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Smith, H. G. P., Bean, D. C., Pitchers, W., Valcanis, M., Clarke, R. H., Loyn, R., Hassell, C. J., & Greenhill, A. R. (2021). Draft Genome Sequences of Four Isolates Recovered from Wild Australian Shorebirds. Microbiology Resource Announcements, 10(1).

Available online at: https://doi.org/10.1128/MRA.01113-20

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Draft Genome Sequences of Four *Citrobacter* Isolates Recovered from Wild Australian Shorebirds

Microbiology[®]

Resource Announcements

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ABSTRACT *Citrobacter* is a ubiquitous bacterial genus whose members inhabit a variety of niches. Some species are clinically important for both antimicrobial resistance (AMR) carriage and as the cause of nosocomial infections. Surveillance of *Citrobacter* species in the environment can provide indicators of the spread of AMR genes outside clinical spaces. In this study, we present draft genome sequences of four *Citrobacter* isolates obtained from three species of wild Australian shorebirds.

The genus *Citrobacter* comprises 11 species. They occupy a broad range of habitats, play a key role in the nitrogen cycle, and are frequently found in food and in the gut of animals, including humans (1). *Citrobacter* is an opportunistic pathogen of humans, most commonly associated with infant meningitis, urinary tract infections, sepsis, and pneumonia (2). The species most commonly isolated from clinical specimens are *C. koseri, C. freundii, C. youngae, C. braakii*, and *C. amalonaticus* (3). *Citrobacter* species can act as reservoirs for antimicrobial resistance (AMR) genes and can transfer these genes to other pathogenic bacteria (4, 5). *Citrobacter* spp. have been isolated from both healthy (6) and diseased birds (7).

We present here draft genomes of four *Citrobacter* isolates collected from Australian shorebirds through 2017 to 2018.

Four *Citrobacter* isolates were recovered from cloacal swabs collected from healthy Australian shorebirds (Table 1). The cloacal swabs were preenriched by incubating overnight in brain heart infusion broth at 35°C, followed by a secondary enrichment by transferring 100 μ l into Mannitol broth and again incubating overnight at 35°C. The broths were subsequently subcultured onto MacConkey II agar (Oxoid) and incubated overnight at 35°C.

Phenotypic testing of antimicrobial resistance was conducted using the disk diffusion method (8). For DNA extraction, organisms were grown overnight on nutrient agar. Genomic DNA (gDNA) was extracted using a Qiagen DNeasy kit and quantified using an Invitrogen Qubit 2 fluorometer. Sequencing was conducted at the Australian Genome Research Facility using the Illumina MiSeq platform, with Illumina gDNA shotgun library preparation with the bead size selection protocol generating 150-bp pairedend reads.

The raw reads were uploaded to the Galaxy Web platform, and the data were analyzed via the public server at usegalaxy.org, version 20.01 (9). The genomes were Citation Smith HGP, Bean DC, Pitchers W, Valcanis M, Clarke RH, Loyn R, Hassell CJ, Greenhill AR. 2021. Draft genome sequences of four *Citrobacter* isolates recovered from wild Australian shorebirds. Microbiol Resour Announc 10:e01113-20. https://doi.org/10 .1128/MRA.01113-20.

Editor Catherine Putonti, Loyola University Chicago

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Received 24 September 2020 Accepted 7 December 2020 Published 7 January 2021

	Data for strain:			
Characteristic	966a	1120a	1241a	1273b
Species	Citrobacter amalonaticus	Citrobacter braakii	Citrobacter freundii	Citrobacter freundii
Sampling location	38.324105 S, 145.517553 E	38.003826 S, 144.596880 E	17.979327 S, 122.336533 E	17.979327 S, 122.336533 E
Host	Double-banded plover (Charadrius bicinctus)	Curlew sandpiper (Calidris ferruginea)	Bar-tailed godwit (<i>Limosa lapponica</i>)	Bar-tailed godwit (<i>Limosa lapponica</i>)
Yr of isolation	2017	2017	2018	2018
Phenotypic resistance	Amoxicillin, ampicillin	Amoxicillin, ampicillin	Amoxicillin	Amoxicillin, ampicillin
AMR ^a gene	None	bla _{CMY-48}	bla _{CMY-48}	bla _{CMY-48}
Total no. of raw paired-end reads	753,330	617,160	728,982	493,188
No. of contigs	91	34	48	54
Total length (bp)	4,903,911	5,098,689	5,280,444	5,279,325
N ₅₀ length (bp)	225,898	851,789	571,851	571,851
Avg depth (\times)	71.7	71.7	71.7	68.5
GC content (%)	53.35	51.49	51.35	54
BioSample accession no.	SAMN13884683	SAMN13884692	SAMN13884697	SAMN13884710
Assembly accession no.	GCF_014333035.1	GCF_014332945.1	GCF_014332855.1	GCF_014332835.1

^a AMR, antimicrobial resistance.

assembled *de novo*, read quality control was performed using Unicycler version 0.4.8.0 (10), and the genome assembly quality was analyzed using QUAST version 5.0.2+galaxy1 (11). The genomes were uploaded to NCBI and annotated using PGAP version 4.12 (12). Further information on the genome parameters is given in Table 1.

