

Effect of gibberellic acid on the queen cell production and queen performance in colonies of *Apis mellifera* L. (Hymenoptera: Apidae)

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Received 17 - I - 2019 | Accepted 01 - VII - 2019 | Published 26 - IX - 2019

<https://doi.org/10.2085/rsea.780303>

Efecto del ácido giberélico en la producción de celdas reales y el rendimiento de la reina en colonias de *Apis mellifera* L. (Hymenoptera: Apidae)

RESUMEN. Para la abeja melífera (*Apis mellifera* L.), la recompensa por la polinización es la obtención de alimento: néctar, una solución azucarada, o polen, que contiene un alto porcentaje de proteínas y lípidos. También hay otras recompensas florales ecológicamente relevantes, las resinas, y/o componentes minoritarios, como las fitohormonas y polifenoles. En la cría de reinas de *A. mellifera* diversos factores afectan el éxito de los traslarves: la edad de las larvas, el diseño y la posición de las celdas reales artificiales, el cebado de las cúpulas con jalea real o la alimentación de la colonia que criará las reinas. El objetivo de nuestro estudio fue evaluar el efecto de dos fitohormonas, ácido giberélico (GA) y ácido indolacético (IAA) en colonias de *A. mellifera* sobre la producción de celdas reales, desarrollo de las glándulas hipofaríngeas de abejas nodrizas y desempeño de abejas reinas. Los resultados indican que el tratamiento con GA en colonias sometidas al método de semiorfandad para la cría de reinas, mejoró la eficiencia de la producción de las mismas y su rendimiento reproductivo en colonias de *A. mellifera*. Estos resultados apoyan el potencial de utilizar metabolitos vegetales como suplementos dietarios para mejorar los métodos de producción de abejas reinas y su desempeño en las colonias.

PALABRAS CLAVE. Abeja melífera. Cría de reinas. Fitomoléculas. Suplemento alimentario.

ABSTRACT. For honey bees (*Apis mellifera* L.), the reward for pollination service is obtaining food: nectar, a sugar solution; or pollen, which contains a high percentage of proteins and lipids. There are also other floral rewards ecologically relevant for pollinators, like resins, and/or minor components, such as phytohormones and polyphenols. When rearing *A. mellifera* queens, several factors affect grafting success: age of the grafted larvae, design and position of the artificial queen cells, priming queen cells with royal jelly before grafting, or nutrition of the queen rearing colony. This study aimed to evaluate the effect of two phytohormones: gibberellic acid (GA) and indole acetic acid (IAA) on queen cell production, the development of hypopharyngeal glands of nurse bees, and the performance of queen bees in colonies of

A. mellifera. The results indicate that GA positively influences queens production efficiency in vertical orphaned colonies and their reproductive performances in honey bee colonies. These results support the potential of plant metabolites as dietary supplements to improve queen bee production methods and their performance in colonies as well.

KEYWORDS. Honey bee. Phytomolecules. Queen rearing. Supplemental feeding.

INTRODUCTION

In natural environments, production of new queen bees can be driven by the aging of the old queen, its poor quality, its accidental death/loss by the beekeeper when managing the colonies, or the creation of new colonies (swarming). In the parthenogenetic life cycle of *Apis mellifera*, each fertilized egg can turn into a potential queen if, throughout its life, larvae are raised entirely on royal jelly secreted by the hypopharyngeal and mandibular glands of nurse bees (Deseyn & Billen, 2005; Spannhoff et al., 2011). This natural behavior allows beekeepers to rear queens from worker-laid larvae in initial stage.

Beekeepers have developed techniques to rear large numbers of queen bees for various practical purposes, such as altering certain genetic characteristics of the colonies, starting new colonies, increasing brood and honey production, etc. (Büchler et al., 2013). Therefore, several tools and methods are currently available for rearing queen bees, being the Doolittle Method the most popular among queen breeders. This method consists in grafting newly-hatched larvae (stage L1) selected from a mother colony and then introduced into the orphan colony for the production of potential queens (Büchler et al., 2013).

