

ASSOCIATE EDITOR: ERIC BARKER

Therapeutic Opportunities and Delivery Strategies for Brain Revascularization in Stroke, Neurodegeneration, and Aging

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This work was supported by the University of the Basque Country (UPV/EHU) [Grant ESPDOC19/47] (postdoctoral fellowship to I.V.B.); and the Basque Country Government (Consolidated Groups) [Grant IT907-16].

No author has an actual or perceived conflict of interest with the contents of this article.

dx.doi.org/10.1124/pharmrev.121.000418.

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Abstract—Central nervous system (CNS) diseases, especially acute ischemic events and neurodegenerative disorders, constitute a public health problem with no effective treatments to allow a persistent solution. Failed therapies targeting neuronal recovery have revealed the multifactorial and intricate pathophysiology underlying such CNS disorders as ischemic stroke, Alzheimer’s disease, amyotrophic lateral sclerosis, vascular Parkinsonism, vascular dementia, and aging, in which cerebral microvasculature impairment seems to play a key role. In fact, a reduction in vessel density and cerebral blood flow occurs in these scenarios, contributing to neuronal dysfunction and leading to loss of cognitive function. In this review, we provide an overview of healthy brain microvasculature structure and function

in health and the effect of the aforementioned cerebral CNS diseases. We discuss the emerging new therapeutic opportunities, and their delivery approaches, aimed at recovering brain vascularization in this context.

Significance Statement—The lack of effective treatments, mainly focused on neuron recovery, has prompted the search of other therapies to treat cerebral central nervous system diseases. The disruption and degeneration of cerebral microvasculature has been evidenced in neurodegenerative diseases, stroke, and aging, constituting a potential target for restoring vascularization, neuronal functioning, and cognitive capacities by the development of therapeutic pro-angiogenic strategies.

I. Introduction

Neurologic disorders are the second leading cause of death and the principal cause of disability in the world (GBD 2015 Neurologic Disorders Collaborator Group, 2017). Increasing life expectancy and population growth worldwide imply that more and more people are reaching ages in which neurologic disorders are more prevalent. The rising incidence and prevalence of these related central nervous system (CNS) diseases have an important socioeconomic impact, so it becomes a real problem not only for patients and families but also for the economy and healthcare systems (Harper, 2014; Wimo et al., 2020). There are no curative pharmacological treatments able to attain a complete neurovascular recovery in CNS diseases; they can only slow down the neurologic degenerative processes. This scenario underscores the difficulty of current pharmacological drugs to target and efficiently act in the brain. One of the main obstacles that lacks the success of such therapies is the blood-brain-barrier (BBB), along with other factors that must be taken into consideration, such as the presence of other extracellular and intracellular barriers and the complexity of the neurovascular network with interactions at several levels. For this reason, huge research efforts are being conducted to find and develop novel therapeutic strategies for CNS diseases (Niu et al., 2019; Poovaiah et al., 2018; Teleanu et al., 2019).

A few years ago, neuroscientists considered the brain as a dichotomized organ comprised of brain cells and cerebral blood vessels, with no relationship among these two entities. Nowadays however, the scientific community is aware of the close connection established and required between neuronal and vascular CNS cells for correct brain functioning. The brain is one of the most highly perfused organs in the body; in fact, nearly every neuron has its own capillary (Zlokovic, 2005), highlighting the pivotal relationship between the neuronal and vascular systems, called the neurovascular network. The neurovascular network in CNS is responsible for supplying the 20% of the cardiac output carrying oxygen and nutrients to the brain (Iadecola, 2013) and thus contributing to a healthy neurologic function. That is why lack of this supply, caused by vessel damage or degeneration, could have a major role in the pathogenesis of CNS diseases. Consequently, it is not surprising that the cognitive impairment that occurs in many CNS diseases could be related to cerebrovascular disruptions, mainly at the microvasculature level, and cerebral blood flow reduction, as in the case of ischemic stroke, amyotrophic lateral sclerosis (ALS), Alzheimer’s disease (AD), vascular Parkinsonism (VP), vascular dementia (VaD) and aging, which will be described in depth in section III, Vascular Disorders in Brain CNS Diseases.

This review provides an overview of the cellular and molecular mechanisms needed to manage the

ABBREVIATIONS: AAV, adeno-associated virus; $A\beta$, amyloid- β ; AD, Alzheimer’s disease; ALS, amyotrophic lateral sclerosis; ANGPTL4, angiopoietin-like 4; AV, adenovirus; BBB, blood-brain barrier; BMSC, bone marrow stromal cells; CNS, central nervous system; EC, endothelial cell; ECM, extracellular matrix; FGF, fibroblast growth factor; HIF-1 α , hypoxia inducible factor-1 α ; IL, interleukin; JAM, junctional adhesion molecules; MCAO, middle cerebral artery occlusion; miR, microRNA; NMN, nicotinamide mononucleotide; NRP1, neuropilin-1; PDGF, platelet-derived growth factor; PEG, polyethylene glycol; PLGA, poly(lactic-co-glycolic acid); PIGF, placental growth factor; PPS, poly(propylene sulfide); RO, retro-orbital; ROS, reactive oxygen species; VaD, vascular dementia; VEGF, vascular endothelial growth factor; VP, vascular Parkinsonism; ZO, zonula occludens.

cerebral microvasculature, as well as an up-to-date perspective of CNS diseases related to cerebral microvasculature damage or deterioration, as is the case of ischemic stroke, Alzheimer's disease (AD), amyotrophic lateral sclerosis (ALS), vascular Parkinsonism (VP), vascular dementia (VaD) and aging, all of which are associated with cognitive impairment. In particular, we describe evidence for microvasculature regeneration as a form of neurologic and cognitive function improvement by looking at progress in the identification of potential therapeutic pro-angiogenic factors and focusing on the nanotechnological approaches, advanced opportunities, and the administration strategies employed.

II. Mechanisms Regulating Brain Microvasculature

The vascular system of CNS originates during embryonic development by angiogenesis as blood vessels on the leptomeningeal surface grow in toward the parenchyma (Marin-Padilla, 1985), and, in this way, CNS vessels and microvasculature are exclusively formed by angiogenic processes from the perineural vascular plexus (Flamme et al., 1997). Angiogenesis involves the creation of new blood vessels by sprouting or diverging from pre-existing vessels, giving rise to the CNS microvasculature network. Both cellular and molecular mechanisms are involved in maintaining a suitable brain microvasculature, in which the cellular scaffold and the molecular exchange are intimately linked. Additionally, epigenetics, gene switching, cytokines, and extracellular matrix (ECM) molecules are also involved in maintaining correct neurovascular unit performance, not only in physiologic conditions but also under a pathologic stimulus.

A. Cellular Mechanisms

Physical and biologic characteristics of the cerebrovasculature are provided by endothelial cells (ECs), pericytes, smooth muscle cells also known as mural cells, or Rouget cells and astroglial foot processes (Giannoni et al., 2018; Sweeney et al., 2019), which together with

neurons constitute the neurovascular unit (Iadecola, 2017; McConnell et al., 2017) (Fig. 1). This concept of neurovascular unit emerged in 2001, during the Stroke Progress Review Group meeting of the National Institute of Neurologic Disorders and Stroke (<https://www.nia.nih.gov/health/vascular-contributions-cognitive-impairment-and-dementia>) to reinforce the close connection between a correct CNS function and its microvasculature, achieved by effective paracrine regulations.

Within the neurovascular unit we find the cerebral endothelium, known as the BBB, which is composed of a monolayer of ECs with more special features than the ECs present in other tissues. In particular, CNS ECs are held together by tight junctions (Kniesel and Wolburg, 2000; Luissint et al., 2012a) regulating the homeostatic movement of ions, molecules, and cells between the blood and the CNS (Daneman and Prat, 2015; Serlin et al., 2015) (Fig. 1). Integral membrane proteins are responsible for these roles in tight junction complexes, namely occludins, claudins, and junctional adhesion molecules (JAM), which interact with cytoplasmic scaffolding proteins, such as actin cytoskeleton, zonula occludens (ZO) proteins, and other associated proteins (Luissint et al., 2012b; Vorbrott and Dobrogowska, 2003). In fact, it has been described that ZO-1, one of the tight junction adaptor proteins, regulates CNS angiogenic potential and EC migration, as well as BBB formation (Tornavaca et al., 2015). JAM family proteins, which are present in tight junctions as mentioned above, have also been linked to angiogenesis, EC migration, and crosstalk with bFGF and $\alpha V\beta 3$ integrin signaling (Cooke et al., 2006; Peddibhotla et al., 2013).

All these features make BBB a dynamic interface; more specifically, this meticulously attuned and specialized system is able to: 1) keep the brain separated and protected from the compounds present in peripheral blood circulation, 2) selectively transport necessary components for the brain, 3) detect changes in blood flow and transmit this information to brain, 4) metabolize substances present in brain and blood, and 5) carry out the clearance of own neurotoxic compounds or xenobiotics generated in brain (Huber et al., 2001) (Fig. 1).

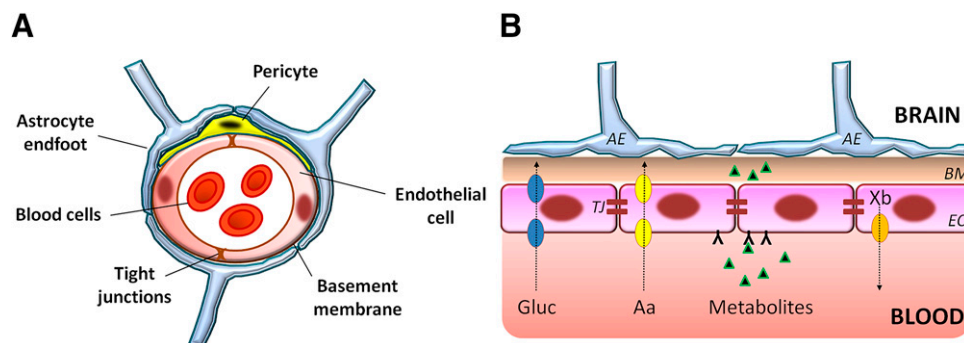


Fig. 1. Cellular components of central nervous system microvasculature and microenvironment integrity are essential for a correct neurovascular function. (A) Schematic representation of the neurovascular unit. (B) Blood-brain barrier exchange function. Gluc, glucose; Aa, aminoacids; Xb, xenobiotics; AE, astrocyte endfoot; TJ, tight junction; EC, endothelial cell; BM, basement membrane.

However, under pathophysiological conditions, the BBB disrupts, altering the biologic function of this key barrier. Changes in BBB permeability can trigger microglial activation and infiltration of immune cells into the brain, alterations in CNS homeostasis, and variable damage to the nearby brain tissue (Thurgur and Pinteaux, 2019), which could ultimately lead to the damage and reduction of brain microvasculature, as happens in many CNS diseases.

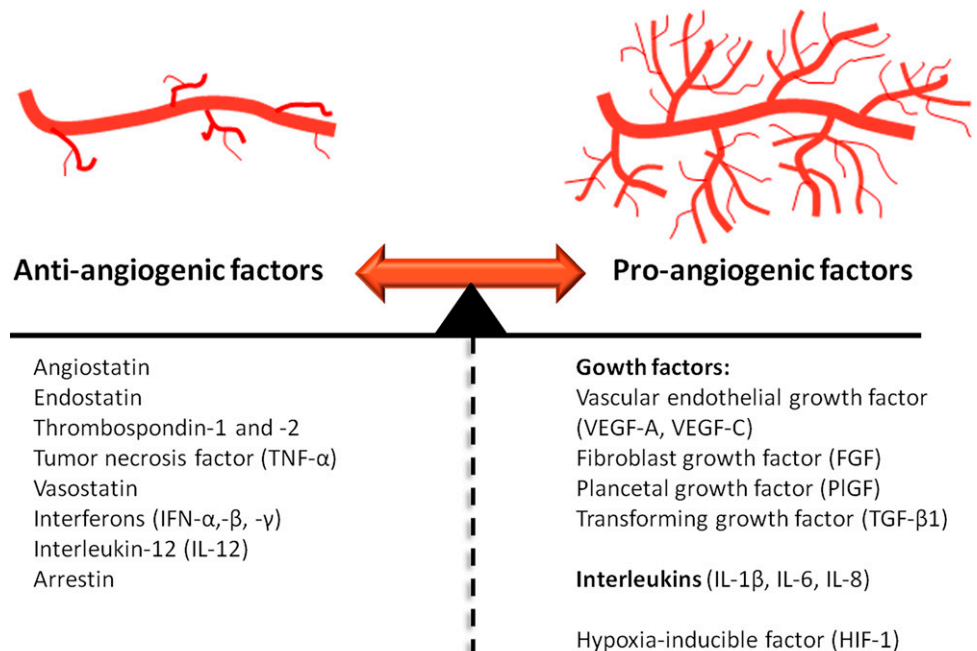
B. Molecular Mechanisms

1. Growth Factors. Contrary to the developing brain, where pro-angiogenic processes are upregulated, angiogenesis in the adult brain is physiologically downregulated. This fact was evidenced in 1985 by a study that analyzed brain capillary proliferation in post-natal rats by measuring [³H] thymidine incorporation (Robertson et al., 1985). In this work, it was demonstrated that only 0.3% of ECs incorporated this substance. Hence, vessel growth in CNS is tightly regulated by pro-angiogenic, for stimulating, and anti-angiogenic, for inhibiting, factors (Chu LH et al., 2012; Harrigan, 2003; Vallon et al., 2014) (Fig. 2). In physiologic conditions, vascular quiescence is preserved in CNS due to angiogenic inhibitors, while the balance changes in favor of pro-angiogenic factors promoting angiogenesis in pathophysiological conditions such as in ischemic stroke or brain hypoxia that occurs in neurodegenerative CNS diseases (Harrigan, 2003; Vallon et al., 2014). Hence, understanding the molecular mechanisms involved in managing brain microvasculature would reveal potential therapeutic candidates.

Most of the factors promoting angiogenesis consist in growth factors (Fig. 2, Fig. 3) and molecules of the ECM (Martino et al., 2015). Among them, the most relevant is the vascular endothelial growth factor (VEGF), followed by the fibroblast growth factor (FGF) (Harrigan, 2003; Logsdon et al., 2014; Mackenzie and Ruhrberg, 2012; Mancuso et al., 2008; Rosenstein et al., 2010). In particular, for the VEGF family, comprised of VEGF-A to VEGF-F and the placental growth factor (PlGF), although some evidence shows angiogenic roles of VEGF-C and PlGF (Freitas-Andrade et al., 2012; Gaál et al., 2013), VEGF-A is the main central actor in angiogenesis. Meanwhile, for the FGF family, grouped into 6 subfamilies, the acidic FGF (aFGF) and the basic FGF (bFGF) FGF1 subfamily members, are involved in the maintenance of CNS vasculature integrity in adults and healing after brain injury (Dordoe et al., 2021). In addition, VEGF-A and bFGF interact synergistically in boosting angiogenesis (Harrigan, 2003) (Fig. 3). Another key factor that promotes angiogenesis is the hypoxia-inducible protein complex (HIF-1) and specifically HIF-1 α transcription factor, which is expressed in hypoxic conditions and enhances VEGF-A gene expression, inducing blood vessel growth (Hoeben et al., 2004) (Fig. 3). Also, the transforming growth factor β 1 seems to play a role in CNS revascularization (Du et al., 2020; Li et al., 2010a). In this sense, a study showed that mice subjected to cortical freeze lesion presented high mRNA and protein levels of these growth factors, leading to neovascularization by 20 days post-injury (Penkowa et al., 2000).

All these growth factors bind to their specific receptors (Fig. 3), triggering the cascade of angiogenic signaling processes. In particular, VEGF family members

Fig. 2. Angiogenesis is governed by a dynamic balance of stimulators and inhibitors. In normal brain, angiogenesis is tightly downregulated but ischemic or hypoxic situations in central nervous system lead to an alteration of the physiologic anti-versus pro-angiogenic balance to try to compensate for vessel damage or degeneration. The most potent pro-angiogenic factor is vascular endothelial growth factor, followed by fibroblast growth factor. Pro-angiogenic factors also include transforming growth factor- β 1, placental growth factor and interleukins, while notable among anti-angiogenic factors are angiostatin, endostatin and thrombospondins-1 and -2 (Harrigan, 2003; Lawler and Lawler, 2012). Identifying the signals that regulate central nervous system vascularization in health and disease offers new insights for therapeutic strategies against central nervous system diseases.



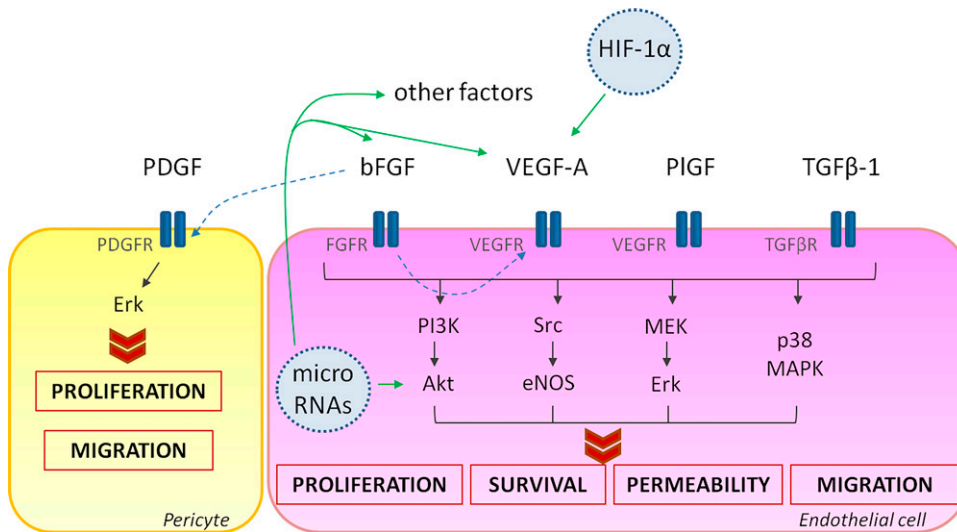


Fig. 3. Overview of the main factors and signaling pathways involved in brain angiogenesis. Schematic representation of the main growth factors and their respective receptors, that trigger the cascade of intracellular signals that induce cell proliferation, survival, permeability, and migration. Especially in hypoxic situations, these pathways, as well as hypoxia-inducible factors (such as HIF-1 α and some microRNAs), are stimulated and promote angiogenesis. As shown, some of these factors can interact with each other, leading to synergistic angiogenic effects.

bind to tyrosine kinase receptors VEGFR1-R3, where VEGFR2 is the main receptor in ECs and mediates mitogenesis and vascular permeability regulation. Even though VEGFR1 is involved in the first stages of vasculature development by promoting EC division, and despite having high affinity for VEGF-A, it has low kinase activity; hence, VEGF-A angiogenic effects are predominantly addressed through its highly homologous VEGFR2 (Abhinand et al., 2016). Meanwhile, VEGF-B and PlGF bind to VEGFR1 and VEGF-C and -D to VEGFR3 (Simons et al., 2016). Additionally, some isoforms of VEGF-A and PlGF have the ability to bind to the co-receptor neuropilin1 (NRP1), which enhances their binding affinity to VEGFR2 (Herzog et al., 2011). Regarding bFGF, this growth factor exerts its angiogenic effects through FGFR1 and, in turn, the signaling cascade can activate *Vegfr2* gene expression via the adapter protein FRS2 α and making ECs more sensitive to VEGF-A stimuli (Simons et al., 2016) (Fig. 3). This fact explains the synergistic angiogenic effect of bFGF and VEGF-A mentioned above. Additionally, it has been described that bFGF can upregulate PDGFR expression in CNS pericytes (Fig. 3) after ischemic stroke, which could contribute to both neuroprotection and angiogenesis (Nakamura et al., 2016).

