

Original Russian text [www.bionet.nsc.ru/vogis/](http://www.bionet.nsc.ru/vogis/)


## Psycho-emotional stress, folliculogenesis, and reproductive technologies: clinical and experimental data

A.L. Levinson<sup>1,3</sup>, T.N. Igonina<sup>2</sup>, I.N. Rozhkova<sup>2</sup>, E.Yu. Brusentsev<sup>2</sup>, S.Ya. Amstislavsky<sup>2,3</sup> 

<sup>1</sup> Novosibirsk Center of Reproductive Medicine, Novosibirsk, Russia

<sup>2</sup> Institute of Cytology and Genetics of the Siberian Branch of the Russian Academy of Sciences, Novosibirsk, Russia

<sup>3</sup> Novosibirsk State University, Novosibirsk, Russia

 amstis@yandex.ru

**Abstract.** Modern life, especially in large cities, exposes people to a high level of noise, high density of population, disrupted sleeping, large amount of excessive and controversial information as well as to other negative factors; all this may cause chronic psycho-emotional stress. The latest publications often use the term "Syndrome of megapolopolis", which means disruption of sleeping, high anxiety, and altered reproductive function. Medical treatment of infertility may also be considered as a stress factor, especially when infertility lasts for years and is aggravated with emotional frustration. Long-lasting distress may worsen health in general and suppress reproductive function, in particular. The review presents the data on the effects of maternal stress on folliculogenesis, especially when assisted reproductive technologies (ARTs) are used. Clinical data are presented alongside data from laboratory animal experiments. Different maternal stress models are taken into account in respect of their influence on oocyte maturation and embryo development. The interfering of psycho-emotional stress and reproductive function is the focus of the review. In these situations, exogenous hormones compensate for the stress-related disruption of the hypothalamic-pituitary-gonadal axis. When ARTs are implemented, stress-induced disruption of oogenesis is realized not via a decrease in hypothalamic and pituitary hormones, but by other ways, which involve paracrine mechanisms described in this review. Based on the literature analysis, one may conclude that stress negatively affects oocyte maturation in the ovary and suppresses subsequent embryo development. The role of some ovarian paracrine factors, such as BDNF, GDF-9, HB-EGF, TNF- $\alpha$ , and some others has been elucidated.

Key words: stress; long-term effects; folliculogenesis; assisted reproductive technologies; preimplantation embryo.

**For citation:** Levinson A.L., Igonina T.N., Rozhkova I.N., Brusentsev E.Yu., Amstislavsky S.Ya. Psycho-emotional stress, folliculogenesis, and reproductive technologies: clinical and experimental data. *Vavilovskii Zhurnal Genetiki i Seleksii* = *Vavilov Journal of Genetics and Breeding*. 2022;26(5):431-441. DOI 10.18699/VJGB-22-53

## Психоэмоциональный стресс, фолликулогенез и репродуктивные технологии: клинические и экспериментальные данные

А.Л. Левинсон<sup>1,3</sup>, Т.Н. Игонина<sup>2</sup>, И.Н. Рожкова<sup>2</sup>, Е.Ю. Брусенцев<sup>2</sup>, С.Я. Амстиславский<sup>2,3</sup> 

<sup>1</sup> Новосибирский центр репродуктивной медицины, Новосибирск, Россия

<sup>2</sup> Федеральный исследовательский центр Институт цитологии и генетики Сибирского отделения Российской академии наук, Новосибирск, Россия

<sup>3</sup> Новосибирский национальный исследовательский государственный университет, Новосибирск, Россия

 amstis@yandex.ru

**Аннотация.** Темп современной жизни, уровень шума, плотность населения, особенно в больших городах, нарушение ритма сна и отдыха, количество получаемой и обрабатываемой разнонаправленной информации и другие факторы современной жизни в мегаполисах могут приводить к хроническому психоэмоциональному стрессу. Все чаще в публикациях последних лет появляется понятие «синдром мегаполиса», объединяющее нарушения сна, проявления тревожности и изменение репродуктивной функции. Медицинские процедуры также считаются стрессовыми факторами; более того, само состояние бесплодия, как и его лечение, нередко является затяжным, длится годами, что в свою очередь способствует субадаптивным и неадаптивным реакциям организма, усугубляющим проблему. Пребывание в состоянии дистресса может отражаться на состоянии здоровья в целом и на реализации репродуктивной функции в частности. В обзоре обсуждаются литературные сведения, касающиеся влияния стресса матери на фолликулогенез, в том числе на фоне применения вспомогательных репродуктивных технологий (ВРТ). Сопоставлены клинические данные и результаты экспериментальных исследований на лабораторных животных о влиянии психоэмоционального стресса до и во время беременности на созревание ооцитов и развитие эмбрионов. Особое внимание уделено влиянию

психоэмоционального стресса на репродуктивную функцию на фоне применения ВРТ, когда введение экзогенных гормонов компенсирует вызванное стрессом нарушение функции гипоталамо-гипофизарно-гонадной оси. При применении ВРТ стресс-индуцированное нарушение оогенеза реализуется не через путь снижения выработки гормонов гипоталамуса и гипофиза, а через другие механизмы, в частности паракринные, которые описаны в этом обзоре. В результате проведенного анализа литературы можно сделать заключение о том, что стресс негативно влияет на развитие ооцитов яичника, а также последующее развитие полученных из них эмбрионов, выявлена роль некоторых паракринных факторов яичника, таких как BDNF, GDF-9, HB-EGF, TNF- $\alpha$  и др., которые задействованы в этих процессах.

Ключевые слова: стресс; отдаленные эффекты; фолликулогенез; вспомогательные репродуктивные технологии; преимплантационный эмбрион.

## Introduction

Adult reproductive function and children's health are in focus of scientific interest and of a great public concern. The implementation of assisted reproductive technologies (ARTs) into clinical practice helps to overcome many types of infertility, miscarriage, and to prevent monogenic diseases in children. At the same time, patients in ART clinics often report that the infertility itself as well as its treatment is the traumatic experience, which may lead to the anxiety and even depression (Cousineau, Domar, 2007; Rockliff et al., 2014). Thus, chronic psycho-emotional stress that affects both women and men during infertility treatment and implementation of the ART in particular, are significant factors affecting fertility.

There are numeric evidences of the negative impact of chronic stress on the human well-being, on the mammalian physiology in general, and on the reproductive function in particular (Louis et al., 2011; Muscatell, Eisenberger, 2012). The effects of maternal stress during pregnancy on the body weight of newborns and on the neurodevelopment in children are reported; moreover, there are evidences that prenatal stress affects the behavior and other phenotypic characteristics of different animals (Weinstock, 2008, 2016; Ragaeva et al., 2018; Fitzgerald et al., 2021).

Although the effects of psycho-emotional stress are described in the medical literature and on the laboratory animal models, these two areas of research are mostly developing independently. It should be noted, that nowadays ART is widely used in medical practice, thus the effects of psycho-emotional stress on reproduction, including stress arising from the use of ART, as well as studying the mechanisms underlying these effects are of great concern. The objective of this article is to review and to systematize the accumulated experimental and clinical data describing the effects of chronic psycho-emotional stress on gametogenesis, fertility, ART outcomes, and the offspring health; to review animal models used in such experiments and to outline possible ways aiming to mitigate the adverse effects of stress associated with the use of ARTs. At the same time, experimental data obtained on animal models are compared with clinical observations published in the medical literature. The effects of stress on folliculogenesis and embryogenesis, as well as on the ART-born offspring, both in humans and in experimental animals, are analyzed.

