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Targeting 4-1BB and PD-L1 induces potent and durable antitumor immunity in B-cell lymphoma

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Introduction: Although PD-1/L1 mAb has demonstrated clinical benefits in certain cancer types, low response rate and resistance remain the main challenges for the application of these immune checkpoint inhibitors (ICIs). 4-1BB is a co-stimulator molecule expressed in T cells, which could enhance T cell proliferation and activation. Herein, the synergetic antitumor effect and underlying mechanism of 4-1BB agonist combined with PD-1/PD-L1 blockade were determined in B-cell lymphoma (BCL).

Methods: Subcutaneous transplantation BCL tumor models and metastasis models were established to evaluate the therapeutic effect of PD-L1 antibody and/or 4-1BB agonist in vivo. For the mechanistic study, RNA-seq was applied to analyze the tumor microenvironment and immune-related signal pathway after combination treatment. The level of IFN- γ , perforin, and granzyme B were determined by ELISA and Real-time PCR assays, while tumor-infiltrating T cells were measured by flow cytometry and immunohistochemical analysis. CD4/ CD8 specific antibodies were employed to deplete the related T cells to investigate the role CD4+ and CD8+ T cells played in combination treatment.

Results: Our results showed that combining anti-PD-L1 ICI and 4-1BB agonists elicited regression of BCL and significantly extended the survival of mice compared to either monotherapy. Co-targeting PD-L1 and 4-1BB preferentially promoted intratumoral cytotoxic lymphocyte infiltration and remodeled their function. RNA-sequence analysis uncovered a series of up-regulated genes related to the activation and proliferation of cytotoxic T lymphocytes, further characterized by increased cytokines including IFN- γ , granzyme B, and perforin. Furthermore, depleting CD8+ T cells not CD4+ T cells totally abrogated the antitumor efficacy, indicating the crucial function of the CD8+ T cell subset in the combination therapy.

Discussion: In summary, our findings demonstrated that 4-1BB agonistic antibody intensified the antitumor immunity of anti-PD-1/PD-L1 ICI via promoting CD8+ T cell infiltration and activation, providing a novel therapeutic strategy to BCL.

KEYWORDS

B-cell lymphoma, anti-PD-L1 ICI, 4-1BB agonist, CD8+ T cells, combination therapy

Introduction

Although PD-1/PD-L1 axis-targeting immune checkpoint inhibitor (ICI) has revolutionized the therapeutic modality of several types of cancers, an obvious fraction of cancer patients showed no responses to ICI monotherapy (1, 2). Multiple factors such as immune cell infiltration, expression of immune checkpoint and cytokine signaling could monitor the response to ICIs and influence their efficacy (3, 4). Elucidation of the underlying immunologic characteristic of the tumor microenvironment (TME) associated with resistance will benefit the patients treated with ICI monotherapy and reveal the determinants for ICIs combination therapy.

As one prominent co-stimulator, 4-1BB is mainly expressed on natural killer T and CD4⁺/CD8⁺ T cells (5-7). Upon conjunction with soluble 4-1BBL or agonistic monoclonal antibody (mAb), 4-1BB forms a heterotrimer and induces Tcell proliferation, cytokine release, and upregulation of antiapoptotic molecules (8). In 4-1BB-deficient models, the function of cytotoxic T lymphocytes is mostly diminished, and 4-1BB on infiltrated T cells in TME could serve as a marker to predict the antitumor effect of immunotherapy (9). A comprehensive study revealed for the first time that targeting 4-1BB has strong antitumor effects via injecting mice bearing Ag104A sarcoma and P815 mastocytoma with anti-4-1BB mAbs (10). Kinetic studies of 4-1BB expression on T cells indicate that although mechanisms for the differential costimulatory ability are not completely elucidated in T cell subsets, 4-1BB is an ideal target on CD8⁺ T cells for immunotherapy. Indeed, while agonic mAb of 4-1BB could reduce the tumor size and increase survival in multiple preclinical studies (11-13), only limited clinical benefit has been observed due to dose-dependent hepatic toxicity (14-16).

