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Chapter

The Role of the Receptor for Advanced Glycation Endproducts (RAGE) in Type 1 Diabetes: An Immune Cell Perspective

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Abstract

Type 1 diabetes (T1DM) is an autoimmune disorder resulting in destruction of the insulin producing pancreatic β -cells that reside in the Islets of Langerhans. Despite significant progress in the understanding of T1DM pathogenesis, some fundamental contributing mechanisms remain to be fully elucidated. The receptor for advanced glycation end products (RAGE) and its ligands are increasingly believed to play a role in the development of T1DM, but this is not well understood. The location of RAGE gene is shared with major T1DM genetic susceptibility loci on chromosome 6 and polymorphism of this region confers risk for T1DM. Furthermore, changes in RAGE expression on and ligand binding by immune cells, in particular T cells, are associated with pro-inflammatory and autoimmune profiles key for T1DM development. Indeed, in murine models for T1DM, targeting of RAGE or its ligands decreased onset and severity of disease including favorable immune cell profiles and infiltration and improved beta cell insulin secretory function. Further understanding of RAGE expression and signaling in immune cells in T1DM will provide valuable insights into disease pathogenesis and therapy development. This chapter will discuss what is currently known about RAGE in the immune cells integral for the pathogenesis of T1DM.

Keywords: type 1 diabetes, RAGE, RAGE ligands, advanced glycation Endproducts, T cells, APCs, NK cells, neutrophils

1. Introduction

Type 1 diabetes (T1DM) is a complex autoimmune disorder resulting from the destruction of pancreatic insulin producing β -cells due to the loss of self-tolerance. There has been a rapid increase in disease incidence worldwide and T1DM is increasingly being diagnosed in even younger individuals and in those from diverse cultural backgrounds. Insulin replacement therapy remains the only viable option for individuals with T1DM. Ensuring any new therapies are meeting a very high safety bar and are superior to insulin replacement therapy remains a challenge. Indeed, treatment with global immunosuppression, while somewhat effective, has significant side

effects and is not a practical solution to prevent T1DM which commonly develops in early life. Also, despite insulin's excellent safety profile, management of insulin replacement therapy involves multiple daily injections, pumps and continuous blood glucose monitoring or closed loop systems which are costly and complex to maintain, particularly in younger children. These therapies also bring increased risk for life-threatening high (hyperglycemia) or low (hypoglycemia) blood glucose concentrations as well as increased risk for chronic complications and shortened lifespan. Despite significant progress made in the development of therapeutics for T1DM and some remarkable results in the pre-clinical models, a successful translation into clinic is yet to occur. Due to the T1DM's multifactorial nature, the necessity to find links between genetic predisposition, immune system abnormalities and environmental triggers is becoming increasingly apparent. Therefore, exploring relationships between these contributing factors is necessary for better understanding of the disease progression and for design of the best therapeutic approaches. More recently, multiple studies have focused on the complex biology and involvement of the receptor for advanced glycation end products (RAGE) and its ligands in inflammation, autoimmunity, diabetes complications, apoptosis, and endoplasmic reticulum stress including the exploration of various ways to alleviate these. This presents an exciting new avenue for the development of targeted RAGE-related therapeutics and their translational potential from pre-clinical models to humans.

2. Preclinical murine models of T1DM

The non-obese diabetic (NOD) mouse model has been a useful tool to examine pathological mechanisms which contribute to and may be targeted in T1DM. Although there are several fundamental differences between murine and human disease [1, 2], the late timing of human disease manifestation, pancreas inaccessibility and lack of biomarkers in the peripheral blood continue to pose a significant challenge. Therefore, NOD mouse models remain instrumental in studying the disease pathophysiology and aids efforts to improve clinical translational potential of identified pathways and a first-line screening for effective therapies. Humanized mouse models, which are mice with a "human-like" immune system have proven to be an excellent platform to bridge the gap between preclinical mouse models and clinical studies by enabling researchers to assess the efficacy of treatments on human immune cells in a more physiological context than provided by cell culture [3]. In the T1DM field several excellent preclinical experimental approaches have also been proposed [4–6], although spontaneous T1DM disease development and sufficient similarity to human disease are still challenges.

3. RAGE in T1DM

RAGE is a member of the immunoglobulin superfamily It is postulated to play a role in host-pathogen defense and its expression increases during inflammation and in chronic inflammatory conditions including T1DM. The location of the RAGE gene (*AGER*) in humans is within the major histocompatibility complex on chromosome 6p21.3, a region implicated in various autoimmune disorders such as T1DM [7–9]. Similarly in mice, the *AGER* gene is located on chromosome 17 where several quantitative trait loci for T1DM have been reported [10]. RAGE is a multifunctional and promiscuous-ligand receptor, binding a wide array of ligands such as such as

advanced glycation endproducts (AGEs), high mobility group box-1 (HMGB1), S100 proteins, β -amyloid fibrils, and others [11]. Upon binding to its ligands, the cytoplasmic domain associates with adaptor proteins including TIRAP and MyD88 followed by activation of downstream signaling of various pathways including NF- κ B and STAT3 (**Figure 1**) [12].

Advanced Glycation Endproducts (AGEs) are canonical RAGE ligands but are low affinity and likely only bind RAGE when increased in concentration [13]. AGEs are formed as the result of non-enzymatic modifications to amino groups on lipids, proteins, peptides, and nucleic acids. Modern industrialization of food, flavor, and color enhancement as well as increased emulsification contribute significantly to formation of AGEs [9, 14, 15]. Prolonged exposure to AGEs has detrimental effects on β -cell function, insulin secretion and sensitivity and disease development in healthy humans and rodents [16]. Moreover, accumulation of AGEs and RAGE is associated with macro and micro vascular complications in diabetic patients [14, 17–19]. In the islet autoantibody positive individuals, the levels of circulating AGEs served as independent predictor for T1DM progression [20]. Chronic exposure of rodents to AGEs led to defects in insulin secretion and beta- cell death as well as defects in mitochondrial function supported by studies in isolated islets and beta cell lines such as MIN6N8. Following the treatment with AGE-lowering agent, the incidence of autoimmune diabetes was reduced in NOD mice [21].

