



Article

Before-During-After Biomonitoring Assessment for a Pipeline Construction in a Coastal Lagoon in the Northern Adriatic Sea (Italy)

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Abstract: During 2006–2008, a pipeline was buried in Vallona lagoon in the Northern Adriatic Sea (Italy). A Before-During-After environmental monitoring programme was scheduled to monitor possible alterations. Bioaccumulation of metal(loid)s, BTs (butyltins) and HMW-PAHs (High Molecular Weight Polycyclic Aromatic Hydrocarbons), and biological responses (Condition index, air Survival—LT₅₀, Acetylcholinesterase, Micronuclei—MN, acyl-CoA oxidase, catalase, malondialdehyde—MDA, and the total oxyradical scavenging capacity—TOSCA) were investigated in Manila clams (*Ruditapes philippinarum*) from November 2005 to June 2015. *In opera* (IO) results showed higher levels of HMW-PAHs (73 ± 13 ng/g), BTs (90 ± 38 ng Sn/g) and increasing levels of Pb (6.7 ± 0.7 mg/kg) and Zn (73.6 ± 6.08 mg/kg) probably linked to works. Other contaminant alterations, especially metal(loid)s, before (AO) and after (PO) the burial, were attributed to a general condition of the area and mostly unrelated to works. In addition, LT₅₀, MN and TOSCA showed alterations, probably due to hotspots occurring in IO. TOSCA and MDA increases, right after the burial, were considered delayed responses of IO, whilst other biological responses detected later were connected to the general condition of the area. Comparisons between results of Principal Component Analyses (PCAs) highlighted partial overlapping of AO and IO, whilst PO differed only for contaminants. Visual correlations between PCAs highlighted the biomarkers' latter response.

Keywords: Po River Delta; Manila clams; polycyclic aromatic hydrocarbons; butyltins; heavy metals; metalloids; biomarkers; LNG Terminal

1. Introduction

Monitoring of biological indicators is very important and useful for detecting impacts from anthropogenic pressures on aquatic ecosystems [1]. Indeed, the use of bioindicators provide better information than physico-chemical variables on ecosystem conditions, as organisms used in biomonitoring assessment can either integrate the biogeochemical changes within the ecosystem, or respond to several stresses over a long period of time [2]. To

assess the potential toxic effects arising from environmental exposure to chemicals, information on their bioaccumulation in biota is essential, since they can provide information on their bioavailability. On the other hand, the measurement of sublethal biological effects also provides information at the molecular, cellular and functional levels, which may be considered as an early-warning tool of adverse effects of environmental stressors, such as contaminants. Indeed, biomarkers allow the assessment of fast responses to external drivers, even under less intense stress levels, and may detect integrated effects from exposure to complex mixtures [3–8]. In this context, bivalves, such as clams, represent the most suitable bioindicators in aquatic environments. They are widely distributed, sessile and filter feeding, and easy to sample. Bivalves are the most representative *taxa* among the benthic biomass in marine and transitional ecosystems, and they are responsible for fundamental ecological processes and functions. Finally, yet importantly, several bivalve species are often of economic value for their human consumption [9–13]. Bivalves are tolerant of, but sensitive to, several chemicals. They reflect environmental exposure levels, due to their limited capability to metabolize contaminants, such as heavy metals, metalloids, polycyclic aromatic hydrocarbons (PAHs) and butyltins (BTs) (see e.g., [4,14–20]).

Heavy metals and metalloids are naturally present in the Earth's crust and can be introduced in the environment through geological, biological and anthropogenic processes [21]. In aquatic environments, metal(loid)s tend to partition between aqueous and solid phases, which may consist of sediments, particulates or biota. Hydrodynamism, biochemical processes and environmental conditions, such as pH, salinity and temperature, regulate their fate in marine and coastal waters [22–24]. Heavy metals and metalloids can be divided into being essential or non-essential to organisms. The former, such as copper, iron, manganese, and zinc, are indispensable at low concentrations for multiple metabolic activities, but at higher concentrations, they can become toxic; the latter, such as arsenic, barium, cadmium, chrome, mercury, nickel and lead, are generally not required for particular metabolic functions and are toxic even at low concentrations [25].

PAHs include a series of compounds of increasing environmental impact with fused aromatic rings [26–28]. Low molecular weight (LMW) PAHs, containing two or three benzene rings show acute toxicity but not chronic toxicity so they are not broad-spectrum carcinogens for marine organisms. High molecular weight (HMW) PAHs present four to six aromatic rings and show lesser acute toxicity, but are potentially more carcinogenic [29,30]. As a result, LMW-PAHs are sometimes classified separately from HMW-PAHs. PAHs released into the environment are due to both natural and, to a much greater extent, anthropogenic sources. Among the factors arising from human activities, pyrolytic and petrogenic sources are the most important. The former includes processes such as burning fossil fuels and emissions from incinerators. On the other hand, petrogenic inputs are considered as contaminations involving the introduction in the environment of petroleum products, such as oil and fuel spills, or coming from the run-off of the road network, industrial discharges from refineries and offshore wells [31]. In petroleum products, HMW PAHs generally show very low concentrations or are undetectable while LMW PAHs and especially their branched homologues are predominant. In combustion processes, HMW PAHs generally predominate over LMW ones [29,30]. So LMW PAHs may be considered petrogenic PAHs and the HMW ones may be considered pyrogenic PAHs.

Tributyltin (TBT) is a synthetic compound and its presence in the marine environment is mainly due to its use as an active component in antifouling paints [32]. TBT was defined as “probably the most toxic chemical compound ever deliberately introduced by societies into natural waters” [33] and it has been totally banned almost all around the world by the AFS Convention since 2008 [34]. Once released into the water, due to its hydrophobicity characteristics, TBT tends to bind to suspended particulate matter. In water, it would be easily degraded by debuthylation into dibutyltin (DBT) and monobutyltin (MBT) [35], but, at the same time, TBT is easily absorbed by organisms due to its liposolubility and accumulates in sediments by binding to organic matter, thus constituting a real reservoir of contamination for several years [36].