## Modeling psycho-emotional stress in laboratory animals

In experimental studies aimed to elucidate the effects of chronic psycho-emotional stress, including the stress associated with the use of ARTs, on the development of oocytes and early embryos, animal model of restriction stress (Burkus et al., 2013; Gao et al., 2016), the predator exposure model (Liu et al., 2012; Di Natale et al., 2019), or the model of chronic unpredictable mild stress – CUMS (Wu L.M. et al., 2012a, b; Gao et al., 2016) are most frequently used. Plasma levels of corticosteroids, adrenocorticotrophic hormone, corticotropin-releasing hormone, adrenaline, noradrenaline, and ghrelin are normally measured in such studies as stress indicators; less often stress-induced analgesia, behavioral characteristics are also taken into account.

The restriction model of stress is one of the most popular (Gao et al., 2016). Sometimes the experimental animal is fixed with tapes, plaster, cloth towel, or other means so that only the head can move freely; however, most often for this purpose animal is settled in a plastic or metal tube, or a special microcell restricting its movements (Zhang et al., 2011; Gao et al., 2016). The duration of the procedure and the number of restriction episodes affect the intensity of the stress response and should be taken into account (Zhang et al., 2011; Wu X.F. et al., 2015; Zhao X.Y. et al., 2020).

Predator model of psychogenic stress is also widely used, for this purpose the natural predators of mice such as cats, ferrets, rats or foxes are normally chosen; sometimes not the predator itself, but its smell is offered to the tested mouse, this causes the fear and anxiety in the experimental animal (Sanchez-Gonzalez et al., 2018; Di Natale et al., 2019). The most commonly used version of the predator stress model for mice is the presentation of a hungry cat or its scent without physical contact between the mouse and the cat (Liu et al., 2012). The presence of the cat affects the stressed animal, and activating its hypothalamic-pituitary-adrenal axis, therefore triggering the secretion of glucocorticoids (Sanchez-Gonzalez et al., 2018).

Another widely used model is CUMS. Rodents are presented with constantly changing variable stressors over several weeks (Campos et al., 2013). According to this model, combination of isolation and overcrowding can be used as stressors, as well as unpredictable changing the situation

in the cage: wet sawdust, tilting the cage, disruption of the day-night cycle, exposure to different temperatures, the use of mobility restrictions, social stress (Haller et al., 1999; Gao et al., 2016; Burstein, Doron, 2018; Gadek-Michalska et al., 2019). After several days of CUMS regimen, animals show an increase in blood corticosterone levels and a reduced response to pleasurable stimuli (Campos et al., 2013; Gadek-Michalska et al., 2019).

### **Influence of stress on the reproductive function of mammals: experimental data**

Animal studies demonstrate that psycho-emotional stress experienced by the female affects the quantity and quality of oocytes, which in turn contributes to further embryonic development (see the Table). Many studies come to the conclusion that stress leads to a decrease in the developmental potential of oocytes (Wiebold et al., 1986; Zhang et al., 2011; Liu et al., 2012; Lian et al., 2013; Wu X.F. et al., 2015; Gao et al., 2016); this, in turn, resulted in a reduced percentage of blastocysts developed from such oocytes. A decrease in the developmental potential of oocytes was associated with the duration and severity of the applied stress treatment (Gao et al., 2016). It was also revealed, that antral follicles are more sensitive to stress than preantral ones (Gao et al., 2016). Moreover, chronic unpredictable stress disrupts ovulation and cyclicity in female mice, these changes in reproductive system correlate with high levels of corticosteroids in the blood and with the increased activity of superoxide dismutase; moreover, after hormonally induced stimulation of superovulation, mature oocytes were not found in the stressed female mice (Kala, Nivsarkar, 2016).

Stress can also affect embryo implantation. It was shown that even a short restriction stress lasting 24 hours, but coinciding in time with the “implantation window” on the fourth day after mating, negatively affects implantation in mice and slows down the onset of hatching in blastocysts (Zhao L.H. et al., 2013). This effect was mediated through a decrease in the blood levels of progesterone and estradiol, and was associated with the level of expression of heparin-binding epidermal growth factor both in the uterus and in the blastocysts (Zhao L.H. et al., 2013).

It is known that stress leads to activation of the hypothalamic-pituitary-adrenal and sympathoadrenal systems; therefore, traditional markers of stress are glucocorticoids and adrenaline. Restriction stress in mice was shown to be accompanied by an increase in plasma cortisol levels (Zhang et al., 2011). Cortisol injections also led to suppression in oocyte development. In addition, stress led to a decrease in the follicle-stimulating hormone (FSH) release, while injections of cortisol did not cause this effect. The researchers concluded that cortisol affects oocytes through a direct effect on the ovary, while stress impairs their competence indirectly, via effects on the hypothalamic-pituitary-adrenal and hypothalamic-pituitary-ovarian axes (Zhang et al., 2011).

One of the ways to implement stressful effects on the female reproductive system is to influence the production of

ovarian regulators of folliculogenesis, including those mediated by corticotropin-releasing hormone. Corticotropin-releasing hormone (CRH), which is identified in the theca and stroma of the ovaries, as well as in the cytoplasm of oocytes and granulosa cells, is involved in the regulation of follicular maturation, ovulation, the formation of *corpus luteum*, and the synthesis of the ovarian steroid hormones (Kiapekou et al., 2010; Zhai et al., 2020).

In female mice, restriction stress caused an increase of CRH concentration in blood serum, ovaries, and oocytes, as well as an increase in the expression of the CRH receptor 1 (CRHR1) in granulosa and theca cells, but a decrease in the expression of the glucocorticoid receptor and brain-derived neurotrophic factor (BDNF) in the ovaries (Liang et al., 2013). All this ultimately led to an imbalance between estradiol and progesterone concentration in blood and negatively affected the developmental competence of oocytes. Besides, the addition of CRH to the culture medium during oocyte's *in vitro* maturation disrupted its development and increased the rate of apoptosis in granulosa cells (Liang et al., 2013). In another study, it was shown that a stress-induced increase of CRH both in blood and in the ovaries of female mice triggers apoptosis in oocytes and in ovarian granulosa cells due to the activation of the TNF- $\alpha$  system, which results in impaired oocyte competence (Zhao X.Y. et al., 2020).

Animal experiments using CUMS stress model demonstrated that inhibition of follicle development is associated not only with gonadotropins, but also with growth factors such as growth differentiation factor 9 (GDF-9) and BDNF (Wu L.M. et al., 2012b). Exposure of female mice to CUMS resulted in the suppressed follicular development, increased level of follicular atresia, and downregulated GDF-9 expression. The introduction of exogenous gonadotropins partially mitigated these negative effects and restored the development of antral follicles, which was suppressed due to chronic stress, but these exogenous gonadotropins exerted no effects on secondary follicles. However, the introduction of recombinant GDF-9 restored the development of secondary follicles. Co-administration of GDF-9 and gonadotropins in stressed mice restored both secondary and antral follicles. Another study of the same research group showed that CUMS reduces BDNF expression in antral follicles but does not affect BDNF expression in primordial, primary, and secondary follicles (Wu L.M. et al., 2012a). Chronic unpredictable mild stress also reduced the number of retrieved oocytes and the percentage of blastocysts formed, which was corrected by the use of exogenous BDNF.

Some studies attempt to elucidate mechanisms of the influence of psycho-emotional stress on the developmental potential of oocytes and preimplantation embryos. One study reported, that the transition of the heterochromatin configuration from the non-surrounded nucleolus (NSN) type to the surrounded nucleolus (SN) type is suppressed at the germinal vesicle (GV) stage preovulatory oocytes exposed to restriction stress, thus, the developmental potential of such oocytes is impaired (Wu X.F. et al., 2015).

