

REVIEW ARTICLE

Novel approaches and perspectives in allergen immunotherapy

H. J. Hoffmann^{1,2} , E. Valovirta^{3,4,5}, O. Pfaar^{6,7} , P. Moingeon⁸, J. M. Schmid^{1,2}, S. H. Skaarup^{1,2}, L.-O. Cardell^{9,10}, K. Simonsen¹¹, M. Larché^{12,13}, S. R. Durham¹⁴ & P. Sørensen^{8,15}

¹Department of Clinical Medicine, HEALTH, Aarhus University; ²Department of Respiratory Diseases and Allergy, Aarhus University Hospital, Aarhus, Denmark; ³Department of Lung Diseases and Clinical Immunology, University of Turku, Turku, Finland; ⁴Filha, Finnish Lung Health Association, Helsinki, Finland; ⁵Terveyystalo Allergy Clinic Turku, Finland; ⁶Department of Otorhinolaryngology, Head and Neck Surgery, Medical Faculty Mannheim, Universitätsmedizin Mannheim, Heidelberg University, Mannheim; ⁷Center for Rhinology and Allergology, Wiesbaden, Germany; ⁸Research and Development, StallergenesGreer, Antony Cedex, France; ⁹Division of ENT Diseases, Department of Clinical Sciences, Intervention and Technology, Karolinska Institutet; ¹⁰Department of ENT Diseases, Karolinska University Hospital, Stockholm, Sweden; ¹¹Anergis SA, BioPole III, Epalinges, Switzerland; ¹²Clinical Immunology & Allergy and Respiratory Divisions, Department of Medicine, McMaster University; ¹³Firestone Institute for Respiratory Health, McMaster University, Hamilton, ON, Canada; ¹⁴Allergy and Clinical Immunology, National Heart and Lung Institute, Imperial College London, London, UK; ¹⁵Department of Biomedicine, HEALTH, Aarhus University & Research, Aarhus, Denmark

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Correspondence

Hans Jürgen Hoffmann, PhD, Department of Clinical Medicine, Aarhus University, Nørrebrogade 44, DK 8000 Aarhus C, Denmark and Department of Respiratory Diseases and Allergy, Aarhus University Hospital, Nørrebrogade 44, DK 8000 Aarhus C, Denmark.
Tel.: +45 78462107/6/5
Fax: +4578462110
E-mail: hjh@clin.au.dk

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Abstract

In this review, we report on relevant current topics in allergen immunotherapy (AIT) which were broadly discussed during the first Aarhus Immunotherapy Symposium (Aarhus, Denmark) in December 2015 by leading clinicians, scientists and industry representatives in the field. The aim of this symposium was to highlight AIT-related aspects of public health, clinical efficacy evaluation, mechanisms, development of new biomarkers and an overview of novel therapeutic approaches. Allergy is a public health issue of high socioeconomic relevance, and development of evidence-based action plans to address allergy as a public health issue ought to be on national and regional agendas. The underlying mechanisms are in the focus of current research that lays the ground for innovative therapies. Standardization and harmonization of clinical endpoints in AIT trials as well as current knowledge about potential biomarkers have substantiated proof of effectiveness of this disease-modifying therapeutic option. Novel treatments such as peptide immunotherapy, intralymphatic immunotherapy and use of recombinant allergens herald a new age in which AIT may address treatment of allergy as a public health issue by reaching a large fraction of patients.

Allergies have become a public health concern of pandemic proportions that affect >150 million Europeans. More alarming, their prevalence and impact are on the rise. It has been predicted

that within the next few decades, up to half of the European population may at some point in their lives experience some type of allergy (1). Allergen immunotherapy (AIT) is the only

Abbreviations

AEC, allergen exposure chamber; AIT, allergen immunotherapy; AR, allergic rhinitis; CSMS, combined symptom and medication score; DC, dendritic cell; EAACI, European Academy of Allergy and Clinical Immunology; EMA, European Medicines Agency; Fet A, fetuin A; GAP, Grazax Asthma Prevention; HRQL, Health-Related Quality of Life; IL, interleukin; ILIT, intralymphatic immunotherapy; IT, immunotherapy; NCT, nasal challenge test; OIT, oral immunotherapy; SCIT, subcutaneous immunotherapy; SLIT, sublingual immunotherapy; SPIRE, synthetic peptide immunoregulatory epitopes; SPT, skin prick test; VAS, visual analog scale; WHO, World Health Organization.

currently available medical intervention that can limit the natural course of the disease (2). Years of preclinical research, clinical trials, systematic reviews and meta-analyses have convincingly shown that AIT can achieve significant reduction of symptoms of patients, improving the allergic individuals' quality of life, changing the course of the disease and reducing the long-term costs and burden of allergies (2–5). These effects of AIT are of utmost importance from the public health point of view.

Nevertheless, despite these advances during the past century (Fig. 1) AIT usage varies across the globe with market penetration rates (the fraction of possible sales achieved) from <1% in emerging markets in Asia to 20% in the United States (6–8), so there is clearly room for improved allergy management and new innovation in the field to unleash this potential. Within this context, leading researchers from academia and industry discussed the latest advances in the field at the first Aarhus Immunotherapy Symposium on 2 December 2015. This review summarizes the main findings from the symposium on the current understanding of allergic disease mechanism, new emerging technologies, development of diagnostic and therapy monitoring technologies and disease management programmes including prevention.

Tolerance induction and prevention in allergy – a public health issue

The prevalence of allergic diseases has increased in many industrialized and urbanized countries during the last

50 years. Although the origin of allergy remains unresolved, increasing evidence indicates that modern living in an urban environment is deprived of environmental protective factors that are fundamental for normal tolerance induction (9). The concept of induction of immune tolerance has become a prime target for prevention and treatment strategies for many chronic inflammatory diseases such as allergy, asthma and autoimmunity in which dysregulation of the immune system plays an essential role.

There are few nationwide, comprehensive public health programmes on allergic disorders with defined goals and systematic follow-up (10). One practical example is the Finnish Allergy Programme 2008–2018. It is based on the idea that the allergy epidemic in modern societies is caused by inadequately developed or broken tolerance (11). Whilst allergies have traditionally been associated with industrial countries, they are also endemic in the developing world. Recent reports indicate that the prevalence of allergic disease in the Asia–Pacific region has reached the highest levels in 50 years (12), a trend also observed in tropical South-East Asia (13, 14). Although a similar set of allergens to those eliciting responses from populations in western regions has been proposed, the course, specificity and complexity of the allergic response in tropical countries may differ substantially from that in temperate zones. In a recent study of two independent cohorts of ethnic Chinese living in Singapore, the allergic response in a tropical urban environment was dominated by house dust mites (15). This monospecific IgE sensitization

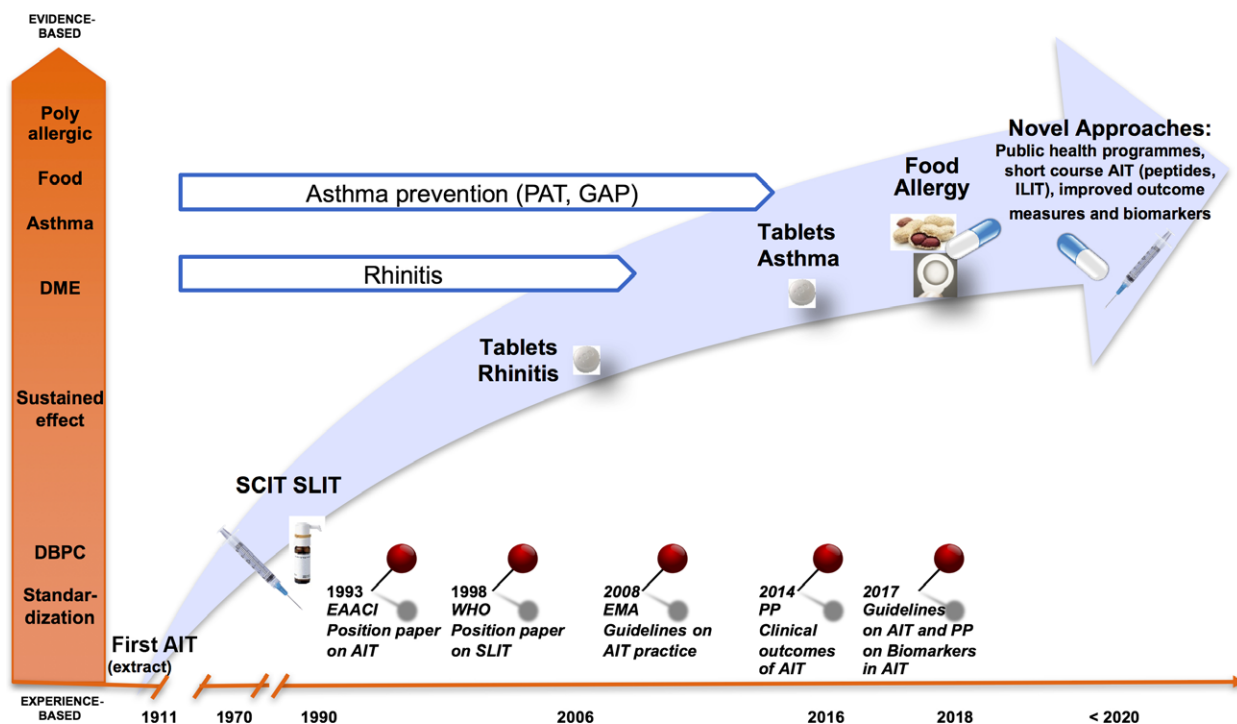


Figure 1 The evolving AIT value proposition. AIT, allergen immunotherapy; DME, disease-modifying effect; DBPC, double-blind placebo-controlled; GAP, Grazax Asthma Prevention; EMA, European Medicines Agency; EAACI, European Academy of Allergy

and Clinical Immunology; ILIT, intralymphatic immunotherapy; PAT, Prevention of Asthma Treatment; PP, position paper; SCIT, subcutaneous immunotherapy; SLIT, sublingual immunotherapy; WHO, World Health Organization.

translates into increased prevalence of allergic airway diseases, which now impact a large proportion of the population in Singapore. It presents a unique opportunity to treat and manage these patients with AIT.