Metal(loid)s, PAHs and BTs are problematic issues for marine ecosystems, and some of them and their derived compounds are listed as priority and/or hazardous priority substances in the European Framework Directive (WFD 2000/60/EC) [37].

Biomarkers can respond to toxic stressors with several levels of specificity. However, most of them respond to environmental stress in general [14]. Various physiological responses, including basic functions, such as changing in growth rate, feeding, and reproduction, have been measured and used as biomarkers. Condition index and air survival are thus used to provide measures of bivalves' well-being status assessments, which are relevant for ecological purposes [14,38–40]. Acetylcholinesterase (AChE) is a neurotoxicity biomarker whose inhibition may lead to behavioural impairment and thus it may affect survival, growth or reproduction of organisms [8]. AChE activity is usually inhibited by organophosphate or carbamate pesticides, but other chemicals (i.e., PAH, PCB, heavy metals, metalloids) have also been shown to modulate its activity [8,15,41–44]. The micronucleus assay is commonly used for evaluating DNA damage. The increase of micronuclei frequency (MN) is indeed a marker of genotoxic pollutants (see [15] and references therein). Peroxisomal enzymes, such as acyl-CoA oxidase (AOX), are involved in lipid metabolism through oxidative reactions and can be considered a specific biomarker of organic xenobiotic in molluscs [16]. Concerning the oxidative stress, the total oxyradical scavenging capacity toward hydroxyl and peroxy radicals (TOSC HO· and ROO·) is a useful biomarker to quantify the whole capability of tissues to neutralize different forms of oxyradicals [8,45]; while catalase (CAT) is an enzyme involved in the antioxidant system, acting on the important precursor of hydroxyl radicals H₂O₂ by reducing the reactive oxygen species (ROS) levels induced by stressors [46]. Damage related to oxidative stress could be quantified by the level of the lipidic peroxidation product malondialdehyde (MDA) [8,45].

Coastal lagoons are priority habitats as declared by the European Union in the Habitats Directive, and their environments are very vulnerable to several anthropogenic pressures [47]. Vallona lagoon is one of the lagoons composing the Delta of the Po River, the most important river in Italy (961 km long), which passes from West to East through different regions and main cities of Northern Italy and flows into the Northern Adriatic Sea. The Po River Delta is a wetland of about 400 km²; it includes several lagoon areas and it is extremely important for its natural environmental variability and complexity, as well as for its economic values [48]. All around the Delta, the use of soil is mostly dedicated to agriculture, while typical activities in the lagoons are the extensive breeding of fish species and intense shellfish production, especially of Manila clams (*Ruditapes philippinarum*) [48,49].

R. philippinarum is often used as a bioindicator in transitional waters as this species can accumulate several pollutants, but can also provide biological responses that are easy to quantify (see e.g., [17,50–52]).

During 2006–2008, the burial of a pipeline took place in the middle of the Vallona lagoon to connect the structures of the first offshore LNG (Liquefied Natural Gas) terminal in Italy, in the Northern Adriatic Sea, to facilities on land [53]. In view of this, an environmental monitoring programme was scheduled from 2005 to 2015, which consisted of three phases: (i) before; (ii) during; and (iii) after the construction of the structures [54]. The monitoring programme for the Vallona lagoon was specifically scheduled to assess the possible impacts of the burial of the pipeline on the environment.

Previous results showed that a contamination probably related to the presence of machinery and working boats during the burying activities of the pipeline occurred [55]. In this context, heavy metal, metalloid and BT bioaccumulation were assessed considering HMW-PAH concentrations in clams as a marker of impacts of the construction activities. Moreover, bioaccumulation values were related to biological responses to monitor possible alterations in *R. philippinarum* populations in the Vallona lagoon by integrating chemical and biological status of the clams.

2. Materials and Methods

2.1. Sampling Strategy

The sampling was carried out in the Vallona lagoon, a transitional water area located in the northern part of the Po River Delta, NE Italy (Figure 1). Manila clams (*R. philippinarum*) were collected by manual rake at four sampling sites located at different distances from the pipeline: L022 and L023 are the closest stations from the pipeline, while L016 and L017 are the furthest.



Figure 1. Maps of the Vallona lagoon (NE, Italy) with localizations of the four sampling sites of Manila clams (*R. philippinarum*).

Manila clams were sampled before, during and after the activities for the burial of the pipeline that were carried out from November 2006 to May 2008 in alternating phases. Dredging activities occurred in November 2006, pipeline-laying operations were in February 2007 and covering was in May 2008. In this context, November 2005, February, April, July 2006 were considered *ante operam* (AO), November 2006, February 2007, June and November 2008 as *in opera* (IO), November 2010, June 2011 and November 2011, June and November 2012, May 2013 and November 2013, and June 2014 as *post operam* (PO).

After the sample collections, the individuals of *R. philippinarum* were quickly transported to the laboratory in a refrigerated and dark container.

For chemical analyses, bivalve specimens were dissected in the laboratory and divided into pools, each consisting of the set of soft parts of several individuals in order to ensure at least three replicates for each sampling station.

The dissected organisms were then homogenized with a dispersing tool (Ultra Turrax T 25, IKA[®] Werke, Germany), lyophilized and then stored at $-20\text{ }^{\circ}\text{C}$ in the dark until the analysis. For each sample, all three replicates were subjected to the analytical procedure.

