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Editorial: Molecular interactions between crops and phytopathogens, volume II: Rice

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Editorial on the Research Topic

Molecular interactions between crops and phytopathogens, volume II: Rice

Crop diseases reduce the yield and quality of agricultural products, seriously threatening human, animal and ecological heath (Dean et al., 2012). Rice (Oryza sativa), as the staple that feeds more than half of the world's population, is often afflicted with multiple devastating diseases, including rice blast caused by Magnaporthe oryzae, false smut caused by Ustilaginoidea virens, and sheath blight caused by Rhizoctonia solani (Ms, 2014). Besides these three aforementioned fungal diseases, there are severe bacterial diseases, including rice bacterial blight caused by Xanthomonas oryzae pv. oryzae (Xoo) and bacterial leaf streak caused by Xanthomonas oryzae pv. oryzicola (Xoc). Among these diseases, rice blast alone results in annual grain losses that are sufficient to feed more than 60 million people globally (Cook et al., 2011). Furthermore, rice false smut not only causes yield losses but also contaminates grains with mycotoxins (Sun et al., 2020), similar to notorious Fusarium head blight of wheat. To understand these diseases for better control strategies, researchers utilize bioinformatics, physiological, molecular, and biochemical tools to elucidate the complex pathogen-host interactions. In total, this Research Topic collects seven articles from researchers in the field and covers a broad range of subjects in the following three themes.

Transcriptional analysis of the *M. oryzae*-rice pathosysterm

The early availability of genome sequences of M. oryzae and rice, including the recently published high-quality genomes (Qin et al., 2021; Wang et al., 2022; Yang et al., 2022), facilitates basic research of the *M. oryzae*-rice pathosystem. Equally important is the accurate genome annotation. In this special issue, Li Z. et al. used RNA-seq data to annotate the M. oryzae genome, leading to the identification of 3,374 additional genes, most of which encode long non-coding RNAs and often show alternative splicing events that could be important for growth, conidiation, and pathogenesis. RNA-seq has long been used in studying pathogen-host interactions and uncovering immunity-related genes. From transcriptomes of the rice cultivar Nipponbare infected by three different M. oryzae strains, Liang et al. identified thousands of conserved differentially expressed genes of rice and uncovered that overexpression of one such gene enhances rice immunity against blast. The above two papers provide rich genomic and transcriptomic information for the M. oryzae-rice interaction study.

Molecular plant pathology of *M. oryzae* and *U. virens*

Three papers in this special issue cover molecular plant pathology of rice fungal pathogens M. oryzae and U. virens. Zhang et al. studied the MoPCS60 gene that encodes a peroxisomal-CoA synthase in M. oryzae. The Mopcs60 mutant is reduced in vegetative growth when the carbon source is limited to oleate and olive oil, and attenuated in virulence. The defects of the Mopcs60 mutant are likely resulted from its defective fatty acid metabolism. The importance of lipids in M. oryzae pathogenesis is also evidenced by another recent publication, which shows that the phosphatidate phosphatase MoPah1 is important for fungal development and pathogenesis (Zhao et al., 2022). In another paper collected this special issue, Liu et al. showed that the calcineurin regulator MoRCN1 is required for full fungal virulence by M. oryzae through regulating the calcineurin pathway that often plays a key role in fungal development and pathogenesis. The Morcn1 mutant is defective in appressorium formation, invasive growth, virulence, and suppressing reactive oxygen species (ROS) of the host. Mechanistically, MoRCN1 interacts directly with the calcineurin subunit A (MoCNA) in the calcineurin pathway of M. oryzae. Rice false smut recently emerges as a serious threat to rice production worldwide, particularly to those highyielding cultivars. Effective resistance genes against rice false smut have not been widely deployed. Recently, molecular studies of U. virens pathogenesis provide valuable information for

the development of novel control strategies. Recognition of fungal chitin by plant receptors triggers immune responses, which has been shown in multiple pathosystems (Gong et al., 2020). As a counteract, fungal pathogens have evolved various approaches to avoid the recognition. Li et al. elegantly showed that the chitin-binding protein UvCBP1 secreted by *U. virens* is important to attenuate chitin-triggered rice immunity including ROS burst, callose deposition, and expression of defense marker genes such as *OsPR10b*. When overexpressed, *UvCBP1* promotes fungal infection. Mechanistically, UvCBP1 competes with OsCEBiP-the rice chitin receptor-for binding to free chitin and suppresses rice immunity. These three papers together deepen our understanding of fungal-rice interactions.