Effects of stress exposures on the development of oocytes and embryos in mice

Stress model	Time of stress exposures	The effect of stress	Reference
The presence of a predator	Within 24 h after the administration of the equine chorionic gonadotropin (eCG)	Impaired <i>in vivo</i> and <i>in vitro</i> development of oocytes and embryos	Liu et al., 2012
Restriction	Daily for 8 h for 4, or 8, or 15, or 23 days immediately prior to the administration of eCG to female mice, i. e. 48 h before collecting early embryos	Decreased <i>in vitro</i> development of embryos collected from mice subjected to stress during 8 or more days	Gao et al., 2016
	Within 24 h, on the 4th day of pregnancy in mice, i. e. during "implantation window"	Decreased number of implantation sites. Slowing down of blastocyst hatching. A decrease in the concentration of progesterone and estradiol in the blood and a decrease in the expression of HB-EGF* in the endometrium of the uterus and in the blastocysts	Zhao L.H. et al., 2013
	Within 48 h, immediately prior to the onset of pregnancy	Increased corticotropin-releasing hormone in the blood and ovaries. Activation of the TNF- $\alpha$ ** system. Triggering of apoptosis in oocytes and ovarian granulosa cells. Disruption of the competence of oocytes	Zhao X.Y. et al., 2020
	24 h, 48 h or 23 days prior to hormonal stimulation and oocyte retrieval	Both 48 h and 23 days of restriction were accompanied by a decrease in histone acetylation and methylation, which ultimately reduced the oocyte development	Wu X.F. et al., 2015
	24 h or 16 h before oocyte retrieval	An increase in the level of corticotropin-releasing hormone in the blood serum, ovaries and oocytes. Increased apoptosis in cumulus cells, leading to a disruption of the oocyte development	Liang et al., 2013
		Disruption of the spindle apparatus at the MI stage. An increase in the proportion of aneuploidy in mature oocytes	Zhou et al., 2012
	Accumulation of ROS in blood serum, ovaries and oocytes. Decrease in the percentage of developing blastocysts and the low cell number in these blastocysts	Lian et al., 2013	
	Within 24 h and 48 h during the period of growth and maturation of oocytes (proestrus in unstimulated mice (24 h), early (0–24 h) and late (24–48 h) stages of oocyte development in hormonally stimulated mice)	Disorder ovulation, decreased percentage of developed blastocysts and the number of cells in them, and fewer live offspring born after the embryo transfer to recipients compared with unstressed controls	Zhang et al., 2011
Chronic unpredictable mild stress (CUMS)	Exposure to various stressors: hot air, swimming in water of different temperatures, shaking (daily, for 4 days, twice a day)	Increase in the percentage of atretic antral follicles; a reduced percentage of 4-cell embryos and blastocysts, fewer cells in blastocysts, and fewer live offspring born after the embryo transfer	Gao et al., 2016
	For five days once a day (30–60 min) exposure to various stressors: restriction; being in a slanted cage or in a cage with dirty bedding; isolation; lack of bedding	Estrous cycle disorders. An increased percentage of atretic antral follicles. Anovulation. The number of oocytes retrieved from stressed mice after hormonal ovarian stimulation was reduced and there were no mature oocyte stages	Kala, Nivsarkar, 2016
	For 30 days, different stressors: isolation; overcrowding and cage tilt; swimming in cold water; hot air; lack of food and water; wet bedding; shaking; shift of the light-dark period	Suppression of development and atresia of follicles. Suppression of GDF-9*** expression  Decreased expression of BDNF**** in antral follicles. No effect on the BDNF expression in primordial, primary and secondary follicles. Reducing the number of oocytes retrieved and the percentage of blastocysts developed	Wu L.M. et al., 2012b  Wu L.M. et al., 2012a

\* HB-EGF, heparin-binding EGF-like growth factor.

\*\* TNF- $\alpha$ , tumor necrosis factor alpha.

\*\*\* GDF-9, growth differentiation factor 9.

\*\*\*\* BDNF, brain-derived neurotrophic factor.



Besides, psycho-emotional stress can lead to disruption of meiotic division in oocytes. It was shown that in stressed females, the proportion of aneuploidy in mature oocytes increases, and the percentage of aneuploid oocytes was three times higher in oocytes with accelerated maturation compared to the delayed ones (Zhou et al., 2012). The authors concluded that maternal stress may cause oxidative stress within oocytes and impair spindle assembly by inactivating the spindle-assembly checkpoint (Zhou et al., 2012).

In addition to hormonal imbalance, psychosocial stress causes an increase in the formation of reactive oxygen species (ROS). High levels of ROS cause oxidative stress, which leads to meiotic cell cycle arrest and resulted in apoptosis (Prasad et al., 2016; Chaudhary et al., 2019). This conclusion was further supported by the observation that oxidative stress induces granulosa cell apoptosis and leads to a decrease in estradiol levels, ovulation frequency, and oocyte quality (Tripathi et al., 2013). Besides, oxidative stress-induced apoptosis of granulosa cells caused the impairments of the contacts of these cells with oocytes, which directly affects the supply of nutrients and the availability of growth factors that affect the quality of oocytes in pre-ovulatory ovarian follicles (Prasad et al., 2016). In experiments using the restriction stress model, it was shown that stress caused the accumulation of ROS in the blood serum of mice, ovaries, and oocytes, and also caused a decrease in the percentage of blastocysts developing *in vitro* with fewer cells observed in these blastocysts (Lian et al., 2013).

It should be noted that the mechanism of the negative impact of chronic stress on the ovary through inhibition of the release of gonadotropins has been well studied. While other ovarian regulatory mechanisms involved in this process are not yet understood. Elucidating these paracrine mechanisms mediating the effects of stress on oogenesis and, subsequently, on the development of embryos is important for more effective use of medical reproductive technologies in patients experiencing chronic psycho-emotional stress.

## Impact of stress on female reproductive function: clinical data

### Clinical data without ART

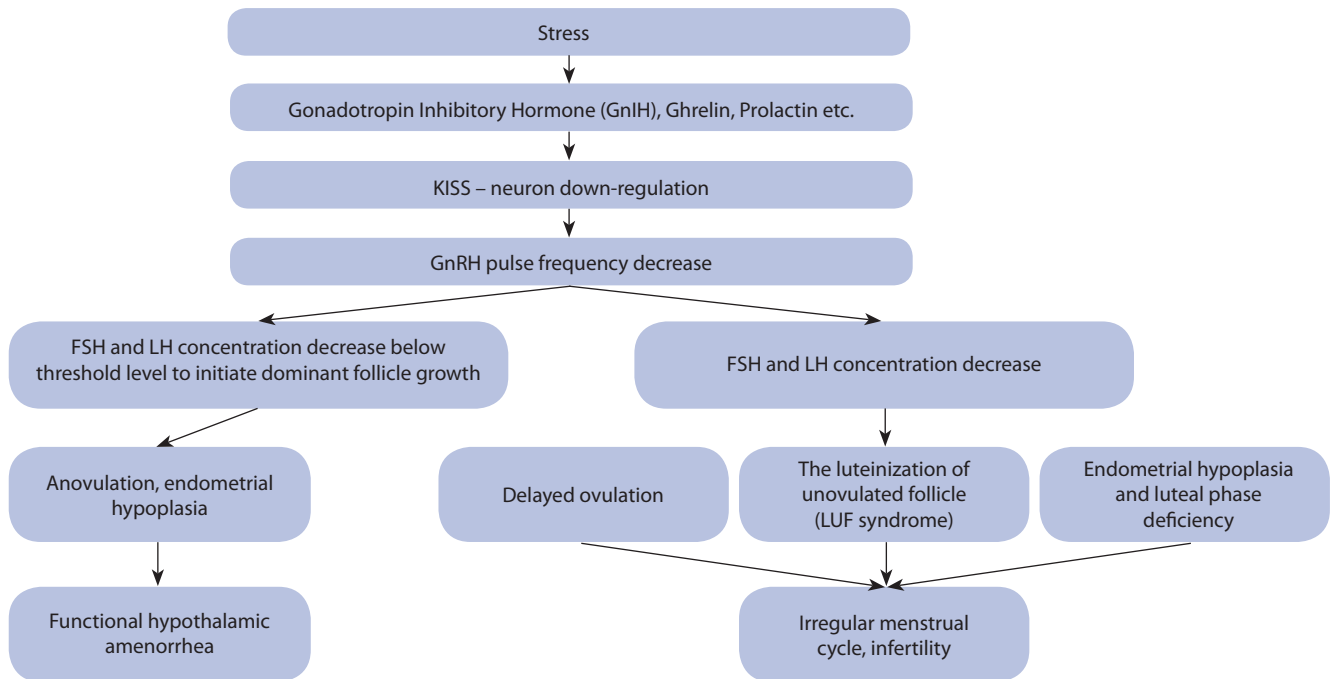
Functional hypothalamic amenorrhea which may be diagnosed in the absence of menstruation for three or more months represents the most striking clinical manifestation of stress-induced disorders of folliculogenesis, with initially intact menstrual function (Warren, Fried, 2001). First of all, functional hypothalamic amenorrhea is characterized by a decrease in the frequency and amplitude of gonadotropin-releasing hormone release peaks, what leads to a decrease in the production of FSH and luteinizing hormone (LH) by the pituitary gland and, as a result, to the absence of hormone-dependent follicle growth. This leads to a disruption of the transition from secondary follicles to antral follicles, a disruption of the formation of a pool of growing follicles, the absence of a dominant follicle and, accordingly, lack of the corpus luteum. Therefore, functional hypothalamic amenorrhea is characterized by a decrease in the produc-

