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Abstract: The development of nanotechnology will inevitably lead to the release of consistent amounts of nanomaterials (NMs) and nanoparticles (NPs) into marine ecosystems, with potential adverse effects for marine biota. Up to date, ecotoxicological studies reported a wide variety of biological injuries in marine species belonging to different trophic levels, with the general aim of identifying potential biological targets of NPs, as well as suitable models for risk assessment and prediction of their impact on the health of the marine environment. From these data, different invertebrate species emerged as significant targets for the impact of NPs, with bivalves representing by large the most studied group, whereas less attention has been focused on sediment dwelling species. However, despite the development of fate models and analytical and biological tools for assessment of marine ecosafety of NPs, still major scientific gaps need to be filled. In this work, information on the factors affecting the behaviour of NPs released in the marine environment, and consequent uptake/accumulation/toxicity by representative invertebrate groups will be summarized. Available data on the effects of different NPs in marine invertebrates will be reviewed, in particular with regards to those reported on immune function, oxidative stress and development. Moreover, the possibility that the effects of NPs in marine invertebrates may be influenced by their interactions with biomolecules both in the external environment and within the body fluids, forming an eco-bio-corona, will be introduced. In the framework of the general need to classify NPs into groups and predict the implications of their release into the marine environment, information on their intrinsic properties is clearly insufficient, and a deeper understanding of NP-eco/bio-interactions is therefore urgently required.

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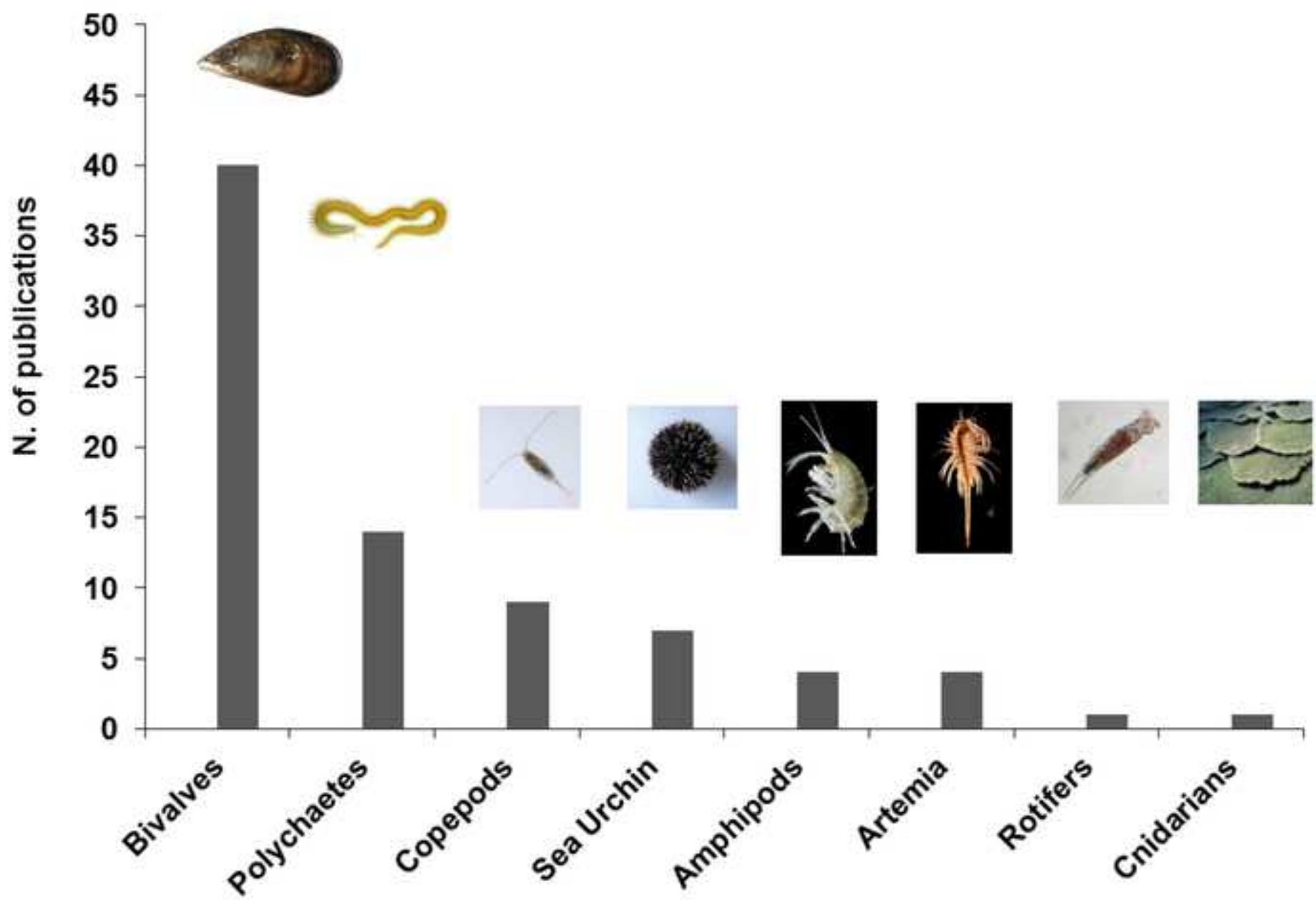
Dear Editor,

I send you the ms. 'Effects of nanomaterials on marine invertebrates' to be considered for publication in the Special Issue of STOTEN: 'Nanomaterials and environment: trends and perspectives'.

I thank you very much for your kind attention

Sincerely Yours

Laura Canesi



## Highlights

- Marine invertebrates are significant targets for NPs released in the environment
- Most data on bivalves and echinoderms, much less in sediment dwelling species
- Main biological responses: immunomodulation, oxidative stress, embryotoxicity
- NPs interactions with environmental/biological components affect their fate/toxicity
- More information on eco/bio-interactions of NPs in the marine environment is needed

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3 **Effects of nanomaterials in marine invertebrates**  
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**Abstract**

The development of nanotechnology will inevitably lead to the release of consistent amounts of nanomaterials (NMs) and nanoparticles (NPs) into marine ecosystems, with potential adverse effects for marine biota. Up to date, ecotoxicological studies reported a wide variety of biological injuries in marine species belonging to different trophic levels, with the general aim of identifying potential biological targets of NPs, as well as suitable models for risk assessment and prediction of their impact on the health of the marine environment. From these data, different invertebrate species emerged as significant targets for the impact of NPs, with bivalves representing by large the most studied group, whereas less attention has been focused on sediment-dwelling invertebrates (benthic infauna). However, despite the development of fate models and analytical and biological tools for assessment of marine ecosafety of NPs, still major scientific gaps need to be filled. In this work, factors affecting NP fate in the marine environment, and consequent uptake/accumulation/toxicity in representative invertebrate groups will be summarized. Available data on the effects of different NPs in marine invertebrates will be reviewed, with particular regard to those reported on immune function, oxidative stress and embryo development. Moreover, the possibility that such effects may be influenced by NP interactions with biomolecules both in the external environment and within the body fluids, forming *eco-bio-coronas*, will be introduced. In the framework of the general need to classify NPs into groups and predict the implications of their release into the marine environment, information on their intrinsic properties is clearly insufficient, and a deeper understanding of NP eco/bio-interactions is therefore urgently required.