For biochemical analyses, haemolymph and digestive glands were rapidly removed from 200 specimens. For each sampling site, digestive glands were pooled in 10 samples (each consisting of tissues from twenty specimens), frozen in liquid nitrogen and maintained at $-80\text{ }^{\circ}\text{C}$; and an aliquot of haemolymph from 10 specimens was fixed with Carnoy's solution (3:1 methanol, acetic acid) and stored at $+4\text{ }^{\circ}\text{C}$ for the microscopic evaluation of micronuclei frequency.

2.2. Analysis of Polycyclic Aromatics Hydrocarbons (PAHs)

The detailed analytical procedures to analyse the High Molecular Weight (HMW) PAHs (pyrene, fluoranthene, chrysene, benzo(a)anthracene, benzo(a)pyrene, dibenzo(a,h)anthracene, benzo(b)fluoranthene, benzo(k)fluoranthene, benzo(g,h,i)perylene, indeno(1,2,3-cd)pyrene) in Manila clams of the present study are reported in [56].

2.3. Analysis of Heavy Metals and Metalloids

Metal(loid)s analysed for this study were arsenic, barium, cadmium, chromium, iron, manganese, mercury, nickel, lead, copper, and zinc.

For all metal(loid)s, the analytical procedure is the same as that reported in [56]. In addition, quantification limits were reported in [56] with the exception of Ba ($2\text{ }\mu\text{g/g d.w.}$), Mn and Zn ($1.5\text{ }\mu\text{g/g d.w.}$), and Cu ($1\text{ }\mu\text{g/g d.w.}$). The recoveries were greater than 90%, except for Zn (80%) and Cr (82%).

2.4. Analysis of Butyltins (BTs)

The determination of BTs was carried out based on modified methods adopted by [57–59].

Briefly, tetrabutyltin (TTBT) was added to 0.5–1 g of each sample as an internal standard. After acidification with hydrochloric acid, BTs were extracted twice with 0.05% *w/v* tropolone solution in methanol, in an ultrasonic bath. The supernatants were separated by centrifugation (10 min at $1000\times g$) and saline solution in Milli-Q water (20%) was added. Methanol–water fraction was extracted with 10 mL of dichloromethane for three times. Dichloromethane extracts were dried through anhydrous Na_2SO_4 and concentrated to about 2 mL under a gentle stream of N_2 . During evaporation, the solvent was changed into n-hexane. The extract was derivatized with 0.5 mL of CH_3MgCl or CH_3MgBr (3M). After elimination of the excess reagent with NH_4Cl (20% in MilliQ water), the organic phase was concentrated and cleaned-up on SPE columns filled with anhydrous Na_2SO_4 and Florisil. The eluate (n-hexane) was concentrated to 0.5 mL for injection into GC-MS/MS. All solvents used were of grade for residual pesticide analysis.

The analytical determination was carried out using a gas chromatograph (Trace GC, Thermo-Finnigan, Rodano-Milan, Italy) equipped with TriPlusAS autosampler (Thermo Fisher Scientific, Rodano-Milan, Italy) and coupled to an ion trap mass spectrometer (PolarisQ, Thermo Finnigan, Austin, TX, USA). GC was fitted with a capillary column Rxi[®]-5ms (length 25 m, i.d. 0.20 mm, film thickness 0.33 μm , Restek, Bellefonte, PA, USA). A splitless mode injection of sample (1.5 μL) was performed at $250\text{ }^{\circ}\text{C}$, with helium flow of 1.5 mL/min. The GC oven temperature was programmed as follows: $50\text{ }^{\circ}\text{C}$ held for 2 min, from $50\text{ }^{\circ}\text{C}$ to $164\text{ }^{\circ}\text{C}$ at $9\text{ }^{\circ}\text{C}/\text{min}$ and held for 5.2 min, from $164\text{ }^{\circ}\text{C}$ to $300\text{ }^{\circ}\text{C}$ at $50\text{ }^{\circ}\text{C}/\text{min}$ and held for 9.5 min. The transfer line was kept at $240\text{ }^{\circ}\text{C}$.

The quantification was performed in MS/MS mode, by employing 137 m/z for TBT, 135 m/z for DBT and 135 m/z for MBT (precursor ions: 193 m/z, 151 m/z and 165 m/z, respectively). For the purpose of quality control of the analytical data, analyses of procedure blanks and certified reference material (ERM[®]-CE 477, EC–DG JRC Institute for Reference Materials and Measurements) were carried out, with recoveries of $92 \pm 12\%$, $86 \pm 6\%$ and $88 \pm 4\%$ ($n = 8$) for TBT, DBT and MBT, respectively. For each compound, the reported concentration data are expressed both as cation per gram of dry weight (re-

spectively $(C_4H_9)_3Sn^+$ for TBT, $(C_4H_9)_2Sn^{2+}$ for DBT and $(C_4H_9)Sn^{3+}$ for MBT), and as tin (Sn) per gram of dry weight. The quantification limit of the method is 4 ng TBT/g d.w., 8 ng DBT/g d.w. and 6 ng MBT/g d.w., corresponding to 2 ng Sn/g d.w., 4 ng Sn/g d.w. and 4 ng Sn/g d.w., respectively.

2.5. Biological Responses

Condition index (CI) and Survival test (LT_{50}) were applied following the methods described by [60]. In order to calculate the two indices, forty specimens per sample per index were used as reported by [55].

Biomarkers in clam tissues were measured following protocols described in [16,61], which included spectrophotometric determination of acetylcholinesterase (AChE) in haemolymph and acyl-CoA oxidase (AOX), catalase activity (CAT) and malondialdehyde levels (MDA) in digestive glands; gas chromatographic assay of total antioxidant capacity (TOSCA) towards peroxy radicals ($ROO\cdot$) and hydroxyl radicals ($HO\cdot$) in digestive glands. Genotoxic damage, as micronuclei (MN) frequency in the haemocytes, was performed as described in [15], scoring 2000 cells with preserved cytoplasm for each specimen.

