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**Determining the Phylogenetic Relationships Among the Northern Pike (*Esox lucius*),
the Muskellunge (*Esox masquinongy*), and the Silver Pike.**

Thesis Submitted to
The Graduate College of
Marshall University

In partial fulfillment of the
Requirements for the Degree of
Masters of Science Program

by

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Determining the Phylogenetic Relationships Among the Northern Pike (*Esox lucius*), the Muskellunge (*Esox masquinongy*), and the Silver Pike.

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Keywords: Silver pike, northern pike (*Esox lucius*), muskellunge (*E. masquinongy*), mitochondrial 16 rRNA, phylogeny.

Prioritization of taxa has become a necessity due to limited resources available for conservation. Conservation efforts are often concentrated on those threatened and endangered species with the highest taxonomic importance. The silver pike is a rare game fish, often accounting for less than 1% of a total pike population where it is found. Although silver pike are normally found in association with northern pike, they have distinct morphological characteristics and have not been shown to hybridize with the northern pike. Still, many fisheries biologists consider the silver pike to be a color variant of the northern pike, and to date the silver pike has not been assigned to a taxonomic unit. This study employs the use of a molecular marker on the mitochondrial genome to determine the phylogenetic relationships between the northern pike, silver pike and muskellunge. Mitochondrial DNA was amplified from individual scales, cloned into the pCR[®]2.1 vector and sequenced at the Marshall Core Facility. The sequences were aligned using ClustalX (Thompson et al., Nuc. Acids Res. 22:4673), and phylogenetic distances and tree topologies were inferred using the programs of the PHYLIP package (Felsenstein, Cladistics 5:164). Sequence data was sufficient to distinguish between the northern pike and muskellunge. No sequence variation was found between the silver pike and northern pike. Resulting tree topologies were unable to distinguish between the silver pike and northern pike in all cases. Molecular data supports the hypothesis that the silver pike is a color variant of the northern pike and not a separate species.

CHAPTER I

INTRODUCTION

Silver Pike

The silver pike is a member of the family Esocidae and most closely resembles the northern pike (*Esox lucius*) in morphological characters. The silver pike is often referred to as a blue pike in Canada. This common name often leads to confusion with a subspecies of the walleye (*Stizostedion vitreum glaucum*) from the Great Lakes region, which is believed to be extinct. Few publications exist on the silver pike. The only peer reviewed journal article from Lawler (1960) stated that the silver pike is a color mutation of the northern pike. Scott and Crossman (1973), Eddy and Underhill (1974), and Becker (1983) briefly discuss the silver pike, mainly pointing out the obvious color difference between the two fish. The majority of citations originally stem from an earlier publication by Eddy and Surber (1943).

The correct common name is the silver pike rather than the silver muskellunge once used by Minnesota fisherman (Eddy and Underhill, 1974). The most distinct difference between the northern pike and the silver pike is that of color (Fig. 1). The northern pike has a basic color arrangement of light yellow to whitish bean shaped spots about as long as the diameter of the eye on a dark, brilliant green to olive background. The body has the appearance of being flecked with gold, caused by a tiny gold spot on the tip of the exposed edge of most body scales. The dorsal, caudal and anal fins are green to yellow and sometimes orange or pale red with large irregular black markings.

Paired fins are usually unmarked and buff to dusky (Scott and Crossman, 1973). The silver pike has deep blue color in the dorsal area, shading through lighter blue and gray devoid of bean shaped spots on the flanks, to white on the ventral surface. The flanks of the silver pike are more specifically dark silver or gray and sometimes flecked with gold. The fins appear to be almost clear with a tint of orange and red and may be speckled with black, particularly the ventral fins (Lawler, 1960). Occasionally, silver pike are found with faint spots on the caudal peduncle, similar to those found on the northern pike but less distinct (Eddy and Underhill, 1974). Silver pike are distinguishable from the northern pike even after death as their silver color intensifies rather than fades (Lawler, 1960).

Although coloration is the most obvious difference between silver pike and northern pike, there are several other subtle morphological differences. First, in a sample of 33 silver pike compared to 189 northern pike, two silver pike had a higher lateral line scale count than the maximum recorded for the northern pike (Lawler, 1960). Though this is not statistically significant, it supports anecdotal information from fisherman that suggest silver pike have finer scales than the northern pike, similar to a rainbow trout (*Onkorhynchus mykiss*). Silver pike have consistently narrow body width in individuals between 150 to 400 mm, while silver pike between 400 to 550 mm were not different from northern pike of the same size. Interorbital width was the most substantial morphological difference between the northern and silver pike. Although the diameter of the eye can not be measured as accurately as a bony structure, interorbital width confirmed that eye diameter was larger in the silver pike than in the northern pike

(Lawler, 1960). When interorbital distances were plotted arithmetically against the standard length, it was possible to draw a line through all the points above which only northern pike occurred and below which only silver pike occurred (Lawler, 1960; Fig. 2). In the same study, measurements of the mandible and maxillary of the silver pike were plotted against the standard lengths to show that 23 of the 28 silver pike had shorter mandibles than did northern pike. It was also shown that 24 of the 26 silver pike had shorter maxillary lengths than northern pike. This suggests that silver pike have a shorter snout length than do northern pike, though no statistical analysis was performed. Bernardo (1998) stated that silver pike have a caudal fin shape similar to that of the muskellunge, the tips being more pointed than those of northern pike, which are noticeably more rounded. This was also noted in observations made during the present study. Although the average size of the silver pike is similar to that of the northern pike, it rarely reaches the maximum sizes of the northern pike. Silver pike seldom exceed ten pounds (Eddy and Underhill, 1974), though larger fish have been reported. Lawler (1960) also noted that at each age the average length and weight of silver pike was greater than that of northern pike.

There are also morphological similarities between the silver pike and the northern pike. As is common with the northern pike, the opercle of the silver pike is only scaled on the upper half while the cheek is completely scaled. The submandibular pore count resembled that of the northern pike with five on both sides totaling ten (Lawler, 1960). Branchiastegal ray counts fell within the range of the counts for northern pike as did fin ray counts and average head lengths for all specimens observed. Stomach contents

showed that silver pike feed on foods similar to that of northern pike (Lawler, 1960). Food items included perch (*Perca flavescens*), common sucker (*Catostomus commersoni*), spottail shiner (*Notropis hudsonius*), nine spine stickleback (*Pungitius pungitius*), and troutperch (*Percopsis omiscomaycus*).

Silver pike captured and placed in aquaria appeared to be exceptionally hardy. One specimen went 22 days before eating, although ample food was supplied. The same fish was dipped twice in formaldehyde to kill *Saprolegnia*, and still lived for 47 days after capture (Lawler, 1960). In another account by Lawler (1960), a silver pike was caught, held by the eyes to dislodge a hook, placed on the deck of the boat where it remained for two hours before being placed in an aquarium, and survived without the appearance of ill effect. Scott and Crossman (1973) also reported that silver pike is similar to northern pike except for an apparent increased hardiness.

Several breeding experiments have been reported but not extensively documented. Silver pike specimens were incidentally obtained while collecting brood stock muskellunge for the Nevis Fish Hatchery from Lake Belletaine, Minnesota. These silver pike were propagated along with the true muskellunge and stocked in nearby lakes for several years. Breeding experiments at the Spirit Lake Hatchery, Iowa (Eddy and Underhill, 1974) found that silver pike breed true in that all offspring were similar to the parents. When silver pike were crossed with northern pike, the offspring had a peculiar black mottling similar to that of the black crappie (*Pomoxis nigromaculatus*) and subsequent generations could not be obtained (Runnström, 1949). This suggests that the offspring from a silver pike-northern pike cross are sterile. When the silver pike was

crossed with the muskellunge the resulting F1 generation resembled a tiger muskellunge which is a cross between the muskellunge and a northern pike. Since no specimens having the black mottled pattern of a silver pike-northern pike hybrid have been taken in the field, it is believed that the silver pike and northern pike do not hybridize in nature.

No research has been done on spawning times or behaviors of silver pike, although silver pike have been captured with spawning muskellunge (Eddy and Underhill, 1974) and spawning pike (Lawler, 1960). All members of the family Esocidae spawn in the same manner. In general, species within this family spawn in the spring in heavily vegetated flood plains. Eggs are scattered at random over the vegetation as large females and smaller males roll bringing vents into proximity while eggs and milt are extruded. Rapid thrusting of tails further scatter the settling eggs, which are not protected by either parent. Because the eggs of these species are scattered in such a manner, the presence of spawning silver pike in the general location of spawning northern pike would lead to an increased occurrence of the silver pike-northern pike hybrids. It is difficult to distinguish the difference between a silver pike-muskellunge and a northern pike-muskellunge hybrid, so occurrence of this hybrid would not be as easily detected. Due to the lack of parental care of fertilized eggs and fingerlings, and the fast growth rate of the northern pike fingerlings, subsequent spawning runs of other fish provide excellent forage for the voracious northern pike fingerlings. This is the fate of many muskellunge fry (Scott and Crossman, 1973) where the muskellunge and northern pike occur together, and may also be the fate of silver pike, explaining decreased population densities.

The most limiting factor on the research of the silver pike is its small population density in relation to the total pike population. Of the total pike population of Heming Lake in Manitoba, Canada, it was estimated that only 0.2 percent were silver pike (Lawler, 1960). From 1952 to 1958, 19,420 pike were taken from Heming Lake, of which only 42 were silver pike. Sixty pike were taken from Angler's Lake, Manitoba, in 1958. Only two of the 60 fish were silver pike. The following year the same ratio applied. In 1959 at Beaverlodge Lake in Mackenzie, Canada, one silver pike was caught out of 151 total pike (Lawler, 1960). From a small lake near the Marten River in Ontario, out of 104 pike caught, four were silver pike (personal observations). Early spawning times, faster growth, better food conversion, and early dominance of forage fish have been cited to explain the northern pike's domination of other Esocids where habitats overlaps (Scott and Crossman, 1973).

Discussions on the classification of the silver pike are clouded by confusion and misunderstanding among fisheries biologists. Becker (1983) stated that hybridization between the silver pike and muskellunge leads to offspring similar to muskellunge-northern pike hybrids, which is speckled like a crappie. The muskellunge-northern pike hybrid, known as the tiger muskellunge, is characterized by having a strong barring pattern on its sides (Page and Burr, 1991). Eddy and Surber (1943) stated that the muskellunge-silver pike hybrids do resemble muskellunge-northern pike hybrids, but it is the silver pike-northern pike hybrids that are characterized by having the crappie-like black mottling. Anecdotal reports from fisheries biologist have reported that silver pike are dominant where they are found and only occur in isolated areas. This contradicts

distribution described by Lawler (1960) and Eddy and Surber (1943) and adds to the confusion among fisheries scientists. The lack of research and familiarity with this fish has resulted in its being arbitrarily categorized as a color mutation of the northern pike based on distribution and morphological characteristics.

Molecular Chronometers

The use of molecules as “documents of evolutionary history,” or “molecular chronometers” was first introduced in 1965 by Emile Zuckerkandl and Linus Pauling (Wolfgang and Schleifer, 1999; Woese, 1987). The advent of molecular chronometers provides taxonomists with a powerful tool to complement phenotypic techniques. Genotypic information is more readily, reliably, and precisely interpreted than phenotypic patterns allowing for increased information on evolutionary relationships (Woese, 1987). In addition, molecular characters are less likely related to adaptive evolution than are morphological characters (Briolay et al., 1998). Despite the benefits inherent in phylogenies derived from sequence data, it is important to remember that genetic information yields hypotheses to be tested and either strengthened or rejected on the basis of other kinds of data (Woese, 1987). In cases where molecules and morphology disagree, provisional morphology-based constraints on the analysis of molecular data offer a practical means of integrating the two types of data (Normark et al., 1991).

Mitochondrial DNA (mtDNA) mutates at a rate that is ten times faster than that of nuclear DNA (Wiesner et al., 1991; Patarnello et al., 1993). This rapid mutation rate

makes mitochondrial DNA a powerful tool in estimating the degree of divergence between closely related species (Wilson et al., 1985; Avise et al., 1987). In addition, the ease with which mitochondrial genes can be isolated and sequenced coupled with special features of mitochondria such as lack of introns, maternal inheritance, absence of recombination events, and haploidy also contribute to its reputation as a reliable tracer of evolutionary history (Zardoya and Meyer, 1996). A marvel of genetic economy, the mitochondrial genome encodes 22 tRNAs, 13 mRNAs, and 2 rRNAs in a 16.5-to 17.5- kb closed circular molecule (Digby et al., 1992; Lake, 1998).

Molecules whose sequences change randomly in time can be considered chronometers (Woese, 1987). In order to yield useful information, the clocklike behavior of molecular chronometers should measure, and be representative of, the overall rate of evolution. The amount of change within a sequence that accumulates (proportionate to genetic distance) is the product of the rate (in occurrence of fixed mutations) multiplied by the time over which these mutations occur. Because distance can not be measured from the original to final states of evolution due to the lack of original states, distances are measured between extant forms. The distance from original states can then be estimated given comparable rates of change in each line.

Ribosomal RNA has been termed the ultimate molecular chronometer (Woese, 1987) and was chosen for this study for several reasons. They are found in all organisms. Their large size provides several domains changing at varying rates; this allows for different levels of evolution to be evaluated. The functional constancy and genetic stability assures a relatively good clocklike behavior. The 16S rRNA contains highly

variable regions that change fast enough to provide information on closely related species (Mangold et al., 1997). There has also been a large sequence database describing a wide spectrum of phylogenetically diverse organisms developed over the past decade.

Mitochondrial 16S rRNA gene has developed the reputation of being a reliable tracer of evolutionary history. Another common molecule used for determining phylogenetic relationships is cytochrome *b*. Some studies indicate, however, that phylogenetic relationships among deep branching lineages are poorly resolved using cytochrome *b* sequences (Briolay et al., 1998). Patarnello et al. (1994) used 16S rRNA gene sequence variation combined with that of cytochrome *b* to infer phylogeny of the brown trout (*Salmo trutta*). They found that the 16S rRNA gene alone gave identical results to those of the combined sequences.

This study employs the use of the mitochondrial 16S rRNA gene as a molecular chronometer to determine the phylogenetic relationships between the northern pike, silver pike and muskellunge. Comparison of mtDNA sequences has become a common way to establish phylogenetic relationships among closely related genera (Woese, 1987; Mangold et al., 1997; Parker and Kornfield, 1996; Normark et al., 1991; Ritchie et al., 1997; Patarnello et al., 1993; Danzmann and Ihssen, 1995; Matthee and Robinson, 1997; Murphy and Collier, 1997; Briolay et al., 1998; Gilles et al., 1998). The use of morphological characteristics such as body size, color, behavior, ecophysiology, and meristics alone to classify higher vertebrates is becoming more questionable, due to the phenotypic plasticity of many similar species. Adding the use of molecular chronometers to existing phenotypic techniques to distinguish between phenotypic variants, subspecies,

and true Linnean species may lead to fewer inappropriate taxonomic divisions.

In recent phylogenetic studies, DNA segments of 300-600 base pairs have been found to be sufficient to infer evolutionary relationships (Kocher et al., 1989; Patarnello et al., 1993). For this study, a 370 bp region of the 16S rRNA gene was used. The northern pike and muskellunge, two clearly distinct species, were used as a control to establish a phylogenetic distance sufficient to delineate species. According to Mangold et al. (1997), a difference of 1.3 percent was sufficient to delineate between two tick species *Rhipicephalus sanguineus* and *R. turanicus*. The high percentage of similarity between these two species suggests they recently diverged within the genus *Rhipicephalus*.

T Vectors

The use of T vectors allows the generation of unambiguous sequence data. T vectors are linearized plasmids containing single 5' T overhangs. This allows for the direct and efficient cloning of PCR products. This is possible due to the inherent activity of *Taq* polymerase to add a single dATP nucleotide to the 3' ends of PCR products under standard PCR conditions. Standard primer annealing sites (M13-20, M13 reverse, T3, and T7) on either side of the inserted amplicon in these vectors allow for efficient sequencing.

Sequence Analysis

Once sequence data have been collected and aligned their meaning or results

must be interpreted. The two most common and accepted ways to develop phylogenetic trees from sequence data, are known as distance matrix and maximum parsimony methods. The distance matrix technique considers all possible pair-wise alignments, i.e., it compares the fraction of positions that are different between the two sequences. The differences are then calculated into a distance and recorded in a matrix. Distance matrices often underestimate true distance due to the occurrence of multiple mutational events at a single site. This has been taken into account by algorithms, which increase the calculated distance as a function of increased sequence divergence. The programs FITCH, KITSCH, and NEIGHBOR are for dealing with data which come in the form of distance matrix. The FITCH and NEIGHBOR programs fit a tree which has the branch lengths unconstrained. The KITSCH program assumes that an "evolutionary clock" is valid, according to which the true branch lengths from the root of the tree to each tip are the same.

