

## Federation University ResearchOnline

<https://researchonline.federation.edu.au>

Copyright Notice

This is the peer reviewed version of the following article:

Benbow, Barton, P. S., Ulyshen, M. D., Beasley, J. C., DeVault, T. L., Strickland, M. S., Tomberlin, J. K., Jordan, H. R., & Pechal, J. L. (2019). Necrobiome framework for bridging decomposition ecology of autotrophically and heterotrophically derived organic matter. *Ecological Monographs*, 89(1), 1–29.

Which has been published in final form at:

<https://doi.org/10.1002/ecm.1331>

This article may be used for non-commercial purposes in accordance with [Wiley Terms and Conditions for use of Self-Archived Versions](#).

See this record in Federation ResearchOnline at:

<http://researchonline.federation.edu.au/vital/access/HandleResolver/1959.17/182679>

# Author Manuscript

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record](#). Please cite this article as [doi: 10.1002/ecm.1331](https://doi.org/10.1002/ecm.1331)

This article is protected by copyright. All rights reserved

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23

**Article Type: Concepts & Synthesis**

**Necrobiome Framework for Bridging Decomposition Ecology of Autotrophically- and Heterotrophically-Derived Organic Matter**

Running Title: The Necrobiome in Decomposition Ecology

M. Eric Benbow<sup>1,2,3\*</sup>, Philip S. Barton<sup>4</sup>, Michael D. Ulyshen<sup>5</sup>, James C. Beasley<sup>6</sup>, Travis L. DeVault<sup>7</sup>, Michael S. Strickland<sup>8</sup>, Jeffery K. Tomberlin<sup>9</sup>, Heather R. Jordan<sup>10</sup>, Jennifer L. Pechal<sup>1</sup>

<sup>1</sup>Department of Entomology, Michigan State University, East Lansing, Michigan, 48824, USA.

<sup>2</sup>Department of Osteopathic Medical Specialties, Michigan State University, East Lansing, Michigan, 48824, USA.

<sup>3</sup>Ecology, Evolutionary Biology and Behavior Program, Michigan State University, East Lansing, Michigan, 48824, USA.

<sup>4</sup>Fenner School of Environment and Society, Australian National University, Canberra, Australian Capital Territory, 2601, Australia.

<sup>5</sup>USDA Forest Service, Southern Research Station, Athens, GA, 30602, USA

<sup>6</sup>University of Georgia, Savannah River Ecology Laboratory and Warnell School of Forestry and Natural Resources, Aiken, South Carolina, 29802, USA

<sup>7</sup>U.S. Department of Agriculture, National Wildlife Research Center, Sandusky, Ohio, 44870, USA

24 <sup>8</sup>Department of Soil & Water Systems, University of Idaho, Moscow, Idaho, 83844, USA

25 <sup>9</sup>Department of Entomology, Texas A&M University, College Station Texas, 77843, USA

26 <sup>10</sup>Department of Biological Sciences, Mississippi State University, Mississippi, 39762, USA

27 <sup>1</sup>Department of Entomology, Michigan State University, East Lansing, Michigan, 48824, USA.

28

29 \*Corresponding author: [benbow@msu.edu](mailto:benbow@msu.edu), T: 517-410-9247, [orcid.org/0000-0003-2630-0282](https://orcid.org/0000-0003-2630-0282).

30

31

32 Manuscript type: Concepts & Synthesis

33 Manuscript received 2 March 2018; revised 11 July 2018; accepted 17 July 2018.

34 Corresponding Editor: Jay A. Rosenheim

35

36 **ABSTRACT**

37 Decomposition contributes to global ecosystem function by contributing to nutrient recycling,  
38 energy flow and limiting biomass accumulation. The decomposer organisms influencing this  
39 process form diverse, complex, and highly dynamic communities that often specialize on  
40 different plant or animal resources. Despite performing the same net role, there is a need to  
41 conceptually synthesize information on the structure and function of decomposer communities  
42 across the spectrum of dead plant and animal resources. A lack of synthesis has limited cross-  
43 disciplinary learning and research in important areas of ecosystem and community ecology. Here  
44 we expound on the ‘necrobiome’ concept and develop a framework describing the decomposer  
45 communities and their interactions associated with plant and animal resource types within  
46 multiple ecosystems. We outline the biotic structure and ecological functions of the necrobiome,  
47 along with how the necrobiome fits into a broader landscape and ecosystem context. The  
48 expanded necrobiome model provides a set of perspectives on decomposer communities across  
49 resource types, and conceptually unifies plant and animal decomposer communities into the  
50 same framework, while acknowledging key differences in processes and mechanisms. This  
51 framework is intended to raise awareness among researchers, and advance the construction of  
52 explicit, mechanistic hypotheses that further our understanding of decomposer community  
53 contributions to biodiversity, the structure and function of ecosystems and global nutrient  
54 recycling and energy flow.

55

56 Key words: biodiversity; carrion; detritus; decomposition; ecosystem; litter; community  
57 assembly; interactions; microbial ecology; nutrient cycling; organic matter

58

59 **INTRODUCTION**

60

61 Decomposer organisms are highly efficient at consuming and transforming dead organic matter  
62 and they play a critical role in shaping ecosystem structure and function. The impact of  
63 decomposition on ecosystems is demonstrated by the relatively small accumulation of the  
64 estimated 150-200 g of carbon/m<sup>2</sup>/yr (Fierer et al. 2009) generated by plant net productivity,  
65 especially considering that 90% of plant biomass is not consumed by herbivores (Cebrian 1999,  
66 Gessner et al. 2010). Early research demonstrated the importance of detritus in communities and  
67 energy flow of both terrestrial and aquatic ecosystems (e.g., Forbes 1887, Summerhayes and  
68 Elton 1923, Lindeman 1942, Teal 1962, Odum 1969, Swift et al. 1979, Wallace et al. 1999). The  
69 detrital pool has significant impact on the structure and function of ecosystems by increasing  
70 system stability and persistence, and the fate of such 'dead plant', 'decaying matter', 'dung' or  
71 'litter' has been described for many ecosystems (Moore et al 2004). Dead plant biomass, defined  
72 here as autotrophically-derived decomposing organic matter, is not the only form of detritus that  
73 is recycled with important ecosystem function. Another often overlooked decomposition  
74 component in ecosystems is carrion, defined here as heterotrophically-derived decomposing  
75 organic matter, or the carcasses of animals and other organisms that do not harvest energy from  
76 the sun but still contribute to the global detritus budget. These two forms of decomposing  
77 biomass may also include transitional forms of decaying organic matter, such as dung or frass,  
78 excreted organic molecules and dissolved and particulate forms that reflect the once living, and  
79 are the partially digested, exuded and egested biomass of plants and animals.

80

81 Dead organic matter of any type is defined as detritus (Petersen et al, 1918; Odum et al, 1963)  
82 but has been primarily perceived in the literature as organic matter resulting from plant death,  
83 with some, but few, instances of dead animal organic matter (or carrion) referred to as detritus  
84 (e.g., Swift et al 1979, Moore et al 2004). In the current literature, the majority of references to  
85 detritus describe decomposing plant biomass, reinforcing a separation with other widely  
86 ubiquitous forms of biomass with relatively higher turnover, such as carrion and dung/feces.  
87 With this understanding we propose to facilitate broader recognition of the commonalities and  
88 differences of dead organic matter of any type, defined in this paper as necromass, a term  
89 previously used in the literature (e.g., Coûteaux et al. 2005, Lomstein et al. 2012), but  
90 understood to be synonymous with the holistic definition of detritus as summarized by Moore et  
91 al. (2004).  
92  
93 Necromass takes a wide range of shapes, sizes and quality (Swift et al. 1979, Carter et al. 2007).  
94 It forms the basis of many foodwebs and consequently has significant bottom-up importance for  
95 ecosystem function (Gessner et al. 2010). It also forms the focal point for interactions and  
96 behaviors among decomposer species comprising complex communities influencing ecosystems  
97 across temporal and spatial scales (Yang et al. 2010); and in freshwater habitats, decomposition  
98 is considered a fundamental ecosystem process that contributes to watershed biodiversity  
99 (Woodward et al. 2012). Despite the importance of decomposer communities there exists no  
100 standard framework to conceptualize their complex and dynamic interactions across both plant  
101 and animal necromass. This lack of a framework is significant because it limits a comprehensive  
102 understanding of the community ecology of decomposition and has implications for defining and

103 testing paradigms related to nutrient recycling, gene flow, population dynamics, and other  
104 ecosystem processes at the frontier of ecological research.

105  
106 There has been a recent expansion of knowledge about both micro- and macro-scale processes  
107 structuring decomposer communities and contributing to the species interactions (e.g., Burkepile  
108 et al. 2006, Gessner et al. 2010, Wilson and Wolkovich 2011, Metcalf et al. 2016); however,  
109 much is still unknown. To date, ecological theory describing decomposition processes has  
110 included succession theory (Payne et al. 1968, Michaud et al. 2015), meta-population theory  
111 (Hanski 1987), and aggregation and coexistence theory (Ives 1991). Carrion and dung, in  
112 particular, have been useful systems to test and develop these theories and mechanistic  
113 hypotheses having relevance to many other areas of ecology (see Box 1). However, a new  
114 synthesis is required to integrate existing knowledge with the new discoveries occurring in the  
115 animal and plant decomposition disciplines. We propose an expansion of a recent framework to  
116 facilitate this synthesis in a way that encompasses all forms, functions, and ecological dynamics  
117 of organic matter decomposition – the necrobiome (Figure 1).

118  
119 The necrobiome was originally defined as “...the community of species (both prokaryotic and  
120 eukaryotic) associated with decomposing remains of heterotrophic biomass, including animal  
121 carrion and human corpses” (sensu Benbow et al. 2013). While the original development of this  
122 term was focused on vertebrate carrion, we argue it can be expanded to include any form of  
123 necromass (e.g., leaves, wood, dung). We also suggest the necrobiome framework is relevant to  
124 both plant (i.e., autotrophically-derived) and animal (i.e., heterotrophically-derived) necromass  
125 decomposition by highlighting the fundamental similarities and differences in the processes that



126 define decomposition. In this paper, we take a conciliatory approach to more fully develop the  
127 necrobiome framework as a unifying construct for guiding research in decomposition, regardless  
128 of how the life form was derived (Figure 2). All life is terminal and the remaining nutrients and  
129 energy are ultimately recycled within and across ecosystems (Polis et al. 1997). Some aspects of  
130 this ecological process are universal across all forms of necromass, such as disintegration,  
131 dispersal, and microbial activity. In contrast, other aspects, such as community dynamics and  
132 decomposition rates, and specific decomposer taxa, are distinct across the different forms of  
133 necromass. The major similarities and differences in necrobiome structure and function have yet  
134 to be synthesized in detail but are influenced by the overall composition and quality of the  
135 specific necromass, and the associated decomposer species that have evolved to exploit the  
136 resource.

137  
138 In this review, we provide an overview of the similarities and differences among decomposer  
139 communities associated with plant and animal necromass. We suggest plant and animal  
140 necromass research findings illustrate a broader generality to the processes structuring  
141 decomposer communities across a range of substrates that, cumulatively, have large-scale  
142 ecosystem level effects. We synthesize the key similarities and differences among these  
143 decomposer communities and place them into a broadened conceptual model of the necrobiome.  
144 We then provide details of the biotic structure of the necrobiome, the different ecological  
145 functions each part performs, and the key interactions occurring among the decomposer  
146 organisms. Further, we explore how the necrobiome provides a useful way to conceptualize how  
147 decomposition delivers nutrients, energy, genomes, and communities across space and time with  
148 ultimate responses in ecosystem biodiversity and function. By presenting a new synthesis of

149 decomposer communities that continues the approach developed by Moore et al. (2004), we aim  
150 to provide an integrative perspective to the key concepts used by researchers that are separated  
151 into plant or animal decomposition disciplines. Rather than continuing to consider each  
152 necromass type in isolation, we stress the importance of synthesizing this knowledge to develop  
153 a holistic understanding of nutrient cycling and food web dynamics across scales.

154

## 155 **AUTOTROPHICALLY- AND HETEROTROPHICALLY-DERIVED BIOMASS:**

### 156 **STANDING STOCKS, PRODUCTION AND TURNOVER**

157

158 Autotrophically-derived necromass refers to the dead biomass produced from organisms that use  
159 energy from sunlight as energy (e.g., plants and algae) or chemical reactions (various  
160 prokaryotes) to produce organic compounds. Heterotrophically-derived necromass, by contrast,  
161 refers to the dead biomass produced from organisms that obtain organic material through the  
162 consumption of other organisms (e.g., animals, fungi, protists and other micro-eukaryotes). We  
163 focus our discussion here on the initial and intact forms of necromass exceeding 8 mm<sup>3</sup> in size  
164 for which there is a richer source of literature supporting a more detailed assessment of the  
165 necrobiome. We acknowledge that all necromass ultimately becomes smaller than this size, and  
166 that the decomposition of microorganisms, such as fungal necromass, is ecologically important  
167 (Fernandez and Kennedy 2018). For instance, fungi and protists produce macroscopic structures  
168 supporting unique necrobiome communities, such as giant kelp (*Macrocystis pyrifera*  
169 [Linnaeus]) (Inglis 1989, Thunes et al. 2000). The importance of bacteria and fungi to  
170 decomposition goes beyond diversity, with reports of soil microbial biomass being an order of  
171 magnitude greater than the biomass of other soil organisms, and two orders of magnitude greater

172 than aboveground animal biomass (Fierer et al. 2009). This large amount of microbial biomass  
173 has the potential to influence ecosystem processes as it dies and becomes microbial necromass  
174 (Throckmorton et al. 2012). In fact, this microbial necromass, in combination with microbial  
175 metabolites, likely drives a significant proportion of soil organic matter formation (Kallenbach et  
176 al. 2015, Brabcová et al. 2016, Fernandez et al. 2016). Soil fungal mycelia production alone  
177 contributes to major functional processes in terrestrial ecosystems, with production (and  
178 associated death and decomposition) reported to range from 100 – 300 kg ha<sup>-1</sup> yr<sup>-1</sup> (Ekblad et al.  
179 2013). The necrobiome of dead fungal mycelium has been reported to be quite diverse (Ekblad et  
180 al. 2013, Brabcová et al. 2016), providing additional evidence that decomposition networks of  
181 microorganisms operate at every biological scale and contribute to complex patterns of  
182 biodiversity and associated functions in all ecosystems. However, there is much less understood  
183 of the species and processes of decomposition of fine particulate necromass and microorganisms  
184 (however, see Alldredge and Silver 1988) across ecosystems, a fruitful area of future research  
185 and review.

186  
187 Based on size and composition, plant material can be divided into herbaceous (e.g., leaves) and  
188 woody forms. Animal necromass produced by vertebrates can be distinguished from invertebrate  
189 based on both size and internal or external skeletal structures. Dung, however, is an intermediate  
190 and highly transformed form of necromass characterized by fragmented and partially digested  
191 plant and/or animal material mixed with host gut-dwelling microbes. Herbivorous invertebrates  
192 (Reynolds and Hunter 2004) and vertebrates (Frank and Evans 1997) are known to generate large  
193 amounts of dung that play a critical role in nutrient cycling and redistribution in terrestrial and  
194 aquatic ecosystems (Wotton and Malmqvist 2001, Carline et al. 2005, Menendez et al. 2016).

195 For the sake of space and focus, we do not elaborate on the dynamics of dung or frass; however,  
196 we encourage those interested in this topic to review the literature on the ecology and evolution  
197 of dung beetles (Hanski and Cambefort 1991) and the additional importance of feces in terrestrial  
198 (Van der Wal et al 2004) and aquatic (Alldredge and Silver 1988, Wotton and Malmqvist 2001,  
199 Joyce et al 2007) ecosystems. Below, we discuss in more detail the differences in the structure  
200 and composition of different forms of plant and animal necromass.

201  
202 To our knowledge, no efforts have been made to compare the relative production or communities  
203 associated with the decomposition of plant and animal necromass in any ecosystem. Several  
204 comparisons of biomass in forested systems, however, provide some insight into the question of  
205 how much detritus is present in a habitat at any given time. Odum (1970), for example,  
206 determined in a Puerto Rican rainforest that plants and animals made up about 58% and 0.03%  
207 of the total biomass (including soil organic matter), respectively. Leaves accounted for only  
208 about 5% of above-ground plant biomass, with the rest consisting of wood and bark. Fittkau and  
209 Klinge (1973) reported animals made up only about 0.02% of the total living biomass in an  
210 Amazonian rainforest, with < 10% of that component being of vertebrate origin. In most  
211 freshwater stream ecosystems leaf litter decomposition is a key process that influences energy  
212 and nutrient flow, biodiversity and links terrestrial and marine ecosystems through a downstream  
213 continuum (Vannote et al. 1980, Webster and Benfield 1986, Wallace et al 1997, Gessner et al.  
214 2010). Detritus in a variety of organic matter forms is thought to represent the dominate energy  
215 pathway in most lakes (Rich and Wetzel 1978, Mann 1988) and other lentic surface waters such  
216 as wetlands (Brinson et al 1981), coastal ecosystems (Duggins et al. 1989) and oceans (Parsons  
217 and Strickland 1962, Alldredge and Silver 1988). The differences in standing necromass

218 between plants and animals are probably even larger than these estimates of biomass given that  
219 animal remains typically decompose much more quickly than plants, even in some aquatic  
220 habitats (Parmenter and Lamarra 1991).

221  
222 In many cases animal necromass will be produced more quickly given the shorter life spans of  
223 most animals compared with many plants, especially when comparing invertebrate necromass  
224 production with the woody components of plants. However, many plants shed biomass  
225 seasonally as leaves and root exudates, thus producing necromass at higher rates than the woody  
226 components that make up a large portion of the standing stock biomass. Furthermore, the total  
227 production of animal necromass per year could be expected to far exceed the standing animal  
228 necromass at any given time due to a combination of short life spans, continuous reproduction  
229 and rapid decay rates such as those for zooplankton (e.g., Tang et al 2014 and references  
230 therein). By contrast, the production of plant necromass produced per year can be expected to be  
231 a small fraction of the standing plant necromass in many ecosystems, and this is especially true  
232 for forests where decomposition is slow and woody debris accumulates over time (Luysaert et  
233 al. 2008). The relative importance of these differences in standing stock, production and rate of  
234 decay of plant and animal necromass is an area of inquiry that could provide broad ecological  
235 understanding of different forms or routes of energy flow and nutrient cycling in ecosystem  
236 function.

237  
238 Focusing on vertebrate animals, Odum (1970) estimated that they accounted for only about 5%  
239 of total animal biomass, with the remaining 95% consisting of invertebrates [see Tables 7 and 22  
240 in Odum (1970)]. Additionally, a meta-analysis by Fierer et al. (2009) determined approximately

241 50% of animal biomass is belowground. The fact that invertebrates are typically much shorter  
242 lived than vertebrates, usually develop to maturity more quickly, and often have multiple  
243 generations per year suggests the annual production of necromass by invertebrates far exceeds  
244 that produced by vertebrates. Indeed, Seastedt and Tate (1981) estimated the standing arthropod  
245 necromass on the forest floor of two oak-hickory forests to be up to double the estimated living  
246 arthropod biomass for the same forests. In other habitats, such as aquatic systems, standing stock  
247 biomass can be estimated for populations and communities (Waters 1966, 1969, Benke et al  
248 1988), but some studies also produce estimates of production, often as secondary production of  
249 macroinvertebrates (Benke et al. 1988, Huryñ and Wallace 2000).

250  
251 Knowledge of animal biomass in aquatic systems can be derived from measures of secondary  
252 production of invertebrates (Hynes 1970, Waters 1977, Benke et al. 1988, Huryñ and Wallace  
253 2000). Secondary production includes the elaboration of heterotrophic population biomass and  
254 assumes a yield to higher trophic levels that ultimately becomes part of the necromass pool in a  
255 given ecosystem. The method employs estimates of individual organismal body mass, often  
256 using length-mass regressions (Benke et al 1999), and densities of those organisms and how they  
257 quantitatively change over time. One such method for calculating secondary production (i.e.,  
258 removal-summation) assumes all biomass produced eventually dies and is equivalent to  
259 estimated production for that cohort of organisms over a given time and under steady-state  
260 conditions (Boysen-Jensen 1919, Waters 1977, Benke and Huryñ 2006). Thus, secondary  
261 production estimates of invertebrates may provide quantitative insight into the contribution of  
262 animal necromass and its turnover to ecosystem energetics, much like it has been done for  
263 measuring the importance of leaf necromass on aquatic invertebrate communities (Cummins et al

264 1973, Wallace et al 1999). Measuring living invertebrate biomass over time, however, does not  
265 allow for estimating other forms of necromass, such as dung or frass.

266  
267 The importance of necrobiomes in the decomposition of entire communities of organisms could  
268 be pivotal in expanding our understanding of biodiversity contributions to global carbon and  
269 nutrient cycling, as has been discussed by Moore et al (2004). As an example, a fascinating study  
270 reported that the quality of grasshopper necromass (affected by fear of predation) mediated  
271 subsequent leaf litter decomposition in old field ecosystems, with estimated significant  
272 ecosystem level consequences (Hawlena et al. 2012). Discovering how animal and plant  
273 necromass decomposition communities interact with each other to drive nutrient and carbon  
274 cycling could lead to paradigm shifts in ecosystem science.

275

## 276 **INTRINSIC VARIABILITY OF NECROMASS TYPES**

277

278 Major forms of necromass (e.g., carrion, dung, leaves and wood) can be distinguished from one  
279 another in three important ways: (i) resource size, (ii) nutrient availability, and (iii) digestibility.  
280 First, the unit volumes of different forms of necromass vary by 5 to 11 orders of magnitude  
281 (Figure 3). While the largest forms of necromass produced globally are the woody stems and  
282 collective dead organic matter shedding of large trees, vertebrate carrion is the largest form in  
283 many non-forested ecosystems (e.g., bison in grasslands, whales in oceans) (Towne 2000, Smith  
284 and Baco 2003). Resource size has important implications for the decomposition process, with  
285 larger resources decomposing more slowly and involving a greater diversity of species than  
286 smaller resources. Small arthropod carcasses, for example, typically disappear within several

287 minutes to hours, being quickly discovered and consumed by ants, wasps or other scavengers  
288 (Fellers and Fellers 1982, Young 1984, Retana et al. 1991). By contrast, the decomposition of  
289 larger carcasses typically involves a much wider variety of species and succession of  
290 decomposer communities (Jones et al. 2015, Turner et al. 2017). Large resources also have less  
291 surface area relative to volume than small resources and this limits initial accessibility by some  
292 decomposers and thus in some cases large vertebrates (e.g., elephants) decompose initially  
293 through microbial decomposition (Coe 1978). Fragmentation and penetration of necromass are  
294 both mediated by the animal community, and thus are important processes with respect to the  
295 acceleration of decomposition (see below).

296

297 Second, different forms of necromass vary in nutritional quality. The carbon:nitrogen (C:N) ratio  
298 is commonly used to indicate the nutritional quality of organic material. The C:N ratio varies  
299 greatly among the various types of necromass, being lowest for carrion and highest for wood  
300 (Figure 3). The C:N ratio also varies widely among necromass types due largely to inter-specific  
301 variability. For example, it was reported up to 16-fold differences in C:N ratios existed among 59  
302 species of tropical wood species (Martin et al. 2014). Intra-specific variability can also be  
303 important. For example, Madritch et al. (2007) reported 2-3-fold differences in C:N ratios of  
304 caterpillar frass depending on the nutritional quality of their host plants. It has also been  
305 suggested that dung beetles are limited by assimilable C and not N (Holter and Scholtz 2007).