## 1. Introduction

Recent environmental fate models underline that *nanowastes* will end up in the aquatic environment, thus potentially affecting natural ecosystems and human health (Bystrzejewska-Piotrowska et al., 2009; Brar et al., 2010; Liu et al., 2014). Marine ecosystems, historically seen as a major sink of anthropogenic contaminants, will receive consistent amounts of nanomaterials (NMs) and nanoparticles (NPs), whose intrinsic and extrinsic complexity are making the prediction and assessment of their fate, exposure and biological effects a major challenge (Klaine et al, 2008). As recently discussed in our overview on fate models and tools (analytical and biological) for *ecosafety* assessment and design of nanomaterials entering the marine environment (Corsi et al., 2014), still major scientific gaps need to be filled. The development for instance of quantitative approaches for integrating more realistic NP exposure scenarios and predicting ecosystem impacts are urgently needed for a proper risk assessment (Gottschalk et al., 2013). Up to date, ecotoxicological studies focusing on marine organisms reported a wide variety of biological injuries in species belonging to different trophic levels, from planktonic to benthic species, including fish, with the general aim to identify potential biological targets as well as suitable models for both risk assessment and prediction of marine environmental implications (Minetto et al., 2014; Baker et al., 2014; Matranga and Corsi, 2012). In order to achieve this goal, basic information on the factors affecting the behaviour of NPs released in the marine environment, and consequent uptake/accumulation/toxicity by representative species must be taken into account. The importance of evaluating the potential impact of NP exposure on aquatic invertebrates has been underlined (Baun et al., 2008; Canesi et al., 2012; Baker et al., 2014; Corsi et al., 2014; Matranga and Corsi, 2014; Canesi et al., 2015; Rocha et al., 2015). The first studies were mainly carried out in freshwater species utilizing standard ecotoxicity tests, with less than 20% of published papers on marine species (Cattaneo et al., 2009). However, the number of publications on NP ecotoxicity in invertebrates has exponentially grown in the last few years, with a strong contribution of those



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3 focused on NP accumulation, sub-lethal effects and mechanisms of action. Among these, the  
4 percentage of studies on marine species has risen considerably (up to 38% of the total) (Fig. 1A).  
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6 Bivalve molluscs are so far the most studied marine invertebrate group (50%), whereas less  
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8 attention has been focused on sediment-dwelling invertebrate (benthic infauna) (about 20%) (Fig.  
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10 1B).

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12 *Mytilus* spp. represents the most utilized bivalve model: special emphasis has been given to  
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14 the effects of different types of NPs on the immune system and on the main tissues involved in NP  
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16 uptake and accumulation, i.e. the gills and the hepatopancreas (reviewed in Canesi et al., 2012,  
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18 2015; Rocha et al., 2015). Scattered information is available on other bivalve species and marine  
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20 invertebrate groups (echinoderms, anellids). In this work, those aspects related to the effects of NPs  
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22 on immune function and oxidative stress in different invertebrate species will be analyzed in more  
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24 detail. Moreover, whether such effects may be influenced by NP interactions with biomolecules  
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26 both in the external environment and within the body fluids, forming *eco-bio-coronas*, will be  
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28 introduced. Finally, available data on the effects of NPs on development of marine invertebrate  
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30 species will be summarized.

## 31 32 33 34 35 **2. Behavior of different particles in SW**

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37 Environmental implications of NPs are strongly linked to their peculiar features such as particle  
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39 surface charge, size, shape, functionalization and coating, all properties that affect their interaction  
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41 with the surrounding media and the resulting fate and toxicity (Klaine et al., 2008). For instance,  
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43 dispersion and aggregation or agglomeration in aqueous solutions are driven by size and surface  
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45 charges of the NPs, but also by several other parameters of the receiving media such as pH, ionic  
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47 strength (osmolarity) and presence of natural organic matter (NOM) (Corsi et al., 2014; Keller et  
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49 al., 2010; Petosa et al., 2010). Natural environmental scenarios are currently barely represented in  
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51 nanoecotoxicological studies, this being probably mainly related to substantial lack of information  
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53 on interactions between specific NPs and the exposure media as, for instance, natural sea water  
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3 (NSW) vs artificial sea water (ASW). The high ionic strength which also affect pH, as well as  
4 organic matter content, have been reported to play a significant role on NP aggregation and thus  
5 affecting their fate in the water column and sediment (Chen et al., 2011). In our recent work with  
6 marine mussels *Mytilus galloprovincialis*, co-exposure to titanium dioxide NPs (nano-TiO<sub>2</sub>) and  
7 CdCl<sub>2</sub> in ASW resulted in reduced Cd toxicity and increases Ti accumulation in selected tissues (i.e.  
8 gills) (Della Torre et al., 2015). On the contrary, in the digestive gland such effect was not  
9 observed, suggesting that the fate of NPs within the organism may be also related to interactions  
10 with biomolecules which affect NP state and effects. Bioavailability, as well as biodistribution and  
11 consequent biological responses, are highly dependent on exposure mechanisms, but also on fate of  
12 NPs inside the body of the organism itself. Further studies on exposure assessment and fate into the  
13 aquatic environment, as well as into the internal environment of exposed organisms are strongly  
14 needed.  
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17 Both heteroaggregation and homoaggregation are driven by NP surface charges and the  
18 chemical composition of the surrounding media, due to the presence of elevated concentrations of  
19 inorganic ions as well as organic molecules such as polysaccharides, proteins and colloids  
20 (Praetorius et al., 2014; Zhou et al., 2013). Moreover, the presence of functional groups on the  
21 surface of NPs, more than their core composition (metal-based, carbon-based, etc) is now being  
22 considered to play a major role in sorption to biofilms in high ionic strength media such as in NSW.  
23 Single NPs may barely adsorb NOM, while “bridging effects” due to colloids interacting with  
24 multiple particles can increase aggregation. For instance, colloidal polymers produced by algae  
25 known as exopolymeric substances (EPS) can influence aggregation and transformation of NPs in  
26 aqueous media, mainly as a function on the hydrophobic and electrostatic interactions between NPs  
27 and EPS (Nevius et al., 2012; Therezien et al., 2014; Adeleye et al., 2014a,b; Kadar et al., 2014).  
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30 Interactions of NPs with biomolecules have been described within biological fluids of  
31 mammalian models, where plasma proteins originate a coating known as protein *corona*, which  
32 affect cellular uptake (biokinetics) and ultimate toxicity (Lundqvist et al., 2011; Monopoli et al.,  
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3 2011; Salvati et al., 2011). The formation of a NP corona has been recently reported also in  
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5 terrestrial and freshwater invertebrate species, i.e. secreted proteins from the coelomic fluid of the  
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7 rugworm *Eisena fetida* and from *Daphnia magna* neonates (Hayashi et al., 2015; Nasser and Linch,  
8  
9 2015), and may also occur in marine invertebrates (see below).