In contrast, maximum parsimony and maximum likelihood techniques analyze primary sequence data without the initial cluster analysis of matrices of binary distance values. Maximum parsimony treats each position individually, assuming that the most correct phylogenetic tree is the most stringent, in terms of differences between sequences. Tree topologies are determined directly from sequence alignments and are based on finding the shortest possible tree length. For each tree topology, parsimony methods calculate the minimum number of nucleotide changes that are required to explain the observed pattern. The number of changes are summed for each tree topology, and the topology having the smallest total number of changes is considered the best

estimate of phylogeny (Yang, 1996). Maximum parsimony is recommended when the rate of evolution is more or less constant among lineages and the amount of evolution is small (Yang, 1996).

This study employs the use of a 370 base pair sequenced region of the 16S rRNA gene from the mitochondrial genome to determine the phylogenetic relationships between the northern pike, muskellunge, and the silver pike. Sequence data will be interpreted using both distance matrix and maximum parsimony methods.

CHAPTER II

TAXONOMY AND DISTRIBUTION

Taxonomy

Eukaryota: Chordata: Vertebrata: Osteichthyes: Actinopterygii: Teleostei:
Esociformes: Esocidae: *Esox: lucius*.

Distribution

In 1897, E. E. Prince gave the first professional assessment of a fish collected from Sharbot Lake, Ontario that resembled a northern pike but lacked the characteristic coloration (Eddy and Surber, 1943). In 1930 biologists collected several specimens of what local fisherman called silver muskellunge from Lake Belletaine near Nevis, Minnesota while collecting brood stocks of muskellunge for the Nevis Fish Hatchery. This was the first time the silver pike appeared while collecting muskellunge at this sight, and local fisherman reported that it had not appeared in their catches until that year (Eddy and Underhill, 1974). For several years afterwards the silver pike was raised along with true muskellunge and stocked in nearby lakes, thus increasing its range throughout much of Minnesota. Years later a specimen was taken from Detroit Lake, where they were not introduced, suggesting silver pike were not confined to Lake Belletaine. In the summer of 1952, a silver pike was found in Manitoba from Heming Lake. Further collections of silver pike from this lake were intensely recorded for seven years. Lawler (1960) noted that the silver pike was also present in several lakes adjacent to Heming

Lake. In 1959 a silver pike was collected from Beaverlodge Lake in Mackenzie, Canada by a field party of the Arctic Unit of the Fisheries Research Board of Canada. This fish represented the northernmost known record of the silver pike range. The silver pike is not confined to North America. Runnström (1949) reported that G. Svårdson had accounts of a silver colored pike in Sweden on which he had performed breeding experiments. Since these early reports, the silver pike has been reported sporadically throughout much of the circum-polar distribution inhabited by the northern pike (Scott and Crossman, 1973). Scott and Crossman (1973) also indicated that the silver pike has been found only in conjunction with the northern pike (Fig. 3)

CHAPTER III

MATERIALS AND METHODS

Primer Design

The mitochondrial genome coding for the 12S rRNA, 16S rRNA, and tRNA-Val genes of ten species, the fin whale (*Balaenoptera physalus*, Arnason et al., 1991), blue whale (*Balaenoptera musculus*, Arnason and Gullberg, 1993), cow (*Bos taurus*, Anderson et al., 1982), rat (*Rattus norvegicus*, Gadaleta et al., 1989), mouse (*Mus musculus*, Bibb et al., 1981), opossum (*Didelphis virginiana*, Janke et al., 1994), chicken (*Gallus gallus*, Desjardins and Morais, 1990), trout (*Oncorhynchus mykiss*, Zardoya et al., 1995), loach (*Crossostoma lacustre*, Tzeng et al., 1992), and carp (*Cyprinus carpio*, Chang et al., 1994) were obtained from GenBank and aligned using ClustalX (Thompson, 1994; Appendix A). Examination of the 16S rRNA gene alignments revealed a number of hypervariable regions flanked by highly conserved sequences. This orientation is important because it allows for the construction of PCR primers to anneal to the adjacent conserved areas and amplify through hypervariable regions.

Once the 16S rRNA gene sequence was chosen, 13 sequences from different genera in the family Cyprinidae were aligned using Clustal X (Thompson et al., 1994) along with one sequence from the northern pike (Appendix B). Sequences were then analyzed to find the region that provided the most variability and still provided enough conservation for primer annealing. Each base pair (bp) location along the 16S rRNA gene (1722 bp long) was compared among all species and assigned a number. If all bases

were the same across all species the row was assigned a zero. If that location contained only two different bases (ex. A and G) that row was assigned a one, if there was three bases (ex. A, G, and T) the row was assigned a two, all four bases scored a three (Fig. 4). If a site within the sequence of the 16S rRNA gene is capable of having any one of the four bases and still remains functional, the site is considered highly variable. Next, all sites that were completely conserved across all species for at least 18 consecutive bp were noted as possible primer locations. Then, the variability between primer sites was calculated. Two sites were found that contained highly variable regions flanked by good primer locations. The two sites were separated by a 23 bp conserved area. It was then decided to combine the two areas to provide a single 338 bp target region. This area had a 30 bp-conserved region on the 5' end and a 45 bp conserved region on the 3' end.

All possible permutations of the upstream and downstream primer sequences were analyzed using the Lazergene Primer Select program (DNASTar, Madison, WI). Of 20 potential primers, one forward primer and one reverse primer were found to have acceptably low self-annealing properties. These primer sequences were used to amplify the DNA from fish tissues. The forward primer was 5'-GGT AGC GCA ATC ACT TGT CT-3'. The reverse primer was 5'-TAT CCC TAG GGT AAC TTG GT-3'. Primers were synthesized at the Marshall University DNA Core Facility. Their designation and properties are listed in Tables 1 and 2. Sequences that would be amplified by the designed primers for members of the family Cyprinidae and the northern pike were then taken from GenBank and aligned using Clustal X (Thompson et al., 1994) to determine efficacy of this DNA region for phylogenetic reconstruction (Fig. 5).

DNA Isolation

DNA samples were prepared for the fish listed in Table 3. All tools used in preparation of fish scales for DNA amplification were first placed in 1 N HCl, rinsed with sterile water, placed in 10 N NaOH, rinsed with sterile water and soaked in 95 percent ethanol until used. All procedures were carried out under a laminar flow hood. Fresh fish specimens were prepared by first swabbing the area where a scale would be removed with 95 percent ethanol to minimize surface contamination. When the area was completely dry, a single scale was removed with a pair of flame-sterilized tweezers and placed in a 1.5 ml sterile centrifuge tube with 100 μ l of 5 percent chelex and 1 μ l of Proteinase K. Tubes were then placed in a dry bath at 56°C for at least 1.5 hours, preferably overnight. Following incubation at 56°C, tubes were boiled for 8 min then immediately centrifuged at 16,200 \times g for 5 min and stored at -20°C until used. Scales were unavailable for silver pikes three and four, northern pikes two and three, and muskellunge four and five. In this case, muscle tissue was used and prepared in a similar manner. A sterile wooden applicator was used to obtain a very small amount of tissue from sample. By twisting the wooden applicator tip in the tissue sample, a small amount of tissue (~10 mg) could be obtained. Subsequent procedures remained the same for both scales and tissue samples.

Amplification

Amplification mixtures were prepared using a PCR core kit as specified by the

manufacturer (Boehringer Mannheim, Germany). Taq DNA polymerase was not added until tubes were placed in the thermal cycler and heated to 94°C for a minimum of 1 min. Cycle one was 5 min at 94°C and allowed ample time for addition of Taq polymerase. Cycle two consisted of 1 min at 94°C for denaturation, 1 min at 37°C to allow primers to anneal, and 1 min at 72°C for primer extension and was repeated 30 times. Cycle three used the same denaturation and primer annealing times and temperatures but had a 6 min extension time. Upon completion of amplification tubes were stored at -20°C. Agarose gel electrophoresis was used to confirm the presence of an amplicon. To aid in determination of band size, one lane contained 0.5 µg/µl of a 1 Kb DNA ladder (Promega, Madison, WI) in 4 µl water with 1 µl of 10X stop buffer (See Table 1). Gels were run with a FisherBiotech Electrophoresis Systems FB 105 voltage regulator and FB MSU-1 Small Horizontal Gel System. Gels were then viewed using a Spectroline® Transilluminator (Model TR-302, 302nm ultraviolet) and photographed using a Fisher Scientific Photo-Documentation Camera (FB-PDC-34), a Tiffen® 40.5 mm deep yellow filter with Polaroid type 667, 3000 iso black and white film.

Cloning

Amplification products were cloned into either the pCR®2.1 (Invitrogen, Carlsbad, CA) vector or the pT-NOT vector (Max-Planck-Institut Für Immunbiologie, Freiburg, Germany; Table 3) using a protocol modified from the TA Cloning Kit® (Invitrogen, Carlsbad, CA). Fresh amplification product was added to 12.5 ng/µl of pCR®2.1 vector in a weight ratio of 3:1 with 1 µl of 10X ligation buffer. The volume of

the mixture was then adjusted to 9 μl with sterile dd water and 1 μl of T4 DNA Ligase was added. Ligation reactions were incubated overnight at 14°C and stored at -20°C.

Plasmids were transformed using a modified protocol included in the TA Cloning Kit®. Competent cells of INV α F' strain designation, 0.5 M β -mercaptoethanol (β -ME), and the ligation mixture were thawed on ice. After thawing, 2 μl of 0.5 M β -ME, and 5 μl of ligation reaction mixture were added to 25 μl of competent cells and mixed gently by stirring with the pipette tip. Competent cells were then incubated on ice for 30 min before being heat shocked at 42°C for 30 sec. Trypticase soy broth (TSB) (1 ml) was added and competent cells were incubated for 1 hr at 37°C. The cells were then spread on Luria Bertani (LB) agar plates containing 50 $\mu\text{g/ml}$ of ampicillin and 40 μl of X-Gal (40 mg/ml) and incubated overnight at 37°C.

The pCR®2.1 vector contains the *lacZ α* gene. This allows for blue-white screening. Bacterial cells that contain an uninterrupted *lacZ α* gene produce colonies that are blue in color. If bacteria take up plasmids that have a *lacZ α* gene that has been interrupted by insertion of PCR amplicon, the colonies are white. Nine white colonies and one blue colony were transferred from LB plates to 5 ml TSB with 50 $\mu\text{g/ml}$ of ampicillin and incubated overnight at 37°C in a waterbath shaker. The cell suspension (1.5 ml) was pelleted in a 1.5 ml microfuge tube by centrifuging 2 min at 10,000 \times g. The pellet was then resuspended in 200 μl of cell resuspension buffer (Table 1) and lysed with 200 μl of NaOH-SDS for 3 min. The suspension was then neutralized with 200 μl of 3M Na-acetate and incubated at room temperature for 4 min before centrifuging 10 min at 10,000 \times g. The supernatant was collected, 1 ml of 95 percent ethanol was added

and the sample was centrifuged at $16,000 \times g$ for 30 min at 4°C . The supernatant was discarded and the pellet was then rinsed three times with 70 percent ethanol and one time with 95 percent ethanol. The pellet was dried and resuspended in $400 \mu\text{l}$ of water and $200 \mu\text{l}$ of 7.5 M Ammonium Acetate. This solution was incubated at room temperature for 10 min then centrifuged for 15 min at $16,000 \times g$. The supernatant was collected, 1 ml of 95 percent ethanol was added and the sample was centrifuged at $16,000 \times g$ for 30 min at 4°C . The supernatant was discarded and the pellet was then rinsed three times with 70 percent ethanol and one time with 95 percent ethanol. The pellet was dried and hydrated in $50 \mu\text{l}$ of water. Agarose gel electrophoresis was used to confirm the presence of plasmids containing an amplicon.

Samples containing plasmids with inserts were desalted using the Wizard[®] *Plus* Minipreps DNA Purification System (Promega, Madison, WI). Purified plasmids were sequenced at the Marshall University DNA Core Facility (Huntington, WV). Both strands were determined for all sequences to assure accuracy. Sequences were aligned using Clustal X (Thompson et al., 1994). Phylogenetic relationships were analyzed using distance matrix and maximum parsimony methods.

All phylogenetic analyses were performed using programs of the PHYLIP package (Felsenstein, 1989). Sequence alignments were bootstrapped using the SEQBOOT program. One hundred bootstrapped data sets were used to assess reproducibility. The resulting output file was analyzed first using DNAPAR. DNADIST was used to generate distance matrices for the NEIGHBOR, FITCH, and KITSCH treeing programs. The distance matrix file was then run in the NEIGHBOR, FITCH and the

KITSCH programs using global rearrangement, randomized input, and 100 multiple data sets with all other settings remaining as default. The resulting files were then run in CONSENSUS to determine the consensus tree topology and trees were visualized using Treeview (Page, 1996; Figs. 7,8,9, and 10). For all trees the four muskellunge sequences were set as outgroup taxa.

Specimen Collection

One northern pike sample was collected from the Great Miami River near Dayton, Ohio and three northern pike specimens were collected from Ramsey Lake in Ontario, Canada. Two northern pike were collected from the Marten River, also in Ontario, which also provided one silver pike. Muscle tissue from two silver pike from Young Lake, Minnesota was obtained from Dr. Loren Miller from the University of Minnesota. Two silver pike specimens were obtained from Mr. Ed Thelend at the Spirit Lake Fish Hatchery, Iowa. Three muskellunge samples were obtained from the Morehead Fish Hatchery, Kentucky and two were collected from the Green River near Cave City, Kentucky (Fig. 6, Table 3).

CHAPTER IV

RESULTS

Alignment of sequences within the Cyprinidae family amplified with designed primers were capable of distinguishing between genera in the same family (Fig. 5). High bootstrap values were found between specimens of the same species. The ability of the primers to distinguish between genera in the same family confirms the efficacy of the primers.

A total of 370 bp were sequenced from each of the 12 specimens. There was one variation observed among the five silver pike, a transversion at site 159. Three sites were not conserved between all northern pike, transitions at sites 143, 207, and 326. There was also a single transition at site 197 found among the muskellunge. There were a total of 32 variable sites found among all species consisting of 21 transitions and 11 transversions. This high ratio of transition mutations to transversion mutations was also found in a study on mitochondrial 16S rDNA sequence variation of brown trout (Patarnello et al., 1994). There were 28 variable sites between the pikes and the muskellunge with a mean sequence difference of 7.6 percent. Nucleotide divergence between the muskellunge and the pikes was calculated by dividing the number of substitutions by the total number of nucleotides examined (Patarnello et al., 1994). Neither the maximum parsimony or distance matrix programs separated the northern pike from the silver pike (Figs. 7,8,9, and 10). Low bootstrap values were found among the silver pike and the northern pike in all trees. The grouping of the silver pike with

northern pike was not consistent through all trees and varied from one tree to another.

No two silver pike or two northern pike were grouped together consistently.

CHAPTER V

DISCUSSION

This study uses mtDNA sequences in an attempt to classify the silver pike. In the eighteenth century, Carolus Linnaeus developed an order for classifying organisms which he termed taxonomy. Linnaeus described species in terms of their morphology. Physiology, biochemistry, life history, and behavior were soon accepted ways to classify organisms. The advent of DNA sequencing for use as a molecular chronometer gives biologist a powerful tool for classifying organisms. The problem comes when deciding what molecules are best or exactly how much divergence is necessary to reclassify an organism. As sequence databases grow and more organisms are examined this technique will grow in acceptance and the parameters will become more defined.

The first objective in primer design was to find a region within the mitochondrial genome that would provide sufficient variability for phylogenetic analysis and still be flanked by highly conserved areas for primer adhesion during the polymerase chain reaction (PCR). Although it was originally hoped to target intergenic regions for sequence determination and analysis, these regions proved to be too small to provide sufficient discriminatory information. The 16S rRNA gene was chosen for analysis after comparison of the 12S and 16S rRNA and the tRNA valine genes. The 16S rRNA gene provided functional constancy and genetic stability to assure a relatively good clocklike behavior, highly variable regions which mutate fast enough to provide information on

closely related species (Mangold et al., 1997), and a comprehensive sequence database describing a wide spectrum of phylogenetically diverse organisms.