2. Extracellular Matrix Signals and Interactions.

The ECM is essential to provide signals for reaching an adequate vascular cell survival, proliferation, migration, differentiation, and maturation to establish a functional vascular network where nascent vessels had matured into durable, stable, non-leaky and functional vessels. During maturation, the new vessel is surrounded by pericytes, which inhibit EC proliferation (Folkman and D'Amore, 1996), but if vessel stabilization does not occur, then the immature vessel undergoes apoptosis (Benjamin et al., 1998). There is evidence that ECM-integrin interactions play an important role in regulating vessel maturation in the CNS (Milner and

Campbell, 2002). In this sense, it has been shown in developing mice that CNS blood vessel maturation was accompanied by an increased expression of β 1 integrin along with α 4 and α 5 integrins in the early development, and with α 1 and α 6 in the late development and in adulthood (Milner and Campbell, 2002). Under pathological conditions, integrins α V β 3 are expressed in the activated microvessels of CNS ECs and have essential functions in the angiogenesis activation that occurs in cerebral ischemia (Abumiya et al., 1999). In the opposite scenario, an antagonism of integrin α V has been employed in experimental models to reduce brain glioma size by the inhibition of angiogenesis (MacDonald et al., 2001).

Also, metalloproteins, which are ECM proteins able to activate growth factors, surface factors, and adhesion molecules, seem to play key roles in maintaining CNS vasculature integrity. In this sense, the use of a metallothionein knockout model has revealed important functions for metallothioneins, a family of cysteine-rich metalloproteins (Penkowa et al., 2000). As a consequence of the lack of these metalloproteins, the levels of trophic factors VEGF, FGF, and transforming growth factor β 1 decreased, which hampered CNS recovery as late as 90 days post-lesion, inflicted by a focal cerebral cryoinjury, due to a reduced angiogenesis and late regeneration.

3. MicroRNAs. Interestingly, vascularization of CNS is also regulated epigenetically by microRNAs (miRs). These small non-coding RNA molecules of 22–24 nucleotides in length play a relevant role in a wide variety of biologic processes through the post-transcriptional modulation of genes. Recent studies have indicated the key function of miRs in the regulation of angiogenesis (Suárez and Sessa, 2009) (Fig. 3). Some of them, especially the miR-9 and miR-30 families, are mainly involved in the development of CNS vasculature (Cho et al., 2019; Madelaine et al., 2017).

In the cases in which cerebral microvasculature is compromised, miR-15a/16-1 cluster (Sun P et al., 2020), miR-124 (Li et al., 2019a), miR-126 (Nammian et al., 2020; Qu et al., 2019), and miR-210 (Zeng LL et al., 2016) seem to be potential candidates for brain revascularization, as will be discussed below.

III. Vascular Disorders in Brain CNS Diseases

Until only a few years ago, research on the design of effective therapies for CNS diseases had been focused mainly on neurons, what has been referred to as the neurocentric view. Although placing great emphasis on neurons has provided deep knowledge about their impaired cell biology during chronic neurodegenerative situations, it fails to address the underlying disease pathology and, in consequence, this strategy has not given rise to disease-modifying therapeutics. The explanation is that non-neuronal cells are also involved in the complex pathogenesis of CNS diseases and play key roles in the neurodegenerative process.

Brain microvasculature has gained attention in recent years in many CNS diseases, such as ischemic stroke, AD, ALS, VP, VaD, and aging (Fig. 4), since it is a pivotal element to maintain or improve the neurovascular network and, therefore, brain functioning. In these CNS diseases, the neurovascular unit (Fig. 2) is damaged, mainly by the injury of ECs, affecting the vascular network and, consequently, the neurovascular coupling due to the deficit in signaling, in neuronal stem cell proliferation/migration, as well as the lack of oxygen and nutrients normally provided by ECs and BBB exchange (Hatakeyama et al., 2020) (Fig. 2). All this leads to a cascade of events (Fig. 4) that contribute to the loss of cognitive function or other basic brain functions. The insults for such injury include BBB disruption, altering not only the homeostatic movement of ions, nutrients, and cells between the blood and the CNS but also the clearance of xenobiotic or neurotoxic components. The impairment of clearance pathways can also foment the accumulation of unwanted injurious molecules prompting proteinopathies. Disruption of the neurovascular unit can also provoke a reduction in cerebral blood flow, which may lead to a diminished brain oxygenation and hence hypoxic microenvironment. On the other hand, less trophic factors are produced and released in response to the insult, which consequently diminishes the chemotactic signals for EC migration and proliferation, contributing to a higher vulnerability and susceptibility of neurons and glial cells to the disease. Additionally, the damaged neurovascular unit can ultimately degenerate, leading to a decrease of the vascular network which would aggravate all the aforementioned issues.

All of the stated CNS diseases (Fig. 4) present a decline in microvasculature and reduced cerebral

blood flow. Additionally, this blood flow typically diminishes at a rate of 0.5% per year during aging; thus, in the elderly, a 20% flow reduction is normally observed (Leenders et al., 1990; Zou et al., 2009). This fact is yet an aggravating factor for neurodegenerative diseases in which the chronic brain hypoperfusion is added to the aging-related decline of cerebral blood flow.

A. Acute Events

Among stroke cases, 80% comprise ischemic stroke, hemorrhagic stroke is responsible of 15% and 5% are of unknown etiology (Beal, 2010). It is particularly a matter of concern that epidemiologic studies report an increasing incidence and proportion of young adult patients with stroke (Ekker et al., 2018). Control of hypertension, along with a moderate or low salt intake, is the major determinant of the long-term declines in cerebrovascular diseases (hemorrhagic and ischemic stroke) mortality (Levi et al., 2009; Petersen et al., 2019). Also, endogenous levels of other substances, such as the anti-oxidant ascorbic acid, also known as vitamin C, have been studied in ischemic stroke models since evidence supports that vitamin C deficiency could be a risk factor for stroke (Chen et al., 2013; Sánchez-Moreno et al., 2004; Yokoyama et al., 2000) as it functions as an inhibitor of the reactive oxygen species (ROS) produced in response to ischemic pathophysiology.

In ischemic stroke, the cerebral artery occlusion provokes a decrease in blood flow and an ischemic environment which leads to brain dysfunction. In this process, the brain tissues are not affected equally, and hence, the cellular response is different in the ischemic core and in the penumbra region (Bandera et al., 2006). In the ischemic core, a severe ischemia occurs, in which necrosis and irreversible cell damage take place, while in the penumbra region, there are metabolically active cells that can be recoverable if reperfusion or increased collateral flow can be provided to sustain the low-flow area. If not, it will evolve/expand into the final injury/infarction as well. Therefore, the penumbra is in the spotlight for therapeutic applications (Baron, 1999; Jung et al., 2013). Angiogenesis and tissue remodeling is highly active following stroke, particularly in penumbral regions. Angiogenesis promotes the delivery of blood flow and cell metabolism recovery to ischemic tissue and is positively correlated with the survival rate of ischemic stroke patients (Arenillas et al., 2007). Therefore, it is reasonable to assume that pro-angiogenic therapies would be beneficial for the improvement or recovery of a correct neurovascular and brain functioning. Currently, the treatment is limited to the intravenous administration of the thrombolytic recombinant tissue plasminogen activator (rt-PA), but this has a narrow therapeutic window (IST-3 collaborative group et al., 2012), within the 6 hours after

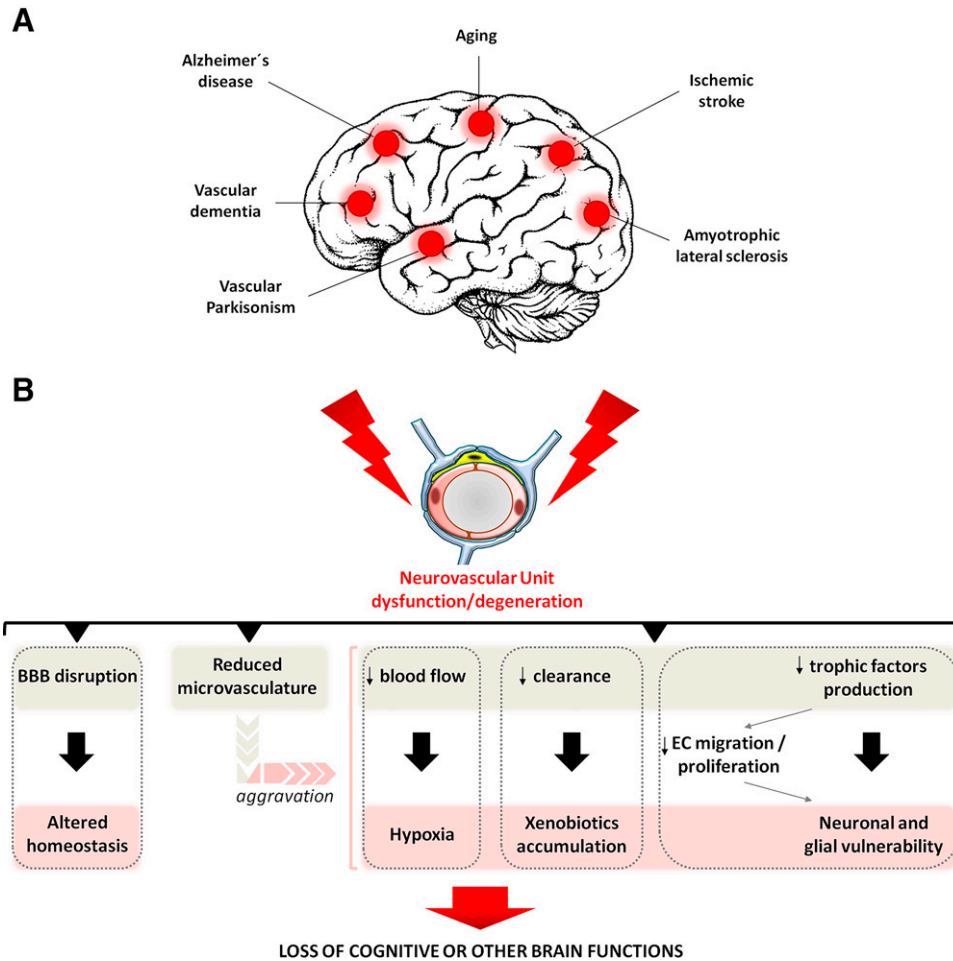


Fig. 4. Brain microvasculature damage/decrease is involved in many central nervous diseases. (A) Main central nervous diseases in which a reduction in brain microvasculature has been described. (B) Cascade of events that occur in the illustrated diseases due to the damage/degeneration of the neurovascular unit. BBB, blood-brain-barrier; EC, endothelial cell.

onset of stroke, and injurious effects, such as intracranial hemorrhage and neurotoxicity (Fan et al., 2016; Kaur et al., 2004). Interventional surgery for mechanical endovascular thrombectomy is another treatment option in the case of acute ischemic stroke due to large-vessel occlusion. In particular, blood clot retrievers, combined or not with rt-PA, have demonstrated improved functional outcomes, achieving reperfusion, and reduced healthcare costs (Campbell et al., 2017; Saver et al., 2015; Zhu et al., 2019). However, additional efforts aimed at longer-term recovery could be facilitated by inducing angiogenesis into penumbra, which is additionally supported by clinical observations demonstrating a strong correlation between neovascularization and functional recovery, suggesting that newly formed vessels contributed to behavioral improvements after ischemic stroke (Gunsilius et al., 2001).

B. Neurodegenerative Diseases

Chronic brain hypoperfusion is present in AD, ALS, VP, VaD and aging, and may be ahead of the appearance of clinical symptoms or neurodegenerative signs,

so it is quite feasible that the pathogenesis of these neurodegenerative diseases could depend on the formation of vascular pathology and/or loss of microvasculature (de la Torre, 2017).

1. Alzheimer's Disease. AD is the most common cause of dementia. The greatest risk factors for late-onset Alzheimer's are older age, up to 65 years of age, (Hebert et al., 2010), genetics (Farrer et al., 1997) and having a family history of Alzheimer's (Green et al., 2002; Lautenschlager et al., 1996; Mayeux et al., 1991). Pathologically, it is characterized by the deposition of extracellular protein aggregates of amyloid- β ($A\beta$), forming senile plaques, and intracellular neurofibrillary tangles containing phosphorylated tau in the brain parenchyma. $A\beta$ also accumulates in the vessels as cerebral amyloid angiopathy (CAA). However, the understanding of the pathophysiology of AD is constantly changing, and many impaired factors seem to play a role in the disease process (Anand et al., 2014). In fact, although many clinical trials have been carried out through immunotherapy by employing monoclonal antibodies against the presumably main

cause of AD, the A β plaques, all of them have failed (Huang et al., 2020). This strengthens the concept of a multifactorial pathophysiology in AD. There is also a genetic predisposition to AD such as inheriting the APOE4 gene. In particular, the APOE4 ϵ 4 allele is present in around 14% of the general population and in 40–60% of all cases of AD as one -showing 3.5-fold higher risk of developing AD, or two copies, increasing the risk by 8–15-fold (Holtzman et al., 2012; Michaelson, 2014).

Clinically, AD manifests itself by a gradual decline of cognitive function. Currently available treatments for AD dementia are aimed at treating the symptoms by manipulating the levels of two neurotransmitters, acetylcholine and glutamate, but do not alter the course of the disease. They consist of four cholinesterase inhibitors, tacrine, donepezil, rivastigmine, and galantamine, and a N-methyl-d-aspartate (NMDA) receptor antagonist, memantine, which are approved by the US Food and Drug Administration (Cummings et al., 2014). Vascular risk factors, as well as aging, stroke atherosclerosis and diabetes mellitus enhance the risk to of developing AD (Gorelick, 2004), which together may result in a decreased response to vasodilators and reduced cerebrovascular blood flow, leading to local hypoxia (Aliev, 2011).

The role of microvasculature damage/dysfunction in AD is increasingly widespread (Bersini et al., 2020; Liesz, 2019; Zlokovic, 2005). The deposition of A β in the cerebrovasculature causes a decline in cerebral perfusion and promotes ischemic damage, accompanied by vascular and neuronal degeneration and subsequent cognitive decline (Di Marco et al., 2015; Roth et al., 2005). Additional evidence suggests that the chronic cerebral hypoperfusion and A β accumulation can boost an impaired angiogenesis that results in dysfunctional microvessels (Biron et al., 2011). Paradoxically, in this scenario, several pro-angiogenic factors are released by brain microvessels in AD (Grammas, 2011), VEGF, IL-1 β , IL-6, IL-8, TNF, TGF β , MCP1, thrombin, angiopoietin 2, α V β 3, and α V β 5 integrins, and hypoxia inducible factor-1 α (HIF-1 α); however, no increase in functional vascularization occurs. In fact, some studies report vascular regression and decreased microvasculature density in AD brain (Brown and Thore, 2011). The explanation is that this endogenous VEGF binds directly to A β peptides, which would result in a local deficiency of accessible VEGF and, consequently, a cerebrovascular degeneration and reduced neuroprotection (Patel et al., 2010; Yang, S. P. et al., 2004). In this regard, current advances in brain vascular vessel imaging techniques, such as micro-optical sectioning tomography, have allowed scientists to precisely visualize and quantify the reduction of the cerebral vascular network, in terms of vessel mean diameter and volume fraction, in AD mouse models

compared with wild-type mouse models (Zhang X et al., 2019).

Some homeobox genes are involved in both physiologic and pathologic scenarios related to vascular remodeling in the adult. In particular, the homeobox gene MEOX2 regulates vascular differentiation (Gorski and Walsh, 2003). In this sense, it has been described that the low expression of the MEOX2 gene (Table 1) found in AD brain is associated with aberrant angiogenesis and vessel regression, ultimately resulting in reductions in brain capillary density and cerebral blood flow (Wu et al., 2005).

2. Amyotrophic Lateral Sclerosis. ALS is a neurodegenerative disorder characterized by progressive motor neuron degeneration in the brain and spinal cord, provoking muscle atrophy, paralysis, and early death since the diagnosis (Haverkamp et al., 1995; Rowland and Shneider, 2001). Most ALS cases are sporadic, and only around 15% of cases have a genetic basis, with more than 20 genes associated with the disease, such as hexanucleotide expansions in chromosome 9 open reading frame 72 (C9orf72), mutations in superoxide dismutase 1 (SOD1), TAR DNA-binding protein 43 (TARDBP), fused in sarcoma (FUS) and TANK-binding kinase 1 (TBK1) (van Es et al., 2017). To date, there is no cure or effective treatment of ALS, and only multidisciplinary care, nutritional and respiratory support, and symptom management are conducted.

Compelling evidence demonstrates the existence of CNS microvascular pathology and neurovascular unit impairment in ALS (Garbuzova-Davis et al., 2011) revealed by the reduced capillary blood flow in spinal cord of ALS mice (Garbuzova-Davis et al., 2007; Zhong Z et al., 2008) and brain of ALS patients (Rule et al., 2010; Waldemar et al., 1992). This would provoke vascular hypoperfusion/dysregulation which occurs before motor neuron degeneration or brain atrophy. Additionally, a significant decrease of circulating ECs has been found in peripheral blood of ALS patients at different disease stages, highlighting an impaired endothelization throughout the course of the illness (Garbuzova-Davis et al., 2010). The microvascular involvement greatly influences the understanding of ALS pathogenesis, demanding reconsiderations in therapeutic approaches and highlighting new treatment strategies to recover or maintain the neurovascular unit function and thus, promote neuron survival and correct CNS functioning (Vangilder et al., 2011).