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11 Whether these processes could take place also in the external environment, for example in  
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13 NSW as consequence of interactions with NOM, as well as within biological fluids of marine  
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15 organisms, deserves further investigation. NP extrinsic properties related to the formation of an *eco-*  
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17 *corona* or bio (protein) *corona* will affect consistently their ultimate fate in the water column and  
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19 therefore biological activity and toxicity (Fig. 2). NP surface charge and size, irrespective to core  
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21 composition, have been reported to play a significant role in cellular adhesion, particle recognition  
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23 as well as efficiency of uptake. We recently showed that polystyrene NP (PS NPs) having cationic  
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25 (-NH<sub>2</sub>) or anionic (-COOH) surface groups undergo different aggregation in NSW affecting their  
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27 biodistribution and ultimate toxicity both in larvae of brine shrimp *Artemia franciscana*, as well as  
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29 sea urchin embryos *Paracentrotus lividus* (Della Torre et al., 2014; Bergami et al., 2015). Anionic  
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31 PS NPs undergo significant aggregation in NSW and retention in the gut lumen of exposed  
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33 organisms, while cationic PS NPs showed far less aggregation in NSW but clear signs of toxicity.

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35 Several mechanisms can be involved in NP uptake but how they are dispersed in exposure  
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37 media may result in the absence or presence of a *eco/bio-corona*, like it may occur in artificial vs  
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39 natural sea water or in different body fluids (Salvati et al., 2011; Liu et al., 2011; Rossi et al., 2014).  
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41 Moreover, in marine coastal areas significant changes in NOM and ion concentrations are  
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43 influenced by the season and river flows, which might consistently affect NP interactions and  
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45 ultimate fate towards both planktonic and benthic invertebrates (Corsi et al., 2014).

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47 In the framework of a general need to classify NPs into groups and predict environmental  
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49 implications once they are released into the marine environment, information on their intrinsic  
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51 properties is clearly insufficient, and a deeper understanding of *eco/bio-interactions* is therefore  
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3 urgently needed. Moreover, ecotoxicity studies based on realistic environmental scenarios could be  
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5 valuable tools for both screening and categorizing NPs for risk assessment purposes.  
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### 8 9 **3. Uptake and accumulation**

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12 NPs surface charges (positive or negative), size and shape, irrespective to core composition, have  
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14 been recently showed to drive cellular adhesion, particle recognition but more important uptake  
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16 efficiency (Lunov et al., 2011; Varela et al., 2012). Moreover, surface charge, more than size, has  
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18 an important role on cellular uptake in human cells, as for instance for polymeric NPs, which are  
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20 consequently more efficiently internalized (Bramini, 2014; Lesniak et al., 2010; Liu et al., 2011;  
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22 Lunov, 2011; Rossi et al., 2014). Cationic polystyrene NPs (PS-NH<sub>2</sub>) have been shown to be  
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24 cytotoxic for mammalian cells *in vitro*, while anionic PS NPs (PS-COOH) exhibited greater and  
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26 rapid accumulation in selected organs such as lung (Bexiga et al., 2011; Wang et al., 2013a,b).  
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28 Recent findings on sea urchin embryos and brine shrimp larvae confirmed such behavior; a  
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30 significant gut retention is observed for anionic PS-COOH NPs while cationic PS-NH<sub>2</sub> NPs showed  
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32 respectively developmental toxicity and increase of molts (Della Torre et al., 2014; Bergami et al.,  
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34 2015). More contributions on other marine invertebrate species confirm a significant accumulation  
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36 of anionic PS NPs in the gut of rotifers (Snell and Hicks, 2011) and in bivalves (Ward and Kach,  
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38 2009; Wegner et al., 2012). Based on our past findings, nanoscale aggregates of PS-NH<sub>2</sub> (~ 100 nm)  
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40 are still present in NSW media while PS-COOH NPs originated microscale aggregates (>900 nm).  
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42 The formation of aggregates as a consequence of interaction with dispersing media has a strong  
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44 influence on their route of uptake and biodistribution. Bioaccumulation seems in general not  
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46 affecting mortality, but it has been associated with several sub-lethal effects (behavioral,  
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48 physiological and biochemical) able to affect species survival in prolonged exposure scenarios.  
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50 Accumulation of NP aggregates in the digestive tract, as observed for PS-COOH NPs, can  
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52 significantly limit food intake and disrupt growth and development of planktonic invertebrate larvae  
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3 (Bergami et al., 2015; Besseling et al., 2014). In addition, the adsorption of hydrophobic aquatic  
4 pollutants by nano-debris may increase their bioavailability and then cellular uptake once  
5 accumulated in the gut (Rochman et al., 2013; Velzeboer et al., 2014). As for inorganic NPs, a  
6 general mechanism of uptake of aggregates by endocytosis in the digestive system and individual  
7 NPs or ionic forms in the gills, respectively, has been hypothesized, despite some recent evidence  
8 of an higher accumulation of nano- compared to micro-metric sized ones has been reported for  
9 soluble metals as gold and copper (both ions and NPs) (Canesi et al., 2010; Gomes et al., 2011,  
10 2012; Dai et al., 2013). Therefore, the ability to act as carriers of toxic contaminants seems affected  
11 by their dispersion in exposure media and linked also to absence or presence of NP-  
12 eco/biomolecular coronas (Monopoli et al., 2011; Lesniak et al., 2013; Salvati et al., 2011).  
13 Moreover, how both type and fate of the corona (hard and soft corona) will affect uptake and  
14 bioaccumulation, as reported in mammalian systems, still represents a major challenge and deserve  
15 further investigation in both adults and larval/embryo stages of marine invertebrate species.  
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30 In bivalves for instance mucus, fecal pellets and gametes can have a significant role in  
31 bioavailability of the NPs but also on uptake by different organs (i.e. gills or digestive gland) (Della  
32 Torre et al., 2015; Rocha et al., 2015). At the level of the cell membrane, ATP-binding cassette  
33 transporters belonging to the ABC family have been shown to have a significant a role in NP  
34 biodistribution. As recently reported in Caco-2 gut epithelial cells, nano-TiO<sub>2</sub> (anatase) significantly  
35 increases the functionality of the cells, then forming a protective barrier against exogenous toxicants  
36 and on nutrient absorption (Dorier et al., 2015). Our recent studies on mussels exposed to nano-  
37 TiO<sub>2</sub> showed similar results with a significant increase in efflux of RhB, a P-glycoprotein substrate,  
38 from the gills, suggesting that this protective role could be also shared by selected barrier organs as  
39 gills in marine invertebrates (Canesi et al., 2014; Della Torre et al., 2015). Further studies are thus  
40 needed in order to clarify the role of efflux activity of ABC transporters in marine invertebrates  
41 upon exposure to selected NP and therefore on their toxicity.  
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#### 4. Effects and mode of action of NPs in marine invertebrate models