Phylogenetic trees constructed from sequence data did not differentiate the silver pike from the northern pike. In all instances the four muskellunge samples were grouped together separate from all other samples (Figs. 7,8,9, and 10). The silver pike and the northern pike sequences were never grouped separate from each other but always intermingled. Low bootstrap numbers between silver pike and northern pike indicate the very low sequence variation between the two. By comparing sequences once they were aligned it was found that none of the differences between the northern pike and the silver pike were conserved throughout all the silver pike or all of the northern pike as was found in the muskellunge. Although there are significant differences between the northern pike and the silver pike in morphological characteristics, these differences are not apparent within the mitochondrial genome. Because the sequenced area was able to differentiate between the northern pike and the muskellunge, it contained enough variability to distinguish between two species within the same genus. This area was unable to differentiate between the northern pike and silver pike. Since none of the differences between the silver pike and northern pike occurred throughout all silver pike, the differences are not the result of divergence from the northern pike as a separate species, but instead are likely to be variations within a single population or individual. Use of specimens from several geographical locations removed the chance that variations among species are do to population variance and not speciation. The geographical distribution of specimens is a possible explanation for intraspecific sequence differences

at single sites. The phylogenetic trees developed in this study confirmed previous research based on morphological characteristics.

The minor sequence differences between the silver pike and the northern pike contrasts with the high degree of phenotypic variation and the inability of these fish to produce fertile hybrids. Similar situations have been reported in several species of fish. Despite at least three phenotypes with very different ecologies, the arctic char (*Salvelinus alpinus*) has very low genetic differentiation between these phenotypes (Vrijenhoek et al., 1987; Snorrason et al., 1988). In contrast, despite their similar morphological characteristics, brown trout from the River Brenta, Ireland, compared to brown trout from the Stura and Ripa Stream, Ireland, display an appreciable degree of genetic differentiation (Patarnello et al., 1994). Cichlid fishes have also displayed differing rates of morphological and molecular evolution (Meyer et al., 1990; Sturmbauer and Meyer, 1992). These studies, present included, further support the idea that phenotypic characteristics evolve at different rates among different species.

The silver or blue color phase of the silver pike may be a similar genetic variation found in other types of fish. In two species of fish, the northern pike and the walleye (*Stizostedion vitreum*) there is a blue color phase which is devoid of many dominant color traits. In addition to this color difference there appears to be other morphological differences, mainly the diameter of eye and overall size. From the late 1800's to the mid 1900's, a species known as the blue pike (*Stizostedion glaucum*) was extensively harvested from the great lakes. The blue pike was similar to the walleye in appearance except its characteristic blue coloration, larger eyes and smaller overall size, the same

characteristics found in the silver color phase of the northern pike. In 1926 Dr. Carl Hubbs classified the blue pike as a separate species of the walleye (Manns and Quinn, 1999). The blue pike, prized for its soft flesh and small size, was quickly fished to extinction. It was placed on the Endangered Species List in 1971 though it was believed to be extinct. As is the case with the silver pike, the blue pike has received little research. Several fish have been collected that appear to be blue pike resulting in research being done to compare archived samples to recently obtained samples. If the blue pike was fished to extinction, it is possible that the color mutation once again occurred, starting another population of the true breeding variant. If the mutation occurs frequently enough to cause repopulation of blue pike in the Great Lakes, it would explain the large range of the silver pike. Recently, the blue pike was reclassified as a subspecies of the walleye (*Stizostedion vitreum glaucum*).

The word species means "kind" or "appearance" in Latin. According to Campbell (1996) a biological species is defined by its reproductive isolation from other species in a natural environment. Mayr (1942) proposed that species are groups of actually or potentially interbreeding natural populations, which are reproductively isolated from other such groups. Although the silver pikes very little divergence from the northern pike in the present study, the inability for the two morphs to hybridize adheres to the basic definition of a species.

Although the silver pike is not considered a subspecies, nor does this study suggest that, it does display several characteristics consistent to a subspecies. The definition of a subspecies according to Campbell (1996) is a population or group of

populations that live in one area and have minor differences from conspecific populations found elsewhere in the species geographical range. Mayr (1953) views a subspecies as being geographically defined aggregates of local populations which differ taxonomically from other subdivisions of a species. Although geographical ranges do not separate the silver pike and northern pikes, they are separated by life history traits (i.e., possible spawning times or niche utilization).

Though no taxonomic importance would be achieved, to classify the silver pike as a sibling species of the northern pike would provide a classification status of the silver pike. A Sibling species according to Mayr (1942) is another class of difficulties caused by pairs of even larger groups of sympatric, related species which are so similar that they are considered as belonging to one species until a more satisfactory analysis clears up the mistake. Mayr (1942) also states that there is no reason to believe that sibling species evolve in a manner that is in the least different from that of other species. Biologists dedicate their lives to defining what amount of evolution constitutes a species, to try to sum it up in a single chapter would be foolish.

The present study concurs with past studies and places the silver pike as a color variant of the northern pike. Nonetheless, fisheries biologists are given a chance to study the exact point at which a new species is evolving from a current species. It is as though we are witnessing the silver pike in its evolutionary descent from the northern pike. Species that are destined not to survive are common, evolving species that die out unstudied because of how they are classified should not be.

CHAPTER VI

SUMMARY AND CONCLUSION

Summary

The silver pike is a member of the family Esocidae and most closely resembles the northern pike (*Esox lucius*) in morphological characteristics. The most distinct difference between the silver pike and the northern pike is color, though other more subtle differences do occur. Breeding experiments have been reported on the silver pike, though not well documented. The silver pike always breeds true when crossed with other silver pike. When the silver pike is crossed with the northern pike, the offspring that survive are unable to produce a F2 generation, suggesting their sterility. The F1 generation of silver pike, northern pike hybrids are mottled like a crappie, a color morph of that pike has not been documented in nature. This suggest that the silver pike and northern pike do not hybridize in nature. The silver pike has only been documented in sympatric populations with northern pike, where it is found it often accounts for less than one percent of the total pike population.

Molecular chronometers provide taxonomists with a powerful tool to complement phenotypic techniques. The rapid mutation rate of mitochondrial DNA provides a reliable tool in estimating the degree of divergence among closely related species. The 16S ribosomal RNA gene has been termed the ultimate molecular chronometer due to its large size, genetic stability, highly variable regions, and large sequence database.

Conclusion

This study employed the use of a 370 base pair sequenced region of the 16S rRNA gene from the mitochondrial genome to determine the phylogenetic relationships between the northern pike, muskellunge, and the silver pike. Sequence data was interpreted using both distance matrix and maximum parsimony methods. The sequenced area was able to delineate between the northern pike and the muskellunge though no differences were found between the northern pike and the silver pike sequences. Phylogenetic trees constructed from sequence data did not differentiate the silver pike from the northern pike. The silver pike and northern pike sequences were never grouped separate from each other but always intermingled. Low bootstrap numbers between the silver pike and northern pike indicate the very low sequence variation between the two. Although there are significant morphological differences between the silver pike and the northern pike in morphological characteristics, these differences are not apparent in the 16S rRNA gene of the mitochondrial genome. The present study concurs with past studies and places the silver pike as a color variant of the northern pike.

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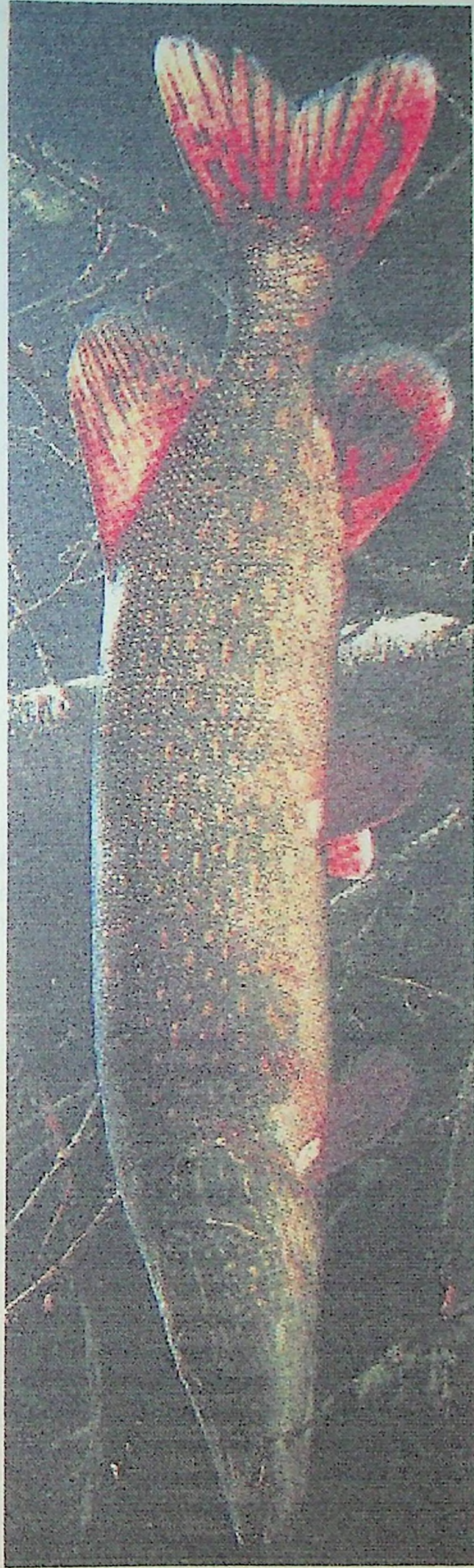
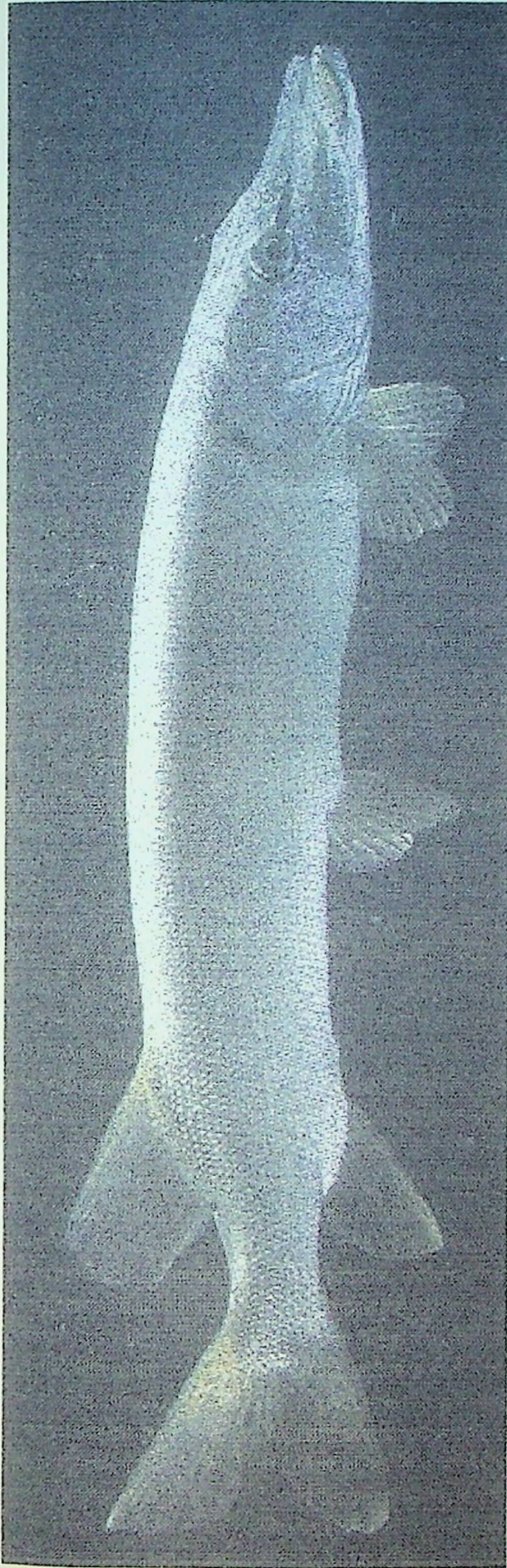


Figure 1. Silver pike (top) in comparison of northern pike (bottom) showing distinct color difference (Stemberg, 1992).

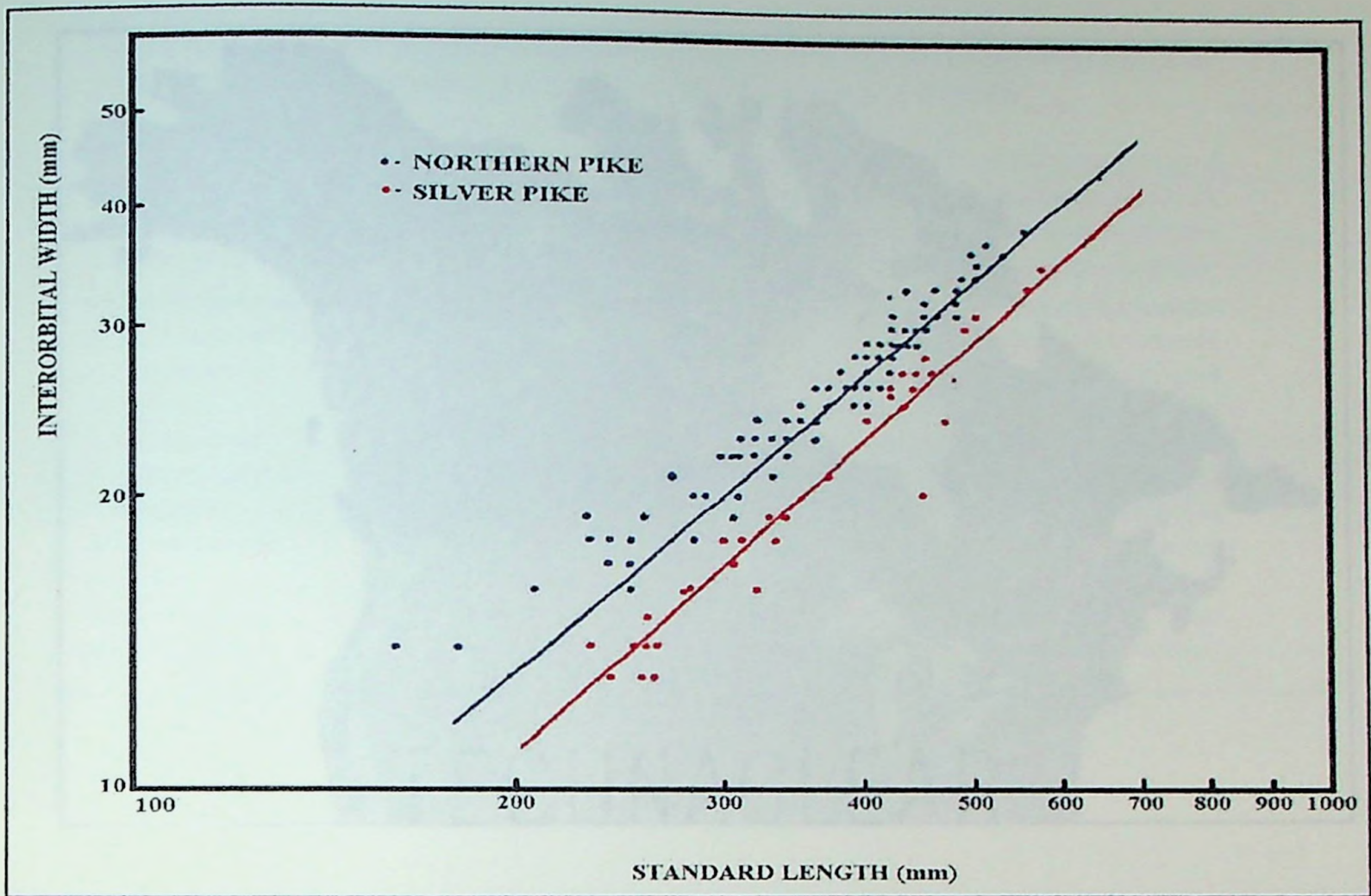


Figure 2. Relation between interorbital width and standard length of normal and silver pike from Heming Lake, Manitoba (Lawler, 1960)