2.6. Statistical Methods

Kruskal–Wallis one way analysis of variance on ranks was applied to detect significant differences in metal(loid)s and BT bioaccumulation, and in biomarkers, comparing monitoring phases, surveys and sites. If significant, pairwise multiple comparisons were also conducted. Significant differences were highlighted with rejection of the null hypothesis at the 5% probability level ($p < 0.05$).

The Spearman's correlation test was performed between TBT and DBT + MBT, and all bioaccumulation and biological response data.

Principal component analyses (PCA) were applied to data of contaminants (HMW-PAHs, Heavy Metals, Metalloids and BTs), and of biological responses (CI, LT_{50} , AChE, AOX, CAT, MN, MDA, TOSC- $HO\cdot$, TOSC- $ROO\cdot$), respectively, to elucidate on bioaccumulation pattern, probable input sources, and biological responses. Finally, biplots of PCA results of bioaccumulations and biological responses were compared.

Software R studio [62] and STATISTICA Software, release 10 (StatSoft Inc., Tulsa, OK, USA) were used for statistical processing.

All values less than the limit of quantification (LOQ) were replaced with zeros to calculate the sum of HMW-PAHs and BTs, and were considered as $\frac{1}{2}$ LOQ in all the other calculations and analyses.

3. Results

3.1. Contaminant Body Burdens

Values of metal(loid)s and BT bioaccumulations for all surveys are reported in Tables S1–S3, respectively.

Comparing the three monitoring phases (AO vs. IO vs. PO), concentrations of all metal(loid)s and BTs were significantly different ($p < 0.05$), except for Hg ($p > 0.05$). Results of Ba, Cr, Fe, Mn, Ni, TBT and DBT showed significantly ($p < 0.05$) lower values (mean \pm s.d.) in PO (3.58 ± 0.46 mg/kg; 3.55 ± 0.53 mg/kg; 680.91 ± 86.6 mg/kg, 19.60 ± 2.58 mg/kg, 7.97 ± 0.87 mg/kg, 33 ± 2.73 ngTBT⁺/g, 8.13 ± 1 ngDBT⁺/g, respectively), whilst AO (6.17 ± 0.85 mg/kg; 5.23 ± 0.83 mg/kg; 902.15 ± 142.83 mg/kg; 29.59 ± 4.57 mg/kg; 8.46 ± 1.01 mg/kg; 164 ± 17.86 ngTBT⁺/g; 28 ± 3.48 ngDBT⁺/g, respectively) and IO (4.58 ± 0.5 mg/kg; 4.43 ± 0.46 mg/kg; 1103.32 ± 119.61 mg/kg; 32.27 ± 3.08 mg/kg; 9.71 ± 0.83 mg/kg; 185 ± 11.29 ngTBT⁺/g; 29 ± 2.44 ngDBT⁺/g, respectively) had similar results ($p > 0.05$). The values of As were significantly ($p < 0.0001$) higher in PO (22.11 ± 2.46 mg/kg) than in AO (12.95 ± 1.67 mg/kg) and IO (15.76 ± 1.93 mg/kg). Levels of Cd were significantly ($p < 0.01$) lower in IO (0.49 ± 0.05 mg/kg) than in AO (0.73 ± 0.1 mg/kg) and PO (0.59 ± 0.07 mg/kg). Concentrations of Pb resulted significantly ($p < 0.0001$) higher in IO (6.69 ± 0.66 mg/kg) rather than in the other two phases

(AO: 0.99 ± 0.13 mg/kg; PO: 1.62 ± 0.18 mg/kg), Zn showed significantly ($p < 0.05$) lower values in AO (73.56 ± 6.02 mg/kg) respect to the following phases (IO: 88.31 ± 6.63 ng/g; PO: 91.18 ± 6.61 mg/kg), and Cu had significantly ($p < 0.05$) higher values in PO (9.55 ± 0.8 mg/kg) than in the previous phases (AO: 8.67 ± 0.78 mg/kg; IO: 9.44 ± 0.53 mg/kg). MBT values were significantly ($p < 0.0001$) higher in AO (26 ± 3.02 ngMBT⁺/g) than in the following phases (IO: 7 ± 0.84 ngMBT⁺/g; PO: <6 ngMBT⁺/g).

No clear and significant differences were observed between spring–summer surveys (April–July) with respect to autumn–winter (October–February) campaigns ($p > 0.05$), and no significant differences in concentrations were observed between sampling sites ($p > 0.05$), except in site L023V, where TBT was significantly higher than L016V ($p < 0.05$), considering all the surveys.

TBT results were significantly correlated with the sum of DBT and MBT (Spearman's $R = 0.78$; $p < 0.05$), also when considering each single monitoring phase, except for IO values (AO: $R = 0.78$; $p < 0.05$; IO: $R = 0-0.17$, $p > 0.05$; PO: $R = 0.38$, $p < 0.05$).

PAH results in Manila clams were presented in [55].

3.2. Biological Responses

Biomarkers results are reported in Table S4.