##### 4.1 Effects on immune function

Increasing evidence supports the hypothesis that in marine invertebrates the immune system represents a significant target for the effects of NPss. Invertebrates lack an adaptive immunity; however, they are endowed with a potent and complex innate immune system (humoral and cellular defenses) similar to that of vertebrates. In most invertebrates, free circulating cells (hemocytes/coelomocytes) are responsible for cell-mediated immunity; when activated, different hemocyte types are capable of phagocytosis, Reactive Oxygen Species (ROS) and NO production, release of hydrolytic enzymes and antimicrobial peptides. Conservation of the general mechanisms of innate immunity from invertebrates to mammals is a key feature that represents an useful basis for evaluating the environmental impact of NPs (Canesi and Procházková, 2013).

In *M. galloprovincialis*, *in vitro* studies showed that different NP types (carbon black, fullerenes, and different nanosized metal-oxides) are rapidly taken up by mussel hemocytes, affecting different functional parameters, from lysosomal function to phagocytic activity and oxyradical production, and also inducing pro-apoptotic processes; the effects were mediated by stress-activated mitogen activated protein kinase (MAPK) signaling, as in mammalian phagocytes, and were observed in the low  $\mu\text{g/ml}$  range (Canesi et al., 2012; Canesi and Procházková, 2013). Recent studies also showed that amino-modified PS NPs (PS-NH<sub>2</sub>) induced significant activation of immune parameters at lower concentrations (1-5  $\mu\text{g/ml}$ ), whereas lysosomal damage and apoptotic process were observed at higher concentrations (50  $\mu\text{g/ml}$ ) (Canesi et al., 2015). Different NPs were shown to target the phagocytic activity also in the hemocytes of the oyster *Crassostrea gigas* (Abbott Chalew et al., 2012) and of the clam *Ruditapes decussatus* (Marisa et al., 2015). Interestingly, induction of functional responses, as well as of cellular damage and apoptotic

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3 processes, were particularly rapid, occurring within 1 h of exposure. This is in line with the  
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5 physiological role of bivalve hemocytes, that represent the first line of defence against non-self  
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7 material (Canesi et al., 2002; Canesi and Procházková, 2013).

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9 Another *in vitro* approach has been successfully utilized to evaluate NP-mediated cytotoxicity in  
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11 *Mytilus* hemocytes after prolonged exposure (24 h) to different types of NPs (CdS quantum dots,  
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13 nanosized Au and Ag NPs, n-ZnO, n-SiO<sub>2</sub>, n-TiO<sub>2</sub>) in a wide concentration range; the effects were  
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15 compared with those observed in isolated gill cells (Katsumiti et al., 2014, 2015a,b,c). These studies  
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17 underlined different cytotoxic effects in terms cell viability (EC<sub>50</sub>), and parameters related to  
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19 oxidative stress, genotoxicity, cytoskeletal alterations, and membrane transport activities.  
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21 Moreover, various degrees of toxicities were observed in hemocytes and gill cells that were related  
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23 to the NP mode of synthesis, crystalline structure and size, presence of additives in the suspensions,  
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25 metal ion release in high ionic strength medium, etc.

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27 Overall, the application of a battery of functional/toxicity tests on bivalve hemocytes proved as  
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29 a powerful tool for the screening of the immunomodulatory/immunotoxic effects of different types  
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31 of NPs in cell models of marine organisms, as well as a robust alternative method for testing the  
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33 toxicity of NPs and a possible basis for a ‘safety by design’ approach (reviewed in Canesi et al.,  
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35 2012; Canesi and Procházková, 2013).

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37 The results obtained *in vitro* on mussel immune cells were confirmed by *in vivo* exposure to  
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39 different NPs, and in particular to n-TiO<sub>2</sub> chosen as a model NP type (Canesi et al., 2010b; Barmo  
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41 et al., 2013). These data allowed formulating an hypothesis on the possible pathways leading to  
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43 nano-induced immunomodulation (Canesi and Procházková, 2013; Corsi et al., 2014). Due to the  
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45 physiological mechanisms involved in the feeding process, n-TiO<sub>2</sub> agglomerates/aggregates formed  
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47 in sea water are taken up by the gills and partly directed to the digestive gland. NPs can be then  
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49 potentially translocated from the digestive system to the hemolymph, and to circulating hemocytes,  
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51 where n-TiO<sub>2</sub> induced changes in functional parameters, as well as in transcription of different  
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53 antimicrobial peptides and downregulation of Toll-like receptors (Barmo et al., 2013; Balbi et al.,  
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3 2014). Interestingly, these effects were observed at concentrations much lower than those usually  
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5 utilized in ecotoxicity tests on aquatic species (1-100 µg/L), and closer to predicted environmental  
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7 concentrations (PEC) (Gottschalk et al., 2013).