3. Vascular Parkinsonism. Vascular changes in the brain are the hallmarks of VP (Korczyn, 2015), which accounts for 3–5% of all patients with parkinsonism (Jellinger, 2003). Although the exact changes in blood vessels have not yet been described in detail, these changes are normally ischemic and affect areas of the brain that are relevant to parkinsonism, including

TABLE 1
Potential candidates for revascularization in brain CNS diseases where cerebral microvasculature is impaired

Target	Name	Function	CNS disease	Ref.
VEGF	Vascular endothelial growth factor	Cooperates with Angiopoietin-1 to mediate angiogenesis and vessel maturation in ischemic brain. Binds to A β peptides present in AD brain microvasculature, which might result in local deficiency of accessible VEGF, leading to cerebrovascular degeneration and reduced neuroprotection. Preliminary evidence suggests a relationship between vasculature, hypoxia and motor neuron survival in ALS.	Focal cerebral ischemia, Alzheimer's disease, Amyotrophic lateral sclerosis and aging	(Evans et al., 2013; Patel et al., 2010; Zhang et al., 2000; Zhang et al., 2002)
PlGF	Placental growth factor	Contributes to neuroprotection, angiogenesis, vessel growth and maturation, maintaining vessel permeability. Synergistic angiogenic role with VEGF-A in hypoxic conditions.	Cerebral ischemia	(Carmeliet et al., 2001; Freitas-Andrade et al., 2012)
ANGPTL4	Angiopoietin-like 4	Potent inducer of cerebral neovascularization under hypoxic conditions. May also serve as diagnostic biomarker in patients with clinically assessed vascular cognitive impairment.	Alzheimer's disease, aging and ischemic stroke	(Bersini et al., 2020; Bouleti et al., 2013; Chakraborty et al., 2018)
bFGF	Basic fibroblast growth factor	Maintains the integrity of cerebral microvasculature. Angiogenic and neuroprotective effects under hypoxia, promotes the proliferation and migration of pericytes via its interaction with PDGF-BB. Interacts with VEGF-A synergistically in promoting angiogenesis.	Cerebral ischemia, ischemic stroke	(Dordoe et al., 2021; Harrigan, 2003; Lyons et al., 1991; Nakamura et al., 2016)
$\alpha v\beta 3$	Integrin $\alpha v\beta 3$	Expressed in the activated microvessels of CNS endothelial cells in response to hypoxia. Essential function in the angiogenesis activation.	Cerebral ischemia	(Abumiya et al., 1999)
NAD ⁺	Nicotinamide adenine dinucleotide	Protects the integrity of cerebral microvasculature by controlling endothelial cells cellular metabolism, energy production and survival.	Aging	(Csiszar et al., 2019)
MEOX2	MEOX2 gene (also known as GAX)	Promotes the angiogenic response of CNS endothelial cells to hypoxia, suppresses apoptosis and increases A β clearance efflux.	Alzheimer's disease	(Wu et al., 2005)
miR- 15a/16-1	microRNA- 15a/16-1 cluster	Controls VEGF and FGF expression regulating angiogenesis and cerebral blood flow.	Cerebral ischemia	(Sun P et al., 2020; Yin et al., 2012)
miR-30a	microRNA-30a	Controls BBB damage, infarct volume and neurovascular deficit caused by zinc accumulation in microvessels under ischemia, targeting the ZnT4 zinc transporter.	Ischemic stroke	(Wang P et al., 2020)
miR-124	microRNA-124	Regulates cerebrovascular impairment, including the decline in microvascular density and reduced angiogenesis.	Alzheimer's disease	(Li et al., 2019a)
miR-126	microRNA-126	Regulates angiogenesis and neurogenesis by the proliferation and migration of endothelial cells.	Focal cerebral ischemia	(Qu et al., 2019)
miR-210	microRNA-210	Acute ischemic stroke patients with higher circulating blood miR-210 show better clinical outcomes. Circulating blood miR-210 level associates with brain miR-210 level in a mouse model of ischemia, representing	Ischemic stroke	(Zeng L et al., 2011; Zeng L et al., 2014)

(continued)

TABLE 1—Continued

Target	Name	Function	CNS disease	Ref.
		a biomarker of brain injury and repair. Promotes vascular endothelial cell migration and tube formation under hypoxia, and associates with local increased levels of VEGF.		

subcortical white matter, basal ganglia, thalamus, and upper brainstem (Foltynie et al., 2002). Chronic ischemic changes that affect the subcortical white matter might result in gait disorders and cognitive impairment (Vale et al., 2012) that are not improved by dopaminergic medication (Benítez-Rivero et al., 2013). In fact, the pathologic vascular features of VP together with the unresponsiveness to treatment with dopaminergic drugs, distinguish this condition from idiopathic Parkinson's disease. Patients with VP have vascular risk factors, such as hypertension, dyslipidemia, and diabetes mellitus. Hence, modification of the VP disease course could be possible through a reduction of vascular risk factors or revascularization strategies.

4. Vascular Dementia. As stated in the National Institute on Aging (<http://www.nia.nih.gov/health/vascular-contributions-cognitive-impairment-and-dementia>), is called a cerebrovascular disease when the damage or pathology of brain microvasculature—also known as small vessel disease—leads to brain tissue injury—due to a decline of blood flow, oxygen or nutrients—contributing to cognitive impairment and dementia (Román, 2004). VaD terminology refers to people with dementia whose brain shows evidence of cerebrovascular disease. VaD is the second most common type of dementia after AD, contributing to nearly 17% of all dementias (Kapasi et al., 2017; O'Brien and Thomas, 2015). The location, number, and size of the brain injuries determine the severity of the dysfunction. In particular, it has been found that cortical microvascular pathology contributes to dementia through pyramidal cell loss (Krill et al., 2002). Therefore, restoration of cerebral microvasculature could potentially alleviate neuronal cell loss and improve cognitive function.

5. Aging. Aging constitutes the main risk factor for most neurodegenerative diseases, increasing their prevalence with increasing age (Hou et al., 2019). This fact could be directly or indirectly associated with the endothelial dysfunction (Bersini et al., 2020; Ungvari et al., 2013; Ungvari et al., 2020), decline in cerebral blood flow, around 20% flow reduction, (Leenders et al., 1990; Zou et al., 2009) and BBB leakage (Verheggen et al., 2020) that occurs with aging. Additionally, scientific consensus exists on the critical role of microvasculature contributions to cognitive impairment in elderly patients (Tarantini et al., 2017).

At the molecular level, age has been shown to impair angiogenesis through various pathways, including

lower endothelial nitric oxide synthase and HIF-1 α activity and decreased availability of VEGF (Lähteenpuo and Rosenzweig, 2012). Additionally, nicotinamide adenine dinucleotide (NAD⁺) deficiency has been associated with aging as a mediator of the endothelial dysfunction (Csiszar et al., 2019; Gomes et al., 2013).

IV. New Opportunities for Brain Revascularization

As explained before, angiogenesis is switched-on under the aforementioned brain hypoxic situations, in which the endogenous pro-angiogenic signals are not enough to achieve sufficient functional vessels to restore brain vascularization. Therefore, pro-angiogenic therapies seem to be a logical solution to this challenge. It is known that boosting brain angiogenesis implies an increase in the number of endothelial cells that can restore the performance of the neurovascular unit and cerebral blood flow. Hence, the extracellular regulatory signals secreted by the generated microvasculature, facilitate the proliferation and migration of neural stem cells as well as nutrients and oxygen supply, promoting neurologic recovery and cognitive function improvement (Hatakeyama et al., 2020).

Among the pro-angiogenic factors employed to achieve brain revascularization, VEGF-A is the most potent and widely employed candidate due to its angiogenic and neuroprotective effects (Korpisalo and Yla-Herttuala, 2010; Lange et al., 2016; Shim and Madsen, 2018). A relevant feature of VEGF function, linked to its therapeutic implications, is its interaction with ECM, which commands its localization in tissues and controls the outcome of the angiogenic process (Martino et al., 2015). Basically, there are three VEGF isoforms with diverse affinity for ECM (Ferrara, 2010), depending on the alternative mRNA splicing of the VEGF-A transcript, comprised of 121, 165, and 189 residues in humans or 120, 164, and 188 in rodents, respectively (Tischer et al., 1991). Among them, VEGF_{164/165} is the isoform of choice for therapeutic delivery since it is the only isoform able to promote physiologic vascular networks in the absence of the other isoforms, by generating intermediate gradients of concentration around cells with balanced matrix affinity (Ruhrberg et al., 2002). Additionally, as stated before, VEGF-A exerts synergistic effects in angiogenesis when combined with bFGF (Harrigan,

2003) (Fig. 3). In this sense, evidence shows that bFGF is able to induce neuroprotective and angiogenic effects in hypoxic situations by promoting the proliferation and migration of pericytes via its interaction with PDGF-BB (Nakamura et al., 2016) (Fig. 3).

For its part, the platelet-derived growth factor (PDGF) (Fig. 3) and its receptor PDGFR- β signaling have been studied in knockdown mouse models. They seem to play a key role in BBB restoration after cerebral ischemia, in part via the regulation of TGF- β signaling (Shen J et al., 2019), through the induction of pericyte recruitment, migration, and proliferation (Shen J et al., 2012). This occurs by the close coordination between ECs and pericytes, allowing the maturation of the emergent new vessels. In fact, the lack of this coordinated signaling occurs in ischemic stroke and neurodegenerative diseases, leading to brain microvasculature impairment and loss of cognitive function (Uemura et al., 2020).

Hence, identifying the specific signals involved in the microvasculature disruption in CNS highlights potential therapeutic angiogenic candidates for recovering the neurovascular network in stroke, neurodegeneration, and aging. Some of the potential candidates for such angiogenic strategy are summarized in Table 1.

A. Therapeutic Factors

1. Growth Factors. As mentioned before, the most potent and widely employed pro-angiogenic factor to achieve brain revascularization is VEGF-A (Table 1) due to its angiogenic and neuroprotective effects (Lange et al., 2016; Shim and Madsen, 2018). It has been shown that systemic or intracerebral early administration of soluble VEGF after stroke increased BBB opening, intensified edema, and promoted disorganized and immature vasculature, while its antagonist reduced the injury (Ma et al., 2012). Importantly, a work by Zhang et al. provided insights into the adequate time point for VEGF therapy, where intravenous VEGF-A administration 48 hours after MCAO increased microvessel density and improved cerebral microvascular perfusion in the ischemic penumbra, while administration at early stage, 1 hour after MCAO increased BBB leakage (Zhang ZG et al., 2000). This provided a starting point for subsequent studies regarding VEGF administration in ischemic stroke animal models, promoting angiogenesis, neurogenesis, reduced infarct size, and improved behavior outcomes (Chu K et al., 2005; Sun Y et al., 2003; Yang JP et al., 2009a).

It is assumed that other ischemia-induced factors are involved in controlling vascular integrity. Among them, angiopoietin-like protein 4 (ANGPTL4) (Table 1) is a secreted vascular growth factor induced by hypoxia in vascular cells in AD, aging, and stroke (Bersini et al., 2020; Bouleti et al., 2013; Chakraborty et al., 2018). It has been found that single intravenous injection in the tail vein of recombinant human ANGPTL4 1 hour

after MCAO in a mouse model of ischemic stroke, led to a reduction in infarct size and improved behavior performance (Bouleti et al., 2013). In this manner, ANGPTL4 seems to counteract the loss of vascular integrity, and consequently diminishes vascular leakage and cerebral edema. Similarly, bFGF (Table 1) intracerebral administration in animal models has evidenced some potential to induce angiogenesis in physiologic conditions (Puumala et al., 1990), but overall, in brain ischemic situations (Lyons et al., 1991) (Table 2), denoting the influence of hypoxia on the angiogenic effect of bFGF.

2. Bioactive Substances. Apart from vascular derived growth factors, bioactive substances, such as ligustilide and astragaloside IV, promote angiogenesis and recovery from ischemic stroke. In particular, it has been recently described that ligustilide, a bioactive substance isolated from *Ligusticum chuanxiong*, increases cerebral vessel number and neuroprotection after MCAO, and that the VEGF-endothelial nitric oxide synthase signaling pathway (Fig. 3) is involved in this process (Ren et al., 2020). Also, some studies have found that astragaloside IV, a triterpenoid saponin bioactive compound of *Astragalus mongholicus* with many beneficial effects in the brain, stimulates angiogenesis, including EC proliferation and migration, by increasing the expression of miRNA-210, which induces the activation of the HIF-VEGF-Notch signaling pathway (Fig. 3) (Liang et al., 2020). Other substances, such as the anti-oxidant ascorbic acid, have been studied in ischemic stroke models, since evidence supports that vitamin C deficiency could be a risk factor for stroke (Chen et al., 2013; Sánchez-Moreno et al., 2004; Yokoyama et al., 2000). Preclinical studies have shown that systemic administration of vitamin C after ischemic stroke, is able to maintain the BBB integrity and ECs tight junctions, protecting from the ischemic damage (Chang et al., 2020). In particular, this study showed that vitamin C reduced brain infarction volume and edema and also prevented neuronal damage; however, these outcomes were not because of angiogenesis. Even therapies for NAD⁺ repletion (Table 1) seem to protect the integrity of the cerebral microvasculature, improving cognitive performance in aging (Csiszar et al., 2019). As aforementioned, age-related NAD⁺ depletion has been associated with endothelial dysfunction. In this regard, restoring the NAD⁺ levels seems to protect the integrity of the cerebral microvasculature, improving cognitive performance in aged mice. In fact, the stimulation of cerebrovascular ECs isolated from aged F344xBN rats with nicotinamide mononucleotide (NMN), has evidenced an improvement in the angiogenic capacity, which could counteract, in part, the adverse effects of aging (Kiss et al., 2019). Some of the approaches for NAD⁺ repletion, aimed at the prevention of vascular

TABLE 2
Administration routes of pro-angiogenic therapies aimed at promoting brain revascularization in preclinical trials

Route	Procedure	Vector	Therapeutic factor	Dose	Duration of treatment	CNS disease	Ref.
Systemic	Tail vein injection/ infusion	-	rhANGPTL4	40 µg/kg	Single dose	IS	(Bouleti et al., 2013)
		-	rhVEGF ₁₆₅	1 mg/kg	5 µl/min, 4 hours	IS	(Zhang ZG et al., 2000)
		-			rhVEGF ₁₆₅ human neural stem cells	50 µg/kg500 µl	1hour10 ⁴ cells/µl, 5min
		IS	(Chu K et al., 2005)				
	AV transfected human mesenchymal stem cells	<i>PIGF</i>	10 ⁷ cells	Single dose	IS	(Liu H et al., 2006)	
Retro-orbital sinus	Fusogenic liposomes	Resveratrol	2 mg/kg/day	4 days	Ag	(Wiedenhoeft et al., 2019)	
Intraperito-neal	-	rhVEGF ₁₆₅	8 µg/kg/day	3 days	AD	(Wang P et al., 2011)	
-		Vitamin C	2000 mg/kg	Single dose	IS	(Chang et al., 2020)	
Intracerebral	Diffusion from cortex	PLGA nanospheres	rhVEGF ₁₆₅	1 µg/mouse	Single dose	AD	(Herran et al., 2013a)
		Encapsulated BHK-VEGF cells	hVEGF ₁₆₅	20–30 microcapsules	Single dose	AD	(Spuch et al., 2010)
		Encapsulated BHK-VEGF cells	hVEGF ₁₆₅	1microcapsule (10 ⁶ cells); 10-20 ng VEGF released/day	Single dose	IS	(Yano et al., 2005)
	Osmotic mini-pumps	-	rhVEGF ₁₆₅	10 µg/ml	1 µl/hour, 3 days	IS	(Sun Y et al., 2003)
		IgG antibody	Anti-Nogo-A	7 mg/ml	7 days	IS	(Rust, et al., 2019b)
	Local injection	AAV	hVegf ₁₆₅	5 µl (5x10 ⁹ TU/ml)	0.2 µl/min, 15 min	IS	(Shen F et al., 2006)
		AAVH9	hVegf ₁₆₅	5 µl (2x10 ⁹ TU/ml)	0.2 µl/min, 15 min	IS	(Shen F et al., 2008)
		AAV	<i>Sestrin2</i>	10 µl (10 ¹⁰ TU/ml)	0.5 µl/min, 20 min	IS	(Li Y et al., 2020)
		AAV	<i>PIGF</i>	3x10 ⁹ AAV particles	2 µl × 4 injections	IS	(Gaal et al., 2013)
		AV	hbFGF	50 µl (1x10 ⁸ TU)	3.3 µl/min, 15 min	IS	(Watanabe et al., 2004)
		Lentivirus	miR-210	2.2 µl (2x10 ⁹ TU/ml)	0.2 µl/min, 15 min	N, IS	(Zeng L et al., 2014; Zeng LL et al., 2016)
		Lentivirus	miR-126	2 µl (5.6x10 ⁸ TU/ml)	0.2 µl/min, 10 min	IS	(Qu et al., 2019)
		Lentivirus	miR-124	0.2 µl (10 ⁸ TU/ml)	Single dose	AD	(Li AD et al., 2019)
		Niosomes	hVegf ₁₆₅	5 µl	Single dose	N	(Gallego et al., 2020)
		PLGA microparticle	hNSCrhVEGF ₁₆₅	20 µg/µl (10 ⁴ cells)	Single dose	IS	(Bible et al., 2012)
	hNSC	hVegf ₁₆₅	2 µl (2x10 ⁵ cells)	Single dose	IS	(Lee et al., 2007)	
	HSV-1 transfected BMSC cells	hbFGF	10 µl (1x10 ⁶ cells)	Single dose	IS	(Ikeda et al., 2005)	
	HA hydrogel	rhVEGF ₁₆₅	6 µl	Single dose	IS	(Ju et al., 2014; Nih et al., 2016)	
	PEG-PPS hydrogel	iPS-NPC cells	-	Single dose	N	(Zhang J et al., 2011)	
Intranasal	Neuro-olfactory pathway	-	rhVEGF ₁₆₅	100 µl/day(200 µg/ml)	10 µl at a time for 18.5min, at intervals of 2min, alternating the nostrils. 3 days	IS	(Yang et al., 2009a)
		RVG29-PEG-PLGA nanoparticles	miR-124	2 drops containing miR-124 at 5 µg/ml	Every 2 days after the modeling during a week	IS	(Hao et al., 2020)

AAV, adeno-associated virus; AD, Alzheimer's disease; Ag, aging; ANGPTL4, angiotensin-like 4; AV, adenovirus; BHK, baby hamster kidney cells; BMSC, bone marrow stromal cells; CNS, central nervous system; HA, hyaluronic acid; HSV-1, herpes simplex virus type 1; iPS-NPC, human induced pluripotent stem-neural progenitor cells; IS, ischemic stroke; hMSCs human mesenchymal stem cells; hNSC, human neural stem cell; N, normal; NSC, neural stem cells; PEG, poly(ethylene glycol); PLGA, poly(lactic-co-glycolic acid); PPS, poly(propylene sulfide); rh, recombinant human; TU, transducing units; VEGF, vascular endothelial growth factor.

cognitive impairment, consist in the intraperitoneal administration of the poly(ADP-ribose) polymerase (PARP-1) inhibitor, named as PJ-34 (Tarantini et al., 2019), or of NMN in drinking water (Mills et al., 2016), among others reviewed by Csiszar et al. (Csiszar et al., 2019).