Comparing all data of each phase, some significant differences ($p < 0.01$) were evidenced in biomarker results, except for AOX ($p > 0.05$). Values of MN, as well as TOSCA HO·, were significantly ($p < 0.0001$) higher in IO (0.54 ± 0.60 ‰, and 485.66 ± 55.73 UTOSC/mg prot., respectively) and PO (0.39 ± 0.54 ‰, and 545.31 ± 85.19 UTOSC/mg prot.) than in AO (0.20 ± 0.36 ‰, and 354.74 ± 60.57 UTOSC/mg prot.). Levels of AChE and CAT were significantly ($p < 0.001$) lower in PO (10.97 ± 3.63 nmol/min/mg prot., and 34.25 ± 7.24 μmol/min/mg prot.) than in the previous phases (AO: 13.62 ± 4.53 nmol/min/mg prot., and 33.93 ± 7.1 μmol/min/mg prot.; IO: 14.59 ± 2.25 nmol/min/mg prot., and 39.27 ± 6.69 μmol/min/mg prot., respectively). Values of TOSC_ROO· and MDA in PO (542.63 ± 86.75 UTOSC/mg prot., and 38.34 ± 9.37 nmol/min/g tissue) were significantly different from those of the other phases ($p < 0.01$), the first being higher than IO (421.02 ± 66.48 UTOSC/mg prot.) and the latter higher than AO (28.26 ± 7.88 nmol/min/g tissue).

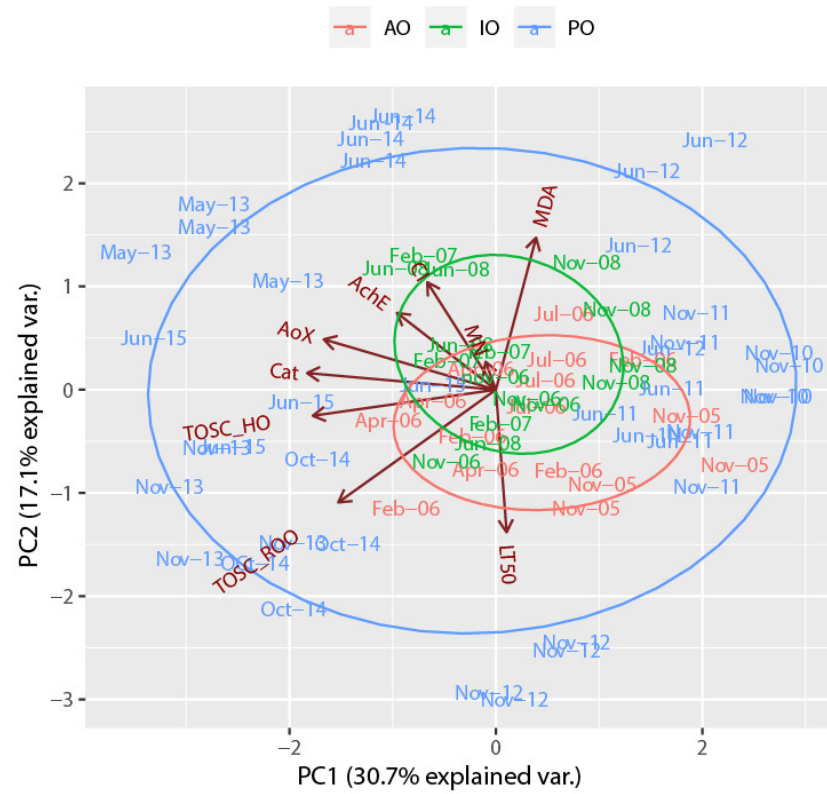
There were no significant differences ($p > 0.05$) between sampling sites. Although a light seasonality was evidenced in the AO phase [63], no clear and significant ($p > 0.05$) differences were highlighted between seasons sampled in the other phases (IO and PO). Results of PO surveys showed higher variability, especially for CAT, AOX, TOSC_HO· and TOSC_ROO· values.

Condition index and LT₅₀ results were presented in [56].

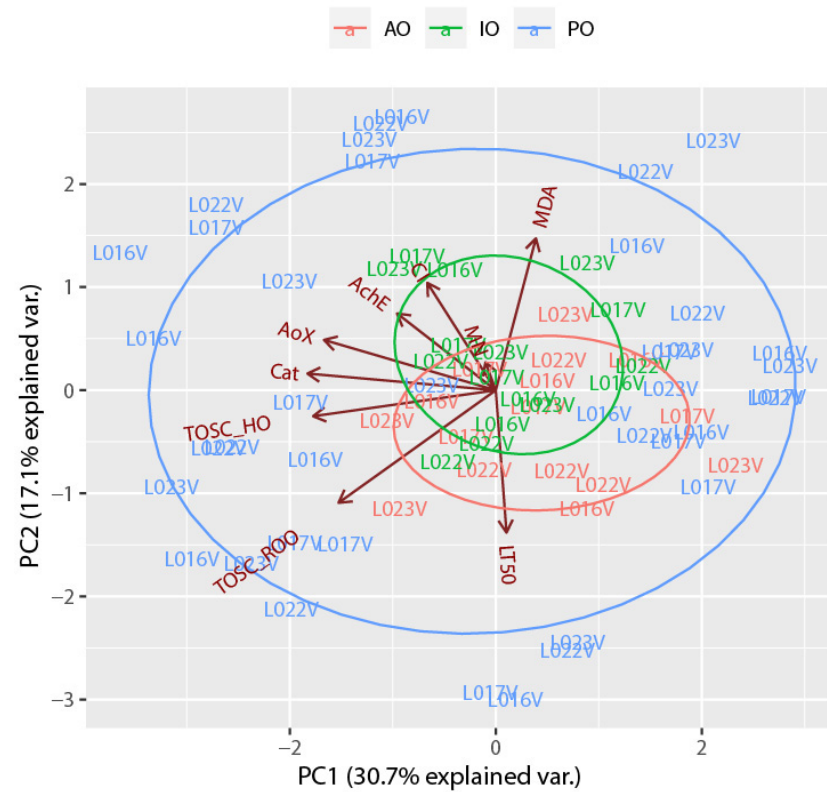
3.3. Bioaccumulation and Biological Response Patterns

Figure 2 depicts the distribution of Manila clam bioaccumulation data according to the first and second components of PCA that accounted for 32.5% and 16% of the total variance, respectively. The first component is mainly related to Ba, Cr, Fe, and Mn, whilst the second component is related to Cu, Zn, DBT, TBT, and HMW-PAHs. In this context, *post operam* samples differed from those collected during *ante operam* and *in opera*. Samples from IO were mostly characterized by HMW-PAHs, TBT and DBT, and slightly also by Cd, Hg, and MBT. Among all samples, those collected in February 2006 and 2007 were separated mostly by HMW-PAHs and BTs. Mn, Fe, Cr, Ni and Ba differed for some AO samples (L022V and L023V of November 2005). Some PO samples from November 2011 are separated from others for Cu and Zn.