Anti-4-1BB mAb has been proved as an enhancer in the antitumor immunity (17). The bispecific Ab targeting 4-1BB and HER2 showed potent therapeutic effects in HER2 positive cancer (18). IL-12 and 4-1BB were found to possess synergistic antitumor effect *via* activating $CD8^+$ T cell response in mouse models (19). As to the impact of 4-1BB on the therapeutic effect of ICIs, researches demonstrated that costimulatory pathways

and immune checkpoint pathways were interdependent (20, 21). Lack of the costimulatory signals led to increased PD-1 expression, which further decreased IL-2 receptors that were necessary for T cell proliferation, whereas 4-1BB co-stimulation potently enhanced reinvigoration of infiltrated T cells in TME (22). Although these findings imply a potential synergy between PD-1 blockade and 4-1BB agonist, the synergistic antitumor effect between PD-L1 blockade and 4-1BB agonist remains unclear (23).

In this context, antitumor immunity of PD-1/PD-L1 blockade and 4-1BB agonist was evaluated in immunocompetent B-cell lymphoma (BCL) models. The transcriptional profile was further analyzed to uncover the underlying mechanisms. Our results highlighted PD-L1 blockade combined with 4-1BB agonist as a potential therapeutic strategy for BCL.

Results

Anti-PD-L1 mAb and 4-1BB agonist elicited synergistic antitumor activity

To investigate the feasibility of dual-targeting PD-L1 and 4-1BB in BCL tumors, mice implanted with A20 or WEHI-231 tumors were treated with 4-1BB agonist, anti-PD-L1 mAb, 4-1BB agonist combined with anti-PD-L1 mAb, respectively. As shown in Figures 1A, B, compared with an equivalent dose of ICI alone, the combined therapy showed a more potent anti-tumor effect in both well-established A20 and WEHI-231 models. A20 tumor-bearing mice were sacrificed on day 18. All mice in the combined group were tumor-free, and the relative tumor weight compared to control in the groups of anti-PD-L1 and 4-1BB agonist antibody were 20.7 \pm 4.1% and 33.9 \pm 27.5% respectively (Figure 1C). Similar results were observed in WEHI-231 model. Compared with the group under monotherapy with anti-PD-L1 or 4-1BB agonist, tumor volume in the combined group began to decrease on day 8 and continued until the end of treatment (Figure 1B). Relative tumor weight in the groups of anti-PD-L1, 4-1BB agonist, anti-PD-L1 plus 4-1BB agonist were $66.8 \pm 7.5\%$, $52.2 \pm 7.0\%$ and $5.0 \pm 4.6\%$ respectively (Figure 1D). To further



explore whether the combined therapy could extend the survival, A20 metastatic model was established. Compared with the 4-1BB agonist or anti-PD-L1 monotherapy, combined therapy significantly extended the median survival. Importantly, half of the mice were still vigorous by the end of this experiment (Figure 1E). These data showed that anti-PD-L1 therapy combined with 4-1BB agonist elicited potent and durable antitumor effect in subcutaneous and metastatic BCL models.