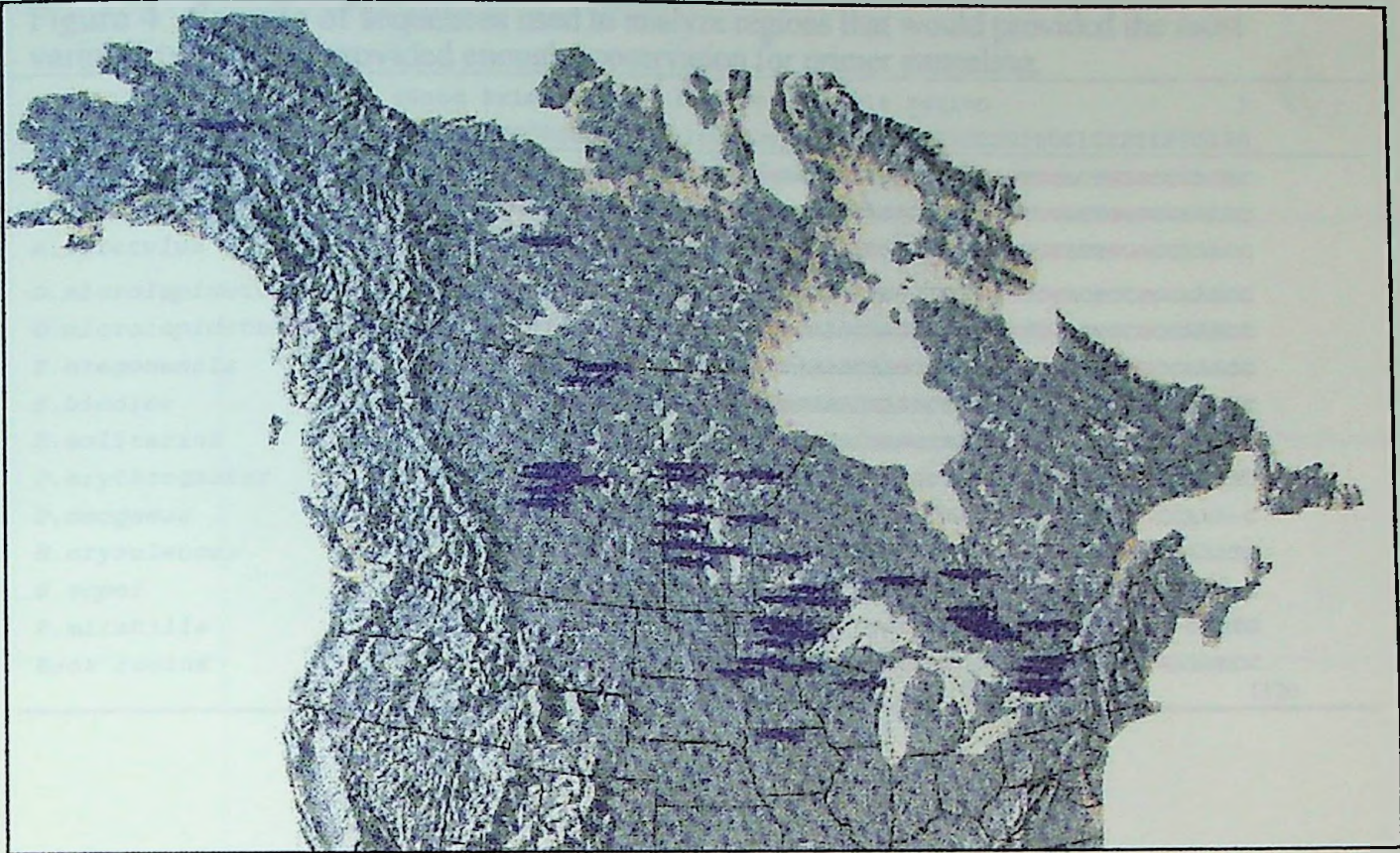


Figure 3. Distribution of silver pike (Bernardo, 1998)

Figure 4. Sample of sequences used to analyze regions that would provided the most variability and still provided enough conservation for primer annealing.

Assigned Number	(Good Primer Area)	(Hypervariable region)					
	0000000000000000000000001101000020023110001002201000102201220120						
<i>N. therinoides</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCGAGAC						
<i>P. gracilis</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCAAAAC						
<i>R. atratulus</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCAAAAC						
<i>O. microlepidotus1</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCAAAAC						
<i>O. microlepidotus2</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCAAAAC						
<i>P. oregonensis</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCAAAAC						
<i>S. bicolor</i>	TACCTTCGGTTGGGGCGACCGGGGAGGAAAAACAAGCCTCCTAGTGGACTGGGCCAAAAC						
<i>R. solitarius</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCAAAAC						
<i>P. erythrogaster</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCAAAAC						
<i>P. neogaeus</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCAAAAC						
<i>N. crysoleucas</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCAAAAC						
<i>S. copei</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCAAAAC						
<i>P. mirabilis</i>	TACCTTCGGTTGGGGCGACCGCGGAGGAAAAACAAGCCTCCGAGTGGACTGGGCCAAAAC						
<i>Esox lucius</i>	TATCTTCGGTTGGGGCGACCGGGGGAAAAACAAGCCCCACGAGGATTAAGGAAAACC						
	1261	1270	1280	1290	1300	1310	1320

Figure 5. Phylogenetic tree of the family Cyprinidae showing positions of sequences obtained with the proposed markers.

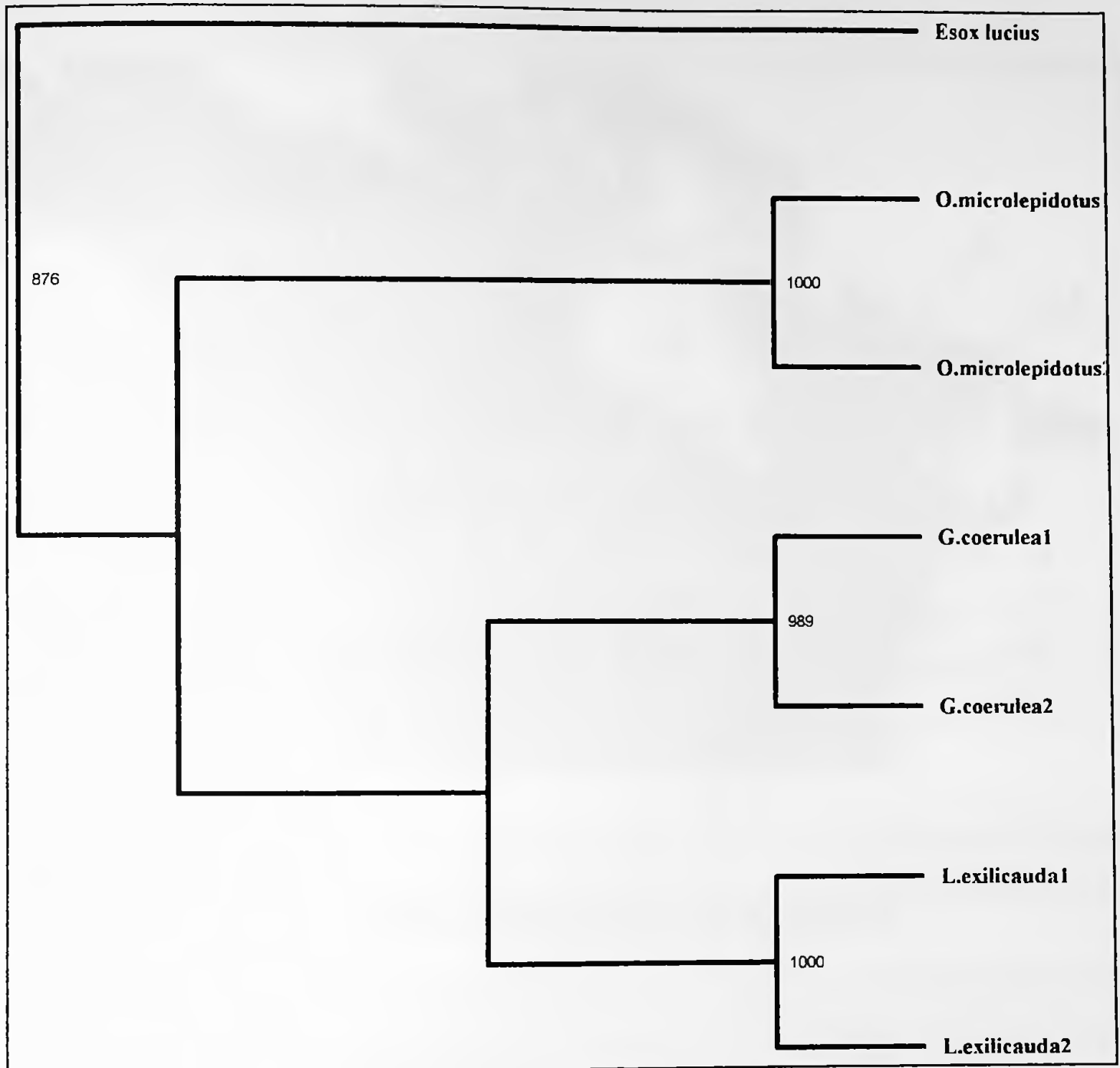


Figure 5. Phylogenetic tree of the family Cyprinidae showing accuracy of clustering obtained with the proposed amplicon.

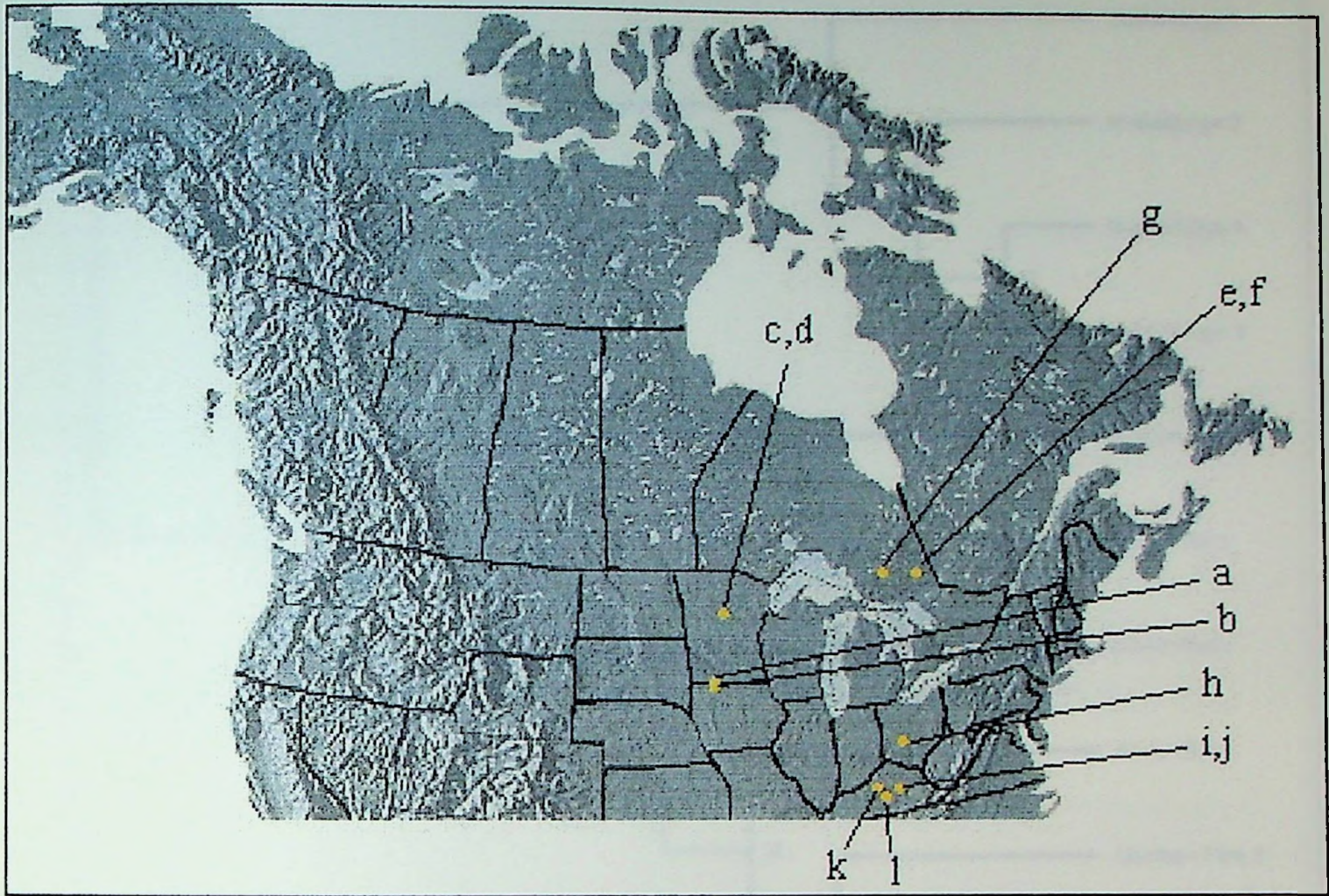


Figure 6. Sample collection locations. Letters represent specimens (Table 3).

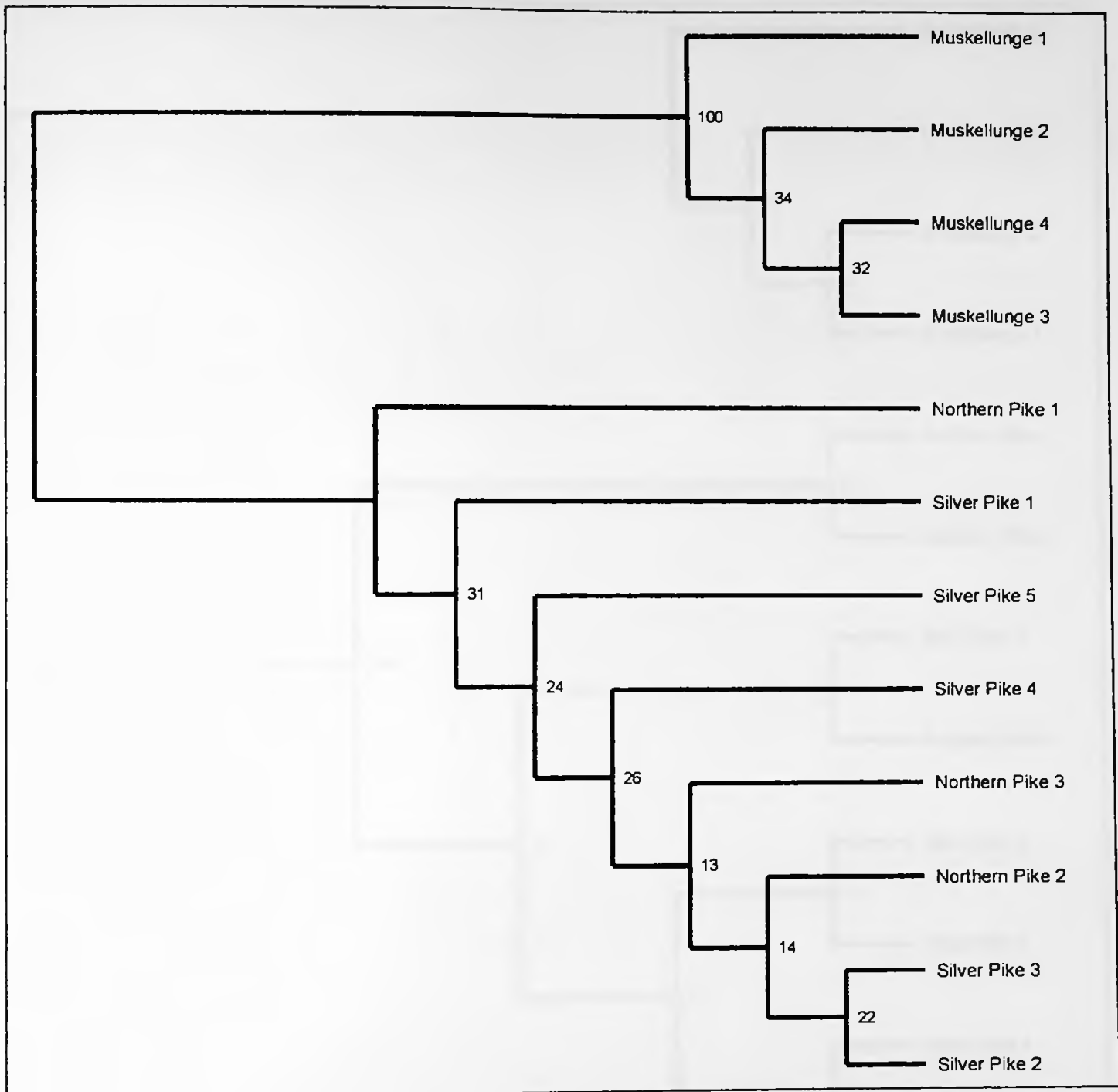


Figure 7. Neighbor joining phylogenetic tree of silver pike, northern pike, and muskellunge using sequenced data. Internal labels are bootstrap values after 100 replications. Muskellunge were set as outgroup.