306

307 The third dimension is digestibility (not shown in Figure 3). For plants, digestibility is largely  
308 determined by the structure of cell walls. Due to the recalcitrance of lignocellulose (i.e., a  
309 complex of lignin, cellulose, and hemicellulose) in plant cell walls, plant matter is much more



310 resistant to decomposition than most animal soft tissues. For example, these recalcitrant  
311 compounds dominate plant tissues, respectively accounting for 20-35%, 40-45% and 20-40% of  
312 dry wood weight (Wilson and White 1986) and are highly variable among leaf litter types that  
313 contribute to stream ecosystem function (Webster and Benfield 1986, Woodward et al. 2012).  
314 Whereas a wide range of organisms can digest animal tissues, the digestion of lignin and  
315 cellulose is almost entirely limited to fungi and prokaryotes that have evolved the enzymes  
316 required to break down these compounds. This dependence on microbes to reduce  
317 autotrophically-derived detritus has given rise to a number of important symbioses between  
318 wood and leaf-feeding insects and fungi (see section on symbioses in the necrobiome).  
319 Secondary plant compounds and other extractives also reduce the digestibility of dead plant  
320 matter and can have toxic effects on decomposers (Käärik 1974, Verhoeven and Liefveld 1997).  
321 Among carrion consumers, there is also specialization on recalcitrant body parts. For example,  
322 the bearded vulture eats only the bones of vertebrates (Margalida et al. 2009), whereas some  
323 tineid moths eat only the horns and hooves of ungulates (Braack 1987). Such specialization,  
324 however, involves digestive enzymes and stomach acids, and not symbioses.  
325  
326 Another important difference between the necromass production from plants and animals  
327 concerns the gradual process by which many perennial plants, especially woody plants, die. In  
328 forest trees, for example, young branches produced at the top cast shade on older branches  
329 below, reducing their ability to capture sunlight and contribute to the growth of the tree. These  
330 overshadowed branches eventually die and fall to the ground, ultimately resulting in the long  
331 limbless trunks characteristic of many mature forest trees: a very long-term shedding of detritus  
332 over the lifetime of the tree. Similarly, growth in tree diameter is accompanied by the senescence

333 and death of the oldest, innermost rings (Figure 2a). Not only are these tissues not living, they  
334 often become colonized by diverse assemblages of decomposers many decades before the rest of  
335 the tree fully dies and falls to the forest floor (Elton 1966). This necromass accumulation co-  
336 occurs for years to decades with living tissue, providing a unique aspect of the necrobiome  
337 concept. Wounds through the bark layer (e.g., caused by extreme weather events, toppled  
338 neighboring trees, injuries from animal activity, etc.) expose the underlying phloem and wood  
339 for colonization and mark the beginning of the decay process in living trees. Although trees  
340 possess a variety of defensive mechanisms to limit the extent to which wounded areas are  
341 colonized by microbes and insects (e.g., flooding the site with water or sap, secondary plant  
342 compounds, etc.), organisms responsible for decay often become established at these sites. The  
343 first to colonize are the more ruderal microbial taxa, such as non-basidiomycetes and bacteria,  
344 whereas wood-rotting fungi arrive later (Boddy 2001). The proportion of dead tissue in otherwise  
345 living trees increases with tree age and the largest and oldest trees, or “veteran trees”, are full of  
346 dead limbs, rot holes and are believed to be particularly important to supporting necrobiome  
347 biodiversity (Speight 1989, Stokland et al. 2012). The decomposition of tree tissue has been  
348 shown to be a substantial source of methane (Covey et al. 2012), suggesting broader effects on  
349 global biogeochemical cycling rates and atmospheric chemistry that require future investigation.

350

## 351 **THE NECROBIOME**

352

### 353 **Necrobiome Structure**

354

355 The necrobiome defines a relatively concentrated set of organisms where many have evolved to  
356 detect, use and ultimately congregate with decaying organic matter either as a food or habitat  
357 resource (Benbow et al. 2013). Below, we describe in more detail the microbial, invertebrate,  
358 and vertebrate components of the decomposer community, as well as important symbioses,  
359 interactions with soil, and abiotic factors that govern the composition and dynamics of the  
360 necrobiome (Figure 1).

361

### 362 **Microbial Communities of the Necrobiome**

363

364 Structurally, the necrobiome consists of the prokaryotic and eukaryotic microbial communities  
365 (microbiome) that are internal (endonectic) and external (epinectic) components of host  
366 necromass. While given different names, these microbial communities have been reported in  
367 both living plants (Bulgarelli et al. 2013) and animals (Dillon and Dillon 2004). These microbial  
368 communities function as symbionts, commensals, pathogens and parasites during life, and the  
369 taxa become the pioneer community of plant detritus or carrion immediately after, or prior to,  
370 death in both terrestrial (Frankland 1966, Latter and Cragg 1967) and aquatic ecosystems  
371 (Preiswerk et al. 2018). The community succession of carrion is rapid after death without the  
372 biotic constraints of host immunology and physiological function (Pechal et al. 2014b, Metcalf et  
373 al. 2016), which determines the taxonomic and functional changes of the microbiota of the  
374 necrobiome through decomposition (Latter and Cragg 1965, Baldy et al 1995, Crippen et al.  
375 2015).

376

377 In aquatic leaf litter decomposition studies in streams, microbial community structural and  
378 functional succession is related to the leaf litter species and quality (Witkamp 1966, Gessner and  
379 Chauvet 1994), similar to other forms of necromass like crayfish (*Procambarus versutus*  
380 [Hagan]) molted exoskeletons (Aumen 1980); particulate aggregates (Grossart and Simon 1988)  
381 lake zooplankton (Yang et al. 2009); and oceanic phytoplankton (Fukami et al. 1985) and  
382 zooplankton (Tang et al. 2006) that harbor complex microbial dynamics that change with water  
383 chemistry, depth and system circulation (Karl et al. 1988). While difficult to study, whale and  
384 other cetacean carcasses that sink to the oceanic abyss attract a high diversity of vertebrate and  
385 invertebrate scavengers (Allison et al. 1991, Jones et al. 1998, Smith and Baco 2003) and also  
386 change the microbial ecology of the local habitat in a way that is dominated by  
387 chemoautotrophic communities (Bennett et al. 1994) in a network of species interactions (Smith  
388 et al. 1998). The microbial community changes associated with the death and deep sea  
389 decomposition of whale carcasses has not, to our knowledge, been studied, but may follow  
390 successional trajectories similar to those reported for other aquatic organisms (Preiswerk et al.  
391 2018).

392  
393 Microbial succession facilitates the proliferation of existing saprophytic taxa of the once living  
394 host through the secretion of enzymes and other compounds used to convert the newly available  
395 organic matter for assimilation. The microbial ecology of decomposing necromass is likely  
396 substantially different between plant and animal forms in terrestrial and aquatic ecosystems;  
397 however, there have been no comparative studies to directly evaluate microbial succession  
398 differences of plant and animal decomposition in either terrestrial or aquatic habitats. When one  
399 descriptively compares microbial succession of plant detritus to carrion there are interesting, and

400 potentially functionally important, differences. For instance, the taxon richness and diversity of  
401 microbial communities were shown to increase over decomposition for beech leaves, with  
402 predominate phyla not including the Firmicutes (Purahong et al. 2016); whereas for carrion,  
403 Firmicutes often make up a significant proportion of the communities and become dominant late  
404 in decomposition when taxon richness and diversity are lowest (Pechal et al. 2014b).  
405 Interestingly, the microbial communities of decomposing fungal mycelia in soil show more  
406 similar characteristics to carrion than leaf litter on soil, with Proteobacteria being most dominant  
407 early and being replaced by Firmicutes later in decomposition (Brabcová et al. 2016). The  
408 microbes inherently a part of the living organism and part of the initial communities present for  
409 decomposition are highly variable depending on the host species, health, habitat and ecological  
410 interactions (Oh et al. 2014, Pechal et al. 2014b, Junker and Keller 2015, Metcalf et al. 2016,  
411 Pechal and Benbow 2016). Using a planktonic crustacean (*Daphnia magna*), Preiswerk et al.  
412 (2018) followed microbial communities of the living host through initial death and subsequent  
413 decomposition, reporting that the communities were highly dynamic and revealed opportunistic  
414 microbes that could exploit the host even before death. Thus, the community assembly dynamics  
415 of necromass is likely dependent on host condition and interactions with the abiotic environment  
416 and other organisms, populations and communities.

417  
418 The soil environment and the microbial communities it harbors are intimately associated with the  
419 decomposition of plant and animal necromass. When considering plant necromass,  
420 decomposition has often been thought to be a product of three hierarchically organized,  
421 interacting factors: climate, substrate quality, and the biota (Swift et al. 1979). Of these factors,  
422 climate has been assumed to be the driving force behind rates of decomposition at broad spatial

423 scales, whereas litter quality, and to a more minor degree, the biota, play a more important role at  
424 finer spatial scales. However, such assumptions have recently been called into question  
425 (Bradford et al. 2016). For instance, the decomposition rate of woody debris is largely  
426 independent of climate and may actually be driven by its proximity to woody debris previously  
427 colonized by wood-decay fungi (Bradford et al. 2014). This interpretation suggests that spatial  
428 proximity of decomposer organisms may be a major driver of decomposition across a landscape  
429 (Ettema and Wardle 2002).

430  
431 The microbial communities in the soil and associated with the rhizosphere that decompose leaf  
432 litter also play a more pronounced role. One such phenomenon is the occurrence of ‘home-field  
433 advantage’ (HFA), whereby decomposer communities sharing a common history with a  
434 particular resource often decompose that resource at a greater than expected rate (Gholz et al.  
435 2000). Such a phenomenon has been found when examining the interaction between climate and  
436 microbial communities (Strickland et al. 2015, Averill et al. 2016). Although HFA has been  
437 observed across a variety of different soil decomposer communities (Ayres et al. 2009,  
438 Strickland et al. 2009a), it has been found absent in others (St John et al. 2011, Bachecha et al.  
439 2016). One explanation for this divergence is HFA only accounts for adaptation to a particular  
440 organic matter resource and does not account for the fact that some decomposer microbial  
441 communities simply decompose an array of necromass types more rapidly due to greater  
442 functional breadth (Keiser et al. 2014). That is, some microbial decomposer communities have a  
443 greater "ability" to decompose a wide range of litter substrates than do others because they have  
444 been historically exposed to chemically complex litter species. Understanding how these two  
445 metrics of microbial function drive patterns of necromass decomposition across a landscape, and

446 how they relate to microbial community characteristics, may prove informative to predictions of  
447 decomposition rates associated with shifts in plant species or changes in necromass quality  
448 (Keiser et al. 2013, Austin et al. 2014, Martiny et al. 2017). Additionally, this HFA and ‘ability’  
449 framework suggests that the function of soil microbial communities is not just a product of the  
450 current input of necromass but that it may also be influenced by past inputs as well.

451

452 Adding to the complex interactions between soil microbial communities, climate, and necromass  
453 quality is the inclusion of the broader soil community in our understanding of decomposition.

454 Soil arthropods are well known for their ability to fragment litter increasing the surface area  
455 available for microbial colonization and thereby increasing litter quality as it passes through the  
456 invertebrates’ gut (Seastedt 1984). Yet there is a growing awareness that trophic interactions  
457 within the soil community may also drive the rate and efficiency of decomposition (Buchkowski  
458 2016). For example, increased soil nitrogen can lead to greater wood decaying fungal biomass  
459 and extracellular enzyme activity in the absence of a fungivore, but the presence of a fungivore  
460 may moderate fungal biomass and wood decay rates (Crowther et al. 2015). More complex  
461 trophic cascades have also been observed, such as the removal of predatory microarthropods  
462 which led to an increase in microbivorous nematodes and a subsequent increase in microbial  
463 biomass (Santos et al. 1981). Such examples indicate that top-down drivers may play an  
464 important role in mediating decomposition (Buchkowski 2016, Hawlena and Zaguri 2016) across  
465 a variety of necromass types. The relative strengths of top-down compared to bottom-up  
466 constraints on necromass decomposition may vary depending on interacting abiotic factors and  
467 climate change (Crowther et al. 2015).

468

469  
470  
471  
472  
473  
474  
475  
476  
477  
478  
479  
480  
481  
482  
483  
484  
485  
486  
487  
488  
489  
490  
491

## **Invertebrates of the Necrobiome**

Invertebrates play a key role in decomposition. For vertebrate carrion, numerous studies have examined the role of arthropods associated with decomposition (Early and Goff 1986, Anderson 2000, Bharti and Singh 2003, Matuszewski et al. 2011, Michaud et al. 2015), and identified a relatively predictable community succession of this group (Payne 1965), which is often used in forensics (Byrd and Castner 2009). Early work on arthropod communities associated with carrion identified key taxa and their roles in decomposition (Fuller 1934, Bornemissza 1957). Later work on this topic examined arthropod communities associated with decomposing swine in a range of contrasting circumstances, including being buried (Payne et al. 1968), on the soil surface (Payne 1965) or submerged in water (Payne and King 1972). A key outcome of this work was the recognition of the important role arthropods play in returning nutrients to the surrounding ecosystem. Preventing arthropod colonization can result in a significant reduction in the rate of biomass removal from animal carcasses (Payne 1965) and can change the community succession of both microbes and necrophagous insects (Pechal et al. 2013, Pechal et al. 2014a). Additional studies have addressed other theoretical aspects of arthropod community responses to carrion resource pulses (Bilby et al. 1996, Yang 2004) or mass mortality events (Fey et al. 2015).

Research has also explored species interactions among carrion arthropods ranging from competition (Goodbrod and Goff 1990), niche partitioning (Denno and Cothran 1975), predator-prey interactions (Brundage et al. 2014), and trophic cascades responsible for variability in succession trajectories (Pechal et al. 2014a). The understanding of carrion arthropods has been facilitated by knowledge of the multiple and distinct functional roles of different taxa at



492 carcasses (Braack 1987). For example, flies (e.g., Diptera: Calliphoridae, Sarcophagidae) are the  
493 primary arthropods responsible for consuming soft tissues of vertebrate carrion and thus are  
494 traditionally recognized as saprophages (Merritt and De Jong 2015). Certain beetles (Coleoptera)  
495 including some rove beetle species (Staphylinidae) (Matuszewski et al. 2008), carrion beetles  
496 (Silphidae) (Trumbo 1990, Trumbo et al. 2016), and skin/hide beetles (Dermestidae, Trogidae)  
497 (Kulshrestha and Satpathy 2001, Barton et al. 2017) are also saprophagous. Other important  
498 functional groups include parasitoid flies (e.g., Diptera: Tachinidae) or wasps (e.g.,  
499 Hymenoptera: Braconidae) (Cammack et al. 2010) and predatory beetles (e.g., Staphylinidae)  
500 and ants (Hymenoptera: Formicidae) (Flores et al. 2014, Barton and Evans 2017). Often  
501 overlooked arthropods found at vertebrate carrion are those species located in the soil  
502 (Bornemissza 1957) or in aquatic environments (Tomberlin and Adler 1998, Merritt and Wallace  
503 2010). In many instances, these arthropods rely on the resource directly as habitat or as a  
504 nutritional resource (e.g., tissue in a stream or lake, or in the case of soil, fungi or liquids seeping  
505 into the soil). Research exploring soil or aquatic associations is limited compared with studies in  
506 above ground terrestrial environments. The ecology and evolution of dung beetles has been  
507 described and tested in detail and demonstrates close evolutionary ties of some invertebrates and  
508 necromass (Hanski and Cambefort 1991, Nichols et al. 2008).

509  
510 Extremely diverse assemblages of invertebrates are associated with decomposing plant material.  
511 Approximately 20-30% of all forest insect species, for example, are directly or indirectly  
512 dependent on dying or dead wood (Stokland et al. 2012). Included among these are a wide range  
513 of phloem- or wood-feeders (e.g., termites, beetles, wood-wasps, etc.), fungus-feeders (e.g.,  
514 beetles, flies, true bugs, etc.) and a variety of predators (Stokland et al. 2012). There is very little

515 overlap known between the invertebrate assemblages involved in the decomposition of animal  
516 and plant necromass, as well as between different forms of plant matter (e.g., leaf litter vs. wood)  
517 (Ferro et al. 2012).

518

519 Among macroinvertebrates, earthworms and termites have by far the strongest direct accelerative  
520 effects on plant biomass decomposition (Lavelle et al. 1997, Bignell and Eggleton 2000). The  
521 importance of earthworms as decomposers was illustrated by the dramatic loss of leaf litter depth  
522 following their introduction into formerly earthworm-free (due to glaciation) forests (Addison  
523 2009 and references therein. Termites are the major consumers of plant debris in soil throughout  
524 the tropics and subtropics, perhaps even exceeding fungi in importance in some areas (Liu et al.  
525 2015, Ulyshen 2016), with the different species feeding preferentially on wood, leaf litter, humus  
526 or soil (Donovan et al. 2001). Invertebrates consuming decaying plant material rely heavily upon  
527 microbes which provide a variety of benefits including digesting and softening the material,  
528 neutralizing allelopathic substances and improving nutritional quality in terrestrial and aquatic  
529 habitats (Cummins et al. 1973, Swift and Boddy 1984, Webster and Benfield 1986). Microbial  
530 biomass is itself quite nutritious relative to most forms of dead plant matter and contributes  
531 greatly to the diets of litter- or wood-feeding invertebrates, many of which are essentially  
532 fungivorous (Tanahashi et al. 2009, Mishima et al. 2016). Predators are a large proportion of the  
533 invertebrate members of the necrobiome [e.g., over a third of saproxylic beetle species in  
534 Germany (Wende et al. 2017)] and exert strong controls on necrophagous populations.

535

536 **Vertebrates of the Necrobiome**

537

538 No vertebrates, to our knowledge, are known to be specialist consumers of decomposing plant  
539 material but many species interact in important ways within the necrobiome of plant necromass.  
540 Many birds (e.g., woodpeckers) and mammals (e.g., aardvarks, armadillos, anteaters, echidnas,  
541 bears), for instance, are specialist or opportunistic predators of wood-feeding insects such as  
542 beetle larvae or termites. The fragmentation of plant matter caused by foraging vertebrates is  
543 assumed to have strong indirect effects on decay rates (Ulyshen 2016) but this remains untested.  
544 By contrast, vertebrates are known to be major consumers of carrion and are thus direct  
545 participants in the decomposition process in addition to any indirect effects they may have as  
546 fragmenters (DeVault et al. 2003).

547

548 Vertebrates consuming carrion are commonly classified into two categories: obligate scavengers  
549 rely entirely on carrion to meet their food requirements, whereas facultative scavengers are  
550 active predators or foragers, as well as scavengers (DeVault et al. 2003). In marine systems there  
551 are some deep-water fish that might also be categorized as obligate scavengers (Smith and Baco  
552 2003). The only obligate vertebrate scavengers in terrestrial systems are Old World and New  
553 World vultures (families Accipitridae and Cathartidae, respectively). Vultures have evolved  
554 several adaptations that allow for dependence on carrion, which is often an ephemeral and  
555 unpredictable food source. First and foremost, obligate scavengers (i.e., vultures) have large  
556 wing spans and other physiological mechanisms that allow them to move quickly and efficiently  
557 to search vast areas for carrion (Ruxton and Houston 2004, Shivik 2006). Vultures also have  
558 well-developed vision, and in some cases olfactory capabilities (e.g., turkey vulture, *Cathartes*  
559 *aura*), which allow them to find carcasses over large distances (Houston 1979, DeVault et al.

560 2003). Due to microbial decomposition, carrion can contain high levels of toxic compounds and  
561 thus vultures have evolved physiological adaptations in their gut that allow them to consume  
562 large volumes of these toxins (Houston and Cooper 1975, Roggenbuck et al. 2014). As a group,  
563 vultures are relatively large-bodied birds and can withstand several days without food (Ruxton  
564 and Houston 2004). The amount of biomass processed by vultures and other vertebrate  
565 scavengers varies across ecosystems (Pereira et al. 2014), but historically has been greatly  
566 underestimated (DeVault et al. 2003, Wilson and Wolkovich 2011). In one example, an  
567 estimated 70% of large ungulates in the Serengeti die from causes other than predation and  
568 become available to scavengers. Thus, collectively Serengeti vultures likely consume more  
569 biomass than all carnivorous mammals combined (Houston 1979), suggesting a large scale  
570 importance of carrion necromass in ecosystem structure and function.

571  
572 Due to their reliance on carrion, populations of obligate scavengers are inextricably linked to the  
573 availability and distribution of carrion on the landscape (Kelly et al. 2007). For some facultative  
574 scavengers the availability of carrion can influence local abundance, species distributions, or  
575 growth rates, especially in ecosystems with limited food resources (Fuglei et al. 2003, Wilmers  
576 et al. 2003, Drazen et al. 2012). For example, local arctic fox (*Alopex lagopus*) populations in  
577 Norway decreased markedly following a decline in reindeer (*Rangifer tarandus platyrhynchus*)  
578 carcasses (Fuglei et al. 2003). In Yellowstone National Park, USA, populations of facultative  
579 scavengers have benefited from the reintroduction of grey wolves (*Canis lupus*), which provide a  
580 more stable subsidy of ungulate carrion (Wilmers et al. 2003). Such effects have also been  
581 reported in aquatic ecosystems, where the abundance and distribution of deep-water fish may be  
582 modulated by carrion availability (Drazen et al. 2012).

583

584 Carrion use by vertebrate scavengers is mediated by a complex suite of biotic and abiotic factors  
585 (DeVault et al. 2003, Selva et al. 2005, Moleón et al. 2015, Turner et al. 2017). As a result, use  
586 of carrion can also vary extensively within a vertebrate species. From a temporal perspective,  
587 variability in food availability and quality can be a key factor and can be a product of intra- and  
588 inter-kingdom competition (Allen et al. 2014, Beasley et al. 2015). These forms of competition  
589 demonstrate some of the ways that very different taxonomic components of the necrobiome  
590 interact to affect decomposition. For example, the activity of invertebrates and microbes is  
591 influenced by temperature, and that means that invertebrates and microbes are more active  
592 during warmer weather and will more rapidly decompose or render carcasses toxic or  
593 unpalatable for vertebrates (Janzen 1977, Burkepile et al. 2006). Consequently, vertebrate  
594 scavengers are typically most successful when temperatures are cooler and decomposition is  
595 slowed (DeVault et al. 2004a, Selva et al. 2005, Parmenter and MacMahon 2009). However, in  
596 some areas the presence of an abundant and efficient vertebrate scavenger species can shift the  
597 balance toward vertebrates, even in warm weather (Houston 1986, DeVault et al. 2011).  
598 Competition for carrion resources also differs across habitat types (Beasley et al. 2015, Turner et  
599 al. 2017), although such differences likely reflect the composition of local vertebrate  
600 communities (DeVault et al. 2004a) and microclimates (Parmenter and MacMahon 2009).  
601  
602 Collectively, obligate and facultative scavengers provide a number of critical ecosystem services  
603 through the removal of animal necromass from landscapes (Moleón et al. 2014, DeVault et al.  
604 2016). Intact scavenging communities (especially the presence of vultures) may reduce risks of  
605 disease transmission among mammals due to rapid removal of carcasses from ecosystems

606 (Jennelle et al. 2009, Ogada et al. 2012, Hill et al. 2018). There is a clear role of vertebrates in  
607 the decomposition of carrion and placing them in the broader necrobiome framework highlights  
608 their links with other taxonomic groups (i.e. insects and microbes) and important environmental  
609 moderators of decomposition and overall ecosystem structure and function.

610

### 611 **Symbioses of the Necrobiome**

612

613 Although symbioses between invertebrates and microbes are common within necrobiomes, the  
614 nature of these interactions differs greatly between decomposers associated with autotrophically-  
615 or heterotrophically-derived biomass. Due to the strength of lignocellulose, dead wood is among  
616 the most recalcitrant forms of plant matter and is also characterized by some of the highest C:N  
617 ratios of any organic material (Figure 3). Although endogenous cellulases are known from some  
618 termites (Blattodea) and may play a particularly important role in the metabolism of Termitidae,  
619 which lack the protists of “lower termites” (Lo et al. 2011), wood-feeding insects are mostly  
620 unable to digest lignocellulose without the enzymes produced by various endo- or ecto-symbiotic  
621 microorganisms. Endo-symbionts include a wide range of gut-dwelling bacteria, archaea and  
622 eukarya (e.g., yeasts and protists) that have been the subjects of intense study in both termites  
623 (O'Brien and Slaytor 1982, Brune 2014) and various beetle species (Reid et al. 2011, Urbina et  
624 al. 2013).

625

626 In addition to digestion, another important function performed by prokaryotes within the guts or  
627 salivary glands of many wood-feeding arthropods is nitrogen fixation, which often provides

628 much of the nitrogen required by these organisms to complete development (Ulyshen 2015, and  
629 references therein). Ecto-symbioses are common between insects and fungi associated with  
630 decomposing wood. Ambrosia beetles, for example, cultivate and consume symbiotic fungi  
631 within galleries these insects create in dead wood. These insect-fungi associations have  
632 repeatedly evolved within scolytine and platypodine Curculionidae and also exist in other beetle  
633 families, such as Lymexylidae and Erotylidae (Farrell et al. 2001, Toki et al. 2012). The bodies  
634 of these insects possess special storage structures, called mycangia, within which their fungal  
635 symbionts are transported. Mycangia also exist in wood-feeding insects that are not technically  
636 ambrosia beetles. For example, female stag beetles (Lucanidae) possess mycangia near the ends  
637 of their abdomens that contain xylose-fermenting yeasts, which are presumably added to  
638 oviposition sites to benefit larval development (Tanahashi et al. 2010). Some of the most  
639 effective forms of symbioses between insects and fungi are exhibited by leaf-cutting ants and  
640 macrotermite termites, both of which cultivate fungal gardens within their nests to perform the  
641 digestion of plant material (Nobre et al. 2011). Macrotermitines are thought to be among the  
642 most efficient wood-consuming insects due to their symbioses with the fungi *Termitomyces*  
643 (Basidiomycetes: Lyophyllaceae) which both allows them to process wood more quickly and  
644 increases assimilation efficiency (Schuurman 2005, Brune and Ohkuma 2011). Other functions  
645 provided by endo- and ecto-symbionts of arthropods associated with decomposing plant material  
646 include detoxification of plant secondary compounds (Dowd 1992), defensive mutualism  
647 (Chouvenc et al. 2013), and protection from pathogens (Peterson and Scharf 2016, and  
648 references therein).