Transcriptional profile involved in the synergistic antitumor effect

To reveal mechanisms underlying the synergistic antitumor effect, we performed RNA-sequencing on mouse A20 tumor tissue at 10 days post-treatment. In brief, Six tumor-bearing mice were divided equally into two groups and treated with PD-L1 antibody or a combination of 4-1bb agonist and PD-L1 antibody, and their tumor tissues were examined ten days later to analyze the transcriptional profile in the TME. Compared to anti-PD-L1 monotherapy, anti-PD-L1 therapy combined with 4-1BB agonist significantly altered the expression profiles of 538 genes (defined as P<0.05, fold change>2) in TME (Figure 2A). Furthermore, the biological functions of the modules were analyzed by KEGG (Figure 2B). Interestingly, most of the increased genes were associated with classical antitumor immunity signal pathways, including cytokine and chemokine pathways (1st and 2nd), and differentiation of Th1, Th2, and Th17 cells (3rd) (adjusted *P*<0.0005). In addition, there were enrichments in Protein digestion and absorption (4th), ECMreceptor interaction (5th), NF-kappa B signaling pathway (7th), PI3K-Akt signaling pathway (8th), and Cell adhesion molecules (9th), demonstrating that the combination therapy had a widespread influence on the metabolism and metastasis of tumor cells (Figure 2B). Integrally, GSEA analysis showed that combined therapy increased T cell activation and IFN- γ signal pathway (Figures 2C, D). These results uncovered the transcriptional landscape in BCL tumors under the cotreatment with anti-PD-L1 ICI and 4-1BB agonist.

4-1BB agonist potentiated anti-PD-L1 mAb-induced T cell immunity

Next, we sought to detect T cells infiltration in TME *via* flow cytometry. Compared to anti-PD-L1 monotherapy, a significant increase of tumor-infiltrating lymphocyte (TIL) was observed in the group of combinatorial therapy (Figure 3A). Furthermore, we determined T cell subsets in the indicated cohorts and found that anti-PD-L1 monotherapy presented modest impact on the infiltration of T cells, whereas there were obvious changes in the combinatorial group with a two-fold increase of CD8⁺ T cells



(Figures 3B, C). Immunohistochemistry staining was also performed to confirm CD8⁺ T cell infiltration. As shown in Figure 3D, compared to anti-PD-L1 alone, PD-L1 blockade plus 4-1BB agonism indeed promoted the infiltration of CD8⁺ T cells. Summary, these data indicated that 4-1BB agonist and PD-L1 blockade resulted in synergistic antitumor efficacy *via* reinforcing T-cell immunity and PD-L1 expression.

4-1BB agonist potentiated cytolytic capacity of infiltrated T cells

Infiltrated T cells in an established TME are mostly dysfunctional. Thus, gene signature and function of the infiltrated T cells were further evaluated after anti-PD-L1 and 4-1BB agonist combined therapy. We examined the genes related to T cell function in TME, and found that the upregulated genes were significantly enriched in T cell chemotaxis, differentiation, proliferation and activation (Figures 4A-C). Importantly, compared to anti-PD-L1 monotherapy, transcriptional levels of *ifng*, *gzma*, *prf1* and *gzmb* were also intensified by the combined therapy. To confirm immune-related gene expression signatures associated with response to T cell cytotoxicity, mRNA expression of key factors related to T cell cytotoxicity were also measured *via* RT-PCR and ELISA assay. As shown in Figure 4D, 4-1BB signaling significantly potent the release of IFN- γ , perforin and granzyme B from T cells induced by anti-PD-L1 mAb. In summary, these results indicated that 4-1BB agonist in combination with anti-PD-L1 ICI potentiated cytolytic capacity of infiltrating T cells.

CD8⁺ T cell was essential to the antitumor efficacy elicited by the combined therapy

To illuminate which T cell subsets were indispensable to the combined therapy-mediated tumor regression, we depleted $CD4^+/CD8^+$ T cells in mice using CD4/8 antibodies, respectively. We then constructed the A20 subcutaneous



tumor model and treated the mice with a combination of anti-PD-L1 antibody and 4-1BB agonist. Depleting $CD8^+$ T cell completely abolished the therapeutic effect, while $CD4^+$ T cell depletion had no obvious effect on the combined therapy (Figure 5). The above results confirmed that $CD8^+$ T cells played an essential role in the combined therapy.