tion of estradiol in the ovaries, which is accompanied by the absence of uterine endometrial proliferation and the absence of menstruation (Fourman, Fazeli, 2015; Prokai, Berga, 2016).

Manifestations of other disorders of folliculogenesis caused by stress may not be as pronounced as amenorrhea. Subclinical manifestations include lengthening and irregularity of the menstrual cycle, insufficiency of the luteal phase of the cycle, luteinization of the non-ovulated follicles (Berga, Loucks, 2007; Palm-Fischbacher, Ehlert, 2014), as well as the absence of a mature oocytes in the ovulated follicle (Tamura et al., 2013). In the case of subclinical manifestations, there is also a decrease in the production of gonadotropin-releasing hormone, however, FSH still reaches a level sufficient to initiate follicle growth. In this case, the growth of the follicles may be slow, with the prolongation of the first phase of the cycle, the late ovulation, and the reduced production of estradiol, which can lead to endometrial hypoplasia (Berga, Loucks, 2007; McEwen et al., 2012). Changes in gonadotropin-releasing hormone pulsation also affect the intensity and frequency of LH peaks, although serum LH levels may remain normal or be only slightly reduced (Krsmanovic et al., 2009). Deficiency in the production of gonadotropin-releasing hormone and estradiol can lead to a reduced LH peak in the preovulatory period and to luteinization of the unovulated follicle, the formation of ovarian cysts, luteal phase insufficiency, and the reduced progesterone production in the second phase of the cycle (Berga, Loucks, 2007). Besides, chronic stress is characterized by increased secretion of cortisol mainly at night, with normal levels of daily and morning secretion (McEwen, 2000), which also contributes to a decrease in the amplitude or even the absence of the ovulatory peak of gonadotropins (Cahill et al., 1998).

Certainly, not all women demonstrate disruption of folliculogenesis and the distortion of the menstrual cycle in the stressful situations (McComb et al., 2006; Ellison et al., 2007). There are both physiological and psychological factors determining tolerance to stress-induced ovarian dysfunction (Wingfield, Sapolsky, 2003; Palm-Fischbacher, Ehlert, 2014). Factors that protect the reproductive system in the situation of stress operate at the level of the central nervous system (the stressor is not perceived), at the level of the hypothalamic-pituitary-adrenal system (impaired secretion of glucocorticoids), at the level of the hypothalamic-pituitary-gonadal system (resistance of the gonads to the action of glucocorticoids), and protection from the action of glucocorticosteroids with the help of a proteins that bind steroids (Wingfield, Sapolsky, 2003).

The development of hypothalamic amenorrhea leads to the reproductive failure due to the absence of follicular growth and the lack of matured oocytes, as well as the lack of appropriate preparation of the endometrium. Sometimes in the case of stressful influences with the formation of luteinization of the unovulated follicle and a deficiency of the luteal phase, the absence of menstruation (amenorrhea) is not observed, but the reproductive potential is significantly reduced (Lynch et al., 2014). Such alterations can be cor-



Effects of stress on female reproductive function.

rected by the proscriptio of appropriate medical treatment that compensate the deficiency of pituitary and/or steroid hormones. However, there is the problem of infertility of unknown origin, with unimpaired folliculogenesis and ovulation. A more detailed study of oogenesis became possible with the introduction of ART.

There are several evidences of a high rate of early pregnancy loss, not associated with chromosomal abnormalities, or an increased anxiety and depression in women with a history of miscarriages. Pregnancy loss itself may be considered as a powerful stressor, which can lead to the recurrent miscarriage (Quenby et al., 2021; Wang et al., 2021). Early pregnancy loss indicates a low viability of the embryo or lack of its interaction with the uterus after implantation, which is probably the result of gametogenesis disruption, distortion in preparation of the endometrium for pregnancy or the development of immunological incompatibility between the maternal organism and the embryo. All of these conditions have been described as possible consequences of stress experienced by a woman during conception and early pregnancy (Wadhwa, 2005; Nepomnaschy et al., 2006). These phenomena can lead to impaired placentation and the development of pregnancy complications typical for later stages of gestation, such as fetoplacental insufficiency, preeclampsia, preterm birth; these conditions may affect the health of children born (Parker, Douglas, 2010; Witt et al., 2012). Mechanisms which cause a change in ovarion function under stress are shown in the Figure.

#### Clinical data obtained with the use of ART

The use of gonadotropins to induce the growth of several follicles for controlled ovarian hyperstimulation is one

of the core ARTs. Thus the stress-induced deficiency of CRH, FSH, and LH, which was discussed in the previous section, is compensated by the administration of exogenous gonadotropins. Moreover, current protocols for controlled ovarian hyperstimulation involve blockade of endogenous gonadotropin-releasing hormone production in order to prevent premature ovulation. Taking all this into account, it can be concluded that during the use of ART, stress-induced disturbance of oogenesis is realized not through a decrease in the production of hormones of the hypothalamus and the pituitary gland, but via other paracrine and autocrine mechanisms described above.

Many reproductologists noticed that unsuccessful ART attempts pretty often take place during adverse life events, such as death of a relative, family problems, etc. This is in practice described by reproductologists as cases of “inexplicably low” quality/quantity of oocytes and embryos in ART programs with an initially good prognosis, and “unexplained” improvement in the quality/quantity of oocytes and embryos in repeated attempts using the same protocols when life situation of the patient was improved (Ebbesen et al., 2009; Meldrum, 2016). A possible explanation for such observations is the inhibitory effect of the stress on gametogenesis.