Grafting success is often modulated by four main factors: priming queen cells with royal jelly before grafting (Pickard & Kither, 1983), the design and position of the artificial queen cells (Al-Fattah et al., 2011), the age of the grafted larvae (Mahbobi et al., 2012) and/or the nutrition of the queen rearing colony (mother colonies) (Gençer et al., 2000; Mahbobi et al., 2012). For example, in colonies destined to queen rearing, feeding with a mixture of vitamins or pollen dissolved in sugar syrup positively affected the cell acceptance rate of grafted larvae for queen cell production (Gençer et al., 2000). On the other hand, a good performance of a bee colony is determined by the quality of the queen, which can be evaluated based on several characteristics, such as weight, number of ovarioles, size of the spermatheca, fecundity rate (number of eggs laid *per* day), as well as brood quality (Kovacic & Puškadija, 2016).

Nurse bees determine the fate of sibling female larvae by controlling food quantity and quality, resulting in two distinct phenotypes: queens and workers (Sagili et al., 2018). These worker bees have hypopharyngeal and mandibular glands that produce and secrete royal

jelly, a complex mixture for the growth and development of young bees (Deseyn & Billen, 2005; Spannhoff et al., 2011). It contains mainly water (~65%), crude protein (~12%), and monosaccharides (glucose and fructose) (~10%). The remainder is composed of an ether-soluble fraction of fatty acids, primarily (E)-10-hydroxy-2-decenoic acid (10HDA), a compound associated epigenetic regulation of queen-bee development (Spannhoff et al., 2011; Wang et al., 2015).

For pollinators such as honey bees the reward for pollination service is, in most cases, obtaining food in the form of nectar, sugar solution, or pollen, which contains a high percentage of proteins and lipids (Chalcoff et al., 2014). There are also other less frequent floral rewards, though ecologically relevant for pollinators, like resins, and/or minor components, such as phytohormones and polyphenols (Mao et al., 2013; Negri et al., 2015; Wang et al., 2017). In recent years, focus has been placed on the study of this type of molecules which have shown to be beneficial in bee immune system (Mao et al., 2013; Negri et al., 2015; Richardson et al., 2015). This study aimed to evaluate the effect of two phytohormones: gibberellic acid (GA) and indoleacetic acid (IAA) on queen cell production, the development of hypopharyngeal glands of nurse bees, and the performance of queen bees in colonies of *A. mellifera*.

MATERIAL AND METHODS

Biological material and characterization of bee ecotype

Field experiments were carried out in the experimental apiary of the Social Bees Research Center located in Santa Paula, National Route 226, Km 10, Mar del Plata city (-37.9348798, -57.682817), Buenos Aires, Argentina. The experiments were conducted between September 2016 and January 2017.

Field research was carried out with a local *A. mellifera* ecotype (*A. mellifera carnica* x *A. mellifera ligustica*). In order to determine bee ecotype, we performed a morphometric analysis of the experimental apiary using geometric morphometrics (Gimenez-Martínez et al., 2017). Twenty individuals were removed *per* colony, whose right posterior wing was individually excised. Photographs were taken using the Leica DM2500 optical microscope with a 4x magnification and DFC295 camera. Images were saved as .JPG files for later

analysis. All the software necessary for the morphometric analysis is available on the Internet (<http://life.bio.sunysb.edu/morph/>), and is of free use. A file was created with the images using tpsUTIL version 1.33 (Adams et al., 2004) for their analysis. Afterward, tpsDIG2 was used to manually digitalize 16 homologous landmarks (Francoy et al., 2008). For statistical analysis of the populations, a canonical variate analysis was performed in MorphoJ software (Klingenberg, 2011). Subsequently, the statistics software package PAST (version 2.17c, <http://folk.uio.no/ohammer/past>) (Hammer & Harper, 2013) was used to calculate the squared Mahalanobis distance; and distances within clusters were determined with Ward's method.

Field Trials

Trial A

We worked with three colonies of *A. mellifera* housed in Langstroth beehives with double brood boxes (two standard hives separated by a queen excluder) for queen cells production, applying the semi-orphaned method (Laidlaw & Page, 1997). This method consists in isolating queen bees in the lower hive box and keeping in the upper box sealed and open worker bee brood. The queen bee is isolated from the box above by placing a queen excluder that prevents the queen from passing through while allowing the passage of worker bees. In the middle of the upper box, there is a graft frame that holds the plastic queen cups where the 12 to 24-hour-old larvae of worker bees are grafted. Since the queen is prevented from reaching the upper box to oviposit eggs, a vertical orphaned condition is created that stimulates nurse bees to accept grafted larvae and produce queen cells. The three colonies (so-called "starter colonies") were equally managed by the same beekeeper during the three months that Trial A lasted.