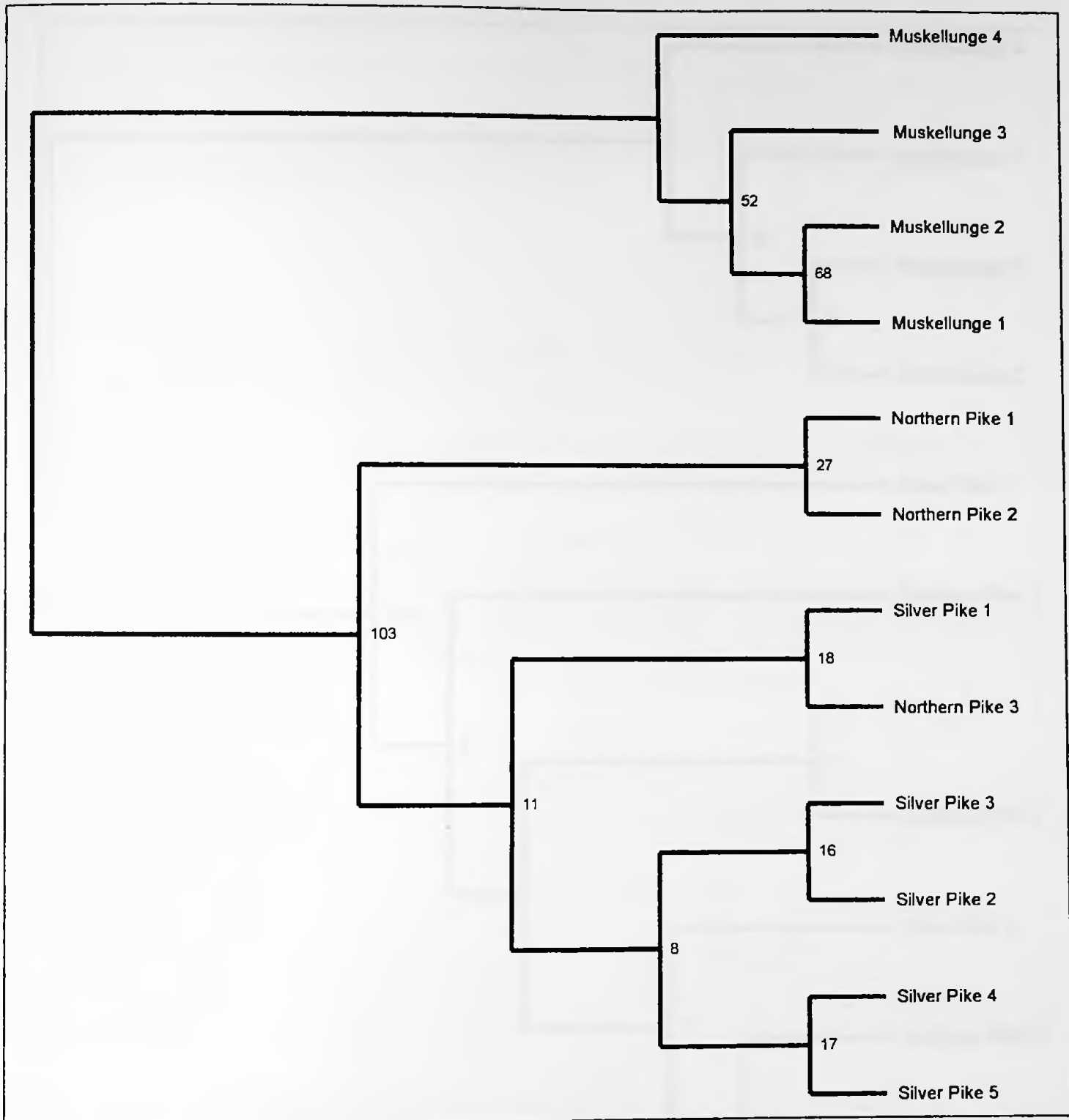


Figure 8: Maximum parsimony phylogenetic tree of silver pike, northern pike, and muskellunge using sequenced data. Internal labels are bootstrap values after 100 replications. Muskellunge were set as outgroup.

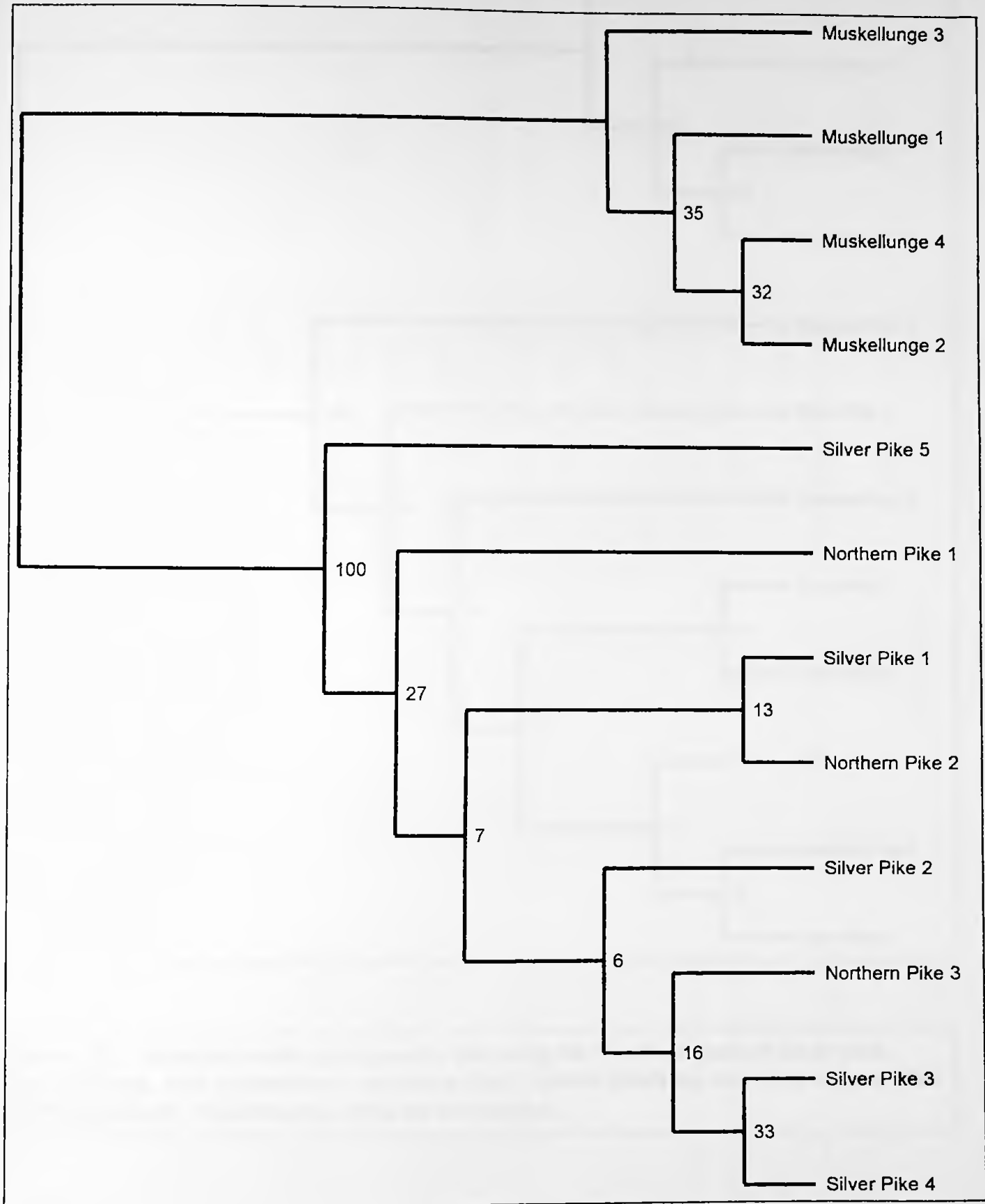


Figure 9. Distance matrix phylogenetic tree using the Fitch program of silver pike, northern pike, and muskellunge sequence data. Internal labels are bootstrap values after 100 replications. Muskellunge were set as outgroup.

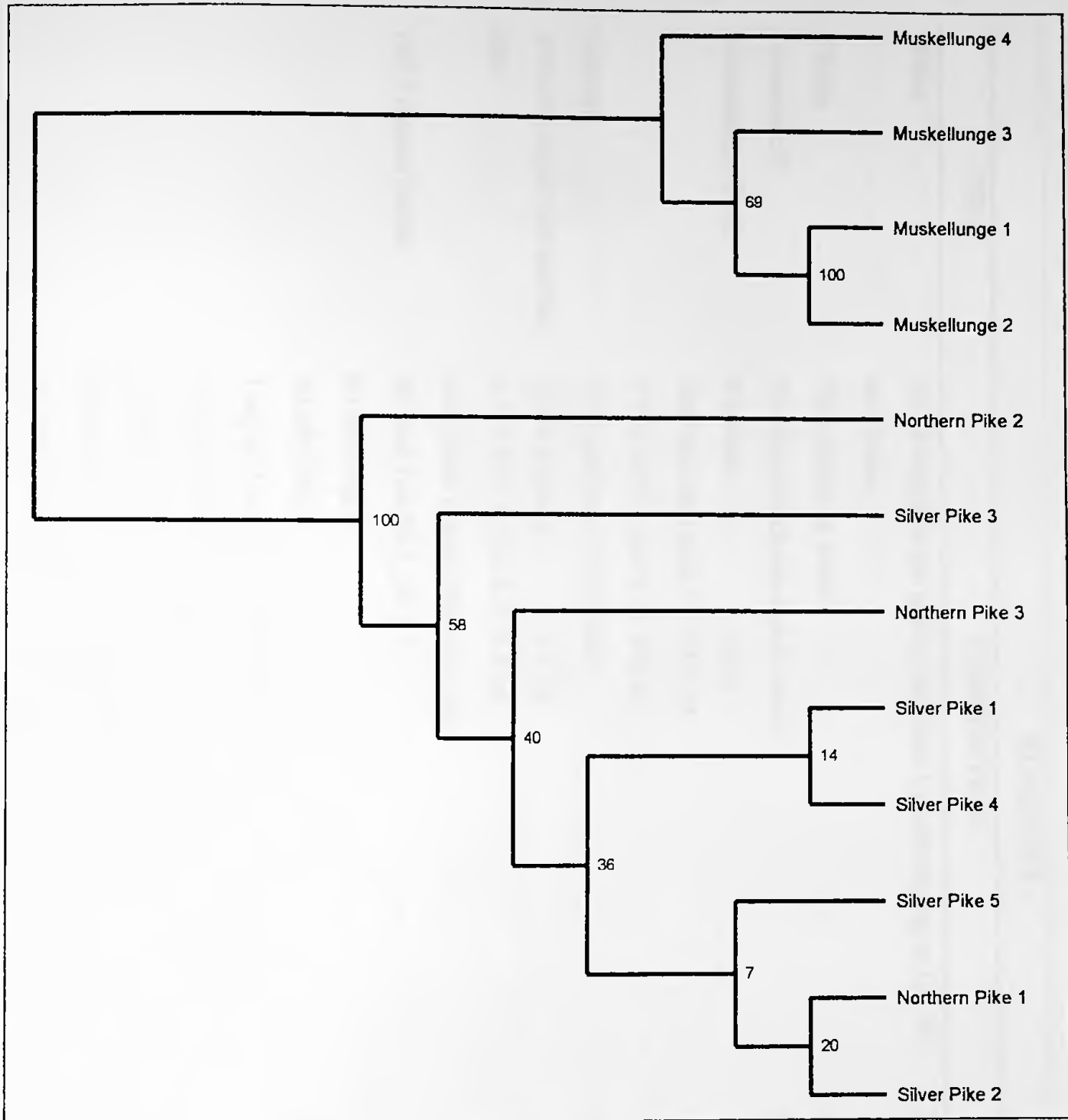


Figure 10. Distance matrix phylogenetic tree using the Kitsch program of silver pike, northern pike, and muskellunge sequence data. Internal labels are bootstrap values after 100 replications. Muskellunge were set as outgroup.

Table 1. Materials used in laboratory experiments.

REAGENTS		
ITEM	COMPOSITION	COMPANY
Water	16-18 megohm-cm purity, sterilized by autoclaving at 125 °C for 20 min	Barnsted E-Pure, Dubuque, IA
Chelex	5% Chelex in water	Carolina Biological Supply Company, NC
Proteinase K	20 mg/ml stock solution in water	Boehringer Mannheim, Germany
Tris-Acetate (TAE)	Tris base 242 g Glacial acetic acid 57.1 ml 0.5 M EDTA (pH 8.0) 100 ml	
Agarose	1% Agarose in TAE buffer	BIO-RAD, Hercules, CA
10X stop buffer for agarose gels	100% glycerol 0.7 ml 0.5 M EDTA (pH 8.0) 0.3 ml Add Brom Phenol Blue for color	
10X Ligation Buffer	60 mM Tris-HCl, pH 7.5 60 mM MgCl ₂ 50 mM NaCl 1 mg/ml bovine serum albumin 70 mM β-Mercaptoethanol 1 mM ATP 20 mM dithiothreitol 10 mM spermidine	

Table 1. Continued.

REAGENTS		COMPANY	
ITEM	COMPOSITION		
T4 DNA Ligase	4.0 Weiss units/ μ l	Invitrogen, Carlsbad, CA	
Cell resuspension buffer	50 mM Tris, (pH 7.5)	Promega, Madison, WI	
	10 mM EDTA		
	100 mg/ml Rnase		
	0.2 M NaOH		
NaOH-SDS	1% Sodium Dodecyl Sulfate (SDS)		

COMPETENT CELLS

Competent cells	<i>E. coli</i> , strain INV α F' (F <i>endA1 recA1 hsdR17</i> (r_k^- , m_k^+) <i>srpE44</i> <i>thi-1 gyrA96 relA1 ϕ80(lacZ)M15</i> (<i>lacZY A-argF</i>)U 169 λ)	Invitrogen, Carlsbad, CA
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Table 2. Oligonucleotide Primers. Oligonucleotide analysis data calculated using OLIGO 4.0 primer analysis software (NBI).

NAME	SEQUENCE	OLIGONUCLEOTIDE ANALYSIS
SF1-68-1 (Forward)	GGT AGC GCA ATC ACT TGT CT	<p>Td = 62.0° (nearest neighbor method)</p> <p>Tm = 68.2° (%GC method)</p> <p>Tm = 60.0° (2*(A+T)+4*(G+C))</p> <p>nmol/OD = 5.35 (nearest neighbor extr. coeff.)</p> <p>µg/OD = 33.1</p> <p>Composition =</p> <p>A + T 10 (50%)</p> <p>C + G 10 (50%)</p>
SF1-68-2 (Reverse)	TAT CCC TAG GGT AAC TTG GT	<p>Td = 57.5° (nearest neighbor method)</p> <p>Tm = 66.2° (%GC method)</p> <p>Tm = 58.0° (2*(A+T)+4*(G+C))</p> <p>nmol/OD = 5.24 (nearest neighbor extr. coeff.)</p> <p>µg/OD = 32.5</p> <p>Composition =</p> <p>A + T 11 (55%)</p> <p>C + G 9 (45%)</p>

Table 3. Locations of specimen collection.

Specimen	Location	Latitude	Longitude
Silver Pike 1 *	Spirit Lake, IA (a)	43.28 N	95.06 W
Silver Pike 2	Okoboji Lake, IA (b)	43.29 N	95.07 W
Silver Pike 3	Young Lake, MN (c)	46.34 N	94.08 W
Silver Pike 4	Young Lake, MN (d)	46.34 N	94.08 W
Silver Pike 5	Marten River, ON, Canada (e)	46.22 N	79.25 W
Northern Pike 1	Marten River, ON, Canada (f)	46.22 N	79.25 W
Northern Pike 2	Lake Ramsey, ON, Canada (g)	46.37 N	80.48 W
Northern Pike 3	Great Miami River, OH (h)	40.07 N	84.14 W
Muskellunge 1	Morehead Fish Hatchery, KY (i)	38.22 N	83.57 W
Muskellunge 2	Morehead Fish Hatchery, KY (j)	38.22 N	83.57 W
Muskellunge 3	Green River, KY (k)	37.11 N	86.07 W
Muskellunge 4	Green River, KY (l)	37.16 N	85.37 W

Letters in parentheses indicate specimen sampling locations shown in Fig. 1. * Indicates specimens cloned into the PT-NOT vector, all others were cloned into the pCR[®]2.1 vector.