3. Considerations. The dosage and the spatio-temporal control of the factor delivery to avoid aberrant vessel formation, BBB leakage, gliomas, and other potential undesirable side effects (Manoonkitiwongsa et al., 2004; Ozawa et al., 2004; Uccelli et al., 2019; von Degenfeld et al., 2006), must be taken into account to achieve better clinical results. Accordingly, it is paradoxical that VEGF levels are usually upregulated after brain insults and how this endogenous molecule, far from being therapeutic, promotes vessel leakage, increases BBB permeability and produces aberrant angiogenesis with vessel regression, in neurodegenerative diseases overall (Dvorak et al., 1995; Manoonkitiwongsa et al., 2004). It has been shown that systemic or intracerebral early administration of soluble VEGF after stroke increased BBB opening, intensified edema, and promoted disorganized and immature vasculature, while its antagonist reduced the injury (Ma et al., 2012). In this sense, the dose of VEGF seems to play a crucial role not only in achieving therapeutic effects, but also in maintaining a correct balance between angiogenesis and brain tissue integrity. In particular, in contrast to non-angiogenic doses of VEGF₁₆₅, the administration of angiogenic doses of this growth factor in rat brain through invasive techniques augmented macrophage density, the typical cell marker of inflammation, in the cortical tissue of ischemic brains (Manoonkitiwongsa et al., 2006). Hence, the authors propose that VEGF monotherapy may not be enough to achieve a therapeutic effect. Accordingly, it has been suggested that a preferable and more complete approach would be the implementation of a cocktail of angiogenic factors rather than VEGF alone. In this sense, a strategy that combines growth factors for mature vessel formation would include VEGF, for promoting angiogenesis, and angiopoietin-1 (ANG-1), for inducing vessel maturation (Greenberg, 2015; Liu J, 2015; Valable et al., 2005; Zhang ZG et al., 2002).

Even though this strategy, consisting of the direct administration of the potential therapeutic factor has been widely explored, in ischemic stroke scenarios by systemic administration overall (Table 2), the approach has critical limitations due to rapid factor degradation and low stability in vivo (Rust et al., 2019a). Moreover, systemic circulation of these factors could affect other organs and, even in the target organ, the sudden local increase of the exogenous substance levels could create an important imbalance. For this reason, nanotechnology based approaches have been proposed in the last

decades to achieve a more sustained and safer delivery of the angiogenic factors, as explained below.

B. Nanotechnology Platforms

Nanotherapies are gaining momentum as a feasible medical option for the treatment of CNS diseases. Nanotechnology platforms, advanced therapies and biomaterials, constitute promising tools for an efficient and safe delivery of therapeutic factors. This approach enables the design and development of nanocarriers that allow the controlled and sustained release of drugs and gene-related products. This approach, protects therapeutic factors from degradation, improves biodistribution, and allows targeting cells/tissues. In fact, there are numerous papers reflecting the high efforts made in this promising field to achieve safe and effective drug or gene therapies targeting CNS (Agrawal et al., 2020; Lombardo et al., 2020; Mulvihill et al., 2020; Teixeira et al., 2020).

Different nanotechnology-based approaches for boosting angiogenesis have been employed, especially in wound healing repair, vectoring pro-angiogenic factors, and in anti-cancer therapies, vectoring anti-angiogenic factors. However, until now, few works have developed systems vectoring pro-angiogenic factors to promote revascularization in brain CNS diseases. Some of these nanotechnology strategies employed as drug, gene and even cell delivery systems focused on achieving therapeutic brain angiogenesis (Fig. 5, Table 2), are discussed below.

1. Nanocarriers. Currently, a nanocarrier-based clinical trial is ongoing for CNS revascularization in ALS patients by the intracerebroventricular administration of a solution containing recombinant human VEGF₁₆₅ using an implanted drug delivery system consisting of an implanted catheter and a SynchroMed II Pump drug infusion system, known as sNN0029 (NCT01384162) (<https://www.clinicaltrials.gov/ct2/show/NCT01384162>). The phase I of this trial revealed that this therapy was safe and well tolerated by ALS patients.

a. Promoting angiogenesis. Regarding experimental studies, drug delivery of VEGF for angiogenic purposes has been carried out employing polymeric poly(lactic-co-glycolic acid) (PLGA) nanospheres in the amyloid precursor protein/presenilin-1 transgenic mouse model of AD (Herrán et al., 2013a) (Table 2). PLGA nanoparticles are colloidal systems combined with the biodegradable and biocompatible PLGA polymer, which has been approved by the US Food and Drug Administration for several therapeutic applications. In the aforementioned work, 3 months after VEGF loaded nanosphere implantation, the vascular density in the cerebral cortex was augmented by 51% compared with control mice group, accompanied by a reduction in β -amyloid deposits, and improvement in memory deficit. This

approach using VEGF-PLGA nanospheres has also been employed also in Parkinson's disease with neuroregenerative results (Herrán et al., 2013b), showing ability to cross the BBB efficiently (Meng et al., 2020). However, the mechanism of VEGF-PLGA nanoparticles for crossing the BBB remains unclear and would require nanoparticle-targeting track, as well as an analysis of the potential side-effects of this therapy in other organs.

b. Facilitating the function of remaining microvasculature. In the case of aging, rather than promoting angiogenesis, some researchers have focused their attention on the recovery of the endothelial function. In this sense, it has been observed that treatment with fusogenic liposomes loaded with resveratrol, a plant-derived polyphenolic stilbene, rescued endothelial function and neurovascular coupling responses in aged mice in vitro in cerebrovascular ECs (Csiszar et al., 2015) and in vivo after retro-orbital (RO) injection (Wiedenhoeft et al., 2019) (Table 2). On the one hand, resveratrol had previously been implemented for endothelial dysfunction recovery by modulating nitric oxide metabolism and protecting against ROS through the activation of Nrf2 antioxidant defense mechanism in EC and vascular smooth muscle cell cultures (Li et al., 2019b). In fact, treatment of aged rodents with resveratrol in food, 200 mg/kg/d for 10 days, exerts protective effects on the cerebral microcirculation and cognitive function (Oomen et al., 2009; Toth et al., 2014). On the other hand, the use of fusogenic liposomes, consisting of the neutral lipid DOPE, the positively charged lipid DOTAP, and the aromatic resveratrol, significantly enhanced the drug uptake into endothelial cells through their fusion with cell membranes, overcoming the limited endocytic uptake, often below 1%, of conventional liposomes (Csiszar et al., 2015; Wiedenhoeft et al., 2019) (Table 2). Other liposome-associated limitations include low stability, quick metabolic degradation of phospholipids, fast systemic clearance, poor sustained release, and moderate efficiency for the entrapment of lipophilic drugs (Wong et al., 2012). Some of these problems have been overcome by surface coating the liposomes with polyethylene glycol (PEG) to extend the liposomes circulating time.

2. Gene Therapy Systems. Therapies based on gene therapy approaches, divided into viral and non-viral vectors, have emerged in recent years as potential tools for the treatment of many congenital, acquired and age-related diseases (Ingusci et al., 2019). Gene therapy strategies cover the chance of developing gene editing, the translation of the gene into endogenous protein/factor of interest, provides the possibility of achieving a stable or inducible expression of the therapeutic gene, as well as a specific expression in target cells.

Clinical trials regarding *Vegf* gene administration are currently ongoing and one of them, known as Neovasculgen (<https://www.clinicaltrials.gov/ct2/show/NCT02538705>) (pl-veg165; NCT02538705), has been commercialized since 2011. It consists of the gene transfer of the *Vegf*₁₆₅ gene embedded in a DNA plasmid vector (pCMV-veg165) for therapeutic angiogenesis in ulcers related to diabetic foot syndrome by intramuscular injections. Particularly, it increases the number of functioning capillaries in ischemic tissues, but it can even be applied in combination with surgical revascularization to improve the long-term results of reconstructive operations. Neovasculgen applicability to CNS disorders is currently being tested in clinical trials for other angiogenic therapies, such as for peripheral nerve injury (NCT02352649) by performing intraneural injections (<https://www.clinicaltrials.gov/ct2/show/NCT02352649>).

a. Pretreatment with viral vectors to protect the vasculature. In the ischemic stroke scenario, it has been shown that intracranial administration of recombinant adeno-associated virus (AAV)-mediated *Vegf* transfer reduces infarct volume by 55% in mouse transient MCAO models (Shen F et al., 2006) and improves focal cerebral blood flow restoration (Shen F et al., 2008) (Table 2). However, these strategies were administered five days prior to the ischemic process, which would constitute more a preventive therapy than a potential treatment post-injury. Along the same line, a recent work has found an angiogenic role for *sestrin2*, a stress-inducible protein that maintains homeostasis, helps in cellular repair, and eliminates toxic metabolites as a result of various insults, including hypoxia. In particular, intracranial injection of adenoviruses and AAVs vectoring the *Sestrin2* gene two weeks before the insult promoted angiogenesis, reduced the infarct volume and brain edema, improving the neurologic function (Li Y et al., 2020) (Table 2).

b. Treatment with viral vectors to promote revascularization. After ischemic stroke, intracerebral administration of adenoviral (AV) vectors expressing bFGF 2 hours after the onset of MCAO can reduce the infarct area by 44.3%, leading to neurologic improvement (Watanabe et al., 2004) (Table 2). It has also been reported that this same strategy, but comparing the potential therapeutic effects of AV encoding the different VEGF family members, promoted the creation of mature microvessels without significant side effects in the case of PlGF, while VEGF-A and VEGF-C promoted angiogenesis accompanied by vessel leakage, and VEGF-B did not induce angiogenesis (Gaál et al., 2013). Hence, the authors proposed PlGF as a potential candidate for therapeutic brain revascularization (Table 2), although no behavioral evaluation was assessed in this study.

In addition to gene delivery, epigenetic approaches have been developed to promote cerebral angiogenesis using viral vectors and microRNAs. Most of them have been applied in ischemic stroke animal models

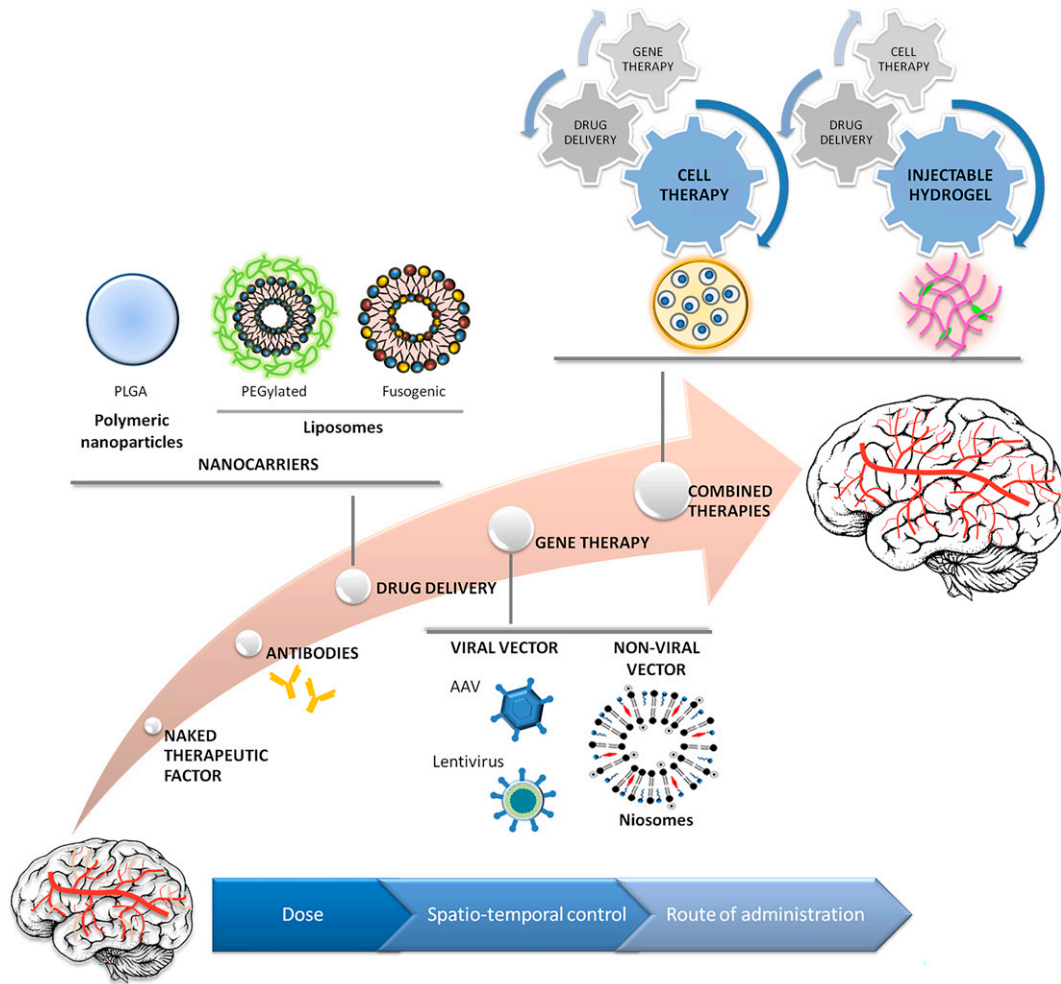


Fig. 5. Therapeutic opportunities employed for achieving brain angiogenesis and neurovascular network recovery. The design of the therapeutic strategy can embrace from the simplest form—with no vector—to the most complex approaches combining various technologic systems. During its implementation, relevant issues that should be considered include dosage of the therapeutic agent, concentration and number of doses, time point of administration, duration of the treatment, and the route of administration.

employing lentivirus vectors, since they can transfect neurons effectively. For instance, angiogenesis was achieved with lentiviral-mediated overexpression of miR-210 in normal mouse brain (Zeng L et al., 2014) and in ischemic mouse models by upregulating brain-derived neurotrophic factor (Zeng LL et al., 2016) (Table 2). Also, lentiviral-mediated overexpression of miR-126 promoted angiogenesis by activating AKT and ERK signaling pathways (Qu et al., 2019) (Fig. 3) (Table 2) or by regulating EC response to VEGF through repressing Sprouty-related EVH1 domain-containing protein 1 and phosphoinositol-3 kinase regulatory subunit 2 (PIK3R2/p85-b) (Nammian et al., 2020). MiR-15a/16-1 cluster endothelium-targeted knockdown models in vitro by lentivirus and in vivo by endothelial cell selective miR-15a/16-1 conditional knockout-enhanced post-stroke angiogenesis and cognitive recovery by upregulating the protein expression of VEGF-A, bFGF, and their receptors VEGFR2 and FGFR1 (Sun P et al., 2020) (Fig. 3). In a transgenic

amyloid precursor protein/presenilin-1 animal model of AD, lentivirus-mediated overexpression of miR-124 promoted angiogenesis and vascular integrity at the hippocampus and cerebral cortex levels by regulating the classic complement of the innate immune system C1ql3 (Li et al., 2019a) (Table 2).

c. Treatment with non-viral vectors to promote revascularization. Even though viral vectors are a good option for efficient gene delivery to CNS, they present many inconveniences related mainly to the relatively small packaging capacity, elevated costs and safety, immunogenicity, mutagenicity, risk of oncogenesis, and persistence of viral vectors in the brain (Kariyawasam et al., 2020; Mingozzi and High, 2013; Provost et al., 2005). Additionally, pre-clinical and clinical trials with viral vectors have evidenced severe symptoms associated with the high and uncontrolled expression of exogenous *Vegf* (Martino et al., 2015). In this regard, non-viral vector-based gene therapy strategies are gaining interest due to their safety profile, high packing capacity, low

cost, and high-scale production potential (Foldvari et al., 2016; Jayant et al., 2016). Additionally, non-viral approaches based on lipid nanoparticles allow a sustained release of the cargo that would also facilitate the desirable transient but prolonged *Vegf* expression needed for at least 4 weeks to allow newly induced vessels to stabilize and persist (Ozawa et al., 2004; Tafuro et al., 2009).

Among lipid nanoparticles, niosomes have been employed for in situ gene therapy purposes in CNS (Al Qtaish et al., 2020). Interestingly, brain angiogenesis has been achieved in vivo by the intracerebral administration of niosomes vectoring the *Vegf*₁₆₅ gene at the cortex level (Gallego et al., 2020) (Table 2). In this work, niosomes were able to efficiently deliver the therapeutic gene, achieved VEGF release to the extracellular medium, and promoted angiogenesis in mouse brain. In fact, vascular density 1 week after intracerebral injection increased 76% in an extensive area in the vicinity of the injection point, compared with controls. Also, polymeric nanoparticles represent an encouraging strategy, as is the case of microRNA-124-loaded RVG29-PEG-PLGA nanoparticles, which have been reported to prevent the ischemic brain injury, contributing to the recovery of neurologic function (Hao et al., 2020) (Table 2).