Discussion

Considering the fact that PD-1/PD-L1 axis confers cancer cells evasion from hosts' immune system, digging excellent anti-PD-1/PD-L1 ICIs will inspire patients against malignancies (24). However, only a small subset of patients received benefits from these ICIs, of which the absence and exhaustion of tumor-infiltrated cytotoxic T lymphocytes are the main symptoms (25, 26). As one of the prominent co-stimulators (19, 27), 4-

1BB signaling could stimulate T cell proliferation and activation, and enhance the cytotoxicity of adaptive T therapy (28–30). Herein, we determined the synergistic effect of anti-PD-L1 therapy in combination with 4-1BB agonists in BCL.

TIL in tumor environment is considered to be the crucial factor determining the antitumor immunity of PD-1/PD-L1 blockade (31, 32). Various efforts have been made to enhance antitumor immunity of tumor infiltrating lymphocytes. Fusion protein consisting of chemokine CCL4 and collagen-binding domain was applied to recruit TILs to improve the antitumor immunity of anti-PD-L1 monotherapy in multiple tumor models (33). Positive correlation has been uncovered in elevated collagen and exhausted T cells in lung cancers. Reducing tumor collagen deposition could increase infiltration of CD8⁺ T cells and overcome resistance to anti-PD-L1 therapy (34). Physical therapies including thermotherapy and radiation therapy were also reported to promote T cell infiltration and



cells. (C) Genes were classified and grouped based on their molecular function. (D) Perforin, IFN- γ , and granzyme B in tumors were measured by RT-PCR or ELISA after combinational treatment (n = 3, mean \pm SD). * means p-value < 0.05, ** means p-value < 0.01, and *** means pvalue < 0.001



FIGURE 5

T cell subsets in the antitumor efficacy of anti-PD-L1 mAb combined with 4-1BB agonist. The antitumor effect of the combined therapy was abolished by CD8⁺ T cell depletion. (A, B) Tumor volume was measured twice a week. (C) Tumor weight was measured at the end of the treatment. (n = 4, mean \pm SD). ** means p-value < 0.01. ns means no significance.

synergize with anti-PD-L1 treatment (35). In addition, activating 4-1BB mAb was found to increase the ratio of tissue-resident T cells in pulmonary and hepatocellular carcinoma (36, 37). Bispecific antibodies MCLA-145 and ABL503 were generated to evaluate the antitumor effect and liver toxicity of co-targeting 4-1BB and PD-L1 (21). Results indicated that high dose of bispecific antibody (10 mg/kg) could induce strong antitumor efficacy with low liver toxicity in MC38 tumor model (38). Our research showed that anti-PD-L1 mAb in combination with low dose of 4-1BB agonistic Ab (1 mg/kg) elicited synergistic antitumor activity with no obvious toxicity, whereas PD-L1 blockade or 4-1BB agonism alone only had modest antitumor effects. Considering the significant discrepancy in dosage, combination therapy targeting 4-1BB and PD-L1 is still a potential option in the clinic.

Analyzing tumor-infiltrating T cells would not only contribute to investigating the T cell subsets, but also provide insights into the function of tumor-specific cytotoxic T lymphocytes. Generally, T cells are essential to eliminate tumor cells and higher cytotoxic T lymphocytes infiltration in tumor correlates with better prognosis (39-42). While characteristics underlying the infiltrated T lymphocytes in TME are still unclear, and functions of TILs in the context of anti-PD-L1 ICI and 4-1BB agonist have not been fully elucidated. Here, we observed that targeting 4-1BB and PD-L1 not only activated T cell immunity, but also promoted recruitment of effector T lymphocytes into TME. We also analyzed the transcriptional profile of TME, and data indicated that anti-PD-L1 mAb in combination with 4-1BB agonist resulted in reinforcing CD8⁺ T-cell immunity via cytokine and chemokine signaling pathway. Furthermore, T cell depletion confirmed that the effect of combinational therapy was dependent on CD8⁺ T cells in BCL.