Data from clinical studies in humans are contradictory. A significant part of these studies indicates the depressing effect of psychological stress on the results of ART. Thus, in a study by Ebbesen et al. (2009) involving 809 women practicing ART for the first time, a decrease in the number of received oocytes, embryo quality and pregnancy rate was shown with an increase in the number of adverse life events that reduce the quality of life, as well as with an increase in the level of perceived stress one month before infertility

treatment with ART (Ebbesen et al., 2009). Another study showed the effect of initial stress on the number of oocytes retrieved and fertilized, as well as on pregnancy and live birth rates (Klonoff-Cohen et al., 2001). Li et al. (2011) found that initial psychological stress was negatively associated with pregnancy rates in ART programs, but intra-follicular concentrations of norepinephrine did not differ between the pregnant and non-pregnant women. In another study, the association of ART results with anxiety levels and serum concentrations of cortisol and noradrenaline was investigated. Elevated levels of cortisol and norepinephrine have been shown to be associated with anxiety levels and negatively correlated with pregnancy and live birth rates (An et al., 2013).

A more recent paper published the results of a study with 135 women involved, that examined the association of salivary and hair cortisol with ART outcomes (Massey et al., 2016). It was shown that salivary cortisol levels were not predictive of ART outcomes. Whereas, lower hair cortisol concentrations predicted the high probability of pregnancy. A recent study of 304 women found that more than 80 % of respondents had elevated levels of anxiety and depression, and these symptoms were inversely correlated with the success of ART implementation (Aimagambetova et al., 2020).

At the same time, some studies do not show an association between anxiety levels, as well as salivary and serum cortisol levels and reproductive outcomes in ART patients (Lovely et al., 2003; Cesta et al., 2018). Miller et al. (2019) assessed the level of anxiety using the Perceived Stress Scale, salivary cortisol concentration at the beginning of the ART cycle, on the day of follicle puncture, and on the day of embryo transfer, and also measured the level of cortisol in the follicular fluid. The authors noted an increase in cortisol and anxiety on the day of follicle puncture, but did not find an association of these stress indicators with pregnancy rates. Besides, elevated follicular cortisol levels correlated with positive ART outcomes. *In vitro* fertilization (IVF) failure has also been shown to predict subsequent psychological distress, but pre-IVF psychological distress did not predict IVF failure (Pasch et al., 2012). The level of stress and the number of oocytes obtained in ART programs for the treatment of infertility was compared. The results of this study showed a significantly higher level of stress in patients with infertility, but the number of oocytes was comparable in both groups (Adeleye et al., 2020).

It can be concluded that despite the large number of publications addressing the effects of stress on the effectiveness of ART in humans, the data obtained are very contradictory. In these studies not only sizes of study groups are variable, but also different approaches to assess experienced stress and anxiety were used. Moreover, many of these studies suffer from the lack of randomization. The majority of these studies do not take into account the fact that the patient's knowledge of his prognosis can significantly affect the assessment of the level of chronic stress and the results of the questionnaire. Due to the heterogeneity of the published data, the conclusions of these works are also

contradictory. Most authors are careful in conclusions about the relationship between stress and reproductive function, based both on the data of their studies and on the general biological considerations suggesting the impossibility of complete suppression of the reproductive function during unfavorable periods due to the need for the survival of the species (Wingfield, Sapolsky, 2003; Rooney, Domar, 2018; Lawson, 2020).

### **Psychotherapy as a way to mitigate the negative effect of psycho-emotional stress on the reproductive system**

The availability of data indicating a significant impact of psychosocial stress on the reproductive system has contributed to an increase in research aimed at studying psychotherapeutic effects in the treatment of infertility.

An early paper addressing this issue highlights the urgent need for quality-compliant research feasible for evaluation (Boivin, 2003). The author analyzed 38 studies, 25 of which were classified as independent, and only eight of them met the research quality standards. In summary, three out of eight good quality studies showed higher pregnancy rates in the psychosocial intervention group compared to the routine care group (Boivin, 2003). In another paper, a meta-analysis of 22 studies was conducted, which indicates that psychotherapy (group and individual/couple) reduced anxiety and depression in infertile patients and possibly affected the success rate of conception (Liz, Strauss, 2005). A review by Campagne (2006) recommends planning of infertility medication taking into account the level of stress, and suggests stress-reducing therapies, prior to initiating infertility treatment (Campagne, 2006).

Subsequent studies presented conflicting results of the use of psychological techniques. Hämmerli et al. (2009) included 21 controlled trials in their meta-analysis and concluded that psychological interventions were not associated with any significant changes in psychological status, but had a positive effect on pregnancy rates in patients receiving treatment without ART (Hämmerli et al., 2009). They also concluded that a therapy of six or more sessions was more effective than a shorter duration of therapy. Frederiksen et al. (2015) performed a meta-analysis of 39 original articles and reported that women receiving some form of psychotherapeutic intervention were about twice as likely to become pregnant compared with women receiving standard treatment (Frederiksen et al., 2015).

Ying et al. (2016) included 20 randomized trials in their systematic review. They concluded that there were methodological problems with studies that reported significant effects of psychological stress on the pregnancy rates, and recommended that a more thorough investigation to be conducted, especially for the most stressful period for infertile patients, in particular, during the time of waiting for the results of a pregnancy test. In a systematic review by Gaitzsch et al. (2020), only two of six studies showed a significant positive effect of psychological interventions on the fertility (Gaitzsch et al., 2020). At the same time, a meta-analysis including 15 studies showed a positive



association between psychosocial interventions, especially long-term ones, and pregnancy rates in infertile women and couples receiving ART treatment (Katyala et al., 2021).

Thus, many researchers emphasize the presence of methodological and practical questions to the currently accumulated data. There is a need for more studies and for unified programs of psychological help. The positive effects of psychotherapy demonstrated in some of the studies indicate that this is a promising area for further research.

## Conclusion

The identification of chronic psycho-emotional stress is challenging both in humans and in experimental studies with laboratory animals. The psychological tests and questionnaires in humans are considered as the “gold standard” for such psycho-emotional stress identifying, however, it requires a lot of time, may not reflect the real physiological situation due to subjective distortions introduced by the interviewee (Slavich, Shields, 2018; Crosswell, Lockwood, 2020). It is also important that the use of psychological tests and questionnaires is not possible in experiments with laboratory animals. Therefore, the search for reliable biomarkers of chronic psycho-emotional stress which can be objectively measured and evaluated is extremely important.

The use of animal models helps to understand the mechanisms underlying the impact of assisted reproductive technology accompanied by stress on the female reproductive function and on the offspring health. Analysis of the literature let to conclude that stress negatively affects the development of ovarian oocytes, as well as the subsequent embryo development. The role of some ovarian paracrine factors that are involved in these processes has been revealed in these studies. Meanwhile, additional experiments on the effect of psycho-emotional stress on the results of *in vitro* fertilization and embryo transfer experiments are warranted, since clinical data are contradictory and only a few experimental works on laboratory animals are available so far.

Available data on the laboratory animals show the effectiveness for the use of such factors as GDF-9 and BDNF to reduce the inhibitory effect of stress on folliculogenesis and embryo development, these factors are promising to be used in the reproductive medicine. Moreover, psychotherapeutic techniques which alleviate effects of stress may increase resistance to stress at the level of the central nervous system, i. e. influence the perception of a stressful event or stimulus. There are reports confirming the effectiveness of psychological techniques in reducing psychological stress, and there are evidences that the use of these techniques is associated with a significant increase in pregnancy rates (Hämmerli et al., 2009; Frederiksen et al., 2015; Katyala et al., 2021). It is important to increase the availability of psychotherapy in reproductive medicine, especially taking into account the level of stress reported by infertile women.