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9 Sea urchin immune cells represent another suitable model for testing the effects of NPs on the  
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11 immune system of marine invertebrates . Well established as sensors of environmental hazards,  
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13 coelomocytes of the mediterranean species *Paracentrotus lividus* have been recently showed to  
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15 activate/express specific cell stress proteins upon n-TiO<sub>2</sub> exposure both in *in vitro* and *ex vivo*  
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17 (Pinsino and Matranga, 2015; Pinsino et al., 2015). Moreover, *in vivo* exposure elicited a  
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19 phagocytic mechanism, involving the TLR/p38 MAPK signaling pathway (Pinsino et al., 2015).  
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21 This species has been used to study the *in vivo* potential toxicity of a few selected metal oxide NPs  
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23 (i. e. SnO<sub>2</sub>, CeO<sub>2</sub>, Fe<sub>3</sub>O<sub>4</sub>), introduced by forced ingestion (Falugi et al., 2012). After 5 days, nano-  
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25 aggregates/agglomerates were found inside the sea urchin immune cells (coelomocytes), causing  
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27 modifications of the trans-Golgi and ER subcellular compartments, inhibiting the activity of the  
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29 three ChE isoforms and affecting the expression levels of the stress proteins HSC70 and GRP78  
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31 (Falugi et al., 2012).. In contrast to what expected by analogy with bivalve molluscs, injection of n-  
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33 TiO<sub>2</sub> into the coelomic cavity did not cause a stress response in the sea urchin, nor immune cell  
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35 death or autophagy and lysosomal dysfunction (Pinsino et al., 2015).  
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38 Phenoloxidase PO and lysozyme activities were induced by AgNP aggregates in the clam  
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40 *Scrobicularia plana* and in the polychaete *Nereis (Hediste) diversicolor*, respectively (Mouneyrac  
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42 et al., 2014). In the coelomocytes of *H. diversicolor*, AgNPs induced lysosomal and DNA damage  
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44 (Cong et al., 2014). These results indicate that NPs may affect the function of immune cells also in  
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46 marine worms.

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48 The availability of the sea urchin genome, shown to be in close phylogenetic relationship to the  
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50 humans (Sea Urchin Genome Sequencing Consortium, 2006), offers the possibility to analyze its  
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52 complex and sophisticated immune system and compare the biological effects observed in sea  
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54 urchins and human phagocytes. Although the *Mytilus* genome is not available yet, the rapidly



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3 expanding application of DNA microarrays and Next Generation Sequencing (NGS) is leading to  
4 the identification of an increasing number of immune-related genes that could represent the target  
5 for different NPs also in mussels (Phillip et al., 2012; Gerdol and Venier, 2015). In this light, both  
6 the mussel and the sea urchin represent promising models for evaluating the effects of NPs on the  
7 immune system of marine invertebrates.  
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#### 10 11 12 13 14 15 16 *4.2 Effects of NPs at tissue level*

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18 Oxidative stress and cell injury in proteins, membrane and DNA damage have been so far  
19 identified as the major modes of action of NPs in bivalve tissues (reviewed in Rocha et al., 2015).  
20 However, the paradigm proposed to explain most of the adverse effects exerted by NP exposure in  
21 mussels, that is all the effects are directly or indirectly mediated by reactive oxygen species (ROS)  
22 and free radical production, cannot be applied to all NP types. Moreover, oxidative damage  
23 induced by NPs in bivalves depends on particle size, composition and concentration, mode and  
24 time of exposure, as well as the target organ analyzed.  
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32 In *Mytilus* gills, short-term exposure (24 h) to both carbon based NPs (Nano carbon black-  
33 nNCB, C60 fullerene), and n-oxides (n-TiO<sub>2</sub> and n-SiO<sub>2</sub>) (0.05, 0.2, 1, 5 mg/L) induced changes in  
34 catalase and GST activities, with both increases and decreases depending on NP type and  
35 concentration; however, antioxidant enzymes did not seem to represent a significant target for these  
36 types of NPs (Canesi et al., 2010b). Longer exposure to n-TiO<sub>2</sub> (0.1 mg/L, 96 h) increased P-gp  
37 efflux activity and NO production; this latter effect may be mainly due to infiltrating hemocytes,  
38 since no changes in antioxidant enzyme activities were observed (Della Torre et al., 2014).  
39 Although the gills of *C. gigas* and *M. galloprovincialis* are susceptible to oxidative stress induced  
40 by ZnO and CuO NPs, this was not the case with other metal-based NPs (reviewed in Rocha et al.,  
41 2015). Gene transcription profiles were not significantly altered in the gills of *M. galloprovincialis*  
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3 exposed to Ag NPs after 15 days, although changes in expression may occur at shorter times of  
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5 exposure (Bebianno et al., 2015).

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7 As described above, NP behavior (aggregation/agglomeration) in both ASW and NSW is a key  
8  
9 factor in their accumulation and effects in different tissues. Although mussel gills are the first  
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11 organ/barrier to surrounding water, being more vulnerable to particle interactions, uptake of  
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13 individual NPs or ionic metal forms seems to mainly occur in this tissue, while NP agglomerates  
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15 may enter in the digestive system of the organism mainly by endocytosis (Corsi et al. 2014; Rocha  
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17 et al., 2015).

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19 In bivalves, the digestive gland is therefore considered the main organ for accumulation of NPs,  
20  
21 where their cellular fate and effect differ according to the NP type and experimental conditions. The  
22  
23 endosomal-lysosomal system of digestive cells, due to its role in intracellular digestion of food  
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25 particles, represents the main subcellular target for NPs in bivalves (Moore, 2006; Canesi et al.,  
26  
27 2012). In *Mytilus* digestive gland, acute exposure (24 h) to NP suspensions (NCB, C60, n-TiO<sub>2</sub>, n-  
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29 SiO<sub>2</sub>) induced lysosomal membrane destabilization, decreases in antioxidant defenses and  
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31 lipofuscin accumulation, indicating oxidative stress conditions only at high concentrations (> 1  
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33 mg/L) (Canesi et al., 2010). At lower concentrations (1-100 µg/L) and longer times of exposure (96  
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35 h), tissue antioxidant defenses were apparently able to cope with potential oxidative stress  
36  
37 conditions induced by n-TiO<sub>2</sub> (Barmo et al., 2013). In whole tissues of the endobenthic bivalve *S.*  
38  
39 *plana*, the response of oxidative stress biomarkers was more important after dietary than waterborne  
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41 exposure to AgNPs; however, defenses were relatively efficient since no biomarkers of damage  
42  
43 were observed. Moreover, labile Ag released from AgNPs was mainly responsible for the observed  
44  
45 effects (Buffet et al., 2012). Mesocosm exposure to low concentrations of n-CuO resulted in  
46  
47 induction of oxidative stress biomarkers but not in lipid peroxidation in both *S. plana* and *H.*  
48  
49 *diversicolor* (Buffet et al., 2013). In *R. decussatus* digestive gland, exposure to predicted  
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51 environmental concentrations (0.75 µg/L) of AuNPs induced oxidative stress and inflammatory  
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53 response markers, as measured by phase II antioxidant enzymes and q-PCR gene expression  
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3 analysis; however, the overall magnitude of responses was low, and oxidative damage was not  
4  
5 observed (Volland et al., 2015).