HMGB1 is a non-histone chromosome protein present in all cells and may serve as a transcription factor in proinflammatory conditions [22]. Both RAGE and toll-like receptor 4 (TLR4) have been reported to serve as HMGB1 receptors, although SPR binding studies suggest that HMGB1 only binds to RAGE if associated with DNA fragments. Therapeutic potential of HMGB1 blockade has been shown in NOD mice, reducing T1DM incidence and autoimmunity [23, 24]. In newly diagnosed children



Figure 1. RAGE cellular expression in immune cells important in T1DM development.

with T1DM, HMGB1 serum concentrations were significantly higher compared to controls suggesting its potential use as an inflammatory biomarker in the disease progression [25]. More recently, it was proposed that increases in HMGB1 impairs the stability of regulatory T cells (Tregs) in NOD mice and increases production of interferon- γ (IFN- γ). In individuals with T1DM increased levels of serum HMGB1 were directly correlated with increases in IFN- γ production by Tregs. Neutralizing HMGB1 antibody rescued Treg function and suppressed autoimmunity [23]. Furthermore, hyperglycemia may contribute to the release of HMGB1 by antigen presenting, natural killer and endothelial cells as well as necrotic and apoptotic cells leading to augmented autoimmunity [26].

Another set of proteins identified as RAGE ligands is S100 calgranulin family containing over 20 members with S100A8/9 and S100B binding RAGE [27]. Although their major site of manufacture is believed to be endothelial cells, they are known to be expressed by myeloid cells such as neutrophils, but expression by lymphocytes has not been reported. This proinflammatory heterodimer is implicated in several conditions including inflammatory bowel disease and rheumatoid arthritis. Furthermore, S100A8/A9 expression is associated with tumorigenesis, suppression of DC function and accumulation of myeloid-derived suppressor cells (MDSCs) [28].

RAGE can exist as both transmembrane protein and a truncated soluble form (sRAGE). The latter is present in serum and is postulated to act as a decoy receptor by competitively binding RAGE ligands and preventing downstream signaling [11, 12]. RAGE expression tends to be low in most tissues except for the skin and alveolar epithelial cells in the lung [29]. RAGE is expressed on a number of immune cells such as T lymphocytes, neutrophils, antigen presenting cells (APCs) including macrophages, dendritic cells (DCs) and B cells as well as endothelial cells [30]. The precise role of RAGE in these cell populations as well its ability to influence cell-cell interactions and behavior still eludes us. However, RAGE continues to serve as one of the major players in inflammatory and autoimmune conditions creating a perfect opportunity to investigate these links.

3.1 RAGE and T cells

Immune tolerance involves a diverse range of processes that prevent potentially harmful immune responses against self-antigens. Given that T1DM is a T cell mediated autoimmune disease, a key event in its development is the failure in the mechanisms of central tolerance, which allows for self-reactive T lymphocytes to escape deletion by the immune system. Once in the periphery, these effector T lymphocytes can exert deleterious effects with CD4⁺ T lymphocytes believed to be important initiators and progressors of autoimmunity. Upon encounter of islet-antigen presented by dendritic cells (DCs), CD4⁺ T lymphocytes become activated which promotes and perpetuates the diabetogenic process. The reasons as to why islet autoantigens are specifically presented to the immune system by DCs to amplify immune responses remains to be fully elucidated. Migration of DCs to pancreatic lymph nodes and isletantigen presentation there amplifies recruitment and activation of CD4⁺ and CD8⁺ T lymphocytes.

An extensive pancreatic β - cell loss or loss of function is hallmark of T1DM. It is now well appreciated that autoreactive T cell are amongst key players in this process. CD4⁺ T cells are thought to be initiators of the disease providing help for B cells in "auto"antibody production as well as enhancing effector activity of CD8⁺ T cells and islet-resident macrophages. CD8⁺ effector T cells are considered to cause pancreatic islet destruction and commonly dominate islet infiltrates [31–33].

Tregs are crucial for the maintenance of the peripheral tolerance where they dampen the effects of any self-reactive cells which have escaped deletion by central tolerance. In T1DM, there is impairment in function of and/or the numbers of Tregs present. This imbalance contributes to greater pathogenic activity of effector T lymphocytes and leads to a loss of peripheral tolerance. Peripheral tolerance is the local backstop to prevent self-antigen production and delete T cells with antigen specificity for self-antigens. Thymic derived Tregs (tTregs) defined as CD4⁺CD25⁺Foxp3⁺ Tregs are responsible for suppression of effector T lymphocytes through secretion of anti-inflammatory cytokines and competition for IL-2. Both function and frequencies of this subset of Tregs is impaired in both humans and NOD mice [34]. Therefore, restoring their functionality and numbers appears to be an attractive option for preventing T1DM development and is certainly widely under investigation. Currently, there are a number of T cell-centered therapeutic approaches for treatment and prevention of T1DM under development. These include Treg enhancement and antigen-specific strategies, as well as strategies that dampen activation of T cells using CTLA-4-Ig (Abatacept) or anti-CD3 monoclonal antibody (Teplizumab) (reviewed here [35]). The latter has produced promising results in at risk antibody positive individuals significantly improving β -cell function [36, 37]. If this therapy progresses to clinic in the near future, it will make an excellent candidate for combination therapies.

RAGE expression is elevated on T cells from "at-risk" islet autoantibody positive (IAb+) individuals and is associated with progression to T1DM and increased effector function of T cells [38, 39]. The soluble isoform of RAGE (sRAGE) can competitively bind RAGE ligands and inhibit RAGE signaling. Previous studies have shown that RAGE gene (AGER) polymorphisms result in reduced sRAGE in the circulation which correlates with increased risk of T1DM and seroconversion to islet autoantibodies [9, 40]. Murine studies have demonstrated a role for RAGE in T cell activation, priming and effector function, where RAGE-deficient T cells showed reduced proliferation and production of pro-inflammatory cytokines such as IFN- γ [41]. RAGE also plays a role in DC maturation, migration, and function as well as T cell priming. In children with acute Kawasaki disease and juvenile idiopathic arthritis, RAGE facilitates recruitment and activation of leuko-cytes and sRAGE is reduced [42].