649

650 Whereas symbioses within plant-based necrobiomes largely stem from the recalcitrance and  
651 nutrient deficiency of dead plant matter, those within animal-based necrobiomes appear to have  
652 arisen largely in response to intense competition with other organisms as well as threats from  
653 pathogens. Necrobiome community interactions may follow a gradient of resource quality where  
654 the most labile forms of necromass facilitate competition and the most recalcitrant forms support  
655 symbioses (Janzen 1977). Microbial competition has been demonstrated to have strong negative  
656 effects on the reproductive success of some carrion-feeding insects (Rozen et al. 2008). Although  
657 carrion-feeding insects produce a variety of antimicrobial peptides and lysozymes to minimize  
658 competition from bacteria (Jordan et al. 2015), they also harbor, within their guts, diverse and  
659 unique microbial assemblages (Kaltenpoth and Steiger 2014). The exact functions provided by  
660 these microbes remain poorly understood but reducing competition from free-living carrion  
661 microbes and other carrion-feeding animals, detoxification and providing protection from  
662 pathogens are several potential benefits (Kaltenpoth and Steiger 2014, Trumbo et al. 2016).  
663 Adult burying beetles, for instance, are known to transmit gut bacteria to their offspring both  
664 directly through regurgitation and anal secretions applied during carcass preparation (Wang and  
665 Rozen 2017). Although the secretions produced by burying beetles have antimicrobial properties,  
666 free living microbes appear to be reduced in favor of microbes found in the guts of these insects  
667 (Duarte et al. 2018). Rather than reducing competition from other microbes, the antimicrobial  
668 secretions produced by *Nicrophorus* may serve primarily to help conceal carcasses from  
669 competitor scavengers by suppressing the bacterial production of olfactory cues (Trumbo et al.  
670 2016). While research in both plant and animal necromass decomposition has shown many  
671 examples of symbioses and cross-domain interactions, including the well-known symbioses of



672 ruminant digestion of plant necromass (Hungate 1966), there are likely many more to be  
673 discovered.

674

### 675 **Abiotic Factors and the Habitat Template**

676

677 Decomposition and the necrobiome constituents are constrained by abiotic factors defined by the  
678 surrounding habitat or ecosystem. Terrestrial, aquatic, and human-modified habitats each differ  
679 in their communities and climate, and this affects the rate and progression of decomposition and  
680 the species diversity that uses necromass (Beasley et al. 2012). Future studies should consider  
681 how changes in abiotic factors described below, in addition to extreme weather events, ocean  
682 acidification, and rising annual global temperatures could alter detritus availability, necrobiome  
683 dynamics, and ultimately decomposition processes in ecosystems.

684

685 Although many decomposition studies have occurred on land, the majority of Earth's surface is  
686 comprised of aquatic habitat. Aquatic habitats include freshwater (e.g., streams, ponds, lakes,  
687 wetlands), marine (e.g., ocean) and brackish (e.g., estuaries) environments. The inherent  
688 physiochemical properties unique to these aquatic habitat types can substantially alter  
689 decomposition through direct or indirect abiotic interactions that constrain necrophagous  
690 physiology or mechanical ability to breakdown organic matter. Further, depending on where a  
691 resource is located within the water column, abiotic factors such as temperature, oxygen  
692 availability, carbon dioxide levels, ion concentrations, turbidity, pressure, salinity, pH, and flow  
693 dynamics will impact the rate and taphonomic changes of decomposition and determine the  
694 biodiversity of consumers that may use the decomposing organic material (Hattenschwiler et al.

695 2005, Wallace 2015). For example, swine carcasses placed in the deep-sea fail to undergo bloat  
696 during the decomposition process at these depths because of the increased pressure within the  
697 environment (Anderson and Bell 2016). Woody debris and leaf litter in aquatic habitats,  
698 primarily in streams, have additional factors to consider that impact the abiotic conditions and  
699 ultimately the decomposition process. These include but are not limited to: where the wood is  
700 located within the stream (e.g., submersion, buried in the substrate, suspended); the morphology  
701 of the streambed (e.g., sand or silt, rock size); and hydrology (e.g., season flooding and/or spate  
702 events) (Scherer 2004, Ruiz- Villanueva et al. 2016).

703  
704 Terrestrial habitats can be separated into broad biomes: tundra, boreal forest, temperate forest,  
705 desert, grassland, and tropical rain forest, with the abiotic factors of temperature, humidity, wind,  
706 solar radiation, and exposure substantially altering decomposition (Benbow et al. 2015a). These  
707 abiotic factors can further interact with landscape configuration and vegetation within each  
708 ecosystem type (García- Palacios et al. 2013). Soil conditions specific to a habitat can alter  
709 decomposition dynamics of plants and animals due to direct and indirect associations with soil  
710 nutrients, pH, salinity, pollution or other contaminants, temperature and moisture content  
711 (Refsnider and Janzen 2010). Further, landscape features, such as land use and cover,  
712 connectivity and complexity can mediate consumer attraction and use of necromass resources.  
713 For example, habitats with increased biodiversity, such as tropical rainforests, have an increased  
714 species pool that can utilize a resource during decomposition when compared to habitats with  
715 slightly decreased complexity, such as an African tropical woodland (Cornaby 1974, Braack  
716 1987).

717

718 The final category of ecosystems, which may not be as apparent as the previous two groups, is  
719 anthropogenic-associated ecosystems. These habitats are comprised of human built (e.g.,  
720 residential structures) or modified areas (e.g., disturbed or change habitats) that impact the  
721 decomposition of organic material and biodiversity of the necrobiome. Specifically, modification  
722 of ecosystems often leads to habitat loss and fragmentation resulting from human development  
723 (e.g., urbanization, deforestation and monoculture farming). Habitat fragmentation typically  
724 elicits negative impacts on biodiversity (MacArthur and Wilson 1967). A recent meta-review,  
725 however, revealed approximately three-quarters of studies showed some measures of positive  
726 response by communities affected by habitat fragmentation (Fahrig 2017). Overall, the impact of  
727 human-modification to decomposition and decomposer communities is underexplored and  
728 poorly understood.

729

## 730 **NECROBIOME ECOLOGICAL FUNCTIONS**

731

732 A key feature of our expanded necrobiome concept is the broad structural and functional  
733 template it provides for better understanding the range of similarities and differences in  
734 decomposer communities found associated with plant and animal detritus. Volatile organic  
735 compound (VOC) emissions result from tissue decomposition and microbial metabolism and are  
736 an important mechanism for the attraction of exogenous macro-saprotrophs, such as blow flies,  
737 bark beetles, and nematodes (see Table 1), from the contiguous substrate (e.g., soil) and the  
738 surrounding local habitat and ecosystem (Ma et al. 2012, Ulyshen 2016). The types of VOCs  
739 produced are determined by necromass composition, quality, and rate of decay; and they are  
740 known to affect the recruitment of exogenous macro-saprotrophs (i.e., invertebrate

741 decomposers), in time frames ranging from minutes to years after death (Mađra et al. 2015,  
742 Ulyshen 2016). In leaf litter decomposition, microbial communities are known to markedly  
743 contribute to the production of associated VOC profiles (Gray et al. 2010). Ultimately,  
744 decomposition is achieved by species that directly consume necromass or produce the enzymes  
745 that catabolize compounds and degrade dead tissues. However, members of the necrobiome also  
746 affect decomposition through physical alterations to the necromass, as well as intra- and inter-  
747 specific species interactions that generate complex inter-dependencies among organisms. Among  
748 the latter, interactions between invertebrates and microbes are particularly important. We  
749 describe below major functions performed by community members of the necrobiome that both  
750 directly and indirectly affect decomposition and compare and contrast functions occurring at  
751 plant and animal necromass.

## 753 **Digestion**

754  
755 Many organisms can perform the primary function of digestion with respect to relatively labile  
756 animal tissues. The primary digesters at animal necromass are specialist carrion flies (Payne  
757 1965), although there can be minor contributions from other insect groups, such as beetles or  
758 ants (Barton and Evans 2017). Vertebrates should also be considered digesters as they will  
759 consume entire carcasses before invertebrates are able to proliferate and participate in this  
760 function (DeVault et al. 2003). For plant necromass, wood-feeding invertebrate symbionts and  
761 saprophytic fungi and microbial communities in the stomachs of ruminant mammals (Hungate  
762 1966) are capable of completely degrading the lignocellulose of plant cell walls to perform a  
763 digestion function (Swift et al. 1979). Digestion by saprophytic fungi and microbes is performed

764 via the production of extra-cellular enzymes in ruminants (Refsnider and Janzen 2010) and is an  
765 external process rather than internal as for insects and vertebrates.

766

### 767 **Fragmentation/penetration**

768

769 Physical alterations brought about by animals that act to fragment or tunnel into dead material  
770 can strongly affect decomposition. The activities of carrion-feeding vertebrate scavengers and  
771 vertebrates foraging for insects in dead wood act to fragment large necromass resources.  
772 Fragments decompose more quickly than intact pieces due to increased surface area and  
773 coverage of microbial biofilms, as well as improved aeration (e.g., Boddy 1983). Penetration of  
774 the outer protective layers of dead animals and plants (e.g., skin or bark) is another important  
775 physical alteration brought about by animal activity, which facilitates colonization by microbes  
776 and other organisms (Connell and Slatyer 1977, Mann et al. 1990). In animal necromass this  
777 activity is performed by larger scavenging animals (e.g., crows, vultures and hyenas) that pick at  
778 parts of a carcass and enable entry by insects and microbes. In decomposing logs, insects that  
779 create holes through bark and tunnel into the heartwood (Leach et al. 1937) play a particularly  
780 important role in facilitating the establishment of rot fungi. Some vertebrates, such as aardvarks  
781 (*Orycteropus afer*) in Africa or echidnas (*Tachyglossus aculeatus*) in Australia, also play a role  
782 in the fragmentation of logs when searching for ant or termite nests. Although there are many  
783 insect species that perform a fragmentation or penetration functional role in terrestrial habitats,  
784 there are fewer major groups of xylophagous macroinvertebrates in freshwater systems  
785 (Anderson et al. 1978), contributing to the slow rates of wood decomposition in these habitats.  
786 Certain molluscs perform this function in marine environments and can cause a great deal of

787 damage to wooden boats and pilings (Nair and Saraswathy 1971). Another consequence of  
788 fragmentation can be changes in microbial community composition. The production of fine  
789 woody particulate material by many wood-boring insects is thought to favor bacterial  
790 communities (Swift and Boddy 1984, van der Wal et al. 2007), including free-living N-fixers  
791 (Ausmus 1977).

792

### 793 **Nutrient and microbe transport and dispersal**

794

795 Organisms within the necrobiome play a role in moving nutrients away from decaying  
796 necromass through both passive and biological means (Figure 1). The microbes and elements of  
797 decaying organic matter can move by direct translocation of microbial cells or along elemental  
798 concentration gradients within the biomass and into, onto and within contiguous substrata. This  
799 transfer is perhaps best visualized by the process of purging, whereby fluids and gases from a  
800 decaying animal escape into soil or the atmosphere. Nutrients and particles also are dispersed by  
801 mobile micro- (e.g., fungi, protists) or macro-saprotrophs (e.g., flies) that act as vectors of the  
802 organic matter, microbes, and elements. The dispersal of these materials can occur in multiple  
803 ways: 1) trophic transfer where the microorganisms and elements are ingested but not  
804 assimilated, and then egested after saprotroph movement away from the necromass source; 2)  
805 trophic transfer where the microorganisms and elements are ingested and assimilated, and  
806 released after saprotroph movement and then death either through predation or decomposition; 3)  
807 external attachment to the saprotroph by physical or chemical means during movement; and 4)  
808 by being aerosolized or passively dispersed by winds, rainfall, water currents, or thermal energy

809 (e.g., convection). Additional discussion is given below of how members of the necrobiome  
810 interact in networks to transport and disperse microbes and nutrients.

811

### 812 **Detoxification**

813

814 Non-structural secondary compounds are often present in heartwood and are known to be toxic  
815 to many microbial decomposers and insects (Käärik 1974). The abundance of these compounds  
816 can vary by as much as 50-fold among angiosperms and 20-fold among gymnosperms (Cornwell  
817 et al. 2009), and the variance is thought to be largely responsible for the differences in decay  
818 resistance observed among different wood species (Bultman and Southwell 1976). The protective  
819 effects of these and other extractives diminish over time (Bultman and Southwell 1976);  
820 however, bacteria and fungi both play a part in neutralizing these potentially toxic compounds  
821 (Burnes et al. 2000, Dorado et al. 2000). Further, microbial symbionts of some insects associated  
822 with detritus provide detoxification function of plant secondary compounds (Dowd 1992). As far  
823 as we are aware, there are no analogous intrinsic toxins present associated with microbial  
824 communities of animal necromass that retard microbial or insect activity. However, a burying  
825 beetle, *Nicrophorus vespilloides* (Silphidae) (Arce et al. 2012), produces antimicrobial secretions  
826 that compete with microbes on carrion. Overall, the detoxification pathways in plants suggest  
827 this function is primarily the activity of microbial components of the plant necrobiome.  
828 However, there are likely undiscovered analogous activities in animal necromass decomposition.

829

### 830 **Predation**

831

832 Predators are conspicuous and important members of the necrobiome. It is not uncommon for  
833 predators to be among the most species-rich guilds in dead wood (Hammond et al. 2001, Wende  
834 et al. 2017), as well as at carcasses (Barton and Evans 2017). Moreover, opportunistic predation  
835 by non-predatory invertebrates is commonplace in decomposing wood. Termites are known to  
836 feed on dead insect larvae and other invertebrate carcasses encountered while foraging in wood  
837 (Thorne and Kimsey 1983, and examples therein), which includes cannibalizing dead or injured  
838 nestmates (Wilson 1971). Many wood-boring beetle larvae are also known to prey on other  
839 larvae encountered in dead wood (Soper and Olson 1963) and this probably relieves the  
840 limitations imposed by the low nutritional quality of wood.

841  
842 For carrion, cannibalism among fly larvae may be an adaptation to intense competition rather  
843 than a nutritional requirement (e.g., the hairy maggot blow fly *Chrysomya rufifacies* Macquart  
844 1842) (Baumgartner 1993). Most predatory beetles and ants at carcasses tend to be generalists  
845 and take advantage of the concentrated and abundant insects associated with carrion (Barton and  
846 Evans 2017). Some species of predatory insect may also opportunistically scavenge on carcass  
847 tissue, possibly due to the similar stoichiometric composition and easy switching between  
848 resources. The function of predators (and parasitoids, not discussed here) in decomposition is  
849 largely restricted to moderating the abundance of other decomposer organisms, and in most cases  
850 probably does not alter the rate or pattern of decomposition to any notable degree. Predatory  
851 ants, however, have been shown to dominate the insect community and retard rates of mass loss  
852 of both wood (Warren and Bradford 2012) and animal carcasses (Barton and Evans 2017).  
853 Indeed, Houston (1985) argued that ant communities in South America prolong carcass  
854 availability to vertebrates by feeding on dipteran larvae that otherwise would quickly consume



855 the carcasses. Further, it is known that some invertebrate predators approach carrion specifically  
856 to prey on other organisms and not necessarily the necromass (Payne 1965).

857

## 858 **NECROBIOME INTERACTIONS AND NETWORKS**

859

860 In addition to discussing the taxonomic and functional organization of the necrobiome, we  
861 expand the necrobiome concept to also consider the complex biotic and abiotic mechanisms of  
862 species and element (e.g., nutrients, carbon) translocation and dispersal from the decaying  
863 necromass into adjacent substrates, the local habitat, and ultimately into the larger ecosystem, as  
864 originally presented by Polis and Strong (1996). We argue that the decaying biomass holds a  
865 community of microbial species that often follow elemental movement into and throughout the  
866 environment, thus providing a way to track the dispersal of species, their genomes, metabolic  
867 diversity and function after host death and through decomposition. While understudied, the  
868 dispersal of genetic and metabolic information into the surrounding environments (Preiswerk et  
869 al 2018) is arguably a potentially transformative way to view decomposition as an ecosystem  
870 process through multiple disciplines and areas of scientific inquiry.

871

872 Many invertebrates act as vectors of microbial digesters from one resource to another. This  
873 transfer by insects can disperse gut symbionts as well as those that passively carry microbes on  
874 their bodies (Persson et al. 2009, Strid et al. 2014). Fungus-farming termites perform the reverse  
875 role, where dead plant particles are transported back to nests containing ecto-symbiotic fungal  
876 gardens (Wood and Thomas 1989). Blow flies and house flies are known to carry and transmit  
877 pathogenic bacteria to and from decomposing organic matter (Greenberg and Bornsetein 1964,

878 Greenberg et al. 1970, Greenberg 1973, Macovei and Zurek 2006, Nayduch 2017). A recent  
879 study by Weatherbee et al. (2017) showed that carrion associated microbial taxa consumed by  
880 blow fly larvae are transferred into the gut of the larvae. Although it is not definitively known  
881 whether microbes are passed transstadially to adults in all species (Singh et al. 2015), evidence  
882 from adult flies suggests a high likelihood that some bacteria ingested from carrion are  
883 assimilated by larvae, persist into the adults, and then are dispersed into the landscape  
884 (Weatherbee et al. 2017).

885  
886 In addition to vectoring microbes, many insects also carry other invertebrates from one  
887 ephemeral or isolated resource patch to another. Phoresy is common among invertebrates  
888 associated with dung and carrion (Krantz 1998), fungal fruiting structures (Fain and Ide 1976),  
889 and dead wood. Examples of saproxylic insects from which phoretic passengers have been  
890 recorded include bark beetles, click beetles, cerambycid beetles, passalid beetles and parasitic  
891 wasps (Haack and Wilkinson 1986). As pointed out by Moser and Roton (1971), virtually all  
892 relatively large insects capable of long distance dispersal serve as hosts for one or more phoretic  
893 species. Many host species carry not just a single phoretic species but a whole community of  
894 passengers. For example, at least eighteen species of mites are known to be phoretic on the bark  
895 beetle *Dendroctonus frontalis* (Coleoptera: Curculionidae) in the southeastern United States  
896 (Moser 1976). Although phoresy is often viewed as a form of commensalism where the phoretic  
897 species benefits and the host is unaffected, this is not always the case with reports of both  
898 negative (Lindquist 1969, Hodgkin et al. 2010) and positive impacts (Purrington and Drake  
899 2008) on the host.

900

901 Interactions between decomposer organisms belonging to different kingdoms (or domains) are  
902 quite common and warrant separate attention. Recent evidence indicates that microscopic species  
903 interacting with higher organisms are playing a far greater role in regulating succession patterns  
904 (Ezenwa et al. 2012). Interkingdom interactions between plants and microbes are prevalent, and  
905 these interactions may also transect invertebrate communities. A specific example is with  
906 Sphagnum mosses. These plant assemblages dominate peatland bog ecosystems and form a  
907 unique and extreme habitat for microbes. This moss genus is well known for its preservative  
908 properties and slow rate of decomposition, occupies one-third of land on the planet, and stores  
909 more carbon than any other single genus of plant (Turetsky 2003). Highly acidic conditions  
910 along with low concentrations of nutrients, together with specialized leaf structures with high  
911 water content, lead to very specialized microbial colonization that is unique to Sphagnum species  
912 (Opelt et al. 2007). Furthermore, Sphagnum moss species produce secondary metabolites, such  
913 as phenolics, terpenoids, and tannins, and accumulate carbohydrates, that influence microbial  
914 colonization and subsequently microbially-mediated decomposition. These plant specific factors  
915 may facilitate peat accumulation through lowering of redox potential that leads to decreased  
916 microbial decomposition rates (Belyea 1996). Methanotropic bacteria within the bogs themselves  
917 are well documented with Burkholderia and Serratia to be among the most dominant bacteria  
918 colonizing Sphagnum plants (Juottonen et al. 2005). Very few fungi colonize the actual plants, as  
919 these hosts are known for their antifungal and antibacterial activity (Opelt et al. 2007). A recent  
920 study showed that bacterial isolates from two Sphagnum species had antagonistic activities  
921 against fungi, while also performing nitrogen fixation and lowering host plant ethylene levels  
922 (Knorr et al. 2015). Despite these documented antimicrobial properties, a relatively large number  
923 of fungal taxa have been identified on *S. fuscum* (Thormann et al. 2001). Thus, the overall

924 functioning of peat bog ecosystems is heavily influenced by interkingdom interactions occurring  
925 among plants, bacteria, and fungi, as well as their associated chemicals associated with high  
926 volumes of necromass.

927  
928 Microbes have evolved complex strategies for outcompeting one another for resources available  
929 within carrion. Although in the early stages of discovery, it is clear microbes associated with  
930 carrion play important roles in regulation of arthropod attraction, colonization, and utilization of  
931 the remains. Furthermore, recent evidence indicates within kingdom interactions (e.g., blow fly  
932 predator v blow fly prey) are partially governed by their associated microbiome in carrion  
933 (Crippen et al. 2015, Weatherbee et al. 2017) and other necromass decomposition networks  
934 (Purahong et al. 2016, Deveau et al. 2018). Some compounds serve as a mechanism of  
935 communication and group responses, better known as quorum sensing (Nealson and Hastings  
936 1979), while others serve to disrupt communication. In both cases, these processes give a  
937 competitive advantage to the producer of these compounds by reducing competition with other  
938 microbes within the environment. For example, indole, a by-product of tryptophan degradation,  
939 is a key molecule used by *Escherichia coli* (gut microbe of many vertebrate species) as a quorum  
940 sensing molecule (Kim and Park 2015) and is a bacterial taxon commonly associated with  
941 vertebrate carrion (Metcalf et al. 2016). In contrast, dimethyl disulphide (DMDS) is detected  
942 during the later stages of vertebrate decomposition (Forbes and Perrault 2014) and is a quorum  
943 quenching molecule for many Gram-negative species. In both instances, these compounds  
944 regulate blow fly attraction (Ma et al. 2012, Tomberlin et al. 2012b). In addition to being an  
945 attractant, the response of blow flies to these compounds tends to be sex and physical state (e.g.,  
946 gravid v non-gravid) specific (Brundage et al. 2017).

947

948 The roles of microbial and invertebrate activity during animal necromass decomposition also  
949 scales to vertebrates and can affect vertebrate scavenger interactions (DeVault et al. 2003,  
950 Beasley et al. 2015). For instance, temperature is known to impact invertebrate and microbial  
951 activity (Vass 2001, Zhou and Byard 2011), and was later found to affect the percentage  
952 consumption of small-mammal carcasses by vertebrate scavengers (DeVault et al. 2004a).  
953 Although the mechanism of this interaction has not been tested, microbial and invertebrate  
954 communities are known to affect the rate of decomposition (Simmons et al. 2010) and quality of  
955 the carcass (Payne et al. 1968, Pechal et al. 2013, Pechal et al. 2014a), which likely influences  
956 the quality and quantity of these resources available to vertebrate scavengers. The reciprocal  
957 effect can be inferred from high vertebrate scavenging rates (DeVault et al. 2011) that eliminate  
958 these resources via consumption of fresh carcasses as they become available and therefore  
959 reducing persistence in the landscape (Jones et al. 2015). Additionally, there is some evidence  
960 that microbes of carcasses and vertebrate scavengers may have co-evolved, as hyenas frequently  
961 feed on ungulate carcasses that have died from anthrax without showing disease characteristics  
962 (Villiers Pienaar 1969, Gasaway et al. 1991). There is precedent for evolved adaptations in avian  
963 scavengers for competing with carrion microbial communities (Houston 1974, Houston and  
964 Cooper 1975). These cross-kingdom scavenger interactions are important to ecosystems but are  
965 often overlooked aspects of the ecology of animal decomposition (Wilson and Wolkovich 2011);  
966 however, they constitute linkages that make up food webs, indirect predatory-prey effects, and  
967 necrobiome community networks.

968

969 The biotic structure and interactions among necrobiome taxa inherently give rise to network  
970 structure that can be used to define the complex and dynamic nature of organic matter  
971 decomposition. Foodwebs are biotic networks of interacting organisms, energy and nutrient flow  
972 and define communities and metacommunities. The complexity of foodweb networks has been  
973 studied in several systems (Polis and Strong 1996, Eklöf et al. 2013, Wende et al. 2017), and has  
974 been proposed as a useful tool for developing a systems approach to understanding ecosystems  
975 (Dunne et al. 2002). The necrobiome framework offers an opportunity to quantify a subset of  
976 overall ecosystem foodweb networks and test novel hypotheses related to the strength of direct  
977 and indirect interactions among species and trophic levels. Furthermore, the necrobiome  
978 approach affords the opportunity to test hypotheses and further theory with evolutionary  
979 importance of the decomposer communities, much like that argued for understanding the broader  
980 importance of scavenging in foodweb research (DeVault et al. 2003, Wilson and Wolkovich  
981 2011). The complexity and stability of plant and animal necromass networks likely arises, in  
982 part, from the indirect (non-physical) interactions that involve evolved sensing, physiology and  
983 behavior associated with finding and consuming decomposing resources (Tomberlin et al. 2011)  
984 and direct interactions of the necrobiome.