As the first allergic sensitization is a strong predictor for ensuing sensitizations, it should be treated with (i) AIT in addition to (ii) individual guided self-management, (iii) symptomatic treatment and possibly with (iv) avoidance of allergens that clearly worsen the symptoms. The WHO AIT guidelines from 1998 state that AIT acts globally on IgE-mediated inflammation in various organs (2). Allergen immunotherapy could be administered in primary health care setting with enough know-how according to the national AIT guidelines based on international guidelines, as well as student and occupational health care where feasible (16). Subcutaneous immunotherapy (SCIT) has proven value but requires specialist supervision due to associated risks of severe allergic reactions. The sublingual route is an effective and safer alternative suitable for daily self-administration.

From the public health point of view, an important fact is that allergic rhinitis is a risk factor for asthma. Allergen immunotherapy is considered to prevent asthma in patients treated with AIT for allergic rhinitis in clinical trials (17) as well as in real-world settings (18). Confounding by indication cannot be excluded but would lead to an underestimation of the true preventive effects of AIT.

The Grazax Asthma Prevention (GAP) trial, investigating the preventive effect on asthma development of grass AIT tablet in children aged from 5 to 12 years with grass pollen-induced allergic rhinitis (AR), is the first double-blind, placebo-controlled randomized trial to assess the preventive effects of AIT (19). Although the trial did not achieve its primary endpoint of preventing asthma, the asthma diagnosis criteria used in the time to onset analysis were rather rigid and dependent on objective tests of reversibility. Even so children treated with the grass tablet AIT had a significantly reduced risk of experiencing asthma symptoms and a reduced use of asthma medication not only during the 3 years of AIT but also for 2 years after discontinuation. They also had a beneficial effect on their allergic nasal and eye symptoms throughout the 5 year study period (20). Thus, the results of the GAP trial confirmed the disease-modifying effect of grass tablet AIT on grass pollen-induced asthma and rhinoconjunctivitis in children.

Early introduction of allergenic food has been shown to be good primary prevention of peanut allergy (21), which may face difficulties when introduced in primary care (22). Despite the lack of effect in the intention-to-treat analysis and reluctance to introduce solids as early as at 3 months of age, the reduction in prevalence by 2/3 in the per-protocol analysis when ingesting 2 g of peanut or egg protein is encouraging. Oral immunotherapy (OIT) of IgE-mediated food allergy has been shown to desensitize individuals at risk of or having experienced severe allergic reactions against peanut (23), egg (24, 25) and cow's milk (26). However, these oral desensitization protocols were experimental, had low success rates, carried significant risks of inducing anaphylaxis and, in contrast to immunotherapy for inhalant allergens, have not been shown to induce long-term tolerance after discontinuation.

Although the data are encouraging, current guidelines confine OIT for food to research protocols in the hands of trained allergy specialists and are not recommended for routine clinical practice.

Allergen immunotherapy has not yet received adequate attention from European institutions, and thus far, too many allergic patients in the general population remain unaware of the benefits of AIT (3). It is time to re-evaluate the allergy paradigm and implement new kinds of actions as allergic individuals are becoming a significant minority of Western populations, and their number is increasing worldwide. National and regional action plans, such as the Finnish Allergy Programme 2008–2018 (11), are needed to meet this challenge.

Evaluation of efficacy in AIT trials

To advocate that AIT resolves the unmet need of allergy in public health, good, standardized assessment of clinical efficacy is mandatory. Throughout the last decade, increasing emphasis has been put on clear guidance in standardization of clinical trials in the field of AIT (27, 28). Besides these academic positions, the European Medicines Agency (EMA) (29) and the Center for Drug Evaluation and Research of the US Food and Drug Administration (30) have published regulatory guidelines which outline standards for the clinical development and documentation of new products in clinical trials. The relevant EMA guideline (29) requires that the primary endpoint in AIT trials has to 'reflect both, symptom severity as well as the intake of rescue medication' and, moreover, specifies several secondary outcome parameters. However, it also highlights that at present 'no validated symptom score exists' and 'different approaches to combine symptom score and intake of rescue medication are possible'. Recently, a Task Force initiative from the European Academy of Allergy and Clinical Immunology (EAACI) recommended a standard for the primary endpoint for future randomized controlled trials in AIT for allergic rhinoconjunctivitis, the 'combined symptom and medication score' (CSMS; Table 1 (31)). Besides these parameters, several secondary endpoints such as Health-Related Quality of Life Questionnaires, visual analog scales or 'Global Assessments' are also used in AIT trials (32, 33). Among these secondary endpoints, allergen provocation tests (conjunctival, bronchial or nasal provocations) can be performed that directly measure the allergen sensitivity (and possible changes throughout the course of AIT) in the allergic target organ (29). Therefore, the EMA also recommends provocation tests to be used as primary endpoints in dose-finding trials of AIT (29) (EMA register of clinical trials at www.clinicaltrialsregister.eu). For pivotal phase III trials, these tests 'can give additional information but are no surrogate markers and cannot replace the measurement of clinical symptoms' (29). Exposure in an allergen exposure chamber (AEC), through standardized protocols under controlled environmental conditions (temperature, humidity), is an attractive alternative. Their technical validation underlines their principal advantage compared to other challenge methods (34). These models have been used

Table 1 The Task Force recommendation providing (A) a homogeneous terminology for nasal and conjunctival symptoms using the six organ-related categories in the daily symptom score (dSS), (B) a stepwise use of rescue medication summed in the daily medication score (dMS) and (C) a scoring system for a combined symptom and medication score (CSMS), which is based on an equal weight of the dSS and of the dMS (reproduced from reference 31 with permission.)

(A) Symptom score		
Nasal symptoms	(Score 0–3)	0 = no symptoms 1 = mild symptoms (sign/symptom clearly present, but minimal awareness; easily tolerated) 2 = moderate symptoms (definite awareness of sign/symptom that is bothersome but tolerable) 3 = severe symptoms (sign/symptom that is hard to tolerate; causes interference with activities of daily living and/or sleeping)
	Itchy nose	0–3
	Sneezing	0–3
	Runny nose	0–3
	Blocked nose	0–3
Conjunctival symptoms	Itchy/red eyes	0–3
	Watery eyes	0–3
(Total) daily symptom score (dSS)*		0–3 (max score is 3, i.e. 18 points/divided by six symptoms)
(B) Medication score		
	Oral and/or topical (eyes or nose) non-sedative H1 antihistamines (H1A)	1
	Intranasal corticosteroids (INS) with/without H1A	2
	Oral corticosteroids with/without INS, with/without H1A	3
(Total) daily medication score (dMS)		0–3 (max score is 3)
(C) Combined symptom and medication score		
CSMS	dSS (0–3) + dMS (0–3)	0–6

*Max score 18/6 (i.e. four nasal symptoms, max score 12; and two conjunctival symptoms, max score 6) is optimal for studies of seasonal pollinosis. This could possibly be modified for studies of perennial allergies (e.g. in mite-allergic patients), for example max score 12/4 (i.e. four nasal symptoms with omission of eye symptoms). By assigning 0–3 for all individual symptoms and dividing by total number of symptoms, the symptom range 0–3 and maximum symptom score 3 would remain the same.

to demonstrate the proof of concept, onset of action and magnitude of clinical effects (35–37). A clear unmet need in the future is a thorough technical standardization and (clinical) validation within and between different AEC models (31, 34).

Mechanisms of AIT

Long-term clinical improvement associates with humoral and cellular modifications to the allergen-specific immune response (38). Allergen immunotherapy may reduce the risk of progression from allergic rhinitis to asthma (17). There are some data to suggest that AIT may reduce the risk of developing new sensitizations to allergens, although further studies are needed to clarify this (39, 40). A greater understanding of the underlying mechanisms (Fig. 2) following the allergen-specific interventions of SCIT and sublingual immunotherapy (SLIT) is important for better understanding of the disease, for the development of predictive biomarkers and to assist the rational design and testing of novel more effective and safer immunotherapy strategies.