Another study explored the effects of dietary AGEs in NOD mice. Here, a T cell receptor (TCR) transgenic NOD 8.3 males with CD8⁺ T cells specific for IGRP₂₀₆₋₂₁₄, one of the main diabetogenic antigens [43], and NOD/ShiLt females and their NOD8.3 female offspring were fed low or high AGEs containing diets from conception to weaning of the offspring. The low AGEs diet resulted in improvements in insulin, proinsulin, and glucagon secretion by the islets as well as reduction in AGEs and RAGE expression in offspring islets. Furthermore, reduced level of immune cell infiltration was seen in the infants whose parents were fed with low AGEs diet in the perinatal period [16]. This is consistent with another previous intergenerational study of low AGE feeding where decreasing rates of T1DM were seen in subsequent generations during feeding with a low AGE diet [44] In the NOD model RAGE antagonism using sRAGE, significantly decreases progression to overt diabetes onset in NOD mice and preserves β -cell mass and insulin secretory function and that sRAGE therapy did not work following specific depletion of Tregs. This was evidenced through reduced islet infiltration, preservation of islet integrity and numbers as well as insulin expression. Moreover, an increased proportion of Tregs in pancreata, pancreatic lymph nodes and spleens of treated animals was demonstrated. sRAGE-induced improvements in human Tregs proliferative and suppressive ability ex vivo [45],

whilst exposure to RAGE ligands, AGEs impairs Treg suppressive capacity [45]. These observations suggest that RAGE inhibition may offer protection against future T1DM development [46].

3.2 RAGE and NK cells

Natural Killer (NK) cells are cytotoxic innate lymphocytes that bridge innate and adaptive immune systems. NK cells' importance is firmly cemented in the field of cancer immunology due to their unique ability to recognize and destroy tumors and virus infected cells. Their killing capacity is driven by the activating and inhibitory receptor-ligand interactions as well as cytolytic granules containing perforin and Granzyme B similar to the CD8⁺ T cells. Some important lectin-like activating receptors include NKG2D and KLRG1 whilst inhibitory include NKG2A and KIRs in humans and Ly49 in mice [47]. Unsurprisingly, NK cells are coming into focus in the context of T1DM as an important effector population for the disease pathogenesis. Their interaction with and ability to suppress other effector cells such as CD8+ T cells is of vital importance, particularly in the setting of autoimmunity [48].

During early human studies it was shown that proportion of NK cells was significantly lower in the peripheral blood of individuals with T1DM compared to controls. This was further linked to reduced lytic and cytotoxic capacity of NK cells and more frequent occurrence of tumors [49–51]. The dysregulation in NKG2D signaling as well as reduction in NK cell proportion was suggested to be a contributing factor to the development of T1DM [52].

HMGB1 a known RAGE ligand plays an important role in NK cell killing ability upon activation. HMGB1 released from NK cells' cytotoxic granules is very effective against oxygen-dependent cancer cells whilst those cells with anaerobic energy metabolism were resistant to HMGB1 mediated killing [53]. These observations are of importance with respect to pancreatic inflammation and associated pathologies. Narumi et al. proposed an NK cell-RAGE dependent suppression mechanism of S100A8/A9 expressing tumors It was proposed that NK cells express RAGE but not TLR4 which is also known to bind S100 family of proteins [54]. The ligation of S100A8/A9 to RAGE led to activation of NK cells, increasing their cytotoxic ability evidenced by elevated IFN- γ production, increased NKG2D activating receptor activity and amelioration of tumor growth. RAGE blockade reversed these effects, highlighting the importance of RAGE-S100 axis for suppression of tumor growth and NK cell cytotoxic ability [55]. Whilst there is as yet no direct evidence linking T1DM, NK cells and RAGE, the clear involvement of RAGE axis and NK cells in T1DM and other autoimmune and proinflammatory disorders warrants future investigation.

3.3 RAGE and neutrophils

Neutrophils are phagocytic leukocytes of innate immune system circulating in the blood in a dormant state. Their activation is initiated in the early stages of inflammation and mechanisms for pathogen clearance include release of cytotoxic granules, cytokines, and reactive oxygen species (ROS). Neutrophils possess a unique ability to form neutrophil extracellular traps (NET) when undergoing altered cell death which have unique antimicrobial properties. Moreover, neutrophils can engage with and modulate activity of other immune cells such as T and B lymphocytes, NK cells and DCs. This can lead to exacerbation of autoimmune disorders such as systemic lupus erythematosus (SLE), multiple sclerosis and autoimmune diabetes [56–58].

In NOD mice activated neutrophils are recruited to the pancreatic islets initiating development of autoimmune diabetes and facilitating recruitment of CD8⁺ T cells and DCs. Studies in NOD mice shown that migration of neutrophils plays pivotal role in disease progression and preventing it halts or reverses disease development [56, 59]. Similarly, neutrophil infiltrates were found in human pancreata prior to T1DM onset and in individuals with overt disease where there was an evidence of NET formation highlighting pathogenicity of the infiltrating neutrophils. Furthermore, neutrophils from the peripheral blood of autoantibody negative at-risk individuals, displayed a unique molecular signature with overexpression of interferon-related genes [60]. These data reinforce the importance of neutrophils in the initiation and progression of T1DM.

It was previously demonstrated that RAGE expression on human neutrophils and binding to AGEs was associated with impaired neutrophil function, in particular bacterial killing [61]. In the murine streptozotocin induced T1DM model the RAGE ligand S100 calcium-binding proteins A8/A9 (S100A8/A9) are released by neutrophils binding to RAGE on hepatic Kupffer cells leading to increase of IL-6 thrombopoietin production associated with atheroprogression in humans [62]. Albeit limited, these data support the importance of RAGE axis in effector immune populations which may be important for the development of T1DM.

3.4 RAGE and antigen presenting cells

Professional antigen presenting cells (APCs) include dendritic cells (DCs), macrophages and B cells. These cell populations have been extensively studied in T1D due to their tolerogenic and immunogenic properties and importance in therapy development [63].