## References

- Adeleye A., Cruz K., Pasch L., Huddleston H. Differences in perceived stress during ovarian stimulation between women with infertility and those pursuing oocyte cryopreservation. *Fertil. Steril.* 2020;114(5):1076-1084. DOI 10.1016/j.fertnstert.2020.05.025.
- Aimagambetova G., Issanov A., Terzic S., Bapayeva G., Ukybassova T., Baikoshkarova S., Aldiyarova A., Shauyen F., Terzic M. The effect of psychological distress on IVF outcomes: reality or speculations? *PLoS One.* 2020;15(12):e0242024. DOI 10.1371/journal.pone.0242024.
- An Y., Sun Z., Li L., Zhang Y., Ji H. Relationship between psychological stress and reproductive outcome in women undergoing *in vitro* fertilization treatment: psychological and neurohormonal assessment. *J. Assist. Reprod. Genet.* 2013;30(1):35-41. DOI 10.1007/s10815-012-9904-x.
- Berga S.L., Loucks T.L. *Stress Induced Anovulation.* Atlanta, Elsevier, USA: Emory Univ. School of Medicine, 2007; 615-631.
- Boivin J. A review of psychosocial interventions in infertility. *Soc. Sci. Med.* 2003;57(12):2325-2341. DOI 10.1016/s0277-9536(03)00138-2.
- Burkus J., Cikos S., Fabian D., Kubandova J., Czikkova S., Koppel J. Maternal restraint stress negatively influences growth capacity of preimplantation mouse embryos. *Gen. Physiol. Biophys.* 2013;32(1):129-137. DOI 10.4149/gpb\_2013010.
- Burstein O., Doron R. The unpredictable chronic mild stress protocol for inducing anhedonia in mice. *J. Vis. Exp.* 2018;140:58184. DOI 10.3791/58184.
- Cahill D.J., Wardle P.G., Harlow C.R., Hull M.G. Onset of the preovulatory luteinizing hormone surge: diurnal timing and critical follicular prerequisites. *Fertil. Steril.* 1998;70(1):56-59. DOI 10.1016/s0015-0282(98)00113-7.
- Campagne D.M. Should fertilization treatment start with reducing stress? *Hum. Reprod.* 2006;21(7):1651-1658. DOI 10.1093/humrep/del078.
- Campos A.C., Fogaca M.V., Aguiar D.C., Guimaraes F.S. Animal models of anxiety disorders and stress. *Braz. J. Psychiatry.* 2013; 35(Suppl.2):S101-S111. DOI 10.1590/1516-4446-2013-1139.
- Cesta C.E., Johansson A.L.V., Hreinsson J., Rodriguez-Wallberg K.A., Olofsson J.I., Holte J., Wramsby H., Wramsby M., Cnattingius S., Skalkidou A., Nyman Iliadou A. A prospective investigation of perceived stress, infertility-related stress, and cortisol levels in women undergoing *in vitro* fertilization: influence on embryo quality and clinical pregnancy rate. *Acta Obstet. Gynecol. Scand.* 2018;97(3):258-268. DOI 10.1111/aogs.13280.
- Chaudhary G.R., Yadav P.K., Yadav A.K., Tiwari M., Gupta A., Sharma A., Pandey A.N., Pandey A.K., Chaube S.K. Necroptosis in stressed ovary. *J. Biomed. Sci.* 2019;26(1):11. DOI 10.1186/s12929-019-0504-2.
- Cousineau T.M., Domar A.D. Psychological impact of infertility. *Best Pract. Res. Clin. Obstet. Gynaecol.* 2007;21(2):293-308. DOI 10.1016/j.bpobgyn.2006.12.003.
- Crosswell A.D., Lockwood K.G. Best practices for stress measurement: how to measure psychological stress in health research. *Health Psychol. Open.* 2020;7(2):2055102920933072. DOI 10.1177/2055102920933072.
- Di Natale M.R., Soch A., Ziko I., De Luca S.N., Spencer S.J., Sominsky L. Chronic predator stress in female mice reduces primordial follicle numbers: implications for the role of ghrelin. *J. Endocrinol.* 2019;241(3):201-219. DOI 10.1530/JOE-19-0109.