Currently, ICI has elicited promising antitumor effects in several clinical trials. PD-1 monoclonal antibody nivolumab achieved an objective remission rate of 87% in a phase I clinical trial against refractory recurrent classic Hodgkin's lymphoma (43). In a phase II clinical trial, the objective remission rate reached 68% with overall survival of 6 months (44). However, the efficacy of PD-1 monoclonal antibodies in the treatment of non-Hodgkin's lymphoma is not satisfactory, with objective remission rates of only 40% and 36% for nivolumab alone in clinical trials for refractory relapsed follicular lymphoma and refractory relapsed diffuse large B-cell tumor (45, 46). Studies have shown that PD-L1 ligands are associated with 9p23-24 gene amplification (47) and that the 9p24 gene has a higher probability of mutation in Hodgkin's lymphoma cells (48), whereas the gene is rarely altered in non-Hodgkin's lymphoma cells (47), thus PD-1/PD-L1 blockade has shown better efficacy in the treatment of Hodgkin's lymphoma.

To further enhance the efficacy of PD-1/PD-L1 monoclonal antibodies in the treatment of B-cell lymphoma, various combination regimens have been proposed, including other immune checkpoint inhibitors, co-stimulatory molecular agonists, and other types of therapeutic antibodies. An objective remission rate of 80% was achieved in clinical trials combining the PD-1 monoclonal antibody pembrolizumab and the CD20 monoclonal antibody rituximab for the treatment of relapsed follicular lymphoma (45). Complete remission rates of 50%-65% were achieved using PD-1 monoclonal antibody nivolumab in combination with brentuximab vedotin, an ADC drug targeting CD30, for refractory relapsed Hodgkin's lymphoma (49).

In conclusion, this paper demonstrated that the combination of 4-1BB agonist and anti-PD-L1 antibody can activate T cell function and increase the expression of cytokines such as IFN- γ . On the other hand, it can release the inhibition of T cells by tumor cells by blocking the PD-1-PD-L1 signaling axis. Thus, eliciting a more significant anti-tumor effect (Figure 6). Our results confirmed the synergetic antitumor effect of anti-PD-L1 mAb and 4-1BB agonist, providing an effective approach to treating BCL.

Materials and methods

Antibodies

Anti-PD-L1 mAb was supplied by Shanghai Hankon Biosciences Co. Ltd (Shanghai, China). 4-1BB agonistic antibody was provided by Eutilex Co., Ltd (Korea). APC anti-CD3 antibody, PE anti-CD8a antibody, and PerCP/Cyanine5.5 anti-CD4 antibody were purchased from BioLegend, Inc.

Cell culture

A20 and WEHI-231 were purchased from Nanjing Cobioer biotechnology company and cultured in the indicated medium with 10% FBS, and 50 μ M B-mercaptoethanol. The cells were incubated at 37 °C and 5% CO₂.

In vivo therapeutic effect

A20 and WEHI-231 subcutaneous graft tumor models were constructed. The cells were collected by low-speed centrifugation (1200 rpm, 5 min), resuspended in sterile phosphate buffer, and the cell concentration was adjusted to a final concentration of 2.5×10^7 cells/mL. BALB/c mice were ready for tumor



inoculation at six weeks of age, weighing about 20 g. Each mouse was injected with 5×10^6 cells, as soon as the tumor volume reached 100 mm³, the mice were randomly divided into groups and treated. Body weight and tumor diameter (a=long diameter, b=short diameter) were measured and recorded twice a week. Tumor volume = $\frac{a \times b^2}{2}$.

Relative tumor weight = $100 \% \times \frac{\text{control group-treatment group}}{\text{control group}}$.

To construct the A20 metastatic tumor model, the cell concentration was adjusted to 1×10^7 cells/mL using sterile PBS. 100 µL of the cell suspension was injected into the mice through the tail vein, i.e., 1×10^6 cells/mouse. Treatment was started the day after injection, Mice were treated for one week and administered twice a week (Anti-PD-L1 10mg/kg, 4-1BB agonist 1mg/kg). The survival of mice was monitored.