Allergic rhinitis is characterized by IgE synthesis, mast cell activation and tissue eosinophilia, events under the regulation of Th2 cytokines that are produced preferentially by CD4⁺ helper Th2 cells but also by mast cells, basophils and innate lymphoid cells (ILC2s). Subcutaneous immunotherapy is associated with a decrease in effector cells in target organs, transient increases in allergen-specific IgE followed by blunting of seasonal increases in IgE and a marked increase in IgG, particularly IgG₄ (41) (Fig. 2). For example, grass pollen SCIT resulted in a decrease in numbers of mast cells (42) and eosinophils (43) in the skin at sites of suppressed allergen-induced late cutaneous responses. Subcutaneous immunotherapy suppressed late nasal responses that paralleled decreases in local c-kit⁺ mast cells (44), eosinophils (45) and basophils (46) in the nasal mucosa. Recent studies have shown suppression of basophil activation (47, 48) and a decrease in circulating ILC2s (49).

Akdis et al. (50) first introduced the concept of early involvement of regulatory T (T reg) cells. Bee venom SCIT resulted in increased IL-10 production from both blood T and B cells. T reg involvement was confirmed by flow

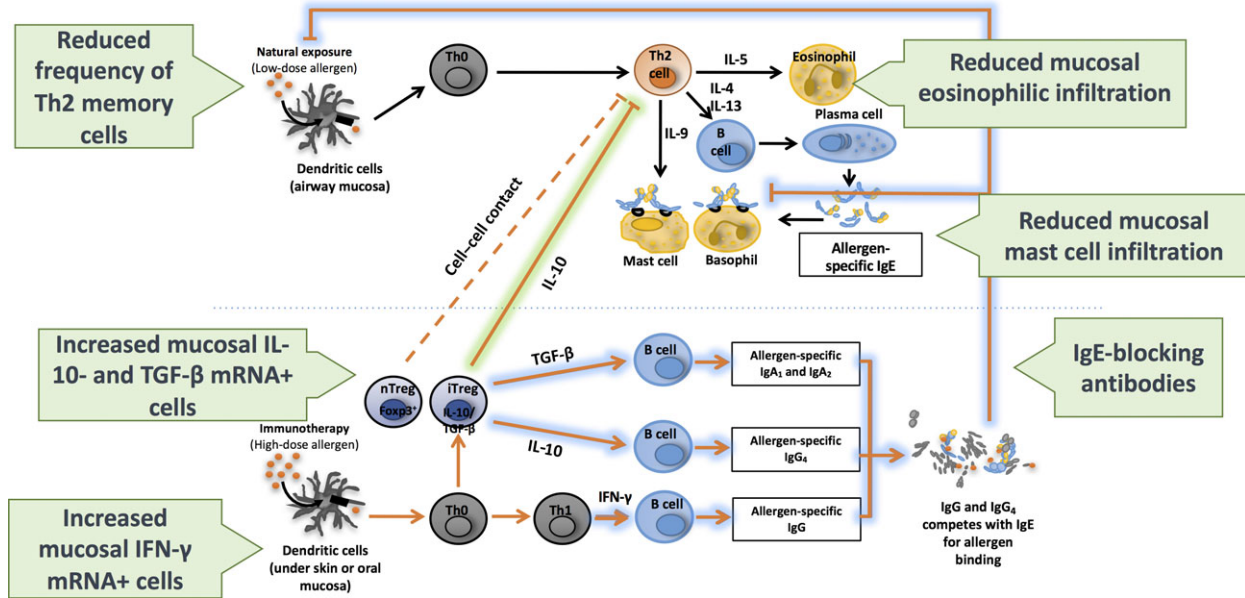


Figure 2 Mechanisms of allergen immunotherapy (AIT). The sites in the allergic response where mechanisms of AIT may intervene are indicated by green text arrows. Adapted from Ref. 118 with permission.

cytometry following grass and house dust mite SCIT (51, 52). Local induction of T regs following grass SCIT was shown by increases in IL-10⁺ (53) and TGF-β⁺ T cells (54) in the nasal mucosa that accompanied increases in serum IgG₄ and IgA (in keeping with their known properties in promoting preferential switching of B cells in favour of IgG₄ and IgA) and an increase in local nasal FOXP3⁺CD25⁺, IL-10-producing T cells (55) by immunofluorescence histology.

The mechanism of SLIT has been shown to be broadly similar to SCIT; in particular, SLIT also induces IL-10 production by T cells (56). Nasal and conjunctival eosinophils and adhesion molecules decrease following mite SLIT (57). Serum allergen-IgG increased after birch SLIT. Higher local numbers of CD4⁺FOXP3⁺ and CD25⁺FOXP3⁺ T cells were found in the sublingual mucosa and increases in peripheral allergen-specific IgG (IgG₁ and IgG₄) and IgA₂ following grass SLIT (58, 59). Blood Th1 cells increased and blood Th2 cells decreased following birch SLIT (60). Peripheral CD25⁺FOXP3⁺, presumed T reg cells, increased after SLIT (61), findings confirmed in relation to grass SLIT (62). Two recent studies showed that birch (63) and mite (64) SLIT resulted in *early* increases in T regs at 4–6 weeks and *delayed* increases in allergen-specific Th1 cells at 12 months. The mechanism of SCIT and SLIT may differ in terms of kinetics, quality and quantity of circulating antibody. Transient early increases in specific IgE are greater after SLIT compared to SCIT, whereas both inhibit seasonal increases in IgE to the same degree. Conversely, immunoreactive IgG levels are approximately 10-fold less for SLIT compared to SCIT (59, 65), whereas both are accompanied by *equivalent* increases in serum IgG-associated IgE-blocking activity. This disparity suggests that IgG antibodies produced by SLIT

may be more functionally active, with greater avidity or affinity for allergen. Although speculative, this could possibly occur as a consequence of more efficient antigen processing and/or T–B cell cooperation in the local environment of the sublingual mucosa and draining cervical lymph glands (66).

The LEAP and LEAP ON studies have brought major advance to our understanding of primary prevention of food allergy, as the number of high risk individuals regularly exposed to allergen acquired fivefold less food allergy than control individuals who avoided allergen, as suggested by international guidelines (21).

The demands to OIT are high as consumption of allergen may result in severe symptoms (67). Adverse side-effects during conventional OIT can be severe, and only half of the enrolled patients reach maintenance dose (23, 68). Half again could be maintained on maintenance dose for planned duration of treatment, and between a quarter and half of these patients had sustained unresponsiveness or developed persistent tolerance (24, 67). To improve this performance, treatment is attempted with hypoallergenic allergens (25) or under a blanket of anti-IgE treatment to limit side-effects (26). Persistent biomarker changes associated with successful primary prevention and OIT are a reduction of skin prick test (SPT) response and improvement in basophil reactivity and sensitivity (67, 69).

Biomarkers for diagnosis and monitoring in AIT trials

Patients with an indication for AIT respond with individual side-effect profiles, degree of symptom relief and persistence of treatment effect (Table 2) (70–74). This may be due to the diversity of their pattern of IgE sensitization (i.e. involvement of seasonal vs perennial allergens, mono- vs polysensitization),

Table 2 Potential biomarkers in support of allergen immunotherapy (AIT)

Categories of biomarkers	Applications	Candidate biomarkers
Biomarkers for diagnosis	Stratify patients to predict disease severity and/or progression	Patterns of IgE sensitization (119) <i>Ex vivo</i> basophil responsiveness to the therapeutic allergen (73)
Biomarkers predictive of AIT safety	Reduce risk and/or severity of side-effects Improve patient compliance	<i>Ex vivo</i> basophil responsiveness to the therapeutic allergen (73)
Biomarkers of AIT efficacy (predictive or follow-up)	Improve efficacy by selecting patients more likely to benefit from AIT Improve patient compliance Reduce cohort size needed for clinical development Confirm or not treatment efficacy after few weeks (early onset of efficacy) Document whether immune protection has been reached (support decision to pursue or stop AIT) Confirm lasting protection after stopping AIT (support decision to resume or not AIT) Adjust treatment modalities (e.g. dosing, immunization scheme)	Fetuin A isoforms (74) Reduction of <i>ex vivo</i> basophil sensitivity to the therapeutic allergen (48, 73) Changes in T-cell or dendritic cell polarization reflecting a reorientation of Th2 responses towards regulatory/suppressive responses (78, 79) Induction of allergen-specific IgG4s and blocking antibodies (76)

varying genetic susceptibility and comorbidities. Identification of biomarkers that predict benefit of AIT for the individual is therefore of high priority. According to the EMA guideline, biomarkers can be analysed in dose-finding studies but need to be validated (29).

Knowledge of mechanisms underlying AIT has been translated into developing biomarkers beyond specific IgE to predict efficacy and side-effects, and to monitor clinical response to and persistence of immunotherapy. Basophil sensitivity (48) and IgG-associated serum inhibitory activity for IgE-facilitated binding of allergen-IgE complexes to B cells (IgE-FAB) correlated closely with clinical response to immunotherapy (75). The latter assay has been standardized (76) and translated into a cell-free solid phase ELISA-based assay (77). Allergen-stimulated basophil activation has been validated (73).