PCA was also performed on biological response data as presented in Figure 3. First and second principal components explained 30.7% and 17.1% of the total variance, respectively, with the first PC being related to AOX, CAT, TOSC_HO· and TOSC_ROO·, whilst PC2 was related to MDA, TOSC_ROO·, CI and LT₅₀. Results displayed similar overlapping between



(a)



(b)

Figure 3. Biplots of the Manila clam (*R. philippinarum*) samples and the loadings of biomarkers and biological indices, labelled as (a) surveys and (b) sites, on the first two principal components. (AO = ante operam, IO = in operam, PO = post operam).

Spearman’s correlation results are reported in Table S5. Positive and significant ($p < 0.05$) correlations resulted between Cu and LT₅₀ and between Cu and all the other metal(loid)s, except for Zn. The results of Ba significantly positive correlated with all the other metal(loid)s, except for As, Cd and Hg, and also correlated with LT₅₀. Significantly positive correlations also resulted between DBT and some heavy metals (Cr, Fe, Mn, Hg, Ni, and Pb), BTs (TBT and MBT), and AChE. Then DBT, along with TBT, were also significantly ($p < 0.05$) positively correlated with HMW-PAHs. Instead, relationships were significantly ($p < 0.05$) negative between DBT and biomarkers, such as AOX and TOSC HO·, and between TBT and AOX, MDA, TOSC_HO·, TOSC_ROO·. Then, HMW-PAHs were significantly ($p < 0.05$) positively correlated with Cr, Fe, Mn, Hg, Ni, Pb, TBT, DBT, and also LT₅₀ and AChE; while significantly negative correlations were found between PAHs and AOX, TOSC_ROO· and TOSC_HO·. TOSC_ROO· and TOSC_HO· values showed negative correlations with Fe, Hg, TBT, in addition to HMW-PAH, and significantly ($p < 0.05$) positive correlations with CAT and with each other. There were positive and significant correlations between AOX and AChE and CAT, and between MN and As; while it was significantly negative between AChE and As, and between CI and all metal(loid)s (except Cr and Mn) and LT₅₀.

The results of visual correlation between two PCA biplots (bioaccumulations vs. biological responses) are reported in Table 1. Both biplots were partially overlapping between AO and IO samples. In the AO phase samples from November 2005 at sites L022V and L023V were separated for higher levels of Mn, Fe, Cr, Ni and Ba, whilst samples at the same sites (L022V and L023V) from February 2006 were grouped for HMW-PAH, TBT and DBT. Similarly in the IO phase all samples from February 2007 were separated for higher levels of HMW-PAH, TBT, DBT, Cd, Hg, and for CI; in this phase, samples from June 2008 were separated for higher values of CI. In the PO phase samples of the initial surveys were characterized first by higher values of Cu, and Zn in November 2011. Later in June 2012, the values of MDA increased, as well as the values of LT₅₀ in November 2012. Samples from May 2013 were very different for lower concentrations of Cd, Hg, and Pb and higher levels of AOX and AChE. In November 2013, samples were grouped for higher level of Cu, Zn and As as well as for increasing of TOSC_ROO· and TOSC_HO· values; samples from June 2014 were characterized by lower values of Fe, Mn, Cu, Zn, and higher of CI, and lower of LT₅₀. Samples from October 2014 were characterized by lower values of PAH, and higher values of Cu and Zn, TOSC_ROO· and TOSC_HO·. Finally, the last surveys of the study showed that in June 2015 samples were separated by lower HMW-PAH, concentrations and higher values of TOSC_HO·, CAT and AOX.

Table 1. Results of visual correlation between PCA biplots of Figures 2 and 3.

	PCA Groups of Samples	Bioaccumulation	Biological Responses
	AO vs. IO vs. PO	AO and IO partially overlapped, PO differed	AO, IO, and PO overlapped
<i>Ante operam</i>	November 2005 February 2006	Mn, Fe, Cr, Ni and Ba (L022V, L023V) (↑) HMW-PAH, TBT, DBT (↑)	-
<i>In opera</i>	February 2007 June 2008	HMW-PAH, TBT, DBT, Cd, Hg (↑) -	CI (↑) CI, (↑)
	November 2011 June 2012 November 2012	Cu, Zn (↑) -	MDA (↑) LT ₅₀ (↑)
<i>Post operam</i>	May 2013 November 2013 June 2014 October 2014 June 2015	Cd, Hg, Pb (↓) Cu, Zn, As (↑) Fe, Mn, Cu, Zn (↓) HMW-PAH (↓) Cu, Zn (↑) HMW-PAH (↓) (↑)	AOX, AChE (↑) TOSC_ROO·, TOSC_HO· (↑) CI (↑) LT ₅₀ (↓) TOSC_ROO·, TOSC_HO· (↑) TOSC_HO·, CAT, AOX (↑)

(↑) and (↓) means increase or decrease, respectively

4. Discussion

Bioaccumulation levels of metal(loid)s, PAHs and BTs, monitored before, during and after the pipeline burying, in Manila clams (*R. philippinarum*) from Vallona lagoon showed dissimilar patterns when considering the three different phases of the monitoring plan.