RNA-seq

RNA extraction

TRIzol[®] Reagent (Invitrogen) and DNAase I (TaKara) were utilized for total RNA extraction and genomic DNA removal respectively. The quantity and quality of RNA were detected by the ND-2000 (NanoDrop Technologies) and 2100 Bioanalyse (Agilent). The next experiment can only be performed if the RNA meets the following requirements. OD260/280 = $1.8 \sim 2.2$, OD260/230≥2.0, RIN≥6.5, 28S:18S≥1.0, >1µg

Library preparation, and Illumina Hiseq xten/ Nova seq 6000 sequencing

Firstly, mRNA was isolated from 1 μg of total RNA and fragmented. Then double-stranded cDNA was synthesized and modified with end-repair, 'A' base addition and

phosphorylation. The 300bp size cDNA was isolated and amplified using PCR. After quantification, sequencing was performed on Illumina Hiseq xten/Nova seq 6000 with read length = 2×150 bp.

Read mapping

SeqPrep and Sickle for quality control. HISAT2 for clean reads aligning to reference genome. StringTie for the assembly of mapped reads.

Differential expression analysis and functional enrichment

The expression level was determined by the transcripts per million reads. Q value ≤ 0.05 (DESeq2) was considered to be significantly different. KEGG pathway analysis was carried out by KOBAS.

GSEA analysis

GSEA was performed with GSEA v3.0 (http://www. broadinstitute.org/gsea/). Gen sets were obtained from MSigDB (http://www.broadinstitute.org/gsea/msigdb).

Flow cytometry

4.5.1 Preparation of single cells suspensions of tumor tissues

Mice were executed to obtain tumor tissue. The tumors were cut into small pieces and placed on a cell sieve with PBS for grinding. The collected grinds were centrifuged to remove the supernatant. After 5 min, the supernatant was discarded by centrifugation, washed once with PBS, and PBS was added again to obtain a single-cell suspension.

Antibody staining

The supernatant was removed by centrifugation according to the instructions, and the blocking antibody was added and blocked for 30 min at 4°C. The blocking antibody was then removed by centrifugation and the flow-labeled antibody (Antimouse CD3 (APC), anti-mouse CD4 (PerCP/Cy5.5), anti-mouse CD8a (PE) (Biolegend)) was added and incubated for a halfhour at 4°C in dark. Centrifuge to remove supernatant, wash twice with PBS, and resuspend in 300µl PBS.

Immunohistochemistry

Mouse tumors were fixed in 4% paraformaldehyde. After the fixed tissues were embedded in wax blocks and sectioned, the tissue sections were stained using CD 8 antibody. The infiltration of CD8-positive cells in the tissues was examined and analyzed by light microscopy.

T cells depletion

All antibodies used for T-cell depletion were purchased from bioxcell. Mice were injected intraperitoneally with an anti-CD4 antibody ($200\mu g$, colone GK1.5), anti-CD8 antibody ($200\mu g$, colone 2.43), and Isotype control (IgG2b, $200\mu g$, colone LTF-2) on days 1, 3, 5, and 7, respectively, and then the antibodies were injected once a week until the end of the experiment.

Statistical analysis

Data in this study were analyzed by GraphPad Prism 9. Comparison was determined by Student's *t*-test and One-Way ANOVA analysis, and P value< 0.05 was considered as statistical significance.

Data availability statement

The original contributions presented in the study are publicly available. The data is deposited in the SRA database, accession number: PRJNA904505.

Ethics statement

The animal study was reviewed and approved by Animal Ethical Committee of School of Pharmacy at Fudan University.

Author contributions

DJ and XZh designed the study. YW, XZh, CX and YN performed the experiments and wrote the paper. YW, XZh, JF, XZe, and BK analyzed the data. All authors contributed to the article and approved the submitted version.

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Conflict of interest

BK is the founder and Chief Executive Officer of Eutilex Co., Ltd.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fimmu.2022.1004475/full#supplementary-material

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