- Ebbesen S.M., Zachariae R., Mehlsen M.Y., Thomsen D., Hojgaard A., Ottosen L., Petersen T., Ingerslev H.J. Stressful life events are associated with a poor in-vitro fertilization (IVF) outcome: a prospective study. *Hum. Reprod.* 2009;24(9):2173-2182. DOI 10.1093/humrep/dep185.
- Ellison P.T., Lipson S.F., Jasienska G., Ellison P.L. Moderate anxiety, whether acute or chronic, is not associated with ovarian suppression in healthy, well-nourished, Western women. *Am. J. Phys. Anthropol.* 2007;134(4):513-519. DOI 10.1002/ajpa.20698.
- Fitzgerald E., Parent C., Kee M.Z.L., Meaney M.J. Maternal distress and offspring neurodevelopment: challenges and opportunities for pre-clinical research models. *Front. Hum. Neurosci.* 2021;15:635304. DOI 10.3389/fnhum.2021.635304.
- Fourman L.T., Fazeli P.K. Neuroendocrine causes of amenorrhea – an update. *J. Clin. Endocrinol. Metab.* 2015;100(3):812-824. DOI 10.1210/jc.2014-3344.
- Frederiksen Y., Farver-Vestergaard I., Skovgaard N.G., Ingerslev H.J., Zachariae R. Efficacy of psychosocial interventions for psychological and pregnancy outcomes in infertile women and men: a systematic review and meta-analysis. *BMJ Open.* 2015;5(1):e006592. DOI 10.1136/bmjopen-2014-006592.
- Gadek-Michalska A., Tadeusz J., Bugajski A., Bugajski J. Chronic isolation stress affects subsequent crowding stress-induced brain nitric oxide synthase (NOS) isoforms and hypothalamic-pituitary-adrenal (HPA) axis responses. *Neurotox. Res.* 2019;36(3):523-539. DOI 10.1007/s12640-019-00067-1.
- Gaitzsch H., Benard J., Hugon-Rodin J., Benzakour L., Streuli I. The effect of mind-body interventions on psychological and pregnancy outcomes in infertile women: a systematic review. *Arch. Womens Ment. Health.* 2020;23(4):479-491. DOI 10.1007/s00737-019-01009-8.
- Gao Y., Chen F., Kong Q.Q., Ning S.F., Yuan H.J., Lian H.Y., Luo M.J., Tan J.H. Stresses on female mice impair oocyte developmental potential: effects of stress severity and duration on oocytes at the growing follicle stage. *Reprod. Sci.* 2016;23(9):1148-1157. DOI 10.1177/1933719116630416.
- Haller J., Fuchs E., Halasz J., Makara G.B. Defeat is a major stressor in males while social instability is stressful mainly in females: towards the development of a social stress model in female rats. *Brain Res. Bull.* 1999;50(1):33-39. DOI 10.1016/s0361-9230(99)00087-8.
- Hämmerli K., Znoj H., Barth J. The efficacy of psychological interventions for infertile patients: a meta-analysis examining mental health and pregnancy rate. *Hum. Reprod. Update.* 2009;15(3):279-295. DOI 10.1093/humupd/dmp002.
- Kala M., Nivsarkar M. Role of cortisol and superoxide dismutase in psychological stress induced anovulation. *Gen. Comp. Endocrinol.* 2016;225:117-124. DOI 10.1016/j.ygcen.2015.09.010.
- Katyala N., Poulsen C.M., Knudsen U.B., Frederiksen Y. The association between psychosocial interventions and fertility treatment outcome: a systematic review and meta-analysis. *Eur. J. Obstet. Gynecol. Reprod. Biol.* 2021;259:125-132. DOI 10.1016/j.ejogrb.2021.02.012.
- Kiapekou E., Zapanti E., Mastorakos G., Loutradis D. Update on the role of ovarian corticotropin-releasing hormone. *Ann. NY Acad. Sci.* 2010;1205:225-229. DOI 10.1111/j.1749-6632.2010.05685.x.
- Klonoff-Cohen H., Chu E., Natarajan L., Sieber W. A prospective study of stress among women undergoing in vitro fertilization or gamete intrafallopian transfer. *Fertil. Steril.* 2001;76(4):675-687. DOI 10.1016/s0015-0282(01)02008-8.
- Krsmanovic L.Z., Hu L., Leung P.K., Feng H., Catt K.J. The hypothalamic GnRH pulse generator: multiple regulatory mechanisms. *Trends Endocrinol. Metab.* 2009;20(8):402-408. DOI 10.1016/j.tem.2009.05.002.
- Lawson A.K. Caution is warranted when discussing correlations between stress and reproductive outcomes. *Fertil. Steril.* 2020;114(5):972-973. DOI 10.1016/j.fertnstert.2020.07.054.
- Li X.H., Ma Y.G., Geng L.H., Qin L., Hu H., Li S.W. Baseline psychological stress and ovarian norepinephrine levels negatively affect the outcome of in vitro fertilisation. *Gynecol. Endocrinol.* 2011;27(3):139-143. DOI 10.3109/09513590.2010.501871.
- Lian H.Y., Gao Y., Jiao G.Z., Sun M.J., Wu X.F., Wang T.Y., Li H., Tan J.H. Antioxidant supplementation overcomes the deleterious effects of maternal restraint stress-induced oxidative stress on mouse oocytes. *Reproduction.* 2013;146(6):559-568. DOI 10.1530/REP-13-0268.
- Liang B., Wei D.L., Cheng Y.N., Yuan H.J., Lin J., Cui X.Z., Luo M.J., Tan J.H. Restraint stress impairs oocyte developmental potential in mice: role of CRH-induced apoptosis of ovarian cells. *Biol. Reprod.* 2013;89(3):64. DOI 10.1095/biolreprod.113.110619.
- Liu Y.X., Cheng Y.N., Miao Y.L., Wei D.L., Zhao L.H., Luo M.J., Tan J.H. Psychological stress on female mice diminishes the developmental potential of oocytes: a study using the predatory stress model. *PLoS One.* 2012;7(10):e48083. DOI 10.1371/journal.pone.0048083.
- Liz de T.M., Strauss B. Differential efficacy of group and individual/couple psychotherapy with infertile patients. *Hum. Reprod.* 2005;20(5):1324-1332. DOI 10.1093/humrep/deh743.
- Louis G.M., Lum K.J., Sundaram R., Chen Z., Kim S., Lynch C.D., Schisterman E.F., Pyper C. Stress reduces conception probabilities across the fertile window: evidence in support of relaxation. *Fertil. Steril.* 2011;95(7):2184-2189. DOI 10.1016/j.fertnstert.2010.06.078.
- Lovely L.P., Meyer W.R., Ekstrom R.D., Golden R.N. Effect of stress on pregnancy outcome among women undergoing assisted reproduction procedures. *South. Med. J.* 2003;96(6):548-551. DOI 10.1097/01.SMJ.0000054567.79881.E9.
- Lynch C.D., Sundaram R., Maisog J.M., Sweeney A.M., Buck Louis G.M. Preconception stress increases the risk of infertility: results from a couple-based prospective cohort study – the LIFE study. *Hum. Reprod.* 2014;29(5):1067-1075. DOI 10.1093/humrep/deu032.
- Massey A.J., Campbell B.K., Raine-Fenning N., Pincott-Allen C., Perry J., Vedhara K. Relationship between hair and salivary cortisol and pregnancy in women undergoing IVF. *Psychoneuroendocrinology.* 2016;74:397-405. DOI 10.1016/j.psyneuen.2016.08.027.
- McComb J.J.R., Qian X.-P., Veldhuis J.D., McGlone J.J., Norman R.L. Neuroendocrine responses to psychological stress in eumenorrheic and oligomenorrheic women. *Stress.* 2006;9(1):41-51. DOI 10.1080/10253890600591678.
- McEwen B.S. The neurobiology of stress: from serendipity to clinical relevance. *Brain Res.* 2000;886(1-2):172-189. DOI 10.1016/s0006-8993(00)02950-4.
- McEwen B.S., Akama K.T., Spencer-Segal J.L., Milner T.A., Waters E.M. Estrogen effects on the brain: actions beyond the hypothalamus via novel mechanisms. *Behav. Neurosci.* 2012;126(1):4-16. DOI 10.1037/a0026708.
- Meldrum D.R. Introduction: examining the many potential reasons why euploid blastocysts do not always result in viable pregnancies: part 1. *Fertil. Steril.* 2016;105(3):545-547. DOI 10.1016/j.fertnstert.2015.12.007.