DCs are categorized into conventional (cDCs), plasmacytoid (pDCs), monocytederived (moDCs) and Langerhans cells (LCs). They are important regulators of immune tolerance, can select specific T cell subsets for anergy or deletion and therefore play pivotal roles in the pathogenesis of T1DM. Their immunomodulatory abilities have been widely explored and are targets for various therapeutic strategies in development including antigen specific immunotherapy [64]. Within the pancreata of NOD mice, DCs and macrophages can be detected as early as 3 weeks of age [65, 66]. Interestingly, studies of chronic high AGE feeding of healthy rats also show the appearance of an islet specific infiltrate that is comprised primarily of macrophages [21]. Other studies suggested that plasmacytoid DCs (pDCs) are recruited to the pancreata of NOD mice where they promote diabetogenic T cell activity and initiate T1DM [56]. Many other studies highlighted the importance of DC function in disease initiation and progression (reviewed here [67, 68]). Their maturation status dictates the level and type of response exerted. The immature or tolerogenic DCs usually exhibit low levels of MHC class II and costimulatory molecules expression such as CD80 and CD86, reduced ability to stimulate T cells and produce proinflammatory cytokines, however their phagocytic and antigen processing and presentation capacity is not affected [68]. Several strategies explored immunomodulatory potential of tolerogenic DCs and maintaining them in their immature state for treatment and prevention of autoimmune disorders including T1DM (reviewed in [64, 68, 69]. DCs will continue to be in the T1DM therapy development spotlight due to their proven ability to induce T cell anergy or deletion while promoting and increasing regulatory T and B cell populations.

The exposure of immature DCs from human PBMCs to a high glucose environment and AGEs results in the upregulation of costimulatory markers, increases production of reactive oxygen species (ROS) and proinflammatory cytokines such as IL-12 and IL-6, decrease in regulatory cytokines such IL-10 and enhances expression of AGEs scavenger receptors SR-A and CD36 and RAGE [70, 71]. In another study by Ge et al., AGE-BSA treatment increased DC expression of both SR-A and RAGE but pre-treatment of DCs with RAGE neutralizing antibody halted maturation by impairing upregulation of costimulatory markers and expression of IL-12 [72]. Another study described the effects of AGEs-stimulation in the presence of the antioxidant resveratrol in immature DCs derived from healthy donor PBMCs. Pre-treatment of DCs with resveratrol prior to AGE exposure prevented their maturation, MAPK and NK-κB activation and production of inflammatory cytokines and reduced RAGE expression [73]. This once more highlights that delineating the AGE-RAGE axis in inflammatory or aging processes may have future benefit in understanding T1DM pathogenesis.

Failure of peripheral and central tolerance not only results in autoreactive T cells but also autoreactive B cells and their migration to pancreas and pancreatic lymph nodes. B cells can present autoantigens to islet specific CD4⁺ and CD8⁺ effector T cells, which in turn causes destruction of pancreatic β -cells. B cells are necessary for the development of autoimmune diabetes in NOD mice in particular autoantibody production and B cell depletion prevents development of the disease [74–76]. In newly diagnosed individuals with T1DM, B cell depletion using anti-CD20 monoclonal antibody rituximab showed initial preservation of β -cell function and the need for exogenous insulin was reduced for up to 1 year post treatment. However, following a two-year follow up period, initial improvements in C-peptide (a marker of beta cell insulin secretion/function) were diminished and the clinical trial was terminated [77, 78]. Nonetheless, despite disappointing therapeutic results, B cell importance in T1DM progression is well appreciated and several improvements to the B cell depletion therapeutic approaches have since been proposed [79].

In murine models of antibody mediated autoimmune disorders such as systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA) the absence of RAGE tended to cause reduction in germinal center B cells along with decreased anti-dsDNA autoantibody titers and increases in follicular B cells [80]. Germinal centers play important role in B cell maturation, clonal expansion, and class-switching as well an antibody production? [81]. Though these modest changes in B cells in RAGE–/– animals may not be sufficient to prevent autoimmunity, they are indicative that RAGE signaling is an important contributor to these processes [80]. Another study explored the effects of RAGE/HMGB1 interactions on activation of autoreactive B cells. In that particular study, it was concluded that HMGB1 binding promotes activation of autoreactive B cells through TLR9 rather than RAGE [82].

Macrophages act alongside DCs for antigen presentation [83, 84]. Though human pancreatic islets have macrophages present, it is unclear whether there are changes between resident and infiltrating populations in T1DM [85]. The phenotype of macrophages can in general be divided into M1/pro-inflammatory cytotoxic and M2/antiinflammatory, alternatively activated cells. However, the local microenvironment plays important role in the process of monocyte to macrophage differentiation. Human macrophages represent less than half of the APC population seen in T1DM and form a mixture of both M1 and M2 phenotypes [86]. In NOD mice APCs are represented almost entirely by macrophages and they have an intricate relationship with lymphocytes, although this may be due to limitations in assessing APC phenotypes in mice. Some studied observed that the absence of resident macrophages arrested T1DM development in NOD mice [87]. Depletion of islet-resident APCs caused complete elimination

of pancreatic lymphocyte infiltration which was restored upon re-introduction of DCs and macrophages [88, 89]. Following the depletion of macrophages, lymphocytes are also unable to initiate T1DM in the NOD-SCID adoptive transfer mouse model [90].

Monocytes, macrophages as well as RAGE- ligands axis are known players in the diabetic vascular complications. RAGE expression is associated with activation of both monocytes and macrophages which has been well explored in diabetes complications such as atherosclerosis [91, 92] and kidney disease [93–95]. Furthermore hypoxic environments can enhance monocyte adhesion and chemotaxis as well as induction of macrophage proinflammatory phenotype mediated by RAGE activity [96]. Furthermore, HMGB1 signaling through RAGE promotes secretion of IL-10 by M2 macrophages is of particular significance in the hypoxic environments of certain metastatic tumors [97]. In the individuals with T1DM suffering from retinopathy and nephropathy, the mRNA RAGE expression in monocytes was significantly reduced compared to controls. Upon exposure of monocyte cultures to glyceraldehyde-derived AGEs both mRNA and protein levels of RAGE were decreased [98]. This was a surprising result considering that upregulation of RAGE is associated with proinflammatory processes. Another study highlighted increased RAGE expression and activation in the M2 macrophages necessary for tumor vascularization and invasion [99]. Despite significant gaps in our knowledge there is unequivocal evidence that upregulation of RAGE upon binding its ligands leads to activation of proinflammatory cascades which likely impart detrimental effects in autoimmunity.