985

## 986 **SPATIAL AND TEMPORAL DIMENSIONS OF THE NECROBIOME**

987

988 Dead wood and animal carcasses vary in size, spatial density, and temporal input and turnover,  
989 adding another layer to necrobiome dynamics. Succession and related theory (Box 1) is well  
990 documented for decomposer communities associated with plant and animal necromass  
991 (Refsnider and Janzen 2010). Yet, scaling up these localized processes and integrating them into

992 the higher-level spatial and temporal dynamics occurring among multiple decomposer  
993 communities at ecosystem scales is a significant challenge. Indeed, very few studies have yet to  
994 document the large-scale dynamics of single sources of detritus (e.g., forest stand blow downs)  
995 beyond ontogeny-based death events of anadromous fishes (e.g., Pacific salmon, *Oncorhynchus*  
996 spp.), despite recent work demonstrating that large animal mass mortality events are increasing  
997 in both magnitude and frequency globally (Fey et al. 2015). Empirical studies of large-scale  
998 carrion effects include the mass emergence of cicadas (*Magicicada* spp.) in North American  
999 forests (Yang 2004), the annual spawning runs of salmon (e.g., *Oncorhynchus* spp.) (Hocking  
1000 and Reynolds 2011), and the mass drowning of wildebeest in the Mara River in Kenya  
1001 (Subalusky et al. 2017). These studies have shown how substantial quantities of carrion can alter  
1002 forest soil microbial communities (Yang 2004), change the biomass and composition of riparian  
1003 plant communities (Hocking and Reynolds 2011), and subsidize 50% of the diet of fish  
1004 communities (Subalusky et al. 2017). Such instances provide evidence of carrion and associated  
1005 necrobiomes changing nutrient cycling in a way that alters ecosystem functioning. Examples of  
1006 plant necromass altering ecosystems are more common, with litter fall and decomposition having  
1007 critical role in the cycling of mega-tons of carbon annually (Gessner et al. 2010). Plant litter is  
1008 ubiquitous in vegetated ecosystems, and associated decomposer communities are similarly  
1009 widespread (Refsnider and Janzen 2010). Similar to the spatially discrete dead trees and coarse  
1010 woody debris, there are fewer instances of analogous, highly recalcitrant, structures of animal  
1011 carcasses (e.g., whale bones) that provide focal points for the assemblages of distinct necrobiome  
1012 communities over long (e.g., years to decades) temporal scales. However, the impact of such  
1013 spatially and temporally unpredictable events on bottom-up or top-down processes are still not  
1014 fully understood.

1015

1016 Across a landscape are many dead logs and carcasses of different sizes and decay stages, with

1017 each stage having a different necrobiome defined by its attendant species and their interactions.

1018 This variability emphasizes the among-necromass dynamics of multiple necrobiomes.

1019 Succession is well documented (see Box 1) and tells us that any single necrobiome exists only

1020 for a certain window of time (Table 1), which might range from hours to days for carcasses in

1021 warm conditions (Payne 1965, Barton and Evans 2017) or months to years for some large fallen

1022 trees (Weedon et al. 2009) or mummified animal necromass in habitats such as dry deserts or

1023 peat bogs (Brothwell et al. 2002, Chapman 2015). Decomposer organisms searching for their

1024 next resource to continue their life-cycle are therefore not only looking for another log or

1025 carcass, but also a specific decay stage, thus further emphasizing the rapid temporal turnover of

1026 the necrobiome within a necromass source. For specialist decomposers in the necrobiome, their

1027 existence is entirely dependent on resource continuity through time, with continuity also

1028 dependent on spatial proximity due to dispersal limitation. Thus, the implications of high-level

1029 among-necromass necrobiome dynamics become apparent across multiple scales in ways that

1030 include: (i) the reproductive success and survival of individuals, and their subsequent dispersal

1031 and contribution to population genetics (Stokland et al. 2012); (ii) competition among species,

1032 coexistence, and community diversity and heterogeneity; (iii) the consumption and metabolic

1033 breakdown of necromass via microbes and saprophytes; and (iv) dispersal of necromass energy,

1034 genomes and elements through the environment as new living biomass.

1035

1036 There are gaps in our knowledge of ecosystem-scale responses to plant and animal necromass,

1037 and many fundamental questions remain. A framework that joins patch-scale phenomena with



1038 ecosystem-scale consequences is therefore needed to fully appreciate the cross-scale dynamics of  
1039 the necrobiome. We synthesize in Figure 4 previous work that has attempted to conceptualize  
1040 among-carcass dynamics (Barton et al. 2013a, Benbow et al. 2015b), but which also applies  
1041 conceptually to leaf litter, dead trees and coarse woody debris. We suggest that the spatial and  
1042 temporal dimensions of the dynamic necrobiome are best conceptualized hierarchically, with  
1043 community dynamics occurring spatially and temporally within necromass sources as well as  
1044 spatially and temporally among multiple sources of necromass (Figure 4).

1045

## 1046 **BUILDING THEORY IN DECOMPOSITION ECOLOGY**

1047

1048 The necrobiome framework illustrates the links and interactions among decomposer organisms  
1049 associated with defined substrates, and the key interactions with each other and the environment  
1050 over a range of spatial and temporal scales. For example, microbes and necrophagous insects  
1051 interact on individual carcasses and come into direct contact with one another (Crippen et al.  
1052 2015). Similarly, phoretic mites interact directly with their insect hosts and carcasses, which  
1053 provides focal points for their movement via necrophilous insects across landscapes (Perotti and  
1054 Braig 2009). Yet these examples of direct contact that occur over relatively small spatial and  
1055 temporal scales can have implications at much larger scales by altering metacommunity structure  
1056 and the rate and pathway of nutrient and genome movement through ecosystems. The  
1057 necrobiome allows for clearer conceptualization of the links between groups of species and their  
1058 environmental context, and we believe this can help to develop empirical tests of ecological  
1059 theory related to decomposition. For example, for many forms of necromass, especially large and  
1060 recalcitrant forms, the decomposition process is characterized by a succession of species (see

1061 Box 1). Succession is, in part, guided by colonization history and priority effects as early  
1062 colonizers can have facilitative or inhibitory effects on later colonizers (Connell and Slatyer  
1063 1977). In addition to direct interactions among decomposers coexisting at the same time and  
1064 place, the indirect priority effects among species across time are known to be of great importance  
1065 to the development of these communities as well as to their collective effects on decomposition.  
1066 In Sweden, for example, Weslien et al. (2011) showed that colonization of Norway spruce (*Picea*  
1067 *abies* L. Karst.) by two beetle species within one year of cutting influenced the occurrence of  
1068 another beetle species a decade later in a way that was mediated through a species of wood-  
1069 decaying fungi.

1070

1071 The examination of species traits can also benefit from the necrobiome concept by understanding  
1072 what traits mediate key functions or processes. Greater general understanding of species and  
1073 assemblage and food web responses to decomposition is necessary to improve the transferability  
1074 of findings across geographic regions with different sets of species, which is a source of variation  
1075 among applied research, such as forensics, using these ecological concepts (Tomberlin et al.  
1076 2012a). For example, dispersal is a key trait that determines species ability to search and  
1077 colonize different forms of necromass (Tomberlin et al. 2011) but has not been widely examined  
1078 among carrion-associated arthropods (Barton et al. 2013b). Additionally, the feeding traits of  
1079 species are closely related to specialization and arrival dynamics (Barton and Evans 2017), can  
1080 be associated with tissue quality (Ulyshen 2016), and drive competition and interactions among  
1081 insects and microbial species (Burkpile et al. 2006). These complex interactions form ecological  
1082 networks upon which ecosystem functions rest. There is a need for a unified ecological theory

1083 building upon the previous knowledge from plant and animal necromass across ecosystems to  
1084 more comprehensively understand decomposition ecology.

1085

## 1086 **IMPLICATIONS AND CONCLUSIONS**

1087

1088 The proposed necrobiome framework provides a general template to conceptualize the structure  
1089 and function of decomposer communities. A set of questions naturally arises from the  
1090 necrobiome framework and its spatial and temporal dimensions. For example, what is the  
1091 functional outcome for decomposition by adding or subtracting different biotic components of  
1092 the necrobiome? What is the effect of altered interactions in the presence or absence of these  
1093 biotic components and are the responses consistent among necromass types? Landscape  
1094 consequences extend from these localized questions, such as: How does altered necrobiome  
1095 structure and function lead to altered rates and pathways of genomic, element, and nutrient  
1096 movement through ecosystems? Manipulative experiments have shown that excluding insects  
1097 from carrion (Payne 1965, Pechal et al. 2014a) and plant matter (Heneghan et al. 1998, Wall et  
1098 al. 2008, Ulyshen 2014, Stoklosa et al. 2015) resulted in changes to necromass loss rates. But  
1099 what about experiments that change various functional components, or enhance fragmentation by  
1100 insects, or manipulate the internal or external microbial communities, such as excluding  
1101 detoxifying bacteria? Necromass varies along important gradients of nutrient quality, size, and  
1102 digestibility, and decomposition progresses as a function of the attendant necrobiome and  
1103 surrounding abiotic template. Clarity in our understanding of these controlling parameters means  
1104 that future research on decomposition can soon be standardized across resource types. Future  
1105 experimental studies that control for these parameters can then focus on manipulating key

1106 functional groups or interactions within the necrobiome to reveal the consequences for  
1107 decomposition, food webs and ecosystem services.

1108  
1109 The necrobiome concept and the further development of decomposition theory have implications  
1110 to disciplines ranging from ecosystem restoration to forensics. In restoration ecology, new  
1111 comparative work could examine the effects of manipulation of plant and animal necromass to  
1112 return key processes critical to decomposition and nutrient cycling and broader ecosystem-scale  
1113 functioning. Using the necrobiome framework, functional and biotic components can be  
1114 identified that enhance or slow nutrient cycling, change community network stability, generate  
1115 ecosystem heterogeneity, or add biodiversity per se. In the field of forensics, insects are the  
1116 primary source of evidence used by practitioners for death cases requiring legal action. Although  
1117 the dynamics of forensically important insects are rooted in ecological theory (e.g., community  
1118 assembly, succession, priority effects, metacommunities, perturbation impacts, biodiversity, and  
1119 patch dynamics) (Benbow et al. 2015a), there must be an expansion of this basic knowledge and  
1120 potential use to other constituents of the necrobiome (e.g., microbes). The necrobiome concept  
1121 provides a strong conceptual basis to further both theoretical and empirical endeavors.

1122

### 1123 **ACKNOWLEDGEMENTS**

1124 The Department of Entomology, AgBioResearch, the College of Agriculture and Natural  
1125 Resources and the College of Osteopathic Medicine at Michigan State University provided  
1126 funding to M.E.B. P.S.B. was funded by an Australian Research Council grant (DE150100026).  
1127 This work was partially funded by a grant from the National Institute of Justice, Office of Justice  
1128 Programs, U.S. Department of Justice awarded (2014-DN-BX-K008) to H.R.J., M.E.B and J.L.P.

1129 M.E.B. and P.S.B contributed equally to leading this paper, and P.S.B. can be contacted as a  
1130 corresponding author via email (philip.barton@anu.edu.au). Points of view in this document are  
1131 those of the authors and do not necessarily represent the official position or policies of the U.S.  
1132 Department of Justice.

1133

1134 **REFERENCES**

1135

1136 Addison, J. A. 2009. Distribution and impacts of invasive earthworms in Canadian forest  
1137 ecosystems. *Biological Invasions* **11**:59-79.

1138 Alldredge, A. L., and M. W. Silver. 1988. Characteristics, dynamics and significance of marine  
1139 snow. *Progress in Oceanography* **20**:41-82.

1140 Allen, M. L., L. M. Elbroch, C. C. Wilmers, and H. U. Wittmer. 2014. Trophic facilitation or  
1141 limitation? Comparative effects of pumas and black bears on the scavenger community.  
1142 *Plos One* **9**:e102257.

1143 Allison, P. A., C. R. Smith, H. Kukert, J. W. Deming, and B. A. Bennett. 1991. Deep-water  
1144 taphonomy of vertebrate carcasses: a whale skeleton in the bathyal Santa Catalina Basin.  
1145 *Paleobiology* **17**:78-89.

1146 Anderson, G. S. 2000. Insect succession on carrion and its relationship to determining time since  
1147 death. Pages 143-176 in J. H. Byrd and J. L. Castner, editors. *Forensic Entomology: The*  
1148 *Utility of Arthropods in Legal Investigations*. CRC Press, Boca Raton, Florida.

1149 Anderson, G. S., and L. S. Bell. 2016. Impact of marine submergence and season on faunal  
1150 colonization and decomposition of pig carcasses in the Salish Sea. *Plos One*  
1151 **11**:e0149107.

- 1152 Anderson, N. H., J. R. Sedell, L. M. Roberts, and F. J. Triska. 1978. The role of aquatic  
1153 invertebrates in processing of wood debris in coniferous forest streams. *American*  
1154 *Midland Naturalist* **100**:64-82.
- 1155 Arce, A., P. Johnston, P. Smiseth, and D. Rozen. 2012. Mechanisms and fitness effects of  
1156 antibacterial defences in a carrion beetle. *Journal of Evolutionary Biology* **25**:930-937.
- 1157 Arnold, A. E., Z. Maynard, G. S. Gilbert, P. D. Coley, and T. A. Kursar. 2000. Are tropical  
1158 fungal endophytes hyperdiverse? *Ecology Letters* **3**:267-274.
- 1159 Aumen, N. G. 1980. Microbial succession on a chitinous substrate in a woodland stream. *Microbial*  
1160 *Ecology* **6**:317-327.
- 1161 Ausmus, B. S. 1977. Regulation of wood decomposition rates by arthropod and annelid  
1162 populations. *Ecological bulletins (Stockholm)* **25**:180-192.
- 1163 Austin, A. T., L. Vivanco, A. Gonzalez-Arzac, and L. I. Perez. 2014. There's no place like home?  
1164 An exploration of the mechanisms behind plant litter- decomposer affinity in terrestrial  
1165 ecosystems. *New Phytologist* **204**:307-314.
- 1166 Averill, C., B. G. Waring, and C. V. Hawkes. 2016. Historical precipitation predictably alters the  
1167 shape and magnitude of microbial functional response to soil moisture. *Global Change*  
1168 *Biology* **22**:1957-1964.
- 1169 Ayres, E., H. Steltzer, B. L. Simmons, R. T. Simpson, J. M. Steinweg, M. D. Wallenstein, N.  
1170 Mellor, W. J. Parton, J. C. Moore, and D. H. Wall. 2009. Home-field advantage  
1171 accelerates leaf litter decomposition in forests. *Soil Biology and Biochemistry* **41**:606-  
1172 610.
- 1173 Bacheaga, L. R., J. P. Bouillet, M. D. C. Piccolo, L. Saint-Andre, J. M. Bouvet, Y. Nouvellon, J.  
1174 L. D. Goncalves, A. Robin, and J. P. Laclau. 2016. Decomposition of *Eucalyptus grandis*

1175 and *Acacia mangium* leaves and fine roots in tropical conditions did not meet the Home  
1176 Field Advantage hypothesis. *Forest Ecology and Management* **359**:33-43.

1177 Baldy, V., M. O. Gessner, and E. Chauvet. 1995. Bacteria, fungi and the breakdown of leaf litter in  
1178 a large river. *Oikos* **74**:93-102.

1179 Barthelmes, A., A. Prager, and H. Joosten. 2006. Palaeoecological analysis of *Alnus* wood peats  
1180 with special attention to non-pollen palynomorphs. *Review of Palaeobotany and*  
1181 *Palynology* **141**:33-51.

1182 Barton, P. S. 2015. The role of carrion in ecosystems. Page 512 in M. E. Benbow, J. K.  
1183 Tomberlin, and A. M. Tarone, editors. *Carrion Ecology, Evolution, and Their*  
1184 *Applications*. CRC Press, Boca Raton, Florida.

1185 Barton, P. S., S. A. Cunningham, D. B. Lindenmayer, and A. D. Manning. 2013a. The role of  
1186 carrion in maintaining biodiversity and ecological processes in terrestrial ecosystems.  
1187 *Oecologia* **171**:761-772.

1188 Barton, P. S., S. A. Cunningham, B. C. T. Macdonald, S. McIntyre, D. B. Lindenmayer, and A.  
1189 D. Manning. 2013b. Species traits predict assemblage dynamics at ephemeral resource  
1190 patches created by carrion. *Plos One* **8**:e53961

1191 Barton, P. S., and M. J. Evans. 2017. Insect biodiversity meets ecosystem function: differential  
1192 effects of habitat and insects on carrion mass loss. *Ecological Entomology* **42**:364-374

1193 Barton, P. S., M. J. Evans, J. L. Pechal, and M. E. Benbow. 2017. Necrophilous insect dynamics  
1194 at small vertebrate carrion in a temperate eucalypt woodland. *Journal of Medical*  
1195 *Entomology* **54**:964-973.

- 1196 Barton, P. S., S. McIntyre, M. J. Evans, J. K. Bump, S. A. Cunningham, and A. D. Manning.  
1197 2016. Substantial long-term effects of carcass addition on soil and plants in a grassy  
1198 eucalypt woodland. *Ecosphere* **7**:e01537
- 1199 Baumgartner, D. L. 1993. Review of *Chrysomya rufifacies* (Diptera: Calliphoridae). *Journal of*  
1200 *Medical Entomology* **30**:338-352.
- 1201 Beasley, J. C., Z. Olson, and T. DeVault. 2012. Carrion cycling in food webs: comparisons  
1202 among terrestrial and marine ecosystems. *Oikos* **121**:1021-1026.
- 1203 Beasley, J. C., Z. H. Olson, and T. L. DeVault. 2015. Ecological role of vertebrate scavengers.  
1204 Pages 107-127 in M. E. Benbow, J. K. Tomberlin, and A. M. Tarone, editors. *Carrion*  
1205 *Ecology, Evolution, and Their Applications*. CRC Press, Boca Raton, Florida.
- 1206 Belyea, L. R. 1996. Separating the effects of litter quality and microenvironment on  
1207 decomposition rates in a patterned peatland. *Oikos* **77**:529–539.
- 1208 Benbow, M. E., A. J. Lewis, J. K. Tomberlin, and J. L. Pechal. 2013. Seasonal necrophagous  
1209 insect community assembly during vertebrate carrion decomposition. *Journal of Medical*  
1210 *Entomology* **50**:440-450.
- 1211 Benbow, M. E., J. K. Tomberlin, and A. M. Tarone. 2015a. *Carrion Ecology, Evolution, and*  
1212 *Their Applications*. CRC Press, Boca Raton, Florida.
- 1213 Benbow, M. E., J. L. Pechal, and R. M. Mohr. 2015b. Community and landscape ecology of  
1214 carrion. Pages 151-185 in M. E. Benbow, J. K. Tomberlin, and A. M. Tarone, editors.  
1215 *Carrion Ecology, Evolution, and Their Applications*. CRC Press, Boca Raton, Florida.
- 1216 Benbow, M. E., J. K. Tomberlin, and A. M. Tarone. 2015c. Introduction to *Carrion Ecology,*  
1217 *Evolution, and Their Applications*. Pages 3-12 *Carrion Ecology, Evolution, and Their*  
1218 *Applications*. CRC Press, Boca Raton, Florida.



- 1219 Benke, A. C., C. A. S. Hall, C. P. Hawkins, R. H. Lowe-McConnell, J. A. Stanford, K.  
1220 Suberkropp, and J. V. Ward. 1988. Bioenergetic considerations in the analysis of stream  
1221 ecosystems. *Journal of the North American Benthological Society* **7**:480-502.
- 1222 Benke, A. C., and A. D. Huryn. 2006. Secondary production of macroinvertebrates. Pages 691-  
1223 710 in F. R. Hauer and G. A. Lamberti, editors. *Methods in stream ecology*. Academic  
1224 Press, New York.
- 1225 Benke, A. C., A. D. Huryn, L. A. Smock, and J. B. Wallace. 1999. Length-mass relationships for  
1226 freshwater macroinvertebrates in North America with particular reference to the  
1227 southeastern United States. *Journal of the North American Benthological Society* **18**:308-343.
- 1228 Bennett, B. A., C. R. Smith, B. Glaser, and H. L. Maybaum. 1994. Faunal community structure of a  
1229 chemoautotrophic assemblage on whale bones in the deep northeast Pacific Ocean. *Marine*  
1230 *Ecology Progress Series* **108**:205-223.
- 1231 Bharti, M., and D. Singh. 2003. Insect faunal succession on decaying rabbit carcasses in Punjab,  
1232 India. *Journal of Forensic Sciences* **48**:1-11.
- 1233 Bignell, D. E., and P. Eggleton. 2000. Termites in ecosystems. Pages 363-387 in T. Abe, D.  
1234 Bignell, and M. Higashi, editors. *Termites: Evolution, Sociality, Symbioses, Ecology*.  
1235 Kluwer Academic Publishers, Dordrecht.
- 1236 Bilby, R. E., B. R. Fransen, and P. A. Bisson. 1996. Incorporation of nitrogen and carbon from  
1237 spawning coho salmon into the trophic system of small streams: evidence from stable  
1238 isotopes. *Canadian Journal of Fisheries and Aquatic Sciences* **53**:164-173.
- 1239 Boddy, L. 1983. Carbon dioxide release from decomposing wood: effect of water content and  
1240 temperature. *Soil Biology and Biochemistry* **15**:501-510.

- 1241 Boddy, L. 2001. Fungal community ecology and wood decomposition processes in angiosperms:  
1242 from standing tree to complete decay of coarse woody debris. *Ecological Bulletins* **49**:43-  
1243 56.
- 1244 Boeskorov, G. G., O. R. Potapova, E. N. Mashchenko, A. V. Protopopov, T. V. Kuznetsova, L.  
1245 Agenbrood, and A. N. Tikhonov. 2014. Preliminary analyses of the frozen mummies of  
1246 mammoth (*Mammuthus primigenius*), bison (*Bison priscus*) and horse (*Equus* sp.) from  
1247 the Yana-Indigirka Lowland, Yakutia, Russia. *Integrative Zoology* **9**:471-480.
- 1248 Bornemissza, G. F. 1957. An analysis of arthropod succession in carrion and the effect of its  
1249 decomposition on the soil fauna. *Australian Journal of Zoology* **5**:1-12.
- 1250 Boysen-Jensen, P. 1919. Valuation of the Limfjord. I. Studies on the fish-food in the Limfjord  
1251 1909-1917, its quantity variation and annual production. *Reports of the Danish Biological*  
1252 *Station* **26**:3-44
- 1253 Braack, L. E. O. 1987. Community dynamics of carrion-attendant arthropods in tropical African  
1254 woodland. *Oecologia* **72**:402-409.
- 1255 Brabcová, V., M. Nováková, A. Davidová, and P. Baldrian. 2016. Dead fungal mycelium in  
1256 forest soil represents a decomposition hotspot and a habitat for a specific microbial  
1257 community. *New Phytologist* **210**:1369-1381.
- 1258 Bradford, M. A., B. Berg, D. S. Maynard, W. R. Wieder, and S. A. Wood. 2016. Understanding  
1259 the dominant controls on litter decomposition. *Journal of Ecology* **104**:229-238.
- 1260 Bradford, M. A., R. J. Warren, P. Baldrian, T. W. Crowther, D. S. Maynard, E. E. Oldfield, W.  
1261 R. Wieder, S. A. Wood, and J. R. King. 2014. Climate fails to predict wood  
1262 decomposition at regional scales. *Nature Climate Change* **4**:625-630.

- 1263 Brinson, M. M., A. E. Lugo, and S. Brown. 1981. Primary productivity, decomposition and  
1264 consumer activity in freshwater wetlands. *Annual Review of Ecology and Systematics*  
1265 **12**:123-161.
- 1266 Brothwell, D., H. Gill-Robinson, and W. Haglund. 2002. Taphonomic and forensic aspects of  
1267 bog bodies. Pages 119-131 in W. D. Haglund and M. H. Sorg, editors. *Advances in*  
1268 *Forensic Taphonomy. Method, Theory and Archaeological Perspectives*, London and  
1269 New York.
- 1270 Brundage, A. L., M. E. Benbow, and J. K. Tomberlin. 2014. Priority effects on the life-history  
1271 traits of two carrion blow fly (Diptera, Calliphoridae) species. *Ecological Entomology*  
1272 **39**:539-547.
- 1273 Brundage, A. L., T. L. Crippen, B. Singh, M. E. Benbow, W. Liu, A. M. Tarone, T. K. Wood, and J.  
1274 K. Tomberlin. 2017. Interkingdom cues by bacteria associated with conspecific and  
1275 heterospecific eggs of *Cochliomyia macellaria* and *Chrysomya rufifacies* (Diptera:  
1276 Calliphoridae) potentially govern succession on carrion. *Annals of the Entomological*  
1277 *Society of America* **110**:73-82.
- 1278 Brune, A. 2014. Symbiotic digestion of lignocellulose in termite guts. *Nature Reviews*  
1279 *Microbiology* **12**:168-180.
- 1280 Brune, A., and M. Ohkuma. 2011. Role of the termite gut microbiota in symbiotic digestion.  
1281 Pages 439–475 in D. E. Bignell, Y. Roisin, and N. Lo, editors. *Biology of Termites: A*  
1282 *Modern Synthesis*. Springer, New York.
- 1283 Buchkowski, R. W. 2016. Top-down consumptive and trait-mediated control do affect soil food  
1284 webs: It's time for a new model. *Soil Biology and Biochemistry* **102**:29-32.

