



**UNIVERSITY
OF TURKU**

This is a self-archived – parallel-published version of an original article. This version may differ from the original in pagination and typographic details. When using please cite the original.

AUTHOR	Tomi Suomi, Ubaid Ullah Kalim, Omid Rasool, Asta Laiho, Henna Kallionpää, Mari Vähä-Mäkilä, Mirja Nurmio, Juha Mykkänen, Taina Härkönen, Heikki Hyöty, Jorma Ilonen, Riitta Veijola, Jorma Toppari, Mikael Knip, Laura L. Elo, Riitta Lahesmaa
TITLE & JOURNAL	Type 1 Diabetes in Children With Genetic Risk May Be Predicted Very Early With a Blood miRNA. - <i>Diabetes Care</i>
YEAR	2022, April
LINK TO ORIGINAL	https://doi.org/10.2337/dc21-2120
THIS VERSION	Final Draft (AAM)
CITATION	Tomi Suomi, Ubaid Ullah Kalim, Omid Rasool, Asta Laiho, Henna Kallionpää, Mari Vähä-Mäkilä, Mirja Nurmio, Juha Mykkänen, Taina Härkönen, Heikki Hyöty, Jorma Ilonen, Riitta Veijola, Jorma Toppari, Mikael Knip, Laura L. Elo, Riitta Lahesmaa; Type 1 Diabetes in Children With Genetic Risk May Be Predicted Very Early With a Blood miRNA. <i>Diabetes Care</i> 1 April 2022; 45 (4): e77–e79. https://doi.org/10.2337/dc21-2120

Type 1 Diabetes in Children with Genetic Risk may potentially be predicted very early with a blood miRNA

Tomi Suomi^{1,2*}, Ubaid Ullah Kalim^{1,2*}, Omid Rasool^{1,2}, Asta Laiho^{1,2}, Henna Kallionpää¹, Mari Vähä-Mäkilä^{3,4}, Mirja Nurmio^{3,4}, Juha Mykkänen^{4,5}, Taina Härkönen^{6,7}, Heikki Hyöty^{8,9}, Jorma Ilonen¹⁰, Riitta Veijola^{11,12}, Jorma Toppari^{2,3,4,13}, Mikael Knip^{6,7,14}, Laura L. Elo^{1,2,15#}, and Riitta Lahesmaa^{1,2#}

¹Turku Bioscience Centre, University of Turku and Åbo Akademi University, Turku, Finland

²InFLAMES Research Flagship Center, University of Turku, Turku Finland

³Research Centre for Integrative Physiology and Pharmacology, Institute of Biomedicine, University of Turku, Turku, Finland

⁴Centre for Population Health Research, University of Turku and Turku University Hospital, Turku, Finland

⁵Research Centre of Applied and Preventive Cardiovascular Medicine, University of Turku, Turku, Finland

⁶Pediatric Research Center, Children's Hospital, University of Helsinki and Helsinki University Hospital, Helsinki, Finland

⁷Research Program for Clinical and Molecular Metabolism, Faculty of Medicine, University of Helsinki, Helsinki, Finland

⁸Department of Virology, Faculty of Medicine and Biosciences, University of Tampere, Tampere, Finland

⁹Fimlab Laboratories, Pirkanmaa Hospital district, Tampere Finland

¹⁰Immunogenetics Laboratory, Institute of Biomedicine, University of Turku, Turku, Finland

¹¹Department of Pediatrics, PEDEGO Research Unit, Medical Research Centre, University of Oulu, Oulu, Finland

¹²Department of Children and Adolescents, Oulu University Hospital, Oulu, Finland

¹³Department of Pediatrics, Turku University Hospital, Turku, Finland

¹⁴Centre for Child Health Research, Tampere University Hospital, Tampere, Finland

¹⁵Institute of Biomedicine, University of Turku, Turku, Finland

*These authors contributed equally

#Correspondence

Professor Riitta Lahesmaa
Turku Bioscience Centre,
Tykistökatu 6A, Turku 20520, FINLAND
Email: rilahes@utu.fi

Professor Laura Elo
Turku Bioscience Centre
Tykistökatu 6A, Turku 20520, FINLAND
Email: laura.elo@utu.fi

Progression to clinical type 1 diabetes is monitored through the appearance of islet

autoantibodies against pancreatic β -cell antigens, and most children with two or more

41 autoantibodies progress to disease (1). However, autoantibodies indicate already active islet
42 autoimmunity, and by that time, loss of immune tolerance may have reached a point of no
43 return. Thus, there is an urgent need for biomarkers that would predict the disease before the
44 appearance of islet autoantibodies and provide a longer window for intervention. New
45 biomarkers might help identify optimal sets of subjects for clinical trials or a subgroup of patients
46 who may more likely benefit from a given therapy.