RAGE is an immunoglobulin type receptor comprised of a ligand binding V-type domain, C1 and C2 domains, transmembrane and cytoplasmic domains. C-truncated soluble RAGE can result from proteolytic cleavage or endogenous splicing of the RAGE gene, AGER. RAGE as a pattern recognition receptor can bind a wide range of ligands including AGEs, HMGB1, S100 calgranulins. Commonly, upon ligation downstream signaling via Diaphenous-1 and JAK-STAT pathways ROS production and NF-kB activation occurs stimulating inflammatory processes [11]. In T1DM, RAGE is postulated as important for various mechanisms of central and peripheral tolerance which fail to suppress the escape and activation of autoreactive T cells. This occurs in the presence of already dampened regulatory mechanisms where both function and number of Tregs are reduced This leads to further activation and expansion of pathogenic islet specific T cells aided by antigen presentation by DCs. CD8⁺ T cells migrate to pancreatic islets facilitating immune destruction and, pancreatic β -cell death and further formation and release of diabetogenic antigens. Pancreatic β -cells may also act as APCs by presenting MHC class I molecules and engaging with cytotoxic CD8⁺ T cells [100]. Inflamed and apoptotic β -cells can also release molecules that act as RAGE ligands further perpetuating the inflammatory cascade by maturation and activation of DCs and macrophages leading to pancreatic injury. NK cells can directly engage with β -cells through NKG2D-RAE1 interaction known to cause pancreatic β -cells death [101]. Furthermore, NK cells can release large quantities of cytotoxic granules and HMGB1 that may directly interact with RAGE expressed on β -cells [53]. Similar to NK cells, neutrophils can also release RAGE ligands and interact with other RAGE expressing cells including pancreatic β -cells [61].

3.5 RAGE, diabetes and COVID-19

The severe acute respiratory syndrome coronavirus (SARS-CoV)-2 pandemic has affected millions of people globally in recent years [102]. Life threatening severe lung inflammation and infections of cardiovascular and central nervous systems as well as the gastrointestinal tract have been reported during SARS-CoV-2 infection [103].

Emerging evidence suggests that individuals with diabetes have poorer prognosis when infected with SARS-CoV-2. Given that the greatest basal expression of RAGE in the body is within the lungs, it is highly likely that RAGE mediated immune processes contribute to these poorer outcomes. In hyperglycemia priming of neutrophils may result in the uncontrolled formation of NETs and release of HMGB1 further increasing vascular permeability. RAGE levels were elevated in both rodent models and humans with acute lung injury and associated with inflammasome formation and IL-1 β release [104–106] Whilst most recent SARS-CoV-2 studies focused on comorbidities associated with type 2 diabetes, a common theme of imbalanced immune responses, upregulation of RAGE ligands such as HMGB1 and S100 proteins, insulin resistance, hyperglycemia and release of pro-inflammatory cytokines is seen [107, 108]. Furthermore, several studies reported a significant increase in diabetic ketoacidosis (DKA) in recent onset T1DM during the pandemic [109–111]. These findings have important implications on future treatment and management of patients with dysglycaemia and severe respiratory conditions.

4. Conclusions

The robust evidence pointing towards involvement of RAGE and its ligands in inflammation and autoimmunity paves a new pathway towards understanding the pathophysiology of T1DM. Valuable lessons can also be learned about approaches to undertake when designing new therapies to target this axis from previous findings. Further understanding of the role of greater RAGE expression on immune cells and pancreatic islets during T1DM pathogenesis is required and is likely to be multifaceted given the myriad of inflammatory diseases which involve RAGE mediated processes. T1DM is incredibly complex and heterogeneous disorder, and it is unlikely for a 'silver bullet' therapy to emerge. This highlights the importance of exploring alternative mechanistic pathways for therapy which have not been previously considered. Indeed, it is likely that the best approach to T1DM may be combination therapies which are targeted towards various aspects of disease and provide greater coverage of the vast immune dysfunction which may be present.

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Conflict of interest

The authors declare no conflict of interest.

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References

[1] Atkinson MA, Leiter EH. The NOD mouse model of type 1 diabetes: as good as it gets? Nature Medicine. 1999;5(6):601-604

[2] Bonifacio E et al. International workshop on lessons from animal models for human type 1 diabetes: Identification of insulin but not glutamic acid decarboxylase or IA-2 as specific autoantigens of humoral autoimmunity in nonobese diabetic mice. Diabetes. 2001;**50**(11):2451-2458

[3] Yin L et al. Humanized mouse model: A review on preclinical applications for cancer immunotherapy. American Journal of Cancer Research.
2020;10(12):4568-4584

[4] Viehmann Milam AA et al. A humanized mouse model of autoimmune insulitis. Diabetes. 2014;**63**(5):1712-1724

[5] Luce S et al. Humanized mouse model to study type 1 diabetes. Diabetes. 2018;**67**(9):1816-1829

[6] Luce S et al. A humanized mouse strain that develops spontaneously immune-mediated diabetes. Frontiers in Immunology. 2021;**12**:748-679

[7] Tripodis N et al. Physical map of human 6p21.2-6p21.3: Region flanking the centromeric end of the major histocompatibility complex. Genome Research. 1998;**8**(6):631-643

[8] Burton PR et al. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. Nature. 2007;**447**(7145):661-678

[9] Forbes JM et al. Receptor for advanced glycation end-products (RAGE) provides

a link between genetic susceptibility and environmental factors in type 1 diabetes. Diabetologia. 2011;**54**(5):1032-1042

[10] Gao P et al. Genetic and molecular basis of QTL of diabetes in mouse: Genes and polymorphisms. Current Genomics. 2008;**9**(5):324-337

[11] Le Bagge S et al. Targeting the receptor for advanced glycation end products (RAGE) in type 1 diabetes. Medicinal Research Reviews. 2020;**40**(4):1200-1219

[12] Chiappalupi S et al. Hyperactivated RAGE in comorbidities as a risk factor for severe COVID-19-the role of RAGE-RAS crosstalk. Biomolecules. 2021;**11**(6):876

[13] Yamamoto Y et al. Short-chain aldehyde-derived ligands for RAGE and their actions on endothelial cells. Diabetes Research and Clinical Practice. 2007;77(Suppl. 1):S30-S40

[14] Forbes JM et al. Role of advanced glycation end products in diabetic nephropathy. Journal of the American Society of Nephrology. 2003;14(suppl. 3): S254-S258

[15] de Courten B et al. Diet low in advanced glycation end products increases insulin sensitivity in healthy overweight individuals: A doubleblind, randomized, crossover trial. The American Journal of Clinical Nutrition. 2016;**103**(6):1426-1433