Plant immune inducers and phytohormones

Chemicals are an important means in disease control, but excessive uses of traditional fungicides lead to environmental pollution and health concerns. Wang et al. identified guanineone nucleobase of the nucleic acids DNA and RNA-as a plant immune inducer from the crude extract of the endophytic fungus Paecilomyces variotii. Guanine induces a series of defense responses, including ROS burst, callose deposition, and activation of mitogen-activated protein kinases in Arabidopsis and rice, which enhance plant resistance to bacterial and fungal pathogens through a mechanism involved in ethylene and jasmonic acid signaling pathways. In another study, Xie et al. showed that salicylic acid (SA), jasmonate, and ethylene are important for rice resistance to the white tip nematode Aphelenchoides besseyi. Authors demonstrated that the SA-related genes are positively associated with rice resistance to the nematode by comparing expression of hormone-responsive genes in resistant and susceptible rice cultivars through quantitative real-time (qRT)-PCR assays. The exogenous application of analogs of the three plant hormones induces rice resistance to the nematode. In contrast, the hormone inhibitors make rice more susceptible to the nematode. The exogenous application results were further confirmed using genetic analyses. Taken together, results from these two papers demonstrate that plant hormones are important for rice resistance to bacteria, fungi and nematodes.

The results reported in this Research Topic provide key insights into pathogen-rice interactions. Yet, some cutting-edge techniques are awaiting researchers in this field to explore, including telomere-to-telomere (T2T) genome sequencing (Nurk et al., 2022), artificial intelligence, and synthetic biology, which will facilitate our deep understanding of pathogenrice interactions and hence development of novel disease control strategies.

Author contributions

All authors have participated in the article writing and have acted as coeditors of this Research Topic. All authors contributed to the article and approved the submitted version.

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References

Cook, D. C., Fraser, R. W., Paini, D. R., Warden, A. C., Lonsdale, W. M., and De Barro, P. J. (2011). Biosecurity and yield improvement technologies are strategic complements in the fight against food insecurity. *PLoS ONE* 6, e26084. doi: 10.1371/journal.pone.0026084

Dean, R., Van Kan, J. A. L., Pretorius, Z. A., Hammond-Kosack, K. E., Di Pietro, A., Spanu, P. D., et al. (2012). The Top 10 fungal pathogens in molecular plant pathology. *Mol. Plant. Pathol.* 13, 414–430. doi: 10.1111/j.1364-3703.2011.00783.x

Gong, B. Q., Wang, F. Z., and Li, J. F. (2020). Hide-and-seek: chitin-triggered plant immunity and fungal counterstrategies. *Trends Plant Sci.* 25, 805–816. doi: 10.1016/j.tplants.2020.03.006

Ms, Y. (2014). Rice sheath blight: a review of disease and pathogen management approaches. *J. Plant Pathol. Microbiol.* 5, 1. doi: 10.4172/2157-7471.1000241

Nurk, S., Koren, S., Rhie, A., Rautiainen, M., Bzikadze, A. V., Mikheenko, A., et al. (2022). The complete sequence of a human genome. *Science* 376, 44–53. doi: 10.1126/science.abj6987

Qin, P., Lu, H. W., Du, H. L., Wang, H., Chen, W. L., Chen, Z., et al. (2021). Pan-genome analysis of 33 genetically diverse rice accessions

reveals hidden genomic variations. *Cell* 184, 3542–3558. doi: 10.1016/j.cell.2021. 04.046

Sun, W., Fan, J., Fang, A. F., Li, Y. J., Tariqjaveed, M., Li, D. Y., et al. (2020). Ustilaginoidea virens: insights into an emerging rice pathogen. Annu. Rev. Phytopathol. 58, 363–385. doi: 10.1146/annurev-phyto-010820-012908

Wang, Y., Yang, L., Ma, C., Zhou, Y., Zhao, M., Bi, R., et al. (2022). Genome sequence of *Magnaporthe oryzae* EA18 virulent to multiple widely used rice varieties. *Mol. Plant Microbe Interact.* MPMI01220030A. doi: 10.1094/MPMI-01-22-0030-A

Yang, L., Zhao, M., Sha, G., Sun, Q., Gong, Q., Yang, Q., et al. (2022). The genome of the rice variety LTH provides insight into its universal susceptibility mechanism to worldwide rice blast fungal strains. *Comput. Struct. Biotechnol. J.* 20, 1012–1026. doi: 10.1016/j.csbj.2022.01.030

Zhao, J., Sun, P., Sun, Q. P., Li, R. J., Qin, Z. T., Sha, G., et al. (2022). The MoPah1 phosphatidate phosphatase is involved in lipid metabolism, development, and pathogenesis in *Magnaporthe oryzae*. *Mol. Plant. Pathol.* 23, 720–732. doi: 10.1111/mpp.13193