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7 Application of proteomics in mussel tissues revealed oxidative changes in response to different  
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9 metal-based NPs (reviewed in Rocha et al., 2015). In particular, tissue redox proteomics indicated  
10  
11 that NP size is a key factor in determining biological responses in mussels exposed to AuNPs  
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13 (Tedesco et al., 2010).

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15 Overall, the results suggest that, although the mechanisms of action of NPs in marine  
16  
17 invertebrates can be ascribed to increased radical production/changes in antioxidant defences,  
18  
19 exposure to different types of NPs, at concentrations compatible with predicted environmental  
20  
21 levels, does not seem to elicit significant oxidative stress at the tissue level. Oxidative stress  
22  
23 conditions observed with mostly metal-based NPs (namely containing Ag, Cu, Zn, Au, Fe) could be  
24  
25 mainly related to dissolution of free metal ions from the particles (Baker et al., 2014).

#### 26 27 28 29 *4.3 NP interactions with biological fluids: the protein corona*

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31 As described above, NPs undergo considerable environmental transformations before reaching  
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33 the target biological system: however, the evaluation of the biological effects of NPs requires  
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35 additional understanding of how, once within the organism, NPs interact at the molecular level  
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37 with cells in a physiological environment.

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39 In mammalian systems, extracellular serum proteins adsorb onto the NP surface, forming a  
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41 protein *corona* which affects particle interactions with target cells (Lundqvist et al., 2008;  
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43 Fleischer and Payne, 2014 and refs. quoted therein). The *corona* proteins control the specific  
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45 cellular receptors used by protein-NP complex, the cellular internalization pathways, and the  
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47 immune response. Cells recognize the biomolecular *corona* around a NP, but the biological identity  
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49 of the complex may be considerably different among mammalian species.

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51 In this light, there is the intriguing possibility that NPs may interact with extracellular medium  
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53 or plasma proteins in lower organisms as marine invertebrates. The formation of a NP protein

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3 *corona* has been described so far only in two invertebrate species. In the earthworm *Eisenia foetida*,  
4 soluble coelomic proteins (EfCP) secreted by coelomocytes, form a long-lived *corona* around  
5 AgNPs (Hayashi et al., 2013). Incubation of AgNPs with coelomic proteins lead to formation over  
6 time of AgNP-EfCP corona complexes that induced significantly greater NP accumulation in  
7 coelomocytes. The effects were species specific and NP-specific. In *D. magna* neonates, proteins  
8 secreted in the growing medium formed a coating around PS NPs; such an eco-bio-*corona* lead to  
9 particle agglomeration and increase in uptake and toxicity (Nasser and Linch, 2015).

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17 Data on formation of a NP-protein *corona* in marine invertebrates are still lacking; however, in  
18 *Mytilus* hemocytes exposed to PS-NH<sub>2</sub> in the presence of hemolymph serum, an increase in  
19 lysosomal membrane damage was observed in comparison with only sea water as incubation  
20 medium (Canesi et al., 2015). Such an effect seems driven by specific type of NP, since it was not  
21 observed for instance in n-TiO<sub>2</sub> exposed hemocytes (Canesi, data not shown), suggesting that a  
22 NP-protein *corona* might also be formed in biological fluids of marine invertebrates that may affect  
23 NP-cell interactions.

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31 In general, the presence of proteins reduces NP surface energy by nonspecific adsorption,  
32 leading to lowered membrane adhesion and uptake efficiency (Lesniak et al., 2013). Therefore,  
33 formation of the protein *corona* in mammalian serum is considered as a general protective effect  
34 from the potential cytotoxicity of NPs. In contrast, in earthworms, it was hypothesized that long-  
35 lived proteins present on the NPs (hard *corona*) function as recognizable molecular patterns for  
36 coelomocytes, making the NP-protein complexes “visible” for clearance by these phagocytic cells  
37 responsible for innate immunity (Hayashi et al., 2013). Moreover, the results obtained in *D. magna*,  
38 a filter feeder that can selectively take up food particles based on size, texture and consistency,  
39 suggested that the larger size of NPs caused by the destabilization of the proteins in the eco-*corona*  
40 could make the NPs a more attractive size food source and therefore cause *D. magna* to take up  
41 protein-coated NPs more readily than bare and monodisperse NPs (Nasser and Linch, 2015). The  
42 results obtained in the hemocytes of marine mussels in the presence of hemolymph serum, although  
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2 preliminary, seem to support this hypothesis. In this light, research is in progress on NP-*corona*  
3 proteins in mussels as suitable model marine invertebrates.  
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6 Overall, these data point out the determinant role of the recognizable biological identity during  
7  
8 *in vitro* testing of NPs with invertebrate immune cells; however, the results obtained so far suggest  
9  
10 that in invertebrate species the formation of a protein *corona* may increase the potential for NP  
11  
12 toxicity, rather than play a protective role as in mammalian systems. However, the protein  
13  
14 composition of extracellular fluids of marine invertebrates is largely unknown, given the large  
15  
16 diversity of phyla and species, and different proteins may be involved in the formation of a stable  
17  
18 corona around different NPs in different invertebrate groups. In earthworms, lysenin was identified  
19  
20 as the major corona protein for AgNP (Hayashi et al., 2013), and different secreted proteins related  
21  
22 to cell-to-cell signalling were identified in *D. magna* (Nasser and Linch, 2015). In *Mytilus*, soluble  
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24 hemolymph proteins have been recently characterized by proteomics (Oliveri et al., 2104). Among  
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26 these the major serum protein, Extrapallial Protein (EP) plays a crucial role in non-self recognition  
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28 (Pezzati et al., 2015) and represents a possible candidate for NP corona formation.  
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31 Overall, the results obtained so far underline the need of understanding how the formation of a  
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33 NP protein *corona* may affect the biological outcome of NP exposure in invertebrates in  
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35 comparison to mammalian systems.  
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### 38 39 **5. Effects on development** 40