[16] Borg DJ et al. Perinatal exposure to high dietary advanced glycation end products in transgenic NOD8.3 mice leads to pancreatic beta cell dysfunction. Islets. 2018;**10**(1):10-24

[17] Ramasamy R, Yan SF, Schmidt AM. Receptor for AGE (RAGE): Signaling

mechanisms in the pathogenesis of diabetes and its complications. Annals of the New York Academy of Sciences. 2011;**1243**:88-102

[18] Forbes JM et al. Advanced glycation end products as predictors of renal function in youth with type 1 diabetes. Scientific Reports. 2021;**11**(1):9422

[19] Diallo AM et al. Association between the tissue and circulating advanced glycation end-products and the microand macrovascular complications in type 1 diabetes: The DIABAGE study. Diabetes Therapy. 2022;**13**(8):1531-1546

[20] Beyan H et al. Glycotoxin and autoantibodies are additive environmentally determined predictors of type 1 diabetes: A twin and population study. Diabetes. 2012;**61**(5):1192-1198

[21] Coughlan MT et al. Advanced glycation end products are direct modulators of β -cell function. Diabetes. 2011;**60**(10):2523-2532

[22] Yang H, Wang H, Andersson U.Targeting inflammation driven by HMGB1. Frontiers in Immunology.2020;11:484

[23] Zhang J et al. Extracellular HMGB1 exacerbates autoimmune progression and recurrence of type 1 diabetes by impairing regulatory T cell stability. Diabetologia. 2020;**63**(5):987-1001

[24] Han J et al. Extracellular highmobility group box 1 acts as an innate immune mediator to enhance autoimmune progression and diabetes onset in NOD mice. Diabetes. 2008;**57**(8):2118-2127

[25] Marjanac I, Lovrić R, Barbić J. Serum levels of the high-mobility group box 1 protein (HMGB1) in children with type 1 diabetes mellitus: Case-control study. Central European Journal of Immunology. 2019;44(1):33-37

[26] Yao D, Brownlee M. Hyperglycemiainduced reactive oxygen species increase expression of the receptor for advanced glycation end products (RAGE) and RAGE ligands. Diabetes.
2009;59(1):249-255

[27] Donato R et al. Functions of S100 proteins. Current Molecular Medicine. 2013;**13**(1):24-57

[28] Huang M et al. S100A9 regulates MDSCs-mediated immune suppression via the RAGE and TLR4 signaling pathways in colorectal carcinoma. Frontiers in Immunology. 2019;**10**:2243

[29] Kierdorf K, Fritz G. RAGE regulation and signaling in inflammation and beyond. Journal of Leukocyte Biology. 2013;**94**(1):55-68

[30] Yan SF et al. Tempering the wrath of RAGE: An emerging therapeutic strategy against diabetic complications, neurodegeneration, and inflammation. Annals of Medicine. 2009;**41**(6):408-422

[31] Walker LS, von Herrath M. CD4 Tcell differentiation in type 1 diabetes.Clinical and Experimental Immunology.2016;**183**(1):16-29

[32] Phillips JM et al. Type 1 diabetes development requires both CD4+ and CD8+ T cells and can be reversed by non-depleting antibodies targeting both T cell populations. The review of diabetic studies : RDS. 2009;6(2):97-103

[33] Willcox A et al. Analysis of islet inflammation in human type 1 diabetes. Clinical and Experimental Immunology. 2009;**155**(2):173-181

[34] James CR et al. Reduced interleukin-2 responsiveness impairs the

ability of T cells to compete for IL-2 in nonobese diabetic mice. Immunology and Cell Biology. 2016;**94**(5):509-519

[35] Marfil-Garza BA et al. Progress in translational regulatory T cell therapies for type 1 diabetes and islet transplantation. Endocrine Reviews. 2020;**42**(2):198-218

[36] Sims EK et al. Teplizumab improves and stabilizes beta cell function in antibody-positive high-risk individuals. Science Translational Medicine. 2021;**13**(583):eabc8980

[37] Volfson-Sedletsky V et al. Emerging therapeutic strategies to restore regulatory T cell control of islet autoimmunity in type 1 diabetes. Frontiers in Immunology. 2021;**12**:635767

[38] Durning SP et al. The receptor for advanced glycation Endproducts drives T cell survival and inflammation in type 1 diabetes mellitus. The Journal of Immunology. 2016;**197**(8):3076-3085

[39] Akirav EM et al. RAGE expression in human T cells: A link between environmental factors and adaptive immune responses. PLoS One. 2012;7(4):e34698

[40] Salonen KM et al. A drop in the circulating concentrations of soluble receptor for advanced glycation end products is associated with seroconversion to autoantibody positivity but not with subsequent progression to clinical disease in children en route to type 1 diabetes. Diabetes/Metabolism Research and Reviews. 2017;**33**(4):e2872

[41] Moser B et al. Receptor for advanced glycation end products expression on T cells contributes to antigen-specific cellular expansion in vivo. Journal of Immunology. 2007;**179**(12):8051-8058 [42] Wittkowski H et al. Acute Kawasaki disease is associated with reverse regulation of soluble receptor for advance glycation end products and its proinflammatory ligand S100A12. Arthritis and Rheumatism. 2007;**56**(12):4174-4181

[43] Krishnamurthy B et al. Autoimmunity to both proinsulin and IGRP is required for diabetes in nonobese diabetic 8.3 TCR transgenic mice. Journal of Immunology. 2008;**180**(7):4458-4464

[44] Peppa M et al. Fetal or neonatal low-Glycotoxin environment prevents autoimmune diabetes in NOD mice. Diabetes. 2003;**52**(6):1441-1448

[45] Leung SS et al. Soluble RAGE prevents type 1 diabetes expanding functional regulatory T cells. Diabetes. 2022;**71**(9):1994-2008

[46] Manfredi AA et al. Maturing dendritic cells depend on RAGE for in vivo homing to lymph nodes. Journal of Immunology. 2008;**180**(4):2270-2275

[47] Buckle I, Guillerey C. Inhibitory receptors and immune checkpoints regulating natural killer cell responses to cancer. Cancers. 2021;**13**(17):4263