47 MicroRNAs (miRNAs) secreted in extracellular vesicles have been detected in blood and may
48 have biomarker potential (2). Several studies have shown the usefulness of miRNAs as
49 biomarkers for many diseases (reviewed in (3)). Aberrant miRNA expression has been
50 observed in sera of type 1 diabetes patients (reviewed in (4)). However, most miRNA studies
51 have analyzed samples at or after the onset of clinical type 1 diabetes.

52 Here we analyzed whether we can detect changes in miRNA levels before and during islet
53 autoimmunity in whole blood samples from children with HLA-conferred risk of type 1 diabetes
54 participating in the Type 1 Diabetes Prediction and Prevention (DIPP) study (5). Children from
55 the DIPP study with high HLA-conferred risk were followed up, and whole blood samples were
56 collected at multiple time points. Case-control matching was based on HLA-DQB1 genotype,
57 date and place of birth, and sex, similarly as described earlier (5). To study genome-wide
58 miRNA profiles before the clinical presentation of type 1 diabetes, we first performed miRNA-
59 sequencing (miRNA-seq) on 87 longitudinal samples collected from four multiple autoantibody-
60 positive cases and their matched autoantibody negative controls (**Fig. 1A: miRNA-seq**
61 **discovery cohort**) using the Illumina HiSeq 2500 platform. A linear mixed-effects model for
62 each miRNA was used to test differential expression between cases and controls. The most
63 significantly upregulated miRNA in cases was hsa-miR-6868-3p ($p < 0.001$), which has not been
64 earlier associated with type 1 diabetes. Interestingly, hsa-miR-6868-3p was upregulated already
65 before seroconversion (**Fig. 1B**). We confirmed the finding ($p < 0.001$) on these and ten

66 additional case-control pairs using miRNA-seq of two time-points before seroconversion (**Fig.**
67 **1A: miRNA-seq validation cohort; Fig. 1C**).

68 We further confirmed the miRNA upregulation by TaqMan qRT-PCR assay in samples collected
69 from 29 case-control pairs, of which 14 were included in the miRNA-seq analysis (**Fig. 1A:**
70 **TaqMan validation**). A strong correlation ($r=0.75$) was observed between the sequencing and
71 TaqMan results for hsa-miR-6868-3p expression. Convincingly, the TaqMan data showed
72 higher hsa-miR-6868-3p expression in cases than controls ($p < 0.001$) across the time points
73 (**Fig. 1D**), recapitulating the miRNA-seq result.

74 Given the early upregulation of hsa-miR-6868-3p, we tested whether the miRNA can classify the
75 29 cases from controls already before seroconversion, using the average expression before
76 seroconversion for a given child. The ΔC_t expression values were adjusted for individual HLA
77 type and TaqMan plates using a linear model. The area under the receiver operating
78 characteristic curve (AUROC) was 0.76 (**Fig. 1E**), suggesting that the miRNA may indeed
79 potentially serve as a screening biomarker for the stratification of children at increased genetic
80 risk for type 1 diabetes.

81 Besides blood, breast and brain, the miRNA is expressed in the pancreas (**Fig. 1F**), suggesting
82 an interesting possibility that its upregulation in blood samples of case children may potentially
83 originate from the pancreas and circulate through blood under inflammatory conditions. It is
84 important to note that this miRNA may also come from blood lymphocytes, given its expression
85 in these cells (**Fig. 1G**).

86 The ROC analysis implied that hsa-miR-6868-3p may serve as a screening biomarker for the
87 stratification of children at risk of islet autoimmunity. However, the small cohort size of 58 study
88 subjects is a limitation of the study, and the finding should be validated in whole-blood samples
89 of an independent, preferably larger cohort. For a screening biomarker, high sensitivity is
90 preferred over high specificity (i.e., false negatives are of more concern than false positives).

91 The current model reaches a higher sensitivity of 0.86 at the specificity of 0.66. It remains to be
92 seen whether combining hsa-miR-6868-3p with other miRNAs or mRNAs will improve the
93 performance of the predictive model. It will also be interesting to determine whether the miRNA
94 expression correlates with the time from seroconversion to clinical disease.

95 **Declarations**

96 *Data availability*

97 Analyzed count data from miRNA-seq of the discovery cohort (87 samples from four case-
98 control pairs) and the validation cohort (56 samples from fourteen pairs) can be accessed from
99 the ArrayExpress database (<https://www.ebi.ac.uk/arrayexpress/>) using the accession codes, E-
100 MTAB-10959 and E-MTAB-10968, respectively. Other data generated during the current study
101 are available from the corresponding author on a reasonable request.