41 The application of early life stage toxicity tests, involving exposure during the most sensitive  
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43 stages of the organism to environmental stress, would greatly help in the identification of those  
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45 chemicals that represent a major threat to marine species. This also applies to studies on the  
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47 biological impact of NPs on marine invertebrates where, complimentary to the use of adult  
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49 specimens, embryos have emerged as valid tools for studies on developmental perturbations  
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51 induced by different types of NMs (metal based NPs and metal-oxides, carbon based NPs,  
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53 nanoplastics etc.).  
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3 Echinoderms have been so far the most studied taxonomic group of marine invertebrates used  
4 for assessing NP developmental toxicity. In the white sea urchin *Lytechinus pictus* no toxicity of n-  
5 TiO<sub>2</sub> and n-SiO<sub>2</sub> was observed. In contrast, n-ZnO was highly toxic (EC<sub>50</sub> = 99.5 µg/L). However,  
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8 toxicity of n-ZnO was similar to that of soluble Zn<sup>2+</sup>, and was shown to be related to particle  
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10 dissolution in sea water (Fairbairn et al., 2011). In *P. lividus*, exposure to AgNPs caused dose-  
11  
12 dependent developmental defects as well as alterations in swimming patterns: however, the cause-  
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14 effect relationship was not demonstrated (Siller et al. 2013). In *Strongylocentrotus droebachiensis*,  
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16 Ag<sup>+</sup>, AgNPs and f-SWCNTs, separately and in mixtures, induced different developmental defects in  
17  
18 exposed embryos (Magesky and Pelletier, 2015).  
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21 Burić et al. (2015) showed that the effects of AgNPs on embryonal development in  
22  
23 Mediterranean sea urchin species (*Arbacia lixula*, *Paracentrotus lividus* and *Sphaerechinus*  
24  
25 *granularis*) were species-specific. The most sensitive species was *A. lixula*, with high numbers of  
26  
27 malformed embryos or arrested development observed at the lowest AgNP concentrations tested  
28  
29 (1-10 µg/L). Moreover, the greatest impact on development was noted for those embryos first  
30  
31 exposed to NPs at 6 and 24 h post fertilization, this underlying how the timing of exposure to NPs  
32  
33 may represent an important factor in the development of abnormalities.  
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36 In *P. lividus*, n-SiO<sub>2</sub> did not affect fertilization, whereas undeveloped and anomalous embryos  
37  
38 were observed at the gastrula stage (EC<sub>50</sub>=0.06 mg/L), and at the pluteus stage, including skeletal  
39  
40 anomalies and delayed larvae (EC<sub>50</sub>=0.27 mg/L) (Gambardella et al., 2015).  
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43 The first data on bivalve molluscs showed that in the oyster *Crassostrea virginica* C<sub>60</sub> fullerene  
44  
45 affected embryonic development and lysosomal membrane stability from concentrations as low as  
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47 10 µg/L (Ringwood et al., 2009). AgNP also induced lysosomal membrane destabilization and up-  
48  
49 regulation of metallothionein (MT) gene expression; however, it was not established whether these  
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51 effects were due to the NP themselves or to Ag<sup>2+</sup> dissociation (Ringwood et al., 2010).  
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54 In *M. galloprovincialis*, n-TiO<sub>2</sub> did not affect embryo development unless at high mg/L  
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56 concentrations (Libralato et al., 2013, Balbi et al., 2014). Similarly, embryos were unaffected by  
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3 Fe<sub>2</sub>O<sub>3</sub> NPs (Kadar et al., 2010). On the other hand, sperm preincubation with stabilized (i.e., coated  
4 with organic polyacrylic stabilizer) and nonstabilized forms of zero-valent nanoiron (nZVI) caused  
5 serious disruption of development (30% mortality among spermatozoa, 20% decline in fertilization  
6 success, and delay in development) (Kadar et al., 2011). The effects of sperm pre-incubation with  
7 coated zero-valent iron (nZVI) were compared on early life stage development of three key marine  
8 invertebrate species: *M. galloprovincialis*, *Ciona intestinalis* and *Psammechinus milliaris*.  
9 Disruption of embryo development was most severe in sea squirts followed by mussels, while the  
10 urchin embryos were not significantly affected. Decreases in fertilisation success and delay in  
11 embryo development were observed, and the effect was more severe with the coated form, possibly  
12 owing to its better colloidal stability (Kadar et al., 2013). The effects of two coated AgNPs (oleic  
13 acid coated AgNPs and polyvinylpyrrolidone coated AgNPs) on marine benthic invertebrate larvae  
14 across three phyla (i.e., the barnacle *Balanus amphitrite*, the slipper-limpet *Crepidula onyx*, and the  
15 polychaete *Hydroides elegans*) were evaluated in terms of growth, development, and  
16 metamorphosis (Chan and Chu, 2015). Chronic exposure to coated AgNPs resulted in a significant  
17 delay in growth and development, and reduction of larval settlement rate, in a species-specific  
18 manner. Types of surface coatings did not affect the sub-lethal toxicity of AgNPs. However, uptake  
19 of coated AgNPs were observed in the vacuoles of epithelial cell in the digestive tract of *C. onyx*,  
20 suggesting that coated AgNPs could be taken up through diet and the toxicity of coated AgNPs  
21 might be mediated through toxic Ag<sup>+</sup> as well as the novel modalities of coated AgNPs.  
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41 In the deposit feeder annelid *Platynereis dumerilii*, exposure to various concentrations of citrate  
42 (cit-Ag NPs) or humic acid (HA-Ag NPs) capped silver NPs (Ag NPs) induced abnormal  
43 development rate, that were higher in younger life stages. Both Ag NPs were more toxic than  
44 AgNO<sub>3</sub> (Garcia-Alonso et al., 2014).  
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49 Overall, the results obtained in different species and experimental conditions with metal based  
50 NPs indicate that, although the observed effects can be largely due to metal ion solubilisation in sea  
51 water, developmental toxicity may be also due to other mechanisms that are particle-specific.  
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3 Moreover, particle coating may contribute to toxicity, possibly owing to its better colloidal stability  
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5 or differences in uptake routes in different types of embryos.