Fin Whale ACTACCACAAATCAA----TCAAATAAAACATTTACCATCCCTTCAA---AGTATAGGAG
 Blue Whale ACTACCATAGACCAA----TCAAATAAAACATTCACCAACCTTCTAA---AGTATAGGAG
 Cow ACAACCAAGATAGAA----TAAAACAAAACATTTAATCCCAATTTAA---AGTATAGGAG
 Rat ACTAA-ACCCCCACA----TAAACTAAAACATTTAACTCA----AAA---AGTATTGGAG
 Mouse ATTAT-ACTATTATA----TAAATCAAACATTTATCCTAC--TAAA---AGTATTGGAG
 Loach AATAAAACAACAAAA-CCTTAAAAAATAAACTAAACCATTTTTTCCACCTTAGTACGGGCG
 Carp AATAAAACAATAAGCCTGACACCAAAAACATAACCATTTTTTTTACCTGAGTATGGGAG
 Trout CATACCCCAATAAAACTTAGAATTAAGTCAACAAACCATTTTTTCCACCTTAGTAGGGGCG
 Chicken AAAAACTTACCTCCCCCTCTTAACCAAAACATTATAAATTGTCCC-----AGTATAGGCG
 Opossum ACTAA--CTACTTTT----TCAATTTAAACCATTTTAATTATCCT-----AGTATAGGTG

1141 1150 1160 1170 1180 1190 1200
 ***** ** *

Fin Whale ATAGAAATT--TAAATATCAGTGGCGCTATAGA----GATAGTACCGTAAGG-AAA-GAT
 Blue Whale ATAGAAATT--TAAACATCAGTGGCGCTATAGA----AATAGTACCGTAAGGGAAA-GAT
 Cow ATAGAAATC--TAAGTAC----GGCGCTATAGA----GAAAGTACCGCAAGGGGAAC-GAT
 Rat AAAGAAATT--TACTTACCA--AGAGCTATAGA----GAAAGTACCGCAAGGGGAAATGAT
 Mouse AAAGAAATTCGTACAT-CTA--GGAGCTATAGA----ACTAGTACCGCAAGGGGAAA-GAT
 Loach ACGGAAAAGGATCCGA----TTAAGCGATAGA----AAAAGTACCGCAAGGGGAAA-GCT
 Carp ACAGAAAAGGTTCC-A-----CAAAGCGATAGA----AATAGTACCGCAAGGGGAAA-GCT
 Trout ACCGAAAAGGAGATAA----TTGAGCAACAGA----AAAAGTACCGCAAGGGGAA-GCT
 Chicken ATAGAAAAGACTACCC-----CGGCGCAATAGAGGCTAACTGTACCGCAAGGGGAAA-GAT
 Opossum ATAGAAAAG-ATATAATA----GGAGCTATAGT--TTATAGTACCGCAAGGGGAAA-AAT
 * **** * * * * * * * * * * * * * * *

1201 1210 1220 1230 1240 1250 1260

Fin Whale GAAAGAAAAAC-CT-----AAAAGTAATAAAAAG-CAAAGCTTACCACTTGTACCT
 Blue Whale GAAAGAAAAAC-CC-----AAAAGTAATAAAAAG-CAAAGCTTACCCCTTGTACCT
 Cow GAAAGAAAAAAACT-----AAAAGTATAAAAAG-CAAAGATTACCCCTTGTACCT
 Rat GAAAGACTAAT--T-----TAAAGTAAAAACAAGACAAAGATTAAACCT-GTACCT
 Mouse GAAAGACTAAT--T-----AAAAGTAAGAACAAG-CAAAGATTAAACCTTGTACCT
 Loach GAAAAAGAAATGAAACAACCCATATAAGCACCACAAAG-CAGAGACACAACCTCGTACCT
 Carp GAAAGAGAAATGAAATAACCCATATAAGCACTAAAAG-CAAAGATTAAACCTCGTACCT
 Trout GAAAGAGAATTGAAATAACCCATTTAAGCCTAGAGAAG-CAGAGATTAAATCTCGTACCT
 Chicken GAAATAGCAATGAAA--ACC--ATAAGCAAAAACAG-CAAAGACCAACCCCTTGTACCT
 Opossum GAAAGATAAAT-----TATAGTAATTTAAAAG-CAAAGATTAACTCTTGTACCT
 **** * * * * * * * * * * * * * * *

1261 1270 1280 1290 1300 1310 1320

Fin Whale TTTGCATAATGACTTAACTAGTAATA-AATTAGCAAAGAGACCTTAAAGTTAAATTACCCG
 Blue Whale TTTGCATAATGACTTAACTAGTAATA-CTTAGCAAAGAGACCTTAAAGTTAAACTACCCG
 Cow TTTGCATAATGAATTAAGTATAAGACTTAACAAAATGAATTTTAGCTAAGCAGCCCG
 Rat TTTGCATAATGAATTAAGTATAAGACTTAACAAAATGAATTTTAGCTAAGCAGCCCG
 Mouse TTTGCATAATGAACTAAGTATAAGACTTAACAAAATGAATTTTAGCTAAGCAGCCCG
 Loach TTTGCATCATGATTTAGCCAGAACAC--CCAAGCAAAGAGACCTTTAGTTTGAACCCCG
 Carp TTTGCATCATGATTTAGCCAGTACAC--CCAAGCAAAGAGACCTTTAGTTTGAACCCCG
 Trout TTTGCATCATGATTTAGCCAGAACAC--CTGAGCAAAGAGAACCTTTAGTTTGAACCCCG
 Chicken TTTGCATCATGATTTAGCAAGAACA--CCAAGCAAAGTGAGCTAAAGTTTGCCTTCCCG
 Opossum TTTGCATAATGATTTAGCCAGTCAAC--ACGGACAAAAGAA-TTATGCCCGACATCCCG
 ***** ** * * * * * * * * * * * * * * *

1321 1330 1340 1350 1360 1370 1380

Fin Whale AAACC-AGACGAGCTACTTATGAGCAGC--ACCTA-GAACGAACCTCATCTATGTGGCAA
 Blue Whale AAACC-AGACGAGCTACTTATGAGCAGT--ACCTA-GGACGAACCTCATCTATGTGGCAA
 Cow AAACC-AGACGAGCTACTCACAAACAGTTTACCAA-GAACTAACTCATCTATGTGGCAA
 Rat AAACC-AAACGAGCTACCTAAAACAAT--TTCAT-GAATCAACCCGTCATGTAGCAA
 Mouse AAACC-AAACGAGCTACCTAAAACAAT--TTTAT-GAATCAACTCGTCATGTGGCAA
 Loach AAACC-AAGTGAGCTACCCCGGACCGCAAC-AT-GGGCCAACCCATCTCTGTGGCAA
 Carp AAACC-AGGTGAGCTACCCCGAGACAGCCTATTAT-GGCCCACCCCGTCCTGTGGCAA
 Trout AAACC-AGACGAGCTACTCCGGGACAGCCTATTGTAGGGCCAACCCGTCCTGTGGCAA
 Chicken AAACCAAGCGAGCTACTTGGGAGCAGCTAAAATTTGAGCGAACCCGTCCTGTGGCAA
 Opossum AAATT-AAGTGAGCTACTATAAGACAGT-TACTAATGAACCAACTCATCTATGTAGCAA
 *** * ***** * * * * * * * * * * * * * * *

1381 1390 1400 1410 1420 1430 1440

Fin Whale CAACGATTAA-AGTCCTACGTGATCTGAGTTCAGACCGGAGTAATCCAGGTCGGTTTCTA
 Blue Whale CAACGATTAA-AGTCCTACGTGATCTGAGTTCAGACCGGAGTAATCCAGGTCGGTTTCTA
 Cow CAACGATTAA-AGTCCTACGTGATCTGAGTTCAGACCGGAGTAATCCAGGTCGGTTTCTA
 Rat CAACGATTAA-AGTCCTACGTGATCTAAGT-----CCGG--CAATCCAGGTCGGTTTCTA
 Mouse CAACGATTAA-AGTCCTACGTGATCTGAGTTCAGACCGGAGCAATCCAGGTCGGTTTCTA
 Loach CAACGATTAA-AGTCCTACGTGATCTGAGTTCAGACCGGAGCAATCCAGGTCAGTTTCTA
 Carp CAACGATTAA-AGTCCTACGTGATCTGAGTTCAGACCGGAGCAATCCAGGTCAGTTTCTA
 Trout CAACGATTAA-AGTCCTACGTGATCTGAGTTCAGACCGGAGTAATCCAGGTCAGTTTCTA
 Chicken CAACGATTAAACAGTCCTACGTGATCTGAGTTCAGACCGGAGCAATCCAGGTCGGTTTCTA
 Opossum CAACGATTAA-AGTCCTACGTGATCTGAGTTCAGACCGGAGAAATCCAGGTCGGTTTCTA
 ***** *

2641 2650 2660 2670 2680 2690 2700

Fin Whale TCTATTA-CGCATTTCTCCAGT-ACGAAAGGACA-AGAGAAATAAGGCCAA--CTTCAA
 Blue Whale TCTATTA-CGCATTTCTCCTAGT-ACGAAAGGACA-AGAGAAATAAGGCCAA--CTTCAA
 Cow TCTATTA-CGTATTTCTCCAGT-ACGAAAGGACA-AGAGAAATAAGGCCAA--CTTTAA
 Rat TCTATTT-ACAATTTCTCCAGTTACGAAAGGACA-AGAGAAATGGAG--AC--CAACCA
 Mouse TCTATTT-ACGATTTCTCCAGT-ACGAAAGGACA-AGAGAAATAGAGCCAC--CTTAA
 Loach TCTGTAACGTTACTTTTCTAGT-ACGAAAGGAACCGGAAAAGAAGGGCCAACTAAA-
 Carp TCTGTAACGCTACTTTTCTAGT-ACGAAAGGATC-GGAAAAGAGGGGCCATACTTAA-
 Trout TCTATGAAGTGATGTTTCTAGT-ACGAAAGGACC-GGAAAAGAAGGGGCCATGCTTGA-
 Chicken TCTATG--GACACTCCTCCTAGT-ACGAAAGGACC-GGAGAAGTGGGGTCAATACCACTG
 Opossum TCTATATATTAATTTCTCCAGT-ACGAAAGGACC-AGAGAAATAAGGCCAA--CATT-
 *** * * *** * * * * *

2701 2710 2720 2730 2740 2750 2760

Fin Whale A-CAAGCGCCTTCAAAC-AATTAATGA-CCTAGTCTCAACTTAATAATTAAGCGCAAACA
 Blue Whale A-CAAGCGCCTTCAAAT-AATTAATGA-CCTAGTCTCAACTTAATAACCAAGCGCAAACA
 Cow ATCAAGCGCCTTAAGAC-AACCAATGA-TAACATCTCAACTGACAA-----CACAAA
 Rat ATCCTAGGCTTCCAACC-AATTTAG----AAAACTTAATAAAGTA--TATATGTACAAT
 Mouse AATAAGCGCTCTCAACTTAATTTATGAATAAAATCTAAATAAAATA--TATAGTACACC
 Loach AGTGCGCCCTACCC--CTAATTAATGA-AACCAACTAAATTAAGCAAAGGTAGAACCC-
 Carp AGCACGCCCCACCC--CTAATTTATGA-AAACAAATAAAATAAAAGGGAGAGCCAAA
 Trout GGCACGCCCCACCC--CCACCTGATGA-AGGCAACTAAAACAGACAAGGGGGCACACCAA
 Chicken AGCACACCCCCAACCTTCTAAGCAATGA-ATACAACCTCAACTGCCAAGAACCCCTCCCCCA
 Opossum TCTATGCGCCCTCATAA-AATTAATGAAATATATCTAAATTAACC-----ATTTAAACT
 * * * * * *

2671 2680 2690 2700 2710 2720 2730

Fin Whale AACCTGCCCA---AGACCAGGGCC
 Blue Whale AGTATGCCCA---AGACCAGGGCC
 Cow ACCCTGCCCT---AGAACAGGGCT
 Rat AAATAACCTT---AGACCCAAGTT
 Mouse CTCTAACCT---AGAGA-AGGTT
 Loach --CTCTACTG-CCAAAATAAGGCC
 Carp ATCCCAGCTGGCCAAAATAAGGAC
 Trout -----GATTGCCTAAAAGAACGGC
 Chicken CACCCGAACCTCCTAGAA-AAGGAT
 Opossum TTATCCACT--CTAGATAAGAGCC
 * * *

2731 2740 2750

APPENDIX B: Sequence alignment of mitochondrial 16S rRNA for members in the family Leuciscidae (*N.therinoides*, *Platygobio gracilis*, *Rhinichthys atratulus*, *Orthodon microlepidotus*, *Ptychocheilus oregonensis*, *S.bicolor*, *R.solitarius*, *P.erythrogaster*, *P.neogaeus*, *Notemigonus crysoleucas*, *S.copei*, and *Phenacobius mirabilis*, with a partial sequence for the northern pike (*Esox lucius*). Conserved base pairs through all species are indicated by *.

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N.therinoides      TGCAAATTGGGTCGCCCTGAGCCAACCAGCTAGCTTAATCACCAGTATAATTCAACAATA
P.gracilis        TGCAAATTGGATCACCCCTGAGCTAACCAGCTAGCTTAATCATCACTATAATTTAACAATA
R.atratulus       TGCAAGTTGGATCACCCCTGAGCTAACCAGCTAGCTTGACTACTAATATAACTAAACTTTA
O.microlepidotus1 TGCAAGTTGGATCACCCCTGAGCCAACAGCTAGCTTAATTACTAATATAACCCAACAATG
O.microlepidotus2 TGCAAGTTGGATCACCCCTGAGCCAACAGCTAGCTTAATTACTAATATAACCCAACAATG
P.oregonensis     TGCAAATTGGGTCGCCCTGAGCCAACAGCTAGCTTAATTACTAATATAATTCAACAATA
S.bicolor         TGCAAATTGGATCACCCCTGAGCCAACAGCTAGCCTAATTACTGATATAATTCAACAATA
R.solitarius      TGCAAATTGGGTCGCCCTGAGCCAACAGCTAGCTTAATTACTAATATAATTCAACAATA
P.erythrogaster   TGCAAATTGGATTACCCCTGAGCCAACAGCTAGCTTAGCCATCAATATAATTCAACAATG
P.neogaeus        TGCAAGTTGGATTACCCCTGAGCCAACAGCTAGCTTAACTACCAATATAATTCAACAATG
N.crysoleucas     TGCAAATTGGATCACCCCTGAGCCAACCAGCTAGCTTGATTATTAATATAATTTAACAATA
S.copei          TGCAAATTGGATCACCCCTGAGCCGACCAGCTAGCCTGATTATTAATATAATTTAACAATA
P.mirabilis       TGCAAATTGGATCACCCCTGAGCCAACCAGCTAGCTTAACTAAAAATATAATGTAGCACTG
Esox lucius       -----
*****  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *
1         10        20        30        40        50        60