A specific treatment was assigned to each starter colony: a) starter colony 1 (i1) was the control group, and was provided with 200 ml of 66% sugar syrup; b) starter colony 2 (i2) received 200 μ M of indole acetic acid (Sigma Aldrich) in 200 ml of 66% sugar syrup; c) and starter colony 3 (i3) was provided with 25 μ M of gibberellic acid (Sigma Aldrich) in 200 ml of 66% sugar syrup. The start of the trial was defined as day 0 (D0). Each starter colony received 10 applications of each treatment that were called T_n, being n the number of applications and T the treatment given. The first eight applications were made every three days and the last two on a weekly basis due to the adverse climate conditions that limited the visits to the experimental apiary (Table I).

Grafting Efficiency. The day after the first treatment was applied (Table I), each colony received 24 grafted larvae in the upper box (semi-orphaned colony). The graft consisted in placing a 12 to 24-hour-old worker bee larva in a plastic bee cup that worked as a receptor for potential queen larvae. All the larvae used for this trial came from a fourth colony defined as the mother

colony. The next graftings were systematically carried out every seven days. A total of seven graftings of 24 larvae each were performed in each colony. Five days after each grafting, the total number of accepted larvae was recorded (Table I), defined as the number of bee cups with a sealed queen cell. Percentage was calculated according to the following formula:

Percentage grafting efficiency in = (Bee cups with sealed queen cells/24) * 100

To analyze the percentage of grafting efficiency, Kruskal Wallis trial ($\alpha = 0.05$) was used.

Effect of diet on the development of the hypopharyngeal glands of nurse bees. Development of the hypopharyngeal glands is associated with the diameter of the acini (cluster of gland cells), and reflects the number of proteins produced (Deseyn & Billen, 2005). A simple method positively correlates the hypopharyngeal gland acini size to the head fresh weight of worker bees (Damiani et al., 2017). To estimate the development of the hypopharyngeal glands, on day five of treatment (13 days from the start of the trial) (Table I), nurse bees that were feeding/attending to open queen cells were sampled at each starter colony. Approximately 250 nurse bees were sampled *per* colony. In this way, bees that were seen to be attending to the queen were sampled. Samples were stored in a freezer (-20 °C) until processing. Then, a sub-sample of 50 bees was randomly chosen from each sample in the laboratory. Each bee was identified according to the treatment applied, and their head was cut with a scalpel. Heads were individually weighed in an analytical balance. The Kruskal Wallis trial ($\alpha = 0.05$) was used to analyze if there were statistical differences in the head weights sampled between treatments.

Trial B

The last grafting was used to produce virgin queens fed with 25 μ M gibberellic acid (GA) in 66% sugar syrup and 66% sugar syrup only (control). Nine (9) nuclei without queens of four frames each (two with sealed cells, one with open brood, and one with honey and pollen) were prepared for this purpose. They remained queenless for three days. The queen cells from the starter control colony (i1) were randomly inserted into three (3) nuclei; while those from the starter colony that had received the differential diet with 25 μ M GA (i3) were randomly inserted into the other six (6) nuclei. The reason why only i3 was chosen, is that it differed in cell production with respect to the control (i1).

When the virgin queens were born, they were allowed to mate freely, and once egg laying was initiated in all the nuclei, population dynamics was registered fortnightly until one of the groups completed the brood chamber. The colony strength parameters were measured according to the subjective mode (Delaplane et al., 2013). Frames with adult bees, open brood areas, sealed brood areas, pollen areas and honey areas were recorded. The method consists in the visual estimation of the parameters defined above by two or more people

	Day of experiment												
	0	1	3	6	8	9	12	15	18	22	29	36	43
T	X		X	X		X	X	X	X	X	X	X	X
G		X			X			X		X	X	X	
NS							X						

Table I. Experimental design on field showing the days of experiment: treatment application (T), grafts (G) and nurse bees sampling (NS)

as a percentage (%) of the covered area and then its conversion into units, *i.e.*, the total number of adult bees and the number of cells covered with sealed brood, open brood, honey and pollen. Beehives were managed by the same two beekeepers during the months that Trial B lasted.