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7 Although these studies demonstrated that, in marine invertebrates, embryo development can  
8  
9 represent a significant target for different types of NP, in a concentration range of  $\mu\text{g/L}$  - low  $\text{mg/L}$ ,  
10  
11 information on the mechanisms underlying the observed developmental effects is still extremely  
12  
13 scarce. In *P. lividus*  $\text{SiO}_2$  NPs caused neurotoxic damage and a decrease in AChE expression in a  
14  
15 non-dose-dependent manner (Gambardella et al., 2015). In the same species, amino modified  
16  
17 polystyrene NPs (PS-NH<sub>2</sub>) caused severe developmental defects at both 24 and 48 h pf, whereas  
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19 carboxylated particles (PS-COOH) were ineffective. Distinct effects were observed on transcription  
20  
21 of selected genes; the Abcb1 transporter was up-regulated at 48 h pf by PS-COOH whereas PS-NH<sub>2</sub>  
22  
23 induced cas8 gene at 24 hpf, suggesting an apoptotic pathway (Della Torre et al., 2014). In the  
24  
25 larvae of the brine shrimp *Artemia salina*, exposure to  $\text{SnO}_2$ ,  $\text{CeO}_2$  and  $\text{Fe}_3\text{O}_4$  NPs (from 0.01 to 1.0  
26  
27  $\text{mg/mL}$ ) induced changes in swimming speed and alterations and cholinesterase, glutathione-S-  
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29 transferase and catalase activities, with different effects depending on the NP type (Gambardella et  
30  
31 al., 2015). Further studies are needed in order to identify the molecular and cellular targets involved  
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33 in the sensitivity of marine invertebrate embryos and larvae to NP exposure.

### 34 35 36 37 **Conclusions and perspectives**

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39 Different invertebrate species are now been recognized as significant biological targets for  
40  
41 NPs. Available data show that laboratory exposure to different types of NPs, in a wide  
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43 concentration range, elicit significant biological responses, from immunomodulation and oxidative  
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45 stress to developmental defects. However, since the majority of studies have been so far obtained in  
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47 species living in the water column, more information is needed in those species living in close  
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49 contact with sediments, where NP settlement/deposition, and consequent concentrations, may be  
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51 higher.



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3 While recognizing the importance of the studies carried out so far, it is becoming clear that  
4 information on the physico-chemical properties of pristine NPs, or on their behavior in low ionic  
5 strength aqueous solutions, is insufficient to predict their behavior in sea water and consequent  
6 uptake/accumulation/effects in marine organisms. Future studies should address NP fate in complex  
7 environmental matrices, such as SW and sediments, as well as their interactions with biomolecules  
8 present in both environmental media and biological fluids. These eco-bio-interactions may play a  
9 crucial role in particle uptake and accumulation, as well as in their distribution, fate and effects  
10 within the organism. In the absence of experimental data on occurrence and concentrations of NPs  
11 in the marine environment, especially in coastal areas that represent the major sites of potential  
12 discharge, this information may be crucial in understanding whether the biological responses of  
13 marine invertebrates to NPs exposure observed in laboratory experiments may be utilized for risk  
14 assessment and prediction of their impact on the health of the marine environment.  
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34  
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38 Marine ecotoxicology of nanomaterials: toxicity and bioaccumulation of nanotitanium dioxide in  
39 edible species in the presence of metals and dioxin.  
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**Figure Legends**

Fig. 1 - Studies on the effects of NPs in aquatic invertebrates. A) Percentage of publications on freshwater and marine invertebrates with respect to total publications in aquatic invertebrates at 31/12/2008 and 31/12/2014 (source <http://www.ncbi.nlm.nih.gov/pubmed>); B) Number of publications on different marine invertebrate groups and genera at 31/12/2014.

Fig. 2 – Schematic representation of the possible interactions of NPs with different natural organic compounds in the external environment (left) and within the organisms (right), leading to the formation of a NP eco-corona and bio-corona, respectively, in the water column and sediment. A bio-corona can be formed from interactions of NPs with components of internal body fluids or with secreted bio-molecules.

Figure1

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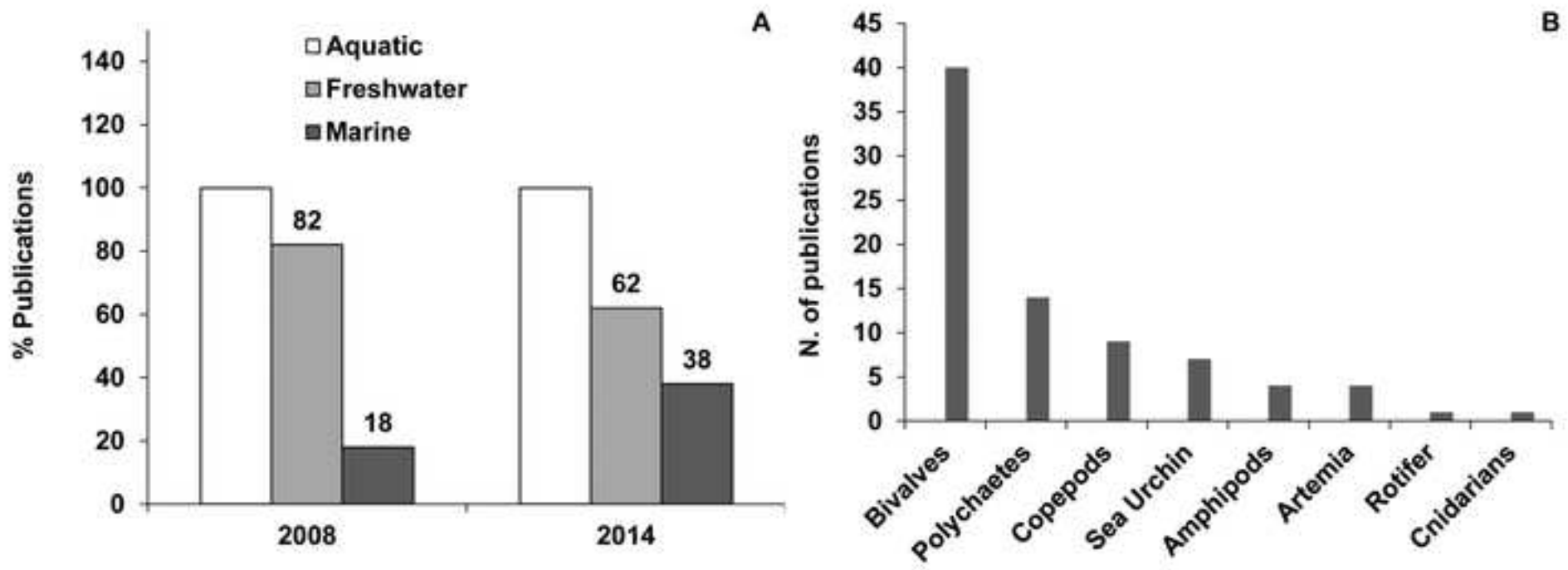


Figure2

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