3. Other Approaches and Combined Systems. To deliver physiologic amounts of the angiogenic factor to the brain tissue in a continuous and localized manner, cell encapsulation provides an interesting approach. In this way, a polymer matrix surrounded by a semi-permeable membrane that encapsulates the engineered cells preserves them against immune cell-mediated and antibody-mediated rejection. Additionally, this membrane regulates the bidirectional diffusion, allowing a controlled and continuous delivery of the therapeutic factor (Gurruchaga et al., 2015; Orive et al., 2003). Even, dual gene therapy and cell therapy strategies, with or without encapsulation, can be implemented. The combination of the three systems, gene therapy, cell therapy, and cell encapsulation matrix, with angiogenic outcome has been applied to an animal model of AD (Spuch et al., 2010). In particular, fibroblast cells transfected to produce human VEGF were encapsulated in an alginate solution and coated with poly-L-lysine plus another alginate layer. These authors found that implantation of VEGF-secreting microcapsules onto the brain for 3 months induced angiogenesis, which contributed to an enhancement of A β clearance and behavioral improvement in amyloid precursor protein/presenilin-1 mice, proposing VEGF microcapsule implementation as part of the treatment in AD patients (Spuch et al., 2010) (Table 2). Another example of this multi-combinational approach has been implemented for brain angiogenesis after stroke by combining gene therapy to develop stable genetically

engineered cells and cell encapsulation in sterile capsules made of polysulfone hollow fibers (Yano et al., 2005) (Table 2). Here, authors first transfected cells with a plasmid encoding for VEGF₁₆₅ and established a VEGF-secreting cell line; then, these encapsulated cells and the capsules containing VEGF-secreting cells were locally administered into the brain parenchyma, achieving a continuous intracerebral release of VEGF₁₆₅. This design exerted neuroprotective and angiogenic effects on focal cerebral ischemia and led to significant functional recovery in rats after stroke. A similar approach, but with no encapsulation, has been also developed in stroke animal models, which involves transplanting into the brain the multipotent human neural stem cells overexpressing VEGF after gene transfection and selection with puromycin (Lee et al., 2007) (Table 2). Similarly, a therapeutic strategy combining viral gene therapy and cell therapy after ischemic stroke has been performed with bFGF gene-transferred bone marrow stromal cells (BMSCs) by the herpes simplex virus type 1 vector (Ikeda et al., 2005) (Table 2). With this approach, BMSCs are intended to secrete several growth factors and have the potential to differentiate into neurons and/or glial cells. Additionally, the enhanced bFGF release from these transduced BMSCs, can induce angiogenesis in hypoxic environments. Thus, intracerebral implantation of bFGF gene-transferred BMSCs 24 hours after MCAO achieved infarct volume reduction, accompanied by a significant brain function recovery 14 days after MCAO. In this same line, the intravenous infusion of human mesenchymal stem cells alone or previously transduced with adenovirus encoding PlGF, 3 hours after MCAO in rats, has evidenced the higher angiogenic effect of this cell therapy when combined with the enhanced expression of PlGF (Liu, H. et al., 2006) (Table 2). Specifically, 7 days after MCAO, the infarct size volume was reduced, angiogenesis was induced without cerebral edema, and this was accompanied by an improvement in behavioral performance.

Another dual strategy used is the combination of cell therapy with a drug delivery system. In this regard, the intracerebral implantation of human neural stem cells attached on the surface of VEGF-releasing PLGA microparticles after stroke, attracted host ECs to invade this neuroscaffolding, and allowed the development of a vascular network (Bible et al., 2012) (Table 2).

The design and development of hydrogels for brain therapy purposes represents an attractive therapeutic opportunity. They can be considered as scaffolds designed to match the mechanical properties of the brain by modulating the crosslinking density and to serve not only as local drug delivery systems (Nih et al., 2016) but also for tissue regeneration and cell transplantation purposes. In this line, some authors have proposed to employ a therapeutic angiogenic material delivering VEGF directly to the stroke cavity

to stimulate angiogenesis and repair brain tissue after stroke (Nih et al., 2018) (Table 2). To this end, they developed a biomaterial consisting of a hyaluronic acid hydrogel containing VEGF and injected it directly within the stroke cavity. The stroke cavity is the resulting damaged brain tissue after stroke which is deficient in extracellular matrix and that becomes a fibrotic scar devoid of neural tissue over time (Fitch et al., 1999). However, it represents a potential transplant space since it can accept a large volume of injection without affecting the unaffected brain tissue (Moshayedi et al., 2016; Nih et al., 2016; Zhong, J. et al., 2010). In this way, the injectable angiogenic biomaterial induced the formation of a vascular and neuronal structure that led to behavioral improvement (Nih et al., 2018). Similarly, a more complex combined approach using HA hydrogel and PLGA microspheres containing VEGF for angiogenesis and ANG-1 for vessel maturation has shown encouraging results in an MCAO mouse model of brain ischemia (Ju et al., 2014) (Table 2). Two weeks after MCAO, the HA-PLGA composite was implanted in the ischemic area, leading to in situ angiogenesis from the sixth week after MCAO and implantation, accompanied with a trend toward better behavior performance. On the other hand, cell transplantation has been carried out, employing an injectable hydrogel scaffold formed by PEG and poly(propylene sulfide) (PPS) (Zhang J et al., 2011) (Table 2). This 3D cell matrix, including human induced pluripotent stem-neural progenitor cells (iPS-NPCs) injected into the brain of naive mice, allowed for angiogenesis even without the addition of pro-angiogenic factors, enhancing stem cell survival. Interestingly, this matrix admits the incorporation of further desired signals, such as proteins, growth factors, and DNA.

Other authors have identified anti-Nogo-A antibodies as potential pro-angiogenic therapy for CNS ischemia that follows stroke, promoting a mature vascular network that, in turn, protects from vascular leakage (Rust et al., 2019b) (Table 2). The neurite outgrowth inhibitor Nogo-A is a membrane protein present in oligodendrocytes and some neurons that limits vascular growth in development and after CNS ischemia. In contrast, its neutralization enhances vascular repair in the ischemic border zone leading to functional outcome (Joly et al., 2018; Walchli et al., 2013).

V. Administration Routes for Targeting Brain Revascularization

Choosing the correct delivery strategy for the administration of the therapeutic systems is critical to ensure the effectiveness, safety, and functionality of the treatment. Each delivery route for the administration of therapies to reach the brain and promote new vessel formation, neurovascular network enhancement and cognitive/behavioral improvement in brain CNS

diseases, involves specific advantages and drawbacks (Fig. 6). Compelling evidence at the preclinical level shows that systemic and intracerebral routes are the most researched. In this regard, emerging administration strategies along with the implementation of nanotechnology-based systems (Fig. 5) are critical factors that will define the efficacy of the angiogenic therapy for the recovery of the neurovascular network (Table 2).

A. Systemic Administration

The route of choice for systemic administration is usually tail vein injection, where the administered factor is normally not associated with any vector to avoid potential interactions with biologic components present in blood flow. In fact, many of the therapies for brain revascularization or microvasculature protection discussed in this review have employed this approach by single injection or infusion with a mini-pump (Bouleti et al., 2013; Chu K et al., 2005; Zhang ZG et al., 2000). However, the RO sinus route in experimental animals has emerged as an alternative systemic route, which seems to be easier, faster, presents fewer complications, and reports lower distress to the animal compared with lateral tail vein injection (Wang F et al., 2015; Yardeni et al., 2011). Regarding the field of cerebral microvasculature impairment, fusogenic liposomes vectoring resveratrol for brain vasculature protection in aged mice have been administered by this RO route (Csiszar et al., 2015; Wiedenhoef et al., 2019). Also, intraperitoneal administration of VEGF in the AD PDGF-hAPP^{V7171} transgenic mouse has been carried out with angiogenic outcome in the hippocampus (Wang P et al., 2011), as well as ascorbic acid parenteral administration 1 hour after MCAO, reducing brain infarction volume (Chang et al., 2020). However, regardless of the systemic administration route, the challenge of affecting other organs still persists together with the therapeutic agent degradation and the difficulty in crossing the BBB (Rust et al., 2019a).

B. Intracerebral Administration

A large number of works have employed intracerebral administration to implement new opportunities for brain angiogenesis by administering naked factors, nanocarriers, gene therapy, cell therapy, biomaterials, antibodies, or systems combining these approaches (Table 2). The methods for intracerebral administration are the following:

Diffusion from cerebral cortex into brain: this approach has been carried out in the amyloid precursor protein/presenilin-1 mouse model of AD after brain craniotomy. 1) VEGF-loaded PLGA nanospheres have been dropped directly onto the surface of the cerebral cortex, enhancing the cerebral vascular density by 51% (Herrán et al., 2013a). 2) Microcapsules of VEGF-producing cells have been placed in the craniotomy site, resulting in a patch of alginate (Spuch

et al., 2010) or polysulfone (Yano et al., 2005) microcapsules overlying the cerebral cortex. Two weeks after their implantation, vascular density augmented gradually during 3 months, accompanied by $A\beta$ clearance and alleviation of the behavioral impairment (Spuch et al., 2010).

Osmotic mini-pumps: these have been employed to infuse the angiogenic factor, 1 day following ischemia, introducing a catheter by the intracerebroventricular route (Rust et al., 2019b; Sun Y et al., 2003).

Intracerebral injection: specially employed for gene therapy, cell transplantation and biomaterials as injectable hydrogels. 1) Viral gene delivery by local injection of AAV vectoring *Vegf* (Shen F et al., 2006) or *Sestrin2* (Li Y et al., 2020) has been carried out in stroke animal models prior to the insult to prevent neurovascular impairment. Also, viral delivery of *PlGF* (Gaal et al., 2013) and *hbFGF* (Watanabe et al., 2004), after MCAO, has been performed with AAV and AV, respectively. 2) Non-viral *Vegf* delivery into the cerebral cortex has been conducted by employing niosomes (Gallego et al., 2020), which enhanced angiogenesis by 55% in normal adult mouse brain, not only at the injection point but also in the vicinity. 3) Epigenetic regulation to achieve an angiogenic response has been developed employing lentiviral-mediated overexpression of miRs injected into the desired area in normal adult mouse brain, with miR-210 (Zeng L et al., 2014), in stroke animal models, with miR-126 (Qu et al., 2019) or miR-210 (Zeng LL et al., 2016), and in AD animal models, with miR-124 (Li et al., 2019a). 4) VEGF (Bible et al., 2012; Lee et al., 2007) and bFGF (Ikeda et al., 2005) secreting cells have been injected into brain parenchyma in stroke mouse models. 5) Injectable or hyaluronic acid hydrogels

containing VEGF are usually placed directly within the stroke cavity 2 weeks after MCAO (Ju et al., 2014; Nih et al., 2016), although 3D cell matrix of PEG and PPS hydrogels containing iPS-NPCs have also been injected into the brain of naive mice promoting angiogenesis and stem cell survival without the addition of pro-angiogenic factors (Zhang J et al., 2011).

C. Intranasal Administration

Many agents and gene vectors have been administered by intranasal administration in a wide number of disease models, some of them related to CNS diseases (Aly and Waszczak, 2015). The intranasal region of administration is also a pivotal item to take into account, since it influences the drug absorption to the brain or to the systemic circulation, in which case would cause side effects in other organs, especially in the lungs. In this sense, the olfactory area, posterior and upper region, is the proper position for drug absorption to target the therapeutic factor to the brain (Dhuria et al., 2010). Focusing on revascularization strategies for brain CNS diseases, it has been observed that the pro-angiogenic VEGF rapidly enters the brain by this route. In a first-step, work comparing intranasal versus intravenous administration of human recombinant VEGF-165 labeled with a radioisotope, it was found that within 30 minutes post-administration, significantly more VEGF was localized in many cerebral sites, including the cerebral cortex, following intranasal administration (Yang et al., 2009b). Although less VEGF was accumulated in peripheral tissues in the nose-brain pathway compared with the systemic pathway, the lungs accumulated the higher levels of the growth factor. In this regard, VEGF overexpression in

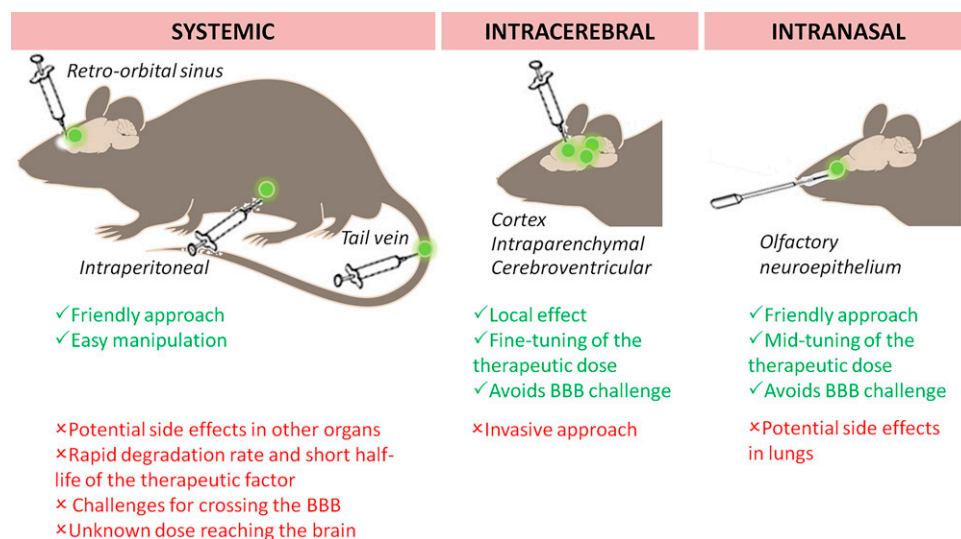


Fig. 6. Features of the potential routes used for the administration of angiogenic factors to brain. The goal of such delivery strategies is to promote angiogenesis, neurovascular network enhancement, and cognitive/behavioral improvement in brain central nervous system diseases, by surpassing the blood-brain-barrier and other biologic barriers. The main routes employed in animal models for therapies focused on brain central nervous system diseases are the systemic intravenous delivery and the intracerebral administration. However, it is now known that the olfactory neuroepithelium provides a non-invasive route of entry into central nervous system, bypassing the blood-brain-barrier.

the lungs can occur following intranasal delivery, which might increase pulmonary vascular permeability and cause pulmonary edema, as observed in mice after administration of an adenoviral vector expressing VEGF-165 through the respiratory tract (Kaner et al., 2000). This fact evidences once more the importance of dosage for achieving therapeutic effects in the target tissue without provoking side effects either in the brain or in peripheral organs. Addressing dose tuning, intranasal therapeutic VEGF dosage has been assessed in a rat model of ischemic stroke, 3 days after MCAO (Yang et al., 2009a) (Table 2). Although in this work authors did not evaluate VEGF accumulation and potential side effects in peripheral organs, with especial significance for lungs, they found an intranasal dosage-dependent VEGF effectiveness on reducing infarct volume and neurologic function, enhancing angiogenesis and improving behavioral recovery.

An interesting approach for intranasal delivery of VEGF to avoid direct contact and potential side effects of the factor on its way to brain, would be nanocarrier-mediated drug delivery or non-viral gene therapy. In particular, non-viral RVG29-PEG-PLGA nanoparticles loaded with miR-124 have been administered intranasally, reducing the ischemic brain injury (Hao et al., 2020) (Table 2).

VI. Conclusions and Future Directions

Although CNS diseases, and in particular the neurodegenerative ones, are an increasing reality in our society, there are still no curative pharmacological treatments able to achieve a complete neurovascular recovery. It is becoming increasingly evident that brain microvasculature impairment and deficiency is a key feature, not only in the development of neurodegenerative diseases, but also in aging. Until a few years ago, scientific research sought therapies focused on neurologic recovery. However, it is now known that vessel integrity is crucial for a correct functioning of the neurovascular unit, promoting neuron survival and preserving cognitive capacities. Thus, new therapeutic approaches must be reconsidered to improve the impaired microvasculature rather than target the affected neurons.

In this review, many brain revascularization strategies have been discussed, showing the great efforts made by the scientific community in this field. Although the majority of brain revascularization approaches are still in the experimental phase, expectations to a translational clinical application is in the process of becoming a reality. For this purpose, several nanotechnology platforms based on nanocarriers, gene therapy, cell therapy and biomaterials, among others, have been developed and have been discussed in detail in the present work.

Currently, some of these approaches are limited to intracerebral administration due to the nature of the vector and/or to avoid potential undesired side-effects in other organs. Although it is possible to reach the brain non-invasively in experimental animals, future translation to clinical practice would require the selective action of the therapeutic factor in the brain to avoid its distribution to other tissues. Taking into account the existence of the selective BBB, strategies to overcome this barrier, to deliver the therapeutic cargo by non-invasive routes of administration, are being developed. Actually, pre-clinically developed non-invasive strategies for brain revascularization require multiple administration doses at high concentrations, augmenting the non-specific systemic absorption and potential damage of other organs. In this sense, the nose-to-brain route of administration seems to be a promising strategy for regular clinical practice, since it represents a less invasive approach able to reach the brain with more precision than systemic administration. In this regard, to minimize the mucociliary clearance of the therapeutic factor in the intranasal vestibular region, optimized delivery devices are being developed. These systems include specially designed intranasal devices, mucoadhesive systems and surface-engineered nanocarriers, among others (Agrawal et al., 2018; Wang Z et al., 2019). Hence, the rise of device manufacturers will enable the benefits of the nose-to-brain delivery route to become translated into approved products for clinical practice. In addition to the development of nanotechnology platforms to recover brain microvasculature and cognitive function, powerful and sensitive imaging system platforms should be implemented. In this regard, cutting-edge techniques are being developed to visualize and analyze the entire brain vascular network to discriminate in detail the decline or increase after a therapeutic angiogenic treatment of microvasculature in the diseases discussed in this review. At the preclinical level, this is made possible by the combination of tissue-clearing methods to render the brain completely transparent (Silvestri et al., 2016) with advanced microscopic techniques, such as micro-optical sectioning tomography, which enables whole-brain imaging at the submicron level (Li et al., 2010b), or light-sheet fluorescence microscopy, employing a fluorescent blood vessel lumen staining for whole-brain vasculature reconstruction at the capillary level (Di Giovanna et al., 2018). At the clinical level, such a detailed vessel analysis it is still not possible; instead, current imaging systems are able to analyze cerebral blood flow. However, it constitutes a promising field in combination with potential pro-angiogenic therapies.

Acknowledgments

The authors wish to thank the ICTS "NANBIOSIS" and the Drug Formulation Unit (U10) of the CIBER in Bioengineering,

Biomaterials, and Nanomedicine (CIBER-BBN) and Charlotte A. White for the English revision.