[48] Pallmer K et al. NK cells negatively regulate CD8 T cells via natural cytotoxicity receptor (NCR) 1 during LCMV infection. PLoS Pathogens. 2019;**15**(4):e1007725

[49] Wilson RG et al. Natural killer cells in insulin dependent diabetes mellitus. British Medical Journal (Clinical Research Ed.). 1986;**293**(6541):244-244

[50] Negishi K et al. Natural killer cell and islet killer cell activities in type 1 (insulin-dependent) diabetes. Diabetologia. 1986;**29**(6):352-357

[51] Negishi K et al. Natural killer cell and islet killer cell activities in human type 1 diabetes. Experimental and Clinical Endocrinology. 1987;**89**(3):345-353

[52] Qin H et al. Natural killer cells from children with type 1 diabetes have defects in NKG2D-dependent function and signaling. Diabetes. 2011;**60**(3):857-866

[53] Cerwenka A et al. HMGB1: The metabolic weapon in the arsenal of NK cells. Molecular & Cellular Oncology.2016;3(4):e1175538

[54] Leung SS, Forbes JM, Borg DJ. Receptor for advanced glycation end products (RAGE) in type 1 diabetes pathogenesis. Current Diabetes Reports. 2016;**16**(10):100

[55] Narumi K et al. Proinflammatory proteins S100A8/S100A9 activate NK cells via interaction with RAGE. Journal of Immunology. 2015;**194**(11):5539-5548

[56] Diana J et al. Crosstalk between neutrophils, B-1a cells and plasmacytoid dendritic cells initiates autoimmune diabetes. Nature Medicine. 2013;**19**(1):65-73

[57] Naegele M et al. Neutrophils in multiple sclerosis are characterized by a primed phenotype. Journal of Neuroimmunology. 2012;**242**(1-2):60-71

[58] Villanueva E et al. Netting neutrophils induce endothelial damage, infiltrate tissues, and expose immunostimulatory molecules in systemic lupus erythematosus. Journal of Immunology. 2011;**187**(1):538-552

[59] Diana J, Lehuen A. Macrophages and β -cells are responsible for CXCR2mediated neutrophil infiltration of the pancreas during autoimmune diabetes. EMBO Molecular Medicine. 2014;**6**(8):1090-1104 [60] Vecchio F et al. Abnormal neutrophil signature in the blood and pancreas of presymptomatic and symptomatic type 1 diabetes. JCI. Insight. 2018;**3**(18):e122146

[61] Collison KS et al. RAGE-mediated neutrophil dysfunction is evoked by advanced glycation end products (AGEs). Journal of Leukocyte Biology.
2002;71(3):433-444

[62] Kraakman MJ et al. Neutrophilderived S100 calcium-binding proteins A8/A9 promote reticulated thrombocytosis and atherogenesis in diabetes. The Journal of Clinical Investigation. 2017;**127**(6):2133-2147

[63] Creusot RJ, Postigo-Fernandez J, Teteloshvili N. Altered function of antigen-presenting cells in type 1 diabetes: A challenge for antigenspecific immunotherapy? Diabetes. 2018;**67**(8):1481-1494

[64] Loaiza Naranjo JD et al. A question of tolerance-antigenspecific immunotherapy for type 1 diabetes. Current Diabetes Reports. 2020;**20**(12):70

[65] Unanue ER, Ferris ST, Carrero JA. The role of islet antigen presenting cells and the presentation of insulin in the initiation of autoimmune diabetes in the NOD mouse. Immunological Reviews. 2016;**272**(1):183-201

[66] Lehuen A et al. Immune cell crosstalk in type 1 diabetes. Nature Reviews Immunology. 2010;**10**(7):501-513

[67] Sandor AM, Jacobelli J, Friedman RS. Immune cell trafficking to the islets during type 1 diabetes. Clinical and Experimental Immunology. 2019;**198**(3):314-325

[68] Khan FU et al. Dendritic cells and their immunotherapeutic potential for

treating type 1 diabetes. International Journal of Molecular Sciences. 2022;**23**(9):4885

[69] Ríos-Ríos WJ, Sosa-Luis SA, Torres-Aguilar H. Current advances in using tolerogenic dendritic cells as a therapeutic alternative in the treatment of type 1 diabetes. World Journal of Diabetes. 2021;**12**(5):603-615

[70] Lu H et al. High glucose induces upregulation of scavenger receptors and promotes maturation of dendritic cells. Cardiovascular Diabetology. 2013;**12**(1):80

[71] Buttari B et al. Advanced glycation end products of human β 2 glycoprotein I modulate the maturation and function of DCs. Blood. 2011;**117**(23):6152-6161

[72] Ge J et al. Advanced glycosylation end products might promote atherosclerosis through inducing the immune maturation of dendritic cells. Arteriosclerosis, Thrombosis, and Vascular Biology. 2005;**25**(10):2157-2163

[73] Buttari B et al. Resveratrol prevents dendritic cell maturation in response to advanced glycation end products. Oxidative Medicine and Cellular Longevity. 2013;**2013**:574029

[74] Akashi T et al. Direct evidence for the contribution of B cells to the progression of insulitis and the development of diabetes in non-obese diabetic mice. International Immunology. 1997;**9**(8):1159-1164

[75] Xiu Y et al. B lymphocyte depletion by CD20 monoclonal antibody prevents diabetes in nonobese diabetic mice despite isotype-specific differences in fc gamma R effector functions. Journal of Immunology. 2008;**180**(5):2863-2875

[76] Serreze DV et al. B lymphocytes are critical antigen-presenting cells

for the initiation of T cell-mediated autoimmune diabetes in nonobese diabetic mice. Journal of Immunology. 1998;**161**(8):3912-3918

[77] Pescovitz MD et al. Rituximab,B-lymphocyte depletion, andpreservation of beta-cell function.The New England Journal of Medicine.2009;361(22):2143-2152

[78] Pescovitz MD et al. B-lymphocyte depletion with rituximab and β -cell function: Two-year results. Diabetes Care. 2014;**37**(2):453-459

[79] Hamad AR et al. B cell-targeted immunotherapy for type 1 diabetes: What can make it work? Discovery Medicine. 2016;**21**(115):213-219

[80] Eichhorst A et al. Relevance of receptor for advanced glycation end products (RAGE) in murine antibodymediated autoimmune diseases. International Journal of Molecular Sciences. 2019;**20**(13):3234