Unbiased identification of new biomarkers of AIT efficacy

Unbiased approaches have been used to find biomarkers for change in dendritic cells (DCs) during AIT. A 4-month SLIT course in grass pollen-allergic patients induced regulatory dendritic cells (DC regs) expressing high levels of C1Q and stabilin (78), paralleled by a decrease in the blood of pro-allergic DC2 markers such as CD141 and OX40L, which support differentiation of Th2 cells (79). This switch of the DC reg/DC2 balance was detected in peripheral blood by quantitative PCR only in patients exhibiting clinical responses.

A sialylated variant of fetuin A (Fet A), which is expressed at high levels in pretreatment sera from grass pollen-allergic patients who benefited from SLIT, but not in samples from nonresponders, was also identified by an unbiased approach (74). Fet A appears to modulate inflammation due to its capacity to interact with a broad variety of ligands including

calcium, TGF- β , the insulin receptor, fatty acids and TLR4 and is involved in multiple inflammatory conditions and cancer (80). The novel link between Fet A and allergy was confirmed in preclinical models where post-translational modifications of Fet A could modulate allergic inflammation (74).

Basophil activation testing as a new biomarker in diagnosis for and monitoring of AIT

Basophil sensitivity and reactivity to allergen in a basophil activation test can measure effect of AIT in response to allergen exposure at the effector cell level in an accessible *ex vivo* assay (73).

In a study in ultrarush venom immunotherapy, AIT induced desensitization of basophils in response to both the treatment allergen and anti-IgE stimulation after 5 days of up dosing (81). This unspecific desensitization was reproduced *in vitro* (82). After 1 year of venom AIT, basophil response to submaximal doses of allergen decreased fourfold in adults (83) and children (84). In subjects suffering from seasonal pollen-allergic rhinitis, SCIT induces a strong decrease in basophil sensitivity during the up dosing phase (85). These early changes in basophil sensitivity were shown to predict clinical outcome in the following pollen season (48) and are maintained throughout maintenance therapy and even after treatment cessation (86). A protocol of preseasonal injections also reduced the basophil response (87).

One study comparing effects of SCIT and SLIT in grass pollen-allergic patients found a slower and smaller, yet still significant decrease in basophil sensitivity in the SLIT-treated group (88), and another using diamine oxidase as a read-out for basophil degranulation found comparable changes in both SCIT- and SLIT-treated patients (47); other studies did not find a clear change in the basophil allergen response (89).

Oral immunotherapy induced a reduction in the basophil reactivity to allergen in children treated with peanut (90) and egg (24).

In SCIT, the decrease in basophil sensitivity seems to reflect the induction of allergen tolerance and may be useful to predict clinical outcome. In SLIT, intralymphatic immunotherapy (ILIT) and OIT, the role of basophil response to allergen is less clear. Further larger studies are needed to standardize basophil testing, to confirm the predictive value of basophil sensitivity during AIT and to establish basophil reactivity or sensitivity as a biomarker.

Novel approaches for AIT

A better understanding of mechanisms has been translated into novel immunotherapy approaches. Targeting IgE by anti-IgE antibody in combination with AIT reduced the incidence of side-effects during immunotherapy and resulted in prolonged suppression of IgE-FAB for longer than the half-life of anti-IgE, suggesting this strategy may have a durable effect (91). The combination of anti-IL-4 with AIT was effective in suppressing circulating Th2 cells and allergen-induced late responses although it had no obvious advantages over allergen extract alone (92). Targeting immune deviation using the TLR4 agonist monophosphoryl lipid A in combination with AIT was effective with four preseasonal injections without an increase in side-effects (93). The use of Bacterial DNA oligonucleotides rich in CpG sequences, covalently linked to the major ragweed allergen Amb a 1, was effective, possibly by inducing T regs and/or immune deviation although this has not been pursued (94). Further strategies such as targeting epithelial cytokines thymic stromal lymphopoietin and IL-33 in combination with AIT or possibly the combined use of probiotics with AIT are yet to be tested. Strategies to reduce allergic adverse events in AIT include the use of engineered recombinant hypoallergenic molecules, allergen multimers and allergen fragments including peptides. These approaches rely on alterations to the structure of the allergen protein reducing the ability to cross-link IgE whilst maintaining the ability to target allergen-specific T cells and induce immune modulation. The following emerging approaches aim at optimizing delivery and immunization schemes towards shorter course therapy to improve compliance by applying targeted delivery to immune compartments or peptide-based technologies.

Peptide immunotherapy

One such approach is the use of short soluble synthetic peptides containing the immunodominant T-cell epitopes of major allergen proteins. Peptides are selected so as to lack the length and three-dimensional structure to cross-link IgE molecules on the surface of mast cells and basophils, whilst retaining the ability to be recognized by and modulate allergen-specific T cells. The mechanisms of action of peptide immunotherapy may involve the induction of antigen-specific hyporesponsiveness (anergy), deviation of the T helper cytokine production profile from Th2 to Th1, clonal deletion

(e.g. through exhaustion) and the induction T cells with regulatory function, perhaps mediated by IL-10 and TGF- β (38).

Clinical efficacy of Fel d 1 synthetic peptides (Cat-SPIRE) was evaluated in a phase 2b clinical trial in an AEC (95). Treatment was safe, well tolerated and reduced symptoms of rhinoconjunctivitis. Administration of eight intradermal injections of 3 nmol synthetic Fel d 1 peptides with 2 week intervals resulted in a greater improvement in symptoms scores than placebo (95).

A further phase 2b trial evaluated efficacy 18–22 and 50–54 weeks after treatment with Fel d 1 synthetic peptides. Reduced symptom scores were observed at both time points with the 50- to 54-week follow-up achieving statistical significance, despite the lack of treatment intervention for the preceding 9 months (36). After approximately 2 years, the changes in TRSS remained at the same level as earlier, providing evidence of enduring efficacy (96). Most recently, the results of a phase 3 trial (EudraCT Number: 2012-001733-13) involving over 1400 subjects showed that treatment with peptides was safe and well tolerated. However, the trial failed to demonstrate clinical efficacy in the field. Although active treatment resulted in an approximately 60% reduction in mean symptom and medication scores, a similar effect was observed in the placebo-treated group.

Safety and efficacy of grass pollen synthetic peptides were evaluated in an AEC. Subjects were exposed to grass pollen for 3 h per day on four consecutive days. Subjects treated with eight administrations of 6 nmol grass-peptide immunotherapy (dosed at 2-weekly intervals) reported a mean 42% change in TRSS compared with the placebo group. Forty-four per cent of subjects in this active treatment group considered themselves 'very much better' after treatment, compared with 22% of subjects in the placebo group ($P < 0.01$) (97).

Contiguous overlapping peptides

AllerT contains three overlapping 49–71 amino acid peptides with no ability to form the original Bet v 1 three-dimensional structure causing birch pollen-induced rhinitis/rhinoconjunctivitis (98). This removes the need of going through the slow progressive dose-escalation process required with conventional AIT and allows the administration of doses up to 10-fold more allergen equivalent than with conventional AIT. These long peptides have markedly reduced IgE binding and do not induce anaphylaxis in sensitized mice (99). A phase I study demonstrated increases in IL-10, IL-5 and IL-13 within weeks followed by 40-fold increases in IgG₄ that remained elevated for more than 3 years (98).

In a phase II b study, treatment with long peptides resulted in significant reductions in the combined rhinoconjunctivitis symptom and medication score compared to placebo, as well as significant reductions in secondary endpoints (100) that persisted in the subgroup that agreed to a second follow-up year (101). Based on data from 1569 injections in 335 participants, long contiguous overlapping peptides seem to be well tolerated. Most reported adverse events were mild to moderate with no anaphylactic reactions and no immediate

systemic reactions within 30 min of injections. Most participants completed the course of five injections. Four subjects experienced WAO grade 3 systemic reactions after 30 min (equivalent to 2.5/1000 injections) and 5% of subjects a >30% drop in FEV1. These reactions occurred between 4 and 6 h after injection and were most pronounced early in the desensitization process. They were likely to be T cell-mediated rather than IgE-dependent anaphylaxis. A new larger European field-based clinical trial is to be implemented during 2016 (EudraCT 2016-000076-23).

Intralymphatic immunotherapy

In the first open randomized study, ILIT was compared with SCIT in 165 patients with grass allergy (102). There was a quick onset of increased tolerance to skin prick tests and nasal provocations. There was a significant reduction in rhinitis symptoms that was long-lasting and comparable to the observed clinical effect of SCIT. Another randomized double-blind trial compared ILIT to placebo in 15 participants grass and birch allergy (103). A significant and clinically relevant reduction in self-reported symptoms was seen in the ILIT group. There was reduction in allergen-specific IgE and a decreased inflammatory response in the nose during provocation tests.

Recombinant cat dander allergen (MAT-Fel d 1) has been investigated with ILIT in a randomized double-blind placebo-controlled study with 20 participants. Primary outcome was response to titrated nasal provocation test. The study demonstrated a 74-fold increase in nasal tolerance and also an increase in specific IgG₄ (104).

Conflicting results were published in 2013 in a randomized double-blind placebo-controlled study in 45 patients with grass allergy (105). There was no difference reduction in the combined symptom medication score (SMS), respiratory quality of life questionnaire (RQLQ) or skin prick test results, whereas an increase in allergen-specific IgG₄ was observed after ILIT. One possible explanation for the lack of clinical efficacy may have been that the dosing interval was 2 weeks, whereas in all other studies the interval was 4 weeks.