ResFinder version 3.2 (13), hosted by the Centre for Genomic Epidemiology (http:// www.genomicepidemiology.org/), was used to identify AMR genes. The AMR gene identified can be seen in Table 1. This gene is thought to have originated in *C. freundii* (14).

Data availability. The whole-genome sequences, assemblies, and raw reads for this project have been deposited in GenBank under the BioProject accession number PRJNA602163. Raw reads are available in the SRA under the accession numbers SRR11613026 (*C. braakii*), SRR11613020 (*C. freundii*), SRR11613006 (*C. freundii*), and SRR11612996 (*C. amalonaticus*).

ACKNOWLEDGMENTS

Funding for this work was provided by the Holsworth Wildlife Research Endowment (grant HLS-18-005) and the Stuart Leslie Bird Research Award provided by Birdlife Australia (grant FOST-18-416). We thank GFN's major funders, the Netherlands Organisation Spinoza Premium Prize for Scientific Research to Theunis Piersma (2014 to 2017), WWF Netherlands (2010 to 2018), and MAVA, Fondation pour la Nature (2018).

We thank the Australasian Wader Study Group (AWSG), Rosalind Jessop, and the late Clive Minton for their help with this project. We acknowledge the Yawuru People, via the offices of Nyamba Buru Yawuru Limited, for permissions granted to AWSG to capture birds on the shores of Roebuck Bay, the traditional lands of the Yawuru people.

REFERENCES

- Lavigne J-P, Defez C, Bouziges N, Mahamat A, Sotto A. 2007. Clinical and molecular epidemiology of multidrug-resistant *Citrobacter* spp. infections in a French university hospital. Eur J Clin Microbiol Infect Dis 26:439–441. https://doi.org/10.1007/s10096-007-0315-3.
- Jorgensen JH, Pfaller MA, Carroll KC, Funke G, Landry ML, Richter SS, Warnock DW (ed). 2015. Manual of clinical microbiology, 11th ed. ASM Press, Washington, DC.
- Samonis G, Karageorgopoulos DE, Kofteridis DP, Matthaiou DK, Sidiropoulou V, Maraki S, Falagas ME. 2009. *Citrobacter* infections in a general hospital: characteristics and outcomes. Eur J Clin Microbiol Infect Dis 28:61–68. https://doi.org/10.1007/s10096-008-0598-z.
- Nawaz M, Khan AA, Khan S, Sung K, Steele R. 2008. Isolation and characterization of tetracycline-resistant *Citrobacter* spp. from catfish. Food Microbiol 25:85–91. https://doi.org/10.1016/j.fm.2007.07.008.
- Jacoby GA, Griffin CM, Hooper DC. 2011. *Citrobacter* spp. as a source of *qnrB* alleles. Antimicrob Agents Chemother 55:4979–4984. https://doi .org/10.1128/AAC.05187-11.
- Giacopello C, Foti M, Mascetti A, Grosso F, Ricciardi D, Fisichella V, Lo Piccolo F. 2016. Antimicrobial resistance patterns of *Enterobacteriaceae* in European wild bird species admitted in a wildlife rescue centre. Vet Ital 52:139–144. https://doi.org/10.12834/Vetlt.327.1374.2.
- Churria CDG, Arias N, Origlia J, Netri C, Marcantoni H, Píscopo M, Loyola MH, Petruccelli M. 2014. *Citrobacter freundii* infection in two captive Australian king parrots (*Alisterus scapularis*). J Zoo Aquar Res 2:52–53. https:// doi.org/10.19227/jzar.v2i2.80.
- CLSI. 2018. Performance standards for antimicrobial susceptibility testing: 22nd informational supplement, 28th ed. CLSI supplement M100. Clinical and Laboratory Standards Institute, Wayne, PA.

- Afgan E, Baker D, Batut B, van den Beek M, Bouvier D, Cech M, Chilton J, Clements D, Coraor N, Grüning BA, Guerler A, Hillman-Jackson J, Hiltemann S, Jalili V, Rasche H, Soranzo N, Goecks J, Taylor J, Nekrutenko A, Blankenberg D. 2018. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2018 update. Nucleic Acids Res 46: W537–W544. https://doi.org/10.1093/nar/gky379.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Mikheenko A, Prjibelski A, Saveliev V, Antipov D, Gurevich A. 2018. Versatile genome assembly evaluation with QUAST-LG. Bioinformatics 34: i142–i150. https://doi.org/10.1093/bioinformatics/bty266.
- 12. Haft DH, DiCuccio M, Badretdin A, Brover V, Chetvernin V, O'Neill K, Li W,

Chitsaz F, Derbyshire MK, Gonzales NR, Gwadz M, Lu F, Marchler GH, Song JS, Thanki N, Yamashita RA, Zheng C, Thibaud-Nissen F, Geer LY, Marchler-Bauer A, Pruitt KD. 2018. RefSeq: an update on prokaryotic genome annotation and curation. Nucleic Acids Res 46:D851–D860. https://doi.org/10.1093/nar/gkx1068.

- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640–2644. https://doi.org/ 10.1093/jac/dks261.
- Verdet C, Gautier V, Chachaty E, Ronco E, Hidri N, Decre D, Arlet G. 2009. Genetic context of plasmid-carried *bla_{CMY-2}*-like genes in Enterobacteriaceae. Antimicrob Agents Chemother 53:4002–4006. https://doi.org/10 .1128/AAC.00753-08.