[81] Chevalier N et al. The role of follicular helper T cell molecules and environmental influences in autoantibody production and progression to inflammatory arthritis in mice. Arthritis & Rhematology. 2016;**68**(4):1026-1038

[82] Avalos AM et al. RAGE-independent autoreactive B cell activation in response to chromatin and HMGB1/DNA immune complexes. Autoimmunity. 2010;**43**(1):103-110

[83] Sica A, Mantovani A. Macrophage plasticity and polarization: in vivo veritas. The Journal of Clinical Investigation. 2012;**122**(3):787-795

[84] Mills CD et al. M-1/M-2 macrophages and the Th1/Th2 paradigm. The Journal of Immunology. 2000;**164**(12):6166-6173

[85] Zirpel H, Roep BO. Islet-resident dendritic cells and macrophages in type 1 diabetes: In search of Bigfoot's print. Frontiers in Endocrinology. Lausanne. 2021;**12**:666795

[86] Wang YJ et al. Multiplexed In situ imaging mass cytometry analysis of the human endocrine pancreas and immune system in type 1 diabetes. Cell Metabolism. 2019;**29**(3):769-783.e4

[87] Hutchings P et al. Transfer of diabetes in mice prevented by blockade of adhesion-promoting receptor on macrophages. Nature. 1990;**348**(6302):639-642

[88] Nikolic T, Roep BO. Regulatory multitasking of tolerogenic dendritic cells - lessons taken from vitamin d3-treated tolerogenic dendritic cells. Frontiers in Immunology. 2013;4:113

[89] Nikolic T et al. Dendritic cells and macrophages are essential for the retention of lymphocytes in (peri)insulitis of the nonobese diabetic mouse: A phagocyte depletion study. Laboratory Investigation. 2005;**85**(4):487-501

[90] Jun HS et al. Absolute requirement of macrophages for the development and activation of beta-cell cytotoxic CD8+ T-cells in T-cell receptor transgenic NOD mice. Diabetes. 1999;**48**(1):34-42

[91] Soro-Paavonen A et al. Receptor for advanced glycation end products (RAGE) deficiency attenuates the development of atherosclerosis in diabetes. Diabetes. 2008;**57**(9):2461-2469

[92] Schmidt AM et al. The multiligand receptor RAGE as a progression factor amplifying immune and inflammatory responses. The Journal of Clinical Investigation. 2001;**108**(7):949-955 [93] Kanter JE, Hsu CC, Bornfeldt KE. Monocytes and macrophages as protagonists in vascular complications of diabetes. Frontiers in Cardiovascular Medicine. 2020;7:10

[94] Kawakami R et al. S100A9-RAGE Axis accelerates formation of macrophage-mediated extracellular vesicle microcalcification in diabetes mellitus. Arteriosclerosis, Thrombosis, and Vascular Biology. 2020;**40**(8):1838-1853

[95] Coughlan MT et al. RAGE-induced cytosolic ROS promote mitochondrial superoxide generation in diabetes. Journal of the American Society of Nephrology. 2009;**20**(4):742-752

[96] Zhou J et al. Intermittent hypoxia enhances THP-1 monocyte adhesion and chemotaxis and promotes M1 macrophage polarization via RAGE. BioMed Research International. 2018;**2018**:1650456

[97] Huber R et al. Tumour hypoxia promotes melanoma growth and metastasis via high mobility group Box-1 and M2-like macrophages. Scientific Reports. 2016;**6**(1):29914

[98] Miura J et al. AGE down-regulation of monocyte RAGE expression and its association with diabetic complications in type 1 diabetes. Journal of Diabetes and its Complications. 2004;**18**(1):53-59

[99] Rojas A et al. HMGB1 enhances the protumoral activities of M2 macrophages by a RAGE-dependent mechanism. Tumour Biology. 2016;**37**(3):3321-3329

[100] Hamilton-Williams EE et al. Beta cell MHC class I is a late requirement for diabetes. Proceedings of the National Academy of Sciences of the United States of America. 2003;**100**(11):6688-6693 [101] Markiewicz MA et al. RAE1ε ligand expressed on pancreatic islets recruits NKG2D receptor-expressing cytotoxic T cells independent of T cell receptor recognition. Immunity. 2012;**36**(1):132-141

[102] Chen Y, Liu Q, Guo D. Emerging coronaviruses: Genome structure, replication, and pathogenesis. Journal of Medical Virology. 2020;**92**(4):418-423

[103] Rojas A et al. Advanced-glycation end-products axis: A contributor to the risk of severe illness from COVID-19 in diabetes patients. World Journal of Diabetes. 2021;**12**(5):590-602

[104] Kim EJ et al. HMGB1 increases IL-1 β production in vascular smooth muscle cells via NLRP3 Inflammasome. Frontiers in Physiology. 2018;**9**:313

[105] De Francesco EM, Vella V,Belfiore A. COVID-19 and diabetes:The importance of controlling RAGE.Frontiers in Endocrinology. 2020;11:526

[106] Uchida T et al. Receptor for advanced glycation end-products is a marker of type I cell injury in acute lung injury. American Journal of Respiratory and Critical Care Medicine. 2006;**173**(9):1008-1015

[107] Egana-Gorrono L et al. Receptor for advanced glycation end products (RAGE) and mechanisms and therapeutic opportunities in diabetes and cardiovascular disease: Insights from human subjects and animal models. Frontiers in Cardiovascular Medicine. 2020;7:37

[108] Sandooja R, Vura N, Morocco M. Heightened ACE activity and unfavorable consequences in COVID-19 diabetic subjects. International Journal of Endocrinology. 2020;**2020**:7847526 [109] Beliard K et al. Increased DKA at presentation among newly diagnosed type 1 diabetes patients with or without COVID-19: Data from a multi-site surveillance registry. Journal of Diabetes. 2021;**13**(3):270-272

[110] Gottesman BL et al. Incidence of new-onset type 1 diabetes among US children during the COVID-19 global pandemic. JAMA Pediatrics. 2022;**176**(4):414-415

[111] Dżygało K et al. Increased frequency of severe diabetic ketoacidosis at type 1 diabetes onset among children during COVID-19 pandemic lockdown: An observational cohort study. Pediatric Endocrinology Diabetes and Metabolism. 2020;**26**(4):167-175