Authorship Contributions

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References

- Abhinand CS, Raju R, Soumya SJ, Arya PS, and Sudhakaran PR (2016) VEGF-A/VEGFR2 signaling network in endothelial cells relevant to angiogenesis. *J Cell Commun Signal* **10**:347–354.
- Abumiya T, Lucero J, Heo JH, Tagaya M, Koziol JA, Copeland BR, and del Zoppo GJ (1999) Activated microvessels express vascular endothelial growth factor and integrin alpha(v)beta3 during focal cerebral ischemia. *J Cereb Blood Flow Metab* **19**:1038–1050.
- Agrawal M, Saraf S, Saraf S, Antimisariis SG, Chougule MB, Shoyele SA, and Alexander A (2018) Nose-to-brain drug delivery: An update on clinical challenges and progress towards approval of anti-Alzheimer drugs. *J Control Release* **281**:139–177.
- Agrawal M, Saraf S, Saraf S, Dubey SK, Puri A, Patel RJ, Ajazuddin, Ravichandiran V, Murty US, and Alexander A (2020) Recent strategies and advances in the fabrication of nano lipid carriers and their application towards brain targeting. *J Control Release* **321**:372–415.
- Al Qtaish N, Gallego I, Villate-Beitia I, Sainz-Ramos M, López-Méndez TB, Grijalvo S, Eritja R, Soto-Sánchez C, Martínez-Navarrete G, Fernández E, et al. (2020) Niosome-based approach for in situ gene delivery to retina and brain cortex as immune-privileged tissues. *Pharmaceutics* **12**:E198 <https://doi.org/10.3390/pharmaceutics12030198>.
- Aliev G (2011) Oxidative stress induced-metabolic imbalance, mitochondrial failure, and cellular hypoperfusion as primary pathogenetic factors for the development of Alzheimer disease which can be used as an alternate and successful drug treatment strategy: past, present and future. *CNS Neurol Disord Drug Targets* **10**:147–148.
- Aly AE and Waszczak BL (2015) Intranasal gene delivery for treating Parkinson's disease: overcoming the blood-brain barrier. *Expert Opin Drug Deliv* **12**:1923–1941.
- Anand R, Gill KD, Mahdi AA. (2014) Therapeutics of alzheimer's disease: Past, present and future. *Neuropharmacology* **76** Pt A:27–50.
- Arenillas JF, Sobrino T, Castillo J, and Dávalos A (2007) The role of angiogenesis in damage and recovery from ischemic stroke. *Curr Treat Options Cardiovasc Med* **9**:205–212.
- Bandera E, Botteri M, Minelli C, Sutton A, Abrams KR, and Latronico N (2006) Cerebral blood flow threshold of ischemic penumbra and infarct core in acute ischemic stroke: a systematic review. *Stroke* **37**:1334–1339.
- Baron JC (1999) Mapping the ischaemic penumbra with PET: implications for acute stroke treatment. *Cerebrovasc Dis* **9**:193–201.
- Beal CC (2010) Gender and stroke symptoms: a review of the current literature. *J Neurosci Nurs* **42**:80–87.
- Benitez-Rivero S, Marín-Oyaga VA, García-Solis D, Huertas-Fernández I, García-Gómez FJ, Jesús S, Cáceres MT, Carrillo F, Ortiz AM, Carballo M, et al. (2013) Clinical features and 123I-FP-CIT SPECT imaging in vascular parkinsonism and Parkinson's disease. *J Neurol Neurosurg Psychiatry* **84**:122–129.
- Benjamin LE, Hemo I, and Keshet E (1998) A plasticity window for blood vessel remodelling is defined by pericyte coverage of the preformed endothelial network and is regulated by PDGF-B and VEGF. *Development* **125**:1591–1598.
- Bersini S, Arrojo E, Drigo R, Huang L, Shokhirev MN, and Hetzer MW (2020) Transcriptional and functional changes of the human microvasculature during physiological aging and Alzheimer disease. *Adv Biosyst* **4**:e2000044.
- Bible E, Qutachi O, Chau DY, Alexander MR, Shakesheff KM, and Modo M (2012) Neo-vascularization of the stroke cavity by implantation of human neural stem cells on VEGF-releasing PLGA microparticles. *Biomaterials* **33**:7435–7446.
- Biron KE, Dickstein DL, Gopaul R, and Jefferies WA (2011) Amyloid triggers extensive cerebral angiogenesis causing blood brain barrier permeability and hypervascularity in Alzheimer's disease. *PLoS One* **6**:e23789.
- Bouleti C, Mathivet T, Coqueran B, Serfaty JM, Lesage M, Berland E, Ardidie-Robouant C, Kauffenstein G, Henrion D, Lapergue B, et al. (2013) Protective effects of angiopoietin-like 4 on cerebrovascular and functional damages in ischaemic stroke. *Eur Heart J* **34**:3657–3668.
- Brown WR and Thore CR (2011) Review: cerebral microvascular pathology in ageing and neurodegeneration. *Neuropathol Appl Neurobiol* **37**:56–74.
- Campbell BCV, Mitchell PJ, Churilov L, Keshtkaran M, Hong KS, Kleinig TJ, Dewey HM, Yassi N, Yan B, Dowling RJ, et al.; EXTEND-IA Investigators (2017) Endovascular thrombectomy for ischemic stroke increases disability-free survival, quality of life, and life expectancy and reduces cost. *Front Neurol* **8**:657.
- Carmeliet P, Moons L, Luttun A, Vincenzi V, Compernelle V, De Mol M, Wu Y, Bono F, Devy L, Beck H, et al. (2001) Synergism between vascular endothelial growth factor and placental growth factor contributes to angiogenesis and plasma extravasation in pathological conditions. *Nat Med* **7**:575–583.
- Chakraborty A, Karamerans A, van Het Hof B, Castricum K, Aanhane E, van Horssen T, Hijssen VL, Scheltens P, Teunissen CE, Fontijn RD, et al. (2018) Angiopoietin like-4 as a novel vascular mediator in capillary cerebral amyloid angiopathy. *Brain* **141**:3377–3388.
- Chang CY, Chen JY, Wu MH, and Hu ML (2020) Therapeutic treatment with vitamin C reduces focal cerebral ischemia-induced brain infarction in rats by attenuating disruptions of blood brain barrier and cerebral neuronal apoptosis. *Free Radic Biol Med* **155**:29–36.
- Chen GC, Lu DB, Pang Z, and Liu QF (2013) Vitamin C intake, circulating vitamin C and risk of stroke: a meta-analysis of prospective studies. *J Am Heart Assoc* **2**:e000329.
- Cho KHT, Xu B, Blenkiron C, and Fraser M (2019) Emerging roles of miRNAs in brain development and perinatal brain injury. *Front Physiol* **10**:227.
- Chu K, Park KI, Lee ST, Jung KH, Ko SY, Kang L, Sinn DI, Lee YS, Kim SU, Kim M, et al. (2005) Combined treatment of vascular endothelial growth factor and human neural stem cells in experimental focal cerebral ischemia. *Neurosci Res* **53**:384–390.
- Chu LH, Rivera CG, Popel AS, and Bader JS (2012) Constructing the angiome: a global angiogenesis protein interaction network. *Physiol Genomics* **44**:915–924.
- Cooke VG, Naik MU, and Naik UP (2006) Fibroblast growth factor-2 failed to induce angiogenesis in junctional adhesion molecule-A-deficient mice. *Arterioscler Thromb Vasc Biol* **26**:2005–2011.
- Csiszár A, Csiszár A, Pinto JT, Gautam T, Kleusch C, Hoffmann B, Tucek Z, Toth P, Sonntag WE, and Ungvari Z (2015) Resveratrol encapsulated in novel fusogenic liposomes activates Nrf2 and attenuates oxidative stress in cerebrovascular endothelial cells from aged rats. *J Gerontol A Biol Sci Med Sci* **70**:303–313.
- Csiszár A, Tarantini S, Yabluchanskiy A, Balasubramanian P, Kiss T, Farkas E, Baur JA, and Ungvari Z (2019) Role of endothelial NAD⁺ deficiency in age-related vascular dysfunction. *Am J Physiol Heart Circ Physiol* **316**:H1253–H1266.
- Cummings JL, Morstorf T, and Zhong K (2014) Alzheimer's disease drug-development pipeline: few candidates, frequent failures. *Alzheimers Res Ther* **6**:37.
- Daneman R and Prat A (2015) The blood-brain barrier. *Cold Spring Harb Perspect Biol* **7**:a020412.
- de la Torre JC (2017) Are major dementias triggered by poor blood flow to the brain? theoretical considerations. *J Alzheimers Dis* **57**:353–371.
- Dhuria SV, Hanson LR, and Frey 2nd WH (2010) Intranasal delivery to the central nervous system: mechanisms and experimental considerations. *J Pharm Sci* **99**:1654–1673.
- Di Giovanna AP, Tibo A, Silvestri L, Mullenbroich MC, Costantini I, Allegra Mascaro AL, Sacconi L, Frascioni P, Pavone FS. (2018) Whole-brain vasculature reconstruction at the single capillary level. *Sci Rep* **8**:12573-018-30533-3.
- Di Marco LY, Farkas E, Martin C, Venneri A, and Frangi AF (2015) Is vasomotion in cerebral arteries impaired in Alzheimer's disease? *J Alzheimers Dis* **46**:35–53.
- Dordoe C, Chen K, Huang W, Chen J, Hu J, Wang X, and Lin L (2021) Roles of fibroblast growth factors and their therapeutic potential in treatment of ischemic stroke. *Front Pharmacol* **12**:671131.
- Du J, Yin G, Hu Y, Shi S, Jiang J, Song X, Zhang Z, Wei Z, Tang C, and Lyu H (2020) Coicis semen protects against focal cerebral ischemia-reperfusion injury by inhibiting oxidative stress and promoting angiogenesis via the TGFβ/ALK1/Smad1/5 signaling pathway. *Aging (Albany NY)* **13**:877–893.
- Dvorak HF, Brown LF, Detmar M, and Dvorak AM (1995) Vascular permeability factor/vascular endothelial growth factor, microvascular hyperpermeability, and angiogenesis. *Am J Pathol* **146**:1029–1039.
- Ekker MS, Boot EM, Singhal AB, Tan KS, Dobbie S, Tuladhar AM, and de Leeuw FE (2018) Epidemiology, aetiology, and management of ischaemic stroke in young adults. *Lancet Neurol* **17**:790–801.
- Evans MC, Couch Y, Sibson N, and Turner MR (2013) Inflammation and neurovascular changes in amyotrophic lateral sclerosis. *Mol Cell Neurosci* **53**:34–41.
- Fan M, Xu H, Wang L, Luo H, Zhu X, Cai P, Wei L, Lu L, Cao Y, Ye R, et al. (2016) Tissue Plasminogen Activator Neurotoxicity is Neutralized by Recombinant ADAMTS 13. *Sci Rep* **6**:25971.
- Farrer LA, Cupples LA, Haines JL, Hyman B, Kukull WA, Mayeux R, Myers RH, Pericak-Vance MA, Risch N, and van Duijn CM; APOE and Alzheimer Disease Meta Analysis Consortium (1997) Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. *JAMA* **278**:1349–1356.
- Ferrara N (2010) Binding to the extracellular matrix and proteolytic processing: two key mechanisms regulating vascular endothelial growth factor action. *Mol Biol Cell* **21**:687–690.
- Fitch MT, Doller C, Combs CK, Landreth GE, and Silver J (1999) Cellular and molecular mechanisms of glial scarring and progressive cavitation: in vivo and in vitro analysis of inflammation-induced secondary injury after CNS trauma. *J Neurosci* **19**:8182–8198.
- Flamme I, Frölich T, and Risau W (1997) Molecular mechanisms of vasculogenesis and embryonic angiogenesis. *J Cell Physiol* **173**:206–210.
- Foldvari M, Chen DW, Nafissi N, Calderon D, Narsineni L, and Rafiee A (2016) Non-viral gene therapy: Gains and challenges of non-invasive administration methods. *J Control Release* **240**:165–190.
- Folkman J and D'Amore PA (1996) Blood vessel formation: what is its molecular basis? *Cell* **87**:1153–1155.
- Foltynie T, Barker R, and Brayne C (2002) Vascular parkinsonism: a review of the precision and frequency of the diagnosis. *Neuroepidemiology* **21**:1–7.
- Freitas-Andrade M, Carmeliet P, Charlebois C, Stanimirovic DB, and Moreno MJ (2012) PlGF knockout delays brain vessel growth and maturation upon systemic hypoxic challenge. *J Cereb Blood Flow Metab* **32**:663–675.
- Gaal EI, Tammela T, Anisimov A, Marbacher S, Honkanen P, Zarkada G, Leppänen VM, Tatlisumak T, Hernesniemi J, Niemelä M, et al. (2013) Comparison of vascular growth factors in the murine brain reveals placenta growth factor as prime candidate for CNS revascularization. *Blood* **122**:658–665.
- Gallego I, Villate-Beitia I, Soto-Sánchez C, Menéndez M, Grijalvo S, Eritja R, Martínez-Navarrete G, Humphreys L, López-Méndez T, Puras G, et al. (2020) Brain angiogenesis induced by nonviral gene therapy with potential therapeutic benefits for central nervous system diseases. *Mol Pharm* **17**:1848–1858.