PAHs were significantly lower in the post operam phase than in the previous two phases. HMW-PAHs characterized the body burden pattern during the pipeline burial phase, which was mostly connected to the presence of machinery and working boats. On the contrary, in the ante operam phase, an increase in LMW-PAHs resulted as a hotspot, probably due to oil pollution, but not related to the pipeline laying [55]. Similar patterns, with AO and IO being higher than PO, resulted also in body burdens of Ba, Cr, Fe, Mn, Ni, TBT and DBT. On the contrary, MBT was higher before the pipeline burial rather than during and after the laying of it. The only substance with higher concentrations in clams during the burial of the pipeline with respect to the other two monitoring phases was Pb, whilst Zn showed an increase during in opera that did not lower in the following phase (PO). Finally, concentrations of Cu increased just at the end of the monitoring activities in PO.

Sediments, which showed HMW-PAHs always higher than LMW-PAHs, were excluded as a possible source of contamination for clams in Vallona lagoon, mostly due to the different contributions of each compound to the two matrices and low values of Biota-Sediment Accumulation Factors (BSAFs) [55]. The same conclusions were also stated by [56] about sediment contamination and bioavailability of metal(loid)s such as As, Cd, Cr, Hg, Ni and Pb. Indeed, they assumed that in Vallona lagoon, metal(loid)s were linked to mineralogical phases of the sediments, which make them not easily available to clams.

To elucidate on the bioaccumulation pattern of Manila clams, principal component analysis (PCA) was performed, considering HMW-PAHs as a marker of effects connected to the burial of the pipeline. Indeed, in previous results, exclusively focusing on PAHs, HMW-PAHs were related to the presence of machinery and working boats during the activities [55]. Performing PCA to all body burden data, three groups were identified according to the monitoring phase. However, while expecting possible overlapping between *ante* and *post operam*, and separation from *in opera*, data collected after pipeline burial (PO) clearly parted from the other two phases (AO and IO), which were partially overlaid. However, the overlapping seems to be due to a spread of data in the AO monitoring phase. Indeed, before the burial of the pipeline, Mn, Fe, Cr, Ni and Ba levels characterized some samples, whilst during the laying of the pipeline, all data differed for HMW-PAHs, TBT and DBT concentrations. On the contrary, the last phase showed more homogenous data with only some samples characterized by As, Cu and Zn. The positive and significant correlation between TBT, DBT and HMW-PAHs could also confirm their associations with boats involved in the burial of the pipeline, while the other alterations, occurring before and after, could be attributed to a general condition of the area and mostly unrelated to the burying activities. Anyway, in samples collected near the future pipeline in February 2006, TBT showed values comparable to those found during the burial activities at all sites. Therefore, a general presence in the area could also be assumed for this compound. Indeed, TBT application was banned by 2003, and the total and global ban has started only since 2008, just at the end of the burial activities that took place, even if not continuously in time, from the end of 2006 to 2008. However, it would not be surprising to find higher BT levels in clams, as old paint layers on the boat hulls continue to act as sources to the environment [64]. Since the positive and significant correlation between TBT and DBT + MBT, it could also be possible to consider the former as the principal compound introduced in environment and the latter as metabolites of TBT [65].

The environmental quality status in Vallona Lagoon was affected by heavy metals contamination [56]; however, the trend in clams was not so clear even considering the burial of the pipeline as a possible impact. Indeed, metal(loid)s mostly characterized samples before and after *in opera* activities, rather than during them, with the exception of Pb and Zn. A possible general source of metal(loid)s contamination is the Po di Levante channel, which flows north of the lagoon and its paths laps it, and collects industrial and sewage discharges

flow from the Po River, the largest river in Italy. Indeed, it runs across the Pianura Padana with a drainage of 74,000 km² and more than 16 million people living around the basin. Moreover, a lot of intensive industry and other economic activities, which represent almost half of Italian GDP, surround the Po River basin [66].

According to the Ecological Quality Standards (EQS) set by Italian Regulation (D.lgs. no. 172/2015) for the implementation of Directive 2013/39/UE, only Hg, Fluoranthene and Benzo[a]pyrene are regulated in biota with EQSs of 20 µg/kg w.w., 30 µg/kg w.w. and 5 µg/kg w.w., respectively. Most of the Hg concentrations detected in samples of the present study are above the standard, whilst levels of the two PAH compounds are all far below the EQSs. Manila clams are both bioindicators of environmental quality status and both important as an economical source for human consumptions. Comparing our results with EC Regulation 1881/2006, which set maximum levels for certain contaminants in foodstuffs, concentrations of Cd, Hg, and benzo[a]pyrene were below the limits (1 mg/kg w.w., 0.5 mg/kg w.w., and 10 ng/g w.w., respectively. Only Pb was above the limit (1.5 mg/kg w.w.), but just in samples collected in November 2006, but it quite quickly returned to below the limit after that. Limits for BTs in biota are lacking both considering regulatory levels for human and/or standards for environmental protection. However, a Tolerable Daily Intake (TDI) for TBT and DBT of 0.1 µg/kg b.w. (expressed as Sn) were set, based on the immunotoxic effects of TBT in humans, and was adopted by EFSA (European Food Safe Authority) [67]. Indeed, the main route for human exposure is diet and, among food, fish products, in particular molluscs, are those with the highest levels of concentration to which humans may be exposed (see e.g., [68–70]). Considering an average person of 70 kg b.w., no sample would exceed TDI for a daily consumption of about 85 g of Manila clams in Vallona lagoon.