In a pilot study with seven patients, the frequency of allergen-specific non-IgE⁺ plasmablasts increased significantly 1 week after allergen injection, and tolerance to nasal provocation as well as titrated skin prick test response increased after the pollen season (106). In a follow-up randomized controlled trial, 36 participants with grass pollen-induced rhinitis were randomized 2 : 1 in favour of the active treatment (three injections of 1000 SQU Alutard). Patients on active treatment had significantly fewer symptoms in the first season after treatment (107). In a more recent double-blind placebo-controlled study with 36 patients, ILIT against birch and grass allergy significantly improved self-reported treatment outcomes (108), whereas there were no differences between the ILIT and placebo group in response to nasal allergen provocation and no differences in specific IgE or IgG₄. A subgroup of treated patients with good clinical response had increased the affinity of IgG₄ for grass allergen to offer better protection. In the first double-blind placebo-controlled trial

of adolescents and the first on the American continent, treatment was safe but did not achieve statistical significance (109). The reason may be low compliance when reporting total symptom score; only 73% of participants entered data on ≥50% of days in the peak pollen season. This underscores the need for a pervasive monitoring tool for AIT.

Given the limited number of studies and conflicting results, there is a need for adequately powered trials of ILIT and studies with prolonged follow-up to assess potential for long-term tolerance.

Recombinant allergens

Molecular allergy has characterized most major allergen components of common inhalants with important implications for diagnosis and therapy (110). For example, Phl p 1 (Phl p 1) and Phl p 5 are the dominant major grass pollen allergens and detection of IgE to Phl p 1 and Phl p 5 confirms clinically relevant sensitivity, whereas specific IgE to Phl p 12 is likely due to cross-reactivity with the birch-derived profilin Bet v 2. Such a patient may exhibit an irrelevant, false-positive skin test to birch pollen extract, due to the presence of IgE to the irrelevant cross-reacting profilin Phl p 12. Such considerations are helpful in selecting suitable patients and the relevant allergen for AIT (111). Recombinant allergens either singly as in the case of Bet v 1, the major birch pollen allergen (112), or as the relevant recombinant grass allergen mixture (Phl p 1, Phl p 2, Phl p 5a, Phl p 5b, Phl p 6) (113) have been shown to be highly effective in randomized controlled trials. Recombinant allergens may also be genetically modified to reduce IgE binding and allergenicity (114) with great potential for safer immunotherapy. The recombinant vaccine protein BM32, encompassing non-IgE binding domains from grass allergens Phl p 1, 2, 5 and 6 fused with a hepatitis B viral surface protein, is currently in phase 2 studies (115, 116). Patients treated with this vaccine raised IgG antibody to allergens and hepatitis B viral sequences encoded by the recombinant proteins. The immune response to both allergen (115) and virus (116) was protective *in vitro*, and patients had reduced allergy symptoms in an AEC challenge (115). Recombinant approaches may ultimately allow tailor-made immunotherapy for individuals (117), which will not induce novel sensitizations to irrelevant allergens found in extracts.

Conclusion

Allergic diseases are underdiagnosed and undertreated. Allergen immunotherapy has not yet received adequate attention from European (public) institutions, and many allergic patients remain unaware of the potential benefit of AIT. Areas for improvement and innovation are found through a better understanding of the mode of action and dose effects, which will lead to improved delivery and immunization schedules. More effective preparations with faster onset and reduced doses are likely to improve compliance. A previous trial of AIT for asthma prevention had methodological limitations, so the recent results from the randomized, double-blind placebo-

controlled GAP trial are encouraging. With improvement in grading of evidence, standardizing clinical outcomes, we can expect better studies in general in this area. Identification of biomarkers for improved patient selection and monitoring will increase the value and usefulness of AIT.

In summary, it is time to re-evaluate the allergy paradigm and implement new treatment approaches with a special focus on AIT as allergic individuals are becoming a significant minority of Western and global populations. Innovation at all levels and national and regional action plans, such as the Finnish Allergy Programme, are needed to meet the challenge. This first interdisciplinary Aarhus Immunotherapy Symposium brought together specialists from different disciplines to document exciting developments in the field of AIT and provided hope for improvement of this therapeutic option for allergic patients in the future.

Author contributions

EV, OP, PM, JMS, SHS, LOC, KS, ML and SRD contributed specific sections of the manuscript. PS and HJH wrote introduction and conclusion. All authors reviewed the final document.

Conflicts of interest

Drs. Hoffmann, Schmid and Cardell have nothing to disclose. Dr. Valovirta reports grants and personal fees from

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References

1. Newson RB, van Ree R, Forsberg B, Janson C, Lötvall J, Dahlén S-E et al. Geographical variation in the prevalence of sensitization to common aeroallergens in adults: the GA² LEN survey. *Allergy* 2014;**69**:643–651.
2. Bousquet J, Lockey R, Malling HJ. Allergen immunotherapy: therapeutic vaccines for allergic diseases. A WHO position paper. *J Allergy Clin Immunol* 1998;**102**:558–562.
3. Calderon MA, Demoly P, Gerth van Wijk R, Bousquet J, Sheikh A, Frew A et al. EAACI: a European declaration on immunotherapy. Designing the future of allergen specific immunotherapy. *Clin Transl Allergy* 2012;**2**:20.
4. Pfaar O, Bachert C, Bufe A, Buhl R, Ebner C, Eng P et al. Guideline on allergen-specific immunotherapy in IgE-mediated allergic diseases: S2k Guideline of the German Society for Allergology and Clinical Immunology (DGAKI), the Society for Pediatric Allergy and Environmental Medicine (GPA), the Medical Association of German Allergologists (AeDA), the Austrian Society for Allergy and Immunology (ÖGAI), the Swiss Society for Allergy and Immunology (SGAI), the German Society of Dermatology (DDG), the German Society of Oto-Rhino-Laryngology, Head and Neck Surgery (DGHNO-KHC), the German Society of Pediatrics and Adolescent Medicine (DGKJ), the Society for Pediatric Pneumology (GPP), the German Respiratory Society (DGP), the German Association of ENT Surgeons (BV-HNO), the Professional Federation of Paediatricians and Youth Doctors (BVKJ), the Federal Association of Pulmonologists (BDP) and the German Dermatologists Association (BVDD). *Allergo J Int* 2014;**23**:282–319.
5. Jutel M, Agache I, Bonini S, Burks AW, Calderon M, Canonica W et al. International consensus on allergy immunotherapy. *J Allergy Clin Immunol* 2015;**136**:556–568.
6. Bauchau V. Prevalence and rate of diagnosis of allergic rhinitis in Europe. *Eur Respir J* 2004;**24**:758–764.
7. Arbes SJ Jr, Gergen PJ, Elliott L, Zeldin DC. Prevalences of positive skin test responses to 10 common allergens in the US population: results from the Third National Health and Nutrition Examination Survey. *J Allergy Clin Immunol* 2005;**116**:377–383.
8. Bousquet P-J, Chinn S, Janson C, Kogevinas M, Burney P, Jarvis D. Geographical variation in the prevalence of positive skin tests to environmental aeroallergens in the European Community Respiratory Health Survey I: prevalence of positive skin tests to environmental aeroallergens. *Allergy* 2007;**62**:301–309.
9. Hanski I, von Hertzen L, Fyhrquist N, Koskinen K, Torppa K, Laatikainen T et al. Environmental biodiversity, human microbiota, and allergy are interrelated. *Proc Natl Acad Sci USA* 2012;**109**:8334–8339.
10. Selroos O, Kupczyk M, Kuna P, Łacwik P, Bousquet J, Brennan D et al. National and regional asthma programmes in Europe. *Eur Respir Rev* 2015;**24**:474–483.
11. Haahtela T, von Hertzen L, Mäkelä M, Hannuksela M, Allergy Programme Working Group. Finnish Allergy Programme 2008–2018—time to act and change the course. *Allergy* 2008;**63**:634–645.
12. Pawankar R, Baena-Cagnani CE, Bousquet J, Canonica W, Cruz A, Kaliner MA et al. State of world allergy report 2008: allergy and chronic respiratory diseases. *World Allergy Organ J* 2008;**2008**:S4–S17.
13. Beasley R, Ellwood P, Asher I. International patterns of the prevalence of pediatric asthma the ISAAC program. *Pediatr Clin North Am* 2003;**50**:539–553.