- Garbuzova-Davis S, Rodrigues MC, Hernandez-Ontiveros DG, Louis MK, Willing AE, Borlongan CV, and Sanberg PR (2011) Amyotrophic lateral sclerosis: a neurovascular disease. *Brain Res* **1398**:113–125.
- Garbuzova-Davis S, Saporta S, Haller E, Kolomey I, Bennett SP, Potter H, and Sanberg PR (2007) Evidence of compromised blood-spinal cord barrier in early and late symptomatic SOD1 mice modeling ALS. *PLoS One* **2**:e1205.
- Garbuzova-Davis S, Woods 3rd RL, Louis MK, Zesiewicz TA, Kuzmin-Nichols N, Sullivan KL, Miller AM, Hernandez-Ontiveros DG, and Sanberg PR (2010) Reduction of circulating endothelial cells in peripheral blood of ALS patients. *PLoS One* **5**:e10614.
- GBD 2015 Neurological Disorders Collaborator Group. (2017) Global, regional, and national burden of neurological disorders during 1990–2015: A systematic analysis for the global burden of disease study 2015. *Lancet Neurol* **16**:877–897.
- Giannoni P, Badaut J, Dargazani C, De Maudave AF, Klement W, Costalat V, and Marchi N (2018) The pericyte-glia interface at the blood-brain barrier. *Clin Sci (Lond)* **132**:361–374.
- Gomes AP, Price NL, Ling AJ, Moslehi JJ, Montgomery MK, Rajman L, White JP, Teodoro JS, Wrann CD, Hubbard BP, et al. (2013) Declining NAD(+) induces a pseudohypoxic state disrupting nuclear-mitochondrial communication during aging. *Cell* **155**:1624–1638.
- Gorelick PB (2004) Risk factors for vascular dementia and Alzheimer disease. *Stroke* **35**(11, Suppl 1):2620–2622.
- Gorski DH and Walsh K (2003) Control of vascular cell differentiation by homeobox transcription factors. *Trends Cardiovasc Med* **13**:213–220.
- Grammas P (2011) Neurovascular dysfunction, inflammation and endothelial activation: Implications for the pathogenesis of Alzheimer's disease. *J Neuroinflammation* **8**:26–2094-8-26.
- Green RC, Cupples LA, Go R, Benke KS, Edeki T, Griffith PA, Williams M, Hipps Y, Graff-Radford N, Bachman D, et al.; MIRAGE Study Group (2002) Risk of dementia among white and African American relatives of patients with Alzheimer disease. *JAMA* **287**:329–336.
- Greenberg DA (2015) Poststroke angiogenesis, pro: making the desert bloom. *Stroke* **46**:e101–e102.
- Gunsilius E, Petzer AL, Stockhammer G, Kähler CM, and Gastl G (2001) Serial measurement of vascular endothelial growth factor and transforming growth factor-beta1 in serum of patients with acute ischemic stroke. *Stroke* **32**:275–278.
- Gurruchaga H, Saenz del Burgo L, Ciriza J, Orive G, Hernández RM, and Pedraz JL (2015) Advances in cell encapsulation technology and its application in drug delivery. *Expert Opin Drug Deliv* **12**:1251–1267.
- Hao R, Sun B, Yang L, Ma C, and Li S (2020) RVG29-modified microRNA-loaded nanoparticles improve ischemic brain injury by nasal delivery. *Drug Deliv* **27**:772–781.
- Harper S (2014) Economic and social implications of aging societies. *Science* **346**:587–591.
- Harrigan MR (2003) Angiogenic factors in the central nervous system. *Neurosurgery* **53**:639–660, discussion 660–661.
- Hatakeyama M, Ninomiya I, and Kanazawa M (2020) Angiogenesis and neuronal remodeling after ischemic stroke. *Neural Regen Res* **15**:16–19.
- Haverkamp LJ, Appel V, and Appel SH (1995) Natural history of amyotrophic lateral sclerosis in a database population. Validation of a scoring system and a model for survival prediction. *Brain* **118**:707–719.
- Hebert LE, Bienias JL, Aggarwal NT, Wilson RS, Bennett DA, Shah RC, and Evans DA (2010) Change in risk of Alzheimer disease over time. *Neurology* **75**:786–791.
- Herrán E, Pérez-González R, Igartua M, Pedraz JL, Carro E, and Hernández RM (2013a) VEGF-releasing biodegradable nanospheres administered by craniotomy: a novel therapeutic approach in the APP/PS1 mouse model of Alzheimer's disease. *J Control Release* **170**:111–119.
- Herrán E, Ruiz-Ortega JA, Aristieta A, Igartua M, Requejo C, Lafuente JV, Ugedo L, Pedraz JL, and Hernández RM (2013b) In vivo administration of VEGF- and GDNF-releasing biodegradable polymeric microspheres in a severe lesion model of Parkinson's disease. *Eur J Pharm Biopharm* **85** (3 Pt B):1183–1190.
- Herzog B, Pellet-Many C, Britton G, Hartzoulakis B, and Zachary JC (2011) VEGF binding to NRP1 is essential for VEGF stimulation of endothelial cell migration, complex formation between NRP1 and VEGFR2, and signaling via FAK Tyr407 phosphorylation. *Mol Biol Cell* **22**:2766–2776.
- Hoeben A, Landuyt B, Highley MS, Wildiers H, Van Oosterom AT, and De Bruijn EA (2004) Vascular endothelial growth factor and angiogenesis. *Pharmacol Rev* **56**:549–580.
- Holtzman DM, Herz J, and Bu G (2012) Apolipoprotein E and apolipoprotein E receptors: normal biology and roles in Alzheimer disease. *Cold Spring Harb Perspect Med* **2**:a006312.
- Hou Y, Dan X, Babbar M, Wei Y, Hasselbalch SG, Croteau DL, and Bohr VA (2019) Ageing as a risk factor for neurodegenerative disease. *Nat Rev Neurol* **15**:565–581.
- Huang LK, Chao SP, Hu CJ. (2020) Clinical trials of new drugs for Alzheimer disease. *J Biomed Sci* **27**:18-019-0609-7.
- Huber JD, Egleton RD, and Davis TP (2001) Molecular physiology and pathophysiology of tight junctions in the blood-brain barrier. *Trends Neurosci* **24**:719–725.
- Iadecola C (2017) The neurovascular unit coming of age: A journey through neurovascular coupling in health and disease. *Neuron* **96**:17–42.
- Iadecola C (2013) The pathobiology of vascular dementia. *Neuron* **80**:844–866.
- Ikeda N, Nonoguchi N, Zhao MZ, Watanabe T, Kajimoto Y, Furutama D, Kimura F, Dezawa M, Coffin RS, Otsuki Y, et al. (2005) Bone marrow stromal cells that enhanced fibroblast growth factor-2 secretion by herpes simplex virus vector improve neurological outcome after transient focal cerebral ischemia in rats. *Stroke* **36**:2725–2730.
- Ingucsi S, Verlengia G, Soukupova M, Zucchini S, and Simonato M (2019) Gene therapy tools for brain diseases. *Front Pharmacol* **10**:724.
- IST-3 Collaborative Group. Sandercock P, Wardlaw JM, Lindley RI, Dennis M, Cohen G, Murray G, Innes K, Venables G, Czlonkowska A, Kobayashi A, et al. (2012) The benefits and harms of intravenous thrombolysis with recombinant tissue plasminogen activator within 6 h of acute ischaemic stroke (the third international stroke trial [IST-3]): A randomised controlled trial. *Lancet* **379**:2352–2363.
- Jayant RD, Sosa D, Kaushik A, Atluri V, Vashist A, Tomitaka A, and Nair M (2016) Current status of non-viral gene therapy for CNS disorders. *Expert Opin Drug Deliv* **13**:1433–1445.
- Jellinger KA (2003) Prevalence of cerebrovascular lesions in Parkinson's disease. A postmortem study. *Acta Neuropathol* **105**:415–419.
- Joly S, Dejda A, Rodriguez L, Sapiéha P, and Pernet V (2018) Nogo-A inhibits vascular regeneration in ischemic retinopathy. *Glia* **66**:2079–2093.
- Ju R, Wen Y, Gou R, Wang Y, and Xu Q (2014) The experimental therapy on brain ischemia by improvement of local angiogenesis with tissue engineering in the mouse. *Cell Transplant* **23** (Suppl 1):S83–S95.
- Jung S, Gilgen M, Slotboom J, El-Koussy M, Zubler C, Kiefer C, Luedi R, Mono ML, Heldner MR, Weck A, et al. (2013) Factors that determine penumbral tissue loss in acute ischaemic stroke. *Brain* **136**:3554–3560.
- Kaner RJ, Ladetto JV, Singh R, Fukuda N, Matthay MA, and Crystal RG (2000) Lung overexpression of the vascular endothelial growth factor gene induces pulmonary edema. *Am J Respir Cell Mol Biol* **22**:657–664.
- Kapasi A, DeCarli C, and Schneider JA (2017) Impact of multiple pathologies on the threshold for clinically overt dementia. *Acta Neuropathol* **134**:171–186.
- Kariyawasam D, Alexander IE, Kurian M, and Farrar MA (2020) Great expectations: virus-mediated gene therapy in neurological disorders. *J Neurol Neurosurg Psychiatry* **91**:849–860.
- Kaur J, Zhao Z, Klein GM, Lo EH, and Buchan AM (2004) The neurotoxicity of tissue plasminogen activator? *J Cereb Blood Flow Metab* **24**:945–963.
- Kiss T, Balasubramanian P, Valcarcel-Ares MN, Tarantini S, Yabluchanskiy A, Csipo T, Lipez A, Reglodi D, Zhang XA, Bari F, et al. (2019) Nicotinamide mononucleotide (NMN) treatment attenuates oxidative stress and rescues angiogenic capacity in aged cerebrovascular endothelial cells: a potential mechanism for the prevention of vascular cognitive impairment. *Geroscience* **41**:619–630.
- Kniessel U and Wolburg H (2000) Tight junctions of the blood-brain barrier. *Cell Mol Neurobiol* **20**:57–76.
- Korczyn AD (2015) Vascular parkinsonism—characteristics, pathogenesis and treatment. *Nat Rev Neurol* **11**:319–326.
- Korpisalo P and Ylä-Herttuala S (2010) Stimulation of functional vessel growth by gene therapy. *Integr Biol* **2**:102–112.
- Kiril JJ, Patel S, Harding AJ, and Halliday GM (2002) Patients with vascular dementia due to microvascular pathology have significant hippocampal neuronal loss. *J Neurol Neurosurg Psychiatry* **72**:747–751.
- Lähteenvuo J and Rosenzweig A (2012) Effects of aging on angiogenesis. *Circ Res* **110**:1252–1264.
- Lange C, Storkebaum E, de Almodóvar CR, Dewerchin M, and Carmeliet P (2016) Vascular endothelial growth factor: a neurovascular target in neurological diseases. *Nat Rev Neurol* **12**:439–454.
- Lautenschlager NT, Cupples LA, Rao VS, Auerbach SA, Becker R, Burke J, Chui H, Duara R, Foley EJ, Glatt SL, et al. (1996) Risk of dementia among relatives of Alzheimer's disease patients in the MIRAGE study: What is in store for the oldest old? *Neurology* **46**:641–650.
- Lawler PR and Lawler J (2012) Molecular basis for the regulation of angiogenesis by thrombospondin-1 and -2. *Cold Spring Harb Perspect Med* **2**:a006627.
- Lee HJ, Kim KS, Park IH, and Kim SU (2007) Human neural stem cells over-expressing VEGF provide neuroprotection, angiogenesis and functional recovery in mouse stroke model. *PLoS One* **2**:e156.
- Leenders KL, Perani D, Lammertsma AA, Heather JD, Buckingham P, Healy MJ, Gibbs JM, Wise RJ, Hatazawa J, Herold S, et al. (1990) Cerebral blood flow, blood volume and oxygen utilization. Normal values and effect of age. *Brain* **113**:27–47.
- Levi F, Chatenoud L, Bertuccio P, Lucchini F, Negri E, and La Vecchia C (2009) Mortality from cardiovascular and cerebrovascular diseases in Europe and other areas of the world: an update. *Eur J Cardiovasc Prev Rehabil* **16**:333–350.
- Li A, Gong H, Zhang B, Wang Q, Yan C, Wu J, Liu Q, Zeng S, and Luo Q (2010b) Micro-optical sectioning tomography to obtain a high-resolution atlas of the mouse brain. *Science* **330**:1404–1408.
- Li AD, Tong L, Xu N, Ye Y, Nie PY, Wang ZY, and Ji LL (2019a) miR-124 regulates cerebrovascular function in APP/PS1 transgenic mice via C1ql3. *Brain Res Bull* **153**:214–222.
- Li H, Xia N, Hasselwander S, and Daiber A (2019b) Resveratrol and vascular function. *Int J Mol Sci* **20**:E2155 <https://doi.org/10.3390/ijms20092155>.
- Li JS, Zhou YL, Liu K, Gao JF, Yang XK, Zhao YW, Liu ZG, and Liu JX (2010a) [Angiogenesis of brain after ischemia/reperfusion injury of brain in aged rats and changes in expressions of basic fibroblast growth factor and transformation growth factor-β1]. *Zhongguo Wei Zhong Bing Ji Jiu Yi Xue* **22**:583–586.
- Li Y, Wu J, Yu S, Zhu J, Zhou Y, Wang P, Li L, and Zhao Y (2020) Sestrin2 promotes angiogenesis to alleviate brain injury by activating Nrf2 through regulating the interaction between p62 and Keap1 following photothrombotic stroke in rats. *Brain Res* **1745**:146948.
- Liang C, Ni GX, Shi XL, Jia L, and Wang YL (2020) Astragaloside IV regulates the HIF/VEGF/Notch signaling pathway through miRNA-210 to promote angiogenesis after ischemic stroke. *Restor Neurol Neurosci* **38**:271–282.
- Liesz A (2019) The vascular side of Alzheimer's disease. *Science* **365**:223–224.
- Liu H, Honmou O, Harada K, Nakamura K, Houkin K, Hamada H, and Kocsis JD (2006) Neuroprotection by PlGF gene-modified human mesenchymal stem cells after cerebral ischaemia. *Brain* **129**:2734–2745.
- Liu J (2015) Poststroke angiogenesis: blood, bloom, or brood? *Stroke* **46**:e105–e106.
- Logsdon EA, Finley SD, Popel AS, and Mac Gabhann F (2014) A systems-biology view of blood vessel growth and remodelling. *J Cell Mol Med* **18**:1491–1508.

- Lombardo SM, Schneider M, Türelı AE, and Günday Türelı N (2020) Key for crossing the BBB with nanoparticles: the rational design. *Beilstein J Nanotechnol* **11**:866–883.
- Luissint AC, Artus C, Glacial F, Ganeshamoorthy K, and Couraud PO (2012a) Tight junctions at the blood brain barrier: Physiological architecture and disease-associated dysregulation. *Fluids Barriers CNS* **9**:23-8118-9-23.
- Luissint AC, Federici C, Guillonneau F, Chrétien F, Camoin L, Glacial F, Ganeshamoorthy K, and Couraud PO (2012b) Guanine nucleotide-binding protein Gzi2: a new partner of claudin-5 that regulates tight junction integrity in human brain endothelial cells. *J Cereb Blood Flow Metab* **32**:860–873.
- Lyons MK, Anderson RE, and Meyer FB (1991) Basic fibroblast growth factor promotes in vivo cerebral angiogenesis in chronic forebrain ischemia. *Brain Res* **558**:315–320.
- Ma Y, Zechariah A, Qu Y, and Hermann DM (2012) Effects of vascular endothelial growth factor in ischemic stroke. *J Neurosci Res* **90**:1873–1882.
- MacDonald TJ, Taga T, Shimada H, Tabrizi P, Zlokovic BV, Cheresh DA, and Laug WE (2001) Preferential susceptibility of brain tumors to the antiangiogenic effects of an alpha(v) integrin antagonist. *Neurosurgery* **48**:151–157.
- Mackenzie F and Ruhrberg C (2012) Diverse roles for VEGF-A in the nervous system. *Development* **139**:1371–1380.
- Madelaine R, Sloan SA, Huber N, Notwell JH, Leung LC, Skariah G, Halluin C, Paşca SP, Bejerano G, Krasnow MA, et al. (2017) MicroRNA-9 couples brain neurogenesis and angiogenesis. *Cell Rep* **20**:1533–1542.
- Mancuso MR, Kuhnert F, and Kuo CJ (2008) Developmental angiogenesis of the central nervous system. *Lymphat Res Biol* **6**:173–180.
- Manoonkitiwongsa PS, Schultz RL, McCreery DB, Whitter EF, and Lyden PD (2004) Neuroprotection of ischemic brain by vascular endothelial growth factor is critically dependent on proper dosage and may be compromised by angiogenesis. *J Cereb Blood Flow Metab* **24**:693–702.
- Manoonkitiwongsa PS, Schultz RL, Whitter EF, and Lyden PD (2006) Contraindications of VEGF-based therapeutic angiogenesis: effects on macrophage density and histology of normal and ischemic brains. *Vascul Pharmacol* **44**:316–325.
- Marin-Padilla M (1985) Early vascularization of the embryonic cerebral cortex: Golgi and electron microscopic studies. *J Comp Neurol* **241**:237–249.
- Martino MM, Brkic S, Bovo E, Burger M, Schaefer DJ, Wolff T, Gürke L, Briquez PS, Larsson HM, Gianni-Barrera R, et al. (2015) Extracellular matrix and growth factor engineering for controlled angiogenesis in regenerative medicine. *Front Bioeng Biotechnol* **3**:45.
- Mayeux R, Sano M, Chen J, Tatemichi T, and Stern Y (1991) Risk of dementia in first-degree relatives of patients with Alzheimer's disease and related disorders. *Arch Neurol* **48**:269–273.
- McConnell HL, Kersch CN, Woltjer RL, and Neuwelt EA (2017) The translational significance of the neurovascular unit. *J Biol Chem* **292**:762–770.
- Meng XY, Huang AQ, Khan A, Zhang L, Sun XQ, Song H, Han J, Sun QR, Wang YD, and Li XL (2020) Vascular endothelial growth factor-loaded poly-lactic-co-glycolic acid nanoparticles with controlled release protect the dopaminergic neurons in Parkinson's rats. *Chem Biol Drug Des* **95**:631–639.
- Michaelson DM (2014) APOE ϵ 4: the most prevalent yet understudied risk factor for Alzheimer's disease. *Alzheimers Dement* **10**:861–868.
- Mills KF, Yoshida S, Stein LR, Grozio A, Kubota S, Sasaki Y, Redpath P, Migaud ME, Apte RS, Uchida K, et al. (2016) Long-term administration of nicotinamide mononucleotide mitigates age-associated physiological decline in mice. *Cell Metab* **24**:795–806.
- Milner R and Campbell IL (2002) Developmental regulation of beta1 integrins during angiogenesis in the central nervous system. *Mol Cell Neurosci* **20**:616–626.
- Mingozzi F and High KA (2013) Immune responses to AAV vectors: overcoming barriers to successful gene therapy. *Blood* **122**:23–36.
- Moshayedi P, Nih LR, Llorente IL, Berg AR, Cinkornpumin J, Lowry WE, Segura T, and Carmichael ST (2016) Systematic optimization of an engineered hydrogel allows for selective control of human neural stem cell survival and differentiation after transplantation in the stroke brain. *Biomaterials* **105**:145–155.
- Mulvihill JJ, Cunnane EM, Ross AM, Duskey JT, Tosi G, and Grabrucker AM (2020) Drug delivery across the blood-brain barrier: recent advances in the use of nanocarriers. *Nanomedicine (Lond)* **15**:205–214.
- Nakamura K, Arimura K, Nishimura A, Tachibana M, Yoshikawa Y, Makihara N, Wakisaka Y, Kuroda J, Kamouchi M, Ooboshi H, et al. (2016) Possible involvement of basic FGF in the upregulation of PDGFR β in pericytes after ischemic stroke. *Brain Res* **1630**:98–108.
- Namman P, Razban V, Tabei SMB, and Asadi-Yousefabad SL (2020). MicroRNA-126: Dual Role in Angiogenesis Dependent Diseases. **163098–108** *Curr Pharm Des.*
- Nih LR, Carmichael ST, and Segura T (2016) Hydrogels for brain repair after stroke: an emerging treatment option. *Curr Opin Biotechnol* **40**:155–163.
- Nih LR, Gogini S, Carmichael ST, and Segura T (2018) Dual-function injectable angiogenic biomaterial for the repair of brain tissue following stroke. *Nat Mater* **17**:642–651.
- Niu X, Chen J, and Gao J (2019) Nanocarriers as a powerful vehicle to overcome blood-brain barrier in treating neurodegenerative diseases: Focus on recent advances. *Asian J Pharm Sci* **14**:480–496.
- O'Brien JT and Thomas A (2015) Vascular dementia. *Lancet* **386**:1698–1706.
- Oomen CA, Farkas E, Roman V, van der Beek EM, Luiten PG, and Meerlo P (2009) Resveratrol preserves cerebrovascular density and cognitive function in aging mice. *Front Aging Neurosci* **1**:4.
- Orive G, Hernández RM, Gascón AR, Calafiore R, Chang TM, De Vos P, Hortelano G, Hunkeler D, Lacić I, Shapiro AM, et al. (2003) Cell encapsulation: promise and progress. *Nat Med* **9**:104–107.
- Ozawa CR, Banfi A, Glazer NL, Thurston G, Springer ML, Kraft PE, McDonald DM, and Blau HM (2004) Microenvironmental VEGF concentration, not total dose, determines a threshold between normal and aberrant angiogenesis. *J Clin Invest* **113**:516–527.
- Patel NS, Mathura VS, Bachmeier C, Beaulieu-Abdelahad D, Laporte V, Weeks O, Mullan M, and Paris D (2010) Alzheimer's beta-amyloid peptide blocks vascular endothelial growth factor mediated signaling via direct interaction with VEGFR-2. *J Neurochem* **112**:66–76.
- Peddibhotla SS, Brinkmann BF, Kummer D, Tuncay H, Nakayama M, Adams RH, Gerke V, and Ebnat K (2013) Tetraspanin CD9 links junctional adhesion molecule-A to α v β 3 integrin to mediate basic fibroblast growth factor-specific angiogenic signaling. *Mol Biol Cell* **24**:933–944.
- Penkowa M, Carrasco J, Giralt M, Molinero A, Hernández J, Campbell IL, and Hidalgo J (2000) Altered central nervous system cytokine-growth factor expression profiles and angiogenesis in metallothionein-I+II deficient mice. *J Cereb Blood Flow Metab* **20**:1174–1189.
- Petersen KS, Rae S, Venos E, Malta D, Trieu K, Santos JA, Thout SR, Webster J, Campbell NRC, and Arcand J (2019) Paucity of high-quality studies reporting on salt and health outcomes from the science of salt: A regularly updated systematic review of salt and health outcomes (April 2017 to March 2018). *J Clin Hypertens (Greenwich)* **21**:307–323.
- Poovalah N, Davoudi Z, Peng H, Schlichtmann B, Mallapragada S, Narasimhan B, and Wang Q (2018) Treatment of neurodegenerative disorders through the blood-brain barrier using nanocarriers. *Nanoscale* **10**:16962–16983.
- Provost N, Le Meur G, Weber M, Mendes-Madeira A, Podevin G, Chel Y, Colle MA, Deschamps JY, Moullier P, and Rolling F (2005) Biodistribution of rAAV vectors following intraocular administration: evidence for the presence and persistence of vector DNA in the optic nerve and in the brain. *Mol Ther* **11**:275–283.
- Puumala M, Anderson RE, and Meyer FB (1990) Intraventricular infusion of HBGF-2 promotes cerebral angiogenesis in Wistar rat. *Brain Res* **534**:283–286.
- Qu M, Pan J, Wang L, Zhou P, Song Y, Wang S, Jiang L, Geng J, Zhang Z, Wang Y, et al. (2019) MicroRNA-126 regulates angiogenesis and neurogenesis in a mouse model of focal cerebral ischemia. *Mol Ther Nucleic Acids* **16**:15–25.
- Ren C, Li N, Gao C, Zhang W, Yang Y, Li S, Ji X, and Ding Y (2020) Ligustilide provides neuroprotection by promoting angiogenesis after cerebral ischemia. *Neurosci Res* **42**:683–692.
- Robertson PL, Du Bois M, Bowman PD, and Goldstein GW (1985) Angiogenesis in developing rat brain: an in vivo and in vitro study. *Brain Res* **355**:219–223.
- Román GC (2004) Brain hypoperfusion: a critical factor in vascular dementia. *Neurosci Res* **26**:454–458.
- Rosenstein JM, Krum JM, and Ruhrberg C (2010) VEGF in the nervous system. *Organogenesis* **6**:107–114.
- Roth AD, Ramirez G, Alarcón R, and Von Bernhardi R (2005) Oligodendrocytes damage in Alzheimer's disease: beta amyloid toxicity and inflammation. *Biol Res* **38**:381–387.
- Rowland LP and Shneider NA (2001) Amyotrophic lateral sclerosis. *N Engl J Med* **344**:1688–1700.
- Ruhrberg C, Gerhardt H, Golding M, Watson R, Ioannidou S, Fujisawa H, Betsholtz C, and Shima DT (2002) Spatially restricted patterning cues provided by heparin-binding VEGF-A control blood vessel branching morphogenesis. *Genes Dev* **16**:2684–2698.
- Rule RR, Schuff N, Miller RG, and Weiner MW (2010) Gray matter perfusion correlates with disease severity in ALS. *Neurology* **74**:821–827.
- Rust R, Gantner C, and Schwab ME (2019a) Pro- and antiangiogenic therapies: current status and clinical implications. *FASEB J* **33**:34–48.
- Rust R, Weber RZ, Gronert L, Mulders G, Maurer MA, Hofer AS, Sartori AM, and Schwab ME (2019b) Anti-nogo-A antibodies prevent vascular leakage and act as pro-angiogenic factors following stroke. *Sci Rep* **9**:20040-019-56634-1.
- Sánchez-Moreno C, Dashe JF, Scott T, Thaler D, Folstein MF, and Martin A (2004) Decreased levels of plasma vitamin C and increased concentrations of inflammatory and oxidative stress markers after stroke. *Stroke* **35**:163–168.
- Saver JL, Goyal M, Bonafe A, Diener HC, Levy EI, Pereira VM, Albers GW, Cognard C, Cohen DJ, Hacke W, et al.; SWIFT PRIME Investigators (2015) Stent-retriever thrombectomy after intravenous t-PA vs. t-PA alone in stroke. *N Engl J Med* **372**:2285–2295.
- Serlin Y, Shelef I, Knyazer B, and Friedman A (2015) Anatomy and physiology of the blood-brain barrier. *Semin Cell Dev Biol* **38**:2–6.
- Shen F, Fan Y, Su H, Zhu Y, Chen Y, Liu W, Young WL, and Yang GY (2008) Adeno-associated viral vector-mediated hypoxia-regulated VEGF gene transfer promotes angiogenesis following focal cerebral ischemia in mice. *Gene Ther* **15**:30–39.
- Shen F, Su H, Fan Y, Chen Y, Zhu Y, Liu W, Young WL, and Yang GY (2006) Adeno-associated viral-vector-mediated hypoxia-inducible vascular endothelial growth factor gene expression attenuates ischemic brain injury after focal cerebral ischemia in mice. *Stroke* **37**:2601–2606.
- Shen J, Ishii Y, Xu G, Dang TC, Hamashima T, Matsushima T, Yamamoto S, Hattori Y, Takatsuru Y, Nabekura J, et al. (2012) PDGFR- β as a positive regulator of tissue repair in a mouse model of focal cerebral ischemia. *J Cereb Blood Flow Metab* **32**:353–367.
- Shen J, Xu G, Zhu R, Yuan J, Ishii Y, Hamashima T, Matsushima T, Yamamoto S, Takatsuru Y, Nabekura J, et al. (2019) PDGFR- β restores blood-brain barrier functions in a mouse model of focal cerebral ischemia. *J Cereb Blood Flow Metab* **39**:1501–1515.
- Shim JW and Madsen JR (2018) VEGF signaling in neurological disorders. *Int J Mol Sci* **19**:275 <https://doi.org/10.3390/ijms19010275>.
- Silvestri L, Costantini I, Sacconi L, and Pavone FS (2016) Clearing of fixed tissue: a review from a microscopist's perspective. *J Biomed Opt* **21**:081205.
- Simons M, Gordon E, and Claesson-Welsh L (2016) Mechanisms and regulation of endothelial VEGF receptor signalling. *Nat Rev Mol Cell Biol* **17**:611–625.
- Spuch C, Antequera D, Portero A, Orive G, Hernández RM, Molina JA, Bermejo-Pareja F, Pedraz JL, and Carro E (2010) The effect of encapsulated VEGF-secreting cells on brain amyloid load and behavioral impairment in a mouse model of Alzheimer's disease. *Biomaterials* **31**:5608–5618.