- Miller N., Herzberger E.H., Pasternak Y., Klement A.H., Shavit T., Yaniv R.T., Ghetler Y., Neumark E., Eisenberg M.M., Berkovitz A., Shulman A., Wiser A. Does stress affect IVF outcomes? A prospective study of physiological and psychological stress in women undergoing IVF. *Reprod. Biomed. Online*. 2019;39(1):93-101. DOI 10.1016/j.rbmo.2019.01.012.
- Muscattell K.A., Eisenberger N.I. A social neuroscience perspective on stress and health. *Soc. Personal. Psychol. Compass*. 2012;6(12):890-904. DOI 10.1111/j.1751-9004.2012.00467.x.
- Nepomnaschy P.A., Welch K.B., McConnell D.S., Low B.S., Strassmann B.I., England B.G. Cortisol levels and very early pregnancy loss in humans. *Proc. Natl. Acad. Sci. USA*. 2006;103(10):3938-3942. DOI 10.1073/pnas.0511183103.
- Palm-Fischbacher S., Ehlert U. Dispositional resilience as a moderator of the relationship between chronic stress and irregular menstrual cycle. *J. Psychosom. Obstet. Gynaecol.* 2014;35(2):42-50. DOI 10.3109/0167482X.2014.912209.
- Parker V.J., Douglas A.J. Stress in early pregnancy: maternal neuroendocrine-immune responses and effects. *J. Reprod. Immunol.* 2010;85(1):86-92. DOI 10.1016/j.jri.2009.10.011.
- Pasch L.A., Gregorich S.E., Katz P.K., Millstein S.G., Nachtigall R.D., Bleil M.E., Adler N.E. Psychological distress and in vitro fertilization outcome. *Fertil. Steril.* 2012;98(2):459-464. DOI 10.1016/j.fertnstert.2012.05.023.
- Prasad S., Tiwari M., Pandey A.N., Shrivastav T.G., Chaube S.K. Impact of stress on oocyte quality and reproductive outcome. *J. Biomed. Sci.* 2016;23:36. DOI 10.1186/s12929-016-0253-4.
- Prokai D., Berga S.L. Neuroprotection via reduction in stress: altered menstrual patterns as a marker for stress and implications for long-term neurologic health in women. *Int. J. Mol. Sci.* 2016;17(12):2147. DOI 10.3390/ijms17122147.
- Quenby S., Gallos I.D., Dhillon-Smith R.K., Podesek M., Stephenson M.D., Fisher J., Brosens J.J., Brewin J., Ramhorst R., Lucas E.S., McCoy R.C., Anderson R., Daher S., Regan L., Al-Memar M., Bourne T., MacIntyre D.A., Rai R., Christiansen O.B., Sugiura-Ogasawara M., Odendaal J., Devall A.J., Bennett P.R., Petrou S., Coomarasamy A. Miscarriage matters: the epidemiological, physical, psychological, and economic costs of early pregnancy loss. *Lancet*. 2021;397(10285):1658-1667. DOI 10.1016/S0140-6736(21)00682-6.
- Ragaeva D.S., Igonina T.N., Brusentsev E.Yu., Rozhkova I.N., Amstislavsky S.Ya. Long-term effects of early prenatal impact on the offspring development, physiology and behavior. *Uspekhi Fiziolicheskikh Nauk = Progress in Physiological Science*. 2018;49(4):30-44. (in Russian)
- Rockliff H.E., Lightman S.L., Rhidian E., Buchanan H., Gordon U., Vedhara K. A systematic review of psychosocial factors associated with emotional adjustment in *in vitro* fertilization patients. *Hum. Reprod. Update*. 2014;20(4):594-613. DOI 10.1093/humupd/dmu010.
- Rooney K.L., Domar A.D. The relationship between stress and infertility. *Dialogues Clin. Neurosci.* 2018;20(1):41-47. DOI 10.31887/DCNS.2018.20.1/krooney.
- Sanchez-Gonzalez B., Planillo A., Navarro-Castilla A., Barja I. The concentration of fear: mice's behavioural and physiological stress responses to different degrees of predation risk. *Naturwissenschaften*. 2018;105(1-2):16. DOI 10.1007/s00114-018-1540-6.
- Slavich G.M., Shields G.S. Assessing lifetime stress exposure using the stress and adversity inventory for adults (adult STRAIN): an overview and initial validation. *Psychosom. Med.* 2018;80(1):17-27. DOI 10.1097/PSY.0000000000000534.
- Tamura H., Takasaki A., Taketani T., Tanabe M., Kizuka F., Lee L., Tamura I., Maekawa R., Asada H., Yamagata Y., Sugino N. Melatonin as a free radical scavenger in the ovarian follicle. *Endocr. J.* 2013;60(1):1-13. DOI 10.1507/endocr.ej12-0263.
- Tripathi A., Shrivastav T.G., Chaube S.K. An increase of granulosa cell apoptosis mediates aqueous neem (*Azadirachta indica*) leaf extract-induced oocyte apoptosis in rat. *Int. J. Appl. Basic Med. Res.* 2013;3(1):27-36. DOI 10.4103/2229-516X.112238.
- Wadhwa P.D. Psychoneuroendocrine processes in human pregnancy influence fetal development and health. *Psychoneuroendocrinology*. 2005;30(8):724-743. DOI 10.1016/j.psyneuen.2005.02.004.
- Wang Y., Meng Z., Pei J., Qian L., Mao B., Li Y., Li J., Dai Z., Cao J., Zhang C., Chen L., Jin Y., Yi B. Anxiety and depression are risk factors for recurrent pregnancy loss: a nested case-control study. *Health Qual. Life Outcomes*. 2021;19(1):78. DOI 10.1186/s12955-021-01703-1.
- Warren M.P., Fried J.L. Hypothalamic amenorrhea. The effects of environmental stresses on the reproductive system: a central effect of the central nervous system. *Endocrinol. Metab. Clin. North Am.* 2001;30(3):611-629. DOI 10.1016/s0889-8529(05)70204-8.
- Weinstock M. The long-term behavioural consequences of prenatal stress. *Neurosci. Biobehav. Rev.* 2008;32(6):1073-1086. DOI 10.1016/j.neubiorev.2008.03.002.
- Weinstock M. Prenatal stressors in rodents: effects on behavior. *Neurobiol. Stress*. 2016;6:3-13. DOI 10.1016/j.ynstr.2016.08.004.
- Wiebold J.L., Stanfield P.H., Becker W.C., Hillers J.K. The effect of restraint stress in early pregnancy in mice. *J. Reprod. Fertil.* 1986;78(1):185-192. DOI 10.1530/jrf.0.0780185.
- Wingfield J.C., Sapolsky R.M. Reproduction and resistance to stress: when and how. *J. Neuroendocrinol.* 2003;15(8):711-724. DOI 10.1046/j.1365-2826.2003.01033.x.
- Witt W.P., Wisk L.E., Cheng E.R., Hampton J.M., Hagen E.W. Preconception mental health predicts pregnancy complications and adverse birth outcomes: a national population-based study. *Matern. Child Health J.* 2012;16(7):1525-1541. DOI 10.1007/s10995-011-0916-4.
- Wu L.M., Hu M.H., Tong X.H., Han H., Shen N., Jin R.T., Wang W., Zhou G.X., He G.P., Liu Y.S. Chronic unpredictable stress decreases expression of brain-derived neurotrophic factor (BDNF) in mouse ovaries: relationship to oocytes developmental potential. *PLoS One*. 2012a;7(12):e52331. DOI 10.1371/journal.pone.0052331.
- Wu L.M., Liu Y.S., Tong X.H., Shen N., Jin R.T., Han H., Hu M.H., Wang W., Zhou G.X. Inhibition of follicular development induced by chronic unpredictable stress is associated with growth and differentiation factor 9 and gonadotropin in mice. *Biol. Reprod.* 2012b;86(4):121. DOI 10.1095/biolreprod.111.093468.
- Wu X.F., Yuan H.J., Li H., Gong S., Lin J., Miao Y.L., Wang T.Y., Tan J.H. Restraint stress on female mice diminishes the developmental potential of oocytes: roles of chromatin configuration and histone modification in germinal vesicle stage oocytes. *Biol. Reprod.* 2015;92(1):13. DOI 10.1095/biolreprod.114.124396.
- Ying L., Wu L.H., Loke A.Y. The effects of psychosocial interventions on the mental health, pregnancy rates, and marital function of infertile couples undergoing in vitro fertilization: a systematic review. *J. Assist. Reprod. Genet.* 2016;33(6):689-701. DOI 10.1007/s10815-016-0690-8.
- Zhai Q.Y., Wang J.J., Tian Y., Liu X., Song Z. Review of psychological stress on oocyte and early embryonic development in

- female mice. *Reprod. Biol. Endocrinol.* 2020;18(1):101. DOI 10.1186/s12958-020-00657-1.
- Zhang S.Y., Wang J.Z., Li J.J., Wei D.L., Sui H.S., Zhang Z.H., Zhou P., Tan J.H. Maternal restraint stress diminishes the developmental potential of oocytes. *Biol. Reprod.* 2011;84(4):672-681. DOI 10.1095/biolreprod.110.087890.
- Zhao L.H., Cui X.Z., Yuan H.J., Liang B., Zheng L.L., Liu Y.X., Luo M.J., Tan J.H. Restraint stress inhibits mouse implantation: temporal window and the involvement of HB-EGF, estrogen and progesterone. *PLoS One.* 2013;8(11):e80472. DOI 10.1371/journal.pone.0080472.
- Zhao X.Y., Li Z.B., Yuan H.J., Han X., Wu J.S., Feng X.Y., Zhang M., Tan J.H. Restraint stress and elevation of corticotrophin-releasing hormone in female mice impair oocyte competence through activation of the tumour necrosis factor  $\alpha$  (TNF- $\alpha$ ) system. *Reprod. Fertil. Dev.* 2020;32(9):862-872. DOI 10.1071/RD20002.
- Zhou P., Lian H.Y., Cui W., Wei D.L., Li Q., Liu Y.X., Liu X.Y., Tan J.H. Maternal-restraint stress increases oocyte aneuploidy by impairing metaphase I spindle assembly and reducing spindle assembly checkpoint proteins in mice. *Biol. Reprod.* 2012;86(3):83. DOI 10.1095/biolreprod.111.095281.

---

**Acknowledgements.** This work was supported by the Russian Foundation for Basic Research, project No. 20-015-00162 and State Budgeted Project ICG SB RAS No. 0259-2021-0015.

**Conflict of interest.** The authors declare no conflict of interest.

Received January 24, 2022. Revised March 21, 2022. Accepted March 31, 2022.