- 1285 Bulgarelli, D., K. Schlaeppi, S. Spaepen, E. V. L. van Themaat, and P. Schulze-Lefert. 2013.  
1286 Structure and Functions of the Bacterial Microbiota of Plants. *Annual Review of Plant*  
1287 *Biology* **64**:807-838.
- 1288 Bultman, J. D., and C. R. Southwell. 1976. Natural resistance of tropical American woods to  
1289 terrestrial wood-destroying organisms. *Biotropica* **8**:71-95.
- 1290 Bump, J. K., R. O. Peterson, and J. A. Vucetich. 2009a. Wolves modulate soil nutrient  
1291 heterogeneity and foliar nitrogen by configuring the distribution of ungulate carcasses.  
1292 *Ecology* **90**:3159-3167.
- 1293 Bump, J. K., C. R. Webster, J. A. Vucetich, R. O. Peterson, J. M. Shields, and M. D. Powers.  
1294 2009b. Ungulate carcasses perforate ecological filters and create biogeochemical hotspots  
1295 in forest herbaceous layers allowing trees a competitive advantage. *Ecosystems* **12**:996-  
1296 1007.
- 1297 Burkepile, D. E., J. D. Parker, C. B. Woodson, H. J. Mills, J. Kubanek, P. A. Sobecky, and M. E.  
1298 Hay. 2006. Chemically mediated competition between microbes and animals: Microbes  
1299 as consumers in food webs. *Ecology* **87**:2821-2831.
- 1300 Burnes, T. A., R. A. Blanchette, and R. L. Farrell. 2000. Bacterial biodegradation of extractives  
1301 and patterns of bordered pit membrane attack in pine wood. *Applied and Environmental*  
1302 *Microbiology* **66**:5201-5205.
- 1303 Cammack, J. A., P. H. Adler, J. K. Tomberlin, Y. Arai, and W. C. Bridges, Jr. 2010. Influence of  
1304 parasitism and soil compaction on pupation of the green bottle fly, *Lucilia sericata*.  
1305 *Entomologia Experimentalis Et Applicata* **136**:134-141.

- 1306 Can, I., G. T. Javan, A. E. Pozhitkov, and P. A. Noble. 2014. Distinctive thanatomicrobiome  
1307 signatures found in the blood and internal organs of humans. *Journal of Microbiological*  
1308 *Methods* **106**:1-7.
- 1309 Carline, K. A., H. E. Jones, and R. D. Bardgett. 2005. Large herbivores affect the stoichiometry  
1310 of nutrients in a regenerating woodland ecosystem. *Oikos* **110**:453-460.
- 1311 Carter, D. O., J. L. Metcalf, A. Bibat, and R. Knight. 2015. Seasonal variation of postmortem  
1312 microbial communities. *Forensic Science, Medicine, and Pathology* **11**:202-207.
- 1313 Carter, D. O., D. Yellowlees, and M. Tibbett. 2007. Cadaver decomposition in terrestrial  
1314 ecosystems. *Naturwissenschaften* **94**:12-24.
- 1315 Cebrian, J. 1999. Patterns in the fate of production in plant communities. *The American*  
1316 *Naturalist* **154**:449-468.
- 1317 Chapman, H. 2015. The Landscape Archaeology of Bog Bodies. *Journal of Wetland*  
1318 *Archaeology* **15**:109-121.
- 1319 Chapman, R., and J. Sankey. 1955. The larger invertebrate fauna of three rabbit carcasses. *Journal*  
1320 *of Animal Ecology* **24**:395-402.
- 1321 Chouvinc, T., C. A. Efstathion, M. L. Elliott, and N.-Y. Su. 2013. Extended disease resistance  
1322 emerging from the faecal nest of a subterranean termite. *Proceedings of the Royal Society*  
1323 *B* **280**:20131885.
- 1324 Coe, M. 1978. The decomposition of elephant carcasses in the Tsavo (East) National Park,  
1325 Kenya. *Journal of Arid Environments* **1**:71-86.
- 1326 Connell, J. H., and R. O. Slatyer. 1977. Mechanisms of succession in natural communities and  
1327 their role in community stability and organization. *American Naturalist* **111**:1119-1144.

- 1328 Cornaby, B. W. 1974. Carrion reduction by animals in contrasting tropical habitats. *Biotropica*  
1329 **6**:51-63.
- 1330 Cornwell, W. K., J. H. C. Cornelissen, S. D. Allison, J. Bauhus, P. Eggleton, C. M. Preston, F.  
1331 A. Scarff, J. T. Weedon, C. Wirth, and A. E. Zanne. 2009. Plant traits and wood fates  
1332 across the globe: rotted, burned, or consumed? *Global Change Biology* **15**:2431-2449.
- 1333 Coûteaux, M.-M., L. Sarmiento, D. Hervé, and D. Acevedo. 2005. Determination of water-soluble  
1334 and total extractable polyphenolics in biomass, necromass and decomposing plant material  
1335 using near-infrared reflectance spectroscopy (NIRS). *Soil Biology and Biochemistry*  
1336 **37**:795-799.
- 1337 Covey, K. R., S. A. Wood, R. J. Warren, X. Lee, and M. A. Bradford. 2012. Elevated methane  
1338 concentrations in trees of an upland forest. *Geophysical Research Letters* **39**:L15705.
- 1339 Crippen, T. L., M. E. Benbow, and J. L. Pechal. 2015. Microbial interactions during carrion  
1340 decomposition. Pages 31-63 in M. E. Benbow, J. K. Tomberlin, and A. M. Tarone,  
1341 editors. *Carrion Ecology, Evolution, and Their Applications*. CRC Press, Boca Raton,  
1342 Florida.
- 1343 Crowther, T. W., S. M. Thomas, D. S. Maynard, P. Baldrian, K. Covey, S. D. Frey, L. T. A. van  
1344 Diepen, and M. A. Bradford. 2015. Biotic interactions mediate soil microbial feedbacks  
1345 to climate change. *Proceedings of the National Academy of Sciences of the United States*  
1346 *of America* **112**:7033-7038.
- 1347 Cummins, K. W., R. C. Petersen, F. O. Howard, J. C. Wuycheck, and V. I. Holt. 1973. The  
1348 utilization of leaf litter by stream detritivores. *Ecology* **54**:336-345.
- 1349 Danell, K., D. Berteaux, and K. A. Brathen. 2002. Effect of muskox carcasses on nitrogen  
1350 concentration in tundra vegetation. *Arctic* **55**:389-392.

- 1351 Dedouit, F., A. Géraut, V. Baranov, B. Ludes, D. Rougé, N. Telmon, and E. Crubézy. 2010.  
1352 Virtual and macroscopical studies of mummies—Differences or complementarity?  
1353 Report of a natural frozen Siberian mummy. *Forensic Science International* **200**:e7-e13.
- 1354 Denno, R., F., and W. R. Cothran. 1975. Niche relationships of a guild of necrophagous flies.  
1355 *Annals of the Entomological Society of America* **68**:741-753.
- 1356 DeVault, T. L., J. C. Beasley, Z. H. Olson, M. Moleón, M. Carrete, A. Margalida, and J. A.  
1357 Sánchez-Zapata. 2016. Ecosystem services provided by avian scavengers. Pages 235-270  
1358 in Ç. H. Sekercioglu, D. G. Wenny, and C. J. Whelan, editors. *Ecosystem Services*  
1359 *Provided by Birds*. The University of Chicago Press, Chicago and London.
- 1360 DeVault, T. L., J. Brisbin, I Lehr, and J. Rhodes, Olin E. 2004a. Factors influencing the  
1361 acquisition of rodent carrion by vertebrate scavengers and decomposers. *Canadian*  
1362 *Journal of Zoology* **82**:502-509.
- 1363 DeVault, T. L., Z. H. Olson, J. C. Beasley, and O. E. Rhodes. 2011. Mesopredators dominate  
1364 competition for carrion in an agricultural landscape. *Basic and Applied Ecology* **12**:268-  
1365 274.
- 1366 DeVault, T. L., B. D. Reinhart, I. L. Brisbin Jr, and O. E. Rhodes Jr. 2004b. Home ranges of  
1367 sympatric black and turkey vultures in South Carolina. *The Condor* **106**:706-711.
- 1368 DeVault, T. L., O. E. Rhodes Jr, and J. A. Shivik. 2003. Scavenging by vertebrates: behavioral,  
1369 ecological, and evolutionary perspectives on an important energy transfer pathway in  
1370 terrestrial ecosystems. *Oikos* **102**:225-234.
- 1371 Deveau, A., G. Bonito, J. Uehling, M. Paoletti, M. Becker, S. Bindschedler, S. Hacquard, V.  
1372 Hervé, J. Labbé, and O. Lastovetsky. 2018. Bacterial-fungal interactions: ecology,  
1373 mechanisms and challenges. *FEMS Microbiology Reviews* **42**:335-352.

- 1374 Dillon, R. J., and V. M. Dillon. 2004. The gut bacteria of insects: nonpathogenic interactions.  
1375 Annual Review of Entomology **49**:71-92.
- 1376 Donovan, S. E., P. Eggleton, and D. E. Bignell. 2001. Gut content analysis and a new feeding  
1377 group classification of termites. Ecological Entomology **26**:356-366.
- 1378 Dorado, J., F. W. Claassen, G. Lenon, T. A. van Beek, J. B. P. A. Wijnberg, and R. Sierra-  
1379 Alvarez. 2000. Degradation and detoxification of softwood extractives by sapstain fungi.  
1380 Bioresource Technology **71**:13-20.
- 1381 Dowd, P. F. 1992. Insect fungal symbionts: A promising source of detoxifying enzymes. Journal  
1382 of Industrial Microbiology **9**:149-161.
- 1383 Drazen, J. C., D. M. Bailey, H. A. Ruhl, and K. L. Smith Jr. 2012. The role of carrion supply in  
1384 the abundance of deep-water fish off California. Plos One **7**:e49332.
- 1385 Duarte, A., M. Welch, C. Swannack, J. Wagner, and R. M. Kilner. 2018. Strategies for managing  
1386 rival bacterial communities: lessons learned from burying beetles. Journal of Animal  
1387 Ecology **87**:424-427.
- 1388 Duggins, D., C. Simenstad, and J. Estes. 1989. Magnification of secondary production by kelp  
1389 detritus in coastal marine ecosystems. Science **245**:170-173.
- 1390 Dunne, J. A., R. J. Williams, and N. D. Martinez. 2002. Network structure and biodiversity loss  
1391 in food webs: robustness increases with connectance. Ecology Letters **5**:558-567.
- 1392 Early, M., and M. L. Goff. 1986. Arthropod succession patterns in exposed carrion on the island  
1393 of O'ahu, Hawaiian Islands, USA. Journal of Medical Entomology **23**:520-531.
- 1394 Ekblad, A., H. Wallander, D. Godbold, C. Cruz, D. Johnson, P. Baldrian, R. Björk, D. Epron, B.  
1395 Kieliszewska-Rokicka, and R. Kjøller. 2013. The production and turnover of



1396 extramatrical mycelium of ectomycorrhizal fungi in forest soils: role in carbon cycling.  
1397 Plant and Soil **366**:1-27.

1398 Eklöf, A., U. Jacob, J. Kopp, J. Bosch, R. Castro-Urgal, N. P. Chacoff, B. Dalsgaard, C. Sassi,  
1399 M. Galetti, and P. R. Guimarães. 2013. The dimensionality of ecological networks.  
1400 Ecology Letters **16**:577-583.

1401 Elton, C. S. 1966. The pattern of animal communities. Methuen and Co. Ltd., London.

1402 Ettema, C. H., and D. A. Wardle. 2002. Spatial soil ecology. Trends in Ecology and Evolution  
1403 **17**:177-183.

1404 Ezenwa, V. O., N. M. Gerardo, D. W. Inouye, M. Medina, and J. B. Xavier. 2012. Animal behavior  
1405 and the microbiome. Science **338**:198-199.

1406 Fahrig, L. 2017. Ecological responses to habitat fragmentation per se. Annual Review of  
1407 Ecology, Evolution, and Systematics **48**:1-23.

1408 Fain, A., and G. S. Ide. 1976. *Ellipsopus ornatus*, a new genus and species of Acaridae (Acari)  
1409 phoretic on the beetle *Bolitotherus cornutus* (Panzer, 1794). Entomological News **87**:233-  
1410 236.

1411 Farrell, B. D., A. S. Sequeira, B. C. O'Meara, B. B. Normark, J. H. Chung, and B. H. Jordal.  
1412 2001. The Evolution of agriculture in beetles (Curculionidae: Scolytinae and  
1413 Platypodinae). Evolution **55**:2011-2027.

1414 Farwig, N., R. Brandl, S. Siemann, F. Wiener, and J. Mueller. 2014. Decomposition rate of  
1415 carrion is dependent on composition not abundance of the assemblages of insect  
1416 scavengers. Oecologia **175**:1291-1300.

1417 Fellers, G. M., and J. H. Fellers. 1982. Scavenging rates of invertebrates in an eastern deciduous  
1418 forest. The American Midland Naturalist **107**:389-392.

- 1419 Fernandez, C. W., and P. G. Kennedy. 2018. Melanization of mycorrhizal fungal necromass  
1420 structures microbial decomposer communities. *Journal of Ecology* **106**:468-479.
- 1421 Fernandez, C. W., J. A. Langley, S. Chapman, M. L. McCormack, and R. T. Koide. 2016. The  
1422 decomposition of ectomycorrhizal fungal necromass. *Soil Biology and Biochemistry*  
1423 **93**:38-49.
- 1424 Ferro, M. L., M. L. Gimmel, K. E. Harms, and C. E. Carlton. 2012. Comparison of the  
1425 Coleoptera communities in leaf litter and rotten wood in Great Smoky Mountains  
1426 National Park, USA. *Insecta Mundi* **259**:1-58.
- 1427 Fey, S. B., A. M. Siepielski, S. Nusslé, K. Cervantes-Yoshida, J. L. Hwan, E. R. Huber, M. J.  
1428 Fey, A. Catenazzi, and S. M. Carlson. 2015. Recent shifts in the occurrence, cause, and  
1429 magnitude of animal mass mortality events. *Proceedings of the National Academy of*  
1430 *Sciences* **112**:1083-1088.
- 1431 Fiene, J. G., G. A. Sword, S. L. Vanlaerhoven, and A. M. Tarone. 2014. The Role of Spatial  
1432 Aggregation in Forensic Entomology. *Journal of Medical Entomology* **51**:1-9.
- 1433 Fierer, N., M. Strickland, D. Liptzin, M. Bradford, and C. Cleveland. 2009. Global patterns in  
1434 belowground communities. *Ecology Letters* **103**:1238–1249.
- 1435 Finley, S. J., J. L. Pechal, M. E. Benbow, B. K. Robertson, and G. T. Javan. 2016. Microbial  
1436 Signatures of Cadaver Gravesoil During Decomposition. *Microbial Ecology*:1-6.
- 1437 Finn, J. A. 2001. Ephemeral resource patches as model systems for diversity-function  
1438 experiments. *Oikos* **92**:363-366.
- 1439 Fittkau, E. J., and H. Klinge. 1973. On biomass and trophic structure of the central Amazonian  
1440 rain forest ecosystem. *Biotropica* **5**:2-14.

- 1441 Flores, M., M. Longnecker, and J. K. Tomberlin. 2014. Effects of temperature and tissue type on  
1442 *Chrysomya rufifacies* (Diptera: Calliphoridae) (Macquart) development. *Forensic Science*  
1443 *International* **245**:24-29.
- 1444 Forbes, S. A. 1887. The lake as a microcosm. Reprinted in *Bulletin of the Illinois State Natural*  
1445 *History Survey* (1925) **15**:537-550.
- 1446 Forbes, S. L., and K. A. Perrault. 2014. Decomposition odour profiling in the air and soil  
1447 surrounding vertebrate carrion. *Plos One* **9**:e95107.
- 1448 Frank, D. A., and R. D. Evans. 1997. Effects of native grazers on grassland N cycling in  
1449 Yellowstone National Park. *Ecology* **78**:2238-2248.
- 1450 Frankland, J. C. 1966. Succession of fungi on decaying petioles of *Pteridium aquilinum*. *The*  
1451 *Journal of Ecology* **54**:41-63.
- 1452 Fuglei, E., N. A. Øritsland, and P. Prestrud. 2003. Local variation in arctic fox abundance on  
1453 Svalbard, Norway. *Polar Biology* **26**:93-98.
- 1454 Fukami, K., U. Simidu, and N. Taga. 1985. Microbial decomposition of phyto- and zooplankton in  
1455 seawater. II. Changes in the bacterial community. *Marine Ecology Progress Series* **21**:7-13.
- 1456 Fuller, M. E. 1934. The insect inhabitants of carrion: a study in animal ecology. *CSIR Bulletin*  
1457 **82**:5-62.
- 1458 García-Palacios, P., F. T. Maestre, J. Kattge, and D. H. Wall. 2013. Climate and litter quality  
1459 differently modulate the effects of soil fauna on litter decomposition across biomes.  
1460 *Ecology Letters* **16**:1045-1053.
- 1461 Gasaway, W., K. Mossestad, and P. Standers. 1991. Food acquisition by spotted hyaenas in  
1462 Etosha National Park, Namibia: predation versus scavenging. *African Journal of Ecology*  
1463 **29**:64-75.

- 1464 Gessner, M. O., and E. Chauvet. 1994. Importance of stream microfungi in controlling breakdown  
1465 rates of leaf litter. *Ecology* **75**:1807-1817.
- 1466 Gessner, M. O., C. M. Swan, C. K. Dang, B. G. McKie, R. D. Bardgett, D. H. Wall, and S.  
1467 Hattenschwiler. 2010. Diversity meets decomposition. *Trends in Ecology and Evolution*  
1468 **25**:372-380.
- 1469 Gholz, H. L., D. A. Wedin, S. M. Smitherman, M. E. Harmon, and W. J. Parton. 2000. Long-  
1470 term dynamics of pine and hardwood litter in contrasting environments: toward a global  
1471 model of decomposition. *Global Change Biology* **6**:751-765.
- 1472 Goodbrod, J. R., and M. L. Goff. 1990. Effects of larval population-density on rates of  
1473 development and interactions between two species of *Chrysomya* (Diptera, Calliphoridae)  
1474 in laboratory culture. *Journal of Medical Entomology* **27**:338-343.
- 1475 Gravel, D., F. Guichard, M. Loreau, and N. Mouquet. 2010. Source and sink dynamics in meta-  
1476 ecosystems. *Ecology* **91**: 2172-2184.
- 1477 Gray, C. M., R. K. Monson, and N. Fierer. 2010. Emissions of volatile organic compounds  
1478 during the decomposition of plant litter. *Journal of Geophysical Research* **115**:G03015.
- 1479 Greenberg, B. 1971a. *Flies and Disease, Vol. 1, Ecology, Classification and Biotic Associations*.  
1480 Princeton University Press, Princeton, NJ.
- 1481 Greenberg, B. 1971b. *Flies and Disease, Vol. 2, Biology and Disease Transmission*. Princeton  
1482 University Press, Princeton, NJ.
- 1483 Greenberg, B. 1973. *Biology and Disease Transmission*. Princeton University Press, Princeton,  
1484 New Jersey.
- 1485 Greenberg, B., and A. A. Bornsetein. 1964. Fly dispersion from a rural Mexican slaughterhouse.  
1486 *The American Journal of Tropical Medicine and Hygiene* **13**:881-886.

- 1487 Greenberg, B., J. A. Kowalski, and M. J. Klowden. 1970. Factors affecting the transmission of  
1488 salmonella by flies: natural resistance to colonization and bacterial interference. *Infection*  
1489 and *Immunity* **2**:800-809.
- 1490 Grossart, H.-P., and M. Simon. 1998. Bacterial colonization and microbial decomposition of  
1491 limnetic organic aggregates (lake snow). *Aquatic Microbial Ecology* **15**:127-140.
- 1492 Haack, R. A., and R. C. Wilkinson. 1986. Phoresy by *Dendrochernes* pseudoscorpions on  
1493 Cerambycidae (Coleoptera) and Aulacidae (Hymenoptera) in Florida. *The American*  
1494 *Midland Naturalist* **117**:369-373.
- 1495 Haila, Y., and J. Niemelä. 1999. Leaf litter and the small-scale distribution of carabid beetles  
1496 (Coleoptera, Carabidae) in the boreal forest. *Ecography* **22**:424-435.
- 1497 Hammond, H. E. J., D. W. Langor, and J. R. Spence. 2001. Early colonization of *Populus* wood  
1498 by saproxylic beetles (Coleoptera). *Canadian Journal of Forest Research* **31**:1175-1183.
- 1499 Handa, I. T., R. Aerts, F. Berendse, M. P. Berg, A. Bruder, O. Butenschoen, E. Chauvet, M. O.  
1500 Gessner, J. Jabiol, M. Makkonen, B. G. McKie, B. Malmqvist, E. T. H. M. Peeters, S.  
1501 Scheu, B. Schmid, J. van Ruijven, V. C. A. Vos, and S. Haettenschwiler. 2014.  
1502 Consequences of biodiversity loss for litter decomposition across biomes. *Nature*  
1503 **509**:218-+.
- 1504 Hanski, I. 1987. Carrion fly community dynamics: patchiness, seasonality and coexistence.  
1505 *Ecological Entomology* **12**:257-266.
- 1506 Hanski, I., and Y. Cambefort, editors. 1991. *Dung Beetle Ecology*. Princeton University Press,  
1507 Princeton, NJ, USA.
- 1508 Hanski, I., and M. Gilpin. 1991. Metapopulation dynamics - Brief history and conceptual domain.  
1509 *Biological Journal of the Linnean Society* **42**:3-16.

1510 Harmon, M. E., J. F. Franklin, F. J. Swanson, P. Sollins, S. V. Gregory, J. D. Lattin, N. H.  
1511 Anderson, S. P. Cline, N. G. Aumen, J. R. Sedell, G. W. Lienkaemper, J. Cromack, K. ,  
1512 and K. W. Cummins. 1986. Ecology of coarse woody debris in temperate ecosystems.  
1513 *Advances in Ecological Research* **15**:133-302.

1514 Hattenschwiler, S., A. V. Tiunov, and S. Scheu. 2005. Biodiversity and litter decomposition in  
1515 terrestrial ecosystems. *Annual Review of Ecology Evolution and Systematics* **36**:191-  
1516 218.

1517 Hawlena, D., M. S. Strickland, M. A. Bradford, and O. J. Schmitz. 2012. Fear of predation slows  
1518 plant-litter decomposition. *Science* **336**:1434-1438.

1519 Hawlena, D., and M. Zaguri. 2016. Fear and below-ground food-webs. *Soil Biology and*  
1520 *Biochemistry* **102**:26-28.

1521 Heard, S. B. 1998. Resource patch density and larval aggregation in mushroom-breeding flies.  
1522 *Oikos* **81**:187-195.

1523 Heneghan, L., D. Coleman, X. Zou, D. Crossley Jr, and B. Haines. 1998. Soil microarthropod  
1524 community structure and litter decomposition dynamics: a study of tropical and temperate  
1525 sites. *Applied Soil Ecology* **9**:33-38.

1526 Hickin, N. E. 1963. The insect factor in wood decay. An account of wood-boring insects with  
1527 particular reference to timber indoors. London, Hutchinson & Co., Ltd., London.

1528 Hill, J. E., T. L. DeVault, J. C. Beasley, J. Rhodes, O.E., and J. L. Belant. 2018. Effects of  
1529 vulture exclusion on carrion consumption by facultative scavengers. *Ecology and*  
1530 *Evolution* **8**:2518-2526.