RESULTS

Production of queen cells in starter colonies with a differential diet supplemented with gibberellic acid (GA) or indole acetic acid (IAA)

Regarding queen cell production in starter colonies with a differential diet, higher values were obtained in i3 colony supplemented with 25 μ M GA (mean 78%), as compared to the control bee colonies (i1), which were provided only with a 66% sugar syrup solution (average 27%) (Kruskal-Wallis trial, $p = 0.013$), and to the colony supplemented with 200 μ M IAA (i2) (approximated mean equal to 32%) (Kruskal-Wallis trial, $p = 0.031$). The i2 colony supplemented with 200 μ M IAA did not show significant differences in terms of percentage of sealed queen cells with respect to control (Kruskal-Wallis trial, $p = 1$) (Fig. 1).

Development of the hypopharyngeal glands of worker bees that attended to the queen cells of the starter colonies i1, i2, and i3

The weight of head of the worker bees that attended to the queen larvae (indirect estimator of hypopharyngeal glands development) was higher in bees from the starter colony i3, which had received the supplemental diet with 25 μ M GA, with respect to colonies i1 (Kruskal Wallis trial, $p < 0.001$) and i2 (Kruskal Wallis trial, $p < 0.001$). The weight of the worker bee heads from i2 colony, which received 200 μ M IAA, did not show significant differences with respect to the bees from the control colony (Kruskal Wallis trial, $p = 1$) (Fig. 2).

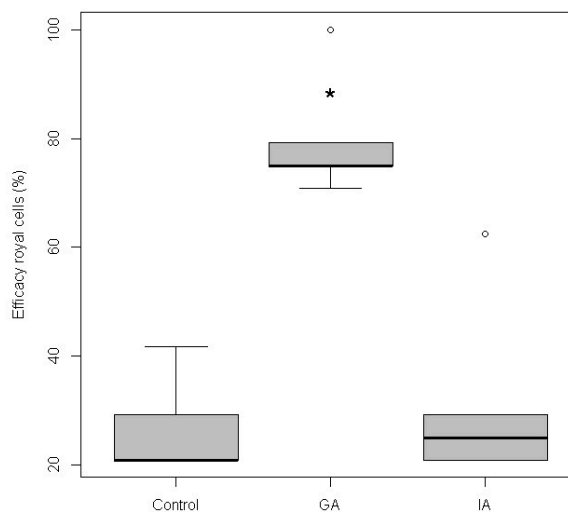


Fig. 1. Efficiency of sealed queen cells (%) in control starter colonies (sugar syrup only), supplemented with 25 μ M gibberellic acid (GA) or 200 μ M indole acetic acid (IA) (* corresponds to the significant difference)

Development of bee colonies with queens produced in control starter colonies (i1) or supplemented with 25 μ M gibberellic acid (GA) (i3)

The population dynamics of the bee colonies whose queens emerged from the colony supplemented with 25 μ M GA (i3) differed with regard to control bee colonies.

Number of adult bees of the colonies with queens from mother colony i3 was higher compared with control on days 2 and 3, respectively (two-way ANOVA, $p < 0.05$) (Fig.3 A). Number of bee brood was higher in colonies whose queens had emerged from the mother beehive i3 with regard to control colonies (ANOVA, $p < 0.001$) (Fig. 3 B).

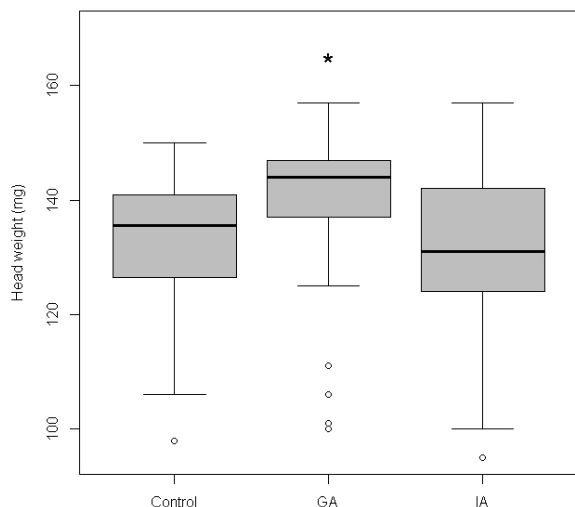


Fig. 2. Weight of head of the worker bees that attended the rearing of queens. Asterisc (*) corresponds to the significant difference.