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N.therinoides      TTTATAACAAAACAAGACCTAACCCCTACAAACTAAACCATTTTTTTTACCTGAGTATGGGA
P.gracilis        TTTATAACAACACAAGACTTAACCCACAAACTAAACCATTTTTTTTACCTGAGTATGGGA
R.atratulus       TTCATAACAAAACAAAACCCAATGCCACAAACTAAACCATTTTTTTTACCTAAGTATGGGA
O.microlepidotus1 TTCATAACAAAACATGACCTGACACCATAAACTAAACCATTTTTTTTACCTAAGTACGGGA
O.microlepidotus2 TTCATAACAAAACATGACCTGACACCATAAACTAAACCATTTTTTTTACCTAAGTACGGGA
P.oregonensis     TTTATAACAAAACATGGCCTAACACCATAAACTAAACCATTTTTTTTACCTAAGTACGGGA
S.bicolor         TTCATAACAAAACATGGCCTAACACCATAAACTAAACCATTTTTTTTACCTGAGTACGGGA
R.solitarius      TTTATAACAAAACATAACCTAACACCATAAACTAAACCATTTTTTTTACCTAAGTACGGGA
P.erythrogaster   TTCATAACAAAACATGGCCCAATATTATAAACTAAACCATTTTTTTTACCTAAGTACGGGA
P.neogaeus        TTCATAACAAAACATGACCTAATATTATAAATTAACCATTTTTTTTACCTAAGTACGGGA
N.crysoleucas     TTCATAACAAAACATGGCTTAACATTACAAACTAAACCATTTTTTTTGCCTAAGTACGGGA
S.copei          TTTATAACAAAACACGGCCTAACGCCACAAATTAACCATTTTTTTTACCTGAGTATGGGA
P.mirabilis       TTTATAACCAACCAAGACCTAACCCACCAACTAAACCATTTTTTTTACCTAAGTATGGGA
Esox lucius       -----
**  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *
61       70       80       90       100      110      120

```


N. therinoides CTACCCCGAGACAGCCTATATAATT-TAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
P. gracilis CTACCCCGAGACAGCCTATGTAATT-TAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
R. atratulus CTACCCCGAGACAGCCTATGTAATT-TAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
O. microlepidotus1 CTACCCCGAGACAGCCTAAACTAAT-TAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
O. microlepidotus2 CTACCCCGAGACAGCCTAAACTAAT-TAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
P. oregonensis CTACCCCGAGACAGCCTATATTATT-TAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
S. bicolor CTACCCCGAGACAGCCTATATTATT-TAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
R. solitarius CTACCCCGAGACAGCCTATATTATT-TAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
P. erythrogaster CTACCCCGAGACAGCCTATATTATT-TAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
P. neogaeus CTACCCCGAGACAGCCTATATTATT-TAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
N. crysoleucas CTACCCCGAGACAGCCTATGTTAATCTAGGGCCAACCCGTCTCTGTGGCAAAGAGTGGG
S. copei CTACCCCGAGACAGCCTATATTAATTTAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
P. mirabilis CTACCCCGAGACAGCCTATAT-ATT-TAGGGCTAACCCGTCTCTGTGGCAAAGAGTGGG
Esox lucius -----
 ***** * * *****
 301 310 320 330 340 350 360

N. therinoides AAGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTGCCTAAGAA
P. gracilis AAGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTGCCTGGGAA
R. atratulus AAGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTGCCTGAGAA
O. microlepidotus1 AAGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTCTCTGAGAA
O. microlepidotus2 AAGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTCTCTGAGAA
P. oregonensis AAGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTATCTGAGAA
S. bicolor AAGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTGTCTGAGAA
R. solitarius AAGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTGTCTGAGAA
P. erythrogaster AGGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTATCTGAGAA
P. neogaeus AAGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTATCTGAGAA
N. crysoleucas AAGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTGCCTGAGAA
S. copei AAGAGCTCCGGGTAGAAGTGACAGACCTACCGAACCTGGTGATAGCTGGTTGCCTGAGAA
P. mirabilis ATGAGCTCCGGGTAGAAGTGATAAACTTACCGAACCTGGTGATAGCTGGTTGCCTAAGAA
Esox lucius -----
 ***** * * ***** * * *
 361 370 380 390 400 410 420

N. therinoides ATGAATAGAAGTTCAGCCTCGCACGCCCT-TAATCAAGAAATACATCATCAAGATA-T-T
P. gracilis ATGAATAGAAGTTCAGCCTCGCACACCCT-TAATCAAGAAACATACCGTCAAGACA-T-T
R. atratulus ATGGATAGAAGTTCAGCCTCGCACCCCC-GAATCAAAAAGAAGTATCACCAGATGCT-T
O. microlepidotus1 GTGGATAGAAGTTCAGCCTCATACGCCCC-AAATCAACATGTATATTATTAAGATAACC-A
O. microlepidotus2 GTGGATAGAAGTTCAGCCTCATACKCCCC-AAATCAACATGTATATTATTAAGATAACC-A
P. oregonensis GTGGATAGAAGTTCAGCCTCATACGCCCC-AGATCAATACGTATGTTATTAAGATACTCA
S. bicolor GTGGATAGAAGTTCAGCCTCATTACCCCC-AAATCAAGA-----TTATTAAGATACT-A
R. solitarius GTGGATAGAAGTTCAGCCTCATACCCCC-AAGTCAACA--TATATTATTAAGATGTT-C
P. erythrogaster GTGGATAGAAGTTCAGCCTCATGTACCCC-AAGTCAAGGAAGATATTATTAAGATACT-A
P. neogaeus CTGGATAGAAGTTCAGCCTCATACCCCCAAATCAAGGAATATATTATGAAGATATT-A
N. crysoleucas ATGGATAGAAGTTCAGCCTCGTATTACCCCCAAACCAAGAACATATCACTAAGGTAAT-T
S. copei GTGGATAGAAGTTCAGCCTTATGCCCCC-GGATCAAGAATATATTATTAAGATACT-C
P. mirabilis ATGAATAGAAGTTCAGCCTCGTCCACCCT-TTGTCAAGAAATATACTACTAAGACACC--
Esox lucius -----
 ** ***** ** ** ***
 421 430 440 450 460 470 480

N. therinoides ATGGATATATACGAGAGTTAGTTGAAGGGGGTACAGCCCCTTTAACAAAGGATACAACCT
P. gracilis ATGGAAAATACGAGAGTTAGTTGAAGGGGGTACAGCCCCTTTAACAAAGGATACAACCT
R. atratulus AAGGGAAAGTGCAGAGAGTTAGTTGAAGGGGGTACAGCCCCTTCAACAAAGGATACAACCT
O. microlepidotus1 GGAGAAATATATGAGAGTTAGTTAAAAGGGGTACAGCCCCTTTAACAAAGGATACAACCT
O. microlepidotus2 GGAGAAATATATGAGAGTTAGTTAAAAGGGGTACAGCCCCTTTAACAAAGGATACAACCT
P. oregonensis AGGGAGATACATGAGAGTTAGTTGAAAGGGGTACAGCCCCTTTAACAAAGGATACAACCT
S. bicolor TGGGAAATATATGAGAGTTAGTTGAAAGGGGTACAGCCCCTTTAACAAAGGATACAACCT
R. solitarius AGGGAAATATATGAGAGTTAGTTGAAAGGGGTACAGCCCCTTTAACAAAGGATACAACCT
P. erythrogaster AGGGAAACATATGAGAGTTAGTTAAAAGGGGTACAGCCCCTTTAACAAAGGATACAACCT
P. neogaeus AGGGCGACATATGAGAGTTAGTTAAAAGGGGTACAGCCCCTTTGACAAAGGATACAACCT
N. crysoleucas AGAGATACATACGAGAGTTAGTTAAAGGGGGTACAGCCCCTTTAACAAAGGATACAACCT
S. copei TGGGCGATACGTGAGAGTTAGTTGAAGGGGGTACAGCCCCTTTAACAAAGGATACAACCT
P. mirabilis TTGGAAATACACGAGAGTTAGTTTAAAGGGGGTACAGCCCCTTTAACAAAGGATACAACCT
Esox lucius -----
* * ***** ** ***** ** ***** *****
481 490 500 510 520 530 540

N. therinoides TA-ACAGGAGGATAAAGATCATAATTAATAAAAATATACTGTTCTAGTGGGCTGAAAGCA
P. gracilis TT-ACAGGAGGATAAAGATCATAATATATAAAAATATACTGTTCTAGTGGGCTGAAAGCA
R. atratulus TC-CCAGGAGGATAAAGATCATAATACATAAGACACACTGTTCTAGTGGGCTGAAAGCA
O. microlepidotus1 TC-ACAGGAGGATAAAGATCATAATATATAAAAACATACTGTTTTAGTGGGCTAAAAGCA
O. microlepidotus2 TC-ACAGGAGGATAAAGATCATAATATATAAAAACATACTGTTTTAGTGGGCTAAAAGCA
P. oregonensis TT-ACAGGAGGATAAAGATCATAATATATAAAAACATACTGTTTTAGTGGGCTAAAAGCA
S. bicolor TTTACAGGAGGATAAAGATCATAATATATAAAAACATACTGTTTTAGTGGGCTAAAAGCA
R. solitarius TT-ACAGGAGGATAAAGATCATAATATATAAAAACATACTGTTTTAGTGGGCTAAAAGCA
P. erythrogaster TCTACAGGAGGATAAAGATCATAATATATAAAAATATACTGTTTTAGTGGGCTGAAAGCA
P. neogaeus TC-ACAGGAGGATAAAGATCATAATATATAAAAATATACTGTTTTAGTGGGCTGAAAGCA
N. crysoleucas TT-ACAGGAGGATAAAGATCATAATATATAAAAACATACTGTTCTAGTGGGCTAAAAGCA
S. copei TC-ACAGGAGGATAAAGATCATAATATACAAGACATACTGTTCTAGTGGGCTGAAAGCA
P. mirabilis TC-ACAGGAGGATAAAGATCATAATTAATAAAAACCTACTGTTCTAGTGGGCTGAAAGCA
Esox lucius -----
* ***** * * * ***** *****
541 550 560 570 580 590 600

N. therinoides GCCATCTAAACAGAAAGCGTTAAAGCTCGGACAGAACGAAGTTTATTATACCGATAA-AA
P. gracilis GCCACCTAAACAGAAAGCGTTAAAGCTCGGACAGAAAGGAAGTTTATTATACCGATAA-AA
R. atratulus GCCATCTAAGCAGAAAGCGTTAAAGCTCAGACAGAAAGTAAATTTATTATACCGATAA-AC
O. microlepidotus1 GCCATCTAAATAGAAAGCGTTAAAGCTCAGACAGAAAGAAGTTTATTATACCGATAG-AA
O. microlepidotus2 GCCATCTAAATAGAAAGCGTTAAAGCTCAGACAGAAAGAAGTTTATTATACCGATAG-AA
P. oregonensis GCCACCTAAATAGAAAGCGTTAAAGCTCAGACAGAAAGAAGTTTATTATACCGATAA-AA
S. bicolor GCCATCTAAATAGAAAGCGTTAAAGCTCAGACAGAAAGAAGTTTATTATACCGATAA-AA
R. solitarius GCCACCTAGATAGAAAGCGTTAAAGCTCAGGACAGAAAGGAAGTTTATTATACCGATAAGAA
P. erythrogaster GCCACCTAATTAGAAAGCGTTAAAGCTCGGACAGACGGAAGTTTATTATACCGATAA-AA
P. neogaeus GCCACCTAAATAGAAAGCGTTAAAGCTCGGACAGAAAGAAGTTTATTATACCGATAA-AA
N. crysoleucas GCCACCTAAACAGAAAGCGTTAAAGCTCAGACAGAAAAAGTTTATTATACCGATAA-AA
S. copei GCCACCTAAGCAGAAAGCGTTAAAGCTCAGACAGAGAGAAGTTTATTATACCGATAA-GT
P. mirabilis GCCACCTAATTAGAAAGCGTTAAAGCTCAGACAGGAAAAAGTTTATTATACCAATAAAAA
Esox lucius -----
***** * * * ***** * * * *
601 610 620 630 640 650 660

START PRIMER →

N. therinoides GCCGCGGTATATTGACCGTGCAAAGGTAGCGCAATCACTTGTCTTTTAAATAGAGACCTG
P. gracilis GCCGCGGTATATTGACCGTGCAAAGGTAGCGCAATCACTTGTCTTTTAAATAGAGACCTG
R. atratulus GCCGCGGTATTTTGGACCGTGCAAAGGTAGCGCAATCACTTGTCTCTTAAATAGAGACCTG
O. microlepidotus1 GCCGCGGTATTTTGGACCGTGCAAAGGTAGCGCAATCACTTGTCTCTTAAATAGAGACCTG
O. microlepidotus2 GCCGCGGTATTTTGGACCGTGCAAAGGTAGCGCAATCACTTGTCTCTTAAATAGAGACCTG
P. oregonensis GCCGCGGTATTTTGGACCGTGCAAAGGTAGCGCAATCACTTGTCTCTTAAATAGAGACCTG
S. bicolor GCCGCGGTATTTTGGACCGTGCAAAGGTAGCGCAATCACTTGTCTCTTAAATAGAGACCTG
R. solitarius GCCGCGGTATTTTGGACCGTGCAAAGGTAGCGCAATCACTTGTCTCTTAAATAGAGACCTG
P. erythrogaster GCCGCGGTATTTTGGACCGTGCAAAGGTAGCGCAATCACTTGTCTCTTAAATAGAGACCTG
P. neogaeus GCCGCGGTATTTTGGACCGTGCAAAGGTAGCGCAATCACTTGTCTCTTAAATAGAGACCTG
N. crysoleucas GCCGCGGTATTTTGGACCGTGCAAAGGTAGCGCAATCACTTGTCTCTTAAATAGAGACCTG
S. copei GCCGCGGTATTTTGGACCGTGCAAAGGTAGCGCAATCACTTGTCTCTTAAATAGAGACCTG
P. mirabilis GCCGCGGTATATTGACCGTGCAAAGGTAGCGCAATCACTTGTCTTTTAAATAGAGACCTG
Esox lucius GCCGCGGTATTTTAAACCGTGCGAAGGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTG
 ***** ** ***** ***** ***** ***** ***** *****
 1021 1030 1040 1050 1060 1070 1080

N. therinoides TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
P. gracilis TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
R. atratulus TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
O. microlepidotus1 TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
O. microlepidotus2 TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
P. oregonensis TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
S. bicolor TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
R. solitarius TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
P. erythrogaster TATGAACGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
P. neogaeus TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
N. crysoleucas TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
S. copei TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTAT
P. mirabilis TATGAATGGCTAGACGAGGGCTTAACTGTCTCCCCCTCCAGTCAGTGAAATTGATCTGC
Esox lucius TATGAATGGCATCACGAGGGCTTAACTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGC
 ***** *** ***** ***** * ***** ***** *****
 1081 1090 1100 1110 1120 1130 1140

N. therinoides CCGTGCAGAAAGCGGGTATAGTACTACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAG
P. gracilis CCGTGCAGAAAGCGGGTATAACAGTACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAA
R. atratulus CCGTGCAGAAAGCGGGTATAATAGTACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAA
O. microlepidotus1 CCGTGCAGAAAGCGGGTATGACCATACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAA
O. microlepidotus2 CCGTGCAGAAAGCGGGTATGACCATACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAA
P. oregonensis CCGTGCAGAAAGCGGGTATGACTATACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAA
S. bicolor CCGTGCAGAAAGCGGGTATGACTCTACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAA
R. solitarius CCGTGCAGAAAGCGGGTATGACCATACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAA
P. erythrogaster CCGTGCAGAAAGCGGGTATAACTATACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAA
P. neogaeus CCGTGCAGAAAGCGGGTATAATTATACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAA
N. crysoleucas CCGTGCAGAAAGCGGGTTAACTATACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAA
S. copei CCGTGCAGAAAGCGGGTATAACTATACAAAGACGAGAAAGACCCTTTGGAGCTTAAAGGTACAA
P. mirabilis CCGTGCAGAAAGCGGGTATAACTATACAAAGACGAGAAAGACCCTTATGGAGCTTAAAGGTACAA
Esox lucius CCGTGCAGAAAGCGGGACATAAAGACATAAGACGAGAAAGACCCTTATGGAGCTTAAAGACACCC
 ***** * * ***** ***** ** **
 1141 1150 1160 1170 1180 1190 1200

← STOP PRIMER

<i>N. therinoides</i>	TAG- AAAGCCGATCAACGG ACCAAGTTACCCTAGGGATAACAGCGCAATCCTCTCCCAGA
<i>P. gracilis</i>	TAATA AAAGCCGATCAACGA ACCAAGTTACCCTAGGGATAACAGCGCAATCCTCTCCCAGA
<i>R. atratulus</i>	TAA- AAAGTCGATCAACGA ACCAAGTTACCCTAGGGATAACAGCGCAATCCTCTCCCAGA
<i>O. microlepidotus1</i>	CAC AAAGAGTCGATCAACGG ACCAAGTTACCCTAGGGATAACAGCGCAATCCCCTCCCAGA
<i>O. microlepidotus2</i>	CAC AAAGAGTCGATCAACGG ACCAAGTTACCCTAGGGATAACAGCGCAATCCCCTCCCAGA
<i>P. oregonensis</i>	CAC AGTGATCGATCAACGA ACCAAGTTACCCTAGGGATAACAGCGCAATCCCCTCCCAGA
<i>S. bicolor</i>	CAC AGGGGTCGATCAACGA ACCAAGTTACCCTAGGGATAACAGCGCAATCCCCTCCCAGA
<i>R. solitarius</i>	CT TAGCAGTCGATCAACGG ACCAAGTTACCCTAGGGATAACAGCGCAATCCCCTCCCAGA
<i>P. erythrogaster</i>	--- AAAAGTCGATCAACGA ACCAAGTTACCCTAGGGATAACAGCGCAATCCCCTCCCAGA
<i>P. neogaeus</i>	--- AGAAGTCGATCAACGG ACCAAGTTACCCTAGGGATAACAGCGCAATCCCCTCCCAGA