102 *Funding*

103 This research was supported by the Academy of Finland including InFLAMES, SYMMyS, and
104 personalized medicine programs (grant numbers: 337530, 292482, 250114, 292482, 294337,
105 292335, 319280 and 314444 329277, 331790, 296801, 304995, 310561, 314443, and 329278);
106 Juvenile Diabetes Research Foundation, including grants 1-SRA-2016-342-M-R and 1-SRA-
107 2019-732-M-B; the Diabetes Research Foundation (Diabetestutkimussäätiö); the Novo Nordisk
108 Foundation; and the Finnish Cancer Foundation; the Sigrid Jusélius Foundation; Pediatric
109 Research Foundation; Turku University Hospital Special Governmental Grants; and the Special
110 Research Funds for University Hospitals in Finland. It was also supported by grants from the
111 European Research Council ERC (677943); European Union's Horizon 2020 research and
112 innovation programme (955321); and the European Foundation for the study of Diabetes.
113 Biocenter Finland and ELIXIR Finland also support our research.

114 Authors' contributions

115 TS and UUK designed experiments, analyzed the data, prepared the figures, and wrote the
116 manuscript. OR designed experiments, analyzed data, and wrote the manuscript. AL, HK, MV-
117 M, MN, JM, and TH contributed to the design of the study and analysis. HH, JI, RV, JT and MK
118 were responsible for the DIPP cohort. RV and MK were responsible for the islet autoantibody
119 analyses. All authors contributed to the final version of the manuscript. RL initiated, designed
120 and supervised the study. LLE participated in the design of the study and analysis, and
121 supervised the study. RL and LLE are the guarantors of this work and, as such, had full access
122 to all the data in the study and take responsibility for the integrity of the data and the accuracy of
123 the data analysis.

124 Acknowledgements

125 The authors are grateful to the families of the DIPP study participants and the study group. We
126 also thank the personnel of the HLA laboratory at the University of Turku and the islet
127 autoantibody laboratory at the University of Oulu. Marjo Hakkarainen and Sarita Heinonen are
128 acknowledged for their skilful assistance in the laboratory. NGS sequencing was performed at
129 the Finnish Functional Genomics Centre (FFGC), Turku, part of the Biocenter Finland network.
130 We acknowledge the Finnish Centre for Scientific Computing (CSC) for data analysis servers.

131 Duality of Interest

132 The authors declare that there is no conflict of interest.

133 References

- 134 1. Ziegler AG, Rewers M, Simell O, Simell T, Lempainen J, Steck A, et al. Seroconversion
135 to multiple islet autoantibodies and risk of progression to diabetes in children. *JAMA*.
136 2013 Jun;309(23):2473–9.
- 137 2. Condrat CE, Thompson DC, Barbu MG, Bugnar OL, Boboc A, Cretoiu D, et al. miRNAs

- 138 as Biomarkers in Disease: Latest Findings Regarding Their Role in Diagnosis and
139 Prognosis. *Cells* [Internet]. 2020;9(2). Available from:
140 <http://dx.doi.org/10.3390/cells9020276>
- 141 3. Das S, Abdel-Mageed AB, Adamidi C, Adelson PD, Akat KM, Alsop E, et al. The
142 Extracellular RNA Communication Consortium: Establishing Foundational Knowledge
143 and Technologies for Extracellular RNA Research. *Cell*. 2019;177(2):231–42.
- 144 4. Ventriglia G, Nigi L, Sebastiani G, Dotta F. MicroRNAs: Novel Players in the Dialogue
145 between Pancreatic Islets and Immune System in Autoimmune Diabetes. *Biomed Res Int*
146 [Internet]. 2015;2015:749734. Available from: <http://dx.doi.org/10.1155/2015/749734>
- 147 5. Kallionpää H, Elo LL, Laajala E, Mykkänen J, Ricano-Ponce I, Vaarma M, et al. Innate
148 immune activity is detected prior to seroconversion in children with HLA-conferred type 1
149 diabetes susceptibility. *Diabetes*. 2014 Jul;63(7):2402–14.