Even if pollution control regulations are only focused on chemical analysis, it is evidently not sufficient to monitor and protect aquatic ecosystem. Indeed, very few substances are regulated for biota, but it must also be considered that sublethal levels of environmental toxicants may cause changes in the physiological condition of molluscs, such as clams *R. philippinarum*, which could be more hazardous for populations and with dangerous consequences for the environment [8,14]. In the present study, the biomonitoring programme also provided an assessment of physiological indices and biomarkers, to better understand the impacts of the pipeline burial on the Manila clam population.

The Condition index showed the typical seasonal fluctuation, reflecting the reproductive and physiological characteristics of the species in Adriatic lagoons [60,71]. However, significantly lower values resulted in a *post operam* phase rather than in the previous ones. Environmental conditions, such as water and air temperatures, may explain the lowering, since spring 2013 was colder than usually reported for the same season [55].

The same seasonal fluctuations were also observed for air survival time of clams, which resulted in a low capacity to survive in summer and spring surveys, and better resistance in colder seasons. However, during the burial of the pipeline, air survival showed a decrease, which made this physiological index more sensitive to the stress conditions occurring during the works [55]. Moreover, this stress condition during the works was also evidenced by biomarkers such as MN and TOSC_HO·, which showed an increasing trend probably related to bioaccumulation hotspots occurring during this phase (IO), still detectable after the burial of the pipeline (PO). The increasing of these biochemical responses, together with the increase of TOSC_ROO· and MDA during the *post operam* phase, indicated the activation of the antioxidant system to counteract the excess of ROS due to the increasing of environmental contaminants, connected with the works [5]; however, this system was not efficient enough to avoid damage at DNA and membrane lipids level. Further damage was evidenced at the nervous level, as suggested by the inhibition of AChE in the *post operam* phase with respect to the previous ones.

As reported for the bioaccumulation pattern, PCA performed on biological responses showed an overlapping of the two first phases as well as in the bioaccumulation biplot, with a greater spread of data in the PO. Although air survival time should have been the marker

of works in our *a priori* hypothesis, the IO group of data was not clearly characterized by some variables, such as in the bioaccumulation biplot, as also confirmed by no correlations between LT_{50} and the other biomarker results.

To compare bioaccumulation data and biomarker results, we first performed correlations matrix and calculated significant relations with Spearman's R. An interesting negative correlation was evidenced between As and AChE, attesting an inhibitor of acetylcholinesterase activity and neurotoxicity related to the exposure to this metalloid, in agreement with what was found in rats [72]; then a positive and expected correlation between CAT, TOSC \cdot ROO \cdot and TOSC \cdot HO \cdot was found, all being biomarkers of antioxidative stress.

To integrate the chemical and biological quality status of the clams, a visual comparison between the two PCA biplots, resulting from analyses of bioaccumulation and biomarker data, respectively, was also proposed. Both biplots depicted a partially overlapping between ante operam and *in opera* phases and a partial separation of the group of bioaccumulation data belonging to *post operam* surveys. In this context, some samples differed from the groups for bioaccumulation data, as shown during burial activity in February 2006. The trend of biological responses in the following phase seemed to be related to the trend of contaminants and human activities carried out in these phases. In the IO phase, although an apparent good condition was underlined by increasing CI values, the high concentrations of HMW-PAH, probably related to the traffic of boats involved in burial of pipeline activities, induced radical formation in clams, partially counteracted by the activation of the antioxidant system in the PO phase. This stress condition, and lipid membrane damage, appeared at the beginning of the PO phase (2012), as evidenced by the increase of MDA, associated with an increase in some heavy metals such as Cu and Zn in November 2011. It partially persisted in November 2013, with an increasing of Cu, followed by an increasing of antioxidant capability towards oxyradicals [5]. In 2014, a partial revitalization occurred as suggested by the CI increasing, associated with a decrease of heavy metals such as Fe, Mn, Cu, and Zn. In the last two surveys of PO (October 2014 and June 2015), even if a partial decrease of HMW-PAHs was detected, the antioxidant system was still active with an increase of total antioxidant capacity (TOSC \cdot ROO \cdot and HO \cdot) and CAT activity, probably relating to the increase of Cu and Zn [5,72,73].

5. Conclusions

The results of biomonitoring of Manila clams (*R. philippinarum*) before, during and after the pipeline burial in Vallona lagoon showed an increase of some contaminants in body burdens connected to the works and that sudden biological responses occurred, which could be also considered as a later reaction. However, conditions improved after the *in opera* phase as reflected by the lowering of bioaccumulation levels and better physiological responses. Indeed, some pollutants also resulted in being at significantly lower levels than before the burial of the pipeline.

Contamination levels were quite comparable to other geographical areas with medium to low pollution impact. As a result, biological responses were related to stress conditions due to working activities, even if a slight effect could be assumed right after the burial. Such a later response was better assessed by visual correlations of PCA biplots that could be adopted as a supporting method to integrate biological responses and bioaccumulation patterns with multiple parameters and compounds to consider.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/environments9070081/s1>, Table S1. Bioaccumulation results (mean \pm dev. std.) of metal(loid)s (As, Ba, Cd, Cr, Fe) in *R. philippinarum*. Table S2. Bioaccumulation results (mean \pm dev. std.) of heavy metals (Mn, Hg, Ni, Pb, Cu, Zn) in *R. philippinarum*. Table S3. Bioaccumulation results (mean \pm dev. std.) of BTs in *R. philippinarum* expressed as ng cation per gram of dry weight and as tin (Sn) per gram of dry weight Table S4. Results of biomarkers (mean \pm dev. std.) analysed in Manila clams (*R. philippinarum*) in Vallona Lagoon. Table S5. Values of R Spearman's correlations resulted significant ($p < 0.05$).

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