14. Pawankar R, Bunnag C, Chen Y, Fukuda T, Kim Y-Y, Le LTT et al. Allergic rhinitis and its impact on asthma update (ARIA 2008)–western and Asian-Pacific perspective. *Asian Pac J Allergy Immunol* 2009;**27**: 237–243.
15. Andiappan AK, Puan KJ, Lee B, Nardin A, Poidinger M, Connolly J et al. Allergic airway diseases in a tropical urban environment are driven by dominant mono-specific sensitization against house dust mites. *Allergy* 2014;**69**:501–509.
16. von Hertzen LC, Savolainen J, Hannuksela M, Klaukka T, Lauerma A, Mäkelä MJ et al. Scientific rationale for the Finnish Allergy Programme 2008–2018: emphasis on prevention and endorsing tolerance. *Allergy* 2009;**64**:678–701.
17. Jacobsen L, Niggemann B, Dreborg S, Ferdousi HA, Halken S, Host A et al. Specific immunotherapy has long-term preventive effect of seasonal and perennial asthma: 10-year follow-up on the PAT study. *Allergy* 2007;**62**:943–948.
18. Schmitt J, Schwarz K, Stadler E, Wüstenberg EG. Allergy immunotherapy for allergic rhinitis effectively prevents asthma: results from a large retrospective cohort study. *J Allergy Clin Immunol* 2015;**136**:1511–1516.
19. Valovirta E, Berstad AKH, de Blic J, Bufer A, Eng P, Halken S et al. Design and recruitment for the GAP trial, investigating the preventive effect on asthma development of an SQ-standardized grass allergy immunotherapy tablet in children with grass pollen-induced allergic rhinoconjunctivitis. *Clin Ther* 2011;**33**:1537–1546.
20. Valovirta E, Cronjäger R, Petersen TH, Piotrowska T, Andersen JS, Sørensen HF et al. Top-line results from the five-year landmark GRAZAX(R) Asthma Prevention (GAP) trial in children. Latebreaking Abstracts #2358: Asthma in Children. Vienna, 2016. eaaci2016.org
21. Du Toit G, Roberts G, Sayre PH, Bahnson HT, Radulovic S, Santos AF et al. Randomized trial of peanut consumption in infants at risk for peanut allergy. *N Engl J Med* 2015;**372**:803–813.
22. Perkin MR, Logan K, Tseng A, Raji B, Ayis S, Peacock J et al. Randomized trial of introduction of allergenic foods in breast-fed infants. *N Engl J Med* 2016;**374**:1733–1743.
23. Clark AT, Islam S, King Y, Deighton J, Anagnostou K, Ewan PW. Successful oral tolerance induction in severe peanut allergy. *Allergy* 2009;**64**:1218–1220.
24. Burks AW, Jones SM, Wood RA, Fleischer DM, Sicherer SH, Lindblad RW et al. Oral immunotherapy for treatment of egg allergy in children. *N Engl J Med* 2012;**367**:233–243.
25. Ballmer-Weber BK, Brockow K, Fiocchi A, Theler B, Vogel L, Ring J et al. Hydrolysed egg displays strong decrease in allergenicity and is well tolerated by egg-allergic patients. *Allergy* 2016;**71**:728–732.
26. Wood RA, Kim JS, Lindblad R, Nadeau K, Henning AK, Dawson P et al. A randomized, double-blind, placebo-controlled study of omalizumab combined with oral immunotherapy for the treatment of cow's milk allergy. *J Allergy Clin Immunol* 2016;**137**:1103–1110.
27. Canonica GW, Baena-Cagnani CE, Bousquet J, Bousquet PJ, Lockey RF, Malling H-J et al. Recommendations for standardization of clinical trials with Allergen Specific Immunotherapy for respiratory allergy. A statement of a World Allergy Organization (WAO) taskforce. *Allergy* 2007;**62**:317–324.
28. Bousquet J, Schünemann HJ, Bousquet PJ, Bachert C, Canonica GW, Casale TB et al. How to design and evaluate randomized controlled trials in immunotherapy for allergic rhinitis: an ARIA-GA(2) LEN statement. *Allergy* 2011;**66**:765–774.
29. European Medicines Agency. Committee for medicinal products for human use (CHMP). Guideline on the clinical development of products for specific immunotherapy for the treatment of allergic diseases (CHMP/EWP/18504/2006). 2009. http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2009/09/WC500003605.pdf; 2008 (accessed 28 June 2016).
30. U.S. Department of Health and Human Services Food and Drug Administration Center for Drug Evaluation and Research (CDER). Allergic rhinitis: developing drug products for treatment guidance for industry. 2016. <http://www.fda.gov/downloads/drugs/guidancecomplianceregulatoryinformation/guidances/ucm071293.pdf>; Revision 1; Feb 2016 (accessed 28 June 2016).
31. Pfaar O, Demoly P, Gerth van Wijk R, Bonini S, Bousquet J, Canonica GW et al. Recommendations for the standardization of clinical outcomes used in allergen immunotherapy trials for allergic rhinoconjunctivitis: an EAACI position paper. *Allergy* 2014;**69**:854–867.
32. Pfaar O, Kleine-Tebbe J, Hörmann K, Klimek L. Allergen-specific immunotherapy: which outcome measures are useful in monitoring clinical trials? *Immunol Allergy Clin North Am* 2011;**31**:289–309.
33. Makatsori M, Pfaar O, Calderon MA. Allergen immunotherapy: clinical outcomes assessment. *J Allergy Clin Immunol Pract* 2014;**2**:123–129.
34. Rösner-Friese K, Kaul S, Vieths S, Pfaar O. Environmental exposure chambers in allergen immunotherapy trials: current status and clinical validation needs. *J Allergy Clin Immunol* 2015;**135**:636–643.
35. Horak F, Ziegler P, Ziegler R, Lemell P, Devillier P, Montagut A et al. Early onset of action of a 5-grass-pollen 300-IR sublingual immunotherapy tablet evaluated in an allergen challenge chamber. *J Allergy Clin Immunol* 2009;**124**:471–477.
36. Patel D, Couroux P, Hickey P, Salapatek AM, Laidler P, Larché M et al. Fel d 1-derived peptide antigen desensitization shows a persistent treatment effect 1 year after the start of dosing: a randomized, placebo-controlled study. *J Allergy Clin Immunol* 2013;**131**:103–109.
37. Nolte H, Maloney J, Nelson HS, Bernstein DI, Lu S, Li Z et al. Onset and dose-related efficacy of house dust mite sublingual immunotherapy tablets in an environmental exposure chamber. *J Allergy Clin Immunol* 2015;**135**:1494–1501.
38. Larché M. Peptide immunotherapy for allergic diseases. *Allergy* 2007;**62**:325–331.
39. Des Roches A, Paradis L, Menardo JL, Bouges S, Daurés JP, Bousquet J. Immunotherapy with a standardized *Dermatophagoides pteronyssinus* extract. VI. Specific immunotherapy prevents the onset of new sensitizations in children. *J Allergy Clin Immunol* 1997;**99**:450–453.
40. Pajno GB, Barberio G, De Luca F, Morabito L, Parmiani S. Prevention of new sensitizations in asthmatic children monosensitized to house dust mite by specific immunotherapy. A six-year follow-up study. *Clin Exp Allergy* 2001;**31**:1392–1397.
41. Wachholz PA, Kristensen Soni N, Till SJ, Durham SR. Inhibition of allergen-IgE binding to B cells by IgG antibodies after grass pollen immunotherapy. *J Allergy Clin Immunol* 2003;**112**:915–922.
42. Durham SR, Varney VA, Gaga M, Jacobson MR, Varga EM, Frew AJ, Kay AB. Grass pollen immunotherapy decreases the number of mast cells in the skin. *Clin Exp Allergy* 1999;**29**:1490–1496.
43. Varney VA, Hamid QA, Gaga M, Ying S, Jacobson M, Frew AJ et al. Influence of grass pollen immunotherapy on cellular infiltration and cytokine mRNA expression during allergen-induced late-phase cutaneous responses. *J Clin Invest* 1993;**92**:644–651.
44. Nouri-Aria KT, Pilette C, Jacobson MR, Watanabe H, Durham SR. IL-9 and c-Kit+ mast cells in allergic rhinitis during seasonal allergen exposure: effect of immunotherapy. *J Allergy Clin Immunol* 2005;**116**:73–79.
45. Wilson DR, Nouri-Aria KT, Walker SM, Pajno GB, O'Brien F, Jacobson MR et al.