- 1531 Hobischak, N. R., S. L. VanLaerhoven, and G. S. Anderson. 2006. Successional patterns of  
1532 diversity in insect fauna on carrion in sun and shade in the Boreal Forest Region of  
1533 Canada, near Edmonton, Alberta. *Canadian Entomologist* **138**:376-383.
- 1534 Hocking, M. D., and J. D. Reynolds. 2011. Impacts of salmon on riparian plant diversity. *Science*  
1535 **331**:1609-1612.
- 1536 Hodgkin, L. K., M. A. Elgar, and M. R. E. Symonds. 2010. Positive and negative effects of  
1537 phoretic mites on the reproductive output of an invasive bark beetle. *Australian Journal of*  
1538 *Zoology* **58**:198-204.
- 1539 Holland, A. E., M. E. Byrne, A. L. Bryan, T. L. DeVault, O. E. Rhodes, and J. C. Beasley. 2017.  
1540 Fine-scale assessment of home ranges and activity patterns for resident black vultures  
1541 (*Coragyps atratus*) and turkey vultures (*Cathartes aura*). *Plos One* **12**:e0179819.
- 1542 Holter, P., and C. H. Scholtz. 2007. What do dung beetles eat? *Ecological Entomology* **32**:690-  
1543 697.
- 1544 Houston, D. 1974. Food searching in griffon vultures. *African Journal of Ecology* **12**:63-77.
- 1545 Houston, D. C. 1979. The adaptations of scavengers. Pages 236-286 in N. Sinclair and N.  
1546 Griffiths, editors. *Serengeti: Dynamics of an ecosystem*. The University of Chicago  
1547 Press, Chicago.
- 1548 Houston, D. C. 1985. Evolutionary ecology of Afrotropical and Neotropical vultures in forests.  
1549 *Ornithological Monographs* **36**:856-864.
- 1550 Houston, D. C. 1986. Scavenging efficiency of turkey vultures in tropical forest. *Condor* **88**:318-  
1551 323.

- 1552 Houston, D. C., and J. Cooper. 1975. The digestive tract of the whiteback griffon vulture and its  
1553 role in disease transmission among wild ungulates. *Journal of Wildlife Diseases* **11**:306-  
1554 313.
- 1555 Hungate, R. E. 1966. *The Rumen and its Microbes*. Academic Press, New York.
- 1556 Huryn, A. D., and J. B. Wallace. 2000. Life history and production of stream insects. *Annual*  
1557 *Review of Entomology* **45**:83-110.
- 1558 Hyde, E., D. Haarmann, J. Petrosino, A. Lynne, and S. Bucheli. 2015. Initial insights into  
1559 bacterial succession during human decomposition. *International Journal of Legal*  
1560 *Medicine* **129**:661-671.
- 1561 Hynes, H. B. N. 1970. *The Ecology of Running Waters*. Liverpool University Press, Liverpool.
- 1562 Inglis, G. 1989. The colonisation and degradation of stranded *Macrocystis pyrifera* (L.) C. Ag.  
1563 by the macrofauna of a New Zealand sandy beach. *Journal of Experimental Marine*  
1564 *Biology and Ecology* **125**:203-217.
- 1565 Ives, A. R. 1991. Aggregation and coexistence in a carrion fly community. *Ecological*  
1566 *Monographs* **61**:75-94.
- 1567 Janzen, D. H. 1977. Why fruits rot, seeds mold, and meat spoils. *American Naturalist* **111**:691-  
1568 713.
- 1569 Javan, G. T., S. J. Finley, I. Can, J. E. Wilkinson, J. D. Hanson, and A. M. Tarone. 2016. Human  
1570 thanatobiome succession and time since death. *Scientific Reports* **6**:29598.
- 1571 Jennelle, C. S., M. D. Samuel, C. A. Nolden, and E. A. Berkley. 2009. Deer carcass  
1572 decomposition and potential scavenger exposure to chronic wasting disease. *Journal of*  
1573 *Wildlife Management* **73**:655-662.



- 1574 Jiménez, M. A., R. Beltran, A. Traveset, M. L. Calleja, A. Delgado-Huertas, and N. Marbà.  
1575 2017. Aeolian transport of seagrass (*Posidonia oceanica*) beach-cast to terrestrial  
1576 systems. *Estuarine, Coastal and Shelf Science* **196**:31-44.
- 1577 Jones, E. G., M. A. Collins, P. M. Bagley, S. Addison, and I. G. Priede. 1998. The fate of cetacean  
1578 carcasses in the deep sea: observations on consumption rates and succession of scavenging  
1579 species in the abyssal north-east Atlantic Ocean. *Proceedings of the Royal Society of*  
1580 *London B: Biological Sciences* **265**:1119-1127.
- 1581 Jones, S. C., E. D. Strauss, and K. E. Holekamp. 2015. Ecology of African carrion. Pages 461-  
1582 491 in M. E. Benbow, J. K. Tomberlin, and A. M. Tarone, editors. *Carrion Ecology,*  
1583 *Evolution and Their Applications.* CRC Press, Boca Raton, Florida.
- 1584 Jordan, H. R., J. K. Tomberlin, T. K. Wood, and M. E. Benbow. 2015. Interkingdom ecological  
1585 interactions of carrion decomposition. Pages 433-460 in M. E. Benbow, J. K. Tomberlin,  
1586 and A. M. Tarone, editors. *Carrion Ecology, Evolution and Their Applications.* CRC  
1587 Press, Boca Raton, Florida.
- 1588 Joyce, P., L. L. Warren, and R. S. Wotton. 2007. Faecal pellets in streams: their binding,  
1589 breakdown and utilization. *Freshwater Biology* **52**:1868-1880.
- 1590 Junker, R. R., and A. Keller. 2015. Microhabitat heterogeneity across leaves and flower organs  
1591 promotes bacterial diversity. *FEMS Microbiology Ecology* **91**:fiv097.
- 1592 Juottonen, H., P. E. Galand, E. Tuittila, J. Laine, H. Fritze, and K. Yrjaelae. 2005. Methanogen  
1593 communities and bacteria along an ecohydrological gradient in a northern raised bog  
1594 complex. *Environmental Microbiology* **7**:1547-1557.
- 1595 Käärik, A. A. 1974. Decomposition of wood. Pages 129-174 in C. H. Dickinson and G. J. F.  
1596 Pugh, editors. *Biology of plant litter decomposition Volume 1.* Academic Press, London.

- 1597 Kallenbach, C. M., A. S. Grandy, S. D. Frey, and A. F. Diefendorf. 2015. Microbial physiology  
1598 and necromass regulate agricultural soil carbon accumulation. *Soil Biology and*  
1599 *Biochemistry* **91**:279-290.
- 1600 Kaltenpoth, M., and S. Steiger. 2014. Unearthing carrion beetles' microbiome: Characterization  
1601 of bacterial and fungal hindgut communities across the Silphidae. *Molecular Ecology*  
1602 **23**:1251-1267.
- 1603 Karl, D. M., G. A. Knauer, and J. H. Martin. 1988. Downward flux of particulate organic matter in  
1604 the ocean: a particle decomposition paradox. *Nature* **332**:438.
- 1605 Keiser, A. D., D. A. Keiser, M. S. Strickland, and M. A. Bradford. 2014. Disentangling the  
1606 mechanisms underlying functional differences among decomposer communities. *Journal*  
1607 *of Ecology* **102**:603-609.
- 1608 Keiser, A. D., J. D. Knoepp, and M. A. Bradford. 2013. Microbial communities may modify how  
1609 litter quality affects potential decomposition rates as tree species migrate. *Plant and Soil*  
1610 **372**:167-176.
- 1611 Kelly, N. E., D. W. Sparks, T. L. DeVault, and O. E. Rhodes Jr. 2007. Diet of black and turkey  
1612 vultures in a forested landscape. *Wilson Journal of Ornithology* **119**:267-270.
- 1613 Kim, J., and W. Park. 2015. Indole: a signaling molecule or a mere metabolic byproduct that alters  
1614 bacterial physiology at a high concentration? *Journal of Microbiology* **53**:421-428.
- 1615 Knorr, K. H., M. A. Horn, and W. Borren. 2015. Significant nonsymbiotic nitrogen fixation in  
1616 Patagonian ombrotrophic bogs. *Global Change Biology* **21**:2357-2365.
- 1617 Kouki, J., and I. Hanski. 1995. Population aggregation facilitates coexistence of many  
1618 competing carrion fly species. *Oikos* **72**:223-227.

- 1619 Krantz, G.W. 1998. Reflections on the biology, morphology and ecology of the  
1620 Macrochelidae. *Experimental & Applied Acarology* **22**: 125-137.
- 1621 Kuiters, A., and H. Sarink. 1986. Leaching of phenolic compounds from leaf and needle litter of  
1622 several deciduous and coniferous trees. *Soil Biology and Biochemistry* **18**:475-480.
- 1623 Kulshrestha, P., and D. K. Satpathy. 2001. Use of beetles in forensic entomology. *Forensic*  
1624 *Science International* **120**:15-17.
- 1625 Kvavadze, E., and K. Kakhiani. 2010. Palynology of the Paravani burial mound (Early Bronze  
1626 Age, Georgia). *Vegetation History and Archaeobotany* **19**:469-478.
- 1627 Latter, P. M., and J. Cragg. 1967. The decomposition of *Juncus squarrosus* leaves and  
1628 microbiological changes in the profile of *Juncus* moor. *The Journal of Ecology* **55**:465-482.
- 1629 Lavelle, P., D. Bignell, M. Lepage, V. Wolters, P. Roger, P. Ineson, O. W. Heal, and S.  
1630 Dhillon. 1997. Soil function in a changing world: the role of invertebrate ecosystem  
1631 engineers. *European Journal of Soil Biology* **33**:159-193.
- 1632 Leach, J. G., L. W. Orr, and C. Christensen. 1937. Further studies on the interrelationship of  
1633 insects and fungi in the deterioration of felled Norway pine logs. *Journal of Agricultural*  
1634 *Research* **55**:129-140.
- 1635 Leff, J. W., and N. Fierer. 2008. Volatile organic compound (VOC) emissions from soil and litter  
1636 samples. *Soil Biology and Biochemistry* **40**:1629-1636.
- 1637 Lewis, A. J., and M. E. Benbow. 2011. When entomological evidence crawls away: *Phormia*  
1638 *regina* en masse larval dispersal. *Journal of Medical Entomology* **48**:1112-1119.
- 1639 Leroux, S. J., and M. Loreau. 2008. Subsidy hypothesis and strength of trophic cascades across  
1640 ecosystems. *Ecology Letters* **11**:1147-1156.

- 1641 Lieutier, F., K. R. Day, A. Battisti, J.-C. Gregoire, and H. F. Evans. 2004. Editors. Bark and  
1642 Wood Boring Insects in Living Trees in Europe: A Synthesis. Springer, Dordrecht.
- 1643 Lindeman, R. L. 1942. The trophic-dynamic aspect of ecology. *Ecology* **23**:399-418.
- 1644 Lindquist, E. E. 1969. Review of holarctic tarsonemid mites (Acari: Prostigmata) parasitizing  
1645 eggs of ipine bark beetles. *Memoirs of the Entomological Society of Canada* **60**:5-111.
- 1646 Liu, G., W. K. Cornwell, K. Cao, Y. Hu, R. S. P. Van Logtestijn, S. Yang, X. Xie, Y. Zhang, D.  
1647 Ye, X. Pan, X. Ye, Z. Huang, M. Dong, and J. H. C. Cornelissen. 2015. Termites amplify  
1648 the effects of wood traits on decomposition rates among multiple bamboo and dicot  
1649 woody species. *Journal of Ecology* **103**:1214-1223.
- 1650 Lo, N., G. Tokuda, and H. Watanabe. 2011. Evolution and function of endogenous termite  
1651 cellulases. Pages 51-67 in D. E. Bignell, Y. Roisin, and N. Lo, editors. *Biology of*  
1652 *Termites: A Modern Synthesis*. Springer, New York.
- 1653 Lomstein, B. A., A. T. Langerhuus, S. D'Hondt, B. B. Jørgensen, and A. J. Spivack. 2012.  
1654 Endospore abundance, microbial growth and necromass turnover in deep sub-  
1655 seafloor sediment. *Nature* **484**:101.
- 1656 Lussenhop, J. 1992. Mechanisms of microarthropod-microbial interactions in soil. *Advances in*  
1657 *Ecological Research* **23**:1-33.
- 1658 Luysaert, S., E.-S. Schulze, A. Börner, A. Knohl, D. Hessenmöller, B. E. Law, P. Ciais, and J.  
1659 Grace. 2008. Old-growth forests as global carbon sinks. *Nature* **455**:213-215.
- 1660 Ma, Q., A. Fonseca, W. Liu, A. T. Fields, M. L. Pimsler, A. F. Spindola, A. M. Tarone, T. L.  
1661 Crippen, J. K. Tomberlin, and T. K. Wood. 2012. *Proteus mirabilis* interkingdom  
1662 swarming signals attract blow flies. *The ISME Journal* **6**:1356-1366.

- 1663 MacArthur, R. H., and E. O. Wilson. 1967. Theory of Island Biogeography. Princeton University  
1664 Press.
- 1665 Macdonald, J. 1992. The decomposition of animal remains in caves. MS Thesis. Nottingham  
1666 Polytechnic, Nottingham, England.
- 1667 Macovei, L., and L. Zurek. 2006. Ecology of antibiotic resistance genes: characterization of  
1668 enterococci from houseflies collected in food settings. Applied and Environmental  
1669 Microbiology **72**:4028-4035.
- 1670 Mądra, A., K. Frątczak, A. Grzywacz, and S. Matuszewski. 2015. Long-term study of pig carrion  
1671 entomofauna. Forensic Science International **252**:1-10.
- 1672 Madritch, M. D., J. R. Donaldson, and R. L. Lindroth. 2007. Canopy herbivory can mediate the  
1673 influence of plant genotype on soil processes through frass deposition. Soil Biology and  
1674 Biochemistry **39**:1192-1201.
- 1675 Mann, K. 1988. Production and use of detritus in various freshwater, estuarine, and coastal marine  
1676 ecosystems. Limnology and Oceanography **33**:910-930.
- 1677 Mann, R. W., W. M. Bass, and L. Meadows. 1990. Time since death and decomposition of the  
1678 human body: variables and observations in case and experimental field studies. Journal of  
1679 Forensic Science **35**:103-111.
- 1680 Margalida, A., J. Bertran, and R. Heredia. 2009. Diet and food preferences of the endangered  
1681 Bearded Vulture *Gypaetus barbatus*: a basis for their conservation. Ibis **151**:235-243.
- 1682 Martin, A. R., D. L. Erickson, W. J. Kress, and S. C. Thomas. 2014. Wood nitrogen  
1683 concentrations in tropical trees: phylogenetic patterns and ecological correlates. New  
1684 Phytologist **204**:484-495.

- 1685 Martiny, J. B. H., A. C. Martiny, C. Weihe, Y. Lu, R. Berlemont, E. L. Brodie, M. L. Goulden,  
1686 K. K. Treseder, and S. D. Allison. 2017. Microbial legacies alter decomposition in  
1687 response to simulated global change. *The ISME Journal* **11**:490-499
- 1688 Matuszewski, S., D. Bajerlein, S. Konwerski, and K. Szpila. 2008. An initial study of insect  
1689 succession and carrion decomposition in various forest habitats of Central Europe.  
1690 *Forensic Science International* **180**:61-69.
- 1691 Matuszewski, S., D. Bajerlein, S. Konwerski, and K. Szpila. 2011. Insect succession and carrion  
1692 decomposition in selected forests of Central Europe. Part 3: Succession of carrion fauna.  
1693 *Forensic Science International* **2007**:150-163.
- 1694 Mégnin, P. 1894. La faune des cadavres application de l'entomologie à la médecine légale.  
1695 Encyclopédie Scientifique des Aide-Mémoire, Paris.
- 1696 Menendez, R., P. Webb, and K. H. Orwin. 2016. Complementarity of dung beetle species with  
1697 different functional behaviours influence dung-soil carbon cycling. *Soil Biology and*  
1698 *Biochemistry* **92**:142-148.
- 1699 Merritt, R. W., and G. D. De Jong. 2015. Arthropod communities in terrestrial environments.  
1700 Pages 65-92 in M. E. Benbow, J. K. Tomberlin, and A. M. Tarone, editors. *Carrion*  
1701 *Ecology, Evolution, and Their Applications*. CRC Press, Boca Raton, Florida
- 1702 Merritt, R. W., and J. R. Wallace. 2010. The Role of Aquatic Insects in Forensic Investigations.  
1703 Pages 271-320 in J. H. Byrd and J. L. Castner, editors. *Forensic Entomology: The Utility*  
1704 *of Arthropods in Legal Investigations*. CRC Press, Boca Raton, FL.
- 1705 Metcalf, J. L., Z. Z. Xu, S. Weiss, S. Lax, W. Van Treuren, E. R. Hyde, S. J. Song, A. Amir, P.  
1706 Larsen, N. Sangwan, D. Haarmann, G. C. Humphrey, G. Ackermann, L. R. Thompson,  
1707 C. Lauber, A. Bibat, C. Nicholas, M. J. Gebert, J. F. Petrosino, S. C. Reed, J. A. Gilbert,

- 1708 A. M. Lynne, S. R. Bucheli, D. O. Carter, and R. Knight. 2016. Microbial community  
1709 assembly and metabolic function during mammalian corpse decomposition. *Science*  
1710 **351**:158-162.
- 1711 Michaud, J. P., K. G. Schoenly, and G. Moreau. 2015. Rewriting ecological succession history:  
1712 Did carrion ecologists get there first? *Quarterly Review of Biology* **90**:45-66.
- 1713 Middeldorp, A. A. 1986. Functional palaeoecology of the Hahnenmoor raised bog ecosystem —  
1714 A study of vegetation history, production and decomposition by means of pollen density  
1715 dating. *Review of Palaeobotany and Palynology* **49**:1-73.
- 1716 Mishima, T., N. Wada, R. Iwata, H. Anzai, T. Hosoya, and K. Araya. 2016. Super-Protective  
1717 Child-Rearing by Japanese Bess Beetles, *Cylindrocaulus patalis*: Adults Provide Their  
1718 Larvae with Chewed and Predigested Wood. *Insects* **7**:18.
- 1719 Moleón, M., J. A. Sanchez-Zapata, N. Selva, J. A. Donazar, and N. Owen-Smith. 2014. Inter-  
1720 specific interactions linking predation and scavenging in terrestrial vertebrate  
1721 assemblages. *Biological Reviews* **89**:1042-1054.
- 1722 Moleón, M., J. A. Sánchez-Zapata, E. Sebastián-González, and N. Owen-Smith. 2015. Carcass  
1723 size shapes the structure and functioning of an African scavenging assemblage. *Oikos*  
1724 **124**:1391-1403.
- 1725 Moore, J. C., E. L. Berlow, D. C. Coleman, P. C. Ruitter, Q. Dong, A. Hastings, N. C. Johnson, K.  
1726 S. McCann, K. Melville, and P. J. Morin. 2004. Detritus, trophic dynamics and biodiversity.  
1727 *Ecology Letters* **7**:584-600.
- 1728 Moreau, G., J.-P. Michaud, and K. G. Schoenly. 2015. Experimental design, inferential statistics,  
1729 and computer modeling. Pages 205-229 in J. K. Tomberlin and M. E. Benbow, editors.

- 1730 Forensic Entomology: International Dimensions and Frontiers. CRC Press, Boca Raton,  
1731 Florida.
- 1732 Moser, J. C. 1976. Surveying mites (Acarina) phoretic on the southern pine beetle (Coleoptera:  
1733 Scolytidae) with sticky traps. *The Canadian Entomologist* **108**:809-813.
- 1734 Moser, J. C., and L. M. Roton. 1971. Mites associated with southern pine bark beetles in Allen  
1735 Parish, Louisiana. *The Canadian Entomologist* **103**:1775-1798.
- 1736 Müller, K., C. Chadeaux, N. Thomas, and I. Reiche. 2011. Microbial attack of archaeological  
1737 bones versus high concentrations of heavy metals in the burial environment. A case study  
1738 of animal bones from a mediaeval copper workshop in Paris. *Palaeogeography,*  
1739 *Palaeoclimatology, Palaeoecology* **310**:39-51.
- 1740 Nair, N. B., and M. Saraswathy. 1971. The Biology of Wood-Boring Teredinid Molluscs.  
1741 *Advances in Marine Biology* **9**:335-509.
- 1742 Nakano, S., H. Miyasaka, and N. Kuhara. 1999. Terrestrial–aquatic linkages: riparian arthropod  
1743 inputs alter trophic cascades in a stream food web. *Ecology* **80**:2435-2441.
- 1744 Nayduch, D. 2017. Special Collection: Filth Fly–Microbe Interactions. *Annals of the*  
1745 *Entomological Society of America* **110**:2-5.
- 1746 Nealson, K. H., and J. W. Hastings. 1979. Bacterial bioluminescence: its control and ecological  
1747 significance. *Microbiological reviews* **43**:496-518.
- 1748 Nelson, J., D. Wubah, M. Whitmer, E. Johnson, and D. Stewart. 1999. Wood-eating catfishes of  
1749 the genus *Panaque*: gut microflora and cellulolytic enzyme activities. *Journal of Fish*  
1750 *Biology* **54**:1069-1082.



1751 Nichols, E., S. Spector, J. Louzada, T. Larsen, S. Amezcuita, M. Favila, and T. S. R. Network.  
1752 2008. Ecological functions and ecosystem services provided by Scarabaeinae dung  
1753 beetles. *Biological conservation* **141**:1461-1474.

1754 Nobre, T., C. Rouland-Lefevre, and D. K. Aanen. 2011. Comparative biology of fungus  
1755 cultivation in termites and ants. Pages 193–210 in D. Bignell, Y. Roisin, and N. Lo,  
1756 editors. *Biology of Termites: A Modern Synthesis*. Springer, New York.

1757 Nowlin, W. H., M. J. Gonzalez, M. J. Vanni, M. H. H. Stevens, M. W. Fields, and J. J. Valentei.  
1758 2007. Allochthonous subsidy of periodical cicadas affects the dynamics and stability of  
1759 pond communities. *Ecology* **88**:2174-2186.

1760 Nykvist, N. 1959a. Leaching and decomposition of litter I. Experiments on leaf litter of *Fraxinus*  
1761 *excelsior*. *Oikos* **10**:190-211.

1762 Nykvist, N. 1959b. Leaching and decomposition of litter II. Experiments on needle litter of *Pinus*  
1763 *silvestris*. *Oikos* **10**:212-224.

1764 Nykvist, N. 1963. Leaching and decomposition of water-soluble organic substances from different  
1765 types of leaf and needle litter. *Studia Forestalia Suecica* No 3, Stockholm.

1766 O'Brien, R. W., and M. Slaytor. 1982. The role of microorganisms in the metabolism of termites.  
1767 *Australian Journal of Biological Sciences* **35**:239-262.

1768 Odum, E. P. 1966. The strategy of ecosystem development. *Science* **164** 262-270.

1769 Odum, H. T. 1970. Summary: An emerging view of the ecological system at El Verde. Pages I-  
1770 191-I-289 in H. T. Odum and R. F. Pigeon, editors. *A tropical rain forest: A study of*  
1771 *irradiation and ecology at El Verde, Puerto Rico*. U.S. Atomic Energy Commission, Oak  
1772 Ridge, Tennessee.

- 1773 Ogada, D., M. Torchin, M. Kinnaird, and V. Ezenwa. 2012. Effects of vulture declines on  
1774 facultative scavengers and potential implications for mammalian disease transmission.  
1775 *Conservation Biology* **26**:453-460.
- 1776 Oh, J., A. L. Byrd, C. Deming, S. Conlan, N. C. S. Program, H. H. Kong, and J. A. Segre. 2014.  
1777 Biogeography and individuality shape function in the human skin metagenome. *Nature*  
1778 **514**:59-64.
- 1779 Opelt, K., V. Chobot, F. Hadacek, S. Schönmann, L. Eberl, and G. Berg. 2007. Investigations of  
1780 the structure and function of bacterial communities associated with Sphagnum mosses. .  
1781 *Environmental Microbiology* **9**:2795–2809.
- 1782 Parmenter, R. R., and V. A. Lamarra. 1991. Nutrient cycling in a freshwater marsh: the  
1783 decomposition of fish and waterfowl carrion. *Limnology and Oceanography* **36**:976-987.
- 1784 Parmenter, R., and J. MacMahon. 2009. Carrion decomposition and nutrient cycling in a  
1785 semiarid shrub-steppe ecosystem. *Ecological Monographs* **79**:637-661.
- 1786 Parsons, T., and J. Strickland. 1962. Oceanic detritus. *Science* **136**:313-314.
- 1787 Payne, J. A. 1965. A summer carrion study of the baby pig *Sus scrofa* Linnaeus. *Ecology*  
1788 **46**:592-602.
- 1789 Payne, J. A., and E. W. King. 1972. Insect succession and decomposition of pig carcasses in  
1790 water. *Journal of the Georgia Entomological Society* **7**:153-162.
- 1791 Payne, J. A., E. W. King, and G. Beinhart. 1968. Arthropod succession and decomposition of  
1792 buried pigs. *Nature* **219**:1180-&.
- 1793 Pechal, J. L., and M. E. Benbow. 2016. Microbial ecology of the salmon necrobiome: evidence  
1794 salmon carrion decomposition influences aquatic and terrestrial insect microbiomes.  
1795 *Environmental Microbiology* **18**:1511-1522.