As far as honey reserves are concerned, control colonies (queens from i1) ended (day 3) with a greater amount of honey as compared to those with queens from colony i3 (two-way ANOVA, $p = 0.001$) (Fig. 3 C). On the other hand, no significant differences were observed regarding pollen reserves between both groups of bee colonies (two-way ANOVA, $p = 0.592$) (Fig. 3 D).

DISCUSSION

To rear *A. mellifera* queens, the number of accepted larvae depends on different factors such as quality, strength, and stage of development of the starter colonies, age of workers, age of grafted larvae, regulation of humidity, temperature in the colony and food supply (nectar flow, pollen and/or supplemental feeding) to the colonies that attend to the grafted queen cells (Büchler et al., 2013). Several studies have suggested that the supplemental feeding of the starter colonies is a key factor on the acceptance rate of the grafted larvae (Krol et al., 1992; Stace & White, 1994; Gençer et al., 2000; Büchler et al., 2013). The acceptance of queen cells after grafting and cell production *per* colony increased when queen rearing colonies were fed a definite amount of iso-leucine in sugar syrup (Stace & White, 1994). In another study, a dietary supplement based on a mixture of vitamins or pollen (in sugar syrup) was tested in starter colonies, obtaining higher acceptance rates of grafted larvae for queen rearing (Gençer et al., 2000).

In our study, the first objective was to evaluate the effect of two phytohormones found naturally in honey (Wang et al., 2017): gibberellic acid (GA) and indoleacetic acid (IAA) on queen cell production in

colonies of *A. mellifera*. With this objective in mind, four colonies (one mother colony and three starter colonies) that had been analyzed by geometric morphometrics to identify bee ecotype were selected. Results indicated that the four colonies belonged to the same ecotype, a hybrid between *A. m. ligustica* and *A. m. carnica*. The production and quality of the queens (weight, egg laying capacity, etc.) are strongly influenced by genetic factors (Büchler et al., 2013). Therefore, the fact that the ecotype of the bee colonies had previously been determined was important to guarantee uniform conditions. The breeding colony that received 25 μ M GA as food supplement (mean = 78%) yielded the greatest efficiency in queen cell production as compared to the control starter colonies (mean = 27%) supplemented with 200 μ M IAA (average = 32%).

There is no literature available on the effect of supplemental feeding with plant hormones on the acceptance rate and production of queen cells. There is the possibility that the concentration of IAA applied was not sufficient to visualize an effect at colony level. Therefore, further studies should be conducted with different concentrations of IAA as dietary supplement. The greater efficiency in queen cell production of the colony supplemented with GA could be explained by the fact that the queen-destined larvae received a better-quality diet than the other colonies did. This could be supported by another parameter also measured in the starter colonies for queen rearing: the head weight of the worker bees that attended to the young larvae. Heads of the nurse bees from the colony that received 25 μ M GA were heavier, indirectly indicating a larger size of the hypopharyngeal glands (Damiani et al., 2017) of these bees compared to the bees coming from control colony to which 200 μ M IAA were supplied. One hypothesis holds that the greater size of the hypopharyngeal glands is related to the quality/quantity of royal jelly, which enhances the acceptance rate of the queen-destined larvae. Several works have dealt with dietary vitamin E supplementation for queen rearing whose conclusions could support the above-raised hypothesis. For instance, weight of the pharyngeal glands of worker bees fed with vitamin E increased by more than 47% as compared to controls (only fed with syrup) (Cheng et al., 1993). A similar study proved that vitamin supplementation not only improved the percentage of acceptance of queen cells, but also increased the royal jelly production of the workers that attended to the larvae with respect to the controls (Sahinler et al., 2005). A recent work showed that the nutritional status of the larvae is meticulously assessed and used by workers in the selection of larvae for queen rearing (Sagili et al., 2018). Authors performed a series of experiments manipulating the nutritional status of one-day-old larvae by depriving them of brood food for a four-hour period. They observed that worker bees selected larvae for rearing queens from nutritionally non-deprived larvae during that time interval. This demonstrates that honey