- Grass pollen immunotherapy: symptomatic improvement correlates with reductions in eosinophils and IL-5 mRNA expression in the nasal mucosa during the pollen season. *J Allergy Clin Immunol* 2001;**107**:971–976.
46. Wilson DR, Irani A-M, Walker SM, Jacobson MR, Mackay IS, Schwartz LB et al. Grass pollen immunotherapy inhibits seasonal increases in basophils and eosinophils in the nasal epithelium. *Clin Exp Allergy* 2001;**31**:1705–1713.
 47. Shamji MH, Layhadi JA, Scadding GW, Cheung DKM, Calderon MA, Turka LA et al. Basophil expression of diamine oxidase: a novel biomarker of allergen immunotherapy response. *J Allergy Clin Immunol* 2015;**135**:913–921.
 48. Schmid JM, Würtzen PA, Dahl R, Hoffmann HJ. Early improvement in basophil sensitivity predicts symptom relief with grass pollen immunotherapy. *J Allergy Clin Immunol* 2014;**134**:741–744.
 49. Lao-Araya M, Steveling E, Scadding GW, Durham SR, Shamji MH. Seasonal increases in peripheral innate lymphoid type 2 cells are inhibited by subcutaneous grass pollen immunotherapy. *J Allergy Clin Immunol* 2014;**134**:1193–1195.
 50. Akdis CA, Akdis M, Blesken T, Wymann D, Alkan SS, Müller U et al. Epitope-specific T cell tolerance to phospholipase A2 in bee venom immunotherapy and recovery by IL-2 and IL-15 in vitro. *J Clin Invest* 1996;**98**:1676–1683.
 51. Francis JN, Till SJ, Durham SR. Induction of IL-10 + CD4 + CD25 + T cells by grass pollen immunotherapy. *J Allergy Clin Immunol* 2003;**111**:1255–1261.
 52. Jutel M, Akdis M, Budak F, Aebischer-Casaulta C, Wrzyszc M, Blaser K et al. IL-10 and TGF- β cooperate in the regulatory T cell response to mucosal allergens in normal immunity and specific immunotherapy. *Eur J Immunol* 2003;**33**:1205–1214.
 53. Nouri-Aria KT, Wachholz PA, Francis JN, Jacobson MR, Walker SM, Wilcock LK et al. Grass pollen immunotherapy induces mucosal and peripheral IL-10 responses and blocking IgG activity. *J Immunol* 2004;**172**:3252–3259.
 54. Pilette C, Nouri-Aria KT, Jacobson MR, Wilcock LK, Detry B, Walker SM et al. Grass pollen immunotherapy induces an allergen-specific IgA2 antibody response associated with mucosal TGF- β expression. *J Immunol* 2007;**178**:4658–4666.
 55. Radulovic S, Jacobson MR, Durham SR, Nouri-Aria KT. Grass pollen immunotherapy induces Foxp3-expressing CD4 + CD25 + cells in the nasal mucosa. *J Allergy Clin Immunol* 2008;**121**:1467–1472.
 56. Ciprandi G, Fenoglio D, Cirillo I, Vizzacaro A, Ferrera A, Tosca MA et al. Induction of interleukin 10 by sublingual immunotherapy for house dust mites: a preliminary report. *Ann Allergy Asthma Immunol* 2005;**95**:38–44.
 57. Passalacqua G, Albano M, Fregonese L, Riccio A, Pronzato C, Mela GS et al. Randomised controlled trial of local allergoid immunotherapy on allergic inflammation in mite-induced rhinoconjunctivitis. *Lancet* 1998;**351**:629–632.
 58. Möbs C, Ipsen H, Mayer L, Slotosch C, Petersen A, Würtzen PA et al. Birch pollen immunotherapy results in long-term loss of Bet v 1-specific TH2 responses, transient TR1 activation, and synthesis of IgE-blocking antibodies. *J Allergy Clin Immunol* 2012;**130**:1108–1116.
 59. Scadding GW, Shamji MH, Jacobson MR, Lee DI, Wilson D, Lima MT et al. Sublingual grass pollen immunotherapy is associated with increases in sublingual Foxp3-expressing cells and elevated allergen-specific immunoglobulin G4, immunoglobulin A and serum inhibitory activity for immunoglobulin E-facilitated allergen binding to B cells. *Clin Exp Allergy* 2010;**40**:598–606.
 60. Ebner C, Siemann U, Bohle B, Willheim M, Wiedermann U, Schenk S et al. Immunological changes during specific immunotherapy of grass pollen allergy: reduced lymphoproliferative responses to allergen and shift from TH2 to TH1 in T-cell clones specific for Phi p 1, a major grass pollen allergen. *Clin Exp Allergy* 1997;**27**:1007–1015.
 61. Nieminen K, Valovirta E, Savolainen J. Clinical outcome and IL-17, IL-23, IL-27 and FOXP3 expression in peripheral blood mononuclear cells of pollen-allergic children during sublingual immunotherapy. *Pediatr Allergy Immunol* 2010;**21**:e174–e184.
 62. Suárez-Fueyo A, Ramos T, Galán A, Jimeno L, Wurtzen PA, Marin A et al. Grass tablet sublingual immunotherapy downregulates the TH2 cytokine response followed by regulatory T-cell generation. *J Allergy Clin Immunol* 2014;**133**:130–138.
 63. Bohle B, Kinaciyan T, Gerstmayr M, Radakovics A, Jahn-Schmid B, Ebner C. Sublingual immunotherapy induces IL-10-producing T regulatory cells, allergen-specific T-cell tolerance, and immune deviation. *J Allergy Clin Immunol* 2007;**120**:707–713.
 64. O'Hehir RE, Gardner LM, de Leon MP, Hales BJ, Biondo M, Douglass JA et al. House dust mite sublingual immunotherapy: the role for transforming growth factor- β and functional regulatory T cells. *Am J Respir Crit Care Med* 2009;**180**:936–947.
 65. James LK, Shamji MH, Walker SM, Wilson DR, Wachholz PA, Francis JN et al. Long-term tolerance after allergen immunotherapy is accompanied by selective persistence of blocking antibodies. *J Allergy Clin Immunol* 2011;**127**:509–516.
 66. Allam J-P, Novak N. Immunological mechanisms of sublingual immunotherapy. *Curr Opin Allergy Clin Immunol* 2014;**14**:564–569.
 67. Ponce M, Diesner SC, Szépfalusi Z, Eiwegger T. Markers of tolerance development to food allergens. *Allergy* 2016;**71**:1393–1404.
 68. Staden U, Rolinck-Werninghaus C, Brewe F, Wahn U, Niggemann B, Beyer K. Specific oral tolerance induction in food allergy in children: efficacy and clinical patterns of reaction. *Allergy* 2007;**62**:1261–1269.
 69. Santos AF, Lack G. Basophil activation test: food challenge in a test tube or specialist research tool? *Clin Transl Allergy* 2016;**6**:10.
 70. Alam R. Biomarkers in asthma and allergy. *Immunol Allergy Clin North Am* 2012;**32**:xi–xii.
 71. Shamji MH, Ljørring C, Würtzen PA. Predictive biomarkers of clinical efficacy of allergen-specific immunotherapy: how to proceed. *Immunotherapy* 2013;**5**:203–206.
 72. Popescu F-D. Molecular biomarkers for grass pollen immunotherapy. *World J Methodol* 2014;**4**:26–45.
 73. Hoffmann HJ, Santos AF, Mayorga C, Nopp A, Eberlein B, Ferrer M et al. The clinical utility of basophil activation testing in diagnosis and monitoring of allergic disease. *Allergy* 2015;**70**:1393–1405.
 74. Moingeon P. Biomarkers for allergen immunotherapy: a 'panoromic' view. *Immunol Allergy Clin North Am* 2016;**36**:161–179.
 75. Shamji MH, Ljørring C, Francis JN, Calderon MA, Larché M, Kimber I et al. Functional rather than immunoreactive levels of IgG4 correlate closely with clinical response to grass pollen immunotherapy: functional IgG4 correlates with clinical efficacy of allergen specific immunotherapy. *Allergy* 2012;**67**:217–226.
 76. Shamji MH, Wilcock LK, Wachholz PA, Dearman RJ, Kimber I, Wurtzen PA et al. The IgE-facilitated allergen binding (FAB) assay: validation of a novel flow-cytometric based method for the detection of inhibitory antibody responses. *J Immunol Methods* 2006;**317**:71–79.
 77. Shamji MH, Francis JN, Würtzen PA, Lund K, Durham SR, Till SJ. Cell-free detection of allergen-IgE cross-linking with immobilized phase CD23: inhibition by blocking antibody responses after immunotherapy. *J Allergy Clin Immunol* 2013;**132**:1003–1005.
 78. Zimmer A, Bouley J, Le Mignon M, Pliquet E, Horiot S, Turfkruyer M et al. A regulatory dendritic cell signature correlates with the clinical efficacy of allergen-specific