- 1796 Pechal, J. L., M. E. Benbow, T. L. Crippen, A. M. Tarone, and J. K. Tomberlin. 2014a. Delayed  
1797 insect access alters carrion decomposition and necrophagous insect community assembly.  
1798 *Ecosphere* **5**:doi: 10.1890/ES1814-00022.00021.
- 1799 Pechal, J. L., T. L. Crippen, M. E. Benbow, A. M. Tarone, S. Dowd, and J. K. Tomberlin. 2014b.  
1800 The potential use of bacterial community succession in forensics as described by high  
1801 throughput metagenomic sequencing. *International Journal of Legal Medicine* **128**:193-  
1802 205.
- 1803 Pechal, J. L., T. L. Crippen, A. M. Tarone, A. J. Lewis, J. K. Tomberlin, and M. E. Benbow.  
1804 2013. Microbial Community Functional Change during Vertebrate Carrion  
1805 Decomposition. *Plos One* **8**:e79035.
- 1806 Pereira, L. M., N. Owen-Smith, and M. Moleon. 2014. Facultative predation and scavenging by  
1807 mammalian carnivores: seasonal, regional and intra-guild comparisons. *Mammal Review*  
1808 **44**:44-55.
- 1809 Perotti, M. A., and H. R. Braig. 2009. Phoretic mites associated with animal and human  
1810 decomposition. *Experimental and Applied Acarology* **49**:85-124.
- 1811 Persson, Y., R. Vasaitis, B. Långström, P. Öhrn, K. Ihrmark, and J. Stenlid. 2009. Fungi vectored  
1812 by the bark beetle *Ips typographus* following hibernation under the bark of standing trees  
1813 and in the forest litter. *Microbial Ecology* **58**:651-659.
- 1814 Peterson, B. F., and M. E. Scharf. 2016. Lower termite associations with microbes: Synergy,  
1815 protection, and interplay. *Frontiers in Microbiology* **7**:422.
- 1816 Polis, G. A., W. B. Anderson, and R. D. Holt. 1997. Toward an integration of landscape and food  
1817 web ecology: The dynamics of spatially subsidized food webs. *Annual Review of*  
1818 *Ecology and Systematics* **28**:289-316.

- 1819 Polis, G. A., and S. D. Hurd. 1995. Extraordinarily high spider densities on islands: flow of  
1820 energy from the marine to terrestrial food webs and the absence of predation. *Proceedings*  
1821 *of the National Academy of Sciences of the United States of America* **92**:4382-4386.
- 1822 Polis, G. A., and S. D. Hurd. 1996. Linking marine and terrestrial food webs: Allochthonous  
1823 input from the ocean supports high secondary productivity on small islands and coastal  
1824 land communities. *American Naturalist* **147**:396-423.
- 1825 Preiswerk, D., J.-C. Walser, and D. Ebert. 2018. Temporal dynamics of microbiota before and after  
1826 host death. *The ISME Journal*: 10.1038/s41396-018-0157-2.
- 1827 Purahong, W., T. Wubet, G. Lentendu, M. Schloter, M. J. Pecyna, D. Kapturska, M. Hofrichter,  
1828 D. Krüger, and F. Buscot. 2016. Life in leaf litter: novel insights into community  
1829 dynamics of bacteria and fungi during litter decomposition. *Molecular Ecology* **25**:4059-  
1830 4074.
- 1831 Purrington, F. F., and C. J. Drake. 2008. Phoretic deutonymphs of *Schwiebea* sp. (Acari:  
1832 *Astigmata*: *Acaridae*) travel in commodious nitid dorsal pits of adult *Lagocheirus*  
1833 *araneiformis stroheckeri* Dillon (Coleoptera: *Cerambycidae*: *Lamiinae*) in Florida,  
1834 U.S.A. *Entomological News* **119**:415-419.
- 1835 Ramirez, K. S., C. L. Lauber, and N. Fierer. 2010. Microbial consumption and production of  
1836 volatile organic compounds at the soil-litter interface. *Biogeochemistry* **99**:97-107.
- 1837 Refsnider, J. M., and F. J. Janzen. 2010. Putting eggs in one basket: ecological and evolutionary  
1838 hypotheses for variation in oviposition-site choice. *Annual Review of Ecology,*  
1839 *Evolution, and Systematics* **41**:39-57.

- 1840 Reid, N. M., S. L. Addison, L. J. Macdonald, and G. Lloyd-Jones. 2011. Biodiversity of active  
1841 and inactive bacteria in the gut flora of wood-feeding Huhu beetle larvae (*Prionopus*  
1842 *reticularis*). *Applied and Environmental Microbiology* **77**:7000–7006.
- 1843 Retana, J., X. Cerdà, and X. Espadaler. 1991. Arthropod corpses in a temperate grassland: a  
1844 limited supply? *Ecography* **14**:63-67.
- 1845 Reynolds, B. C., and M. D. Hunter. 2004. Nutrient cycling. Pages 387-396 in M. D. Lowman  
1846 and H. B. Rinker, editors. *Forest Canopies*. Elsevier, New York.
- 1847 Rich, P. H., and R. G. Wetzel. 1978. Detritus in the lake ecosystem. *The American Naturalist*  
1848 **112**:57-71.
- 1849 Richards, E. N., and M. L. Goff. 1997. Arthropod succession on exposed carrion in three  
1850 contrasting tropical habitats on Hawaii island, Hawaii. *Journal of Medical Entomology*  
1851 **34**:328-339.
- 1852 Rode, K. D., C. A. Chapman, L. J. Chapman, and L. R. McDowell. 2003. Mineral resource  
1853 availability and consumption by colobus in Kibale National Park, Uganda. *International*  
1854 *Journal of Primatology* **24**:541-573.
- 1855 Roggenbuck, M., I. B. Schnell, N. Blom, J. Bælum, M. F. Bertelsen, T. Sicheritz-Pontén, S. J.  
1856 Sørensen, M. T. P. Gilbert, G. R. Graves, and L. H. Hansen. 2014. The microbiome of  
1857 New World vultures. *Nature communications* **5**:5498.
- 1858 Ronkainen, T., E. L. McClymont, M. Väiliranta, and E.-S. Tuittila. 2013. The n-alkane and sterol  
1859 composition of living fen plants as a potential tool for palaeoecological studies. *Organic*  
1860 *Geochemistry* **59**:1-9.
- 1861 Rothman, J. M., P. J. Van Soest, and A. N. Pell. 2006. Decaying wood is a sodium source for  
1862 mountain gorillas. *Biology Letters* **2**:321-324.

- 1863 Rozen, D. E., D. J. P. Engelmoer, and P. T. Smiseth. 2008. Antimicrobial strategies in burying  
1864 beetles breeding on carrion. *Proceedings of the National Academy of Sciences of the*  
1865 *United States of America* **105**:17890-17895.
- 1866 Ruiz-Villanueva, V., H. Piégay, A. A. Gurnell, R. A. Marston, and M. Stoffel. 2016. Recent  
1867 advances quantifying the large wood dynamics in river basins: New methods and  
1868 remaining challenges. *Reviews of Geophysics* **54**:611-652.
- 1869 Ruxton, G. D., and D. C. Houston. 2004. Obligate vertebrate scavengers must be large soaring  
1870 fliers. *Journal of Theoretical Biology* **228**:431-436.
- 1871 Santos, P. F., J. Phillips, and W. G. Whitford. 1981. The role of mites and nematodes in early  
1872 stages of buried litter decomposition in a desert. *Ecology* **62**:664-669.
- 1873 Scherer, R. 2004. Decomposition and longevity of in-stream woody debris: a review of literature  
1874 from North America. Pages 127-133 in *Forest Land–Fish Conference–Ecosystem*  
1875 *Stewardship through Collaboration. Proc. Forest-Land-Fish Conf. II. Citeseer.*
- 1876 Schoenly, K., and W. Reid. 1987. Dynamics of heterotrophic succession in carrion arthropod  
1877 assemblages: discrete series or a continuum of change? *Oecologia* **73**:192-202.
- 1878 Schuurman, G. 2005. Decomposition rates and termite assemblage composition in semiarid  
1879 Africa. *Ecology* **86**:1236-1249.
- 1880 Seastedt, T. R. 1984. The role of microarthropods in decomposition and mineralization  
1881 processes. *Annual Review of Entomology* **29**:25-46.
- 1882 Seastedt, T. R., and C. M. Tate. 1981. Decomposition rates and nutrient contents of arthropod  
1883 remains in forest litter. *Ecology* **62**:13-19.

- 1884 Selva, N., B. Jędrzejewska, W. Jędrzejewski, and A. Wajrak. 2005. Factors affecting carcass use  
1885 by a guild of scavengers in European temperate woodland. *Canadian Journal of Zoology*  
1886 **83**:1590-1601.
- 1887 Shivik, J. A. 2006. Are vultures birds, and do snakes have venom, because of macro-and  
1888 microscavenger conflict? *AIBS Bulletin* **56**:819-823.
- 1889 Simmons, T., R. E. Adlam, and C. Moffatt. 2010. Debugging decomposition data—comparative  
1890 taphonomic studies and the influence of insects and carcass size on decomposition rate.  
1891 *Journal of Forensic Sciences* **55**:8-13.
- 1892 Singh, J., and S. Gupta. 1977. Plant decomposition and soil respiration in terrestrial ecosystems.  
1893 *The Botanical Review* **43**:449-528.
- 1894 Singh, B., T. L. Crippen, L. Zheng, A. T. Fields, Z. Yu, Q. Ma, T. K. Wood, S. E. Dowd, M.  
1895 Flores, and J. K. Tomberlin. 2015. A metagenomic assessment of the bacteria associated  
1896 with *Lucilia sericata* and *Lucilia cuprina* (Diptera: Calliphoridae). *Applied Microbiology*  
1897 *and Biotechnology* **99**:869-883.
- 1898 Smith, C. R., H. L. Maybaum, A. R. Baco, R. H. Pope, S. D. Carpenter, P. L. Yager, S. A. Macko,  
1899 and J. W. Deming. 1998. Sediment community structure around a whale skeleton in the deep  
1900 Northeast Pacific: macrofaunal, microbial and bioturbation effects. *Deep Sea Research Part*  
1901 *II: Topical Studies in Oceanography* **45**:335-364.
- 1902 Smith, C. R., and A. R. Baco. 2003. Ecology of whale falls at the deep-sea floor. *Oceanography*  
1903 *and Marine Biology* **41**:311-354
- 1904 Smith, J. B., L. J. Laatsch, and J. C. Beasley. 2017. Spatial complexity of carcass location  
1905 influences vertebrate scavenger efficiency and species composition. *Scientific Reports*  
1906 **7**:10250.

- 1907 Song, Z., P. G. Kennedy, F. J. Liew, and J. S. Schilling. 2017. Fungal endophytes as priority  
1908 colonizers initiating wood decomposition. *Functional Ecology* **31**:407-418.
- 1909 Soper, R. S., and R. E. Olson. 1963. Survey of biota associated with *Monochamus* (Coleoptera:  
1910 Cerambycidae) in Maine. *The Canadian Entomologist* **95**:83-95.
- 1911 Speight, M. C. D. 1989. Saproxylic invertebrates and their conservation. Council of Europe,  
1912 Strasbourg.
- 1913 St John, M. G., K. H. Orwin, and I. A. Dickie. 2011. No 'home' versus 'away' effects of  
1914 decomposition found in a grassland-forest reciprocal litter transplant study. *Soil Biology  
1915 & Biochemistry* **43**:1482-1489.
- 1916 Stokland, J. N., J. Siitonen, and B. G. Jonsson. 2012. *Biodiversity in Dead Wood*. Cambridge  
1917 University Press, Cambridge.
- 1918 Stoklosa, A. M., M. D. Ulyshen, Z. Fan, M. Varner, S. Seibold, and J. Müller. 2015. Effects of  
1919 mesh bag enclosure and termites on fine woody debris decomposition in a subtropical  
1920 forest. *Basic and Applied Ecology* **17**:463-470.
- 1921 Strickland, M., A. Keiser, and M. Bradford. 2015. Climate history shapes contemporary leaf  
1922 litter decomposition. *Biogeochemistry* **122**:165-174.
- 1923 Strickland, M. S., C. Lauber, N. Fierer, and M. A. Bradford. 2009a. Testing the functional  
1924 significance of microbial community composition. *Ecology* **90**:441-451.
- 1925 Strickland, M. S., E. Osburn, C. Lauber, N. Fierer, and M. A. Bradford. 2009b. Litter quality is  
1926 in the eye of the beholder: initial decomposition rates as a function of inoculum  
1927 characteristics. *Functional Ecology* **23**:627-636.



- 1928 Strid, Y., M. Schroeder, B. Lindahl, K. Ihrmark, and J. Stenlid. 2014. Bark beetles have a  
1929 decisive impact on fungal communities in Norway spruce stem sections. *Fungal Ecology*  
1930 **7**:47-58.
- 1931 Subalusky, A. L., C. L. Dutton, E. J. Rosi, and D. M. Post. 2017. Annual mass drownings of the  
1932 Serengeti wildebeest migration influence nutrient cycling and storage in the Mara River.  
1933 *Proceedings of the National Academy of Sciences* **114**:7647-7652.
- 1934 Summerhayes, V., and C. S. Elton. 1923. *Contributions to the Ecology of Spitsbergen and*  
1935 *Bear Island*. University Press.
- 1936 Sun, L., Z. Xie, and J. Zhao. 2000. A 3,000-year record of penguin populations. *Nature* **407**:858.
- 1937 Swift, M. J., and L. Boddy. 1984. Animal-Microbial Interactions in Wood Decomposition. Pages  
1938 89-131 in J. M. Anderson, A. D. M. Rayner, and D. W. H. Walton, editors. *Invertebrate-*  
1939 *microbial interactions*. Cambridge University Press, Cambridge.
- 1940 Swift, M. J., O. W. Heal, and J. M. Anderson. 1979. *Decomposition in Terrestrial Ecosystems.*  
1941 *Studies in Ecology*. Blackwell Scientific, Oxford, UK.
- 1942 Tanahashi, M., K. Kubota, N. Matsushita, and K. Togashi. 2010. Discovery of mycangia and the  
1943 associated xylose-fermenting yeasts in stag beetles (Coleoptera: Lucanidae).  
1944 *Naturwissenschaften* **97**:311-317.
- 1945 Tanahashi, M., N. Matsushita, and K. Togashi. 2009. Are stag beetles fungivorous? *Journal of*  
1946 *Insect Physiology* **55**:983-988.
- 1947 Tang, K. W., C. S. Freund, and C. L. Schweitzer. 2006. Occurrence of copepod carcasses in the  
1948 lower Chesapeake Bay and their decomposition by ambient microbes. *Estuarine, Coastal*  
1949 *and Shelf Science* **68**:499-508.

- 1950 Tang, K. W., S. L. Bickel, C. Dziallas, and H.-P. Grossart. 2009. Microbial activities  
1951 accompanying decomposition of cladoceran and copepod carcasses under different  
1952 environmental conditions. *Aquatic Microbial Ecology* **57**:89-100.
- 1953 Tang, K. W., M. I. Gladyshev, O. P. Dubovskaya, G. Kirillin, and H.-P. Grossart. 2014.  
1954 Zooplankton carcasses and non-predatory mortality in freshwater and inland sea  
1955 environments. *Journal of Plankton Research* **36**:597-612.
- 1956 Teal, J. M. 1962. Energy flow in the salt marsh ecosystem of Georgia. *Ecology* **43**:614-624.
- 1957 Thormann, M. N., R. S. Currah, and S. E. Bayley. 2001. Microfungi isolated from *Sphagnum*  
1958 *fuscum* from a southern boreal bog in Alberta, Canada. *Bryologist* **104**:548-559.
- 1959 Thorne, B. L., and R. B. Kimsey. 1983. Attraction of neotropical *Nasutitermes* termites to  
1960 carrion. *Biotropica* **15**:295-296.
- 1961 Throckmorton, H. M., J. A. Bird, L. Dane, M. K. Firestone, and W. R. Horwath. 2012. The  
1962 source of microbial C has little impact on soil organic matter stabilisation in forest  
1963 ecosystems. *Ecology Letters* **15**:1257-1265.
- 1964 Thunes, K. H., F. Midtgaard, and I. Gjerde. 2000. Diversity of coleoptera of the bracket fungus  
1965 *Fomitopsis pinicola* in a Norwegian spruce forest. *Biodiversity and Conservation* **9**:833-  
1966 852.
- 1967 Toki, W., M. Tanahashi, K. Togashi, and T. Fukatsu. 2012. Fungal farming in a non-social  
1968 beetle. *Plos One* **7**:e41893.
- 1969 Tomberlin, J. K., and P. H. Adler. 1998. Seasonal colonization and decomposition of rat carrion  
1970 in water and on land in an open field in South Carolina. *Journal of Medical Entomology*  
1971 **35**:704-709.

- 1972 Tomberlin, J. K., J. Byrd, J. R. Wallace, and E. M. Benbow. 2012a. Assessment of  
1973 decomposition studies indicates need for standardized and repeatable research methods in  
1974 forensic entomology. *Journal of Forensic Research* **3**:147.
- 1975 Tomberlin, J. K., T. L. Crippen, A. M. Tarone, B. Singh, K. Adams, Y. H. Rezenom, M. E.  
1976 Benbow, M. Flores, M. Longnecker, J. L. Pechal, D. H. Russell, R. C. Beier, and T. K.  
1977 Wood. 2012b. Interkingdom responses of flies to bacteria mediated by fly physiology and  
1978 bacterial quorum sensing. *Animal Behaviour* **84**:1449-1456.
- 1979 Tomberlin, J. K., R. Mohr, M. E. Benbow, A. M. Tarone, and S. VanLaerhoven. 2011. A  
1980 roadmap for bridging basic and applied research in forensic entomology. *Annual Review*  
1981 *of Entomology* **56**:401-421.
- 1982 Towne, E. G. 2000. Prairie vegetation and soil nutrient responses to ungulate carcasses.  
1983 *Oecologia* **122**:232-239.
- 1984 Trumbo, S. T. 1990. Reproductive success, phenology and biogeography of burying beetles  
1985 (*Silphidae*, *Nicrophorus* spp.). *American Midland Naturalist* **124**:1-11.
- 1986 Trumbo, S. T., D. S. Sikes, and P. K. B. Philbrick. 2016. Parental care and competition with  
1987 microbes in carrion beetles: a study of ecological adaptation. *Animal Behaviour* **118**:47-  
1988 54.
- 1989 Tscharrntke, T., J. M. Tylianakis, T. A. Rand, R. K. Didham, L. Fahrig, P. Batáry, J. Bengtsson,  
1990 Y. Clough, T. O. Crist, C. F. Dormann, R. M. Ewers, J. Fründ, R. D. Holt, A. Holzschuh,  
1991 A. M. Klein, D. Kleijn, C. Kremen, D. A. Landis, W. F. Laurance, D. Lindenmayer, C.  
1992 Scherber, N. S. Sodhi, I. Steffan-Dewenter, C. Thies, W. H. van der Putten, and C.  
1993 Westphal. 2012. Landscape moderation of biodiversity patterns and processes - eight  
1994 hypotheses. *Biological Reviews* **87**:661-685.

- 1995 Turetsky, M. R. 2003. The role of bryophytes in carbon and nitrogen cycling. *Bryologist*
- 1996 **106**:395–409.
- 1997 Turner, K. L., E. F. Abernethy, M. L. Conner, O. E. Rhodes, and J. C. Beasley. 2017. Abiotic
- 1998 and biotic factors modulate carrion fate and vertebrate scavenging communities. *Ecology*.
- 1999 Ulyshen, M. D. 2014. Interacting effects of insects and flooding on wood decomposition. *Plos*
- 2000 *One* **9**:e101867.
- 2001 Ulyshen, M. D. 2015. Insect-mediated nitrogen dynamics in decomposing wood. *Ecological*
- 2002 *Entomology* **40**:97-112.
- 2003 Ulyshen, M. D. 2016. Wood decomposition as influenced by invertebrates. *Biological Reviews*
- 2004 **91**:70-85.
- 2005 Urbina, H., J. Schuster, and M. Blackwell. 2013. The gut of Guatemalan passalid beetles: a
- 2006 habitat colonized by cellobiose- and xylose-fermenting yeasts. *Fungal Ecology* **6**:339-
- 2007 355.
- 2008 Van der Wal, R., R. D. Bardgett, K. A. Harrison, and A. Stien. 2004. Vertebrate herbivores and
- 2009 ecosystem control: cascading effects of faeces on tundra ecosystems. *Ecography* **27**:242-252.
- 2010 Van der Wal, A., W. de Boer, W. Smant, and A. van Veen. 2007. Initial decay of woody
- 2011 fragments in soil is influenced by size, vertical position, nitrogen availability and soil
- 2012 origin. *Plant and Soil* **301**:189-201.
- 2013 Van Geel, B. 1978. A palaeoecological study of holocene peat bog sections in Germany and The
- 2014 Netherlands, based on the analysis of pollen, spores and macro- and microscopic remains
- 2015 of fungi, algae, cormophytes and animals. *Review of Palaeobotany and Palynology* **25**:1-
- 2016 120.

- 2017 Vannote, R. L., G. W. Minshall, K. W. Cummins, J. R. Sedell, and C. E. Cushing. 1980. River  
2018 Continuum Concept. *Canadian Journal of Fisheries and Aquatic Sciences* **37**:130-137.
- 2019 Vass, A. A. 2001. Beyond the grave: understanding human decomposition. *Microbiology Today*  
2020 **28**:190-192.
- 2021 Verhoeven, J. T. A., and W. M. Liefveld. 1997. The ecological significance of organochemical  
2022 compounds in Sphagnum. *Acta Botanica Neerlandica* **46**:117-130.
- 2023 Villiers Pienaar, U. d. 1969. Predator-prey relationship amongst the larger mammals of the  
2024 Kruger National Park. *Koedoe* **12**:108-176.
- 2025 Voříšková, J., and P. Baldrian. 2013. Fungal community on decomposing leaf litter undergoes  
2026 rapid successional changes. *The ISME Journal* **7**:477-486.
- 2027 Wall, D. H., M. A. Bradford, M. G. ST. JOHN, J. A. Trofymow, V. BEHAN-PELLETIER, D. E.  
2028 Bignell, J. M. Dangerfield, W. J. Parton, J. Rusek, and W. Voigt. 2008. Global  
2029 decomposition experiment shows soil animal impacts on decomposition are  
2030 climate-dependent. *Global Change Biology* **14**:2661-2677.
- 2031 Wallace, J. B., S. L. Eggert, J. L. Meyer, and J. R. Webster. 1997. Multiple trophic levels of a  
2032 forest stream linked to terrestrial litter inputs. *Science* **277**:102-104.
- 2033 Wallace, J. B., S. Eggert, J. L. Meyer, and J. Webster. 1999. Effects of resource limitation on a  
2034 detrital-based ecosystem. *Ecological Monographs* **69**:409-442.
- 2035 Wallace, J. R. 2015. Aquatic vertebrate carrion decomposition. Pages 247-271 in M. E. Benbow,  
2036 J. K. Tomberlin, and A. M. Tarone, editors. *Carrion Ecology, Evolution and Their*  
2037 *Applications*. CRC Press, Boca Raton, FL.

- 2038 Wang, Y., and D. E. Rozen. 2017. Gut microbiota colonization and transmission in the burying  
2039 beetle *Nicrophorus vespilloides* throughout development. *Applied and Environmental*  
2040 *Microbiology* **83**.
- 2041 Warren, R. J., and M. A. Bradford. 2012. Ant colonization and coarse woody debris  
2042 decomposition in temperate forests. *Insectes Sociaux* **59**:215-221.
- 2043 Waters, T. F. 1966. Production rate, population density, and drift of a stream invertebrate. *Ecology*  
2044 **47**:595-604.
- 2045 Waters, T. F. 1969. The turnover ratio in production ecology of freshwater invertebrates. *The*  
2046 *American Naturalist* **103**:173-185.
- 2047 Waters, T. F. 1977. Secondary production in inland waters. *Advances in Ecological Research*  
2048 **10**:91-164.
- 2049 Weatherbee, C., J. Pechal, and M. Benbow. 2017. The dynamic maggot mass microbiome.  
2050 *Annals of the Entomological Society of America* **110**:45-53.
- 2051 Webster, J., and E. Benfield. 1986. Vascular plant breakdown in freshwater ecosystems. *Annual*  
2052 *Review of Ecology and Systematics* **17**:567-594.
- 2053 Weedon, J. T., W. K. Cornwell, J. H. C. Cornelissen, A. E. Zanne, C. Wirth, and D. A. Coomes.  
2054 2009. Global meta-analysis of wood decomposition rates: a role for trait variation among  
2055 tree species? *Ecology Letters* **12**:45-56.
- 2056 Wei, H., Q. Xu, L. E. Taylor, J. O. Baker, M. P. Tucker, and S.-Y. Ding. 2009. Natural  
2057 paradigms of plant cell wall degradation. *Current Opinion in Biotechnology* **20**:330-338.
- 2058 Wende, B., M. M. Gossner, I. Grass, T. Arnstadt, M. Hofrichter, A. Floren, K. E. Linsenmair, W.  
2059 W. Weisser, and I. Steffan-Dewenter. 2017. Trophic level, successional age and trait

2060 matching determine specialization of deadwood-based interaction networks of saproxylic  
2061 beetles. *Proceedings of the Royal Society B: Biological Sciences* **284**:20170198.