- Suárez Y and Sessa WC (2009) MicroRNAs as novel regulators of angiogenesis. *Circ Res* **104**:442–454.
- Sun P, Zhang K, Hassan SH, Zhang X, Tang X, Pu H, Stetler RA, Chen J, and Yin KJ (2020) Endothelium-targeted deletion of microRNA-15a/16-1 promotes poststroke angiogenesis and improves long-term neurological recovery. *Circ Res* **126**:1040–1057.
- Sun Y, Jin K, Xie L, Childs J, Mao XO, Logvinova A, and Greenberg DA (2003) VEGF-induced neuroprotection, neurogenesis, and angiogenesis after focal cerebral ischemia. *J Clin Invest* **111**:1843–1851.
- Sweeney MD, Zhao Z, Montagne A, Nelson AR, and Zlokovic BV (2019) Blood-brain barrier: From physiology to disease and back. *Physiol Rev* **99**:21–78.
- Tafuro S, Ayuso E, Zacchigna S, Zentilin L, Moimas S, Dore F, and Giacca M (2009) Inducible adeno-associated virus vectors promote functional angiogenesis in adult organisms via regulated vascular endothelial growth factor expression. *Cardiovasc Res* **83**:663–671.
- Tarantini S, Tran CHT, Gordon GR, Ungvari Z, and Csiszar A (2017) Impaired neurovascular coupling in aging and Alzheimer's disease: Contribution of astrocyte dysfunction and endothelial impairment to cognitive decline. *Exp Gerontol* **94**:52–58.
- Tarantini S, Yabluchanskiy A, Csipo T, Fulop G, Kiss T, Balasubramanian P, DelFavero J, Ahire C, Ungvari A, Nyúl-Tóth A, et al. (2019) Treatment with the poly(ADP-ribose) polymerase inhibitor PJ-34 improves cerebrovascular endothelial function, neurovascular coupling responses and cognitive performance in aged mice, supporting the NAD⁺ depletion hypothesis of neurovascular aging. *Geroscience* **41**:533–542.
- Teixeira MI, Lopes CM, Amaral MH, and Costa PC (2020) Current insights on lipid nanocarrier-assisted drug delivery in the treatment of neurodegenerative diseases. *Eur J Pharm Biopharm* **149**:192–217.
- Teleanu DM, Negut I, Grumezescu V, Grumezescu AM, and Teleanu RI (2019) Nanomaterials for drug delivery to the central nervous system. *Nanomaterials (Basel)* **9**:E371 <https://doi.org/10.3390/nano9030371>.
- Thurgur H and Pinteaux E (2019) Microglia in the neurovascular unit: Blood-brain barrier-microglia interactions after central nervous system disorders. *Neuroscience* **405**:55–67.
- Tischer E, Mitchell R, Hartman T, Silva M, Gospodarowicz D, Fiddes JC, and Abraham JA (1991) The human gene for vascular endothelial growth factor. Multiple protein forms are encoded through alternative exon splicing. *J Biol Chem* **266**:11947–11954.
- Tornavaca O, Chia M, Dufton N, Almagro LO, Conway DE, Randi AM, Schwartz MA, Matter K, and Balda MS (2015) ZO-1 controls endothelial adherens junctions, cell-cell tension, angiogenesis, and barrier formation. *J Cell Biol* **208**:821–838.
- Toth P, Tarantini S, Tucek Z, Ashpole NM, Sosnowska D, Gautam T, Ballabh P, Koller A, Sonntag WE, Csiszar A, et al. (2014) Resveratrol treatment rescues neurovascular coupling in aged mice: role of improved cerebrovascular endothelial function and downregulation of NADPH oxidase. *Am J Physiol Heart Circ Physiol* **306**:H299–H308.
- Uccelli A, Wolff T, Valente P, Di Maggio N, Pellegrino M, Gürke L, Banfi A, and Gianni-Barrera R (2019) Vascular endothelial growth factor biology for regenerative angiogenesis. *Swiss Med Wkly* **149**:w20011.
- Uemura MT, Maki T, Ihara M, Lee VMY, and Trojanowski JQ (2020) Brain microvascular pericytes in vascular cognitive impairment and dementia. *Front Aging Neurosci* **12**:80.
- Ungvari Z, Tarantini S, Sorond F, Merkely B, and Csiszar A (2020) Mechanisms of vascular aging, a geroscience perspective: JACC focus seminar. *J Am Coll Cardiol* **75**:931–941.
- Ungvari Z, Tucek Z, Sosnowska D, Toth P, Gautam T, Podlutzky A, Csiszar A, Losonczy G, Valcarcel-Ares MN, Sonntag WE, et al. (2013) Aging-induced dysregulation of dicer1-dependent microRNA expression impairs angiogenic capacity of rat cerebrovascular endothelial cells. *J Gerontol A Biol Sci Med Sci* **68**:877–891.
- Valable S, Montaner J, Bellail A, Berezowski V, Brillault J, Cecchelli R, Divoux D, Mackenzie ET, Bernaudin M, Roussel S, et al. (2005) VEGF-induced BBB permeability is associated with an MMP-9 activity increase in cerebral ischemia: both effects decreased by Ang-1. *J Cereb Blood Flow Metab* **25**:1491–1504.
- Vale TC, Barbosa MT, Caramelli P, and Cardoso F (2012) Vascular Parkinsonism and cognitive impairment: literature review, Brazilian studies and case vignettes. *Dement Neuropsychol* **6**:137–144.
- Vallon M, Chang J, Zhang H, and Kuo CJ (2014) Developmental and pathological angiogenesis in the central nervous system. *Cell Mol Life Sci* **71**:3489–3506.
- van Es MA, Hardiman O, Chio A, Al-Chalabi A, Pasterkamp RJ, Veldink JH, and van den Berg LH (2017) Amyotrophic lateral sclerosis. *Lancet* **390**:2084–2098.
- Vangilder RL, Rosen CL, Barr TL, and Huber JD (2011) Targeting the neurovascular unit for treatment of neurological disorders. *Pharmacol Ther* **130**:239–247.
- Verheggen ICM, de Jong JJA, van Boxtel MPJ, Gronenschild EHB, Palm WM, Postma AA, Jansen JFA, Verhey FRJ, and Backes WH (2020) Increase in blood-brain barrier leakage in healthy, older adults. *Geroscience* **42**:1183–1193.
- von Degenfeld G, Banfi A, Springer ML, Wagner RA, Jacobi J, Ozawa CR, Merchant MJ, Cooke JP, and Blau HM (2006) Microenvironmental VEGF distribution is critical for stable and functional vessel growth in ischemia. *FASEB J* **20**:2657–2659.
- Vorbrodt AW and Dobrogowska DH (2003) Molecular anatomy of intercellular junctions in brain endothelial and epithelial barriers: electron microscopist's view. *Brain Res Brain Res Rev* **42**:221–242.
- Wälchli T, Pernet V, Weimann O, Shiu JY, Guzik-Kornacka A, Decrey G, Yüksel D, Schneider H, Vogel J, Ingber DE, et al. (2013) Nogo-A is a negative regulator of CNS angiogenesis. *Proc Natl Acad Sci USA* **110**:E1943–E1952.
- Waldemar G, Vorstrup S, Jensen TS, Johnsen A, and Boysen G (1992) Focal reductions of cerebral blood flow in amyotrophic lateral sclerosis: a [99mTc]-l-HMPAO SPECT study. *J Neurol Sci* **107**:19–28.
- Wang F, Nojima M, Inoue Y, Ohtomo K, and Kiryu S (2015) Assessment of MRI contrast agent kinetics via retro-orbital injection in mice: Comparison with tail vein injection. *PLoS One* **10**:e0129326.
- Wang P, Pan R, Weaver J, Jia M, Yang X, Yang T, Liang J, Liu KJ. (2020) MicroRNA-30a regulates acute cerebral ischemia-induced blood-brain barrier damage through ZnT4/zinc pathway. *J Cereb Blood Flow Metab*: 271678X/20926787.
- Wang P, Xie ZH, Guo YJ, Zhao CP, Jiang H, Song Y, Zhu ZY, Lai C, Xu SL, and Bi JZ (2011) VEGF-induced angiogenesis ameliorates the memory impairment in APP transgenic mouse model of Alzheimer's disease. *Biochem Biophys Res Commun* **411**:620–626.
- Wang Z, Xiong G, Tsang WC, Schätzlein AG, and Uchegbu IF (2019) Nose-to-Brain Delivery. *J Pharmacol Exp Ther* **370**:593–601.
- Watanabe T, Okuda Y, Nonoguchi N, Zhao MZ, Kajimoto Y, Furutama D, Yukawa H, Shibata MA, Otsuki Y, Kuroiwa T, et al. (2004) Postischemic intraventricular administration of FGF-2 expressing adenoviral vectors improves neurologic outcome and reduces infarct volume after transient focal cerebral ischemia in rats. *J Cereb Blood Flow Metab* **24**:1205–1213.
- Wiedenhoeft T, Tarantini S, Nyúl-Tóth A, Yabluchanskiy A, Csipo T, Balasubramanian P, Lipez A, Kiss T, Csiszar A, Csiszar A, et al. (2019) Fusogenic liposomes effectively deliver resveratrol to the cerebral microcirculation and improve endothelium-dependent neurovascular coupling responses in aged mice. *Geroscience* **41**:711–725.
- Wimo A, Handels R, Winblad B, Black CM, Johansson G, Salomonsson S, Eriksdotter M, and Khandker RK (2020) Quantifying and Describing the Natural History and Costs of Alzheimer's disease and Effects of Hypothetical Interventions. *J Alzheimers Dis* **75**:891–902.
- Wong HL, Wu XY, and Bendayan R (2012) Nanotechnological advances for the delivery of CNS therapeutics. *Adv Drug Deliv Rev* **64**:686–700.
- Wu Z, Guo H, Chow N, Sallstrom J, Bell RD, Deane R, Brooks AI, Kanagala S, Rubio A, Sagare A, et al. (2005) Role of the MEOX2 homeobox gene in neurovascular dysfunction in Alzheimer disease. *Nat Med* **11**:959–965.
- Yang JP, Liu HJ, Cheng SM, Wang ZL, Cheng X, Yu HX, and Liu XF (2009b) Direct transport of VEGF from the nasal cavity to brain. *Neurosci Lett* **449**:108–111.
- Yang JP, Liu HJ, Wang ZL, Cheng SM, Cheng X, Xu GL, and Liu XF (2009a) The dose-effectiveness of intranasal VEGF in treatment of experimental stroke. *Neurosci Lett* **461**:212–216.
- Yang SP, Bae DG, Kang HJ, Gwag BJ, Gho YS, and Chae CB (2004) Co-accumulation of vascular endothelial growth factor with beta-amyloid in the brain of patients with Alzheimer's disease. *Neurobiol Aging* **25**:283–290.
- Yano A, Shingo T, Takeuchi A, Yasuhara T, Kobayashi K, Takahashi K, Muraoka K, Matsui T, Miyoshi Y, Hamada H, et al. (2005) Encapsulated vascular endothelial growth factor-secreting cell grafts have neuroprotective and angiogenic effects on focal cerebral ischemia. *J Neurosurg* **103**:104–114.
- Yardeni T, Eckhaus M, Morris HD, Huizing M, and Hoogstraten-Miller S (2011) Retrol-oral injections in mice. *Lab Anim (NY)* **40**:155–160.
- Yin KJ, Olsen K, Hamblin M, Zhang J, Schwendeman SP, and Chen YE (2012) Vascular endothelial cell-specific microRNA-15a inhibits angiogenesis in hindlimb ischemia. *J Biol Chem* **287**:27055–27064.
- Yokoyama T, Date C, Kokubo Y, Yoshiike N, Matsumura Y, and Tanaka H (2000) Serum vitamin C concentration was inversely associated with subsequent 20-year incidence of stroke in a Japanese rural community. The Shibata study. *Stroke* **31**:2287–2294.
- Zeng L, He X, Wang Y, Tang Y, Zheng C, Cai H, Liu J, Wang Y, Fu Y, and Yang GY (2014) MicroRNA-210 overexpression induces angiogenesis and neurogenesis in the normal adult mouse brain. *Gene Ther* **21**:37–43.
- Zeng L, Liu J, Wang Y, Wang L, Weng S, Tang Y, Zheng C, Cheng Q, Chen S, and Yang GY (2011) MicroRNA-210 as a novel blood biomarker in acute cerebral ischemia. *Front Biosci (Elite Ed)* **3**:1265–1272.
- Zeng LL, He XS, Liu JR, Zheng CB, Wang YT, and Yang GY (2016) Lentivirus-mediated overexpression of MicroRNA-210 improves long-term outcomes after focal cerebral ischemia in mice. *CNS Neurosci Ther* **22**:961–969.
- Zhang J, Tokatlian T, Zhong J, Ng QK, Patterson M, Lowry WE, Carmichael ST, and Segura T (2011) Physically associated synthetic hydrogels with long-term covalent stabilization for cell culture and stem cell transplantation. *Adv Mater* **23**:5098–5103.
- Zhang X, Yin X, Zhang J, Li A, Gong H, Luo Q, Zhang H, Gao Z, and Jiang H (2019) High-resolution mapping of brain vasculature and its impairment in the hippocampus of Alzheimer's disease mice. *Natl Sci Rev* **6**:1223–1238.
- Zhang ZG, Zhang L, Jiang Q, Zhang R, Davies K, Powers C, Bruggen Nv, and Chopp M (2000) VEGF enhances angiogenesis and promotes blood-brain barrier leakage in the ischemic brain. *J Clin Invest* **106**:829–833.
- Zhang ZG, Zhang L, Tsang W, Soltanian-Zadeh H, Morris D, Zhang R, Goussev A, Powers C, Yeich T, and Chopp M (2002) Correlation of VEGF and angiopoietin expression with disruption of blood-brain barrier and angiogenesis after focal cerebral ischemia. *J Cereb Blood Flow Metab* **22**:379–392.
- Zhong J, Chan A, Morad L, Kornblum HI, Fan G, and Carmichael ST (2010) Hydrogel matrix to support stem cell survival after brain transplantation in stroke. *Neurorehabil Neural Repair* **24**:636–644.
- Zhong Z, Deane R, Ali Z, Parisi M, Shapovalov Y, O'Banion MK, Stojanovic K, Sagare A, Boillee S, Cleveland DW, et al. (2008) ALS-causing SOD1 mutants generate vascular changes prior to motor neuron degeneration. *Nat Neurosci* **11**:420–422.
- Zhu Y, Zhang H, Zhang Y, Wu H, Wei L, Zhou G, Zhang Y, Deng L, Cheng Y, Li M, et al. (2019) Endovascular metal devices for the treatment of cerebrovascular diseases. *Adv Mater* **31**:e1805452.
- Zlokovic BV (2005) Neurovascular mechanisms of Alzheimer's neurodegeneration. *Trends Neurosci* **28**:202–208.
- Zou Q, Wu CW, Stein EA, Zang Y, and Yang Y (2009) Static and dynamic characteristics of cerebral blood flow during the resting state. *Neuroimage* **48**:515–524.