- sublingual immunotherapy. *J Allergy Clin Immunol* 2012;**129**:1020–1030.
79. Gueguen C, Bouley J, Moussu H, Luce S, Duchateau M, Chamot-Rooke J et al. Changes in markers associated with dendritic cells driving the differentiation of either TH2 cells or regulatory T cells correlate with clinical benefit during allergen immunotherapy. *J Allergy Clin Immunol* 2016;**137**:545–558.
 80. Mori K, Emoto M, Inaba M. Fetuin-A: a multifunctional protein. *Recent Pat Endocr Metab Immune Drug Discov* 2011;**5**:124–146.
 81. Ciprandi G, Klersy C, Cirillo I, Marseglia GL. Quality of life in allergic rhinitis: relationship with clinical, immunological, and functional aspects. *Clin Exp Allergy* 2007;**37**:1528–1535.
 82. Witting Christensen SK, Kortekaas Krohn I, Thuraiayah J, Skjold T, Schmid JM, Hoffmann HJH. Sequential allergen desensitization of basophils is non-specific and may involve p38 MAPK. *Allergy* 2014;**69**:1343–1349.
 83. Eržen R, Košnik M, Šilar M, Korošec P. Basophil response and the induction of a tolerance in venom immunotherapy: a long-term sting challenge study. *Allergy* 2012;**67**:822–830.
 84. Žitnik SEK, Vesel T, Avčini T, Šilar M, Košnik M, Korošec P. Monitoring honeybee venom immunotherapy in children with the basophil activation test. *Pediatr Allergy Immunol* 2012;**23**:166–172.
 85. Nopp A, Cardell LO, Johansson SGO, Oman H. CD-sens: a biological measure of immunological changes stimulated by ASIT. *Allergy* 2009;**64**:811–814.
 86. Zidarn M, Košnik M, Šilar M, Bajrović N, Korošec P. Sustained effect of grass pollen subcutaneous immunotherapy on suppression of allergen-specific basophil response; a real-life, nonrandomized controlled study. *Allergy* 2015;**70**:547–555.
 87. Kepil Özdemiir S, Sin BA, Gülođlu D, İkin-çiođulları A, Gençtürk Z, Mısırlıgil Z. Short-term preseasonal immunotherapy: is early clinical efficacy related to the basophil response? *Int Arch Allergy Immunol* 2014;**164**:237–245.
 88. Aasbjerg K, Backer V, Lund G, Holm J, Nielsen NC, Hølse M et al. Immunological comparison of allergen immunotherapy tablet treatment and subcutaneous immunotherapy against grass allergy. *Clin Exp Allergy* 2014;**44**:417–428.
 89. Van Overtvelt L, Baron-Bodo V, Horiot S, Moussu H, Ricarte C, Horak F et al. Changes in basophil activation during grass-pollen sublingual immunotherapy do not correlate with clinical efficacy. *Allergy* 2011;**66**:1530–1537.
 90. Jones SM, Pons L, Roberts JL, Scurlock AM, Perry TT, Kulis M et al. Clinical efficacy and immune regulation with peanut oral immunotherapy. *J Allergy Clin Immunol* 2009;**124**:292–300.
 91. Klunker S, Saggari LR, Seyfert-Margolis V, Asare AL, Casale TB, Durham SR et al. Combination treatment with omalizumab and rush immunotherapy for ragweed-induced allergic rhinitis: inhibition of IgE-facilitated allergen binding. *J Allergy Clin Immunol* 2007;**120**:688–695.
 92. Chaker AM, Shamji MH, Dumitru FA, Calderon MA, Scadding GW, Makatsori M et al. Short-term subcutaneous grass pollen immunotherapy under the umbrella of anti-IL-4: a randomized controlled trial. *J Allergy Clin Immunol* 2016;**137**:452–461.
 93. Mothes N, Heinzkill M, Drachenberg KJ, Sperr WR, Krauth MT, Majlesi Y et al. Allergen-specific immunotherapy with a monophosphoryl lipid A-adjuvanted vaccine: reduced seasonally boosted immunoglobulin E production and inhibition of basophil histamine release by therapy-induced blocking antibodies. *Clin Exp Allergy* 2003;**33**:1198–1208.
 94. Creticos PS, Schroeder JT, Hamilton RG, Balcer-Whaley SL, Khattignavong AP, Lindblad R et al. Immunotherapy with a Ragweed-Toll-like receptor 9 agonist vaccine for allergic rhinitis. *N Engl J Med* 2006;**355**:1445–1455.
 95. Worm M, Patel D, Creticos PS. Cat peptide antigen desensitisation for treating cat allergic rhinoconjunctivitis. *Expert Opin Investig Drugs* 2013;**22**:1347–1357.
 96. Couroux P, Patel D, Armstrong K, Larché M, Hafner RP. Fel d 1-derived synthetic peptide immuno-regulatory epitopes show a long-term treatment effect in cat allergic subjects. *Clin Exp Allergy* 2015;**45**:974–981.
 97. Ellis AK, Frankish CW, O'Hehir RE, Armstrong K, Steacy L, Larché M et al. Treatment with grass allergen peptides improves symptoms of grass pollen-induced allergic rhinoconjunctivitis. *J Allergy Clin Immunol* 2017: <http://dx.doi.org/10.1016/j.jaci.2016.11.043>
 98. Spertini F, Perrin Y, Audran R, Pellaton C, Boudousquie C, Barbier N et al. Safety and immunogenicity of immunotherapy with Bet v 1-derived contiguous overlapping peptides. *J Allergy Clin Immunol* 2014;**134**:239–240.
 99. Pellaton C, Perrin Y, Boudousquie C, Barbier N, Wassenberg J, Corradin G et al. Novel birch pollen specific immunotherapy formulation based on contiguous overlapping peptides. *Clin Transl Allergy* 2013;**3**:17.
 100. Spertini F, de Blay F, Jacobsen L. Ultrafast hypoallergenic birch pollen allergy vaccine AllerT is efficient and safe: result of a phase IIb study. *J Allergy Clin Immunol* 2014;**133**:AB290.
 101. Spertini F, Jutel M, Jacobsen L. Sustained efficacy of AllerT allergy vaccine after a second birch pollen season: a phase IIb study. San Diego 2015:AB143.
 102. Senti G, Prinz Vavricka BM, Erdmann I, Diaz MI, Markus R, McCormack SJ et al. Intralymphatic allergen administration renders specific immunotherapy faster and safer: a randomized controlled trial. *Proc Natl Acad Sci USA* 2008;**105**:17908–17912.
 103. Hylander T, Latif L, Petersson-Westin U, Cardell LO. Intralymphatic allergen-specific immunotherapy: an effective and safe alternative treatment route for pollen-induced allergic rhinitis. *J Allergy Clin Immunol* 2013;**131**:412–420.
 104. Senti G, Cramer R, Kuster D, Johansen P, Martinez-Gomez JM, Graf N et al. Intralymphatic immunotherapy for cat allergy induces tolerance after only 3 injections. *J Allergy Clin Immunol* 2012;**129**:1290–1296.
 105. Witten M, Malling H-J, Blom L, Poulsen BC, Poulsen LK. Is intralymphatic immunotherapy ready for clinical use in patients with grass pollen allergy? *J Allergy Clin Immunol* 2013;**132**:1248–1252.
 106. Schmid JM, Nezam H, Madsen HHT, Schmitz A, Hoffmann HJ. Intralymphatic immunotherapy induces allergen specific plasmablasts and increases tolerance to skin prick testing in a pilot study. *Clin Transl Allergy* 2016;**6**:19.
 107. Helbo Skaarup S, Schmid JM, Skjold T, Graumann O, Hoffmann HJ. ILIT is a similarly effective treatment of grass pollen induced rhinitis for patients with moderate/severe and mild symptoms. *Allergy*, **71**(S102):60–61.
 108. Hylander T, Larsson O, Petersson-Westin U, Eriksson M, Kumlien Georén S, Winqvist O et al. Intralymphatic immunotherapy of pollen-induced rhinoconjunctivitis: a double-blind placebo-controlled trial. *Respir Res* 2016;**17**:10.
 109. Patterson AM, Bonny AE, Shiels WE, Erwin EA. Three-injection intralymphatic immunotherapy in adolescents and young adults with grass pollen rhinoconjunctivitis. *Ann Allergy Asthma Immunol* 2016;**116**:168–170.
 110. Valenta R, Ferreira F, Focke-Tejkl M, Linhart B, Niederberger V, Swoboda I et al. From allergen genes to allergy vaccines. *Annu Rev Immunol* 2010;**28**:211–241.
 111. Stringari G, Tripodi S, Caffarelli C, Dondi A, Asero R, Di Rienzo Businco A et al. The effect of component-resolved diagnosis on specific immunotherapy prescription in children with hay fever. *J Allergy Clin Immunol* 2014;**134**:75–81.

112. Pauli G, Larsen TH, Rak S, Horak F, Pastorello E, Valenta R et al. Efficacy of recombinant birch pollen vaccine for the treatment of birch-allergic rhinoconjunctivitis. *J Allergy Clin Immunol* 2008;**122**: 951–960.
113. Jutel M, Jaeger L, Suck R, Meyer H, Fiebig H, Cromwell O. Allergen-specific immunotherapy with recombinant grass pollen allergens. *J Allergy Clin Immunol* 2005;**116**:608–613.
114. Focke-Tejkl M, Weber M, Niespodziana K, Neubauer A, Huber H, Henning R et al. Development and characterization of a recombinant, hypoallergenic, peptide-based vaccine for grass pollen allergy. *J Allergy Clin Immunol* 2015;**135**:1207–1217.
115. Ziegelmayer P, Focke-Tejkl M, Schmutz R, Lemell P, Ziegelmayer R, Weber M et al. Mechanisms, safety and efficacy of a B cell epitope-based vaccine for immunotherapy of grass pollen allergy. *EBioMedicine* 2016;**11**:43–57.
116. Cornelius C, Schöneweis K, Georgi F, Weber M, Niederberger V, Ziegelmayer P et al. Immunotherapy with the PreS-based grass pollen allergy vaccine BM32 induces antibody responses protecting against hepatitis B infection. *EBioMedicine* 2016;**11**:58–67.
117. Valenta R, Campana R, Focke-Tejkl M, Niederberger V. Vaccine development for allergen-specific immunotherapy based on recombinant allergens and synthetic allergen peptides: lessons from the past and novel mechanisms of action for the future. *J Allergy Clin Immunol* 2016;**137**:351–357.
118. Shamji MH, Durham SR. Mechanisms of immunotherapy to aeroallergens. *Clin Exp Allergy* 2011;**41**:1235–1246.
119. Matricardi PM, Kleine-Tebbe J, Hoffmann HJ, Valenta R, Hilger C, Hofmaier S et al. EAACI Molecular Allergology User's Guide. *Pediatric Allergy and Immunology* 2016;**27**:1–250.