2062 Weslien, J., L. B. Djupström, M. Schroeder, and O. Widenfalk. 2011. Long-term priority effects  
2063 among insects and fungi colonizing decaying wood. *Journal of Animal Ecology* **80**:1155-  
2064 1162.

2065 Wiens, J. A. 1995. Landscape mosaics and ecological theory. Pages 1-26 in L. Hannsson, L. Fahrig,  
2066 and G. Merriam. Editors. *Mosaic landscapes and ecological processes*. Springer, Dordrecht.

2067 Wilmers, C. C., R. L. Crabtree, D. W. Smith, K. M. Murphy, and W. M. Getz. 2003. Trophic  
2068 facilitation by introduced top predators: grey wolf subsidies to scavengers in Yellowstone  
2069 National Park. *Journal of Animal Ecology* **72**:909-916.

2070 Wilson, E. E., and E. M. Wolkovich. 2011. Scavenging: how carnivores and carrion structure  
2071 communities. *Trends in Ecology and Evolution* **26**:129-135.

2072 Wilson, E. O. 1971. *The insect societies*. Harvard University Press, Cambridge.

2073 Wilson, K., and D. J. B. White. 1986. *The Anatomy of Wood: Its Diversity and Variability*.  
2074 Stobart and Son Ltd, London.

2075 Witkamp, M. 1966. Decomposition of leaf litter in relation to environment, microflora, and  
2076 microbial respiration. *Ecology* **47**:194-201.

2077 Wood, T. G., and R. J. Thomas. 1989. The mutualistic association between Macrotermitinae and  
2078 Termitomyces. Pages 69–92 in N. Wilding, N. M. Collins, P. M. Hammond, and J. F.  
2079 Webber, editors. *Insect-Fungus Interactions* Academic Press, London.

2080 Woodcock, B. A., A. D. Watt, and S. R. Leather. 2002. Aggregation, habitat quality and  
2081 coexistence: a case study on carrion fly communities in slug cadavers. *Journal of Animal*  
2082 *Ecology* **71**:131-140.

- 2083 Woodward, G., M. O. Gessner, P. S. Giller, V. Gulis, S. Hladyz, A. Lecerf, B. Malmqvist, B. G.  
2084 McKie, S. D. Tiegs, H. Cariss, M. Dobson, A. Elozegi, V. Ferreira, M. A. S. Graça, T.  
2085 Fleituch, J. O. Lacoursière, M. Nistorescu, J. Pozo, G. Risnoveanu, M. Schindler, A.  
2086 Vadineanu, L. B.-M. Vought, and E. Chauvet. 2012. Continental-scale effects of nutrient  
2087 pollution on stream ecosystem functioning. *Science* **336**:1438-1440.
- 2088 Wotton and Malmqvist. 2001. Feces in aquatic ecosystems. *BioScience* **51**:537-544.
- 2089 Yang, L. H. 2004. Periodical cicadas as resource pulses in North American forests. *Science*  
2090 **306**:1565-1567.
- 2091 Yang, L. H., K. F. Edwards, J. E. Byrnes, J. L. Bastow, A. N. Wright, and K. O. Spence. 2010. A  
2092 meta-analysis of resource pulse–consumer interactions. *Ecological Monographs* **80**:125-  
2093 151.
- 2094 Young, O. P. 1984. Utilization of Dead Insects on the Soil Surface in Row Crop Situations.  
2095 *Environmental Entomology* **13**:1346-1351.
- 2096 Zak, D. R., W. E. Holmes, D. C. White, A. D. Peacock, and D. Tilman. 2003. Plant diversity, soil  
2097 microbial communities, and ecosystem function: are there any links? *Ecology* **84**:2042-2050.
- 2098 Zhou, C., and R. W. Byard. 2011. Factors and processes causing accelerated decomposition in  
2099 human cadavers—An overview. *Journal of Forensic and Legal Medicine* **18**:6-9.

2100

2101

2102 **DATA AVAILABILITY**

2103 Data are available from the Dryad Digital Repository: <https://doi.org/10.5061/dryad.pp5bs35>

2104



2105 **Table 1.** Decomposer components and characteristics of heterotrophically- and autotrophically-  
 2106 derived necromass decomposition with representative studies for each component. Microbial  
 2107 communities include prokaryotes and microscopic protists, fungi and eukaryotes. Temporal  
 2108 scales range from ecologically relevant scales related to the life history traits of decomposer  
 2109 species to plant and animal biomass protected from decomposer species over geological time  
 2110 scales of decades or centuries (e.g., peat bog mummies).

Component/Characteristic	Heterotrophically-derived biomass (Invertebrate and Vertebrate Animals)	Autotrophically-derived biomass (Plant Leaves and Wood and Algal Detritus)
Communities		
Endonecrotic Microbial Communities	(Can et al. 2014, Hyde et al. 2015, Javan et al. 2016)	(Arnold et al. 2000, Wei et al. 2009, Song et al. 2017)
Epinecrotic Microbial Communities	(Burkepile et al. 2006, Pechal et al. 2013, Pechal et al. 2014b, Metcalf et al. 2016)	(Swift and Boddy 1984, Arnold et al. 2000, Persson et al. 2009, Voříšková and Baldrian 2013)
Macroinvertebrate-Saprotrophs	Blow flies (e.g., Diptera: Calliphoridae, Sarcophagidae) and Beetles (e.g., Coleoptera:	Wood boring and detrital insects (e.g., Coleoptera: Curculionidae): (Hickin 1963, Wallace et al. 1997, Haila and Niemelä 1999, Lieutier et

---

	Staphylinidae and	al. 2004, Hattenschwiler et al.
	Silphidae): (Fuller 1934,	2005)
	Chapman and Sankey 1955,	
	Bornemissza 1957, Payne	
	1965, Greenberg 1971a, b,)	
Vertebrate-Saprotrophs	(DeVault et al. 2003, Selva	(Nelson et al 1999, Rode et al 2003,
	et al. 2005, Wilson and	Rothman et al 2006)
	Wolkovich 2011, Turner et	
	al. 2017)	
Soil/Habitat Communities	(Lauber et al 2014, Carter et	(Singh and Gupta 1977, Zak et al
	al. 2015, Finley et al. 2016)	2003, Hawlena et al. 2012)
Taxa and Element		
Translocation and Dispersal		
Spatial Scale Context	Localized (0.1 – 100 m):	Localized (0.1 – 100 m):
	(Payne 1965, Moreau et al.	(Lussenhop 1992, Strickland et al.
	2015)	2009a)
	Diffuse (100-1000 m):	Diffuse (100 – 10,000 m): (Polis
	(Houston 1974, Holland et	and Hurd 1996, Jiménez et al.
	al. 2017)	2017)
Temporal Scale Context	Ecologically Short (minutes	Ecologically Short (minutes to
	to days): (DeVault et al.	days): Leaching rates (Nykqvist

---

---

2004b, Farwig et al. 2014, 1959 a,b, 1963, Cummins et al  
Smith et al. 2017) 1973, Kuiters and Sarink 1986)

Ecologically Long (weeks Ecologically Long (weeks to years):  
to years): (Danell et al. (Swift et al. 1979, Handa et al.  
2002, Barton et al. 2016) 2014)

Geological (decades to Geological (decades to centuries):  
centuries): (Macdonald (Van Geel 1978, Middeldorp 1986,  
1992, Sun et al. 2000, Barthelmes et al. 2006, Kvavadze  
Dedout et al. 2010, and Kakhiani 2010, Ronkainen et  
Kvavadze and Kakhiani al. 2013)  
2010, Müller et al. 2011,  
Boeskorov et al. 2014)

---

2111

2112

Both plant and animal necromass has been used extensively as model systems to test and develop ecological theory, and while historically often operating independently these areas of study are where ecologists on both sides of the decomposition ‘divide’ have done well to learn from each other. This is perhaps due to the implicit recognition by theorists of the similarities in fundamental processes occurring during decomposition. There has been research that included both autotrophically- and heterotrophically-derived necromass in food web studies (Polis and Hurd 1996, Polis et al 1997, Nakano et al 1999), but comparisons and similarities in processes, community ecology and mechanisms has received less attention in the literature. **Community succession** of carrion has a long history (Méglin 1894, Schoenly and Reid 1987, Michaud et al. 2015), and has many important similarities with community succession on dead plant matter, such as leaf and fruit litter and dead wood (Ulyshen 2016). Indeed, succession theory is one of the most widely used theories relating to decomposition, especially with regard to the study of the diversity and structure of arthropod communities (Richards and Goff 1997, Hobischak et al. 2006), and more recently for microbial communities (Pechal et al. 2013, Metcalf et al. 2016).

**Meta-population** and **meta-community** theory has been tested using animal carcasses and dung pads as model resource patches to simulate the focal points of insect assemblage aggregation and dispersal (Hanski 1987). This work has led to validation of competitive exclusion and dispersal limitation as mechanisms underpinning population dynamics (Hanski and Gilpin 1991) but could also be tested using decaying plant material. **Aggregation and coexistence theory** has been tested using fly communities at carrion (Ives 1991, Fiene et al. 2014), as well as mushrooms and rotting fruit (Heard 1998, Finn 2001). The theory predicts that if the distribution of a competitively superior species is spatially aggregated, this frees other patches to be colonized by

competitively inferior species, thus leading to differences in community composition and greater overall diversity across multiple patches (Kouki and Hanski 1995, Woodcock et al. 2002).

**Resource subsidy theory** describes the spatial movement of energy and nutrients between two spatially disjunct ecosystems (Leroux and Loreau 2008, Gravel, et al. 2010, Polis, et al. 1997).

This theory has been effective at integrating plant and animal necromass into one resource pool (Moore et al 2004), and at demonstrating links between terrestrial and aquatic systems (Polis and Hurd 1996). For example, the mass emergence of cicadas from forest ecosystems can lead to the deposition of animal necromass into ponds, increasing pond productivity and altering their communities (Nowlin, et al. 2007). Measurement of marine-derived plant and animal necromass washing up on islands has also been shown to underpin island terrestrial arthropod biomass (Polis and Hurd 1995). **Landscape heterogeneity theory** has emerged as a way to link patterns and processes occurring at individual patches to large-scale consequences for landscapes (Wiens 1995). For example, two patches of dead wood or carrion at different temporal decay stages or spatial locations will support a greater range of species than two carcasses at the same decay stage or location. Ultimately, the state of the resource has consequences for supporting overall biodiversity (Barton et al. 2013a), driving soil nutrient heterogeneity (Harmon et al. 1986, Bump et al. 2009a), and generating novel combinations of species (Bump et al. 2009b, Barton et al. 2013b). Application of other landscape ecology principles, such as patch composition vs. configuration theory (Tscharntke et al. 2012), has the potential to yield further insights into the large-scale role of necromass in ecosystem structure and function. Our necrobiome framework can act as a catalyst for further testing and synthesis of theory describing the interactions among organisms and their environment. Especially, for interactions and processes with clear temporal or spatial components in saprotrophic communities as resource substrates degrade over time,

such as dispersal of insects or microbes among patches and the succession of necrophagous communities.

2114

2115

Author Manuscript

2116 **Figure Captions**

2117

2118 **Figure 1.**

2119 The necrobiome is the community of organisms associated with necromass decomposition, and  
2120 includes their interactions with the necromass, with each other, and with their surrounding  
2121 habitat and ecosystem. This concept is general across plant, fungi, dung/frass/feces or animal  
2122 necromass types, and in both terrestrial and aquatic realms. (a) Various qualities of necromass  
2123 have strong controlling effects on its associated necrobiome. (b) The decomposing necromass is  
2124 host to a microbiome while alive, both internal (endonecrotic) and external (epinecrotic), and  
2125 these communities begin to change after death and during the course of decomposition. The  
2126 changing microbiomes use the organic matter as habitat and for energy and nutrients. As decay  
2127 progresses, the organic matter and associated activities of the microbes facilitate the attraction of  
2128 invertebrate and vertebrate saprophages that directly consume the necromass and the microbial  
2129 communities. (c) All of these biotic components of the necrobiome interact in ways that give rise  
2130 to pathways of microbial and elemental dispersal away from the decaying necromass into  
2131 adjacent soil or water media, as well as to the surrounding ecosystem at both localized and broad  
2132 spatial scales. (d) The necrobiome and its functions operate within a complex network of  
2133 interactions that are constrained by its terrestrial or aquatic context, the regional species pool,  
2134 and gradients of abiotic factors. Critically, all these aspects of the necrobiome are interlinked,  
2135 and the relative importance of each component of the necrobiome, and their function, will  
2136 depend on the intrinsic qualities of decaying necromass and the extrinsic controlling  
2137 environmental factors (e.g., temperature, moisture, latitude).

2138

2139 **Figure 2.** Examples of necromass types occurring in ecosystems showing differences in size and  
2140 heterogeneity. (a) Log cross section showing different rates of decay at edges versus center, (b)  
2141 millipede carcass exoskeleton remains, (c) leaf litter, and (d) vertebrate carcass showing  
2142 endoskeleton remains. Despite the contrasting physical appearance of necromass types, the broad  
2143 structure of the necrobiome at each necromass type can be conceptualized in a similar way.

2144

2145 **Figure 3.** Some examples of the range of necromass sizes (L) and nutritional quality (C:N) for  
2146 carrion, dung, leaf and wood substrates. Red points represent examples where there is volume  
2147 data available in the literature, whereas white points are examples for which there is no volume  
2148 data available. Data used for this figure were deposited the Dryad Digital Repository  
2149 (doi:10.5061/dryad.pp5bs35).

2150

2151 **Figure 4.** The spatial and temporal dimensions of the necrobiome among multiple necromass  
2152 patches introduce an additional level of dynamics important to understanding ecosystem-scale  
2153 processes. (1) Each item of necromass decomposes from fresh (F) to dry (D), and the attendant  
2154 necrobiome changes in diversity and composition through time. (2) This creates time windows  
2155 for colonization and dispersal by specialist decomposers and their predators as they search for  
2156 the next similar decay stage. (3) Multiple necromass resources occur at any one time and  
2157 generates a range of decay states with different necrobiomes. (4) This creates a shifting spatial  
2158 mosaic of necrobiome patches over time, each with unique levels of diversity, composition, and  
2159 types of interactions and processes.

2160

2161



(d) **Habitat and ecosystem** determines the substrate context, the species pool, and key abiotic moderating processes

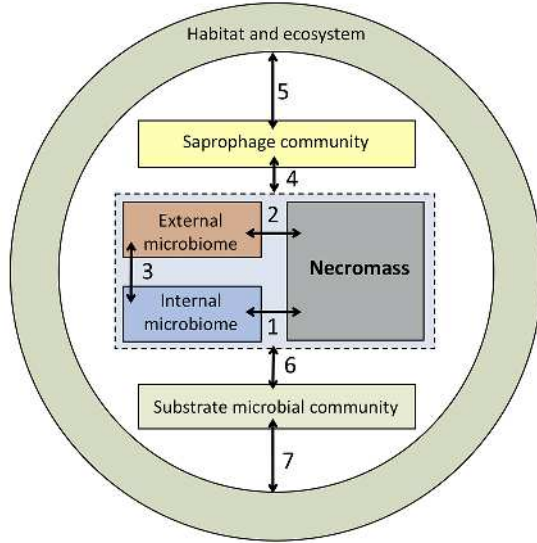
(b) **Necrobiome structure**

**Internal microbial communities** (endonecrotic microbiome)

**External microbial communities** (epinecrotic microbiome)

**Substrate microbial communities** (epigeic, soil, or water microbiome)

**Saprophage communities** (invertebrate and vertebrate consumers)

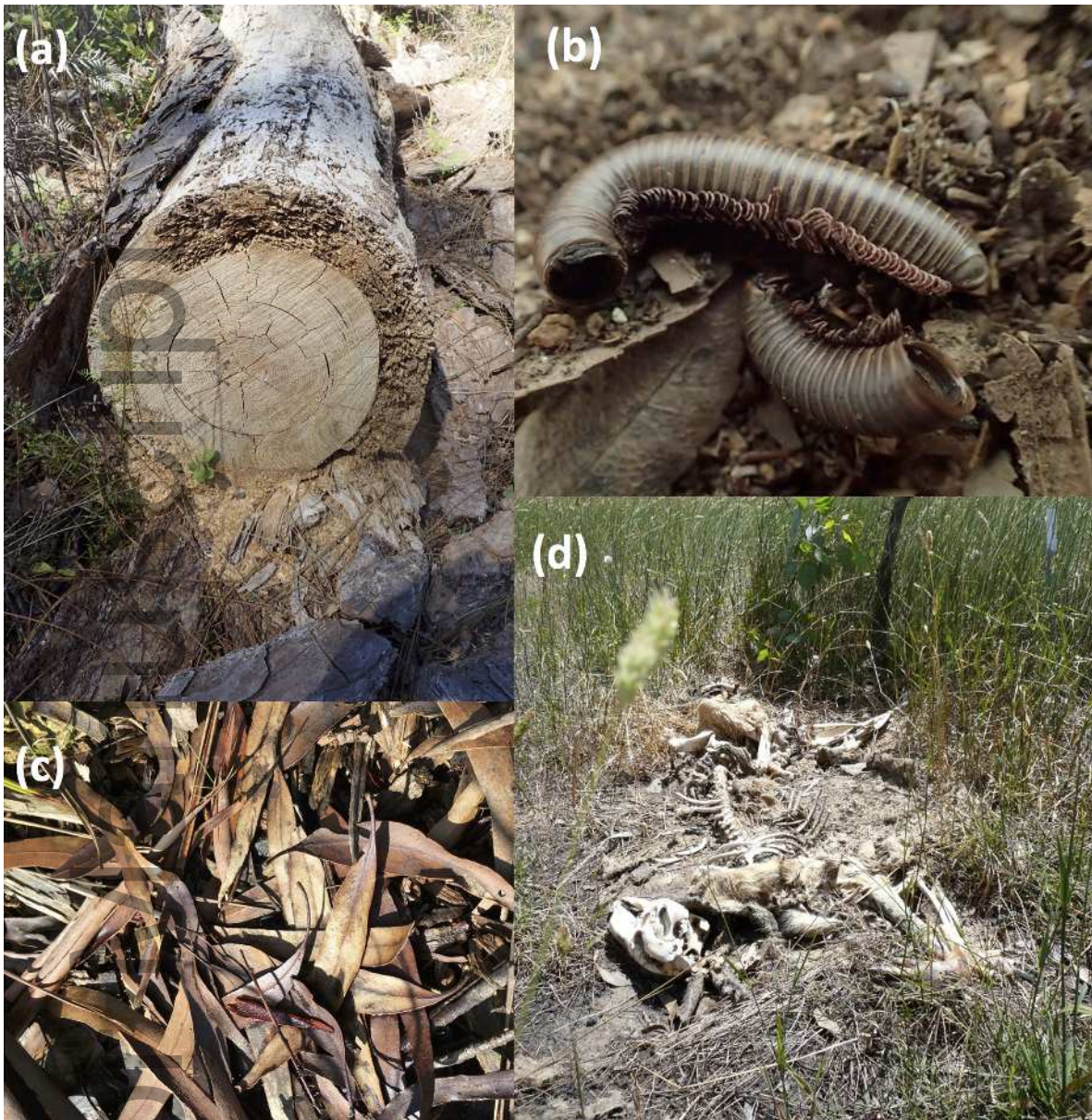


(c) **Necrobiome interactions and functions**

1. Internal microbe and element translocation, molecular catabolism
2. External surface microbe and element translocation
3. Mixing of microbes and elements via trophic and physical pathways
4. Translocation of elements and microbes via invertebrate and vertebrate trophic pathways
5. Saprophage-mediated dispersal of microbes and elements into broader ecosystem
6. Microbe and element mixing across the necromass-substrate interface via trophic and physical pathways
7. Dispersal of microbes and elements into broader ecosystem via trophic and physical pathways

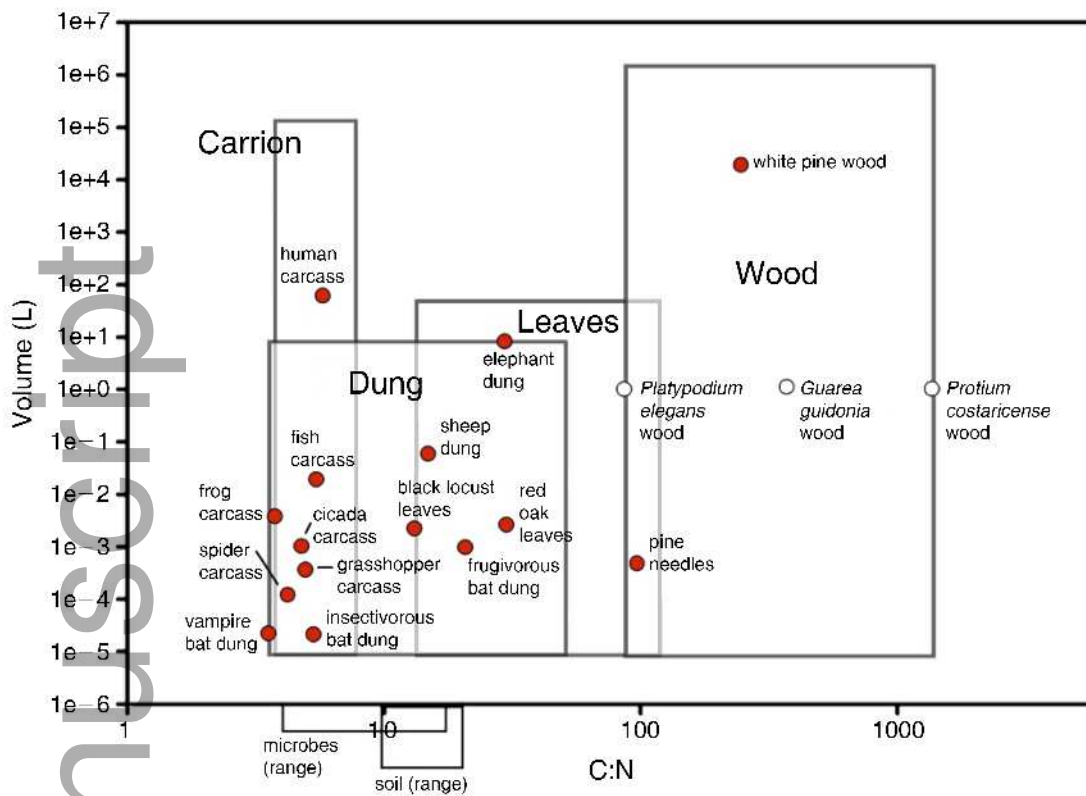
(a) **Necromass qualities** include the size, nutrient content, and digestibility of the resource

ecm\_1331\_f1.tif

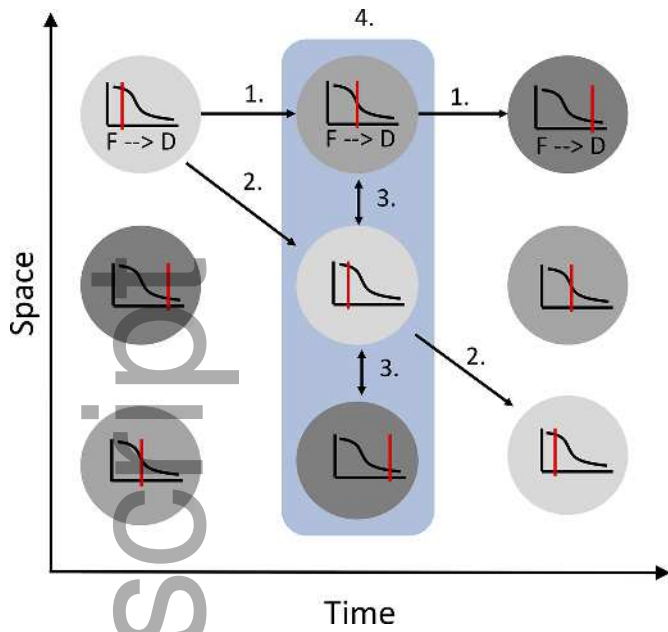


ecm\_1331\_f2.tif

Autho



ecm\_1331\_f3.tif



1. Each item of necromass decomposes from fresh (F) to dry (D), and the attendant necrobiome changes in diversity and composition through time.
2. This creates time windows for colonization and dispersal by specialist decomposers as they search for the next similar decay stage.
3. Multiple necromass resources occur at any one time, and generates a range of decay states with different necrobiomes.
4. The creates a shifting spatial mosaic of necrobiomes over time, each with unique levels of diversity, composition, and types of interactions and processes.

ecm\_1331\_f4.tif