martín-Fernández, J.A.; Pawlowsky-Glahn, V. Logratio Analysis and Compositional Distance. *Math. Geol.* **2000**, *32*, 271–275. [[CrossRef](#)]
44. McMurdie, P.J.; Holmes, S. phyloseq: An R Package for Reproducible Interactive Analysis and Graphics of Microbiome Census Data. *PLoS ONE* **2013**, *8*, e61217. [[CrossRef](#)] [[PubMed](#)]
45. Oksanen, J.; Blanchet, F.G.; Kindt, R.; Legendre, P.; Minchin, P.; O’hara, R.; Simpson, G.; Solymos, P.; Stevens, M.H.H.; Wagner, H. Community Ecology Package. R Package Version 2.2-0. Available online: <http://CRAN.Rproject.org/package=vegan> (accessed on 1 April 2022).
46. Chao, A.; Gotelli, N.J.; Hsieh, T.C.; Sander, E.L.; Ma, K.H.; Colwell, R.K.; Ellison, A.M. Rarefaction and extrapolation with Hill numbers: A framework for sampling and estimation in species diversity studies. *Ecol. Monogr.* **2014**, *84*, 45–67. [[CrossRef](#)]
47. Fernandes, A.D.; Macklaim, J.M.; Linn, T.G.; Reid, G.; Gloor, G.B. ANOVA-Like Differential Expression (ALDEx) Analysis for Mixed Population RNA-Seq. *PLoS ONE* **2013**, *8*, e67019. [[CrossRef](#)] [[PubMed](#)]
48. von Lampe, F. Goeveg R-Package. In *Functions for Community Data and Ordinations*. A Collection of Functions Useful in (Vegetation) Community Analyses and Ordinations. 2021. Available online: <https://github.com/fvlampe/goeveg> (accessed on 1 April 2022).
49. Moitinho-Silva, L.; Forslund, S.K.; Chakaroun, R.; Zimmermann-Kogadeeva, M.; Markó, L.; Aron-Wisniewsky, J.; Nielsen, T.; Birkner, T. Grp-bork/vpthemall: Release of Vpthemall—Assistant Functions for Variation Partition with dbRDA Method. Available online: <https://zenodo.org/record/6242715#.Yzra43bMInI> (accessed on 1 April 2022).
50. Forslund, S.K.; Chakaroun, R.; Zimmermann-Kogadeeva, M.; Markó, L.; Aron-Wisniewsky, J.; Nielsen, T.; Birkner, T. Data Analysis Pipeline for Investigating Drug-Host-Microbiome Relationships in Cardiometabolic Disease (MetaCardis Cohort). Available online: <https://zenodo.org/record/5463864#.YzrbsHbMInI> (accessed on 1 April 2022).

51. Benjamini, Y.; Drai, D.; Elmer, G.; Kafkafi, N.; Golani, I. Controlling the false discovery rate in behavior genetics research. *Behav. Brain Res.* **2001**, *125*, 279–284. [[CrossRef](#)]
52. Ticinesi, A.; Milani, C.; Lauretani, F.; Nouvenne, A.; Mancabelli, L.; Lugli, G.A.; Turrone, F.; Duranti, S.; Mangifesta, M.; Viappiani, A.; et al. Gut microbiota composition is associated with polypharmacy in elderly hospitalized patients. *Sci. Rep.* **2017**, *7*, 11102. [[CrossRef](#)]
53. Shanahan, F.; Ghosh, T.S.; O’Toole, P.W. The Healthy Microbiome—What Is the Definition of a Healthy Gut Microbiome? *Gastroenterology* **2021**, *160*, 483–494. [[CrossRef](#)]
54. Deleu, S.; Machiels, K.; Raes, J.; Verbeke, K.; Vermeire, S. Short chain fatty acids and its producing organisms: An overlooked therapy for IBD? *eBioMedicine* **2021**, *66*, 103293. [[CrossRef](#)]
55. Buttó, L.F.; Haller, D. Dysbiosis in intestinal inflammation: Cause or consequence. *Int. J. Med. Microbiol.* **2016**, *306*, 302–309. [[CrossRef](#)]
56. Mirsepasi-Lauridsen, H.C.; Vallance, B.A.; Krogfelt, K.A.; Petersen, A.M. Escherichia coli Pathobionts Associated with Inflammatory Bowel Disease. *Clin. Microbiol. Rev.* **2019**, *32*, e00060-18. [[CrossRef](#)]
57. Sokol, H.; Pigneur, B.; Watterlot, L.; Lakhdari, O.; Bermudez-Humaran, L.G.; Gratadoux, J.J.; Blugeon, S.; Bridonneau, C.; Furet, J.P.; Corthier, G.; et al. Faecalibacterium prausnitzii is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 16731–16736. [[CrossRef](#)] [[PubMed](#)]
58. Melgar, S.; Shanahan, F. Inflammatory bowel disease—from mechanisms to treatment strategies. *Autoimmunity* **2010**, *43*, 463–477. [[CrossRef](#)] [[PubMed](#)]
59. Effenberger, M.; Reider, S.; Waschina, S.; Bronowski, C.; Enrich, B.; Adolph, T.E.; Koch, R.; Moschen, A.R.; Rosenstiel, P.; Aden, K.; et al. Microbial Butyrate Synthesis Indicates Therapeutic Efficacy of Azathioprine in IBD Patients. *J. Crohn’s Colitis* **2021**, *15*, 88–98. [[CrossRef](#)]
60. Estevinho, M.M.; Rocha, C.; Correia, L.; Lago, P.; Ministro, P.; Portela, F.; Trindade, E.; Afonso, J.; Peyrin-Biroulet, L.; Magro, F.; et al. Features of Fecal and Colon Microbiomes Associate with Responses to Biologic Therapies for Inflammatory Bowel Diseases: A Systematic Review. *Clin. Gastroenterol. Hepatol.* **2020**, *18*, 1054–1069. [[CrossRef](#)]
61. Olaisen, M.; Spigset, O.; Flatberg, A.; Granlund, A.V.B.; Brede, W.R.; Albrektsen, G.; Royset, E.S.; Gilde, B.; Sandvik, A.K.; Martinsen, T.C.; et al. Mucosal 5-aminosalicylic acid concentration, drug formulation and mucosal microbiome in patients with quiescent ulcerative colitis. *Aliment. Pharmacol. Ther.* **2019**, *49*, 1301–1313. [[CrossRef](#)] [[PubMed](#)]