<i>N. crysoleucas</i>	- TGAAAAGGC-ATCAACGA ACCAAGTTACCCTAGGGATAACAGCGCAATCCTCTCCCAGA
<i>S. copei</i>	C-- GGAAGCCGATCAACGG ACCAAGTTACCCTAGGGATAACAGCGCAATCCTCTCCCAGA
<i>P. mirabilis</i>	T- CTAGGGCCGATCAACGA ACCAAGTTACCCTAGGGATAACAGCGCAATCCTCTCCCAGA
<i>Esox lucius</i>	--- ACCAGTCGATCAACGG ACCAAGTTACCCTAGGGATAACAGCGCAATCCCCTCCCAGA

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1381	1390	1400	1410	1420	1430	1440
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<i>N. therinoides</i>	GCCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTGGTGGTGCA
<i>P. gracilis</i>	GCCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTGGTGGTGCA
<i>R. atratulus</i>	GCCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA
<i>O. microlepidotus1</i>	GTCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA
<i>O. microlepidotus2</i>	GTCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA
<i>P. oregonensis</i>	GTCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA
<i>S. bicolor</i>	GTCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA
<i>R. solitarius</i>	GTCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA
<i>P. erythrogaster</i>	GGCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA
<i>P. neogaeus</i>	GGCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA
<i>N. crysoleucas</i>	GTCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA
<i>S. copei</i>	GCCCGTATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA
<i>P. mirabilis</i>	GCCCATATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA
<i>Esox lucius</i>	GTCCCTATCGACGAGGGGGTTTACGACCTCGATGTTGGATCAGGACATCCTAATGGTGGCA

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1441	1450	1460	1470	1480	1490	1500
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<i>N. therinoides</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>P. gracilis</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>R. atratulus</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>O. microlepidotus1</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>O. microlepidotus2</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>P. oregonensis</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>S. bicolor</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>R. solitarius</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>P. erythrogaster</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>P. neogaeus</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>N. crysoleucas</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>S. copei</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>P. mirabilis</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTAAAGTCCTACGTGATCTGAGTTCAGACC
<i>Esox lucius</i>	GCCGCTATTAAGGGTTCGTTTGTTCACGATTA-----

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1501	1510	1520	1530	1540	1550	1560
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N. therinoides GGAGCAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCAGTACGAAAGGATCGGA
P. gracilis GGAGCAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCTAGTACGAAAGGATCGGA
R. atratulus GGAGCAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCAGTACGAAAGGATCGGA
O. microlepidotus1 GGAGCAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCTAGTACGAAAGGATTGGA
O. microlepidotus2 GGAGCAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCTAGTACGAAAGGATTGGA
P. oregonensis GGAGTAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCTAGTACGAAAGGATTGGA
S. bicolor GGAGTAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCTAGTACGAAAGGATTGGA
R. solitarius GGAGTAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCTAGTACGAAAGGATTGGA
P. erythrogaster GGAGTAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCTAGTACGAAAGGATTGGA
P. neogaeus GGAGTAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCTAGTACGAAAGGATTGGA
N. crysoleucas GGAGCAATCCAGGTCAGTTTCTATCTGTAGCGCTACTTTTCCCTAGTACGAAAGGATCGGA
S. copei GGAGTAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCTAGTACGAAAGGATCGGA
P. mirabilis GGAGTAATCCAGGTCAGTTTCTATCTGTAACGCTACTTTTCCCTAGTACGAAAGGATCGGA
Esox lucius -----

 1561 1570 1580 1590 1600 1610 1620

N. therinoides AAAGAGGGGGCC TACTTAACGCATGCCCCACCCCTAATTCATGAAAACAAATAAATCAA
P. gracilis AAAGAGGGGGCC CATACTTAGAGCATGCCCCACCCCTAATTCATGAAAACAAATAAATCAA
R. atratulus AAAGAGGGGGCC TATGCTTAAGGCATGCCCCGCCCCAATTGATGAAAACAAATAAATCAA
O. microlepidotus1 AAAGAGGGGGCC TATACCTCAGGCACGCCCCGCCCCAATTAATGAAAACAAATAAATCAA
O. microlepidotus2 AAAGAGGGGGCC TATACCTCAGGCACGCCCCGCCCCAATTAATGAAAACAAATAAATCAA
P. oregonensis AAAGAGGGGGCC CATACTTAAGGCACGCCCCGCCCCAATTGATGAAAACAAATAAATCAA
S. bicolor AAAGAGGGGGCC CATACTTAAGGCACGCCCCGCCCCAATTAATGAAAACAAATAAATCAA
R. solitarius AAAGAGGGGGCC CATACTTAAGGCACGCCCCGCCCCAATTAATGAAAACAAATAAATCAA
P. erythrogaster AAAGAGGGGGCC CATACTTAGAGCATGCCCCGCCCCAATTAATGAAAACAAATAAATGGA
P. neogaeus AAAGAGGGGGCC CATACTTAAAGCAGCCCCGCCCCAATTAATGAAAACAAATAAATGGA
N. crysoleucas AAAGAGGGGGCC TACTTTAGGCATGCCCCACCCCTAATTGATGAAAACAAATAAATCAA
S. copei AAAGAGGGGGCC TATGCTCAAAGCATGCCCCGCCCCAATTGATGAGGACAAATAAATCAA
P. mirabilis AAAGAGGGGGCC TATGCTTAAAGCAGCCCCACCCCTAATTTATGAAACAAATAAATCAA
Esox lucius -----
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 1621 1630 1640 1650 1660 1670 1680

N. therinoides AT-AAGGGAGGGCCAAAACCCCTGCCGCCCAAGATAAGGGCA
P. gracilis AT-AAGGGAGGGCCAGAACCCCTGCCGCCCAAGTTAAGGGCA
R. atratulus GTGAAGGGAGGGCCAAAACCCCTACCGCCCGATAGAAGGGCA
O. microlepidotus1 GTAAAGGGAGGGCCAAAACCCCTGCCGTTCAAGATAAAGGACA
O. microlepidotus2 GTAAAGGGAGGGCCAAAACCCCTGCCGTTCAAGATAAAGGACA
P. oregonensis GTAAAGGGAGGGCCAAAACCCCTGCCGTTCAAGATAAAGGACA
S. bicolor GTAAAGGGAGGGCCAAAACCCCTGCCGTTCAAGATAAAGGACA
R. solitarius GTAAAGGGAGGGCTAAAACCCCTACCGCTCAAGATAAAGGGAA
P. erythrogaster GTAAAGGGAGGGCC TAAACCCCGCCGCCCAAGAGAAGGGCA
P. neogaeus GTAAAGGGGTGGGCCAAAACCCCGCCGCCCAAGAGAAGGGCA
N. crysoleucas GTAAAGGGAGGGCTAAAACCCCTACCGTCCGAAATAAAGGACA
S. copei GCAAAGGGAGGGCC-AAAGCCCTACCGTCCGAAATAAAGGACA
P. mirabilis AC-AAGGGCGGGCC--AAACCCTGCCGCCCAAAATAAAGGGCA
Esox lucius -----
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 1681 1690 1700 1710 1720

APPENDIX C: The sequenced area of the 16S rDNA from the silver pike, northern pike, and muskellunge.

Silver Pike 1	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAATGGCATCACGAGGGCTTA
Silver Pike 2	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAANGGCATCACGAGGGCTTA
Silver Pike 3	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAATGGCATCACGAGGGCTTA
Silver Pike 4	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAATGGCATCACGAGGGCTTA
Silver Pike 5	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAATGGCATCACGAGGGCTTA
Northern Pike 2	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAATGGCATCACGAGGGCTTA
Northern Pike 3	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAATGGCATCACGAGGGCTTA
Northern Pike 5	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAATGGCATCACGAGGGCTTA
Muskellunge 1	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAATGGCATCACGAGGGCTTA
Muskellunge 2	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAATGGCATCACGAGGGCTTA
Muskellunge 4	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAATGGCATCACGAGGGCTTA
Muskellunge 5	GGTAGCGCAATCACTTGTCTTTTAAATGAAGACCTGTATGAATGGCATCACGAGGGCTTA

1 10 20 30 40 50 60

Silver Pike 1	GCTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGAAC
Silver Pike 2	GCTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGAAC
Silver Pike 3	GCTGTCTCCTCTNTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGAAC
Silver Pike 4	GCTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGANC
Silver Pike 5	GCTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGAAC
Northern Pike 2	GCTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGAAC
Northern Pike 3	GCTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGAAC
Northern Pike 5	GCTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGAAC
Muskellunge 1	GCTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGAAC
Muskellunge 2	GCTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGAAC
Muskellunge 4	GCTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGAAC
Muskellunge 5	GCTGTCTCCTCTTTCAAGTCAATGAAATTGATCTGCCCGTGCAGAAGCGGACATAAGAAC

61 70 80 90 100 110 120

Silver Pike 1	ATAAGACGAGAAGACCCTATGGAGCTTTAGACACCCGGCAGACCCTGTTAAGTAGCTGAA
Silver Pike 2	ATAAGACGAGAAGACCCTATGGAGCTTTAGACACCCGGCAGACCCTGTTAAGTAGCTGAA
Silver Pike 3	ATAAGACGAGAAGACCCTATGGAGCTTTAGACACCCGGGAGACCCTGTTAAGTAGCTGAA
Silver Pike 4	ATAAGACGAGAAGACCCTATGGAGCTTTAGACACCCGGCAGACCCTGTTAAGTAGCTGAA
Silver Pike 5	ATAAGACGAGAAGACCCTATGGAGCTTTAGACACCCGGCAGACCCTGTTAAGTAGCTGAA
Northern Pike 2	ATAAGACGAGAAGACCCTATGGAGCTTTAGACACCCGGCAGACCCTGTTAAGTAGCTGAA
Northern Pike 3	ATAAGACGAGAAGACCCTATGGGGCTTTAGACACCCGGCAGACCCTGTTAAGTAGCTGAA
Northern Pike 5	ATAAGACGAGAAGACCCTATGGAGCTTTAGACACCCGGCAGACCCTGTTAAGTAGCTGAA
Muskellunge 1	ATAAGACGAGAAGACCCTATGGAGCTTTAGACACCCGGCAGACCATGTCAAGTAACCTG
Muskellunge 2	ATAAGACGAGAAGACCCTATGGAGCTTTAGACACCCGGCAGACCATGTCAAGTAACCTG
Muskellunge 4	ATAAGACGAGAAGACCCTATGGAGCTTTAGACACCCGGCAGACCATGTCAAGTAACCTG
Muskellunge 5	ATAAGACGAGAAGACCCTATGGAGCTTTAGACACCCGGCAGACCATGTCAAGTAACCTG

121 130 140 150 160 170 180

Silver Pike 1	CTATCAGATTAAAACAAAGCGGCCCTGGCCTACATGTCTTCGGTTGGGGCGACCACGGG
Silver Pike 2	CTATCAGATTAAAACAAAGCGGCCCTGGCCTACATGTCTTCGGTTGGGGCGACCACGGG
Silver Pike 3	CTATCAGATTAAAACAAAGCGGCCCTGGCCTACATGTCTTCGGTTGGGGCGACCACGGG
Silver Pike 4	CTATCAGATTAAAACAAAGCGGCCCTGGCCTACATGTCTTCGGTTGGGGCGACCACGGG
Silver Pike 5	CTATCAGATTAAAACAAAGCGGCCCTGGCCTACATGTCTTCGGTTGGGGCGACCACGGG
Northern Pike 2	CTATCAGATTAAAACAAAGCGGCCCTGGCCTACATGTCTTCGGTTGGGGCGACCACGGG
Northern Pike 3	CTATCAGATTAAAACAAAGCGGCCCTGGCCTACATGTCTTCGGTTGGGGCGACCACGGG
Northern Pike 5	CTATCAGATTAAAACAAAGCGGCCCTGGCCTACATGTCTTCGGTTGGGGCGACCACGGG
Muskellunge 1	TTATTGGATTAAAACAAAGCGGCTCCTGGCCACATGTCTTCGGTTGGGGCGACCACGGG
Muskellunge 2	TTATTGGATTAAAACAAAGCGGCTCCTGGCCACATGTCTTCGGTTGGGGCGACCACGGG
Muskellunge 4	TTATTGGATTAAAACAAAGCGGCTCCTGGCCACATGTCTTCGGTTGGGGCGACCACGGG
Muskellunge 5	TTATTGGATTAAAACAGAACGGCTCCTGGCCACATGTCTTCGGTTGGGGCGACCACGGG

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181 190 200 210 220 230 240

Silver Pike 1 GGAAAACAAAGCCCCACGAGGATTAAGGAAAACCTCCTTATAACCACGAGCGACAGCTC
Silver Pike 2 GGAAAACAAAGCCCCACGAGGATTAAGGAAAACCTCCTTATAACCACGAGCGACAGCTC
Silver Pike 3 GGAAAACAAAGCCCCACGAGGATTAAGGAAAACCTCCTTATAACCACGAGCGACAGCTC
Silver Pike 4 GGAAAACAAAGCCCCACGAGGATTAAGGAAAACCTCCTTATAACCACGAGCGACAGCTC
Silver Pike 5 GGAAAACAAAGCCCCACGAGGATTAAGGAAAACCTCCTTATAACCACGAGCGACAGCTC
Northern Pike 2 GGAAAACAAAGCCCCACGAGGATTAAGGAAAACCTCCTTATAACCACGAGCGACAGCTC
Northern Pike 3 GGAAAACAAAGCCCCACGAGGATTAAGGAAAACCTCCTTATAACCACGAGCGACAGCTC
Northern Pike 5 GGAAAACAAAGCCCCACGAGGATTAAGGAAAACCTCCTTATAACCACGAGCGACAGCTC
Muskellunge 1 GGAAAATATAGCCCCATGCGGACTAAGGGTAACACCCTTATAACCATGAACTACAGCTC
Muskellunge 2 GGAAAATATAGCCCCATGCGGACTAAGGGTAACACCCTTATAACCATGAACTACAGCTC
Muskellunge 4 GGAAAATATAGCCCCATGCGGACTAAGGGTAACACCCTTATAACCATGAACTACAGCTC
Muskellunge 5 GGAAAATATAGCCCCATGCGGACTAAGGGTAACACCCTTATAACCATGAACTACAGCTC
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241 250 260 270 280 290 300

Silver Pike 1 TAAGTCTCAGAACTTCTGACCAAAAAGATCCGACACCAGTCGATCAACGGACCAAGTTAC
Silver Pike 2 TAAGTCTCAGAACTTCTGACCAAAAAGATCCGACACCAGTCGATCAACGGACCAAGTTAC
Silver Pike 3 TAAGTCTCAGAACTTCTGACCAAAAAGATCCGACACCAGTCGATCAACGGACCAAGTTAC
Silver Pike 4 TAAGTCTCAGAACTTCTGACCAAAAAGATCCGACACCAGTCGATCAACGGACCAAGTTAC
Silver Pike 5 TAAGTCTCAGAACTTCTGACCAAAAAGATCCGACACCAGTCGATCAACGGACCAAGTTAC
Northern Pike 2 TAAGTCTCAGAACTTCTGACCAAAAAGATCCGACACCAGTCGATCAACGGACCAAGTTAC
Northern Pike 3 TAAGTCTCAGAACTTCTGACCAAAAAGATCCGACACCAGTCGATCAACGGACCAAGTTAC
Northern Pike 5 TAAGTCTCAGAACTTCTGACCAAAAAGATCCGACACCAGTCGATCAACGGACCAAGTTAC
Muskellunge 1 TAAGTCTCAGAAATTCTGACCAAAAAGATCCGACACCCGTCGATCAACGGACCAAGTTAC
Muskellunge 2 TAAGTCTCAGAAATTCTGACCAAAAAGATCCGACACCCGTCGATCAACGGACCAAGTTAC
Muskellunge 4 TAAGTCTCAGAAATTCTGACCAAAAAGATCCGACACCCGTCGATCAACGGACCAAGTTAC
Muskellunge 5 TAAGTCTCAGAAATTCTGACCAAAAAGATCCGACACCCGTCGATCAACGGACCAAGTTAC
***** ***** ***** ***** ***** ***** ***** ***** *****

301 310 320 330 340 350 360

Silver Pike 1 CCTAGGGATA
Silver Pike 2 CCTAGGGATA
Silver Pike 3 CCTAGGGATA
Silver Pike 4 CCTAGGGATA
Silver Pike 5 CCTAGGGATA
Northern Pike 2 CCTAGGGATA
Northern Pike 3 CCTAGGGATA
Northern Pike 5 CCTAGGGATA
Muskellunge 1 CCTAGGGATA
Muskellunge 2 CCTAGGGATA
Muskellunge 4 CCTAGGGATA
Muskellunge 5 CCTAGGGATA

361 370