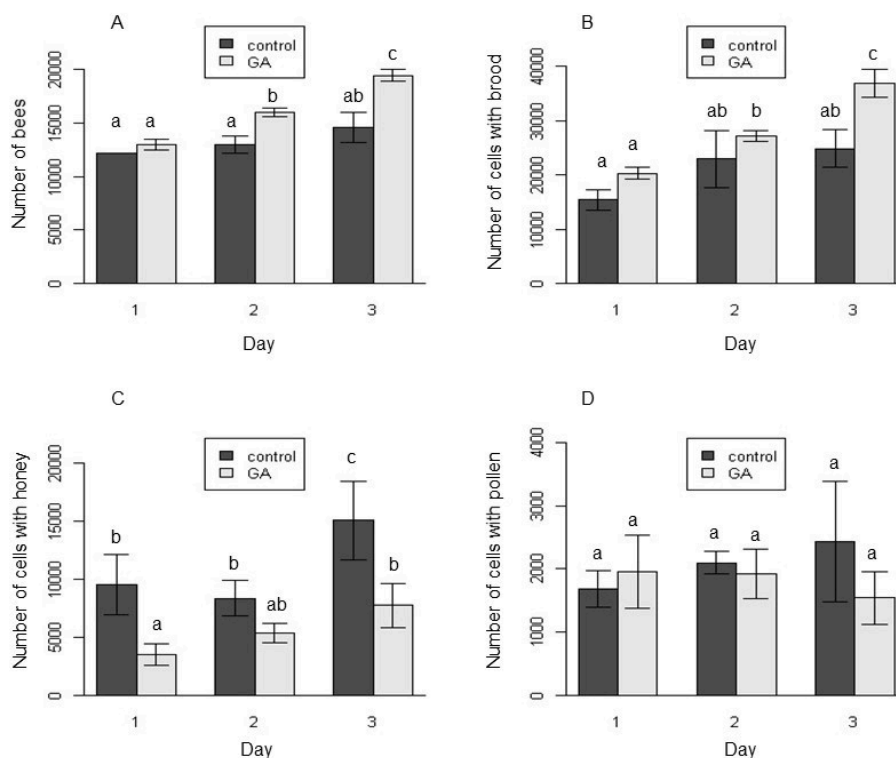


Fig. 3. Development of bee colonies with queens produced in starter hives control and supplemented with 25 μ M gibberellic acid (GA). A. number of adult bees; B. number of cells with breeding; C. number of cells with honey; D. number of cells with pollen, for days 1-2-3 corresponding to the three survey points. Significant differences are indicated by letters

bees perceive the nutritional status of the larvae, and use that information when selecting those that will be destined for queen rearing.

Studies report that a supplemental diet in starter colonies improves the queen quality (Mahbobi et al., 2012) and heavier queens performed better in the colony, which is strongly linked to greater brood production (Gençer et al., 2000). For the second part of our study, we selected the queens reared in the colonies that had received 25 μ M GA supplemental feeding (i3), due to the better responses obtained in the variables studied, along with the queens from the control colony (only fed with sugar syrup). Then, nuclei were organized with the selected queens standardized in population of adult bees, brood, and reserve in order to study queen performance in these new bee colonies. The population dynamics of the beehives with queens emerging from the starter colony and supplemented with GA showed significant differences relative to control colonies, mainly in terms of the number of adult bees and frames covered with brood. In this way, it was demonstrated that the performance of queens produced in colonies provided with GA was better than that of queens produced in control colonies.

The results indicate that gibberellic acid (GA) in colonies of *A. mellifera* under a vertical orphaned colony method for queen rearing, would influence queen production efficiency. This could be related to the differences found in the head weight of the nurse bees,

which could be ascribed to a greater development of the hypopharyngeal glands. We believe that the evidence presented here supports and reinforces the potential of plant metabolites as dietary supplements to improve queen bee production methods and their performance in honey bee colonies.

ACKNOWLEDGEMENTS

The authors would like to thank the National University of Mar del Plata (UNMdP), the Agencia Nacional de Promoción Científica y Tecnológica (ANPCYT) and the Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) for financial support.

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