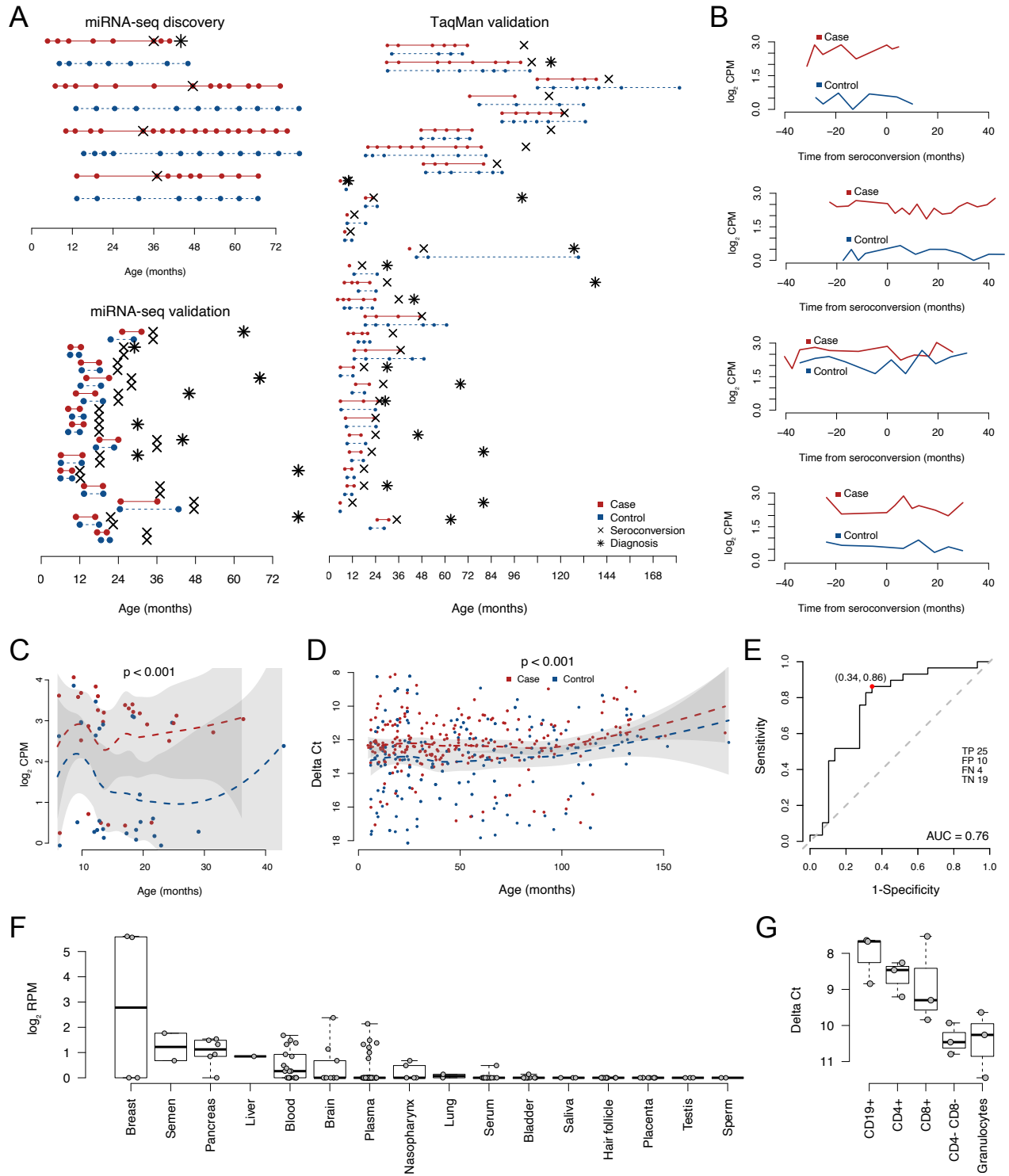
150

151

152

153

154 Figure Legend



155

156 Figure 1. hsa-miR-6868-3p as early marker for type 1 diabetes

157 (A) The samples from the miRNA-seq discovery cohort (top left), the miRNA-seq validation cohort
158 (bottom left), and the TaqMan validation cohort (right). Each line is an individual, and each dot is
159 a PAXgene sample. (B) The line plots showing the expression profiles of hsa-miR-6868-3p for
160 the four case-control pairs of the discovery cohort. The plots are seroconversion centered. CPM
161 stands for counts per million. (C) Line plots showing the average longitudinal case-control profiles
162 of hsa-miR-6868-3p in the validation cohort. Each red and blue dot shows a case or a control
163 sample, respectively. The dashed lines show average expression, and the grey area shows a
164 95% confidence interval. (D) TaqMan expression of hsa-miR-6868-3p over time in 29 case-control
165 pairs. The p value shown at the top was obtained from the linear mixed-effects model. (E)
166 Receiver operating characteristic (ROC) analysis showing the performance of hsa-miR-6868-3p
167 in differentiating the DIPP cases from controls in samples before the appearance of islet
168 autoantibodies. The delta Ct values were adjusted using linear regression with HLA-type
169 (DR3/DR4, DR3/DR3, DR4/DR4, DR3/other, DR4/other) and TaqMan plate as explanatory
170 variables. The resulting residuals were used to normalize the possible effects of HLA and plate.
171 True positives (TP), true negatives (TN), false positives (FP), and false negatives (FN) as
172 predicted by the model are shown on the plot. (F) The expression profile of hsa-miR-6868-3p
173 across different tissues. The data was taken from miRmine database
174 (<https://guanfiles.dcmf.med.umich.edu/mirmine/index.html>). (G) Expression of hsa-miR-6868-3p
175 in blood cells of healthy donors (n=3). Each dot represents an individual.

176